

Supporting Information

***Trypanosoma cruzi* Malic Enzyme is the target for sulfonamide hits from GSK Chagas Box**

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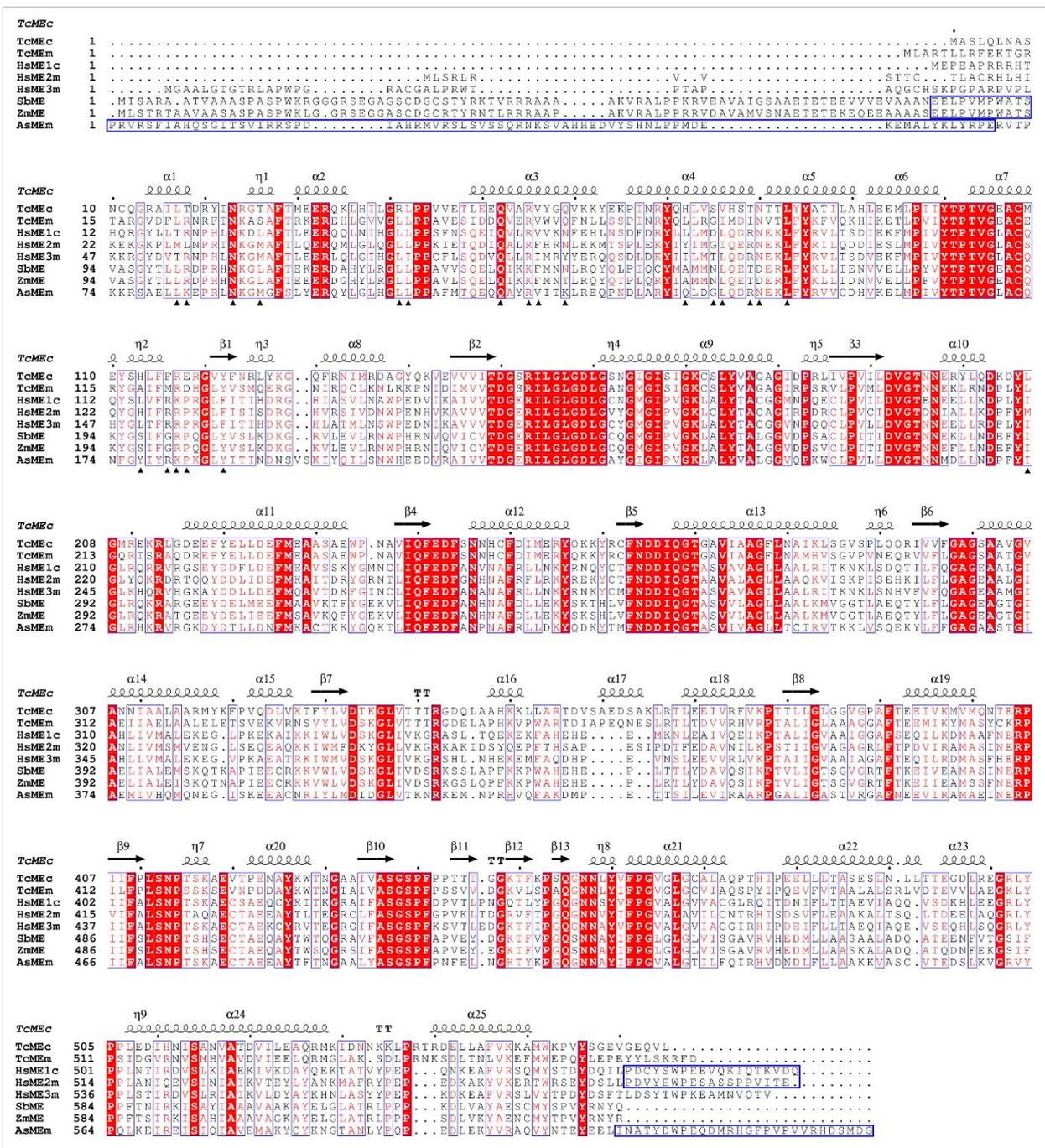


Figure S1. Alignment of amino acid sequences of eukaryotic malic enzymes (ME).

Residues of the cytosolic ME from *Trypanosoma cruzi* that delimit the allosteric inhibition site are indicated (▲). N- or C-terminal extensions known to stabilize the tetrameric structure in eukaryotic enzymes are highlighted (blue box). **Abbreviations:** *TcMEc*, cytosolic *T. cruzi* ME (UniProt Q4DJ68); *TcMEem*, mitochondrial *T. cruzi* ME (UniProt Q4DJ69); *HsME1c*, cytosolic human ME1 (UniProt P48163); *HsME2m*, mitochondrial human ME2 (UniProt P23368); *HsME3m*, mitochondrial human ME3 (UniProt Q16798); *SbME*, *Sorghum bicolor* ME (UniProt Q84LQ5); *ZmME*, *Zea mays* ME (UniProt P16243); *AsMEem*, mitochondrial *Ascaris suum* ME (UniProt P27443). Figure prepared using Clustal Omega¹ and ESPrict 3.0².

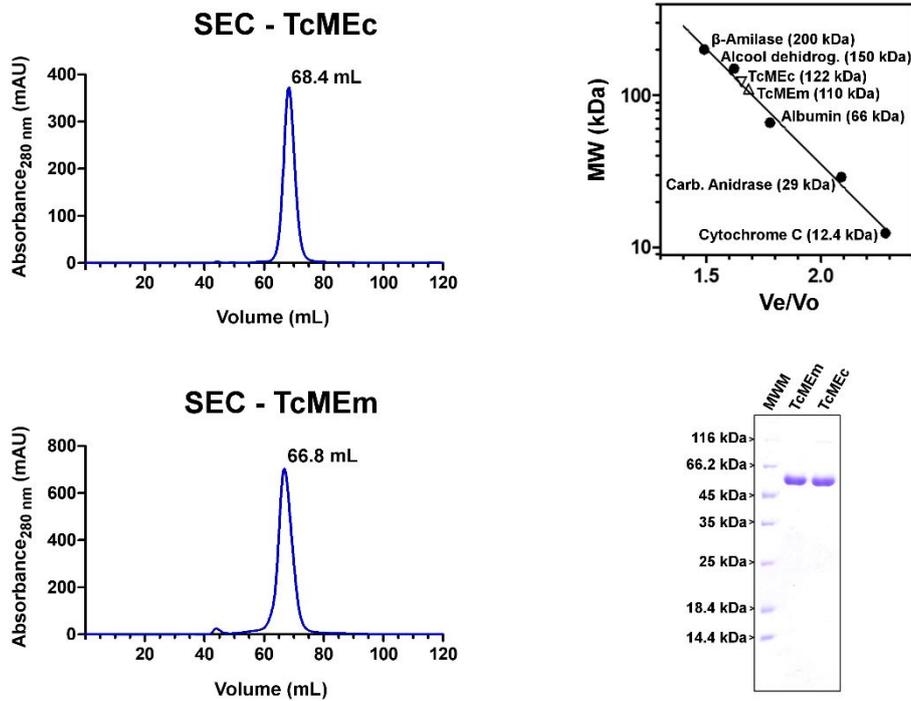


Figure S2. Size exclusion chromatography (SEC) and oligomeric state analysis of *Trypanosoma cruzi* malic enzymes, cytosolic (TcMEc) and mitochondrial (TcMEM) isoforms. Chromatograms obtained for TcMEc and TcMEM (left side) using a Superdex 200 16/600 column (GE Healthcare) show that both proteins eluted with similar elution volumes (V_e). Analysis of the oligomeric state with protein standards of known molecular weights indicate that TcMEc and TcMEM eluted from column in major peaks correspond to dimers (top right). Electrophoretic analysis in denaturing conditions (SDS-PAGE) shows that the protomers of TcMEc and TcMEM have molecular weight of 64.8 and 65.8 kDa, respectively (bottom-right). Figure adapted from ref.³.

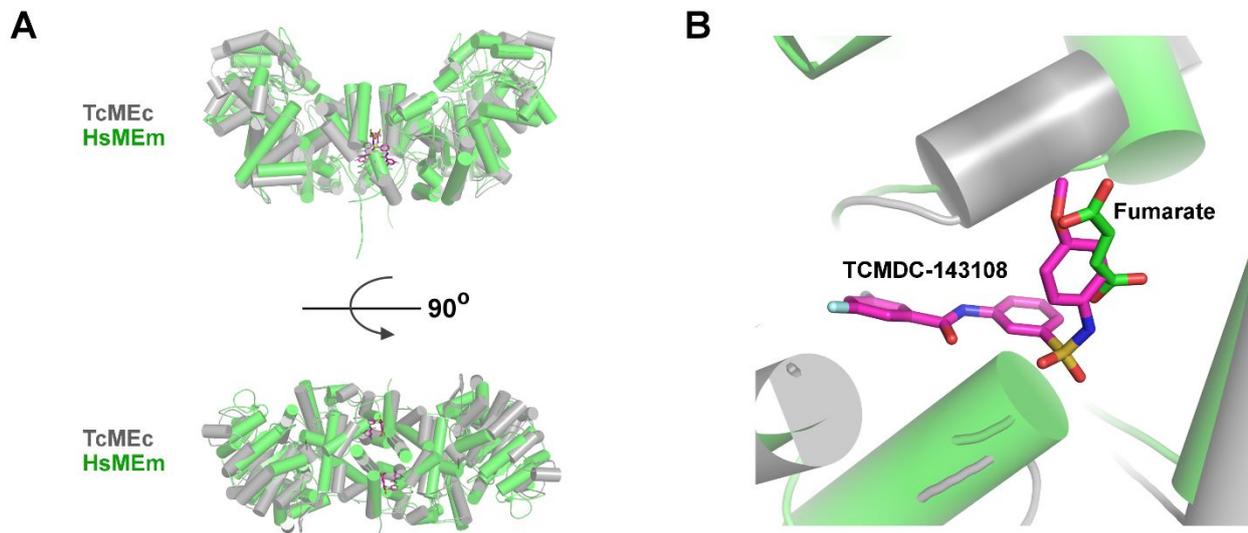


Figure S3. Superposition of Malic Enzymes from *Trypanosoma cruzi* (TcMEc, cytosolic isoform) and *Homo sapiens* (mitochondrial isoform, HsME2m).

Structural alignment of TcMEc dimer (gray cartoon, PDB 6W29) with a HsME2m dimer (green cartoon, PDB 1GZ3) (**A**) showing that the sulfonamide TCMDC-143108 overlaps with fumarate (**B**), the allosteric activator of the human enzyme. Both the sulfonamide and allosteric activator are located at the dimer interface, apart from the enzyme catalytic center.

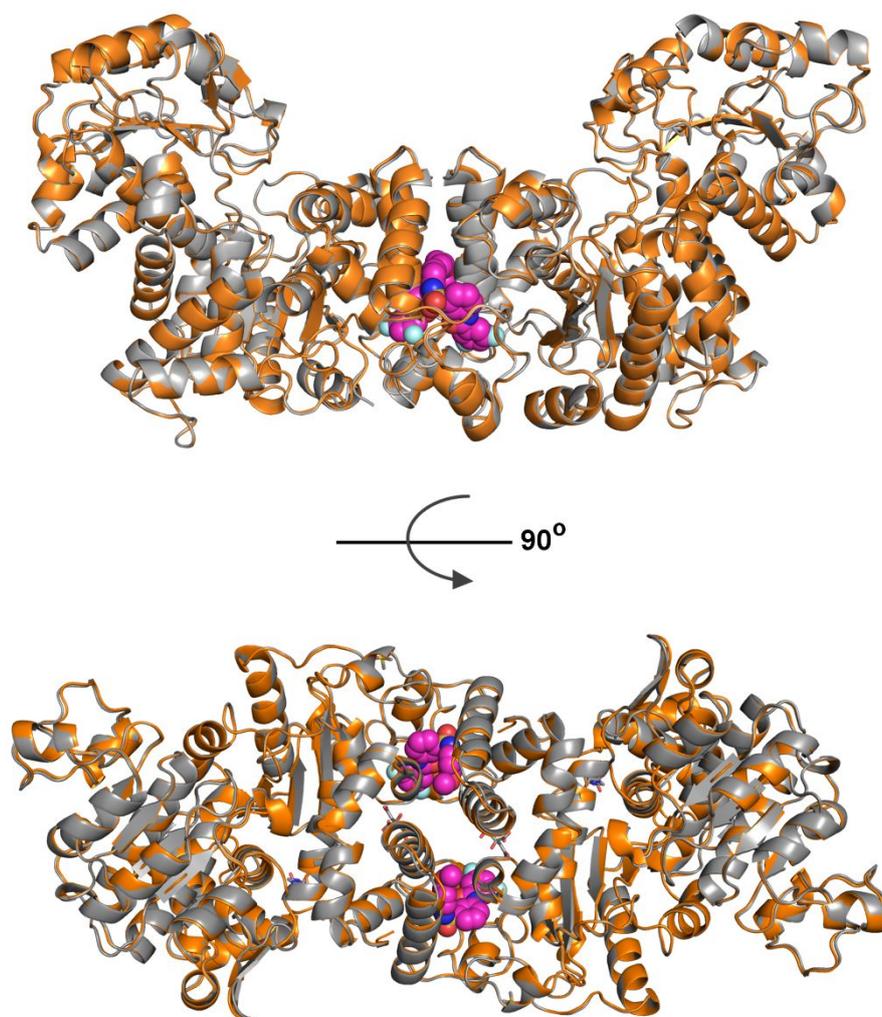


Figure S4. Superposition of structures of TcMEc dimer in ligand-free form (orange cartoon) and bound to sulfonamide inhibitor (gray cartoon with ligand as magenta spheres).

The alignment of TcMEc structures in ligand-bound and ligand-free form shows that the the enzyme has exactly the same domain and quaternary structure arrangement in these two forms (RMSD of 0.24 Å). This supports the idea that the binding of the inhibitor to the allosteric site locks the enzyme in an inactive conformation.

Table S1. Effect of TcMEc sulfonamide inhibitors over *T. cruzi* (Tulahuen strain) intracellular amastigotes infecting H9c2. Molecules described in this table refers to previously reported compounds identified by HTS.⁴

ID	Modifier (#1)	pIC50 <i>T. cruzi</i> INF CELL ^a	Modifier (#2)	pIC50 <i>T. cruzi</i> AM CELL ^b	Modifier (#3)	pIC50 <i>T. cruzi</i> H9C2 ^c	SI (AMCELL - H9C2) ^d
ATR3-001	=	4,6	=	4,6	=	5,2	-0,6
ATR3-002	<	4,3	<	4,3	<	4,3	0
ATR3-005	=	4,7	=	4,9	=	4,7	0,2
ATR3-006	=	4,6	=	4,8	<	4,3	0,5
ATR3-007	<	4,3	<	4,3	<	4,3	0
ATR3-008	<	4,3	<	4,3	<	4,3	0
ATR3-014	<	4,3	<	4,3	<	4,3	0
ATR3-016	<	4,3	<	4,3	<	4,3	0
ATR3-021	=	4,5	=	4,6	<	4,3	0,3
ATR3-023	=	4,4	=	4,8	=	4,4	0,4
ATR3-028	=	5,1	=	5,1	=	5,3	-0,2
ATR3-030	=	4,9	=	4,9	=	5,2	-0,3
ATR3-033	=	4,4	=	4,5	=	4,5	0
ATR3-034	=	4,8	=	5,1	=	5,1	0
ATR3-036	=	5,1	=	5,1	=	5	0,1
ATR3-040	<	4,3	<	4,3	<	4,3	0
ATR3-043	=	4,8	=	5	=	5	0
ATR3-045	=	4,8	=	5	=	5	0
ATR3-049	=	4,6	=	4,4	=	4,6	-0,2
ATR3-053	=	5	=	4,9	<	4,3	0,6
ATR3-057	=	4,3	=	4,5	=	4,5	0
ATR3-065	<	4,3	<	4,3	=	4,4	-0,1
ATR3-072	=	4,9	=	5	=	5,2	-0,2
ATR3-073	=	4,7	=	4,8	=	4,6	0,2
ATR3-074	=	5	=	5	=	5,2	-0,2
ATR3-075	=	4,7	=	4,9	=	4,8	0,1
ATR3-076	<	4,3	=	4,4	=	4,4	0
ATR3-080	=	4,8	=	4,9	=	4,9	0
ATR3-084	=	4,6	=	4,5	=	4,6	-0,1
ATR3-086	=	4,8	=	5	=	4,8	0,2
ATR3-087	<	4,3	<	4,3	<	4,3	0
ATR3-088	=	4,4	=	4,5	<	4,3	0,2
ATR3-089	=	4,7	=	4,7	=	4,7	0
ATR3-090	=	4,7	=	4,5	=	4,6	-0,1
ATR3-091	=	5	=	5,2	=	5	0,2
ATR3-092	=	4,8	=	4,9	<	4,3	0,6
ATR3-093	=	4,5	=	4,5	<	4,3	0,2
ATR3-095	<	4,3	<	4,3	<	4,3	0
ATR3-098	=	4,6	=	4,8	=	4,8	0
ATR3-100	=	4,6	=	4,8	=	4,6	0,2
ATR3-102	<	4,3	<	4,3	<	4,3	0

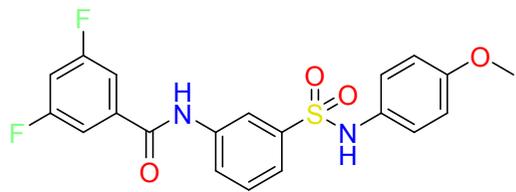
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Table S1. Continuation...

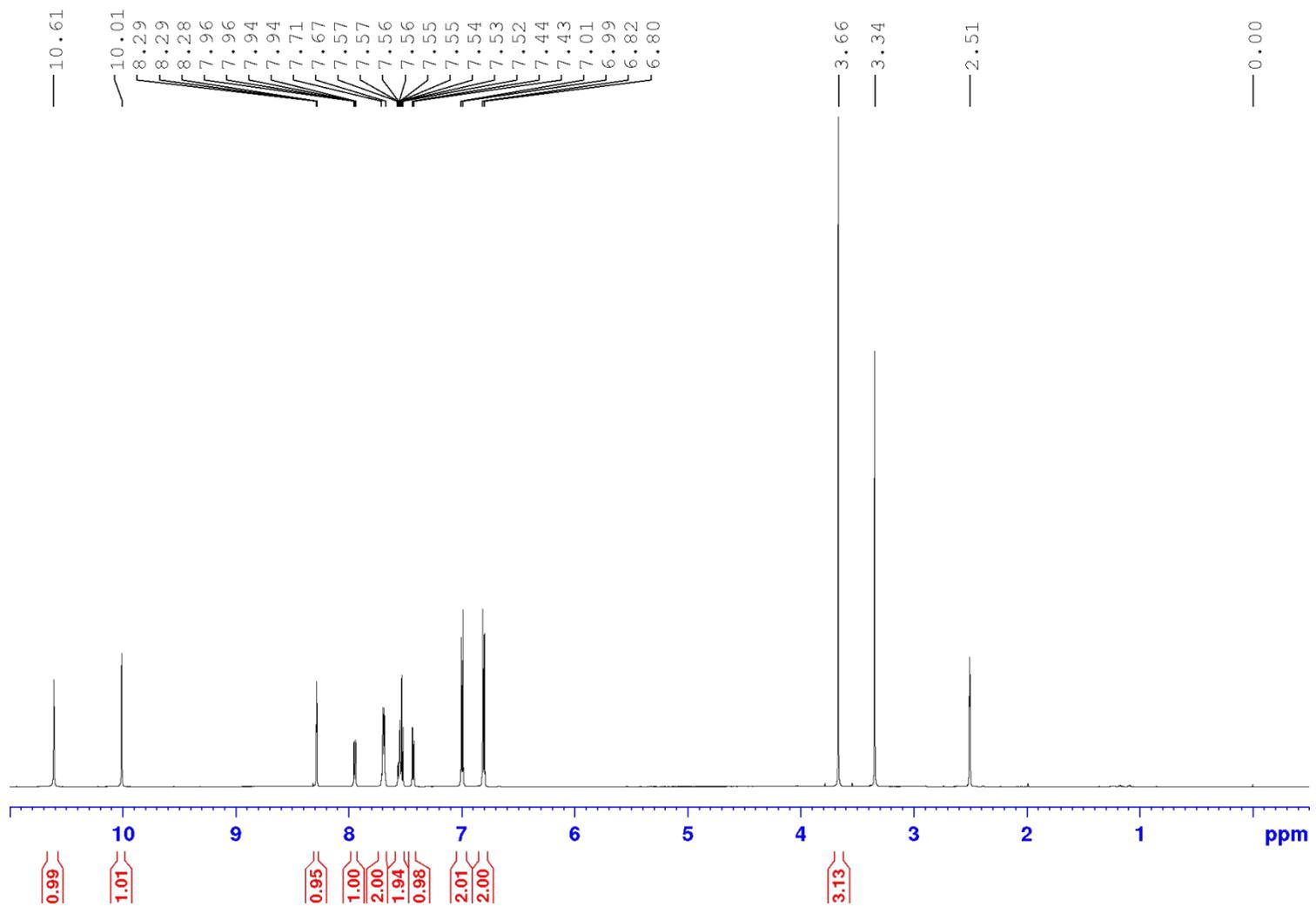
ID	Modifier (#1)	pIC50 <i>T. cruzi</i> INF CELL ^a	Modifier (#2)	pIC50 <i>T. cruzi</i> AM CELL ^b	Modifier (#3)	pIC50 <i>T. cruzi</i> H9C2 ^c	SI (AMCELL - H9C2) ^d
ATR3-103	=	4,5	=	4,7	=	4,5	0,2
ATR3-105	=	4,7	=	4,8	=	4,7	0,1
ATR3-107	<	4,3	=	4,3	<	4,3	0
ATR3-112	<	4,3	<	4,3	<	4,3	0
ATR3-114	=	4,7	=	4,8	=	6	-1,2
ATR3-115	=	4,5	=	4,8	=	4,6	0,2
ATR3-124	=	4,3	=	4,4	=	4,3	0,1
ATR3-125	=	4,8	=	5,1	=	5,1	0
ATR3-128	=	4,4	=	4,7	<	4,3	0,4
ATR3-130	=	4,5	=	4,6	=	4,4	0,2
ATR3-132	=	4,6	=	4,6	=	4,5	0,1
ATR3-133	=	5	=	5,4	=	5,1	0,3
ATR3-139	<	4,3	<	4,3	<	4,3	0
ATR3-141	<	4,3	<	4,3	<	4,3	0
ATR3-142	=	4,5	=	4,7	<	4,3	0,4
ATR3-143	<	4,3	<	4,3	<	4,3	0
ATR3-144	=	4,4	=	4,6	<	4,3	0,3
ATR3-146	=	4,3	=	4,5	=	4,5	0
ATR3-152	=	5,2	=	5,5	=	5,3	0,2
ATR3-156	=	4,7	=	5,1	=	5	0,1

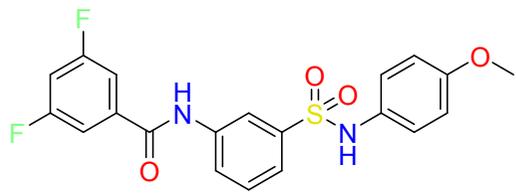
^a - pIC50 *Trypanosoma cruzi* Imaging: Infected Cells; ^b - pIC50 *Trypanosoma cruzi* Imaging: Amastigotes/Cell; ^c - pIC50 H2c9 Imaging: H9c2 Host Cardiomyocytes; ^d – Selective Index in log scale, calculated by the subtraction pIC50 *T. cruzi* AM CELL less pIC50 *T. cruzi* H9c2.

^1H and ^{13}C NMR spectra for compounds **1**, **6-43** and **3,5-difluorobenzoyl chloride**

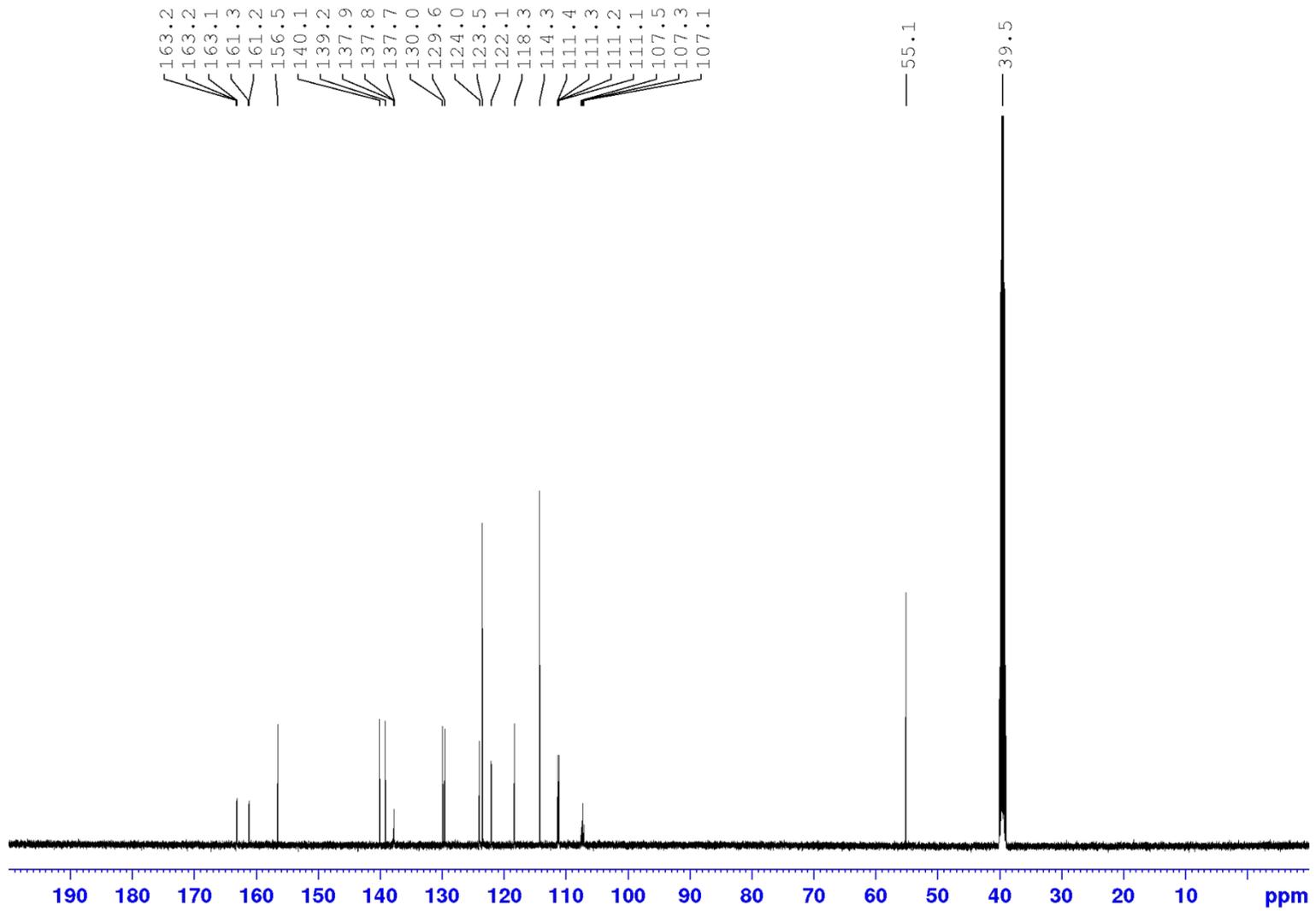


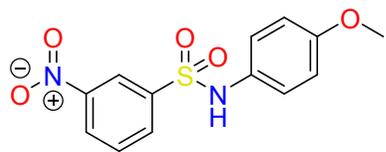
Compound 1 – ^1H NMR (600 MHz) – $\text{DMSO-}d_6$



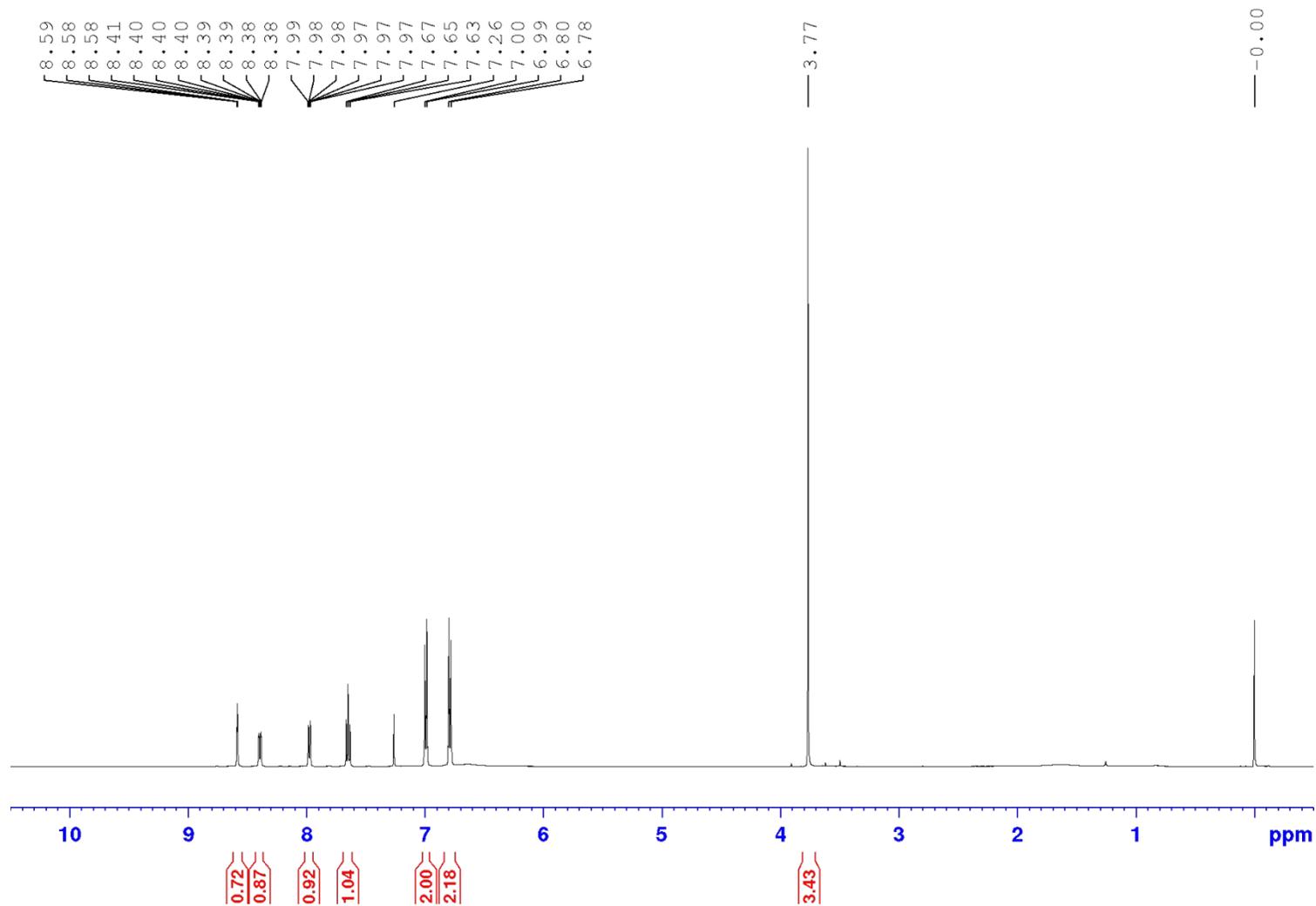


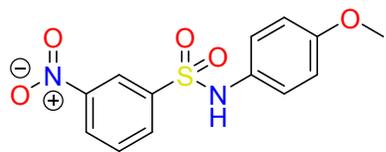
Compound 1 – ^{13}C NMR (150 MHz) – $\text{DMSO-}d_6$



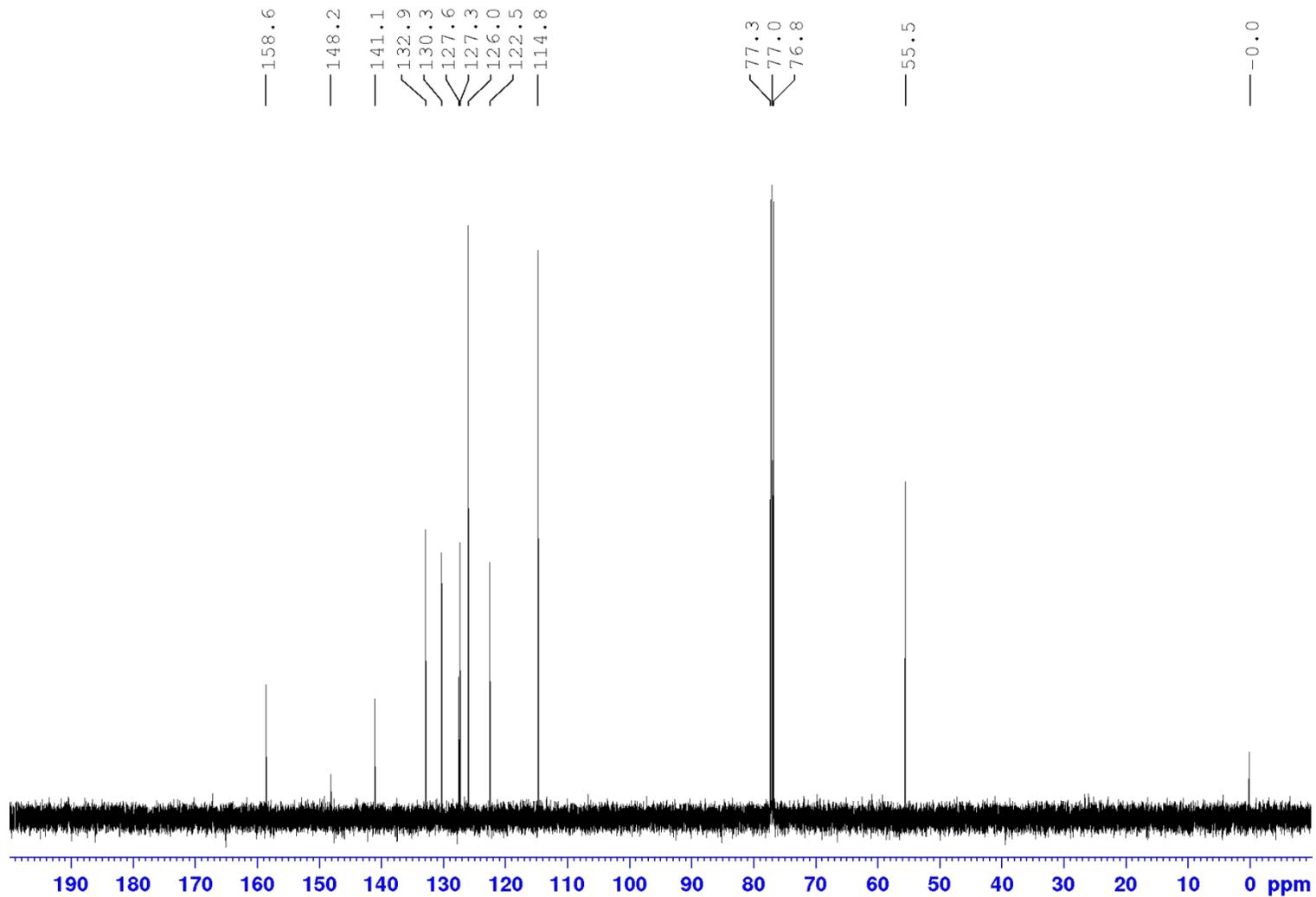


Compound 6 – ^1H NMR (500 MHz) – CDCl_3



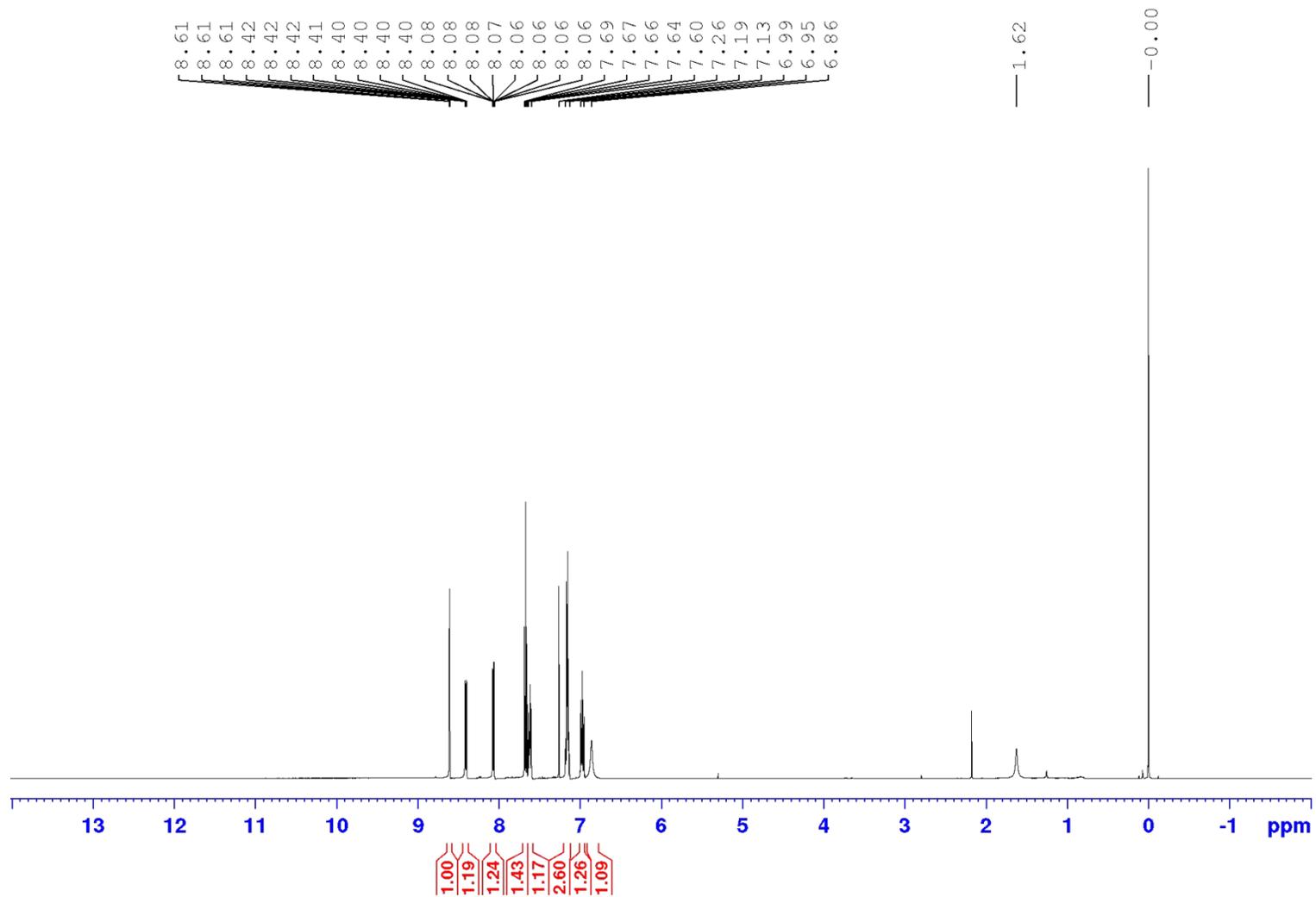


Compound 6 – ^{13}C NMR (125.5 MHz) – CDCl_3



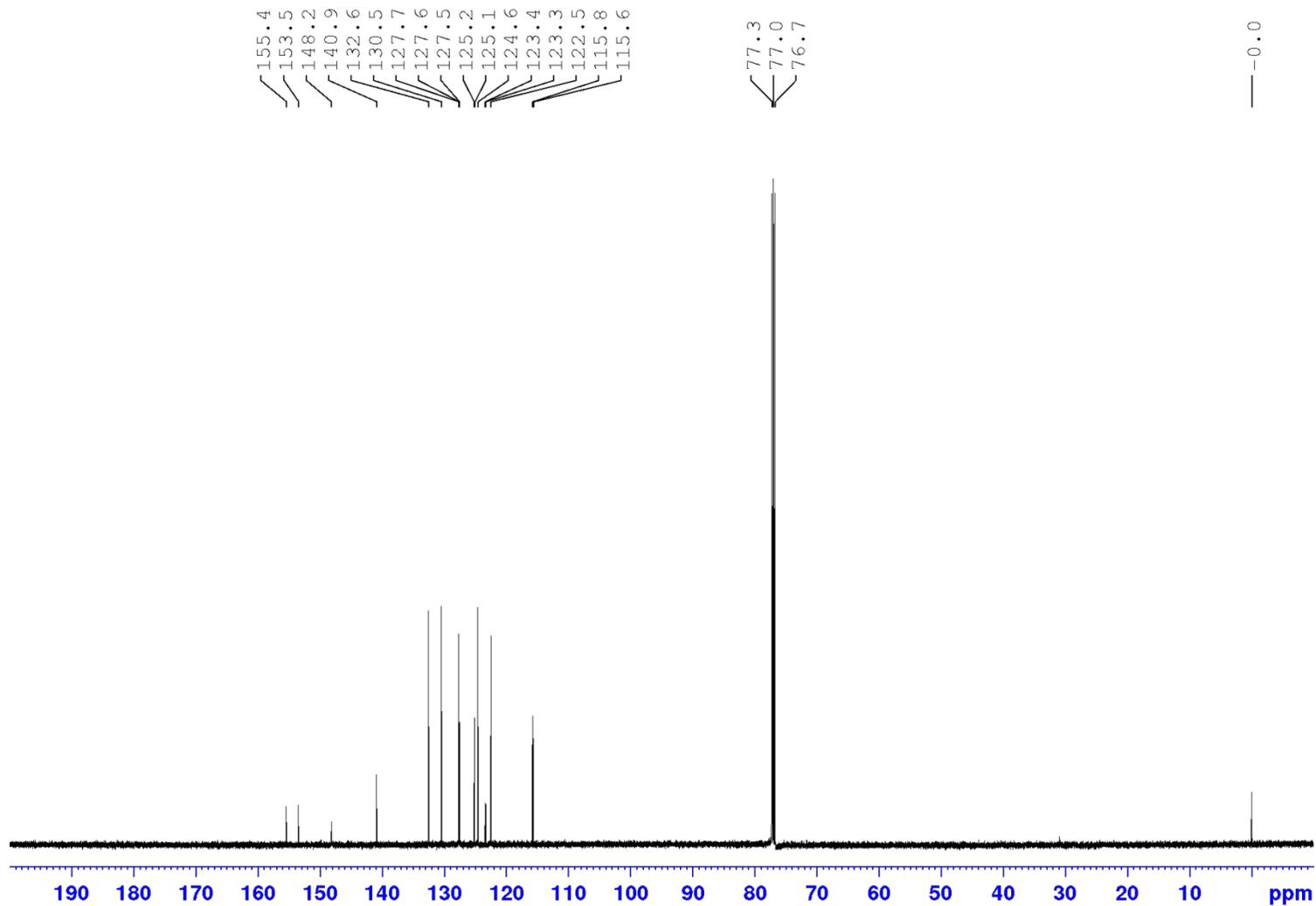


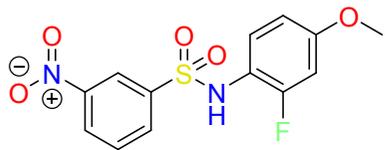
Compound 7 - ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



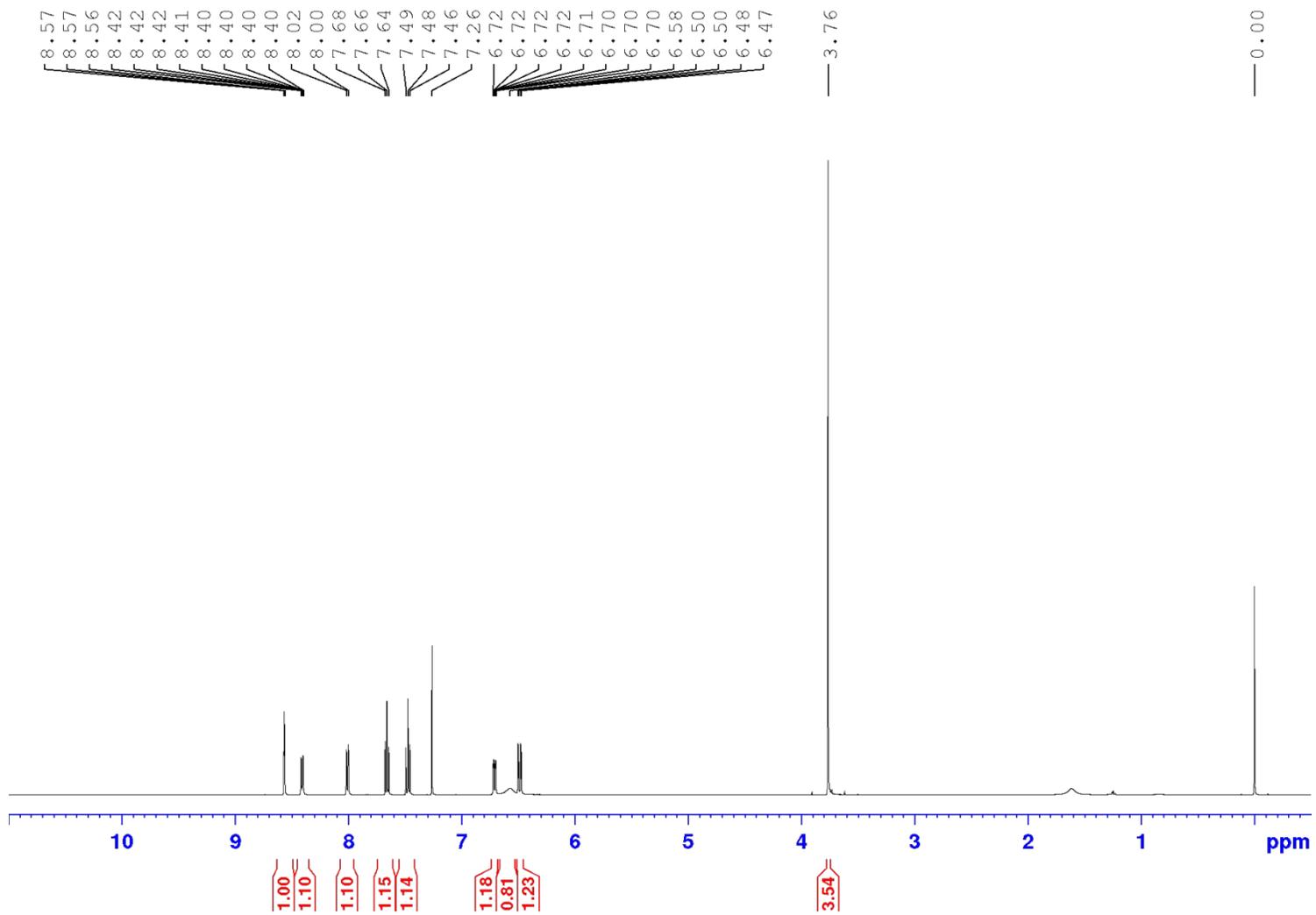


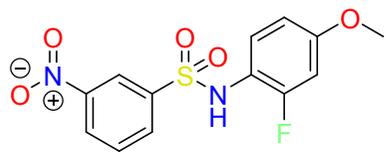
Compound 7 – ^{13}C NMR (125.5 MHz) – $\text{DMSO-}d_6$



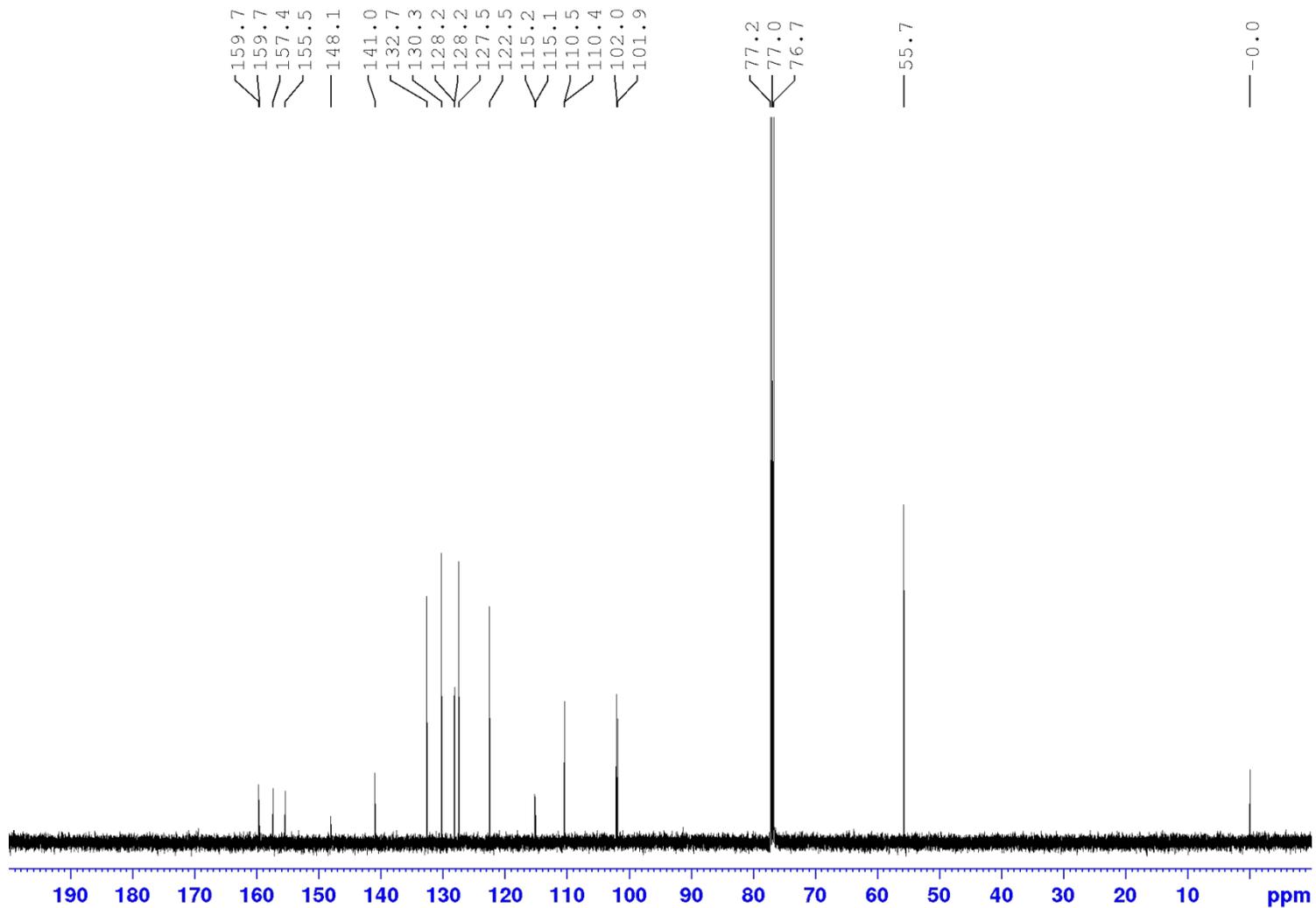


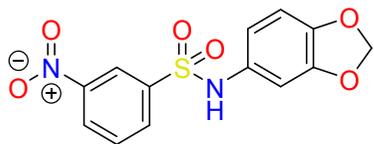
Compound **8** – ^1H NMR (500 MHz) – CDCl_3



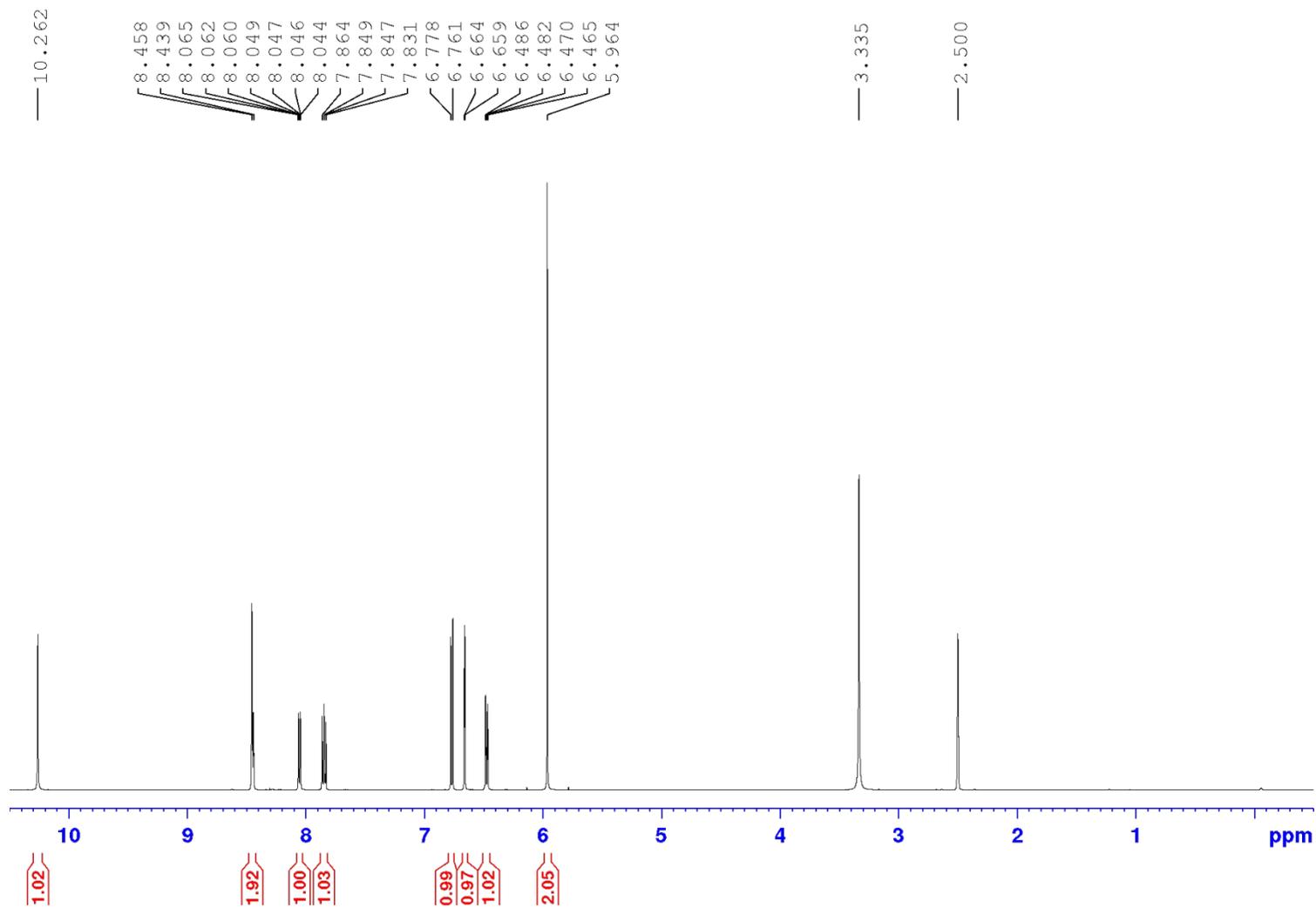


Compound 8 – ¹³C NMR (125.5 MHz) – CDCl₃



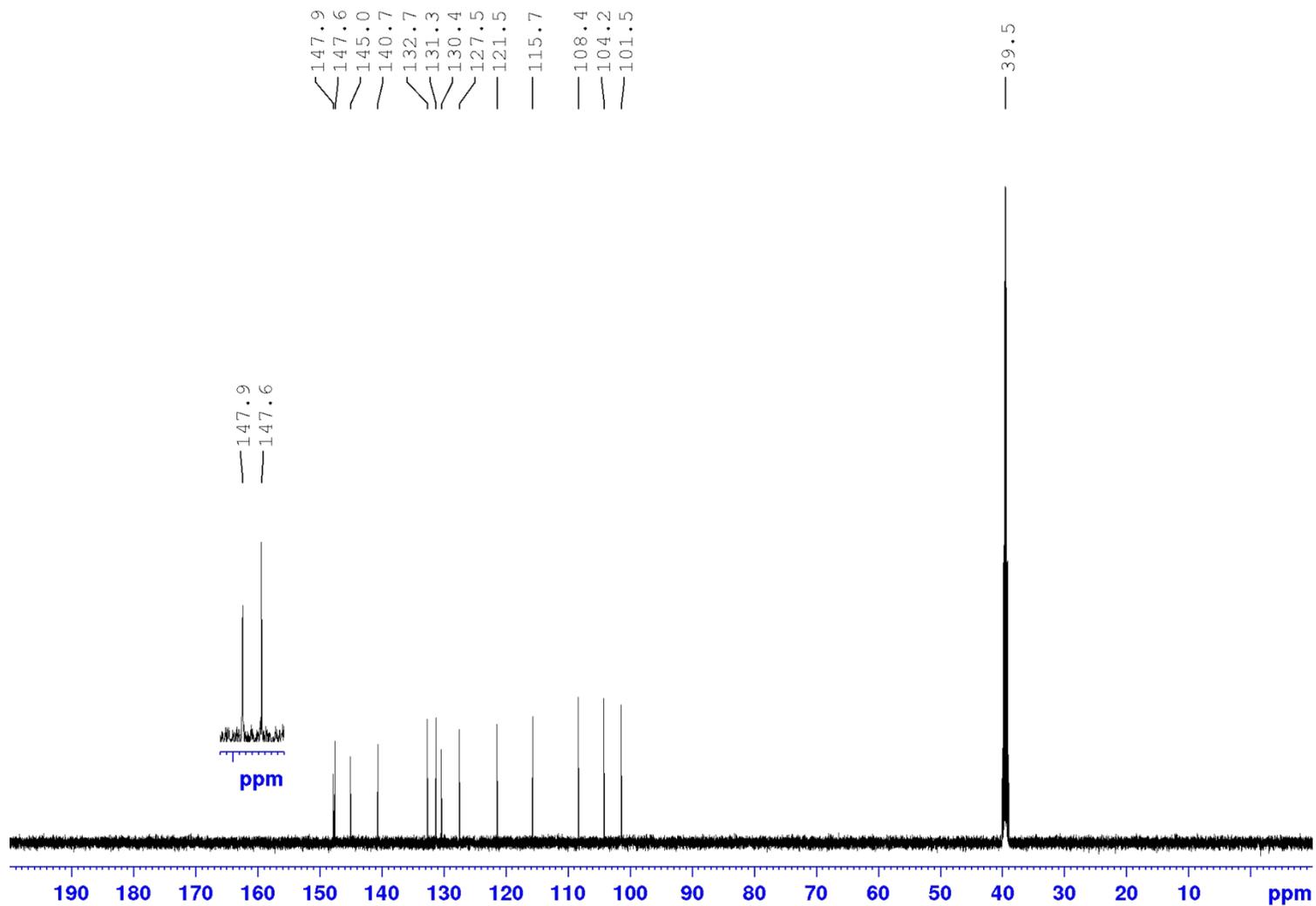


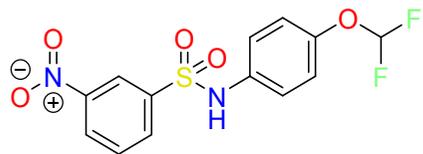
Compound **9** – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



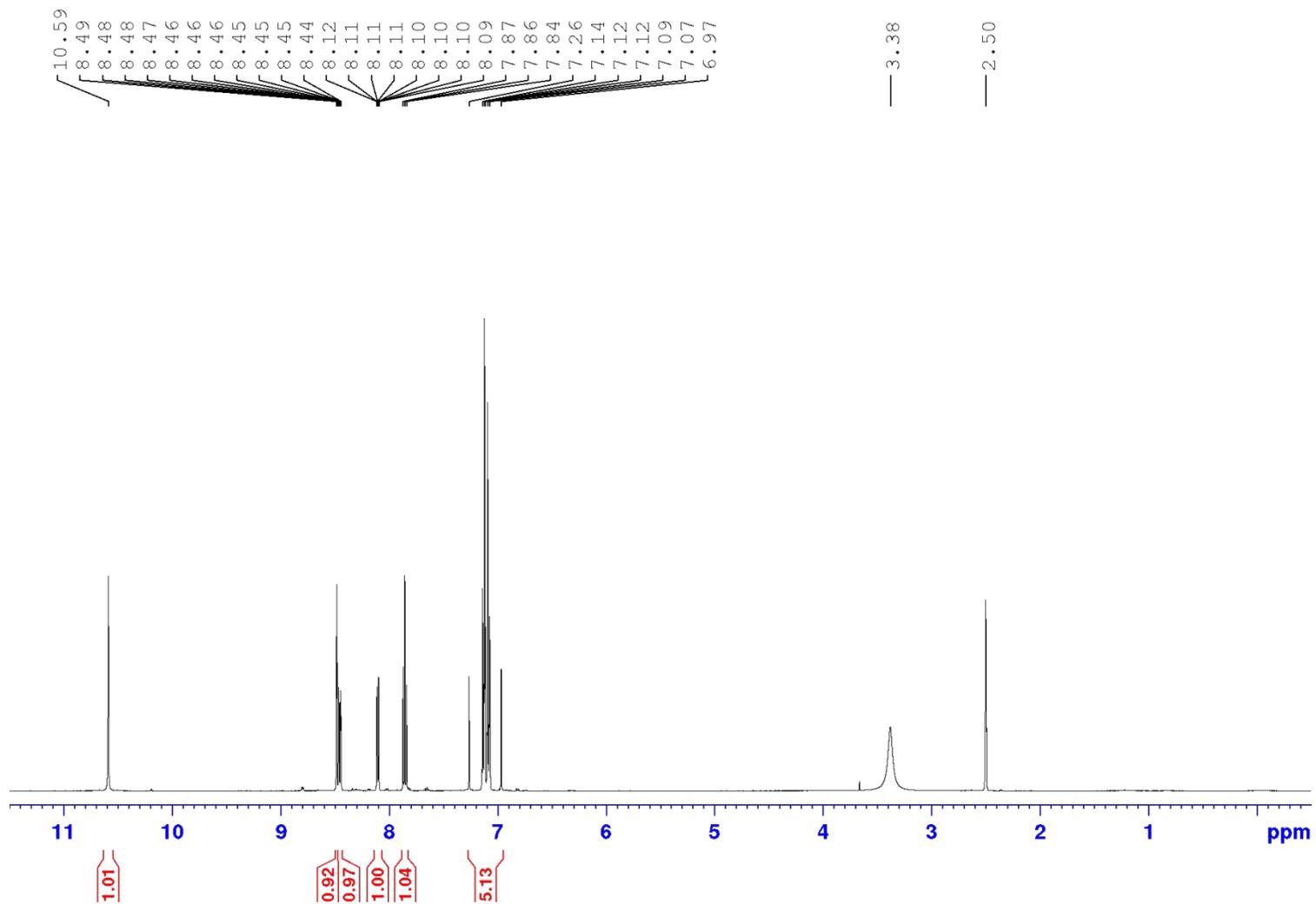


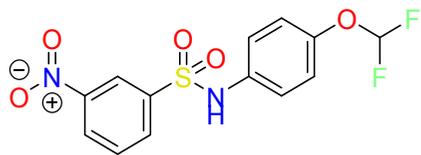
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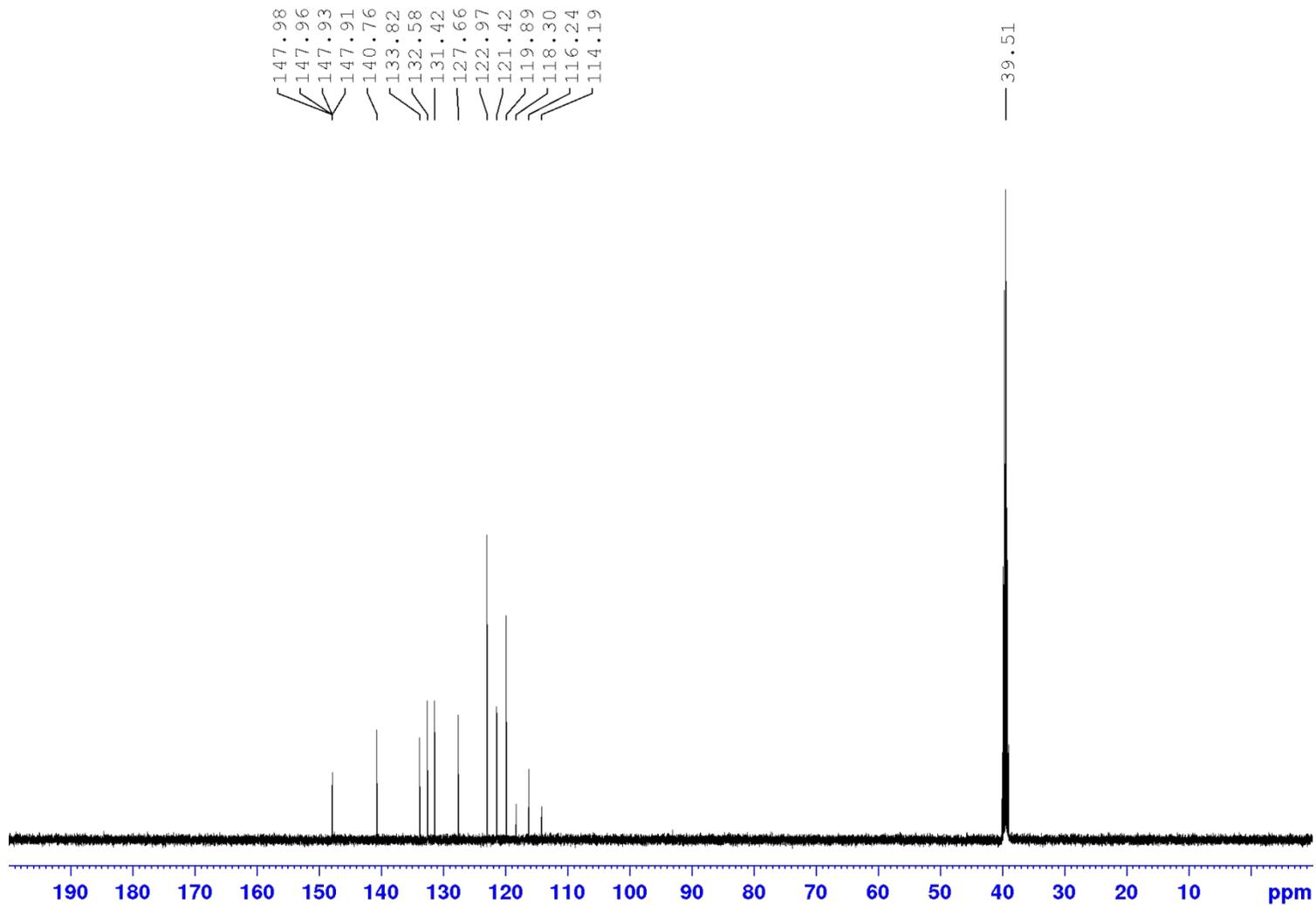


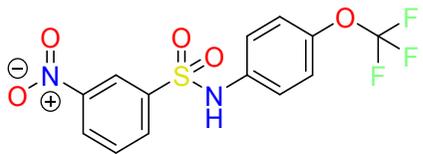
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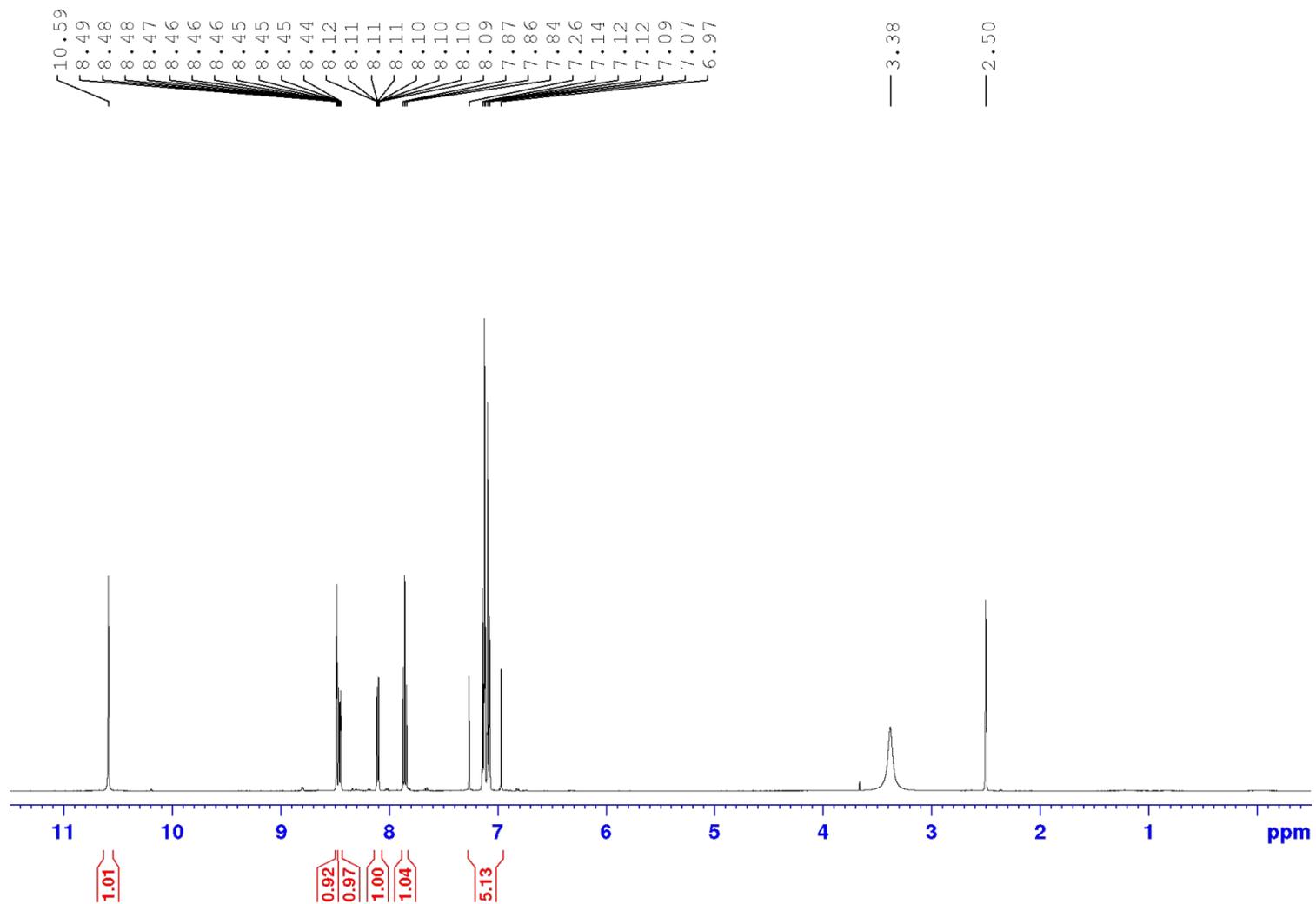


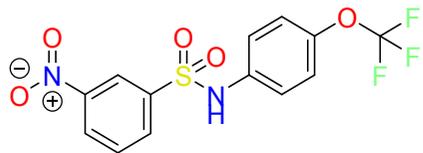
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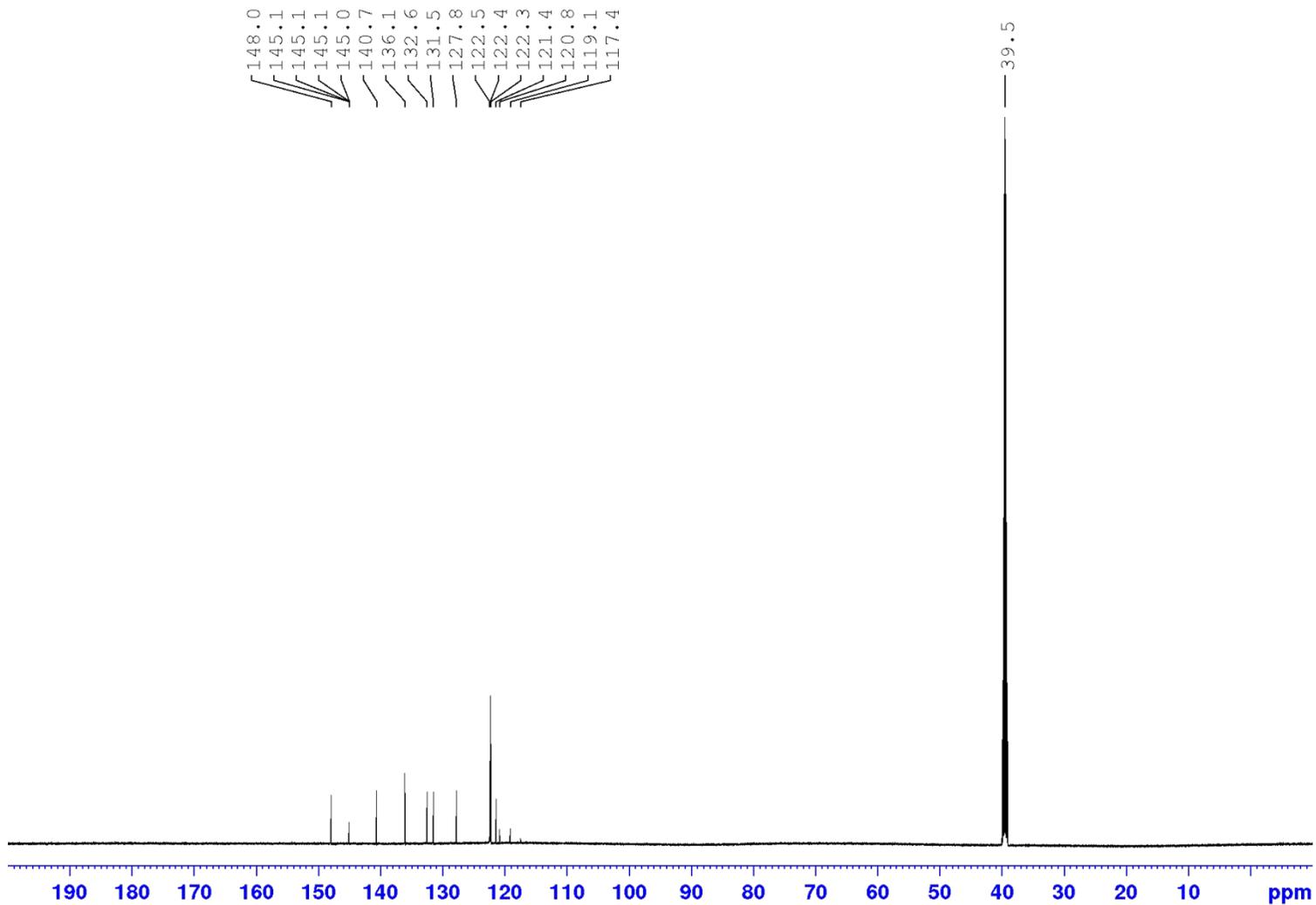


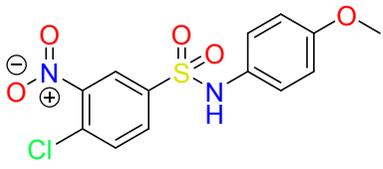
Compound 11 – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



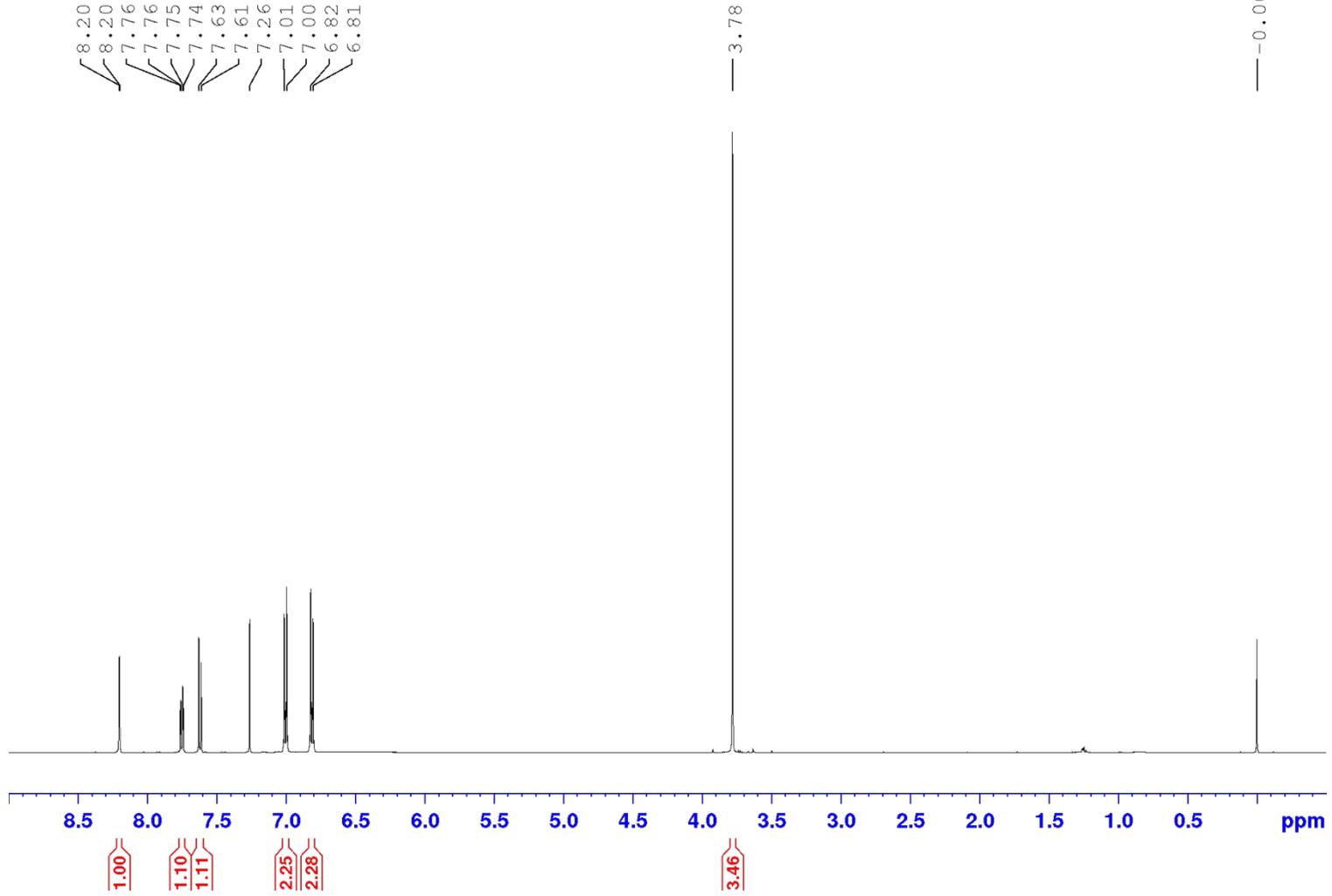


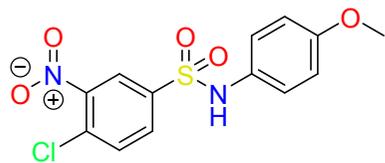
Compound **11** – ^{13}C NMR (150 MHz) – $\text{DMSO-}d_6$



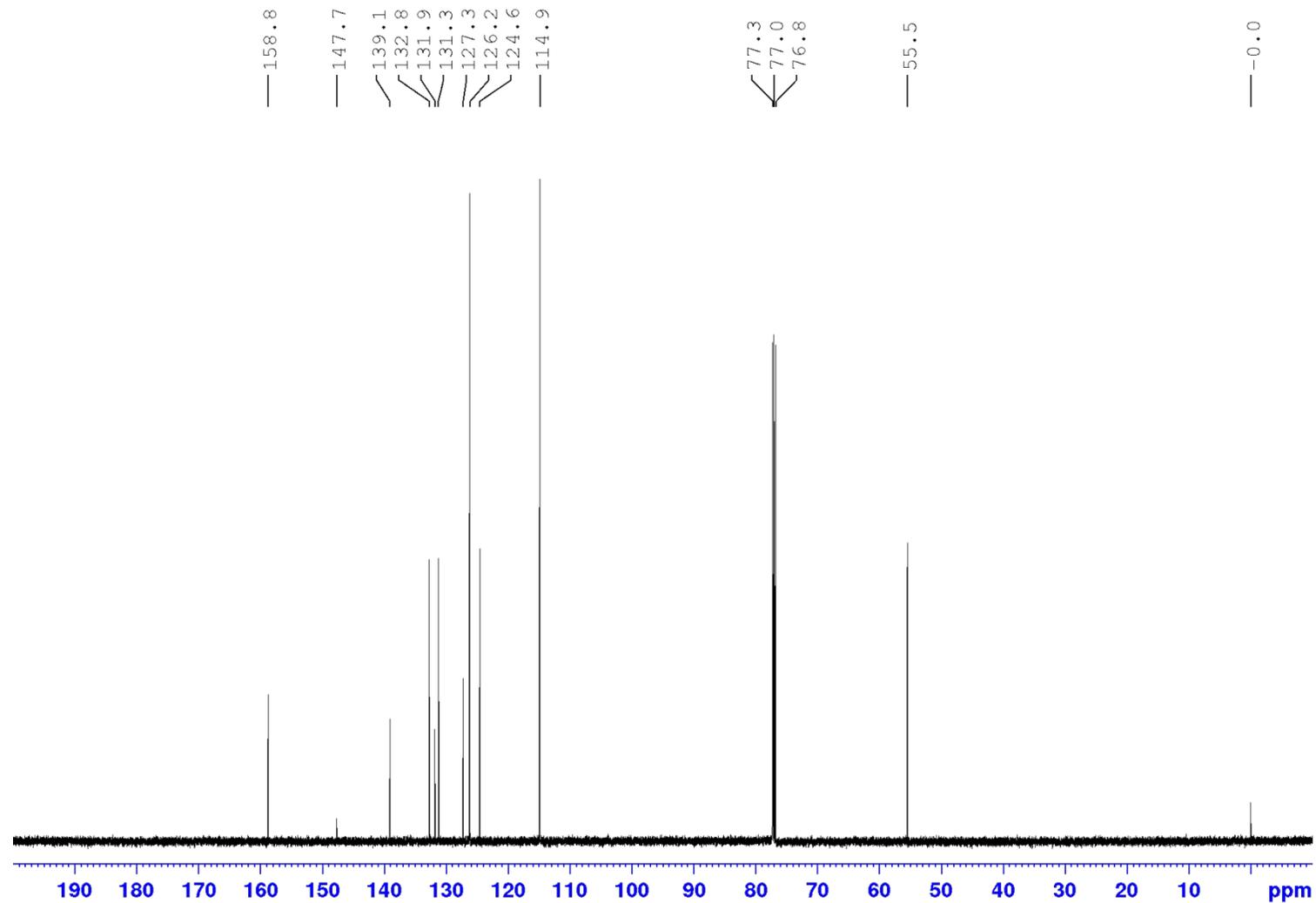


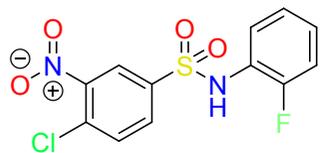
Compound 12 – ¹H NMR (500 MHz) – CDCl₃



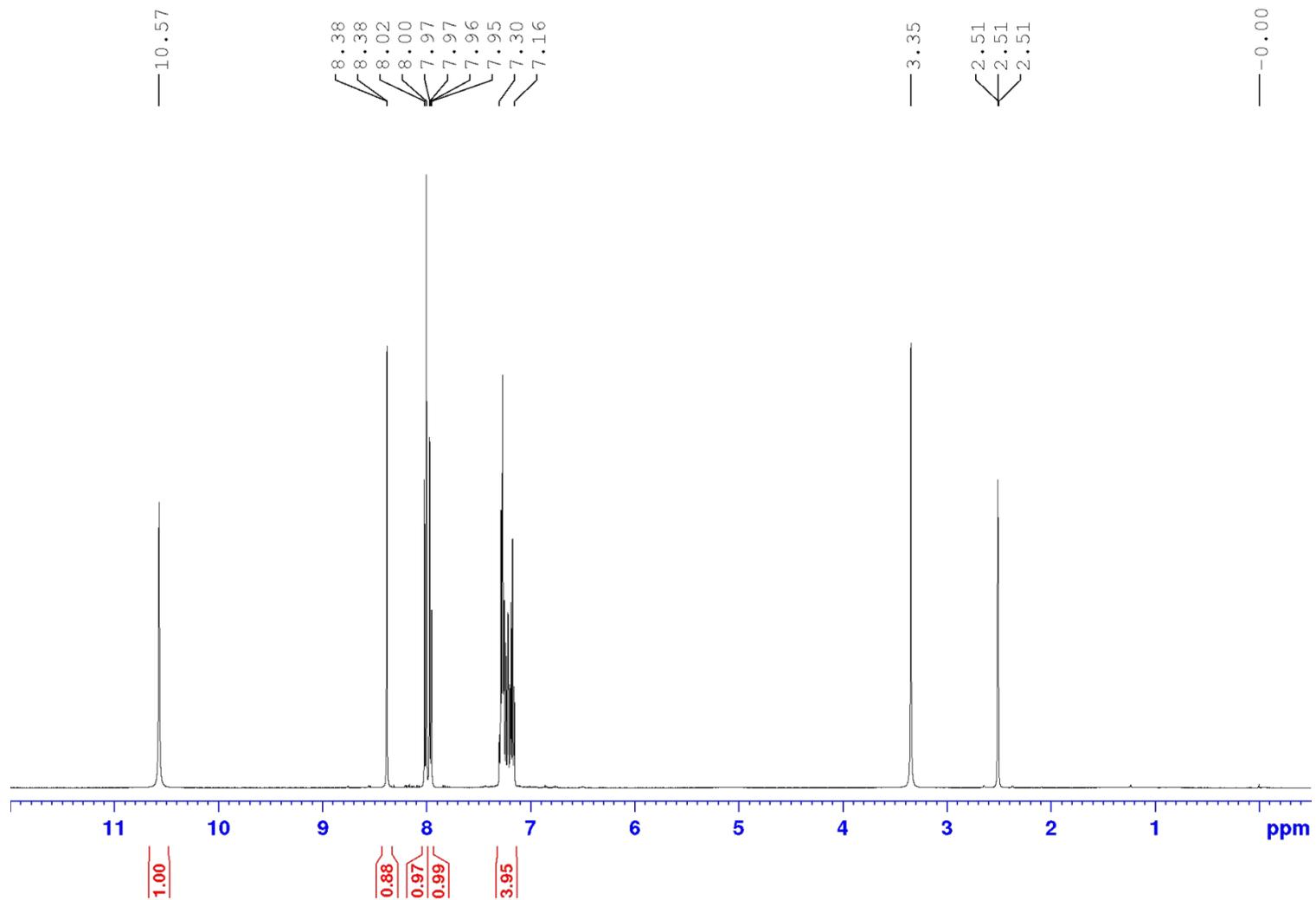


Compound **12** – ^{13}C NMR (125.5 MHz) – CDCl_3



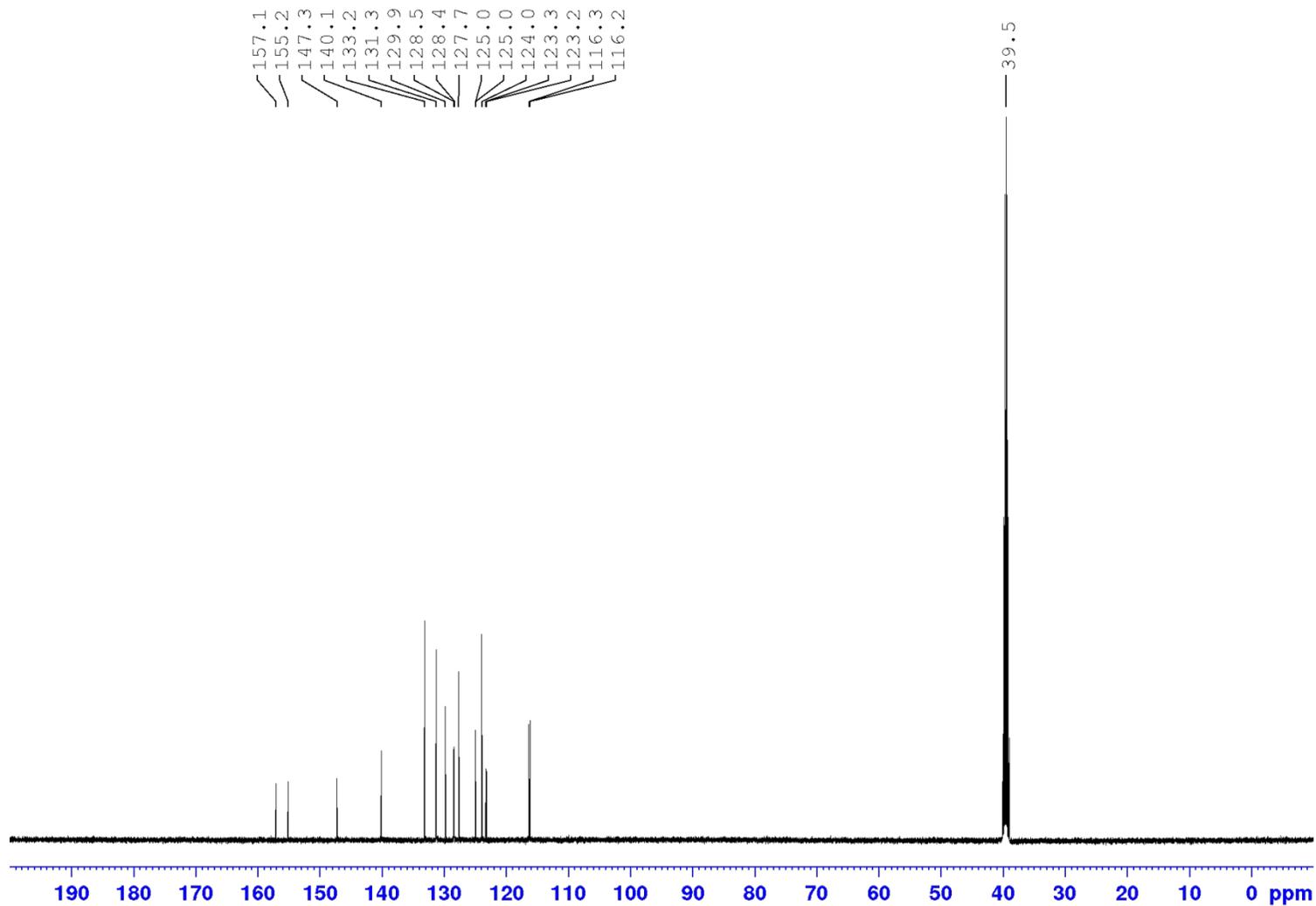


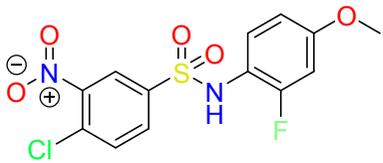
Compound **13** – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



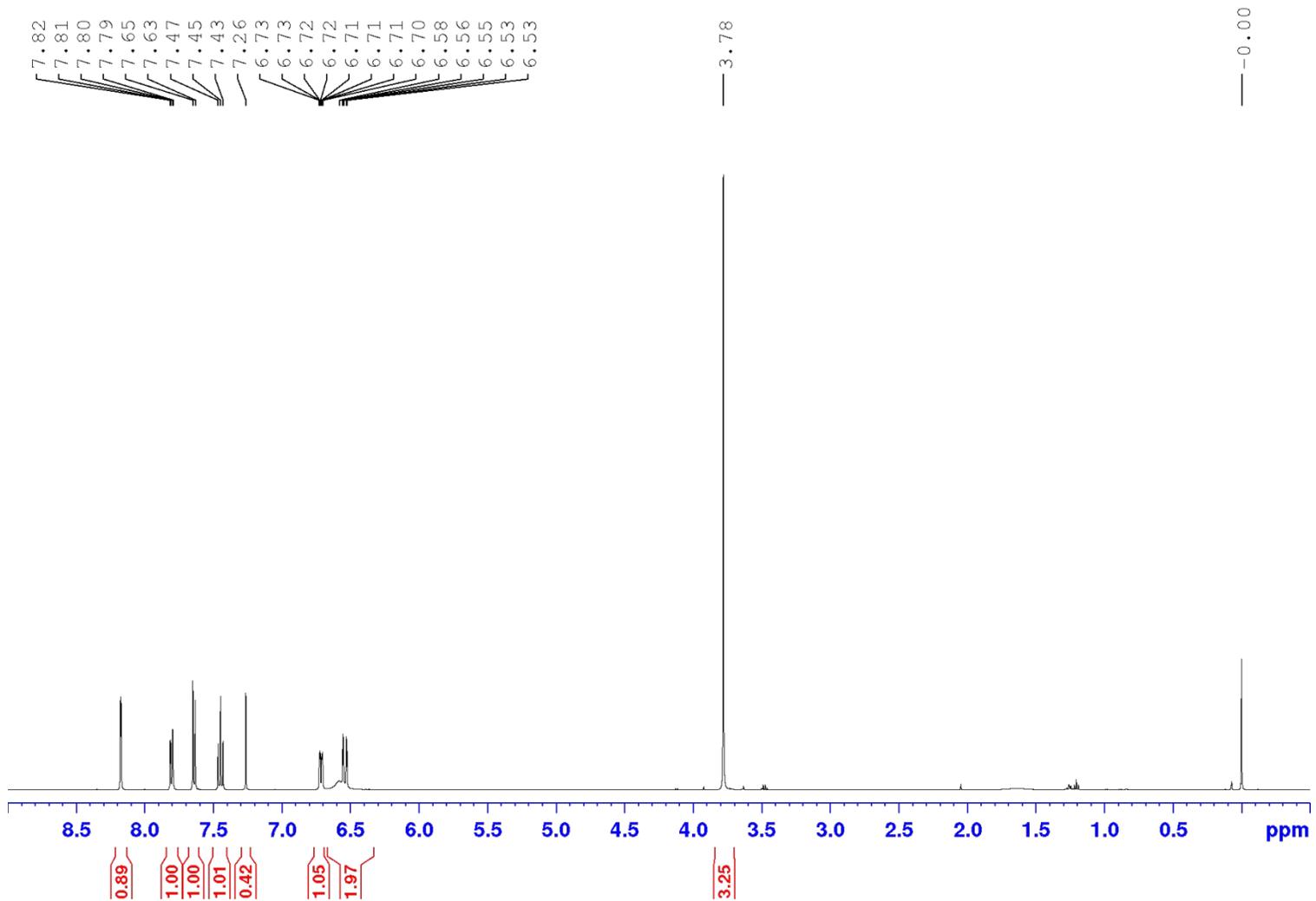


Compound 13 – ^{13}C NMR (125.5 MHz) – $\text{DMSO-}d_6$



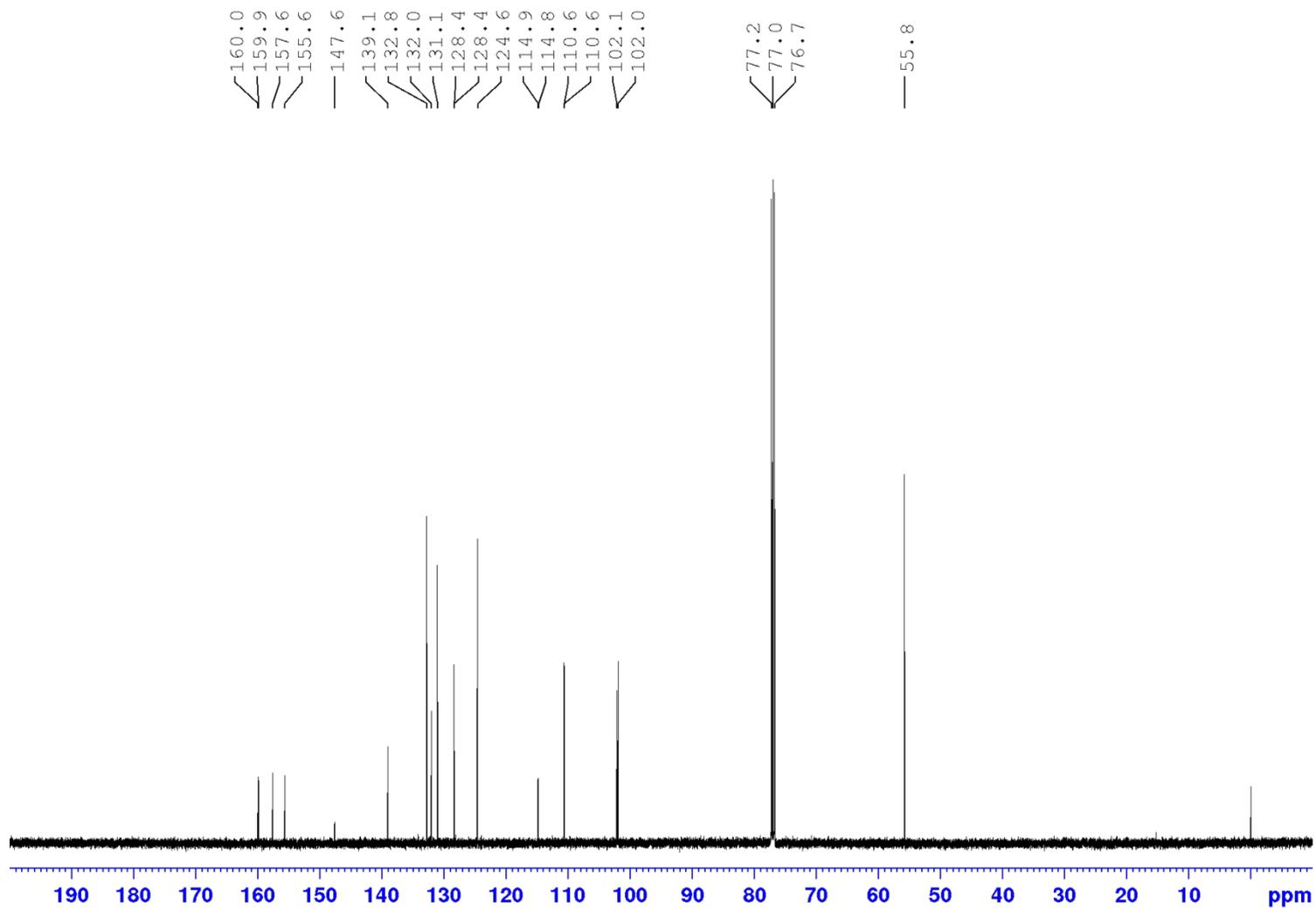


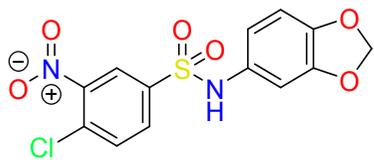
Compound 14 – ¹H NMR (500 MHz) – CDCl₃



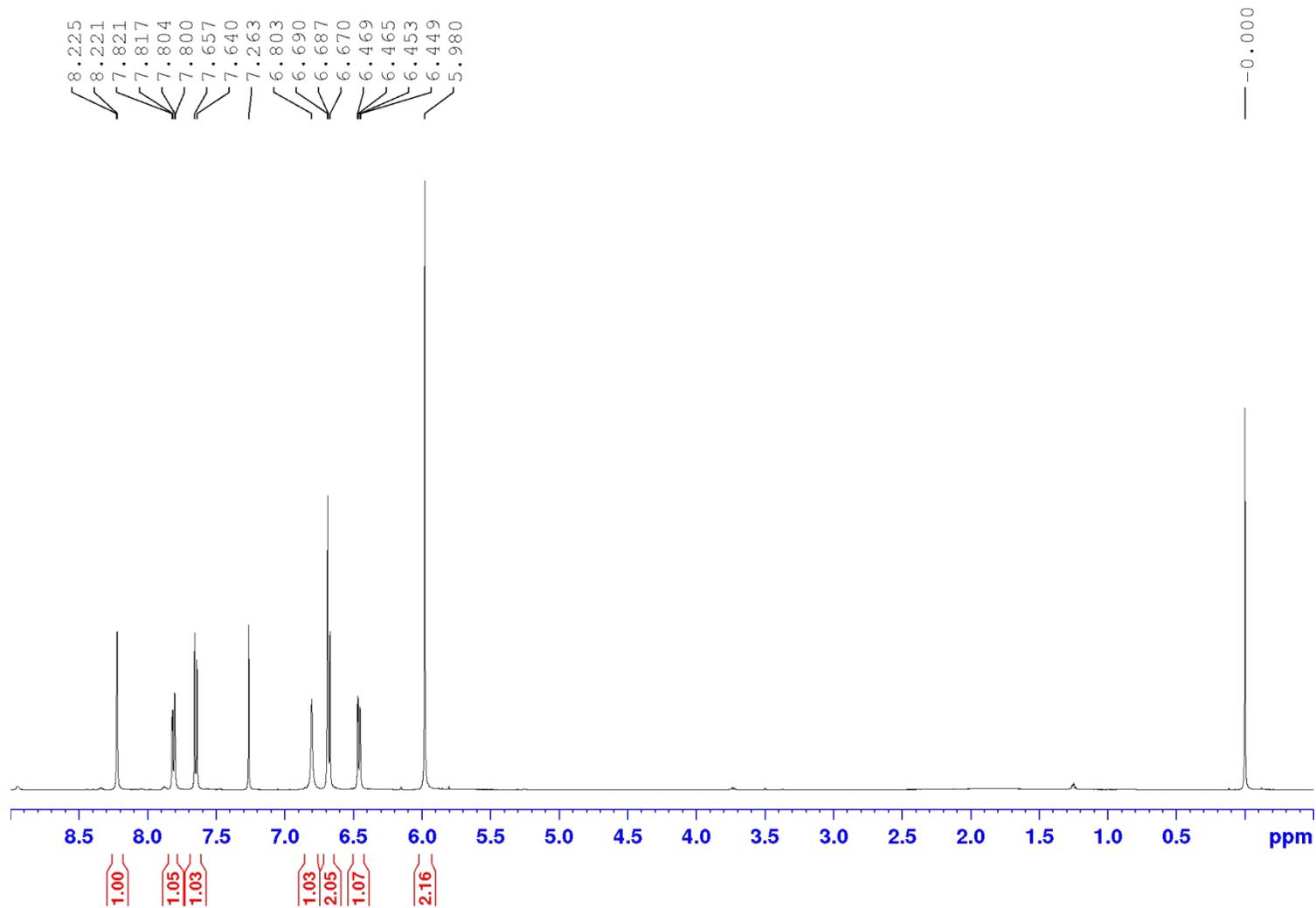


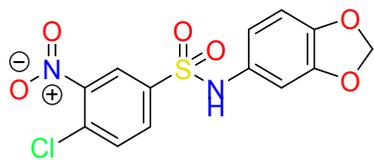
Compound **14** – ^{13}C NMR (125.5 MHz) – CDCl_3



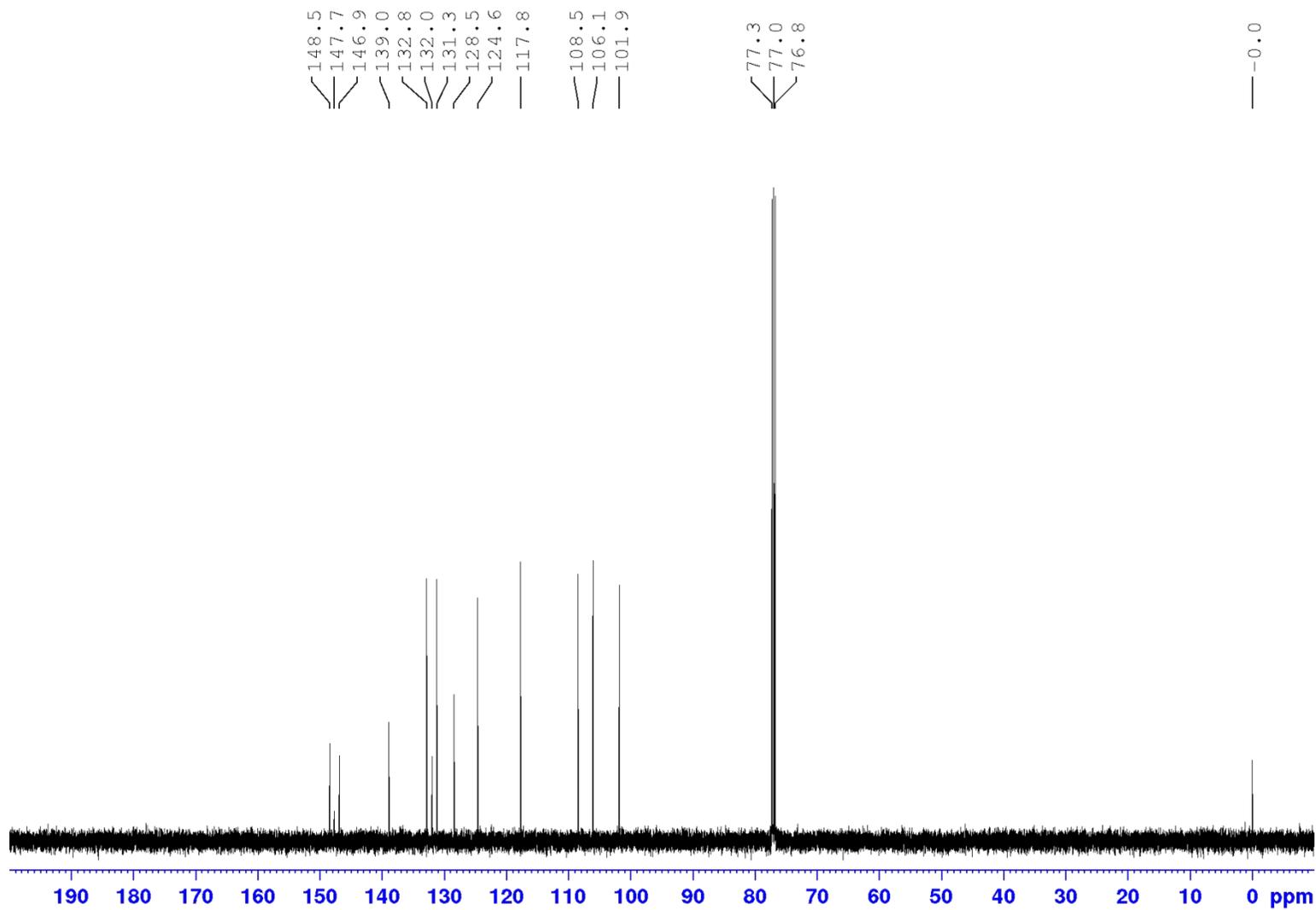


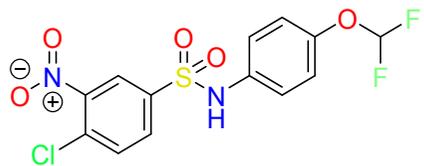
Compound 15 – ¹H NMR (500 MHz) – CDCl₃



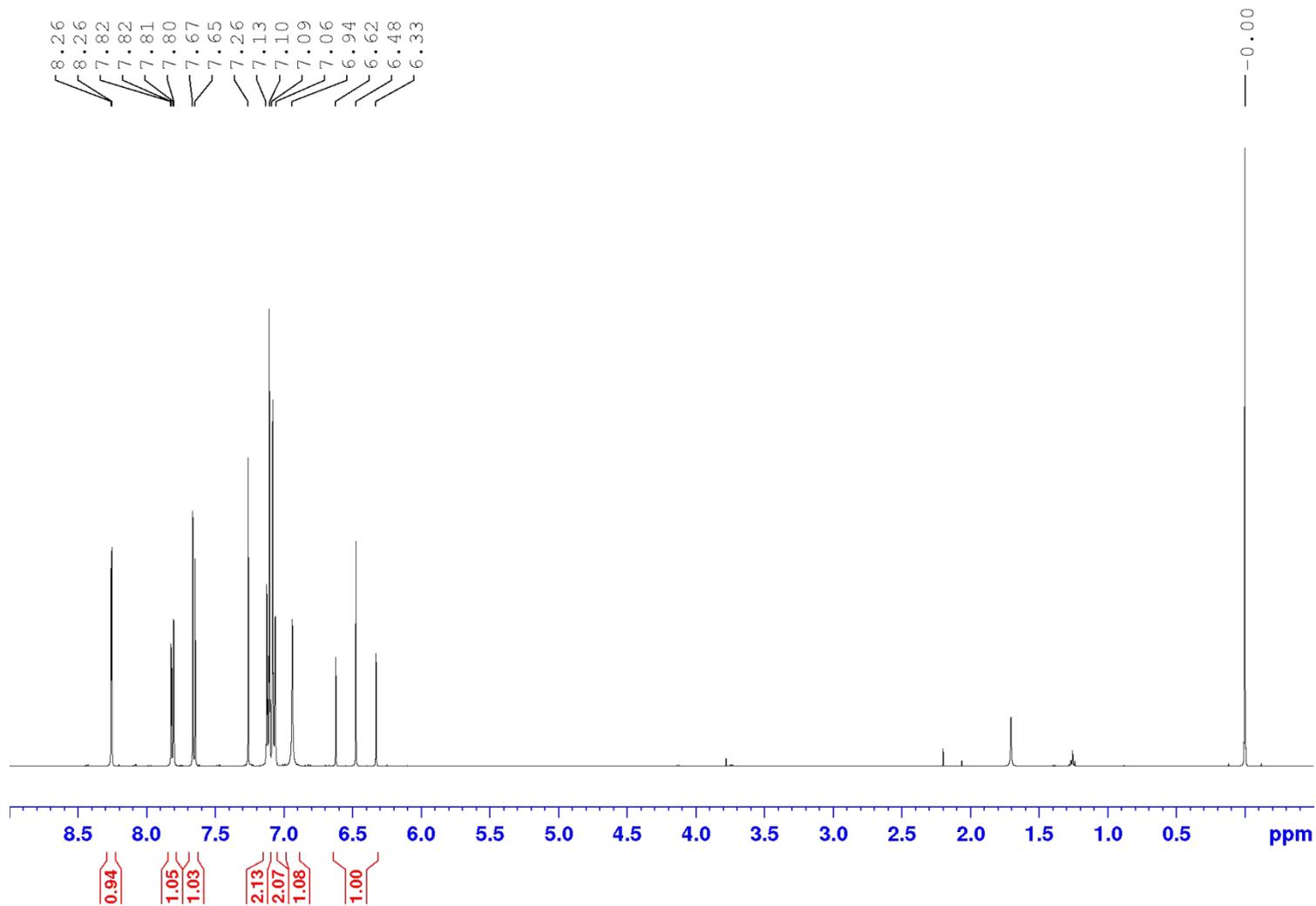


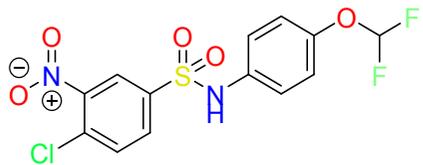
Compound **15** – ^{13}C NMR (125.5 MHz) – CDCl_3



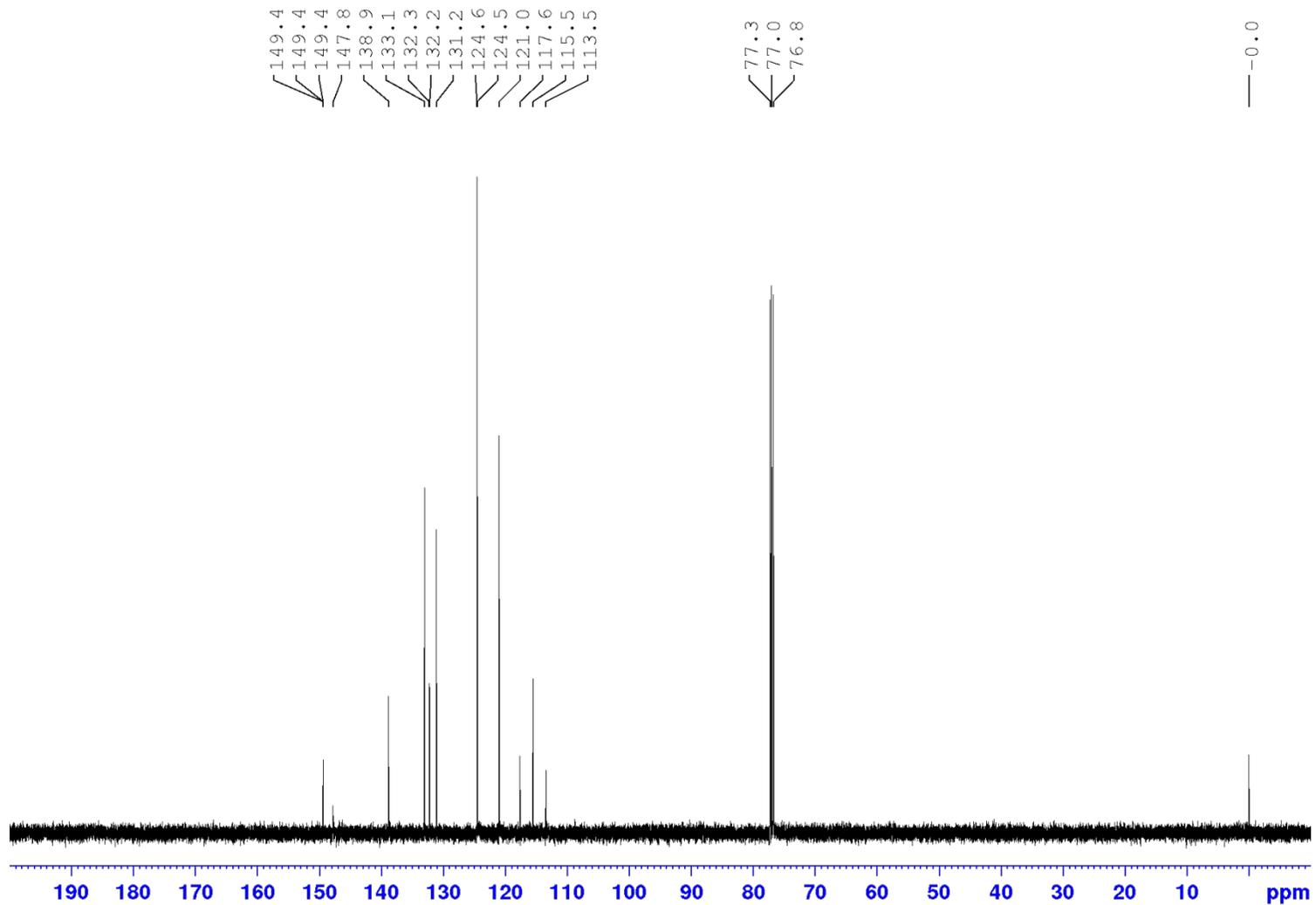


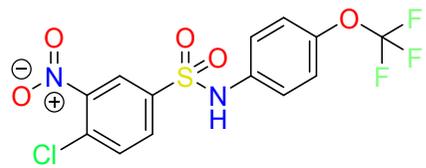
Compound 16 – ^1H NMR (500 MHz) – CDCl_3



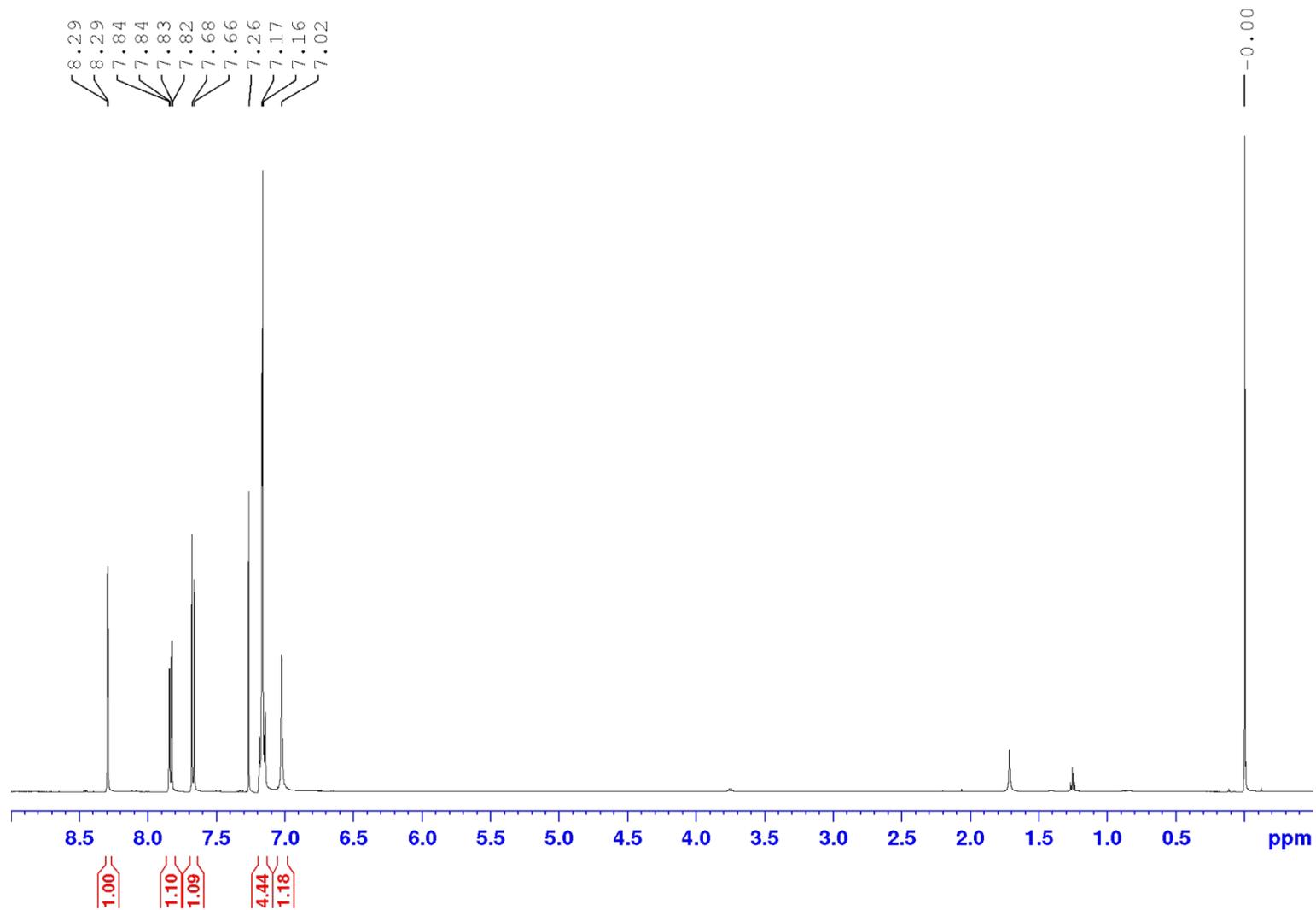


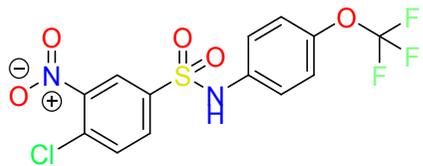
Compound **16** – ^{13}C NMR (125.5 MHz) – CDCl_3



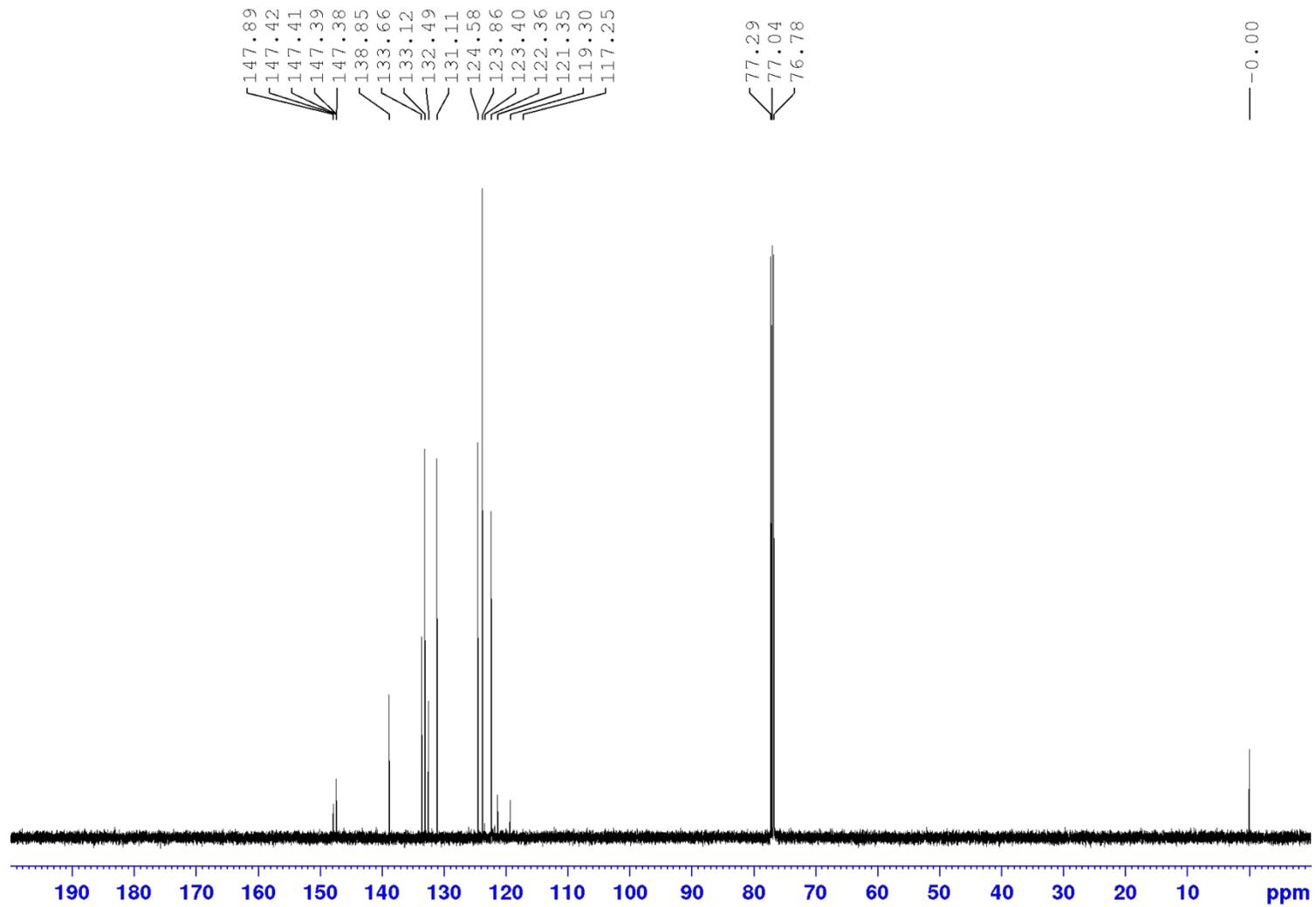


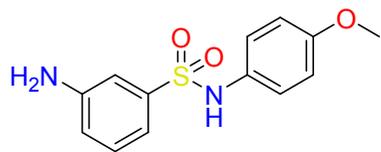
Compound 17 – ^1H NMR (500 MHz) – CDCl_3



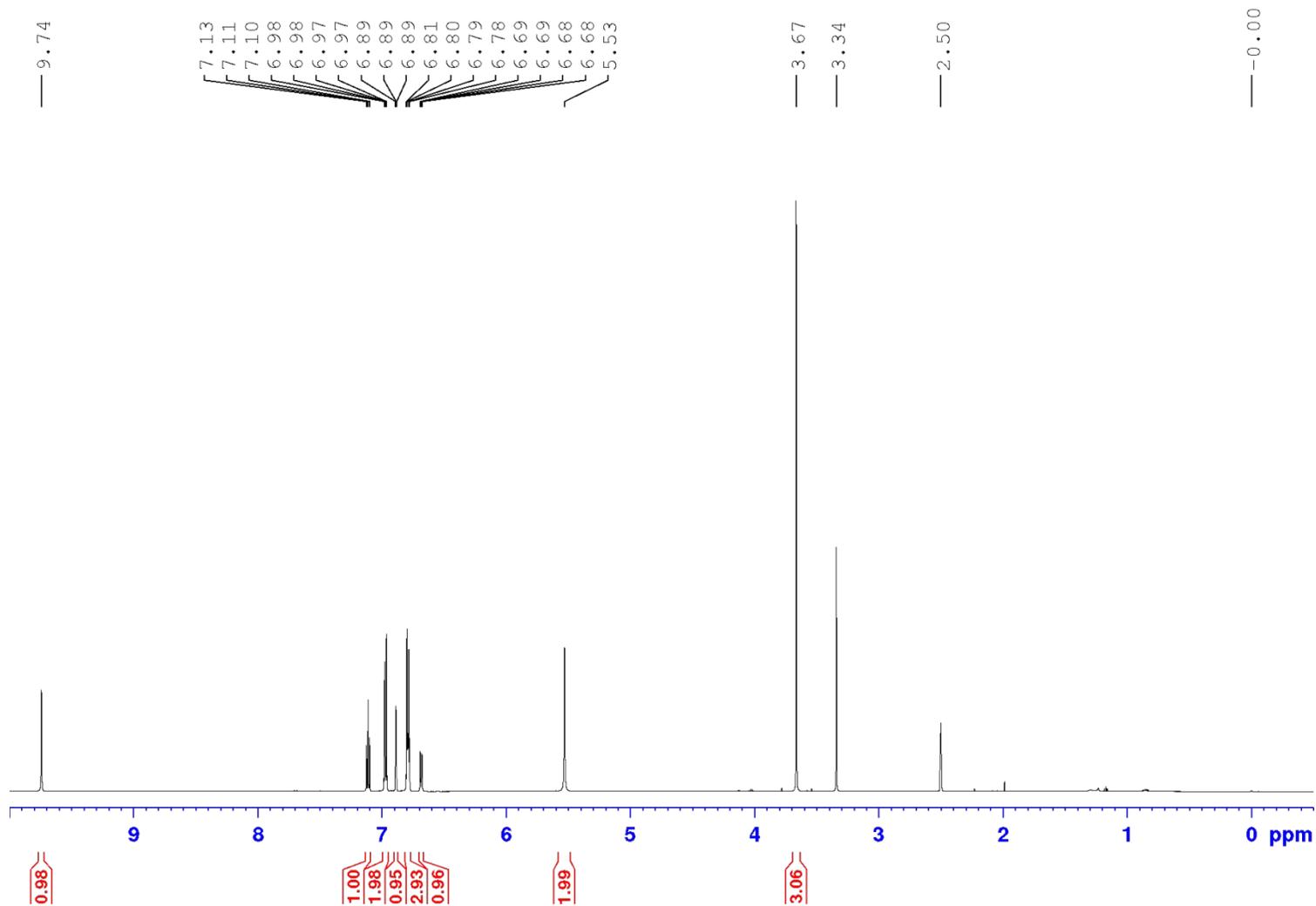


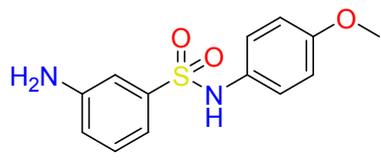
Compound **17** – ^{13}C NMR (125.5 MHz) – CDCl_3



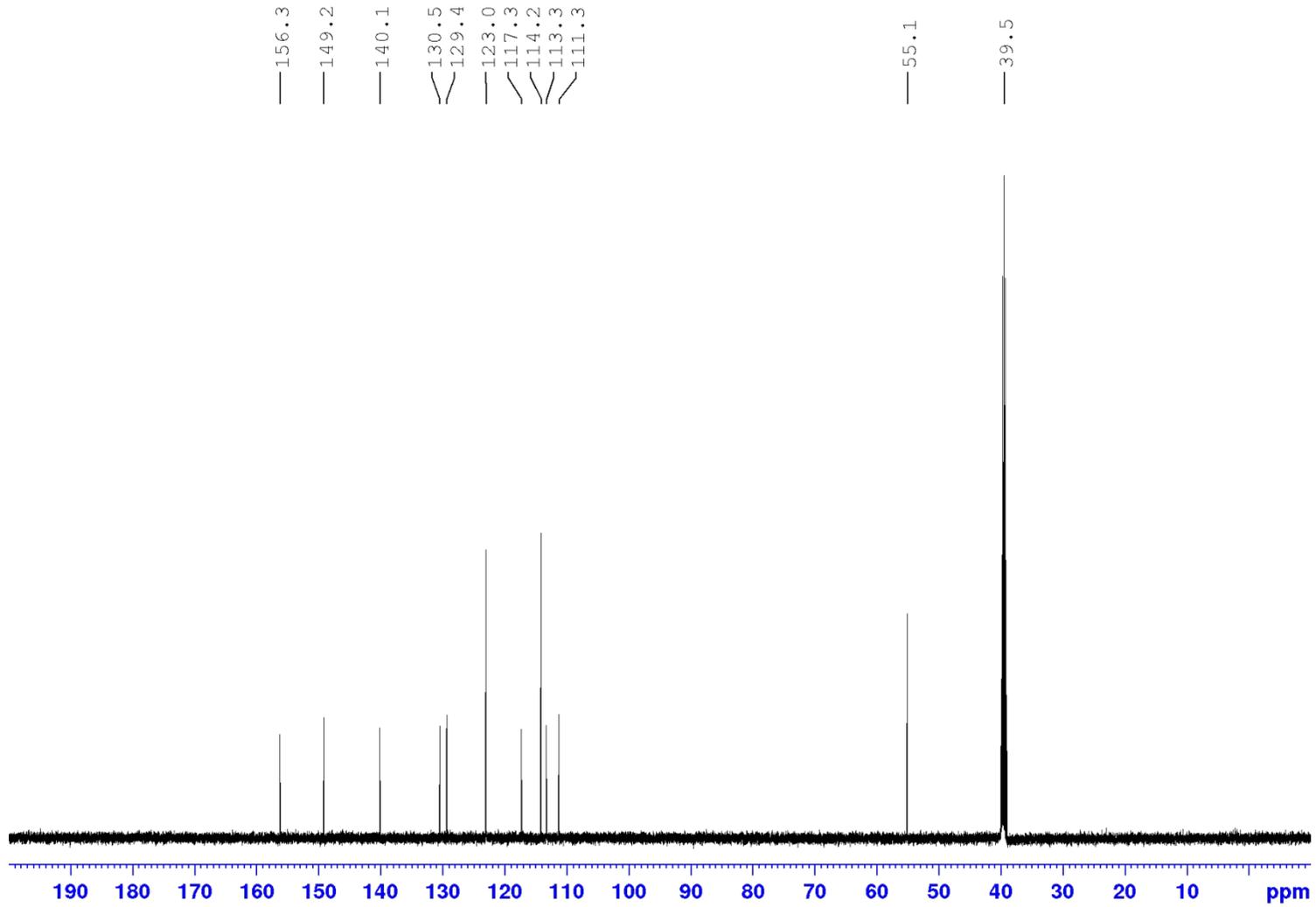


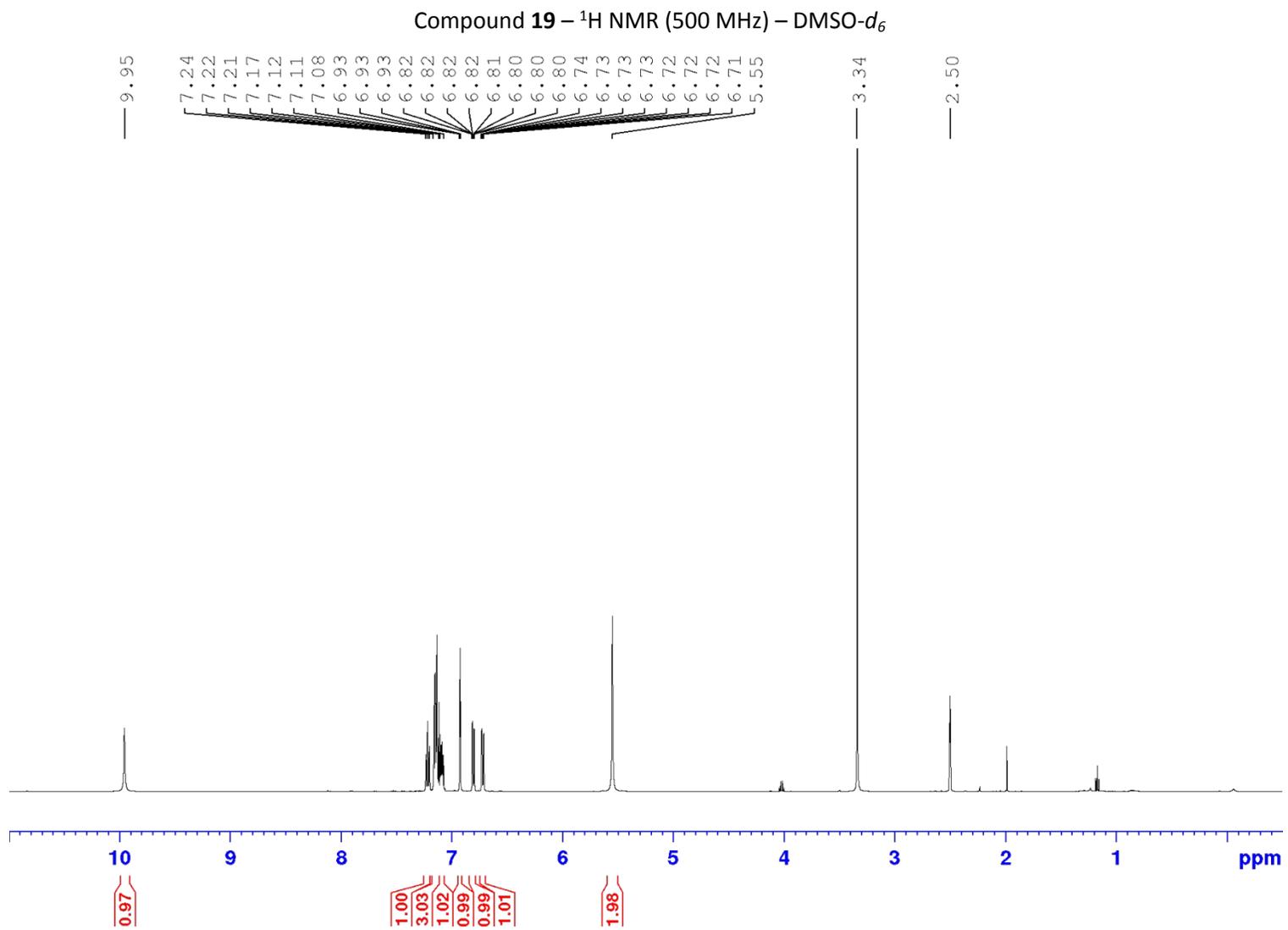
Compound **18** – ^1H NMR (600 MHz) – $\text{DMSO-}d_6$





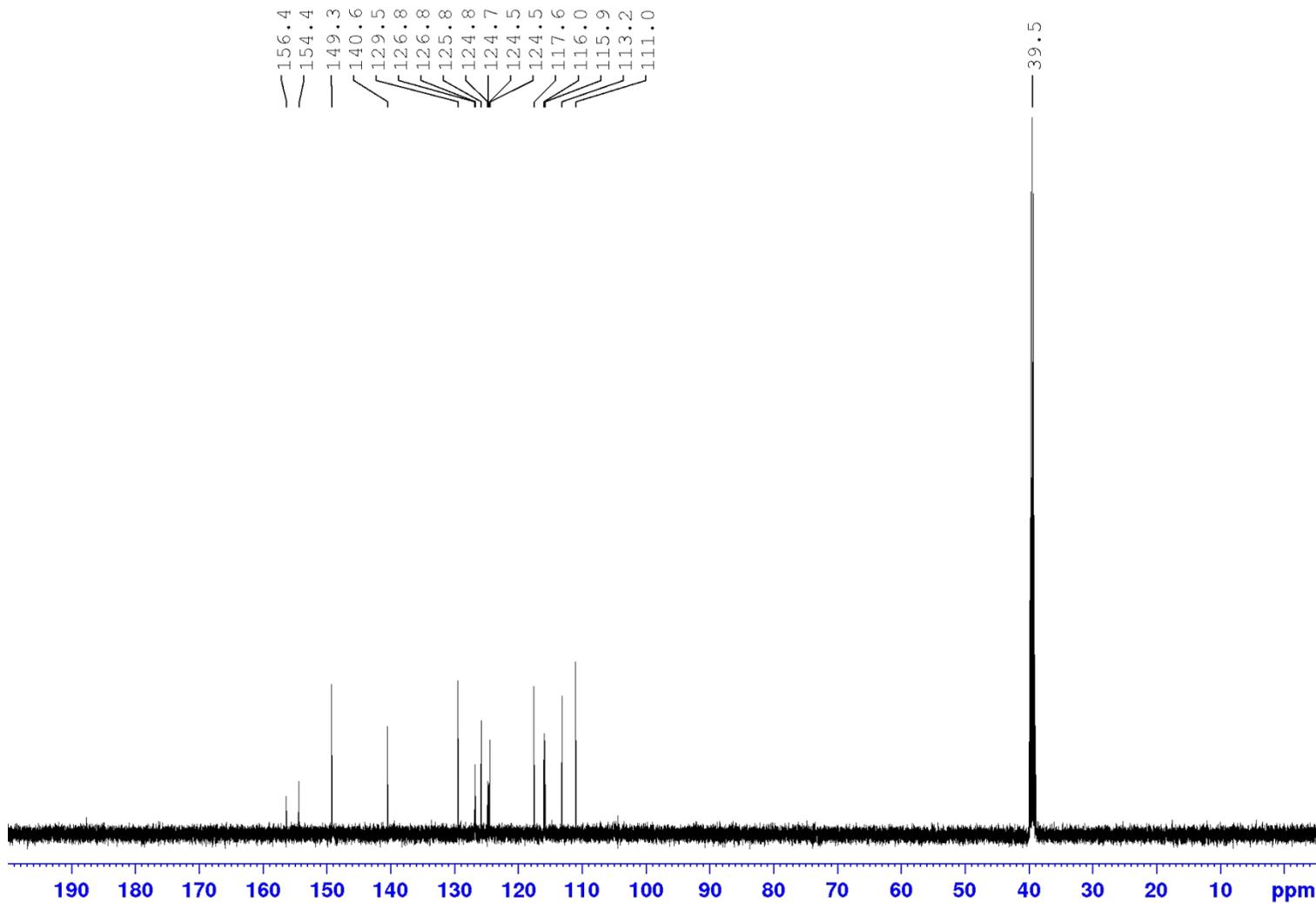
Compound **18** – ^{13}C NMR (150 MHz) – $\text{DMSO-}d_6$

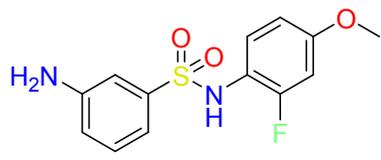




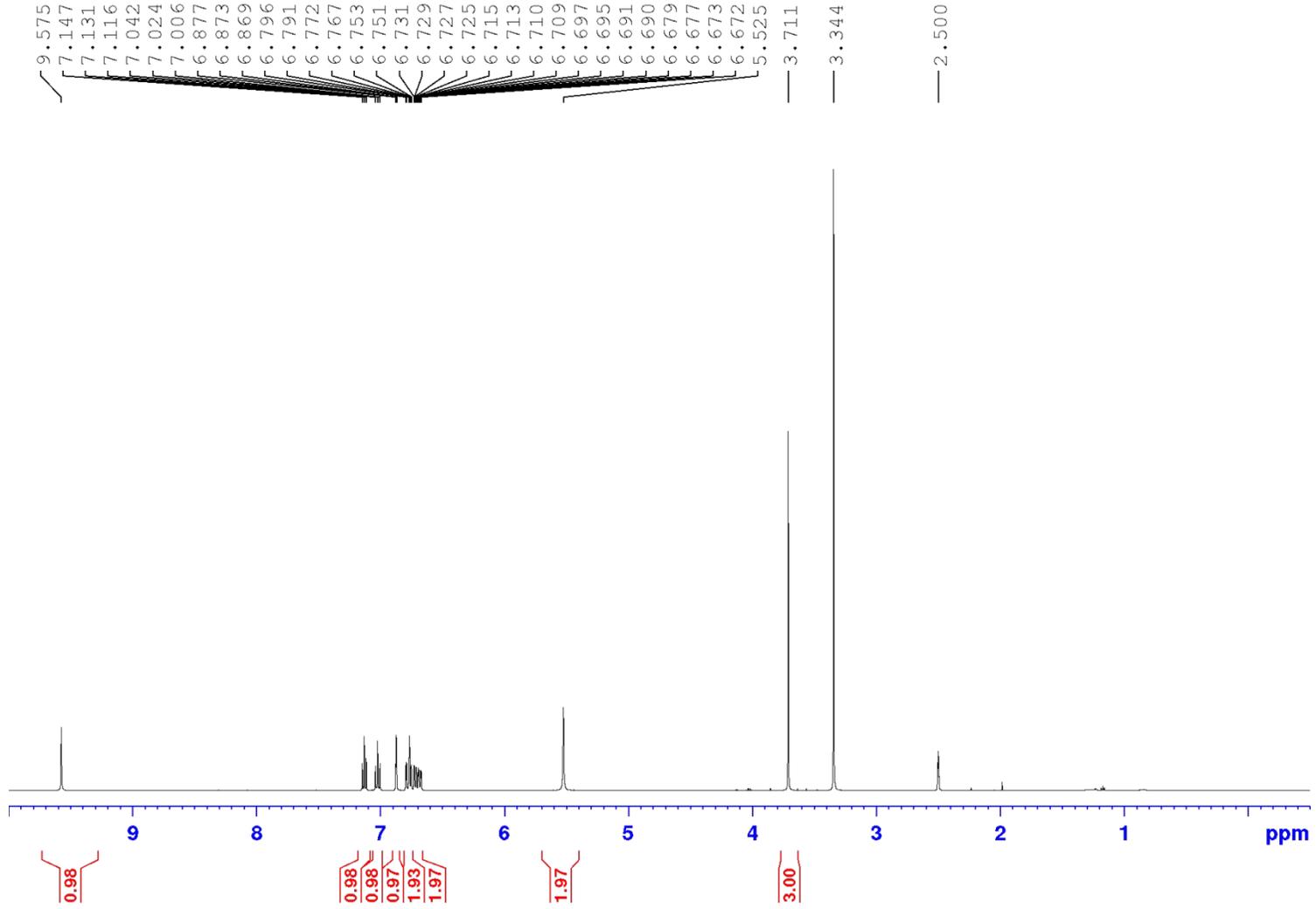


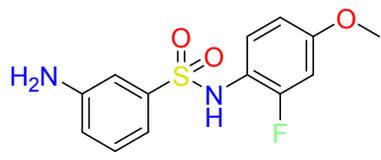
Compound 19 – ¹³C NMR (125.5 MHz) – DMSO-d₆



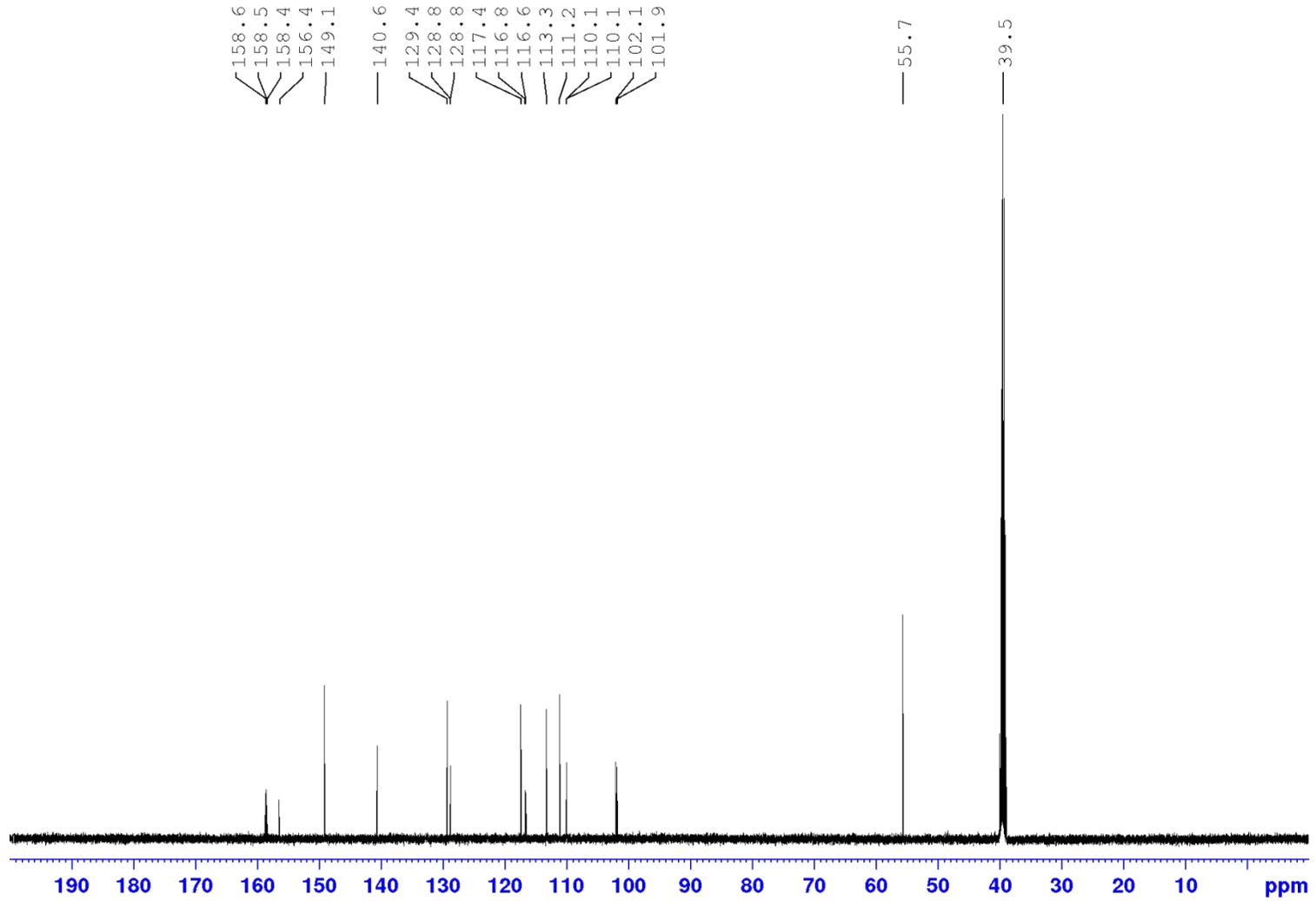


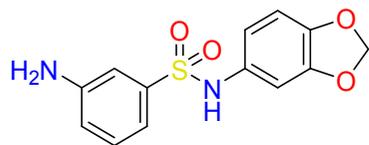
Compound 20 – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



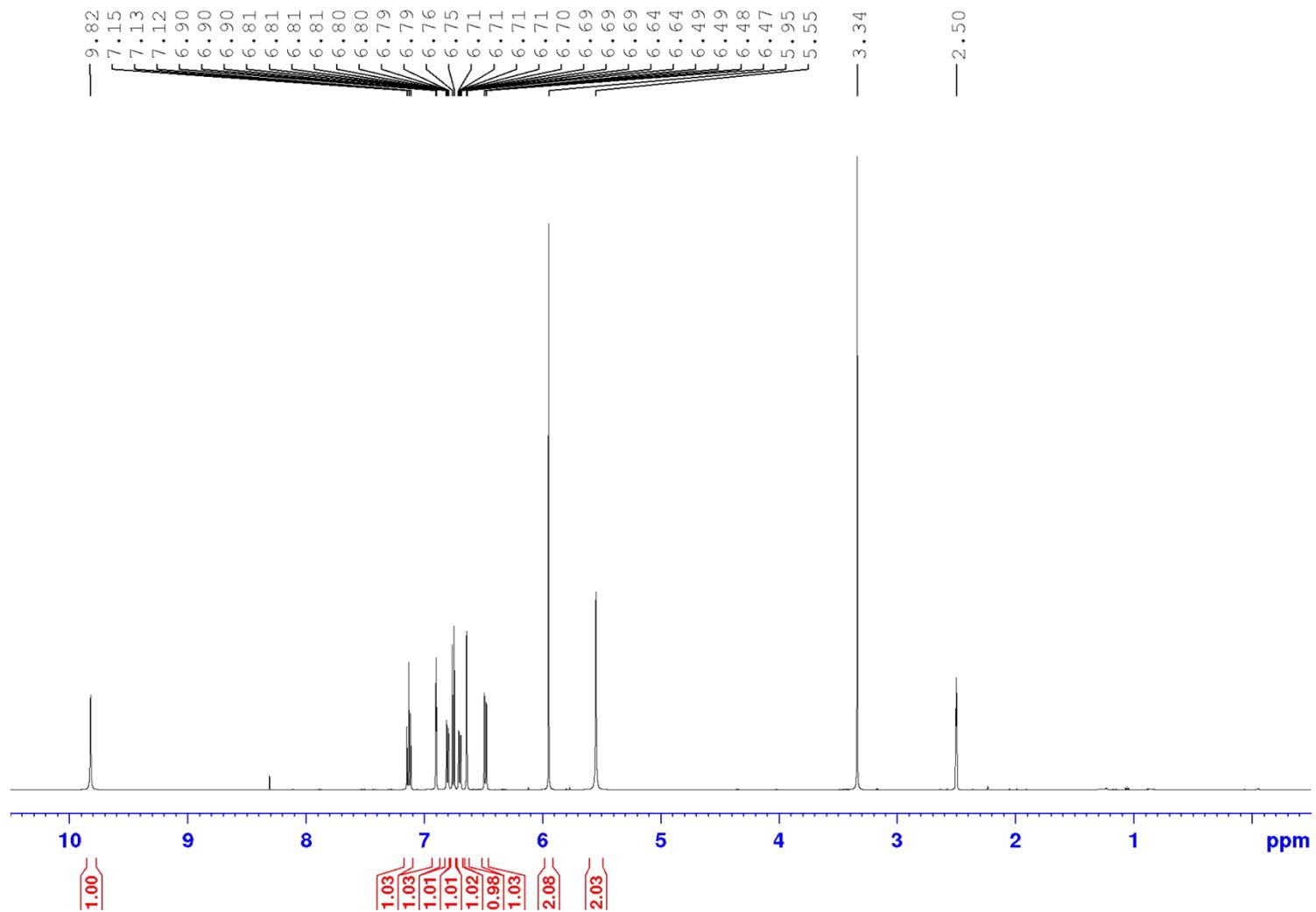


Compound 20 – ^{13}C NMR (125.5 MHz) – $\text{DMSO-}d_6$



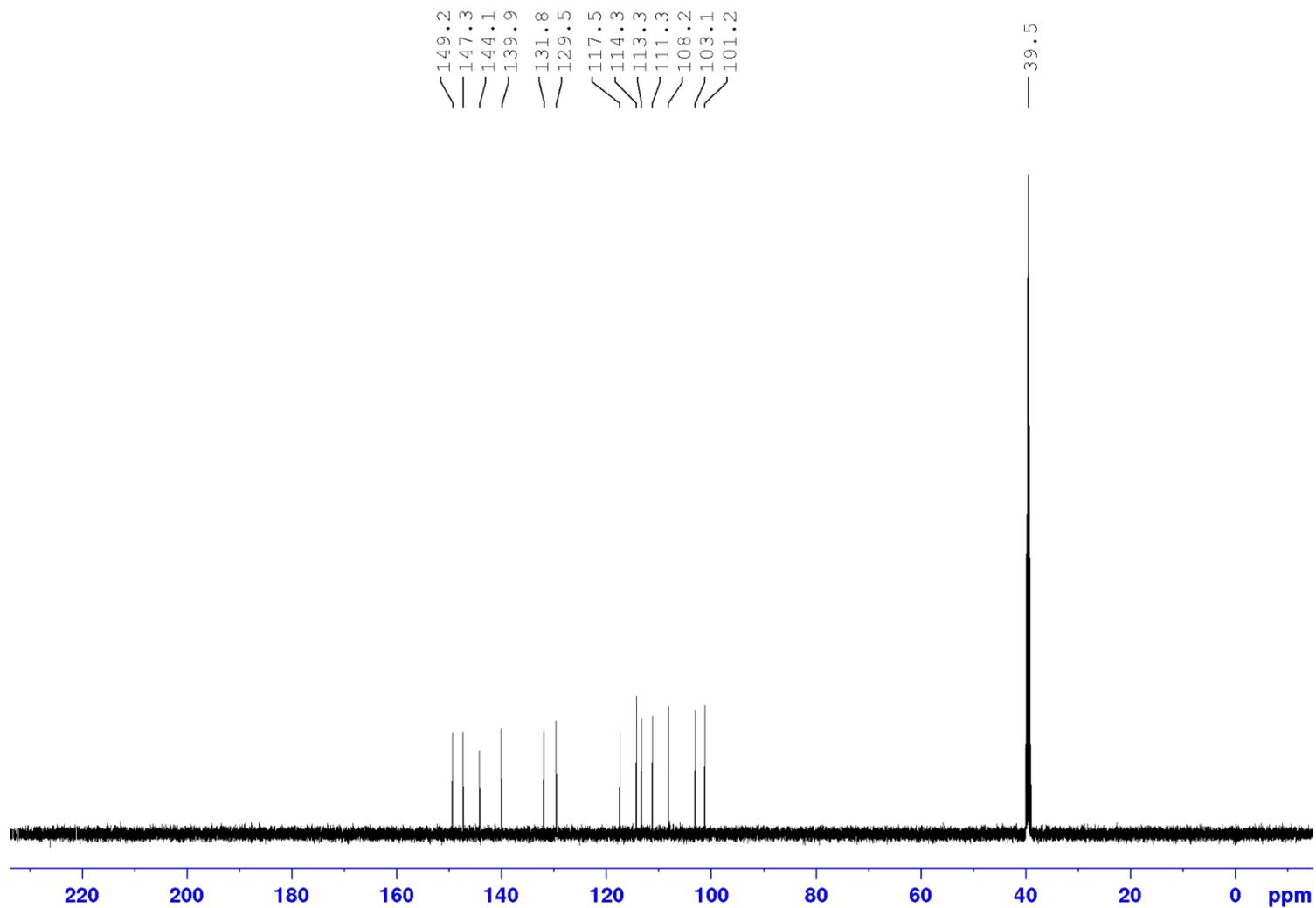


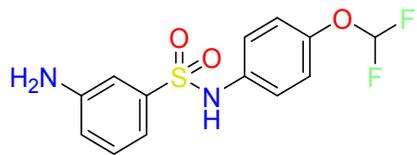
Compound 21 – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



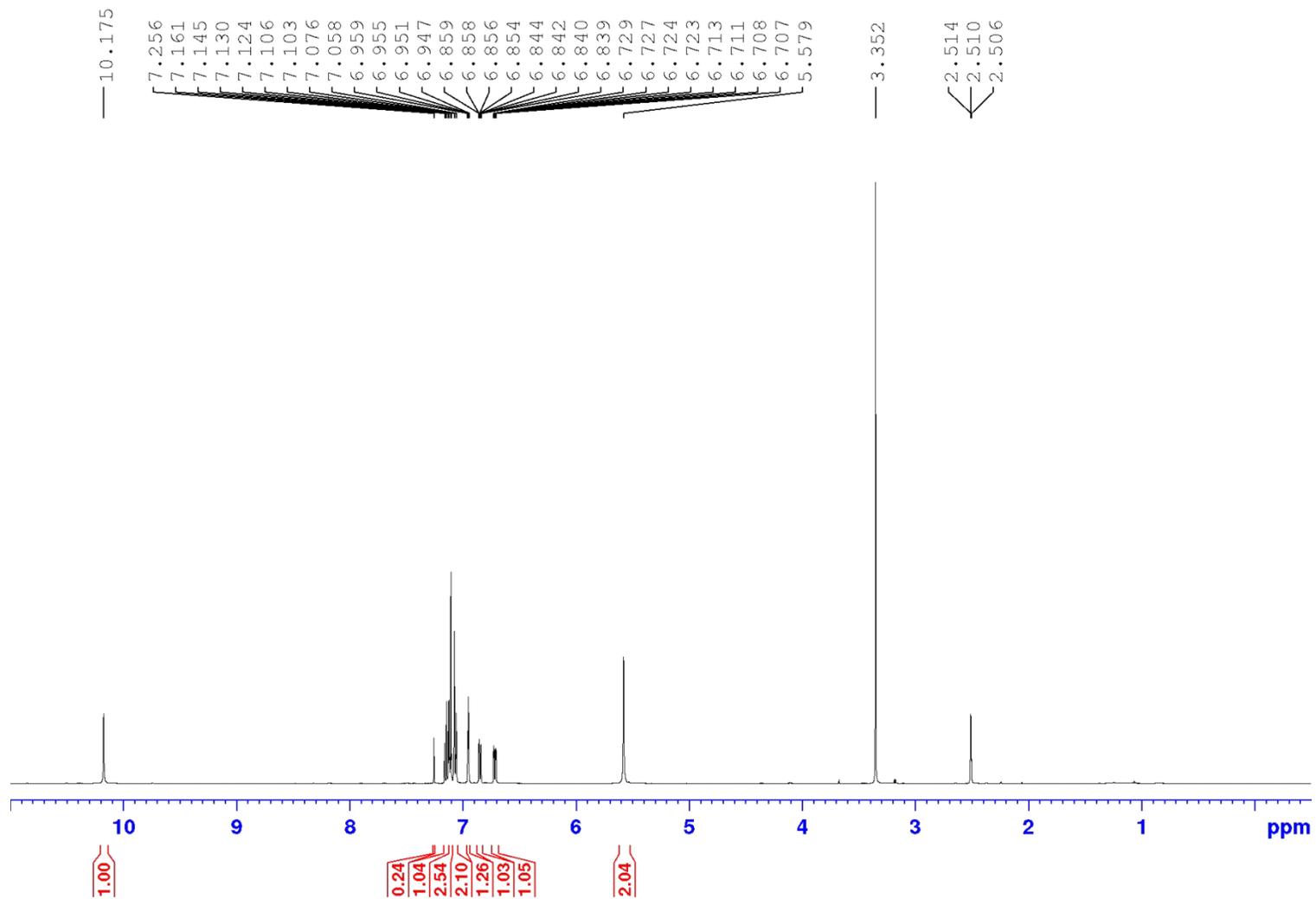


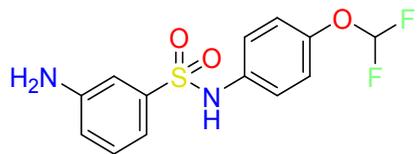
Compound **21** – ^{13}C NMR (125.5 MHz) – $\text{DMSO-}d_6$



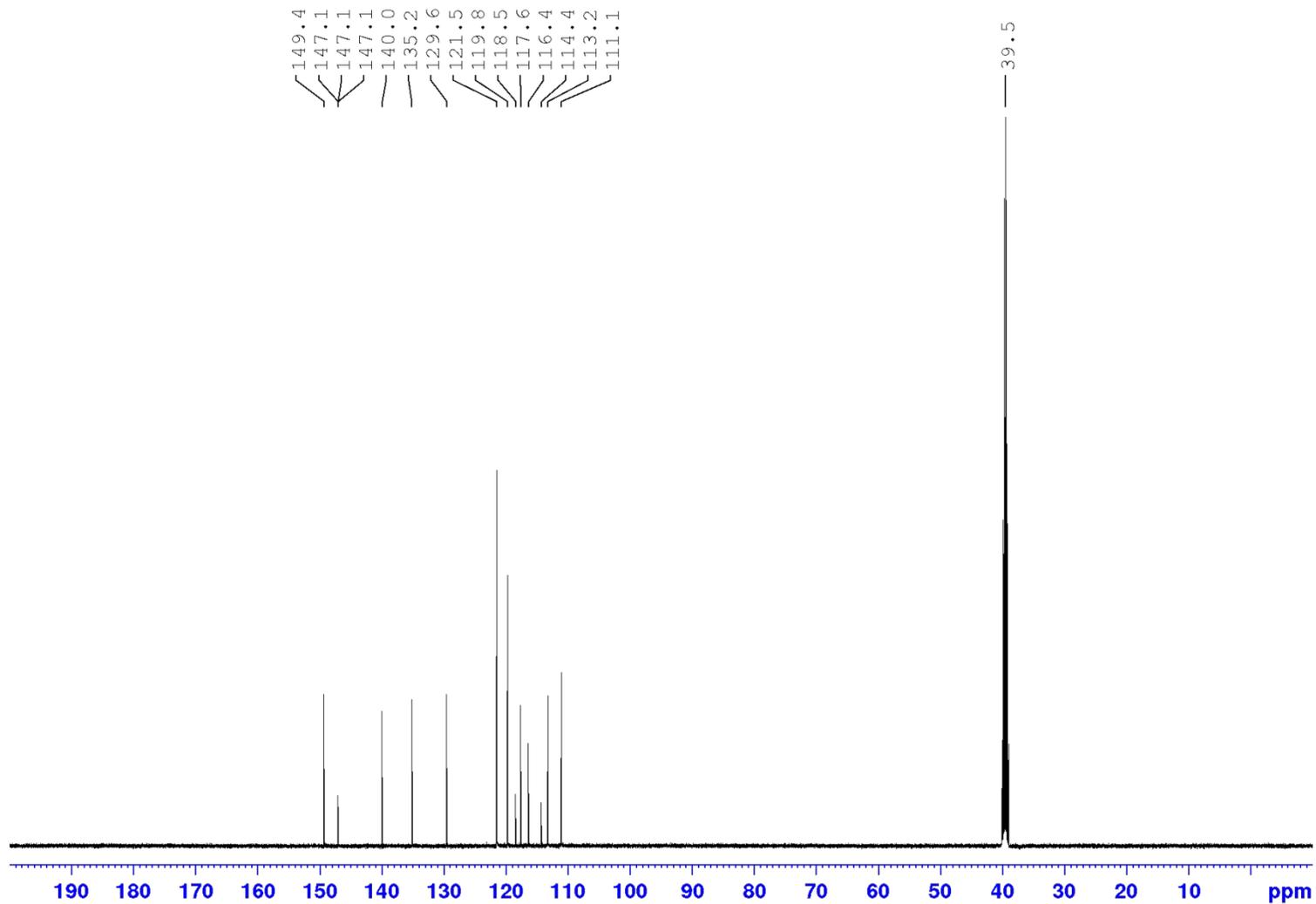


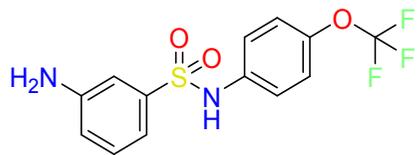
Compound 22 – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



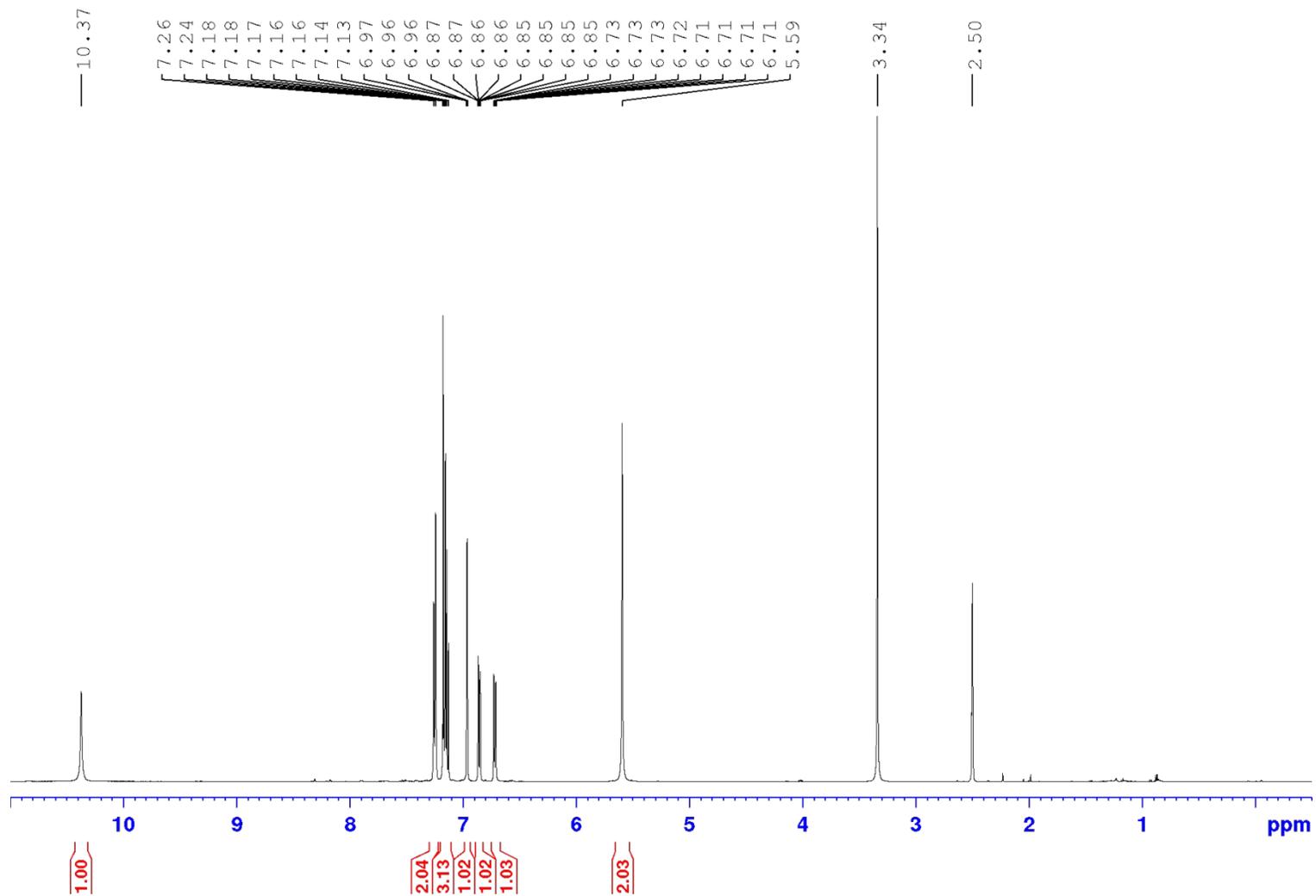


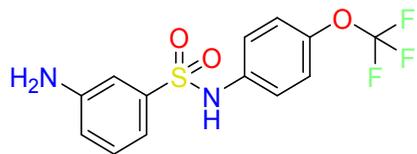
Compound **22** – ^{13}C NMR (125.5 MHz) – $\text{DMSO-}d_6$



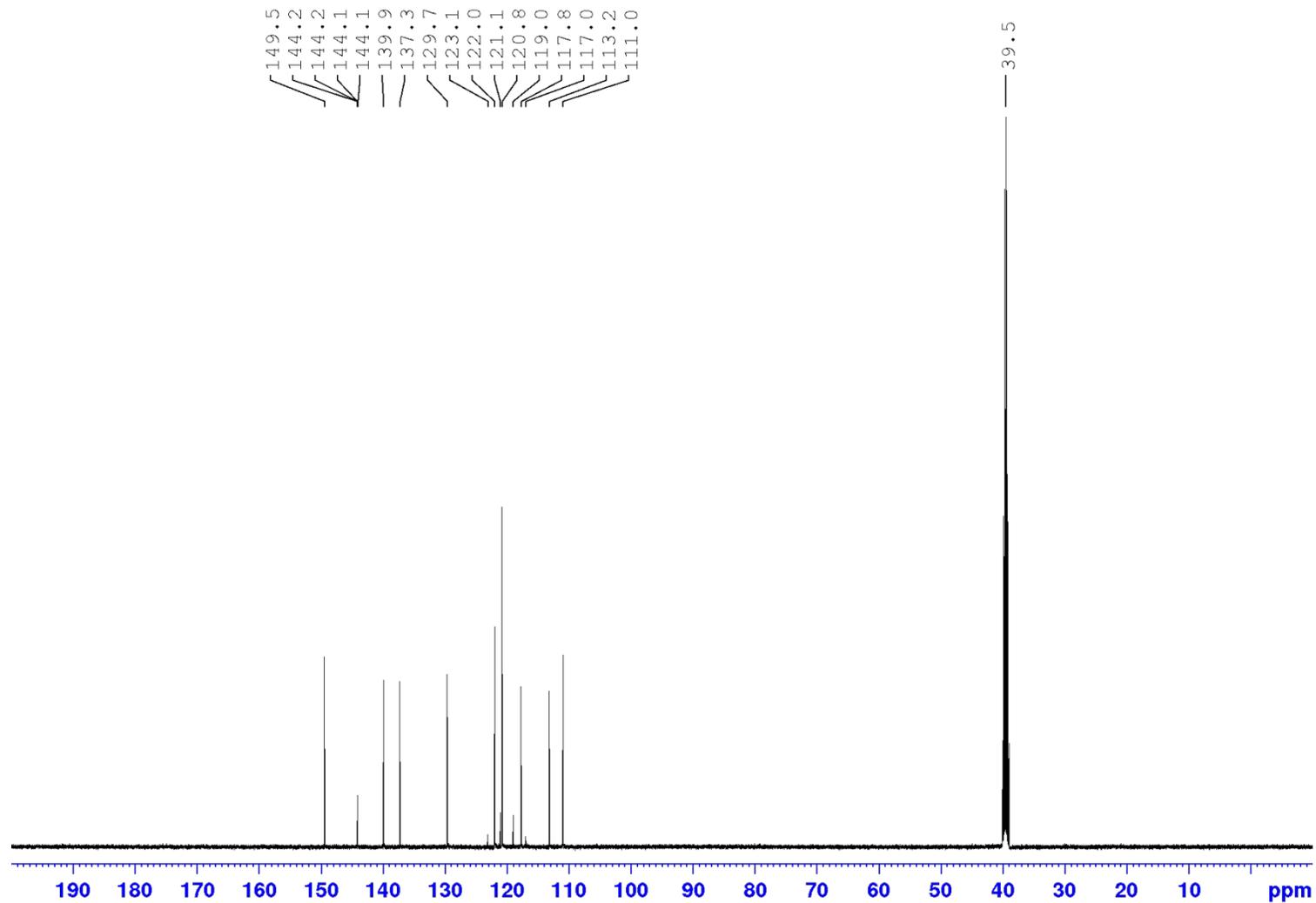


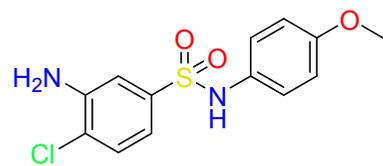
Compound **23** – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



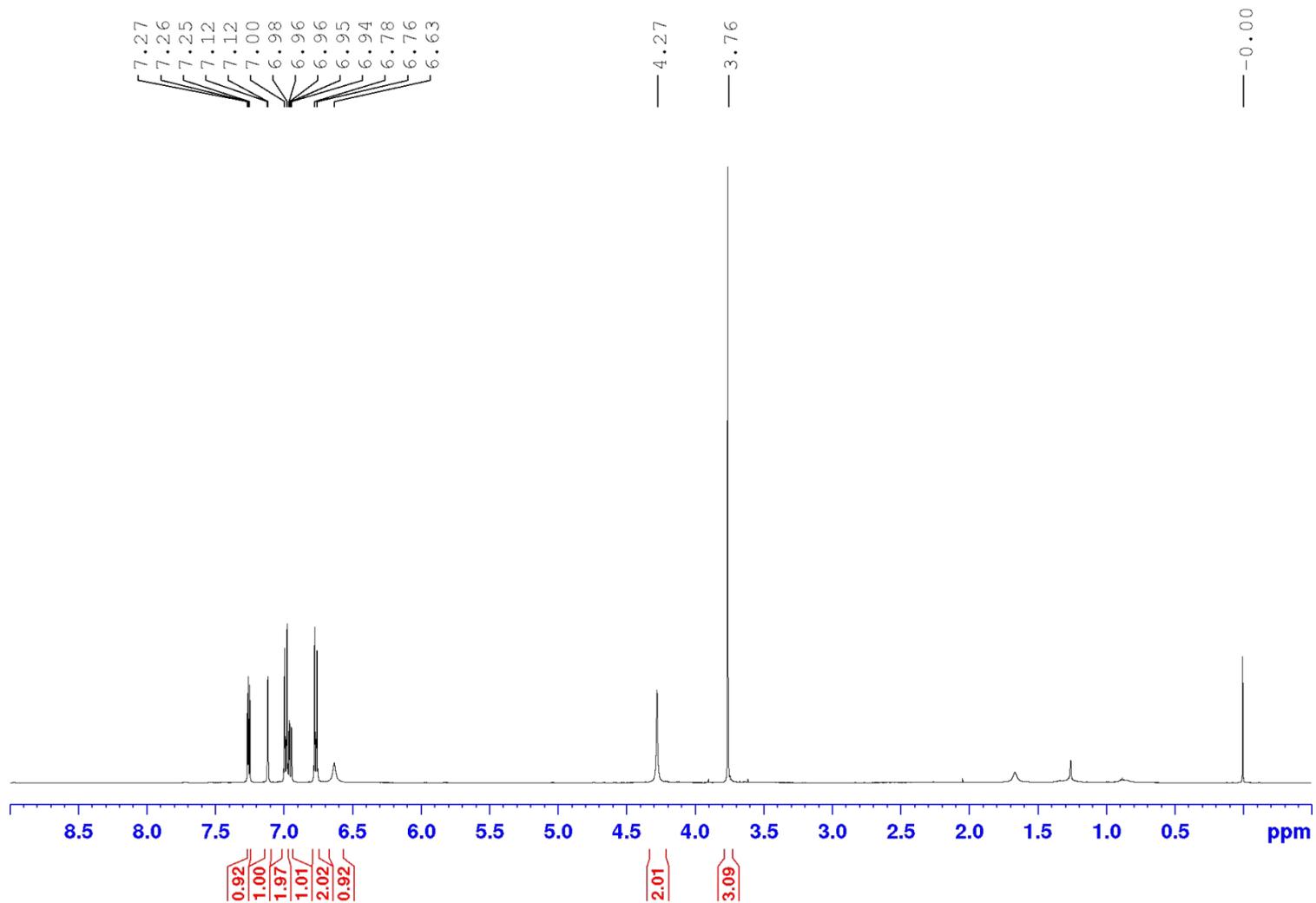


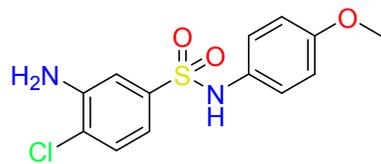
Compound **23** – ^{13}C NMR (125.5 MHz) – $\text{DMSO-}d_6$



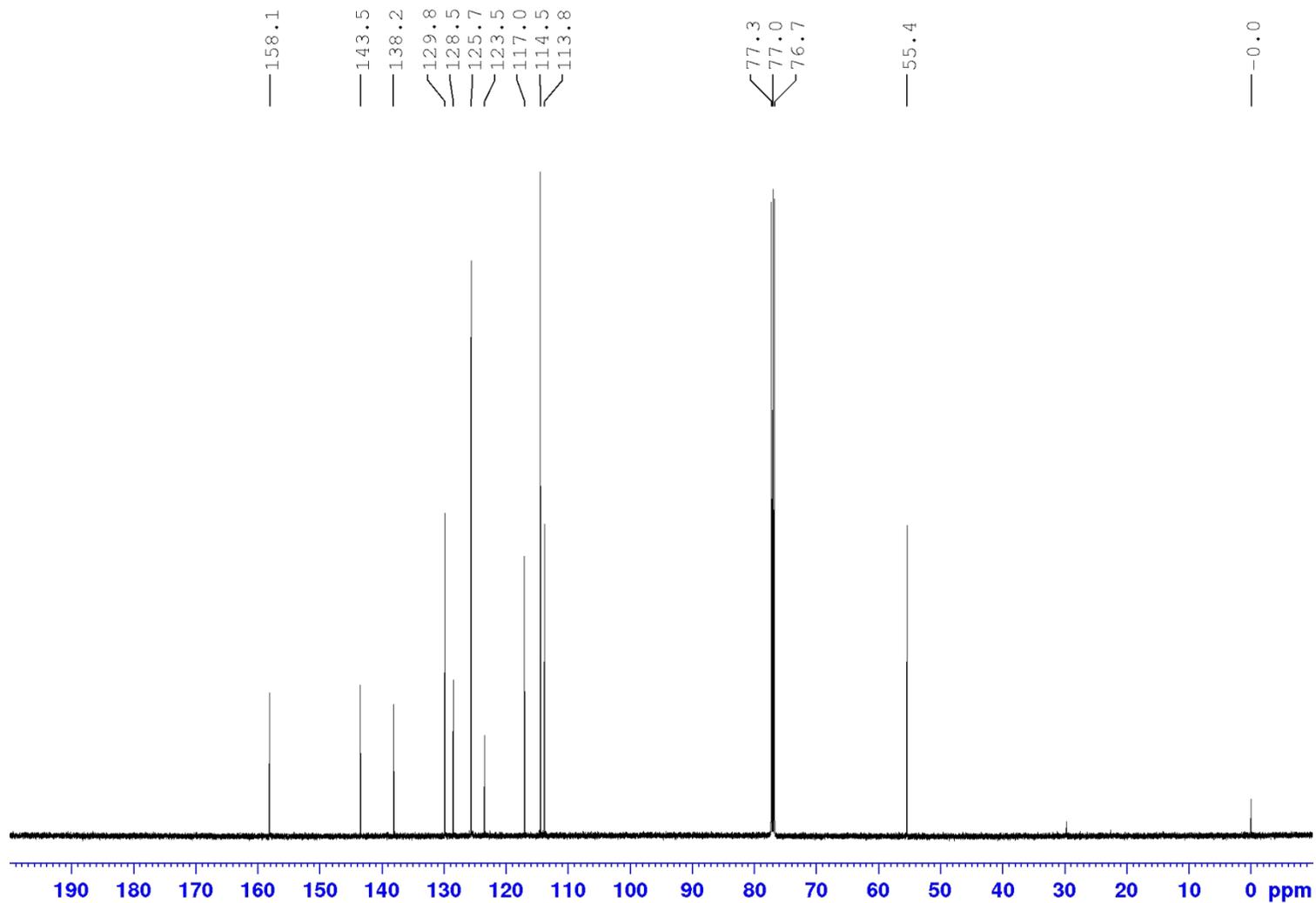


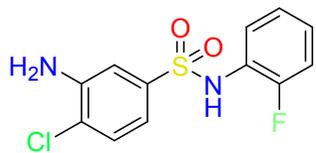
Compound 24 – ¹H NMR (500 MHz) – CDCl₃



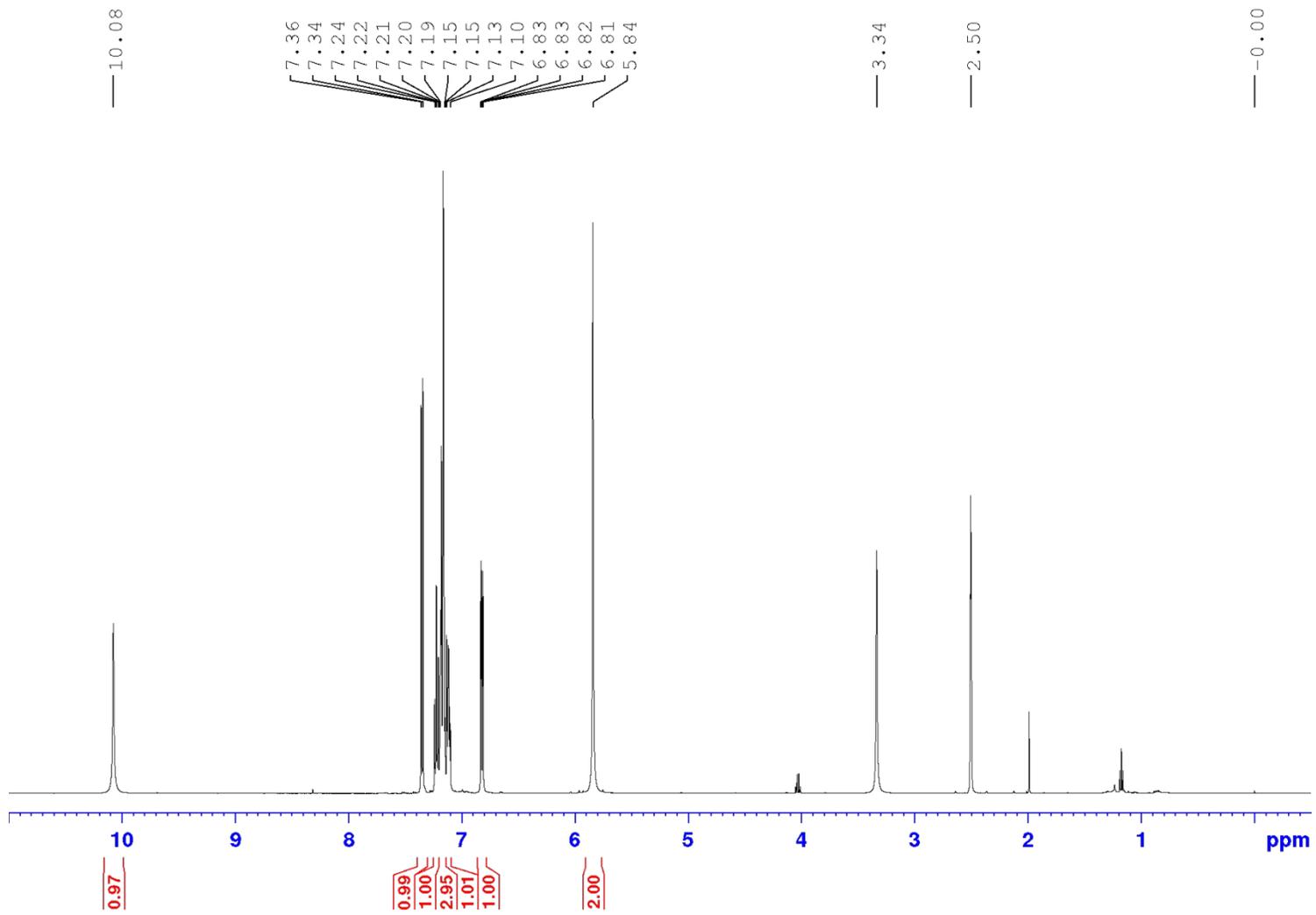


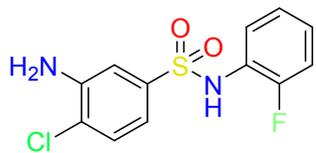
Compound **24** – ^{13}C NMR (125.5 MHz) – CDCl_3



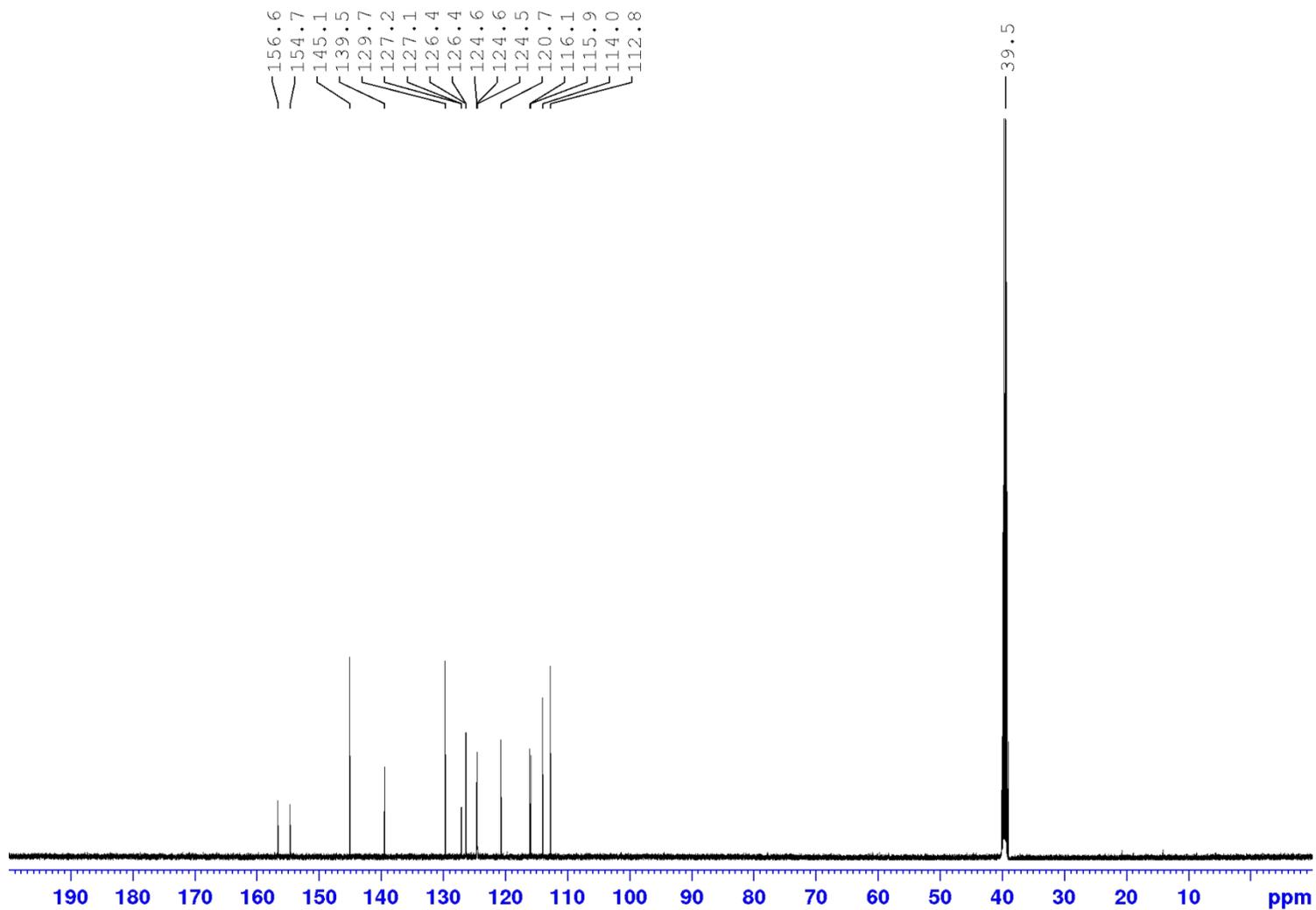


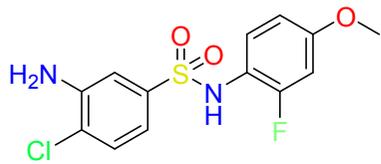
Compound 25 – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



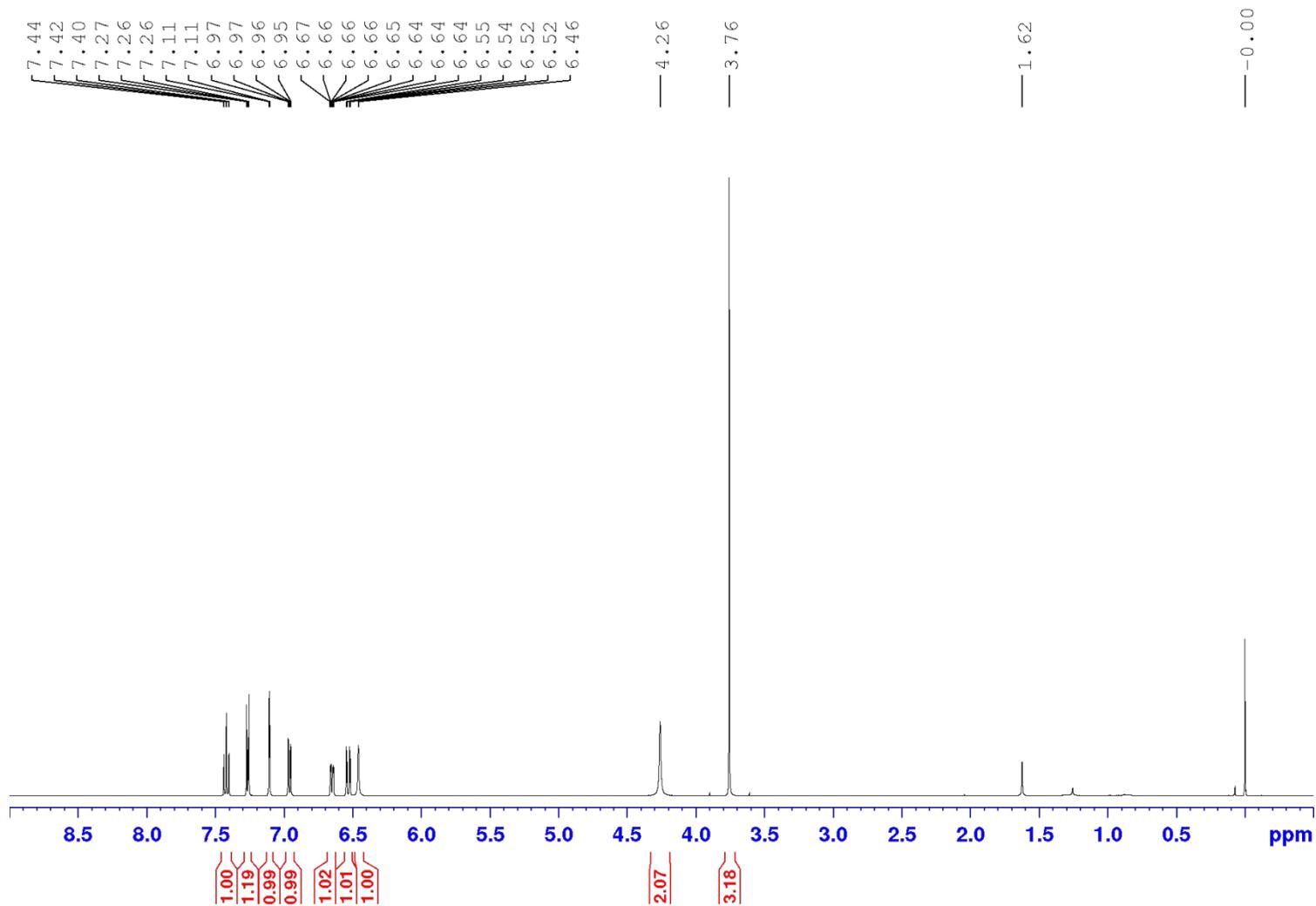


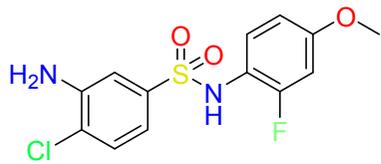
Compound 25 – ^{13}C NMR (125.5 MHz) – $\text{DMSO-}d_6$



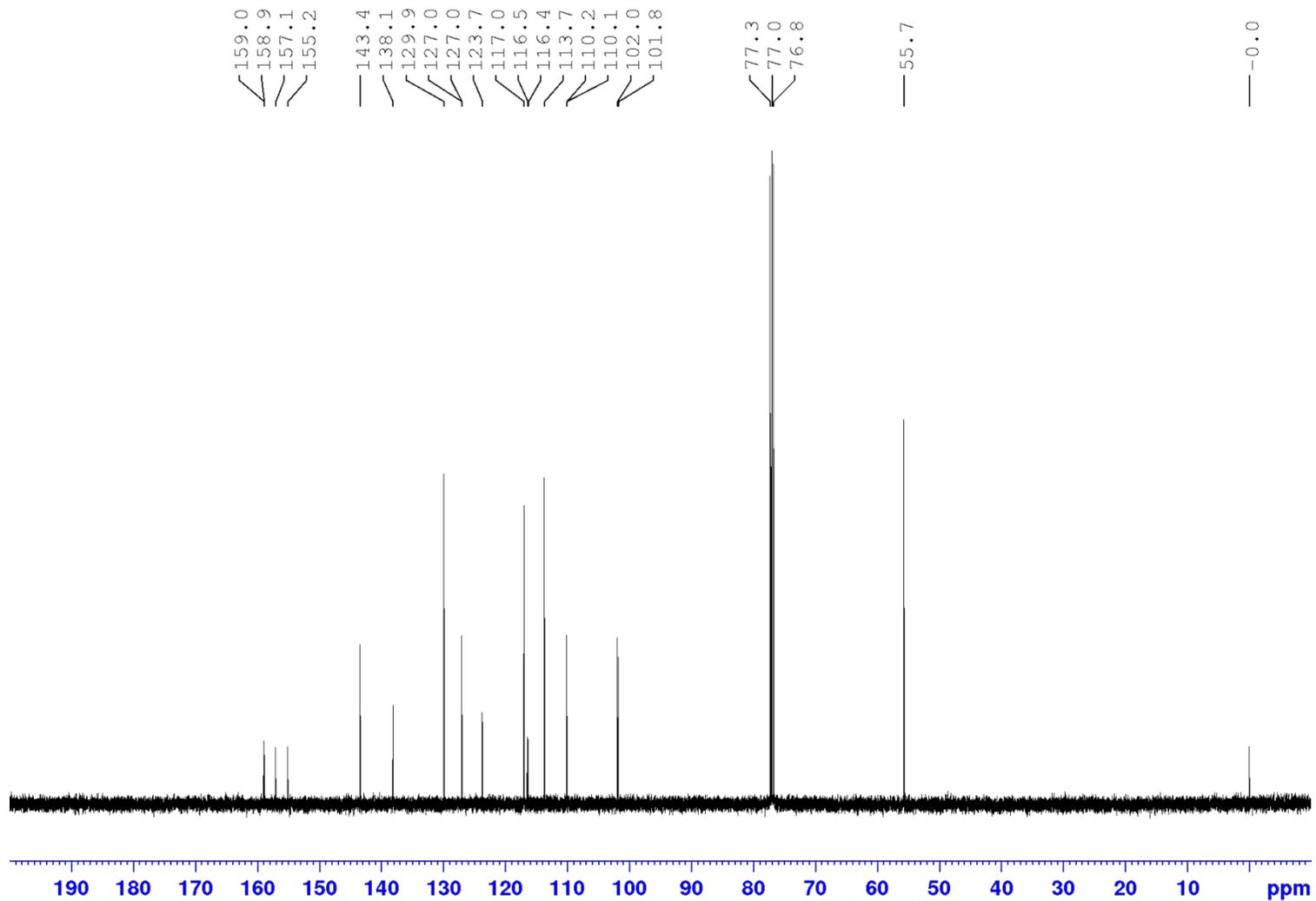


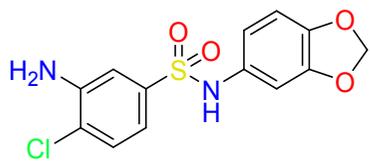
Compound 26 – ^1H NMR (500 MHz) – CDCl_3



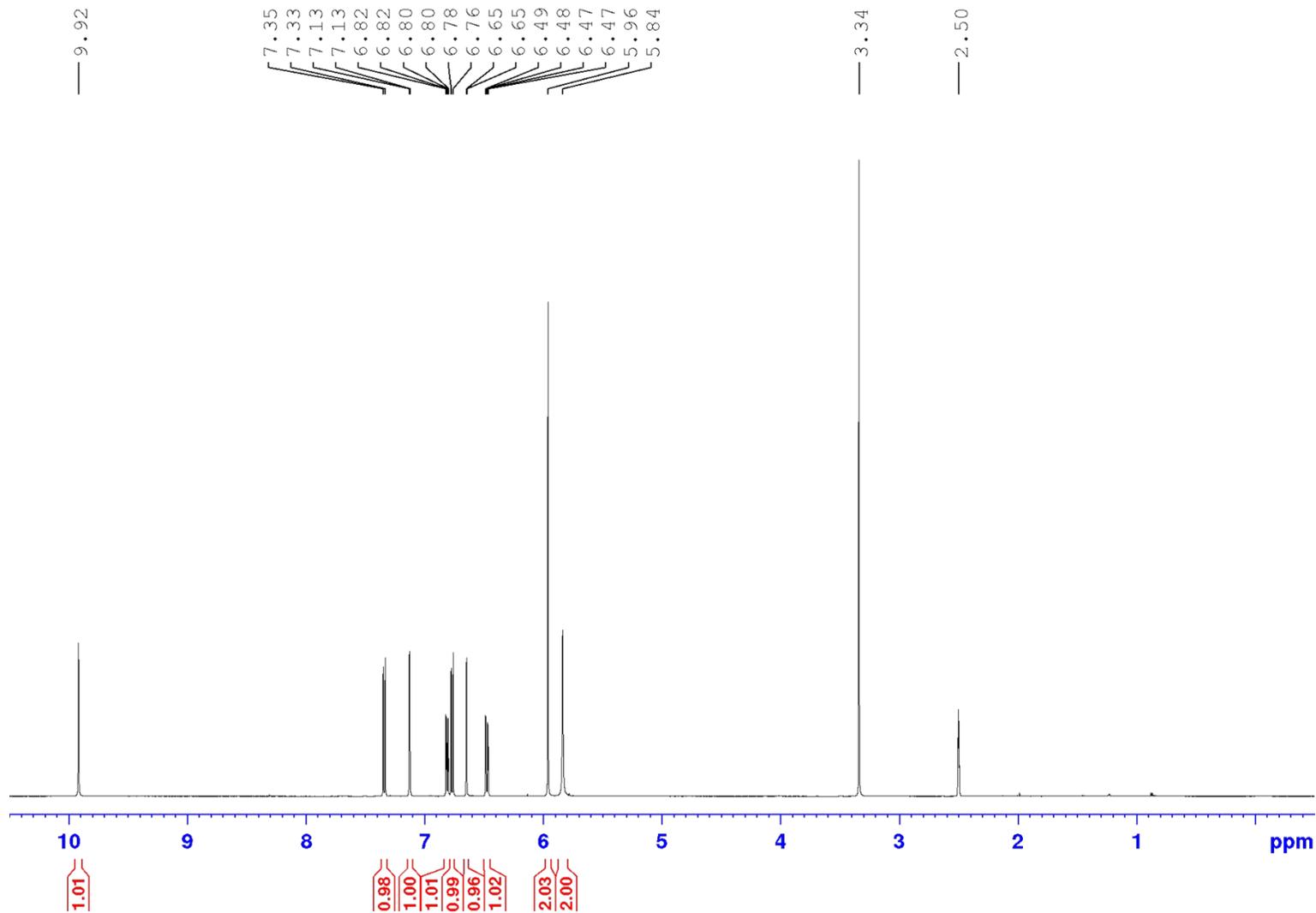


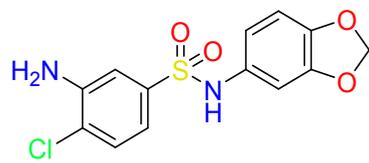
Compound 26 – ¹³C NMR (125.5 MHz) – CDCl₃



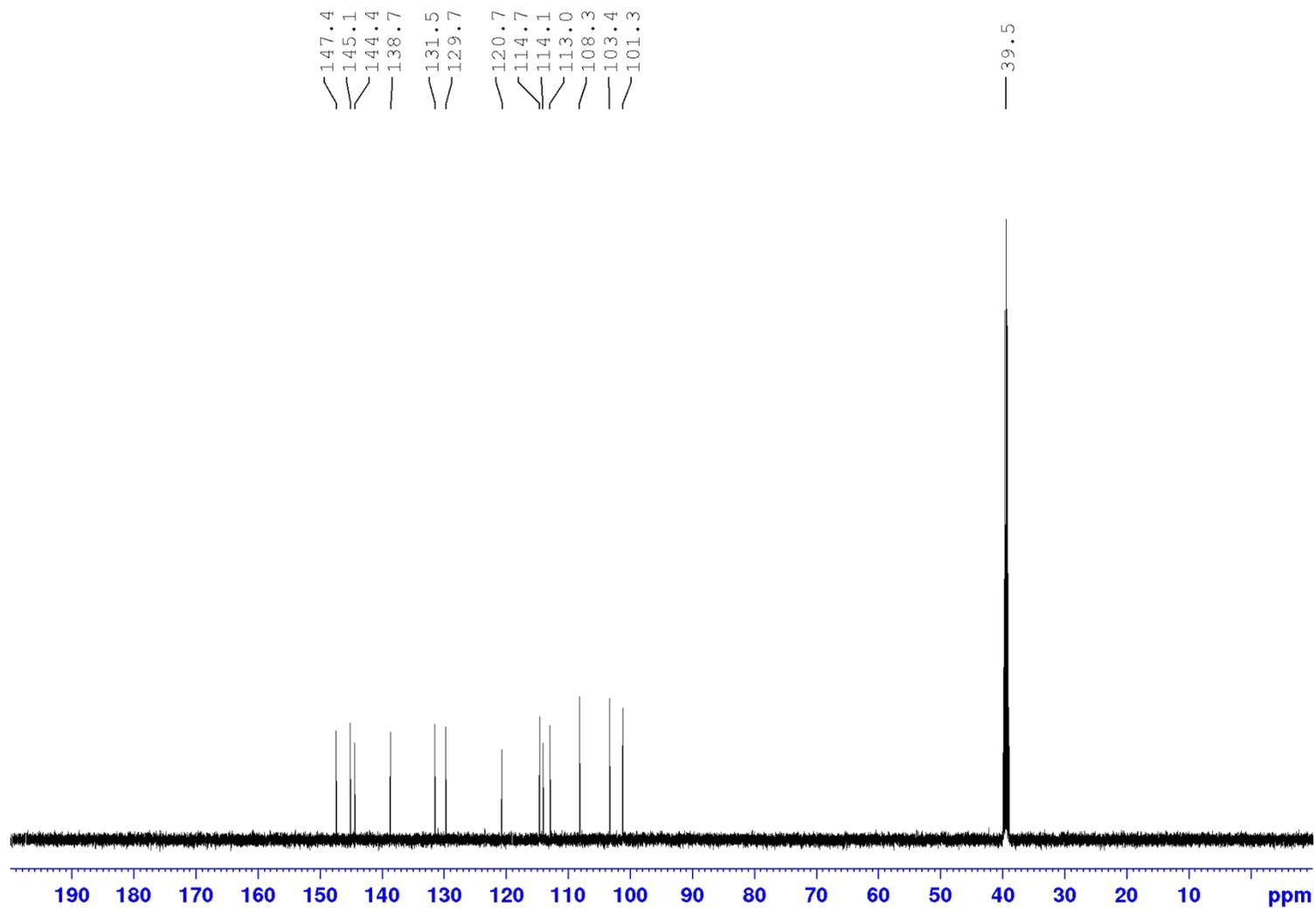


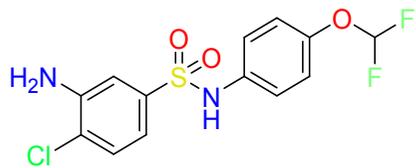
Compound 27 – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



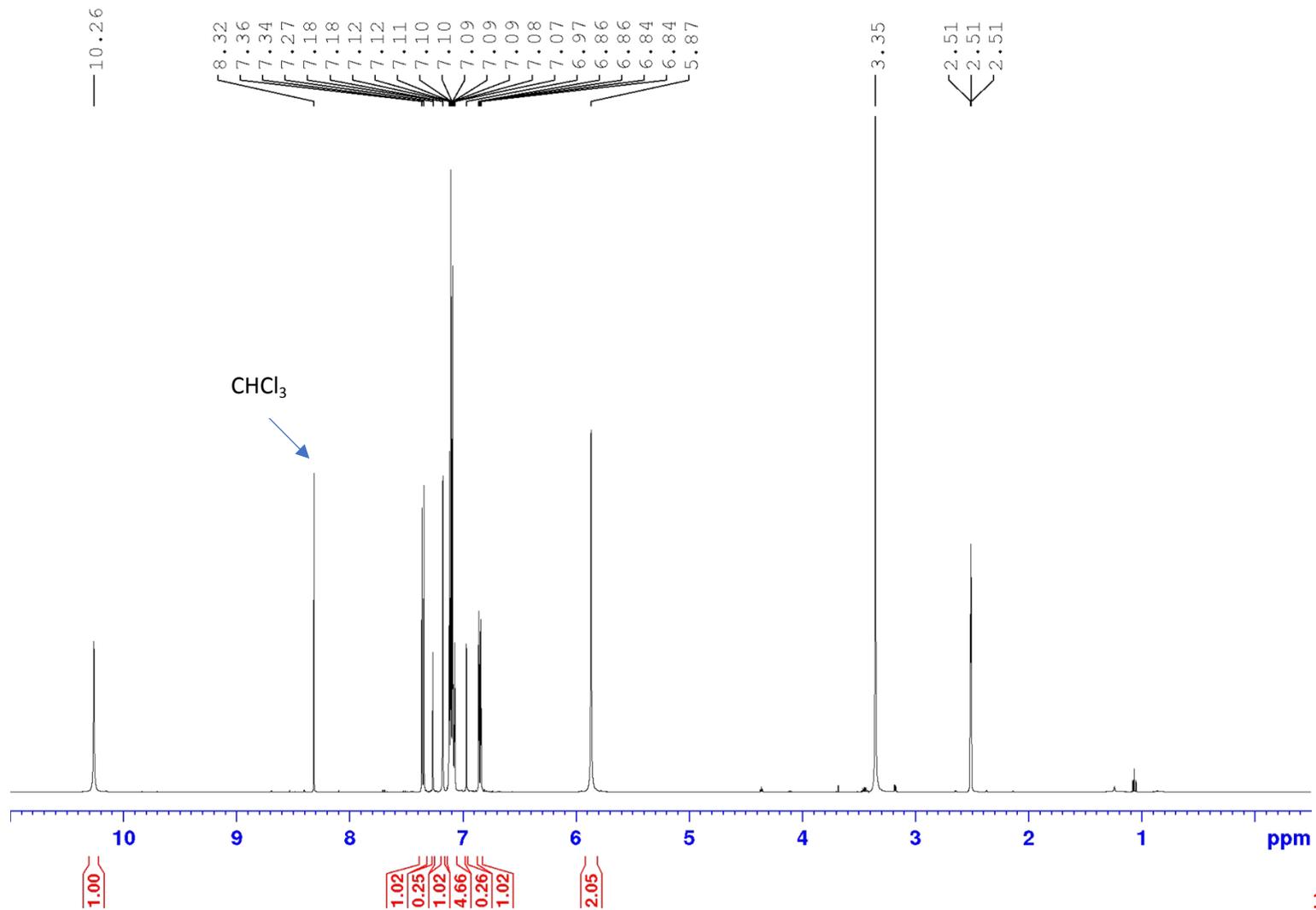


Compound 27 – ^{13}C NMR (125.5 MHz) – $\text{DMSO-}d_6$

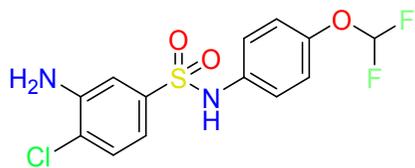




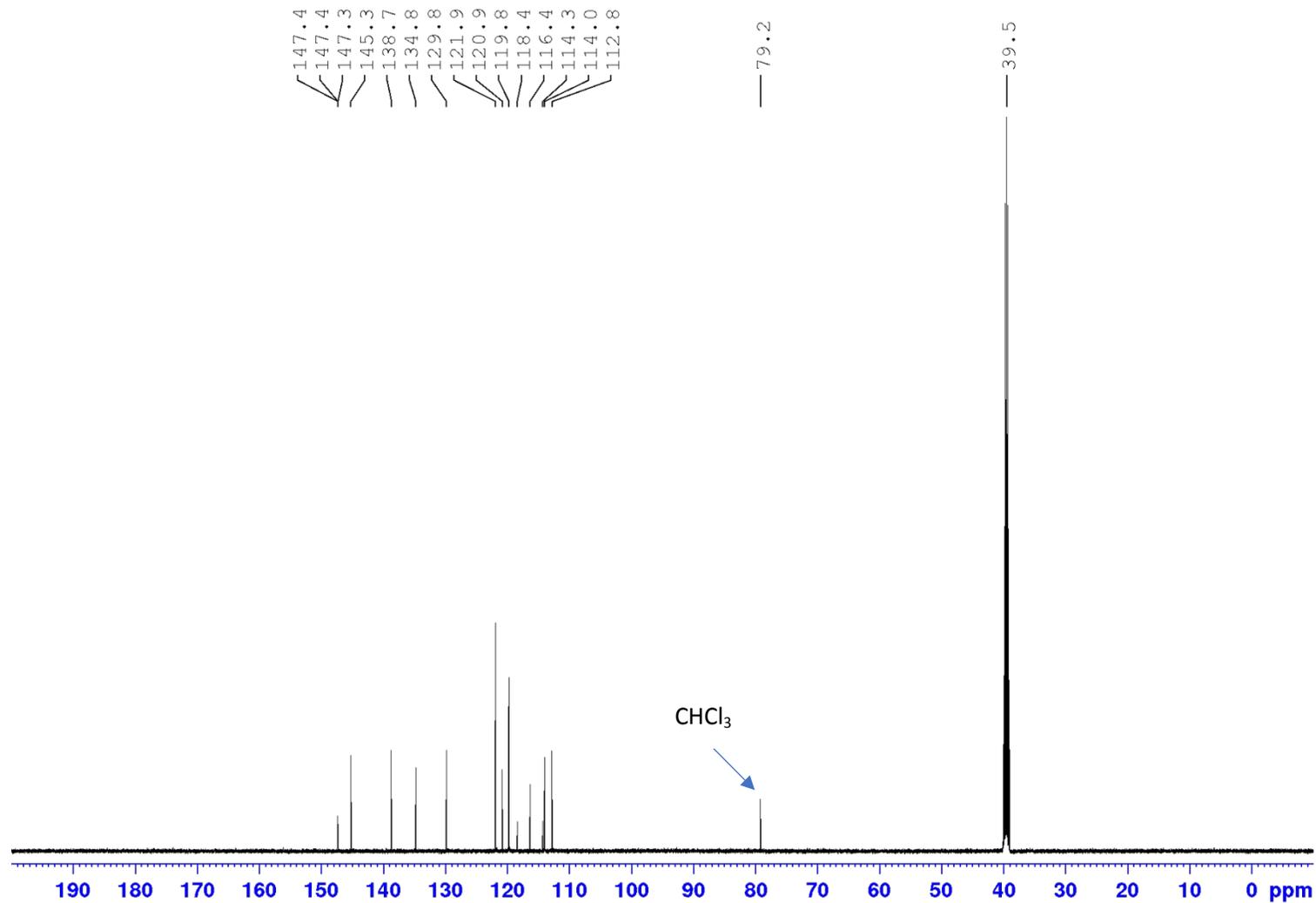
Compound **28** – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$

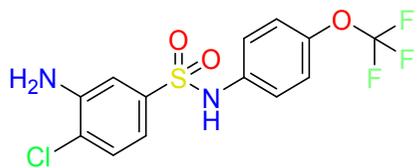


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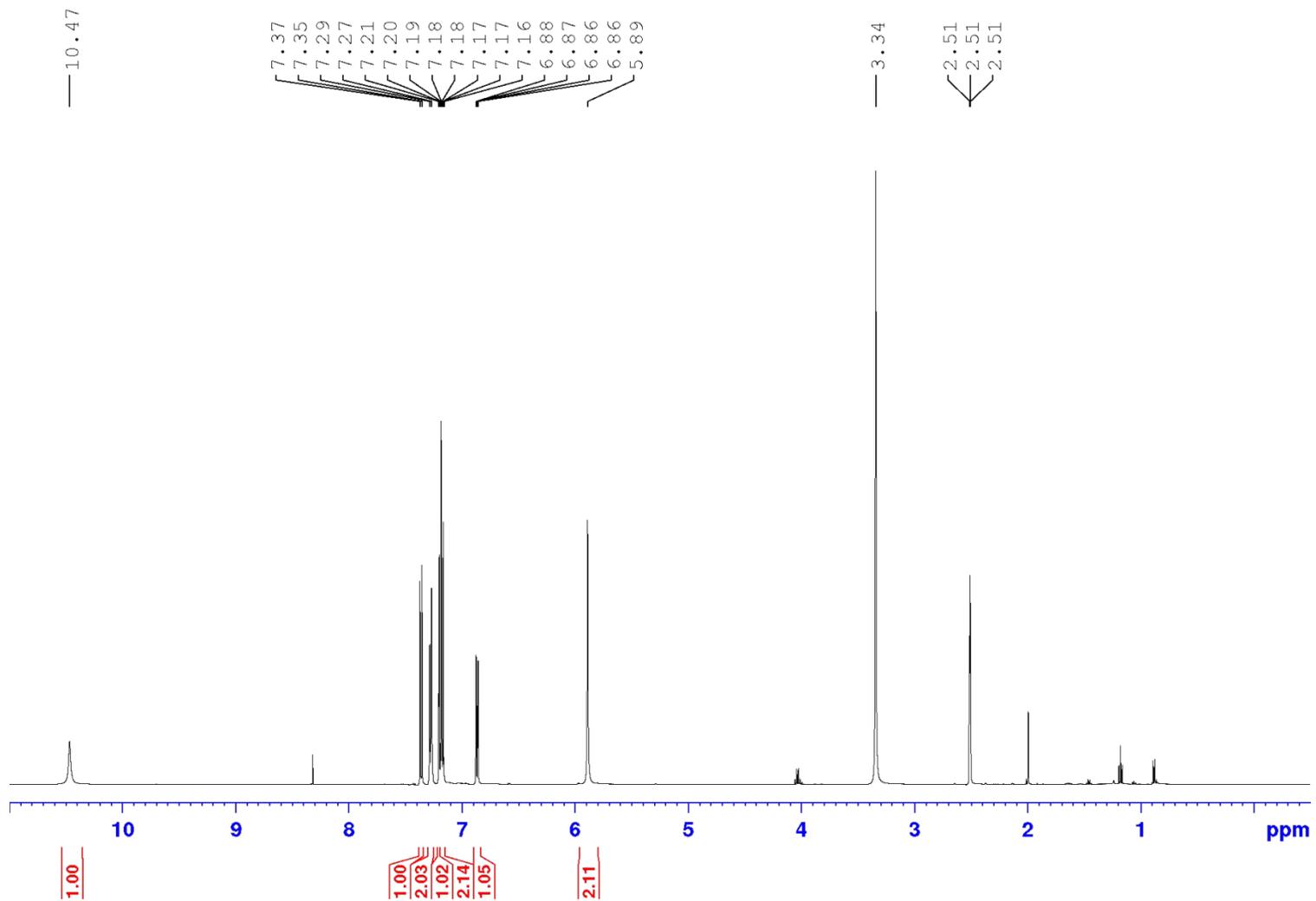


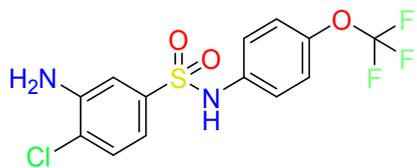
Compound **28** – ^{13}C NMR (125.5 MHz) – $\text{DMSO-}d_6$



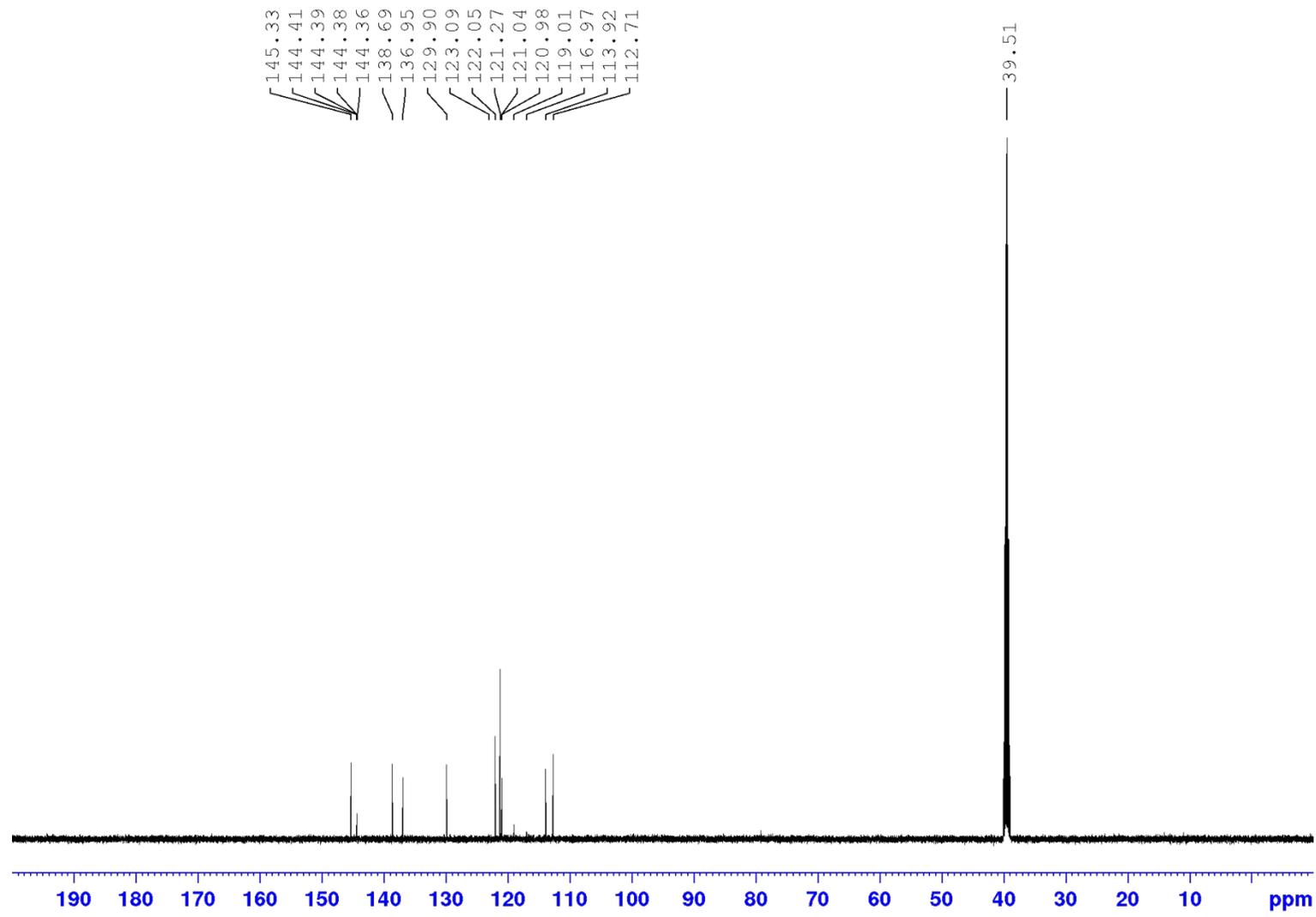


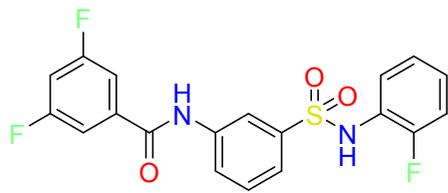
Compound 29 – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



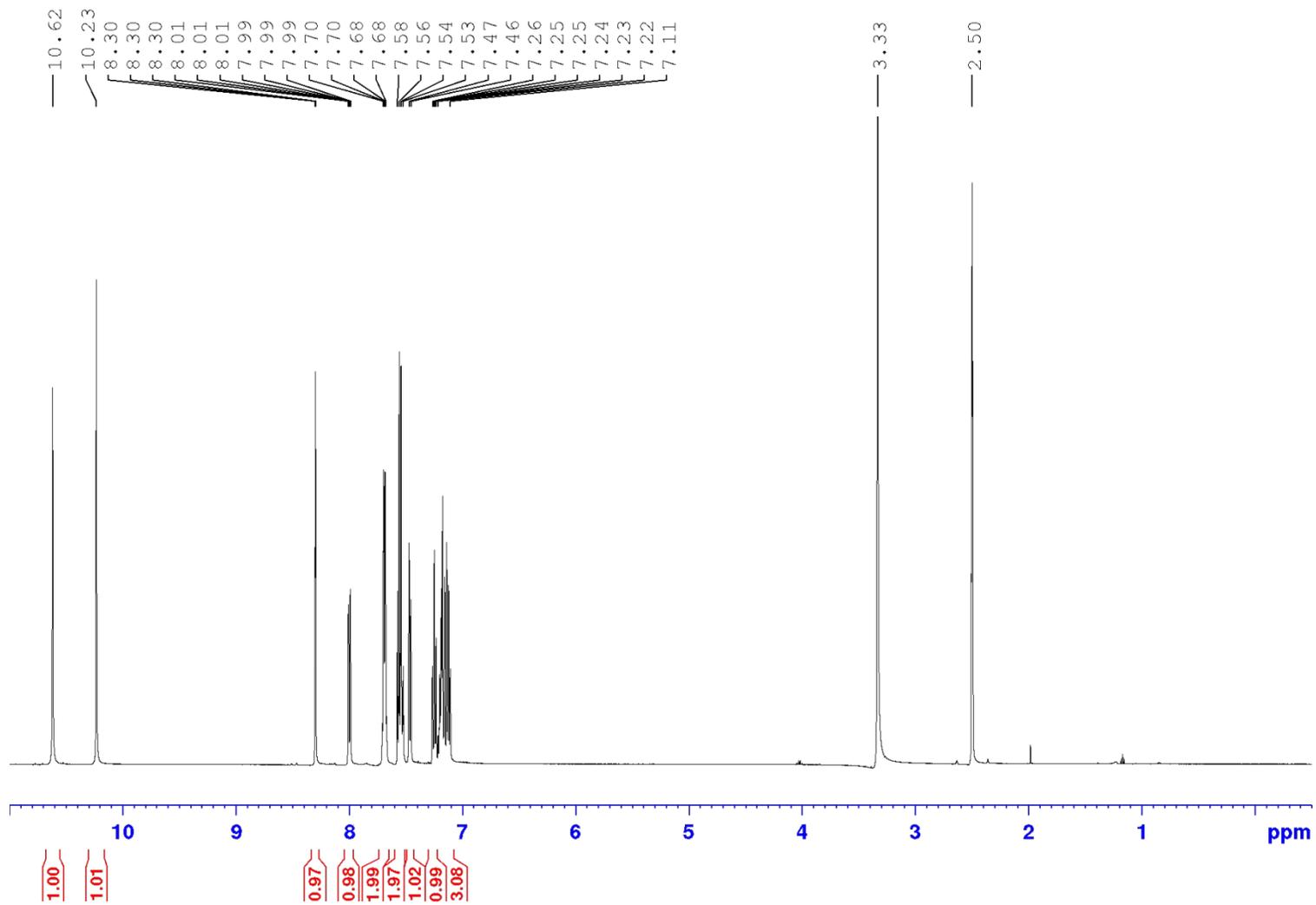


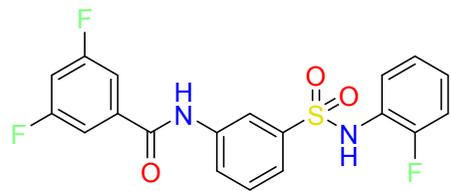
Compound 29 – ¹³C NMR (125.5 MHz) – DMSO-*d*₆



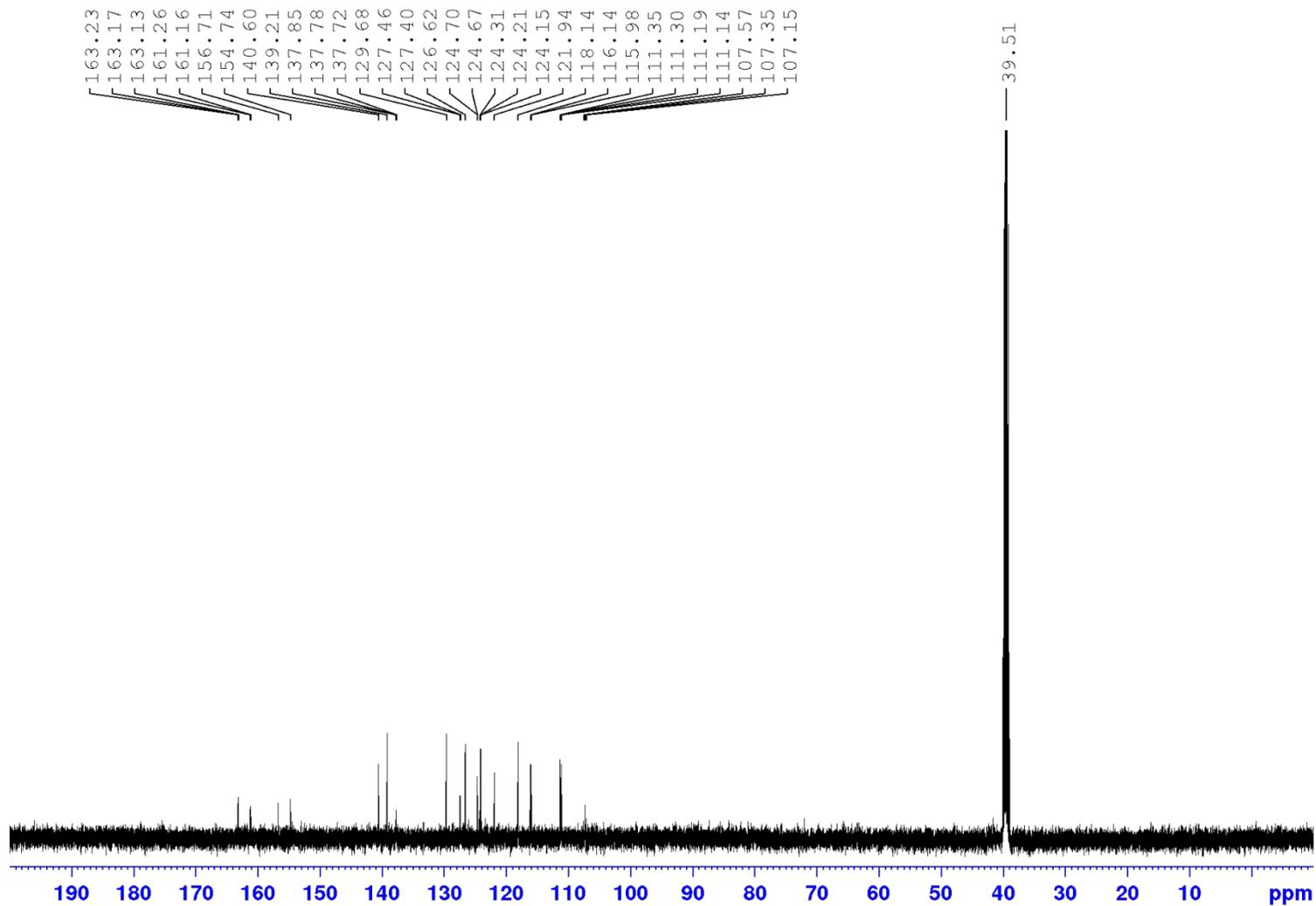


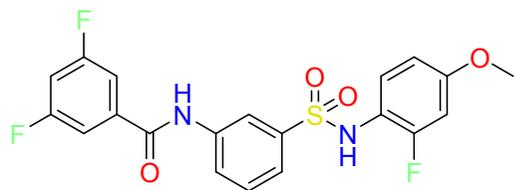
Compound **30** – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



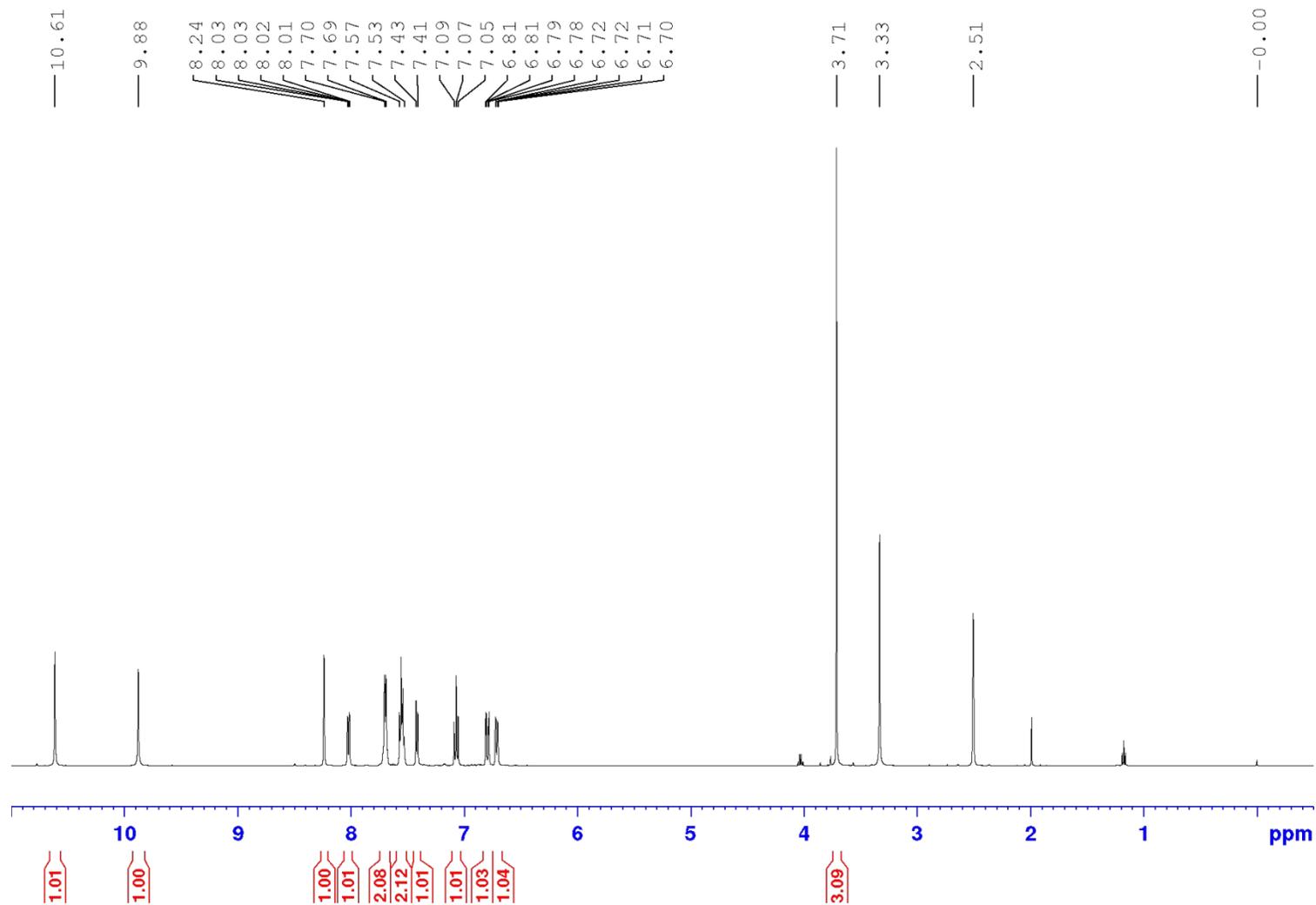


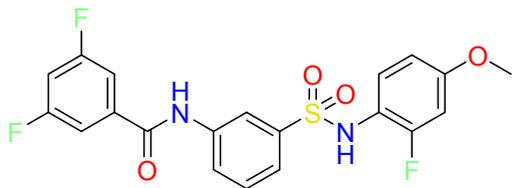
Compound 30 – ^{13}C NMR (125.5 MHz) – $\text{DMSO-}d_6$



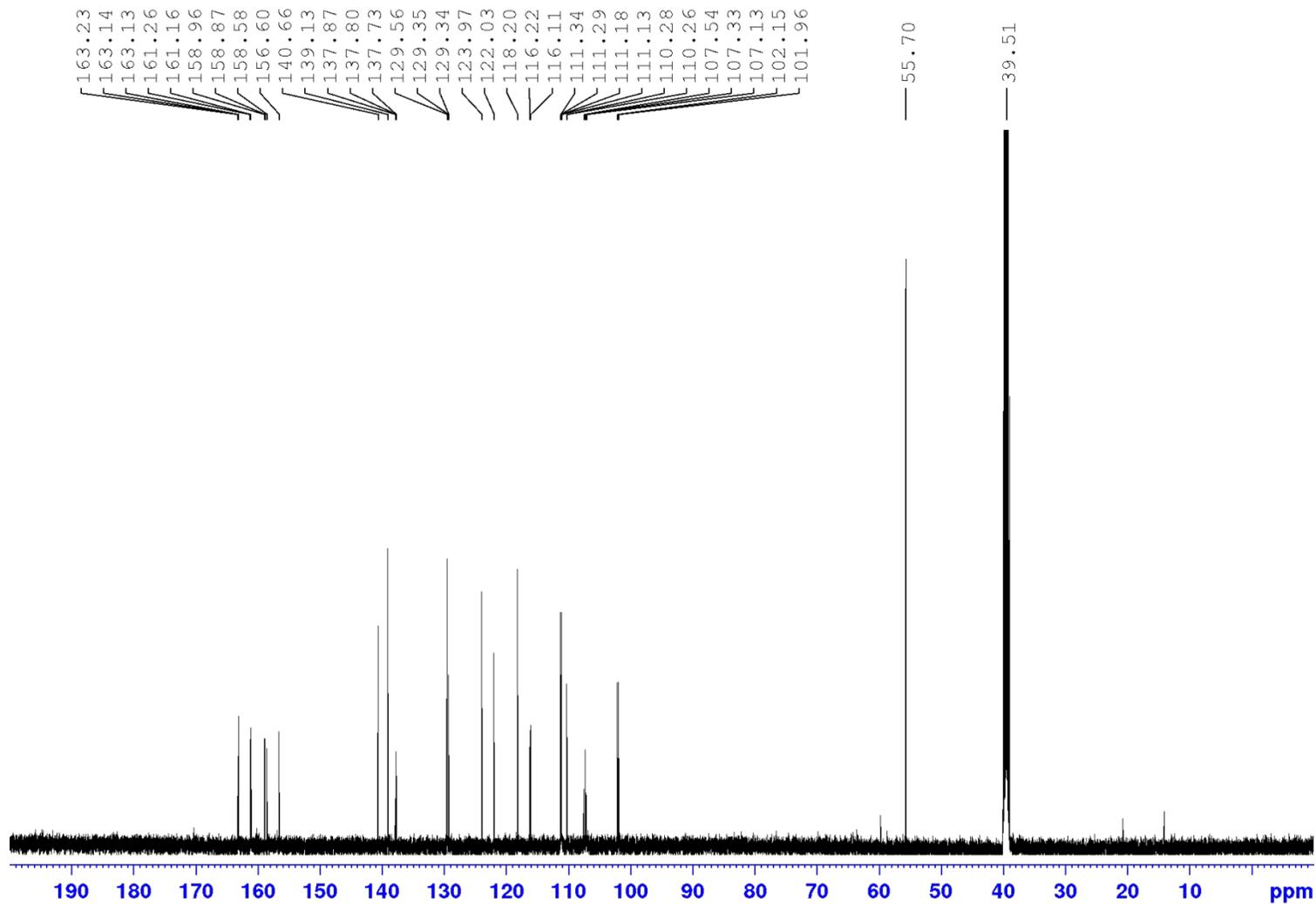


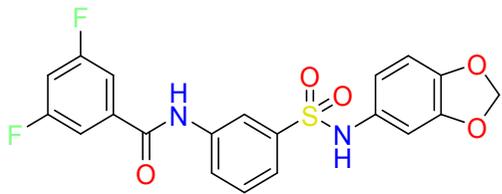
Compound **31** – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



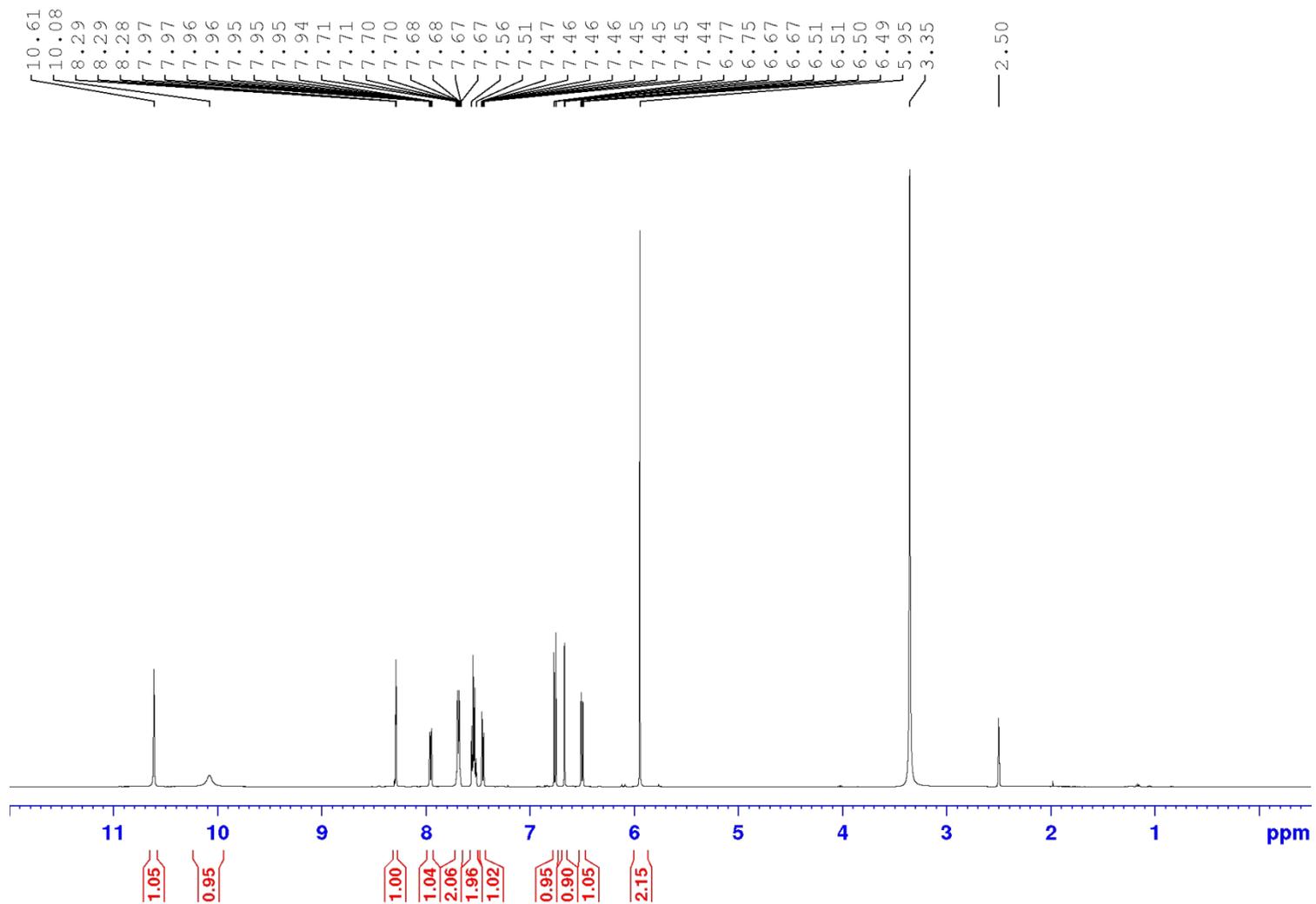


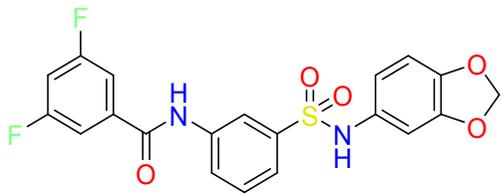
Compound 31 – ¹³C NMR (125.5 MHz) – DMSO-*d*₆



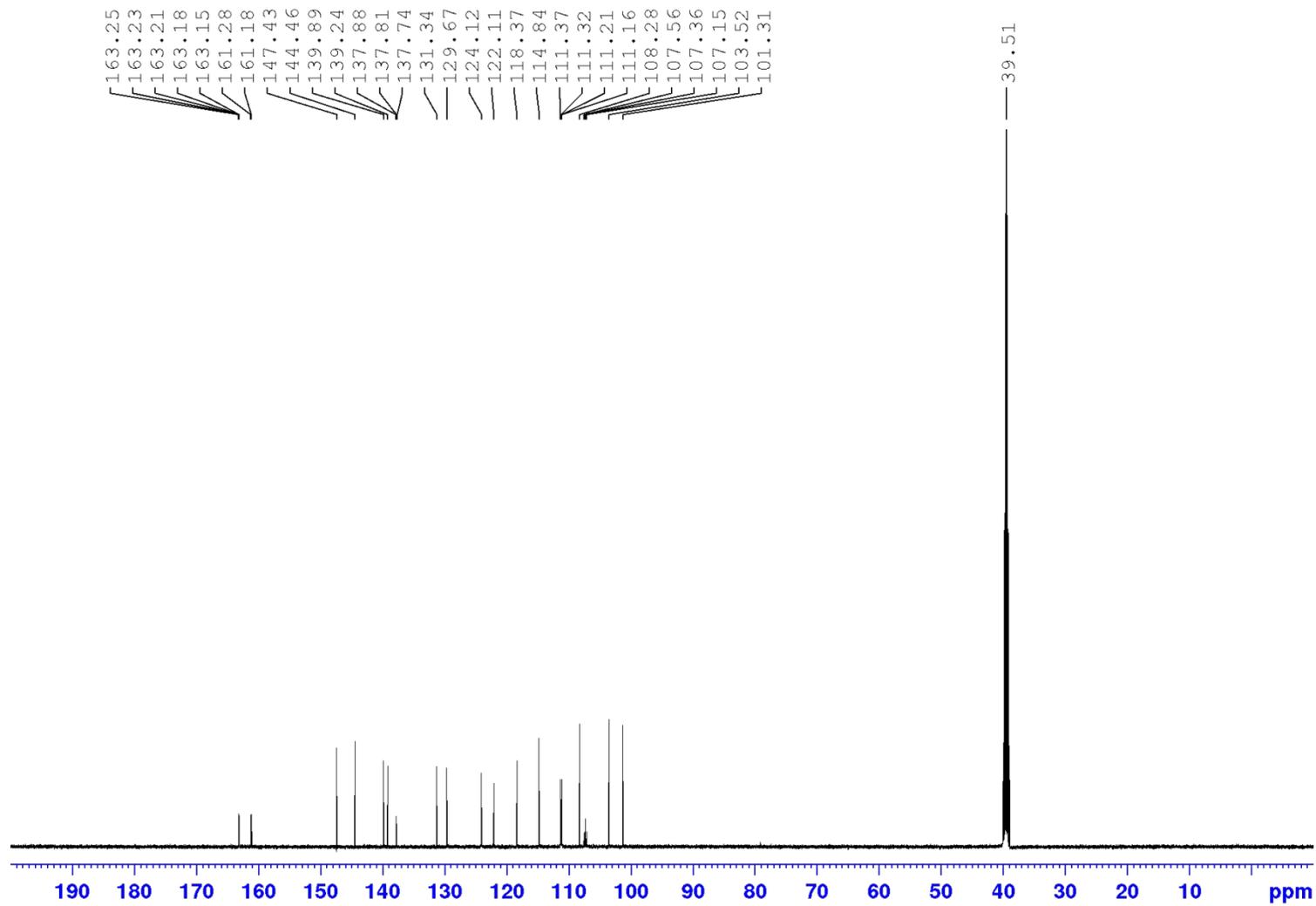


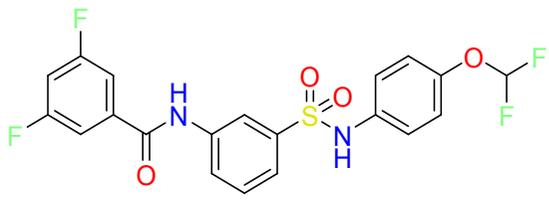
Compound **32** – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



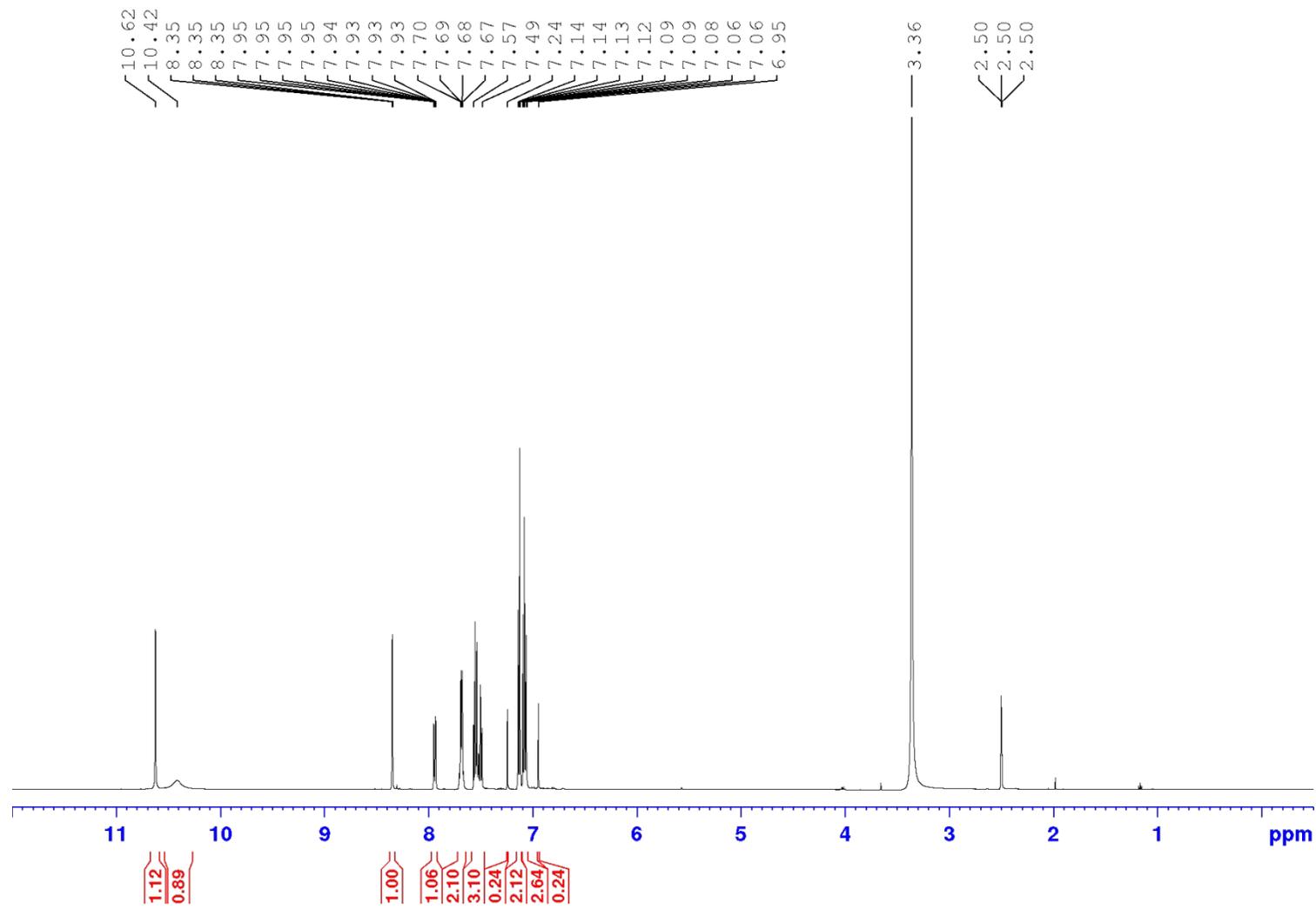


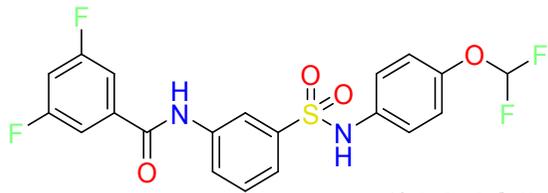
Compound **32** – ^{13}C NMR (125.5 MHz) – $\text{DMSO-}d_6$



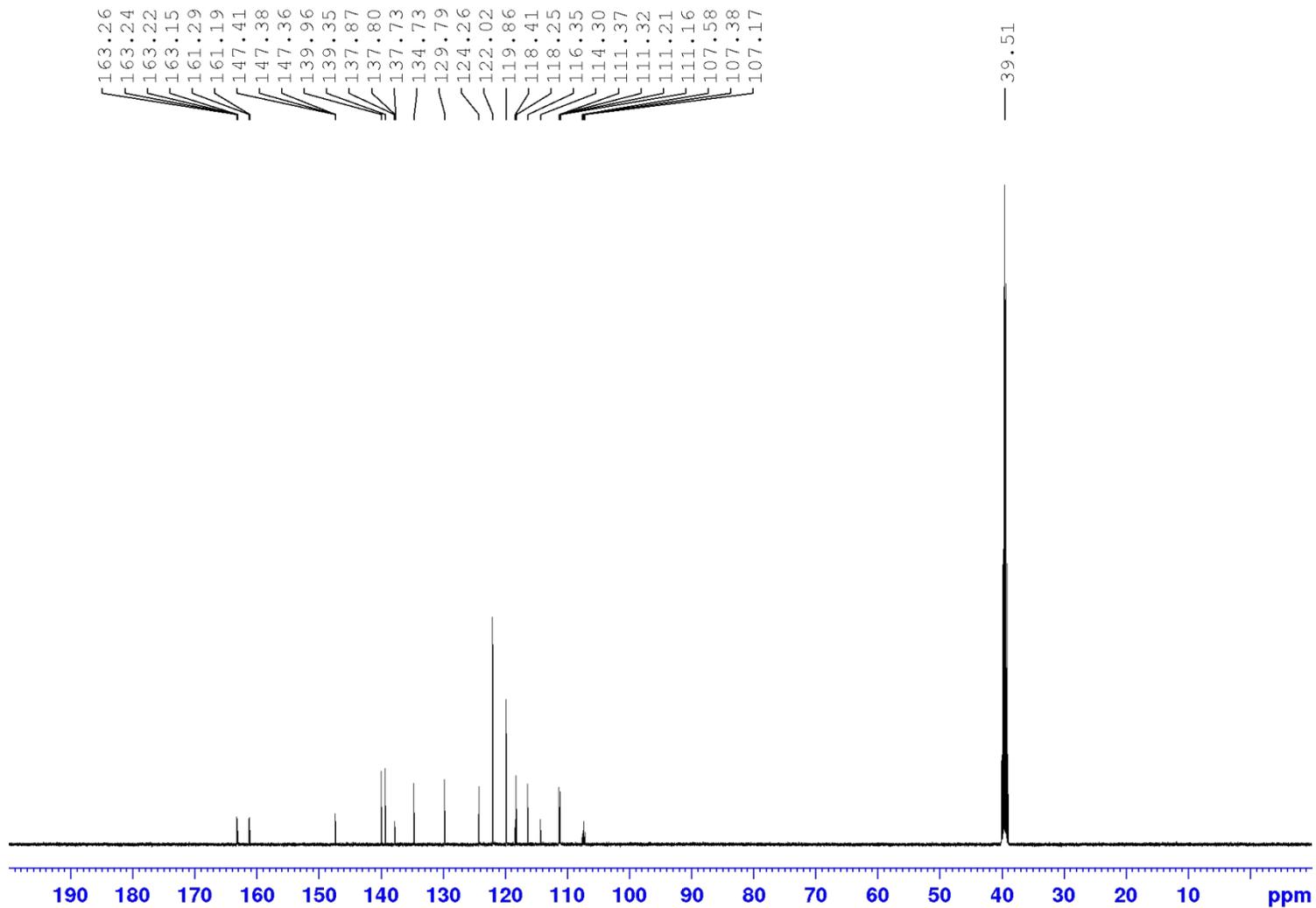


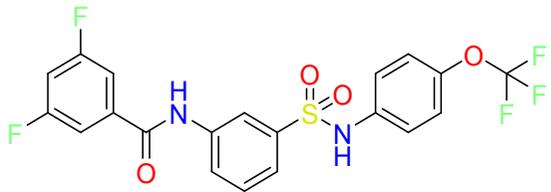
Compound 33 – ¹H NMR (500 MHz) – DMSO-d₆



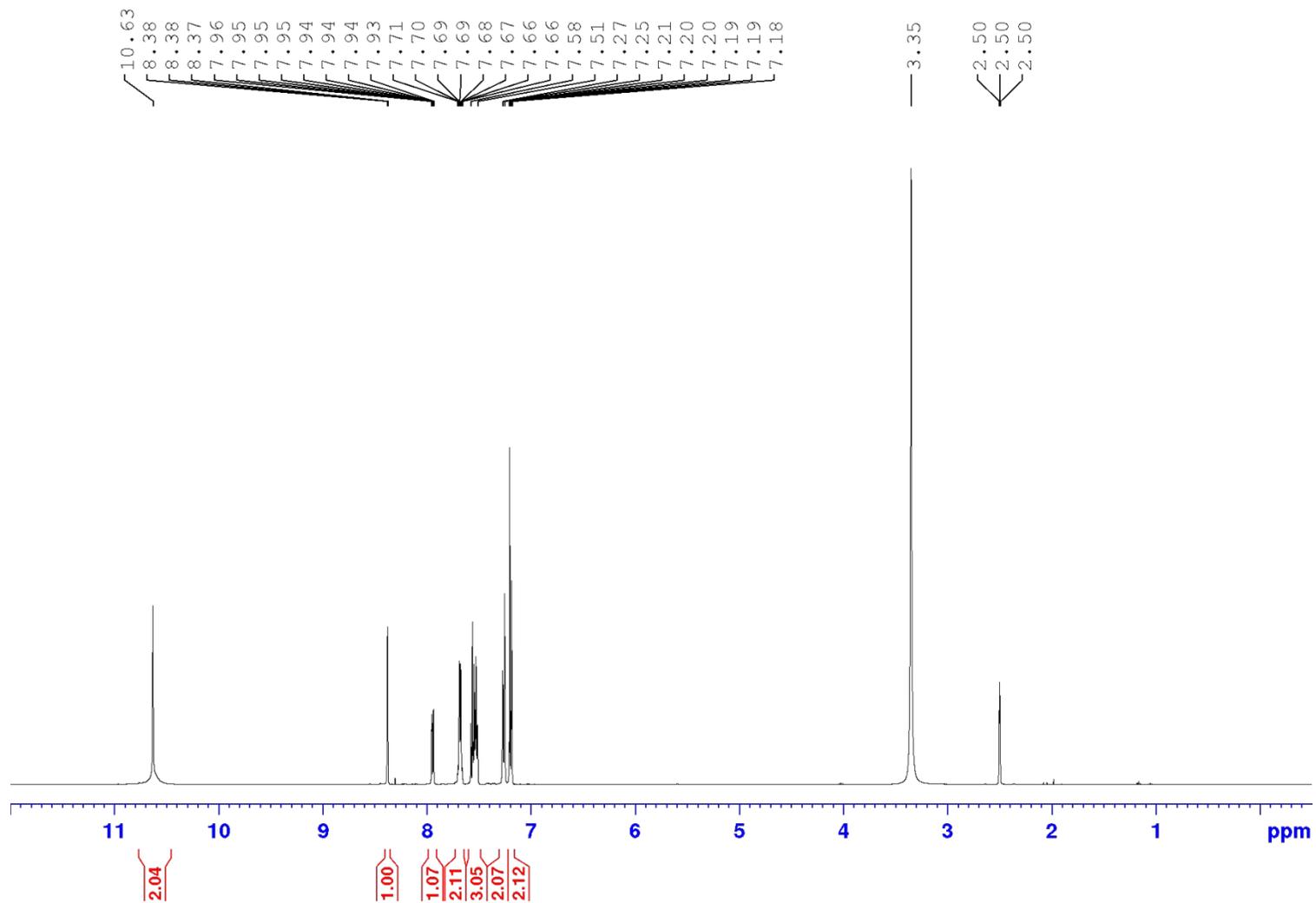


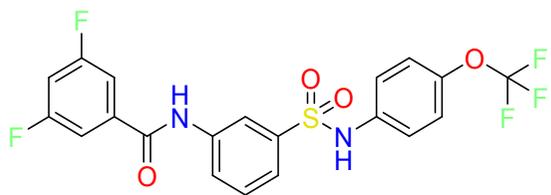
Compound **33** – ^{13}C NMR (125.5 MHz) – $\text{DMSO-}d_6$



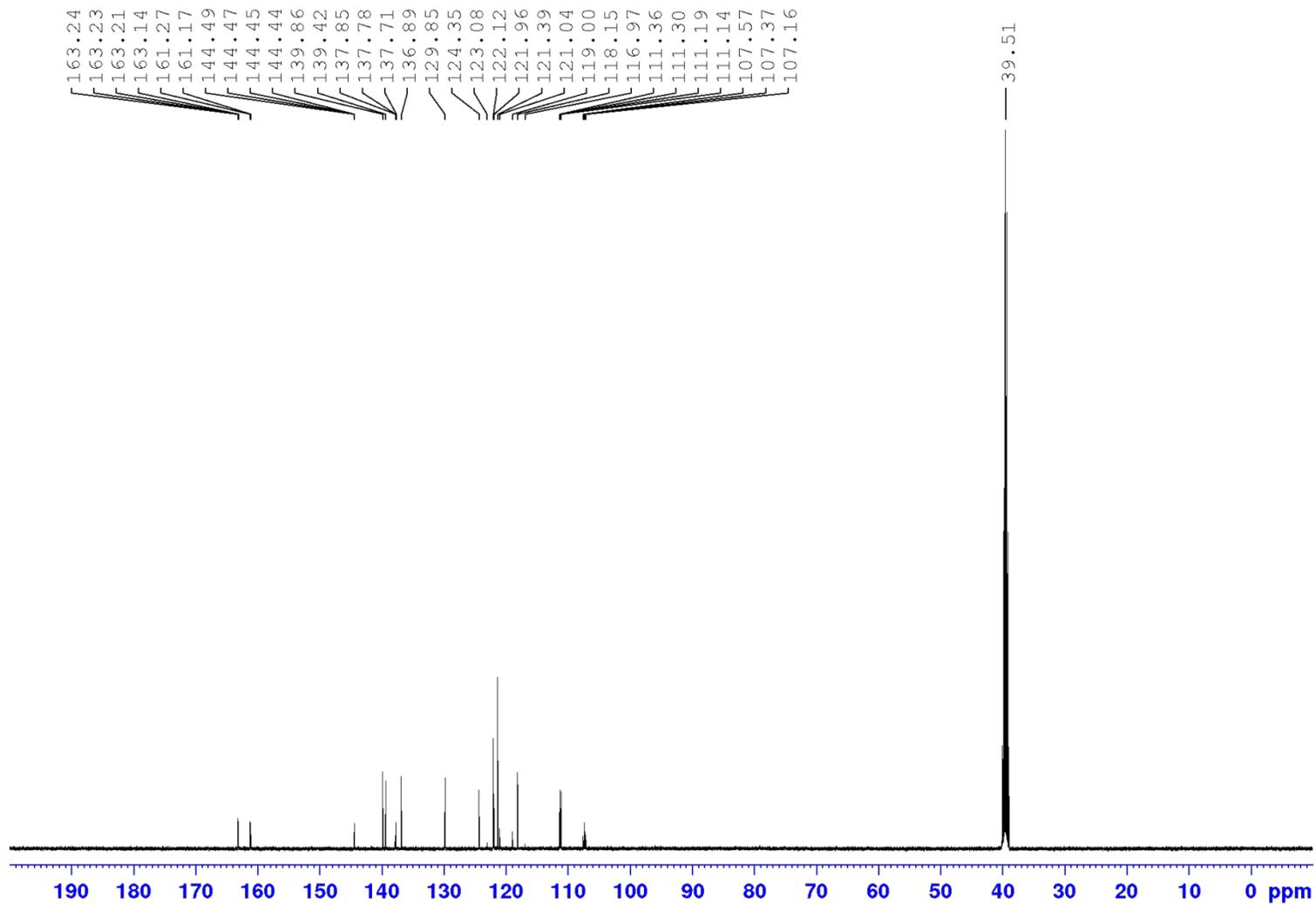


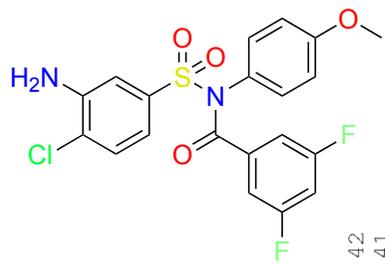
Compound **34** – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



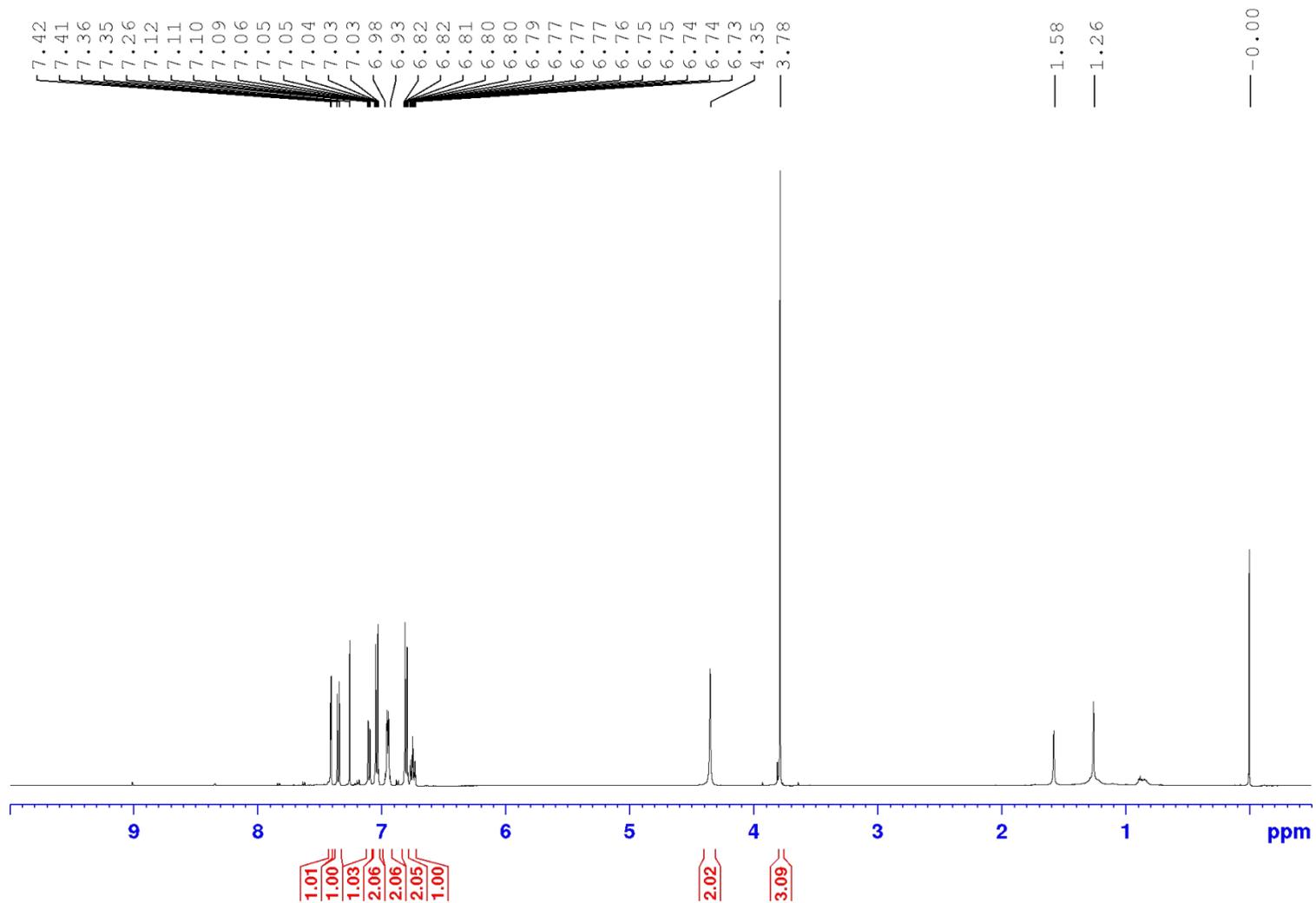


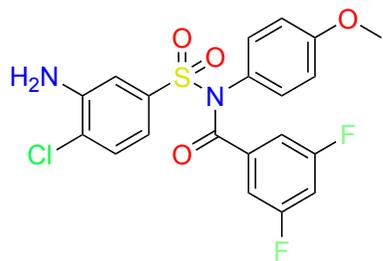
Compound **34** – ^{13}C NMR (125.5 MHz) – $\text{DMSO-}d_6$



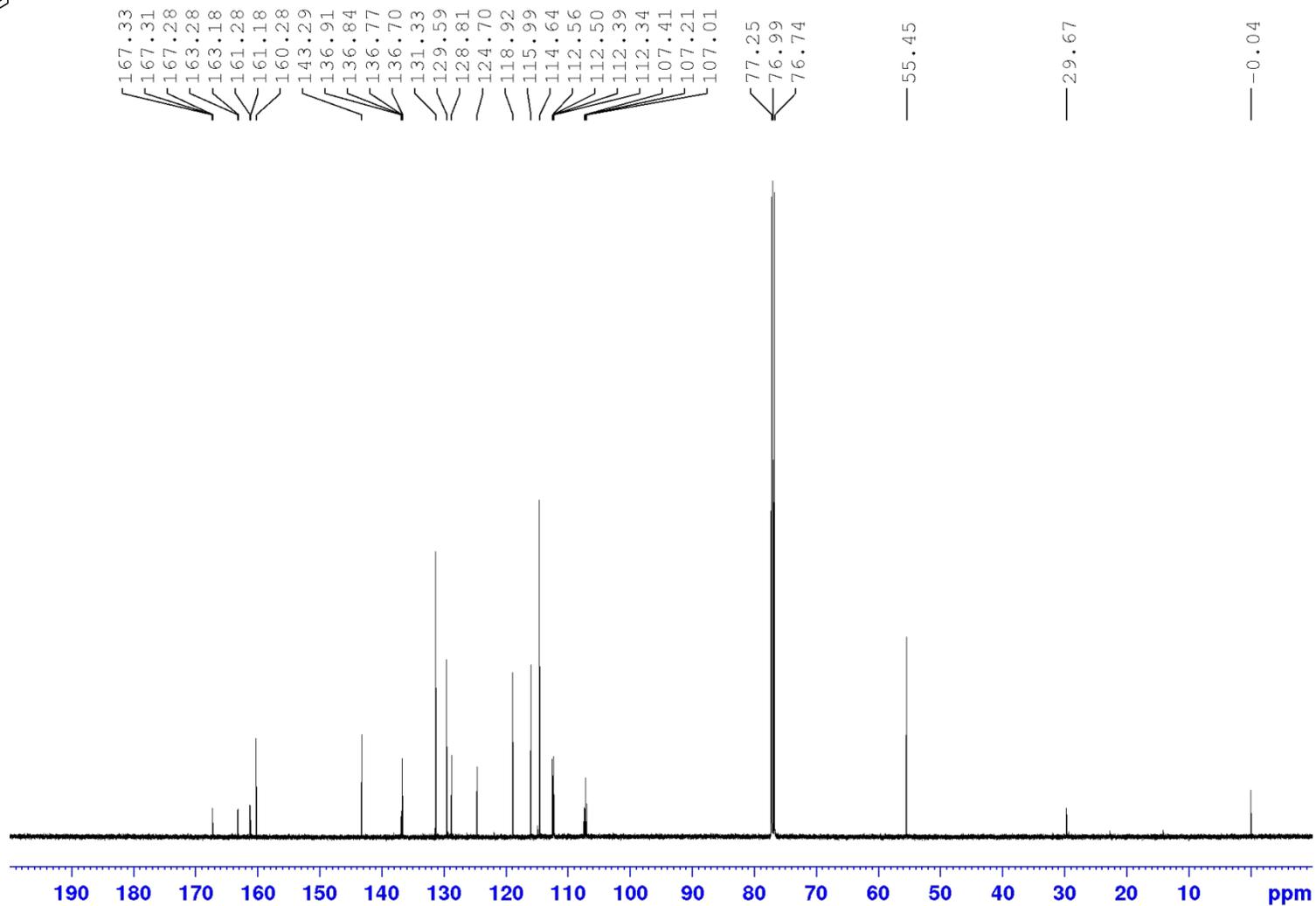


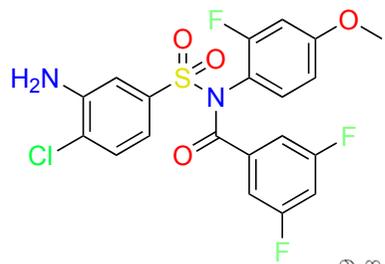
Compound 35 – ¹H NMR (500 MHz) – CDCl₃



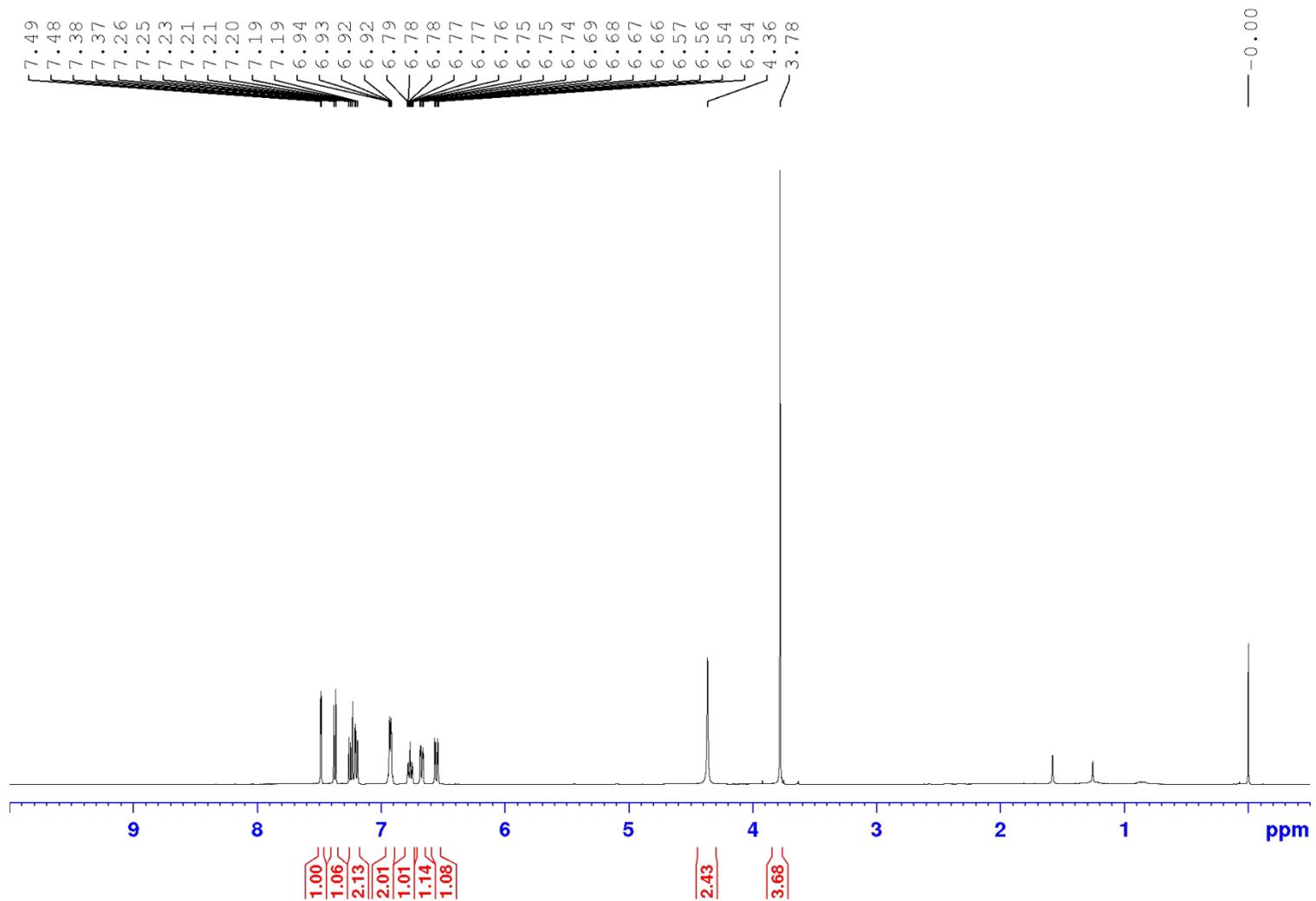


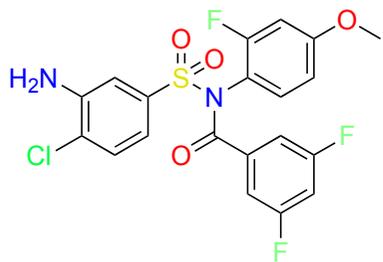
Compound 35 – ¹³C NMR (125.5 MHz) – CDCl₃



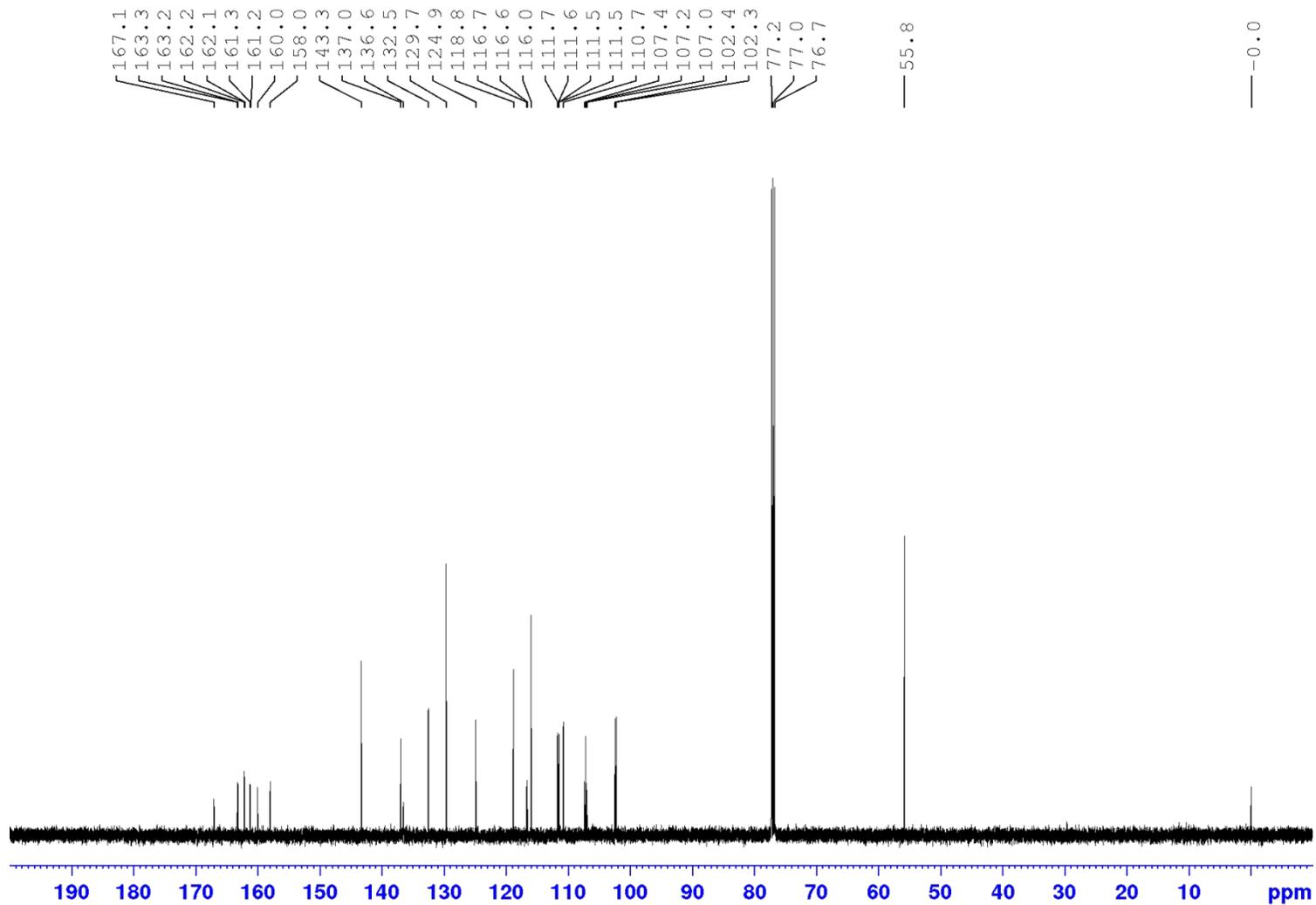


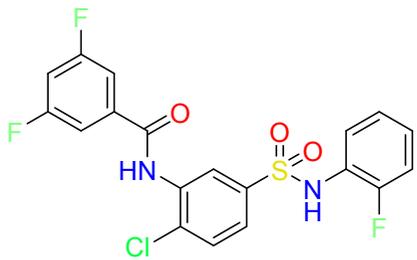
Compound 36 – ¹H NMR (500 MHz) – CDCl₃



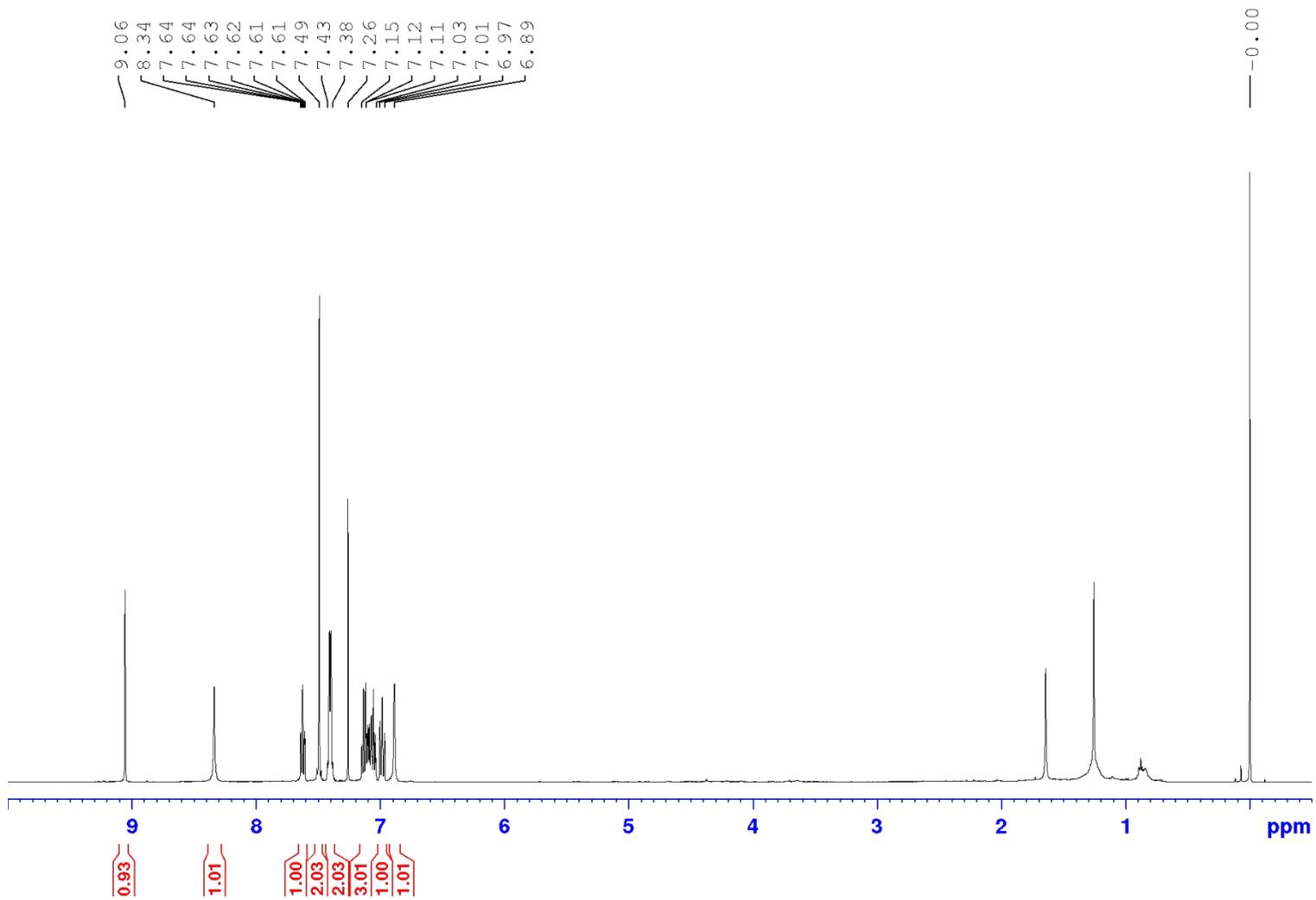


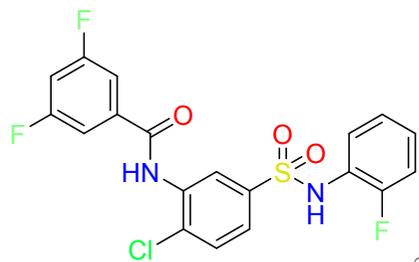
Compound 36 – ¹³C NMR (125.5 MHz) – CDCl₃



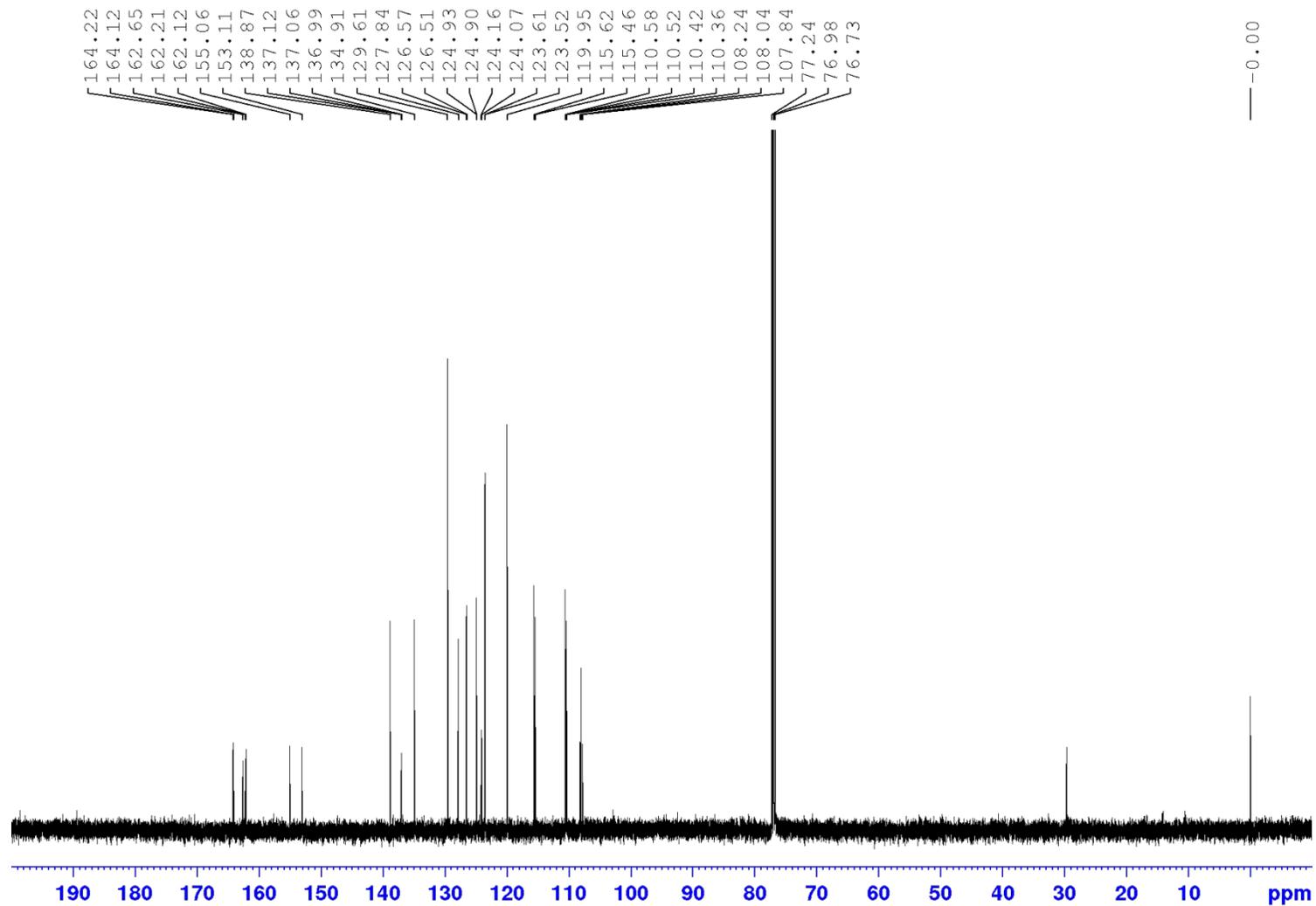


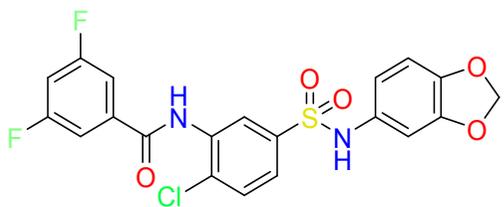
Compound 37 – ^1H NMR (500 MHz) – CDCl_3



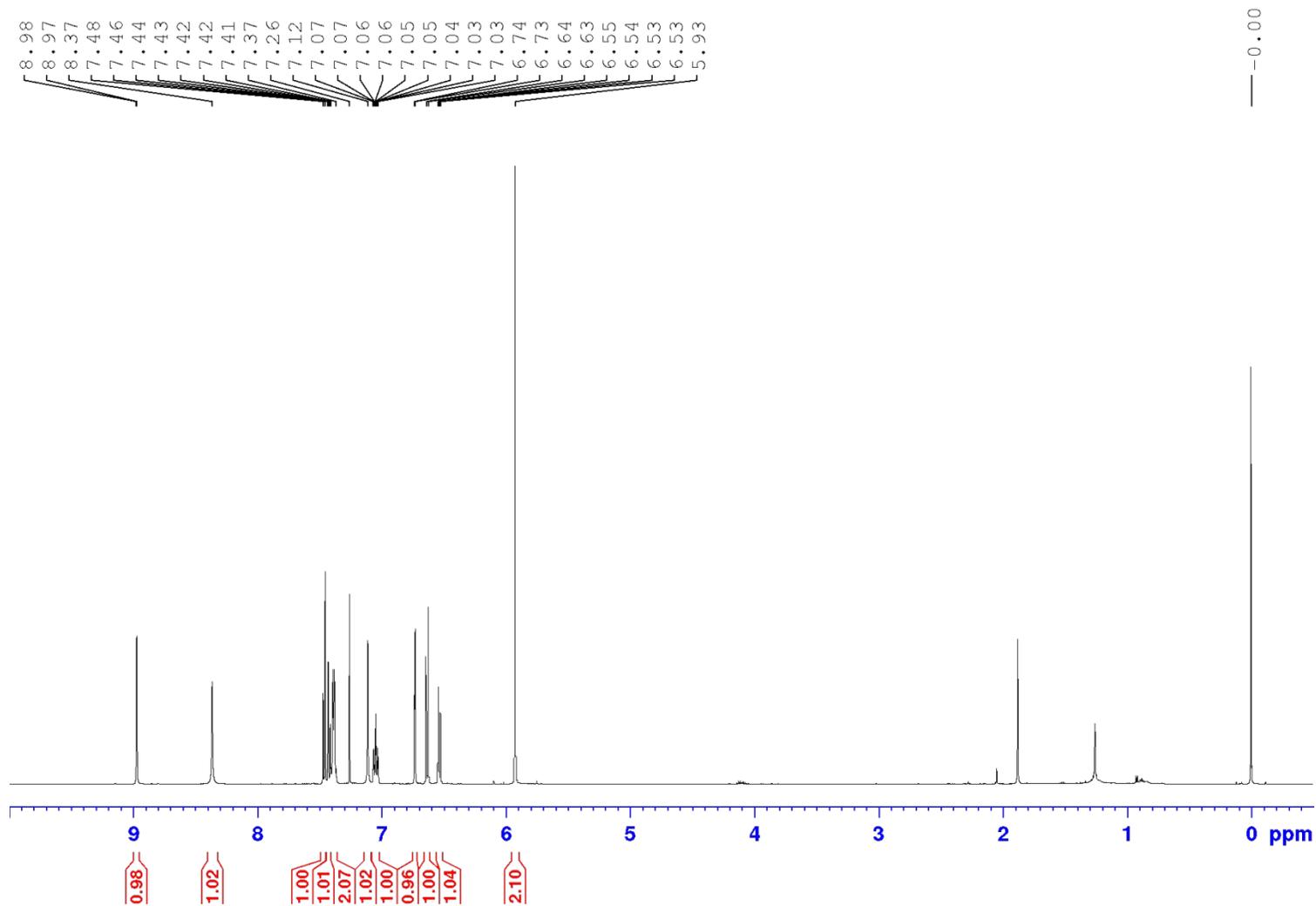


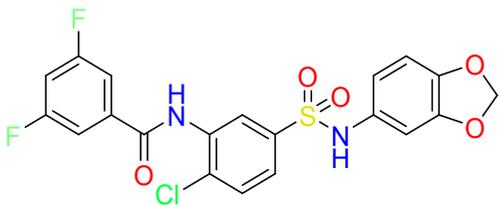
Compound 37 – ¹³C NMR (125.5 MHz) – CDCl₃



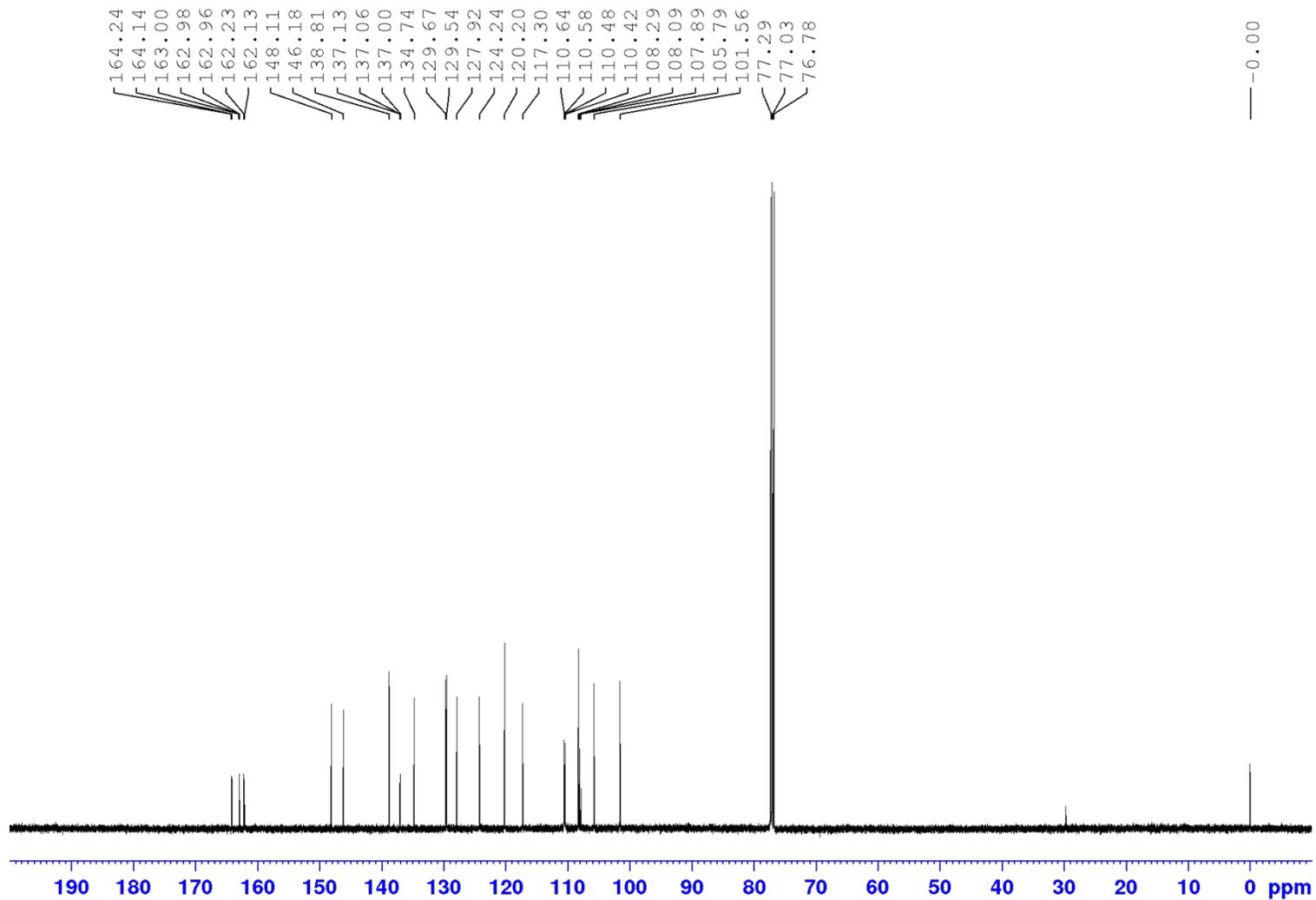


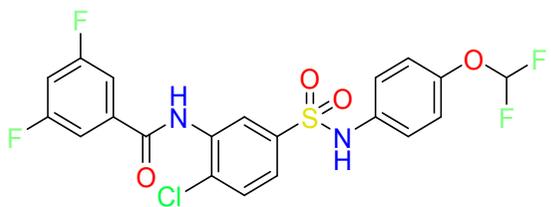
Compound **38** – ^1H NMR (500 MHz) – CDCl_3



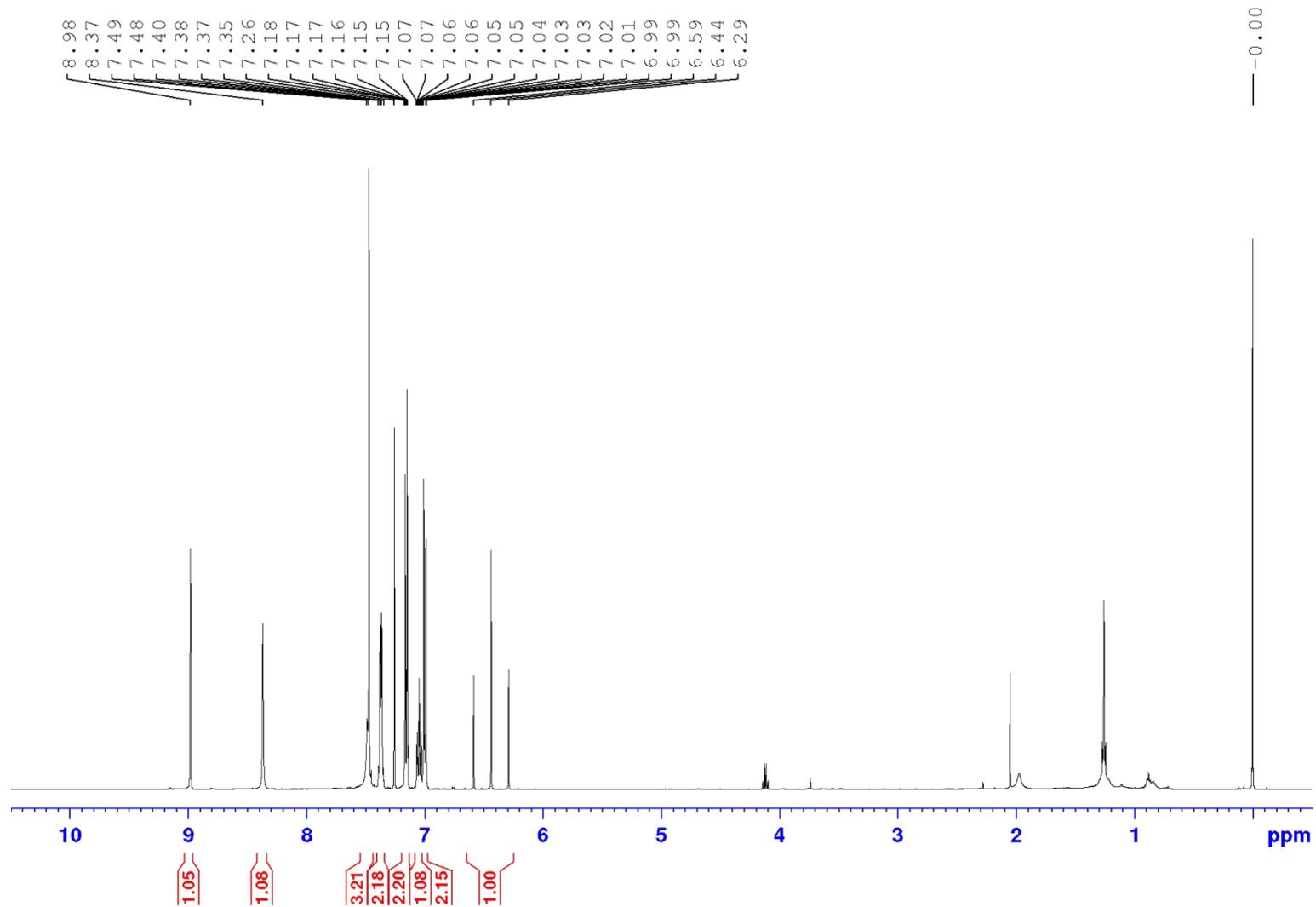


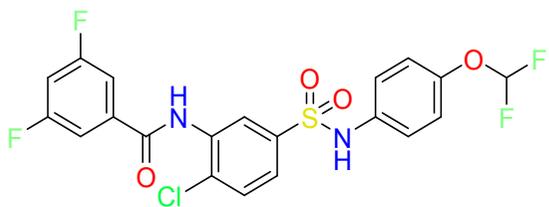
Compound **38** – ^{13}C NMR (125.5 MHz) – CDCl_3



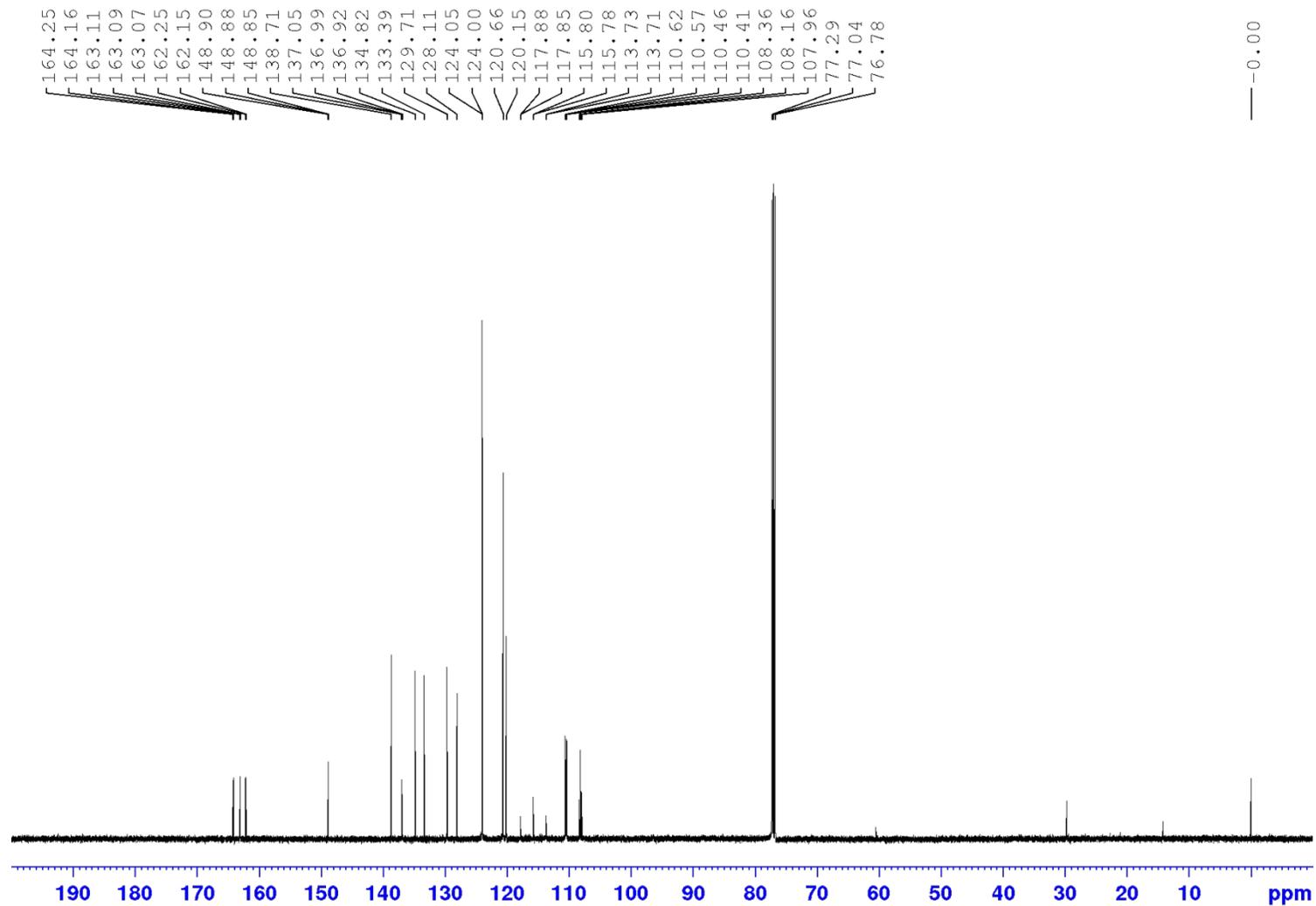


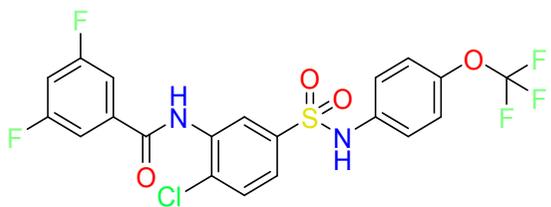
Compound 39 – ^1H NMR (500 MHz) – CDCl_3



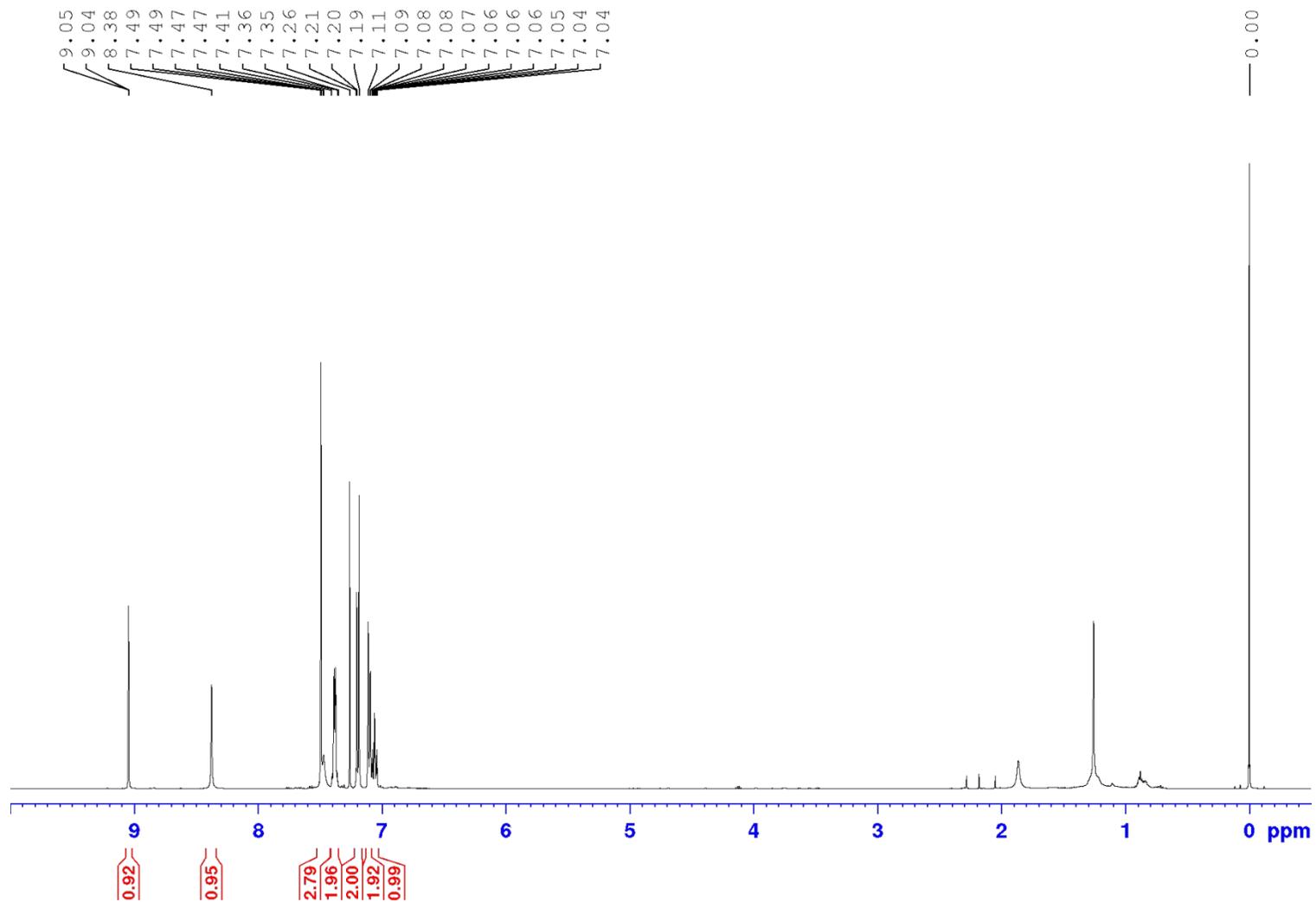


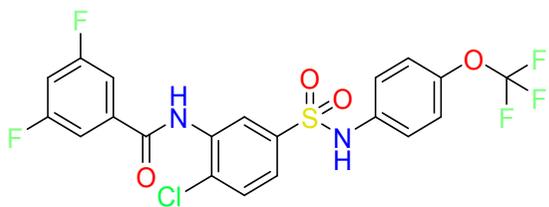
Compound 39 – ^1H NMR (500 MHz) – CDCl_3



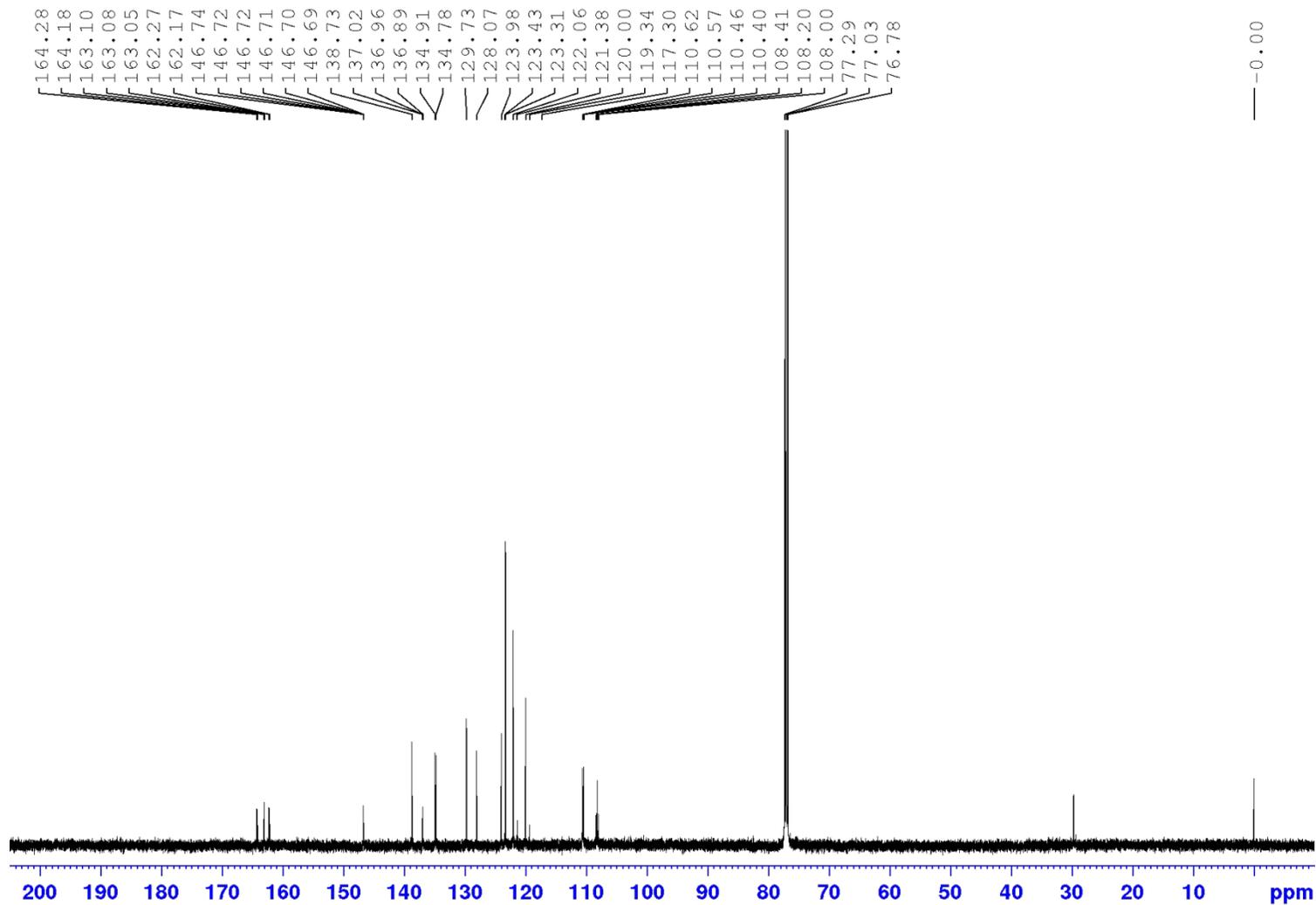


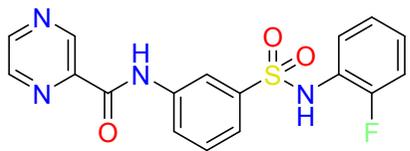
Compound **40** – ^1H NMR (500 MHz) – CDCl_3



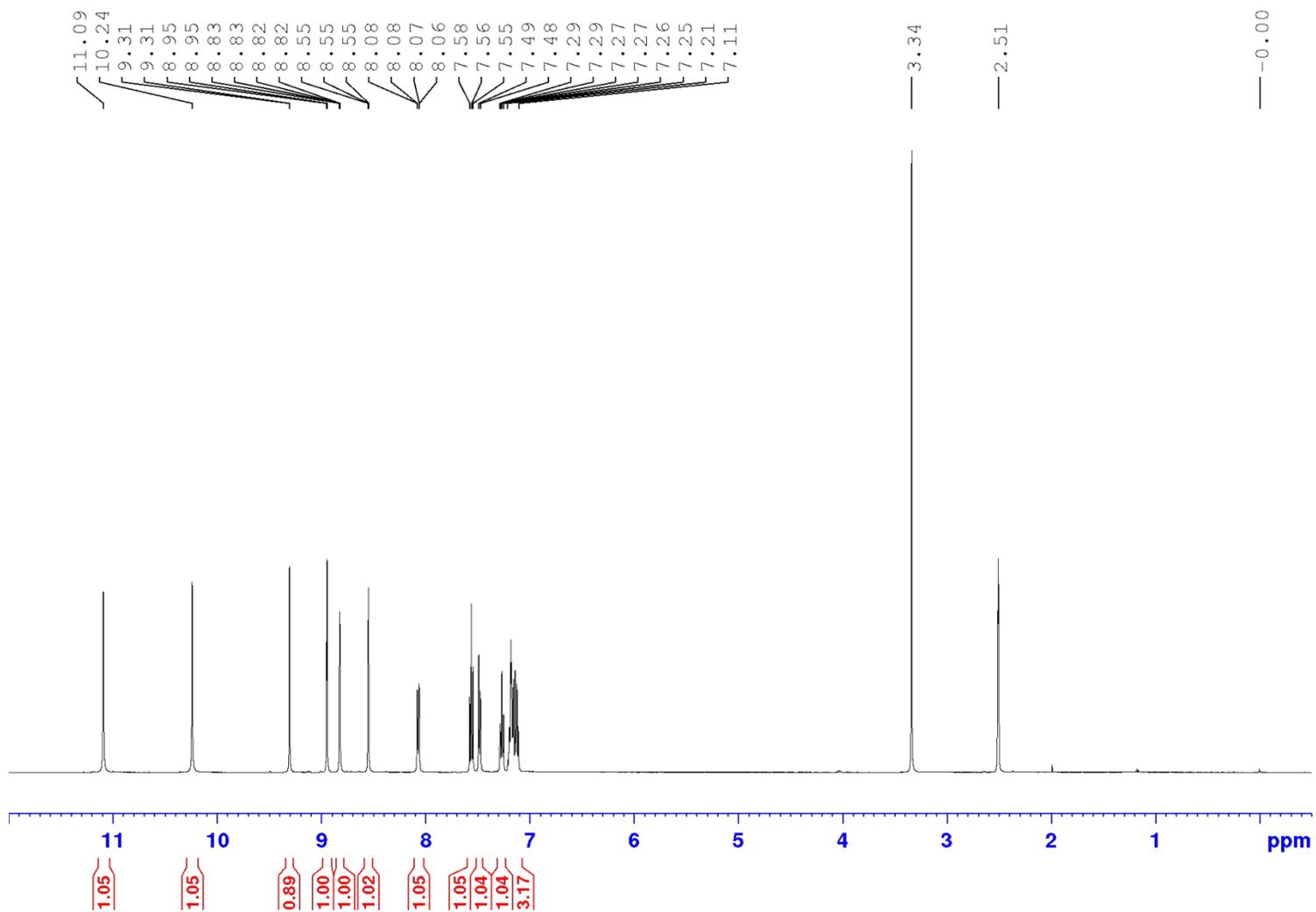


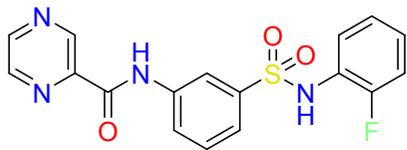
Compound **40** – ^{13}C NMR (125.5 MHz) – CDCl_3



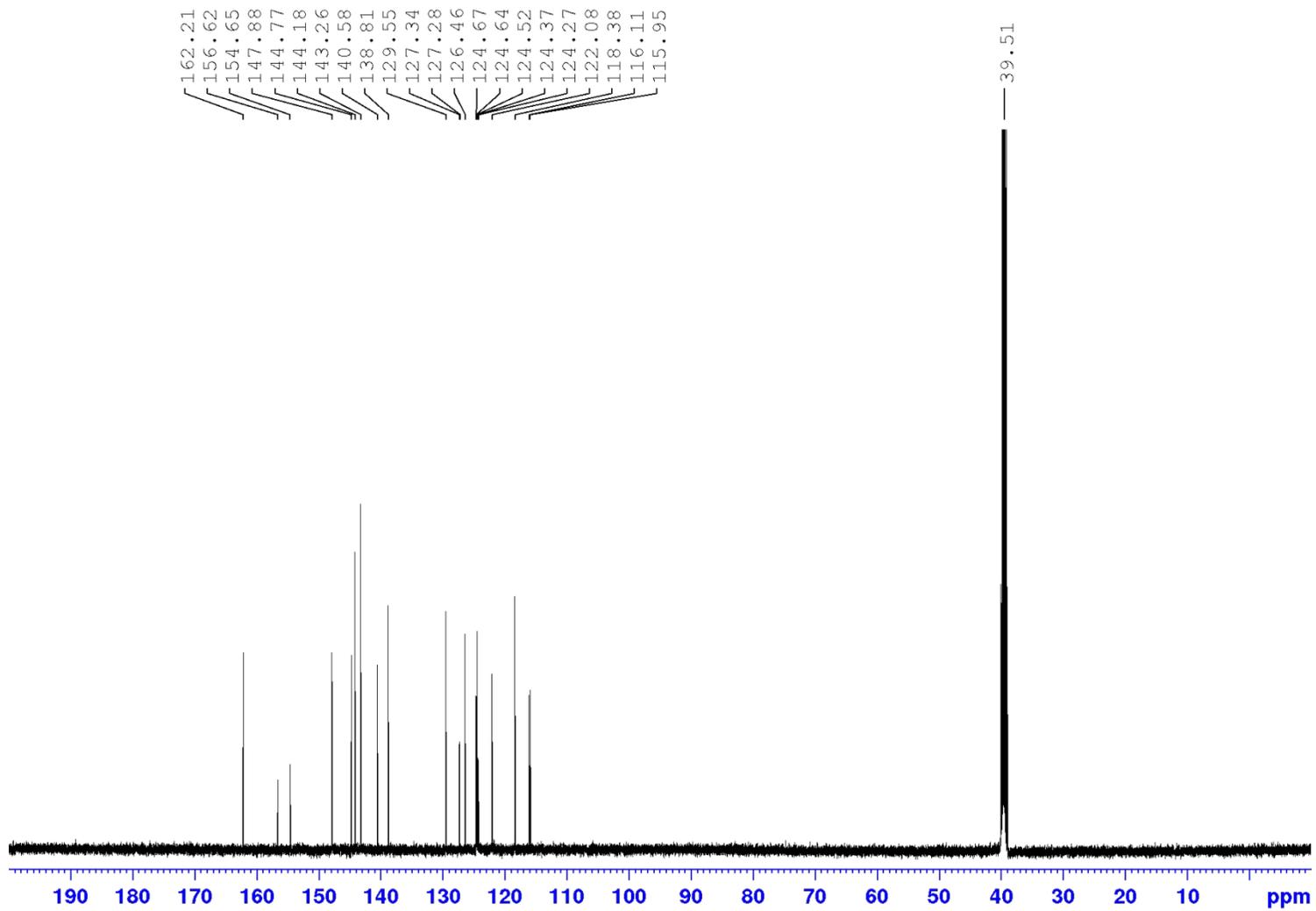


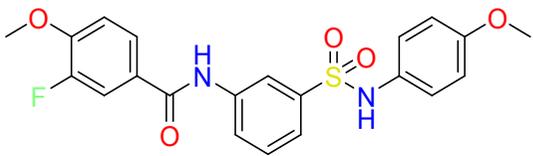
Compound **41** – ^1H NMR (500 MHz) – $\text{DMSO-}d_6$



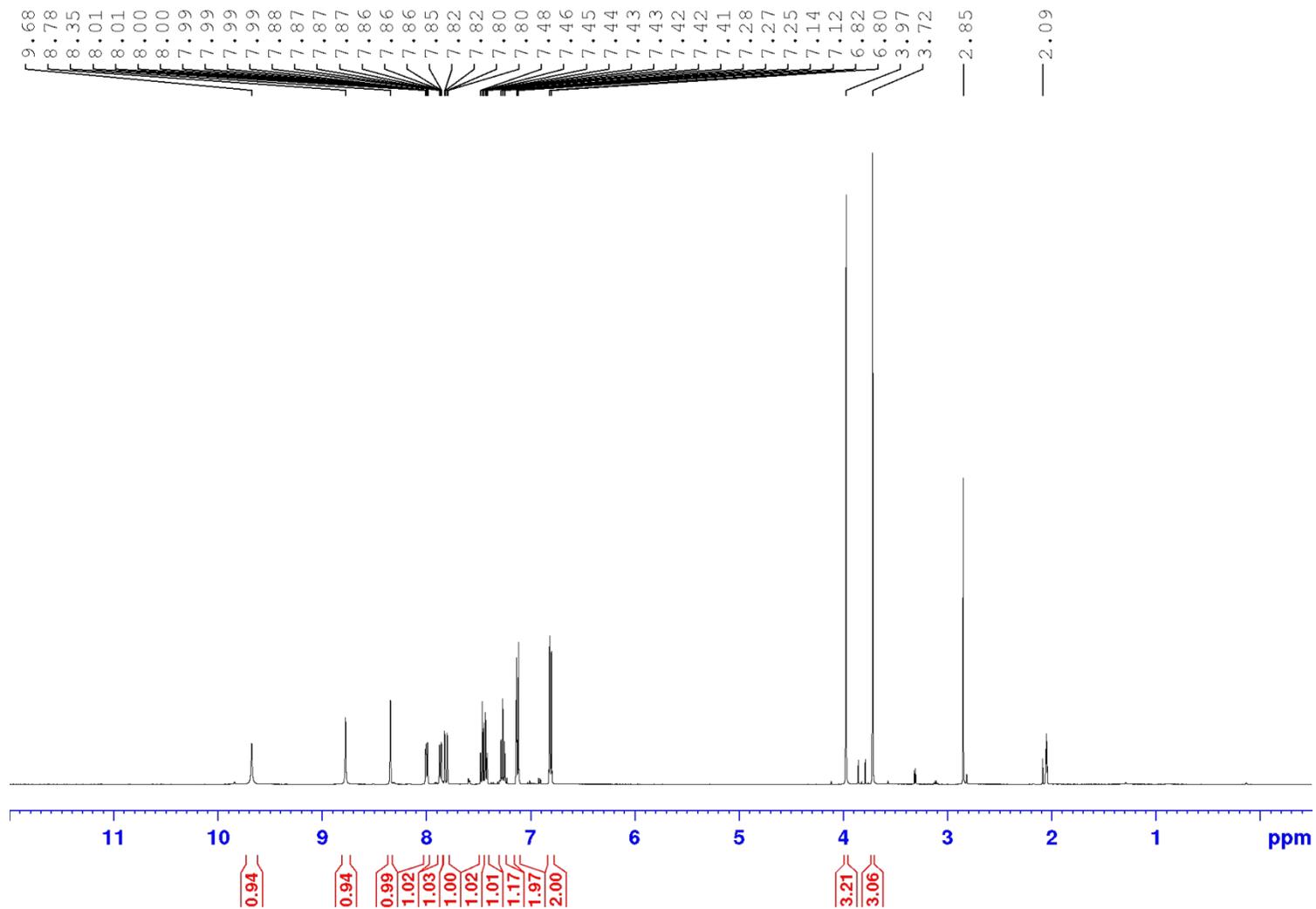


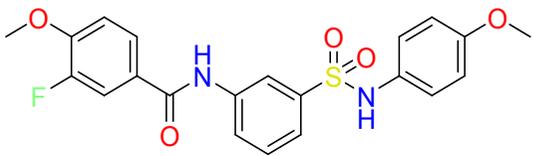
Compound **41** – ^{13}C NMR (125.5 MHz) – $\text{DMSO-}d_6$



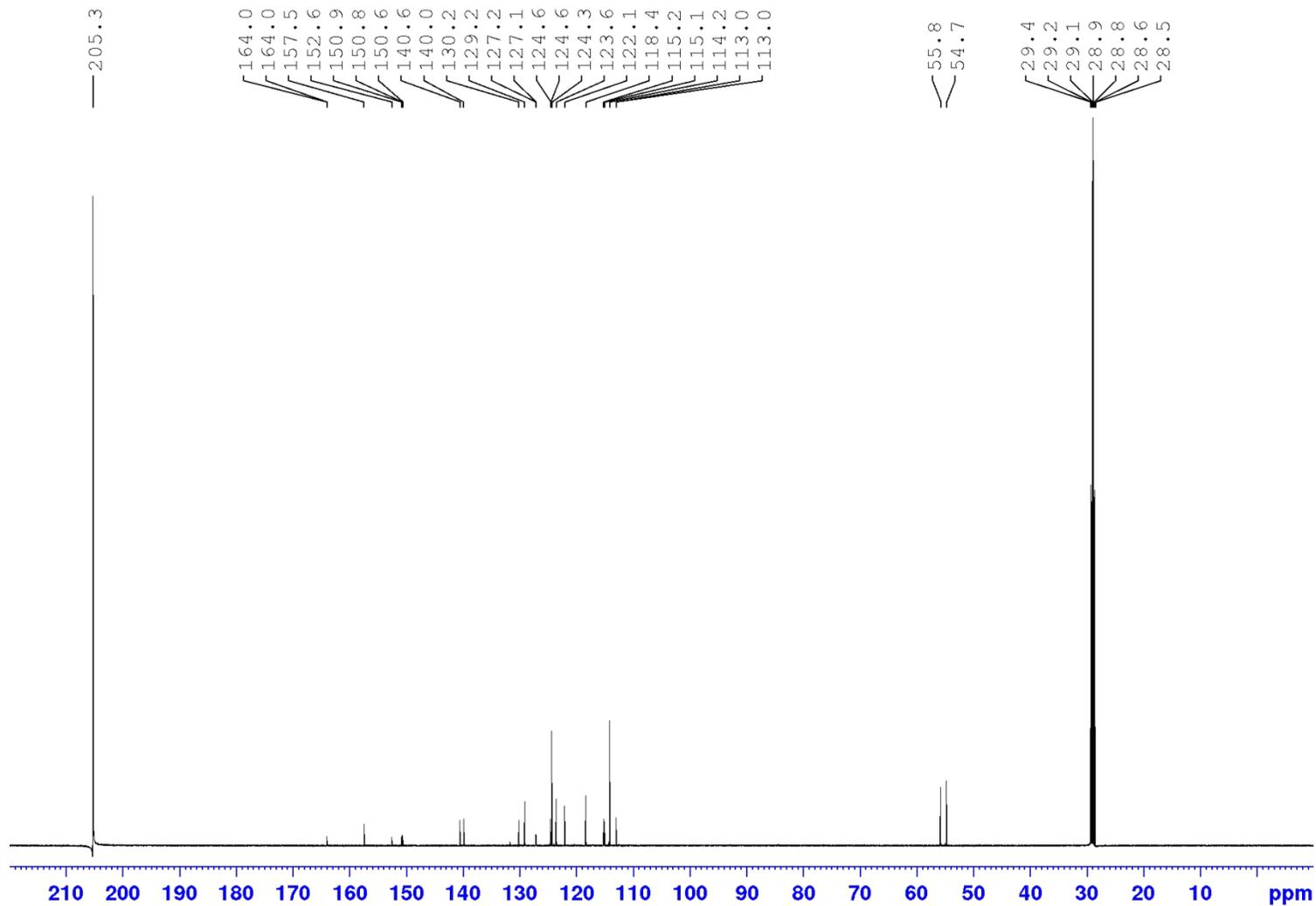


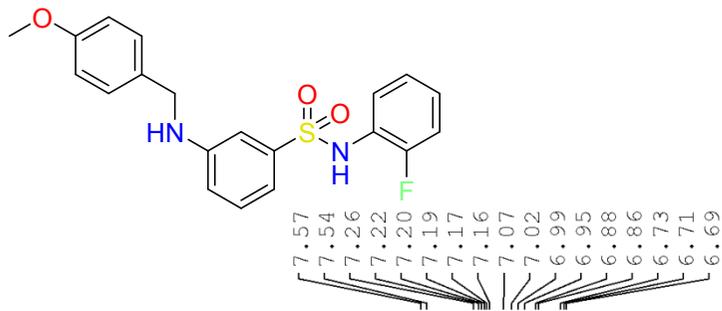
Compound 42 – ^1H NMR (500 MHz) – Acetone- d_6



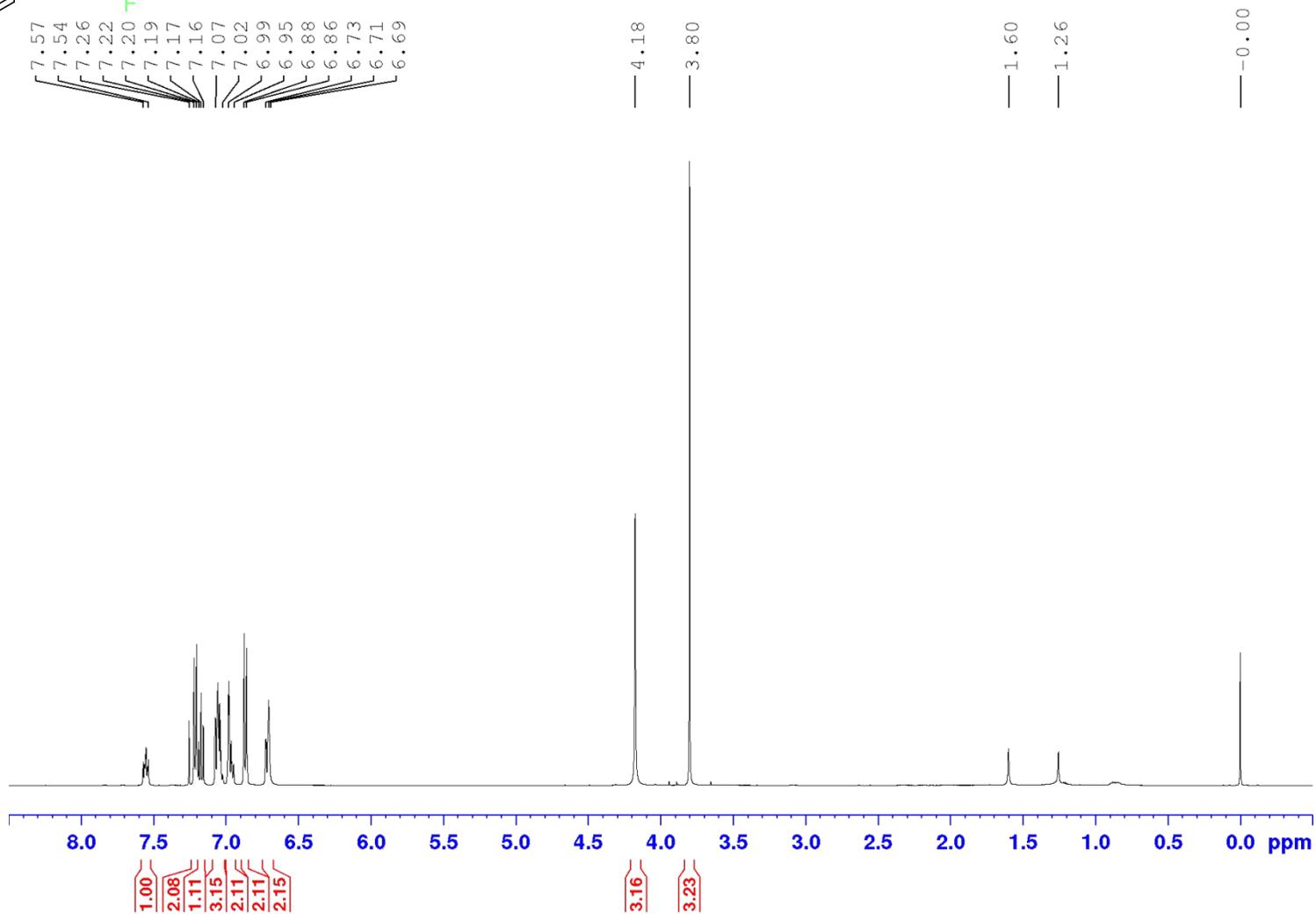


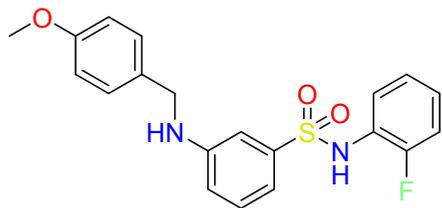
Compound **42** – ^{13}C NMR (125.5 MHz) – Acetone- d_6



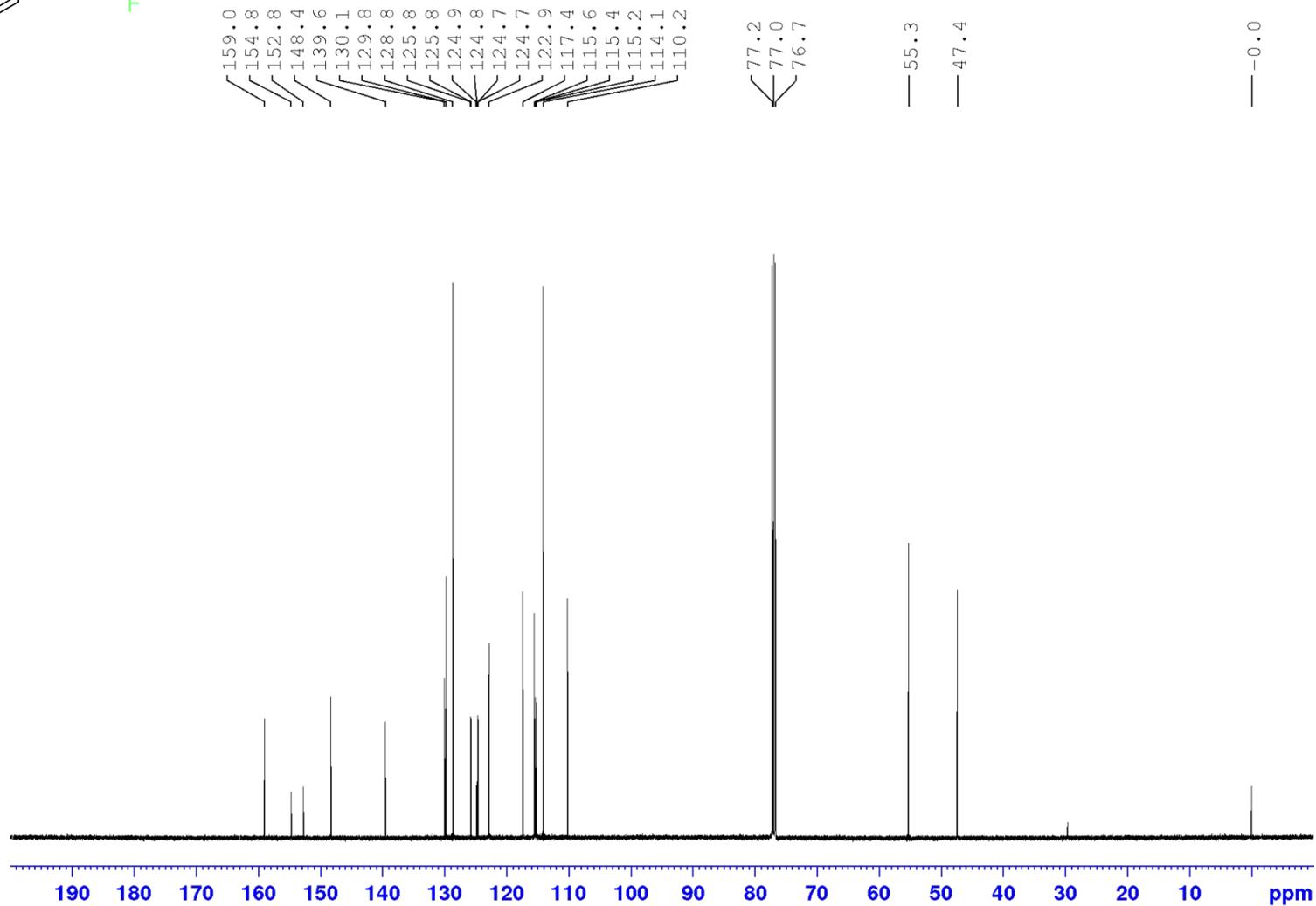


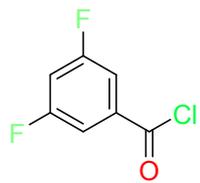
Compound 43 – ¹H NMR (500 MHz) – CDCl₃



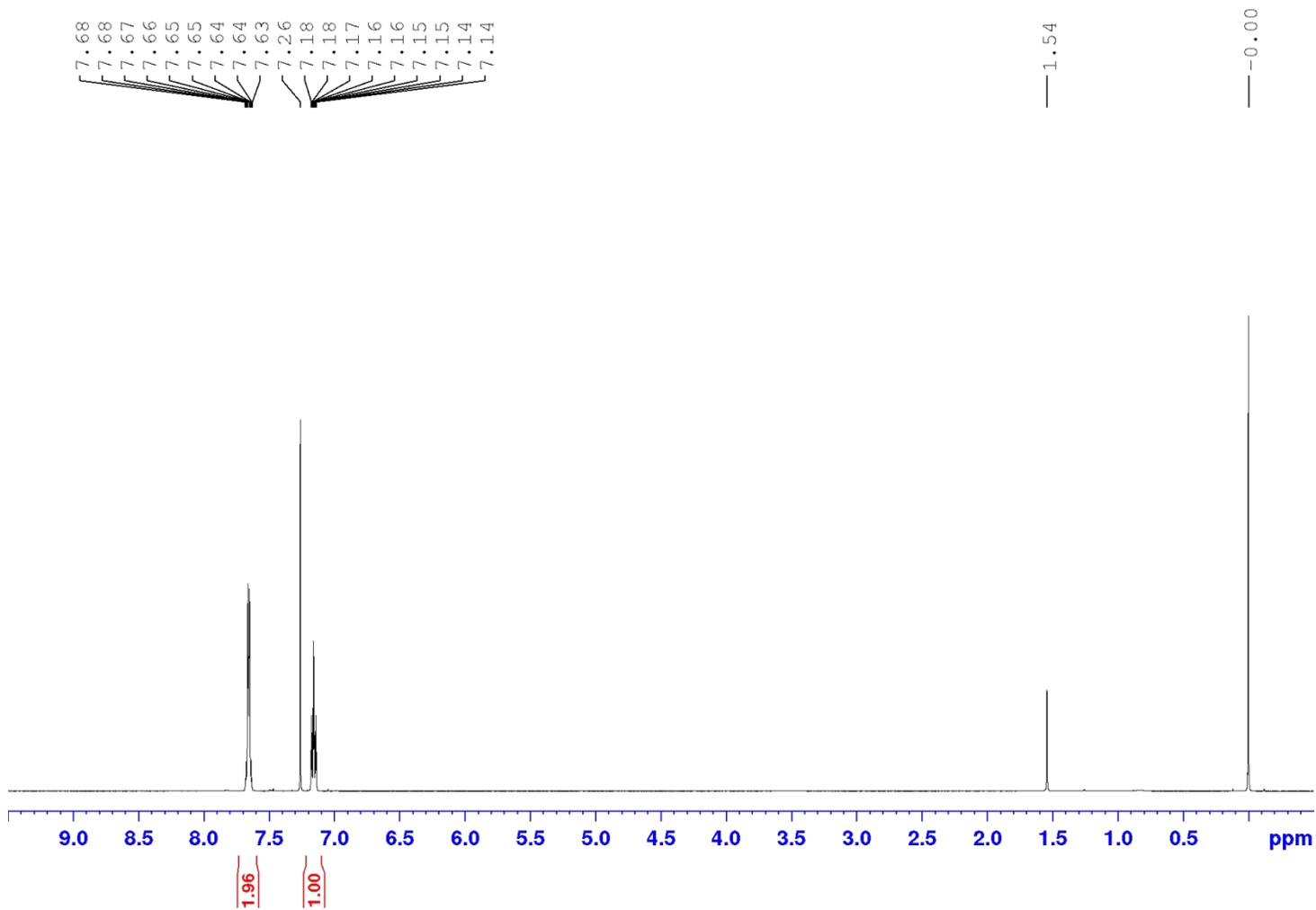


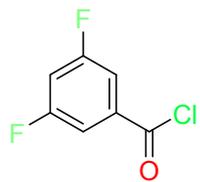
Compound **43** – ^{13}C NMR (125.5 MHz) – CDCl_3



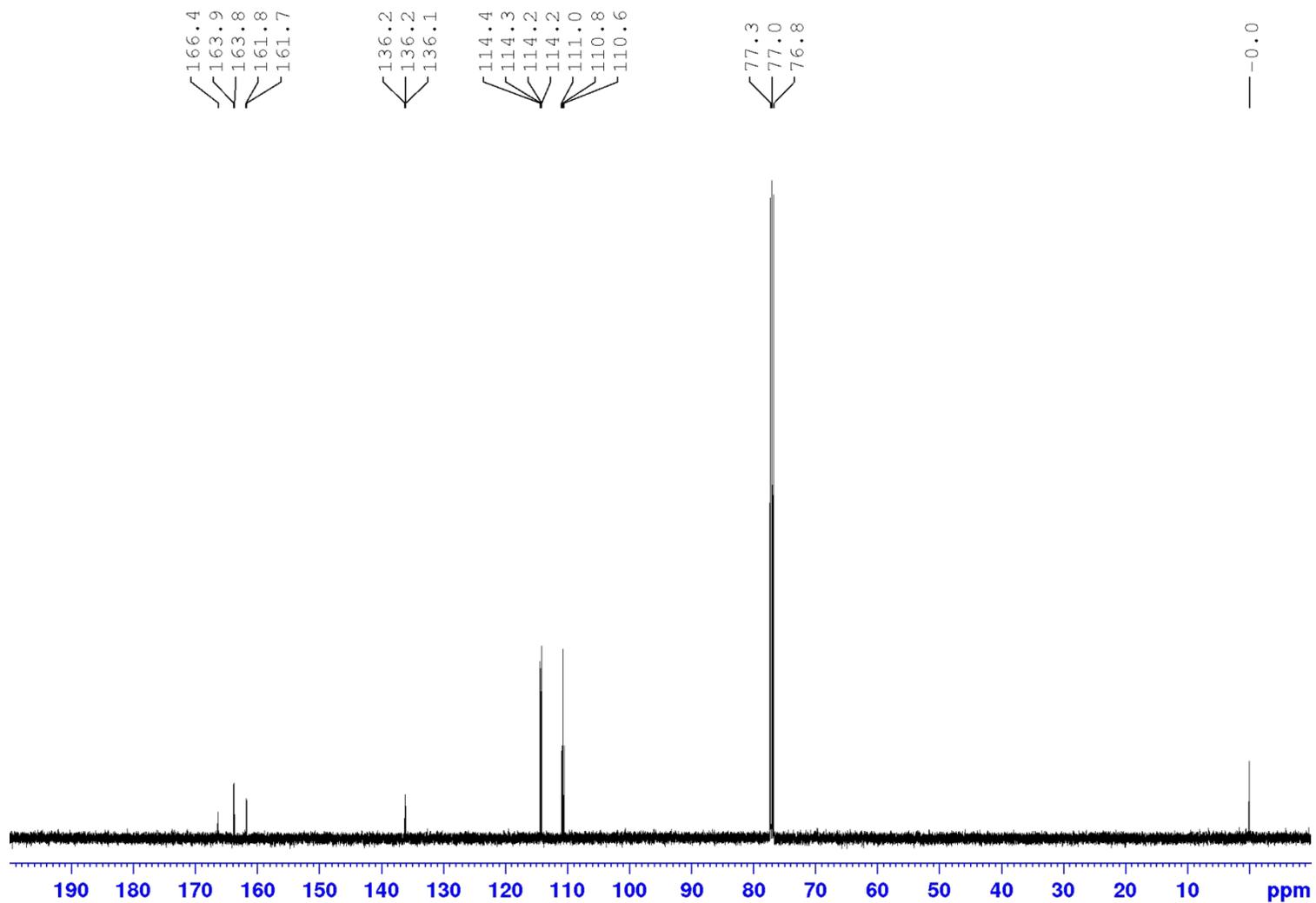


3,5-Difluorobenzoyl chloride – ^1H NMR (500 MHz) – CDCl_3





3,5-Difluorobenzoyl chloride – ^{13}C NMR (125.5 MHz) – CDCl_3



SUPPORTING REFERENCES

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- (2) Robert, X.; Gouet, P. Deciphering key features in protein structures with the new ENDscript server. *Nucleic Acids Res.* **2014**, 42, W320-324.
- (3) Ranzani, A. T. Discovery of Inhibitors and Structural Studies of the Malic Enzyme from *Trypanosoma cruzi* [dissertation] . Campinas: University of Campinas, 2017.
- (4) Ranzani, A. T.; Nowicki, C.; Wilkinson, S. R.; Cordeiro, A. T. Identification of Specific Inhibitors of *Trypanosoma cruzi* Malic Enzyme Isoforms by Target-Based HTS. *SLAS Discov.* **2017**, 22 (9), 1150–1161.