Modulating Polymeric Amphiphiles Using Thermoand pH-Responsive Copolymers with Cyclodextrin Pendant Groups through Molecular Recognition of Lipophilic Dye

Shintaro Kawano, \*<sup>†</sup>Jenni Lie, <sup>‡§</sup> Ryusei Ohgi, <sup>‡</sup> Motohiro Shizuma, <sup>†</sup> and Masahiro Muraoka <sup>‡</sup>

<sup>†</sup>Osaka Research Institute of Industrial Science and Technology (ORIST), 1-6-50 Morinomiya, Joto-ku, Osaka 536-8553, JAPAN

‡Department of Applied Chemistry, Faculty of Engineering, Osaka Institute of Technology (OIT),5-16-1 Ohmiya, Asahi-ku, Osaka 535-8585, JAPAN

§Department of Chemical Engineering, National Taiwan University of Science and Technology (NTUST), No. 43, Keelung Rd., Sec. 4, Da'an Dist., Taipei 10607, TAIWAN

#### Materials.

All the reagents and solvents were commercially available and were used as received unless otherwise specified. All aqueous solutions were prepared with distilled water.

### Measurements.

The product was characterized by proton nuclear magnetic resonance (<sup>1</sup>H-NMR), Fouriertransform infrared spectroscopy (FT-IR), and matrix-assisted laser desorption ionization-time-offlight mass spectrometry (MALDI-TOF-MS). The <sup>1</sup>H-NMR spectra (600 MHz) were obtained using a JEOL delta ECA system with DMSO- $d_6$  as the solvent for TsO- $\beta$ -CD and EDA- $\beta$ -CD. The <sup>1</sup>H-NMR spectra were analyzed by Delta NMR processing and control software v5.3 (JEOL RESONANCE, Inc). The FT-IR spectra of EDA- $\beta$ -CD and AEA- $\beta$ -CD were obtained using an FT/IR-4100 system (JASCO) with attenuated total reflectance (ATR). The molecular weights of the  $\beta$ -CD derivatives were determined by an AXIMA Confidence MALDI-TOF-MS spectrometer (Shimadzu Co. Ltd) with sodium trifluoroacetate as the ionizing agent and 2,5-dihydroxybenzoic acid as the matrix agent. DMSO and water were used as the solvents for TsO- $\beta$ -CD and EDA- $\beta$ -CD, respectively. Electrospray ionization-mass spectrometry (ESI-MS) was conducted by an LCMS-IT-TOF spectrometer (Shimadzu Corp.) to determine molecular weight of AEA- $\beta$ -CD. 1%w/v sample dissolved in water was injected into the LC-MS-IT-TOF spectrometer.





Scheme S1. Synthetic routes for TsO- $\beta$ -CD (A) and EDA- $\beta$ -CD (B).

#### Synthesis and characterization of the β-cyclodextrin-substituted monomer.

The mono-6-*O*-(*p*-toluenesulfonyl)- $\beta$ -cyclodextrin (TsO- $\beta$ -CD) was synthesized as described in the literature<sup>1</sup>. The reaction is described in **Scheme S1**. The reaction of  $\beta$ -CD and TsCl was conducted at a molar ratio of 1: 5 in 750 mL of 0.4 M NaOH solution. The  $\beta$ -CD (34.1 g) was mixed with the NaOH solution at 0 °C for 1 h in a round-bottomed flask equipped with a magnetic stirring bar and a thermometer. The cooling bath was then removed from the system, and a portion of TsCl (18.7 g) was added to the mixed solution, which was stirred vigorously for 2 h. Another portion of TsCl (10.0 g) was added and the mixture was further stirred for 2 h. The mixture was filtered to remove the unreacted TsCl, and the filtrate was neutralized using 10% HCl solution to produce a pH of 6–7, then stored overnight in a fridge (< 4 °C). The white precipitate of TsO- $\beta$ -

CD was filtered and dried under vacuum in an oven at 40 °C for 24 h. The product was recrystallized three times by dissolving in 300 mL of boiling water and cooling at < 4 °C. The purified product was dried under vacuum in an oven at 40 °C for 24 h to produce mono-TsO- $\beta$ -CD at a yield of 19%. <sup>1</sup>H-NMR (600 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  (ppm) 7.76 (d, *J* = 8.2 Hz, 2H, Ar-*H*), 7.44 (d, *J* = 7.6 Hz, 2H, Ar-*H*), 5.84–5.64 (m, 14H, 2, 3-O*H* in  $\beta$ -CD), 4.84 (d, *J* = 46.1 Hz, 7H, 1-*H* in  $\beta$ -CD), 4.52–4.35 (m, 6H, 6-O*H* in  $\beta$ -CD), 4.20–4.17 (m, 2H, 6'-*H* in  $\beta$ -CD), 3.65–3.20 (m, 40H, 2, 3, 4, 5, 6-*H* in  $\beta$ -CD), 2.43 (s, 3H, Ar-*CH*<sub>3</sub>); the purity was 97%. MALDI-TOF-MS: *m/z* calculated for [M-Na]<sup>+</sup> C<sub>49</sub>H<sub>76</sub>O<sub>37</sub>S<sup>+</sup>, 1311.16 found 1311.34.

The ethylene diamino β-CD (EDA-β-CD) was synthesized by mixing 5.16 g of TsO-β-CD in 30 mL of EDA at 75 °C under degassed N<sub>2</sub> for 5 h (**Scheme S1B**). To terminate the reaction, the mixed solution was cooled to room temperature, then added to cold acetone (300 mL). The resulting solid precipitate of EDA-β-CD was filtered under vacuum. Subsequently the precipitate was dissolved in a mixed solution comprising 40 mL of water and methanol (1:3), and was reprecipitated by adding dropwise to a solution of cold acetone. The reprecipitation and filtration processes were repeated three times, and the resulting EDA-β-CD was collected and dried at 60 °C under vacuum in an oven to produce the purified product (4.22 g, 90% yield). <sup>1</sup>H-NMR (600 MHz, D<sub>2</sub>O) δ (ppm) 4.92 (s, 7H, 1-*H* in β-CD), 3.83–3.68 (m, 26H, 3, 5, 6-*H* in β-CD), 3.51–3.41 (m, 14H, 2, 4-*H* in β-CD), 3.32 (t, *J* = 9.3 Hz, 1H, -CH<sub>2</sub>-CH<sub>2</sub>-NH-CH<sub>2</sub>-), 2.91 (dd, *J* = 12.7, 2.4 Hz, 1H, 6'-*H* in β-CD), 2.73–2.67 (m, 3H, 6'-*H* in β-CD and NH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-NH-), 2.60 (t, *J* = 7.2 Hz, 2H, NH<sub>2</sub>-CH<sub>2</sub>-CH<sub>2</sub>-NH-). MALDI-TOF-MS: *m/z* calculated for [M-Na]<sup>+</sup> C<sub>44</sub>H<sub>76</sub>O<sub>34</sub>N<sub>2</sub><sup>+</sup>, 1199.1 found 1200.4.



**Figure S1.** <sup>1</sup>H-NMR spectrum (600 MHz, DMSO- $d_6$ , 25 °C) of TsO- $\beta$ -CD.



**Figure S2.** MALDI-TOF-MS of TsO-β-CD.



Figure S3. <sup>1</sup>H-NMR spectrum (600 MHz,  $D_2O$ , 25 °C) of EDA- $\beta$ -CD.





Figure S6. ESI-MS (positive) of AEA-β-CD.



Figure S7. FT-IR spectra of EDA- $\beta$ -CD (A) and AEA- $\beta$ -CD (B).

# Synthesis of poly (*N*-isopropylacrylamide-*co*-acrylic acid-*co*-AEA-β-CD) (poly(NIPAM-AAβ-CD)) and poly (*N*-isopropylacrylamide-*co*-acrylic acid) (poly(NIPAM-AA)).

Poly(NIPAM-*co*-AA) without the CD moiety in the polymer segment was synthesized by the free radical polymerization of the NIPAM and AA monomers dissolved in toluene (the feed monomer ratio was 4:1 for NIPAM to AA). AIBN was added at a concentration of 0.5 mol% relative to the total concentration of the monomers. Another procedure was carried out in essentially the same manner as that described for the production of poly(NIPAM-AA- $\beta$ -CD). After dialysis, the purified solution was freeze-dried for 3 days and a white solid was produced (78.1% yield). <sup>1</sup>H-NMR (600 MHz, D<sub>2</sub>O).  $\delta$  (ppm) 3.74 (br, 1H, -NH-CH-(CH<sub>3</sub>)<sub>2</sub> in PNIPAM), 1.97–1.43 (br, 4H, -CH-CH<sub>2</sub>- in the poly(NIPAM-*co*-AA) chain), 0.99 (br, 6H, -NHCH(CH<sub>3</sub>)<sub>2</sub> in PNIPAM).



Figure S9. <sup>1</sup>H-NMR spectrum (600 MHz, D<sub>2</sub>O, 25 °C) of Poly(NIPAM-AA).

Characterization of copolymer, solution property and the functionality.



Figure S10. GPC curves of (A) Poly(NIPAM-AA- $\beta$ -CD) and (B) Poly(NIPAM-AA).



**Figure S11.** DLS size distributions (intensity% and number%) of Poly(NIPAM-AA- $\beta$ -CD) at each pH as function of temperature at 25 (pH 4.0) and 50 °C.



**Figure S12.** DLS count rates versus the concentrations of the Poly(NIPAM-AA- $\beta$ -CD) solutions at each pH (25 °C).



**Figure S13.** UV-Vis absorption spectra of NR-incorpolated complex solutions of Poly(NIPAM-AA- $\beta$ -CD) at each pH,  $\beta$ -CD and Poly(NIPAM-AA) (< pH= 4.0) after subtracting the absorption base line of each naked sample.



**Figure S14.** Fluorescence emission spectra of the NR-incorporated complex solutions of Poly(NIPAM-AA- $\beta$ -CD) (pH 5.8) (A, B) and  $\beta$ -CD (C, D). Spectra B and D show the magnified images at the concentrations from 0.001 to 0.1 mg/ mL.

## REFERENCE

 Brady, B.; Lynam, N.; O'Sullivan, T.; Ahern, C. ; Darcy, R. 6A-O-p-Toluenesulfonyl-β-Cyclodextrin. Org Synth. 2000, 77, 220-222.