

Synthesis of mannosidase-stable Man₃ and Man₄ glycans containing S-linked Man α 1 \rightarrow 2Man termini

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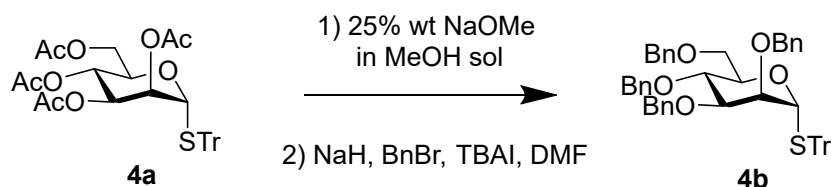
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General Experimental Methods

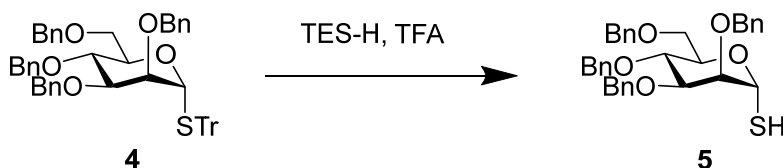
Reactions were carried out in oven-dried glassware under a nitrogen atmosphere and were stirred magnetically. All reagents were purchased from Sigma-Aldrich, Acros Organics, Fluka, Alfa Aesar or TCI, and used without further purification unless otherwise noted. Molecular sieves were crushed and freshly activated prior to use by heating on gas flame in quartz round bottom flask under < 1 Torr on a vacuum manifold attached to a Welch 1400 vacuum pump. The word “concentrated” refers to removal of solvents by means of rotary evaporator attached to a Welch 1400 oil pump (bled to 5-300 mm Hg as needed) followed by removal of residual solvents at < 1 Torr on a vacuum manifold attached to a Welch 1400 vacuum pump. SiliaFlash® F60 (230-400 mesh) from Silicycle® was used for flash column chromatography unless specifically indicated. Analytical thin layer chromatography (TLC) was performed using silica gel F-354 pre-coated glass plates (0.25 mm). TLC plates were analyzed by short wave UV illumination or dipping in cerium-ammonium-molybdate (CAM) stain (40g of ammonium pentamolybdate, 1.6g of Cerium (IV) sulfate, 800 mL of diluted sulfuric acid (1:9, with water, v/v) and heating on a hot plate. Tetrahydrofuran (THF), dichloromethane (DCM) and toluene, were purified by degassing with argon and passage through activated alumina columns. ¹H, ¹³C, ¹⁹F and HSQC NMR were recorded on a Varian Innova 400 MHz and Bruker 800 MHz in deuterated solvents at ambient temperature unless otherwise noted. Spectra acquired in CDCl₃ were internally referenced to TMS (0 ppm) or residual CDCl₃ (7.26 for ¹H and 77.06 for ¹³C). ¹⁹F Fluorine spectra were referenced to trifluorotoluene (-63.72 ppm), internally or by substituting a tube containing standard. Spectra acquired in D₂O were referenced to sodium 3-(trimethylsilyl) propane sulfonate by the substitution method, or referenced to the HDO peak using temperature-specific values. Chemical shifts are reported in parts per million (ppm). Coupling constants are reported in Hz with multiplicities denoted as s (singlet), d (doublet), dd (doublet of doublets), t (triplet), q (quartet), m (multiplet), app (apparent) and br (broad). High Resolution mass spectra (HRMS) were performed by Mass Spectrometry Laboratory, University of Illinois at Urbana-Champaign using electron impact (EI) or electrospray (ESI) ionization and a TOF analyzer. LC/MS analysis was performed at Brandeis University on a Waters Acquity UPLC chromatograph with reverse phase C18 and C4 column, and Waters Photodiode Array and Micro mass ZQ4000 mass detectors. IR spectra were recorded on a Varian 640-IR FT-IR spectrometer and are reported in wavenumbers (cm⁻¹). Optical rotation was measured on JASCO-polarimeter.

Experimental Section



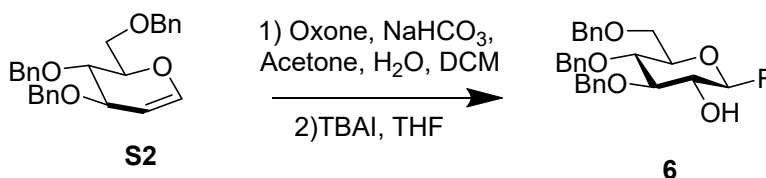
Triphenylmethyl 2,3,4,6-tetra-*O*-benzyl-1-thio- α -D-mannopyranoside (**4b**)

NaOMe in methanol (25% wt; 565 μ L; 2.47 mmol; 0.3 equiv.) was added to a solution of mannoside **4a**¹ (5 g; 8.24 mmol; 1 equiv.) dissolved in dry dichloromethane and methanol (1:1; 50 mL) at rt with stirring for 4h. The solution was adjusted to pH 6-7 with Amberlite IR-120 resin (H^+ form), filtered, and the filtrate evaporated to dryness under reduced pressure to give the tetraol, which was coevaporated with toluene two times and used without purification for next step. Crude tetraol (3.6 g; 8.21 mmol; 1 equiv.) was dissolved in dry DMF (30 ml) and cooled in ice bath. Sodium hydride (1.97 g; 49.25 mmol; 6 equiv; 60%) was added to reaction mixture and stirred for 30 minutes at 0 $^{\circ}\text{C}$, followed by dropwise addition of benzyl bromide (5.85 mL; 49.25 mmol; 6 equiv.) and TBAI (2.1 g; 2.21 mmol; 1 equiv.). The reaction mixture was stirred for 6 hours at rt and quenched with 2 mL of methanol at 0 $^{\circ}\text{C}$. The reaction mixture was then diluted with EtOAc and washed with brine, then dried over MgSO_4 , filtered and the solvent evaporated under reduced pressure. The crude was purified by column chromatography (1:5; EtOAc: hexane) to give compound **4b** as a yellow syrup (4.4 g; 5.58 mmol) in 68% yield. ^1H NMR (400 MHz, CDCl_3) δ 7.50 – 7.10 (multiple signals, 35H + residual CHCl_3), 4.87 (d, J = 10.8 Hz, 1H), 4.83 (d, J = 1.6 Hz, 1H), 4.69 (d, J = 12.1 Hz, 1H), 4.58 – 4.41 (m, 4H), 4.25 – 4.17 (m, 1H), 4.15 (d, J = 12.6 Hz, 1H), 3.99 (t, J = 9.6 Hz, 1H), 3.89 – 3.70 (m, 4H), 3.61 (app. t, J = 2.4 Hz, 1H). ^{13}C NMR (100 MHz, CDCl_3 , Selected signals): δ 144.7, 138.6, 138.4, 138.3, 138.2, 130.1, 128.3, 128.2, 128.1, 127.9, 127.9, 127.8, 127.6, 127.5, 127.4, 126.9, 81.8, 80.4, 78.1, 75.1, 74.6, 74.5, 71.9, 70.6, 69.8, 69.3. $[\alpha]_{\text{D}}^{25}$: +76.1 (c1.0, CHCl_3). IR (cm^{-1}): 3028, 2863, 1949, 1451, 1087. HRMS (ESI-TOF) m/z : $[\text{M}+\text{Na}]^+$ Calcd. for $\text{C}_{53}\text{H}_{50}\text{O}_5\text{SNa}$: 821.3277, found 821.3294.



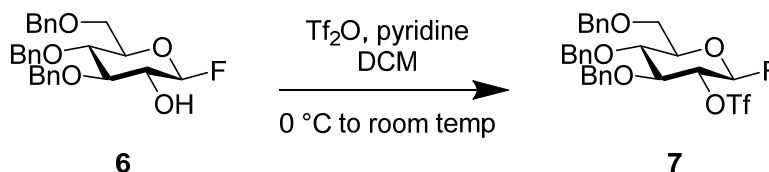
2,3,4,6-Tetra-*O*-benzyl- α -D-mannopyranosyl thiol (**5**)

Triethyl silane (1.5 mL; 9.39 mmol; 5 equiv.) and TFA (3 mL, 30.04 mmol; 16 equiv.) were added to a solution of mannosyl trityl thiol **4** (1.5g; 1.88 mmol; 1 equiv.) dissolved in dry dichloromethane (30 mL) under a N₂ atm. After 30 minutes the reaction had reached completion and the solvents were evaporated followed by co-evaporation with toluene and kept for high vacuum for 3 hours to afford crude of compound **5** as yellow oil, which was used in next steps without purification. ¹H NMR (400 MHz, CDCl₃) δ 7.67 – 6.76 (multiple signals, 20H + residual CHCl₃+ triphenylmethane), 5.66 (dd, *J* = 7.0, 1.8 Hz, 1H), 4.87 (d, *J* = 10.7 Hz, 1H), 4.71 (d, *J* = 12.5 Hz, 1H), 4.67 – 4.45 (multiple signals, 6H), 4.11 (ddd, *J* = 9.4, 5.0, 1.8 Hz, 1H), 4.01 (app t, *J* = 9.4 Hz, 1H), 3.86 (dd, *J* = 9.4, 3.0 Hz, 1H), 3.81 (dd, *J* = 3.0, 1.8 Hz, 1H), 3.68 (dd, *J* = 10.8, 1.8 Hz, 1H), 2.02 (d, *J* = 7.0 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃, Selected Signals) δ 138.2, 138.1, 138.0, 137.8, 128.3, 128.3, 128.3, 128.2, 128.2, 127.9, 127.8, 127.8, 127.8, 127.7, 127.7, 127.6, 127.5, 127.5, 79.0, 75.1, 76.2, 74.8, 73.3, 72.5, 72.1, 72.0, 68.9, 67.9, 56.8. HRMS (ESI-TOF) *m/z*: [M+Na]⁺: Calcd. for C₃₄H₃₆O₅SNa: 579.2181, found 579.2173.



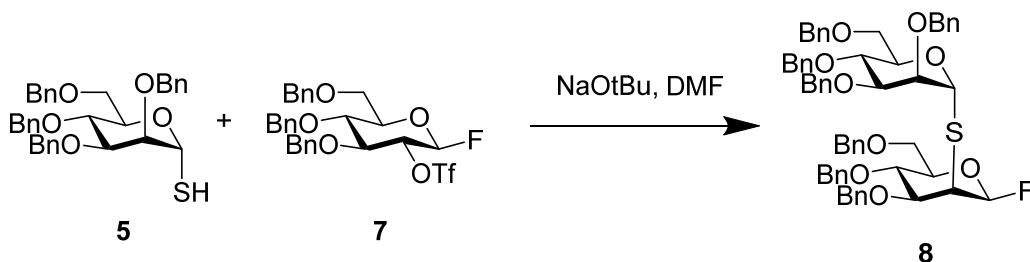
3,4,6-Tri-*O*-benzyl-β-D-glucopyranosyl fluoride (**6**)

To a vigorously stirred, cooled biphasic solution of 3,4,6-tri-*O*-benzyl-D-glucal **S2**² (6 g, 14.42 mmol; 1 equiv.) in dichloromethane (60 mL), acetone (6 mL) and sat. aq. NaHCO₃ (100 mL), a solution of oxone (17.74 g; 57.68 mmol; 4 equiv.) dissolved in water (36 mL) was added dropwise over 20 minutes. The reaction mixture was stirred vigorously for 2 hours at room temperature. The organic layer was separated, and aqueous layer was extracted twice with 140 mL of dichloromethane. The combined organic layers were dried by MgSO₄ and concentrated under reduced pressure to afford white solid 1,2-anhydro-3,4,6-tri-*O*-benzyl-α/β-D-glucopyranose, which was used for next reaction without purification and dissolved in anhydrous THF (60 mL) and cooled to 0 °C. 1M Tetrabutylammonium fluoride in THF³ (28.2 mL; 28.21 mmol; 2 equiv.) was added dropwise and stirred for 6 hours. The reaction mixture was extracted with dichloromethane 150 mL two times and combined organic layers were dried over MgSO₄, filtered and concentrated under reduced pressure. The crude was purified by column chromatography (1:4; EtOAc: hexane) to afford compound **6** (1.8 g; 8.32 mmol) as yellow amorphous solid in 29% yield over two steps. ¹H NMR (400 MHz, CDCl₃) δ 7.42 – 7.12 (multiple signals, 15H + residual CHCl₃), 5.14 (dd, ¹*J*_{FH} = 53.1, *J*_{H1H2} = 6.4 Hz, 1H), 4.81 (app s, 2H), 4.76 (d, *J* = 10.9 Hz, 1H), 4.59 (d, *J* = 12.1 Hz, 1H), 4.56 (d, *J* = ~11 Hz, 1H), 4.53 (d, *J* = 12.1 Hz, 1H), 3.80 – 3.63 (m, 5H), 3.58 (app t, *J* = 8.2 Hz, 1H), 2.56 (d, *J* = 3.9 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 138.1, 137.8, 137.6, 128.5, 128.4, 128.4, (127.92, 127.91, 127.84, 127.82, 127.71, 6 resonances with an overlap), 109.0 (C-F, d, ¹*J*_{CF} = 215.1 Hz), 81.1 (C-F d, ³*J*_{CF} = 9.3 Hz), 76.3, 74.86, 74.82, 74.5, 73.5, 73.0 (C-F, d, ²*J*_{CF} = 23.7 Hz), 68.48. ¹⁹F NMR (376 MHz, CDCl₃) δ -140.13 (dd, *J*_{HF} = 53.2, 11.7). [α]_D²⁵: +59.76 (c1.0, CHCl₃). IR (cm⁻¹): 3431.3, 2872.1, 2362.2, 2019.1, 1736.4, 1080.1, 1027.7. HRMS (ESI-TOF) *m/z*: [M+Na]⁺: Calcd. for C₂₇H₂₉FO₅Na: 475.1897, found 475.1898.



3,4,6-tri-*O*-benzyl-2-*O*-trifluoromethylsulfonyl-β-D-glucopyranosyl fluoride (7)

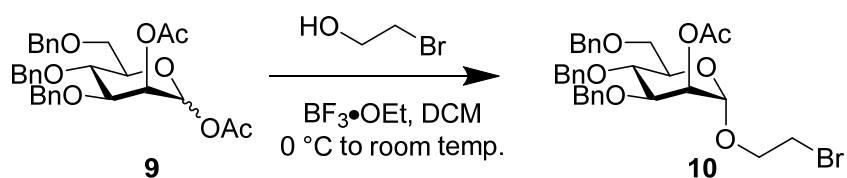
To a solution of compound **6** (1 g; 2.21 mmol; 1 equiv.) and pyridine (2.19 mL, 22.1 mmol, 10 equiv.) in 15 mL of CH₂Cl₂ was added Tf₂O (408 μL; 2.43 mmol; 1.1 equiv.) at 0 °C under N₂ atm. After 0.5 hour at 0 °C and 1 h at ambient temperature, the solution was diluted with 30 mL of CH₂Cl₂ and washed with ice-cold sat aq. NaHCO₃, ice-cold solution of CuSO₄, and brine. The organic layer was then dried over MgSO₄, filtered and concentrated in vacuo to afford 1.21 g (2.06 mmol; 93%) of compound **7** as yellow liquid which was used for next reaction without further purification. ¹H NMR (400 MHz, CDCl₃) δ 7.34 – 7.12 (multiple signals, 15H + residual CHCl₃), 5.301 (dd, ¹J_{FH} = 52.4 Hz, ¹J_{H1H2} = 7.0 Hz, 1H), 4.87 – 4.70 (multiple signals, 4H), 4.60 – 4.48 (multiple signals, 3H), 3.84 (app t, *J* = 9.1 Hz, 1H), 3.77 (app t, *J* = 9.1 Hz, 1H), 3.739 – 3.668 (m, 2H), 3.61 (ddd, *J* = 9.4 Hz, 2.9 Hz, 2.9 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 137.27, 136.98, 136.56, (128.31, 128.26, 127.89, 127.67, 127.65, 127.62; 9 resonances with overlaps), 118.17 (C-F, q, ¹J_{CF} = 319.54 Hz), 105.15 (C-F, d, ¹J_{CF} = 219.07 Hz), 83.90 (C-F, d, ²J_{CF} = 24.7 Hz), 80.32 (C-F, d, ³J_{CF} = 8.4 Hz), 75.56, 75.14 (C-F, d, long range *J*_{CF} = 4.3 Hz), 74.89, 73.40, 67.38. ¹⁹F NMR (376 MHz, CDCl₃) δ -74.38 (d, ⁶J_{FF} = 9.7 Hz), -140.11 (ddq, ¹J_{HF} = 52.4, ²J_{HF} = 9.7 Hz, ⁶J_{FF} = 9.7 Hz). LRMS (ESI-single quad) *m/z*: [M+NH₄]⁺. Calcd. for C₂₈H₃₂F₄O₇NS: 602.1836, Found: 602.15 (this compound was too unstable to send for HRMS).



3,4,6-tri-*O*-benzyl-2-*S*-(2,3,4,6-tetra-*O*-benzyl-α-D-mannopyranosyl-(1→2))-β-D-mannopyranosyl fluoride (8)

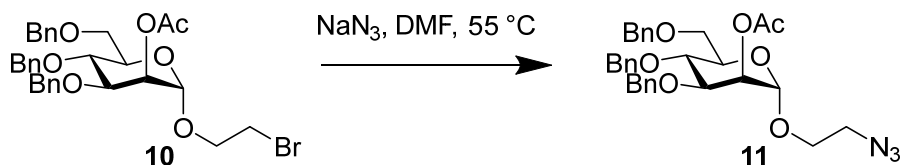
To a solution of freshly prepared crude compound **5** and compound **7** (1.21 g; 2.07 mmol; 1.1 equiv.) in 20 mL of dry DMF was added NaO^tBu (181 mg; 1.89 mmol; 1 equiv.) under N₂ atm. After 30 min, the reaction mixture was diluted with 50 mL of EtOAc and washed with ice-cold brine solution. Combined organic layers were dried over MgSO₄, filtered and evaporated under reduced pressure maintaining water bath temperature 30 °C. The crude was purified by column chromatography by using triethyl amine quenched silica gel (1:5; EtOAc: hexane) to afford compound **8** (1.18 g; 1.19 mmol) as yellow amorphous solid in 63% yield. ¹H NMR (400 MHz,

CDCl₃) δ 7.33 – 7.16 (multiple signals, 35H + residual CHCl₃), 5.48 (dd, $J_{F,H}$ = 53.15 Hz, $J_{H,H}$ = 2.2 Hz, 1H), 5.50 (s, 1H), 4.88 (d, J = 11.0 Hz, 1H), 4.72–4.63 (multiple signals, 2H), 4.62 – 4.43 (multiple signals, 12H), 4.27 (dd, J = 9.6, 4.1 Hz, 1H), 4.03 (app t, J = 9.5 Hz, 1H), 3.92 (dd, J = 9.3, 3.1 Hz, 1H), 3.88 – 3.79 (multiple signals, 5H), 3.77 – 3.65 (multiple signals, 3H), 3.60 (app dt, J = 16.8, 2–3 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 138.5, 138.3, 138.2, 138.1, 138.0, 137.7, 137.3, 128.41 (2C), 128.38 (2C), 128.29 (2C), 128.26 (2C), 128.21 (2C), 128.19 (4C), 127.86 (2C), 127.85, 127.84, 127.79 (2C), 127.69 (2C), 127.67 (2C), 127.66 (4C), 127.56 (2C), 127.54, 127.52, 127.49, 127.4, 127.3, 106.1 (C-F, d, $^1J_{CF}$ = 221.9 Hz), 82.1, 80.2, 77.6 (C-F, d, $^3J_{CF}$ = 3.32 Hz), 76.2, 74.9 (2C), 74.2, 73.31, 73.30, 73.1, (72.7, 72.5, 72.2, 71.9, 71.8, 6C with 2 overlapping signals), 69.4, 69.0, 46.79 (C-F, d, $^2J_{CF}$ = 22.10 Hz). ¹⁹F NMR (376 MHz, CDCl₃) δ -129.83 (dd, J = 53, 16.8 Hz). $[\alpha]_D^{25}$: +12.65 (c1.0, CHCl₃). IR (cm⁻¹): 3029, 2866, 2251, 1973, 1363, 1087. HRMS (ESI-TOF) m/z : $[M+Na]^+$: Calcd. for C₆₁H₆₃FO₉SNa: 1013.4075, found 1013.4073.



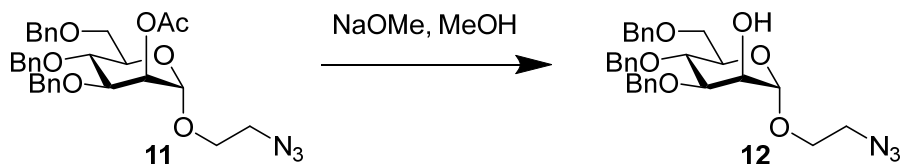
2-Bromoethyl-2-O-acetyl-3,4,6-tri-O-benzyl- α -D-mannopyranoside (**10**)

A solution of 1,2-di-O-acetyl-3,4,6-tri-O-benzyl- α/β -D-mannopyranose **9**⁴ (1.1g; 2.06 mmol; 1 equiv.) and bromoethanol (365 μ L; 5.14 mmol; 2.5 equiv.) in dry dichloromethane (11 mL) was cooled to 0 °C and BF₃·OEt₂ (1.41 mL; 10.29 mmol; 5 equiv.) was added dropwise. The ice bath was allowed to melt and warm to room temperature as the stirred overnight. The reaction mixture was diluted with 40 mL of dichloromethane and washed with cold aq. sat. NaHCO₃ and brine solution. The organic layers were dried over MgSO₄, concentrated and crude was purified by column chromatography (ethyl acetate: hexane; 3:1) to afford compound **10** (925 mg; 1.54 mmol) as a syrup in 75% yield. ¹H NMR (400 MHz, CDCl₃) δ 7.35 – 7.21 (multiple signals, 13H + residual CHCl₃), 7.18 – 7.13 (m, 2H), 5.38 (dd, J = 3.4, 1.8 Hz, 1H), 4.89 (d, J = 1.8 Hz, 1H), 4.86 (d, J = 10.8 Hz, 1H), 4.71 (d, J = 11.2 Hz, 1H), 4.67 (d, J = 12.4 Hz, 1H), 4.55 (d, J = 11.2 Hz, 1H), 4.51 (d, J = 12.4 Hz, 1H), 4.47 (d, J = 10.8 Hz, 1H), 4.02 – 3.91 (m, 2H), 3.91 – 3.84 (m, 2H), 3.84 – 3.74 (m, 2H), 3.71 (d, J = 10.7 Hz, 1H), 3.47 (t, J = 6.1 Hz, 2H), 2.15 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ (carbonyl C not observed), 138.3, 138.1, 137.9, 128.4 (2C), 128.3 (4C), 128.1 (2C), 127.9 (2C), 127.74 (2C), 127.73, 127.63, 127.58, 98.0, 78.1, 75.2, 74.2, 73.4, 71.9, 71.8, 68.8, 68.6, 67.9, 29.9, 21.1. $[\alpha]_D^{25}$: +3.2 (c1.0, CHCl₃). IR (cm⁻¹): 2915, 2015, 1745, 1237, 1095. HRMS (ESI-TOF) m/z : $[M+Na]^+$: Calcd. for C₃₁H₃₅BrO₇Na : 621.1464, found 621.1453.



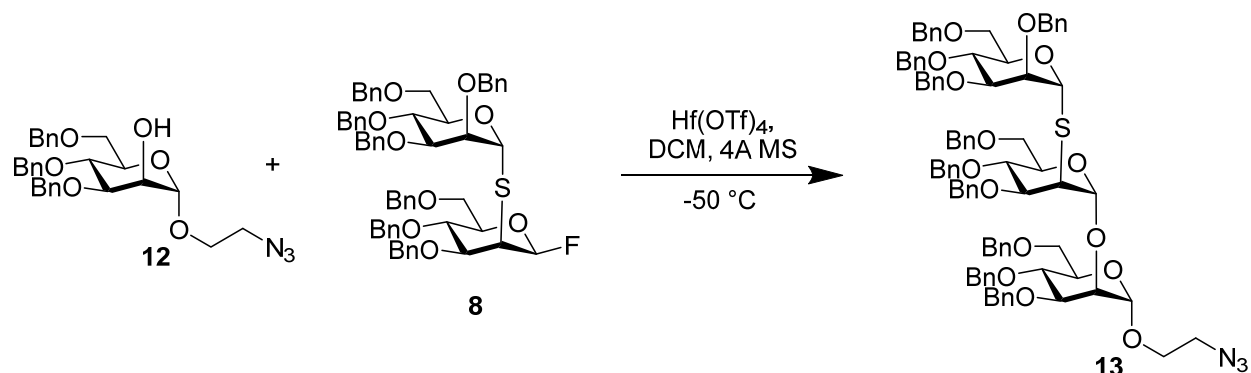
2-azidoethyl-2-*O*-acetyl-3,4,6-tri-*O*-benzyl- α -D-mannopyranoside (**11**)

A solution of the compound **10** (910 mg; 1.52 mmol; 1 equiv.) and sodium azide (493 mg; 7.59 mmol; 5 equiv.) in dry DMF (25 mL) was stirred at 55 °C for 6 hours. After completion of the reaction, solvent was co-evaporated with toluene under reduced pressure, and the residue was dissolved in the mixture of water and ethyl acetate and extracted with ethyl acetate. The organic layers were dried over MgSO₄, concentrated and residue was purified by column chromatography (ethyl acetate and hexane; 2:1) to afford compound **11** as colorless syrup (826 mg; 1.47 mmol) in 97% yield. ¹H NMR (400 MHz, CDCl₃) δ 7.30 (multiple signals + residual CHCl₃, 13H), 7.15 (dd, J = 7.3, 2.2 Hz, 2H), 5.39 (app t, J = 1.9 Hz, 1H), 4.89 (d, J = 1.8 Hz, 1H), 4.86 (d, J = 10.8 Hz, 1H), 4.68 (dd, J = 11.6, 9.6 Hz, 2H), 4.01 (dd, J = 8.9, 3.3 Hz, 1H), 3.94 – 3.75 (m, 4H), 3.71 (dd, J = 10.4, 1.5 Hz, 1H), 3.65 – 3.57 (m, 1H), 3.44 – 3.31 (m, 2H), 2.14 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 170.4, 138.3, 138.1, 137.8, 128.4, 128.3, 128.3, 128.1, 127.8, 127.7, 98.0, 78.0, 75.1, 74.1, 73.4, 71.9, 71.7, 68.8, 68.6, 66.7, 50.4, 21.1. $[\alpha]_D^{25}$: +1.9 (c1.0, CHCl₃). IR (cm⁻¹): 3030, 2866, 2103, 1743, 1233, 1088. HRMS (ESI-TOF) m/z : $[M+Na]^+$: Calcd. for C₃₁H₃₅N₃O₇Na: 584.2373, found 584.2372.



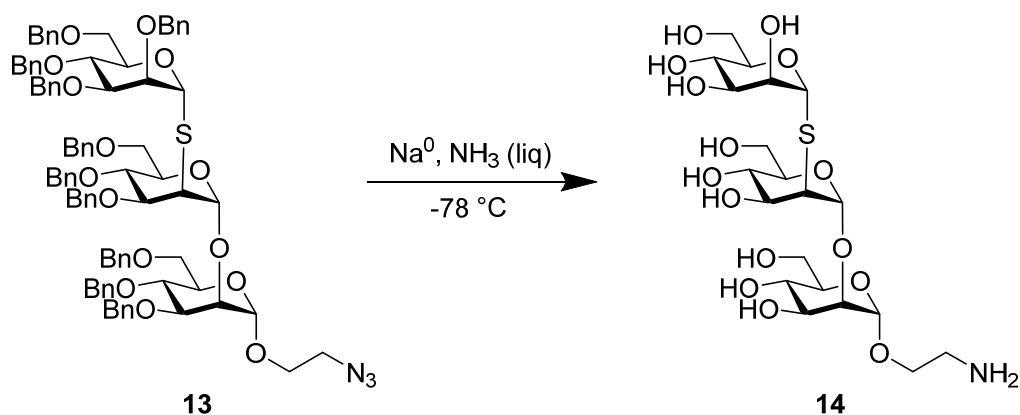
2-azidoethyl-3,4,6-tri-*O*-benzyl- α -D-mannopyranoside (**12**)

Compound **11** (810 mg; 1.44 mmol; 1 equiv.) was dissolved in methanol (4 mL) and dichloromethane (4 mL) and 65 μ L (289 μ mol; 0.2 equiv.) 25% sodium methoxide in methanol solution was added and stirred reaction mixture for 2 hours at room temperature. Reaction was neutralized with Amberlite IR-120 H⁺ form ion exchange resin. Resin was filtered off and the reaction mixture was concentrated under reduced pressure. The residue was purified by column chromatography to afford compound **12** (704 mg; 1.36 mmol) as colorless syrup in 94% yield. ¹H NMR (400 MHz, CDCl₃) δ 7.35 – 7.23 (multiple signals, 13H + residual CHCl₃), 7.21–7.14 (m, 2H), 4.94 (d, J = 1.7 Hz, 1H), 4.82 (d, J = 10.8 Hz, 1H), 4.68 (app s, 2H), 4.63 (d, J = 12.2 Hz, 1H), 4.52 (d, J = 12.2 Hz, 1H), 4.50 (d, J = 10.8 Hz, 1H), 4.05 (br s, 1H), 3.92 – 3.66 (multiple signals, 6H), 3.61 (ddd, J = 10.6, 6.7, 3.7 Hz, 1H), 3.44 – 3.27 (m, 2H), 2.54 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 138.3, 138.2, 137.8, 128.5, 128.4, 128.3, 127.95, 127.93, 127.92, 127.8, 127.7, 127.6, 99.4, 80.0, 75.1, 74.1, 73.5, 72.1, 71.4, 68.9, 68.1, 66.6, 50.5. $[\alpha]_D^{25}$: +22.3 (c1.0, CHCl₃). IR (cm⁻¹): 3436, 3030, 2914, 2101, 1960, 1495, 1060. HRMS (ESI-TOF) m/z : $[M+Na]^+$: Calcd. for C₂₉H₃₃N₃O₇Na: 542.2267, found 542.2265.



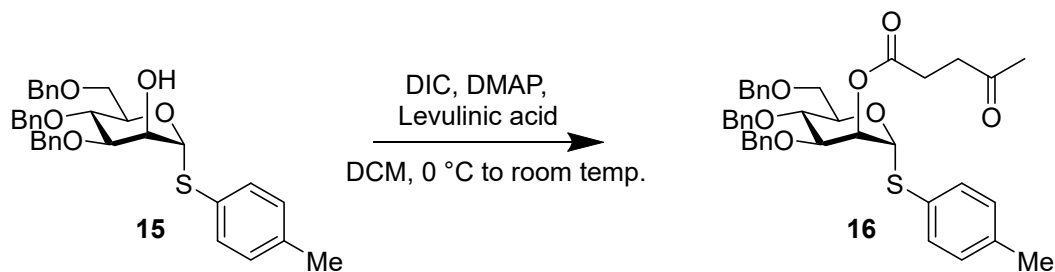
2-azidoethyl-3,4,6-tri-*O*-benzyl-2-*O*-[3,4,6-tri-*O*-benzyl-2-*S*--α-*D*-mannopyranosyl-(1→2)]-α-*D*-mannopyranoside (13**)**

Disaccharide donor **8** (300 mg; 303 μmol; 1 equiv.) and acceptor **12** (235 mg; 454 μmol; 1.5 equiv.) were dissolved in of anhydrous dichloromethane (3 mL), and freshly crushed and flame dried 4 Å molecular sieves were added and stirred for 15 min at room temperature. The mixture was cooled at -50 °C, then Hf(OTf)₄ (234 mg; 303 μmol; 1 equiv.) was added and allowed to stir for next 1 h. The reaction was neutralized with a few drops of triethylamine, diluted with CH₂Cl₂, filtered through a celite pad and concentrated. Purification by column chromatography (EtOAc:Hexane, 1:5) afforded trisaccharide **13** as white foam in 64% yield (287 mg, 194 μmol). ¹H NMR (400 MHz, CDCl₃) δ 7.40 – 7.08 (multiple signals, 50H + residual CHCl₃), 5.59 (s, 1H), 5.33 (s, 1H), 4.97 (d, *J* = 1.8 Hz, 1H), 4.94 – 4.80 (multiple signals, 3H), 4.71 – 4.45 (multiple signals, 15H), 4.40-4.32 (multiple signals, 2H), 4.23 (dd, *J* = 9.3, 4.3 Hz, 1H), 4.18 – 4.10 (m, 1H), 4.09 – 4.02 (m, 2H), 4.01 – 3.95 (m, 1H), 3.94 – 3.89 (m, 2H), 3.88 – 3.64 (multiple signals, 11H), 3.59 (dd, *J* = 11.0, 1.7 Hz, 1H), 3.32 (ddd, *J* = 10.5, 6.0, 4.1 Hz, 1H), 3.18 (m, 2H). ¹³C NMR (100 MHz, CDCl₃) δ 138.7, 138.6, 138.50, 138.47, 138.45, 138.39, 138.37, 138.3, 138.1 (two resonances), 128.5, 128.4, 128.38 (multiple resonances), 128.37, 128.35, 128.30 (multiple resonances), 128.26, 128.00, 127.95, 127.93, 127.92, 127.90, 127.83, 127.79, 127.77, 127.72, 127.68, 127.67, 127.66, 127.63, 127.57, 127.55, 127.54, 127.50, 127.44, 127.41, 102.7, 98.9, 83.1, 80.5, 79.4, 78.9, 75.8, 75.7, 75.17, 75.14, 74.96, 74.90, 74.7, 73.5, 73.3, 73.1, 72.45, 72.40, 72.1, 72.00, 71.98, 71.4, 69.6, 69.3, 69.0, 66.4, 50.4, 49.5. [α]_D²⁵: +5.1 (c1.0, CHCl₃). IR (cm⁻¹): 3035, 2358, 2167, 1981, 1219; HRMS (ESI-TOF) *m/z*: [M+Na]⁺: Calcd. for C₉₀H₉₅N₃O₁₅SNa: 1512.6382, found 1512.6356.



2-aminoethyl-[α -D-mannopyranosyl-(1 \rightarrow 2)-2-*S*-{ α -D-mannopyranosyl-(1 \rightarrow 2)}]- α -D-mannopyranoside (14**)**

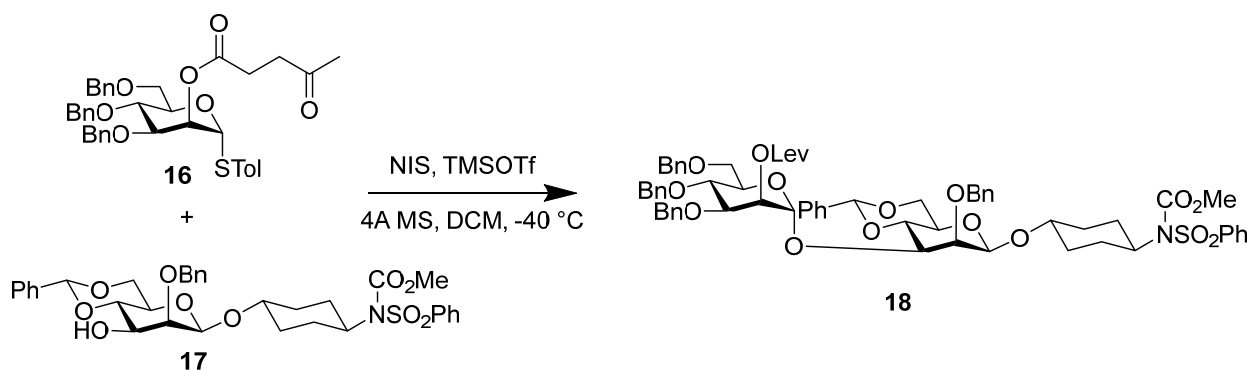
Into an oven dried 50 mL 3-necked flask, ~10 mL ammonia was condensed under stream of nitrogen at $-78\text{ }^{\circ}\text{C}$. Next, 23 mg of (1.0 mmol, 100 equiv.) Na^0 was added, and the bright blue reaction was allowed to stir for 1 hour to ensure that the color stably persisted. 15 mg (10 μmol ; 1equiv.) of trisaccharide **13** in 1 mL of dry THF was added by syringe, and the reaction was allowed to stir for 4 hours. When reaction was finished (monitored by direct-infusion LC-MS), solid NH_4Cl was added portionwise until the disappearance of blue color and the reaction mixture was warmed to room temperature. The crude product was desalted on a Biogel P-2 size exclusion gel column followed by lyophilization, affording 3.5 mg (6.2 μmol) of deprotected trisaccharide **14** as a glassy white solid in 62% yield. ^1H NMR (400 MHz, D_2O) δ 5.41 (s, 1H), 5.27 (s, 1H), 5.12 (s, 1H), 4.26 (dd, $J = 9.6, 4.6$ Hz, 1H), 4.12 (dd, $J = 3.4, 1.5$ Hz, 1H), 4.06 – 3.59 (m, 17H), 3.51 (t, $J = 9.7$ Hz, 1H), 3.33 – 3.20 (m, 2H). ^{13}C NMR (100 MHz, D_2O) δ 103.0, 98.1, 86.8, 78.4, 73.4 (multiple resonances), 72.9, 71.3, 70.8, 69.9, 69.0, 67.8, 67.0, 66.8, 63.7, 60.8 (multiple resonances), 52.2, 39.0. $[\alpha]_{\text{D}}^{25}$: -1133.9 (c1.0, H_2O) IR (cm^{-1}): 3422(b), 3016, 2360, 2051, 1625, 1437; HRMS (ESI-TOF) m/z : $[\text{M}+\text{Na}]^+$: Calcd. for $\text{C}_{20}\text{H}_{37}\text{NO}_{15}\text{SNa}$: 586.1782, found 586.1770.



***p*-Tolyl-3,4,6-tri-*O*-benzyl-2-*O*-(4-oxopentanoyl)-1-thio- α -D-mannopyranoside (**16**)**

To a solution of alcohol **15**⁵ (600 mg; 1.077 mmol; 1 equiv.) in anhydrous CH_2Cl_2 (10 mL), DMAP (26 mg; 0.215 mmol; 0.2 equiv.), levulinic acid (0.21 mL; 2.1 mmol; 2 equiv.) and N,N' -diisopropylcarbodiimide (0.33 mL; 2.1 mmol; 2 equiv.) were added sequentially at $0\text{ }^{\circ}\text{C}$. The reaction mixture was gradually warmed up to ambient temperature and stirred for 5 hours. After

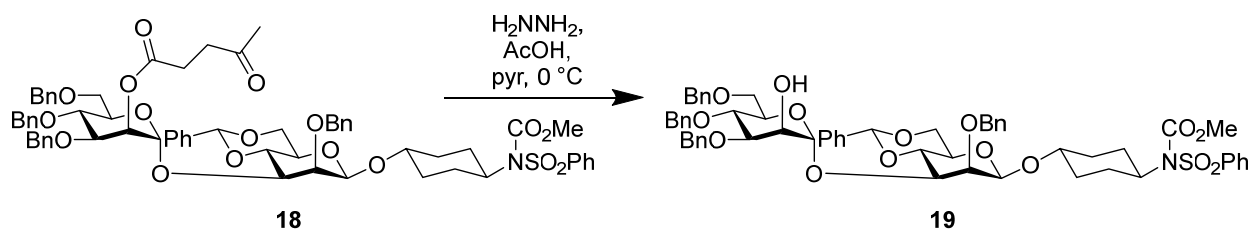
complete consumption of the starting alcohol, the reaction was concentrated in vacuo and purified by flash chromatography (30% ethyl acetate in hexane) to afford corresponding levulinoate ester **16** (648 mg; 0.99 mmol; 92%) as white foam. ^1H NMR (400 MHz, CDCl_3) δ 7.42 – 7.22 (multiple signals, 15H + residual CHCl_3), 7.20 (m, 2H), 7.04 (d, $J = 7.9$ Hz, 2H), 5.58 (dd, $J = 2.8, 1.7$ Hz, 1H), 5.44 (d, $J = 1.7$ Hz, 1H), 4.88 (d, $J = 10.8$ Hz, 1H), 4.70 (d, $J = 11.2$ Hz, 1H), 4.63 (d, $J = 11.9$ Hz, 1H), 4.54 (d, $J = 11.2$ Hz, 1H), 4.52 (d, $J = 10.8$ Hz, 1H), 4.46 (d, $J = 11.9$ Hz, 1H), 4.34 (m, 1H), 3.94 (dd, $J = 10.7, 2.8$ Hz, 1H), 3.91 (app t, 9 Hz, 1H), 3.84 (dd, $J = 10.9, 4.6$ Hz, 1H), 3.71 (dd, $J = 10.9, 1.9$ Hz, 1H), 2.82 – 2.62 (m, 4H), 2.29 (s, 3H), 2.11 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 206.2, 171.9, 138.3, 138.2, 137.9, 137.7, 132.4, 129.8, 129.7, 128.4, 128.3, 128.3, 128.1, 127.9, 127.8, 127.7, 127.7, 127.5, 86.4, 78.4, 75.2, 74.5, 73.3, 72.3, 71.7, 70.4, 68.9, 37.9, 29.7, 28.1, 21.1. $[\alpha]_{\text{D}}^{25}$: +99.4 (c1.0, CHCl_3). IR (cm^{-1}): 2908, 2861, 2660, 1739, 1718, 1101. HRMS (ESI-TOF) m/z : $[\text{M}+\text{Na}]^+$: Calcd. for $\text{C}_{39}\text{H}_{42}\text{O}_7\text{SNa}$: 677.2549, found: 677.2542.



2-*O*-benzyl-3-*O*-(3,4,6-tri-*O*-benzyl-2-*O*-(4-oxopentanoyl)- α -D-mannopyranosyl)-4,6-*O*-benzylidene- β -D-mannopyranoside (**18**)

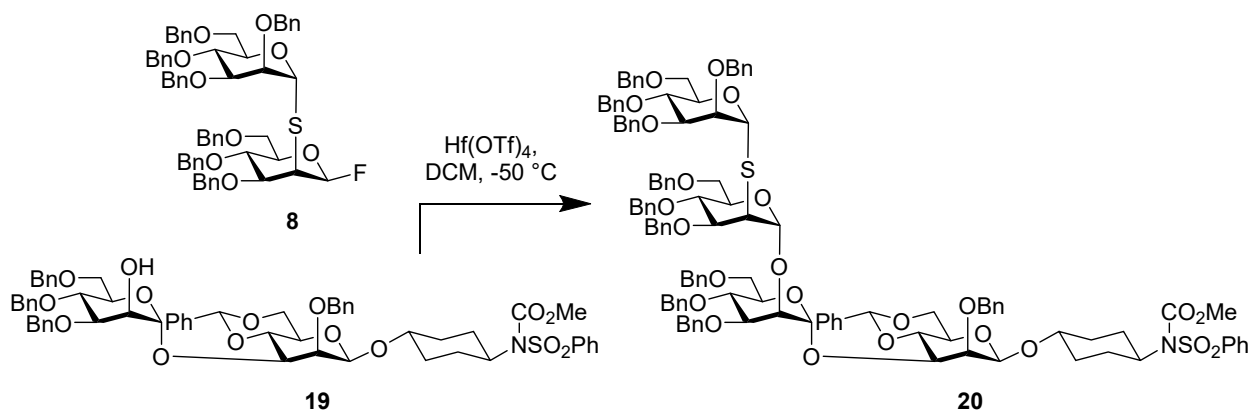
500 mg (0.763 mmol; 1 equiv.) of **16** and 598 mg (0.915 mmol; 1.2 equiv.) of **17**⁶ in a 50 mL flask were dissolved in toluene and cooled to -78 °C. Vacuum was applied and the cooling bath was removed and allowed to warm to room temperature as the toluene evaporated. This procedure was repeated twice. The dry residue was dissolved in 14 mL of CH_2Cl_2 and activated 4Å molecular sieves were added stirred for 20 minutes at room temperature. The reaction mixture was cooled to -40 °C and stirred another 15 minutes more. 429 mg of NIS (1.91 mmol; 2.5 equiv.) was added, and after 15 minutes, 27 μL of TMSOTf (0.15 mmol; 0.2 equiv.) dissolved in 2 mL of CH_2Cl_2 was added dropwise. The reaction was stirred at -40 °C for 2 h and quenched with aq. sat. NaHCO_3 and solid $\text{Na}_2\text{S}_2\text{O}_3$. The reaction mixture was filtered, and the filtrate was washed with aq. sat. $\text{Na}_2\text{S}_2\text{O}_3$, aq. sat. NaHCO_3 and brine, then the organic layer was dried (MgSO_4) and concentrated prior to flash column chromatography (25% ethyl acetate and hexane). Compound **18** (741 mg; 0.626 mmol; 82%) was obtained as a white amorphous solid. ^1H NMR (400 MHz, CDCl_3) δ 7.92 (app d, $J = 8$ Hz, 2H), 7.62 (app t, $J = 7.2$ Hz, 1H), 7.53 (app t, $J = 8$ Hz, 2H), 7.5 (m, 2H), 7.41 – 7.10 (multiple signals, 23H), 5.60 (s, 1H), 5.58 (dd, $J = 2.8, 1.3$ Hz, 1H), 5.23 (d, $J = 1.3$ Hz, 1H), 4.92 – 4.83 (m, 2H), 4.78 (d, $J = 12.3$ Hz, 1H), 4.66 (d, $J = 11.3$ Hz, 1H), 4.57 – 4.38 (multiple signals, 6H), 4.32 (dd, $J = 10.4, 4.9$ Hz, 1H), 4.18 (app t, $J = 9.7$ Hz, 1H), 3.90 (multiple signals,

3H), 3.82 – 3.54 (multiple signals, 6H), 3.65 (s, 3H), 3.31 (app td, $J = 9.7, 4.9$ Hz, 1H), 2.73 – 2.58 (m, 4H), 2.37–2.13 (m, 3H), 2.10 (s, 3H), 2.0 – 1.91 (m, 1H), 1.89 – 1.79 (m, 2H), 1.59 – 1.48 (m, 1H), 1.40 – 1.28 (m, 1H). ^{13}C NMR (100 MHz, CDCl_3) δ 206.1, 202.2 171.7, 152.6, 140.3, 138.6, 138.3, 138.1, 137.9, 137.3, 133.4, 128.8, 128.7, 128.36 (two resonances), 128.34, 128.3, 128.2, 128.13, 128.10, 127.9, 127.75, 127.71, 127.67, 127.63, 127.61, 127.56, 126.0, 101.2, 100.1, 98.7, 78.6, 77.81, 77.75, 76.4, 75.2, 75.0, 74.9, 74.3, 73.4, 72.0, 71.5, 69.2, 68.5, 68.4, 67.4, 58.5, 53.5, 38.1, 33.2, 31.5, 29.8, 28.46, 28.44, 28.2. $[\alpha]_{\text{D}}^{25}$: -3.6 (c1.0, CHCl_3). IR (cm^{-1}): 3030, 2924, 2861, 1962, 1732, 1496, 1356, 1088. HRMS (ESI-TOF) m/z : $[\text{M}+\text{Na}]^+$: Calcd. for $\text{C}_{66}\text{H}_{73}\text{NO}_{17}\text{SNa}$: 1206.4497, found 1206.4469.



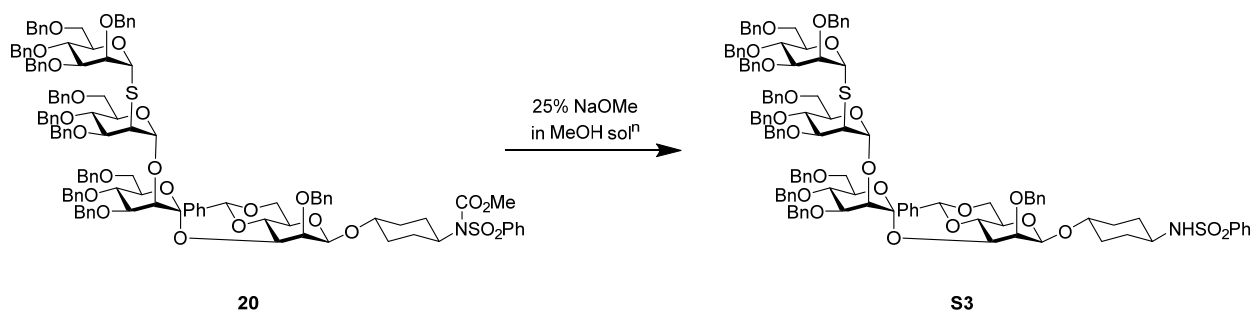
2-*O*-benzyl-3-*O*-(3,4,6-tri-*O*-benzyl- α -D-mannopyranosyl)-4,6-*O*-benzylidene- β -D-mannopyranoside (19)

Compound **18** (700 mg; 0.591 mmol; 1 equiv.) was dissolved in 10 mL of CH_2Cl_2 and cooled to 0 °C. Pyridine (190 μL ; 2.364 mmol; 4 equiv.) and acetic acid (34 μL ; 0.591 mmol; 1 equiv.) was mixed in a 1 mL ice cooled vial and added to reaction mixture dropwise, followed by addition of hydrazine (91 μL ; 2.955 mmol; 5 equiv.) to reaction mixture. The reaction was stirred for 2 h at 0 °C and quenched with 2 mL of acetone. The reaction mixture was concentrated under reduced pressure and the crude was purified by column chromatography in 30% ethyl acetate and hexane to afford compound **19** (430 mg; 0.395 mmol; 67%) as white glassy solid. ^1H NMR (400 MHz, CDCl_3) δ 7.92 (app d, $J = 8$ Hz, 2H), 7.62 (app t, $J = 7.4$ Hz, 1H), 7.53 (app t, $J = 7.8$ Hz, 2H), 7.47 – 7.12 (multiple signals, 25H + residual CHCl_3), 5.56 (s, 1H), 5.33 (d, $J = 1.7$ Hz, 1H), 4.91 (d, $J = 12.4$ Hz, 1H), 4.85 (d, $J = 11.1$ Hz, 1H), 4.80 (d, $J = 12.4$ Hz, 1H), 4.67 (d, $J = 11.4$ Hz, 1H), 4.61 (d, $J = 11.4$ Hz, 1H), 4.54 – 4.46 (m, 4H), 4.41 (tt, $J = 12.1, 3.8$ Hz, 1H), 4.30 (dd, $J = 10.4, 4.8$ Hz, 1H), 4.18 (dd, $J = 2.8, 1.7$ Hz, 1H), 4.13 (app t, $J = 9.7$ Hz, 1H), 4.00 (dd, $J = 10.1, 3.1$ Hz, 1H), 3.91 (app t, $J = 10.3$ Hz, 1H), 3.87 – 3.80 (m, 2H), 3.79 – 3.52 (m, 5H), 3.66 (s, 3H), 3.33 (app td, $J = 9.7, 4.9$ Hz, 1H), 2.40 (br s, 1H), 2.35 – 2.10 (m, 3H), 2.00 – 1.9 (m, 1H), 1.9 – 1.8 (m, 2H), 1.59 – 1.45 (m, 1H + H_2O), 1.39 – 1.26 (m, 1H). ^{13}C NMR (101 MHz, CDCl_3) δ 152.5, 140.3, 138.5, 138.33, 138.28, 137.8, 137.4, 133.4, 128.95, 128.8, 128.5, 128.4, 128.33, 128.30, 128.28, 128.2, 127.94, 127.90, 127.88, 127.72, 127.71, 127.59, 127.58, 127.56, 126.0, 101.5, 100.3, 100.1, 79.9, 78.7, 78.1, 76.4, 75.3, 74.93, 74.90, 74.3, 73.4, 71.9, 71.8, 69.3, 68.6, 68.0, 67.4, 58.4, 53.5, 33.2, 31.5, 30.9, 28.4. $[\alpha]_{\text{D}}^{25}$: -2.01 (c1.0, CHCl_3). IR (cm^{-1}): 3540, 2919, 1732, 1452, 1666, 1359, 1086. HRMS (ESI-TOF) m/z : $[\text{M}+\text{Na}]^+$: Calcd. for $\text{C}_{61}\text{H}_{67}\text{NO}_{15}\text{SNa}$: 1108.4129, found 1108.4065.



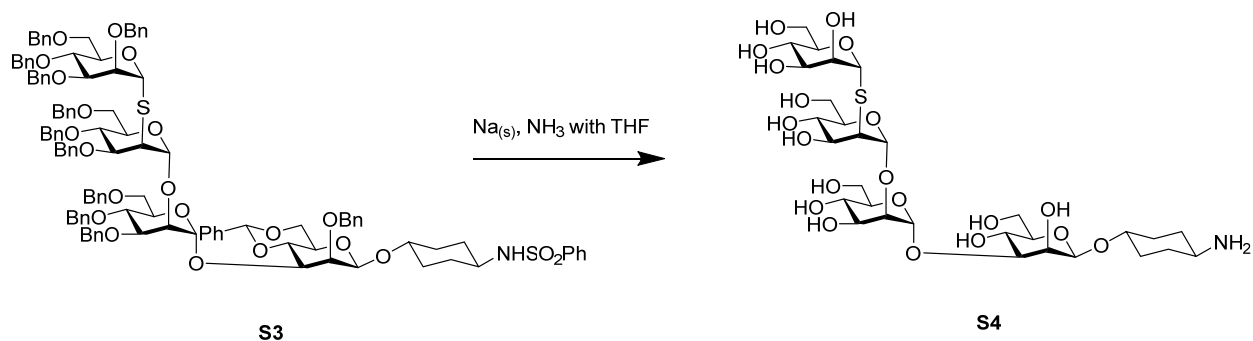
***Trans*-(*N*-methylcarbonate-*N*-1-phenylsulfonyl)-cyclohexyl-2-*O*-benzyl-4,6-*O*-benzylidene-3-*O*-[*3,4,6*-tri-*O*-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)]-2-*O*-[*3,4,6*-tri-*O*-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)]-2-*S*-[*3,4,6*-tri-*O*-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)]- β -D-mannopyranoside (**20**)**

Disaccharide donor **8** (280 mg; 282 μ mol; 1 equiv.) and acceptor **19** (230 mg; 212 μ mol; 0.75 equiv.) were dissolved in anhydrous dichloromethane (5 mL), and freshly crushed and flame dried 4 Å molecular sieves was added and stirred for 15 min at room temperature. The mixture was cooled at -50 °C, then Hg(OTf)₄ (219 mg; 282 μ mol; 1 equiv.) was added and allowed to stir for 1 h. The reaction was neutralized with a few drops of triethylamine, diluted with CH₂Cl₂ and filtered through celite pad and concentrated, and then purification by column chromatography by 35% of EtOAc in Hexane, afforded trisaccharide **20** as white foam in 77% yield based on acceptor **19** (335 mg, 163.84 μ mol). ¹H NMR (400 MHz, CDCl₃) δ 7.92 (app d, *J* = 7.4 Hz, 2H), 7.61 (app t, *J* = 7.4 Hz, 1H), 7.40 (t, *J* = 7.7 Hz, 2H), 7.44 – 6.98 (multiple signals, 60H + residual CHCl₃), 5.57 (s, 1H), 5.36 (s, 2H), 5.33 (s, 1H), 5.01 – 4.73 (multiple signals, 5H), 4.62 – 4.30 (multiple signals, 17H), 4.29 – 4.15 (multiple signals, 5H), 4.08 – 3.97 (multiple signals, 3H), 3.96 – 3.39 (m, 20H), 3.30 (d, *J* = 11.0 Hz, 1H), 3.12 (app td, *J* = 9.6, 4.9 Hz, 1H), 2.34 – 2.16 (m, 2H), 2.16 – 2.08 (m, 1H), 2.00 – 1.69 (m, 3H), 1.57 – 1.43 (m, 1H), 1.34 – 1.19 (m, 1H). ¹³C NMR (100 MHz, CDCl₃) δ (selected signals) 152.5, 140.3, 138.9, 138.76, 138.63, 138.52, 138.50, 138.32, 138.28, 138.22 (two resonances), 138.1 (two resonances), 137.3, 133.4, 128.8, 128.46, 128.40, 128.38, 128.34 (multiple resonances), 128.31, 128.28 (multiple resonances), 128.23, 128.21, 128.17, 128.12, 128.0, 128.0, 127.9, 127.8, 127.75, 127.71, 127.66, 127.64, 127.59, 127.54, 127.51, 127.49, 127.47, 127.40, 127.3, 127.1, 125.95, 102.1, 101.3, 99.9, 99.8, 83.0, 80.4, 78.9, 78.3, 78.1, 76.3, 75.7, 75.5, 75.19, 75.14, 75.0, 74.9, 74.8, 74.0, 73.2, 73.1, 72.9, 72.5, 72.41, 72.46, 72.2, 71.9, 71.3, 69.7, 68.9, 68.8, 68.5, 67.3, 58.5, 53.4, 49.4, 33.1, 31.4, 28.4. [α]_D²⁵: +12.0 (c1.0, CHCl₃). IR (cm⁻¹): 3030, 2853, 1732, 1495, 1260. HRMS (ESI-TOF) *m/z*: [M+Na]⁺: Calcd. for C₁₂₂H₁₂₉NO₂₅S₂Na: 2078.8244, found 2078.8289.



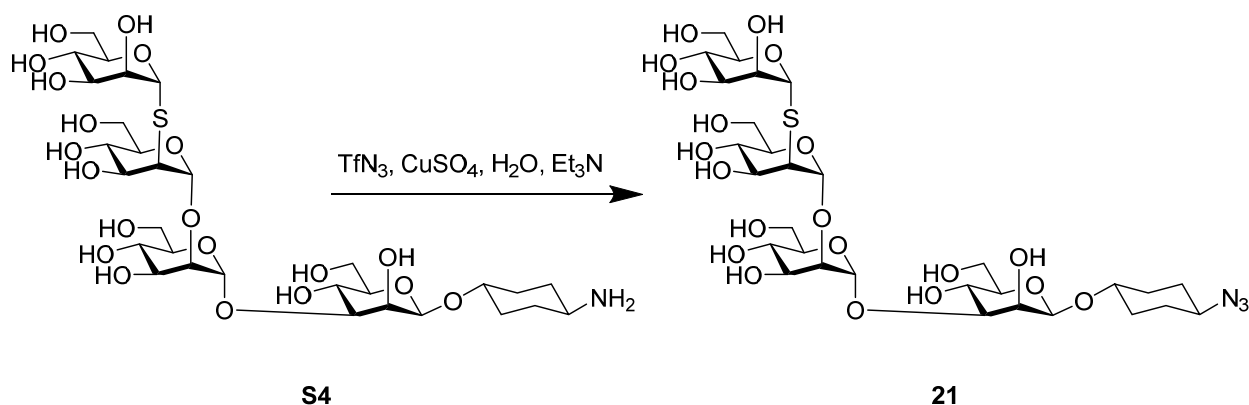
***Trans*-(*N*-1-phenylsulfonyl)-cyclohexyl-2-*O*-benzyl-4,6-*O*-benzylidene-3-*O*-[3,4,6-tri-*O*-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)]-2-*O*-[3,4,6-tri-*O*-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)]-2-*S*-[3,4,6-tri-*O*-benzyl- α -D-mannopyranosyl-(1 \rightarrow 2)]- β -D-mannopyranoside (S3)**

Compound **20** (280 mg; 136 μ mol; 1 equiv.) was dissolved in methanol (2 mL) and dichloromethane (2 mL) and 10 μ L (40.8 μ mol; 0.3 equiv.) 25% sodium methoxide in methanol solution was added. After 3 hours at room temperature, the reaction was neutralized with Amberlite IR-120 H⁺ form ion exchange resin with caution to avoid acidifying below pH 6. The resin was filtered off and the filtrate was concentrated under reduced pressure. The concentrate was purified by column chromatography to afford compound **S3** (244 mg; 122 μ mol) as colorless syrup in 90% yield. ¹H NMR (400 MHz, CDCl₃) δ 7.88 (app d, *J* = 7.1 Hz, 2H), 7.59 (app t, *J* = 7 Hz, 1H), 7.52 (app t, *J* = 7 Hz, 2H), 7.47 – 7.09 (multiple signals, 58H + residual CHCl₃), 7.08 – 6.98 (m, 2H), 5.56 (s, 1H), 5.33 (app s, 2H), 5.31 (s, 1H), 4.95 – 4.77 (multiple signals, 4H), 4.71 (d, *J* = 12.3 Hz, 1H), 4.62 – 4.39 (multiple signals, 13H), 4.39 – 4.29 (multiple signals, 4H), 4.27 – 4.11 (multiple signals, 5H), 4.08 – 3.97 (m, 3H), 3.93 – 3.54 (multiple signals, 14H), 3.50 – 3.36 (m, 3H), 3.28 (d, *J* = 10.6 Hz, 1H), 3.20 – 3.00 (m, 2H), 2.25 – 1.62 (m, 4H), 1.41 – 1.13 (m, 4H). ¹³C NMR (100 MHz, CDCl₃) δ (selected signals) 141.1, 139.0, 138.8, 138.7, 138.56, 138.54, 138.36, 138.34, 138.25, 138.23, 138.16, 138.13, 137.3, 132.7, 129.2, 129.0, 128.48, 128.43, 128.41, 128.35, 128.33, 128.31, 128.29, 128.25, 128.22, 128.19, 128.12, 128.05, 127.9, 127.8, 127.75, 127.74, 127.71, 127.69, 127.61, 127.59, 127.57, 127.50, 127.45, 127.40, 127.3, 127.2, 126.9, 126.0, 101.3, 99.84, 99.82, 83.0, 80.5, 78.9, 78.3, 78.1, 75.8, 75.7, 75.5, 75.2, 75.1, 75.0, 74.95, 74.85, 74.0, 73.2, 73.15, 73.0, 72.54, 72.47, 72.37, 72.30, 71.9, 71.3, 69.7, 69.0, 68.9, 68.5, 67.4, 51.7, 49.4, 31.0 (multiple signals), 29.3. $[\alpha]_D^{25}$: +8.0 (c1.0, CHCl₃). IR (cm⁻¹): 3029, 2862, 1731, 1496, 1452, 1090. HRMS (ESI-TOF) *m/z*: [M+Na]⁺: Calcd. for C₁₂₀H₁₂₇NO₂₅S₂Na : 2020.8189, found 2020.8182.



***Trans*-aminocyclohexyl-3-*O*-[α -D-mannopyranosyl-(1 \rightarrow 2)]-2-*O*-[α -D-mannopyranosyl-(1 \rightarrow 2)]-2-*S*-[α -D-mannopyranosyl-(1 \rightarrow 2)]- β -D-mannopyranoside (**S4**)**

Into an oven dried 500 mL 3-necked flask, ~200 mL ammonia was condensed under stream of nitrogen at -78 °C. 471 mg of (20.5 mmol) Na^o was added, and the bright blue reaction was allowed to stir for 1 hour to observe persistence of the blue color. Next, 205 mg (103 μ mol; 1 equiv.) of tetrasaccharide **S3** in 2 mL of dry THF was added by syringe, and the reaction was allowed to stir for 2 hours. When reaction was finished (monitored by direct-infusion LCMS), solid NH₄Cl (548 mg; 10.3 mmol) was added portionwise until the disappearance of blue color and the ice bath was removed, allowing the mixture to warm to room temperature. The crude product was desalted on a Biogel P-2 size exclusion gel column and lyophilization afforded ~105 mg of deprotected trisaccharide **S4** (including some residual salt) as a hazy glassy white solid, which was used for the next step without further purification. ¹H NMR (400 MHz, D₂O) δ 5.13 (s, 1H), 5.06 (s, 1H), 5.00 (s, 1H), 4.54 (s, 1H), 3.99 (dd, *J* = 9.7, 4.5 Hz, 1H), 3.84 (d, *J* = 2.8 Hz, 1H), 3.81 – 3.77 (m, 2H), 3.76 – 3.68 (m, 2H), 3.68 – 3.33 (m, 17H), 3.28 (app t, *J* = 9.5 Hz, 1H), 3.15 – 3.08 (m, 1H), 2.97 – 2.86 (m, 1H), 1.92 – 1.73 (m, 4H), 1.35 – 1.05 (m, 4H). ¹³C NMR (100 MHz, D₂O) δ 103.0, 100.4, 97.7, 86.7, 80.4, 78.4, 75.9, 73.4, 73.3, 73.2, 71.3, 70.8, 70.6, 69.9, 69.1, 67.5, 67.0, 66.8, 66.1, 60.82, 60.78, 60.6, 52.1, 49.0, 46.6, 30.2, 28.9, 28.1, 28.0, 8.1. HRMS (ESI-TOF) *m/z*: [M+Na]⁺: Calcd. for C₃₀H₅₃NO₂₀SNa : 802.2779, found 802.2761.



***Trans*-azidocyclohexyl-3-*O*-[α -D-mannopyranosyl-(1 \rightarrow 2)]-2-*O*-[α -D-mannopyranosyl-(1 \rightarrow 2)]-2-*S*-[α -D-mannopyranosyl-(1 \rightarrow 2)]- β -D-mannopyranoside (**21**)**

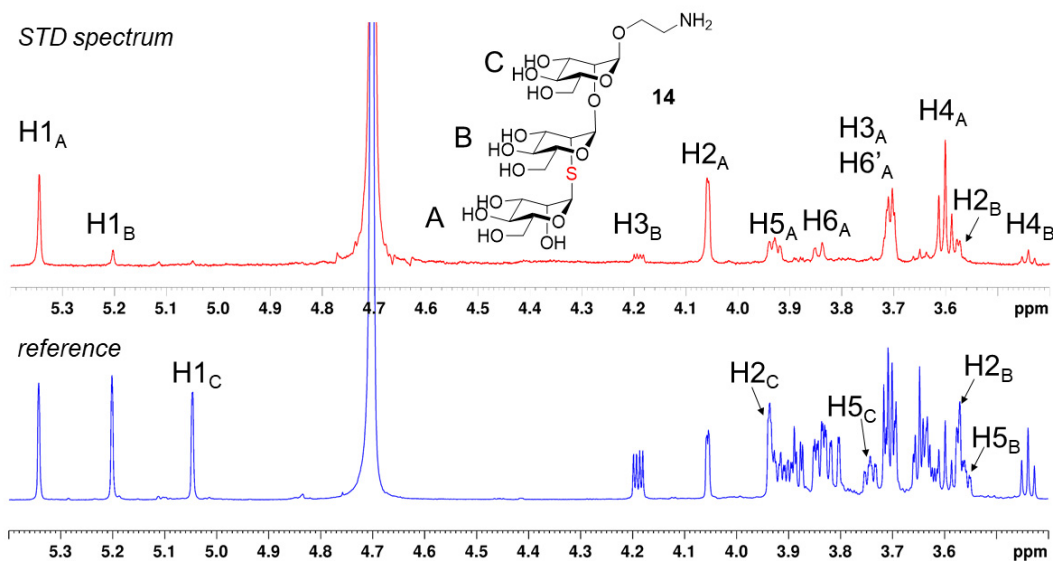
In a 5 mL oven-dried reaction vial with magnetic stir bar, 105 mg (1.62 mmol; 1 equiv.) of sodium azide was cooled to 0 °C and dissolved in 1.3 mL of pyridine, followed by treatment with 327 μ L triflic anhydride (1.94 mmol; 1.2 equiv.). While this stirred for 2 hours at 0 °C, in other 5 mL RB, ~105 mg of crude tetrasaccharide amine **S4** (used directly from previous step) was dissolved in 1.4 mL of water and cooled to 0 °C. 5.1 mg of cupric sulphate (20.5 mmol; 0.2 equiv.) dissolved in 1 mL of H₂O was added to reaction mixture followed addition of 142 μ L (143 mmol; 10 equiv.) of triethyl amine. The triflyl azide solution prepared above was added dropwise to tetrasaccharide reaction mixture at 0 °C and stirred for 1 hour. The crude material was desalted on a Biogel P-2 size exclusion gel column to yield 70 mg of crude product. The crude material was then purified

by reverse phase HPLC (Column: Waters Xbridge Prep, C₁₈, 5 μ m, 19 x 250 mm, 150Å pore diameter. Method: 16 mL/ min flow rate, A = H₂O; B = Acetonitrile without buffer; 1 % B for 10 minutes, then 1-20% B over 55 minutes. Product was detected by UV at 220 nm. Product containing fractions from 3 injections were lyophilized to yield 33 mg (41.04 μ mol) in 40% yield based on **S3** as a colorless glass. ¹H NMR (800 MHz, D₂O, 293 K) δ 5.40 (s, 1H), 5.33 (s, 1H), 5.28 (s, 1H), 4.80 (s, 1H), 4.26 (dd, J = 9.7, 4.6 Hz, 1H), 4.12 (dd, J = 3.4, 1.5 Hz, 1H), 4.08 – 4.06 (m, 2H), 4.02 – 3.97 (m, 2H), 3.94 – 3.86 (m, 4H), 3.85 – 3.65 (m, 12H), 3.63 (dd, J = 4.7, 1.5 Hz, 1H), 3.55 (app t, J = 9.5 Hz, 1H), 3.53 – 3.48 (m, 1H), 3.39 (ddd, J = 9.7, 6.2, 2.3 Hz, 1H), 2.12 – 1.94 (m, 4H), 1.52 – 1.30 (m, 4H). ¹³C NMR (200 MHz, D₂O) δ 103.1, 100.5, 97.7, 86.8, 80.5, 78.5, 76.0, 75.9, 73.41, 73.36, 73.26, 71.4, 70.9, 70.8, 70.0, 69.1, 67.6, 67.1, 66.9, 66.1, 60.9 (two resonances), 60.8, 60.6, 58.8, 52.2, 30.0, 28.6, 28.3, 28.1. $[\alpha]_D^{25}$: -1533 (c1.0, H₂O). IR (cm⁻¹): 3370, 3266, 2236, 2173, 1970, 1063. HRMS (ESI-TOF) m/z : $[M+Na]^+$: Calcd. for C₃₀H₅₁N₃O₂₀SNa: 828.2684, found 828.2676.

STD NMR experiments of trisaccharide **14** and tetrasaccharide **21**

STD NMR experiments were carried out with oligosaccharides **14** and **21** in the presence of the IgG antibody 2G12 (200:1 sugar:2G12, with 2G12 at 25 μ M) in pH 6.7 deuterated phosphate buffered saline. The temperature was 298 K for trisaccharide **14** and 293 K for tetrasaccharide **21** in order to move the HDO peak to avoid covering an anomeric resonance. The experiments were performed without suppression of the residual HDO signal. On-resonance frequencies of 0.86 ppm δ (protein aliphatic) and 7 ppm δ (protein aromatic) were applied for trisaccharides **14** and tetrasaccharide **21** respectively, in both cases with a saturation time of 3s and an off-resonance frequency of 40 ppm δ .

Figure S1. STD for S-Man₃ (**14**) with HIV IgG antibody 2G12. For S-Man₄ (**21**), see Main Text, Figure 1.



Mannosidase experiments

SPAAC labeling with LC-MS/UV tag: To facilitate separation and detection of degradation products via LC/MS, Man₄-cyclohexyl derivatives **21** and **22** were labeled by strain-promoted azide/alkyne cycloaddition (SPAAC) with DBCO amine linker **23**. Man₄-cyclohexyl-N₃ **22** or the thioether derivative **21** were dissolved in degassed H₂O to afford a 25 mM stock, while the DBCO-NH₂ SPAAC reagent **23** was dissolved in acetonitrile (29 mM). Under a N₂ atmosphere, 1.1 equiv. of **23** was added to the Man₄ derivative, followed by heating to 37°C and reaction monitoring by LC/MS. At 30 minutes, both SPAAC reactions were complete by LC/MS, showing two regioisomers, separable by LC/MS. Crude reactions were analyzed with an analytical C4 column (ACQUITY UPLC Protein BEH C4, 2.1mm X 150 mm, 300Å, 1.7µm) using the method in the table below with mobile phases A: H₂O (0.7% formic acid) and B: MeCN (0.1% formic acid).

Time (minutes)	Flow (mL/min)	%A	%B
0	0.3	90	10
1	0.3	90	10
7	0.3	70	30
7.05	0.3	5	95
10	0.3	5	95
10.05	0.3	90	10
13	0.3	90	10

Mannosidase Digestion: 100µg/mL DBCO-NH₂-labeled Man₄-derivative was digested with an α1-2,3 Mannosidase cloned from the pathogenic bacterium *Xanthomonas manihotis* (NEB: P0729S) according to manufacturer's instructions. Briefly, either Man₄-derivative was diluted into 1X glycobuffer I (provided with the enzyme) supplemented with 100µg/mL BSA and finally 80 units of α1-2,3 Mannosidase. Reactions were monitored by LC/MS by the same method detailed above, over the course of 48 hr (see Figures S2 and S3).

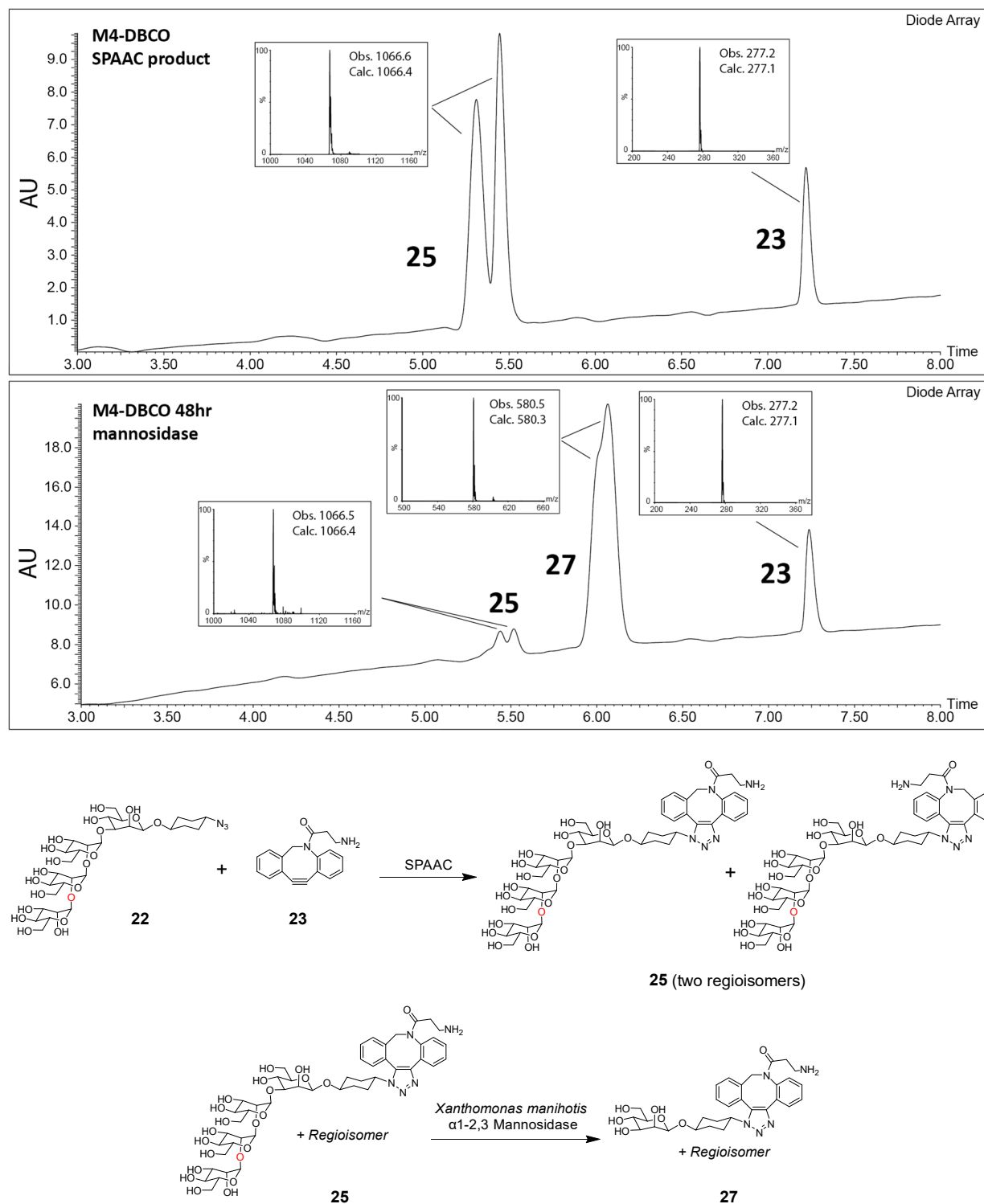


Figure S2. SPAAC labeling and mannosidase digestion of Man₄ derivative 22.

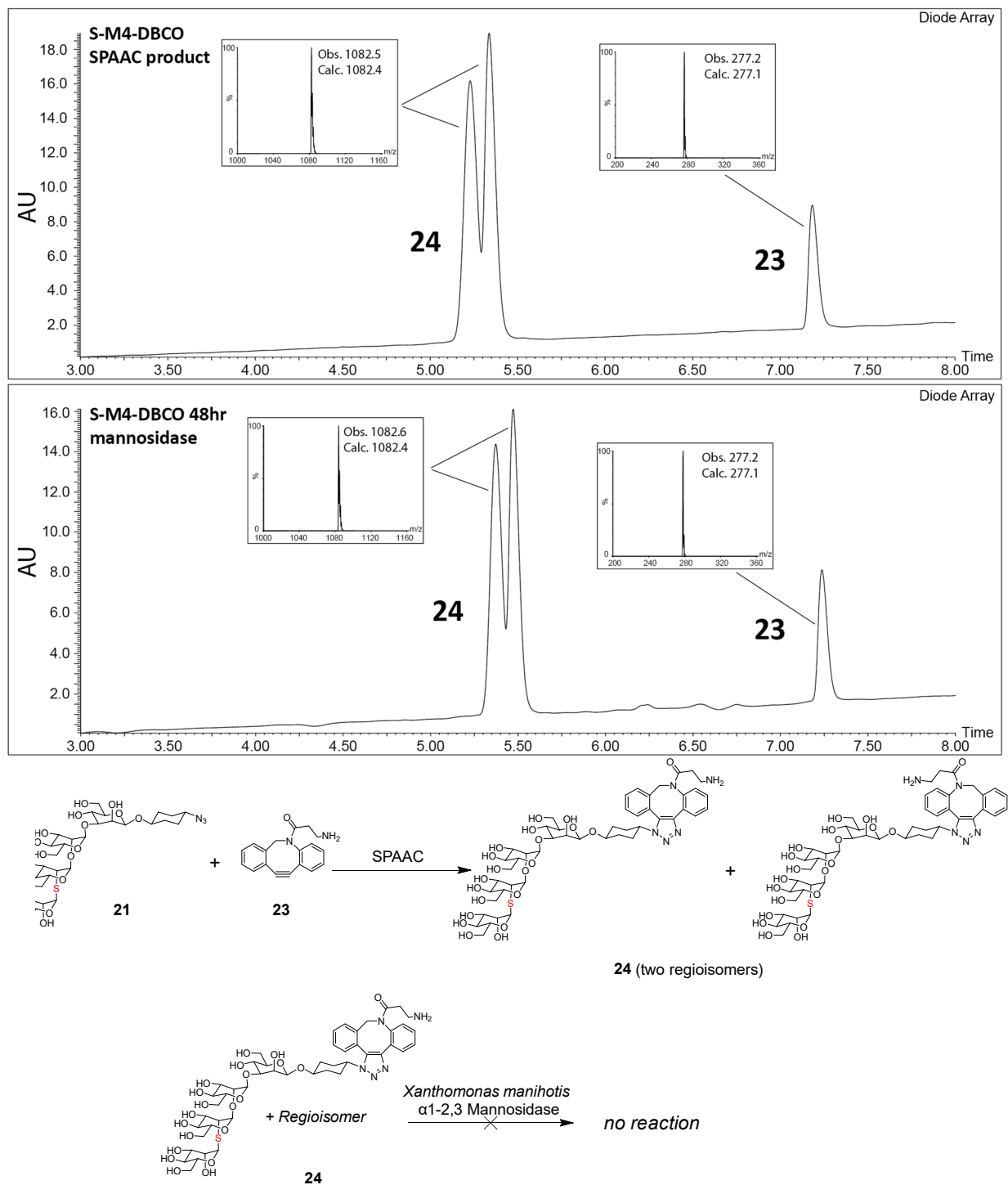
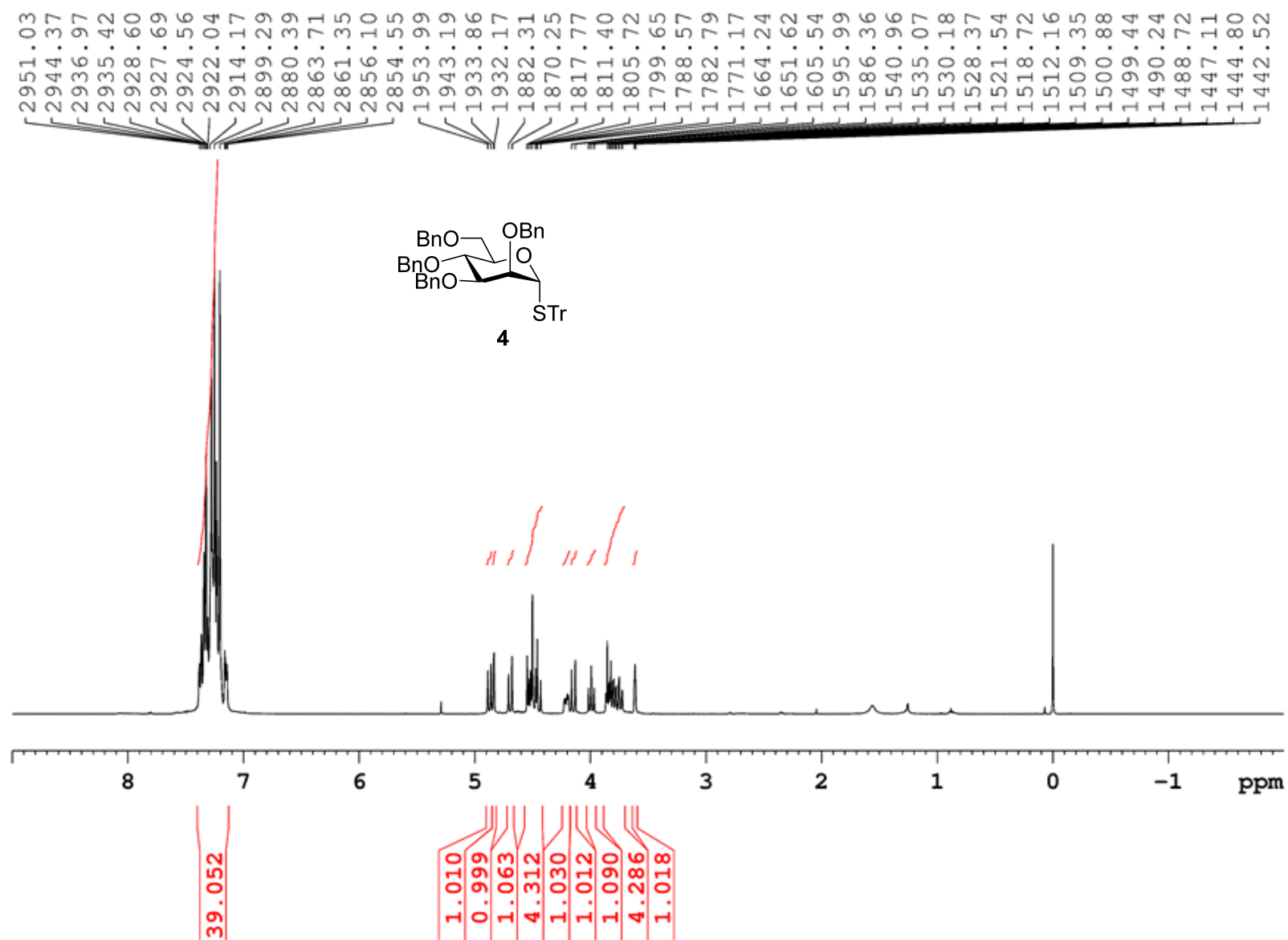


Figure S3. SPAAC labeling and mannosidase digestion of Man₄ derivative 21.

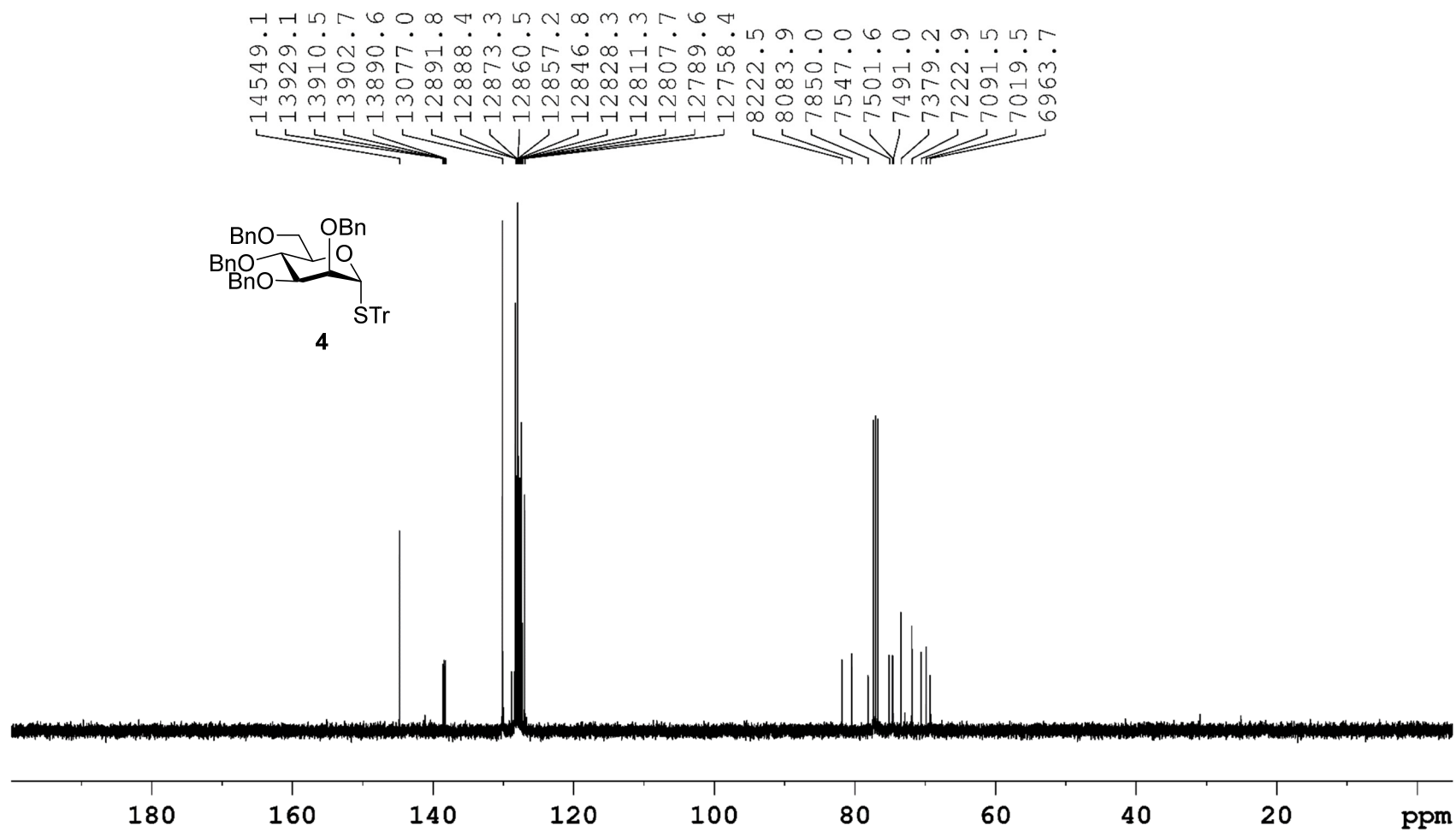
References:

1. Matta, K. L.; Girotra, R. N.; Barlow, J. J., Synthesis of p-nitrobenzyl and p-nitrophenyl 1-thioglycopyranosides. *Carbohydr Res* **1975**, *43*, 101-9.
2. Cheshev, P.; Marra, A.; Dondoni, A., Direct epoxidation of D-glucal and D-galactal derivatives with in situ generated DMDO. *Carbohydr Res* **2006**, *341*, 2714-6.
3. Gordon, D. M.; Danishefsky, S. J., Displacement reactions of a 1,2-anhydro- α -D-hexopyranose: installation of useful functionality at the anomeric carbon. *Carbohydr Res* **1990**, *206*, 361-6.
4. John, F.; Hendrickson, T. L., Synthesis of Truncated Analogues for Studying the Process of Glycosyl Phosphatidylinositol Modification. *Organic Letters* **2010**, *12*, 2080-2083.
5. Alam, S. M.; Aussedat, B.; Vohra, Y.; Meyerhoff, R. R.; Cale, E. M.; Walkowicz, W. E.; Radakovich, N. A.; Anasti, K.; Armand, L.; Parks, R.; Sutherland, L.; Searce, R.; Joyce, M. G.; Pancera, M.; Druz, A.; Georgiev, I. S.; Von Holle, T.; Eaton, A.; Fox, C.; Reed, S. G.; Louder, M.; Bailer, R. T.; Morris, L.; Abdool-Karim, S. S.; Cohen, M.; Liao, H. X.; Montefiori, D. C.; Park, P. K.; Fernandez-Tejada, A.; Wiehe, K.; Santra, S.; Kepler, T. B.; Saunders, K. O.; Sodroski, J.; Kwong, P. D.; Mascola, J. R.; Bonsignori, M.; Moody, M. A.; Danishefsky, S.; Haynes, B. F., Mimicry of an HIV broadly neutralizing antibody epitope with a synthetic glycopeptide. *Sci Transl Med* **2017**, *9*.
6. MacPherson, I. S.; Temme, J. S.; Habeshian, S.; Felczak, K.; Pankiewicz, K.; Hedstrom, L.; Krauss, I. J., Multivalent Glycocluster Design through Directed Evolution. *Angewandte Chemie International Edition* **2011**, *50*, 11238-11242.

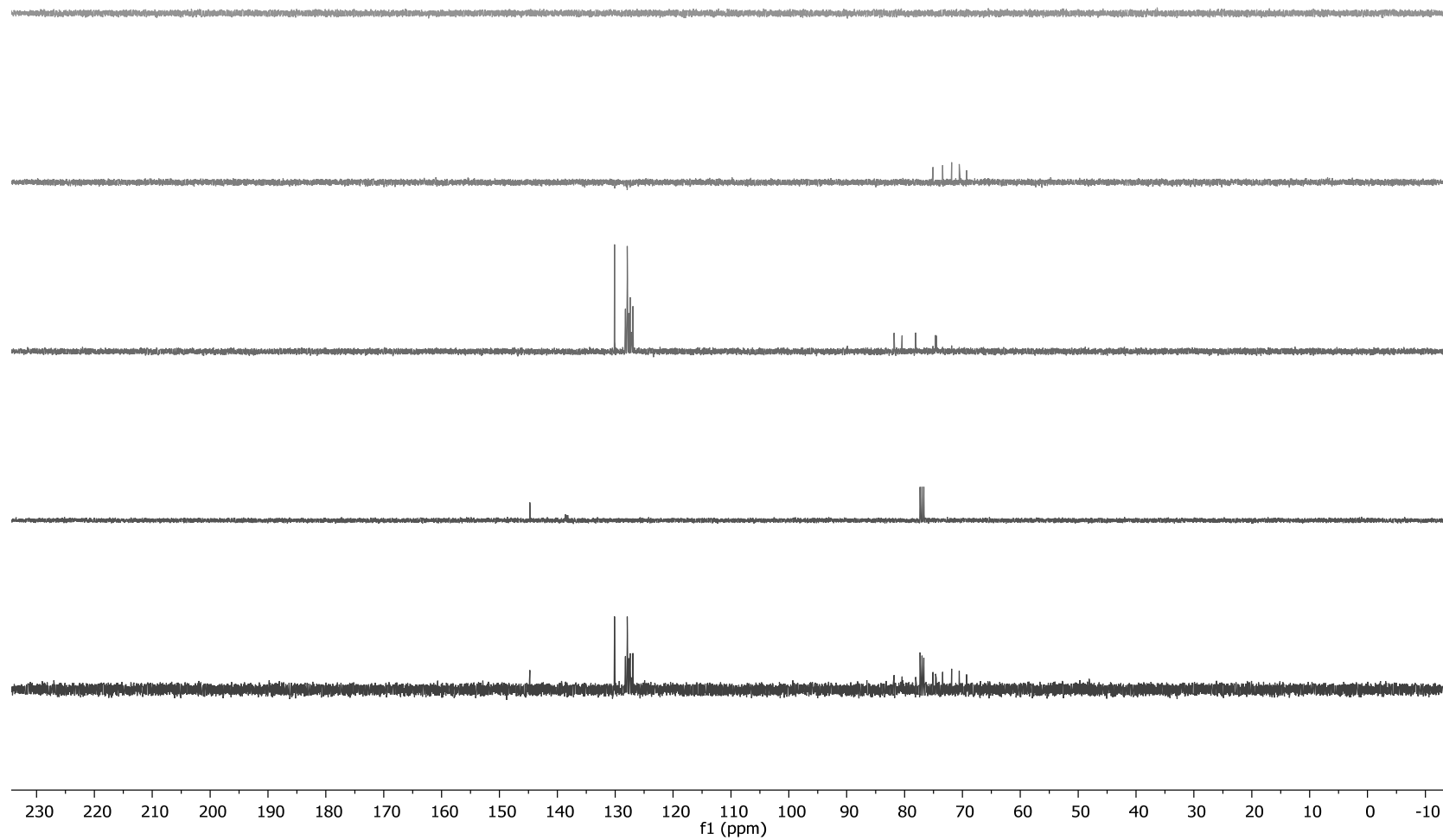
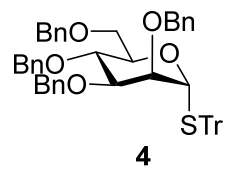
^1H NMR (400 MHz) of **4** in CDCl_3



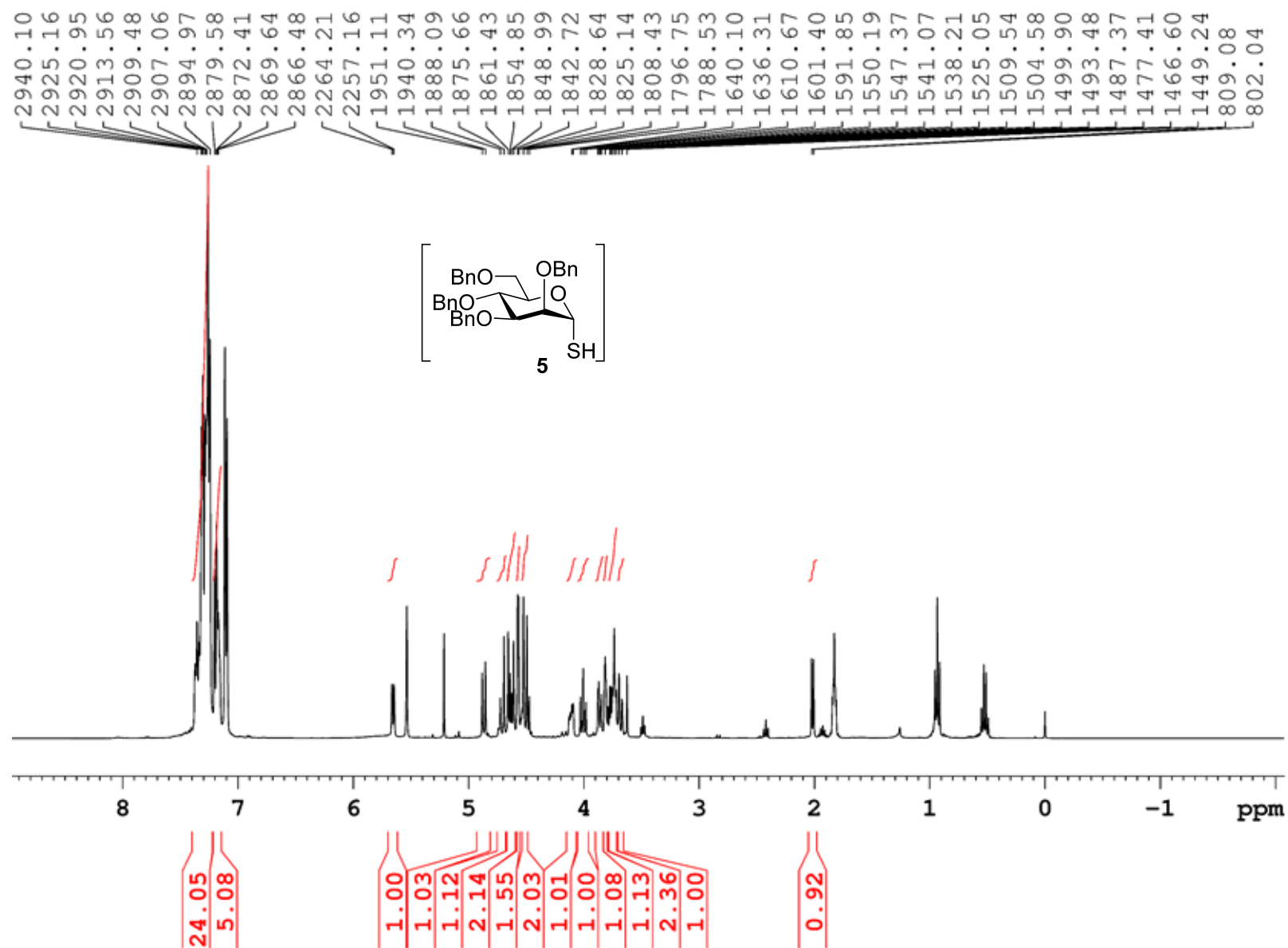
^{13}C NMR (100 MHz) of **4** in CDCl_3



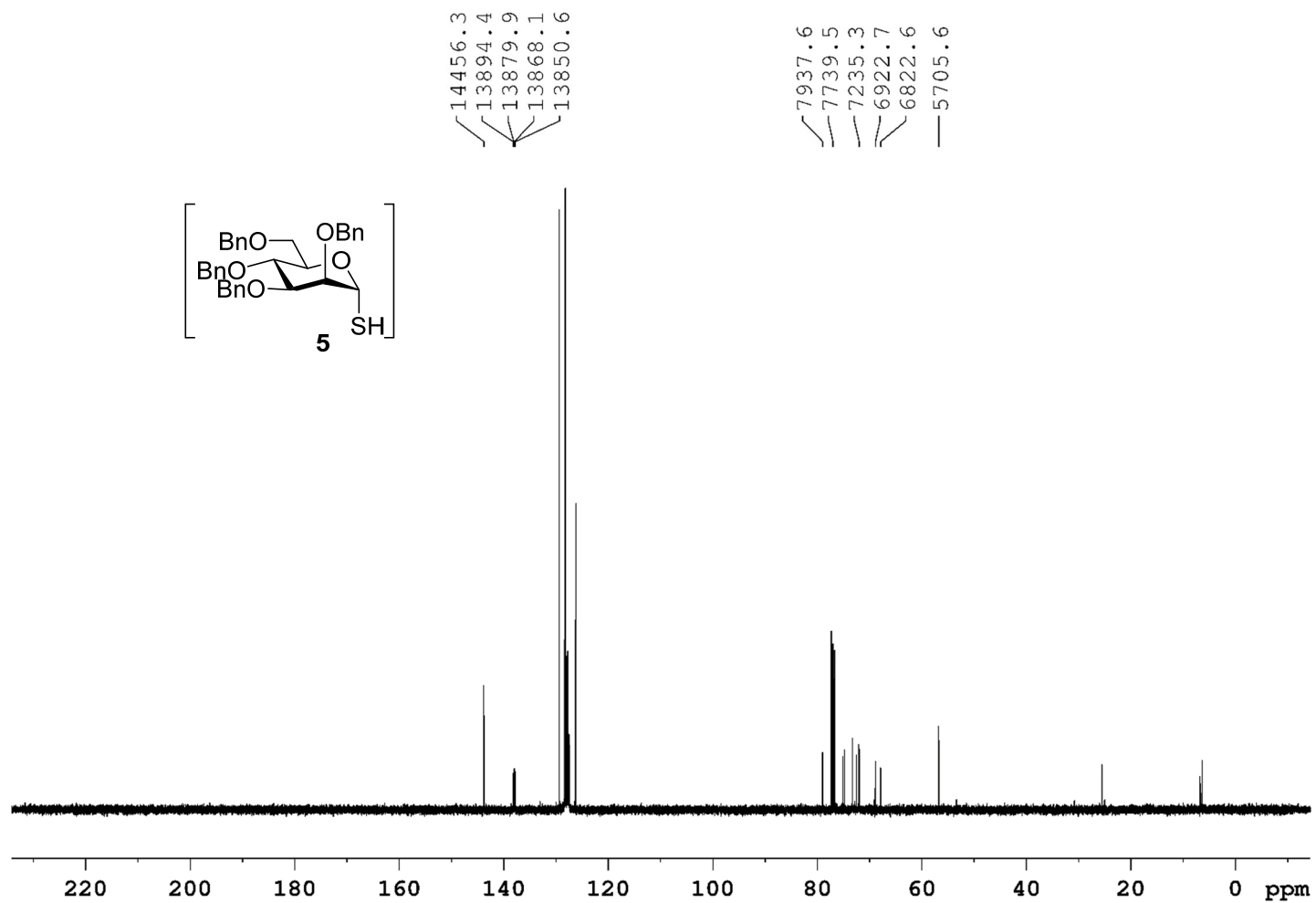
DEPT NMR (100 MHz) of **4** in CDCl₃



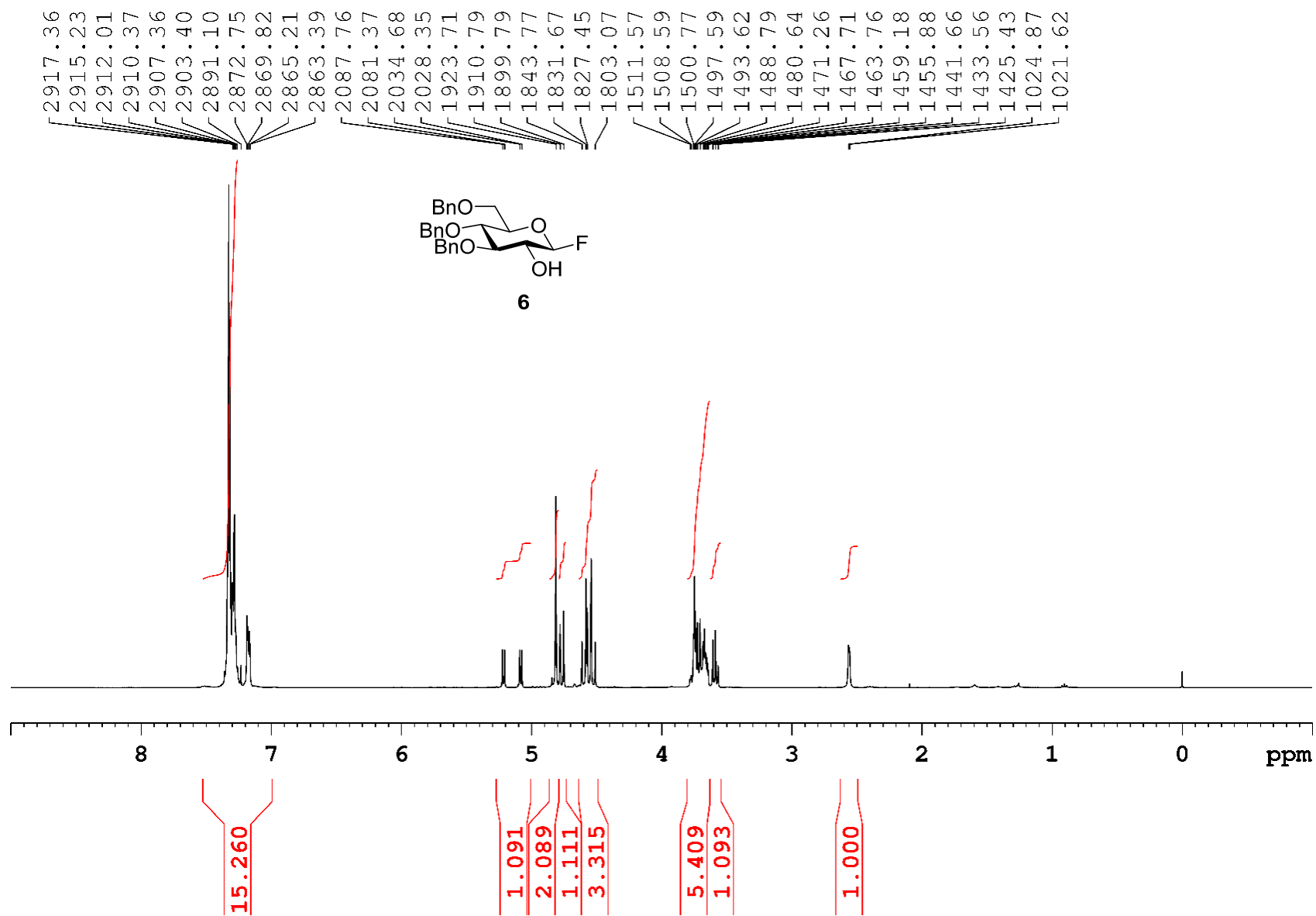
^1H NMR (400 MHz) of crude **5** in CDCl_3



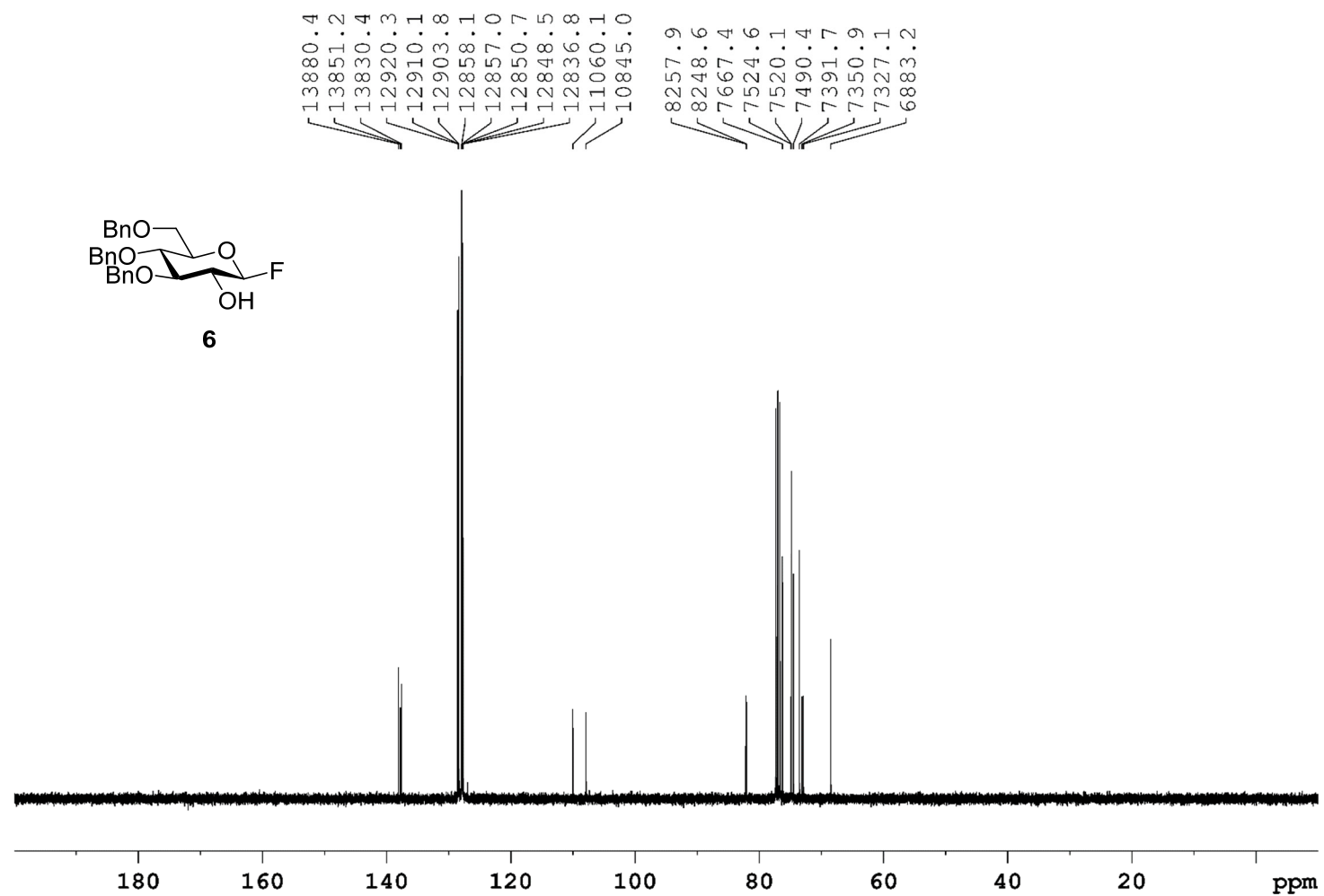
^{13}C NMR (100 MHz) of crude **5** in CDCl_3



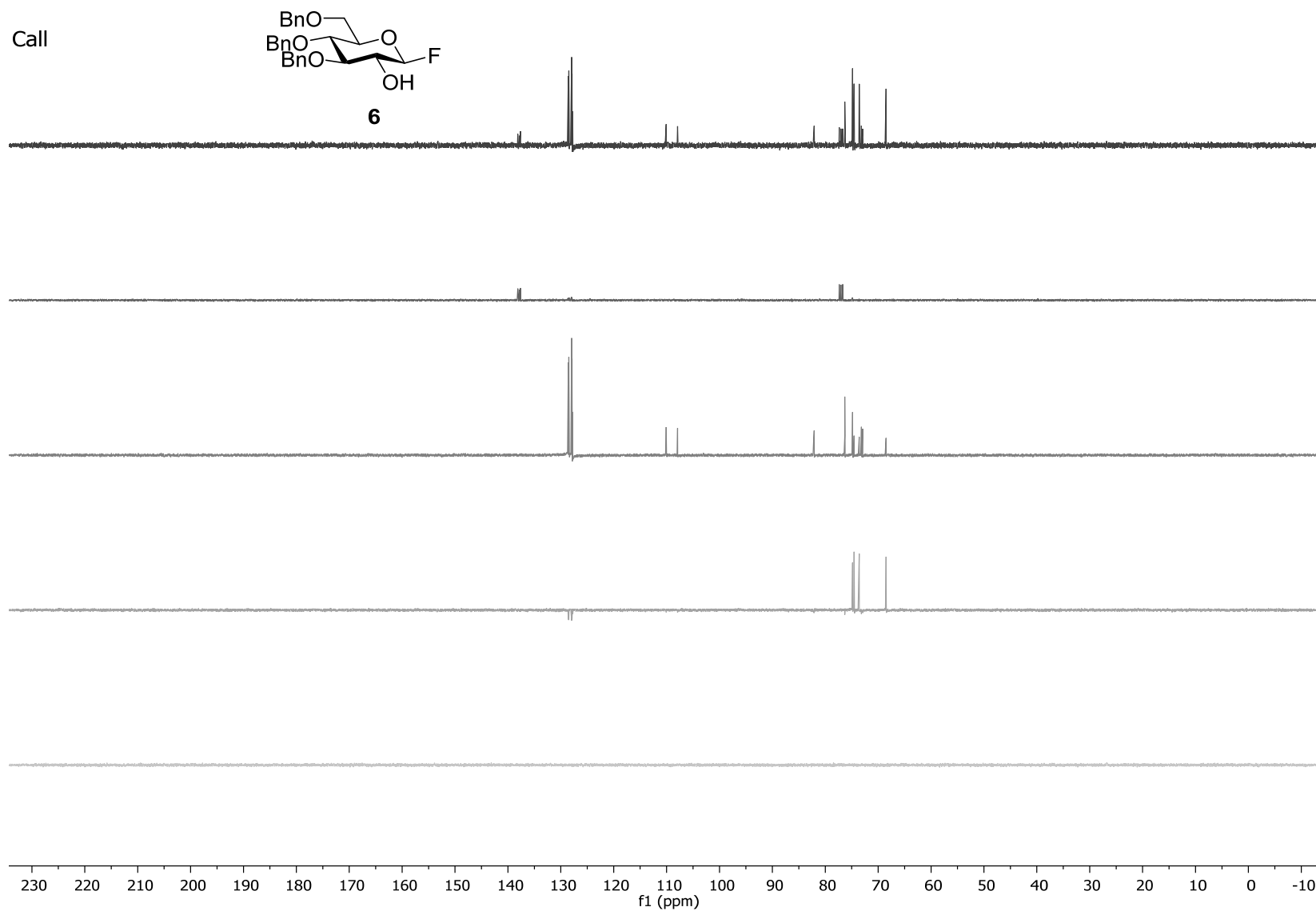
¹H NMR (400 MHz) of **6** in CDCl₃



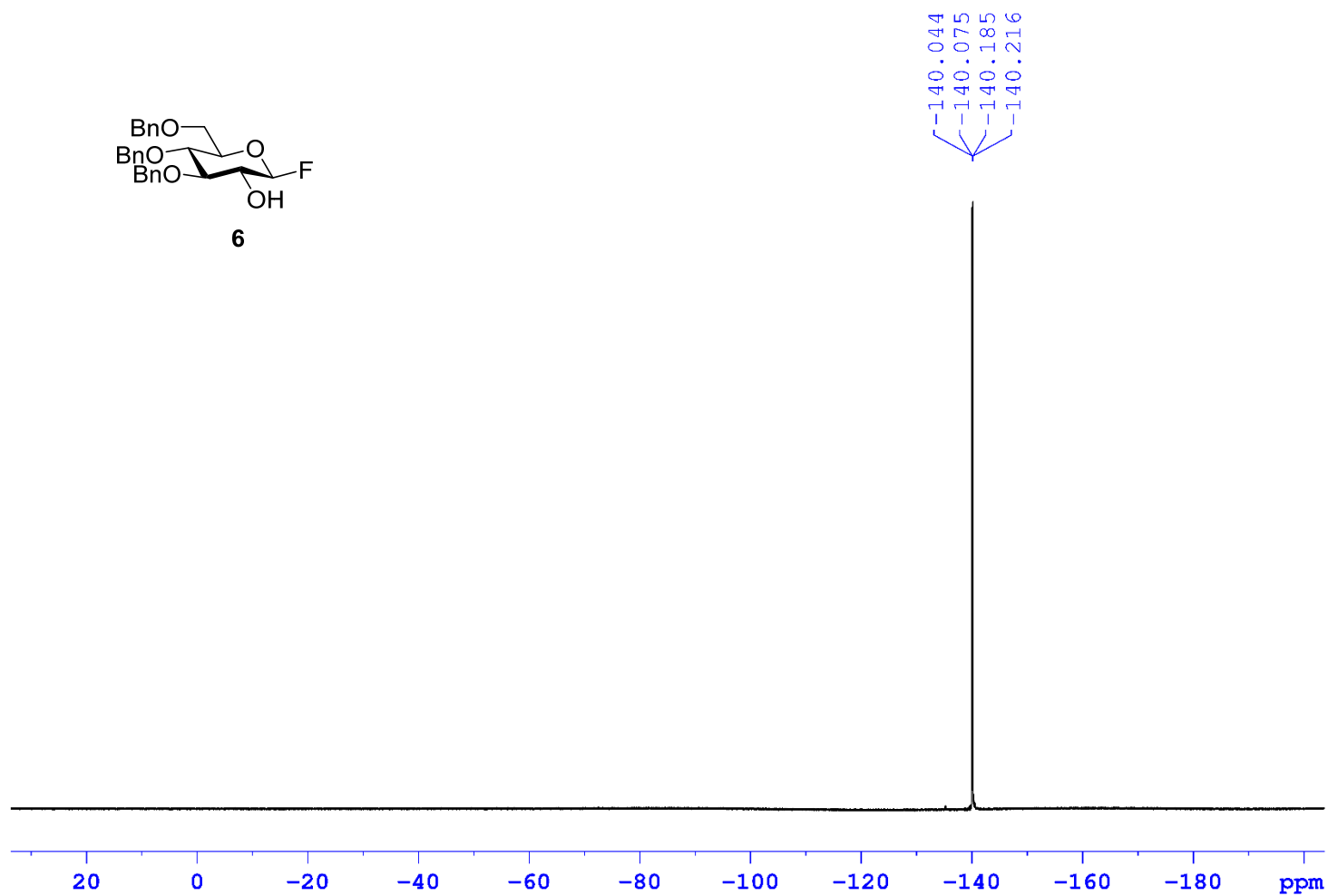
^{13}C NMR (100 MHz) of **6** in CDCl_3



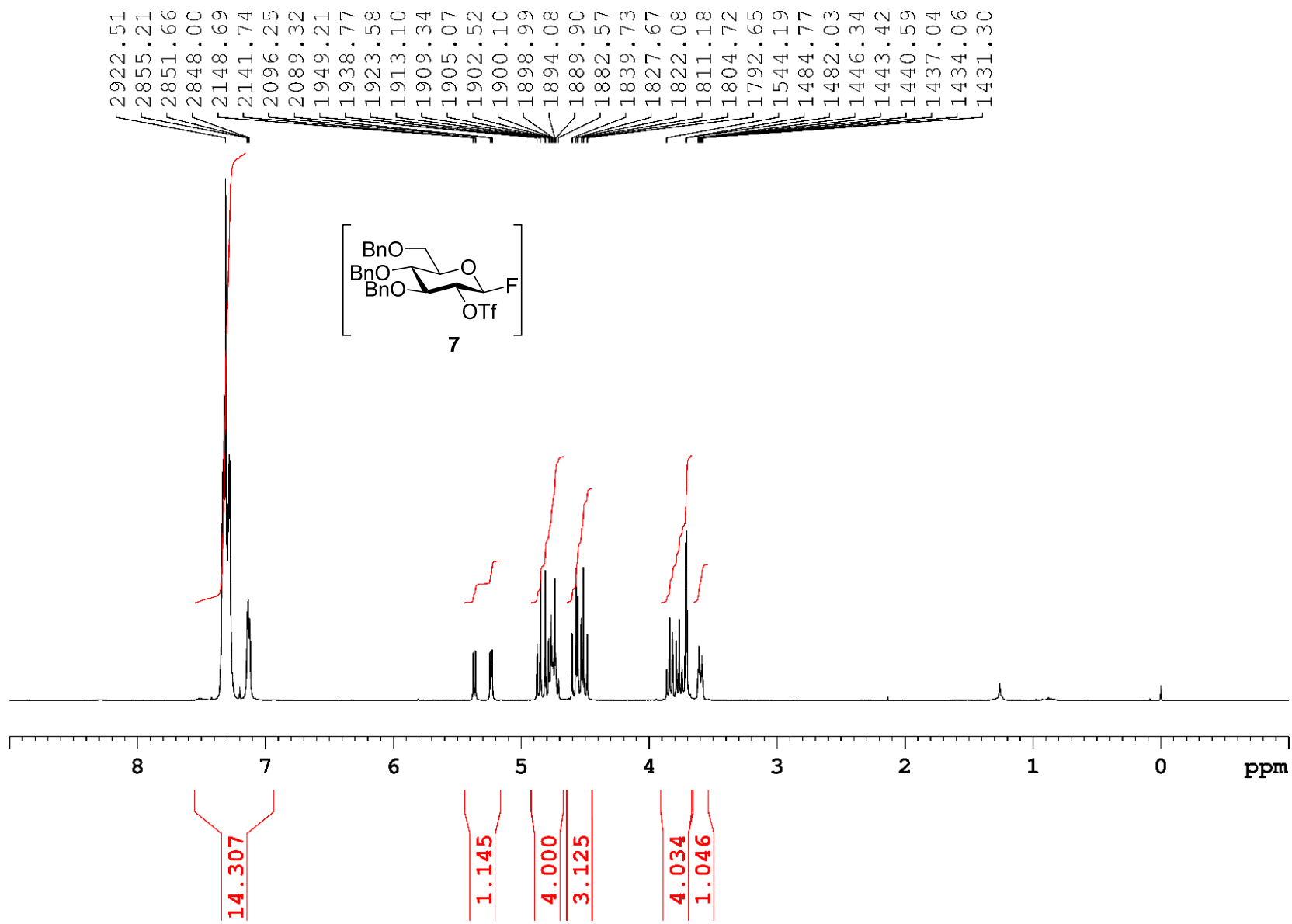
DEPT NMR (100 MHz) of **6** in CDCl₃



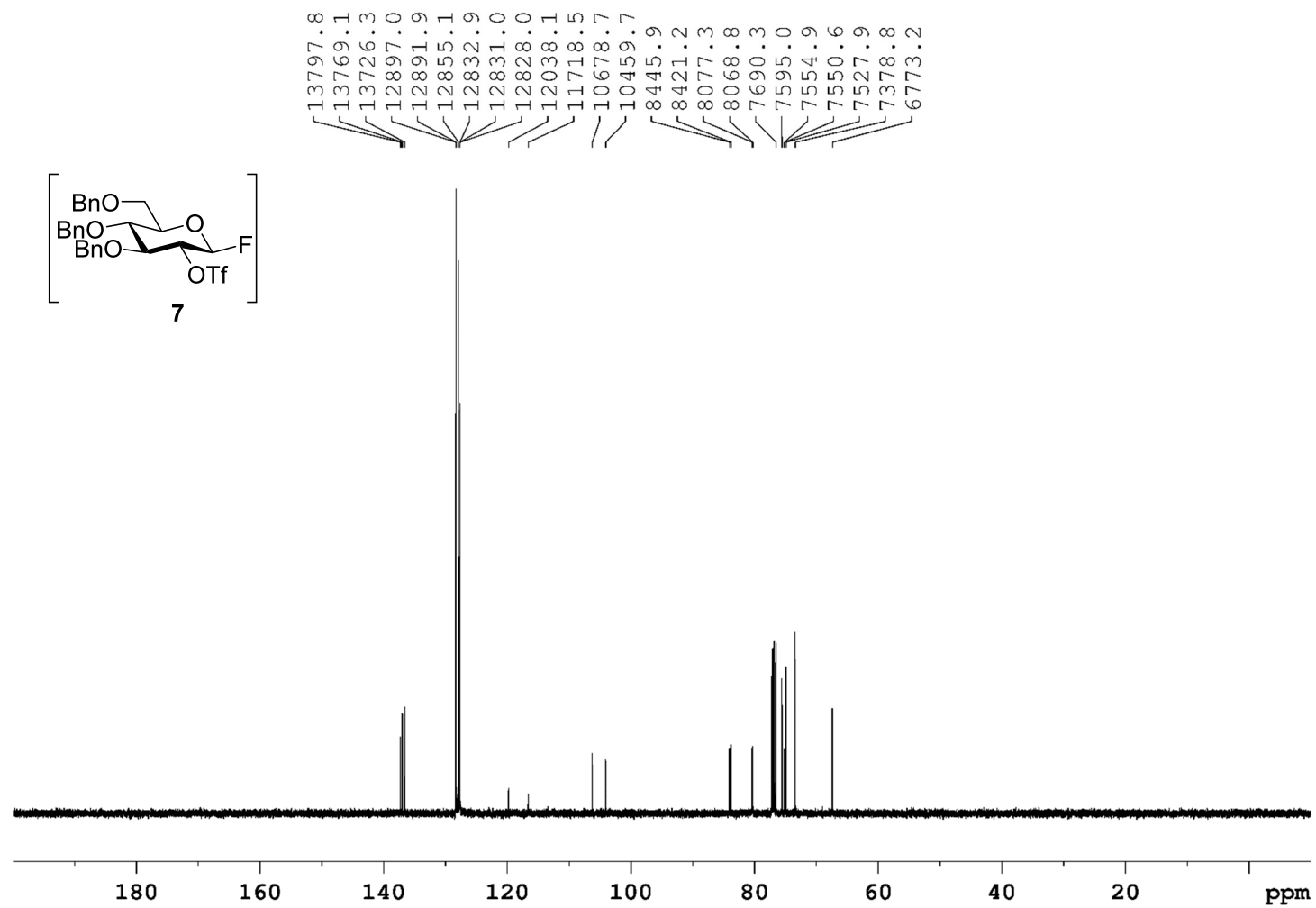
^{19}F NMR (376 MHz) of **6** in CDCl_3



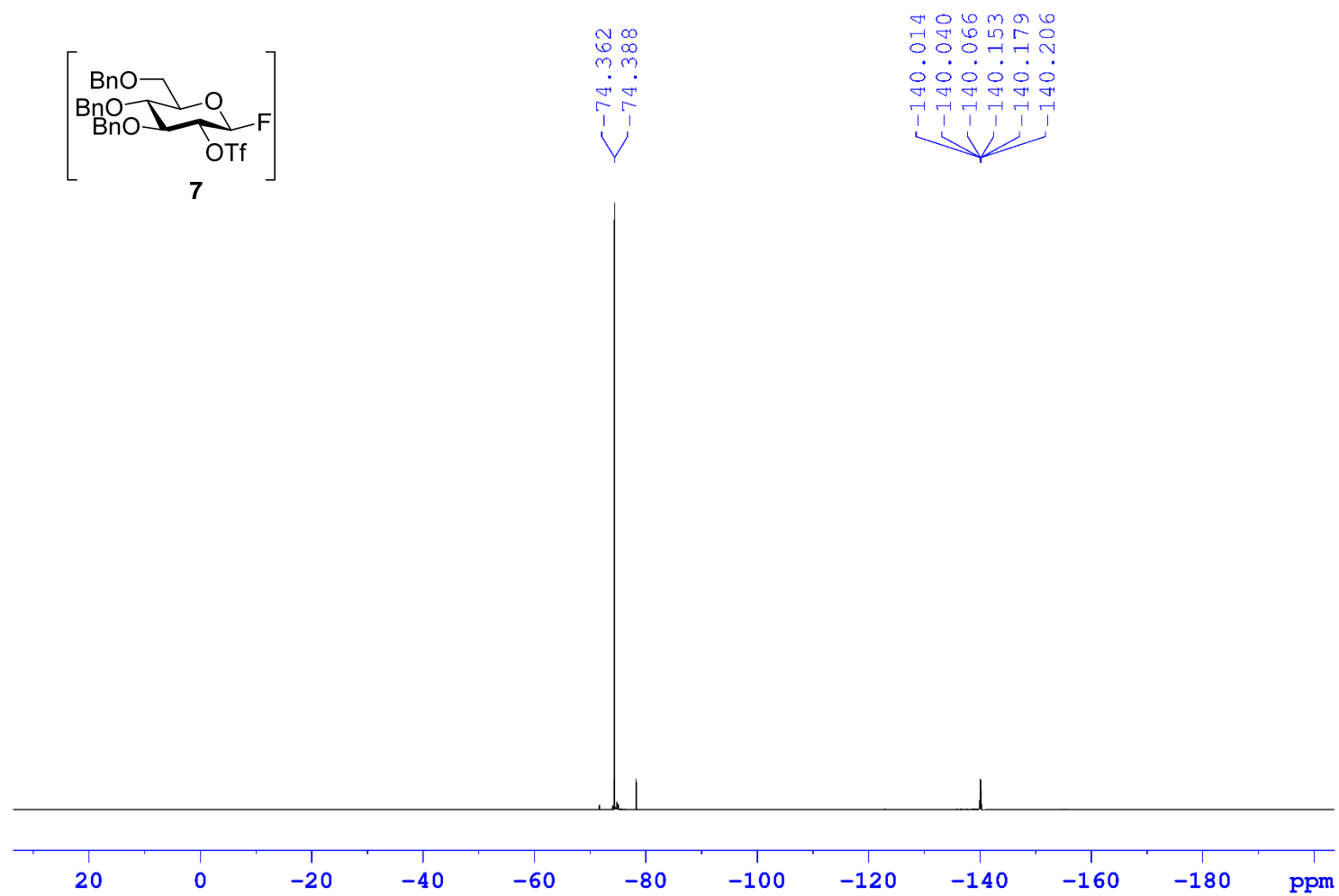
¹H NMR (400 MHz) of **7** in CDCl₃



^{13}C NMR (100 MHz) of **7** in CDCl_3



^{19}F NMR (376 MHz) of **7** in CDCl_3

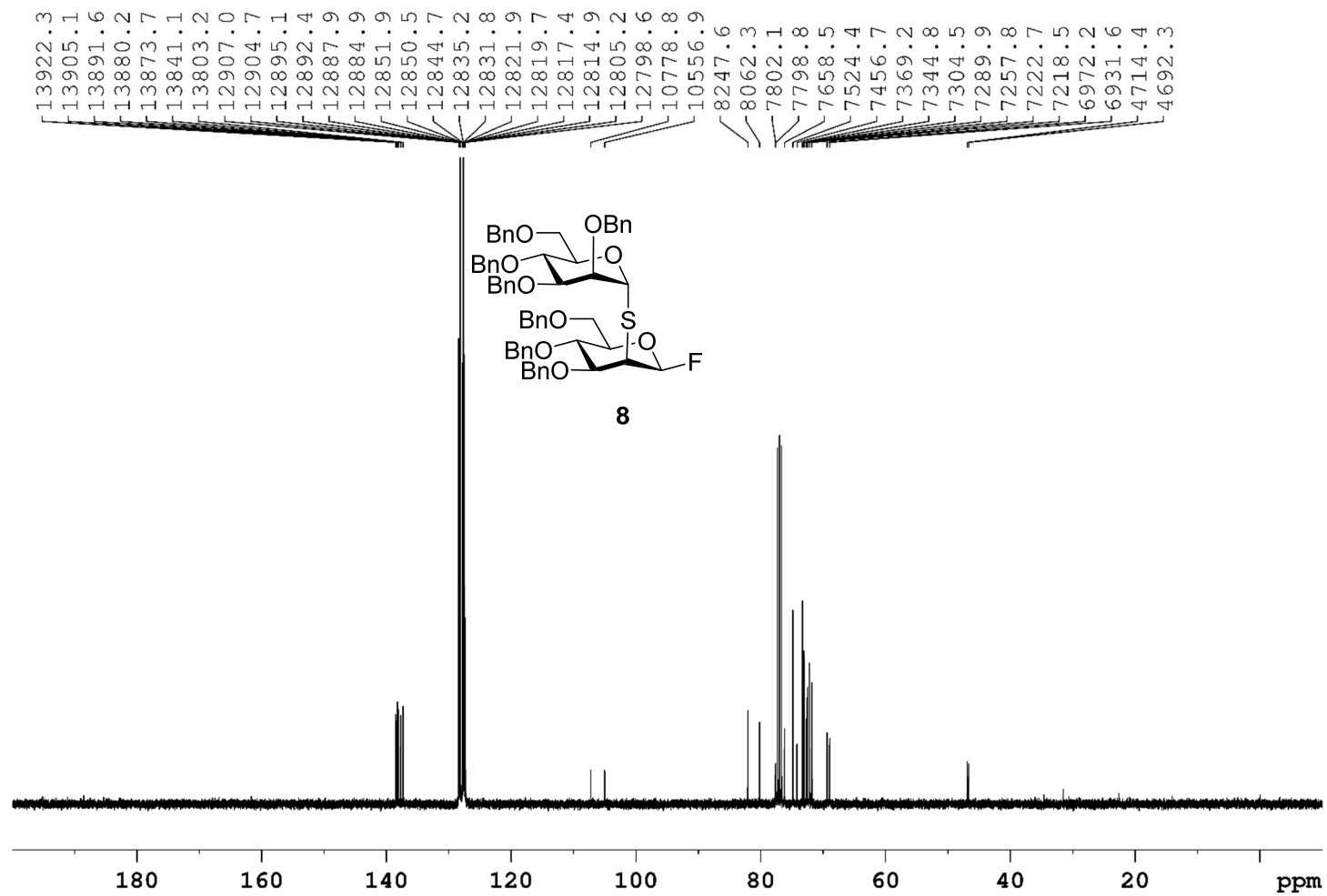


Chemical structure of compound 8:

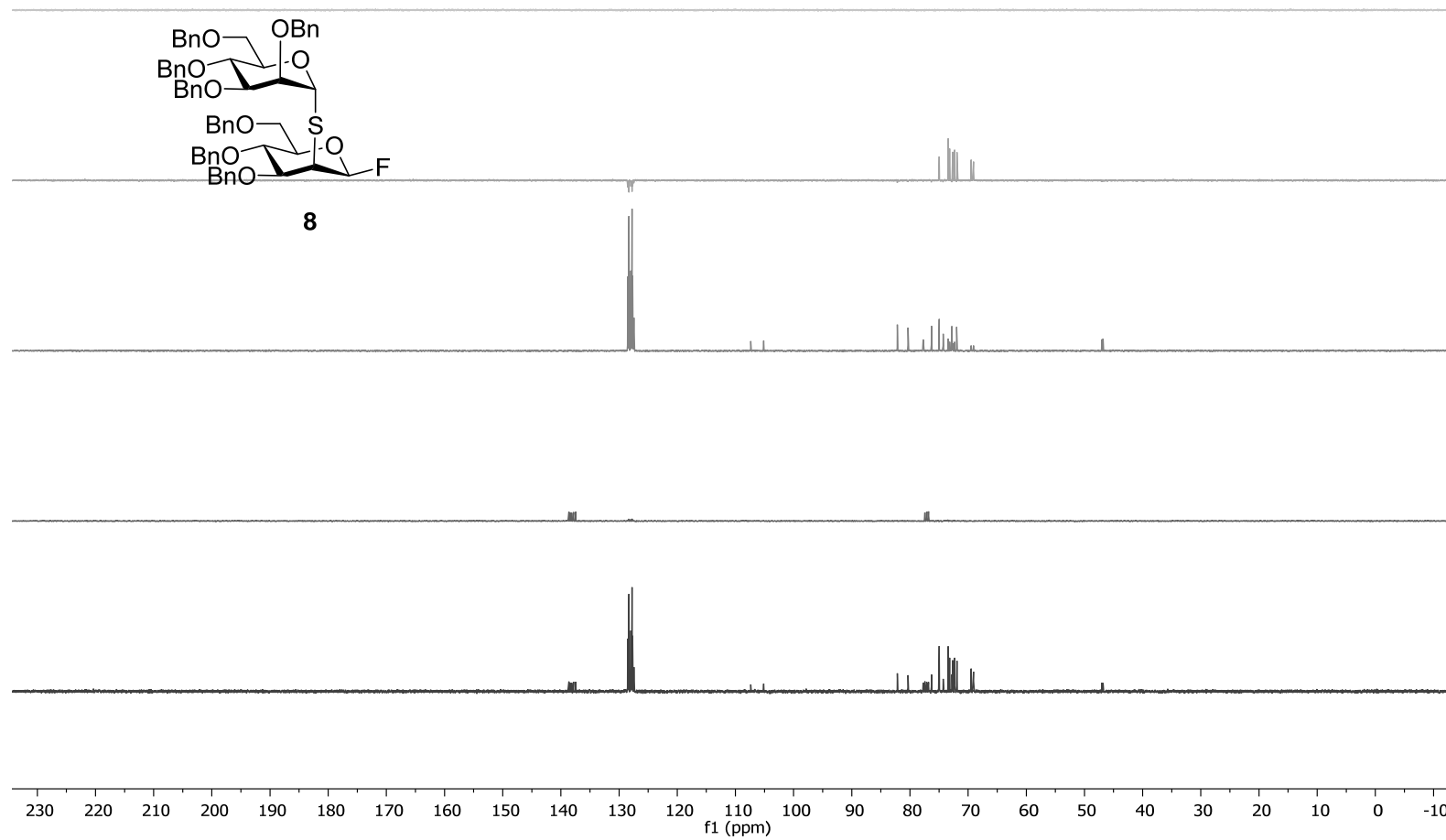
O[C@H]1[C@@H](OC(=O)c2ccccc2)[C@H](OC(=O)c3ccccc3)[C@H](OC(=O)c4ccccc4)[C@H](OC(=O)c5ccccc5)[C@H](OC(=O)c6ccccc6)[C@H]1OC(F)(C)C

¹H NMR spectrum (CDCl₃):

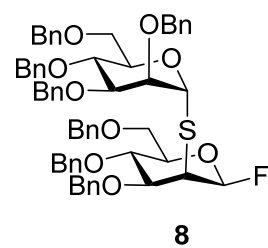
Chemical Shift (ppm)	Integration
7.25 - 7.45	34.49
5.50 - 5.65	2.00
4.80 - 5.00	1.06
4.60 - 4.75	2.15
4.40 - 4.55	11.12
4.20 - 4.35	1.02
4.00 - 4.15	1.09
3.80 - 3.95	1.05
3.60 - 3.75	5.04
3.40 - 3.55	3.05
3.20 - 3.35	1.05

^{13}C NMR (100 MHz) of **8** in CDCl_3 

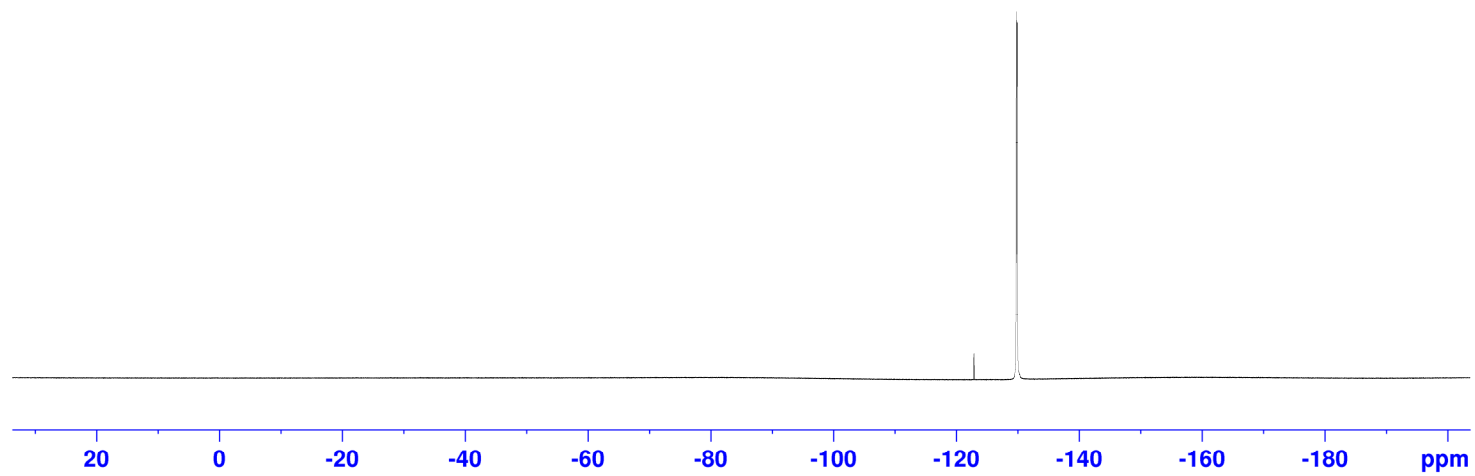
DEPT NMR (100 MHz) of **8** in CDCl₃



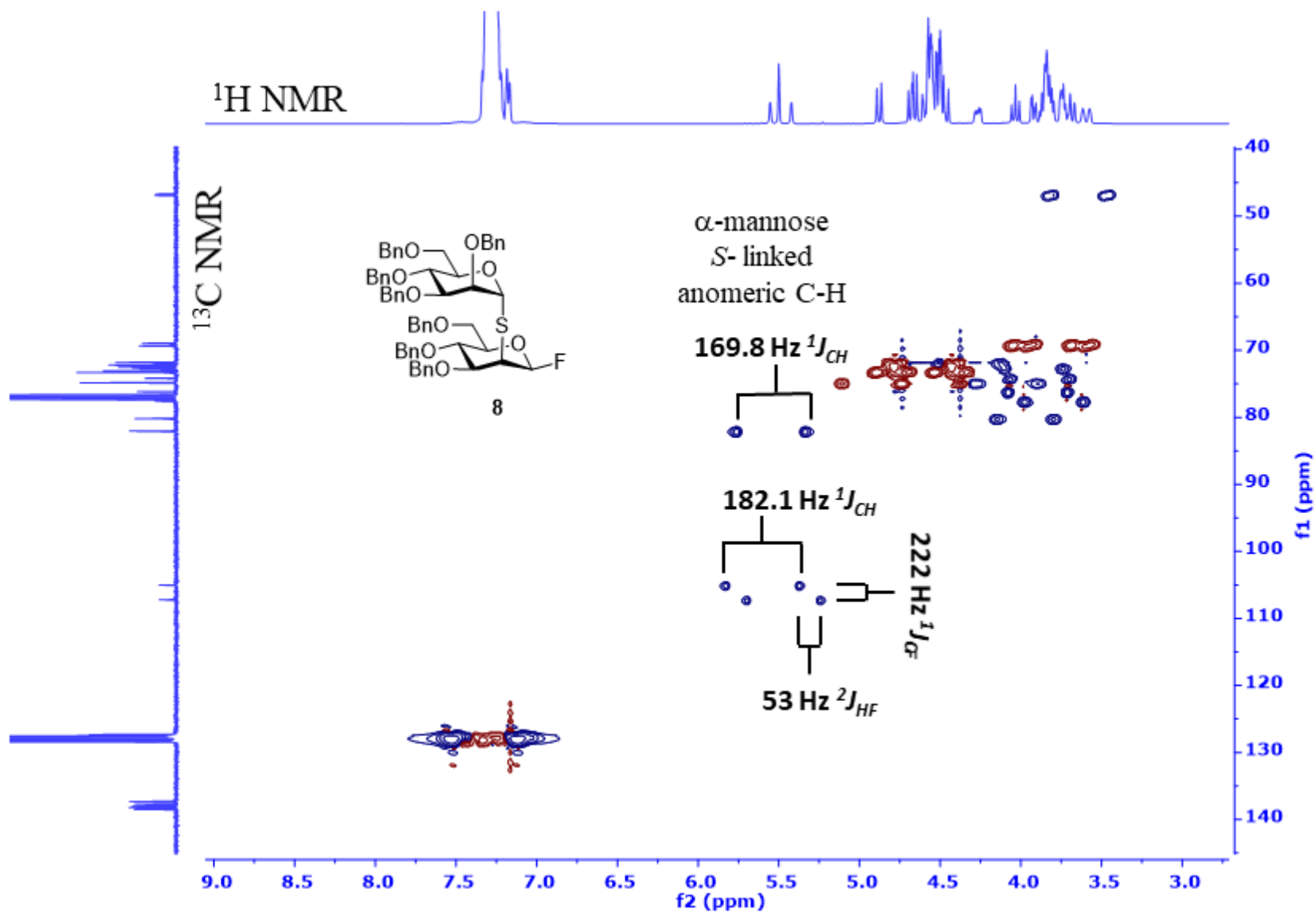
^{19}F NMR (376 MHz) of **8** in CDCl_3

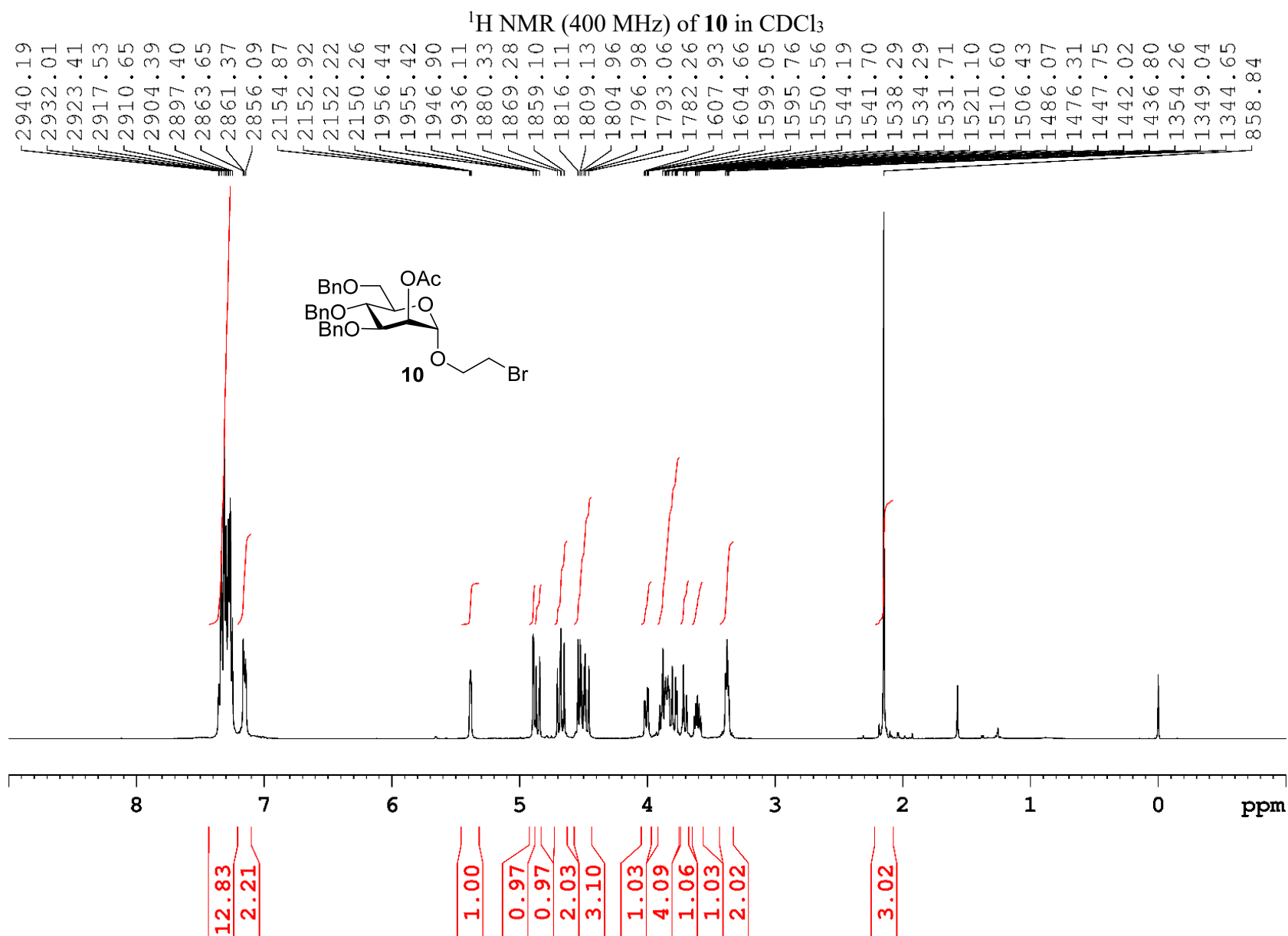


-129.749
-129.789
-129.887
-129.928

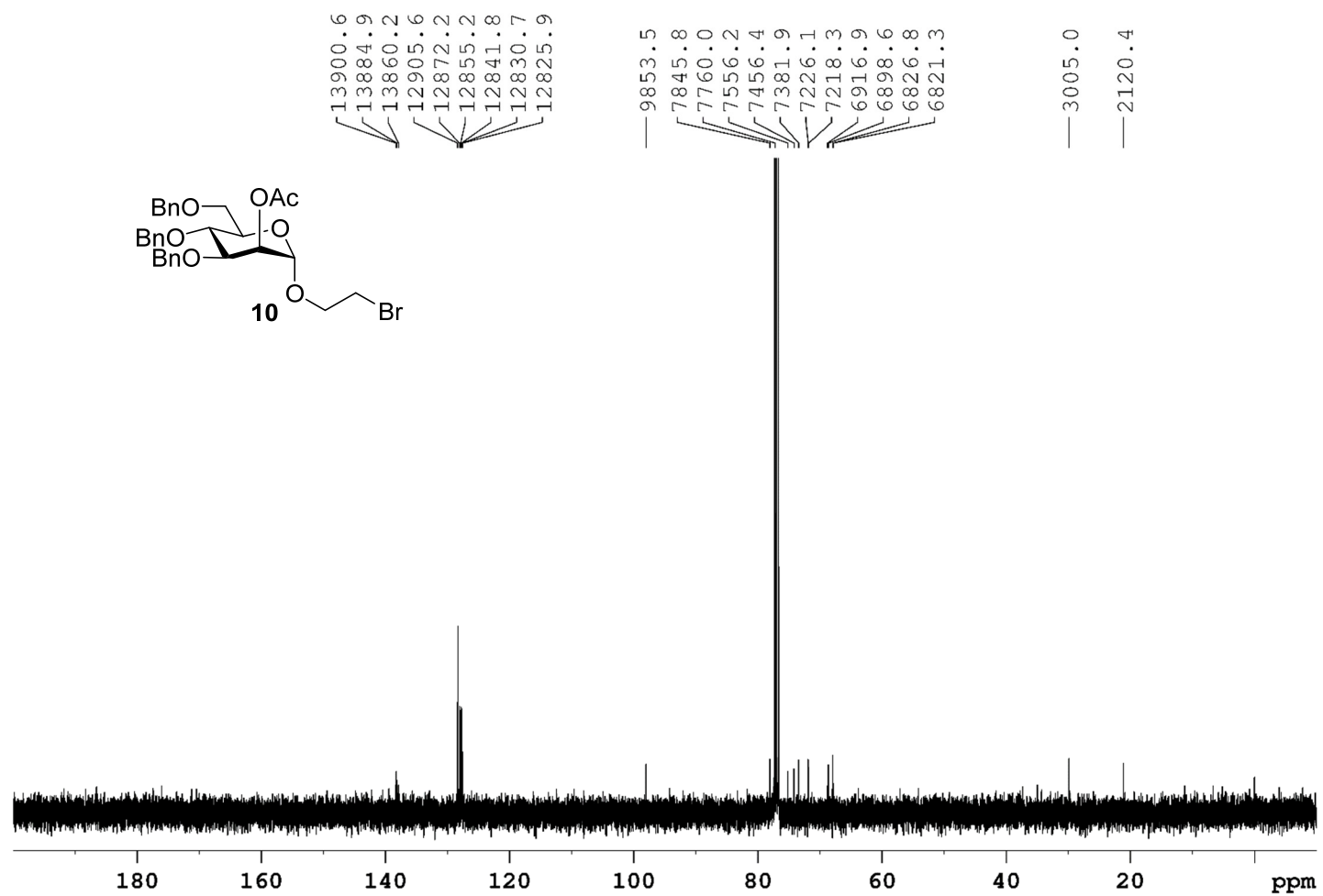


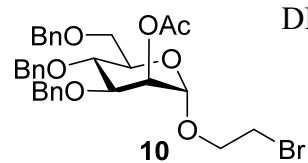
^1H - ^{13}C coupled HSQC NMR (400 MHz) of **8** in CDCl_3





^{13}C NMR (100 MHz) of **10** in CDCl_3





DEPT NMR (100 MHz) of **10** in CDCl₃

CH₃



CH₂



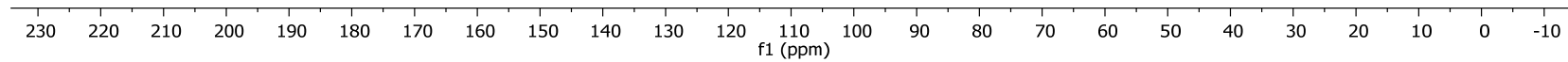
CH



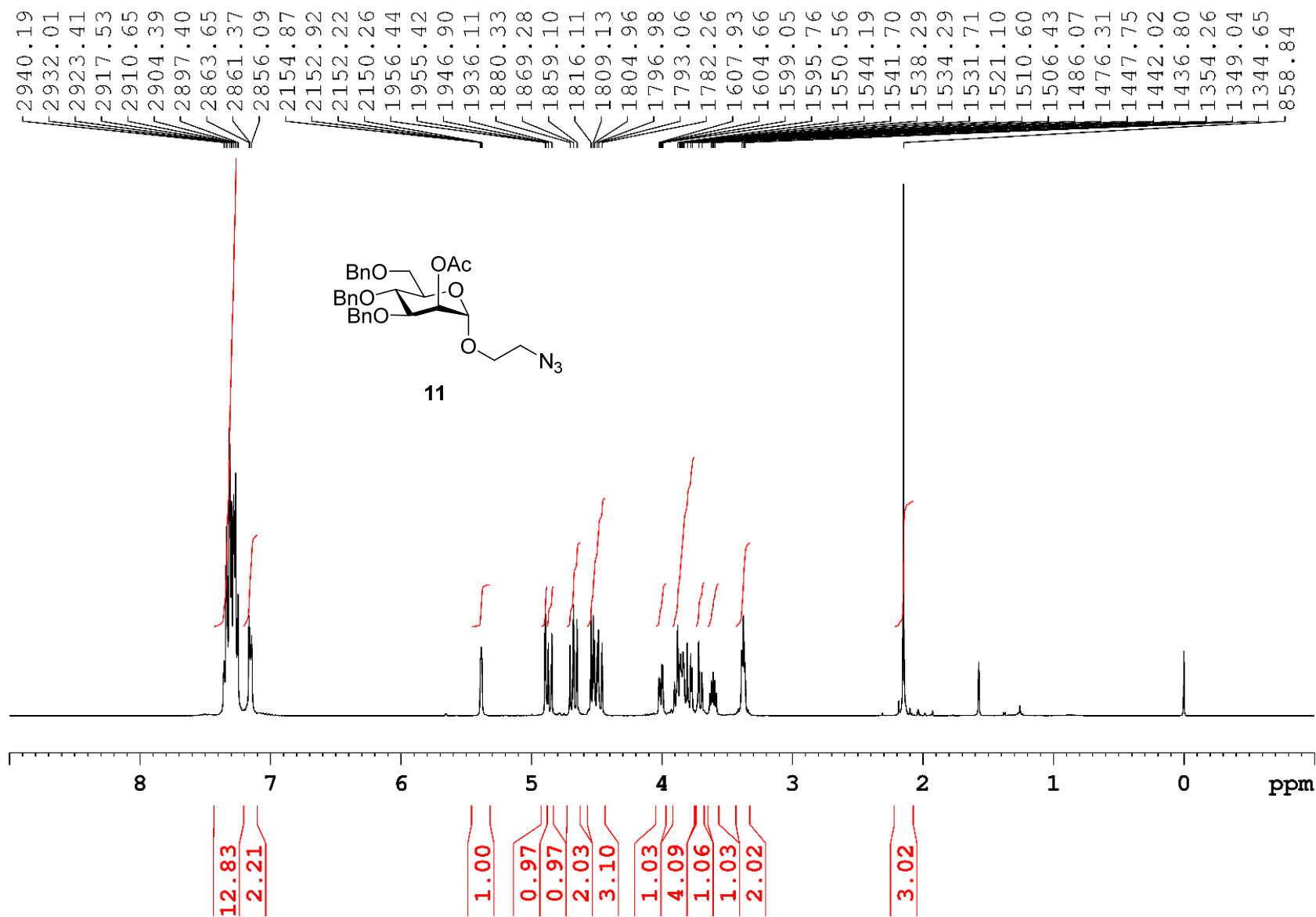
Cq



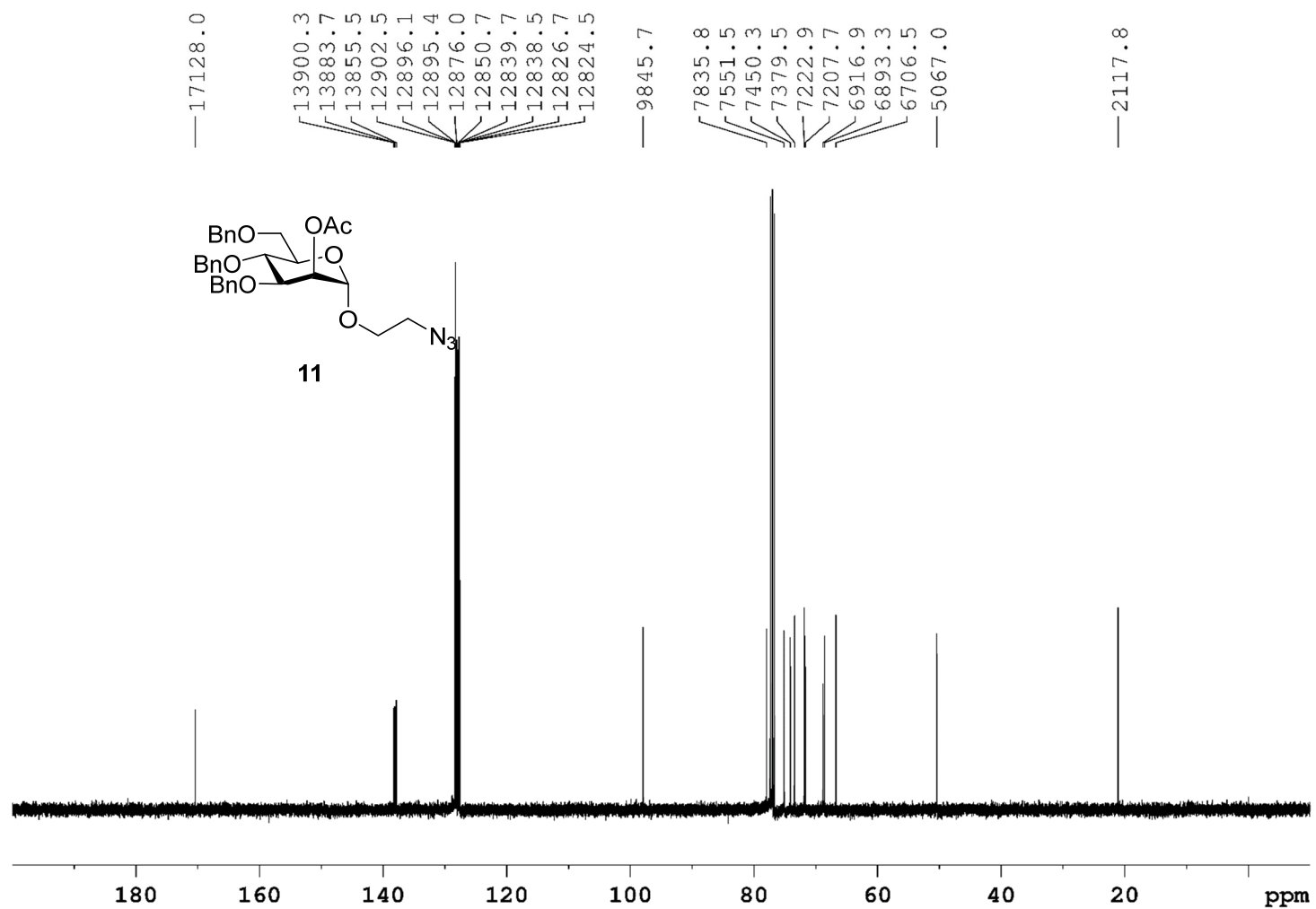
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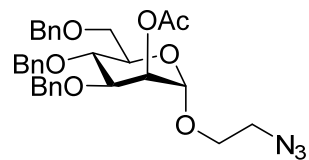
¹H NMR (400 MHz) of **11** in CDCl₃



^{13}C NMR (100 MHz) of **11** in CDCl_3



DEPT NMR (100 MHz) of **11** in CDCl₃



11

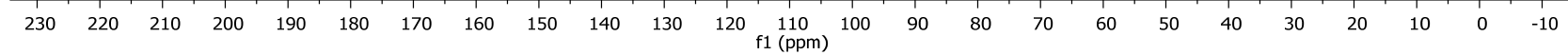
CH₃

CH₂

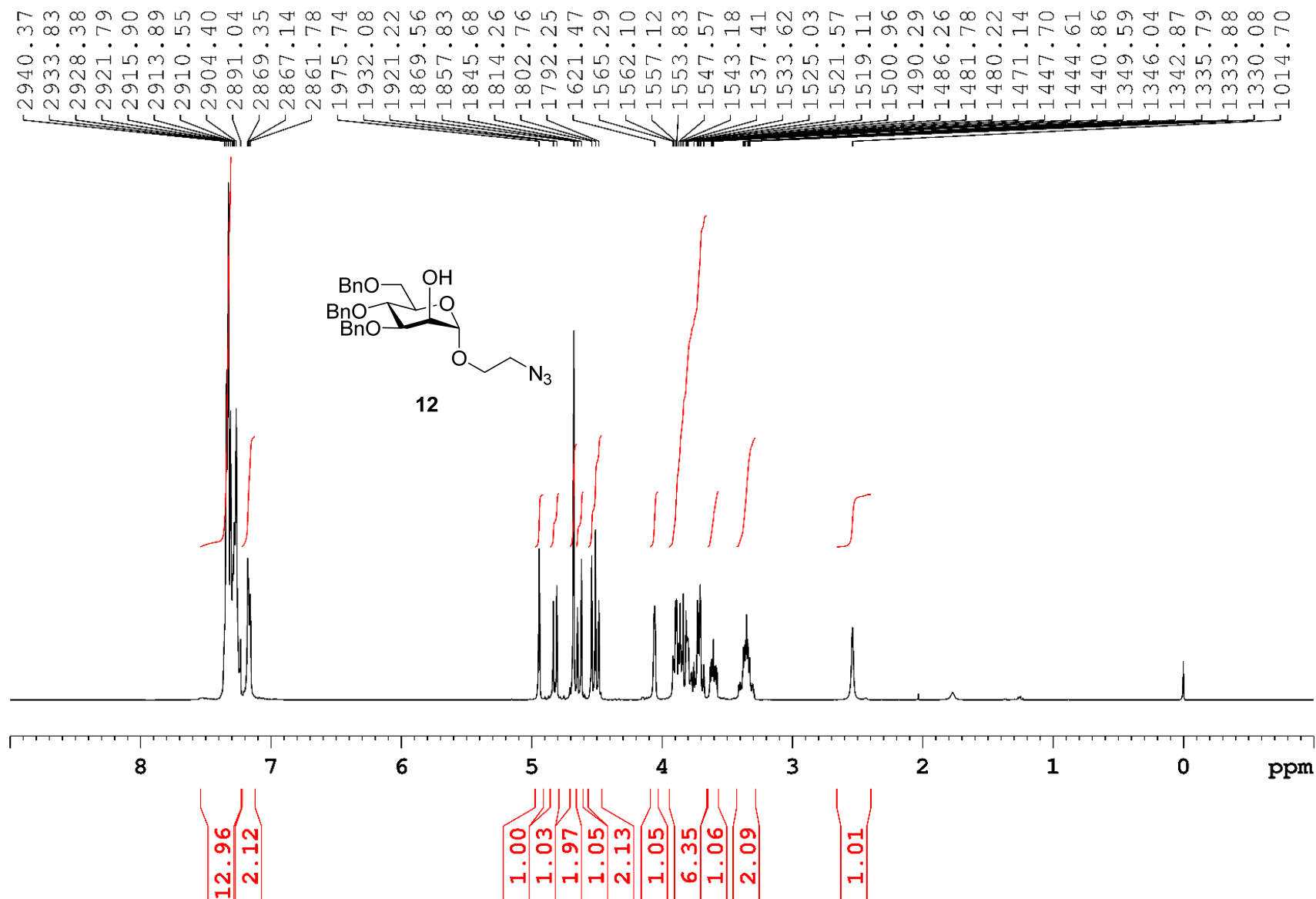
CH

Cq

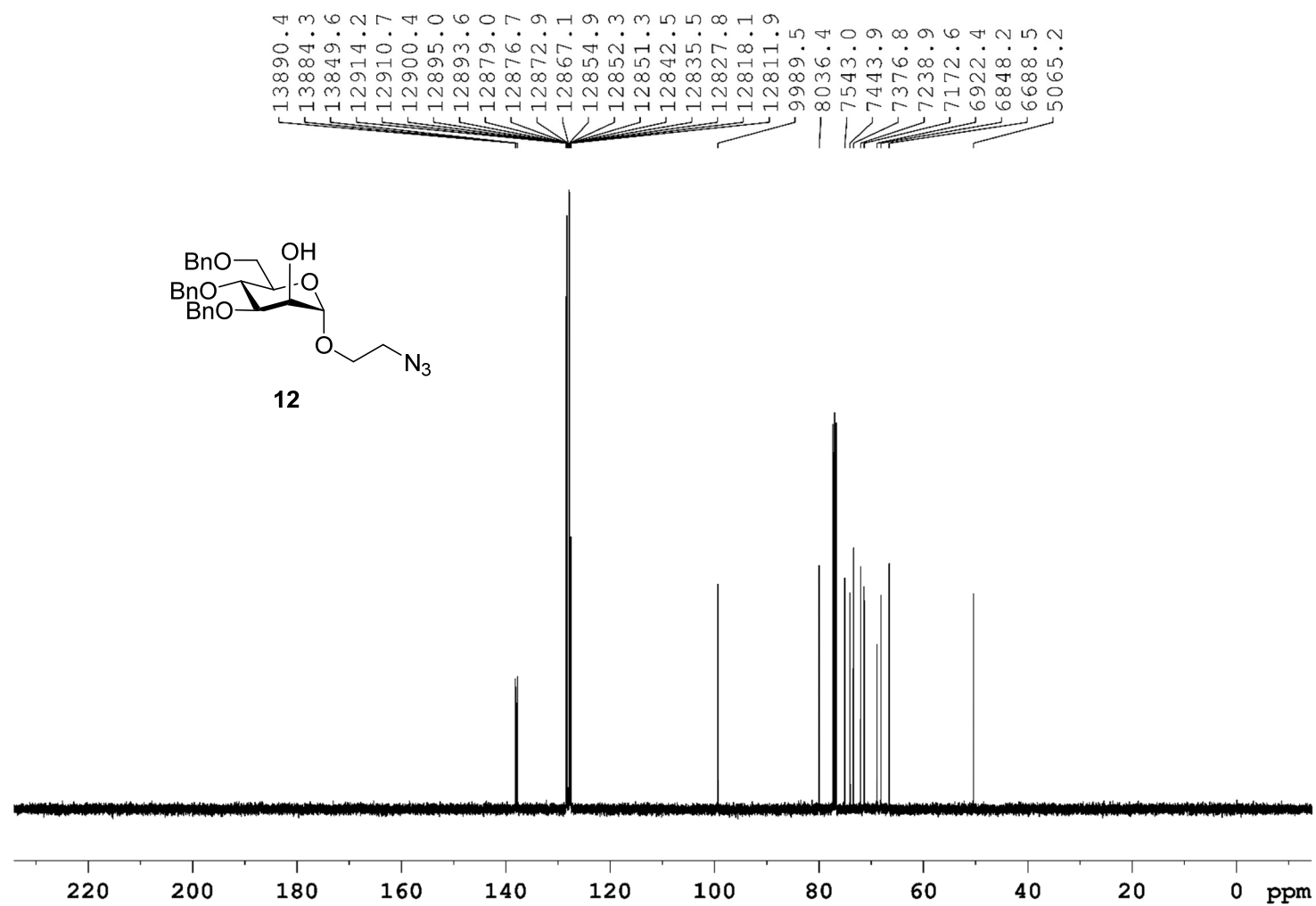
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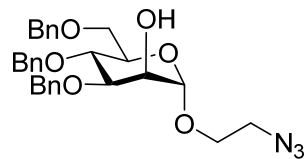
¹H NMR (400 MHz) of **12** in CDCl₃



^{13}C NMR (100 MHz) of **12** in CDCl_3



DEPT NMR (100 MHz) of **12** in CDCl₃



12

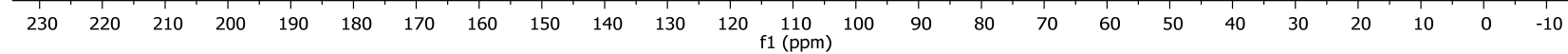
CH₃

CH₂

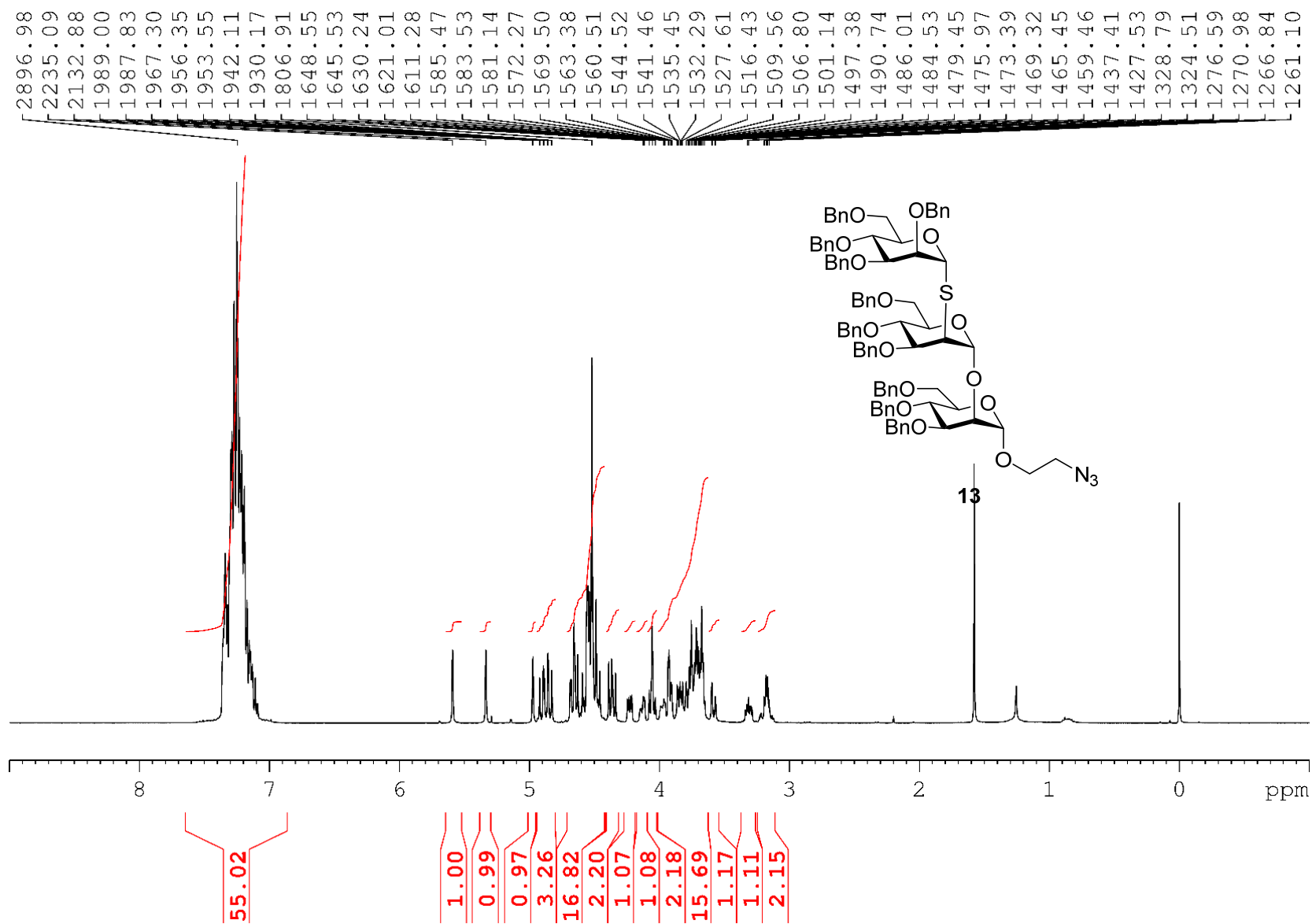
CH

Cq

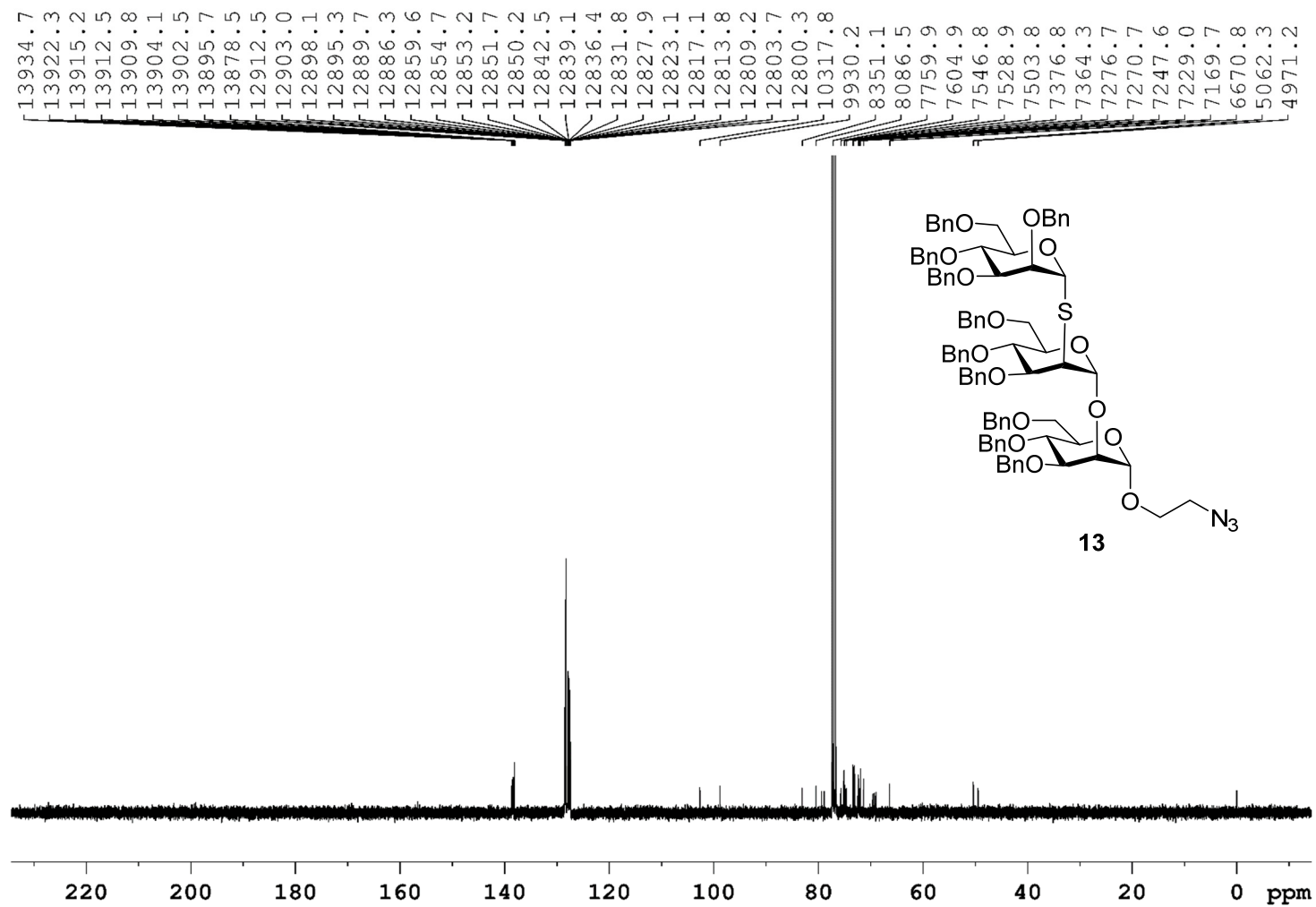
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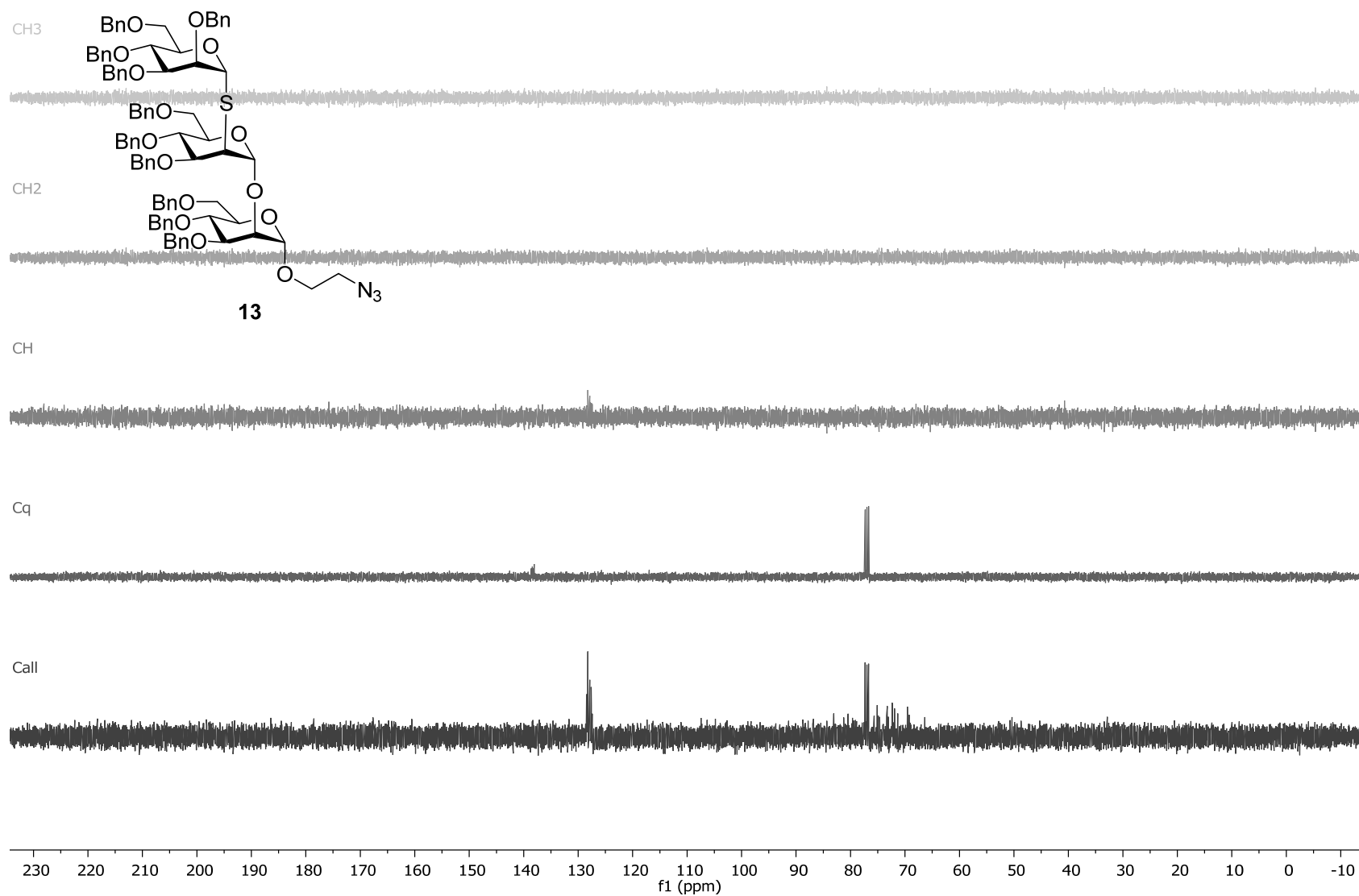
¹H NMR (400 MHz) of **13** in CDCl₃



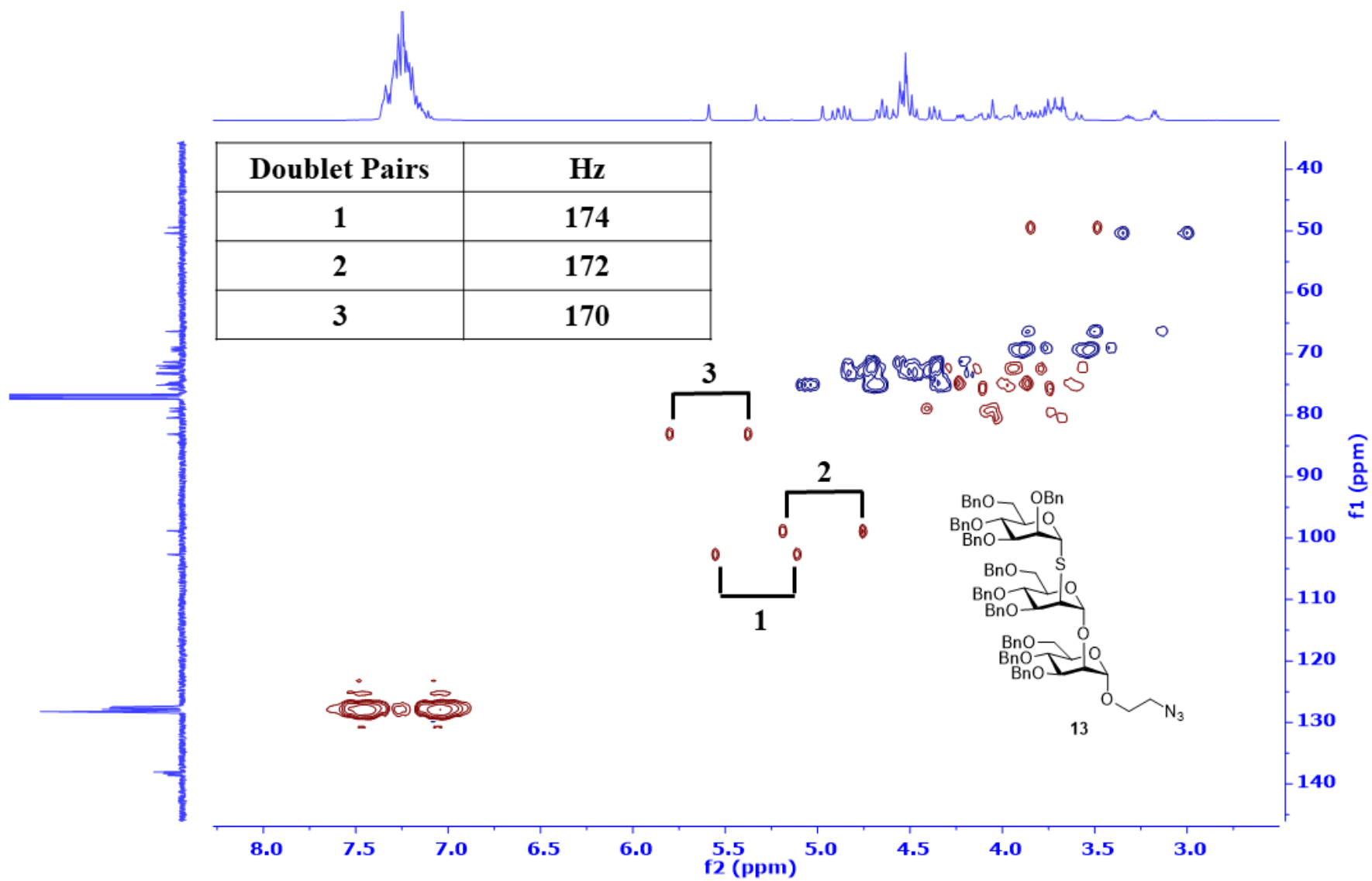
^{13}C NMR (100 MHz) of **13** in CDCl_3

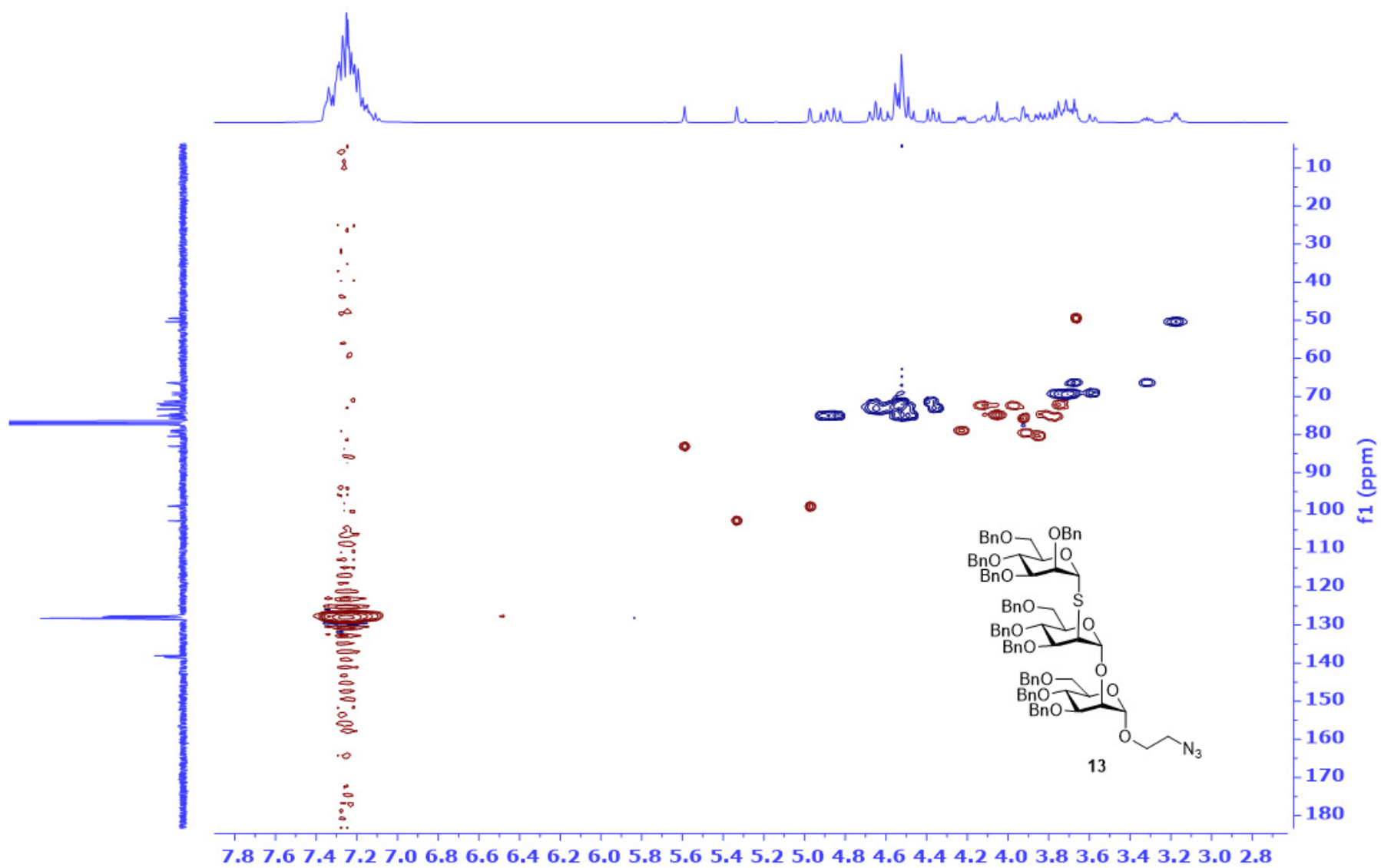


DEPT NMR (100 MHz) of **13** in CDCl₃

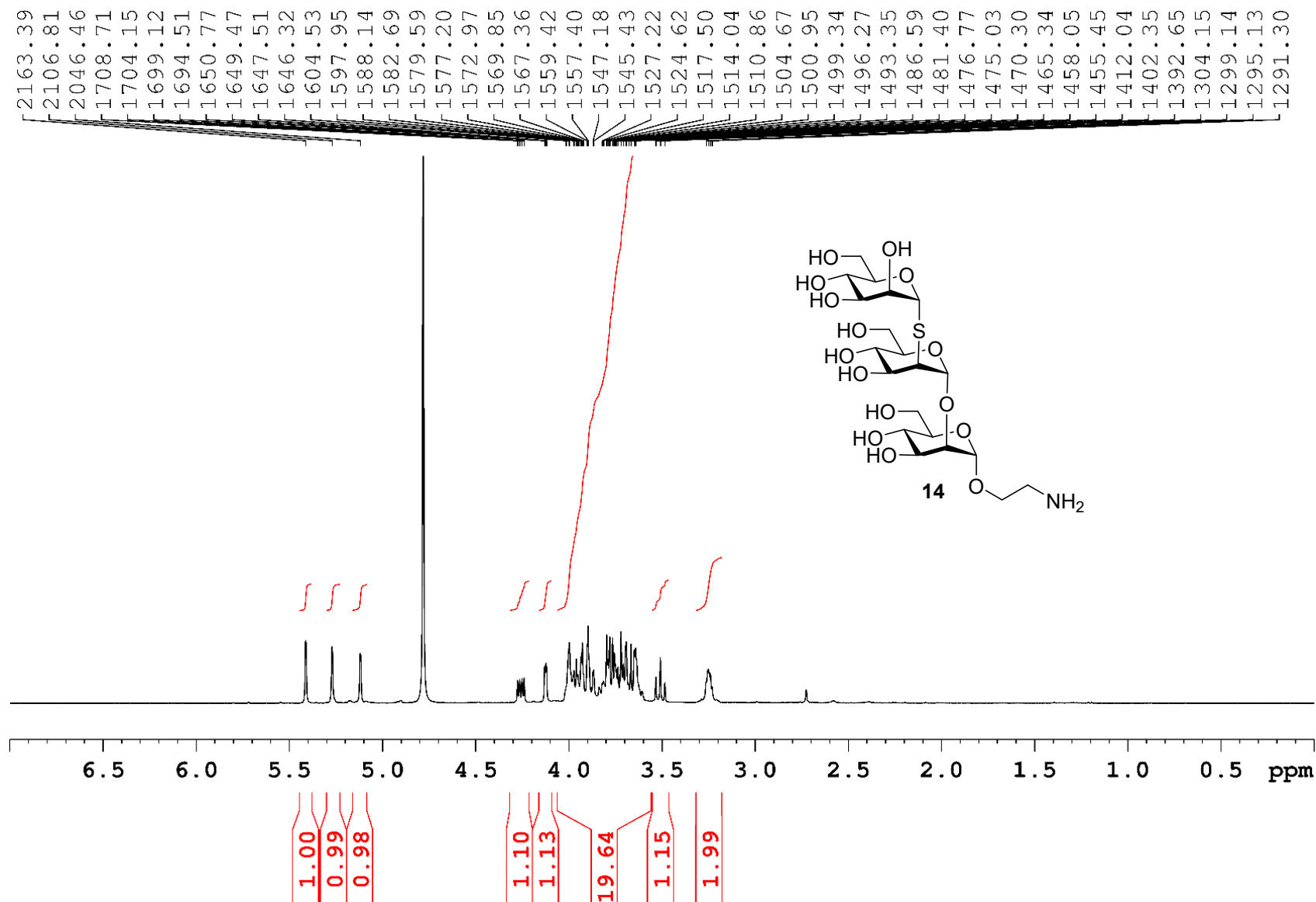


^1H - ^{13}C coupled HSQC NMR (400 MHz) of **13** in CDCl_3

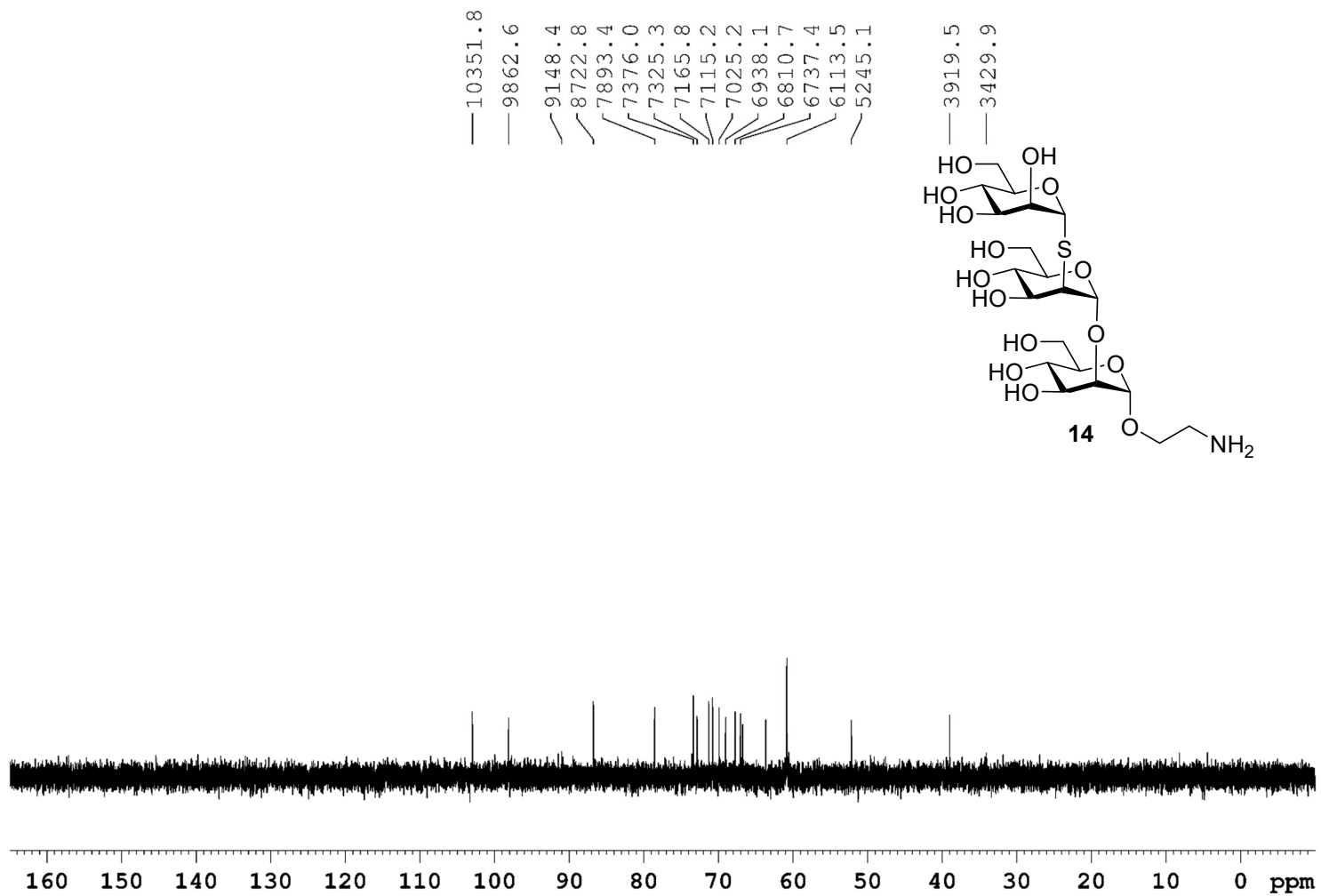


$^1\text{H} - ^{13}\text{C}$ HSQC NMR (400 MHz) of **13** in CDCl_3 

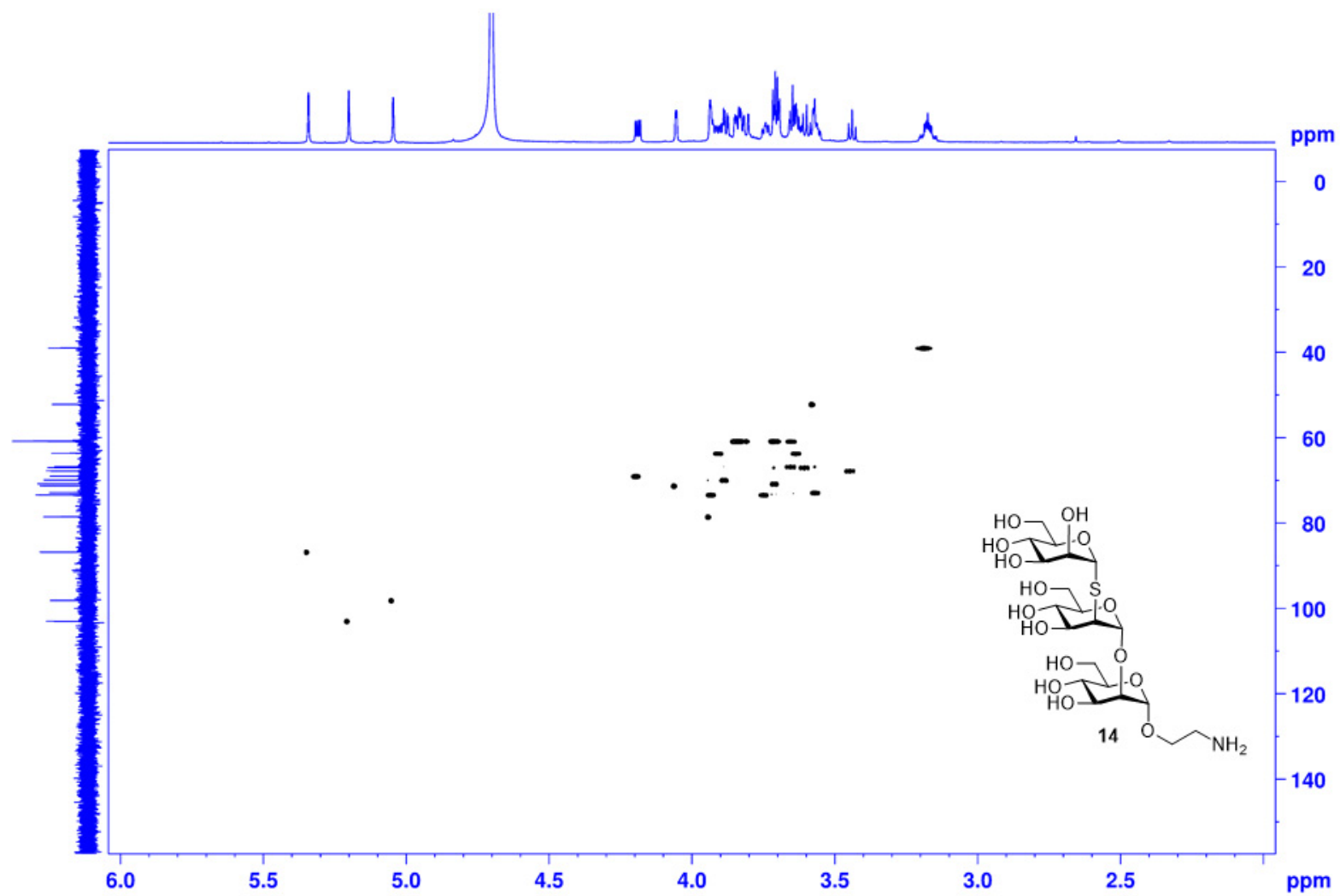
¹H NMR (400 MHz) of **14** in D₂O



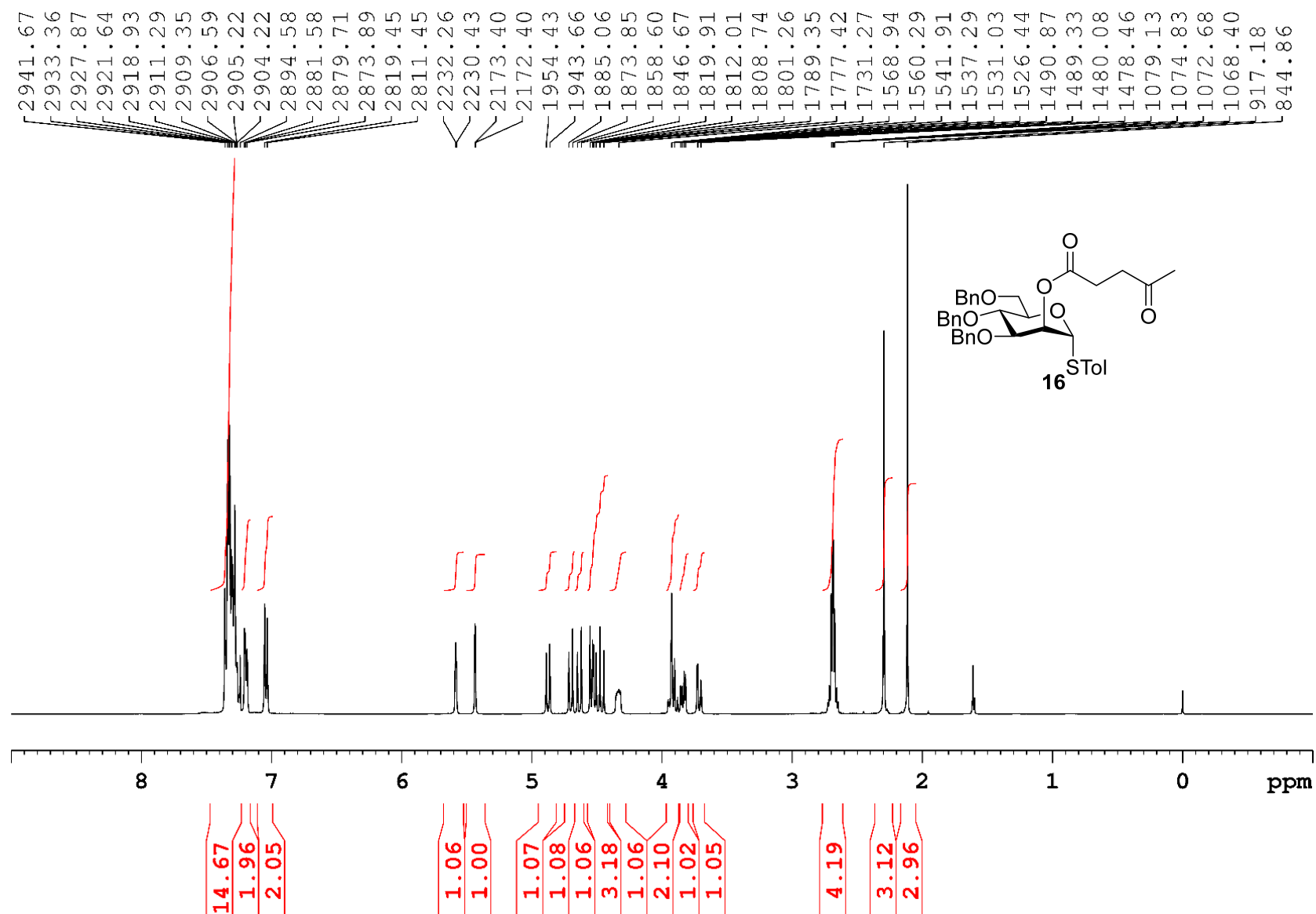
^{13}C NMR (100 MHz) of **14** in D_2O



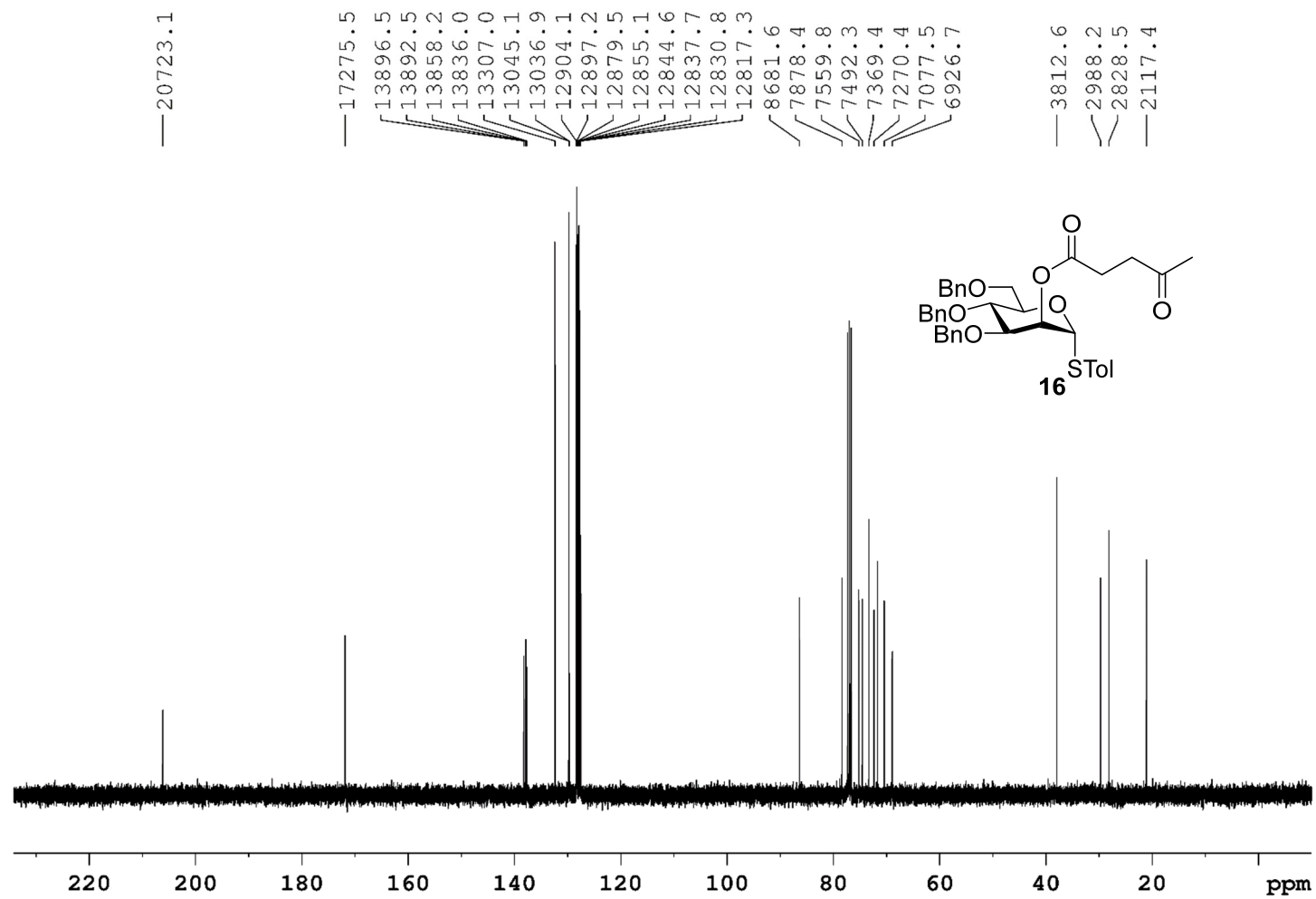
^1H - ^{13}C HSQC NMR (800 MHz) of **14** in D_2O



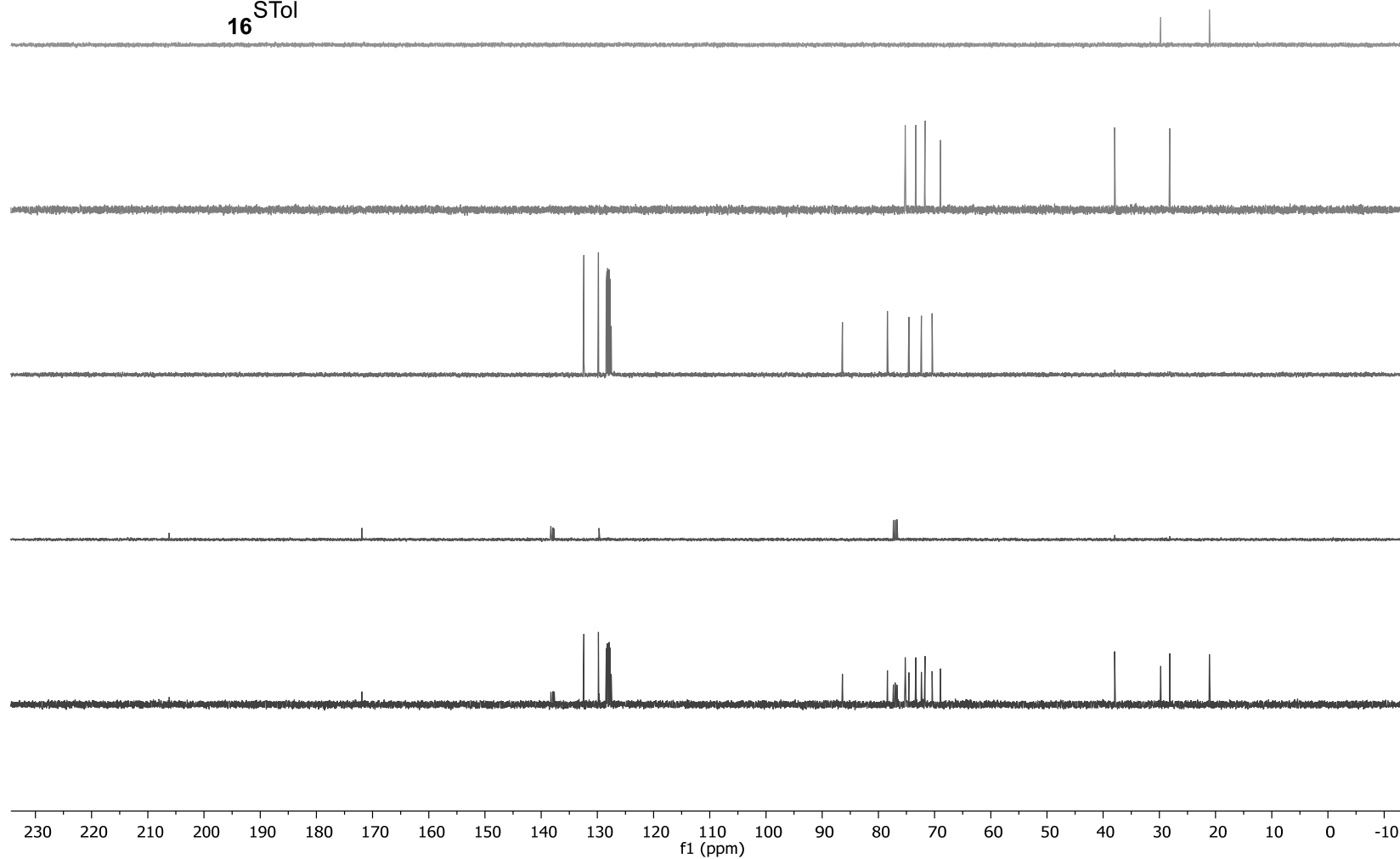
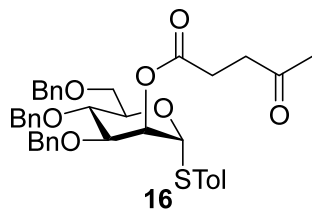
¹H NMR (400 MHz) of **16** in CDCl₃



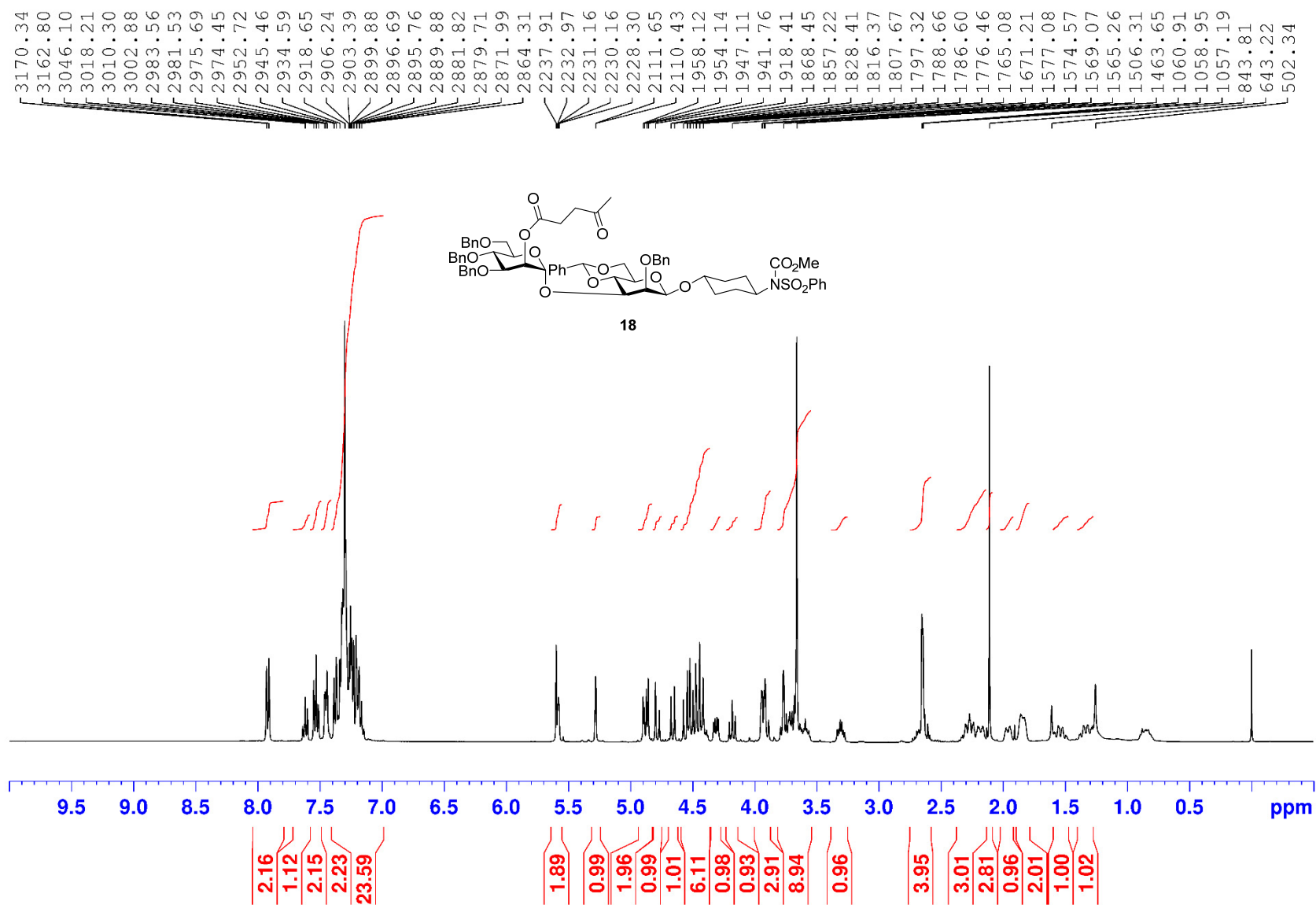
^{13}C NMR (100 MHz) of **16** in CDCl_3



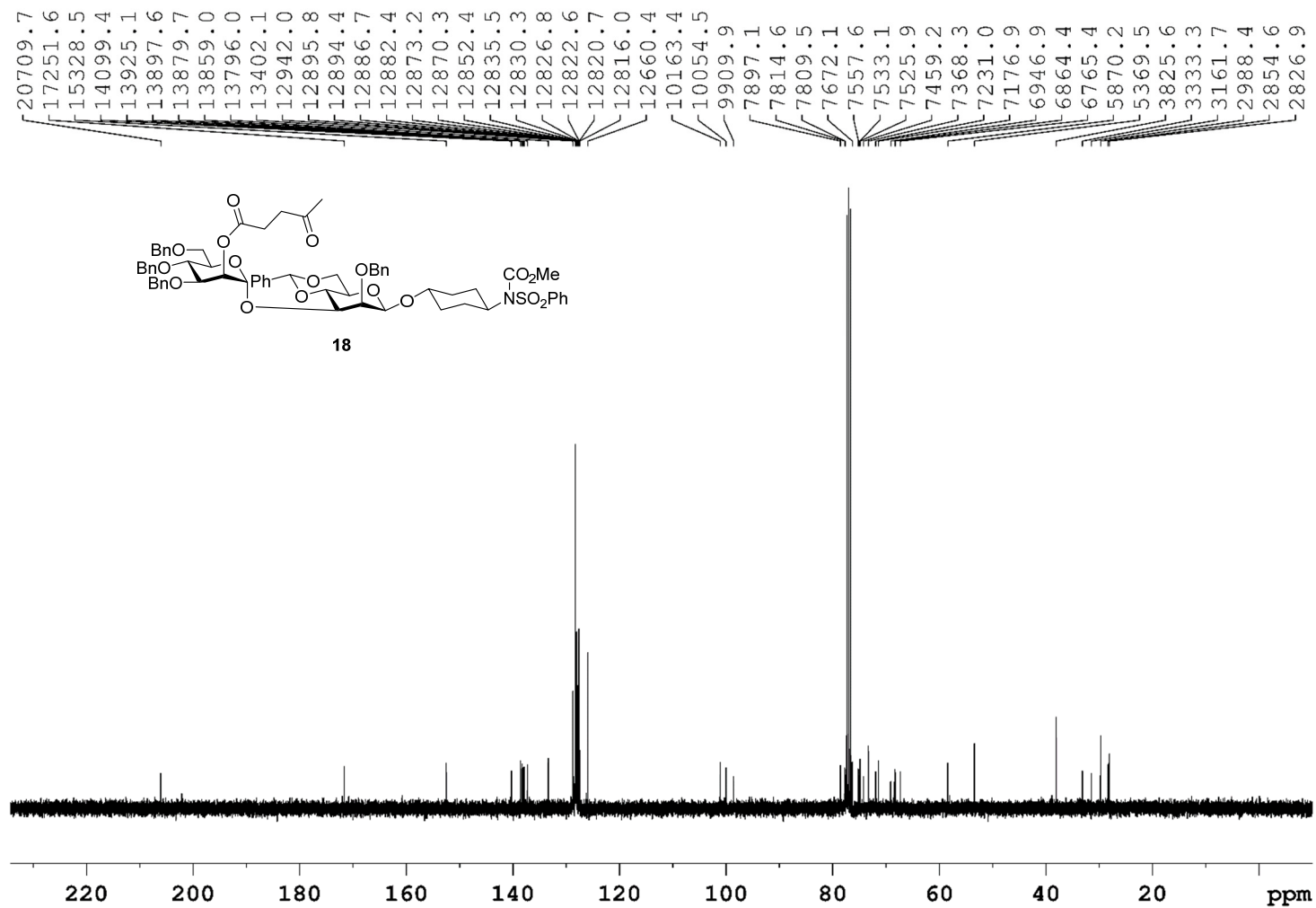
DEPT NMR (100 MHz) of **16** in CDCl₃



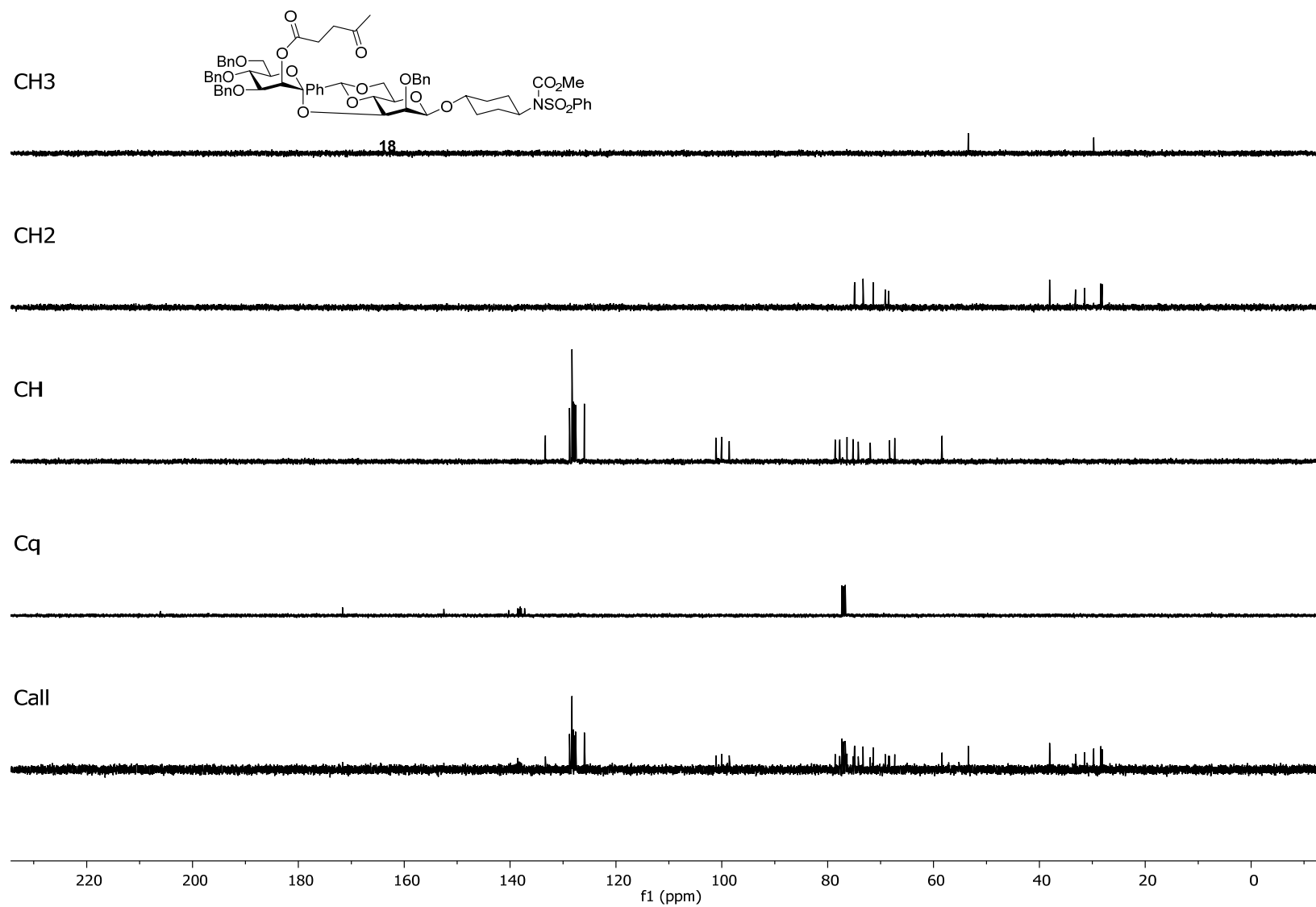
¹H NMR (400 MHz) of **18** in CDCl₃



^{13}C NMR (100 MHz) of **18** in CDCl_3



DEPT NMR (100 MHz) of **18** in CDCl₃



Chemical structure of 19: COC(=O)[C@H]1CC[C@@H](CO[C@@H]2[C@@H](OC(=O)c3ccccc3)[C@H](OC(=O)c4ccccc4)[C@H](OC(=O)c5ccccc5)O2)[C@H](O)[C@H]1O

¹H NMR spectrum (CDCl₃):

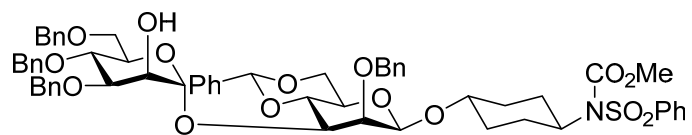
Chemical Shift (ppm)	Integration
7.26, 7.21, 7.16, 7.12, 7.07, 7.02, 6.97, 6.92, 6.87, 6.82, 6.77, 6.72, 6.67, 6.62, 6.57, 6.52, 6.47, 6.42, 6.37, 6.32, 6.27, 6.22, 6.17, 6.12, 6.07, 6.02, 5.97, 5.92, 5.87, 5.82, 5.77, 5.72, 5.67, 5.62, 5.57, 5.52, 5.47, 5.42, 5.37, 5.32, 5.27, 5.22, 5.17, 5.12, 5.07, 5.02, 4.97, 4.92, 4.87, 4.82, 4.77, 4.72, 4.67, 4.62, 4.57, 4.52, 4.47, 4.42, 4.37, 4.32, 4.27, 4.22, 4.17, 4.12, 4.07, 4.02, 3.97, 3.92, 3.87, 3.82, 3.77, 3.72, 3.67, 3.62, 3.57, 3.52, 3.47, 3.42, 3.37, 3.32, 3.27, 3.22, 3.17, 3.12, 3.07, 3.02, 2.97, 2.92, 2.87, 2.82, 2.77, 2.72, 2.67, 2.62, 2.57, 2.52, 2.47, 2.42, 2.37, 2.32, 2.27, 2.22, 2.17, 2.12, 2.07, 2.02, 1.97, 1.92, 1.87, 1.82, 1.77, 1.72, 1.67, 1.62, 1.57, 1.52, 1.47, 1.42, 1.37, 1.32, 1.27, 1.22, 1.17, 1.12, 1.07, 1.02, 0.97, 0.92, 0.87, 0.82, 0.77, 0.72, 0.67, 0.62, 0.57, 0.52, 0.47, 0.42, 0.37, 0.32, 0.27, 0.22, 0.17, 0.12, 0.07, 0.02	2.00, 1.09, 2.20, 26.01, 1.03, 1.00, 3.08, 2.09, 4.18, 1.03, 1.06, 2.07, 1.02, 1.12, 2.09, 2.53, 2.93, 2.05, 1.10, 0.86, 3.37, 3.20, 2.21, 1.69

Chemical structure of compound **19** is shown above the spectrum. The structure is a complex molecule with three sugar units: a 2,3,4-tri-O-benzyl-6-O-(2,3,4-tri-O-benzyl-6-O-(2,3,4-tri-O-benzyl-6-O-methoxycarbonyl-2,3,4-tri-O-benzyl-6-O-phenylsulfonyl)-D-glucopyranosyl)-D-glucopyranoside.

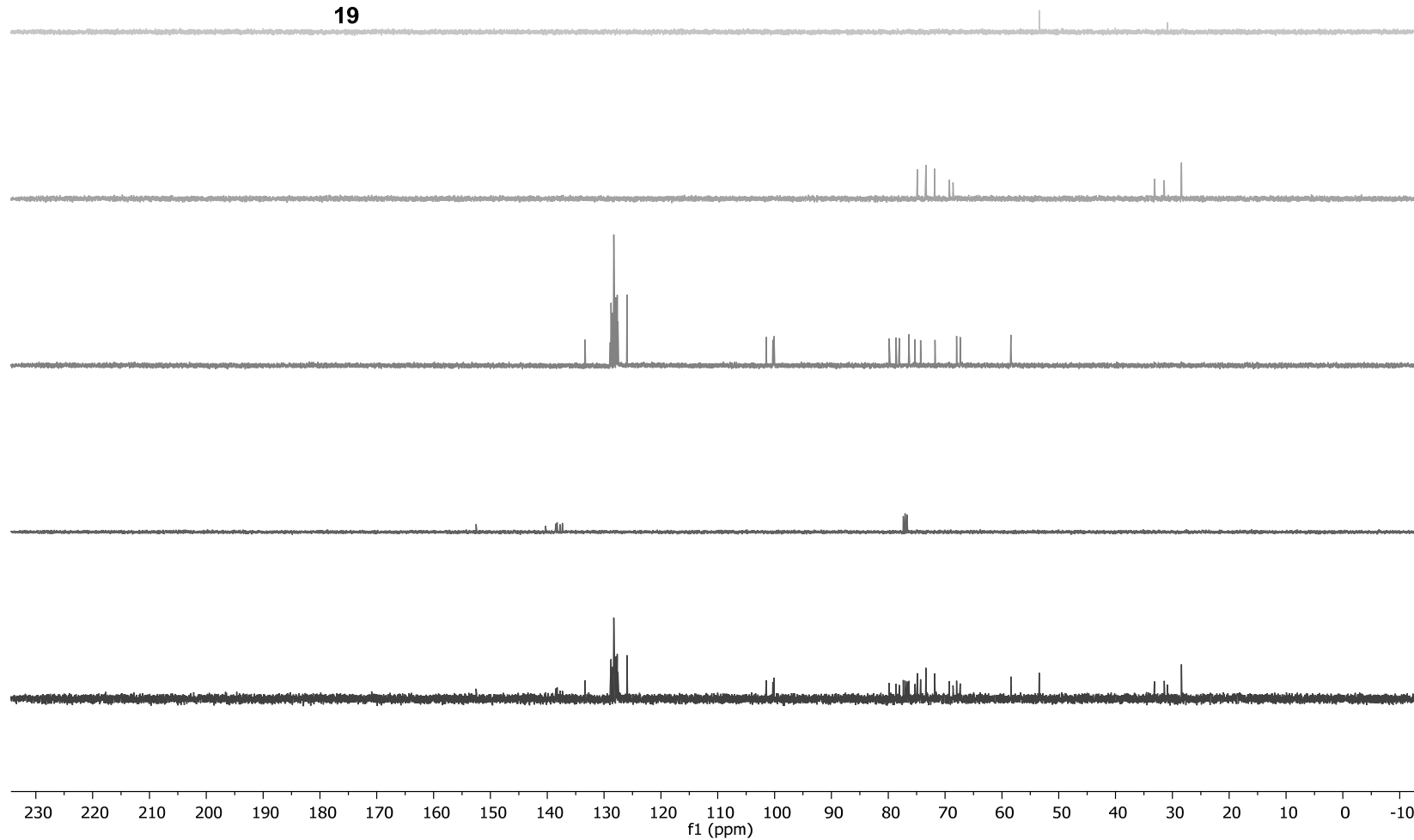
The ^{13}C NMR spectrum shows peaks from 31 to 153 ppm. Key peaks are labeled with their chemical shifts:

- 153.26.0
- 140.96.4
- 139.14.1
- 138.97.9
- 138.93.4
- 138.41.3
- 137.99.7
- 133.99.9
- 129.55.7
- 129.39.6
- 129.12.4
- 128.93.9
- 128.90.4
- 128.87.8
- 128.79.3
- 128.54.1
- 128.49.7
- 128.47.6
- 128.32.3
- 128.30.5
- 128.18.6
- 128.15.9
- 126.57.2
- 101.94.9
- 100.79.6
- 100.57.6
- 80.21.1
- 79.00.6
- 78.42.1
- 76.68.7
- 75.61.4
- 75.25.5
- 75.22.7
- 74.64.2
- 73.71.4
- 72.17.3
- 72.08.7
- 69.57.4
- 68.91.3
- 68.27.8
- 67.64.3
- 58.67.6
- 53.67.2
- 33.30.2
- 31.56.1
- 31.02.1

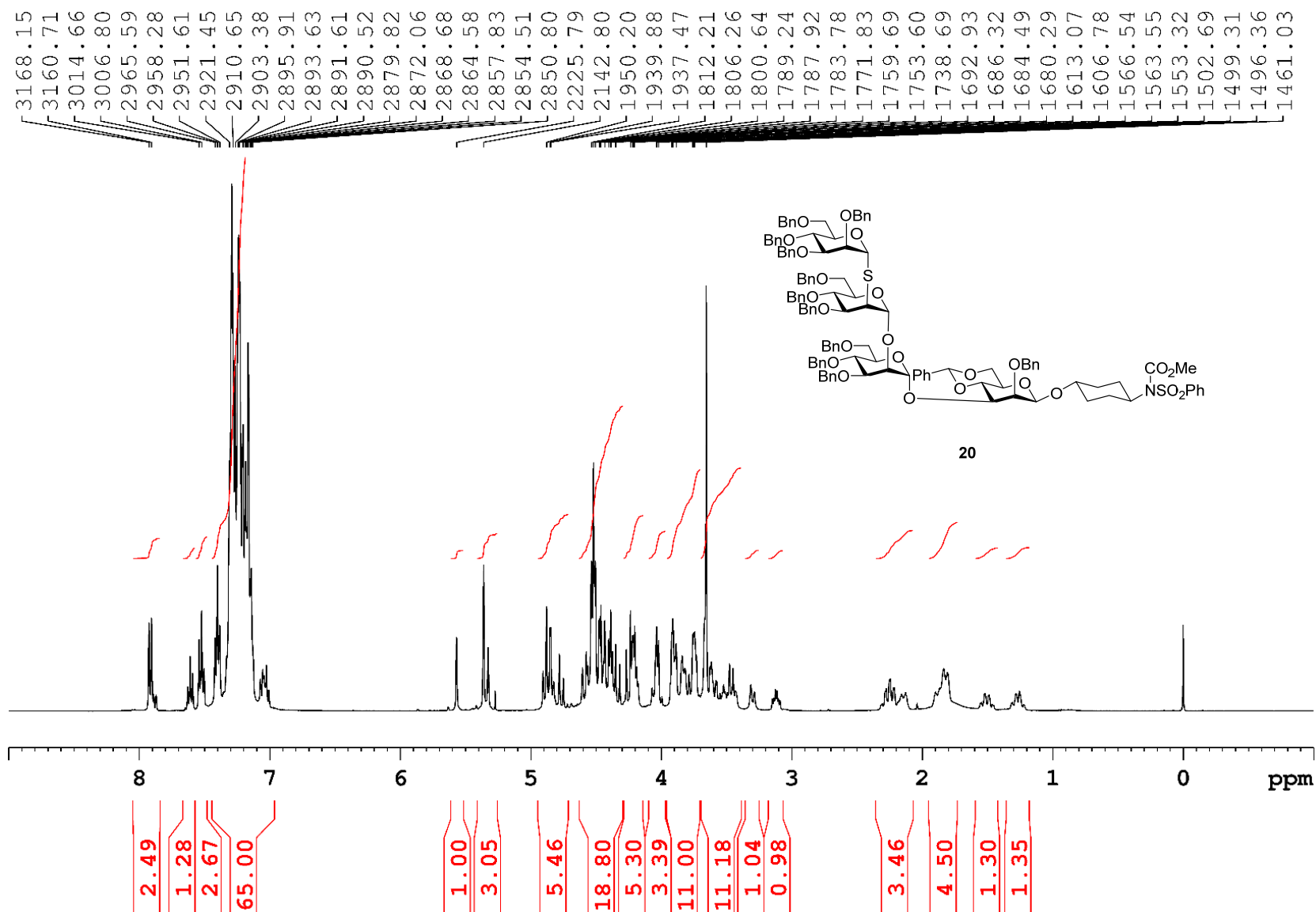
DEPT NMR (100 MHz) of **19** in CDCl₃



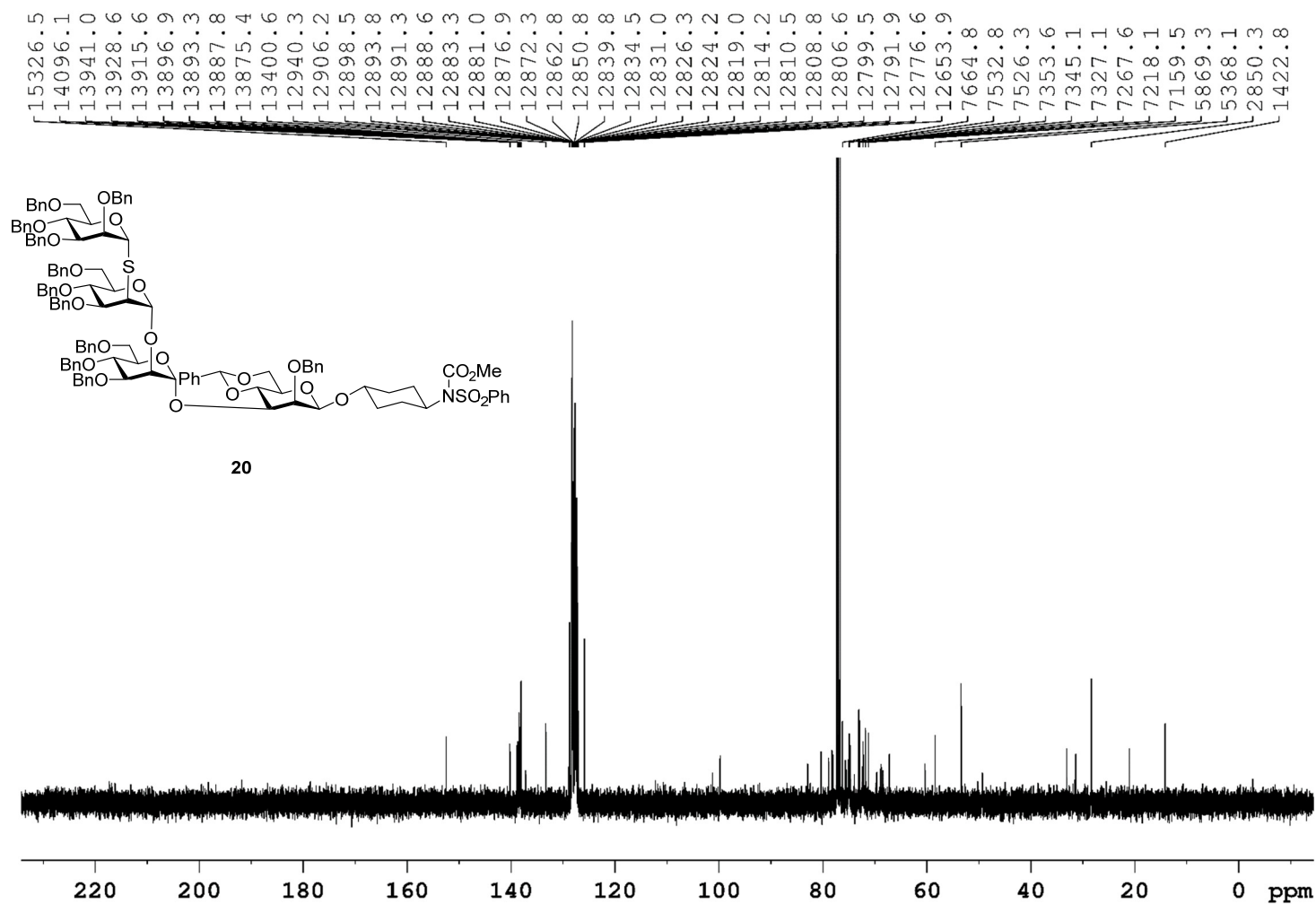
19



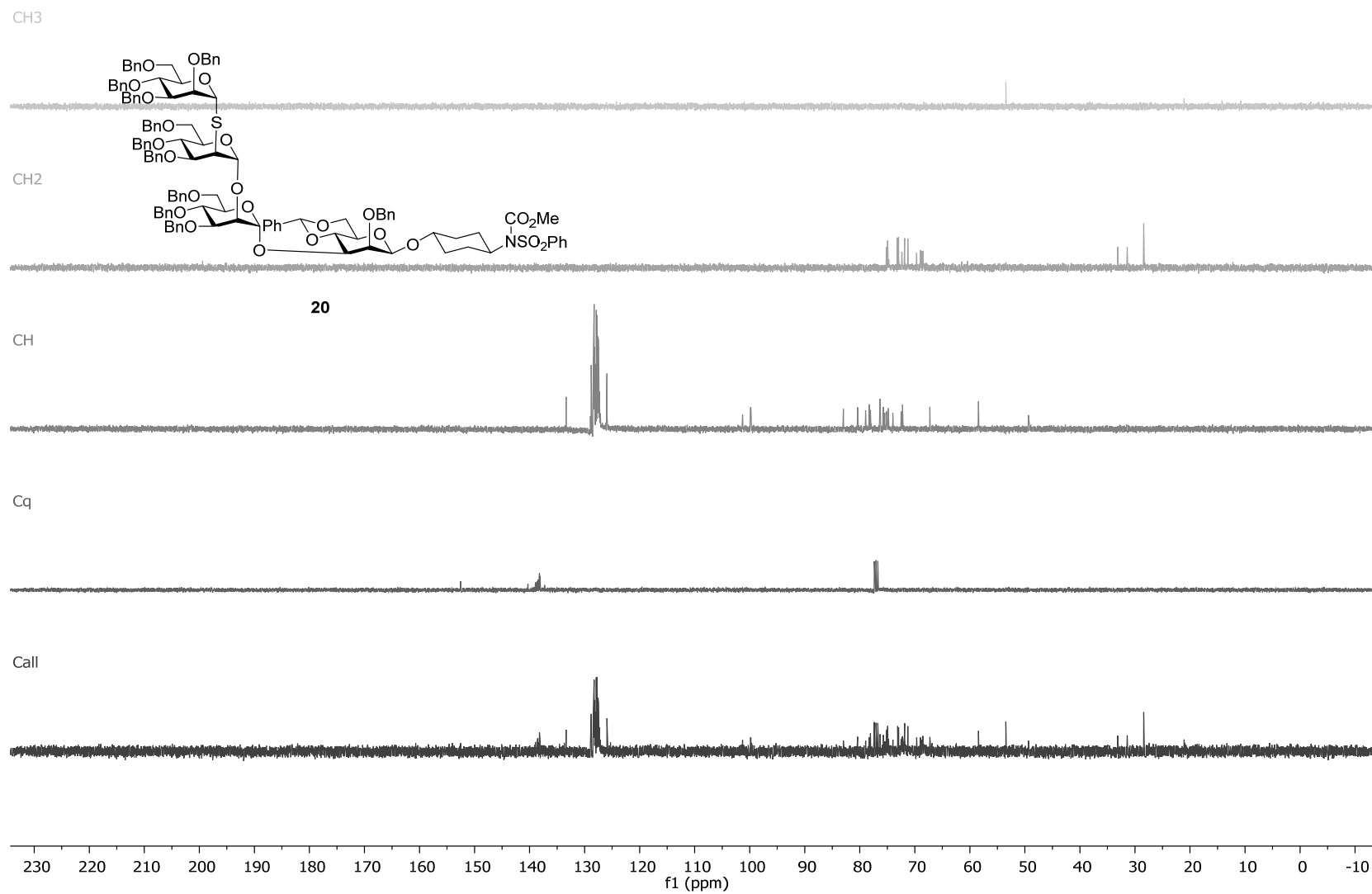
¹H NMR (400 MHz) of **20** in CDCl₃



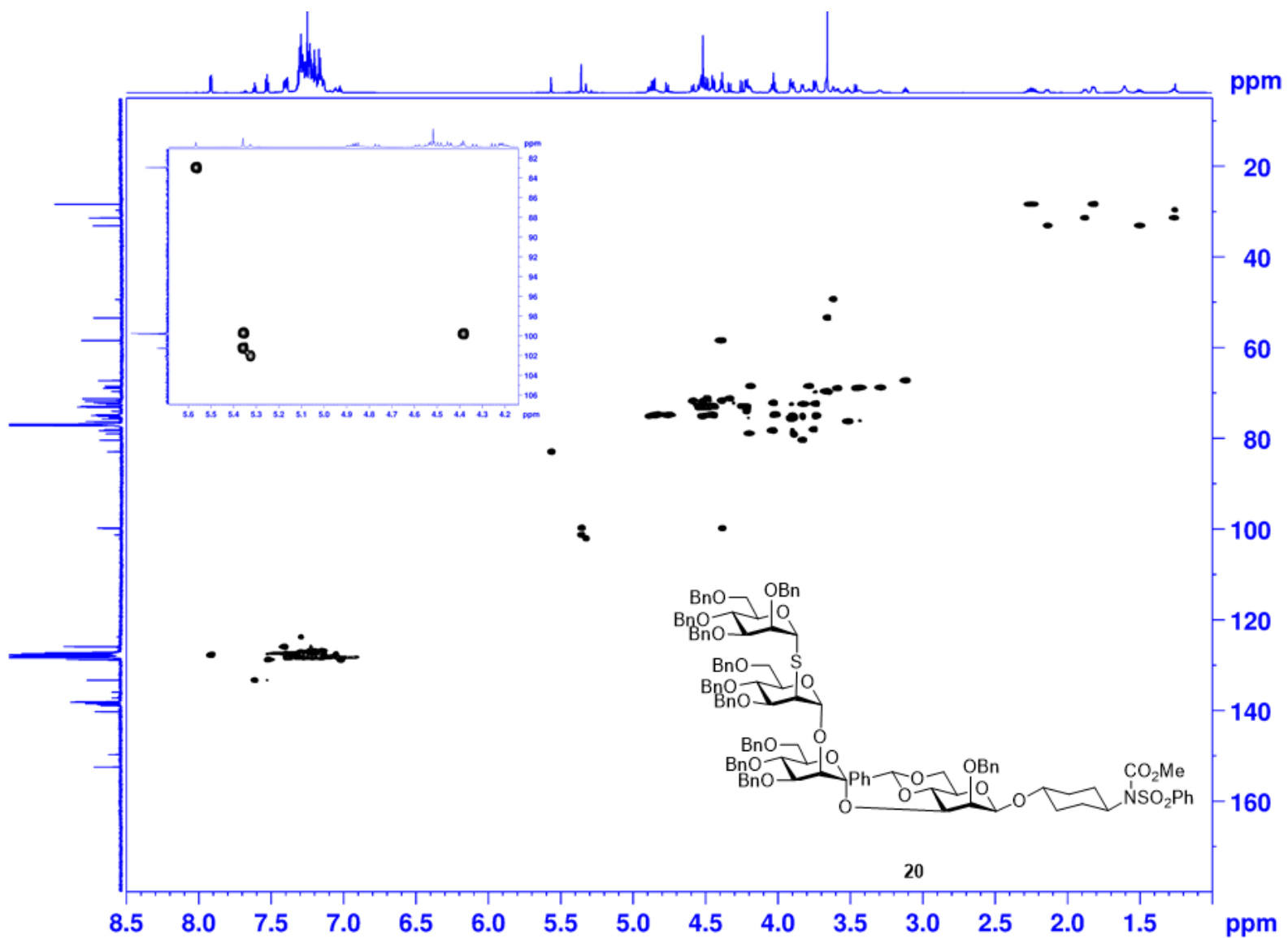
^{13}C NMR (100 MHz) of **20** in CDCl_3



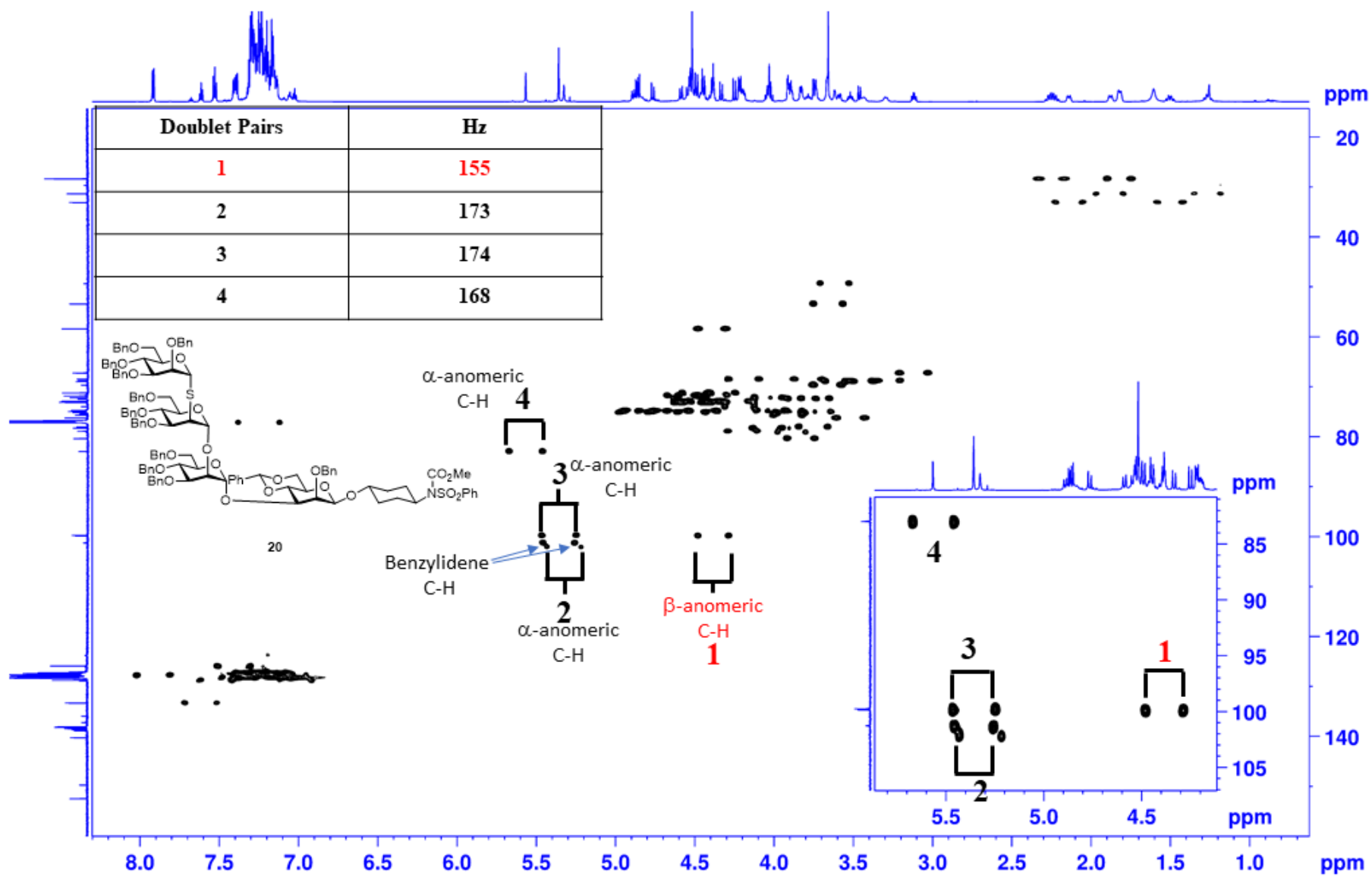
DEPT NMR (100 MHz) of **20** in CDCl₃



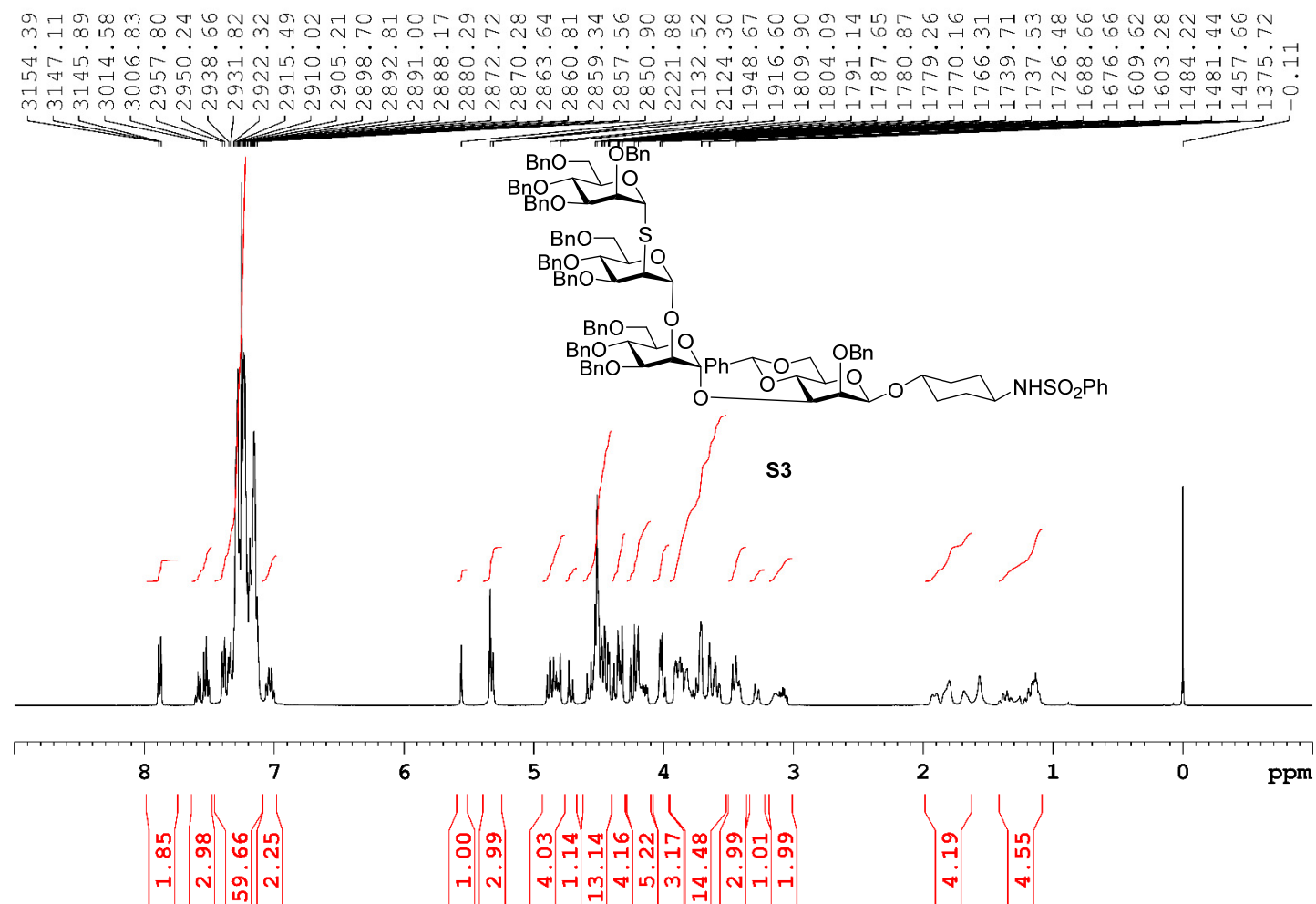
^1H - ^{13}C HSQC NMR (800 MHz) of **20** in CDCl_3



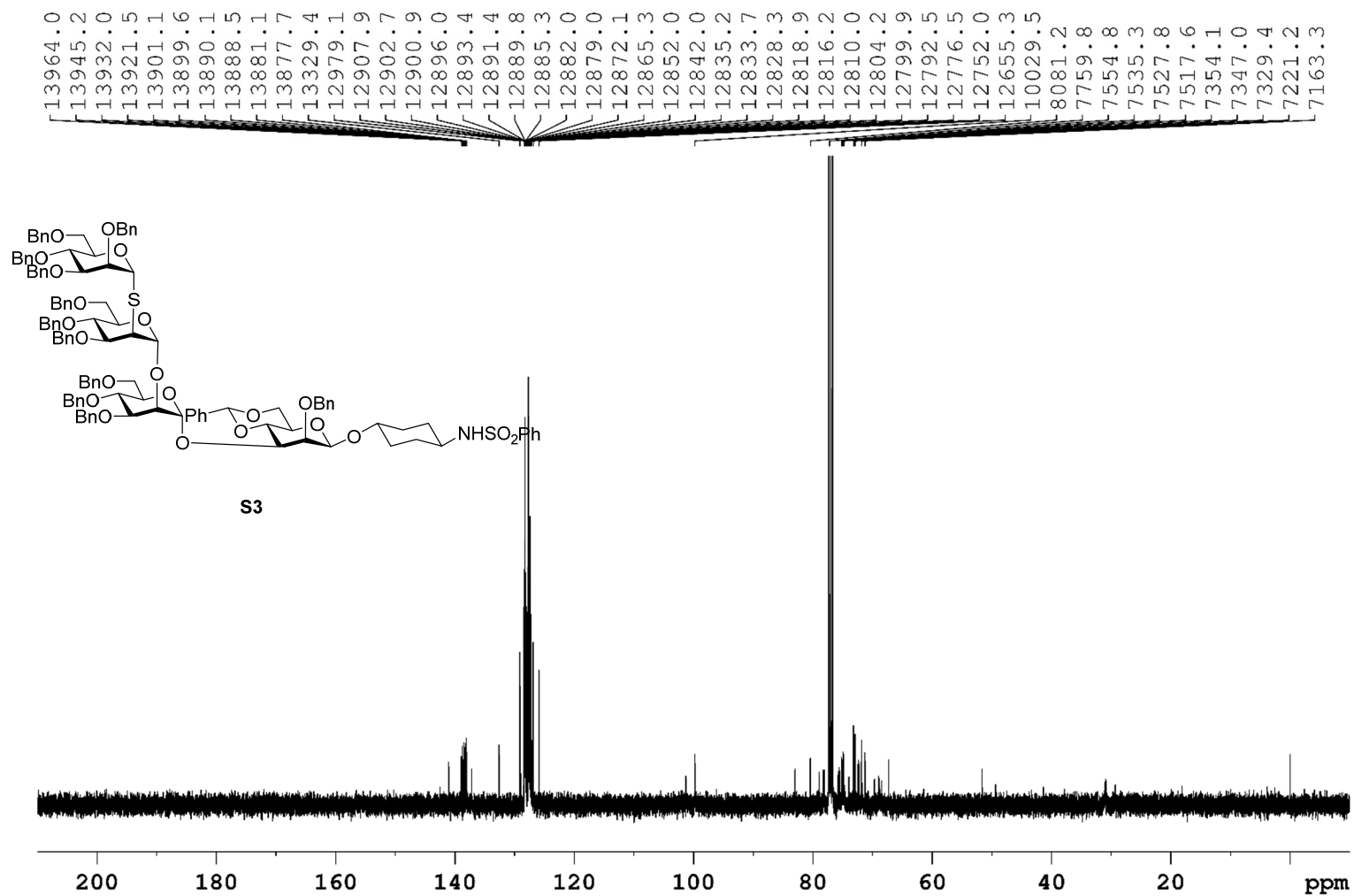
^1H - ^{13}C coupled HSQC NMR (800 MHz) of **20** in CDCl_3



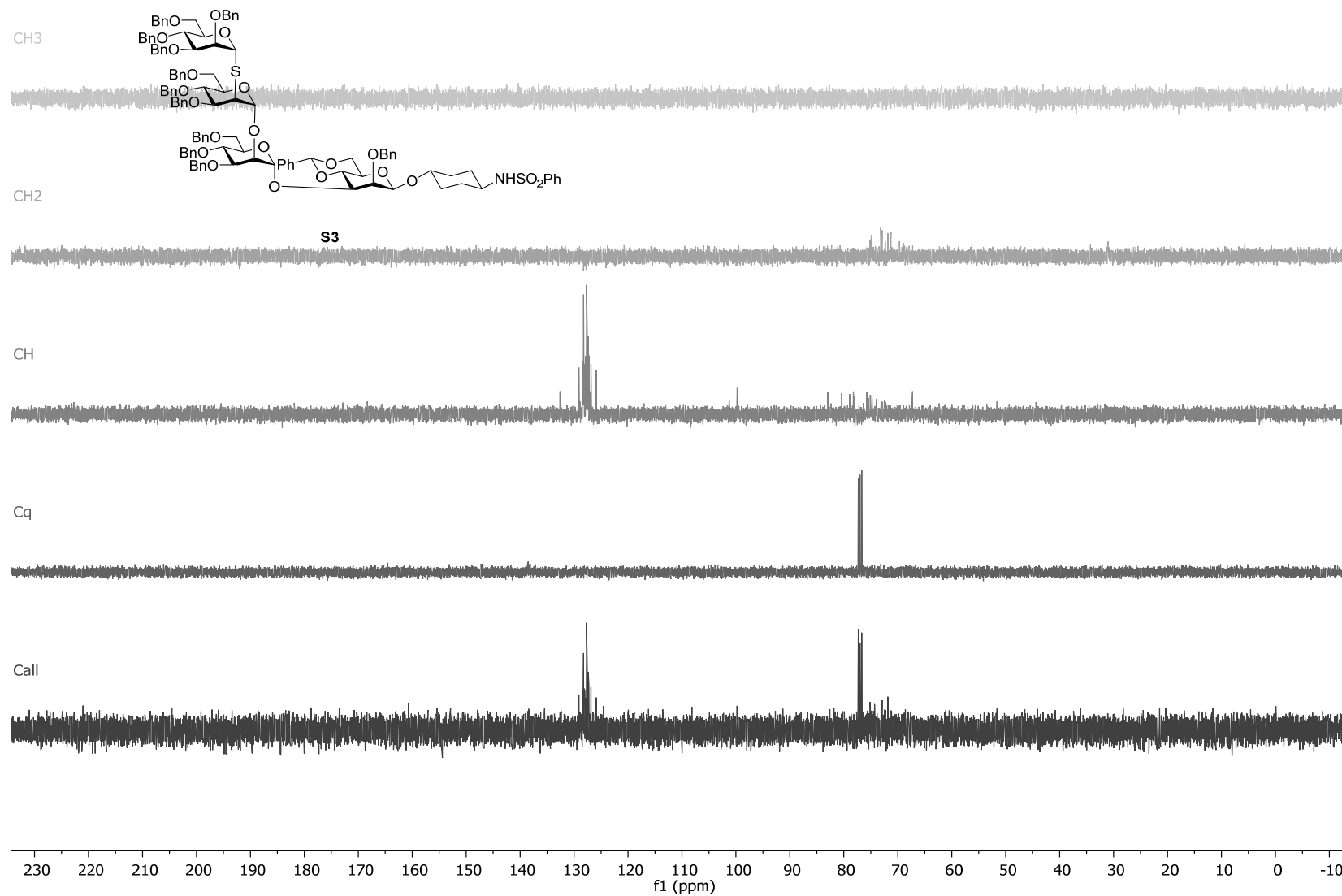
^1H NMR (400 MHz) of **S3** in CDCl_3



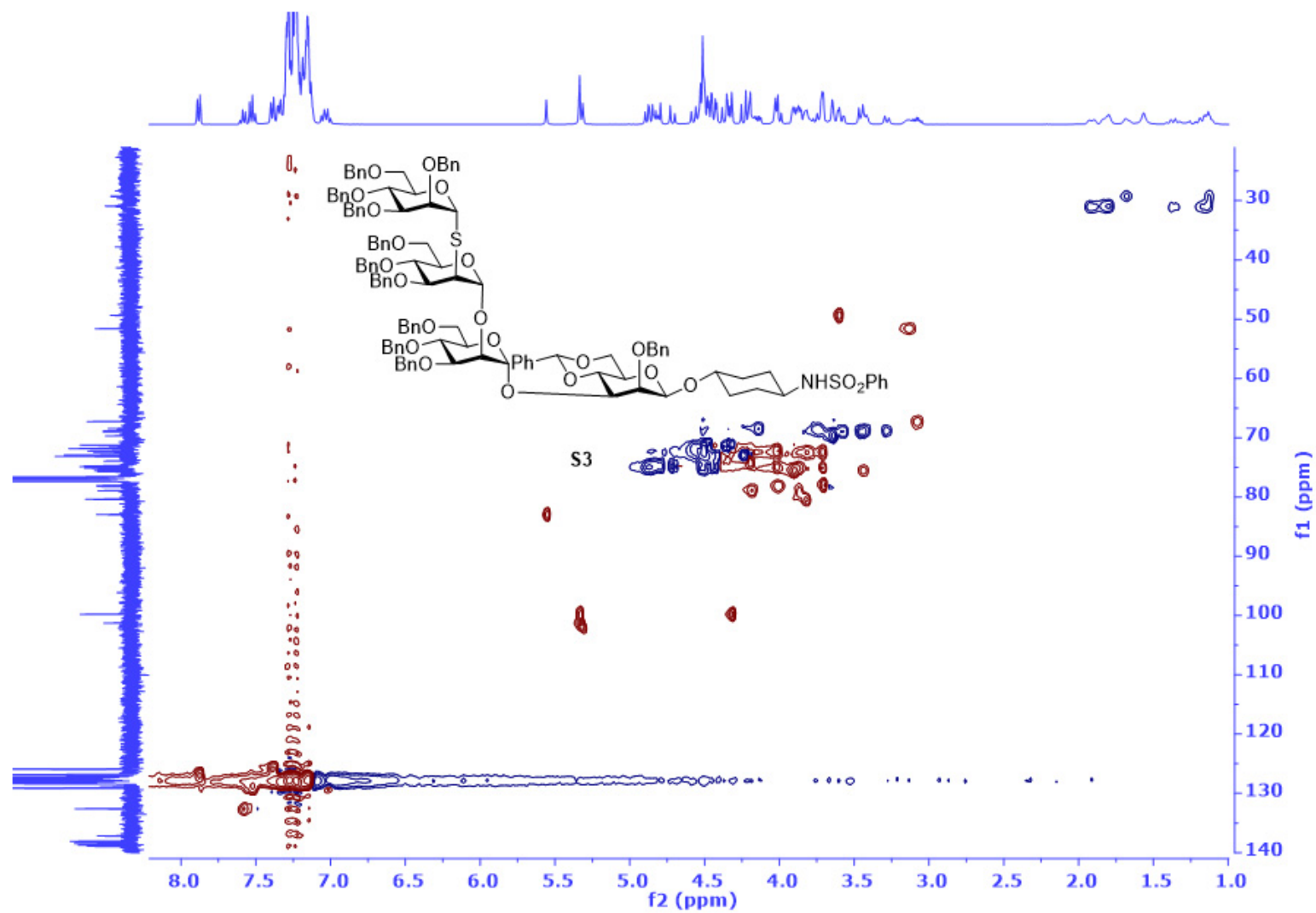
¹³C NMR (100 MHz) of **S3** in CDCl₃



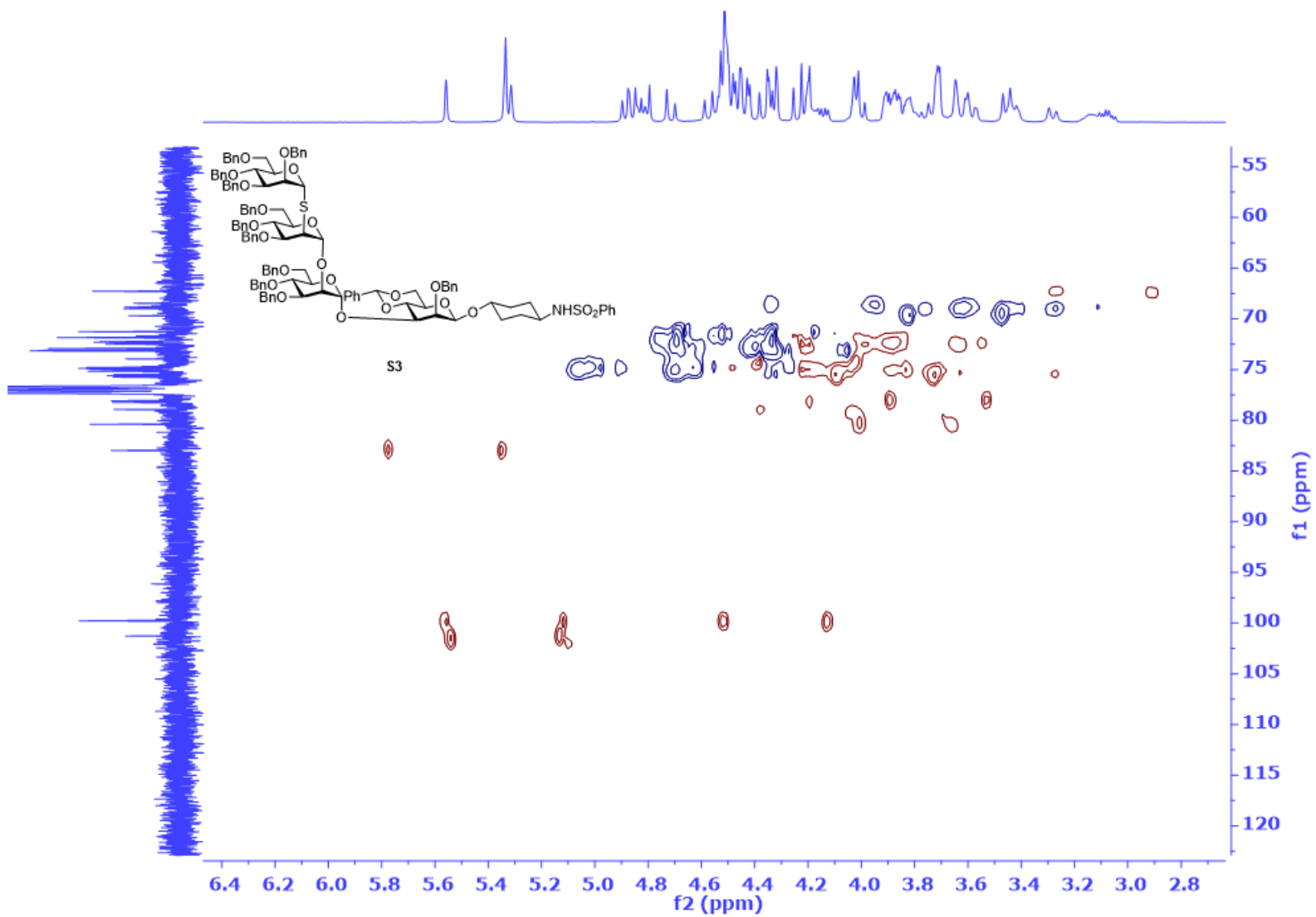
DEPT NMR (100 MHz) of **S3** in CDCl₃



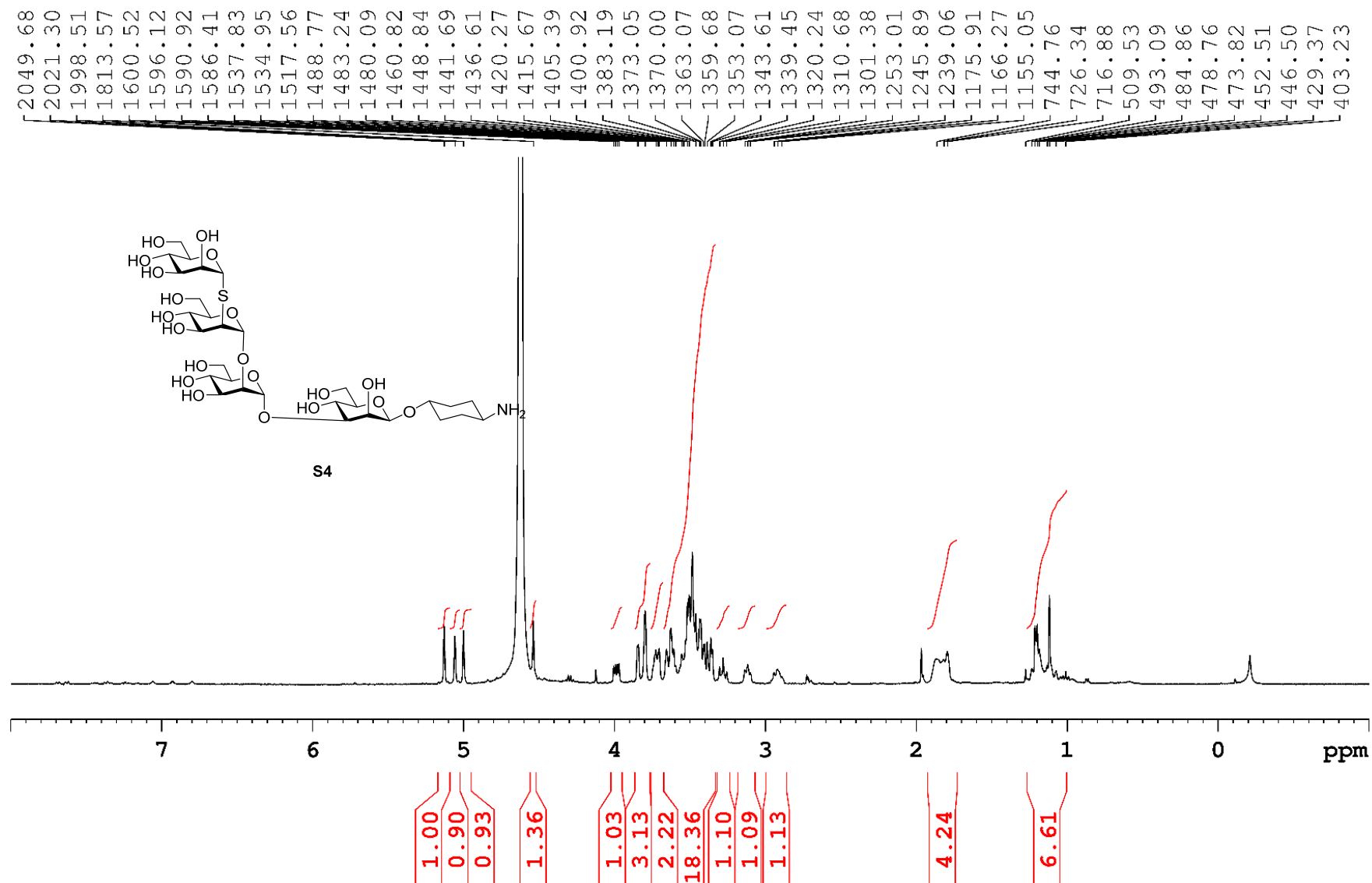
^1H - ^{13}C HSQC NMR (400 MHz) of **S3** in CDCl_3



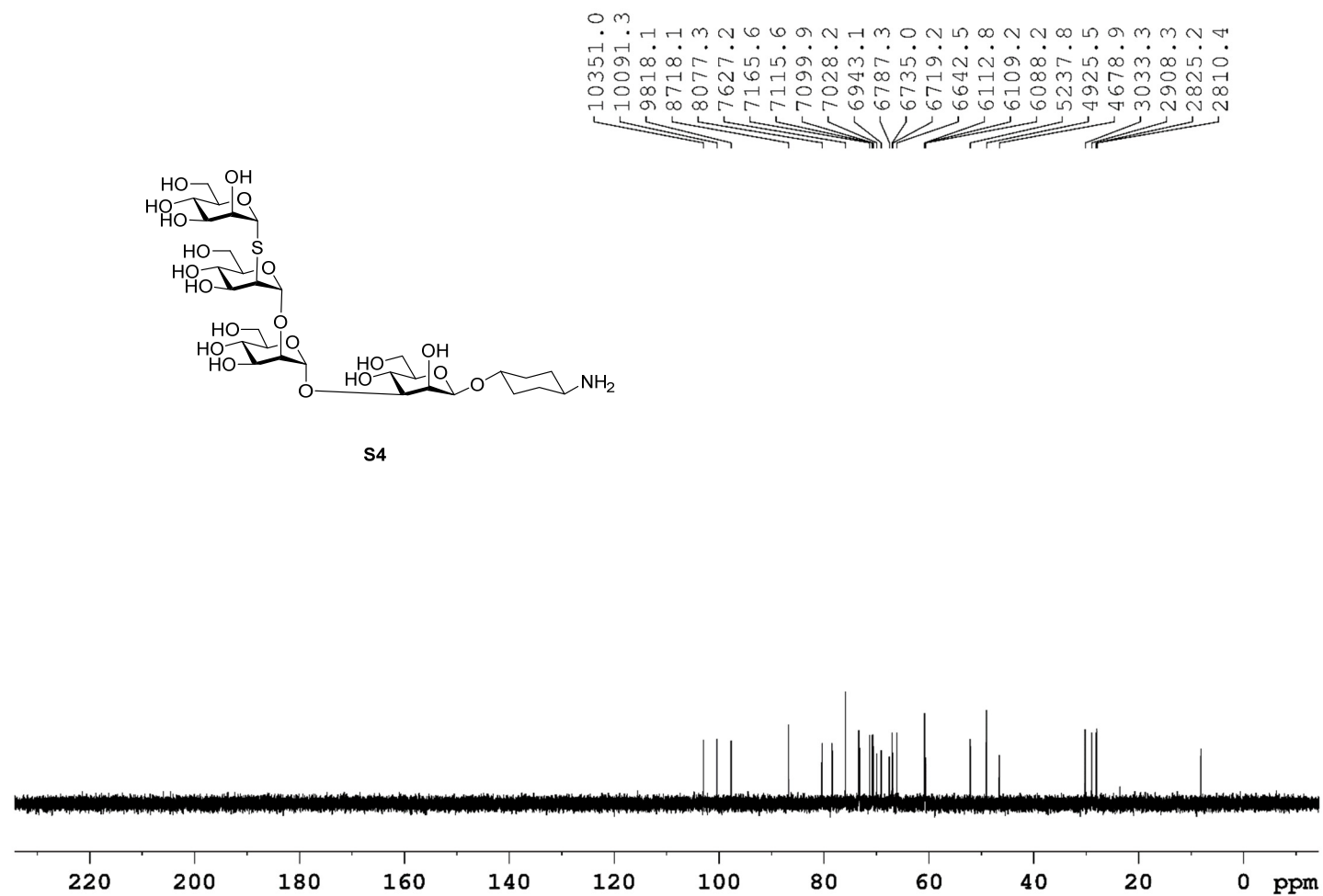
^1H - ^{13}C coupled HSQC NMR (400 MHz) of **S3** in CDCl_3



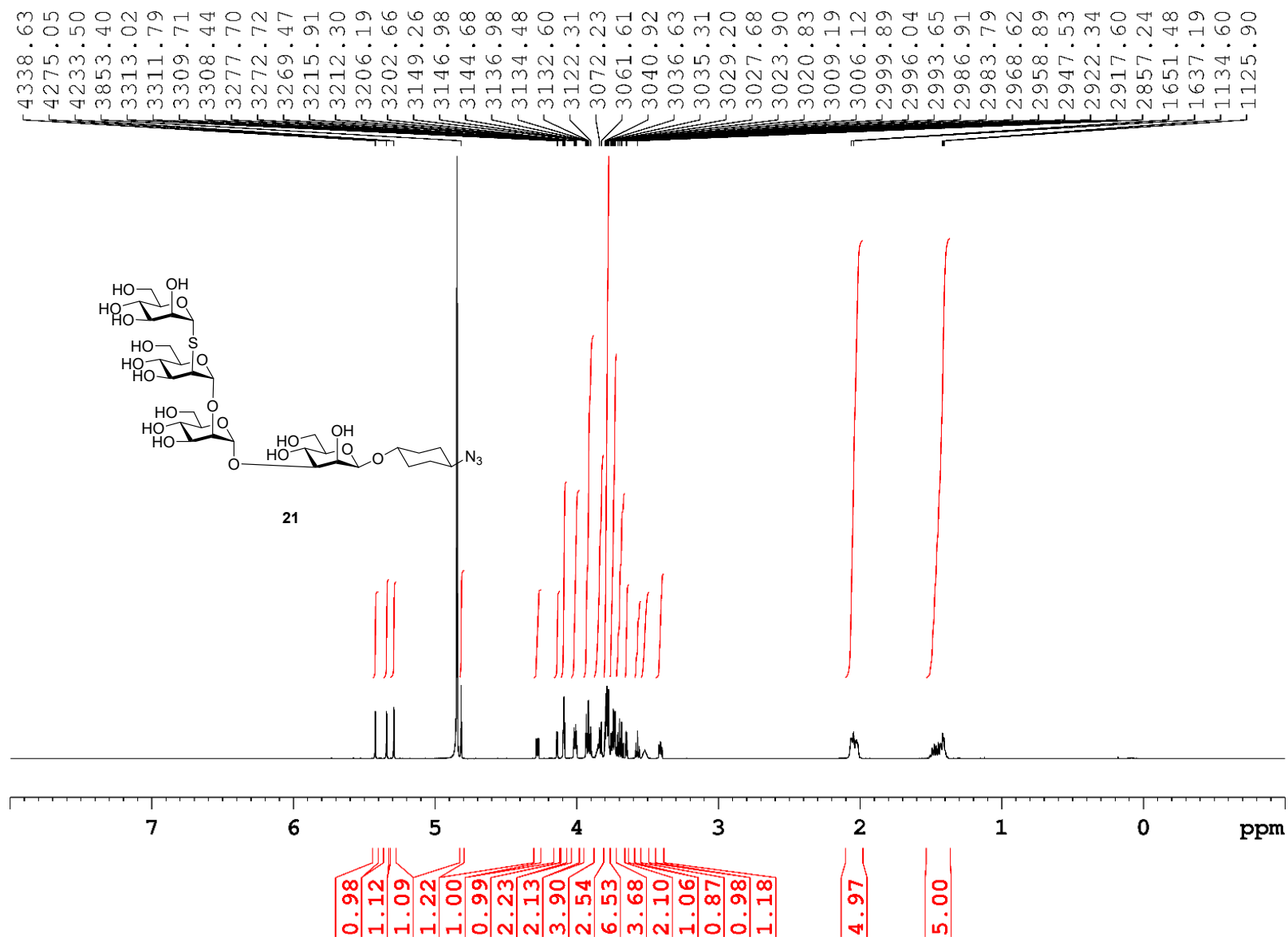
¹H NMR (400 MHz) of S4 in D₂O



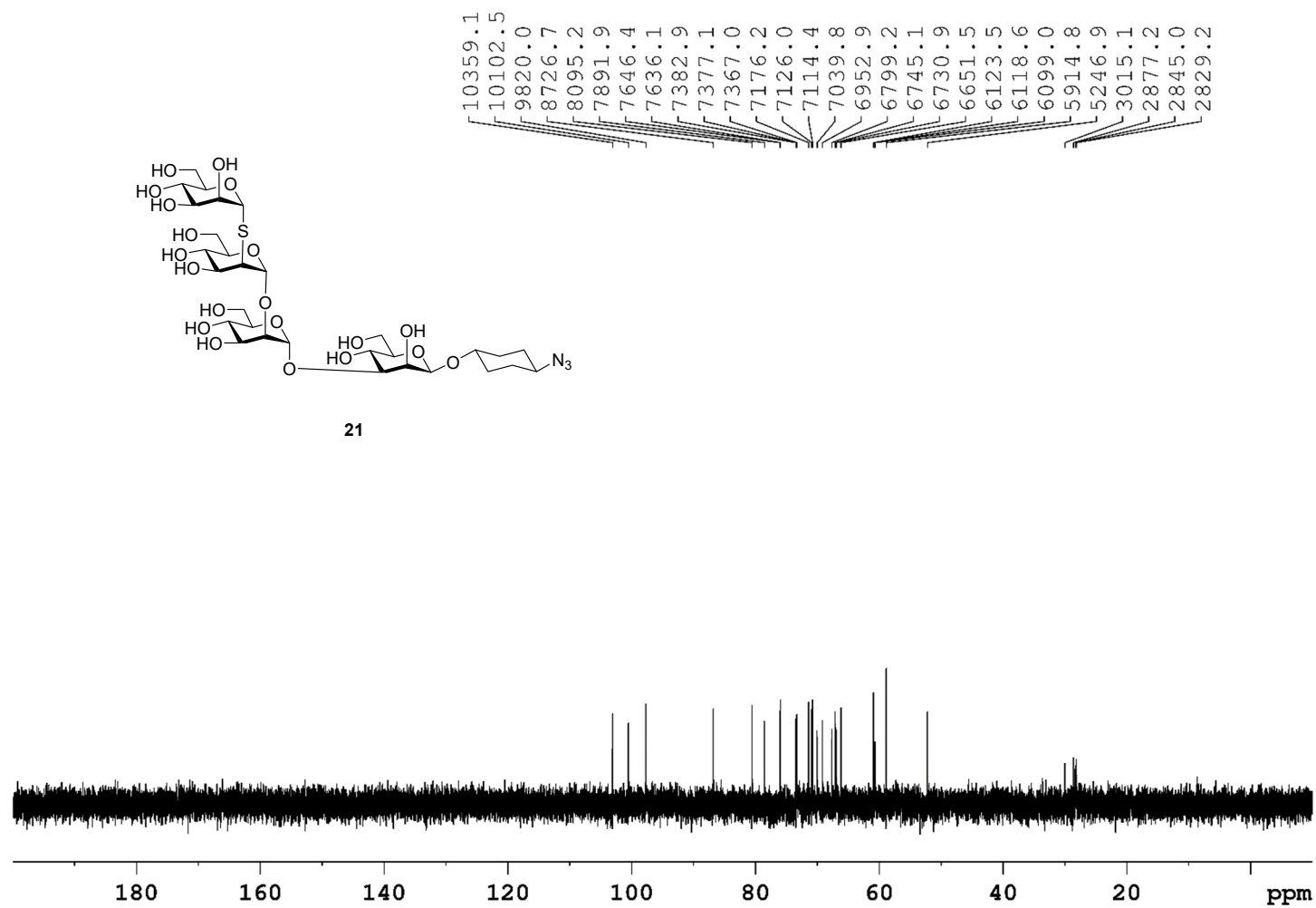
^{13}C NMR (100 MHz) of **S4** in D_2O



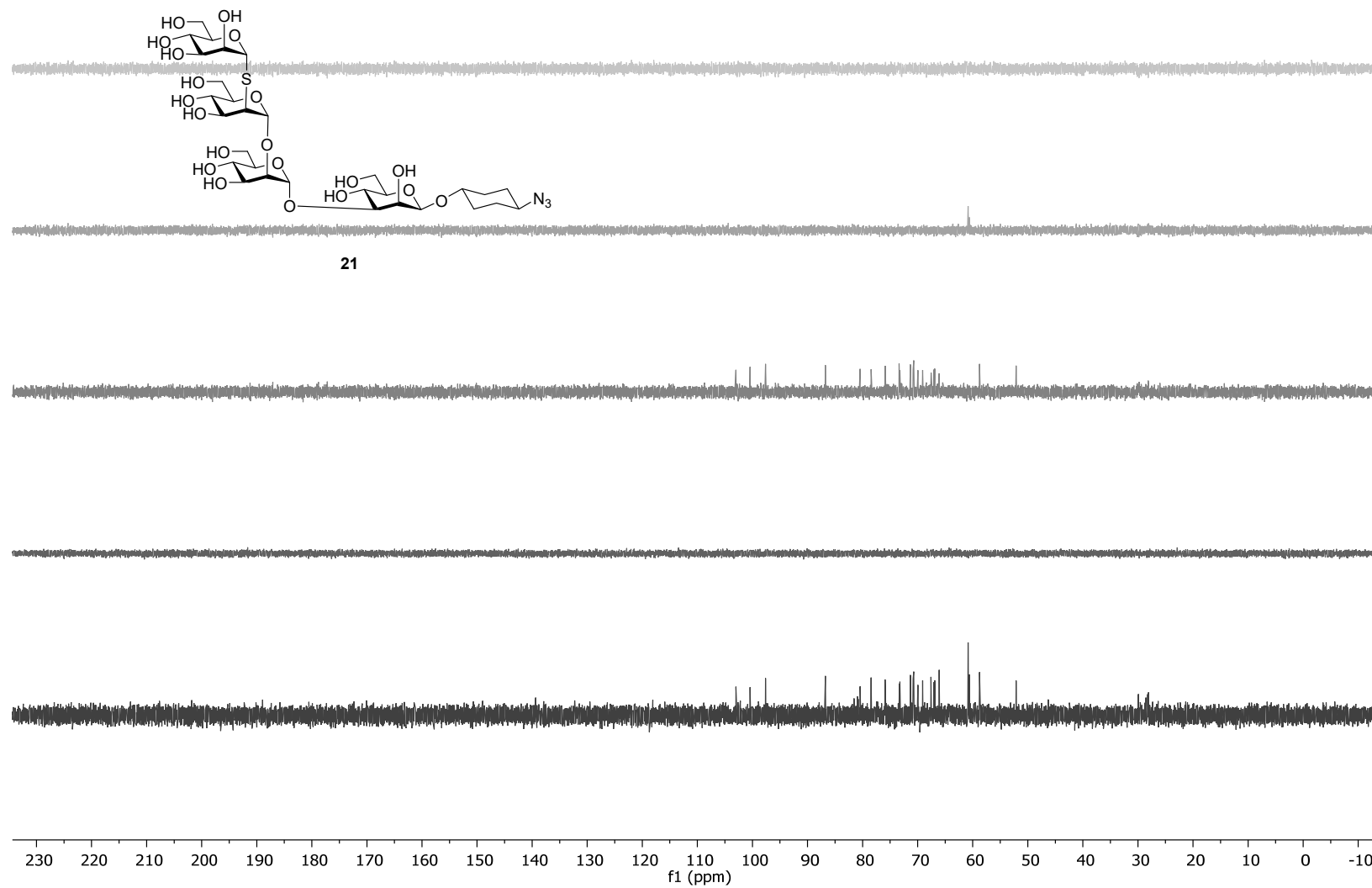
¹H NMR (800 MHz) of **21** in D₂O



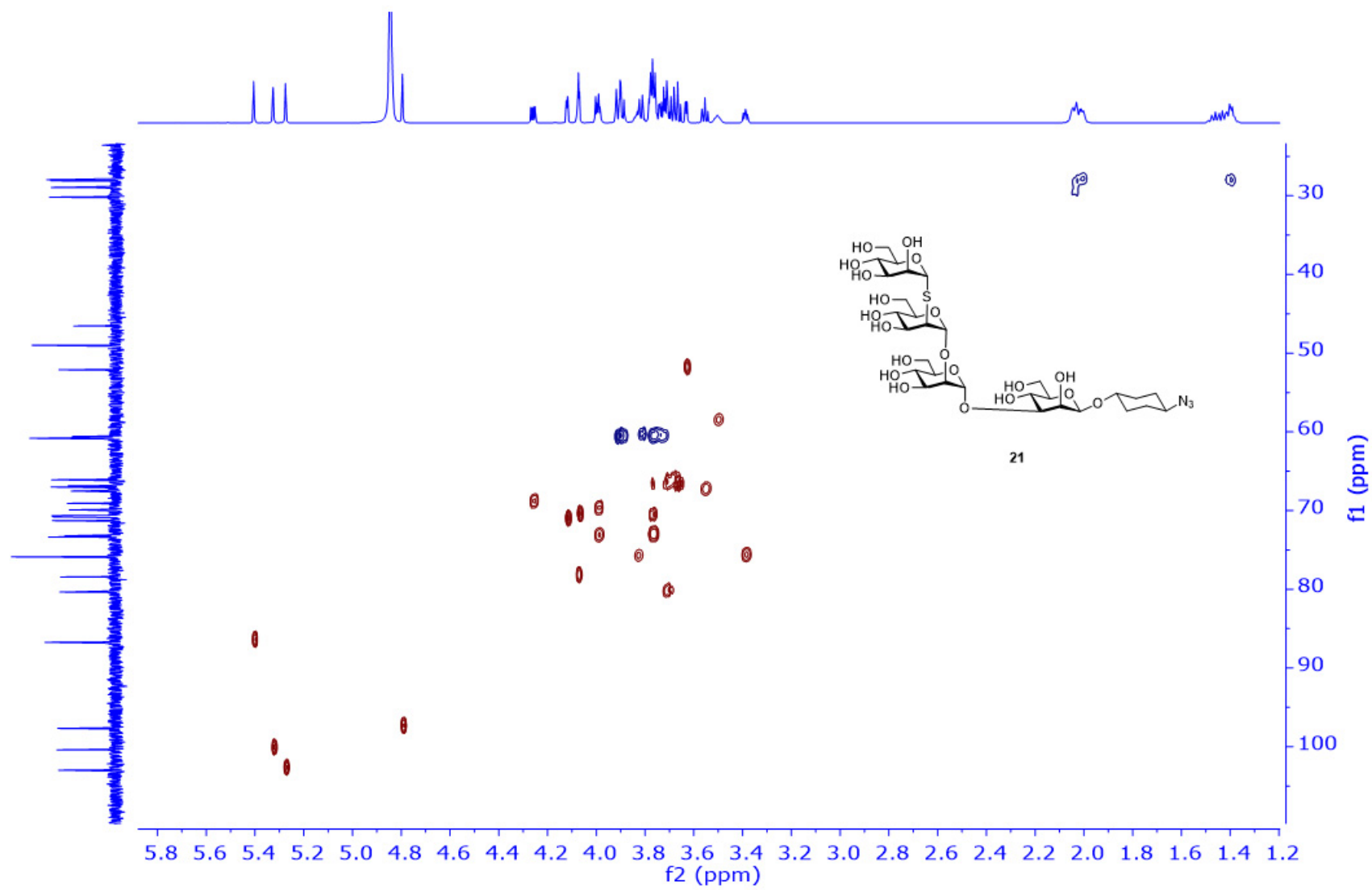
^{13}C NMR (200 MHz) of **21** in D_2O



DEPT NMR (100 MHz) of **21** in D₂O



^1H - ^{13}C HSQC NMR (800 MHz) of **21** in D_2O



^1H - ^{13}C coupled HSQC NMR (800 MHz) of **21** in D_2O

