

## Supporting Information

### Enantioselective Conjugate Addition of Catalytically Generated Zinc Homoenolate

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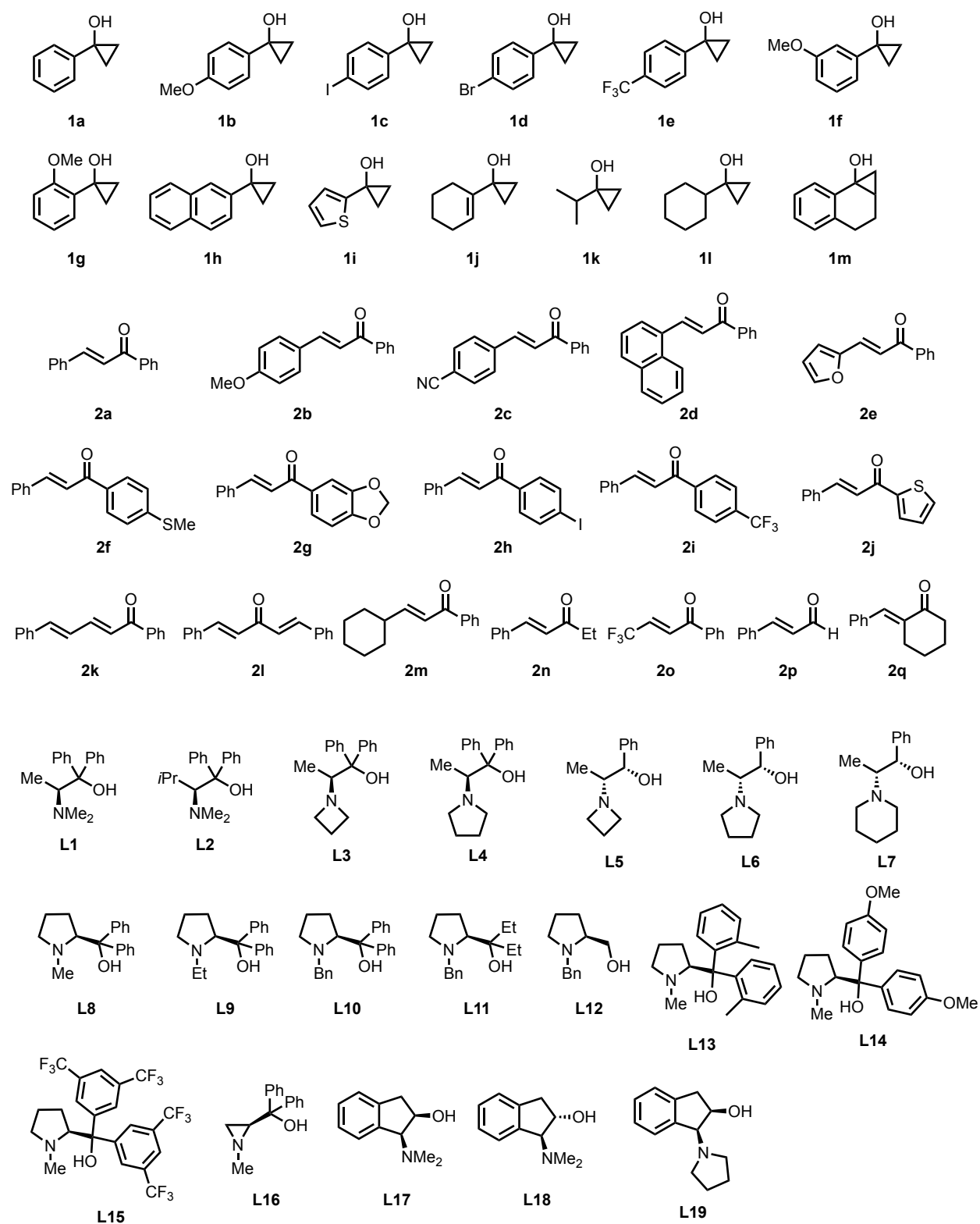
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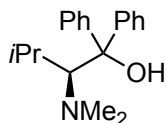
## Material and Methods

**General.** All reactions dealing with air- or moisture-sensitive compound were performed by standard Schlenk techniques in oven-dried reaction vessels under nitrogen atmosphere or in the argon-filled glove box. Analytical thin-layer chromatography (TLC) was performed on Merck 60 F254 silica gel plates. Flash chromatography was performed using 40–63  $\mu\text{m}$  silica gel (Si 60, Merck).  $^1\text{H}$ ,  $^{13}\text{C}$  and  $^{19}\text{F}$  nuclear magnetic resonance (NMR) spectra were recorded on Bruker AV-400 (400 MHz) NMR spectrometers.  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra are reported in parts per million (ppm) downfield from an internal standard, tetramethylsilane (0.00 ppm) and  $\text{CHCl}_3$  (77.0 ppm), respectively.  $^{19}\text{F}$  NMR spectra are referenced to external standard ( $\text{CF}_3\text{CO}_2\text{H}$ ,  $-76.6$  ppm). Gas chromatographic (GC) analysis was performed on a Shimadzu GC-2010 system equipped with an FID detector and a capillary column, DB-5 (Agilent J&W, 0.25 mm i.d. x 30 m, 0.25  $\mu\text{m}$  film thickness). Chiral HPLC analysis was performed on a Shimadzu LC-20AD instrument using Daicel Chiralpak columns at room temperature (25–29  $^\circ\text{C}$ ). Optical rotations were recorded on an Anton Paar MCP 150 machine. High-resolution mass spectra (HRMS) were obtained with a Q-ToF Premier LC HR mass spectrometer.

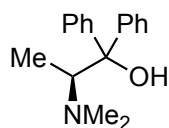
**Materials.** Unless otherwise noted, commercial reagents were purchased from Aldrich, Alfa Aesar, and other commercial suppliers and were used as received.  $\text{Et}_2\text{Zn}$  was purchased from TCI as a hexane solution (1 M) and was used as received. Molecular sieves 4 $\text{\AA}$  was purchased from Aldrich (powder, activated, -325 mesh particle size) and activated under vacuum ( $<0.06$  mbar) at 265  $^\circ\text{C}$  for more than 10 h, every time before the reaction. DMPU was distilled over  $\text{CaH}_2$  and stored under Ar. Anhydrous DMSO (Aldrich) were used without further purification and stored under Ar. Figure S1 summarizes the substrates and the ligands used in this study. The substrates **1a**,<sup>1</sup> **1b**,<sup>1</sup> **1c**,<sup>2</sup> **1d**,<sup>1</sup> **1e**,<sup>3</sup> **1f**,<sup>1</sup> **1g**,<sup>3</sup> **1h**,<sup>3</sup> **1i**,<sup>4</sup> **1j**,<sup>5</sup> **1k**,<sup>6</sup> **1l**,<sup>1</sup> **1m**,<sup>7</sup> **2b**,<sup>8</sup> **2c**,<sup>8</sup> **2d**,<sup>9</sup> **2e**,<sup>8</sup> **2f**,<sup>10</sup> **2g**,<sup>11</sup> **2h**,<sup>12</sup> **2i**,<sup>13</sup> **2j**,<sup>14</sup> **2k**,<sup>15</sup> **2m**,<sup>16</sup> **2n**,<sup>17</sup> **2o**,<sup>18</sup> and **2q**<sup>19</sup> were prepared according to the literature procedure. The chiral ligands **L5**–**L7**<sup>20</sup> were prepared from (1*S*, 2*R*)-(+)-norephedrine. (1*R*, 2*S*)-**L6** was purchased from Aldrich and used only for preparing racemic and scalemic **L6** for nonlinear effect study. Additional chiral ligands **L8**,<sup>21</sup> **L9**,<sup>22</sup> **L10**,<sup>21</sup> **L11**,<sup>23</sup> **L12**,<sup>24</sup> **L13**,<sup>25</sup> **L14**,<sup>21</sup> **L15**,<sup>21</sup> **L16**,<sup>26</sup> **L17**,<sup>27</sup> **L18**,<sup>28</sup> and **L19**<sup>27</sup> were prepared according to the literature procedure and used during the optimization study.



**Figure S1.** Substrates and ligands used in this study.

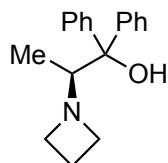


**(S)-2-(Dimethylamino)-3-methyl-1,1-diphenylbutan-1-ol (L2):**<sup>29</sup> Prepared according to the literature procedure.<sup>30</sup> A 25-mL round bottom flask equipped with a magnetic stir bar was charged with (S)-2-amino-3-methyl-1,1-diphenylbutan-1-ol<sup>31</sup> (255 mg, 1.00 mmol), paraformaldehyde (300 mg, 10.0 mmol), methanol (6 mL), and acetic acid (0.1 mL), followed by the addition of NaBH<sub>3</sub>CN (189 mg, 3.00 mmol). The resultant black suspension was stirred at 23 °C for 48 h and then filtered through a pad of Celite. The organic solution was concentrated under reduced pressure, and to the residue were added Et<sub>2</sub>O (5 mL) and H<sub>2</sub>O (5 mL). After the aqueous phase was acidified (ca. pH 2) by adding 1 M hydrochloric acid, the layers were separated. The aqueous layer was washed with Et<sub>2</sub>O (3 mL), and then 1 M NaOH aq. was added until brown precipitates formed. The aqueous solution was then extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 × 2 mL). The organic solution was concentrated under reduced pressure, and the residue was purified by flash chromatography to afford the product as a pale yellow solid (222 mg, 78%); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.66-7.64 (m, 2H), 7.41-7.37 (m, 4H), 7.33-7.29 (m, 1H), 7.26-7.17 (m, 3H), 6.10 (brs, 1H), 3.04 (d, *J* = 10.4 Hz, 1H), 2.25-2.18 (m, 7H), 0.97 (d, *J* = 6.4 Hz, 3H), 0.61 (d, *J* = 6.6 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 146.2, 142.8, 128.3 127.7, 127.7, 127.6, 126.9, 126.8, 80.1, 78.6, 43.2, 29.3, 23.4, 23.2; HRMS (ESI) Calcd for C<sub>19</sub>H<sub>26</sub>NO [M + H]<sup>+</sup> 284.2014, found 284.2006; [α]<sup>25</sup><sub>D</sub> = +129 (*c* = 1.94 in CHCl<sub>3</sub>).

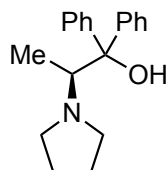


**(S)-2-(Dimethylamino)-1,1-diphenylpropan-1-ol (L1):** Prepared from (S)-2-amino-1,1-diphenylpropan-1-ol<sup>31</sup> (227 mg 1.0 mmol), according to the synthetic procedure for L2. Pale yellow oil (250 mg, 98%); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.44-7.42 (m, 4H), 7.30-7.26 (m, 4H), 7.24-7.18 (m, 2H), 6.10 (brs, 1H), 3.52 (q, *J* = 7.1 Hz, 1H), 2.10 (s, 6H), 1.09 (d, *J* = 7.1 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 146.5, 144.6, 127.9, 127.9, 127.2, 126.9, 126.4, 78.0, 65.7, 42.9, 9.2; HRMS (ESI) Calcd for C<sub>17</sub>H<sub>22</sub>NO [M + H]<sup>+</sup> 256.1701, found 256.1694; [α]<sup>25</sup><sub>D</sub> = -40.9 (*c* = 2.25 in CHCl<sub>3</sub>).

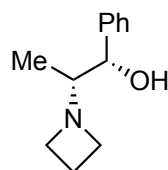




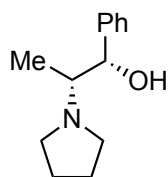
**(S)-2-(Azetidin-1-yl)-1,1-diphenylpropan-1-ol (L3):** Prepared according to the literature procedure.<sup>20</sup> A 25-mL round bottom flask equipped with a magnetic stir bar was charged with (*S*)-2-amino-1,1-diphenylpropan-1-ol<sup>31</sup> (1.00 g, 4.40 mmol), NaHCO<sub>3</sub> (0.81 g, 9.68 mmol), and toluene (12 mL), followed by the addition of 1,3-dibromopropane (0.98 g, 4.84 mmol). After 48 h of reflux, the mixture was filtered through a pad of Celite. The organic solution was concentrated under reduced pressure, and the residue was purified by flash chromatography to afford the product as a white solid (859 mg, 73%); m.p. 63-65 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.71-7.69 (m, 2H), 7.54-7.52 (m, 2H), 7.26-7.22 (m, 4H), 7.13-7.09 (m, 2H), 4.81 (brs, 1H), 3.54 (q, *J* = 6.4 Hz, 1H), 2.95 (q, *J* = 6.7 Hz, 2H), 2.75 (q, *J* = 6.7 Hz, 2H), 1.74 (quint, *J* = 6.9 Hz, 2H), 0.82 (d, *J* = 6.4 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 148.1, 145.8, 127.9, 127.7, 126.3, 126.0, 125.7, 125.3, 77.1, 67.5, 54.9, 17.6, 12.3; HRMS (ESI) Calcd for C<sub>18</sub>H<sub>22</sub>NO [M + H]<sup>+</sup> 268.1701, found 268.1692; [α]<sub>D</sub><sup>25</sup> = +12.5 (*c* = 1.69 in CHCl<sub>3</sub>).



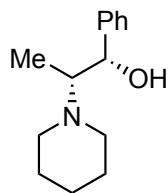
**(S)-1,1-Diphenyl-2-(pyrrolidin-1-yl)propan-1-ol (L4):** Prepared from (*S*)-2-amino-1,1-diphenylpropan-1-ol<sup>31</sup> (227 mg 1.0 mmol) and 1,4-dibromobutane (237 mg, 1.1 mmol), according to the synthetic procedure for L3. Pale yellow oil (142 mg, 50%); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.57-7.55 (m, 2H), 7.48-7.46 (m, 2H), 7.29-7.24 (m, 4H), 7.20-7.15 (m, 2H), 3.74 (q, *J* = 6.9 Hz, 1H), 2.42-2.39 (m, 4H), 1.62-1.59 (m, 4H), 1.04 (d, *J* = 6.9 Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 147.7, 145.7, 127.9, 127.5, 126.9, 126.6, 126.5, 126.2, 77.9, 63.6, 51.3, 23.5, 12.7; HRMS (ESI) Calcd for C<sub>19</sub>H<sub>24</sub>NO [M + H]<sup>+</sup> 282.1858, found 282.1851; [α]<sub>D</sub><sup>25</sup> = -51.2 (*c* = 1.64 in CHCl<sub>3</sub>).



**(1*S*,2*R*)-2-(Azetidin-1-yl)-1-phenylpropan-1-ol (L5):** Prepared from (1*S*, 2*R*)-(+)-norephedrine (1.89 g 12.5 mmol) and 1,3-dibromopropane (2.78 g, 13.8 mmol), according to the literature procedure.<sup>20</sup> Colorless oil (1.57 g, 65%);<sup>i</sup> **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.33-7.31 (m, 4H), 7.26-7.20 (m, 1H), 4.71(d, *J* = 2.9 Hz, 1H), 3.41 (brs, 1H), 3.31 (q, *J* = 6.7Hz, 2H), 3.24 (q, *J* = 6.7 Hz, 2H), 2.40 (qd, *J* = 6.5, 3.0 Hz, 1H), 2.05 (quint, *J* = 7.0 Hz, 2H), 0.63 (d, *J* = 6.5 Hz, 3H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 141.3, 128.0, 126.8, 125.8, 70.9, 68.5, 53.4, 16.8, 8.9; **HRMS** (ESI) Calcd for C<sub>12</sub>H<sub>18</sub>NO [M + H]<sup>+</sup> 192.1388, found 192.1388; [α]<sup>25</sup><sub>D</sub> = -32.9 (*c* = 2.07 in CHCl<sub>3</sub>).



**(1*S*,2*R*)-1-Phenyl-2-(pyrrolidin-1-yl)propan-1-ol (L6):**<sup>32</sup> Prepared from (1*S*, 2*R*)-(+)-norephedrine (2.27 g 15.0 mmol) and 1,4-dibromobutane (3.56 g, 16.5 mmol) according to the literature procedure.<sup>20</sup> Pale orange solid (2.54 g, 82%); m.p. 44-45 °C, Lit 44-45 °C<sup>20</sup>; **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.35-7.30 (m, 4H), 7.25-7.20 (m, 1H), 5.00 (d, *J* = 3.1 Hz, 1H), 3.61 (brs, 1H), 2.81-2.76 (m, 2H), 2.66-2.61 (m, 2H), 2.48 (qd, *J* = 6.6, 3.1 Hz, 1H), 1.83-1.77 (m, 4H), 0.79 (d, *J* = 6.6 Hz, 3H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>) δ 141.7, 127.9, 126.6, 125.7, 72.6, 65.3, 51.8, 23.5, 12.0; **HRMS** (ESI) Calcd for C<sub>13</sub>H<sub>20</sub>NO [M + H]<sup>+</sup> 206.1545, found 206.1549; [α]<sup>25</sup><sub>D</sub> = -12.5 (*c* = 2.15 in CHCl<sub>3</sub>). Lit [α]<sup>24</sup><sub>D</sub> = -7.25 (*c* = 2.00 in CHCl<sub>3</sub>),<sup>32</sup> [α]<sup>20</sup><sub>D</sub> ((**1*R***, **2*S***)-**L6**) = +15.2 (*c* = 2.00 in CHCl<sub>3</sub>).<sup>20</sup>



**(1*S*,2*R*)-1-Phenyl-2-(piperidin-1-yl)propan-1-ol (L7):**<sup>33</sup> Prepared from (1*S*, 2*R*)-(+)-norephedrine (756 mg 5.0 mmol) and 1,5-diiodopentane (1.78 g, 5.5 mmol), according to the

<sup>i</sup> This compound was not stable at room temperature for long-time storage. It should be stored in a fridge, and its purity should be checked by <sup>1</sup>H NMR before use.

literature procedure.<sup>20</sup> White solid (883 mg, 81%); m.p. 108-109 °C; **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.31-7.30 (m, 4H), 7.25-7.19 (m, 1H), 4.81 (d,  $J$  = 4.2 Hz, 1H), 4.09 (brs, 1H), 2.69 (qd,  $J$  = 6.9, 4.3 Hz, 1H), 2.55-2.45 (m, 4H), 1.60-1.54 (m, 4H), 1.46-1.40 (m, 2H), 0.82 (d,  $J$  = 4.2 Hz, 3H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>)  $\delta$  142.3, 127.8, 126.6, 126.0, 72.2, 64.6, 51.7, 26.6, 24.6, 10.2; **HRMS** (ESI) Calcd for C<sub>14</sub>H<sub>22</sub>NO [M + H]<sup>+</sup> 220.1701, found 220.1705;  $[\alpha]^{25}_{\text{D}} = +0.471$  ( $c$  = 2.55 in CHCl<sub>3</sub>).

## Early-Stage Optimization of Reaction Conditions

During initial attempts to render the racemic reaction between **1a** and **2a** (eq 1 in the paper) enantioselective, the proline-derived amino alcohol **L8** was found to induce modest enantioselectivity to afford, after 3 h at 80 °C, the cyclopentene product **4aa** in 68% yield with 67:33 er (Table S1, entry 1). Upon screening of solvents (entries 2–6), we observed improvement of the enantioselectivity up to 77:23 er in solvents such as DMA, NMP, NEt<sub>3</sub>, and DMPU, albeit with diminished yields (10–40%). The reaction in DMPU at a lower temperature (60 °C) for a prolonged time (12 h) did not improve the yield and the enantioselectivity (entry 7). Efforts to improve the yield at this stage led us to find a NEt<sub>3</sub>/DMPU (6:1) mixed solvent. Though somewhat peculiar, the reaction in this mixed solvent afforded **4aa** in 50% yield with 90:10 er, which was accompanied by uncyclized **3aa** (26%, 69:31 er; entry 8). *The difference between ers of 4aa and 3aa indicates that the aldol condensation of 3aa to 4aa is promoted by the chiral zinc catalyst and thus involves kinetic resolution.* With this observation, we modified the conditions to ensure full conversion of **3aa** to **4aa** (entries 9 and 10). By performing the conjugate addition at 30 °C and the aldol condensation at 100 °C, **4aa** was obtained in 79% yield with 83:17 er (entry 10).

**Table S1.** Initial Screening Experiments using **L8** as Ligand<sup>a</sup>

| entry | solvent                      | temp/time                    | yield(%) <sup>b</sup> | er                 |
|-------|------------------------------|------------------------------|-----------------------|--------------------|
| 1     | DMSO                         | 80 °C/3 h                    | 68                    | 67:33              |
| 2     | DMF                          | 80 °C/3 h                    | 12                    | 65:35              |
| 3     | DMA                          | 80 °C/3 h                    | 10                    | 74:26              |
| 4     | NMP                          | 80 °C/3 h                    | 26                    | 75:25              |
| 5     | NEt <sub>3</sub>             | 80 °C/3 h                    | 16                    | 74:26              |
| 6     | DMPU                         | 80 °C/3 h                    | 40                    | 77:23              |
| 7     | DMPU                         | 60 °C/12 h                   | 33                    | 79:21              |
| 8     | NEt <sub>3</sub> /DMPU (6:1) | 60 °C/12 h                   | 50                    | 90:10 <sup>c</sup> |
| 9     | NEt <sub>3</sub> /DMPU (6:1) | 60 °C/12 h, then 100 °C/12 h | 75                    | 81:19              |
| 10    | NEt <sub>3</sub> /DMPU (6:1) | 30 °C/24 h, then 100 °C/12 h | 79                    | 83:17              |

<sup>a</sup>The reaction was performed using 0.15 mmol of **1a** and 0.1 mmol of **2a** (0.33 M). <sup>b</sup>Determined by GC using mesitylene as an internal standard. <sup>c</sup>**3aa** was obtained in 26% yield with 69:31 er.

Given the best result using **L8** in Table S1, entry 10, we screened a series of chiral  $\beta$ -amino alcohol ligands using the  $\text{NEt}_3$ /DMPU solvent system (Table S2). Among the proline-derived ligands **L9–L15**, only **L14** bearing *p*-methoxyphenyl groups displayed reaction efficiency and enantioselectivity comparable to that of **L8** (entries 2–8). The aziridine-containing ligand **L16** also gave a comparable result as **L8** (entry 9). The aminoindanol-type ligands **L17–L19** induced only modest enantioselectivities (entries 10–12). Finally, the alanine-derived ligands **L1** and **L3** were found to show similar or slightly better enantioselectivity compared with **L8** (entries 13 and 14). With this finding, we decided to revisit the reaction in more convenient single solvent (DMSO or DMPU) using **L1**, **L3** and their structurally related amino alcohols (i.e. **L2** as a valinol analogue of **L1**, **L4** as pyrrolidine analogue of **L3**, and **L5–L7** as norephedrine analogues of **L3**; Table 1). As a result, **L6** was found to not only display high enantioselectivity but also promote the reaction smoothly, in contrast to the poor performance of **L8** in DMPU (Table S1, entries 6 and 7).

**Table S2.** Screening of Amino Alcohol Ligands<sup>a</sup>

| entry | ligand     | yield(%) <sup>b</sup> | er (%) |
|-------|------------|-----------------------|--------|
| 1     | <b>L8</b>  | 79                    | 83:17  |
| 2     | <b>L9</b>  | 86                    | 71:29  |
| 3     | <b>L10</b> | 83                    | 55:45  |
| 4     | <b>L11</b> | 73                    | 53:47  |
| 5     | <b>L12</b> | 25                    | 53:47  |
| 6     | <b>L13</b> | 72                    | 54:46  |
| 7     | <b>L14</b> | 80                    | 84:16  |
| 8     | <b>L15</b> | 53                    | 61:39  |
| 9     | <b>L16</b> | 80                    | 82:18  |
| 10    | <b>L17</b> | 53                    | 64:36  |
| 11    | <b>L18</b> | 40                    | 52:48  |
| 12    | <b>L19</b> | 11                    | 58:42  |
| 13    | <b>L1</b>  | 73                    | 84:16  |
| 14    | <b>L3</b>  | 79                    | 88:12  |

**L8:** R = Me  
**L9:** R = Et

**L10:** R = Ph  
**L11:** R = Et  
**L12:** R = H

**L13:** Ar = *o*-MeC<sub>6</sub>H<sub>4</sub>  
**L14:** Ar = *p*-OMeC<sub>6</sub>H<sub>4</sub>  
**L15:** Ar = 3,5-(CF<sub>3</sub>)<sub>2</sub>C<sub>6</sub>H<sub>3</sub>

**L16**

**L17**

**L18**

**L19**

**L1**

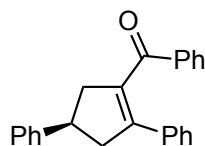
**L3**

<sup>a</sup>The reaction was performed using 0.15 mmol of **1a** and 0.1 mmol of **2a** (0.33 M).

<sup>b</sup>Determined by GC using mesitylene as an internal standard.

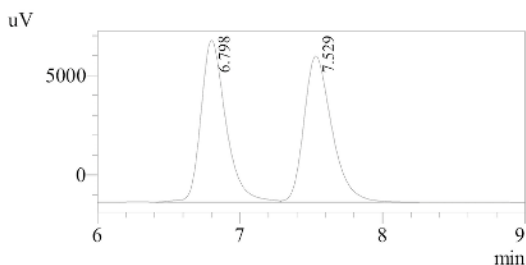
## Zn-Catalyzed Enantioselective Ring-Opening Conjugate Addition of Cyclopropanols

**General procedure:** In an argon-filled glove box, a 4-mL vial equipped with a magnetic stir bar was charged sequentially with **L6** (9.2 mg, 0.045 mmol), molecular sieves 4 Å (300 mg), and DMPU (0.9 mL, stored in -20 °C fridge). To this mixture was added Et<sub>2</sub>Zn (1 M in hexane, 45 µL, 0.045 mmol), enone **2** (0.30 mmol), and cyclopropanol **1** (0.45 mmol) sequentially, without a particular break (longer than 15 seconds) between additions. The vial was closed and removed from the glove box, and the mixture was stirred at 0 °C for 48 h. After confirmation of the full conversion of **2** by TLC or GC, the mixture was stirred at 100 °C for 6 h. Upon cooling to room temperature, the reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (3 mL) and filtered through a pad of silica gel with additional CH<sub>2</sub>Cl<sub>2</sub> (10 mL) as an eluent. The organic solution was concentrated under reduced pressure, and the residue was purified by flash chromatography on silica gel or preparative TLC on silica gel to afford the desired product.



**(R)-(2,4-Diphenylcyclopent-1-en-1-yl)(phenyl)methanone (4aa):** Pale yellow solid (78.3 mg, 80%); *R<sub>f</sub>* 0.31 (hexane/EtOAc = 19/1); m.p. 79-80 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.79-7.76 (m, 2H), 7.40-7.32 (m, 5H), 7.27-7.21 (m, 3H), 7.19-7.16 (m, 2H), 7.12-7.08 (m, 3H), 3.79 (quint, *J* = 8.3 Hz, 1H), 3.49-3.33 (m, 2H), 3.23-3.08 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 197.6, 145.3, 145.0, 136.5, 136.3, 135.6, 132.8, 129.3, 128.6, 128.2, 128.0, 127.9, 127.9, 126.9, 126.3, 45.7, 45.2, 42.4; HRMS (ESI) Calcd for C<sub>24</sub>H<sub>21</sub>O [M + H]<sup>+</sup> 325.1592, found 325.1592; [α]<sub>D</sub><sup>25</sup> = +14.4 (*c* = 1.31 in CHCl<sub>3</sub>, 95:5 er sample).

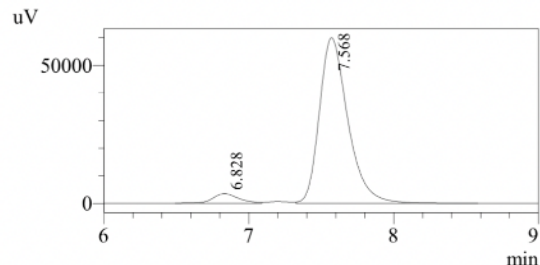
HPLC analysis: Daicel CHIRALPAK AS-H; hexane:*i*-PrOH = 95:5; detection wavelength = 254 nm; flow rate = 1.0 mL/min. *t<sub>R</sub>* = 6.8 min (minor) and 7.6 min (major), 95:5 er.



1 Det.A Ch1 / 254nm

PeakTable

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 6.798     | 49.818  |
| 2     | 7.529     | 50.182  |
| Total |           | 100.000 |

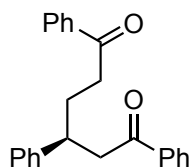


1 Det.A Ch1 / 254nm

PeakTable

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 6.828     | 4.949   |
| 2     | 7.568     | 95.051  |
| Total |           | 100.000 |

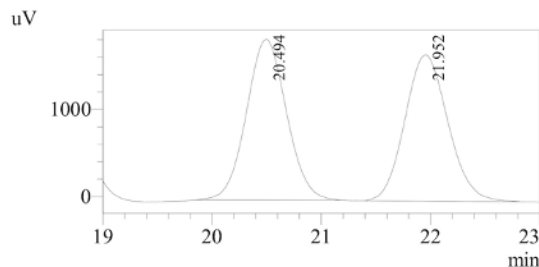
**3 mmol-scale reaction:** In an argon-filled glove box, a 50-mL Schlenk tube (pre-cooled in  $-20\text{ }^{\circ}\text{C}$  fridge) equipped with a magnetic stir bar was charged sequentially with **L6** (92.4 mg, 0.45 mmol), molecular sieves  $4\text{ \AA}$  (3.0 g), and DMPU (9 mL, stored in  $-20\text{ }^{\circ}\text{C}$  fridge). To the mixture was added  $\text{Et}_2\text{Zn}$  (1 M in hexane, 450  $\mu\text{L}$ , 0.45 mmol), **2a** (625 mg, 3.0 mmol), and **1a** (604 mg, 4.5 mmol) sequentially. After stirring for 30 seconds, the Schlenk tube was closed and removed from the glove box, and the mixture was stirred at  $0\text{ }^{\circ}\text{C}$  for 48 h and then at  $100\text{ }^{\circ}\text{C}$  for 6 h. Upon cooling to room temperature,  $\text{CH}_2\text{Cl}_2$  (30 mL) and  $\text{H}_2\text{O}$  (30 mL) were added to the reaction mixture. The aqueous layer was extracted with  $\text{CH}_2\text{Cl}_2$  ( $2 \times 30\text{ mL}$ ) and the combined organic layers were dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated under reduced pressure. The residue was purified by flash chromatography on silica gel to afford the product as a pale yellow solid (683 mg, 70%, 95:5 er).



**(S)-1,3,6-Triphenylhexane-1,6-dione (3aa):** The reaction was performed at  $0\text{ }^{\circ}\text{C}$  for 48 h. White solid (88.1 mg, 86%);  $R_f$  0.32 (hexane/EtOAc = 9/1); m.p.  $146\text{--}148\text{ }^{\circ}\text{C}$ ;  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.90–7.88 (m, 2H), 7.83–7.81 (m, 2H), 7.55–7.48 (m, 2H), 7.44–7.37 (m, 4H), 7.32–7.25 (m, 4H), 7.22–7.18 (m, 1H), 3.48–3.41 (m, 1H), 3.39–3.28 (m, 2H), 2.93 (ddd,  $J = 17.1, 10.2, 5.9\text{ Hz}$ , 1H), 2.75 (ddd,  $J = 17.1, 10.1, 4.9\text{ Hz}$ , 1H), 2.27–2.19 (m, 1H), 2.12–2.02 (m, 1H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  199.9, 198.6, 143.8, 137.1, 136.8, 133.0, 132.9, 128.7, 128.5, 128.5, 128.0, 127.9, 127.6, 126.6, 46.0, 40.7, 36.7, 30.5; **HRMS** (ESI) Calcd for  $\text{C}_{24}\text{H}_{23}\text{O}_2$   $[\text{M} + \text{H}]^+$  343.1698,

found 343.1695;  $[\alpha]^{25}_D = -11.7$  ( $c = 2.30$  in  $\text{CHCl}_3$ , 95:5 er sample).

HPLC analysis: Daicel CHIRALPAK ID; hexane:*i*-PrOH = 90:10; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R = 20.5$  min (minor) and 22.0 min (major), 95:5 er.

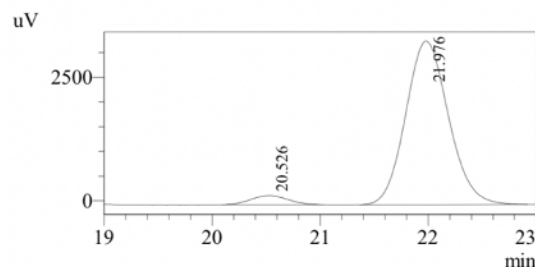


1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 20.494    | 50.482  |
| 2     | 21.952    | 49.518  |
| Total |           | 100.000 |

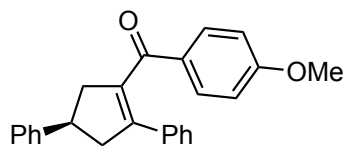


1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

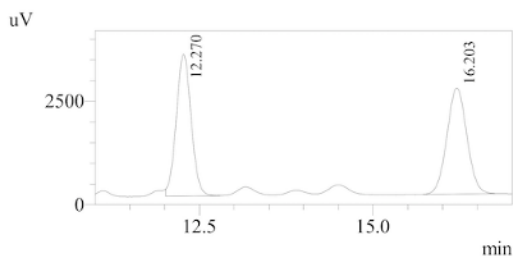
| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 20.526    | 4.687   |
| 2     | 21.976    | 95.313  |
| Total |           | 100.000 |



**(R)-(2,4-Diphenylcyclopent-1-en-1-yl)(4-methoxyphenyl)methanone (4ba):** Pale yellow oil (71.1 mg, 67%);  $R_f$  0.21 (hexane/EtOAc = 19/1);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.82-7.78 (m, 2H), 7.39-7.31 (m, 4H), 7.25-7.21 (m, 3H), 7.15-7.08 (m, 3H), 6.78-6.74 (m, 2H), 3.82-3.74 (m, 4H), 3.47-3.30 (m, 2H), 3.21-3.05 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  196.5, 163.5, 145.4, 142.7, 136.6, 135.6, 131.7, 129.3, 128.5, 128.1, 127.8, 127.7, 126.9, 126.3, 113.6, 55.3, 45.5, 45.3, 42.4; **HRMS** (ESI) Calcd for  $\text{C}_{25}\text{H}_{23}\text{O}_2$   $[\text{M} + \text{H}]^+$  355.1698, found 355.1699;  $[\alpha]^{25}_D = +13.1$  ( $c = 1.12$  in  $\text{CHCl}_3$ , 95:5 er sample).

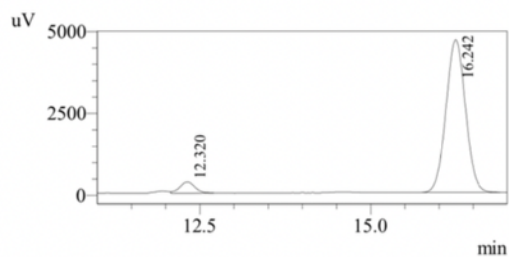
HPLC analysis: Daicel CHIRALPAK ID; hexane:*i*-PrOH = 90:10; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R = 12.3$  min (minor) and 16.2 min (major), 95:5 er.





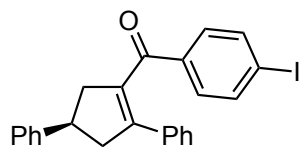
1 Det.A Ch1 / 254nm

| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 12.270    | 50.097  |
| 2                    | 16.203    | 49.903  |
| Total                |           | 100.000 |



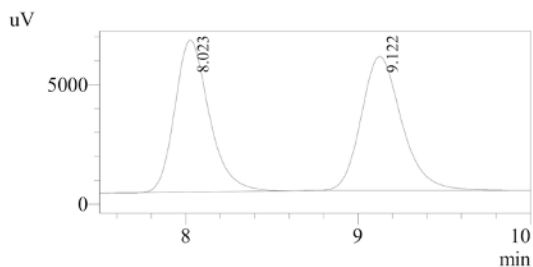
1 Det.A Ch1 / 254nm

| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 12.320    | 5.265   |
| 2                    | 16.242    | 94.735  |
| Total                |           | 100.000 |



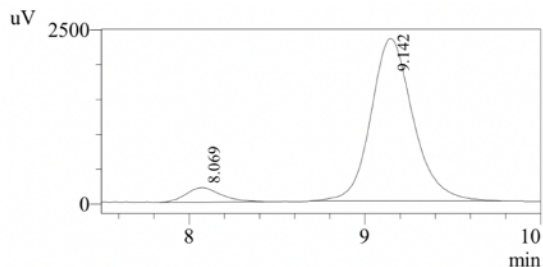
**(R)-(2,4-Diphenylcyclopent-1-en-1-yl)(4-iodophenyl)methanone (4ca):** White solid (115.4 mg, 85%);  $R_f$  0.40 (hexane/EtOAc = 19/1); m.p. 131-133 °C;  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.61-7.58 (m, 2H), 7.46-7.43 (m, 2H), 7.38-7.32 (m, 4H), 7.26-7.22 (m, 1H), 7.16-7.11 (m, 5H), 3.78 (quint,  $J = 8.2$  Hz, 1H), 3.49-3.42 (m, 1H), 3.38-3.32 (m, 1H), 3.21-3.06 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  196.7, 146.0, 145.2, 137.5, 135.9, 135.8, 135.4, 130.7, 128.6, 128.2, 128.2, 127.9, 126.9, 126.4, 100.7, 45.8, 45.0, 42.3; **HRMS** (ESI) Calcd for  $\text{C}_{24}\text{H}_{20}\text{OI}$   $[\text{M} + \text{H}]^+$  451.0559, found 451.0558;  $[\alpha]_D^{25} = +14.5$  ( $c = 1.84$  in  $\text{CHCl}_3$ , 93:7 er sample).

HPLC analysis: Daicel CHIRALPAK AS-H; hexane:*i*-PrOH = 98:2; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 8.1 min (minor) and 9.1 min (major), 93:7 er.



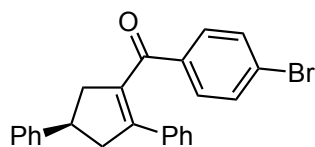
1 Det.A Ch1 / 254nm

| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 8.023     | 49.028  |
| 2                    | 9.122     | 50.972  |
| Total                |           | 100.000 |



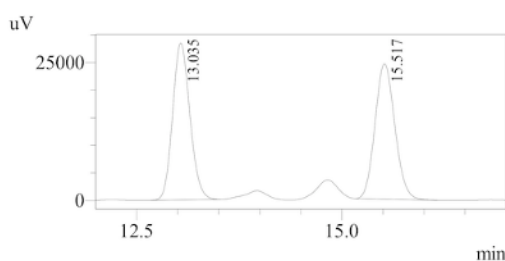
1 Det.A Ch1 / 254nm

| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 8.069     | 6.837   |
| 2                    | 9.142     | 93.163  |
| Total                |           | 100.000 |



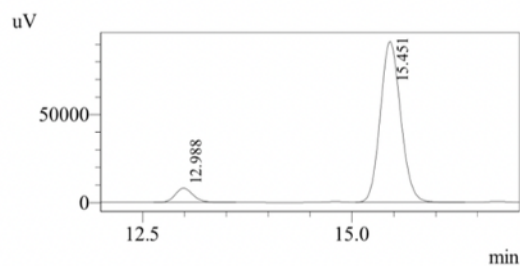
**(R)-(4-Bromophenyl)(2,4-diphenylcyclopent-1-en-1-yl)methanone (4da):** The reaction was performed at 0 °C for 72 h, followed by further heating at 100 °C for 6 h. White solid (109.6 mg, 91%);  $R_f$  0.42 (hexane/EtOAc = 19/1); m.p. 99-101 °C;  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.61-7.58 (m, 2H), 7.38-7.32 (m, 6H), 7.26-7.23 (m, 1H), 7.16-7.11 (m, 5H), 3.79 (quint,  $J$  = 8.2 Hz, 1H), 3.49-3.43 (m, 1H), 3.39-3.32 (m, 1H), 3.22-3.07 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  196.4, 146.1, 145.2, 135.8, 135.4, 135.4, 131.5, 130.8, 128.6, 128.2, 128.2, 127.9, 127.8, 126.9, 126.4, 45.8, 45.0, 42.3; **HRMS** (ESI) Calcd for  $\text{C}_{24}\text{H}_{20}\text{OBr}$   $[\text{M} + \text{H}]^+$  403.0696, found 403.0698;  $[\alpha]_D^{25} = +16.2$  ( $c$  = 1.25 in  $\text{CHCl}_3$ , 93:7 er sample).

HPLC analysis: Daicel CHIRALPAK ID; hexane:*i*-PrOH = 90:10; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 13.0 min (minor) and 15.5 min (major), 93:7 er.



1 Det.A Ch1 / 254nm

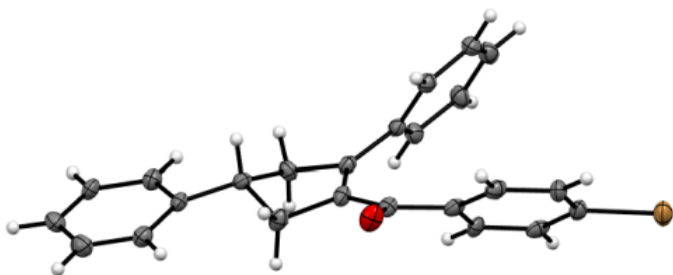
| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 13.035    | 50.218  |
| 2                    | 15.517    | 49.782  |
| Total                |           | 100.000 |



1 Det.A Ch1 / 254nm

| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 12.988    | 7.159   |
| 2                    | 15.451    | 92.841  |
| Total                |           | 100.000 |

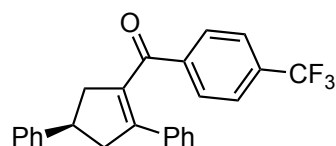
Recrystallization of **4da** (sample of 93:7 er) from THF/pentane afforded single crystals of racemate first. The supernatant of this first recrystallization contained enantiopure **4da** (>99:1 er, confirmed by HPLC). The second recrystallization of this enantiopure sample from THF/pentane afforded single crystals suitable for X-ray diffraction analysis, which unambiguously confirmed the molecular structure of **4da** and its absolute stereochemistry (**Figure S2**).<sup>34</sup>



**Figure S2.** ORTEP drawing of **4da** (thermal ellipsoids set at 50% probability; CCDC 2056575).

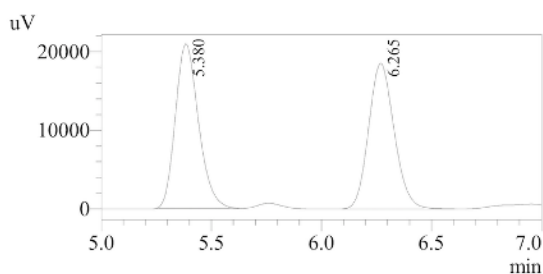
**Table S3.** Crystal data and structure refinement for **4da**

|                                   |  |                              |
|-----------------------------------|--|------------------------------|
| Identification code               | yoshi108m_0m_5   |                              |
| Chemical formula                  | C <sub>24</sub> H <sub>19</sub> BrO  |                              |
| Formula weight                    | 403.30 g/mol   |                              |
| Temperature                       | 100(2) K   |                              |
| Wavelength                        | 0.71073 Å  |                              |
| Crystal size                      | 0.060 x 0.140 x 0.200 mm   |                              |
| Crystal habit                     | colorless plate  |                              |
| Crystal system                    | triclinic  |                              |
| Space group                       | P 1  |                              |
| Unit cell dimensions              | a = 7.6759(4) Å  | $\alpha = 87.9511(19)^\circ$ |
|                                   | b = 9.4634(5) Å  | $\beta = 79.5409(19)^\circ$  |
|                                   | c = 13.2999(8) Å   | $\gamma = 74.096(2)^\circ$   |
| Volume                            | 913.59(9) Å <sup>3</sup>   |                              |
| Z                                 | 2  |                              |
| Density (calculated)              | 1.466 g/cm <sup>3</sup>  |                              |
| Absorption coefficient            | 2.259 mm <sup>-1</sup>   |                              |
| F(000)                            | 412  |                              |
| Theta range for data collection   | 2.24 to 33.73°   |                              |
| Index ranges                      | -11 ≤ h ≤ 11, -14 ≤ k ≤ 14, -20 ≤ l ≤ 20   |                              |
| Reflections collected             | 12460  |                              |
| Independent reflections           | 12460 [R(int) = 0.0410]  |                              |
| Coverage independent reflections  | 99.7%  |                              |
| Absorption correction             | Multi-Scan   |                              |
| Max. and min. transmission        | 0.8760 and 0.6610  |                              |
| Structure solution technique      | direct methods   |                              |
| Structure solution program        | XT, VERSION 2018/2   |                              |
| Refinement method                 | Full-matrix least-squares on F <sup>2</sup>                                      |                              |
| Refinement program                | SHELXL-2018/3 (Sheldrick, 2018)  |                              |
| Function minimized                | $\Sigma w(F_o^2 - F_c^2)^2$  |                              |
| Data / restraints / parameters    | 12460 / 3 / 470  |                              |
| Goodness-of-fit on F <sup>2</sup> | 1.052  |                              |
| Final R indices                   | 9274 data; I > 2σ(I)   | R1 = 0.0525, wR2 = 0.1137    |
|                                   | all data   | R1 = 0.0781, wR2 = 0.1268    |
| Weighting scheme                  | $w = 1/[\sigma^2(F_o^2) + (0.0225P)^2 + 0.1052P]$ where $P = (F_o^2 + 2F_c^2)/3$ |                              |
| Absolute structure parameter      | 0.017(8)   |                              |
| Largest diff. peak and hole       | 0.827 and -1.094 eÅ <sup>-3</sup>  |                              |
| R.M.S. deviation from mean        | 0.098 eÅ <sup>-3</sup>   |                              |



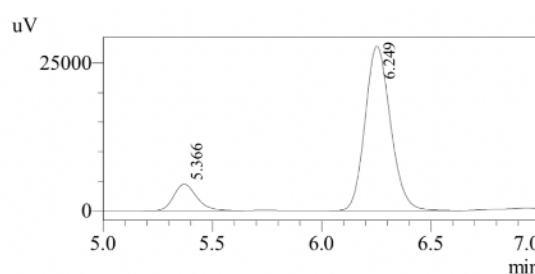
**(R)-(2,4-Diphenylcyclopent-1-en-1-yl)(4-(trifluoromethyl)phenyl)methanone (4ea):** The reaction was performed at 30 °C for 12 h and then at 100 °C for 6 h. White solid (100.2 mg, 85%);  $R_f$  0.43 (hexane/EtOAc = 19/1); m.p. 128-129 °C;  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.78-7.76 (m, 2H), 7.46-7.44 (m, 2H), 7.39-7.33 (m, 4H), 7.26-7.22 (m, 1H), 7.12-7.06 (m, 5H), 3.79 (quint,  $J$  = 8.2 Hz, 1H), 3.51-3.36 (m, 2H), 3.23-3.10 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  196.0, 148.2, 145.1, 139.7, 135.8, 135.4, 133.5 (q,  $^2J_{\text{C-F}}$  = 32.5 Hz), 129.5, 128.6, 128.4, 128.1, 128.0, 126.9, 126.4, 125.0 (q,  $^3J_{\text{C-F}}$  = 3.7 Hz), 123.5 (q,  $^1J_{\text{C-F}}$  = 272.6 Hz), 46.2, 44.7, 42.2;  $^{19}\text{F NMR}$  (376 MHz,  $\text{CDCl}_3$ )  $\delta$  -63.1; **HRMS** (ESI) Calcd for  $\text{C}_{25}\text{H}_{20}\text{OF}_3$   $[\text{M} + \text{H}]^+$  393.1466, found 393.1463;  $[\alpha]^{25}_{\text{D}} = +15.7$  ( $c$  = 1.52 in  $\text{CHCl}_3$ , 87:13 er sample).

HPLC analysis: Daicel CHIRALPAK ID; hexane:*i*-PrOH = 90:10; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 5.4 min (minor) and 6.2 min (major), 87:13 er.



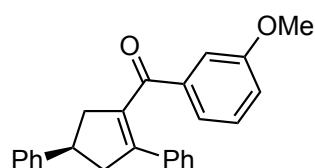
1 Det.A Ch1 / 254nm

| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 5.380     | 50.478  |
| 2                    | 6.265     | 49.522  |
| Total                |           | 100.000 |



1 Det.A Ch1 / 254nm

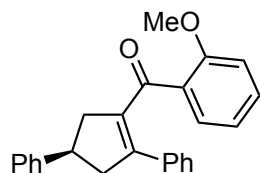
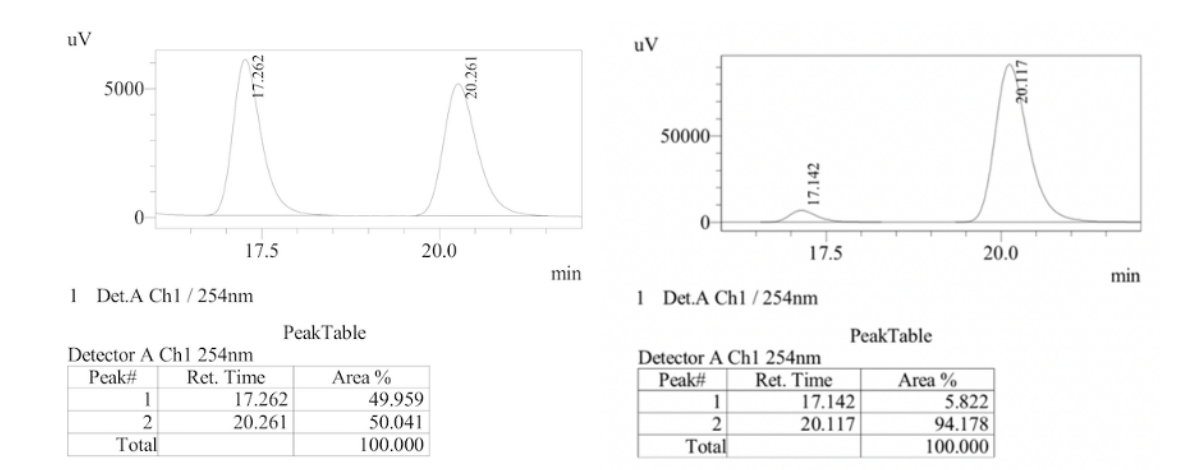
| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 5.366     | 12.745  |
| 2                    | 6.249     | 87.255  |
| Total                |           | 100.000 |



**(R)-(2,4-Diphenylcyclopent-1-en-1-yl)(3-methoxyphenyl)methanone (4fa):** The reaction was performed at 0 °C for 72 h and then at 100 °C for 6 h. Pale yellow oil (82.0 mg, 77%);  $R_f$  0.26 (hexane/EtOAc = 19/1);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.38-7.31 (m, 6H), 7.25-7.11 (m, 7H),

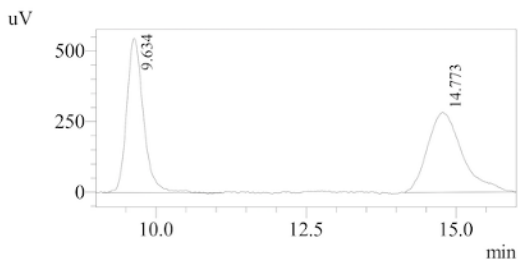
6.95-6.92 (m, 1H), 3.83-3.72 (m, 4H), 3.48-3.33 (m, 2H), 3.22-3.07 (m, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  197.4, 159.5, 145.3, 144.7, 137.8, 136.3, 135.6, 129.2, 128.6, 128.0, 127.9, 127.8, 126.9, 126.3, 122.3, 119.7, 113.1, 55.3, 45.6, 45.3, 42.3; **HRMS** (ESI) Calcd for  $\text{C}_{25}\text{H}_{23}\text{O}_2$   $[\text{M} + \text{H}]^+$  355.1698, found 355.1695;  $[\alpha]_D^{25} = +6.8$  ( $c = 0.38$  in  $\text{CHCl}_3$ , 94:6 er sample).

HPLC analysis: Daicel CHIRALPAK AS-H; hexane:*i*-PrOH = 95:5; detection wavelength = 254 nm; flow rate = 0.5 mL/min.  $t_R$  = 17.1 min (minor) and 20.1 min (major), 94:6 er.



**(R)-(2,4-Diphenylcyclopent-1-en-1-yl)(2-methoxyphenyl)methanone (4ga):** The reaction was performed at 30 °C for 48 h and then at 100 °C for 6 h. Pale yellow oil (73.7 mg, 69%);  $R_f$  0.15 (hexane/EtOAc = 19/1);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.38-7.31 (m, 5H), 7.24-7.20 (m, 1H), 7.18-7.14 (m, 1H), 7.10-7.07 (m, 2H), 7.06-7.02 (m, 3H), 6.78-6.75 (m, 1H), 6.58-6.55 (m, 1H), 3.72-3.64 (m, 4H), 3.42-3.31 (m, 2H), 3.12-3.04 (m, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  195.6, 157.4, 149.0, 145.9, 138.1, 136.1, 132.2, 130.1, 129.3, 128.5, 127.8, 127.6, 127.4, 126.8, 126.2, 120.0, 110.7, 55.4, 47.6, 43.5, 41.7; **HRMS** (ESI) Calcd for  $\text{C}_{25}\text{H}_{23}\text{O}_2$   $[\text{M} + \text{H}]^+$  355.1698, found 355.1697;  $[\alpha]_D^{25} = -4.3$  ( $c = 0.35$  in  $\text{CHCl}_3$ , 91:9 er sample).

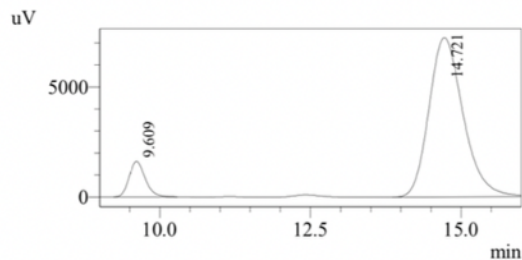
HPLC analysis: Daicel CHIRALPAK ID; hexane:*i*-PrOH = 80:20; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 9.6 min (minor) and 14.7 min (major), 91:9 er.



1 Det.A Ch1 / 254nm

PeakTable

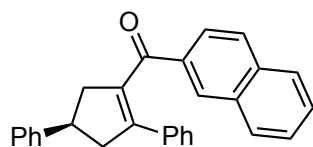
| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 9.634     | 48.120  |
| 2     | 14.773    | 51.880  |
| Total |           | 100.000 |



1 Det.A Ch1 / 254nm

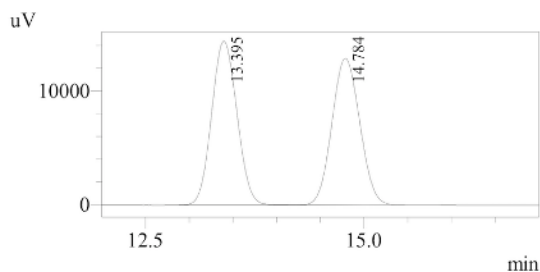
PeakTable

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 9.609     | 9.459   |
| 2     | 14.721    | 90.541  |
| Total |           | 100.000 |



**(R)-(2,4-Diphenylcyclopent-1-en-1-yl)(naphthalen-2-yl)methanone (4ha):** The reaction was performed at 0 °C for 72 h and then at 100 °C for 6 h. Colorless oil (105.0 mg, 93%);  $R_f$  0.31 (hexane/EtOAc = 19/1);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.26 (s, 1H), 7.93-7.90 (m, 1H), 7.75-7.70 (m, 3H), 7.50-7.46 (m, 1H), 7.43-7.33 (m, 5H), 7.27-7.22 (m, 3H), 7.06-6.96 (m, 3H), 3.82 (quint,  $J$  = 8.0 Hz, 1H), 3.52-3.38 (m, 2H), 3.26-3.12 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  197.6, 145.3, 144.5, 136.5, 135.5, 135.4, 133.7, 132.3, 131.6, 129.4, 128.6, 128.2, 128.2, 128.0, 127.8, 127.7, 127.6, 126.9, 126.4, 126.3, 124.5, 45.5, 45.4, 42.4; **HRMS** (ESI) Calcd for  $\text{C}_{28}\text{H}_{23}\text{O}$   $[\text{M} + \text{H}]^+$  375.1749, found 375.1747;  $[\alpha]_D^{25} = +18.7$  ( $c$  = 3.00 in  $\text{CHCl}_3$ , 95:5 er sample).

HPLC analysis: Daicel CHIRALPAK AD-H; hexane:*i*-PrOH = 90:10; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 13.5 min (minor) and 14.9 min (major), 95:5 er.

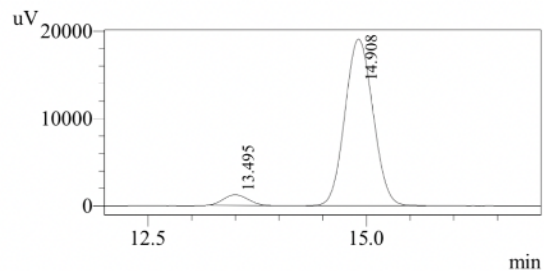


1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 13.395    | 50.403  |
| 2     | 14.784    | 49.597  |
| Total |           | 100.000 |

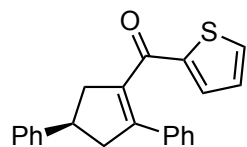


1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

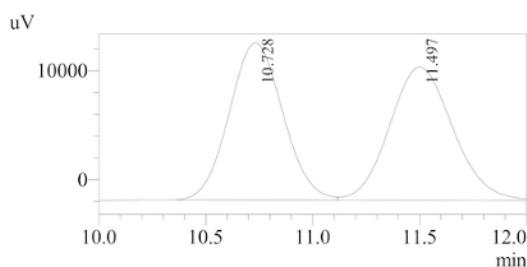
| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 13.495    | 5.295   |
| 2     | 14.908    | 94.705  |
| Total |           | 100.000 |



**(R)-(2,4-Diphenylcyclopent-1-en-1-yl)(thiophen-2-yl)methanone (4ia):** Pale yellow oil (81.2 mg, 82%);  $R_f$  0.28 (hexane/EtOAc = 19/1);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.50-7.49 (m, 1H), 7.41-7.32 (m, 5H), 7.29-7.13 (m, 6H), 6.88-6.86 (m, 1H), 3.77 (quint,  $J = 8.1$  Hz, 1H), 3.47-3.35 (m, 2H), 3.20-3.08 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  189.6, 145.3, 144.1, 143.6, 136.3, 135.6, 134.1, 134.0, 128.6, 128.2, 128.0, 127.9, 127.8, 126.9, 126.3, 45.5, 45.3, 42.3; **HRMS** (ESI) Calcd for  $\text{C}_{22}\text{H}_{19}\text{OS}$   $[\text{M} + \text{H}]^+$  331.1157, found 331.1155;  $[\alpha]_D^{25} = +9.23$  ( $c = 2.15$  in  $\text{CHCl}_3$ , 96:4 er sample).

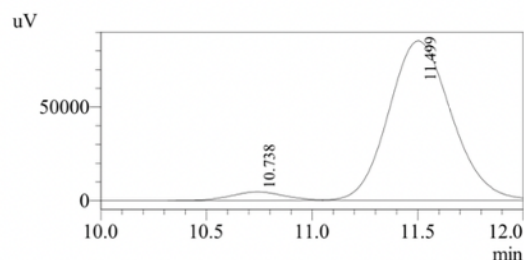
HPLC analysis: Daicel CHIRALPAK ID; hexane:*i*-PrOH = 80:20; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 10.7 min (minor) and 11.5 min (major), 96:4 er.





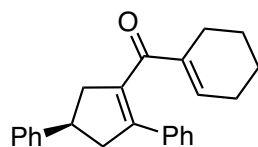
1 Det.A Ch1 / 254nm

| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 10.728    | 49.918  |
| 2                    | 11.497    | 50.082  |
| Total                |           | 100.000 |



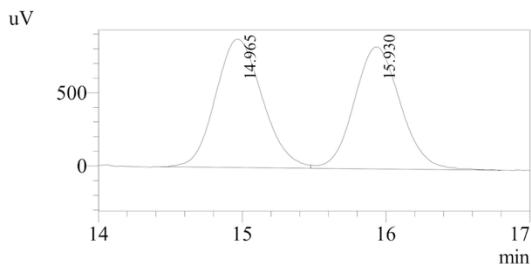
1 Det.A Ch1 / 254nm

| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 10.738    | 4.270   |
| 2                    | 11.499    | 95.730  |
| Total                |           | 100.000 |



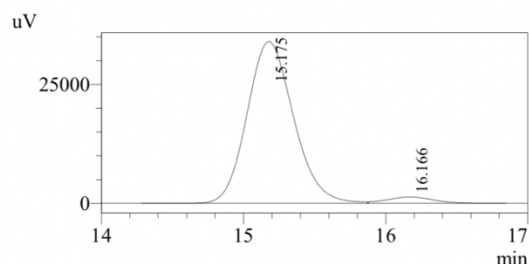
**(R)-Cyclohex-1-en-1-yl(2,4-diphenylcyclopent-1-en-1-yl)methanone (4ja):** White solid (53.4 mg, 54%);  $R_f$  0.31 (hexane/EtOAc = 19/1); m.p. 73-75 °C;  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.35-7.17 (m, 10H), 6.65-6.63 (m, 1H), 3.71 (quint,  $J$  = 8.2 Hz, 1H), 3.38-3.32 (m, 1H), 3.27-3.21 (m, 1H), 3.12-3.05 (m, 1H), 3.01-2.94 (m, 1H), 2.24-2.21 (m, 2H), 1.94-1.91 (m, 2H), 1.54-1.48 (m, 2H), 1.44-1.38 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  199.5, 145.5, 143.8, 142.8, 138.4, 136.9, 136.4, 128.5, 128.1, 127.6, 127.5, 126.9, 126.2, 45.3, 45.2, 42.3, 26.0, 23.0, 21.7, 21.4; **HRMS** (ESI) Calcd for  $\text{C}_{24}\text{H}_{25}\text{O}$   $[\text{M} + \text{H}]^+$  329.1903, found 329.1905;  $[\alpha]_D^{25} = +10.12$  ( $c$  = 2.10 in  $\text{CHCl}_3$ , 96:4 er sample).

HPLC analysis: Daicel CHIRALPAK OJ-H; hexane:*i*-PrOH = 95:5; detection wavelength = 254 nm; flow rate = 0.5 mL/min.  $t_R$  = 15.2 min (major) and 16.2 min (minor), 96:4 er.



PeakTable

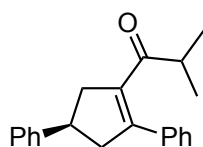
| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 14.965    | 51.573  |
| 2     | 15.930    | 48.427  |
| Total |           | 100.000 |



1 Det.A Ch1 / 254nm

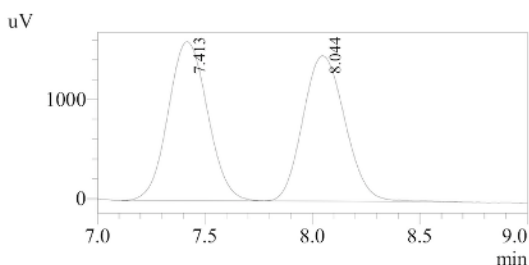
PeakTable

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 15.175    | 96.277  |
| 2     | 16.166    | 3.723   |
| Total |           | 100.000 |



**(R)-1-(2,4-Diphenylcyclopent-1-en-1-yl)-2-methylpropan-1-one (4ka):** The reaction was performed at 30 °C for 48 h and then at 100 °C for 6 h. Pale yellow oil (41.8 mg, 48%);  $R_f$  0.41 (hexane/EtOAc = 39/1);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.38-7.21 (m, 10H), 3.65 (quint,  $J = 8.2$  Hz, 1H), 3.35-3.23 (m, 2H), 3.09-2.94 (m, 2H), 2.63 (sept,  $J = 6.9$  Hz, 1H), 0.98-0.96 (m, 6H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  208.5, 147.1, 145.5, 137.7, 136.9, 128.6, 128.3, 128.3, 127.6, 126.8, 126.3, 47.3, 44.2, 41.9, 38.9, 18.6, 18.3; **HRMS** (ESI) Calcd for  $\text{C}_{21}\text{H}_{23}\text{O}$   $[\text{M} + \text{H}]^+$  291.1749, found 291.1743;  $[\alpha]_D^{25} = -9.59$  ( $c = 0.61$  in  $\text{CHCl}_3$ , 97:3 er sample).

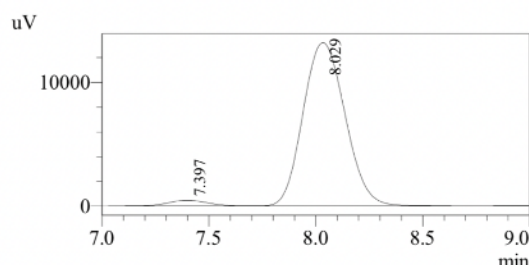
HPLC analysis: Daicel CHIRALPAK AD-H; hexane:*i*-PrOH = 99:1; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R = 7.4$  min (minor) and 8.0 min (major), 97:3 er.



1 Det.A Ch1 / 254nm

PeakTable

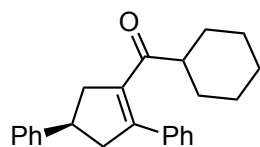
| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 7.413     | 50.667  |
| 2     | 8.044     | 49.333  |
| Total |           | 100.000 |



1 Det.A Ch1 / 254nm

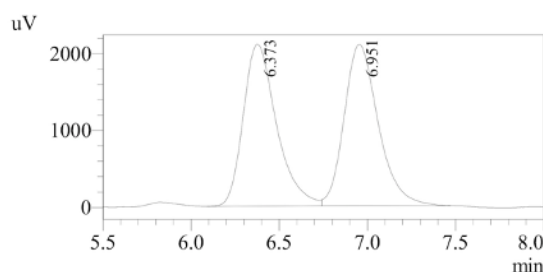
PeakTable

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 7.397     | 3.140   |
| 2     | 8.029     | 96.860  |
| Total |           | 100.000 |



**(R)-Cyclohexyl(2,4-diphenylcyclopent-1-en-1-yl)methanone (4la):** The reaction was performed at 30 °C for 48 h and then at 100 °C for 6 h. Colorless oil (73.3 mg, 74%);  $R_f$  0.19 (hexane/EtOAc = 39/1);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.38-7.19 (m, 10H), 3.63 (quint,  $J$  = 8.2 Hz, 1H), 3.33-3.21 (m, 2H), 3.08-2.92 (m, 2H), 2.31 (tt,  $J$  = 11.5, 3.2 Hz, 1H), 1.70-1.61 (m, 4H), 1.54-1.50 (m, 1H), 1.35-1.22 (m, 2H), 1.15-1.04 (m, 1H), 0.97-0.84 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  207.2, 147.5, 145.4, 137.8, 137.0, 128.5, 128.3, 128.1, 127.5, 126.8, 126.2, 48.8, 47.7, 43.8, 41.9, 28.9, 28.6, 25.7, 25.6, 25.5; **HRMS** (ESI) Calcd for  $\text{C}_{24}\text{H}_{27}\text{O}$   $[\text{M} + \text{H}]^+$  331.2062, found 331.2065;  $[\alpha]_D^{25}$  = -5.28 ( $c$  = 2.35 in  $\text{CHCl}_3$ , 97:3 er sample).

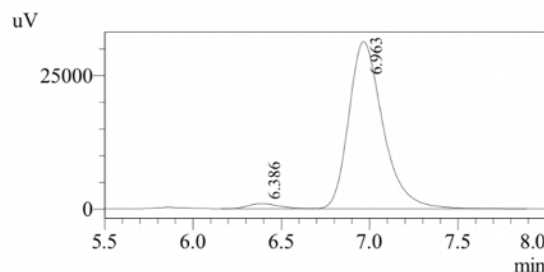
HPLC analysis: Daicel CHIRALPAK AS-H; hexane:*i*-PrOH = 99:1; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 6.4 min (minor) and 7.0 min (major), 97:3 er.



1 Det.A Ch1 / 254nm

PeakTable

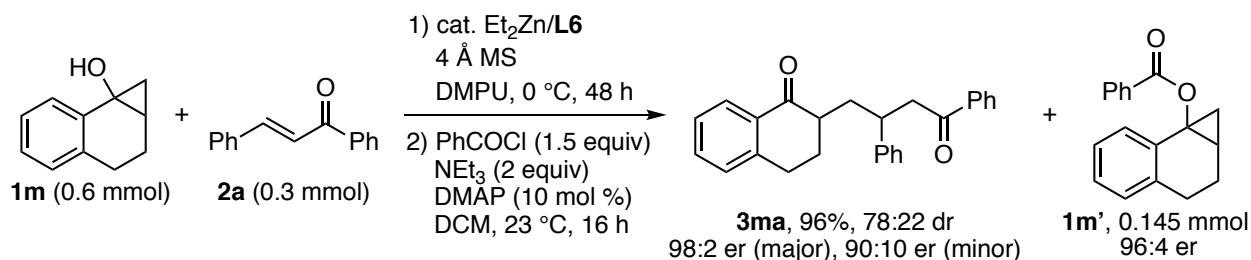
| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 6.373     | 49.574  |
| 2     | 6.951     | 50.426  |
| Total |           | 100.000 |



1 Det.A Ch1 / 254nm

PeakTable

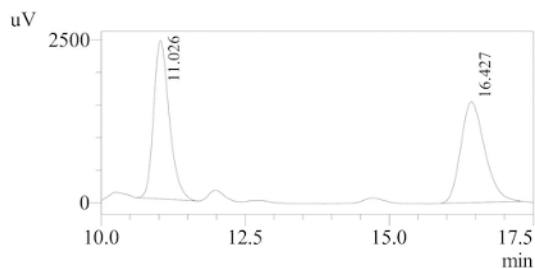
| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 6.386     | 2.766   |
| 2     | 6.963     | 97.234  |
| Total |           | 100.000 |



**2-(4-Oxo-2,4-diphenylbutyl)-3,4-dihydronaphthalen-1(2H)-one (3ma):** The reaction was

conducted using 2 equiv of **1m** at 0 °C for 48 h. A mixture of the unreacted cyclopropanol (**1m**) and the product (**3ma**) was obtained by silica gel chromatography. For the separation of these compounds and detection of **1m** by the UV detector in HPLC, the unreacted **1m** was transformed to the corresponding benzoyl derivative (**1m'**) by the following procedure: To a solution of a mixture of **1m** and **3ma** in CH<sub>2</sub>Cl<sub>2</sub> (0.5 mL) was added benzoyl chloride (63.3 mg, 0.45 mmol) and triethylamine (84 µL, 0.6 mmol), and DMAP (3.7 mg, 0.03 mmol). The reaction mixture was stirred at 23 °C for 16 h. The reaction was quenched with sat. aq. NH<sub>4</sub>Cl and extracted with CH<sub>2</sub>Cl<sub>2</sub> (3 times), and the combined organic extracts were dried over MgSO<sub>4</sub> and concentrated under reduced pressure. The residue was purified with silica gel chromatography to afford the title compound as a white solid (105.9 mg, 96%, 78:22 dr) along with **1m'** (38.4 mg, 0.145 mmol, 24%). Pure samples of both the diastereomers of **3ma** could be obtained by partial separation of the diastereomer mixture on silica gel using toluene as an eluent; *R<sub>f</sub>* 0.31 (single spot, hexane/EtOAc = 19/1), 0.26 (major diastereomer, toluene), 0.18 (minor diastereomer, toluene); m.p. 152-153 °C (major diastereomer); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) major diastereomer: δ 7.97-7.96 (m, 1H), 7.89-7.87 (m, 2H), 7.54-7.50 (m, 1H), 7.43-7.38 (m, 3H), 7.34-7.14 (m, 7H), 3.61-3.54 (m, 1H), 3.32 (d, *J* = 6.8 Hz, 2H), 3.00 (dt, *J* = 16.8, 4.4 Hz, 1H), 2.87-3.79 (m, 1H), 2.60-2.53 (m, 1H), 2.42-2.35 (m, 1H), 2.21-2.13 (m, 1H), 1.93-1.83 (m, 1H), 1.80-1.73 (m, 1H); minor diastereomer: 8.00-7.98 (m, 1H), 7.89-7.87 (m, 2H), 7.54-7.49 (m, 1H), 7.44-7.38 (m, 3H), 7.30-7.24 (m, 5H), 7.18-7.14 (m, 2H), 3.75-3.67 (m, 1H), 3.38 (dd, *J* = 16.5, 7.7 Hz, 1H), 3.27 (dd, *J* = 16.5, 6.2 Hz, 1H), 2.91-2.87 (m, 2H), 2.52-2.46 (m, 1H), 2.43-2.35 (m, 1H), 2.13-2.06 (m, 1H), 1.88-1.78 (m, 1H), 1.72-1.65 (m, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) major diastereomer: δ 200.1, 198.7, 143.9, 143.4, 137.1, 133.1, 133.0, 132.3, 128.7, 128.6, 128.5, 127.9, 127.7, 127.3, 126.6, 126.4, 46.6, 45.1, 37.8, 34.7, 28.4, 27.4; minor diastereomer: δ 200.1, 198.8, 144.8, 143.7, 137.1, 133.1, 132.9, 132.5, 128.6, 128.5, 128.5, 128.1, 127.7, 127.4, 126.5, 126.4, 46.0, 45.6, 39.4, 37.2, 29.6, 28.4; HRMS (ESI) Calcd for C<sub>26</sub>H<sub>25</sub>O<sub>2</sub> [M + H]<sup>+</sup> 369.1855, found 369.1852 (major diastereomer), 369.1857 (minor diastereomer); [α]<sub>D</sub><sup>25</sup> = +7.48 (*c* = 0.99 in CHCl<sub>3</sub>, 78:22 dr, 98:2 er (major), 90:10 er (minor) sample of **3ma**); [α]<sub>D</sub><sup>25</sup> = -151 (*c* = 1.79 in CHCl<sub>3</sub>, 96:4 er sample of **1m'**).

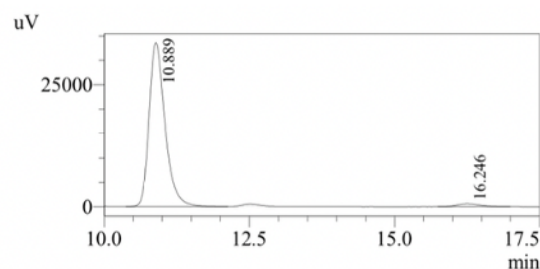
HPLC analysis (major diastereomer of **3ma**): Daicel CHIRALPAK AS-H; hexane:*i*-PrOH = 70:30; detection wavelength = 254 nm; flow rate = 0.5 mL/min. *t<sub>R</sub>* = 10.9 min (major) and 16.2 min (minor), 98:2 er.



1 Det.A Ch1 / 254nm

PeakTable

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 11.026    | 50.849  |
| 2     | 16.427    | 49.151  |
| Total |           | 100.000 |

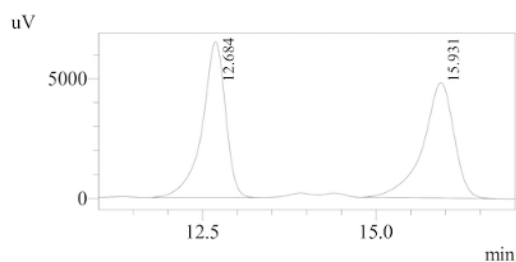


1 Det.A Ch1 / 254nm

PeakTable

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 10.889    | 97.534  |
| 2     | 16.246    | 2.466   |
| Total |           | 100.000 |

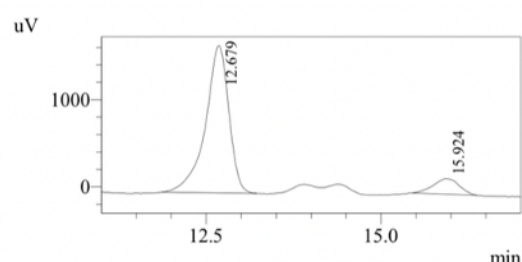
HPLC analysis (minor diastereomer of **3ma**): Daicel CHIRALPAK OD; hexane:*i*-PrOH = 98:2; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 12.7 min (major) and 15.9 min (minor), 90:10 er.



1 Det.A Ch1 / 254nm

PeakTable

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 12.684    | 50.243  |
| 2     | 15.931    | 49.757  |
| Total |           | 100.000 |

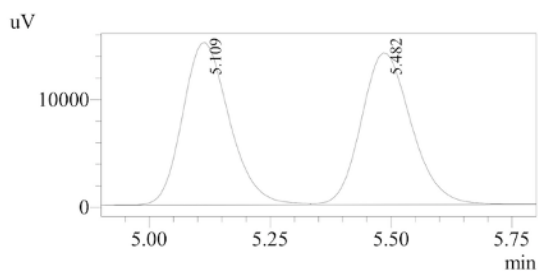


1 Det.A Ch1 / 254nm

PeakTable

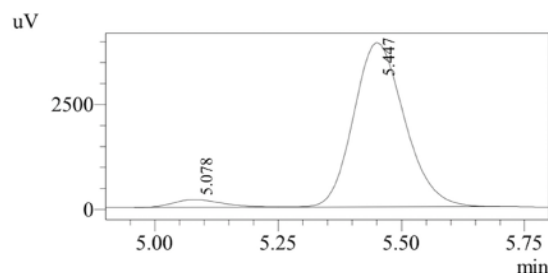
| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 12.679    | 89.563  |
| 2     | 15.924    | 10.437  |
| Total |           | 100.000 |

HPLC analysis (**1m'**): Daicel CHIRALPAK ID; hexane:*i*-PrOH = 95:5; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 5.1 min (minor) and 5.4 min (major), 96:4 er.



1 Det.A Ch1 / 254nm

| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 5.109     | 49.918  |
| 2                    | 5.482     | 50.082  |
| Total                |           | 100.000 |



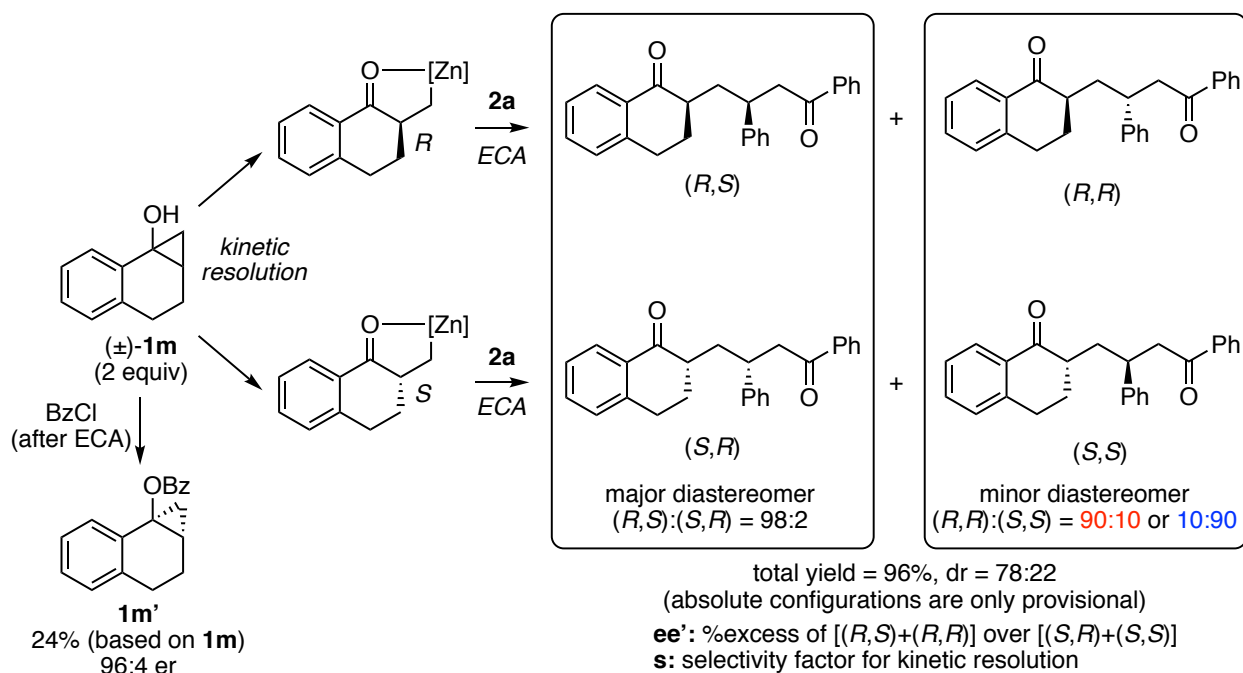
1 Det.A Ch1 / 254nm

| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 5.078     | 4.333   |
| 2                    | 5.447     | 95.667  |
| Total                |           | 100.000 |

Based on the observed dr and ers of **3ma**, one can formulate the pathways for the formation of the diastereomers/enantiomers as shown in Scheme S1, which involve enantioselective ring-opening (kinetic resolution) of **1m** and enantioselective conjugate addition (ECA) of each of the resulting zinc homoenolate to **2a** (the absolute configurations shown are only provisional, assigning (*R,S*) as the dominant enantiomer of the major diastereomer). Here, two possible cases (Cases 1 and 2) can be conceived for the origin of the minor diastereomer, where (*R,R*) and (*S,S*) are assumed as the dominant enantiomer of the minor diastereomer, respectively. For Cases 1 and 2, %excess of [(*R,S*)+(*R,R*)] over [(*S,R*)+(*S,S*)] (**ee'**), which should reflect the kinetic resolution step, is calculated to be 92.5% and 56.9%, respectively. In light of near quantitative yield of **3ma** and these **ee'** values, the selectivity factor (**s**) of the kinetic resolution can be approximated to be 85.6 and 6.3 for Cases 1 and 2, respectively. On the basis of the isolation of benzoyl ester (**1m'**) in a substantial yield (24%)<sup>ii</sup> with high enantiomeric excess (96:4 er; 92% ee), we conclude that Case 1 is the more likely scenario (see the simulated graph in Scheme S1), which corresponds to highly selective kinetic resolution and moderately selective ECA.

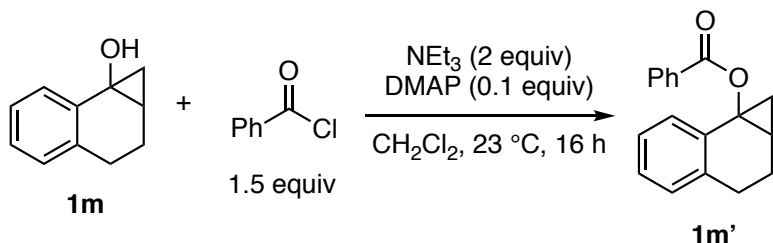
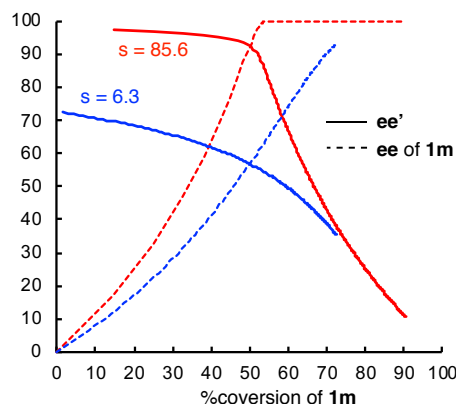
<sup>ii</sup> The isolated yield should be lower than the actual recovery of **1m** because of the loss during the benzoylation step and the purification process.

**Scheme S1.** Kinetic resolution/ECA process in the reaction between **1m** and **2a**



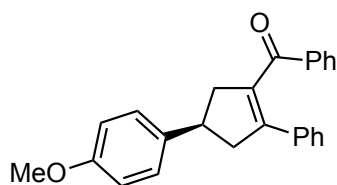
**Case 1:**  $(R,R)$  is the major enantiomer of the minor diastereomer  
**ee'** =  $(0.78 \times 0.98 + 0.22 \times 0.90) - (0.78 \times 0.02 + 0.22 \times 0.10) = 92.5\%$   
 assuming 50% conversion of **1m** and simple first-order kinetics,  
**s**  $\approx \ln[1 - 0.5 \times (1 + 0.925)] / \ln[1 - 0.5 \times (1 - 0.925)] \approx 85.6$

**Case 2:**  $(S,S)$  is the major enantiomer of the minor diastereomer  
**ee'** =  $(0.78 \times 0.98 + 0.22 \times 0.10) - (0.78 \times 0.02 + 0.22 \times 0.90) = 56.9\%$   
 assuming 50% conversion of **1m** and simple first-order kinetics,  
**s**  $\approx \ln[1 - 0.5 \times (1 + 0.569)] / \ln[1 - 0.5 \times (1 - 0.569)] \approx 6.3$



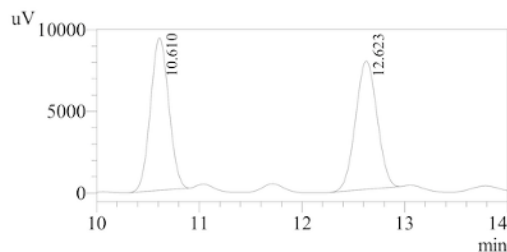
**1,1a,2,3-Tetrahydro-7bH-cyclopropa[a]naphthalen-7b-yl benzoate (**1m'**):** To a solution of **1m** (48.1 mg, 0.30 mmol), benzoyl chloride (63.3 mg, 0.45 mmol) and triethylamine (84  $\mu\text{L}$ , 0.60 mmol) in  $\text{CH}_2\text{Cl}_2$  (0.3 mL) was added DMAP (3.7 mg, 0.030 mmol), and the reaction mixture was stirred at 23 °C for 16 h. The reaction was quenched with sat. aq.  $\text{NH}_4\text{Cl}$  and extracted with  $\text{CH}_2\text{Cl}_2$ ,

and the combined organic extracts were dried over MgSO<sub>4</sub> and concentrated under reduced pressure. The residue was purified with silica gel chromatography to afford the product as a colorless oil (61.0 mg, 77%); *R*<sub>f</sub> 0.59 (hexane/EtOAc = 19/1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.13-8.11 (m, 2H), 7.58-7.55 (m, 1H), 7.47-7.43 (m, 2H), 7.33-7.31 (m, 1H), 7.14-7.07 (m, 3H), 2.73 (dt, *J* = 16.1, 3.4 Hz, 1H), 2.48-2.39 (m, 1H), 2.10-2.05 (m, 2H), 1.94-1.90 (m, 1H), 1.46 (dd, *J* = 10.0, 6.4 Hz, 1H), 1.24 (t, *J* = 6.7 Hz, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 166.1, 136.8, 133.1, 132.5, 130.1, 129.7, 128.5, 128.4, 126.2, 125.8, 123.5, 58.2, 25.8, 23.5, 18.3, 15.2; HRMS (ESI) Calcd for C<sub>18</sub>H<sub>17</sub>O<sub>2</sub> [M + H]<sup>+</sup> 265.1229, found 265.1227.



**(R)-4-(4-Methoxyphenyl)-2-phenylcyclopent-1-en-1-yl(phenyl)methanone (4ab):** White solid (72.7 mg, 68%); *R*<sub>f</sub> 0.24 (hexane/EtOAc = 19/1); m.p. 119-120 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.78-7.75 (m, 2H), 7.40-7.35 (m, 1H), 7.31-7.23 (m, 4H), 7.19-7.15 (m, 2H), 7.11-7.08 (m, 3H), 6.90-6.87 (m, 2H), 3.79-3.70 (m, 4H), 3.46-3.29 (m, 2H), 3.17-3.03 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 197.7, 158.1, 145.0, 137.4, 136.5, 136.4, 135.6, 132.8, 129.3, 128.2, 128.0, 127.9, 127.8, 113.9, 55.3, 45.9, 45.4, 41.7; HRMS (ESI) Calcd for C<sub>25</sub>H<sub>23</sub>O<sub>2</sub> [M + H]<sup>+</sup> 355.1698, found 355.1693; [α]<sub>D</sub><sup>25</sup> = +12.3 (*c* = 1.11 in CHCl<sub>3</sub>, 96:4 er sample).

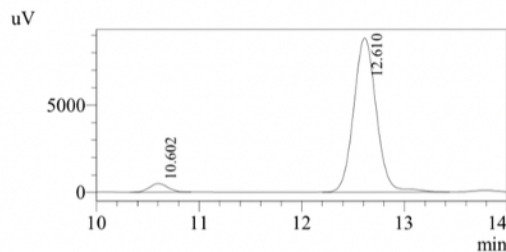
HPLC analysis: Daicel CHIRALPAK ID; hexane:*i*-PrOH = 90:10; detection wavelength = 254 nm; flow rate = 1.0 mL/min. *t*<sub>R</sub> = 10.6 min (minor) and 12.6 min (major), 96:4 er.



1 Det.A Ch1 / 254nm

PeakTable

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 10.610    | 50.267  |
| 2     | 12.623    | 49.733  |
| Total |           | 100.000 |

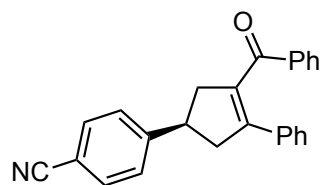


1 Det.A Ch1 / 254nm

PeakTable

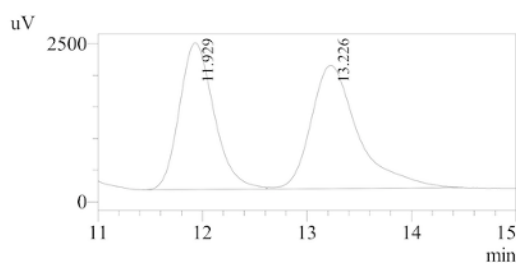
| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 10.602    | 4.340   |
| 2     | 12.610    | 95.660  |
| Total |           | 100.000 |





**(R)-4-(3-Benzoyl-4-phenylcyclopent-3-en-1-yl)benzonitrile (4ac):** Pale yellow oil (73.2 mg, 70%);  $R_f$  0.12 (hexane/EtOAc = 19/1);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.76-7.73 (m, 2H), 7.65-7.62 (m, 2H), 7.49-7.47 (m, 2H), 7.42-7.37 (m, 1H), 7.28-7.24 (m, 2H), 7.18-7.11 (m, 5H), 3.87-3.79 (m, 1H), 3.56-3.48 (m, 1H), 3.44-3.37 (m, 1H), 3.18-3.06 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  197.2, 151.0, 144.7, 136.3, 135.9, 135.1, 133.0, 132.5, 129.3, 128.3, 128.2, 128.1, 127.9, 127.8, 118.9, 110.3, 45.4, 44.8, 42.3; **HRMS** (ESI) Calcd for  $\text{C}_{25}\text{H}_{20}\text{NO}$   $[\text{M} + \text{H}]^+$  350.1545, found 350.1545;  $[\alpha]_D^{25} = +6.4$  ( $c = 0.64$  in  $\text{CHCl}_3$ , 92:8 er sample).

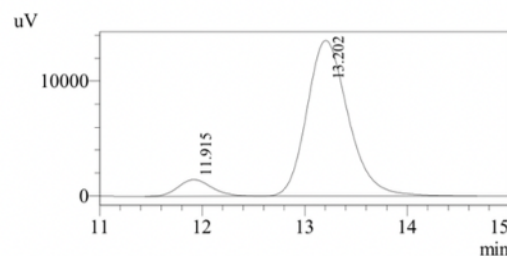
HPLC analysis: Daicel CHIRALPAK ID; hexane:*i*-PrOH = 70:30; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 11.9 min (minor) and 13.2 min (major), 92:8 er.



1 Det.A Ch1 / 254nm

PeakTable

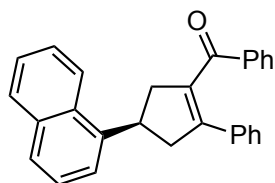
| Detector A Ch1 254nm |           |         |
|----------------------|-----------|---------|
| Peak#                | Ret. Time | Area %  |
| 1                    | 11.929    | 46.770  |
| 2                    | 13.226    | 53.230  |
| Total                |           | 100.000 |



1 Det.A Ch1 / 254nm

PeakTable

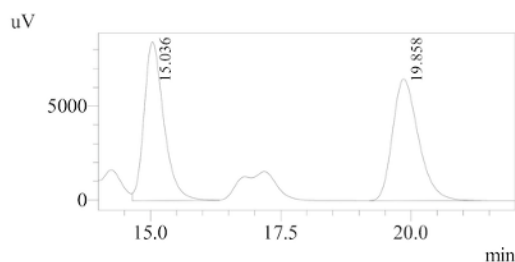
| Detector A Ch1 254nm |           |         |
|----------------------|-----------|---------|
| Peak#                | Ret. Time | Area %  |
| 1                    | 11.915    | 8.021   |
| 2                    | 13.202    | 91.979  |
| Total                |           | 100.000 |



**(R)-4-(Naphthalen-1-yl)-2-phenylcyclopent-1-en-1-yl(phenyl)methanone (4ad):** The reaction was performed at 0 °C for 48 h and then at 50 °C for 16 h. Pale yellow oil (89.7 mg, 80%);  $R_f$  0.29 (hexane/EtOAc = 19/1);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  8.17-8.15 (m, 1H), 7.87-7.85 (m,

1H), 7.79-7.73 (m, 3H), 7.59-7.42 (m, 4H), 7.37-7.32 (m, 1H), 7.24-7.17 (m, 4H), 7.11-7.06 (m, 3H), 4.54-4.46 (m, 1H), 3.63-3.49 (m, 2H), 3.34-3.21 (m, 2H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 197.6, 145.0, 140.8, 136.5, 136.4, 135.5, 134.1, 132.8, 131.5, 129.3, 129.0, 128.2, 128.0, 127.9, 127.9, 127.0, 125.9, 125.5, 125.5, 123.4, 122.8, 45.0, 44.4, 37.8; HRMS (ESI) Calcd for C<sub>28</sub>H<sub>23</sub>O [M + H]<sup>+</sup> 375.1749, found 375.1751; [α]<sub>D</sub><sup>25</sup> = -16.7 (*c* = 1.50 in CHCl<sub>3</sub>, 96:4 er sample).

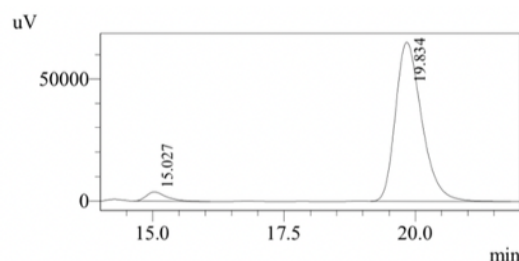
HPLC analysis: Daicel CHIRALPAK AS-H; hexane:*i*-PrOH = 95:5; detection wavelength = 254 nm; flow rate = 1.0 mL/min. *t*<sub>R</sub> = 15.0 min (minor) and 19.8 min (major), 96:4 er.



1 Det.A Ch1 / 254nm

PeakTable

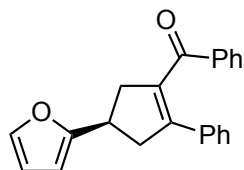
| Detector A Ch1 254nm |           |         |
|----------------------|-----------|---------|
| Peak#                | Ret. Time | Area %  |
| 1                    | 15.036    | 49.805  |
| 2                    | 19.858    | 50.195  |
| Total                |           | 100.000 |



1 Det.A Ch1 / 254nm

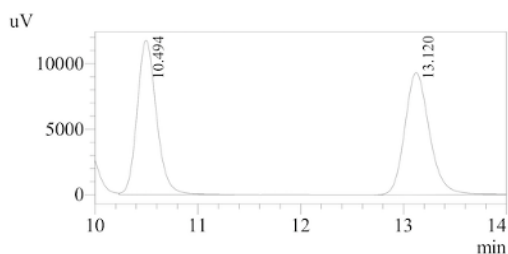
PeakTable

| Detector A Ch1 254nm |           |         |
|----------------------|-----------|---------|
| Peak#                | Ret. Time | Area %  |
| 1                    | 15.027    | 4.154   |
| 2                    | 19.834    | 95.846  |
| Total                |           | 100.000 |



**(R)-(4-(Furan-2-yl)-2-phenylcyclopent-1-en-1-yl)(phenyl)methanone (4ae):** Colorless oil (72.4 mg, 77%); *R*<sub>f</sub> 0.31 (hexane/EtOAc = 19/1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.75-7.73 (m, 2H), 7.38-7.34 (m, 2H), 7.25-7.21 (m, 2H), 7.17-7.13 (m, 2H), 7.10-7.07 (m, 3H), 6.33-6.31 (m, 1H), 6.14-6.14 (m, 1H), 3.81 (quint, *J* = 7.9 Hz, 1H), 3.39-3.21 (m, 3H), 3.17-3.11 (m, 1H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 197.5, 158.0, 144.7, 141.3, 136.5, 136.0, 135.4, 132.8, 129.3, 128.2, 128.0, 127.9, 127.9, 110.1, 104.2, 43.0, 42.5, 35.7; HRMS (ESI) Calcd for C<sub>22</sub>H<sub>19</sub>O<sub>2</sub> [M + H]<sup>+</sup> 315.1385, found 315.1383; [α]<sub>D</sub><sup>25</sup> = +23.3 (*c* = 2.63 in CHCl<sub>3</sub>, 97:3 er).

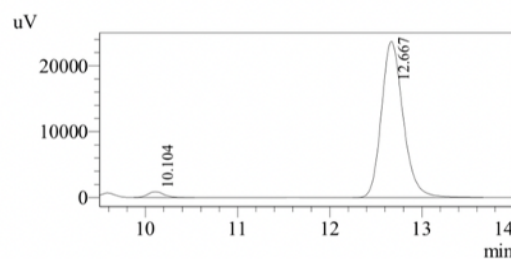
HPLC analysis: Daicel CHIRALPAK ID; hexane:*i*-PrOH = 98:2; detection wavelength = 254 nm; flow rate = 1.0 mL/min. *t*<sub>R</sub> = 10.1 min (minor) and 12.7 min (major), 97:3 er.



1 Det.A Ch1 / 254nm

PeakTable

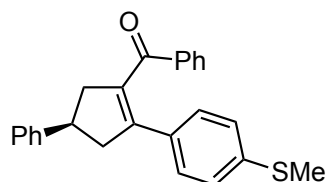
| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 10.494    | 49.742  |
| 2     | 13.120    | 50.258  |
| Total |           | 100.000 |



1 Det.A Ch1 / 254nm

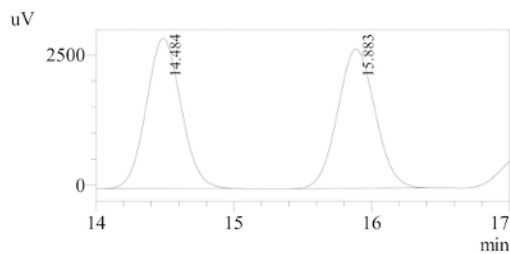
PeakTable

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 10.104    | 2.740   |
| 2     | 12.667    | 97.260  |
| Total |           | 100.000 |



**(R)-2-(4-(Methylthio)phenyl)-4-phenylcyclopent-1-en-1-yl(phenyl)methanone (4af):** Pale yellow oil (89.5 mg, 81%);  $R_f$  0.26 (hexane/EtOAc = 19/1);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.79-7.77 (m, 2H), 7.44-7.22 (m, 8H), 7.12-7.10 (m, 2H), 7.00-6.98 (m, 2H), 3.78 (quint,  $J = 8.2$  Hz, 1H), 3.46-3.31 (m, 2H), 3.19-3.06 (m, 2H), 2.38 (s, 3H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  197.8, 145.3, 144.0, 138.6, 136.5, 136.0, 132.9, 132.2, 129.3, 128.6, 128.4, 128.3, 126.9, 126.4, 125.8, 45.5, 45.3, 42.4, 15.4; **HRMS** (ESI) Calcd for  $\text{C}_{25}\text{H}_{23}\text{OS}$   $[\text{M} + \text{H}]^+$  371.1470, found 371.1465;  $[\alpha]^{25}_{\text{D}} = +21.5$  ( $c = 0.60$  in  $\text{CHCl}_3$ , 97:3 er sample).

HPLC analysis: Daicel CHIRALPAK ID; hexane:*i*-PrOH = 95:5; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 14.4 min (minor) and 15.8 min (major), 97:3 er.

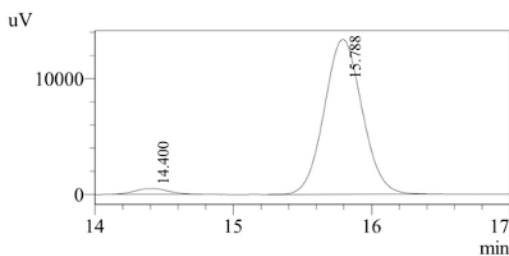


1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 14.484    | 50.114  |
| 2     | 15.883    | 49.886  |
| Total |           | 100.000 |

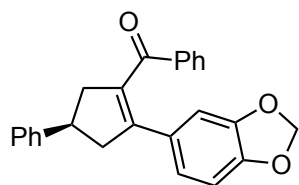


1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

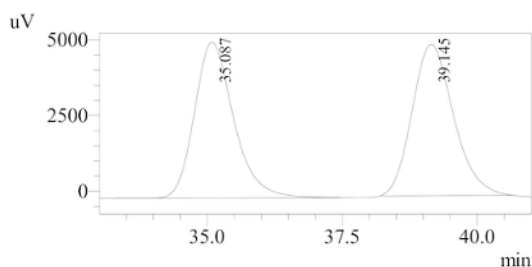
| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 14.400    | 3.207   |
| 2     | 15.788    | 96.793  |
| Total |           | 100.000 |



**(R)-2-(2-(Benzo[d][1,3]dioxol-5-yl)-4-phenylcyclopent-1-en-1-yl)(phenyl)methanone (4ag):**

Pale yellow oil (92.7 mg, 84%);  $R_f$  0.33 (hexane/EtOAc = 9/1);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.79-7.77 (m, 2H), 7.43-7.21 (m, 8H), 6.69-6.67 (m, 2H), 6.55-6.53 (m, 1H), 5.82 (s, 2H), 3.75 (quint,  $J = 8.2$  Hz, 1H), 3.42-3.29 (m, 2H), 3.15-3.04 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  197.7, 147.3, 147.3, 145.3, 144.5, 136.6, 135.4, 132.8, 129.7, 129.3, 128.6, 128.3, 126.9, 126.3, 122.1, 108.1, 107.9, 101.0, 45.8, 45.1, 42.3; **HRMS** (ESI) Calcd for  $\text{C}_{25}\text{H}_{21}\text{O}_3$   $[\text{M} + \text{H}]^+$  369.1491, found 369.1493;  $[\alpha]_D^{25} = +7.18$  ( $c = 3.39$  in  $\text{CHCl}_3$ , 95:5 er sample).

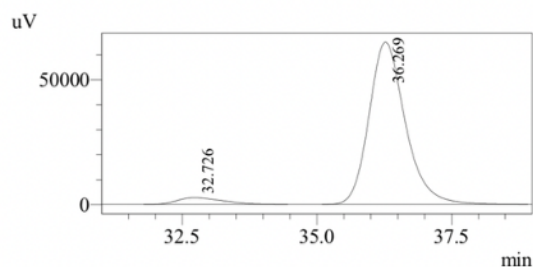
HPLC analysis: Daicel CHIRALPAK AS-H; hexane:*i*-PrOH = 98:2; detection wavelength = 254 nm; flow rate = 0.5 mL/min.  $t_R = 32.7$  min (minor) and 36.3 min (major), 95:5 er.



1 Det.A Ch1 / 254nm

PeakTable

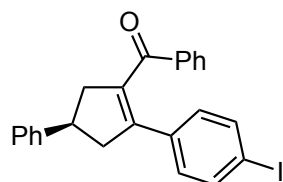
| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 35.087    | 49.416  |
| 2     | 39.145    | 50.584  |
| Total |           | 100.000 |



1 Det.A Ch1 / 254nm

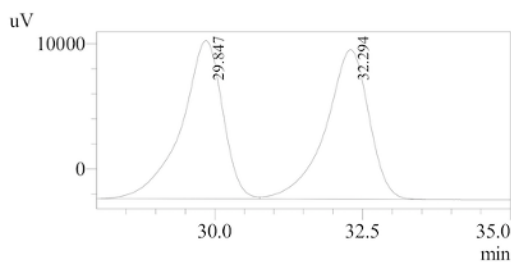
PeakTable

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 32.726    | 5.092   |
| 2     | 36.269    | 94.908  |
| Total |           | 100.000 |



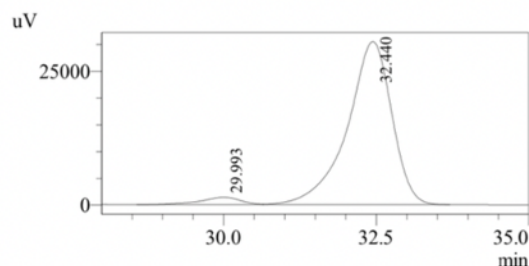
**(R)-2-(4-Iodophenyl)-4-phenylcyclopent-1-en-1-yl(phenyl)methanone (4ah):** Pale yellow oil (114.5 mg, 85%);  $R_f$  0.29 (hexane/EtOAc = 19/1);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.78-7.76 (m, 2H), 7.47-7.43 (m, 3H), 7.37-7.23 (m, 7H), 6.94-6.91 (m, 2H), 3.79 (quint,  $J = 8.2$  Hz, 1H), 3.44-3.30 (m, 2H), 3.19-3.05 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  197.5, 145.1, 143.2, 137.4, 137.2, 136.2, 135.0, 133.2, 129.5, 129.3, 128.6, 128.5, 126.9, 126.4, 93.7, 45.4, 42.4; **HRMS** (ESI) Calcd for  $\text{C}_{24}\text{H}_{20}\text{OI}$   $[\text{M} + \text{H}]^+$  451.0559, found 451.0557;  $[\alpha]^{25}_D = +7.0$  ( $c = 0.46$  in  $\text{CHCl}_3$ , 96:4 er sample).

HPLC analysis: Daicel CHIRALPAK OD; hexane:*i*-PrOH = 99:1; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 30.0 min (minor) and 32.4 min (major), 96:4 er.



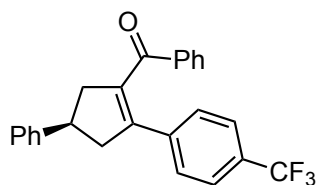
1 Det.A Ch1 / 254nm

| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 29.847    | 50.257  |
| 2                    | 32.294    | 49.743  |
| Total                |           | 100.000 |

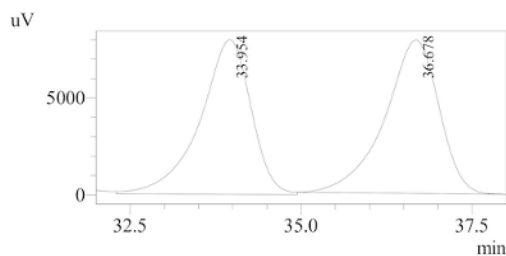


1 Det.A Ch1 / 254nm

| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 29.993    | 4.081   |
| 2                    | 32.440    | 95.919  |
| Total                |           | 100.000 |



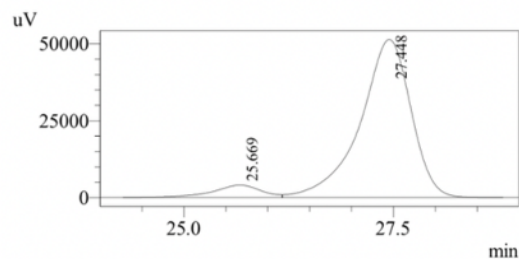
**(*R*)-Phenyl(4-phenyl-2-(4-(trifluoromethyl)phenyl)cyclopent-1-en-1-yl)methanone (4ai):** The reaction was performed at 0 °C for 48 h and then at 50 °C for 16 h. White solid (109.8 mg, 93%);  $R_f$  0.35 (hexane/toluene = 1/1); m.p. 82-84 °C;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.78-7.76 (m, 2H), 7.46-7.42 (m, 1H), 7.39-7.23 (m, 11H), 3.83 (quint,  $J$  = 8.2 Hz, 1H), 3.49-3.35 (m, 2H), 3.24-3.10 (m, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  197.3, 144.9, 142.8, 139.1, 138.8, 136.2, 133.3, 129.7 (q,  $^2J_{\text{C-F}}$  = 32.7 Hz), 129.2, 128.7, 128.5, 128.0, 126.9, 126.5, 125.1 (q,  $^3J_{\text{C-F}}$  = 3.8 Hz), 123.9 (q,  $^1J_{\text{C-F}}$  = 271.9 Hz), 45.5, 45.4, 42.5;  $^{19}\text{F}$  NMR (376 MHz,  $\text{CDCl}_3$ )  $\delta$  -63.2; **HRMS** (ESI) Calcd for  $\text{C}_{25}\text{H}_{20}\text{OF}_3$   $[\text{M} + \text{H}]^+$  393.1466, found 393.1461;  $[\alpha]^{25}_{\text{D}}$  = +13 ( $c$  = 0.29 in  $\text{CHCl}_3$ , 93:7 er sample). HPLC analysis: Daicel CHIRALPAK OD; hexane:*i*-PrOH = 99:1; detection wavelength = 254 nm; flow rate = 0.5 mL/min.  $t_R$  = 25.7 min (minor) and 27.4 min (major), 93:7 er.



1 Det.A Ch1 / 254nm

PeakTable

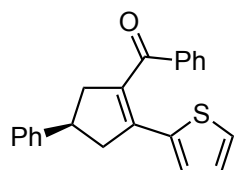
| Detector A Ch1 254nm |           |         |
|----------------------|-----------|---------|
| Peak#                | Ret. Time | Area %  |
| 1                    | 33.954    | 48.674  |
| 2                    | 36.678    | 51.326  |
| Total                |           | 100.000 |



1 Det.A Ch1 / 254nm

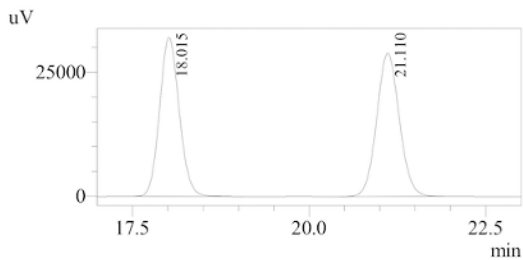
PeakTable

| Detector A Ch1 254nm |           |         |
|----------------------|-----------|---------|
| Peak#                | Ret. Time | Area %  |
| 1                    | 25.669    | 6.963   |
| 2                    | 27.448    | 93.037  |
| Total                |           | 100.000 |



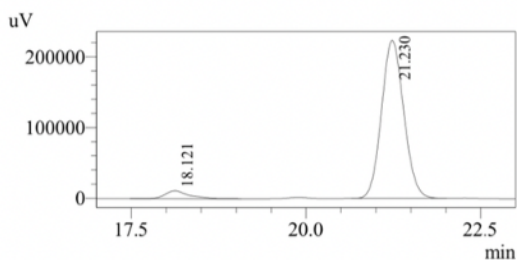
**(S)-Phenyl(4-phenyl-2-(thiophen-2-yl)cyclopent-1-en-1-yl)methanone (4aj):** Pale yellow oil (82.2 mg, 83%);  $R_f$  0.49 (hexane/EtOAc = 9/1);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.88-7.86 (m, 2H), 7.50-7.45 (m, 1H), 7.37-7.31 (m, 6H), 7.25-7.20 (m, 1H), 7.16-7.15 (m, 1H), 7.06-7.04 (m, 1H), 6.90-6.88 (m, 1H), 3.76 (quint,  $J = 8.2$  Hz, 1H), 3.46-3.39 (m, 1H), 3.33-3.27 (m, 1H), 3.18-3.02 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  197.9, 145.3, 138.1, 136.4, 136.4, 135.1, 133.2, 129.3, 128.6, 128.5, 127.2, 126.9, 126.3, 125.4, 124.1, 45.4, 45.3, 42.3; **HRMS** (ESI) Calcd for  $\text{C}_{22}\text{H}_{19}\text{OS}$   $[\text{M} + \text{H}]^+$  331.1157, found 331.1159;  $[\alpha]_D^{25} = +14$  ( $c = 0.61$  in  $\text{CHCl}_3$ , 95:5 er sample).

HPLC analysis: Daicel CHIRALPAK ID; hexane:*i*-PrOH = 90:10; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 18.1 min (minor) and 21.3 min (major), 95:5 er.



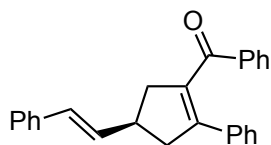
1 Det.A Ch1 / 254nm

| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 18.015    | 49.026  |
| 2                    | 21.110    | 50.974  |
| Total                |           | 100.000 |



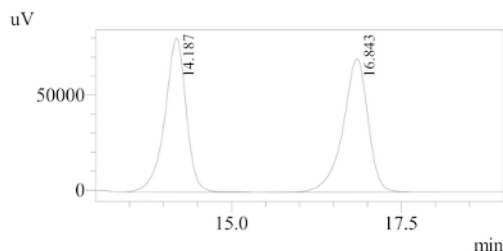
1 Det.A Ch1 / 254nm

| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 18.121    | 4.941   |
| 2                    | 21.230    | 95.059  |
| Total                |           | 100.000 |



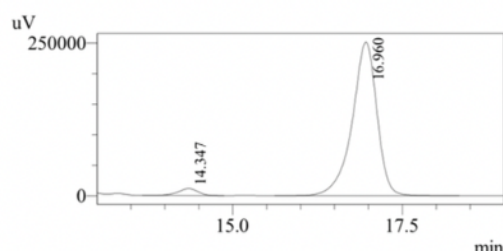
**(*R,E*)-Phenyl(2-phenyl-4-styrylcyclopent-1-en-1-yl)methanone (4ak):** Yellow oil (79.0 mg, 75%);  $R_f$  0.32 (hexane/EtOAc = 19/1);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.77-7.75 (m, 2H), 7.39-7.07 (m, 13H), 6.52-6.38 (m, 2H), 3.41-3.31 (m, 1H), 3.27-3.12 (m, 2H), 3.01-2.87 (m, 2H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  197.6, 145.0, 137.3, 136.5, 136.3, 135.6, 133.3, 132.7, 129.4, 129.3, 128.5, 128.2, 128.0, 127.8, 127.8, 127.1, 126.0, 44.1, 43.7, 40.7; HRMS (ESI) Calcd for  $\text{C}_{26}\text{H}_{23}\text{O}$   $[\text{M} + \text{H}]^+$  351.1749, found 351.1744;  $[\alpha]^{25}_D = +14.7$  ( $c = 0.985$  in  $\text{CHCl}_3$ , 97:3 er sample).

HPLC analysis: Daicel CHIRALPAK ID; hexane:*i*-PrOH = 95:5; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 14.3 min (major) and 17.0 min (minor), 97:3 er.



1 Det.A Ch1 / 254nm

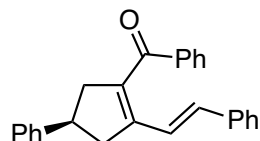
| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 14.187    | 49.548  |
| 2                    | 16.843    | 50.452  |
| Total                |           | 100.000 |



1 Det.A Ch1 / 254nm

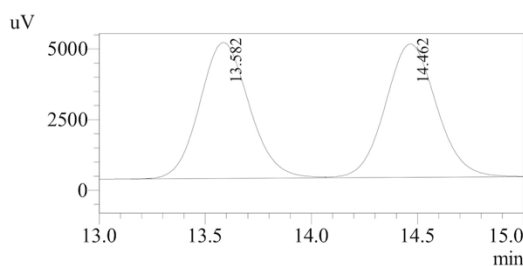
| PeakTable            |           |         |
|----------------------|-----------|---------|
| Detector A Ch1 254nm |           |         |
| Peak#                | Ret. Time | Area %  |
| 1                    | 14.347    | 3.486   |
| 2                    | 16.960    | 96.514  |
| Total                |           | 100.000 |





**(*R,E*)-Phenyl(4-phenyl-2-styrylcyclopent-1-en-1-yl)methanone (4a):** Pale yellow oil (63.1 mg, 60%);  $R_f$  0.28 (hexane/EtOAc = 19/1);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.83-7.81 (m, 2H), 7.57-7.54 (m, 1H), 7.47-7.44 (m, 2H), 7.34-7.33 (m, 4H), 7.25-7.18 (m, 6H), 7.02 (d,  $J = 16.1$  Hz, 1H), 6.66 (d,  $J = 16.1$  Hz, 1H), 3.65 (quint,  $J = 8.2$  Hz, 1H), 3.38-3.23 (m, 2H), 3.11-2.96 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  195.9, 147.3, 145.3, 139.3, 138.1, 136.7, 134.1, 132.6, 129.1, 128.7, 128.7, 128.6, 128.4, 127.0, 126.9, 126.4, 123.3, 44.3, 42.1, 42.0; **HRMS** (ESI) Calcd for  $\text{C}_{26}\text{H}_{23}\text{O}$   $[\text{M} + \text{H}]^+$  351.1749, found 351.1748;  $[\alpha]_D^{25} = +79.9$  ( $c = 2.10$  in  $\text{CHCl}_3$ , 96:4 er sample).

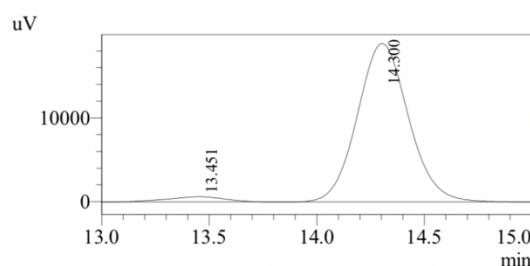
HPLC analysis: Daicel CHIRALPAK IA; hexane:*i*-PrOH = 95:5; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 13.5 min (minor) and 14.3 min (major), 96:4 er.



1 Det.A Ch1 / 254nm

PeakTable

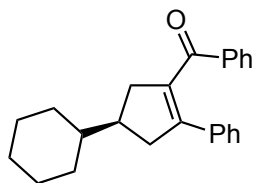
| Detector A Ch1 254nm |           |         |
|----------------------|-----------|---------|
| Peak#                | Ret. Time | Area %  |
| 1                    | 13.582    | 49.718  |
| 2                    | 14.462    | 50.282  |
| Total                |           | 100.000 |



1 Det.A Ch1 / 254nm

PeakTable

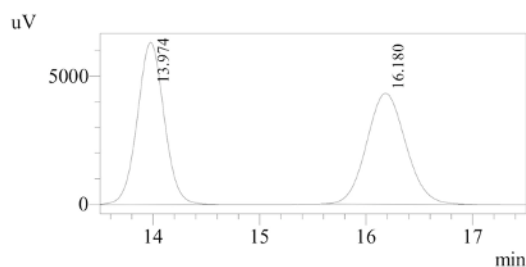
| Detector A Ch1 254nm |           |         |
|----------------------|-----------|---------|
| Peak#                | Ret. Time | Area %  |
| 1                    | 13.451    | 3.734   |
| 2                    | 14.300    | 96.266  |
| Total                |           | 100.000 |



**(*R*)-(4-Cyclohexyl-2-phenylcyclopent-1-en-1-yl)(phenyl)methanone (4am):** The reaction was performed using **L3** as the ligand in DMSO at 40 °C for 48 h and then at 100 °C for 6 h. Colorless oil (82.4 mg, 83%);  $R_f$  0.39 (hexane/EtOAc = 19/1);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.73-7.71 (m, 2H), 7.37-7.33 (m, 1H), 7.24-7.20 (m, 2H), 7.14-7.05 (m, 5H), 3.07-2.91 (m, 2H), 2.79-2.65 (m, 2H), 2.29 (sext,  $J = 8.7$  Hz, 1H), 1.84-1.66 (m, 5H), 1.41-1.13 (m, 4H), 1.07-0.96 (m, 2H);  $^{13}\text{C NMR}$

**NMR** (100 MHz, CDCl<sub>3</sub>)  $\delta$  198.1, 145.7, 136.9, 136.6, 136.1, 132.6, 129.3, 128.1, 127.9, 127.8, 127.6, 43.6, 43.2, 42.0, 41.6, 31.5, 31.3, 26.5, 26.3, 26.2; **HRMS** (ESI) Calcd for C<sub>24</sub>H<sub>27</sub>O [M + H]<sup>+</sup> 331.2062, found 331.2066;  $[\alpha]^{25}_D = +13.2$  ( $c = 2.17$  in CHCl<sub>3</sub>, 78:22 er sample). The absolute configuration was deduced as (*R*) based on the result that **L3** gave (*R*)-**4aa** as the major enantiomer in the reaction between **1a** and **2a**.

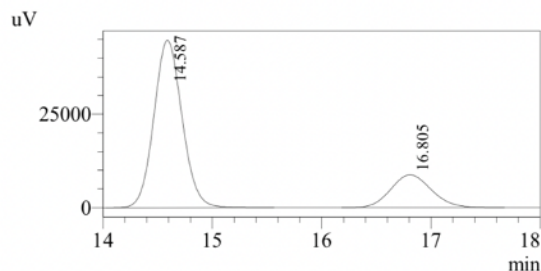
HPLC analysis: Daicel CHIRALPAK IA; hexane:*i*-PrOH = 99:1; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 14.6 min (major) and 16.8 min (minor), 78:22 er.



1 Det.A Ch1 / 254nm

PeakTable

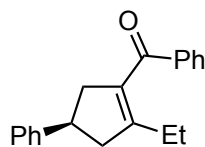
| Detector A Ch1 254nm |           |         |
|----------------------|-----------|---------|
| Peak#                | Ret. Time | Area %  |
| 1                    | 13.974    | 50.111  |
| 2                    | 16.180    | 49.889  |
| Total                |           | 100.000 |



1 Det.A Ch1 / 254nm

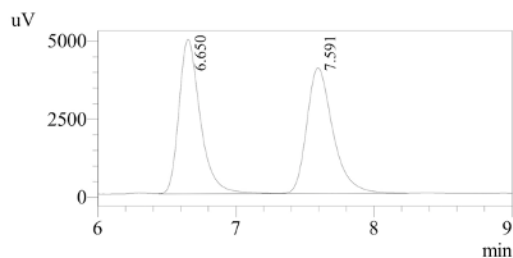
PeakTable

| Detector A Ch1 254nm |           |         |
|----------------------|-----------|---------|
| Peak#                | Ret. Time | Area %  |
| 1                    | 14.587    | 77.924  |
| 2                    | 16.805    | 22.076  |
| Total                |           | 100.000 |



**(*R*)-(2-Ethyl-4-phenylcyclopent-1-en-1-yl)(phenyl)methanone (4an):** The reaction was performed using **L3** as the ligand at 30 °C for 48 h and then at 100 °C for 6 h. Colorless oil (47.9 mg, 58%);  $R_f$  0.40 (hexane/EtOAc = 19/1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.82-7.79 (m, 2H), 7.56-7.52 (m, 1H), 7.46-7.43 (m, 2H), 7.35-7.29 (m, 4H), 7.24-7.19 (m, 1H), 3.58 (quint,  $J = 8.2$  Hz, 1H), 3.19-3.13 (m, 1H), 3.04-2.91 (m, 2H), 2.76-2.69 (m, 1H), 2.15 (q,  $J = 7.5$  Hz, 2H), 0.99 (t,  $J = 7.6$  Hz, 3H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>)  $\delta$  196.8, 153.3, 145.6, 139.0, 134.5, 132.5, 128.8, 128.5, 128.4, 126.9, 126.2, 44.6, 43.6, 42.2, 23.5, 12.4; **HRMS** (ESI) Calcd for C<sub>20</sub>H<sub>21</sub>O [M + H]<sup>+</sup> 277.1592, found 277.1590;  $[\alpha]^{25}_D = -7.17$  ( $c = 0.600$  in CHCl<sub>3</sub>, 79:21 er sample).

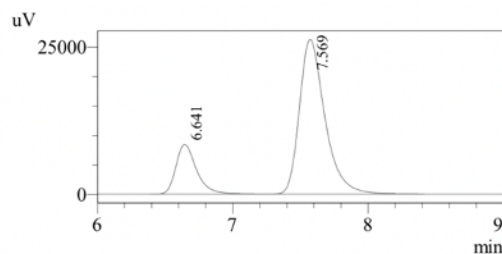
HPLC analysis: Daicel CHIRALPAK AS-H; hexane:*i*-PrOH = 99:1; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 6.6 min (minor) and 7.6 min (major), 79:21 er.



1 Det.A Ch1 / 254nm

PeakTable

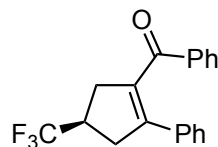
| Detector A Ch1 254nm |           |         |
|----------------------|-----------|---------|
| Peak#                | Ret. Time | Area %  |
| 1                    | 6.650     | 50.079  |
| 2                    | 7.591     | 49.921  |
| Total                |           | 100.000 |



1 Det.A Ch1 / 254nm

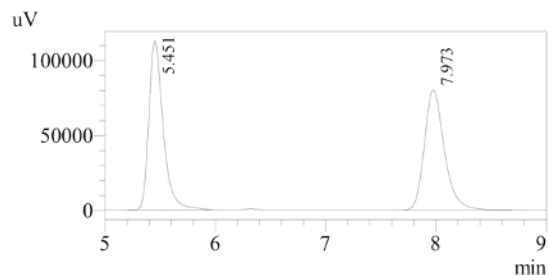
PeakTable

| Detector A Ch1 254nm |           |         |
|----------------------|-----------|---------|
| Peak#                | Ret. Time | Area %  |
| 1                    | 6.641     | 20.669  |
| 2                    | 7.569     | 79.331  |
| Total                |           | 100.000 |



**Phenyl(2-phenyl-4-(trifluoromethyl)cyclopent-1-en-1-yl)methanone (4ao):** The reaction was performed using **L3** as the ligand at 30 °C for 24 h and then at 100 °C for 6 h. Pale yellow oil (36.8 mg, 39%);  $R_f$  0.27 (hexane/EtOAc = 19/1);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.72-7.70 (m, 2H), 7.40-7.37 (m, 1H), 7.27-7.23 (m, 2H), 7.14-7.09 (m, 5H), 3.28-3.08 (m, 5H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  196.6, 143.7, 136.1, 134.8, 134.6, 133.0, 129.3, 128.4, 128.3, 128.1, 127.9 (q,  $^1J_{\text{C-F}} = 277.1$  Hz), 127.9, 39.5 (q,  $^2J_{\text{C-F}} = 28.3$  Hz), 37.7 (q,  $^3J_{\text{C-F}} = 2.7$  Hz), 37.1 (q,  $^3J_{\text{C-F}} = 2.6$  Hz);  $^{19}\text{F}$  NMR (376 MHz,  $\text{CDCl}_3$ )  $\delta$  -73.2; **HRMS** (ESI) Calcd for  $\text{C}_{19}\text{H}_{16}\text{OF}_3$   $[\text{M} + \text{H}]^+$  317.1153, found 317.1154;  $[\alpha]_D^{25} = +2.2$  ( $c = 0.49$  in  $\text{CHCl}_3$ , 77:23 er sample).

HPLC analysis: Daicel CHIRALPAK AS-H; hexane:*i*-PrOH = 95:5; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 5.4 min (minor) and 8.0 min (major), 77:23 er.

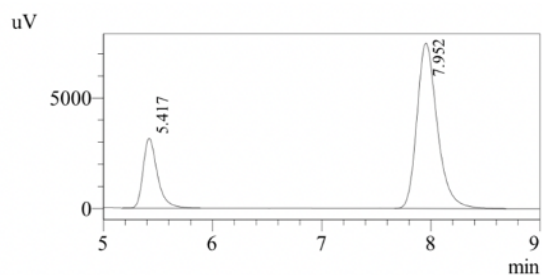


1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 5.451     | 50.299  |
| 2     | 7.973     | 49.701  |
| Total |           | 100.000 |

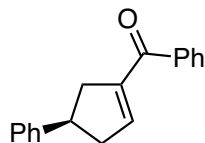


1 Det.A Ch1 / 254nm

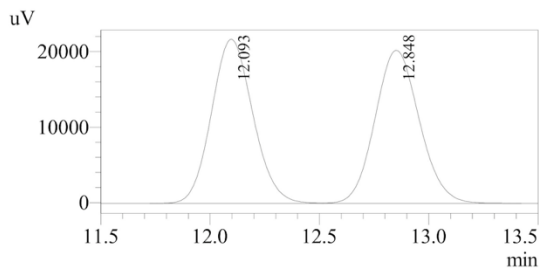
PeakTable

Detector A Ch1 254nm

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 5.417     | 22.781  |
| 2     | 7.952     | 77.219  |
| Total |           | 100.000 |



**(R)-Phenyl(4-phenylcyclopent-1-en-1-yl)methanone (4ap):** The reaction was performed at 60 °C for 12 h. Pale yellow oil (25.5 mg, 34%);  $R_f$  0.16 (hexane/EtOAc = 19/1);  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.79-7.76 (m, 2H), 7.56-7.52 (m, 1H), 7.47-7.43 (m, 2H), 7.34-7.28 (m, 4H), 7.25-7.20 (m, 1H), 6.58-6.56 (m, 1H), 3.66 (quint,  $J$  = 8.3 Hz, 1H), 3.27-3.20 (m, 1H), 3.15-3.07 (m, 1H), 2.96-2.88 (m, 1H), 2.78-2.70 (m, 1H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  193.8, 145.5, 145.1, 143.5, 138.7, 131.9, 128.9, 128.5, 128.2, 126.9, 126.2, 42.9, 42.5, 39.8; **HRMS** (ESI) Calcd for  $\text{C}_{18}\text{H}_{17}\text{O}$   $[\text{M} + \text{H}]^+$  249.1279, found 249.1278;  $[\alpha]_D^{25} = -3.07$  ( $c$  = 1.76 in  $\text{CHCl}_3$ , 62:38 er sample). HPLC analysis: Daicel CHIRALPAK OJ-H; hexane:*i*-PrOH = 95:5; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 12.1 min (major) and 12.8 min (minor), 62:38 er.

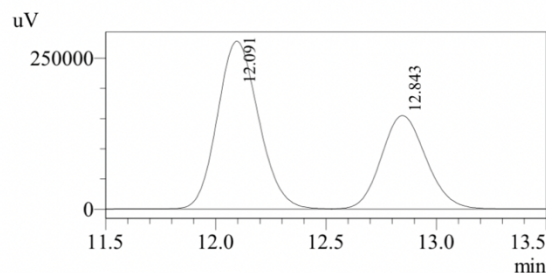


1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 12.093    | 50.056  |
| 2     | 12.848    | 49.944  |
| Total |           | 100.000 |



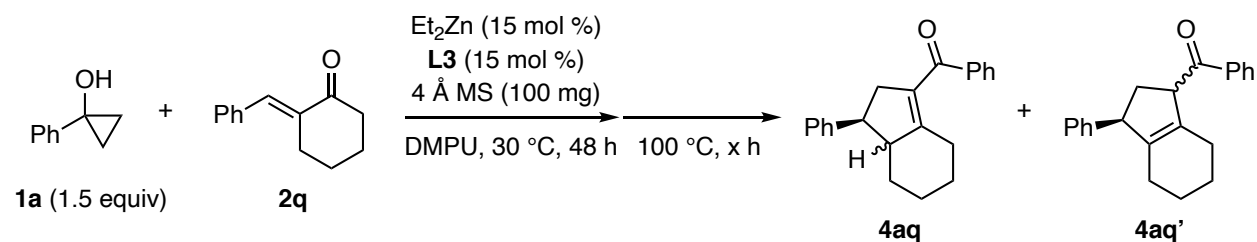
1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

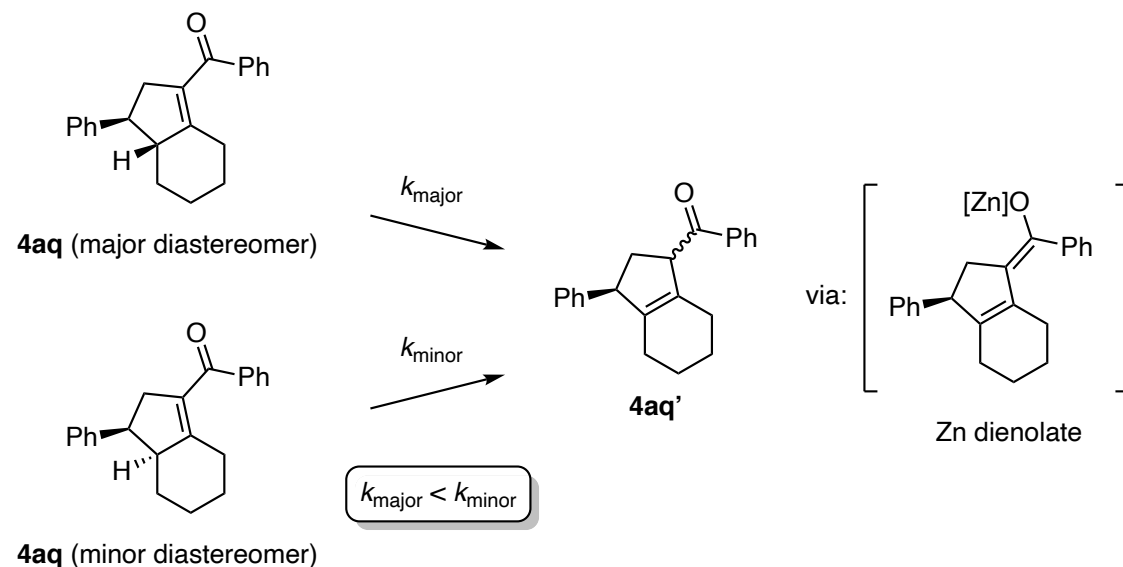
| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 12.091    | 62.305  |
| 2     | 12.843    | 37.695  |
| Total |           | 100.000 |

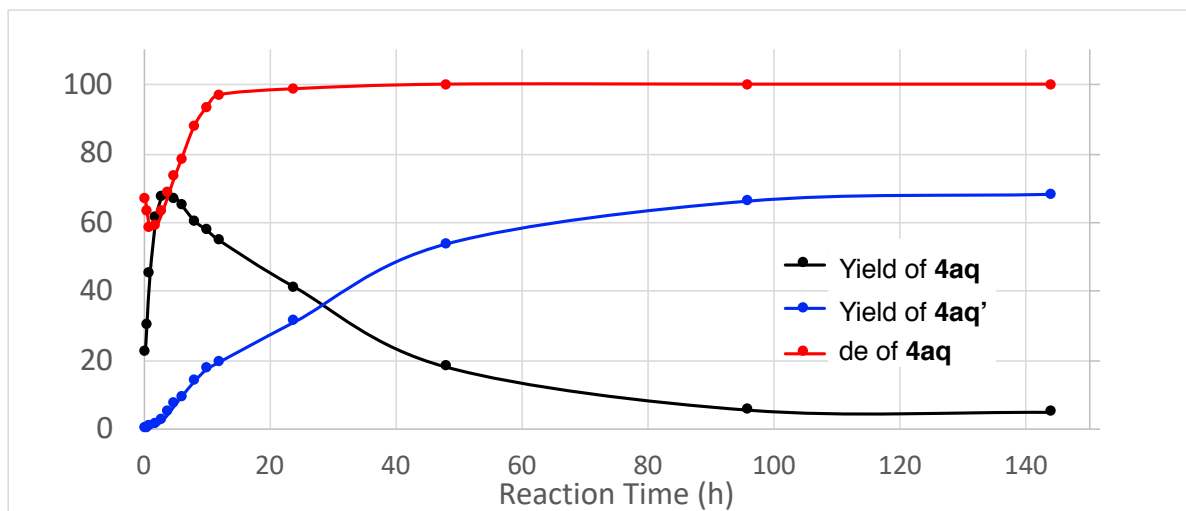
## The reaction between **1a** and **2q**



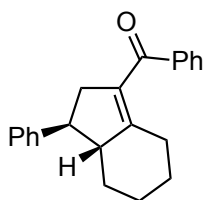
The reaction was performed using **L3** as the ligand at 30 °C for 48 h and then at 100 °C. The time profile of the aldol condensation step showed that **4aq** gradually transformed to **4aq'**, presumably via zinc dienolate intermediate (Scheme S2 and Figure S3). As the minor diastereomer of **4aq** had a higher rate of conversion than the major diastereomer, the de of **4aq** increased as the reaction proceeded. When the reaction was quenched after 3 h at 100 °C, **4aq** was obtained as a colorless oil (57.0 mg, 63%, 80:20 dr; The relative stereochemistry of the major diastereomer was determined by X-ray crystallographic analysis of its anilide derivative **5aq** (vide infra)). When the reaction was quenched after 6 h at 100 °C, **4aq** was obtained in 51% yield (46.6 mg, as a single diastereomer). When the reaction was quenched after 144 h at 100 °C, **4aq'** was obtained as a colorless oil (0.6 mmol scale, 120.5 mg, 66%, 50:50 dr).

## Scheme S2. Conversion of **4aq** to **4aq'**

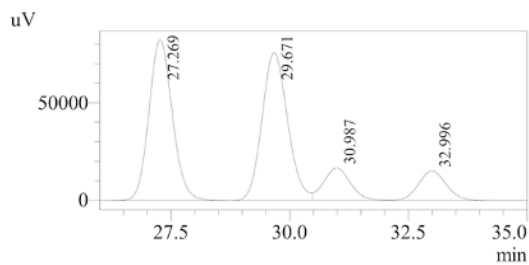




**Figure S3.** Time profile of the aldol condensation–isomerization step at 100 °C.



**Phenyl(1-phenyl-2,4,5,6,7,7a-hexahydro-1H-inden-3-yl)methanone (4aq):**  $R_f$  0.34 (hexane/EtOAc = 19/1);  $^1\text{H NMR}$  (major diastereomer, 500 MHz,  $\text{CDCl}_3$ )  $\delta$  7.83-7.81 (m, 2H), 7.55-7.52 (m, 1H), 7.47-7.44 (m, 2H), 7.34-7.29 (m, 4H), 7.24-7.20 (m, 1H), 3.17-3.12 (m, 1H), 3.07 (q,  $J$  = 8.4 Hz, 1H), 2.99-2.93 (m, 1H), 2.73-2.68 (m, 1H), 2.47-2.44 (m, 1H), 2.13-2.10 (m, 1H), 1.84-1.77 (m, 2H), 1.72-1.69 (m, 1H), 1.36-1.19 (m, 3H);  $^{13}\text{C NMR}$  (major diastereomer, 126 MHz,  $\text{CDCl}_3$ )  $\delta$  196.5, 153.5, 144.7, 139.2, 132.4, 131.8, 128.8, 128.5, 128.4, 127.3, 126.2, 56.3, 50.6, 42.6, 34.4, 28.6, 26.1, 25.2; **HRMS** (ESI) Calcd for  $\text{C}_{22}\text{H}_{23}\text{O}$   $[\text{M} + \text{H}]^+$  303.1749, found 303.1746;  $[\alpha]^{25}_{\text{D}} = -14.0$  ( $c$  = 2.36 in  $\text{CHCl}_3$ , 80:20 dr, 84:16 er (major), 70:30 er (minor) sample). HPLC analysis (the sample obtained after 3 h at 100 °C, 80:20 dr): Daicel CHIRALPAK ID; hexane:*i*-PrOH = 80:20; detection wavelength = 254 nm; flow rate = 0.2 mL/min. Major diastereomer  $t_R$  = 25.6 min (minor) and 27.7 min (major), 84:16 er. Minor diastereomer  $t_R$  = 29.0 min (minor) and 30.8 min (major), 70:30 er.

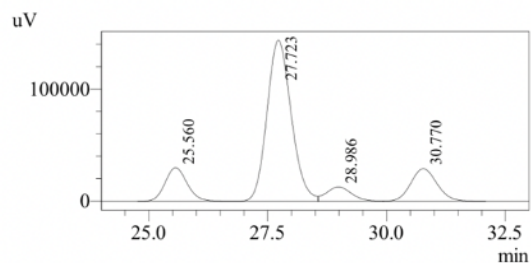


1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 27.269    | 40.587  |
| 2     | 29.671    | 40.811  |
| 3     | 30.987    | 9.573   |
| 4     | 32.996    | 9.029   |
| Total |           | 100.000 |



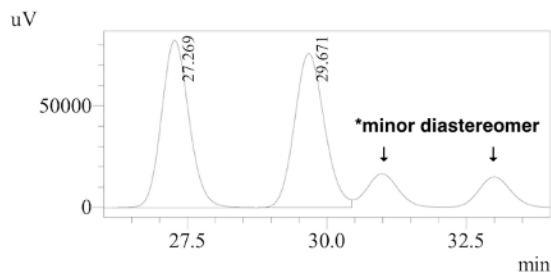
1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 25.560    | 12.588  |
| 2     | 27.723    | 66.789  |
| 3     | 28.986    | 6.131   |
| 4     | 30.770    | 14.492  |
| Total |           | 100.000 |

HPLC analysis (the sample obtained after 6 h at 100 °C, single diastereomer): Daicel CHIRALPAK ID; hexane:*i*-PrOH = 80:20; detection wavelength = 254 nm; flow rate = 0.2 mL/min.  $t_R$  = 27.3 min (minor) and 29.7 min (major), 82:18 er.

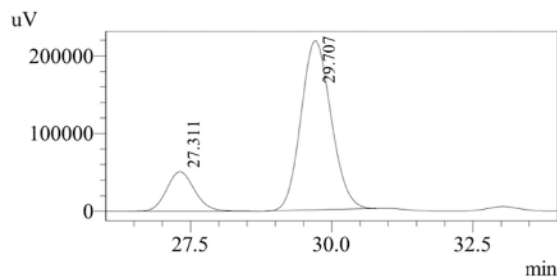


1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 27.269    | 50.000  |
| 2     | 29.671    | 50.000  |
| Total |           | 100.000 |

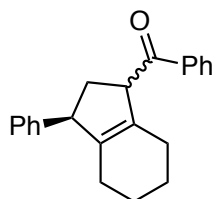


1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

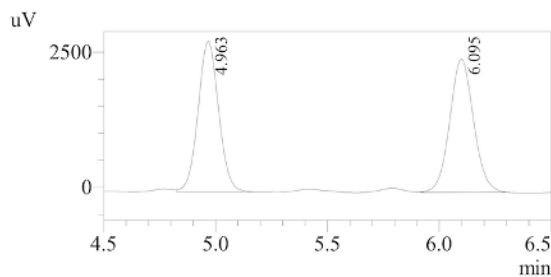
| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 27.311    | 17.733  |
| 2     | 29.707    | 82.267  |
| Total |           | 100.000 |



**Phenyl(3-phenyl-2,3,4,5,6,7-hexahydro-1H-inden-1-yl)methanone (4aq')**: Colorless oil (120.5 mg, 66%, 50:50 dr);  $R_f$  0.49, 0.44 (hexane/EtOAc = 19/1);  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ) the diastereomer of  $R_f$  0.49:  $\delta$  8.02-8.00 (m, 2H), 7.57-7.52 (m, 1H), 7.48-7.44 (m, 2H), 7.30-7.22 (m, 4H), 7.19-7.14 (m, 1H), 4.51-4.47 (m, 1H), 3.76 (brs, 1H), 2.80 (dt,  $J$  = 13.4, 9.2 Hz, 1H), 2.17-

2.13 (m, 1H), 1.98 (dt,  $J = 13.6, 6.8$  Hz, 1H), 1.89-1.85 (m, 1H), 1.75 (brs, 2H), 1.72-1.54 (m, 4H); the diastereomer of  $R_f$  0.44:  $\delta$  8.02-7.99 (m, 2H), 7.58-7.54 (m, 1H), 7.48-7.44 (m, 2H), 7.33-7.29 (m, 2H), 7.23-7.19 (m, 1H), 7.17-7.15 (m, 2H), 4.61-4.58 (m, 1H), 3.89 (brs, 1H), 2.60 (ddd,  $J = 13.3, 8.8, 5.1$  Hz, 1H), 2.15 (ddd,  $J = 13.3, 9.2, 5.3$  Hz, 1H), 2.00 (brs, 2H), 1.89-1.85 (m, 1H), 1.77-1.55 (m, 5H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ ) the diastereomer of  $R_f$  0.49:  $\delta$  202.3, 145.0, 140.7, 137.3, 134.9, 132.9, 128.5, 128.5, 128.4, 128.0, 126.1, 55.0, 54.3, 38.0, 25.2, 24.5, 22.8, 22.6; the diastereomer of  $R_f$  0.44:  $\delta$  202.5, 145.3, 141.2, 137.3, 135.0, 132.9, 128.6, 128.5, 127.6, 126.2, 55.1, 53.9, 38.5, 25.4, 24.5, 22.9, 22.6; **HRMS** (ESI) Calcd for  $\text{C}_{22}\text{H}_{23}\text{O}$   $[\text{M} + \text{H}]^+$  303.1749, found 303.1739;  $[\alpha]^{25}_{\text{D}} = -56.9$  ( $c = 1.80$  in  $\text{CHCl}_3$ , 80:20 er sample of the diastereomer of  $R_f$  0.49), -111 ( $c = 1.52$  in  $\text{CHCl}_3$ , 80:20 er sample of the diastereomer of  $R_f$  0.44).

HPLC analysis (the diastereomer of  $R_f$  0.49): Daicel CHIRALPAK IA; hexane:*i*-PrOH = 90:10; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R = 5.0$  min (minor) and 6.1 min (major), 80:20 er.

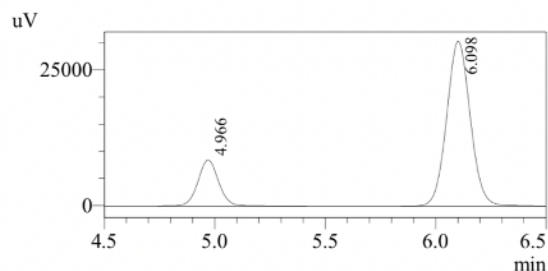


1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 4.963     | 49.289  |
| 2     | 6.095     | 50.711  |
| Total |           | 100.000 |



1 Det.A Ch1 / 254nm

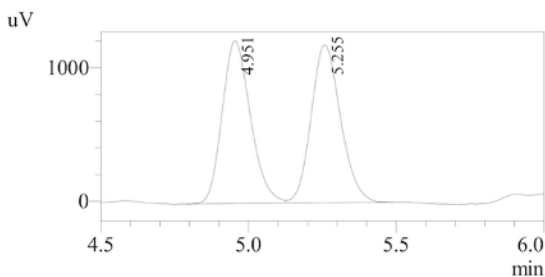
PeakTable

Detector A Ch1 254nm

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 4.966     | 19.560  |
| 2     | 6.098     | 80.440  |
| Total |           | 100.000 |

HPLC analysis (the diastereomer of  $R_f$  0.44): Daicel CHIRALPAK ID; hexane:*i*-PrOH = 90:10; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R = 5.0$  min (minor) and 5.3 min (major), 80:20 er.



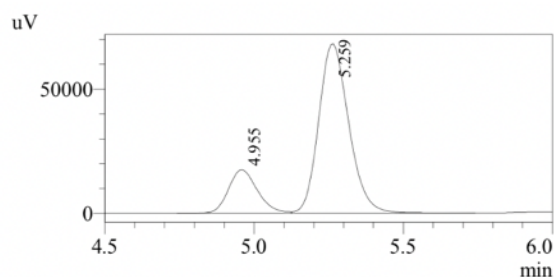


1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 4.951     | 49.777  |
| 2     | 5.255     | 50.223  |
| Total |           | 100.000 |

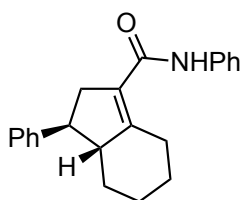


1 Det.A Ch1 / 254nm

PeakTable

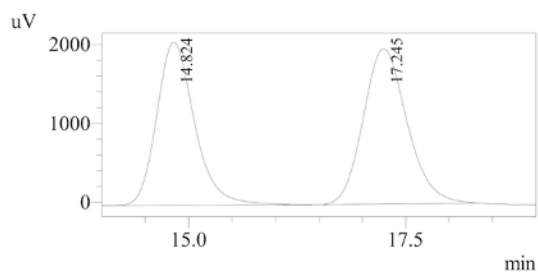
Detector A Ch1 254nm

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 4.955     | 19.626  |
| 2     | 5.259     | 80.374  |
| Total |           | 100.000 |



**(1S,7aR)-N,1-Diphenyl-2,4,5,6,7,7a-hexahydro-1H-indene-3-carboxamide (5aq):** Prepared according to the synthetic procedure for **5** (vide infra). Pale yellow solid (59.2 mg, 62%);  $R_f$  0.18 (hexane/EtOAc = 19/1); m.p. 148-150 °C;  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.56-7.54 (m, 2H), 7.33-7.20 (m, 8H), 7.10-7.06 (m, 1H), 3.63-3.60 (m, 1H), 3.09-2.97 (m, 2H), 2.83-2.76 (m, 1H), 2.67-2.61 (m, 1H), 2.07-1.97 (m, 2H), 1.88-1.76 (m, 2H), 1.38-1.13 (m, 3H);  $^{13}\text{C}$  NMR (100 MHz,  $\text{CDCl}_3$ )  $\delta$  164.6, 155.5, 144.5, 137.9, 128.9, 128.5, 127.3, 126.3, 125.5, 124.1, 120.0, 55.9, 50.4, 41.3, 34.1, 27.6, 26.0, 25.2; HRMS (ESI) Calcd for  $\text{C}_{22}\text{H}_{24}\text{NO}$   $[\text{M} + \text{H}]^+$  318.1858, found 318.1857;  $[\alpha]_D^{25} = -20.8$  ( $c = 2.86$  in  $\text{CHCl}_3$ , 84:16 er sample).

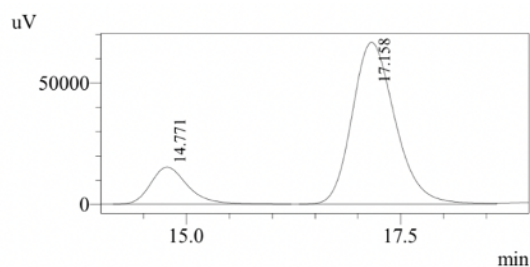
HPLC analysis: Daicel CHIRALPAK ID; hexane:*i*-PrOH = 80:20; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R$  = 14.8 min (minor) and 17.2min (major), 84:16 er.



1 Det.A Ch1 / 254nm

PeakTable

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 14.824    | 47.583  |
| 2     | 17.245    | 52.417  |
| Total |           | 100.000 |

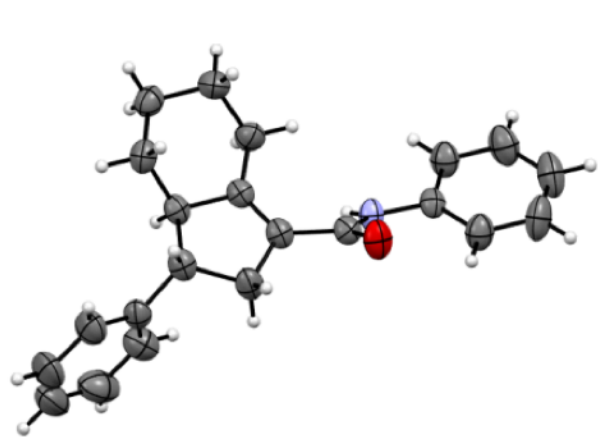


1 Det.A Ch1 / 254nm

PeakTable

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 14.771    | 16.495  |
| 2     | 17.158    | 83.505  |
| Total |           | 100.000 |

Recrystallization of **5aq** from THF/pentane afforded single crystals suitable for X-ray diffraction analysis, which unambiguously confirmed the molecular structure of **5aq** (major diastereomer) and its relative configuration (Figure S4).<sup>31</sup>

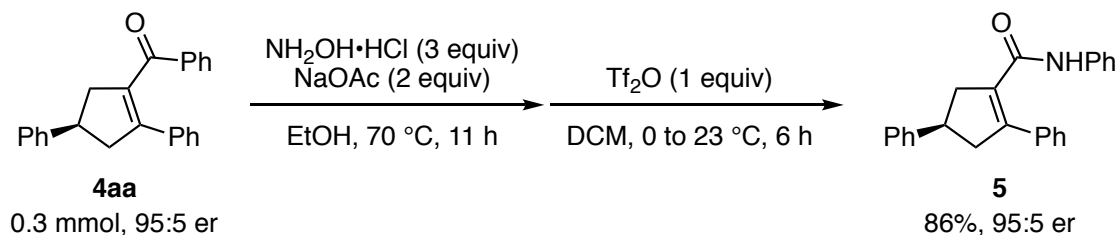


**Figure S4.** ORTEP drawing of **5aq** (thermal ellipsoids set at 50% probability; CCDC 2056576).

**Table S4.** Crystal data and structure refinement for **5aq**

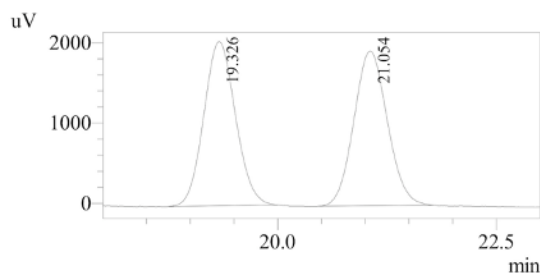
|                                   |  |                              |
|-----------------------------------|--|------------------------------|
| Identification code               | yoshi107s  |                              |
| Chemical formula                  | C <sub>22</sub> H <sub>23</sub> NO                                     |                              |
| Formula weight                    | 317.41 g/mol   |                              |
| Temperature                       | 296(2) K   |                              |
| Wavelength                        | 1.54178 Å  |                              |
| Crystal size                      | 0.010 x 0.040 x 0.240 mm   |                              |
| Crystal habit                     | colorless needle   |                              |
| Crystal system                    | monoclinic   |                              |
| Space group                       | C 1 2 1  |                              |
| Unit cell dimensions              | a = 24.6438(8) Å   | $\alpha = 90^\circ$          |
|                                   | b = 5.1210(2) Å  | $\beta = 123.8273(17)^\circ$ |
|                                   | c = 16.5702(6) Å   | $\gamma = 90^\circ$          |
| Volume                            | 1737.18(11) Å <sup>3</sup>   |                              |
| Z                                 | 4  |                              |
| Density (calculated)              | 1.214 g/cm <sup>3</sup>  |                              |
| Absorption coefficient            | 0.569 mm <sup>-1</sup>   |                              |
| F(000)                            | 680  |                              |
| Theta range for data collection   | 3.21 to 68.04°   |                              |
| Index ranges                      | -29<= <i>h</i> <=29, -5<= <i>k</i> <=5, -19<= <i>l</i> <=19            |                              |
| Reflections collected             | 24610  |                              |
| Independent reflections           | 3078 [R(int) = 0.0648]   |                              |
| Coverage independent reflections  | 98.8%  |                              |
| Absorption correction             | Multi-Scan   |                              |
| Max. and min. transmission        | 0.9940 and 0.8760  |                              |
| Structure solution technique      | direct methods   |                              |
| Structure solution program        | XT, VERSION 2014/5   |                              |
| Refinement method                 | Full-matrix least-squares on F <sup>2</sup>                            |                              |
| Refinement program                | SHELXL-2018/3 (Sheldrick, 2018)  |                              |
| Function minimized                | $\Sigma w(F_o^2 - F_c^2)^2$  |                              |
| Data / restraints / parameters    | 3078 / 1 / 193   |                              |
| Goodness-of-fit on F <sup>2</sup> | 1.045  |                              |
| Final R indices                   | 2832 data; I>2σ(I)   | R1 = 0.0429, wR2 = 0.1154    |
|                                   | all data   | R1 = 0.0470, wR2 = 0.1203    |
| Weighting scheme                  | $w=1/[\sigma^2(F_o^2)+(0.0783P)^2+0.3359P]$ where $P=(F_o^2+2F_c^2)/3$ |                              |
| Absolute structure parameter      | 0.0(2)   |                              |
| Largest diff. peak and hole       | 0.183 and -0.165 eÅ <sup>-3</sup>                                      |                              |
| R.M.S. deviation from mean        | 0.036 eÅ <sup>-3</sup>   |                              |

## Product Transformations



**(R)-N,2,4-Triphenylcyclopent-1-ene-1-carboxamide (5):** A 4-mL vial equipped with a magnetic stir bar was charged with **4aa** (97.3 mg, 0.30 mmol, 95:5 er), hydroxylammonium chloride (62.6 mg, 0.90 mmol, 3 equiv) and sodium acetate (49.2 mg, 0.6 mmol, 2 equiv), followed by the addition of ethanol (0.9 mL). The mixture was stirred at 70 °C for 11 h. After the reaction mixture was concentrated under the reduced pressure, the residue was diluted with  $\text{H}_2\text{O}$  (1.5 mL) and  $\text{CH}_2\text{Cl}_2$  (2 mL). The layers were separated, and the aqueous layer was extracted with  $\text{CH}_2\text{Cl}_2$  ( $3 \times 2$  mL). The combined organic layers were dried  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated to give the crude oxime intermediate as a colorless oil. The crude oxime product was dissolved in anhydrous  $\text{CH}_2\text{Cl}_2$  (4 mL) and the solution was stirred at 0 °C. To the mixture was added triflic anhydride solution (84.3 mg in  $\text{CH}_2\text{Cl}_2$  (2 mL)) dropwise over 15 min, and the resulting mixture was stirred and allowed to warm to room temperature. After 6 h, the reaction mixture was washed with sat.  $\text{NaHCO}_3$  aq (2 mL) and the organic layers were dried over  $\text{Na}_2\text{SO}_4$ , filtered, and concentrated. The crude product was purified by flash column chromatography to afford the product as a pale yellow solid (88.0 mg, 86%);  $R_f$  0.23 (hexane/EtOAc = 9/1); m.p. 126-127 °C;  $^1\text{H NMR}$  (400 MHz,  $\text{CDCl}_3$ )  $\delta$  7.41-7.32 (m, 9H), 7.24-7.15 (m, 6H), 7.04-7.00 (m, 1H), 3.66 (quint,  $J = 8.2$  Hz, 1H), 3.43-3.28 (m, 2H), 3.12-3.02 (m, 2H);  $^{13}\text{C NMR}$  (100 MHz,  $\text{CDCl}_3$ )  $\delta$  164.3, 145.3, 145.2, 137.7, 135.9, 133.3, 128.9, 128.8, 128.6, 128.5, 127.8, 126.8, 126.3, 124.0, 119.4, 47.8, 43.0, 41.6; **HRMS** (ESI) Calcd for  $\text{C}_{24}\text{H}_{22}\text{NO}$   $[\text{M} + \text{H}]^+$  340.1701, found 340.1703;  $[\alpha]^{25}_{\text{D}} = +8.18$  ( $c = 2.29$  in  $\text{CHCl}_3$ , 95:5 er sample).

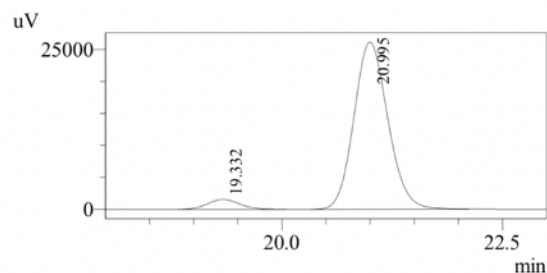
HPLC analysis: Daicel CHIRALPAK ID; hexane:*i*-PrOH = 90:10; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R = 19.3$  min (minor) and 21.0 min (major), 95:5 er.



1 Det.A Ch1 / 254nm

PeakTable

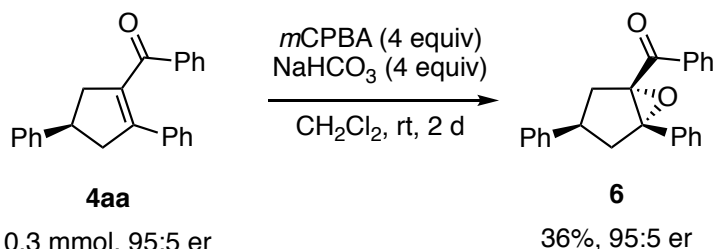
| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 19.326    | 50.037  |
| 2     | 21.054    | 49.963  |
| Total |           | 100.000 |



1 Det.A Ch1 / 254nm

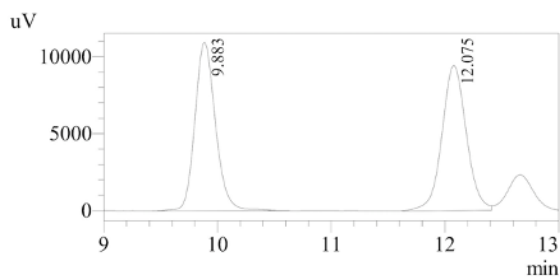
PeakTable

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 19.332    | 5.248   |
| 2     | 20.995    | 94.752  |
| Total |           | 100.000 |



**((1S,3R,5S)-3,5-Diphenyl-6-oxabicyclo[3.1.0]hexan-1-yl)(phenyl)methanone (6):** A 25 mL round bottom flask equipped with a magnetic stir bar was charged with **4aa** (97.3 mg, 0.30 mmol, 95:5 er), *m*-chloroperoxybenzoic acid (50-55 wt%, 414 mg, ca. 1.2 mmol, 4 equiv) and sodium bicarbonate (100.8 mg, 1.20 mmol, 4 equiv), followed by the addition of CH<sub>2</sub>Cl<sub>2</sub> (9 mL). The mixture was stirred at rt for 48 h. The reaction progress was monitored by GC-MS as *R<sub>f</sub>* of the product was very close to that of the starting material. The reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub> (15 mL) and washed with sat. NaHCO<sub>3</sub> aq (2 × 10 mL) and brine (10 mL). The organic layers were dried over Na<sub>2</sub>SO<sub>4</sub>, filtered, and concentrated. The crude product was purified by flash column chromatography to afford the product as a colorless oil (36.9 mg, 36%); *R<sub>f</sub>* 0.31 (hexane/EtOAc = 19/1); <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.86-7.84 (m, 2H), 7.53-7.49 (m, 1H), 7.41-7.31 (m, 8H), 7.27-7.17 (m, 4H), 3.41-3.32 (m, 1H), 2.80-2.64 (m, 4H); <sup>13</sup>C NMR (100 MHz, CDCl<sub>3</sub>) δ 195.8, 142.2, 135.8, 134.5, 133.3, 129.0, 128.6, 128.4, 128.2, 128.1, 127.4, 126.7, 126.0, 75.9, 72.5, 39.5, 38.9, 38.7; **HRMS** (ESI) Calcd for C<sub>24</sub>H<sub>21</sub>O<sub>2</sub> [M + H]<sup>+</sup> 341.1542, found 341.1544; [α]<sub>D</sub><sup>25</sup> = -7.80 (*c* = 1.30 in CHCl<sub>3</sub>, 95:5 er sample).

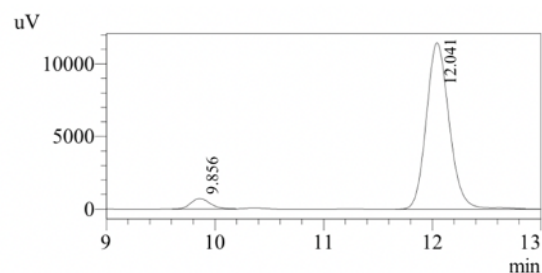
HPLC analysis: Daicel CHIRALPAK ID; hexane:*i*-PrOH = 98:2; detection wavelength = 254 nm; flow rate = 1.0 mL/min. *t<sub>R</sub>* = 9.9 min (minor) and 12.0 min (major), 95:5 er.



1 Det.A Ch1 / 254nm

PeakTable

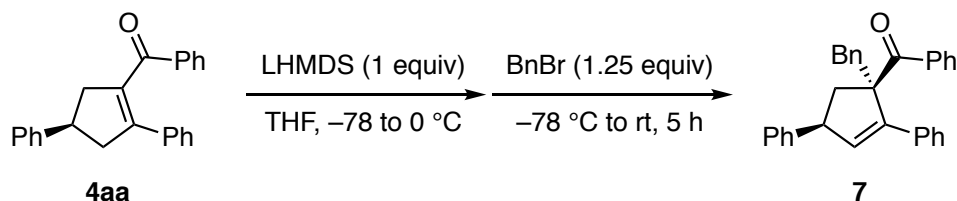
| Detector A Ch1 254nm |           |         |
|----------------------|-----------|---------|
| Peak#                | Ret. Time | Area %  |
| 1                    | 9.883     | 49.464  |
| 2                    | 12.075    | 50.536  |
| Total                |           | 100.000 |



1 Det.A Ch1 / 254nm

PeakTable

| Detector A Ch1 254nm |           |         |
|----------------------|-----------|---------|
| Peak#                | Ret. Time | Area %  |
| 1                    | 9.856     | 5.122   |
| 2                    | 12.041    | 94.878  |
| Total                |           | 100.000 |



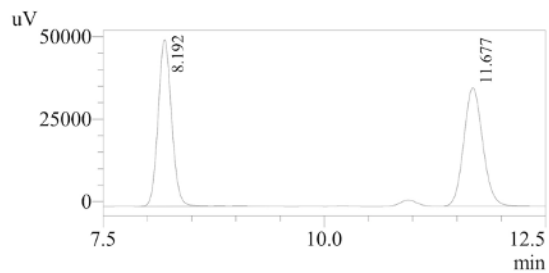
0.3 mmol, 95:5 er

75%, 95:5 er

**((1*S*,4*S*)-1-Benzyl-2,4-diphenylcyclopent-2-en-1-yl)(phenyl)methanone (7):** A 10 mL schlenk flask equipped with a magnetic stir bar was charged with **4aa** (97.3 mg, 0.30 mmol, 95:5 er) and anhydrous THF (3 mL) under nitrogen atmosphere. The mixture was cooled to -78 °C, and lithium bis(trimethylsilyl)amide (1.3 M in THF, 0.23 mL, 0.30 mmol) was added slowly. The resulting pale yellow mixture was stirred at 0 °C for 1 h. The mixture was cooled again to -78 °C, followed by dropwise addition of benzyl bromide (1.25 equiv, 64.4 mg, 0.375 mmol). The reaction mixture was gradually warmed to room temperature. After 5 h, the reaction mixture was concentrated in vacuo. The crude product was purified by flash column chromatography to afford the product as a colorless oil (93.5 mg, 75%);  $R_f$  0.15 (hexane/EtOAc = 39/1); **<sup>1</sup>H NMR** (400 MHz, CDCl<sub>3</sub>) δ 8.08-8.06 (m, 2H), 7.53-7.51 (m, 2H), 7.42-7.38 (m, 1H), 7.33-7.28 (m, 4H), 7.27-7.10 (m, 9H), 6.97-6.95 (m, 2H), 6.44 (d,  $J$  = 2.2 Hz, 1H), 3.56 (d,  $J$  = 13.8 Hz, 1H), 3.38 (d,  $J$  = 13.8 Hz, 1H), 2.89-2.84 (m, 1H), 2.64 (dd,  $J$  = 13.5, 8.2 Hz, 1H), 2.48 (ddd,  $J$  = 13.5, 9.5, 0.9 Hz, 1H); **<sup>13</sup>C NMR** (100 MHz, CDCl<sub>3</sub>; peaks showing positive and negative DEPT 135 signals are indicated with + and -, respectively) δ 204.2, 145.0, 144.2, 138.0, 137.5, 135.2 (+), 134.6, 131.9 (+), 131.0 (+), 129.5 (+), 128.9 (+), 128.5 (+), 128.1 (+), 127.8 (+), 127.6 (+), 127.2 (+), 126.7 (+), 126.4 (+), 126.4 (+), 66.7, 49.9 (+), 45.2 (-), 40.7 (-); **HRMS** (ESI) Calcd for C<sub>31</sub>H<sub>27</sub>O [M + H]<sup>+</sup> 415.2062, found

415.2067;  $[\alpha]^{25}_D = -62.7$  ( $c = 1.10$  in  $\text{CHCl}_3$ , 95:5 er sample).

HPLC analysis: Daicel CHIRALPAK IA; hexane:*i*-PrOH = 98:2; detection wavelength = 254 nm; flow rate = 1.0 mL/min.  $t_R = 8.2$  min (minor) and 11.4 min (major), 95:5 er.

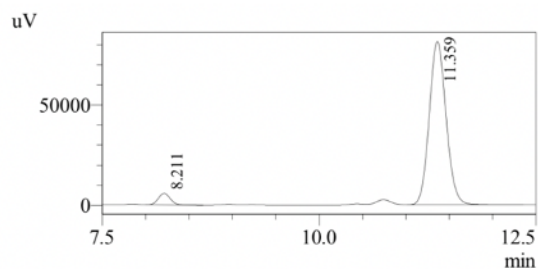


1 Det.A Ch1 / 254nm

PeakTable

Detector A Ch1 254nm

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 8.192     | 50.007  |
| 2     | 11.677    | 49.993  |
| Total |           | 100.000 |



1 Det.A Ch1 / 254nm

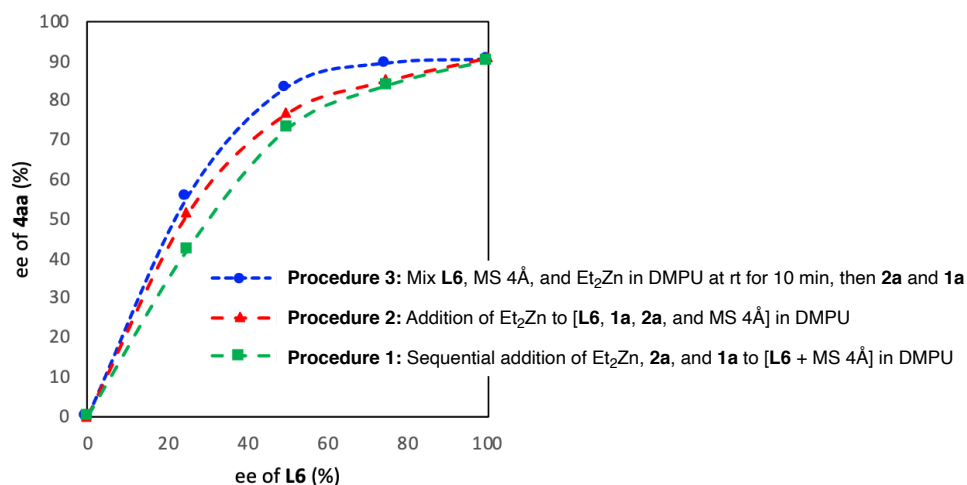
PeakTable

Detector A Ch1 254nm

| Peak# | Ret. Time | Area %  |
|-------|-----------|---------|
| 1     | 8.211     | 4.874   |
| 2     | 11.359    | 95.126  |
| Total |           | 100.000 |

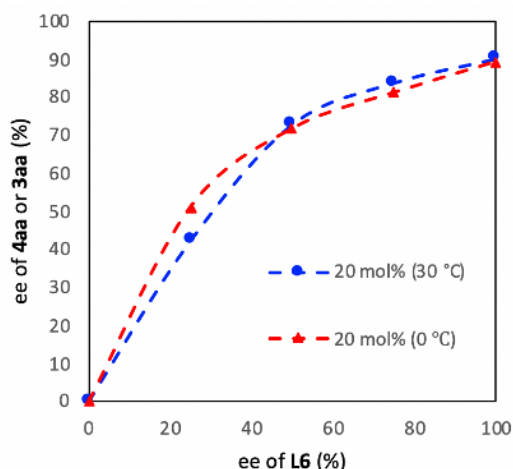
## Mechanistic Experiments

**Nonlinear effect (Figures 1a and S5-S7).** The model reaction between **1a** and **2a** was performed using ligand **L6** of different ees (25%, 50%, 75%, and 100%) with variation of other reaction parameters including the catalyst loading (10% (33 mM), 15% (50 mM), or 20% (67 mM); Figure 1a), the order of reagent addition (Figure S5), the temperature for the ECA step (0 °C or 30 °C; Figure S6), and the solvent composition (DMPU or DMPU/THF (1:1); Figure S7), and the ee of the product determined by chiral HPLC.

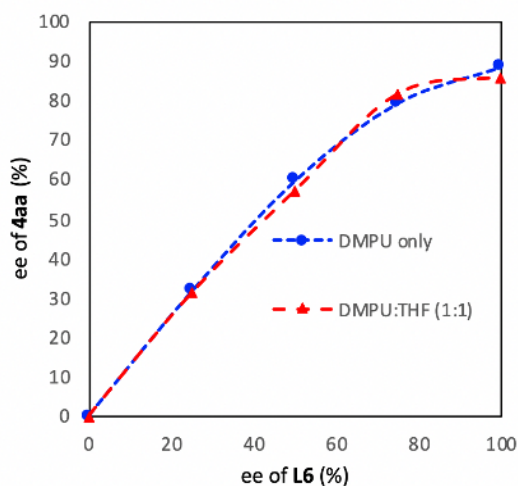


**Figure S5.** Nonlinear effects depending on the procedure for the reaction setup: Procedure 1, sequential addition of Et<sub>2</sub>Zn, **2a**, and **1a** (in this order) to a mixture of **L6** and 4 Å MS in DMPU (standard procedure); Procedure 2, addition of Et<sub>2</sub>Zn to a mixture of **L6**, **1a**, **2a**, and 4 Å MS in DMPU; Procedure 3, a mixture of **L6**, 4 Å MS, and Et<sub>2</sub>Zn stirred at room temperature for 10 min, followed by addition of **2a** and **1a**. The reactions were conducted with 20 mol % catalyst loading at 30 °C for 30 h, then 100 °C for 6 h.





**Figure S6.** Nonlinear effects with the different reaction temperature (30 °C and 0 °C). The reactions were conducted with 20 mol % catalyst loading. Ee of **3aa** was determined for the reaction conducted at 0 °C.



**Figure S7.** Nonlinear effects with different solvent composition (DMPU and DMPU/THF (1:1)). The reactions were conducted with 15 mol % catalyst loading at 30 °C for 30 h, then 100 °C for 6 h.

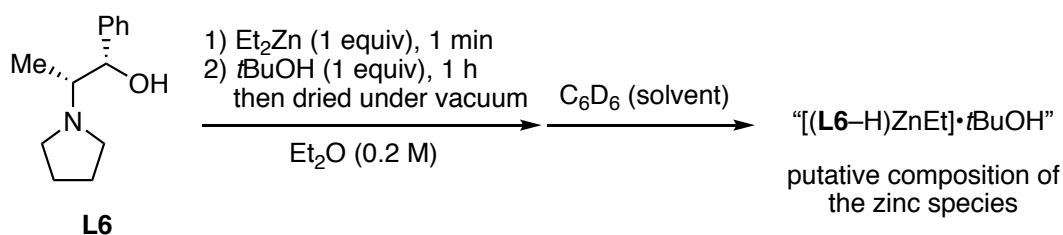
**Initial rate study (Figure 1b).** The model reaction between **1a** and **2a** was performed according to the standard procedure (15 mol % catalyst loading, 0 °C) using racemic, scalemic (50% ee), or enantiopure **L6**. Each reaction was tracked by GC analysis of periodically taken aliquots using mesitylene as an internal standard.

**Control reaction of 1a (Figure 1c).** The cyclopropanol **1a** was subjected to the standard conditions in the absence of an enone reaction partner. The conversion of **1a** to propiophenone was tracked by GC analysis of periodically taken aliquots using mesitylene as an internal standard. Full and clean conversion was observed after 48 h. Analogous experiments were also performed using DABCO instead of **L6** or simply omitting **L6**.

**Solvent effect (Figure 1d).** The model reaction between **1a** and **2a** was performed according to the standard procedure (15% catalyst, 0 °C for 48 h; without aldol condensation step) using THF/DMPU mixed solvents with different ratios (%THF = 0, 25%, 50%, 75%, and 100%). The crude reaction mixture was analyzed by GC to determine the yield of **3aa**. The ee of **3aa** was determined using a pure sample obtained by preparative TLC of the crude mixture. When the reaction was performed in pure THF, we observed the formation of a substantial amount of 2-methyl-1,3,5-triphenylpentane-1,5-dione (21% GC yield) as a result of Michael addition of propiophenone enolate to **2a**.<sup>35</sup>

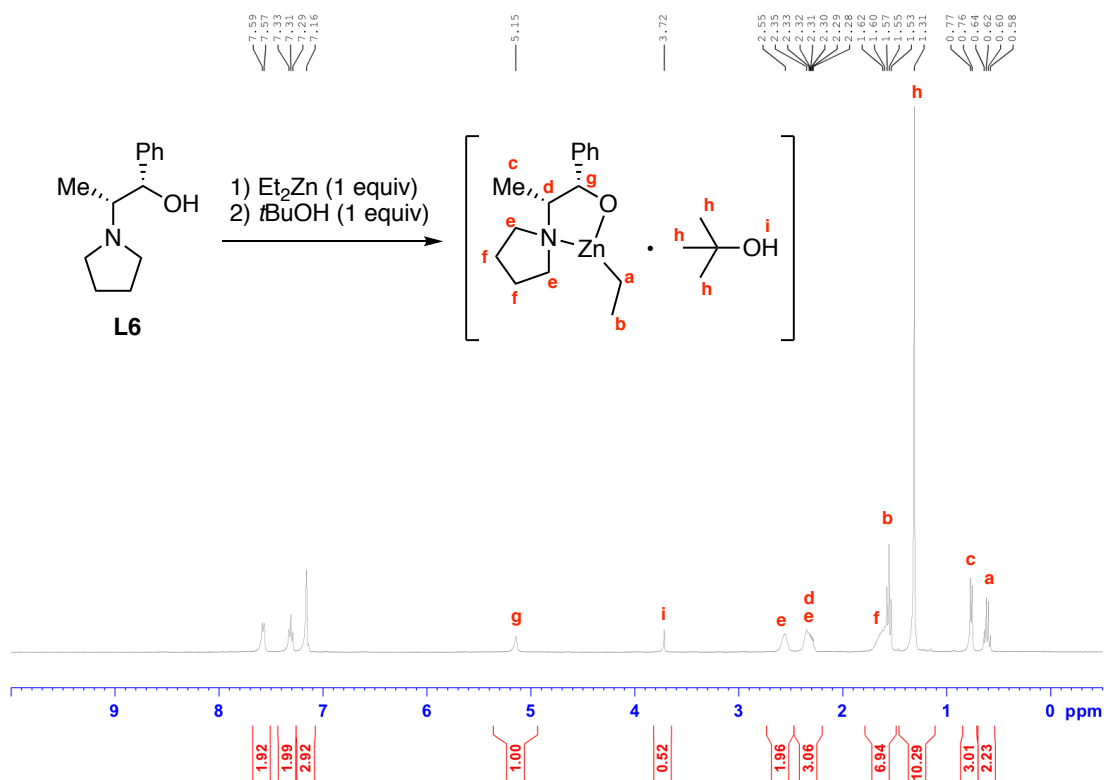
#### **<sup>1</sup>H NMR analysis of zinc species.**

**Typical procedure for sample preparation:** To a solution of **L6** (205 mg, 1.0 mmol) in Et<sub>2</sub>O (5 mL) was added Et<sub>2</sub>Zn (1 M in hexane, 1.0 mL, 1.0 mmol) at room temperature. After stirring for 1 min, *t*BuOH (95 μL, 1.0 mmol) was added, and the resulting solution was stirred for 1 h. The solution was then concentrated into dryness under vacuum. Inside a glove box, the resulting solid material was dissolved in benzene-*d*<sub>6</sub> and the solution was transferred to a J. Young NMR tube. <sup>1</sup>H NMR spectrum of the solution was recorded at room temperature.

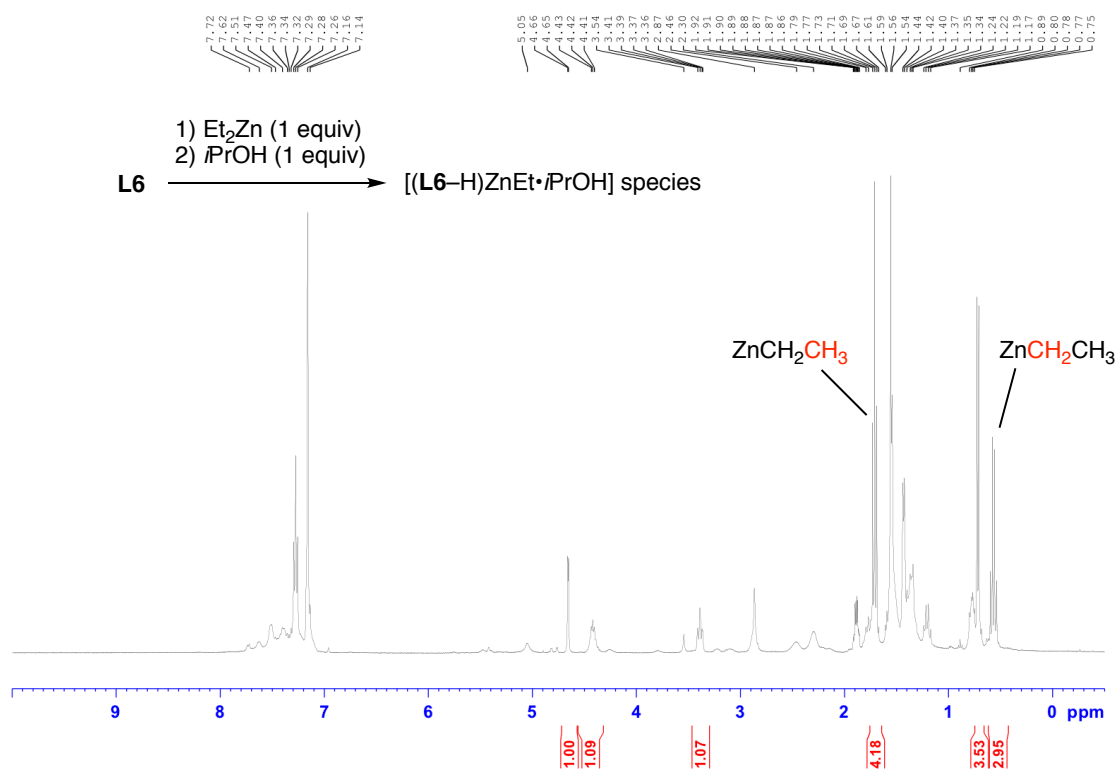


As shown in Figures S8 and S9, the samples prepared using *t*BuOH or *i*PrOH displayed well-resolved quartet (~0.6 ppm) and triplet (~1.6 ppm) that could be assigned to an ethyl group on zinc. In accordance with this signal, we observed gas evolution when adding Et<sub>2</sub>Zn to **L6** but did not do

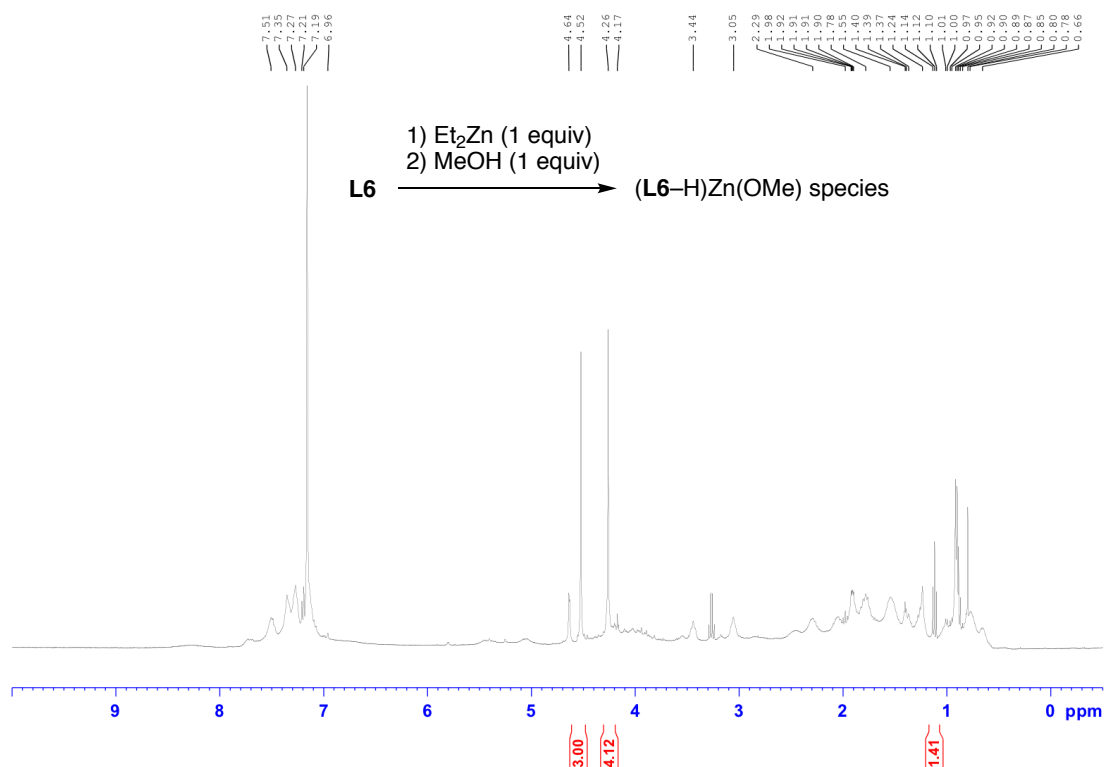
so upon the following addition of the alcohol. The former spectrum was particularly clean and allowed assignment of other signals to the respective protons with reasonable integrations, while the latter spectrum was more complex and suggested the generation of multiple aggregate species. Note that the reaction using *i*PrOH was also conducted in DMSO, which gave a spectrum identical to the one shown in Figure S9. The sample prepared using MeOH gave a much more complicated spectrum with no sign of a Zn–Et bond (Figure S10), which implied that both the ethyl groups of Et<sub>2</sub>Zn had been mostly lost by protonation and a mixture of zinc alkoxide aggregates had formed. In addition, <sup>1</sup>H NMR spectrum of the sample prepared by swapping **L6** and *t*BuOH in the above procedure (Figure S11) turned out to be close to the one shown in Figure S8, suggesting the formation of the same zinc species. Similar to the observation made in the standard sample preparation, we observed gas evolution when adding Et<sub>2</sub>Zn to *t*BuOH but did not do so upon the following addition of **L6**.



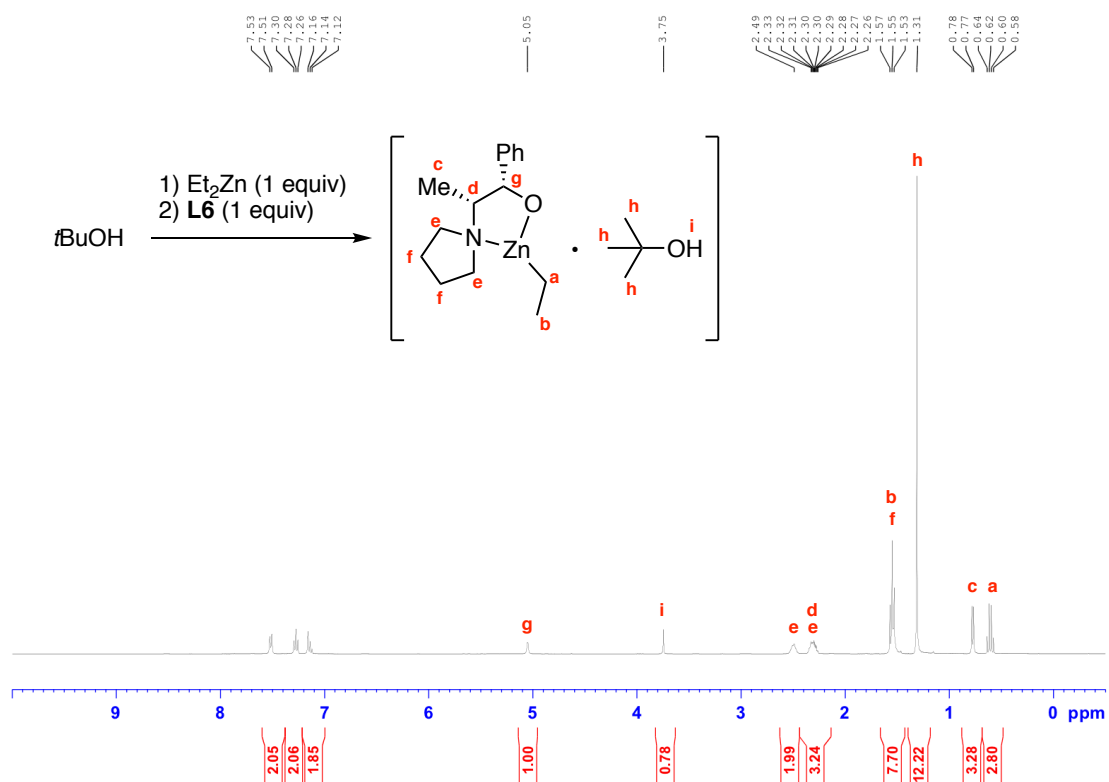
**Figure S8.** <sup>1</sup>H NMR spectrum (C<sub>6</sub>D<sub>6</sub>, 400 MHz) of zinc species prepared from **L6**, Et<sub>2</sub>Zn, and *t*BuOH.



**Figure S9.**  $^1\text{H}$  NMR spectrum ( $\text{C}_6\text{D}_6$ , 400 MHz) of zinc species prepared from **L6**,  $\text{Et}_2\text{Zn}$ , and  $i\text{PrOH}$ .



**Figure S10.**  $^1\text{H}$  NMR spectrum ( $\text{C}_6\text{D}_6$ , 400 MHz) of zinc species prepared from **L6**,  $\text{Et}_2\text{Zn}$ , and MeOH.



**Figure S11.** <sup>1</sup>H NMR spectrum (C<sub>6</sub>D<sub>6</sub>, 400 MHz) of zinc species prepared from *t*BuOH, Et<sub>2</sub>Zn, and **L6** (the order of reagent addition: *t*BuOH, Et<sub>2</sub>Zn, and then **L6**).

## References

- (1) Jia, K.; Zhang, F.; Huang, H.; Chen, Y. *J. Am. Chem. Soc.* **2016**, *138*, 1514.
- (2) Ye, Z.; Cai, X.; Li, J.; Dai, M. *ACS Catal.* **2018**, *8*, 5907.
- (3) He, X.-P.; Shu, Y.-J.; Dai, J.-J.; Zhang, W.-M.; Feng, Y.-S.; Xu, H.-J. *Org. Biomol. Chem.* **2015**, *13*, 7159.
- (4) Mills, L. R.; Zhou, C.; Fung, E.; Rousseaux, S. A. L. *Org. Lett.* **2019**, *21*, 8805.
- (5) Quan, L. G.; Lee, H. G.; Cha, J. K. *Org. Lett.* **2007**, *9*, 4439.
- (6) Li, J.; Zheng, Y.; Huang, M.; Li, W. *Org. Lett.* **2020**, *22*, 5020.
- (7) Mills, L. R.; Monteith, J. J.; dos Passos Gomes, G.; Aspuru-Guzik, A.; Rousseaux, S. A. L. *J. Am. Chem. Soc.* **2020**, *142*, 13246.
- (8) Mazzanti, S.; Kurpil, B.; Pieber, B.; Antonietti, M.; Savateev, A. *Nat. Commun.* **2020**, *11*, 1.
- (9) Chong, J. M.; Shen, L.; Taylor, N. J. *J. Am. Chem. Soc.* **2000**, *122*, 1822.
- (10) Wang, S.-K.; You, X.; Zhao, D.-Y.; Mou, N.-J.; Luo, Q.-L. *Chem. Eur. J.* **2017**, *23*, 11757.
- (11) Parambi, D. G. T.; Oh, J. M.; Baek, S. C.; Lee, J. P.; Tondo, A. R.; Nicolotti, O.; Kim, H.; Mathew, B. *Bioorg. Chem.* **2019**, *93*, 103335.
- (12) Ono, M.; Hori, M.; Haratake, M.; Tomiyama, T.; Mori, H.; Nakayama, M. *Bioorg. Med. Chem.* **2007**, *15*, 6388.
- (13) Martelli, L. S. R.; Vieira, L. C. C.; Paixão, M. W.; Zukerman-Schpector, J.; de Souza, J. O.; Aguiar, A. C. C.; Oliva, G.; Guido, R. V. C.; Corrêa, A. G. *Tetrahedron* **2019**, *75*, 3530.
- (14) Lator, A.; Gaillard, S.; Poater, A.; Renaud, J.-L. *Chem. Eur. J.* **2018**, *24*, 5770.
- (15) Yang, X.-Y.; Tay, W. S.; Li, Y.; Pullarkat, S. A.; Leung, P.-H. *Organometallics* **2015**, *34*, 5196.
- (16) Thomson, C. J.; Barber, D. M.; Dixon, D. J. *Angew. Chem. Int. Ed.* **2019**, *58*, 2469.
- (17) Arisawa, M.; Torisawa, Y.; Kawahara, M.; Yamanaka, M.; Nishida, A.; Nakagawa, M. *J. Org. Chem.* **1997**, *62*, 4327.
- (18) Xiong, H.-Y.; Yang, Z.-Y.; Chen, Z.; Zeng, J.-L.; Nie, J.; Ma, J.-A. *Chem. Eur. J.* **2014**, *20*, 8325.
- (19) Qiu, R.; Qiu, Y.; Yin, S.; Xu, X.; Luo, S.; Au, C.-T.; Wong, W.-Y.; Shimada, S. *Adv. Synth. Catal.* **2010**, *352*, 153.
- (20) Zhao, D.; Chen, C.-Y.; Xu, F.; Tan, L.; Tillyer, R.; Pierce, M. E.; Moore, J. R. *Org. Synth.* **2000**, *77*, 12.
- (21) Haraguchi, R.; Matsubara, S. *Org. Lett.* **2013**, *15*, 3378.

- (22) Kessar, S. V.; Singh, P.; Singh, K. N.; Venugopalan, P.; Kaur, A.; Bharatam, P. V.; Sharma, A. *K. J. Am. Chem. Soc.* **2007**, *129*, 4506.
- (23) Putatunda, S.; Alegre-Requena, J. V.; Meazza, M.; Franc, M.; Rohal'ová, D.; Vemuri, P.; Císařová, I.; Herrera, R. P.; Rios, R.; Veselý, J. *Chem. Sci.* **2019**, *10*, 4107.
- (24) Wommack, A. J.; Kingsbury, J. S. *J. Org. Chem.* **2013**, *78*, 10573.
- (25) Wu, X.; Liu, X.; Zhao, G. *Tetrahedron: Asymmetry* **2005**, *16*, 2299.
- (26) Bulman Page, P. C.; Allin, S. M.; Maddocks, S. J.; Elsegood, M. R. J. *J. Chem. Soc. Perkin Trans. I* **2002**, 2827.
- (27) Qin, D.-D.; Lai, W.-H.; Hu, D.; Chen, Z.; Wu, A.-A.; Ruan, Y.-P.; Zhou, Z.-H.; Chen, H.-B. *Chem. Eur. J.* **2012**, *18*, 10515.
- (28) Rodríguez-Escrich, S.; Solà, L.; Jimeno, C.; Rodríguez-Escrich, C.; Pericàs, M. A. *Adv. Synth. Catal.* **2008**, *350*, 2250.
- (29) Delair, P.; Einhorn, C.; Einhorn, J.; Luche, J. L. *Tetrahedron* **1995**, *51*, 165.
- (30) García-Delgado, N.; Fontes, M.; Pericàs, M. A.; Riera, A.; Verdaguer, X. *Tetrahedron: Asymmetry* **2004**, *15*, 2085.
- (31) Zhang, W.; Tang, R.; Yu, H.; Gao, S. *Appl. Organomet. Chem.* **2014**, *28*, 545.
- (32) Soai, K.; Yokoyama, S.; Hayasaka, T. *J. Org. Chem.* **1991**, *56*, 4264.
- (33) Mlynarski, J.; Rakiel, B.; Stodulski, M.; Syszczynska, A.; Frelek, J. *Chem. Eur. J.* **2006**, *12*, 8158.
- (34) CCDC 2056575 and 2056576 contain the supplementary crystallographic data of this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre.
- (35) Nakagawa, T.; Fujisawa, H.; Nagata, Y.; Mukaiyama, T. *Bull. Chem. Soc. Jpn.* **2005**, *78*, 236.



# NMR Spectra

