

Supporting information

Nucleophilic Fluorination of Heteroaryl Chlorides and Aryl Triflates Enabled by Cooperative Catalysis

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Purification and analytical details

Purification of desired compounds was performed by direct crystallization or chromatographic purification. Column chromatography was performed using a Teledyne ISCO CombiFlash® Rf+ chromatography system using prepacked single-use silica packed cartridges (RediSep® Rf Gold Normal-Phase Silica, 20–40 micron average particle size, 60 Å average pore size, with varying cartridge sizes ranging from 24 g (part number 69-2203-346), 40 g (part number 69-2203-347), 80 g (part number 69-2203-348) to 120 g SiO₂ cartridges, part number 69-2203-349) operating with a flow rate of 35, 40, 80 and 85 mL/min, respectively, depending on the size of the cartridge.

Proton nuclear magnetic resonance (¹H NMR) spectra, proton decoupled carbon nuclear magnetic resonance (¹³C NMR) spectra, and proton decoupled fluorine nuclear magnetic resonance (¹⁹F NMR) spectra were recorded at 25 °C unless stated otherwise on a Bruker DRX-500 spectrometer or a Bruker Neo 600 MHz spectrometer equipped with an iProbe TBO. Chemical shifts for protons are reported in parts per million (ppm) downfield from tetramethylsilane and are referenced to residual proton signals of the NMR solvent according to values reported in the literature.¹ Chemical shifts for carbon are reported in parts per million (ppm) downfield from tetramethylsilane and are referenced to the carbon resonances of the NMR solvent according to values reported in the literature.¹ Data are presented as follows: chemical shift, integration, multiplicity (br = broad, s = singlet, d = doublet, t = triplet, q = quartet, quint = quintet, m = multiplet, dt = doublet of triplets), coupling constants (J) in Hertz (Hz). High resolution mass spectra (HRMS) were obtained on an Agilent 1290 Infinity 2 equipped with a Binary Pump and DAD coupled to a Agilent 6230 TOF. High throughput screening was performed in a 24 well plate using stock solutions and multichannel pipettors.

Screening results

A 24-well reaction plate was used to evaluate initial conditions and performance of two tetramethylammonium salts for cooperative catalysis with various crown ethers on 0.05 mmol scale with respect to substrate (**1a**). These wells were charged with stir bars and 3 equiv. of KF or CsF. Stock solutions of (**1a**), tetramethylammonium chloride, tetramethylammonium hexafluorophosphate, 18-crown-6 ether, 15-crown-5 ether, dibenzo-18-crown-6 ether were prepared in MeCN in a nitrogen glovebox. These were dispensed accordingly to screen all combinations of tetramethylammonium salts (0.1 equiv) for cooperative catalysis in the presence of a phase transfer catalyst (0.1 equiv).

Well	F ⁻ salt	TMA salt	Crown Ether	Evaluated	LC area % 2a
1	KF	-	-	Control	0
2	KF	-	18-c-6	18-c-6	4
3	KF	TMACl	-	TMACl	0
4	KF	TMACl	15-c-5	TMACl + 15-c-5	2
5	KF	TMACl	18-c-6	TMACl + 18-c-6	11
6	KF	TMACl	BZ-18-c-6	TMACl + BZ-18-c-6	1
7	KF	TMABF ₄	-	TMABF ₄	3
8	KF	TMABF ₄	15-c-5	TMABF ₄ + 15-c-5	2
9	KF	TMABF ₄	18-c-6	TMABF ₄ + 18-c-6	18
10	KF	TMABF ₄	BZ-18-c-6	TMABF ₄ + BZ-18-c-6	1
11	CsF	-	-	TMABF ₄ + Control	5
12	CsF	-	18-c-6	TMABF ₄ + 18-c-6	47
13	CsF	TMACl	-	TMACl	12
14	CsF	TMACl	15-c-5	TMACl + 15-c-5	95
15	CsF	TMACl	18-c-6	TMACl + 18-c-6	97
16	CsF	TMACl	BZ-18-c-6	TMACl + BZ-18-c-6	88
17	CsF	TMABF ₄	-	TMABF ₄	31
18	CsF	TMABF ₄	15-c-5	TMABF ₄ + 15-c-5	81
19	CsF	TMABF ₄	18-c-6	TMABF ₄ + 18-c-6	94
20	CsF	TMABF ₄	BZ-18-c-6	TMABF ₄ + BZ-18-c-6	95

Hygroscopicity studies of raw materials

The primary challenges with using highly hygroscopic reagents such as TMAF on manufacturing scale are two-fold. First, a reagent's ability to rapidly and efficiently absorb water from atmospheric conditions poses a challenge to water-sensitive chemistry by introducing a point of entry for uncontrolled amounts of water. Second, reagents that are hygroscopic to the degree of deliquescing at ambient humidities pose challenges from the perspective of raw material handling, even if the chemistry is tolerant to water. While solid, liquid, slurry, and gas reagents are well established for handling in a plant setting, deliquescing reagents can form wet clumpy solids, gels, or thick liquids, which pose a significant operational challenge.

To quantify the relative degrees of water uptake, samples of raw materials for this method and TMAF were aged open to the atmosphere under ambient laboratory conditions (40% relative humidity, 72 °F). These conditions can be used to approximate the conditions under which pilot plant subdivisions are typically carried out. A

representative subdivision operation to prepare 50 kg of a solid reagent can require time cycles of up to six hours of exposure of the raw material to ambient conditions. It was observed that at 22 hours of aging, TMAF had completely deliquesced into a thick gel, while TMACl, 18-crown-6 ether, and CsF were still well-behaved (albeit visibly wet) solids.

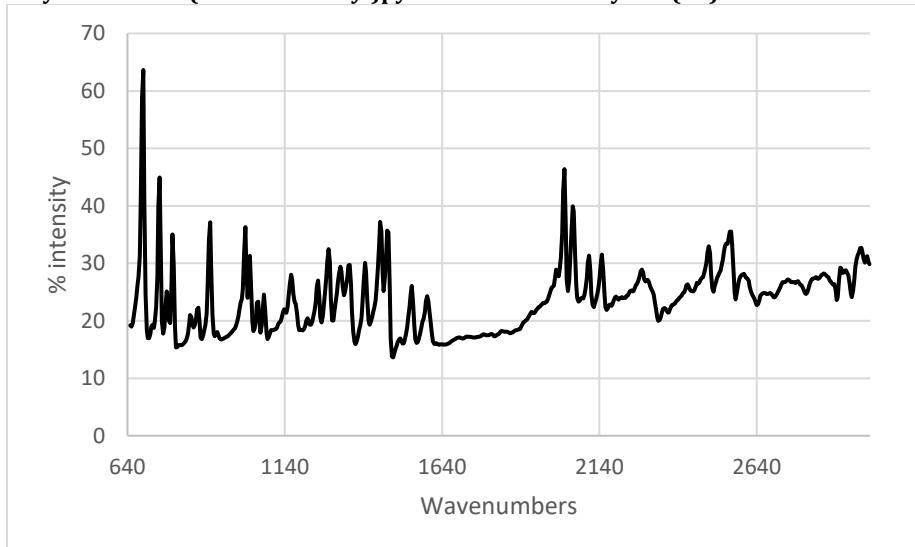
Water uptake in mg / g of reagent			
Reagent	1 h	4 h	22 h
TMAF	66	162	547
TMACl	23	49	148
18-c-6	1	2	2
CsF	7	13	43

Impact of water content

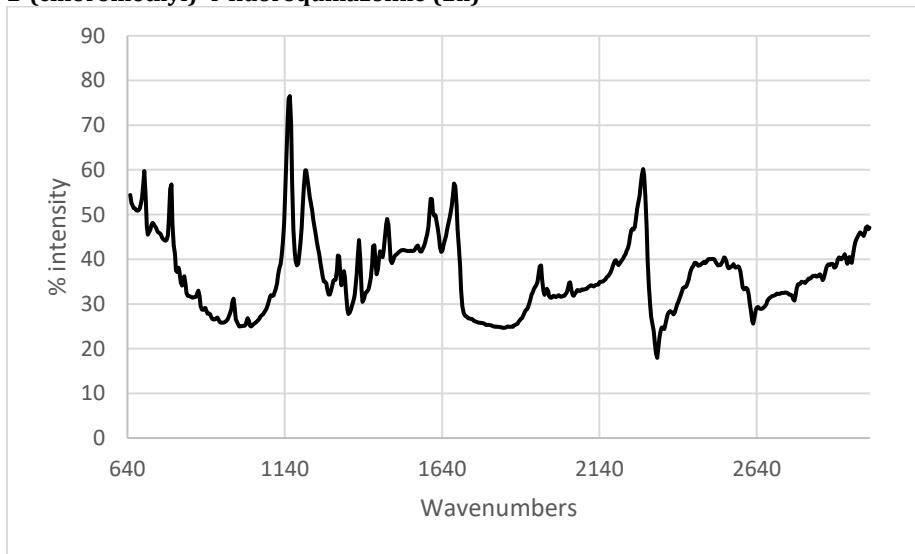
To understand the tolerance of this method to water intrusion, the fluorination of **1a** was evaluated with varying degrees of water content in the acetonitrile used (measured in duplicate *via* Karl Fischer titration). Reaction profiles were evaluated at 16 hours by HPLC (210 nm). The reaction is notably tolerant to water intrusion, showing only minor deterioration in the impurity profile and a slightly slowed reaction rate at high levels of water intrusion.

Water content (ppm) measured in duplicate	Product (LC area %)	Starting Material (LC area %)	Impurities (LC area %)
72.25	87.1	0.0	12.9
943.05	89.8	0.0	10.2
1955.70	89.1	0.0	10.9
3093.80	85.5	0.0	14.5
4094.50	84.6	4.0	11.4

IR data for previously unreported compounds
ethyl 4-fluoro-2-(trifluoromethyl)pyrimidine-5-carboxylate (2k)

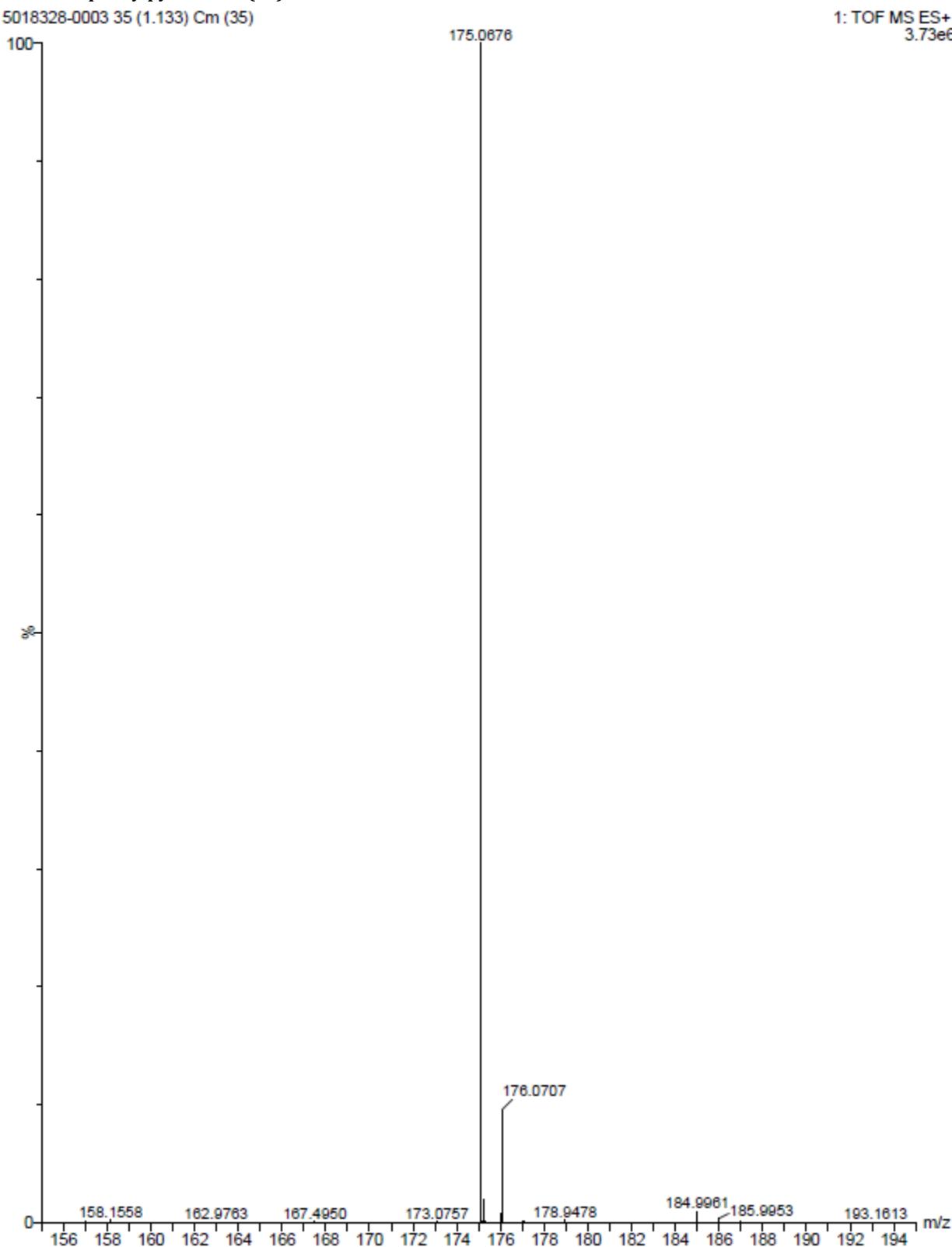


2-(chloromethyl)-4-fluoroquinazoline (2n)



HRMS data for previously unreported and selected compounds

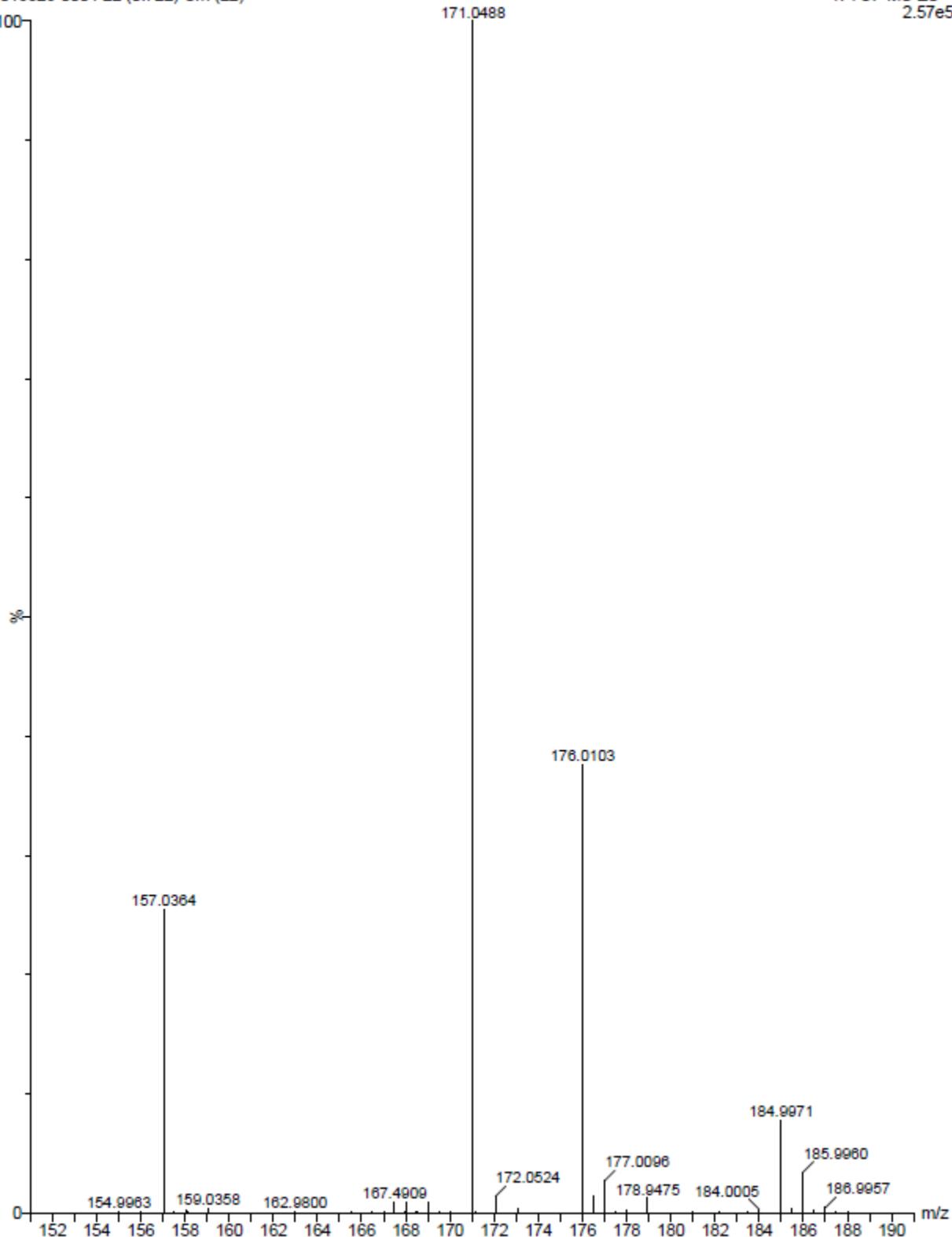
2-fluoro-4-phenylpyrimidine (2a)



2,6-difluoro-9-methyl-9H-purine (2b)

5018328-0004 22 (0.722) Cm (22)

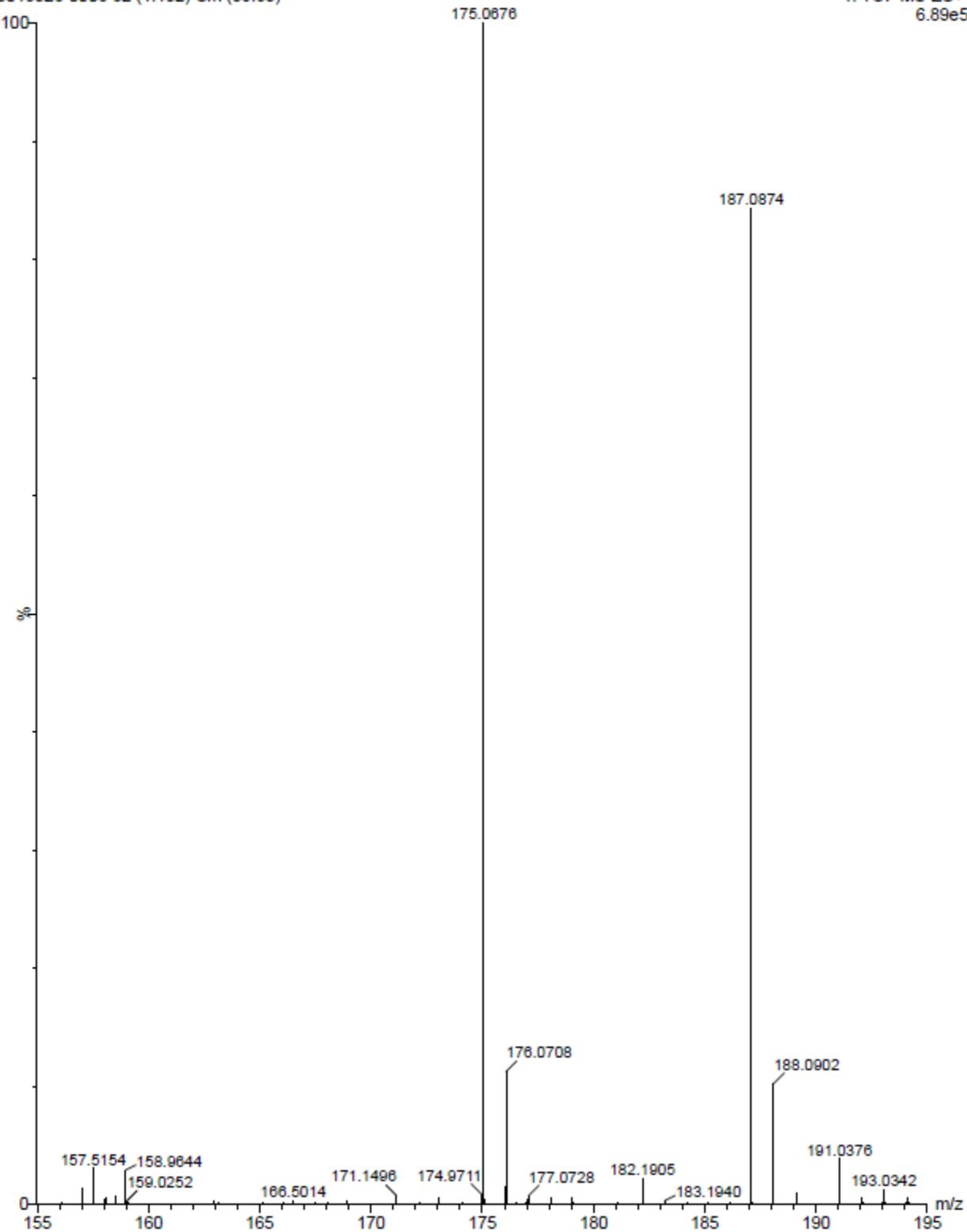
1: TOF MS ES+
2.57e5



3-fluoro-6-phenylpyridazine (2d)

5018328-0006 62 (1.152) Cm (59:63)

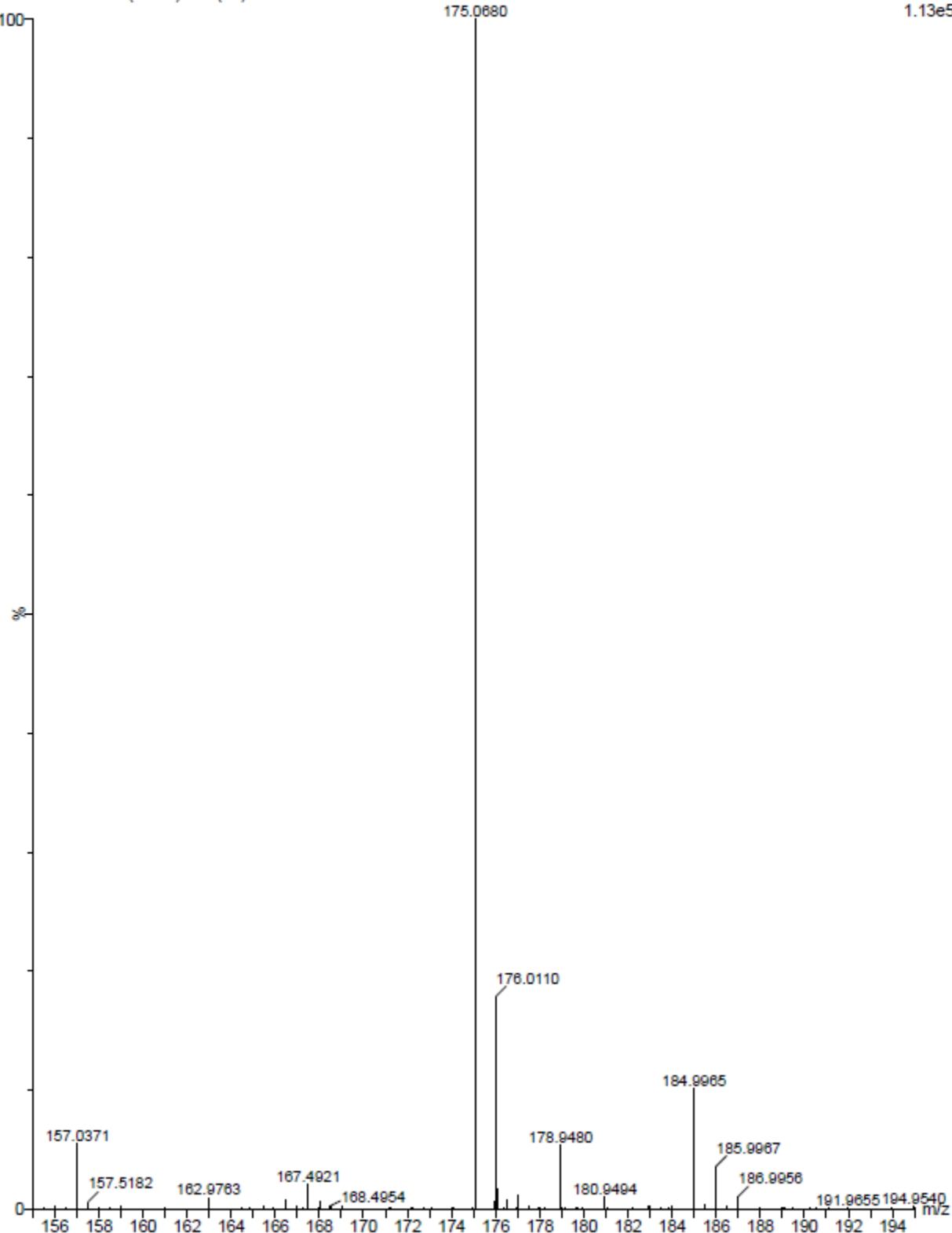
1: TOF MS ES+
6.89e5



2-fluoro-5-phenylpyrazine (2g)

5018328-0007 38 (1.221) Cm (38)

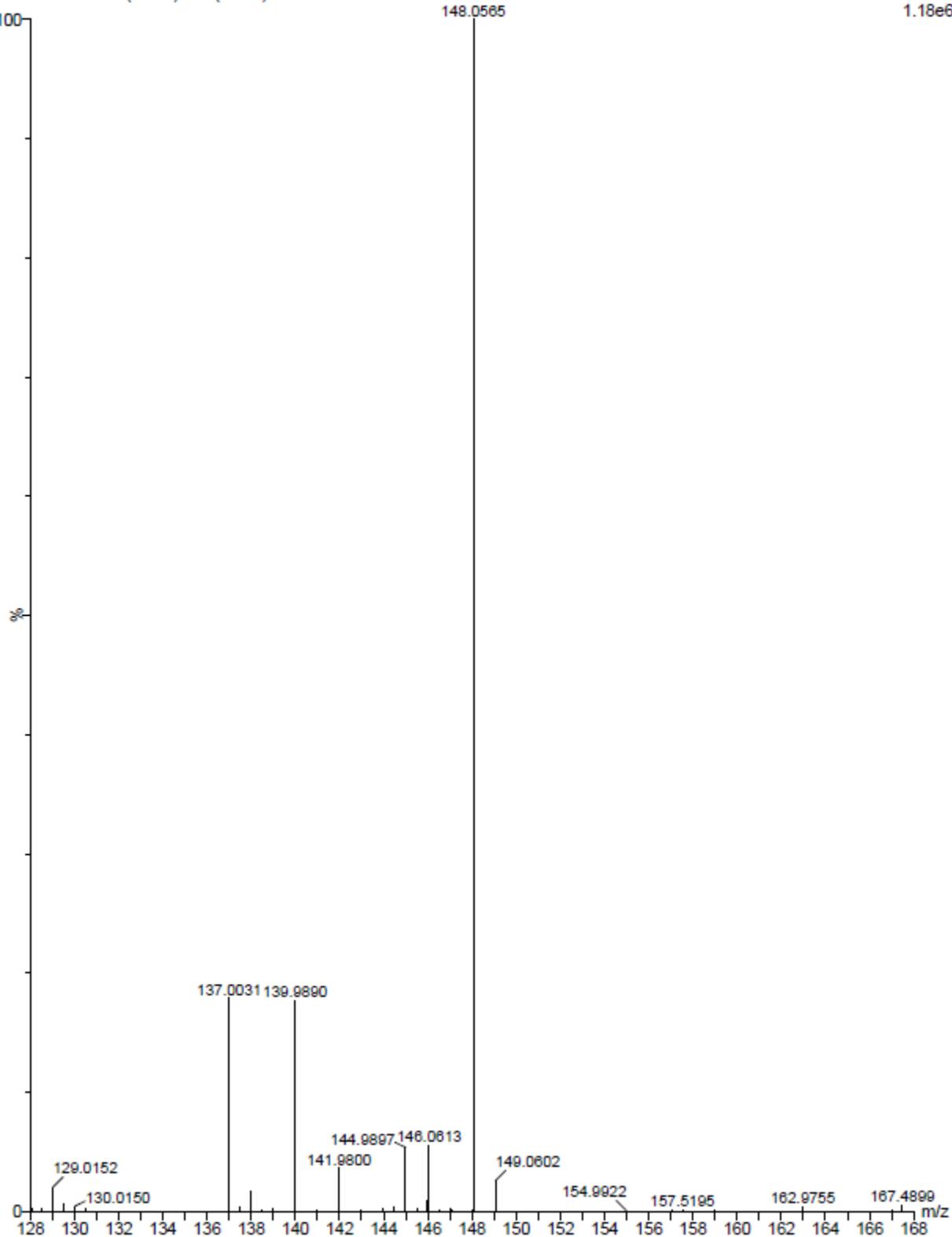
1: TOF MS ES+
1.13e5



2-fluoroquinoline (2h)

5018328-0020 33 (1.066) Cm (32:34)

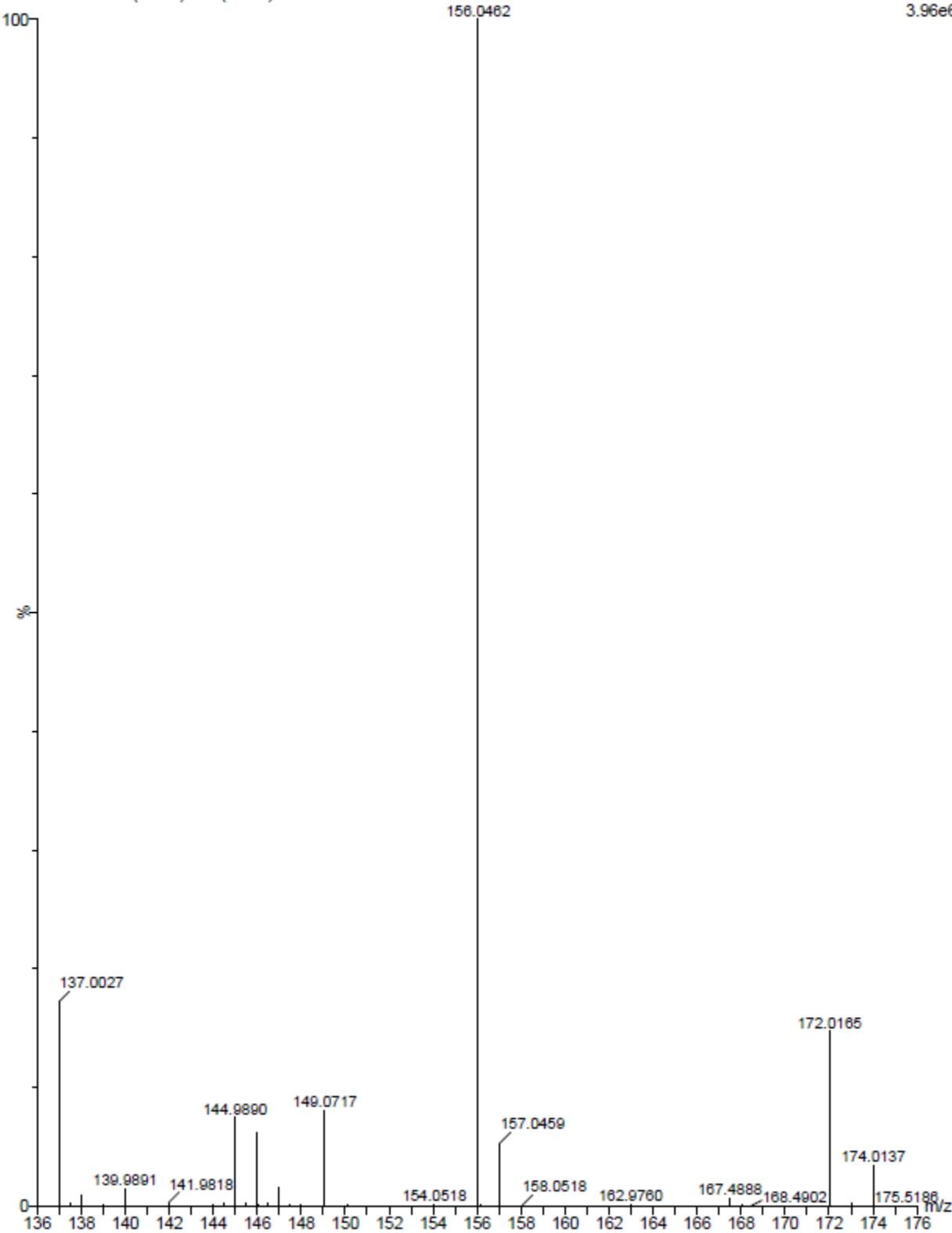
1: TOF MS ES+
1.18e6



methyl 2-fluoronicotinate (2j)

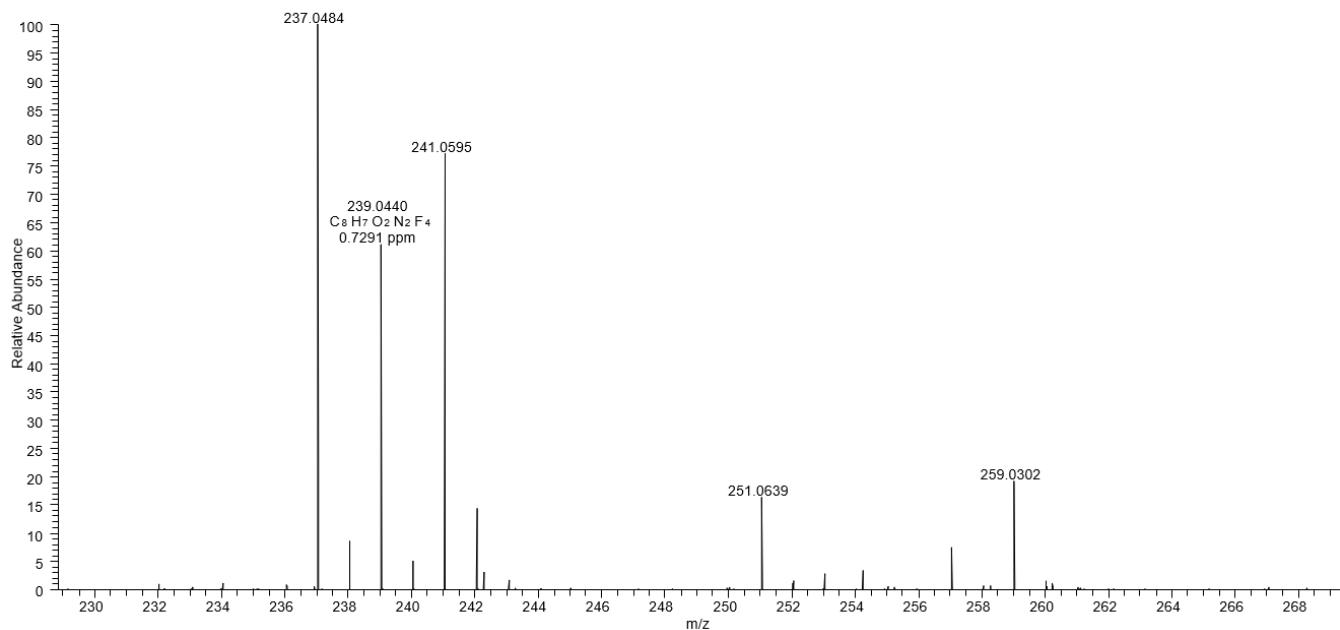
5018328-0028 25 (0.809) Cm (25:27)

1: TOF MS ES+
3.96e6

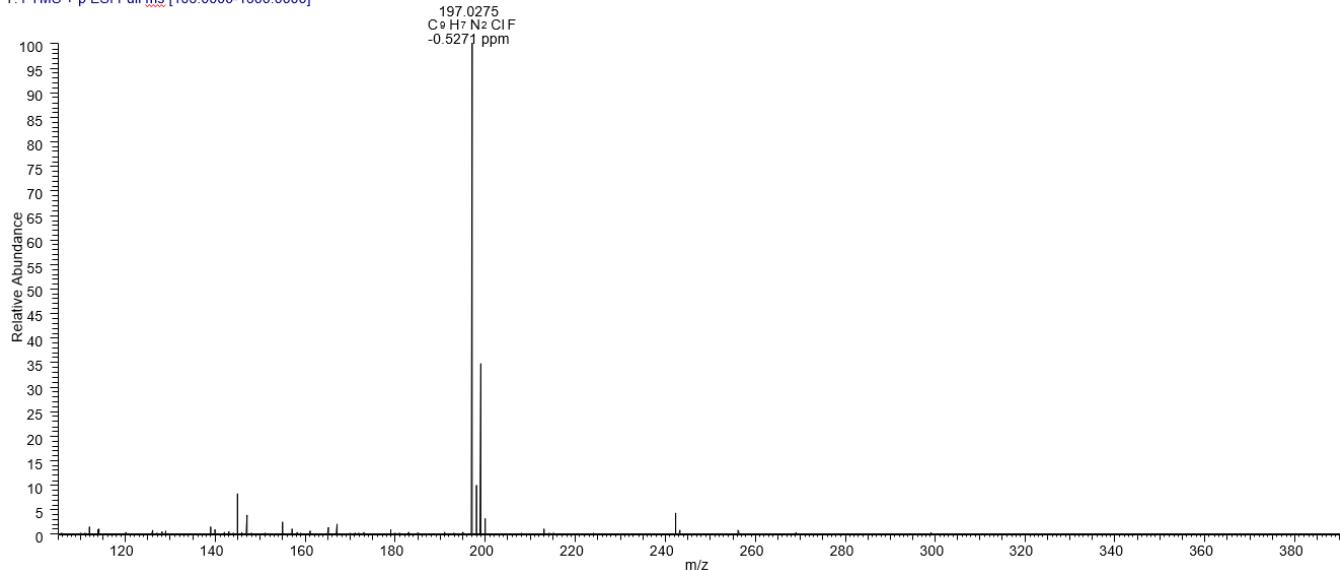


ethyl 4-fluoro-2-(trifluoromethyl)pyrimidine-5-carboxylate (2k)

T: FTMS + p ESI Full ms [105.0000-1500.0000]

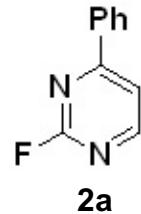
**2-(chloromethyl)-4-fluoroquinazoline (2n)**

T: FTMS + p ESI Full ms [105.0000-1500.0000]

**References**

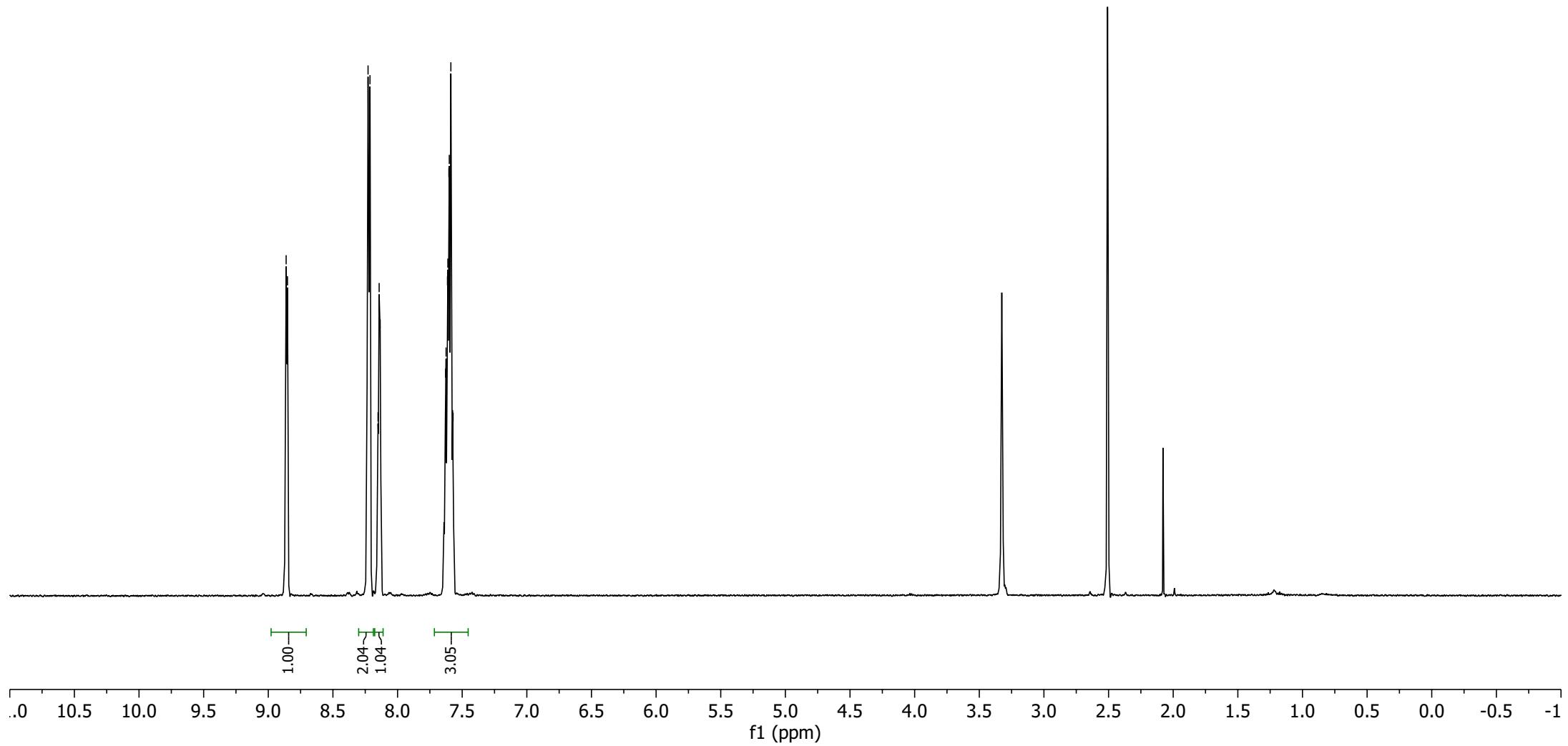
1. Fulmer, G. R.; Miller, A. J. M.; Sherden, N. H.; Gottlieb, H. E.; Nudelman, A.; Stoltz, B. M.; Bercaw, J. E.; Goldberg, K. I. *Organometallics*. **2010**, *29*, 2176–2179.

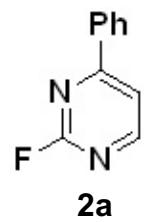
Copies of NMR data



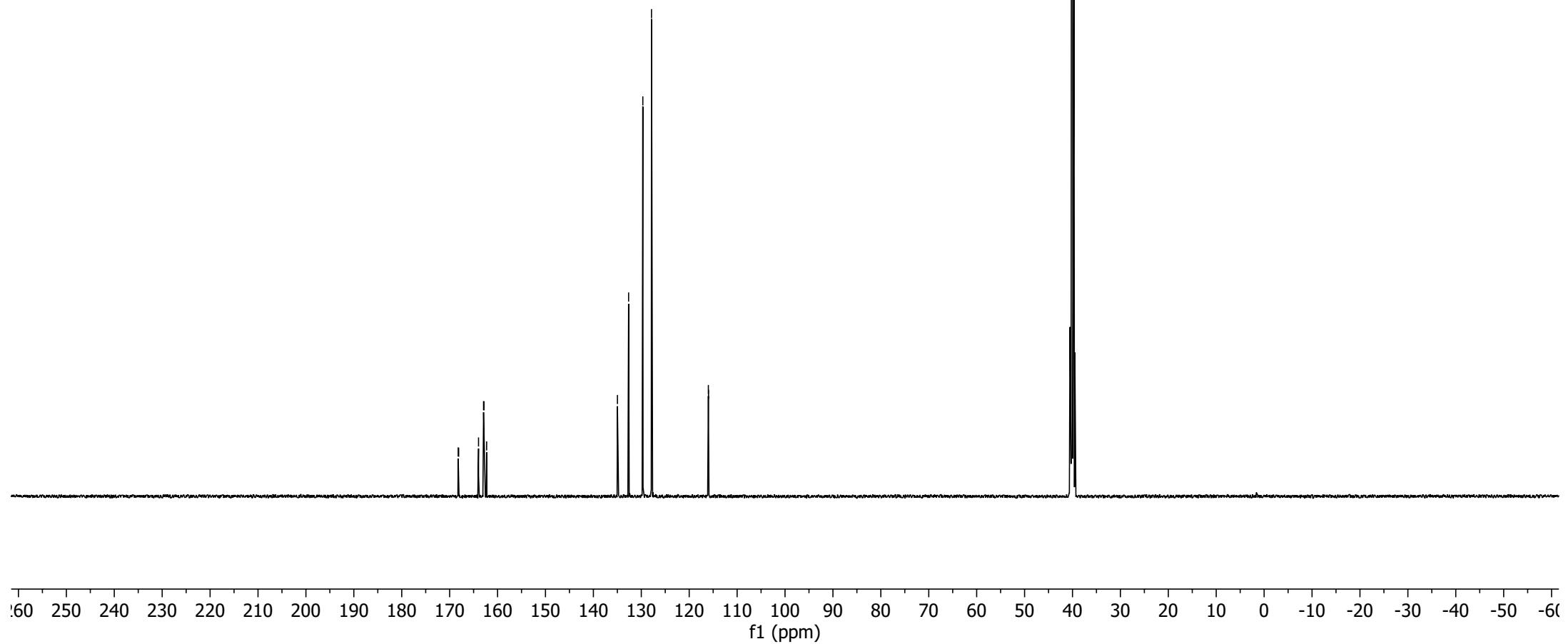
8.86
8.86
8.85
8.23
8.21
8.15
8.14
8.13
7.63
7.62
7.61
7.61
7.60
7.60
7.59
7.57

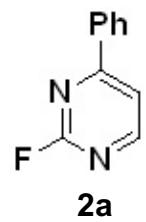
500 MHz, DMSO-d6



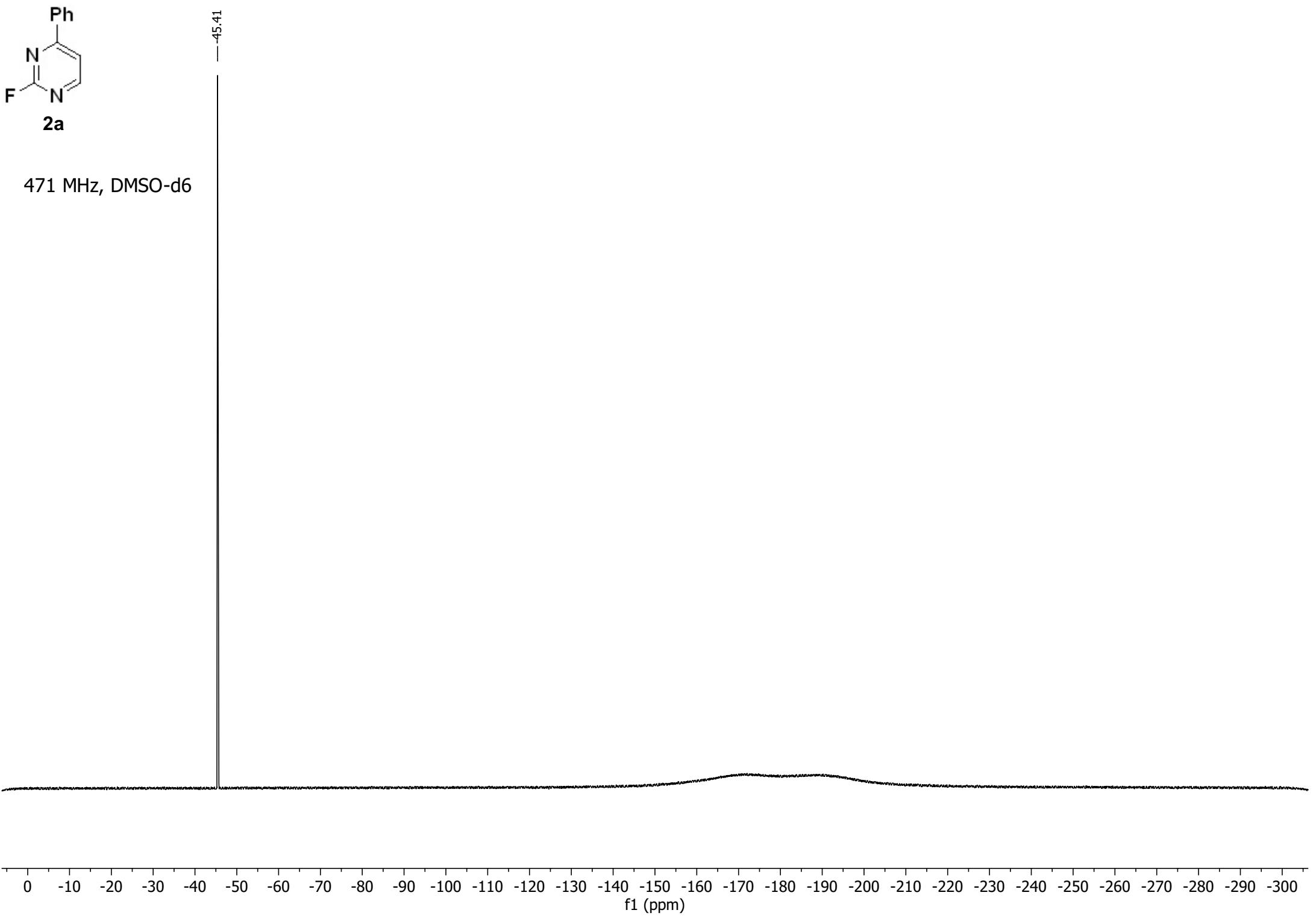


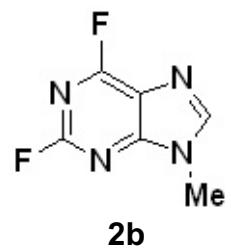
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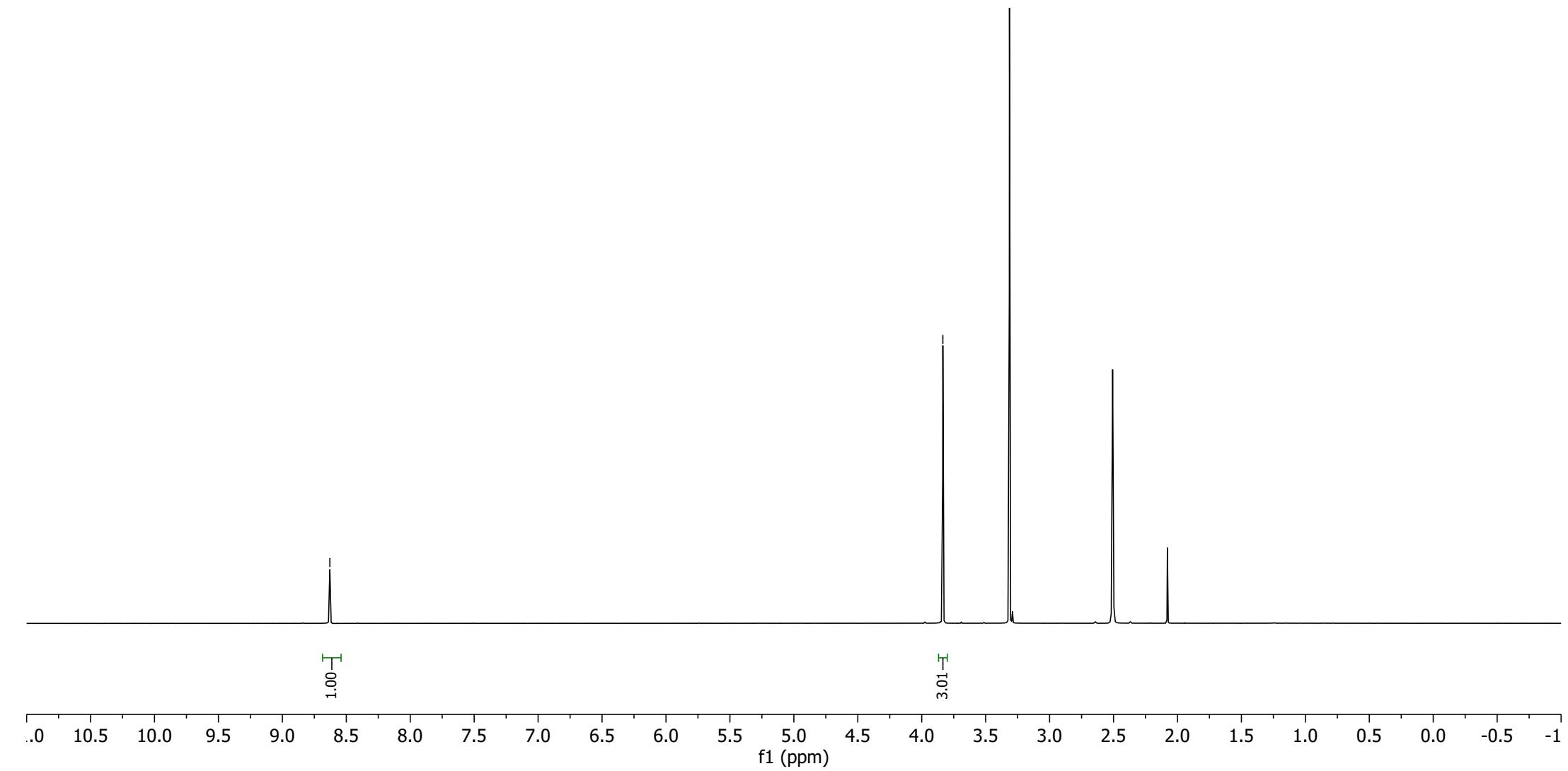


471 MHz, DMSO-d6



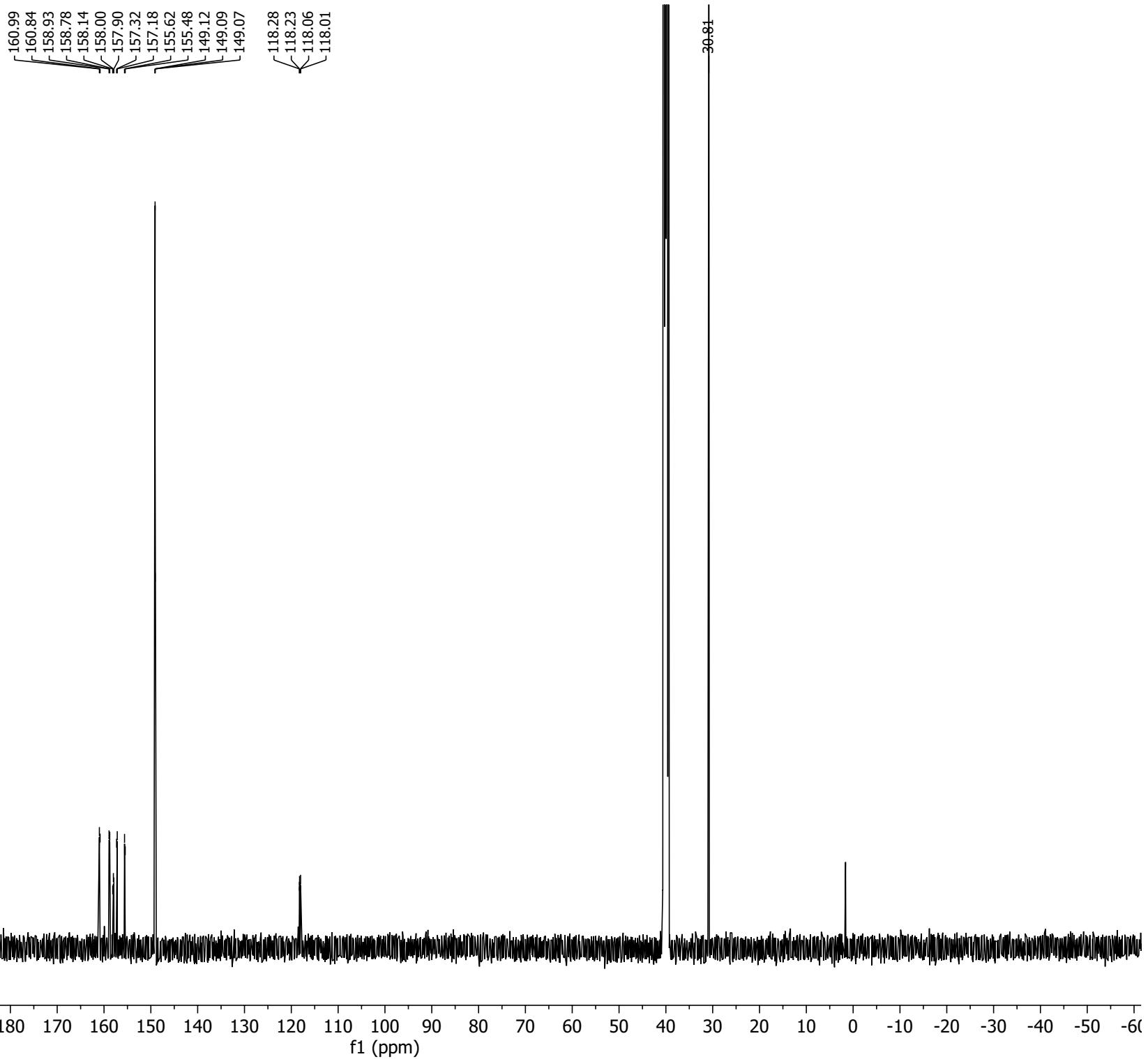


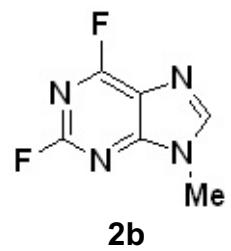
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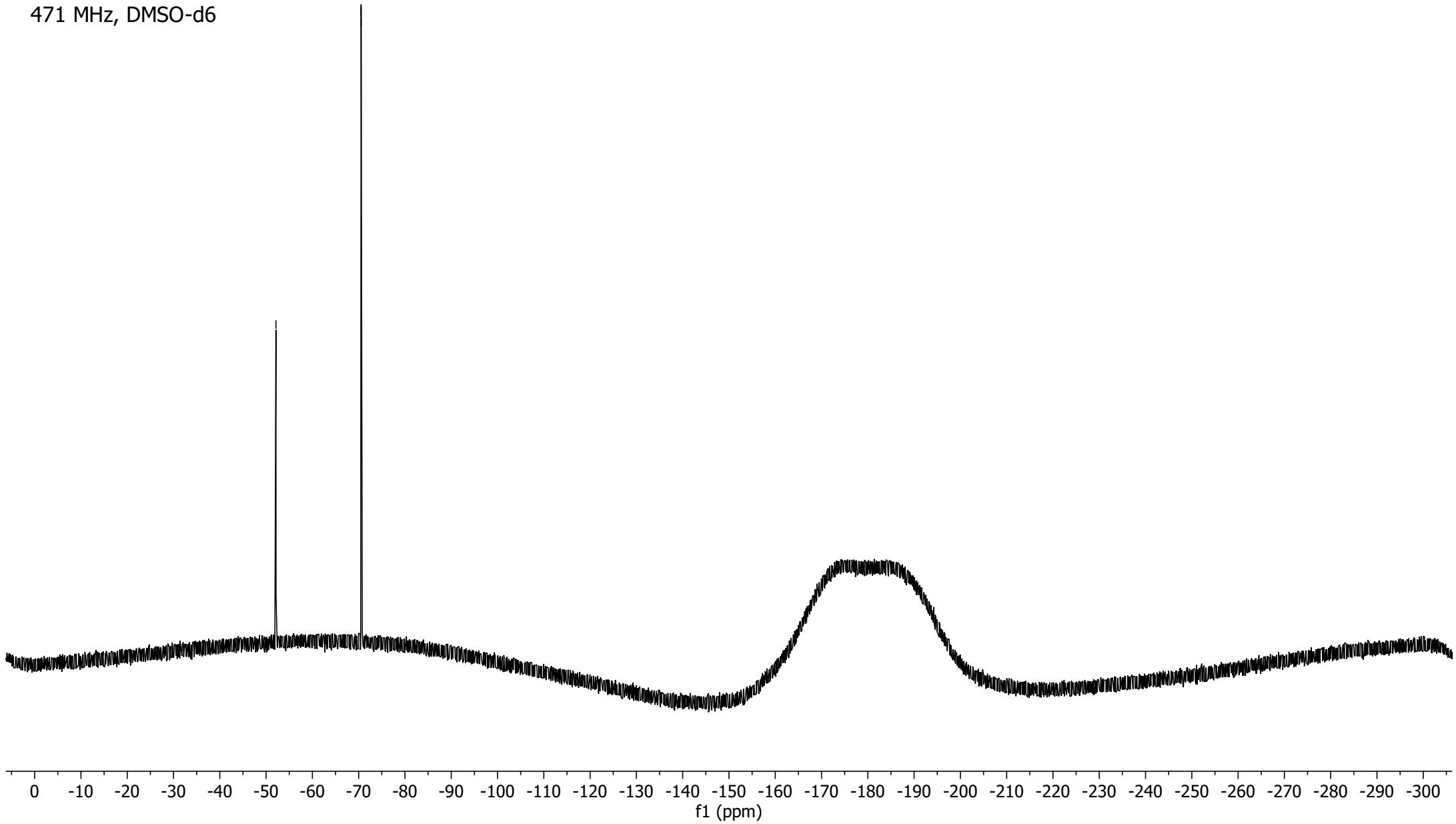
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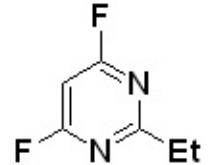




—52.13
-70.53
-70.54

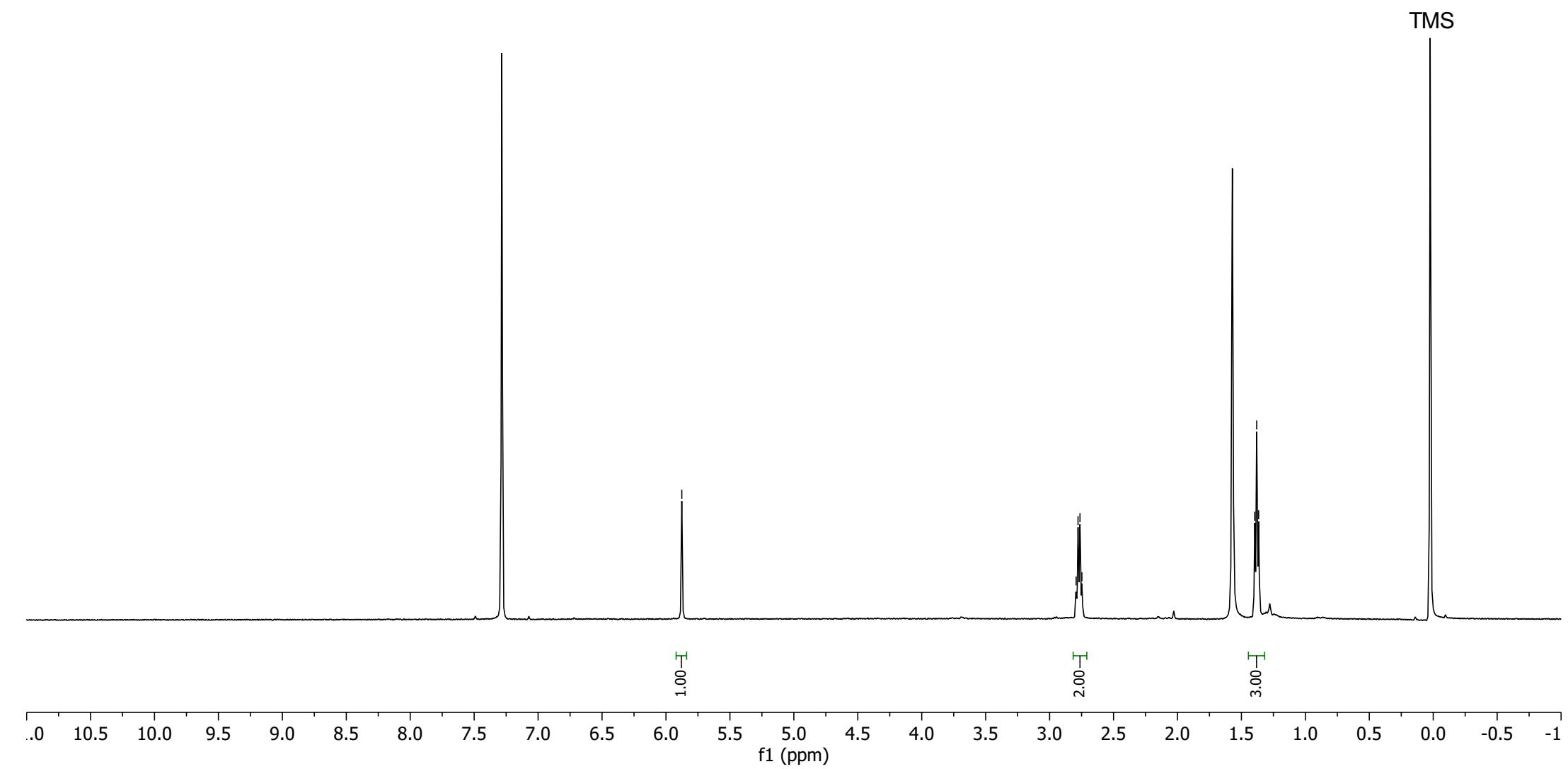
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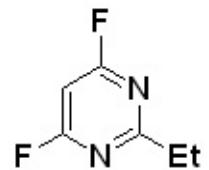




2c

500 MHz, CDCl₃





2c

126 MHz, CDCl₃

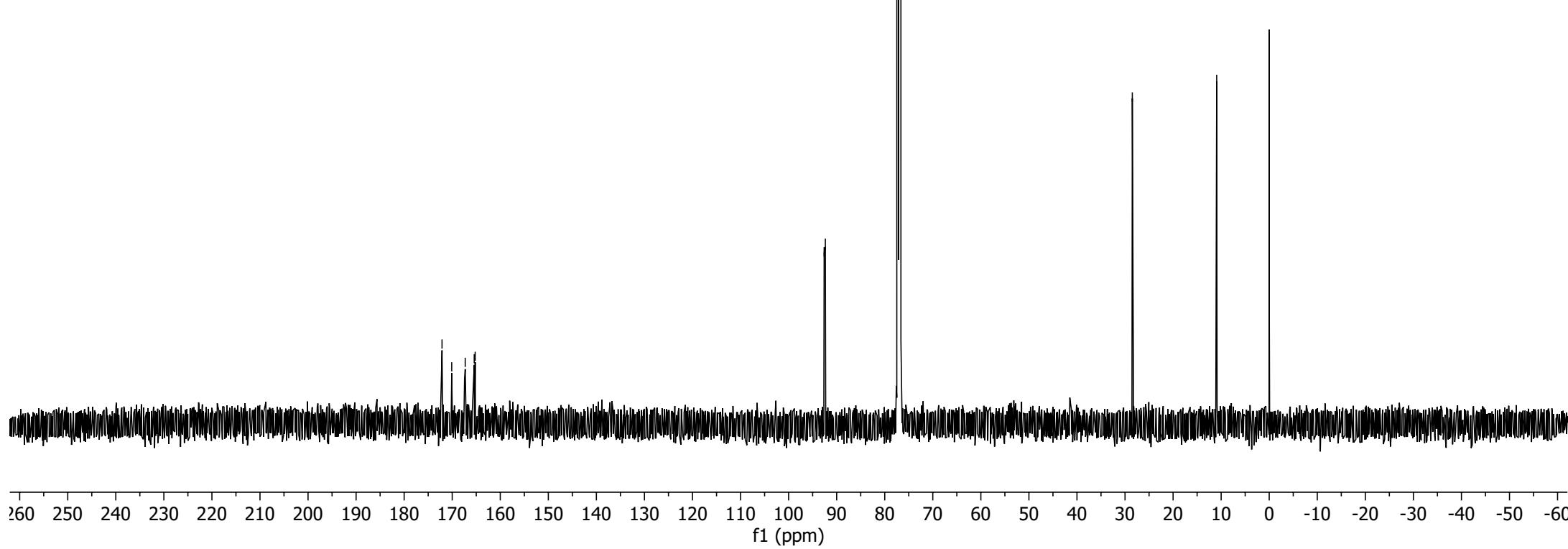
172.12
170.10
167.29
165.42
165.22

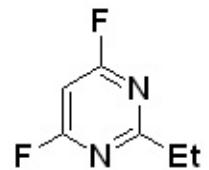
92.61
92.36

-28.48

-10.93

TMS

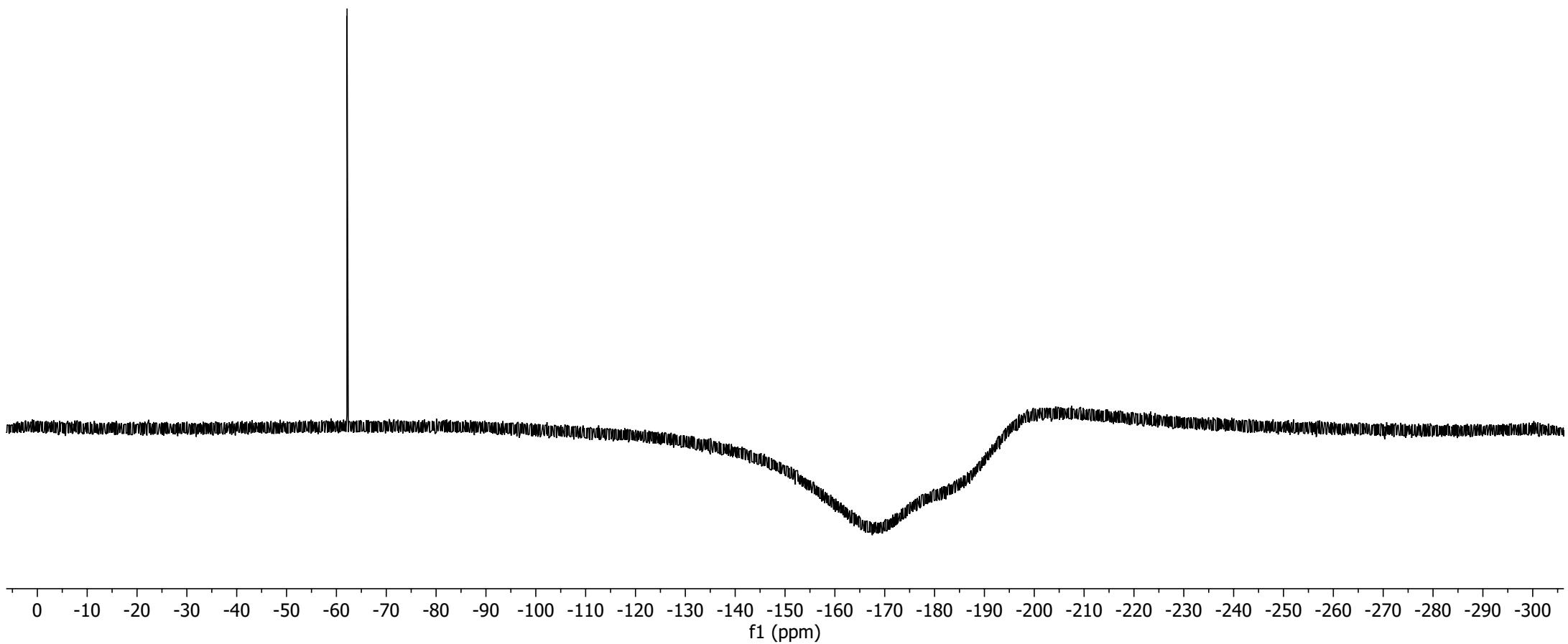


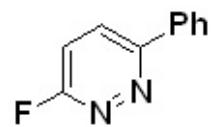


2c

471 MHz, CDCl₃

--62.13

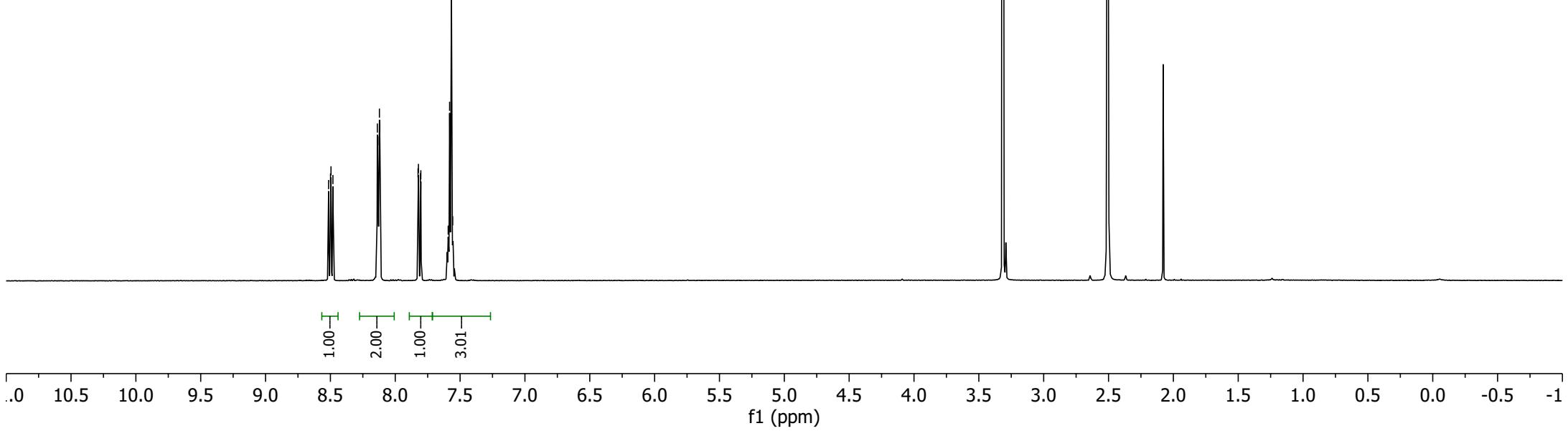


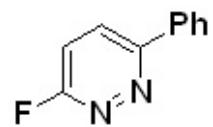


2d

8.51
8.50
8.48
8.14
8.13
8.12
8.11
7.82
7.82
7.81
7.80
7.59
7.58
7.57
7.56

500 MHz, DMSO-d6

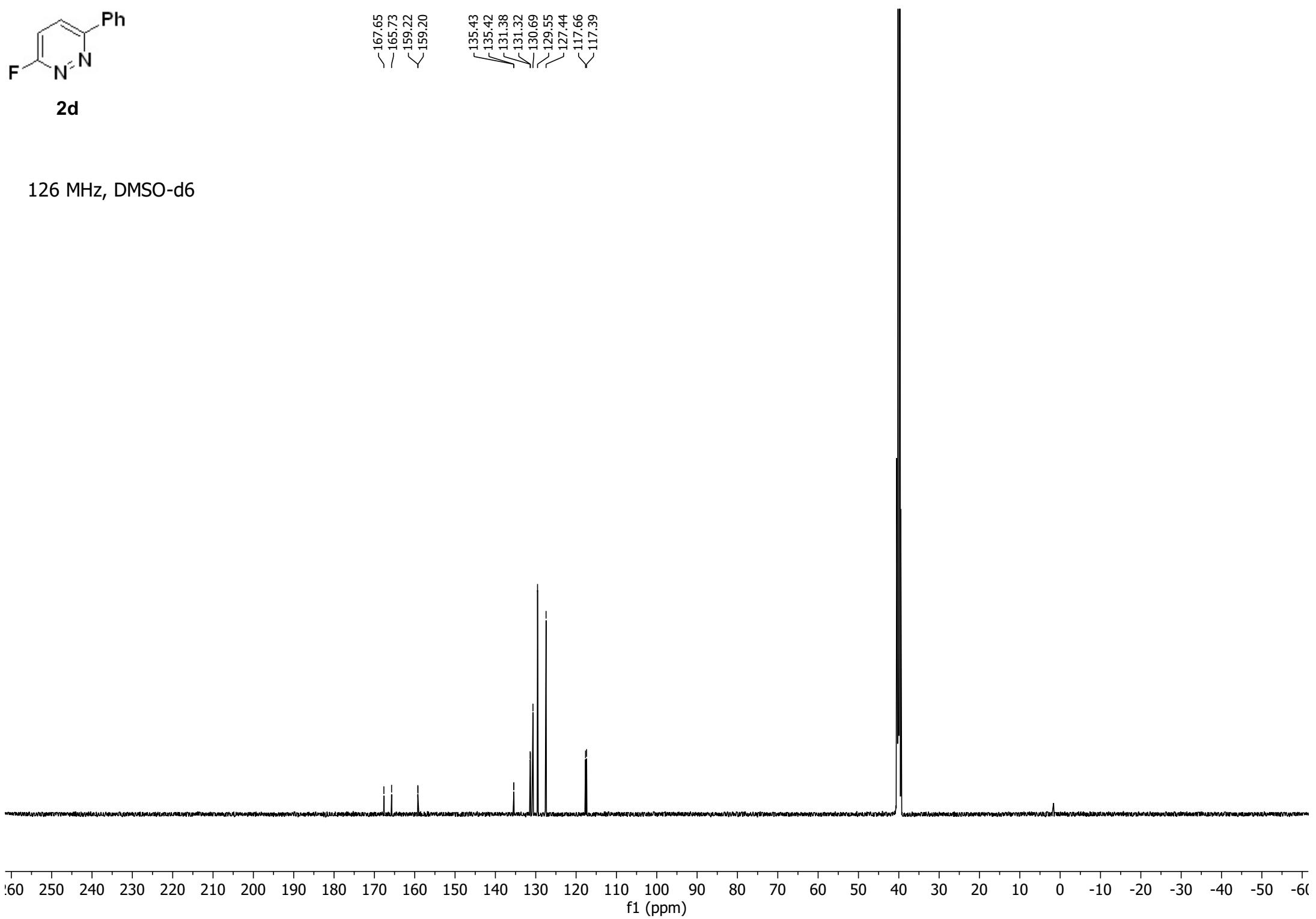


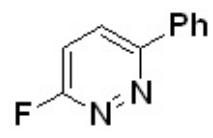


2d

126 MHz, DMSO-d6

167.65
165.73
159.22
159.20
135.43
135.42
131.38
131.32
130.69
129.55
127.44
117.66
117.39



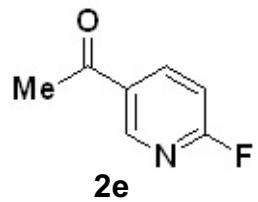


2d

471 MHz, DMSO-d6

-83.37

f1 (ppm)

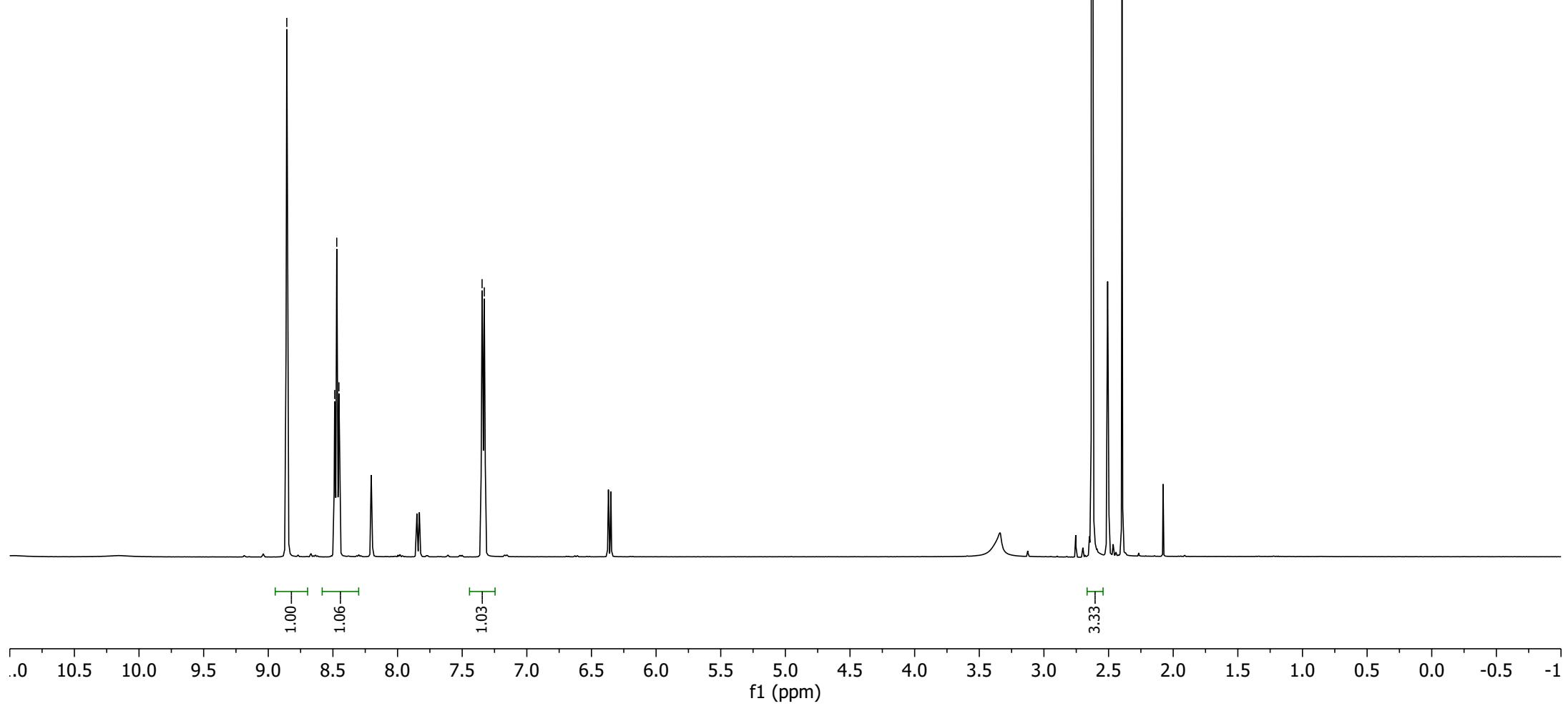


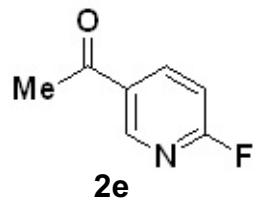
8.86
8.49
8.48
8.47
8.45
8.45

7.35
7.33

2.63

500 MHz, DMSO-d₆





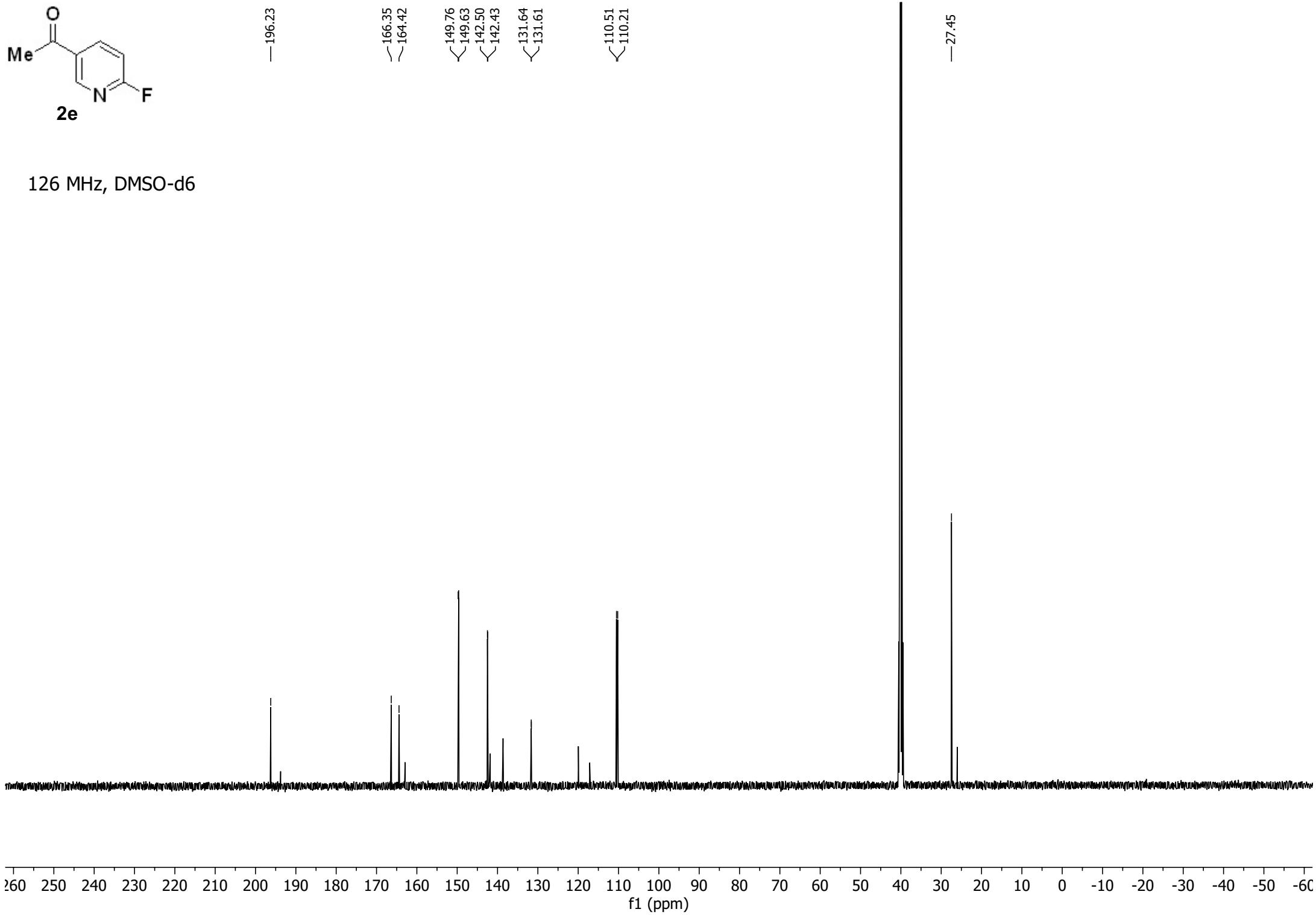
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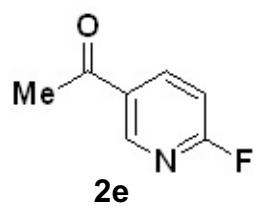
—166.35
—164.42
149.76
149.63
142.50
142.43
131.64
131.61

110.51
110.21

—27.45

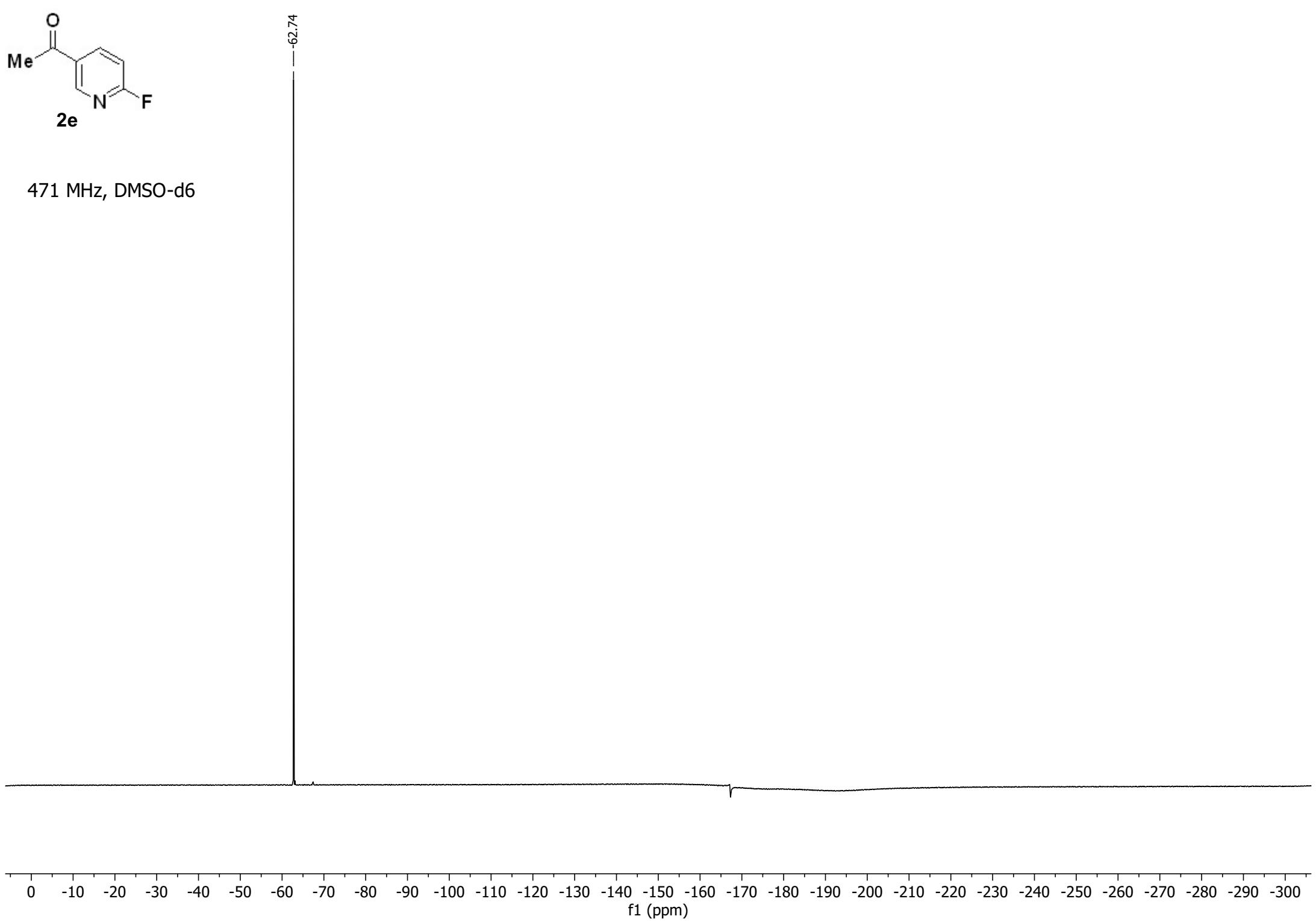
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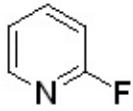




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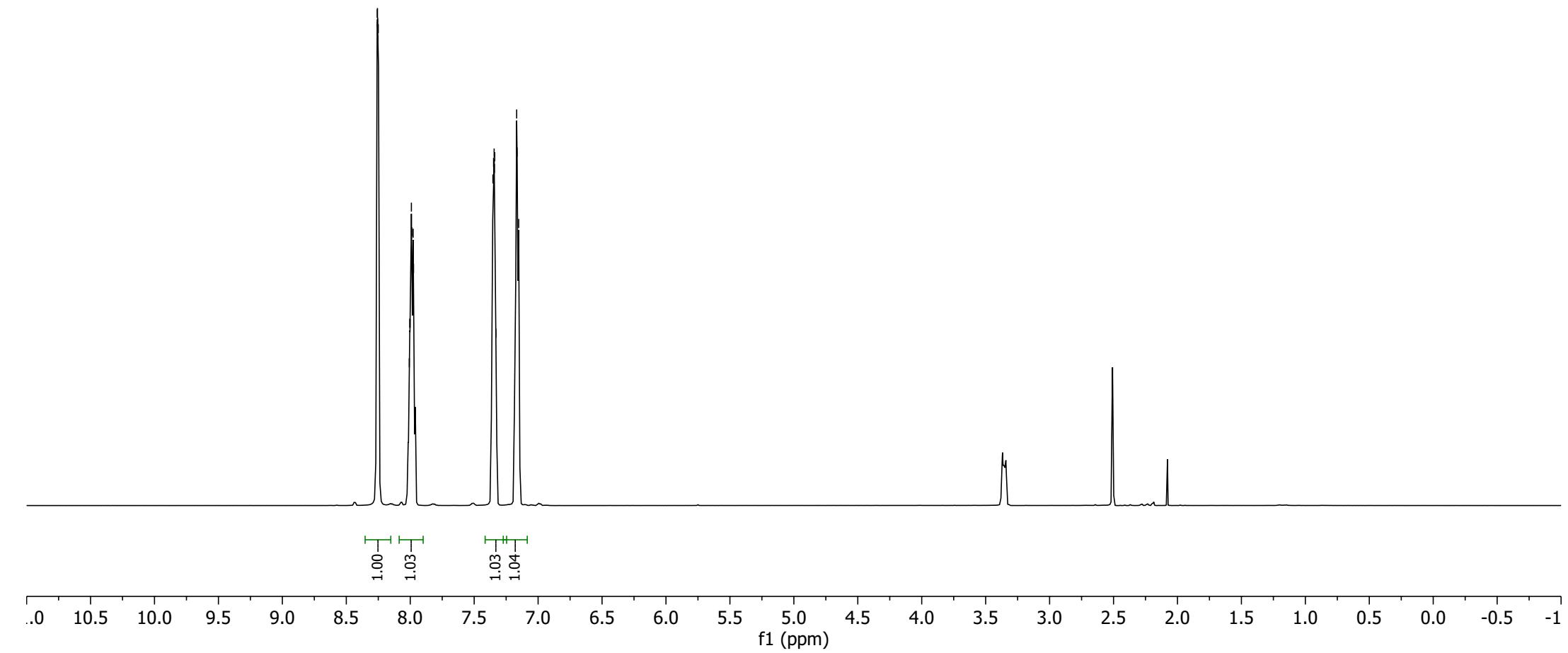
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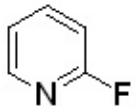




2f

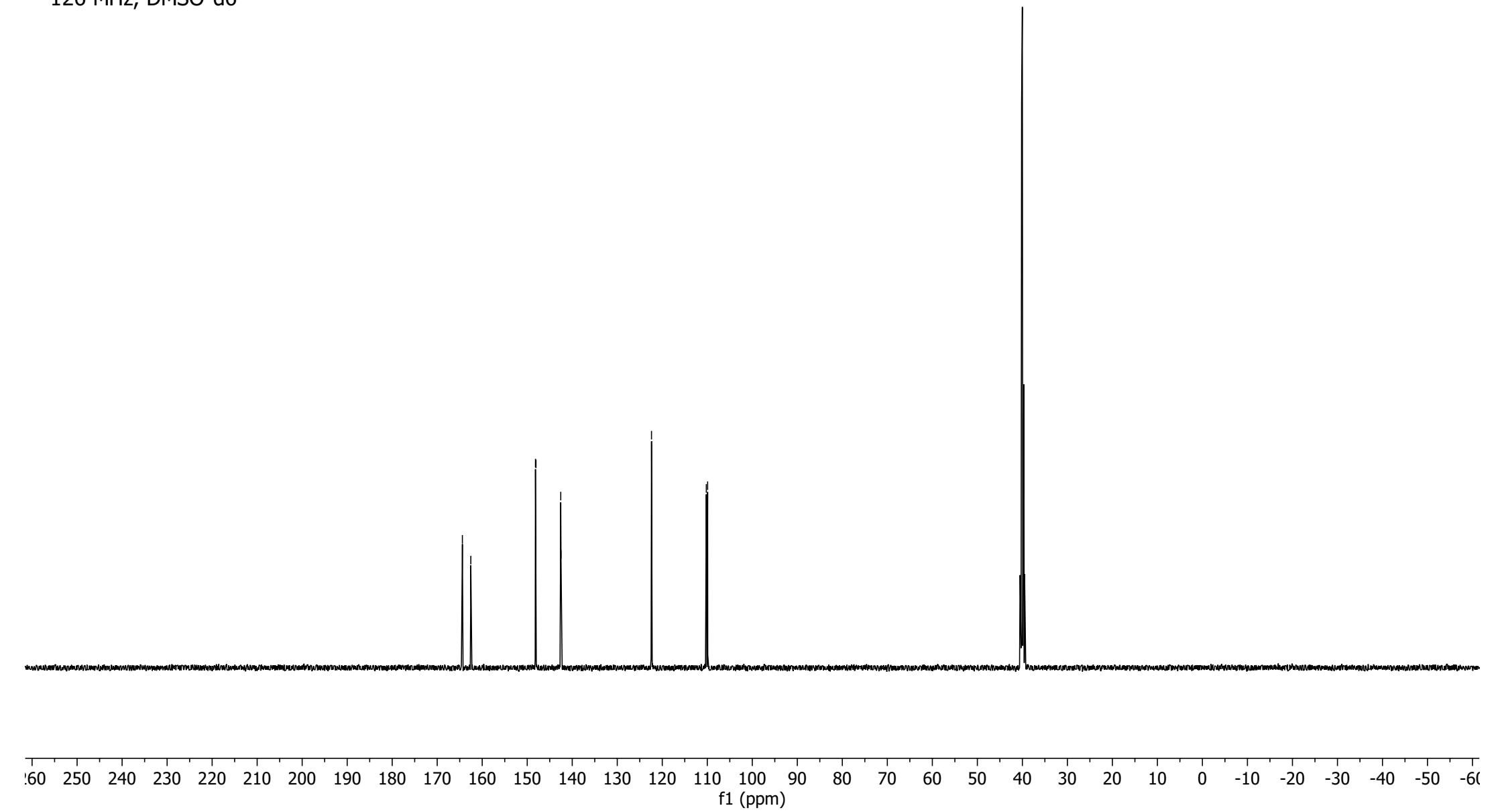
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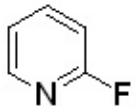




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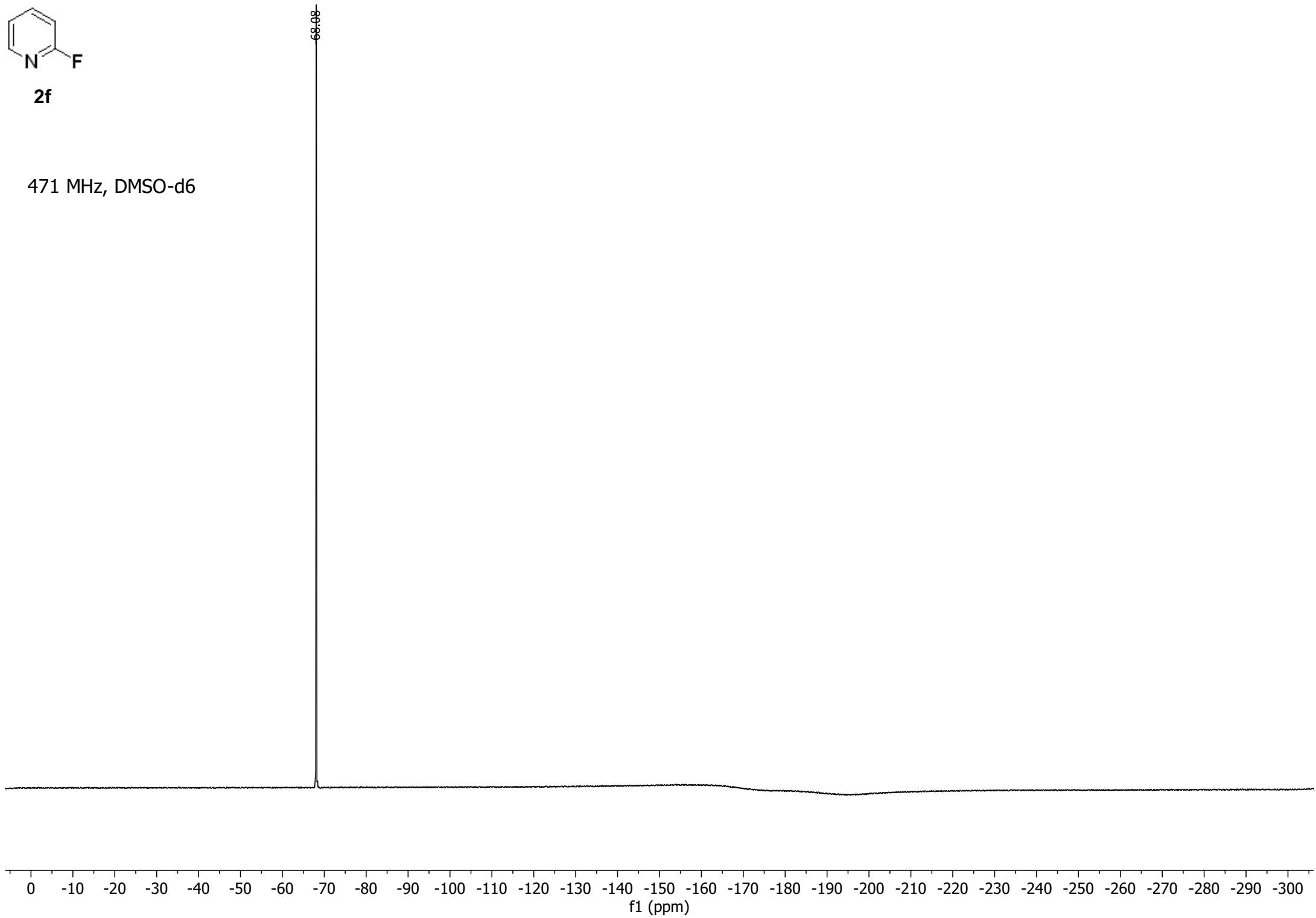
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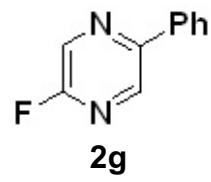




2f

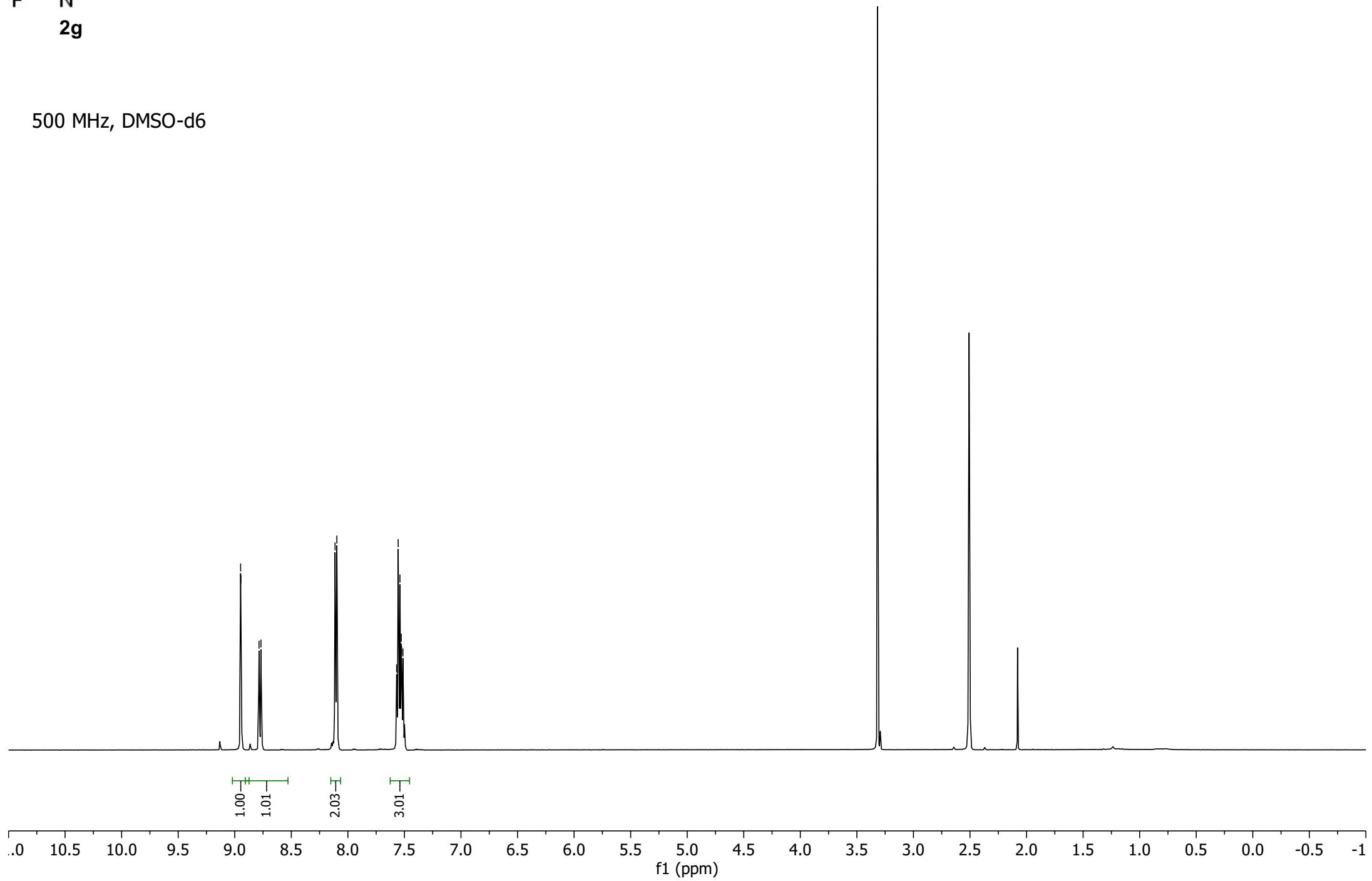
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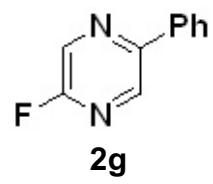




8.95
8.95
8.79
8.77
8.11
8.10
7.57
7.56
7.54
7.53
7.53
7.51

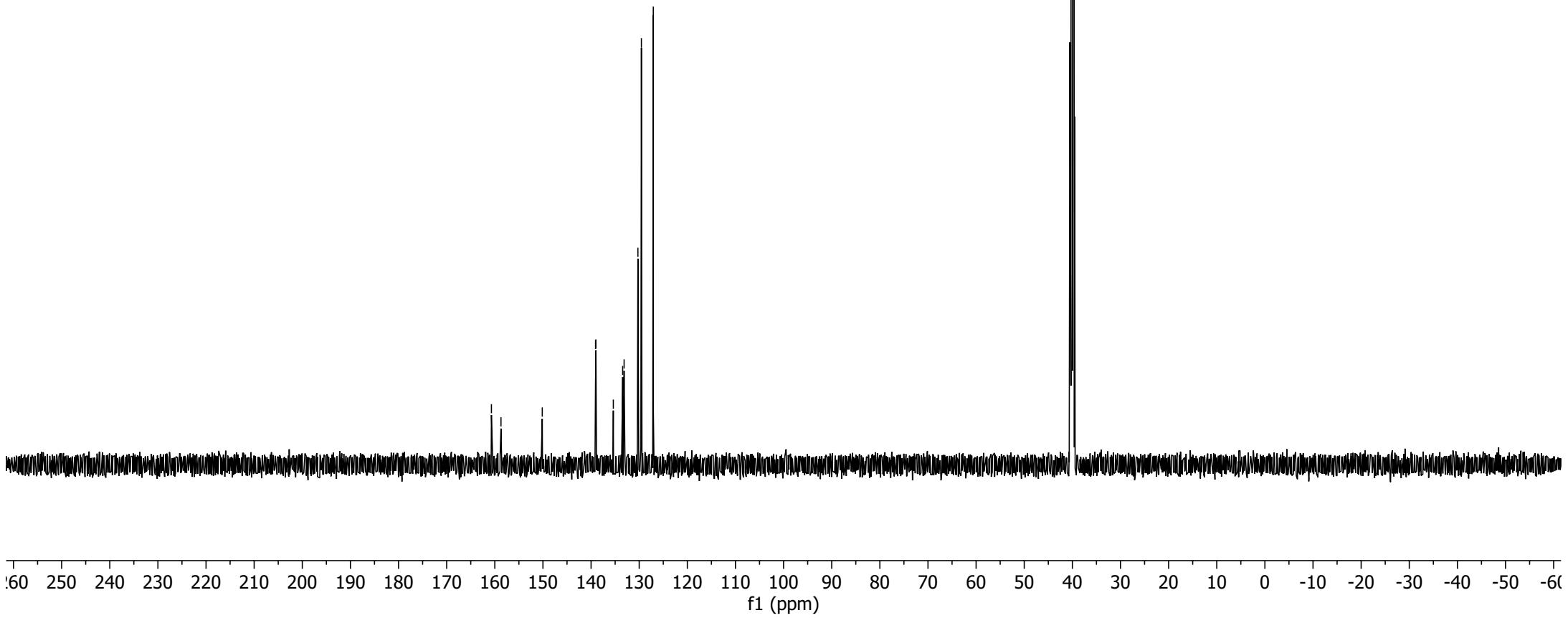
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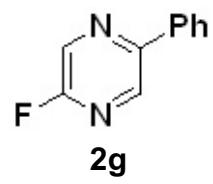




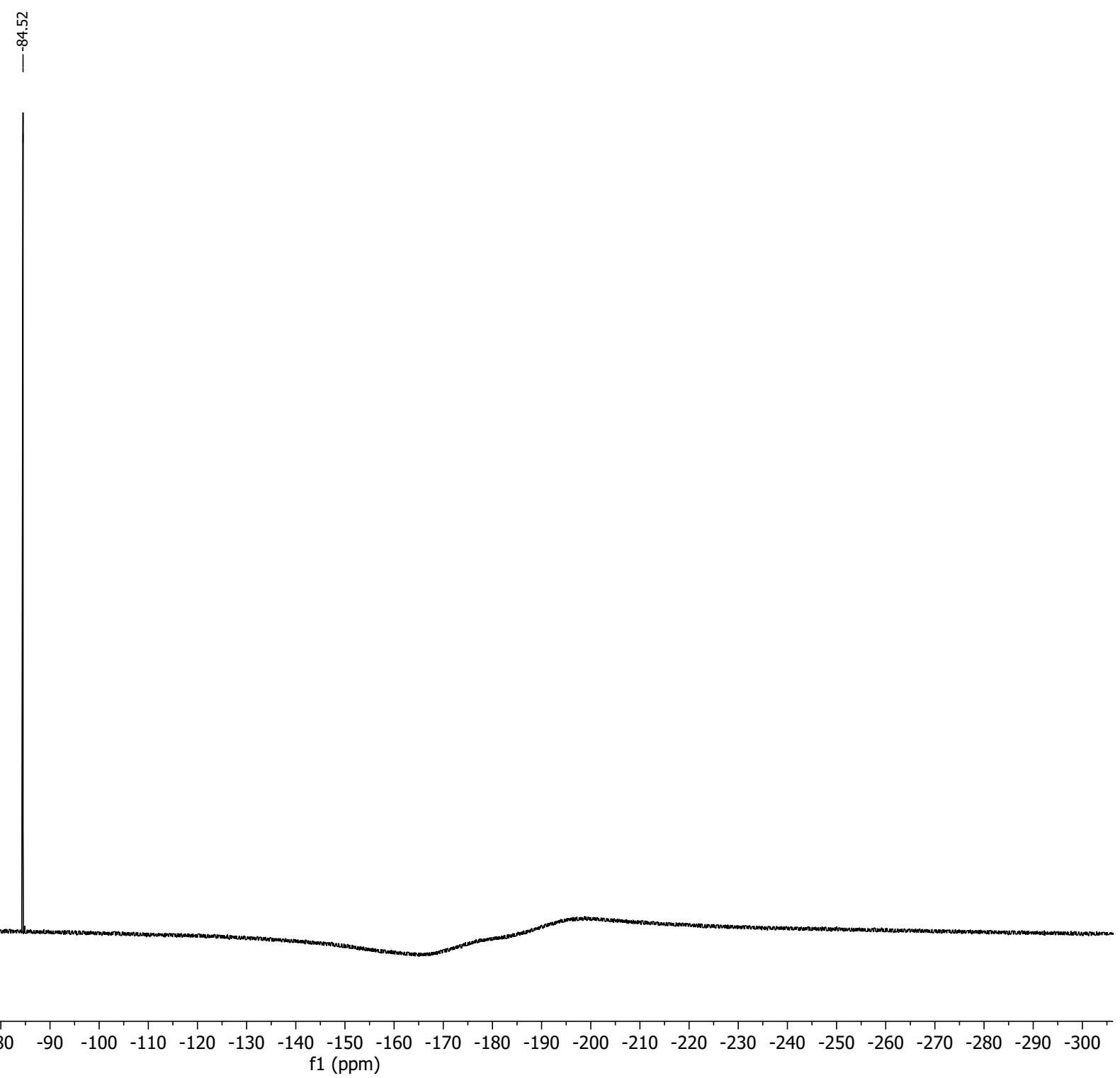
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-150.14
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139.00
135.38
133.44
133.14
130.27
129.55
127.08

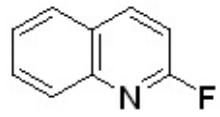
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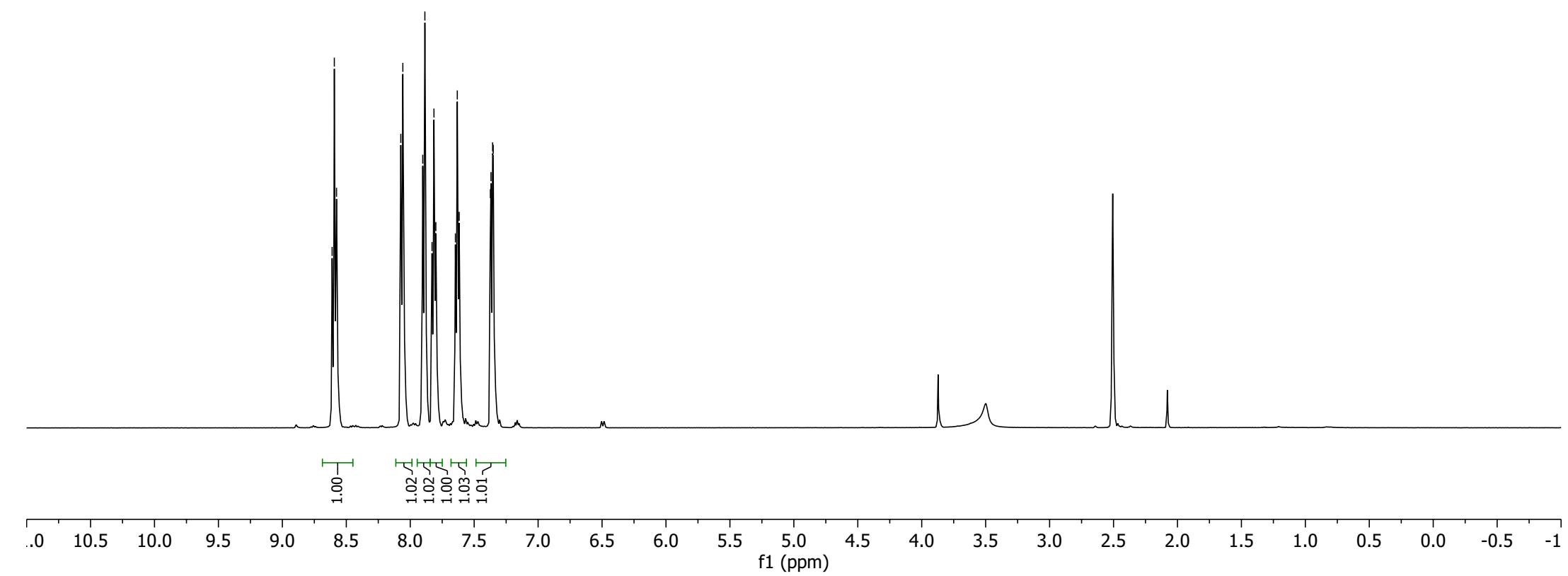
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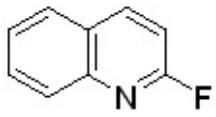




2h

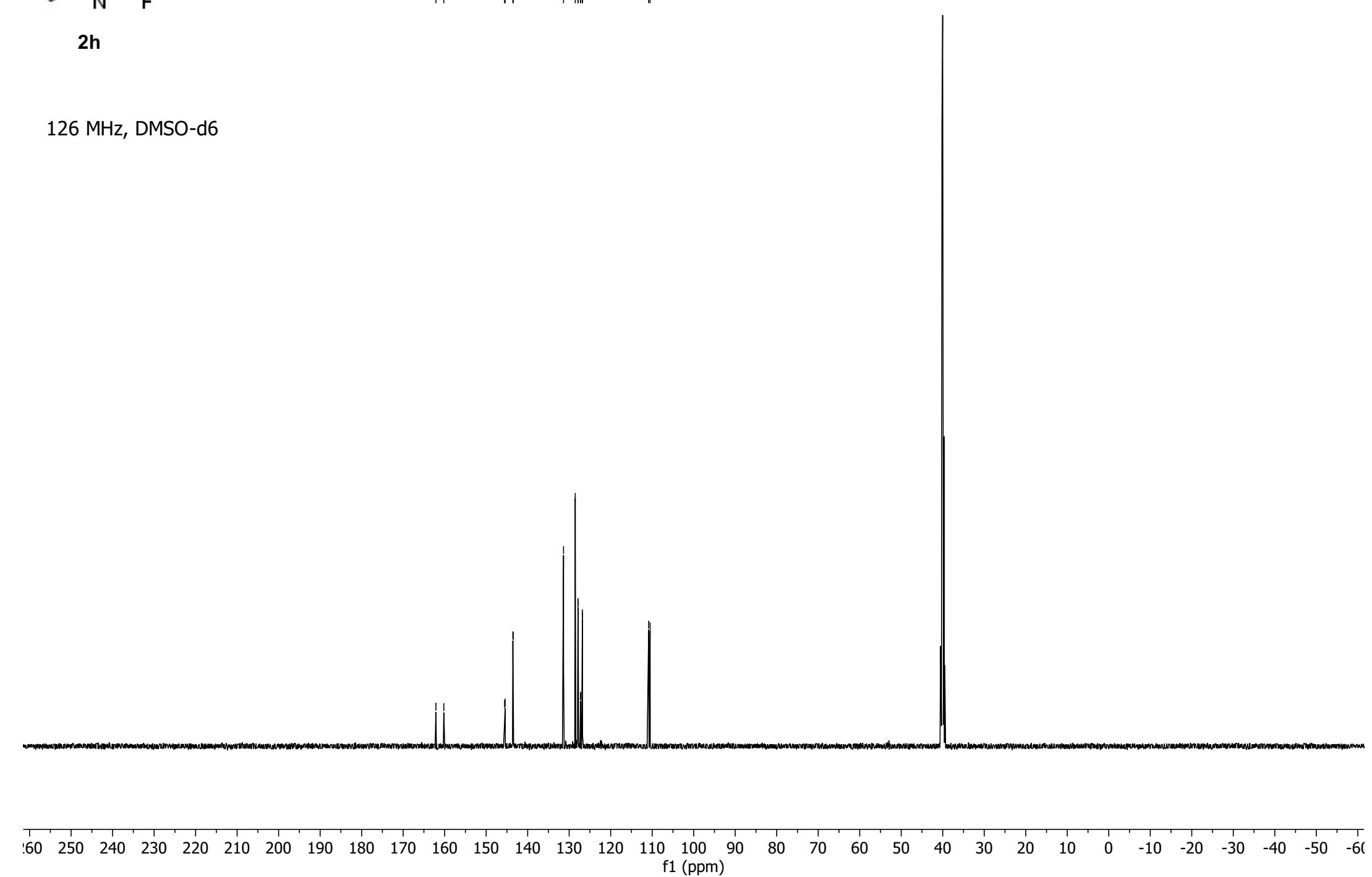
500 MHz, DMSO-d6

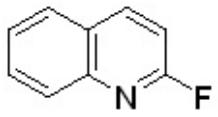




2h

126 MHz, DMSO-d6





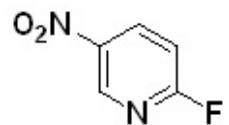
2h

471 MHz, DMSO-d6

-61.96

0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 -230 -240 -250 -260 -270 -280 -290 -300

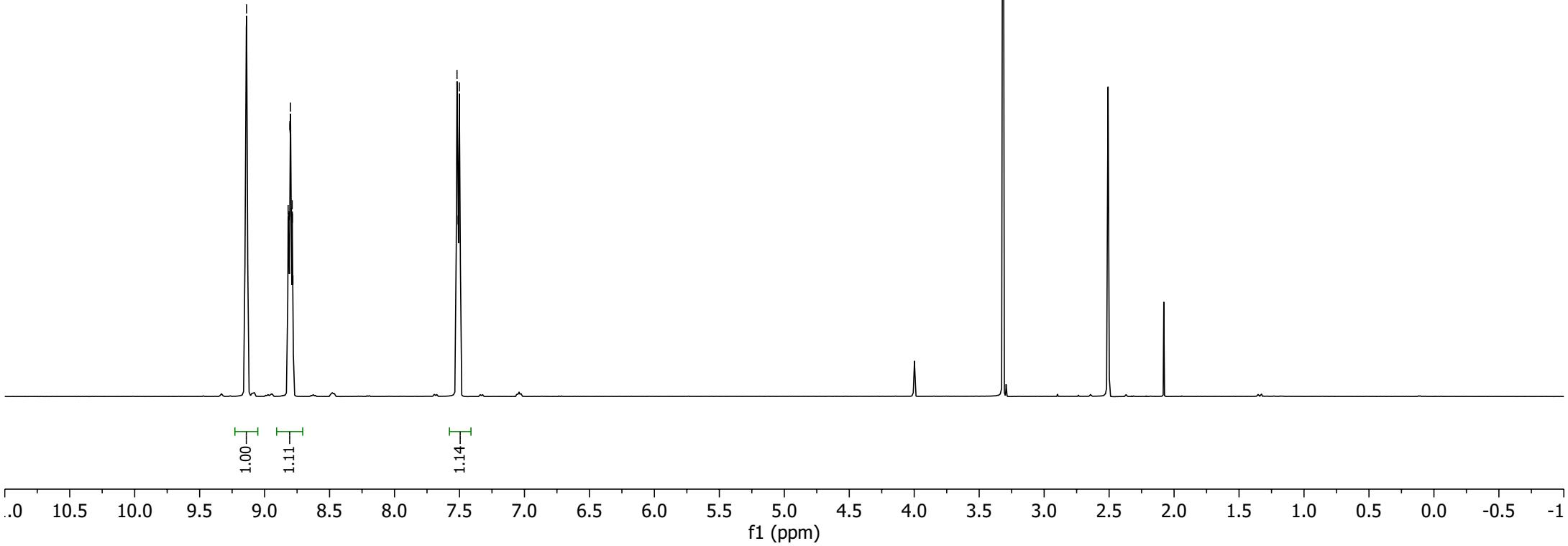
f1 (ppm)

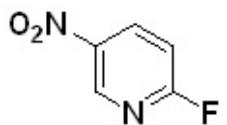


2i

—9.14
8.82
8.82
8.81
8.80
8.80
8.79
8.79
8.79
8.78
7.52
7.51
7.51
7.50

500 MHz, DMSO-d6

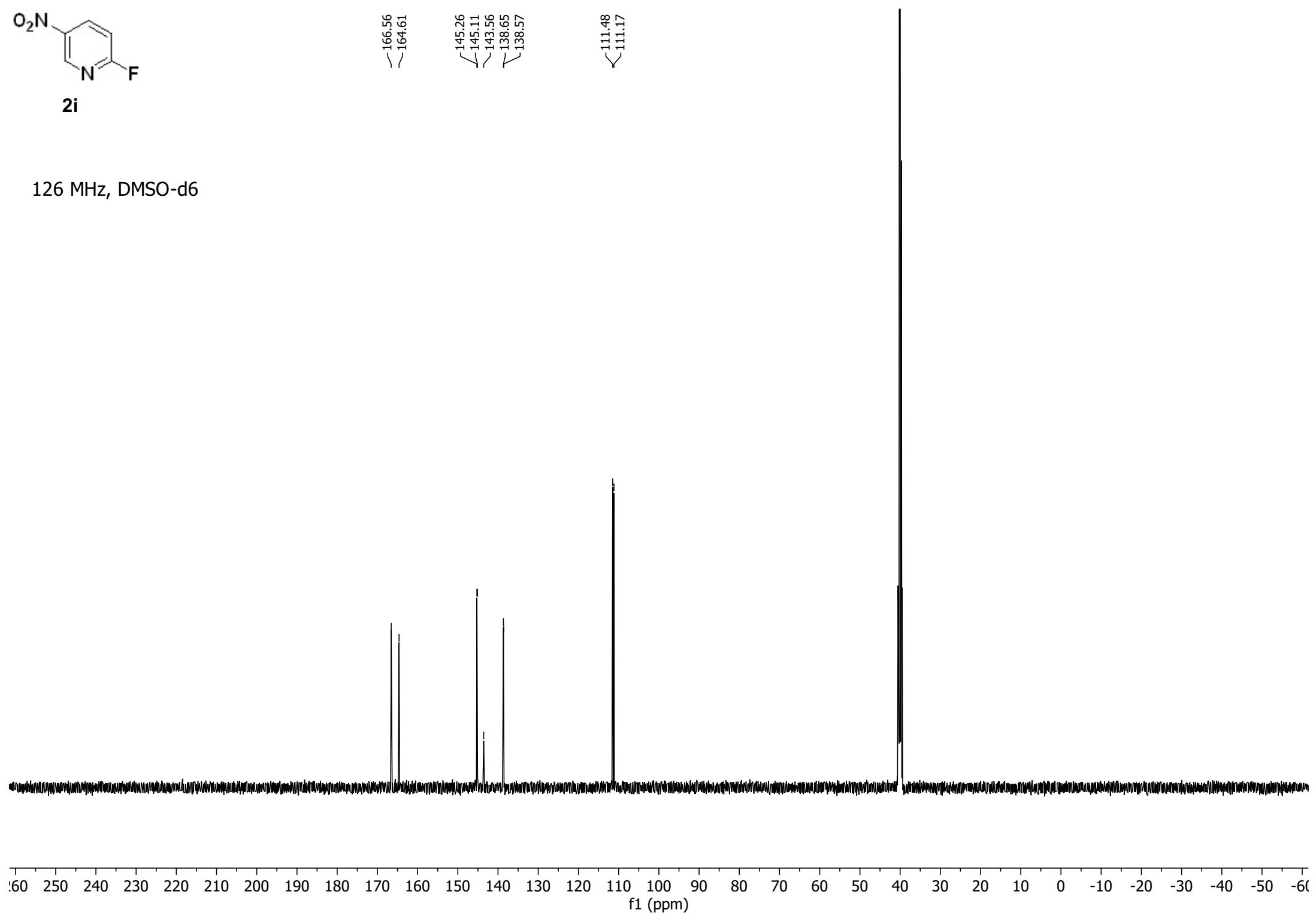




2i

126 MHz, DMSO-d6

166.56
~164.61
145.26
145.11
143.56
138.65
138.57
111.48
111.17



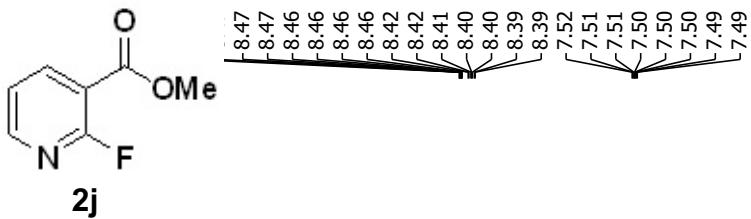


471 MHz, DMSO-d6

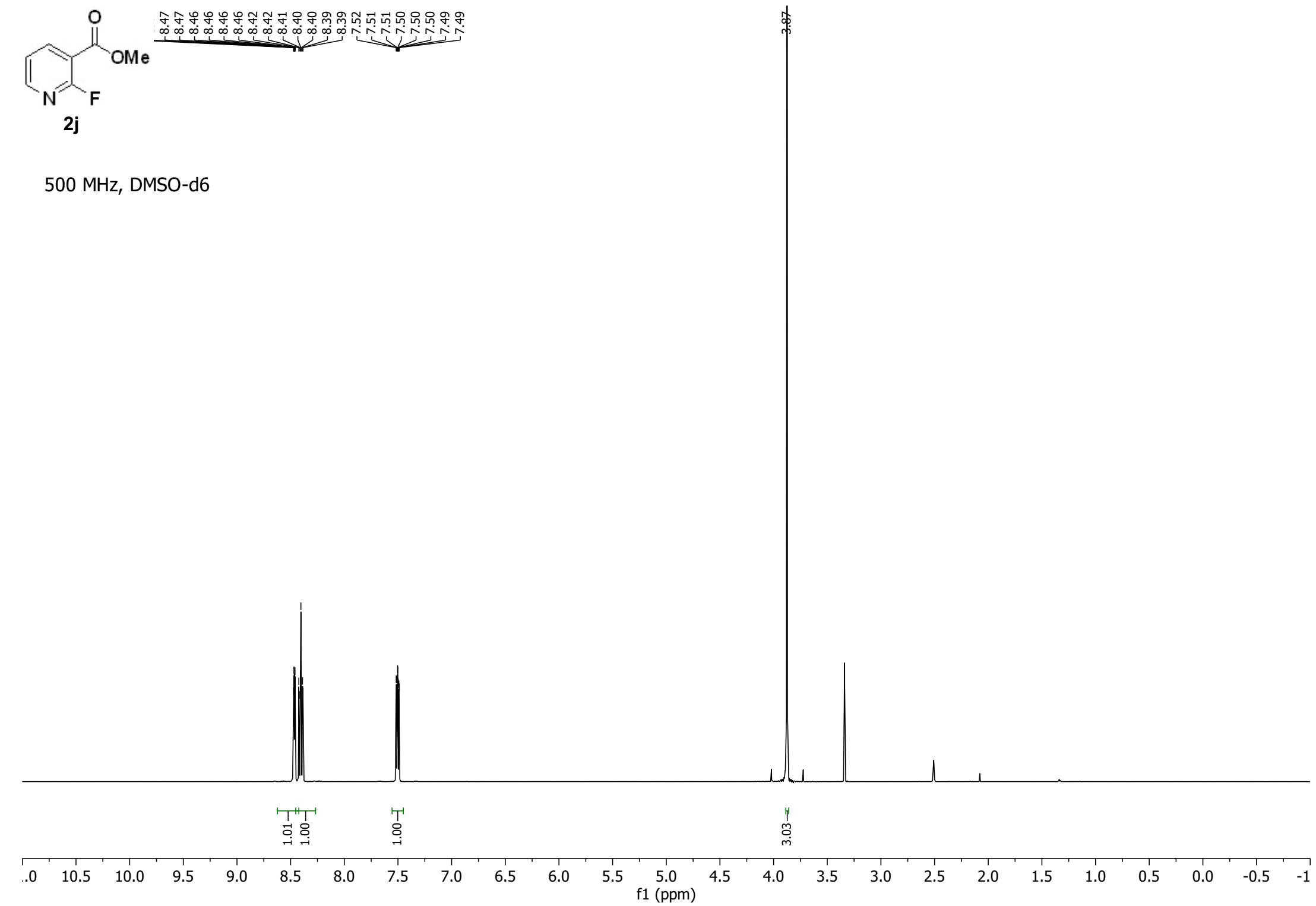
-59.84

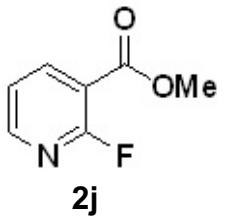
0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 -230 -240 -250 -260 -270 -280 -290 -300

f1 (ppm)

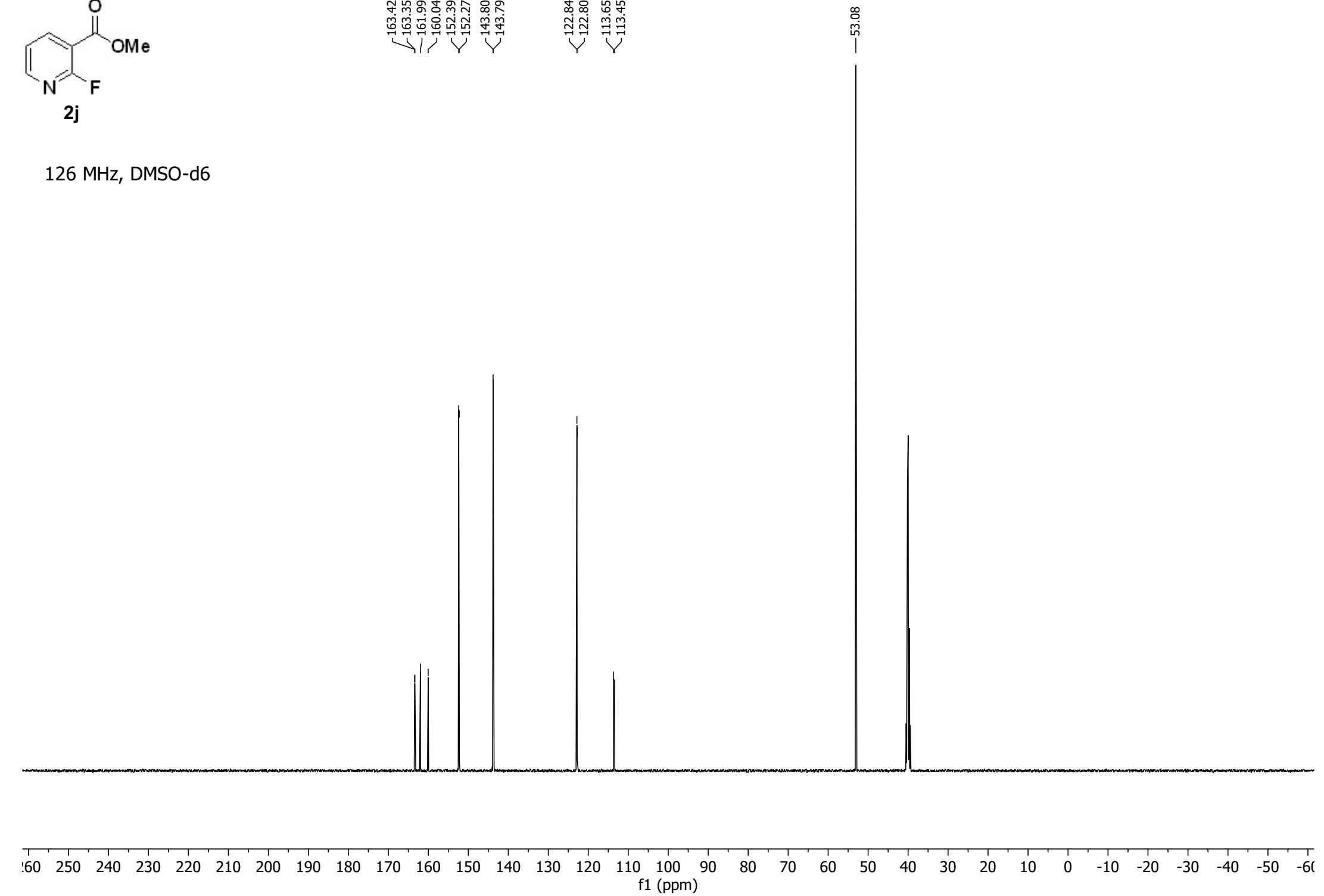


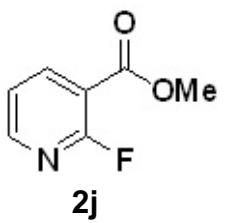
500 MHz, DMSO-d6





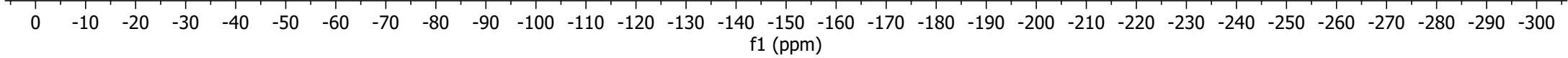
126 MHz, DMSO-d6

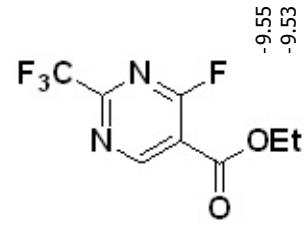




471 MHz, DMSO-d6

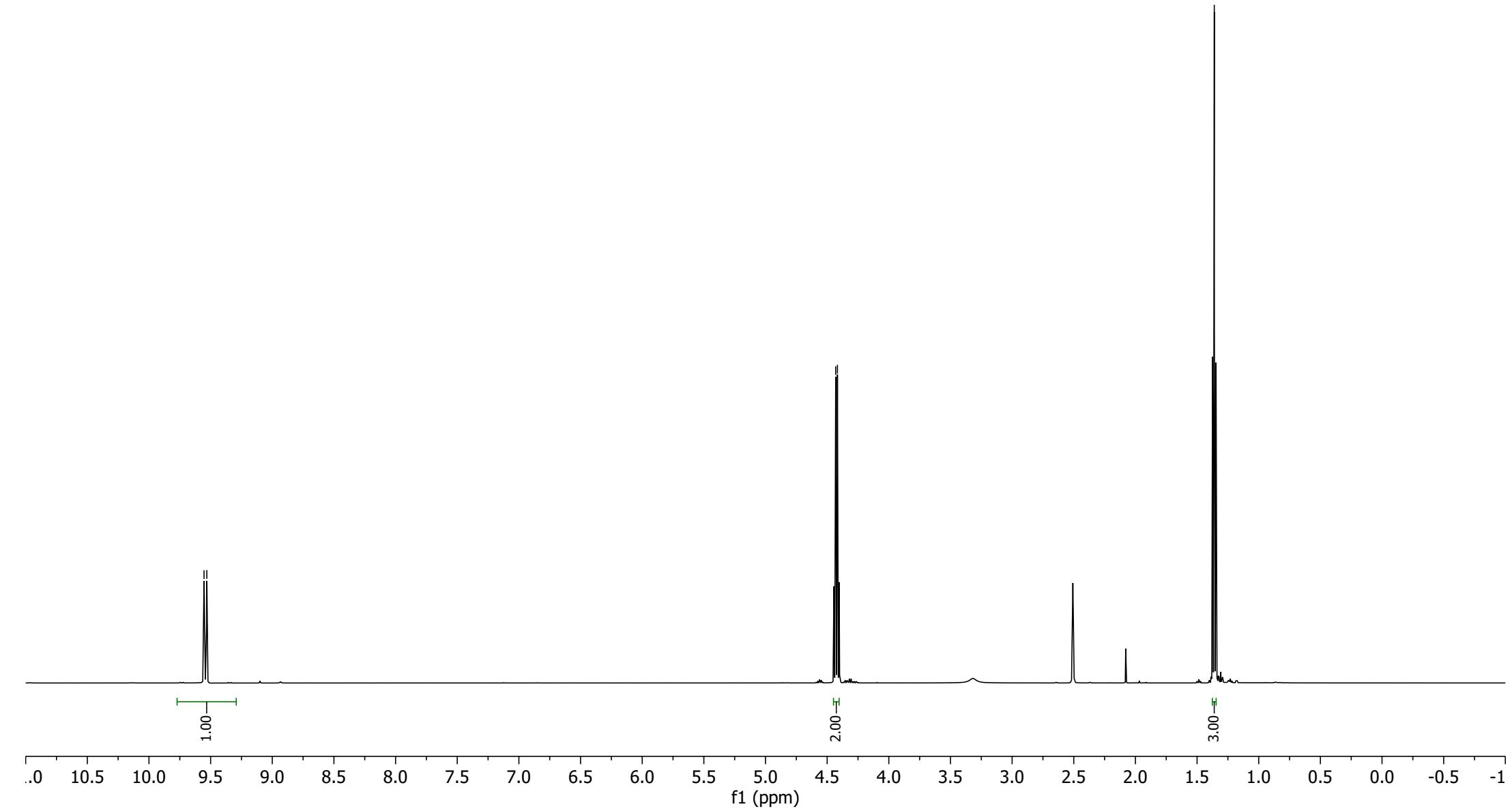
-64.12

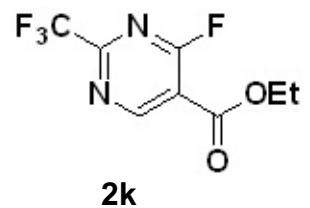




2k

500 MHz, DMSO-d6





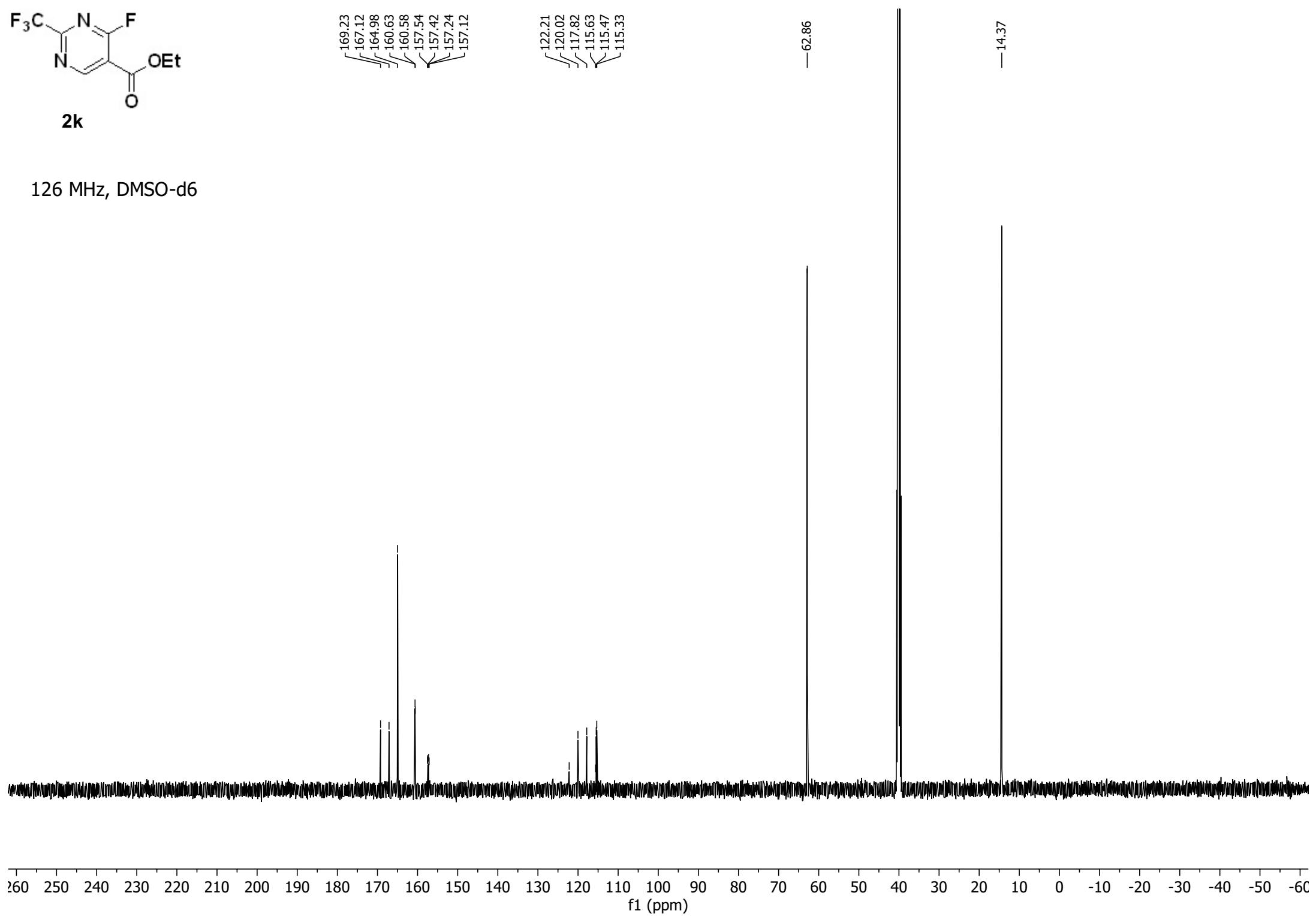
169.23
167.12
164.98
160.63
160.58
157.54
157.42
157.24
157.12

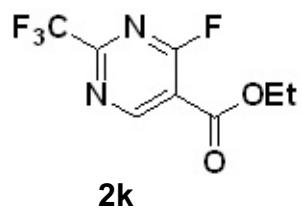
122.21
120.02
117.82
115.63
115.47
115.33

—62.86

—14.37

126 MHz, DMSO-d₆

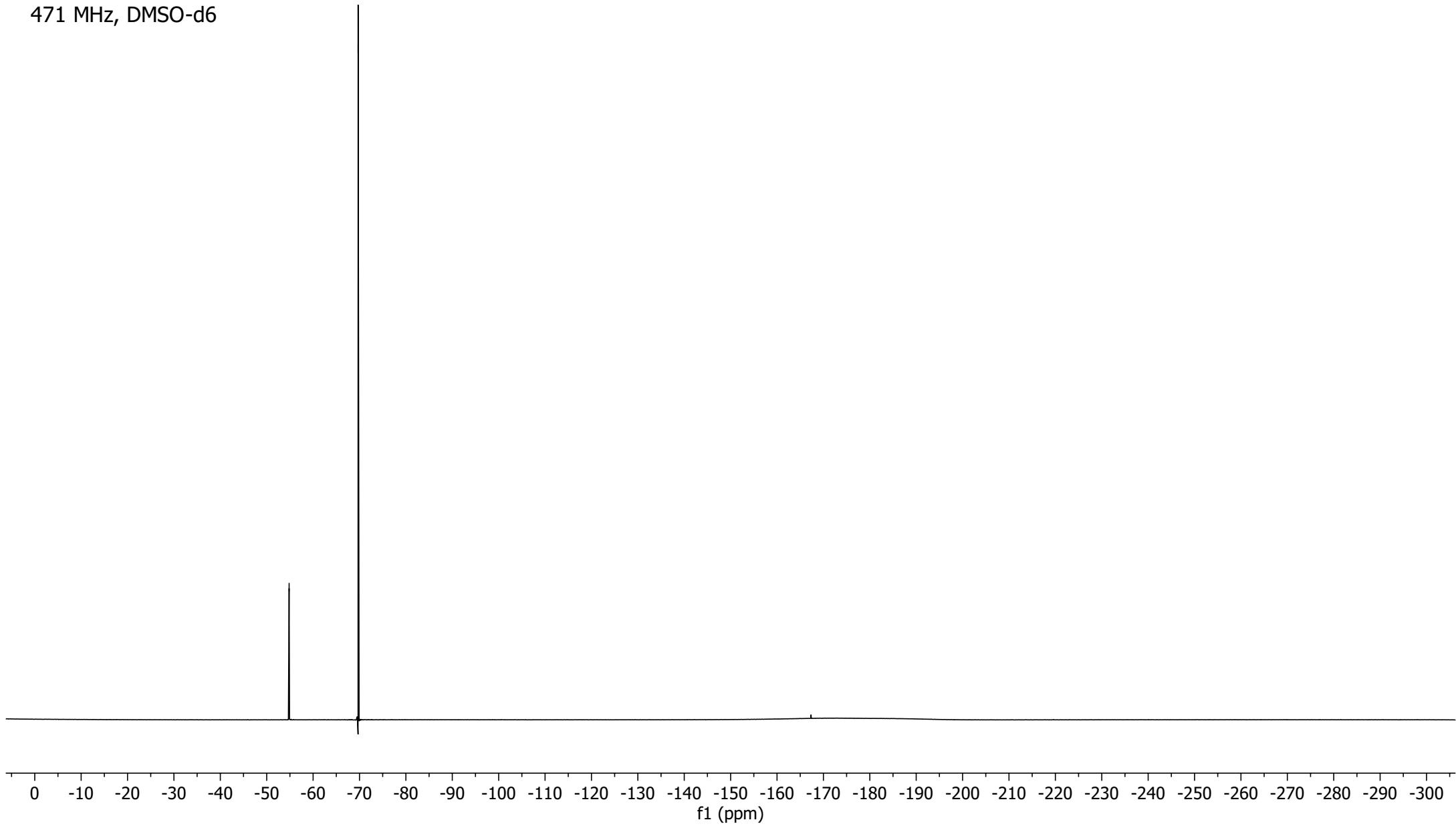


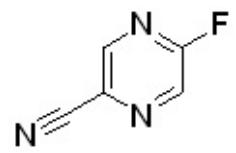


-54.83

-69.70

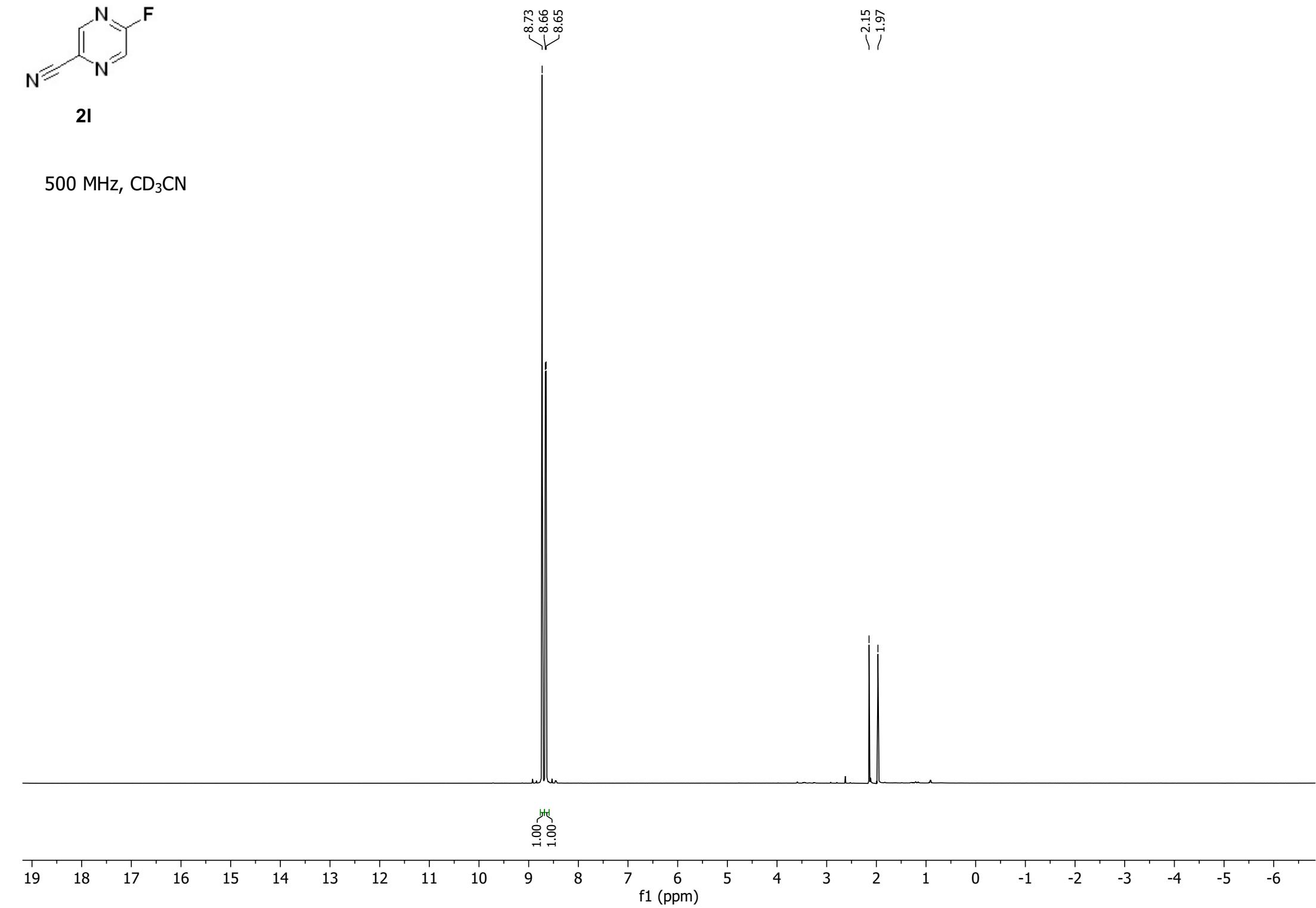
471 MHz, DMSO-d6

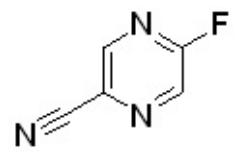




2l

500 MHz, CD₃CN





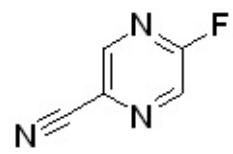
2l

126 MHz, CD₃CN

161.71
~159.65
147.00
146.90
136.08
135.76
127.62
127.58
117.30
115.07
115.06

260 250 240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10 -20 -30 -40 -50 -60

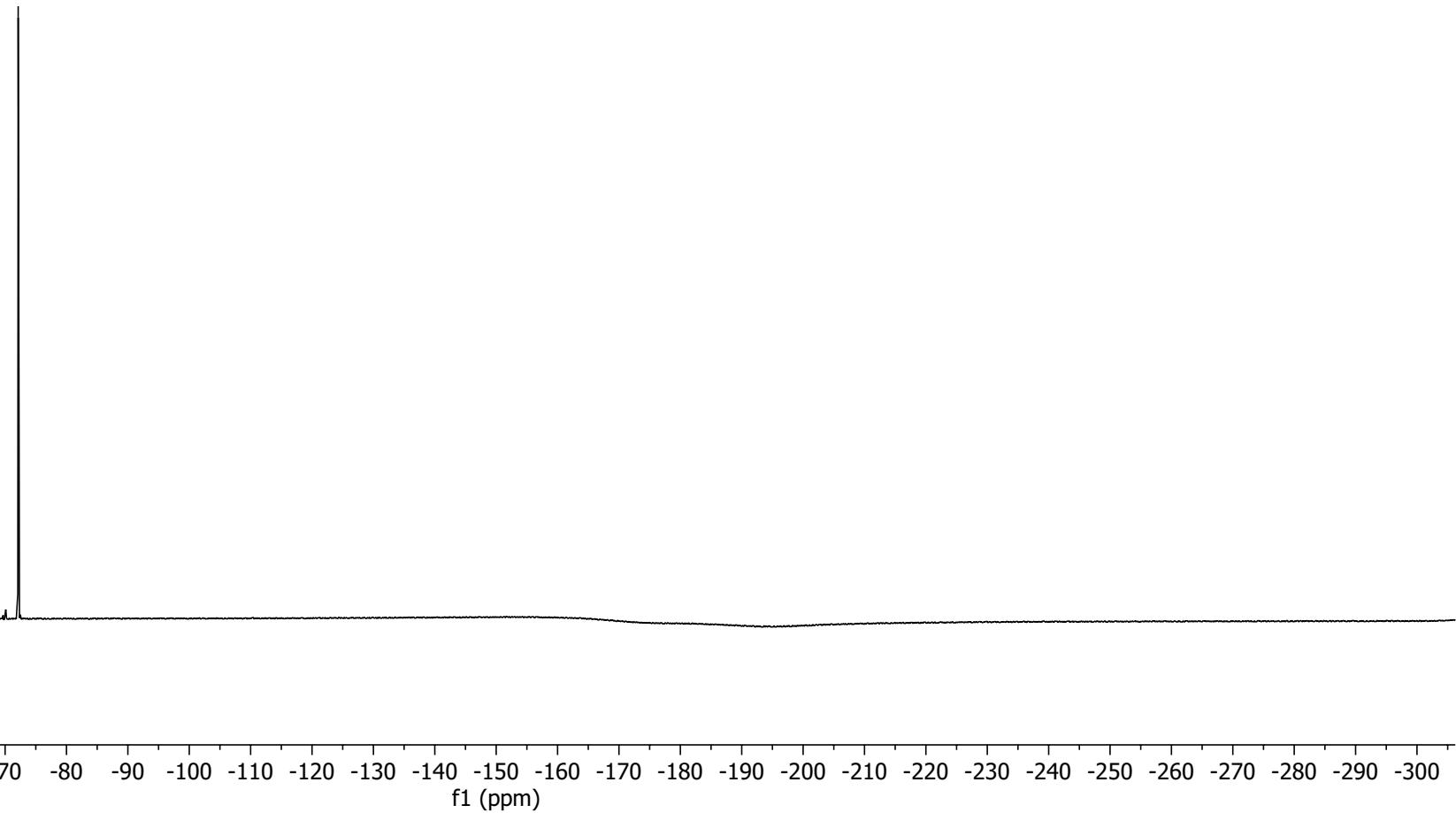
f1 (ppm)



-72.13

2l

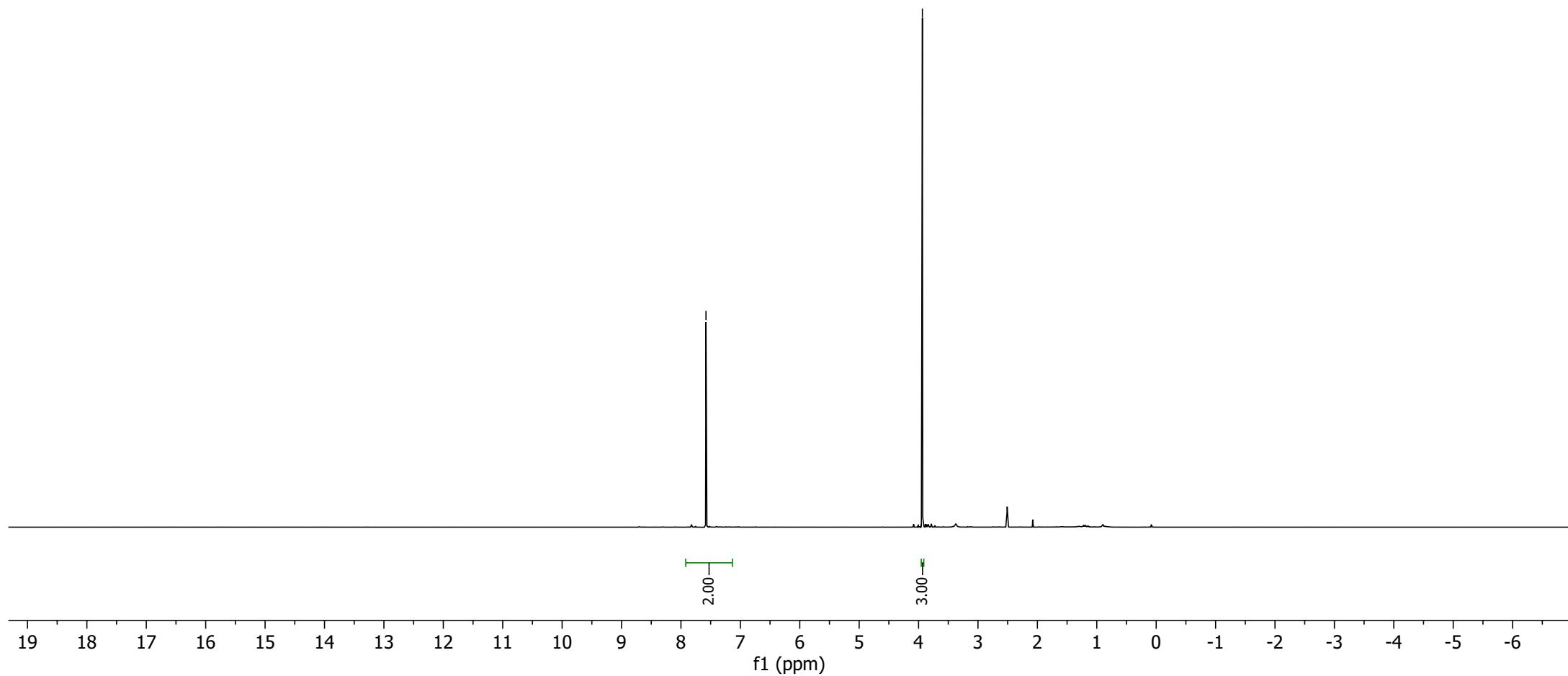
471 MHz, CD₃CN





2m

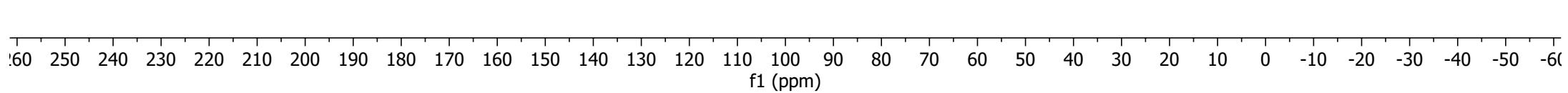
500 MHz, DMSO-d6





2m

126 MHz, DMSO-d6

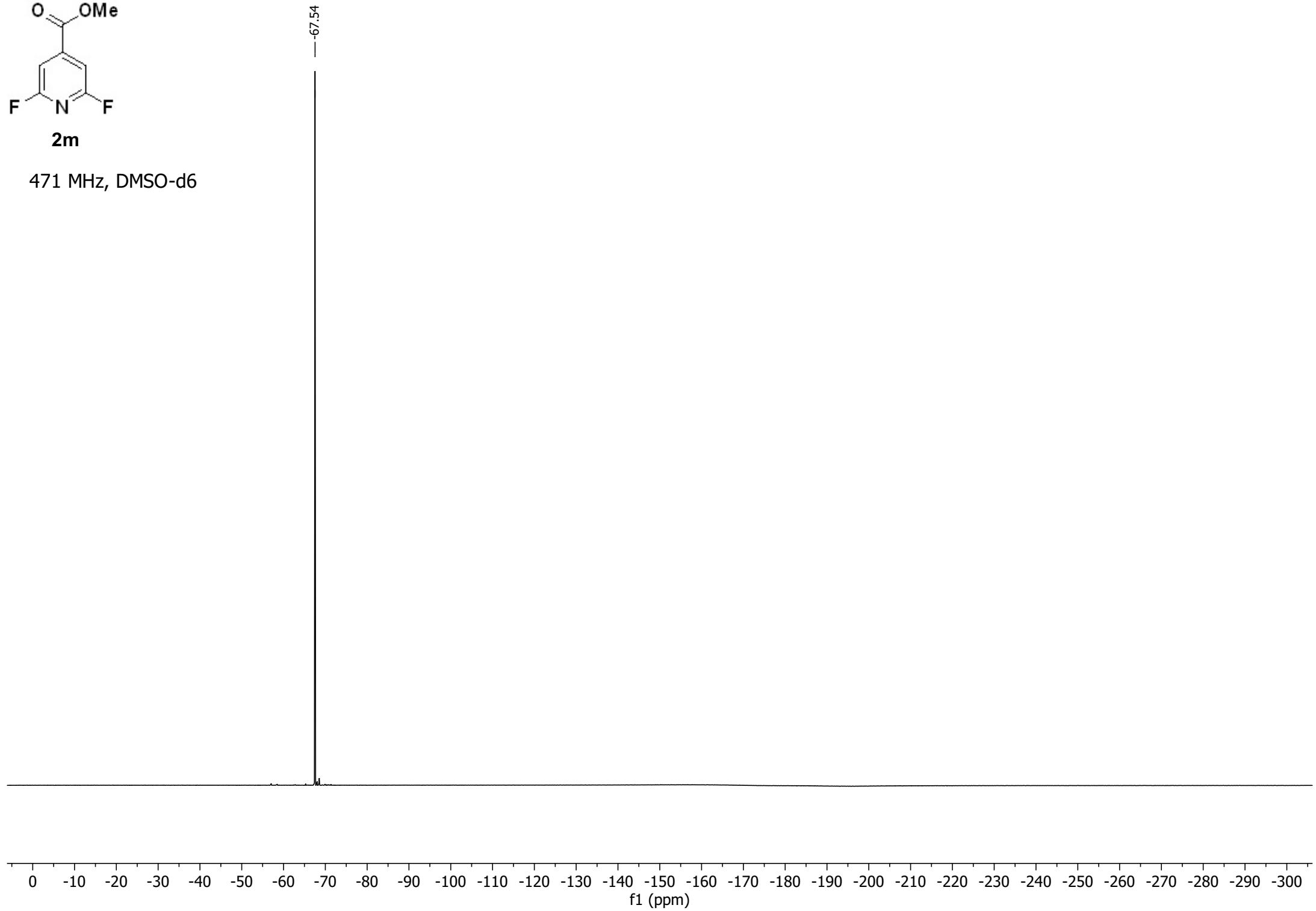


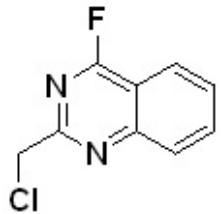


2m

471 MHz, DMSO-d6

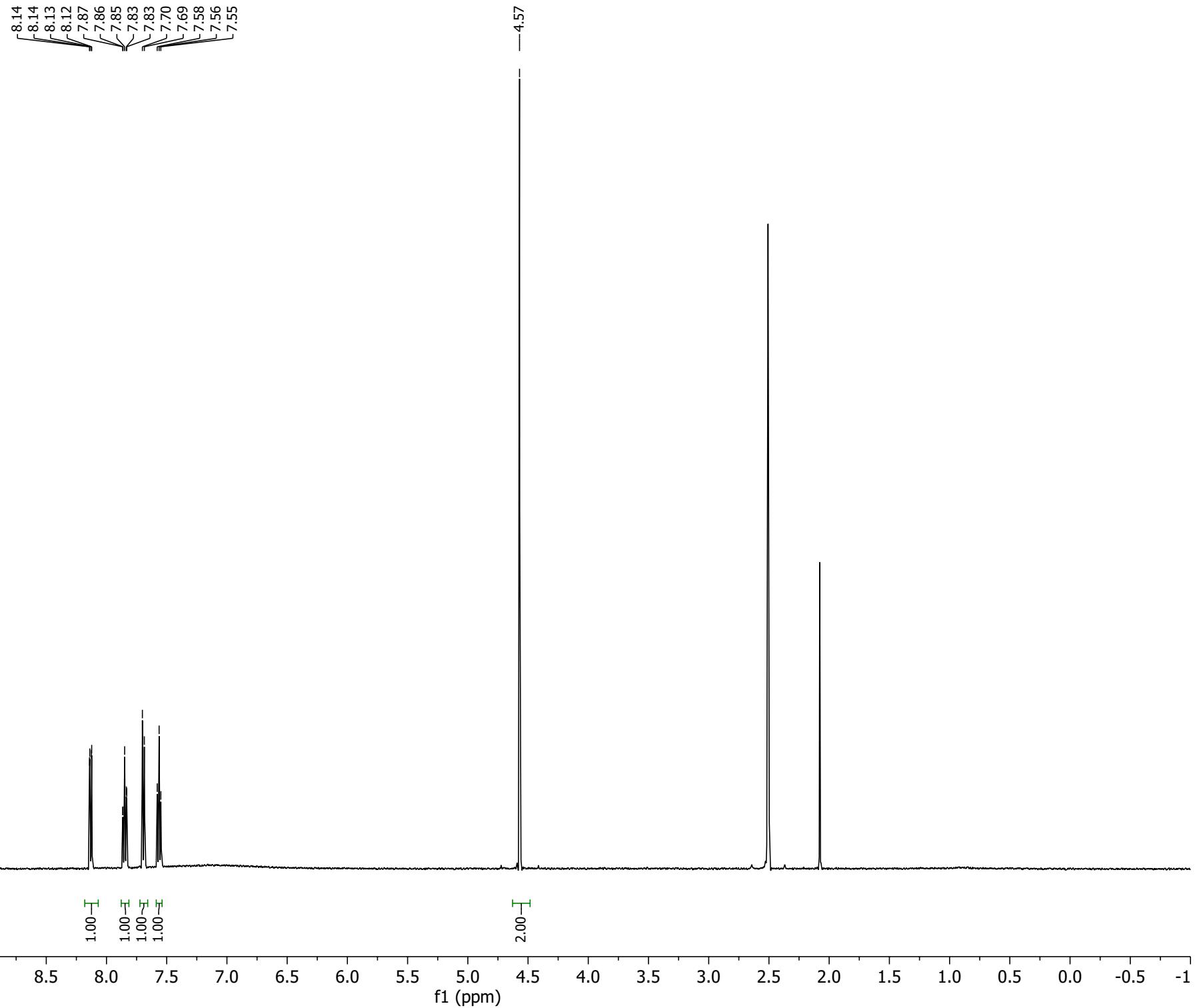
-67.54

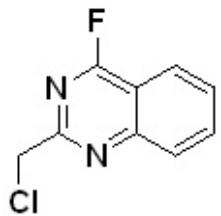




2n

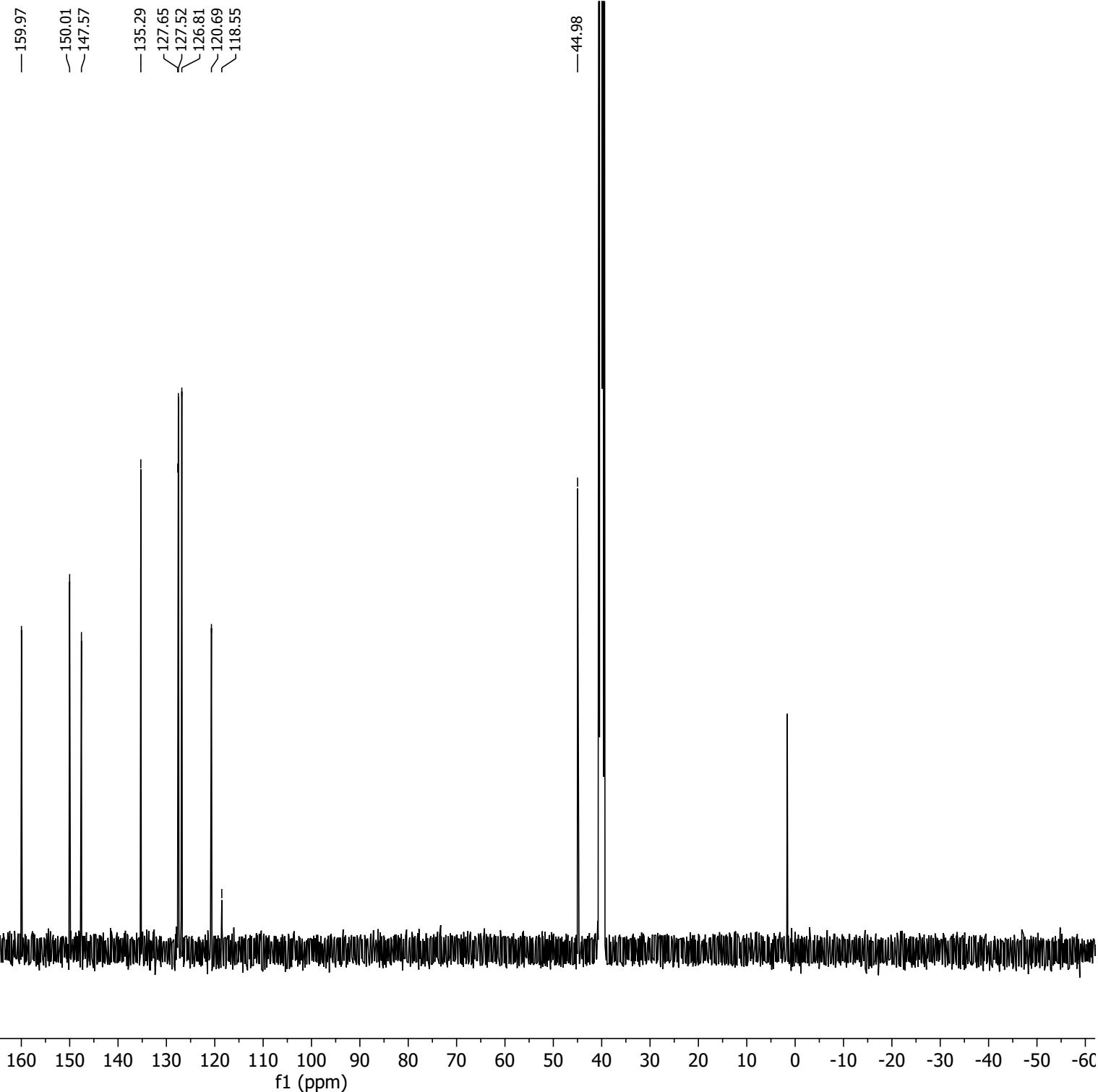
500 MHz, DMSO-d6





2n

126 MHz, DMSO-d6

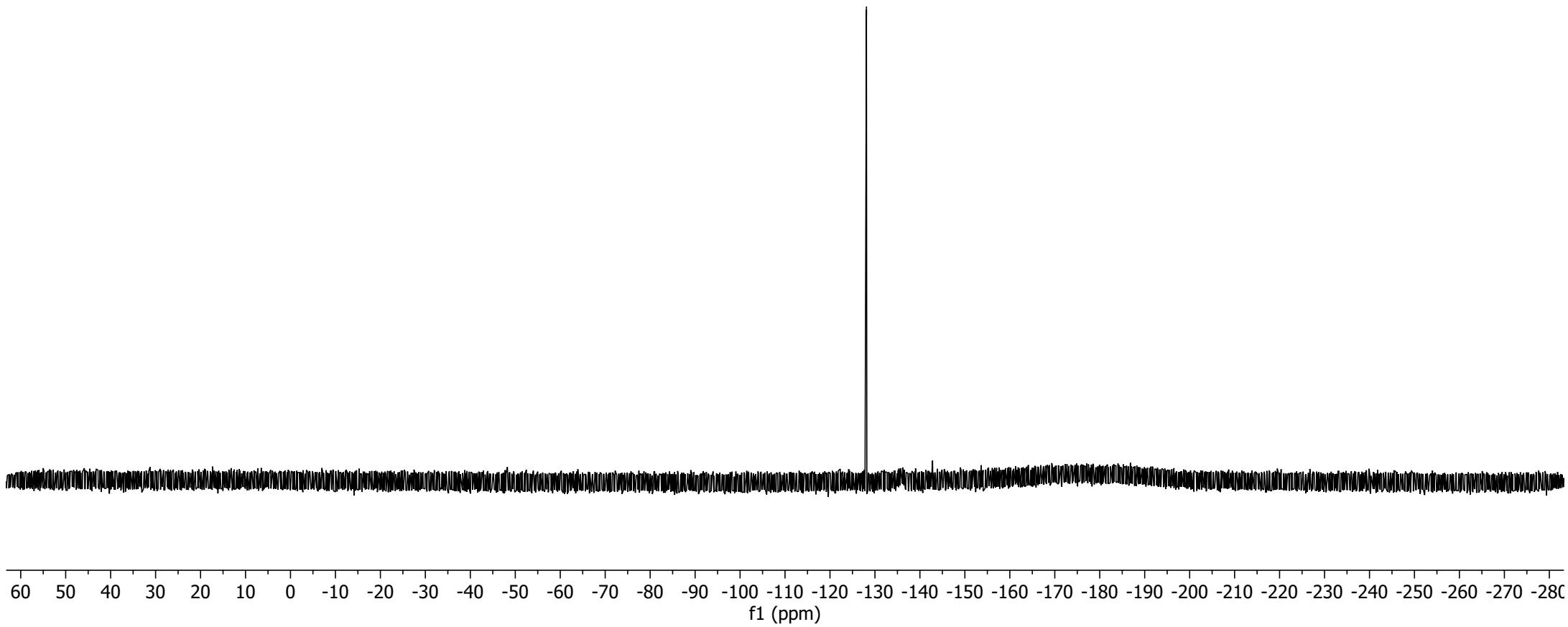


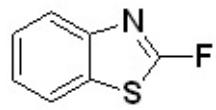


2n

471 MHz, DMSO-d6

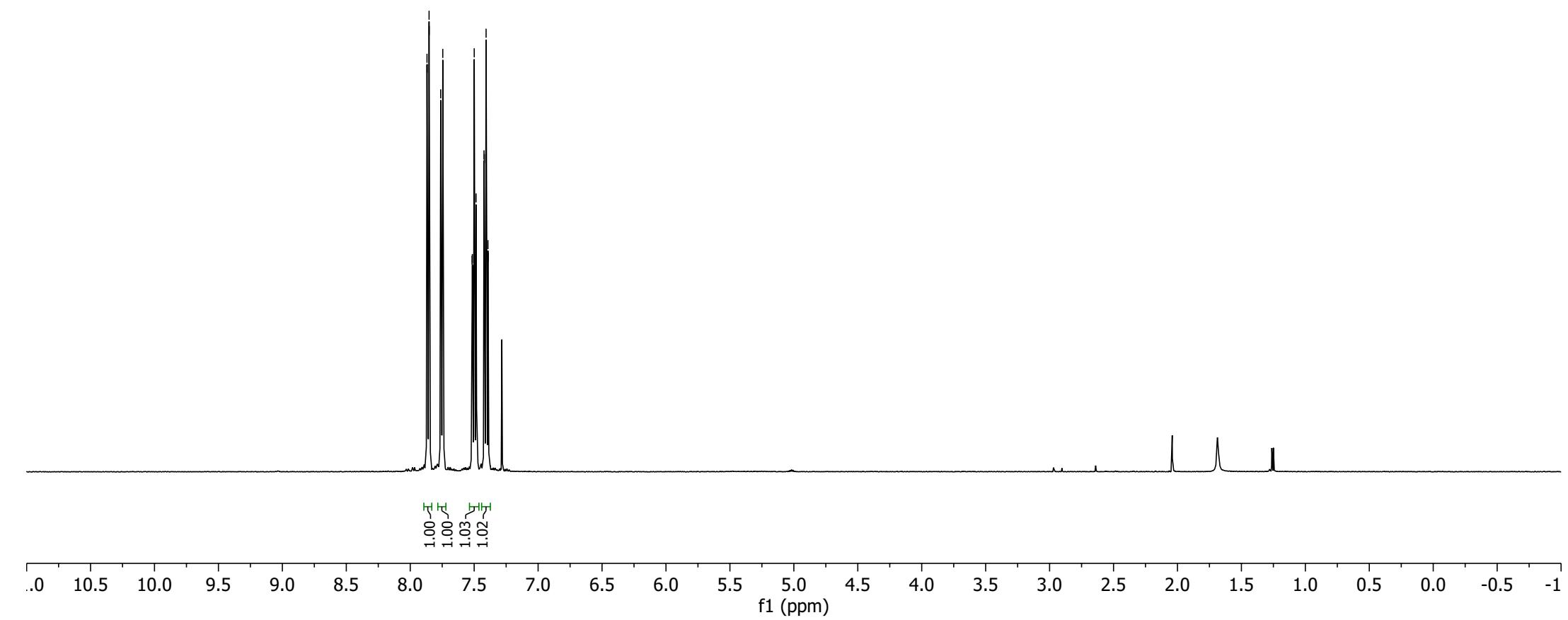
-128.11

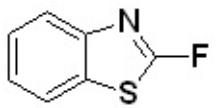




2o

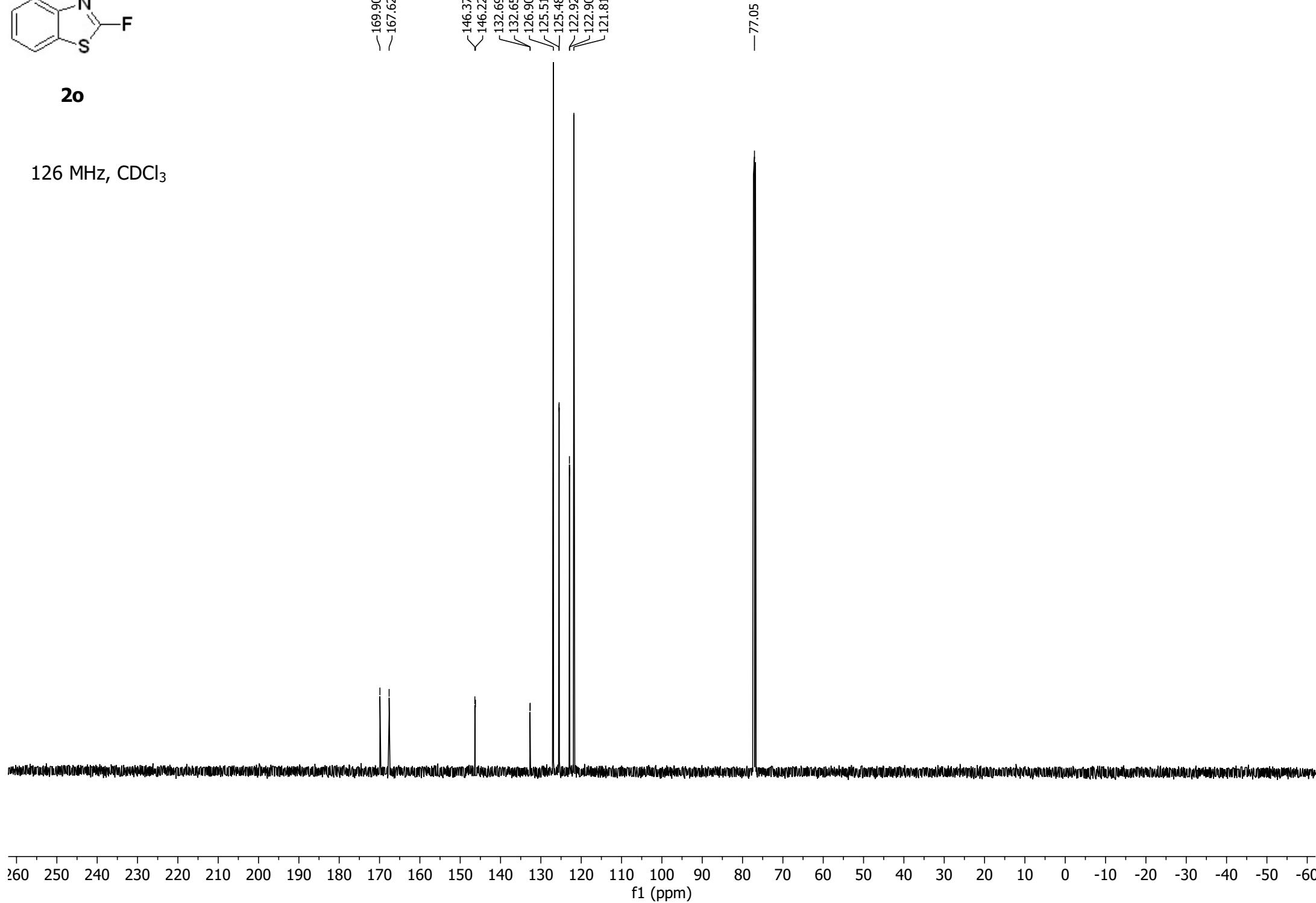
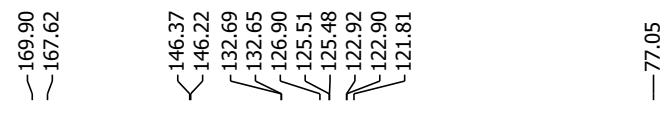
500 MHz, CDCl₃

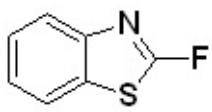




2o

126 MHz, CDCl₃

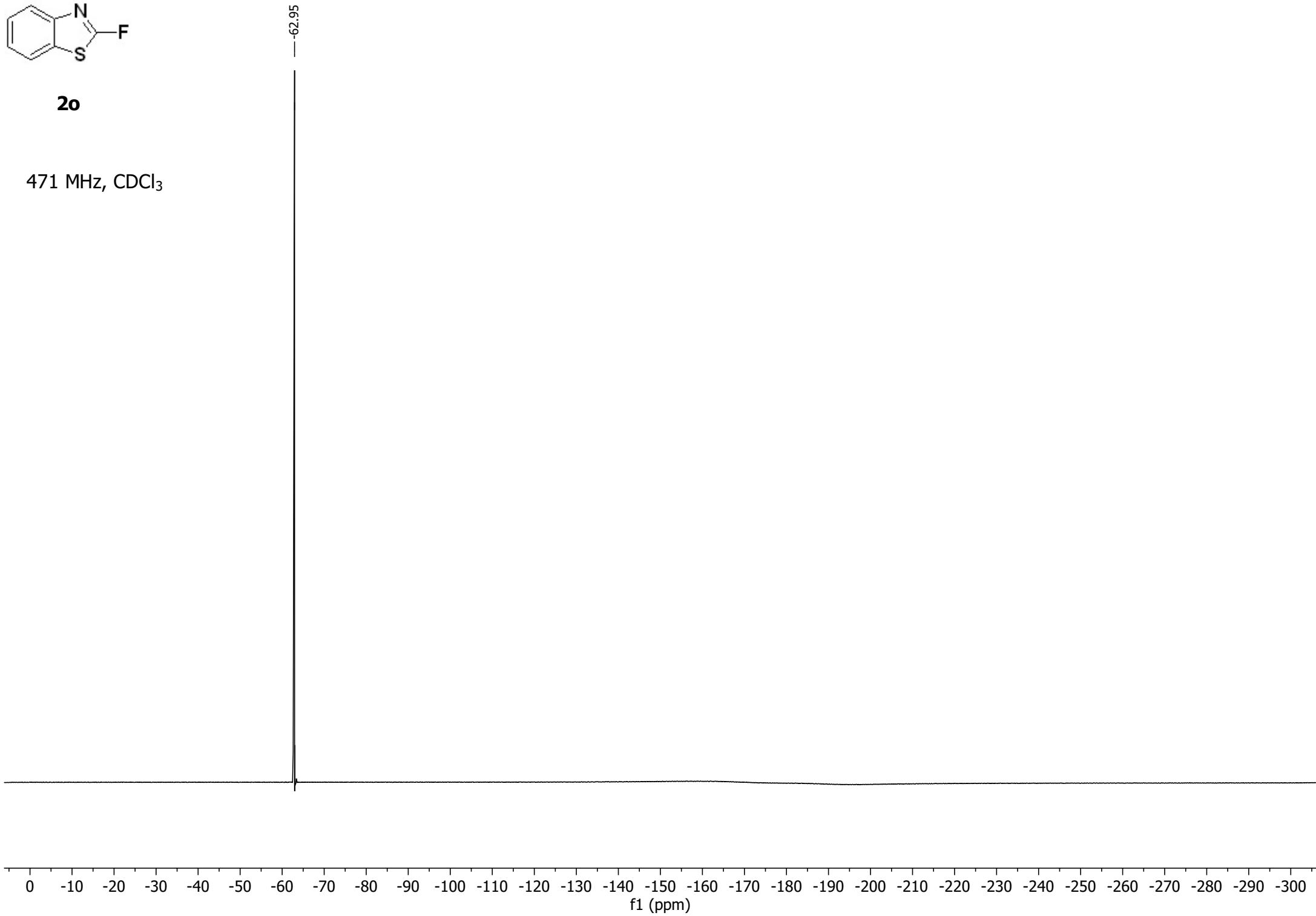


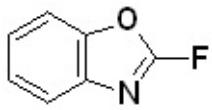


2o

471 MHz, CDCl₃

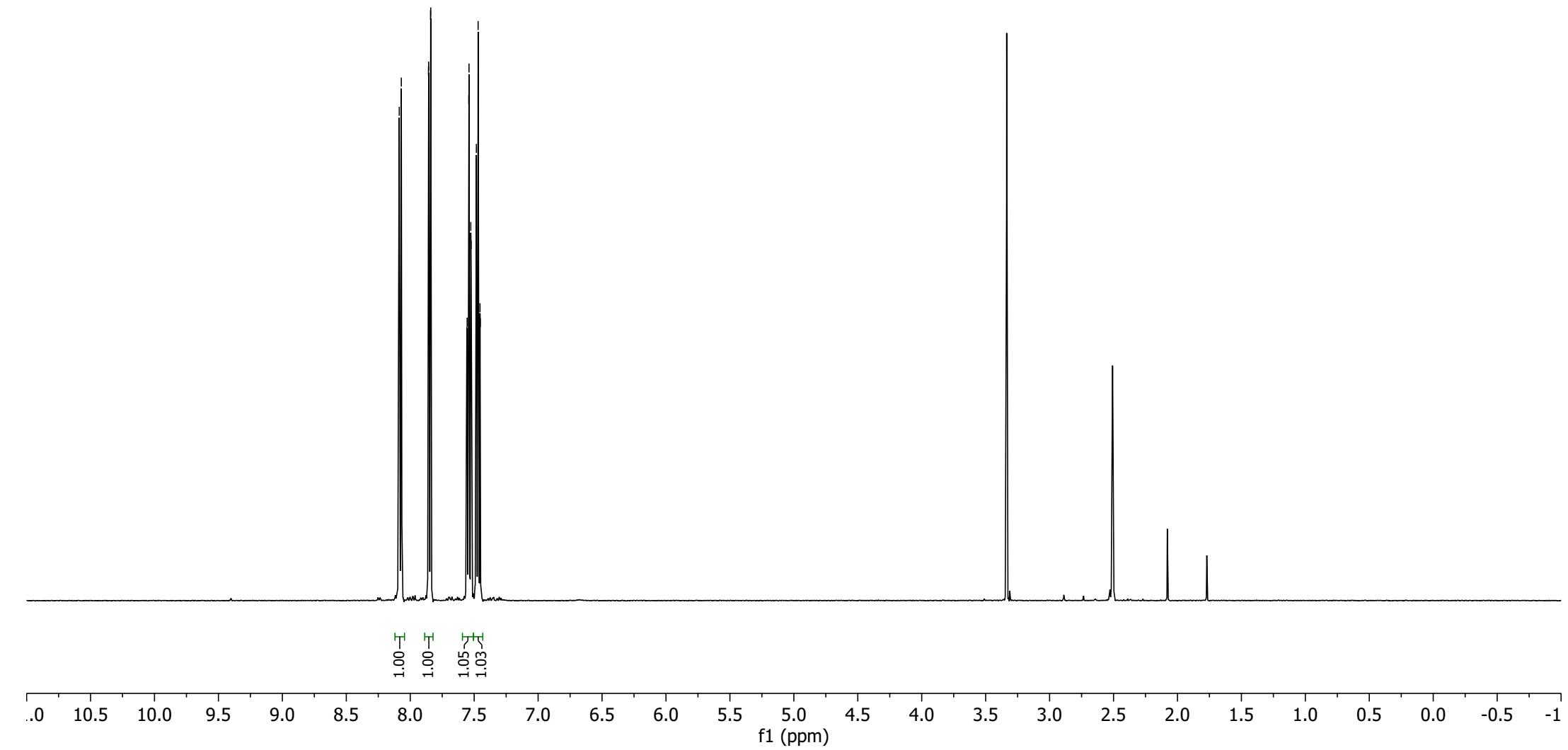
-62.95

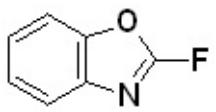




2p

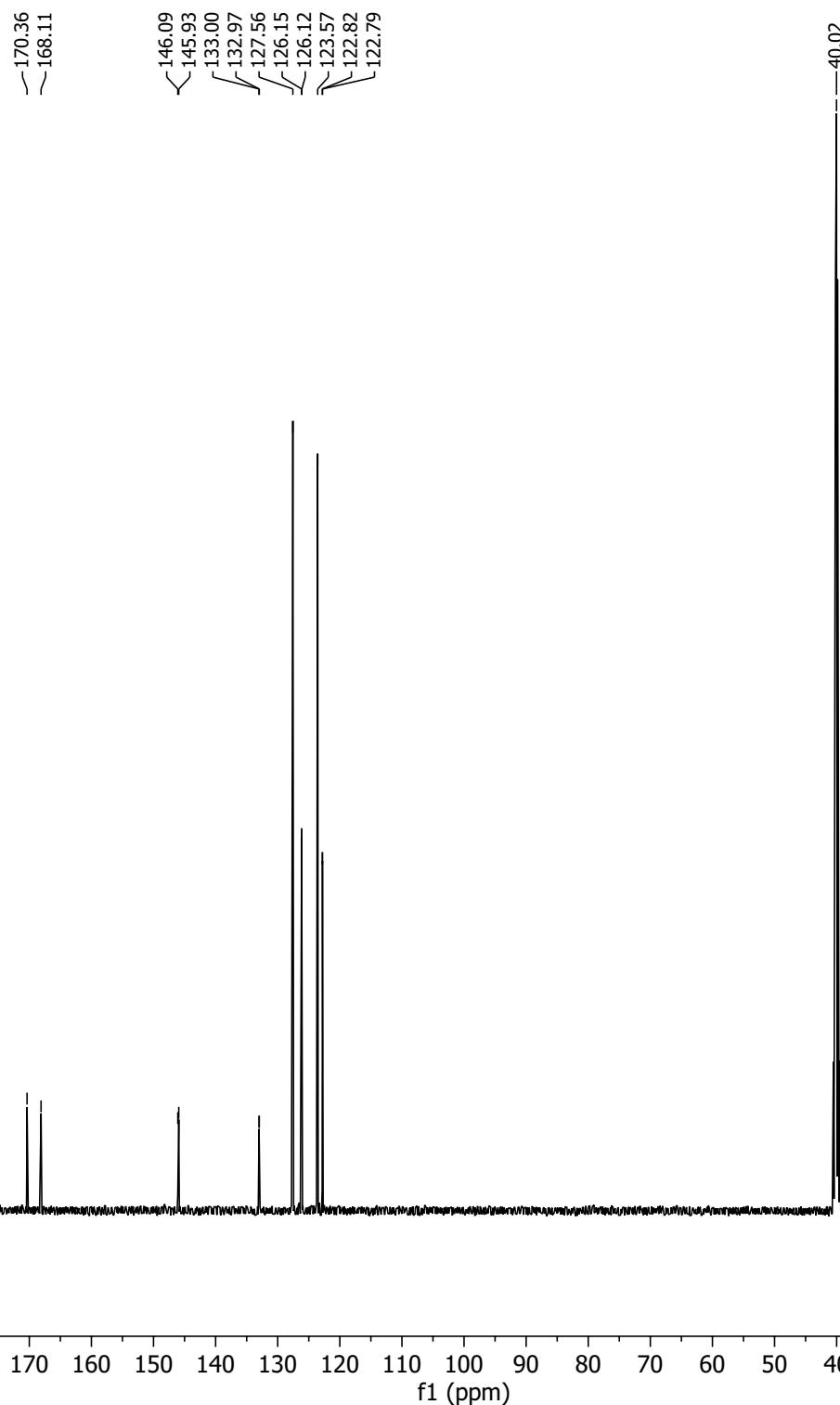
500 MHz, DMSO-d6

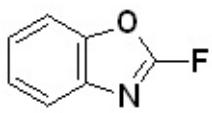




2p

126 MHz, DMSO-d6





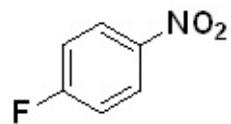
2p

471 MHz, DMSO-d6

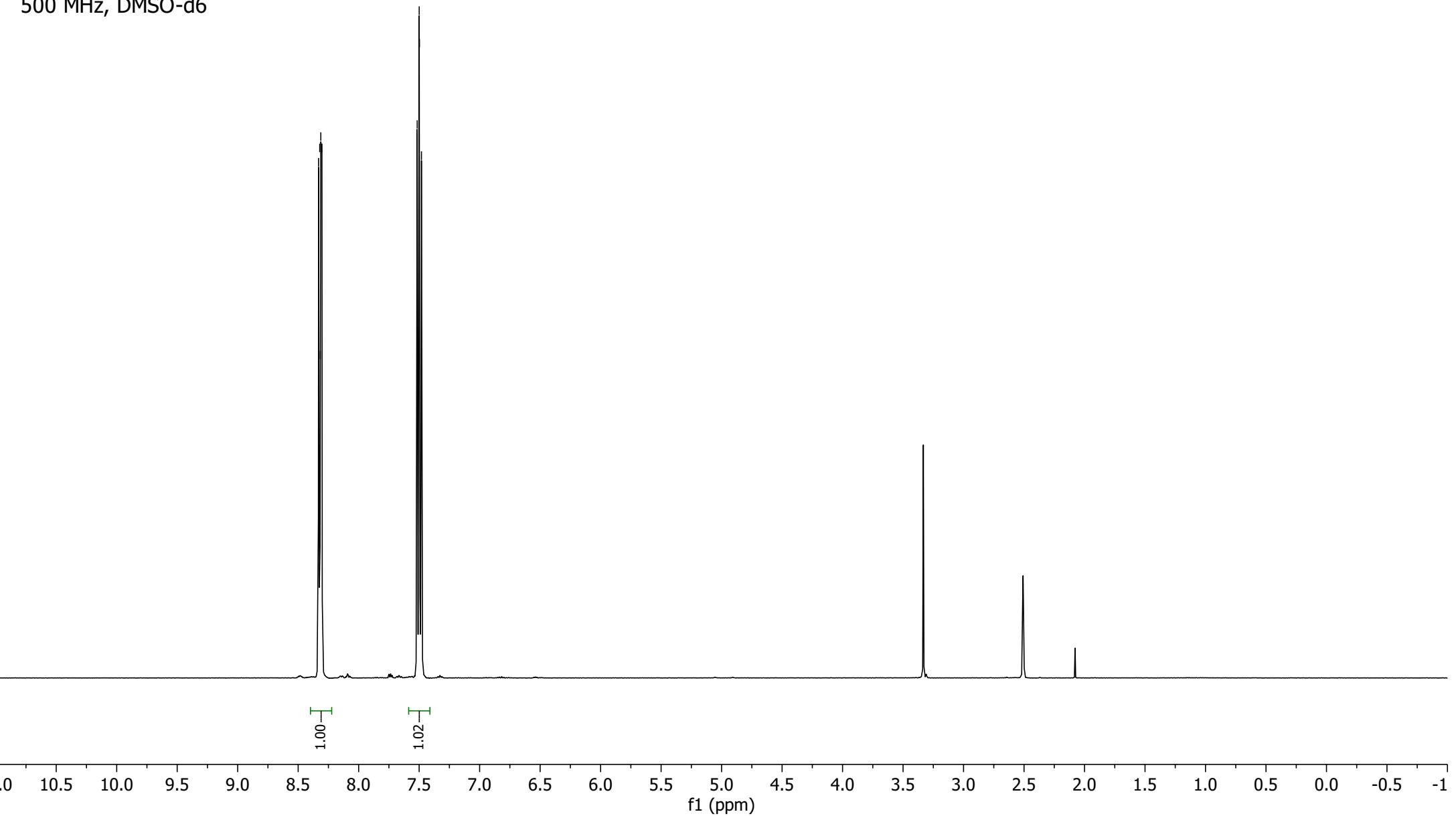
-73.64

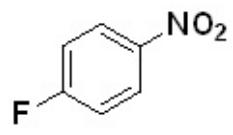
0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 -230 -240 -250 -260 -270 -280 -290 -300

f1 (ppm)

**4a**

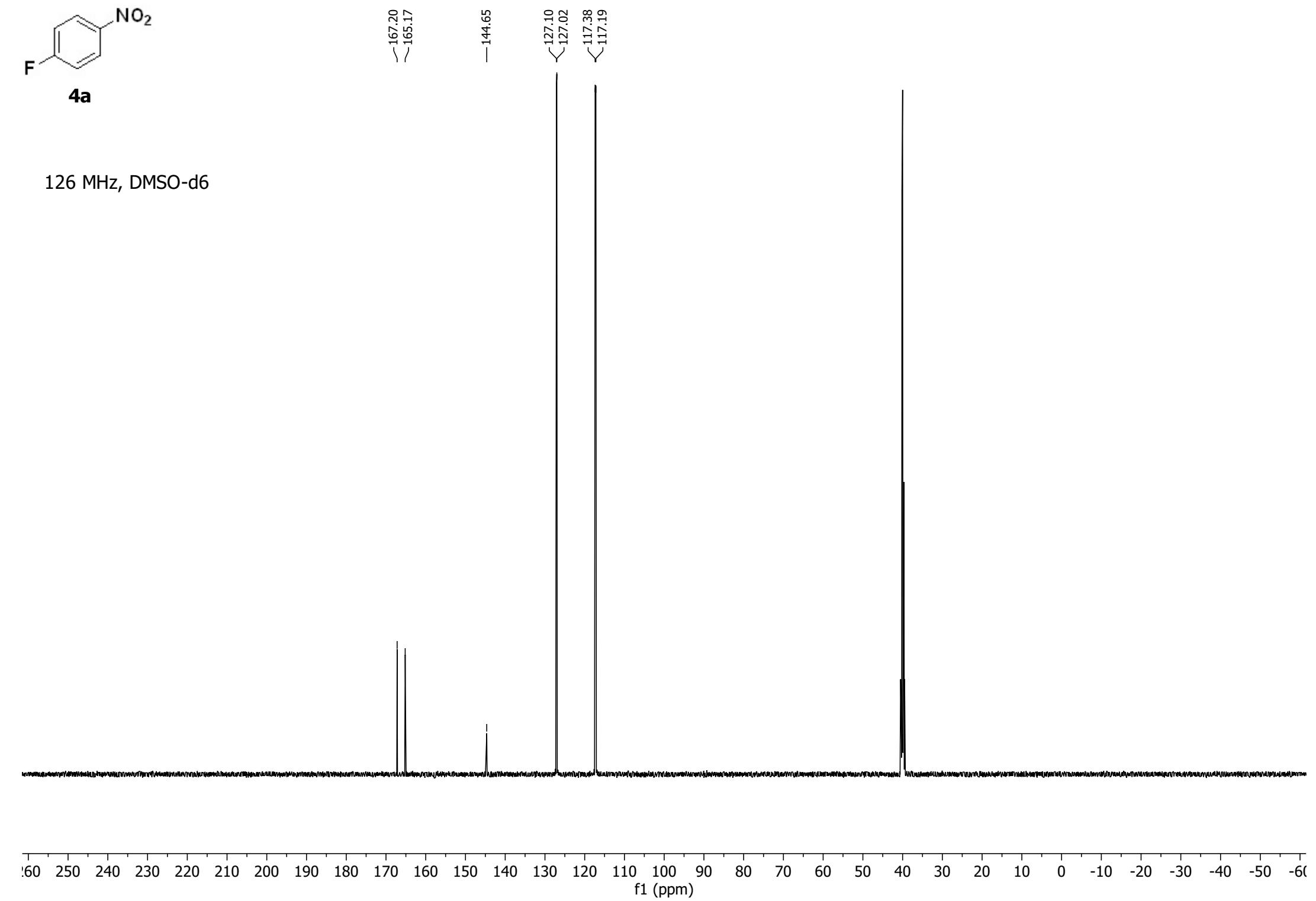
500 MHz, DMSO-d6

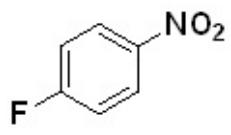




4a

126 MHz, DMSO-d6





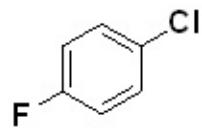
4a

471 MHz, DMSO-d6

-102.81

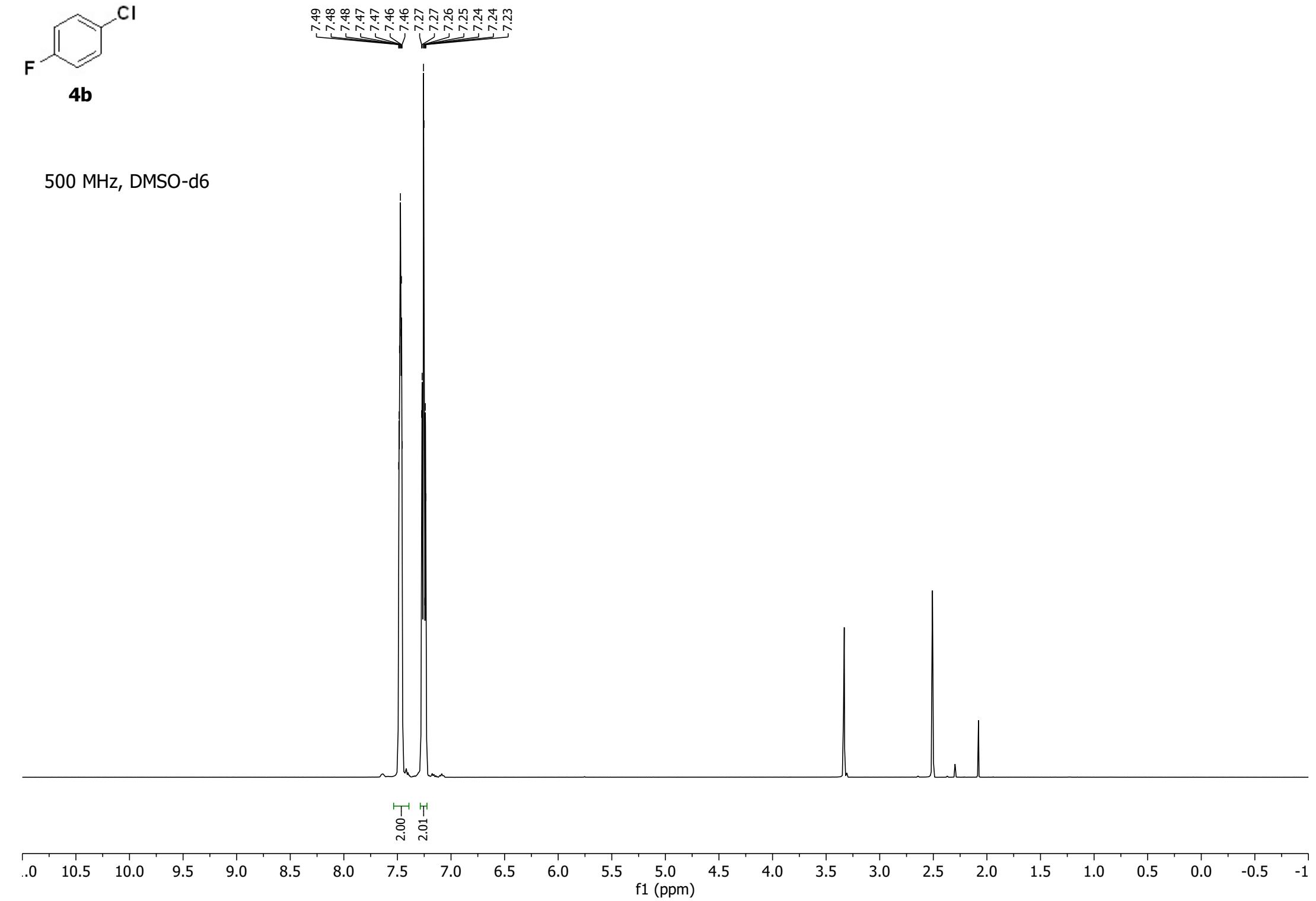
0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 -230 -240 -250 -260 -270 -280 -290 -300

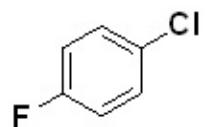
f1 (ppm)



4b

500 MHz, DMSO-d6

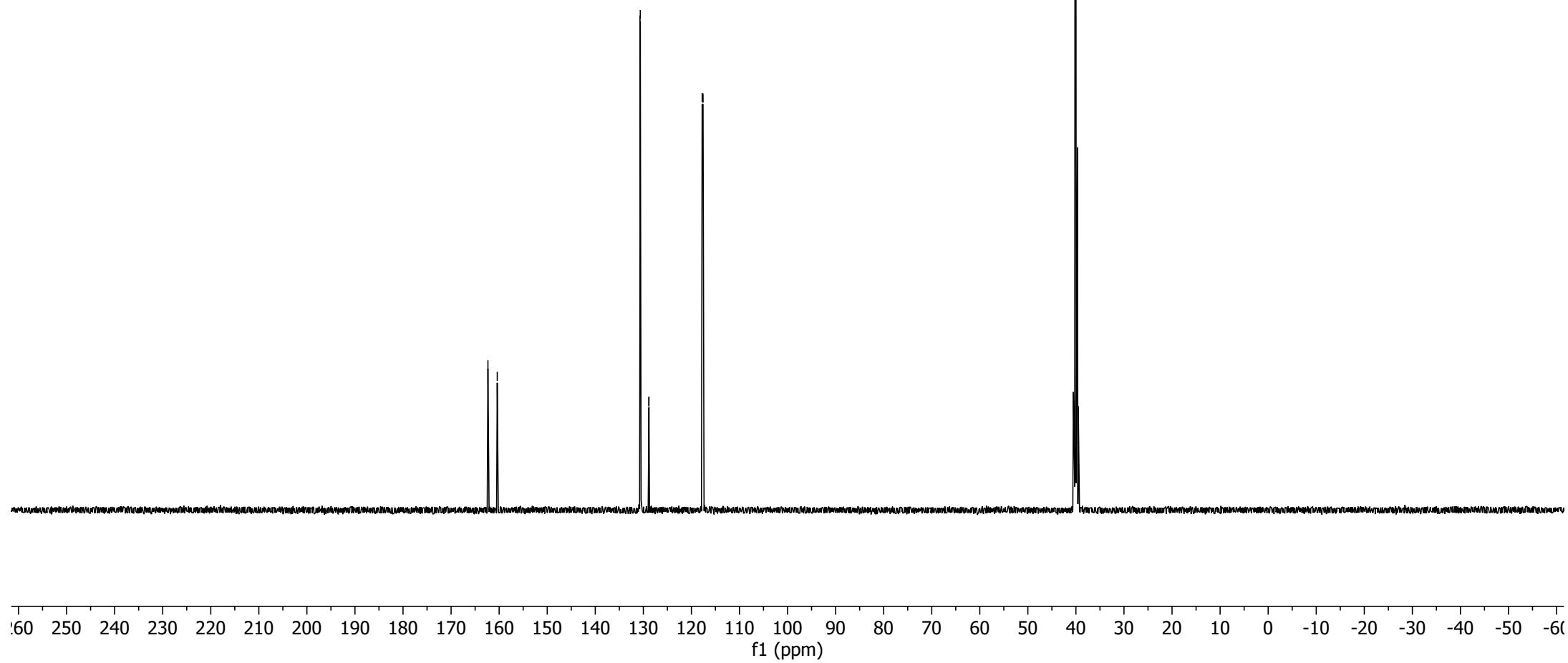


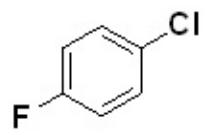


4b

126 MHz, DMSO-d6

162.33
~160.39
130.70
130.64
128.86
128.83
117.76
117.58





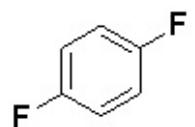
4b

471 MHz, DMSO-d6

-115.54
-115.55

0 -10 -20 -30 -40 -50 -60 -70 -80 -90 -100 -110 -120 -130 -140 -150 -160 -170 -180 -190 -200 -210 -220 -230 -240 -250 -260 -270 -280 -290 -300

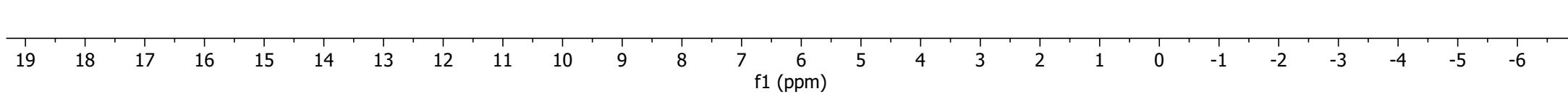
f1 (ppm)

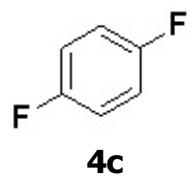


4c

500 MHz, DMSO-d6

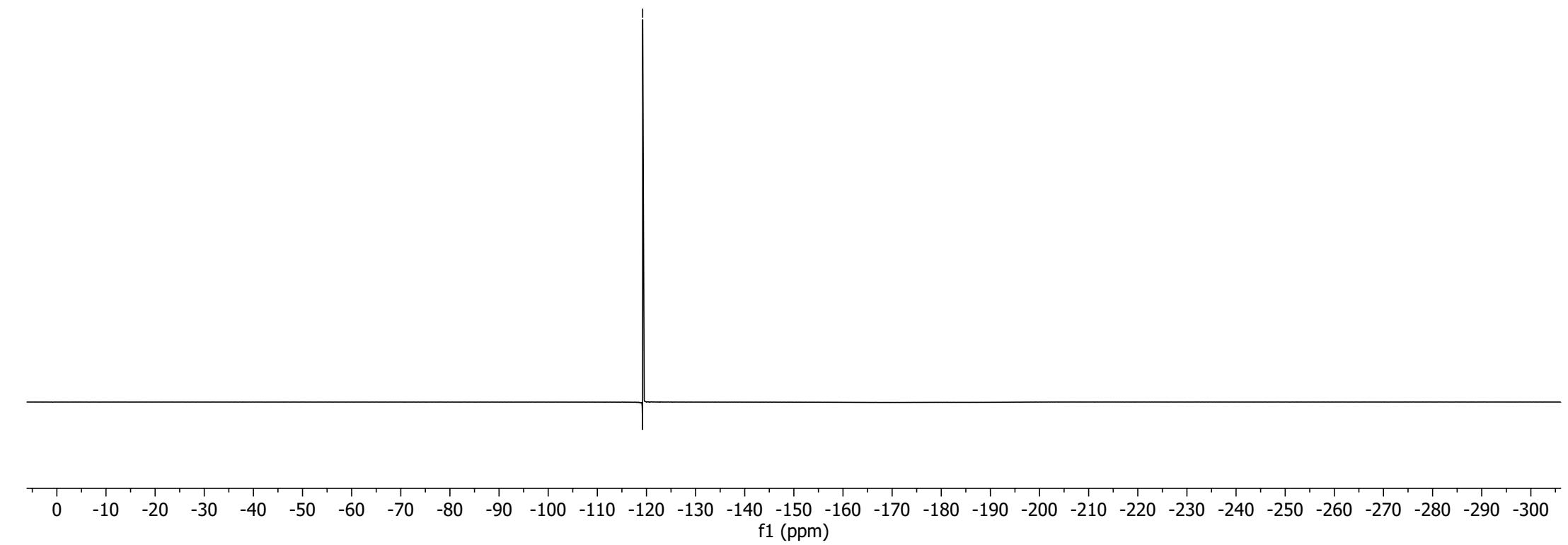
7.26
7.25
7.23

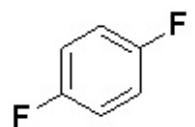




-119.23

126 MHz, DMSO-d6



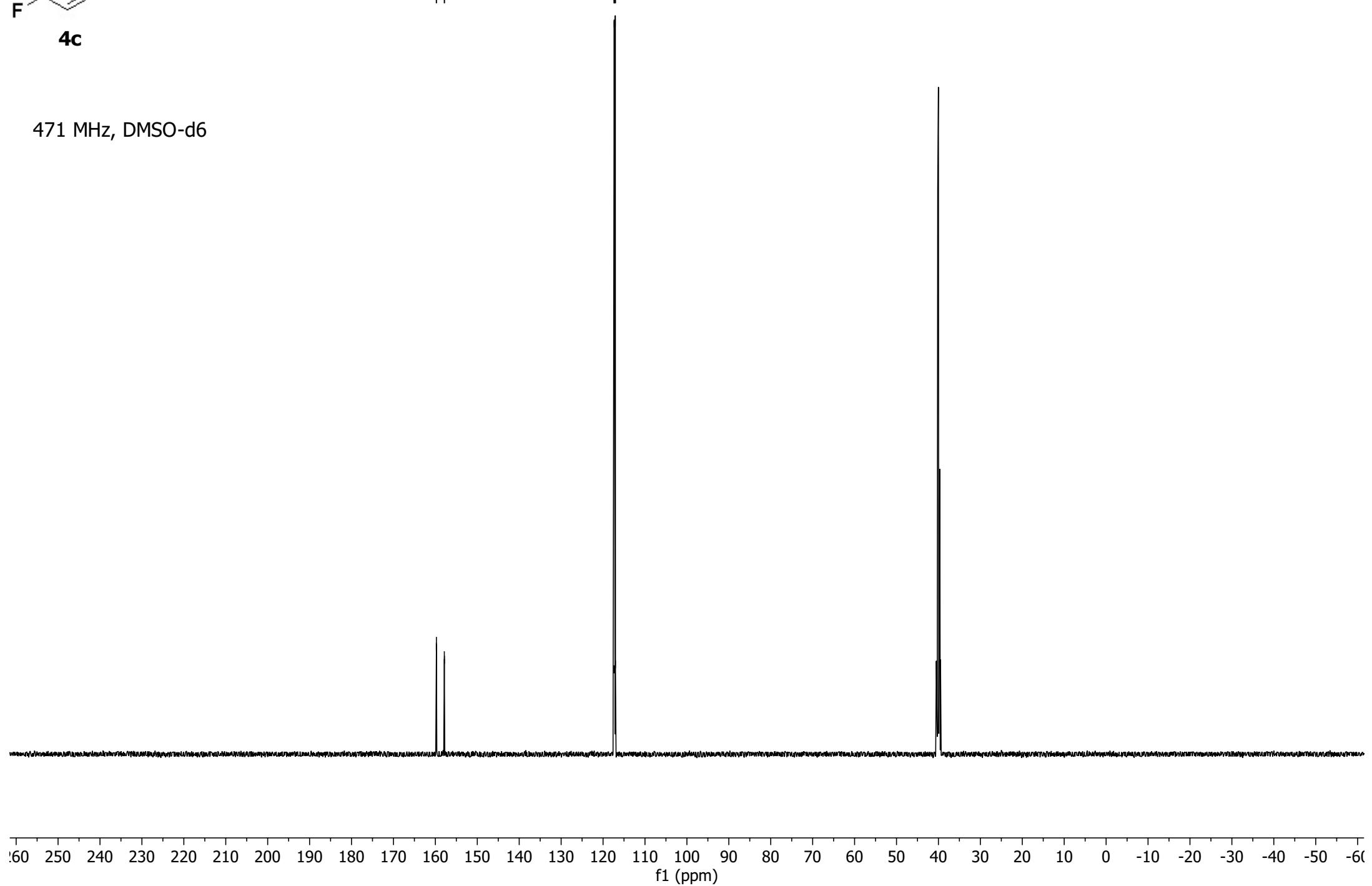


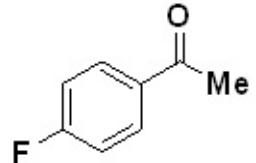
4c

471 MHz, DMSO-d6

159.78
159.75
157.86
157.83

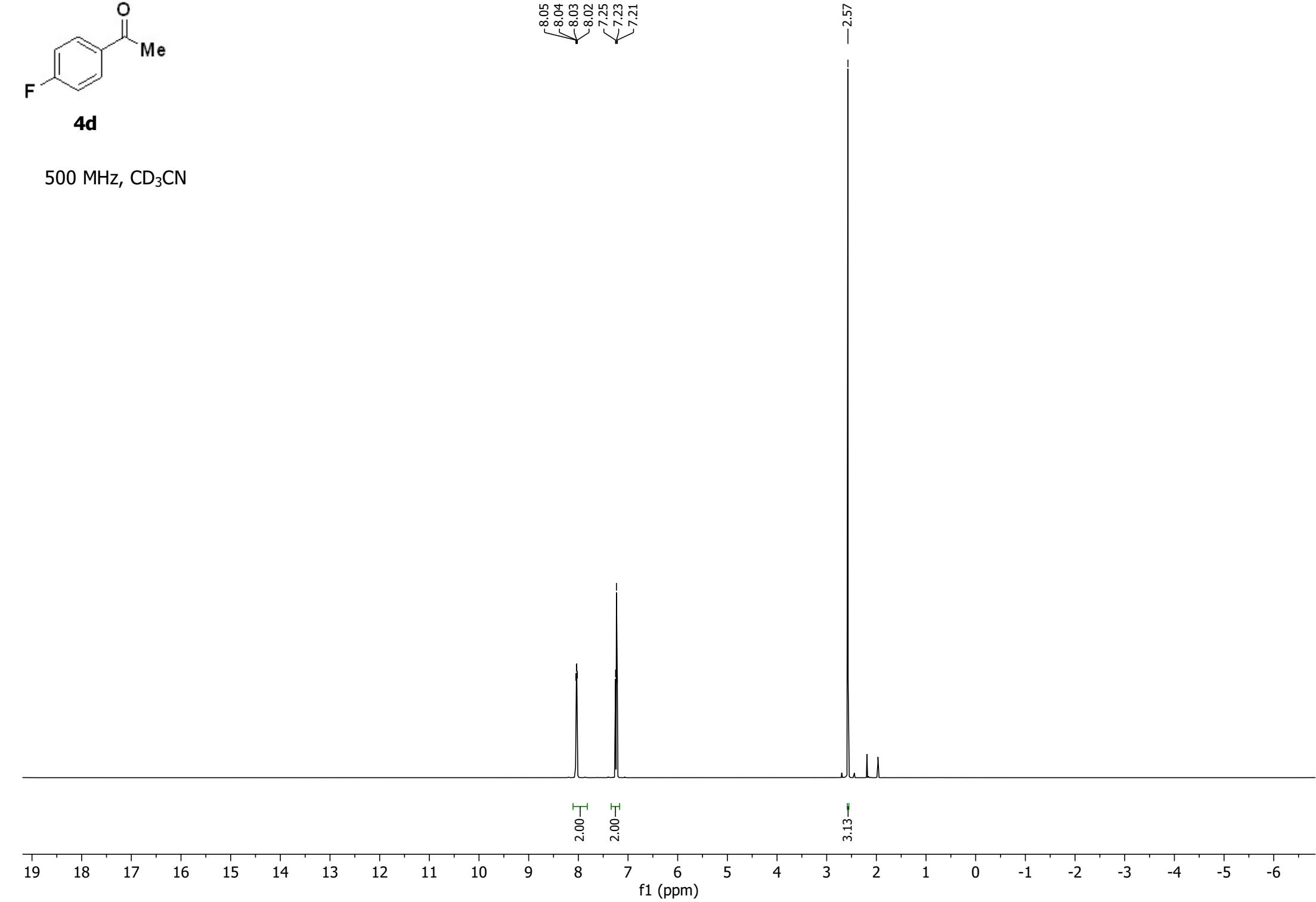
117.38
117.34
117.23
117.18
117.08
117.04

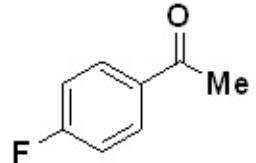




4d

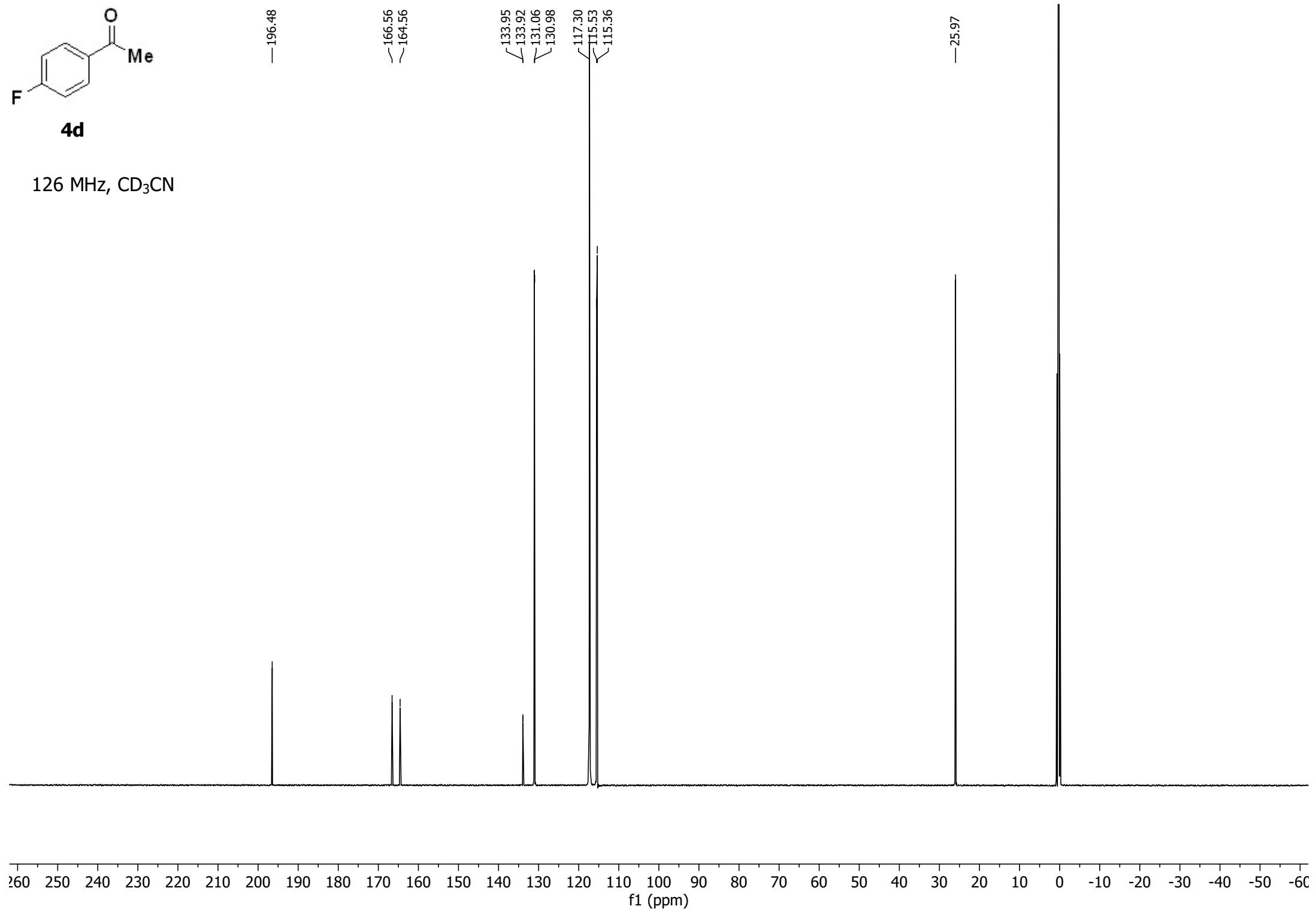
500 MHz, CD₃CN

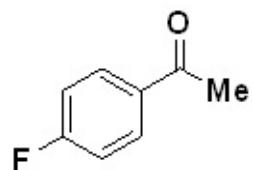




4d

126 MHz, CD_3CN





4d

471 MHz, CD_3CN

