

## Supporting Information

### **Half-Sandwich Iridium Complexes for One-Pot Synthesis of Amides: Preparation, Structure and Diverse Catalytic Activity**

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## General information

All reagents obtained from Admas-beta were used without further purification.  $^1\text{H}$  NMR and  $^{13}\text{C}$  spectra were recorded on Bruker AVANCE III 500 MHz (500 MHz for proton) spectrometer with tetramethylsilane as the internal reference using  $\text{CDCl}_3$  as solvent in all cases, and chemical shifts were reported in parts per million (ppm,  $\delta$ ). FT-IR spectra were recorded on a Thermo fisher Nicolet 6700. GC analyses were performed on Shimadzu GC-2014 with a flame ionization detector equipped with an Rtx-1 capillary column (internal diameter = 0.25 mm, length = 30 m) or a Stabilwax capillary column (internal diameter = 0.25 mm, length = 30 m). GC mass spectra were recorded on Shimadzu GCMS-QP2010 with a capillary column (0.25 mm $\times$  30 m). Column chromatography was performed using 200-300 mesh base-washed silica gel.

## Characterizations of typical products

**5a:**  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.78–7.74 (m, 2H), 7.50 (t,  $J$  = 7.5 Hz, 1H), 7.48 (t,  $J$  = 7.5 Hz, 2H), 5.83 (brs, 2H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5b:**  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.45 (d,  $J$  = 7.5 Hz, 1H), 7.35 (d,  $J$  = 7.5 Hz 1H), 7.25–7.20 (m, 2H), 6.45 (brs, 1H), 6,10 (brs, 1H), 2.45 (s, 3H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5c:**  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  8.13 (s, 1H), 7.61 (s, 1H), 7.43 (s, 1H), 7.38 (d,  $J$  = 8.0 Hz, 1H), 7.31 (d,  $J$  = 7.5 Hz, 1H), 7.20 (d,  $J$  = 7.5 Hz, 1H), 2.36 (s, 3H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5d:**  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  8.10 (s, 2H), 7.52 (d,  $J$  = 7.5 Hz, 2H), 6.93 (d,  $J$  = 7.5 Hz, 2H), 3.84 (s, 3H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5e:**  $^1\text{H}$  NMR (500 MHz,  $\text{CDCl}_3$ ):  $\delta$  7.95 (s, 1H), 7.62 (s, 1H), 7.48–7.45 (m, 4H). ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5f:** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 8.15 (s, 1H), 8.02 (d, J = 6.5 Hz, 1H), 7.85 (d, J = 7.0 Hz, 1H), 7.73–7.69 (m, 1H), 7.55 (s, 1H), 7.40 (t, J = 7.5 Hz, 1H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5g:** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 8.05 (s, 1H), 7.82 (d, J = 8.0 Hz, 2H), 7.62 (d, J = 8.0 Hz, 2H), 7.47 (s, 1H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5h:** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 8.00 (s, 1H), 7.89 (d, J = 7.5 Hz, 2H), 7.60 (d, J = 7.5 Hz, 2H), 7.41 (s, 1H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5i:** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 7.81 (d, J = 7.5 Hz, 2H), 7.53 (s, 1H), 7.03 (s, 1H), 6.99 (d, J = 7.5 Hz, 2H) 5.46 (s, 1H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5j:** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 7.71 (d, J = 8.0 Hz, 2H), 7.61 (s, 1H), 6.98 (s, 1H), 6.64 (d, J = 7.5 Hz, 2H), 2.90 (s, 6H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5k:** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 8.25 (d, J = 7.5 Hz, 2H), 8.23 (s, 1H), 7.98 (s, 1H), 7.79 (d, J = 7.0 Hz, 2H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5l:** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 8.65–8.61 (m, 1H), 8.12 (brs, 1H), 8.02–7.97 (m, 2H), 7.63 (brs, 1H), 7.62–7.58 (m, 1H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5m:** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 7.82 (s, 1H), 7.78 (d, J = 5.0 Hz, 1H), 7.36 (s, 1H), 7.12 (d, J = 7.0 Hz, 1H), 6.56 (dd, J = 7.0, 3.0 Hz, 1H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5n:** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 7.95 (s, 1H), 7.79–7.73 (m, 2H), 7.39 (s, 1H), 7.12 (t, J = 7.0 Hz, 1H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5o:** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 6.16 (s, 1H), 5.90 (s, 1H), 2.09 (s, 3H) ppm. Data are in

accordance with that previously published.<sup>1-5</sup>

**5p:** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 5.67 (brs, 1H), 5.48 (brs, 1H), 2.17 (t, J = 10.5 Hz, 1H), 1.87 (d, J = 12.0 Hz, 2H), 1.75 (d, J = 10.0 Hz, 2H), 1.65 (d, J = 10.5 Hz, 1H), 1.43 (q, J = 12.0 Hz, 2H), 1.23 (m, 3H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5q:** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 7.95 (s, 1H), 7.76 (d, J = 7.5 Hz, 2H), 7.29 (s, 1H), 7.22 (d, J = 7.0 Hz, 2H), 2.33 (s, 3H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5r:** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 9.12 (d, J = 7.0 Hz, 1H), 8.73 (s, 1H), 7.95 (d, J = 7.0 Hz, 1H), 7.83 (dd, J = 7.0 Hz, J = 4.0 Hz, 1H), 7.82 (d, J = 7.5 Hz, 1H), 7.59–7.55 (m, 1H) 7.51–7.45 (m, 2H), 7.12 (s, 1H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5s:** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 10.05 (s, 1H), 8.17 (s, 1H), 8.03 (d, J = 7.0 Hz, 2H), 7.96 (d, J = 7.0 Hz, 2H), 7.62 (s, 1H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

**5u:** <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>): δ 8.11 (s, 1H), 7.95 (s, 2H), 7.51 (s, 1H) ppm. Data are in accordance with that previously published.<sup>1-5</sup>

## References

- (1) Ambreen, N.; Wirth, T. High-Temperature Synthesis of Amides from Alcohols or Aldehydes by Using Flow Chemistry. *Eur. J. Org. Chem.* **2014**, 7590–7593.
- (2) Zhang, S.; Xu, H.; Lou, C.; Senan, A. M.; Chen, Z.; Yin, G. Efficient Bimetallic Catalysis of Nitrile Hydration to Amides with a Simple Pd(OAc)<sub>2</sub>/Lewis Acid Catalyst at Ambient Temperature. *Eur. J. Org. Chem.* **2017**, 2017, 1870–1875.
- (3) Crestani, M. G.; García, J. J. Catalytic hydration of mono and dinitriles using nickel(0) and PTSA. *J. Mol. Catal. A. Chem.* **2009**, 299, 26–36.
- (4) Guo, B.; deVaries, J. G.; Otten, E. Hydration of nitriles using a metal–ligand cooperative ruthenium pincer catalyst. *Chem. Sci.* **2019**, 10, 10647–10652.
- (5) Lee, J.; Kim, M.; Chang, S.; Lee, H. -Y. Anhydrous Hydration of Nitriles to Amides using

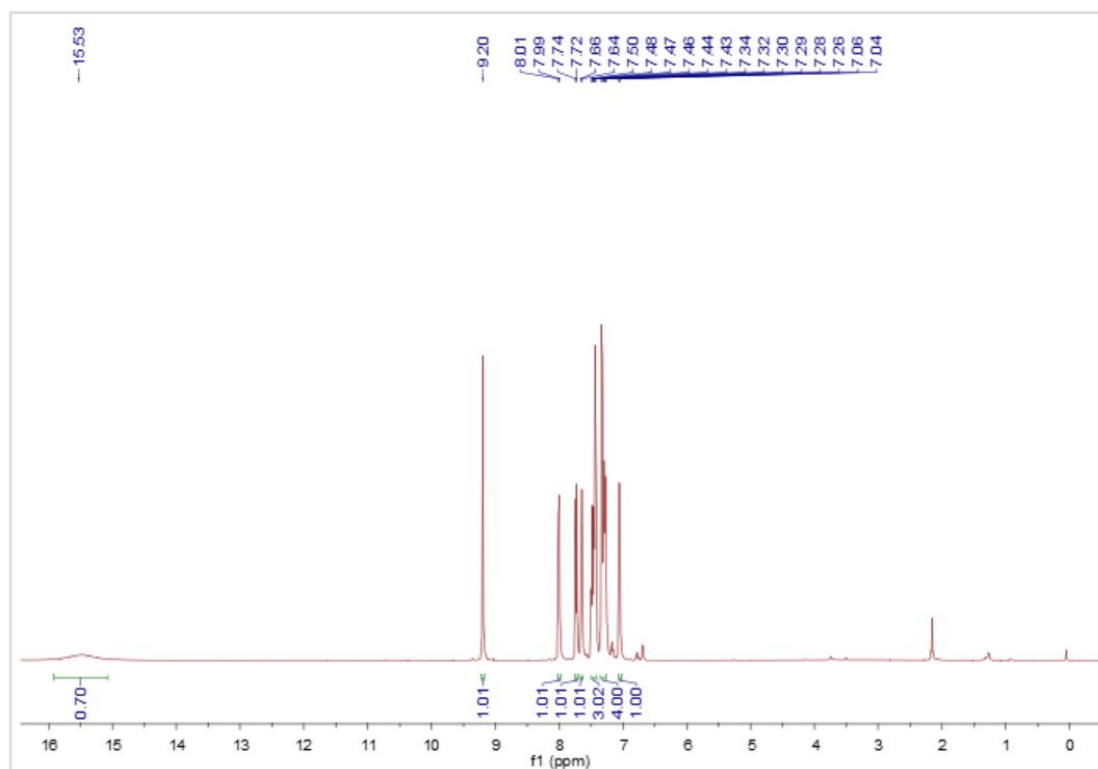


Figure S1. <sup>1</sup>H NMR spectra of **L1**

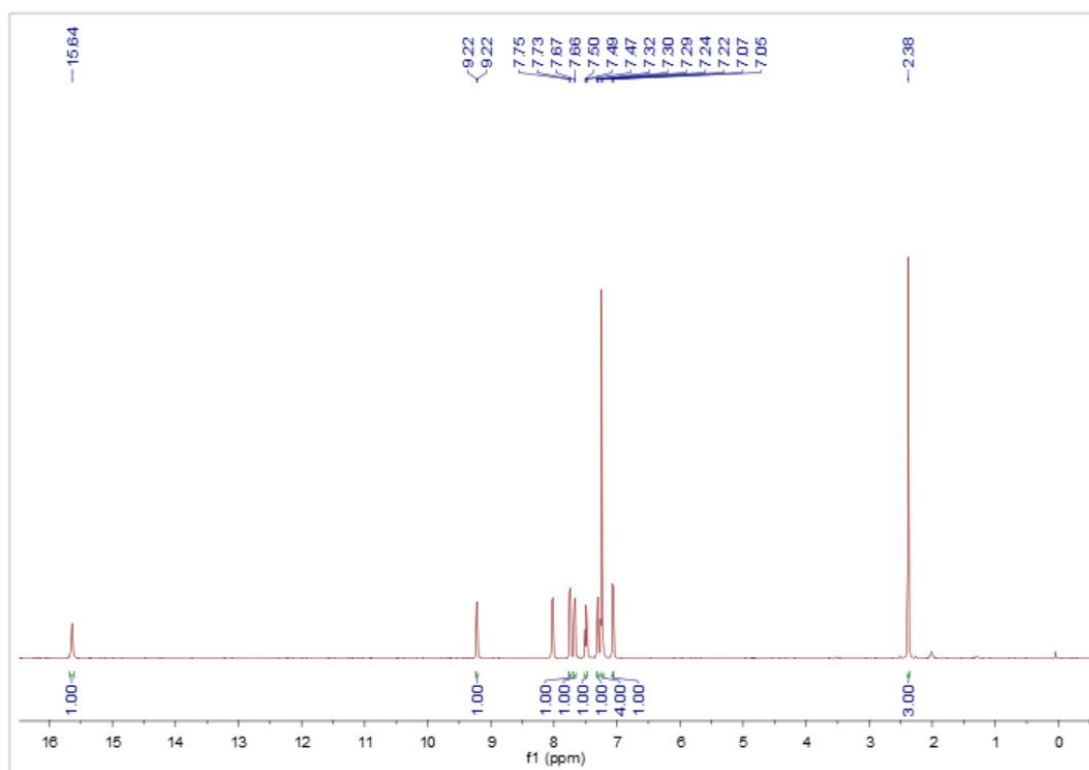


Figure S2. <sup>1</sup>H NMR spectra of **L2**

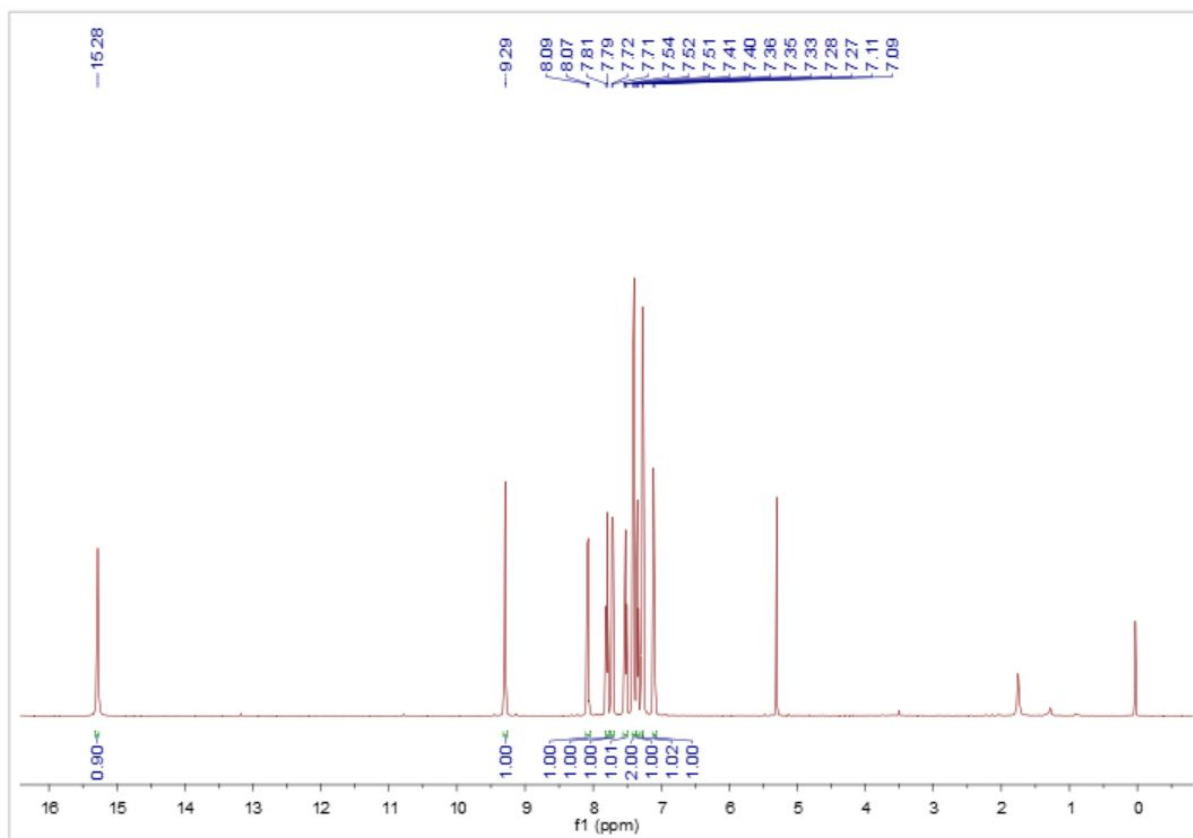


Figure S3. <sup>1</sup>H NMR spectra of L3

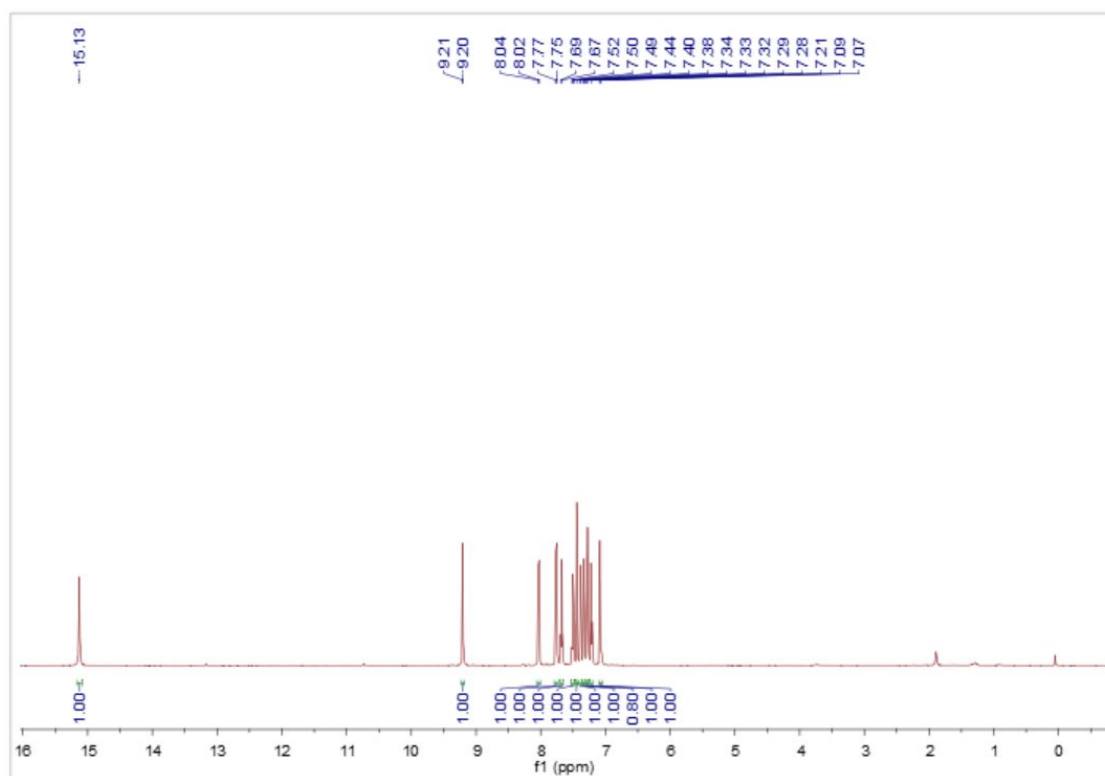


Figure S4. <sup>1</sup>H NMR spectra of L4

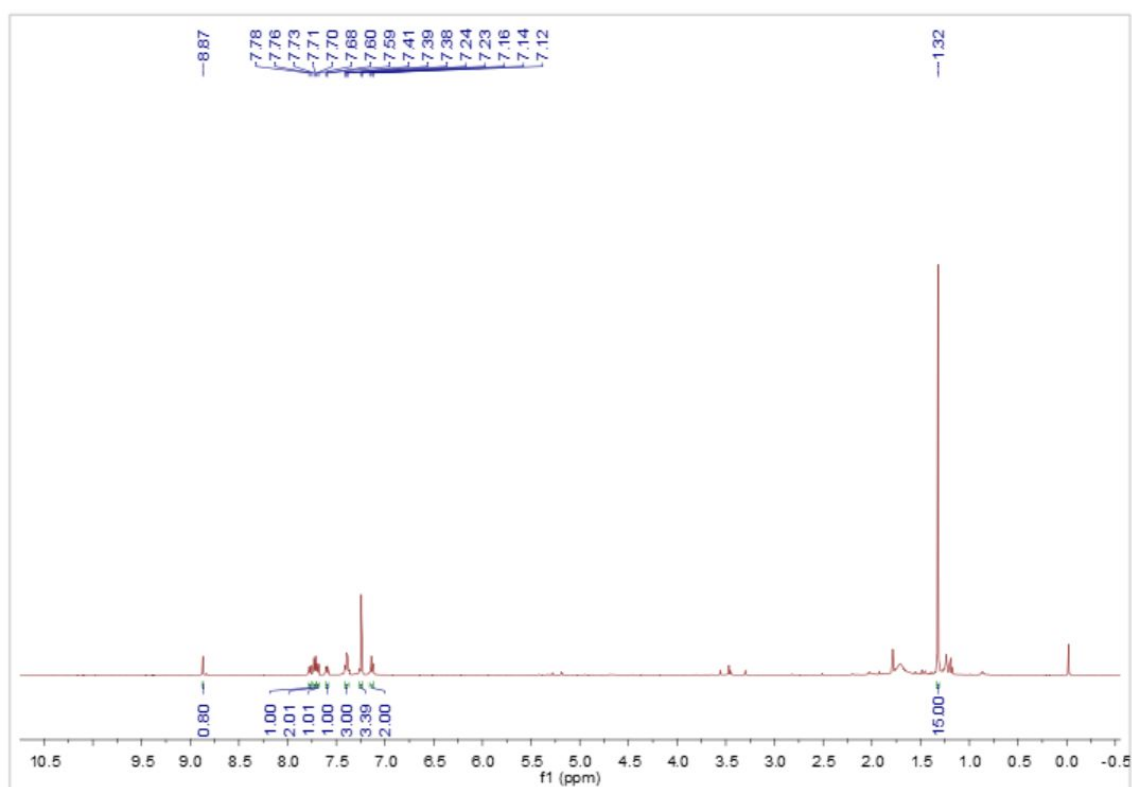


Figure S5. <sup>1</sup>H NMR spectra of **1**

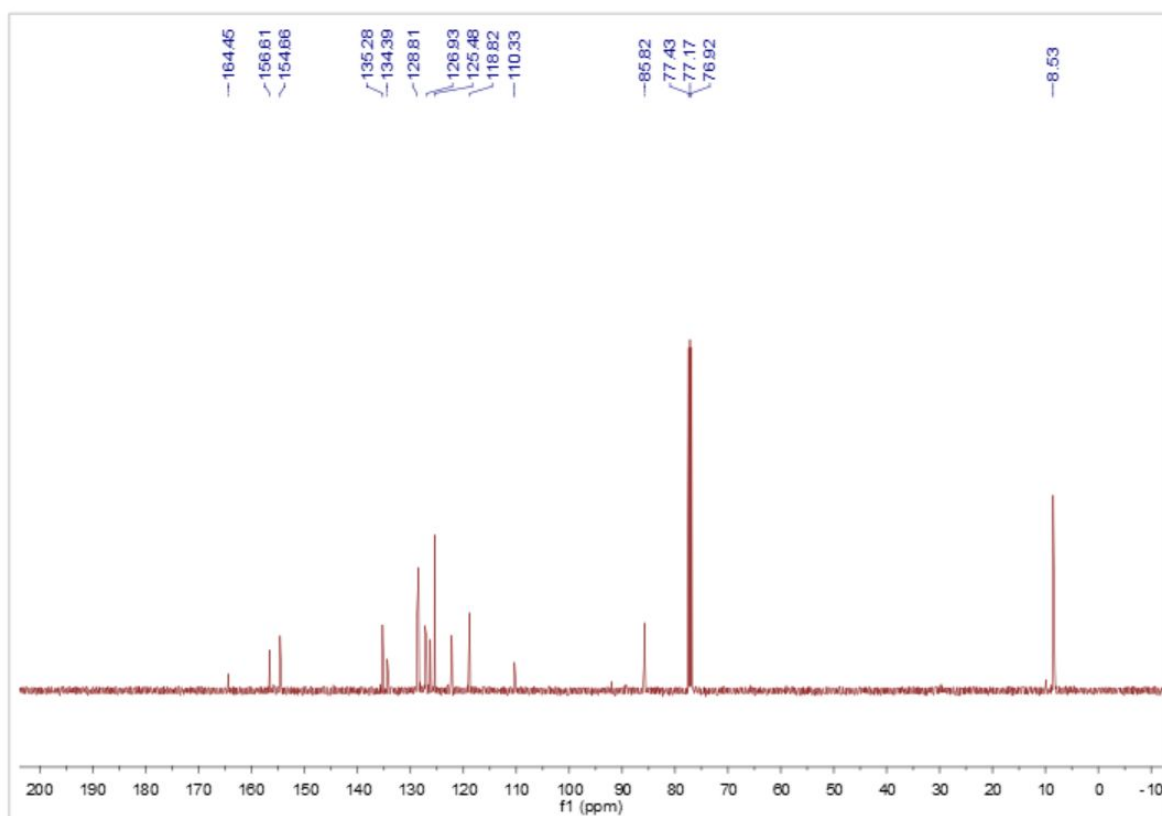


Figure S6. <sup>13</sup>C NMR spectra of **1**

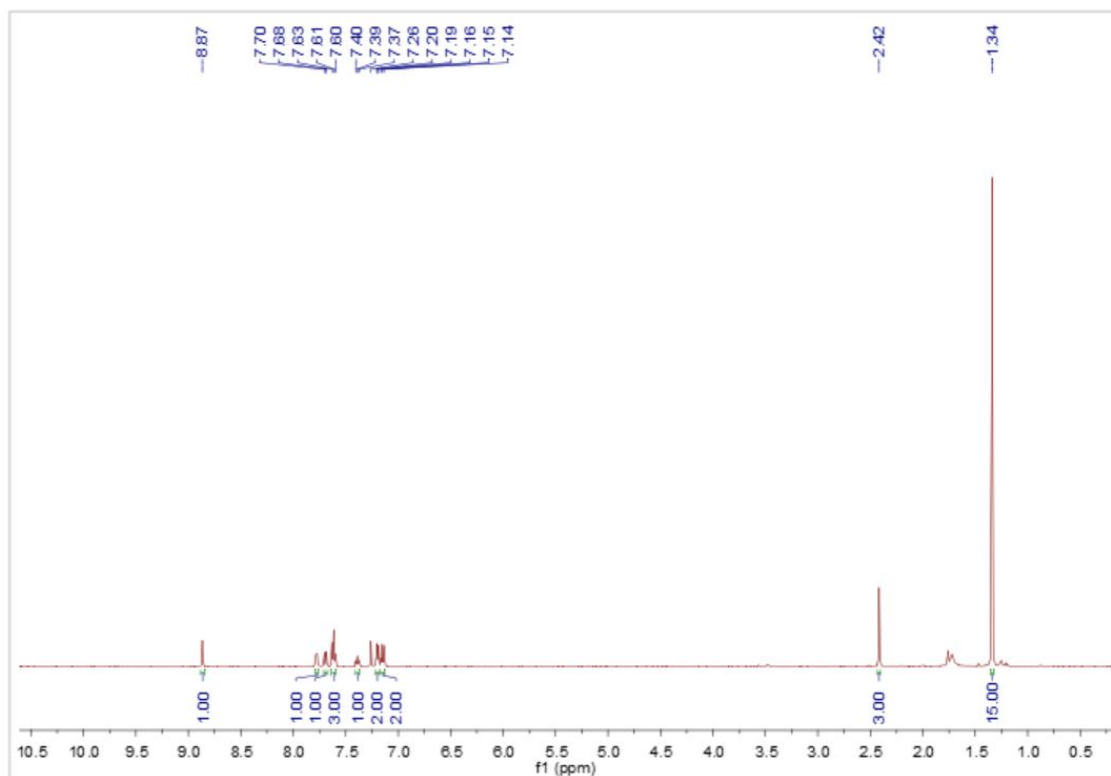


Figure S7. <sup>1</sup>H NMR spectra of **2**

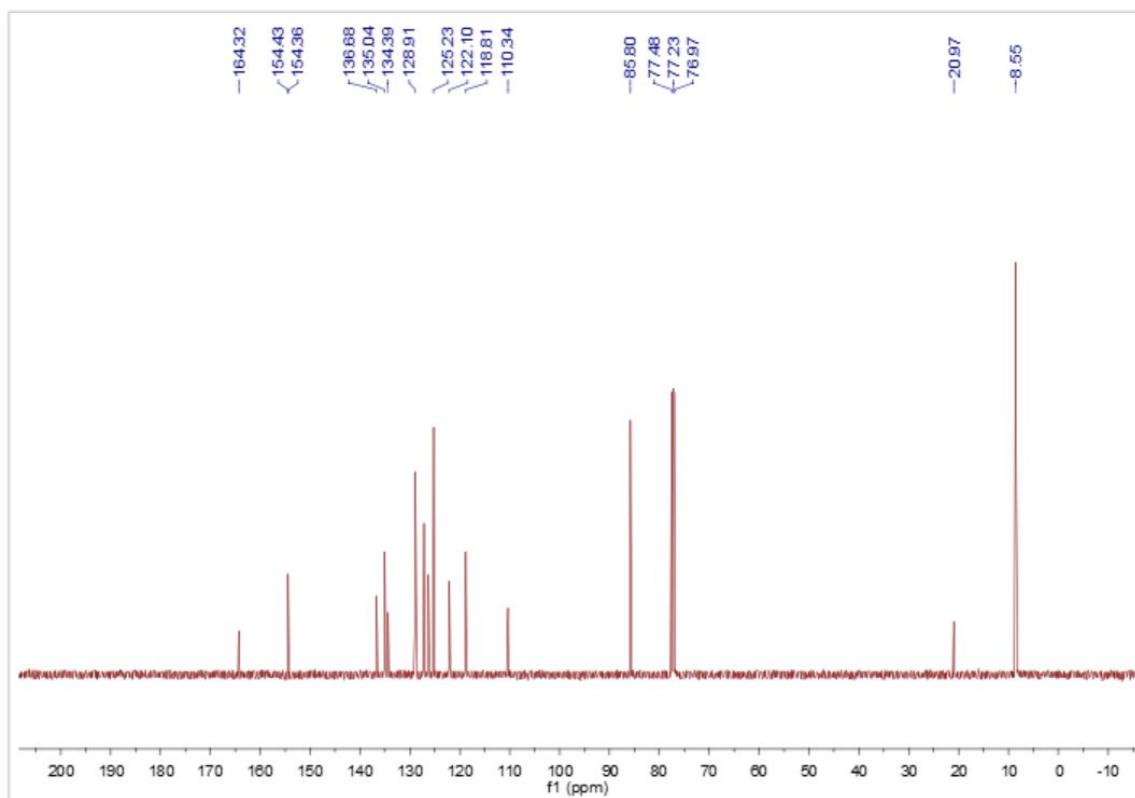


Figure S8. <sup>13</sup>C NMR spectra of **2**



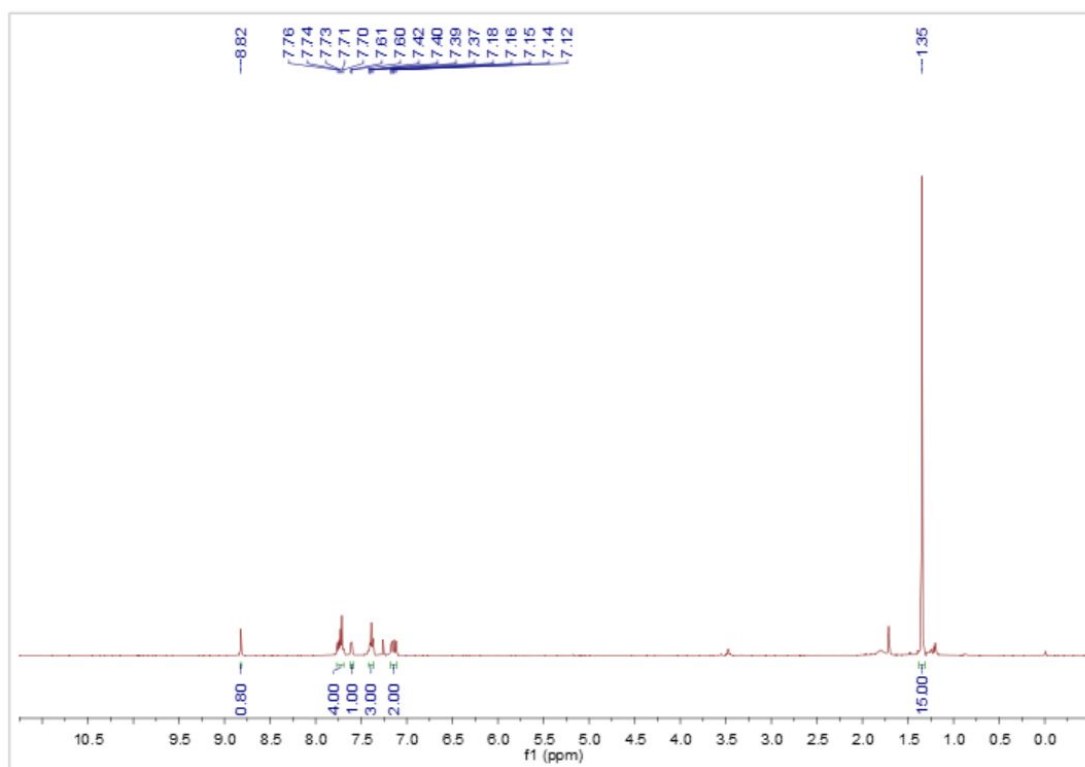


Figure S9.  $^1\text{H}$  NMR spectra of **3**

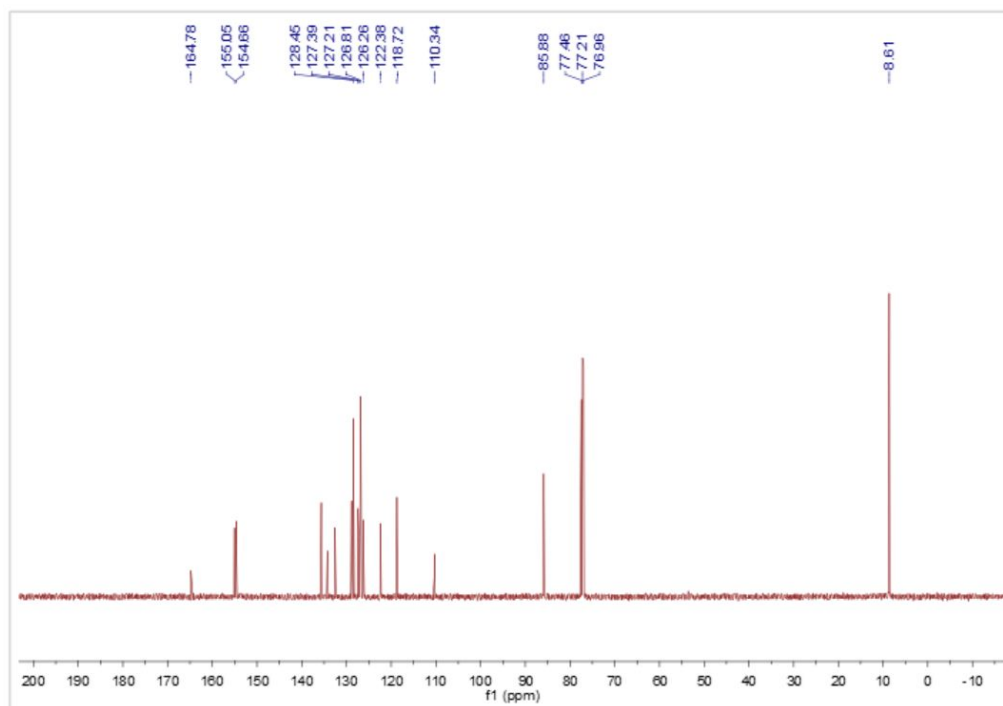


Figure S10.  $^{13}\text{C}$  NMR spectra of **3**

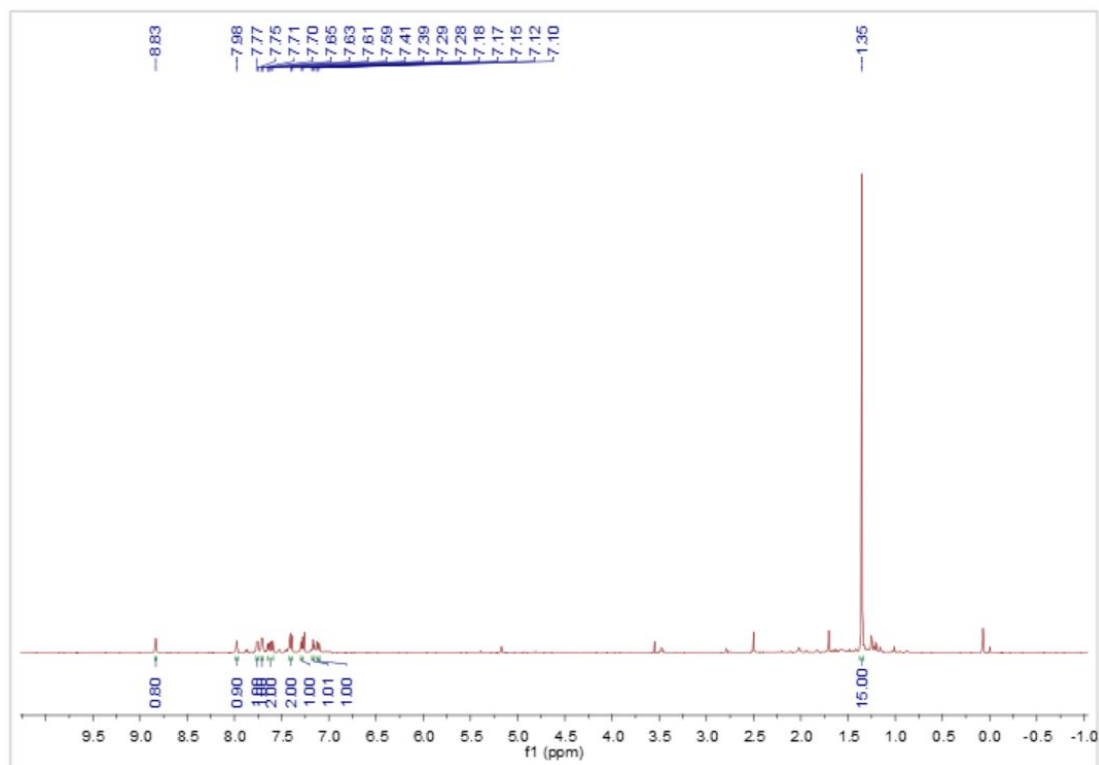


Figure S11. <sup>1</sup>H NMR spectra of 4

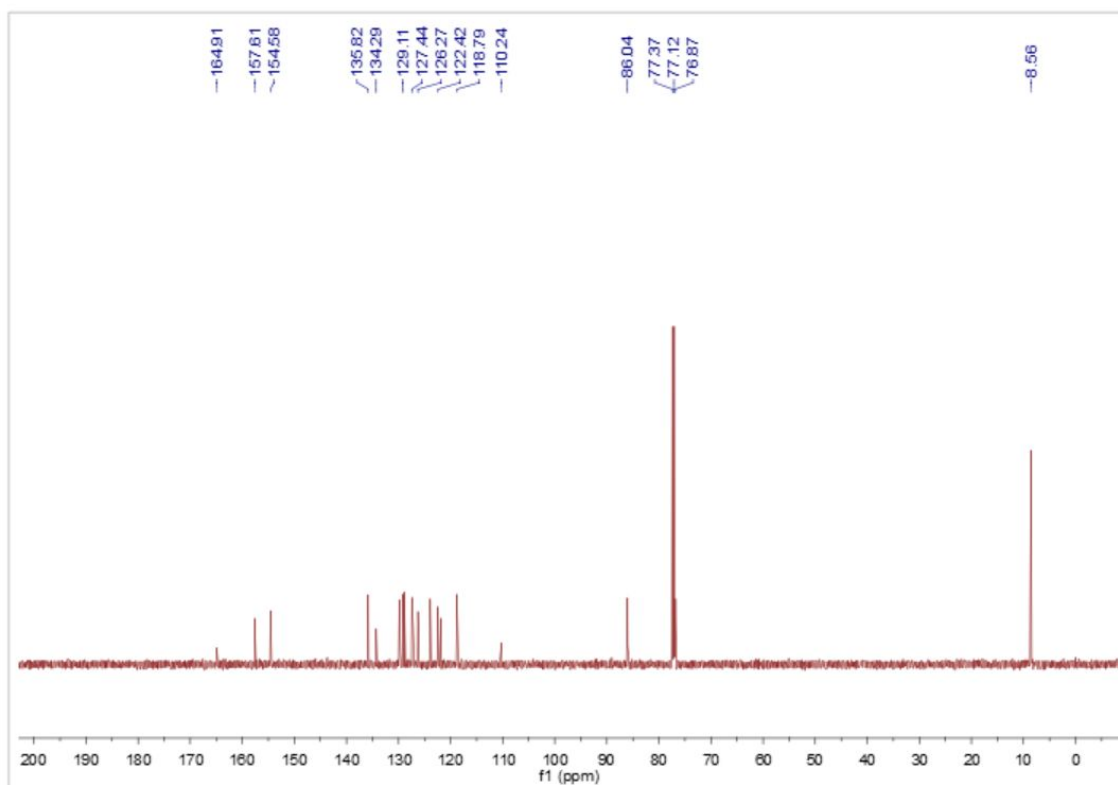


Figure S12. <sup>13</sup>C NMR spectra of 4

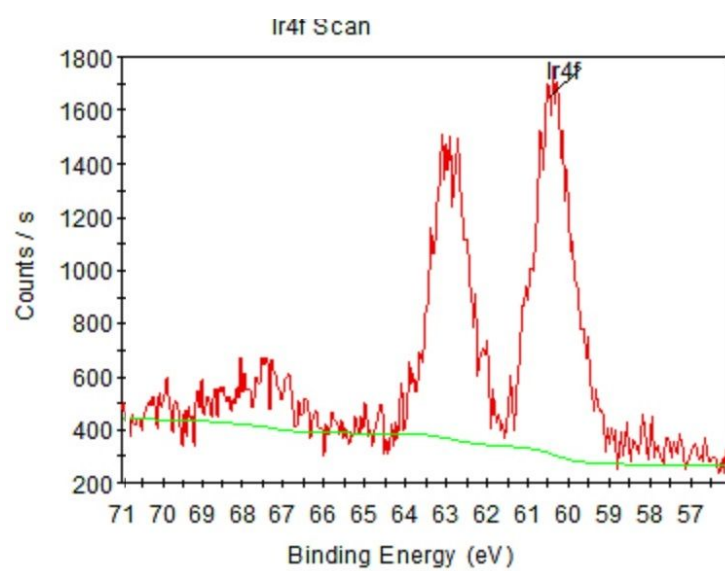


Figure S13. XPS of complex **1**