Supporting Information

Unified Total Synthesis of Five Bufadienolides

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1. Experimental procedures

General method: All reactions sensitive to air or moisture were carried out under argon atmosphere in dry solvents unless otherwise noted. CH₂Cl₂, DMF, THF, and toluene were purified by Glass Contour solvent dispensing system (Nikko Hansen & Co., Ltd.). All other reagents were used as supplied. All reactions under heating conditions were performed in an oil bath. Analytical thin-layer chromatography (TLC) was performed using E. Merck Silica gel 60 F₂₅₄ pre-coated plates (0.25 mm). Flash column chromatography was performed using 40-50 µm Silica Gel 60N (Kanto Chemical Co., Inc.), 40-63 µm Silica Gel 60 (Merck), or 32-53 µm Silica Gel BW-300 (Fuji Silysia Chemical Ltd.). Preparative thin-layer chromatography (PTLC) was performed using Merck silica gel 60 F254 pre-coated plates (0.5 mm). Preparative gel permeation chromatography (GPC) was performed on LaboACE LC-5060 (Japan Analytical Industry Co., Ltd.) equipped with JAIGEL-2HR column using UV (254 nm) and Refractive Index (RI) detectors in CHCl₃ with flow rate of 10.0 mL/min. High performance liquid chromatography (HPLC) was performed on a JASCO HPLC system (pump: JASCO PU-2086 Plus x2, detector: JASCO MD-4010, degasser: ERC Inc. ERC-3325, data analysis by JASCO ChromNAV 1.5.2.). Melting points were measured on Yanaco MP-J3 micro melting point apparatus and were uncorrected. Optical rotations were measured on JASCO P-2200 polarimeter at ambient temperature using the sodium D line. Infrared (IR) spectra were recorded on JASCO FT/IR-4100 spectrometer as a thin film on KBr or CaF₂. ^{1}H and ¹³C{¹H} NMR spectra were recorded on JEOL JNM-ECX-500, JNM-ECZ-500, or JNM-ECS-400 spectrometer. Chemical shifts were denoted in ppm on the δ scale relative to residual solvent peaks as internal standard: CHCl₃ (δ 7.26 for ¹H NMR), CDCl₃ (δ 77.0 for ¹³C{¹H} NMR), C₆D₅H (δ 7.16 for ¹H NMR), C₆D₆ (δ 128.0 for ¹³C{¹H} NMR), CD₂HOD (δ 3.31 for ¹H NMR), CD₃OD (δ 49.0 for ¹³C{¹H} NMR), D₃CSOCD₂H (δ 2.50 for ¹H NMR), and D₃CSOCD₃ (δ 39.52 for ¹³C{¹H} NMR). Signal patterns are indicated as s, singlet; d, doublet; t, triplet; m, multiplet; br, broaden peak. The carbon numbering of compounds corresponds to that of vulgarobufotoxin (4) and 3-(N-succinyl argininyl) bufotalin (5), unless otherwise noted. High resolution mass spectra were measured on JEOL JMS-T100LP (ESI-TOF) or BRUKER DALTONICS microTOF II (ESI-TOF). X-ray crystallographic analysis was conducted using Inorganic Fine Crystal Structural Diffractometer VariMax Dual (Rigaku Co., Ltd.) with Mo radiation ($\lambda = 0.71073$ Å).

Supporting Information



Figure S1. Numbering system of vulgarobufotoxin (4) and 3-(*N*-succinyl argininyl) bufotalin (5)



Diketones 10a [CAS: 1229-12-5]^{S1} and 10b [CAS: 846-46-8].^{S2} A suspension of Pd/C (10 wt% Pd on carbon, 499 mg) in pyridine (15 mL) was stirred at room temperature for 30 min. Compound 9 (5.53 g, 19.3 mmol) was added to the suspension, and the resultant suspension was stirred under H₂ atmosphere (1 atm) at room temperature for 12 h. The reaction mixture was then filtered through a pad of Celite with CH₂Cl₂ (15 mL) and concentrated. After 1 M aqueous HCl (15 mL, 15 mmol) was added to the residue, the resultant mixture was extracted with CH₂Cl₂ (30 mL x6). The combined organic layers were dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (100 g, hexane/CH₂Cl₂ = 3/1 to 0/1) to afford a 9.3:1 diastereometric mixture of diketones **10a** and **10b** (5.34 g, 18.5 mmol) in 96% yield. The C5-stereochemistry of **10a** was determined by the Xray crystallographic analysis of epoxide 6: white solid. m.p. 121-123 °C. $[\alpha]_D^{27}$ +98 (c 0.96, CHCl₃). IR (film) 2934, 2861, 1738, 1712, 1452, 1377, 1266, 1248, 1051 cm⁻¹. ¹H NMR (500 MHz, CD₃OD) peaks of the major product 10a: δ 0.89 (3H, s, H18), 1.07 (3H, s, H19), 1.30-1.80 (13H, m), 1.95-2.10 (6H, m), 2.41-2.48 (2H, m), 2.84 (1H, dd, J = 14.6, 14.6 Hz). ¹³C{¹H} NMR (125 MHz, CD₃OD) peaks of the major product 10a: δ 14.2, 21.5, 22.7, 22.9, 25.7, 27.5, 32.8, 36.2, 36.4, 36.7, 37.9, 38.1, 41.8, 43.1, 45.8, 49.0 (deduced from HMBC), 52.4, 215.6, 223.5. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₁₉H₂₈O₂Na 311.1987; Found 311.1976.

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Ketones S1a [CAS: 571-31-3]^{S3} and S1b. KBH(*s*-Bu)₃ (1.0 M THF solution, 18.5 mL, 18.5 mmol) was added to a solution of a 9.3:1 diastereomeric mixture of diketones 10a and 10b (5.34 g, 18.5 mmol) in THF (185 mL) at -78 °C. After the reaction mixture was stirred at -78 °C for 5.5 h, 2.6 M aqueous NaOH (7.0 mL, 18 mmol) and aqueous H₂O₂ (30 wt%, 9.8 M, 7.0 mL, 69 mmol) were added to the mixture. The resultant mixture was concentrated and extracted with EtOAc (50 mL x5). The combined organic layers were dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (150 g, hexane/EtOAc = 1/0 to 20/1 to 10/1) to afford a 9.3:1 diastereometric mixture of ketones S1a and S1b (5.06 g, 17.4 mmol) in 94% yield. The C3-stereochemistry of S1a was determined by the X-ray crystallographic analysis of epoxide 6: white solid. m.p. 163-166 °C. $[\alpha]_D^{27}$ +62 (c 1.10, CHCl₃). IR (film) 3218, 2929, 2859, 1738, 1451, 1405, 1324, 1197, 1031 cm⁻¹. ¹H NMR (500 MHz, CDCl₃) peaks of the major product S1a: δ 0.85 (3H, s, H18), 0.98 (3H, s, H19), 1.07-1.17 (1H, m), 1.19-1.42 (7H, m), 1.44-1.66 (7H, m), 1.73-1.82 (2H, m), 1.88-1.99 (3H, m), 2.06 (1H, ddd, J = 19.2, 9.4, 9.4 Hz), 2.43 (1H, dd, J = 19.2, 8.9 Hz), 4.11 (1H, dd, J = 2.8, 2.8 Hz, H3). ¹³C{¹H} NMR (125 MHz, CDCl₃) peaks of the major product S1a: δ 13.8, 20.3, 21.8, 23.8, 25.2, 26.3, 27.8, 29.9, 31.7, 33.4, 35.2, 35.3, 35.9, 36.5, 40.0, 47.9, 51.6, 66.9, 221.5. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₁₉H₃₀O₂Na 313.2143; Found 313.2140.



Ketone 11 [CAS: 65598-74-5].^{S4} *t*-Butyldimethylsilyl chloride (TBSCl, 15.7 g, 104 mmol) was added to a solution of a 9.3:1 diastereomeric mixture of ketones **S1a** and **S1b** (15.1 g, 52.0

mmol), and imidazole (14.1 g, 207 mmol) in DMF (73 mL) at room temperature. After the reaction mixture was stirred at 70 °C for 15 h, saturated aqueous NaHCO₃ (100 mL) and H₂O (50 mL) were added to the mixture at 0 °C. The resultant mixture was extracted with Et₂O (100 mL x4). The combined organic layers were washed with brine (30 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (250 g, hexane/Et₂O = 100/1 then hexane/EtOAc = 3/1) to afford ketone **11** (18.2 g, 45.0 mmol) in 87% yield: white solid. m.p. 121-123 °C. $[\alpha]_D^{27}$ +70 (c 0.47, CHCl₃). IR (film) 2927, 2857, 1742, 1449, 1406, 1373, 1253, 1163, 1057, 1005 cm⁻¹. ¹H NMR (400 MHz, C₆D₆) δ 0.10 (6H, s, CH₃ of TBS), 0.62 (3H, s, H18), 0.88 (3H, s, H19), 0.94-1.41 (13H, m), 1.02 (9H, s, *t*-Bu of TBS), 1.47-1.56 (2H, m), 1.61-1.96 (6H, m), 2.16 (1H, dd, *J* = 19.2, 9.2 Hz, H16a), 4.02 (1H, brs, H3). ¹³C{¹H} NMR (125 MHz, C₆D₆) δ -4.66, 13.7, 18.4, 20.6, 21.8, 24.0, 25.5, 26.1, 26.9, 28.8, 30.4, 32.4, 34.7, 35.3, 35.4, 35.7, 37.0, 40.4, 47.6, 51.4, 67.9, 218.2. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₂₅H₄₄O₂SiNa 427.3008; Found 427.3014.



Enone 12 [CAS: 64647-72-9].^{S4} Trimethylsilyl trifluoromethanesulfonate (TMSOTf, 24 mL, 0.13 mol) was added to a solution of ketone 11 (18.2 g, 45.0 mmol) and Et₃N (25 mL, 0.18 mol) in CH₂Cl₂ (900 mL) at 0 °C. After the reaction mixture was stirred at 0 °C for 20 min and at room temperature for 40 min, saturated aqueous NaHCO₃ (300 mL) was added to the mixture. The resultant mixture was extracted with hexane (600 mL x3). The combined organic layers were washed with brine (100 mL x2), dried over Na₂SO₄, filtered, and concentrated to afford the crude silyl enol ether S2 [CAS: 914644-97-6]^{S4} (19.9 g), which was used in the next reaction without further purification.

 $Pd(OAc)_2$ (12.1 g, 53.9 mmol) was added to a solution of the above crude silyl enol ether **S2** (19.9 g) in CH₃CN (720 mL) and CH₂Cl₂ (180 mL) at room temperature. After the reaction mixture was stirred at room temperature for 25 h, saturated aqueous NaHCO₃ (300 mL) was added to the mixture. The resultant mixture was filtered through a pad of Celite with EtOAc

(300 mL) and extracted with EtOAc (300 mL x3). The combined organic layers were washed with brine (200 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (500 g, hexane/EtOAc = 100/1 to 20/1) to afford enone **12** (13.2 g, 32.8 mmol) and ketone **11** (2.66 g, 6.57 mmol) in 73% and 15% yields over 2 steps, respectively: white solid. m.p. 122-124 °C. $[\alpha]_{D}^{25}$ -17 (*c* 1.74, CHCl₃). IR (film) 2931, 2858, 1711, 1455, 1369, 1252, 1058, 1003 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 0.01 (6H, s, CH₃ of TBS), 0.88 (9H, s, *t*-Bu of TBS), 1.00 (3H, s), 1.04 (3H, s), 1.16-1.27 (3H, m), 1.38-1.58 (8H, m), 1.70-1.74 (1H, m), 1.81-1.99 (5H, m), 2.33 (1H, d, *J* = 11.2 Hz), 4.03 (1H, s, H3), 6.01 (1H, dd, *J* = 5.5, 3.0 Hz, H16), 7.52 (1H, d, *J* = 5.5 Hz, H15). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ -4.88, -4.86, 18.1, 20.0, 20.7, 23.9, 25.4, 25.8, 26.5, 28.5, 29.4, 29.9, 32.7, 34.4, 35.4, 36.5, 41.6, 51.2, 57.2, 67.1, 131.6, 158.8, 213.6. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₂₅H₄₂O₂SiNa 425.2852; Found 425.2849.



Ketone 13 [CAS: 914644-98-7].^{S4} *i*-Pr₂NEt (160 mL, 918 mmol) was added to a suspension of enone **12** (13.5 g, 33.5 mmol) and SiO₂ (50.3 g) in C₆F₅CF₃ (85 mL) at room temperature. After being stirred at 80 °C for 4 h, the reaction mixture was filtered through a pad of Celite with EtOAc (500 mL) and concentrated. The residue was purified by flash column chromatography on silica gel (500 g, hexane/EtOAc = 1/0 to 100/1 to 20/1) to afford ketone **13** (6.81 g, 16.9 mmol) and enone **12** (6.13 g, 15.2 mmol) in 50% and 45% yields, respectively: colorless oil. [α]p²⁶+111 (*c* 0.38, CHCl₃). IR (film) 2930, 2860, 1747, 1639, 1449, 1376, 1254, 1158, 1057, 1009 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 0.00 (6H, s, *CH*₃ of TBS), 0.87 (9H, s, *t*-Bu of TBS), 0.98 (3H, s), 1.09 (3H, s), 1.20-1.26 (3H, m), 1.32-1.43 (5H, m), 1.48-1.55 (2H, m), 1.56-1.61 (2H, m), 1.74-1.82 (3H, m), 1.91-1.99 (1H, m), 2.17 (1H, brs), 2.79-2.85 (1H, m, H16a), 2.95-3.02 (1H, m, H16b), 4.00 (1H, s, H3), 5.47 (1H, d, *J* = 1.6 Hz, H15). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ -4.90, -4.87, 18.1, 19.9, 20.8, 23.1, 23.7, 25.8, 26.3, 28.6, 29.7, 33.5, 34.2, 35.5, 35.6, 36.4, 41.1, 41.4, 51.0, 67.2, 112.6, 154.1, 222.9. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₂₅H₄₂O₂SiNa 425.2852; Found 425.2835.

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Ketones 14a and 14b. A solution of ketone **13** (10.1 g, 25.1 mmol) in 1,4-dioxane (450 mL) was bubbled with O₂ for 25 min at room temperature. Then cobalt(II) acetylacetonate (Co(acac)₂, 1.30 g, 5.06 mmol) was added to the mixture at room temperature. A solution of PhSiH₃ (9.3 mL, 75 mmol) in 1,4-dioxane (50 mL) was added to the mixture at room temperature over 1 h via a syringe pump under O₂ atmosphere (1 atm). After the reaction mixture was stirred under O₂ atmosphere (1 atm) at room temperature for 4 h, saturated aqueous Na₂S₂O₃ (200 mL) was added to the mixture. The resultant mixture was extracted with EtOAc (200 mL x4). The combined organic layers were washed with brine (50 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (500 g, hexane/EtOAc = 10/1 to 3/1) to afford a 2.3:1 mixture of ketones 14a and 14b (8.71 g, 20.7 mmol) in 82% yield. The C14-stereochemistry of 14a was determined by the X-ray crystallographic analysis of epoxide 6: white solid. m.p. 148-149 °C. [a]_D²⁵+21 (*c* 1.65, CHCl₃). IR (film) 3426, 2931, 2888, 2858, 1732, 1469, 1447, 1376, 1361, 1253, 1064 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 0.01 (6H, s, CH₃ of TBS), 0.87 (9H, s, *t*-Bu of TBS), 0.93 (3H x7/10, s, H19), 0.96 (1H x3/10, s, H19), 0.97 (3H x3/10, s, H18), 1.04 (3H x7/10, s, H18), 1.10-1.91 (18H and 1H x3/10, m), 2.13-2.21 (1H x7/10, m, H15a), 2.33-2.47 (2H, m, H16), 3.94-4.09 (1H, m, H3). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ -4.9, 12.8 (1C x7/10), 18.0 (1C x3/10), 18.1 (1C x7/10), 19.1 (1C x3/10), 19.6 (1C x7/10), 19.8 (1C x7/10), 19.9 (1C x3/10), 23.7 (1C x3/10), 23.8 (1C x7/10), 25.0 (1C x3/10), 25.8, 26.3 (1C x3/10), 26.4 (1C x7/10), 27.3 (1C x7/10), 28.5 (1C x3/10), 28.6 (1C x7/10), 29.7 (1C x7/10), 30.0 (1C x3/10), 30.2 (1C x3/10), 32.1 (1C x7/10), 33.0 (1C x7/10), 33.1 (1C x3/10), 33.2 (1C x3/10), 34.2, 35.2 (1C x7/10), 35.3 (1C x3/10), 35.9, 36.4 (1C x3/10), 38.0 (1C x3/10), 41.5 (1C x7/10), 52.7 (1C x3/10), 53.6 (1C x7/10), 67.0 (1C x7/10), 67.1 (1C x3/10), 81.5 (1C x3/10), 82.7 (1C x7/10), 219.2 (1C x3/10), 221.6 (1C x7/10), one ${}^{13}C{}^{1}H$ peak of 14a was not observed. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₂₅H₄₄O₃SiNa 443.2957; Found 443.2943.



Vinyl iodide 8. NH₂NH₂•H₂O (7.2 mL, 0.15 mol) and Et₃N (21 mL, 0.15 mol) were added to a solution of a 2.3:1 diastereomeric mixture of ketones **14a** and **14b** (3.14 g, 7.46 mmol) in EtOH (750 mL) at room temperature. After being stirred at 50 °C for 13 h, the reaction

mixture was concentrated. A solution of I₂ (5.73 g, 22.6 mmol) in THF (50 mL) was added dropwise to a solution of the residue and Et₃N (21 mL, 0.15 mol) in THF (300 mL) at 0 °C. After the reaction mixture was stirred at 0 °C for 40 min, saturated aqueous Na₂S₂O₃ (150 mL) and saturated aqueous NaHCO₃ (100 mL) were added to the mixture. The resultant mixture was extracted with EtOAc (150 mL x3). The combined organic layers were washed with brine (100 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (250 g, hexane/EtOAc = 1/0 to 70/1 to 10/1) to afford vinyl iodide **8** (1.98 g, 3.73 mmol) in 50% yield: white foam. [α] p^{24} +12 (*c* 0.54, CHCl₃). IR (film) 3458, 2931, 2885, 2857, 1462, 1378, 1359, 1254, 1062 cm⁻¹. ¹H NMR (500 MHz, CDCl₃) δ 0.01 (6H, s, CH₃ of TBS), 0.80-1.05 (2H, m), 0.87 (9H, s, *t*-Bu of TBS), 0.93 (3H, s, H19), 1.03 (3H, s, H18), 1.15-1.26 (3H, m), 1.35-1.55 (6H, m), 1.61-1.97 (6H, m), 2.21 (1H, d, *J* = 16.6 Hz, H15a), 2.55 (1H, d, *J* = 16.6 Hz, H15b), 4.03 (1H, s, H3), 6.09 (1H, s, H16). ¹³C{¹H} NMR (125 MHz, CDCl₃) δ -4.89, -4.87, 17.9, 18.0, 19.7, 21.1, 24.0, 25.8, 26.5, 28.7, 29.7, 34.2, 35.1, 36.0, 36.5, 37.5, 41.4, 42.6, 54.7, 67.1, 82.5, 111.5, 133.5. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₂₅H₄₃IO₂SiNa 553.1975; Found 553.1955.



Bromide S5 [CAS: 19978-33-7].^{\$5} N-bromosuccinimide (NBS, 39.5 g, 222 mmol) and benzoyl peroxide ((BzO)₂, 75% purity, 1.36 g, 4.21 mmol) were successively added to a solution of lactone S3 (9.0 mL, 0.11 mol) in CCl₄ (500 mL) at room temperature. After being stirred at 95 °C for 14.5 h, the reaction mixture was concentrated. The residue was purified by flash column chromatography on silica gel (250 g, hexane/EtOAc = 1/0 to 3/1) to afford the crude dibromide S4 (24.6 g), which was used in the next reaction without further purification. Et₃N (74 mL, 0.53 mol) was added to a solution of the above crude dibromide S4 (24.6 g) in CHCl₃ (500 mL) at 0 °C. After being stirred at 0 °C for 4 h, the reaction mixture was concentrated. The resulting mixture was filtered through a pad of silica gel (150 g, The residue was purified by flash column hexane/EtOAc = 1/1) and concentrated. chromatography on silica gel (500 g, hexane/EtOAc = 1/0 to 30/1 to 20/1) to afford bromide S5 (6.31 g, 36.1 mmol) in 33% yield over 2 steps as white solid. The analytical data of S5 were identical to those reported previously.^{S6}



2-Pyrone 7 [CAS: 181866-08-0].^{S6} Pd(PPh₃)₄ (1.91 g, 1.65 mmol) and (Me₃Sn)₂ (4.4 mL, 21 mmol) were added to a solution of bromide S5 (2.84 g, 16.2 mmol) in 1,4-dioxane (41 mL), and then the mixture was degassed by the freeze-thaw procedure (x3). After being stirred at 100 °C for 12 h, the reaction mixture was concentrated. The residue was purified by flash column chromatography on silica gel (150 g, hexane/EtOAc = 1/0 to 20/1 to 10/1) to afford 2-pyrone **7** (3.34 g, 12.9 mmol) in 80% yield: colorless oil. IR (film) 2958, 2919, 2850, 1739, 1716, 1522, 1411, 1341, 1216, 1131 cm⁻¹. ¹H NMR (400 MHz, C₆D₆) δ -0.05 (9H, s, CH₃ of SnMe₃), 6.04 (1H, d, *J* = 9.2 Hz, H23), 6.45 (1H, dd, *J* = 9.2, 1.6 Hz, H22), 6.66 (1H, s, H21). ¹³C{¹H} NMR (100 MHz, C₆D₆) δ -10.0, 112.0, 117.4, 147.1, 155.0, 160.8. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₈H₁₂O₂SnNa 282.9751; Found 282.9756.



Pentacycle 15. Pd(PPh₃)₄ (3.24 g, 2.80 mmol), CuCl (2.75 g, 27.8 mmol), and LiCl (1.41 g, 33.3 mmol) were added to a mixture of vinyl iodide 8 (2.95 g, 5.56 mmol) and 2-pyrone 7 (4.80 g, 18.5 mmol). DMSO (49 mL) and THF (6.2 mL) were added to the mixture, and then the mixture was degassed by the freeze-thaw procedure (x3). After being stirred at 60 $^{\circ}$ C for 13 h, the reaction mixture was filtered through a pad of silica gel (200 g, hexane/EtOAc = 1/1) and Then pH 7 phosphate buffer (200 mL) was added to the residue. concentrated. The resultant mixture was extracted with Et₂O (200 mL x5). The combined organic layers were washed with brine (100 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography (a column consecutively packed with silica gel 150 g and 10 wt% KF contained silica gel 50 g, hexane/EtOAc = 1/0 to 10/1 to 2/1 to 1/1) to afford pentacycle **15** (1.87 g, 3.75 mmol) in 67% yield: white solid. m.p. 260 °C. $[\alpha]_D^{27}$ -58 (*c* 0.60, CHCl₃). IR (film) 3454, 2935, 2880, 2856, 1727, 1694, 1528, 1444, 1374, 1252, 1070, 833 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 0.01 (6H, s, CH₃ of TBS), 0.87 (9H, s, *t*-Bu of TBS), 0.94 (3H, s, H19), 1.03-1.09 (1H, m), 1.12 (3H, s, H18), 1.17-1.28 (4H, m), 1.35-1.49 (4H, m), 1.50-1.58 (2H, m),

1.62-1.73 (1H, m), 1.77-1.83 (3H, m), 1.86-1.93 (2H, m), 2.24 (1H, dd, J = 17.4, 3.2 Hz, H15a), 2.68 (1H, dd, J = 17.4, 1.8 Hz, H15b), 4.04 (1H, s, H3), 5.77 (1H, dd, J = 3.2, 1.8 Hz, H16), 6.31 (1H, dd, J = 9.8, 1.2 Hz, H23), 7.38 (1H, dd, J = 9.8, 2.8 Hz, H22), 7.438-7.444 (1H, m, H21). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ -4.92, -4.88, 16.2, 18.0, 19.8, 21.2, 24.0, 25.8, 26.5, 28.7, 29.7, 34.2, 35.1, 35.9, 36.1, 38.6, 39.8, 40.9, 52.3, 67.1, 85.8, 115.9, 116.3, 125.0, 144.3, 145.1, 147.5, 161.2. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₃₀H₄₆O₄SiNa 521.3058; Found 521.3060.



Epoxide 6. *m*-Chloroperoxybenzoic acid (*m*-CPBA, 77% purity, 965 mg, 4.31 mmol) was added to a solution of pentacycle 15 (1.65 g, 3.31 mmol) and Na₂CO₃ (465 mg, 4.39 mmol) in CH₂Cl₂ (83 mL) at -78 °C. After the reaction mixture was stirred at -20 °C for 5 h, saturated aqueous $Na_2S_2O_3$ (50 mL) was added to the mixture. The resultant mixture was extracted with The combined organic layers were washed with saturated aqueous EtOAc (50 mL x3). NaHCO₃ (20 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (100 g, hexane/EtOAc = 1/0 to 10/1 to 5/1 to 2/1) to afford epoxide 6 (1.32 g, 2.56 mmol) in 77% yield. The structure of 6 was determined by the X-ray crystallographic analysis (Page S39): colorless prism crystal (recrystallization from hexane/EtOAc by vapor diffusion method). m.p. 240-242 °C. $[\alpha]_D^{25}$ +10 (c 0.85, CHCl₃). IR (film) 3535, 3491, 3080, 2925, 2886, 2859, 1730, 1713, 1539, 1453, 1376, 1254, 1161, 1121 cm⁻¹. ¹H NMR (400 MHz, C₆D₆) δ 0.105 (3H, s, CH₃ of TBS), 0.112 (3H, s, CH₃ of TBS), 0.41-0.48 (1H, m), 0.82-0.94 (2H, m), 0.88 (3H, s, H19), 0.89 (3H, s, H18), 1.02 (9H, s, t-Bu of TBS), 1.05-1.13 (1H, m), 1.19-1.24 (3H, m), 1.29-1.33 (1H, m), 1.45-1.65 (5H, m), 1.69-1.86 (3H, m), 1.87-1.96 (2H, m), 2.10-2.14 (1H, m), 2.99 (1H, s, H16), 3.28 (1H, brs, OH), 4.11 (1H, brs, H3), 5.89 (1H, dd, J = 9.6, 1.2 Hz, H23), 6.61 (1H, dd, J = 9.6, 2.7 Hz, H22), 6.77 (1H, dd, J = 2.7, 1.2 Hz, H21). ¹³C{¹H} NMR (100 MHz, C₆D₆) δ -4.7, -4.6, 13.9, 18.3, 20.0, 21.4, 24.2, 26.1, 26.9, 29.2, 30.1, 34.6, 34.8, 35.0, 35.4, 35.9, 36.4, 40.8, 46.4, 61.2, 67.7, 71.2, 80.6, 113.0, 115.9, 143.0, 151.1, 159.9. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₃₀H₄₆O₅SiNa 537.3007; Found 537.3005.



Hexacycle 16. A solution of InCl₃ (40.1 mg, 181 µmol) in THF (0.60 mL) was added to a solution of epoxide 6 (24.6 mg, 47.8 µmol) in THF (1.0 mL) at room temperature. After the reaction mixture was stirred at 50 °C for 5 h, saturated aqueous NaHCO₃ (5 mL) was added to the mixture at room temperature. The resultant mixture was extracted with EtOAc (5 mL x3). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, filtered, and The residue was purified by flash column chromatography on silica gel (5 g, concentrated. hexane/EtOAc = 1/0 to 2/1 to 1/1) to afford alcohol **16** (14.9 mg, 28.9 µmol) in 60% yield: white solid. m.p. 235-237 °C. [a]D²³ +24 (c 0.37, CHCl₃). IR (film) 3398, 2932, 2858, 1685, 1626, 1452, 1253, 1128, 1066 cm⁻¹. ¹H NMR (500 MHz, CDCl₃) δ 0.01 (3H, s, CH₃ of TBS), 0.02 (3H, s, CH3 of TBS), 0.88 (9H, s, t-Bu of TBS), 0.91 (3H, s, H19), 1.04 (3H, s, H18), 1.08-1.87 (17H, m), 2.08 (1H, d, J = 15.2 Hz, H15a), 2.50 (1H, dd, J = 15.2, 6.6 Hz, H15b), 4.05 (1H, brs, H3), 5.18 (1H, d, J = 6.6 Hz, H16), 5.72 (1H, d, J = 9.4 Hz, H23), 6.75 (1H, s, H21), 7.29 (1H, d, J = 9.4 Hz, H22). ¹³C{¹H} NMR (125 MHz, CDCl₃) δ -4.9, 16.3, 18.1, 20.6, 20.8, 23.7, 25.8, 26.6, 28.6, 29.5, 33.2, 34.2, 35.2, 35.5, 35.9, 37.0, 42.0, 54.3, 67.1, 86.8, 93.3, 101.6, 111.8, 115.4, 136.7, 149.5, 163.6. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₃₀H₄₆O₅SiNa 537.3007; Found 537.2983.



p-Bromobenzoate 18. 1 M aqueous HCl (270 μ L, 270 μ mol) was added to a solution of alcohol 16 (7.4 mg, 14 μ mol) in MeOH (3.2 mL) at room temperature. After the reaction

mixture was stirred at room temperature for 1.5 h, saturated aqueous NaHCO₃ (3 mL) was added to the mixture at 0 °C. The resultant mixture was extracted with EtOAc (3 mL x4). The combined organic layers were washed with brine (3 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (1 g, hexane/EtOAc = 1/0 to 1/1 to 1/2) to afford the crude diol **17** (4.8 mg), which was used in the next reaction without further purification: colorless amorphous. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₂₄H₃₂O₅Na 423.2142; Found 423.2157.

p-BrBzCl (13.2 mg, 60.1 µmol) was added to a solution of the above crude diol **17** (4.8 mg) and N,N-dimethyl-4-aminopyridine (DMAP, 7.8 mg, 64 µmol) in CH₂Cl₂ (0.40 mL) at room temperature. After the reaction mixture was stirred at room temperature for 14 h, DMAP (7.5 mg, 61 µmol) and p-BrBzCl (12.5 mg, 57.0 µmol) were successively added to the mixture. After the reaction mixture was stirred at room temperature for 3 h, saturated aqueous NaHCO₃ (3 mL) was added to the mixture at 0 °C. The resultant mixture was extracted with EtOAc (3 mL x4). The combined organic layers were washed with brine (3 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (1 g, hexane/EtOAc = 1/0 to 2/1 to 1/2) to afford *p*-bromobenzoate **18** (3.3 mg, 5.7 µmol) in 41% yield over 2 steps. The structure of **18** was determined by the X-ray crystallographic analysis (Page S40): colorless plate crystal (recrystallization from hexane/EtOAc by vapor diffusion method). m.p. 201-203 °C. [a]_D²⁴ +6.4 (*c* 0.17, CHCl₃). IR (film) 3496, 2938, 2872, 1711, 1633, 1453, 1389, 1277, 1115, 1012 cm⁻¹. ¹H NMR (500 MHz, DMSO-*d*₆) δ 0.86-2.09 (18H, m), 0.917 (3H, s, H18), 0.924 (3H, s, H19), 2.58 (1H, dd, J = 15.5, 6.3 Hz, H15a), 3.98 (1H, s, OH), 5.07 (1H, d, J = 6.3 Hz, H16), 5.23 (1H, brs, H3), 5.57 (1H, d, J = 9.2 Hz, H23), 7.07 (1H, s, H21), 7.50 (1H, d, J = 9.2 Hz, H22), 7.75 (2H, d, J = 7.8 Hz, aromatic), 7.89 (2H, d, J = 7.8 Hz, aromatic). ¹³C{¹H} NMR (100 MHz, DMSO-*d*₆) δ 16.8, 20.1, 20.4, 23.6, 24.5, 26.1, 30.0, 30.5, 32.6, 34.4, 34.9, 36.4, 36.9, 41.3, 53.8, 71.5, 84.7, 92.6, 101.9, 111.2, 113.5, 127.2, 129.6, 131.1, 132.0, 138.4, 151.2, 163.2, 164.4. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₃₁H₃₅BrO₆Na 605.1509 and 607.1489; Found 605.1493 and 607.1489.



Ketone 19. TMSOTf (1.1 mL, 6.1 mmol) was added to a solution of epoxide **6** (1.32 g, 2.56 mmol) and 2,6-lutidine (1.5 mL, 13 mmol) in CH₂Cl₂ (26 mL) at -78 °C. After the reaction mixture was stirred at room temperature for 1 h, saturated aqueous NaHCO₃ (50 mL) was added to the mixture at -78 °C. The resultant mixture was extracted with EtOAc (50 mL x3). The

combined organic layers were washed with brine (50 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (50 g, hexane/EtOAc = 1/0 to 10/1 to 5/1) to afford ketone **19** (1.03 g, 1.75 mmol) in 68% yield. The C17-stereochemistry of **19** was determined by the NOE experiment: white solid. m.p. 138-140 °C. $[\alpha]_{D^{25}}$ +44 (*c* 0.54, CHCl₃). IR (film) 2930, 2858, 1747, 1540, 1448, 1380, 1253, 1062, 984, 838 cm⁻¹. ¹H NMR (400 MHz, C₆D₆) δ -0.01 (9H, s, *CH*₃ of TMS), 0.13 (3H, s, *CH*₃ of TBS), 0.14 (3H, s, *CH*₃ of TBS), 0.62 (3H, s, H18), 0.85-1.12 (4H, m), 0.87 (3H, s, H19), 1.04 (9H, s, *t*-Bu of TBS), 1.16-1.51 (7H, m), 1.55-1.82 (5H, m), 1.91-1.94 (1H, m), 2.14 (1H, d, *J* = 17.4 Hz, H15a), 2.36 (1H, s, H17), 2.42 (1H, d, *J* = 17.4 Hz, H15b), 4.08 (1H, brs, H3), 6.00 (1H, dd, *J* = 9.6, 1.4 Hz, H23), 6.72 (1H, brs, H21), 6.74 (1H, dd, *J* = 9.6, 2.8 Hz, H22). ¹³C{¹H} NMR (100 MHz, C₆D₆) δ -4.7, -4.6, 2.8, 18.3, 18.4, 20.6, 23.6, 24.1, 26.1, 26.5, 28.8, 30.1, 34.6, 35.9, 36.2, 37.2, 39.9, 41.5, 47.97, 48.04, 61.4, 67.6, 88.3, 115.3, 116.1, 146.1, 151.8, 160.4, 214.1. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₃₃H₅₄O₅Si₂Na 609.3402; Found 609.3374.



Alcohol 24. NaBH₄ (80.6 mg, 2.13 mmol) was added to a solution of ketone 19 (1.03 g, 1.75 mmol) in MeOH (17 mL) and THF (17 mL) at 0 °C. After the reaction mixture was stirred at 0 °C for 1.5 h, saturated aqueous NH₄Cl (50 mL) was added to the mixture. The resultant mixture was extracted with EtOAc (50 mL x3). The combined organic layers were washed with brine (20 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (50 g, hexane/EtOAc = 1/0 to 5/1 to 3/1) to afford alcohol 24 (954 mg, 1.62 mmol) in 93% yield. The C16-stereochemistry of 24 was determined by converting 24 to bufogenin B (2): colorless oil. $[\alpha]_D^{24} + 13$ (c 0.70, CHCl₃). IR (film) 3460, 2931, 2859, 1714, 1634, 1538, 1447, 1379, 1251, 1116, 1061, 994 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 0.02 (6H, s, CH₃ of TBS), 0.26 (9H, s, CH₃ of TMS), 0.81 (3H, s), 0.88 (9H, s, t-Bu of TBS), 0.91 (3H, s), 1.03-1.87 (17H, m), 1.93 (1H, d, J = 14.6 Hz, H15a), 2.24 (1H, dd, J = 14.6, 6.0 Hz, H15b), 2.54 (1H, d, J = 6.4 Hz, H17), 4.05 (1H, brs, H3), 4.29 (1H, dd, J = 6.4, 6.0 Hz, H16), 6.21 (1H, d, J = 9.8 Hz, H23), 7.30 (1H, brs, H21), 7.56 (1H, dd, J = 9.8, 1.7 Hz, H22). ¹³C{¹H} NMR (100 MHz, C₆D₆) δ -4.7, -4.6, 3.1, 18.4, 18.7, 20.9, 23.9, 24.1, 26.1, 26.8, 28.9, 30.2, 34.7, 36.0, 36.3, 37.3, 41.4, 42.8, 43.1, 49.7, 59.1, 67.7, 73.7, 92.7, 113.2, 117.4, 149.1, 150.4, 161.5. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₃₃H₅₆O₅Si₂Na 611.3558; Found 611.3537.



Bufogenin B (2) [CAS: 465-19-0]. A mixture of HF (70 wt% pyridine solution, 1.6 mL, 61 mmol) and pyridine (1.6 mL) was added to a solution of alcohol 24 (183 mg, 311 µmol) in THF (3.1 mL) at room temperature. After the reaction mixture was stirred at room temperature for 77 h, saturated aqueous NaHCO₃ (30 mL) was added to the mixture. The resultant mixture was extracted with EtOAc (30 mL x4). The combined organic layers were washed with brine (10 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (5 g, hexane/EtOAc = 1/1 to 1/5 to 1/2 then CHCl₃/MeOH = 5/1) to afford bufogenin B (2) (113 mg, 281 μ mol) in 90% yield: colorless oil. $[\alpha]_D^{17}$ +44 (c 0.54, DMSO). IR (film) 3397, 2930, 2861, 1712, 1631, 1537, 1448, 1379, 1258, 1029 cm⁻¹. ¹H NMR (500 MHz, DMSO-d₆) δ 0.64 (3H, s, H18), 0.85 (3H, s, H19), 1.02-1.84 (18H, m), 2.44 (1H, dd, *J* = 14.4, 8.3 Hz, H15a), 2.63 (1H, d, *J* = 8.6 Hz, H17), 3.89 (1H, brs, H3), 4.16 (1H, s, OH), 4.19 (1H, brs, OH), 4.41-4.44 (1H, m, H16), 4.52 (1H, d, J = 4.1 Hz, OH), 6.12 (1H, d, J = 9.8 Hz, H23), 7.47 (1H, d, J = 2.3 Hz, H21), 8.08 (1H, dd, J = 9.8, 2.3 Hz, H22). ¹³C{¹H} NMR (100 MHz, DMSO-*d*₆) δ 16.9, 20.9, 21.0, 23.9, 26.5, 27.6, 29.6, 33.2, 34.8, 35.0, 35.7, 39.6 (deduced from HMQC), 41.5, 42.8, 48.8, 57.6, 64.7, 70.6, 83.3, 111.0, 118.8, 150.3, 151.5, 161.7. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₂₄H₃₄O₅Na 425.2298; Found 425.2293.



Acetate 25. Ac₂O (0.20 mL, 2.1 mmol) was added to a solution of alcohol 24 (157 mg, 267 μ mol) in pyridine (2.7 mL) at room temperature. After the reaction mixture was stirred at room temperature for 18 h, saturated aqueous NH₄Cl (30 mL) was added to the mixture. The resultant mixture was extracted with EtOAc (30 mL x3). The combined organic layers were washed with brine (30 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (10 g, hexane/EtOAc = 1/0 to 10/1) to afford acetate 25 (153 mg, 242 µmol) in 91% yield: white solid. m.p. 160-163 °C. [α] $_{D}^{24}$ +9.5

(*c* 0.76, CHCl₃). IR (film) 3523, 3439, 2926, 2858, 1746, 1730, 1714, 1638, 1539, 1448, 1378, 1361, 1249, 1059 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 0.01 (6H, s, *CH*₃ of TBS), 0.18 (9H, s, *CH*₃ of TMS), 0.76 (3H, s, H18), 0.87 (9H, s, *t*-Bu of TBS), 0.90 (3H, s, H19), 1.07-1.87 (18H, m), 1.89 (3H, s, *CH*₃CO), 2.45 (1H, dd, *J* = 15.6, 8.7 Hz, H15a), 2.79 (1H, d, *J* = 8.7 Hz, H17), 4.04 (1H, brs, H3), 5.46 (1H, dd, *J* = 8.7, 8.7 Hz, H16), 6.16 (1H, d, *J* = 9.8 Hz, H23), 7.22 (1H, d, *J* = 2.5 Hz, H21), 7.75 (1H, dd, *J* = 9.8, 2.5 Hz, H22). ¹³C{¹H} NMR (125 MHz, CDCl₃) δ - 4.89, -4.86, 3.1, 18.06, 18.12, 20.5, 21.0, 23.4, 23.9, 25.8, 26.6, 28.5, 29.7, 34.4, 35.7, 35.8, 37.0, 41.0, 41.56, 41.65, 50.5, 57.2, 67.1, 74.0, 90.3, 112.7, 116.8, 149.5, 150.7, 162.0, 170.0. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₃₅H₅₈O₆Si₂Na 653.3670; Found 653.3643.



Bufotalin (3) [CAS: 471-95-4]. A mixture of HF (70 wt% pyridine solution, 3.1 mL, 0.12 mol) and pyridine (3.1 mL) was added to a solution of acetate 25 (153 mg, 242 µmol) in THF (2.4 mL) at room temperature. After the reaction mixture was stirred at room temperature for 124 h, saturated aqueous NaHCO₃ (50 mL) was added to the mixture. The resultant mixture was extracted with EtOAc (50 mL x4). The combined organic layers were washed with brine (20 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (10 g, hexane/EtOAc = 1/1 to 1/3) to afford bufotalin (3) (100 mg, 225 μ mol) in 93% yield: colorless oil. [α] $_{D}^{24}$ +5.3 (c 1.28, CHCl₃). IR (film) 3468, 3057, 2935, 1713, 1633, 1537, 1448, 1378, 1250, 1136, 1092, 1037, 736 cm⁻¹. ¹H NMR (500 MHz, CDCl₃) δ 0.78 (3H, s, H18), 0.94 (3H, s, H19), 1.15-1.32 (5H, m, H1a, H6a, H7a, H11a, H12a), 1.37 (1H, d, J = 13.5 Hz, H4a), 1.42-1.61 (7H, m, H1b, H2, H8, H9, H11b, H12b), 1.75-1.78 (2H, m, H5, H7b), 1.81-1.92 (2H, m, H4b, H6b), 1.83 (1H, d, J = 15.8 Hz, H15a), 1.86 (3H, s, CH₃CO), 2.64 (1H, dd, *J* = 15.8, 8.9 Hz, H15b), 2.86 (1H, d, *J* = 8.9 Hz, H17), 4.14 (1H, brs, H3), 5.53 (1H, dd, J = 8.9, 8.9 Hz, H16), 6.19 (1H, d, J = 9.8 Hz, H23), 7.24 (1H, s, H21), 8.03 (1H, d, J = 9.8 Hz, H22). ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 16.4, 20.9, 21.0, 21.1, 23.7, 26.3, 27.9, 29.5, 33.2, 35.3, 35.5, 35.9, 40.4, 40.8, 42.3, 49.4, 57.1, 66.7, 73.5, 84.4, 113.1, 116.8, 149.1, 150.9, 162.0, 170.0. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₂₆H₃₆O₆Na 467.2404; Found 467.2396.



Bromide 26 and olefin 27. Imidazole (548 mg, 8.05 mmol), CBr₄ (1.31 g, 3.95 mmol), and PPh₃ (1.05 g, 4.00 mmol) were successively added to a solution of alcohol 24 (584 mg, 992 µmol) in CH₂Cl₂ (5.0 mL) at room temperature. After the reaction mixture was stirred at 40 °C for 21 h, imidazole (533 mg, 7.83 mmol), CBr₄ (1.35 g, 4.07 mmol), and PPh₃ (1.06 g, 4.04 mmol) were successively added to the mixture at room temperature. After the reaction mixture was stirred at 40 °C for 20 h, imidazole (542 mg, 7.96 mmol), CBr₄ (1.31 g, 3.95 mmol), and PPh₃ (1.04 g, 3.97 mmol) were successively added to the mixture at room temperature. After the reaction mixture was stirred at 40 °C for 9 h, saturated aqueous NaHCO₃ (50 mL) was added to the mixture at room temperature. The resultant mixture was extracted with EtOAc (50 mL x3). The combined organic layers were washed with brine (30 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (50 g, hexane/EtOAc = 1/0 to 50/1 to 10/1) to afford a 1:1.1 mixture of bromide 26 and olefin 27 (411 mg, 317 µmol for 26, 357 µmol for 27) in 32% and 36% yields, respectively. For characterizations of 26 and 27, a small amount of the mixture The C16-stereochemistry of **26** was determined by the NOE was purified by GPC (CHCl₃). experiment.

26: colorless oil. $[\alpha]_{D}^{32}$ -31 (*c* 0.18, CHCl₃). IR (film) 2931, 2858, 1747, 1641, 1543, 1466, 1380, 1253, 1065, 990 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 0.02 (6H, s, CH₃ of TBS), 0.20 (9H, s, CH₃ of TMS), 0.70 (3H, s, H18), 0.88 (9H, s, *t*-Bu of TBS), 0.90 (3H, s, H19), 1.04-1.16 (1H, m), 1.23-1.34 (3H, m), 1.39-1.42 (3H, m), 1.46-1.58 (4H, m), 1.67-1.73 (3H, m), 1.81-1.89 (3H, m), 2.27-2.41 (2H, m, H15), 2.73 (1H, d, *J* = 8.0 Hz, H17), 4.07 (1H, brs, H3), 4.24 (1H, ddd, *J* = 11.2, 8.0, 8.0 Hz, H16), 6.29 (1H, d, *J* = 9.6 Hz, H23), 7.27-7.28 (1H, m, H21), 7.41 (1H, dd, *J* = 9.6, 2.5 Hz, H22). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ -4.8, 3.1, 18.1, 18.6, 20.9, 23.6, 23.8, 25.8, 26.6, 28.5, 29.7, 34.3, 35.7, 35.8, 37.1, 40.6, 43.4, 45.1, 50.6, 52.1, 63.2, 67.1, 90.8, 115.6, 118.1, 145.0, 149.4, 161.7. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₃₃H₅₅BrO4Si₂Na 673.2714 and 675.2694; Found 673.2744 and 675.2686.

27: colorless oil. $[\alpha]_D^{23}$ +34 (*c* 0.39, CHCl₃). IR (film) 2933, 2887, 2858, 1747, 1447, 1249, 1225, 1153, 1088, 1062 cm⁻¹. ¹H NMR (400 MHz, C₆D₆) δ 0.08 (9H, s, *CH*₃ of TMS), 0.12 (6H, s, *CH*₃ of TBS), 0.82-0.99 (2H, m), 0.90 (3H, s, H19), 0.92 (3H, s, H18), 1.03 (9H, s, *t*-Bu of TBS), 1.14-1.52 (9H, m), 1.65-2.00 (6H, m), 2.18 (1H, dd, *J* = 17.4, 3.2 Hz, H15a), 2.35 (1H, dd, *J* = 17.4, 1.4 Hz, H15b), 4.04 (1H, brs, H3), 5.21 (1H, brs, H16), 5.97 (1H, dd, *J* = 9.8, 0.9 Hz, H23), 6.75 (1H, dd, *J* = 9.8, 2.5 Hz, H22), 7.07 (1H, d, *J* = 2.5 Hz, H21). ¹³C{¹H} NMR

 $\begin{array}{l} (100 \text{ MHz}, C_6D_6) \, \delta \, \text{-}4.7, \, \text{-}4.6, \, 2.9, \, 16.7, \, 18.4, \, 20.1, \, 21.9, \, 24.2, \, 26.1, \, 27.0, \, 29.0, \, 30.2, \, 34.6, \, 35.3, \\ 36.5, \, 36.6, \, 38.6, \, 39.0, \, 42.7, \, 53.0, \, 67.8, \, 90.7, \, 115.6, \, 116.4, \, 123.9, \, 143.1, \, 146.0, \, 147.9, \, 160.0. \\ \text{HRMS} \, (\text{ESI-TOF}) \, [\text{M+Na}]^+ \, \text{m/z: Calcd for } C_{33}\text{H}_{54}\text{O}_4\text{Si}_2\text{Na} \, 593.3453; \, \text{Found} \, 593.3429. \end{array}$



Bufalin (1) [CAS: 465-21-4] and diol 28. Et₃B (1.00 M hexane solution, 1.15 mL, 1.15 mmol) was added to a solution of a 1:1.1 mixture of bromide 26 and olefin 27 (297 mg, 230 μmol for 26, 258 μmol for 27), and *n*-Bu₃SnH (185 μL, 0.687 mmol) in hexane (8.1 mL) and CH₂Cl₂ (8.1 mL) at room temperature. The mixture was bubbled with O₂ for 3 min at room temperature. After the reaction mixture was stirred under O₂ atmosphere (1 atm) at room temperature for 2 h, Et₃B (1.00 M hexane solution, 1.15 mL, 1.15 mmol) and *n*-Bu₃SnH (185 µmol, 0.687 mmol) were successively added to the mixture. After the reaction mixture was stirred under O₂ atmosphere (1 atm) at room temperature for 2 h, a mixture of 3 M aqueous HCl (2.5 mL, 7.5 mmol) and MeOH (30 mL) was added to the mixture at room temperature. After the reaction mixture was stirred at room temperature for 20 h, saturated aqueous NaHCO₃ (50 mL) was added to the mixture. The resultant mixture was extracted with EtOAc (50 mL x4). The combined organic layers were washed with brine (30 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography (a column consecutively packed with silica gel 50 g and 10 wt% KF contained silica gel 25 g, hexane/EtOAc = 1/0 to 3/2 to 1/1 to 1/3) to afford bufalin (1) (65.2 mg, 169 μ mol) and diol 28 (30.6 mg, 79.6 µmol) in 35% and 16% yields, respectively.

1: colorless oil. $[\alpha]_D^{20}$ +6.3 (*c* 0.97, CHCl₃). IR (film) 3456, 3059, 2933, 1711, 1538, 1450, 1255, 1134 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 0.70 (3H, s, H18), 0.95 (3H, s, H19), 1.10-1.65

(13H, m, H1, H2, H4a, H6a, H7a, H8, H9, H11, H12), 1.66-1.80 (4H, m, H5, H7b, H15a, H16a), 1.86-1.96 (2H, m, H4b, H6b), 2.02-2.11 (1H, m, H15b), 2.15-2.23 (1H, m, H16b), 2.46 (1H, dd, J = 9.6, 6.4 Hz, H17), 4.14 (1H, brs, H3), 6.26 (1H, d, J = 9.8 Hz, H23), 7.23 (1H, d, J = 2.8 Hz, H21), 7.84 (1H, dd, J = 9.8, 2.8 Hz, H22). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 16.5, 21.4 (2C), 23.7, 26.5, 27.9, 28.7, 29.6, 32.7, 33.2, 35.3, 35.6, 35.9, 40.9, 42.3, 48.3, 51.2, 66.8, 85.4, 115.3, 122.7, 146.8, 148.5, 162.4. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₂₄H₃₄O₄Na 409.2349; Found 409.2343.

28: white solid. m.p. 128-131 °C. $[\alpha]_D^{18}$ +73 (*c* 0.12, CHCl₃). IR (film) 3449, 3052, 2934, 2879, 1719, 1638, 1532, 1449, 1379, 1265, 1226, 1153, 1125, 1038 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ 0.98 (3H, s, H18), 1.06-1.99 (16H, m), 1.14 (3H, s, H19), 2.26 (1H, dd, *J* = 17.6, 3.2 Hz, H15a), 2.70 (1H, d, *J* = 17.6 Hz, H15b), 4.14 (1H, brs, H3), 5.77-5.79 (1H, m, H16), 6.33 (1H, dd, *J* = 9.6, 1.2 Hz, H23), 7.37-7.40 (1H, m, H22), 7.447-7.452 (1H, m, H21). ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 16.2, 19.8, 21.1, 23.9, 26.2, 27.9, 29.6, 33.3, 35.2, 35.93, 35.95, 38.6, 39.8, 40.7, 52.3, 66.8, 85.7, 116.0, 116.2, 125.0, 144.3, 145.1, 147.5, 161.2. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₂₄H₃₂O₄Na 407.2193; Found 407.2197.



Alcohol 15 from diol 28. TBSCl (123 mg, 816 μ mol) was added to a solution of diol 28 (15.1 mg, 39.3 μ mol) and imidazole (104 mg, 1.53 mmol) in DMF (0.78 mL) at room temperature. After the reaction mixture was stirred at room temperature for 16 h, saturated aqueous NaHCO₃ (3 mL) was added to the mixture. The resultant mixture was extracted with Et₂O (3 mL x₃). The combined organic layers were washed with brine (3 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (5 g, hexane/EtOAc = 1/0 to 1/1) to afford alcohol 15 (18.4 mg, 36.9 μ mol) in 94% yield.



Carboxylic acid S7 [CAS: 15570-39-5].^{S7} KOH (79.2 mg, 1.41 mmol) was added to a solution of suberic acid (**S6**, 258 mg, 1.48 mmol) in MeOH (11.4 mL) at room temperature. After being stirred at room temperature for 20 min, the reaction mixture was concentrated and dissolved in toluene (2.5 mL). n-Bu₄NBr (48.1 mg, 0.149 mmol) and benzyl bromide (BnBr,

0.20 mL, 1.7 mmol) were successively added to the solution at room temperature. After the reaction mixture was stirred at 130 °C for 13 h, 1 M aqueous HCl (1.7 mL, 1.7 mmol) was added to the mixture at room temperature. The resultant mixture was extracted with Et₂O (10 mL x3). The combined organic layers were washed with H₂O (10 mL) and brine (10 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (10 g, hexane/EtOAc = 1/0 to 3/1) to afford carboxylic acid **S7** (233 mg, 882 µmol) in 60% yield: colorless oil. IR (film) 3033, 2937, 2862, 1735, 1706, 1458, 1414, 1240, 1171 cm⁻¹. ¹H NMR (500 MHz, CDCl₃) δ 1.33-1.36 (4H, m, H4', H5'), 1.61-1.68 (4H, m, H3', H6'), 2.33 (2H, t, *J* = 7.5 Hz, H2'), 2.36 (2H, t, *J* = 7.5 Hz, H7'), 5.11 (2H, s, PhC*H*₂), 7.30-7.38 (5H, m, aromatic). ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 24.4, 24.6, 28.56, 28.61, 33.9, 34.1, 66.1, 128.1, 128.5, 136.0, 173.6, 179.9. HRMS (ESI-TOF) [M-H+2Na]⁺ m/z: Calcd for C₁₅H₁₉O₄Na₂ 309.1073; Found 309.1067.



Benzvl **S9**. DMAP ester (14.9)mg, 122 μmol), 1-ethyl-3-(3dimethylaminopropyl)carbodiimide hydrochloride (EDCI•HCl, 87.6 mg, 457 µmol), and Et₃N (0.10 mL, 0.72 mmol) were successively added to a solution of amine hydrochloride S8 (159 mg, 306 µmol) and carboxylic acid S7 (121 mg, 458 µmol) in CH₂Cl₂ (1.5 mL) at room temperature. After the reaction mixture was stirred at room temperature for 19 h, saturated aqueous NaHCO₃ (5 mL) was added to the mixture. The resultant mixture was extracted with EtOAc (5 mL x4). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (10 g, hexane/EtOAc = 1/0 to 2/1 then CHCl₃/MeOH = 20/1) to afford benzyl ester **S9** (211 mg, 289 μ mol) in 94% yield: colorless oil. [α]_D³⁰ +5.4 (c 0.31, CHCl₃). IR (film) 3441, 3337, 2933, 2863, 1734, 1622, 1551, 1455, 1370, 1253, 1157, 1108 cm⁻¹. ¹H NMR (500 MHz, CDCl₃) δ 1.25-1.31 (4H, m), 1.44 (9H, s, *t*-Bu), 1.45 (6H, s, H4'''), 1.57-1.65 (7H, m), 1.78-1.84 (1H, m), 2.08 (3H, s, H3'''), 2.17-2.20 (2H, m), 2.34 (2H, t, J = 7.5 Hz), 2.51 (3H, s, H1'''), 2.57 (3H, s, H2'''), 2.94 (2H, s, H5'''), 3.21 (1H, brs, H5''a), 3.30 (1H, brs, H5''b), 4.37-4.41 (1H, m, H2''), 5.10 (2H, s, PhCH₂), 6.29 (1H, brs, NH), 6.33 (1H,

Supporting Information

brs, N*H*), 6.39 (1H, d, J = 7.8 Hz, N*H*), 7.31-7.36 (5H, m, aromatic). ¹³C{¹H} NMR (100 MHz, CDCl₃) δ 12.4, 17.9, 19.2, 24.7, 25.1, 25.3, 27.9, 28.55, 28.65, 28.7, 34.1, 36.2, 40.6, 43.2, 66.1, 82.4, 86.3, 117.4, 124.5, 128.11, 128.15, 128.5, 132.2, 132.9, 135.9, 138.3, 156.1, 158.6, 171.4, 173.6 (2C), two ¹³C{¹H} peaks were not observed. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₃₈H₅₆N₄O₈SNa 751.3711; Found 751.3694.



Carboxylic acid 29a. A suspension of benzyl ester **S9** (53.4 mg, 73.3 µmol) and Pd/C (5 wt% Pd on carbon, 10.7 mg) in MeOH (1.5 mL) was stirred under H₂ atmosphere (1 atm) at room temperature for 1 h. The reaction mixture was then filtered through a pad of Celite with EtOAc (10 mL) and concentrated. The residue was purified by flash column chromatography on silica gel (5 g, CHCl₃/MeOH = 1/0 to 30/1 to 10/1) to afford carboxylic acid **29a** (41.5 mg, 65.0 µmol) in 89% yield: colorless oil. $[\alpha]_{D}^{28}$ +7.6 (*c* 0.91, CHCl₃). IR (film) 3442, 3337, 2932, 2863, 1728, 1629, 1552, 1456, 1370, 1252, 1155, 1105 cm⁻¹. ¹H NMR (400 MHz, CD₃OD) δ 1.34-1.37 (4H, m), 1.44 (9H, s, *t*-Bu), 1.46 (6H, s, H4^{***}), 1.53-1.67 (7H, m), 1.74-1.83 (1H, m, H3^{**}a), 2.08 (3H, s, H3^{***}), 2.23 (2H, t, *J* = 7.3 Hz), 2.26-2.30 (2H, m), 2.51 (3H, s, H1^{****}), 2.58 (3H, s, H2^{****}), 3.00 (2H, s, H5^{****}), 3.12-3.20 (2H, m, H5^{****}), 4.21 (1H, brs, H2^{****}), 8.16 (1H, d, *J* = 7.8 Hz, N*H*). ¹³C{¹H} NMR (125 MHz, CD₃OD) δ 12.5, 18.4, 19.6, 25.91, 25.94, 26.8, 27.1, 28.3, 28.7, 29.8, 29.9, 34.9, 36.6, 41.5, 44.0, 54.2, 82.7, 87.7, 118.4, 126.0, 133.5, 134.4, 139.4, 158.1, 159.9, 172.8, 176.3, 177.6. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₃₁H₅₀N4O₈SNa 661.3242; Found 661.3260.



Vulgarobufotoxin (4) [CAS: 464-81-3]. Carboxylic acid **29a** (21.7 mg, 34.0 µmol), DMAP (1.0 mg, 8.2 µmol), and *N*,*N*'-diisopropylcarbodiimide (DIC, 5.0 µL, 32 µmol) were added to a solution of bufotalin (**3**) (12.6 mg, 28.3 µmol) in CH₂Cl₂ (0.28 mL) at room temperature. After the reaction mixture was stirred at room temperature for 15 h, saturated aqueous NaHCO₃ (3 mL) was added to the mixture. The resultant mixture was extracted with EtOAc (3 mL x3). The combined organic layers were washed with brine (3 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (1 g, hexane/EtOAc = 1/0 to 1/1 to 1/2 then CHCl₃/MeOH = 20/1) to afford the crude steroid **30a** (29.8 mg), which was used in the next reaction without further purification: colorless oil. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₅₇H₈₄N₄O₁₃SNa 1087.5648; Found 1087.5644.

A mixture of CF₃CO₂H (720 µL), PhOMe (16 µL), PhSMe (40 µL), and HSCH₂CH₂SH (24 µL) was added to a solution of a part of the above crude steroid **30a** (16.6 mg) in CH₂Cl₂ (1.5 mL) at room temperature. After the reaction mixture was stirred at room temperature for 4.5 h, toluene (20 mL) was added to the mixture and concentrated. The residue was purified by reversed-phase HPLC (column: Inertsil ODS-4 10 x 250 mm, eluent A: MeOH+0.05% CF₃CO₂H, eluent B: H₂O+0.05% CF₃CO₂H, A/B = 60/40, flow rate: 2.0 mL/min, detection: UV 254 nm, $t_R = 21.3$ min) to afford CF₃CO₂H salt of vulgarobufotoxin (**4**) (5.47 mg, 6.28

μmol) in 40% yield over 2 steps. Vulgarobufotoxin (**4**) was further purified by flash column chromatography on silica gel (0.5 g, CH₂Cl₂/MeOH/28 wt% aqueous NH₃ = 28/8/1) to compare the NMR data with those prepared by the Pettit's route:^{S8} colorless oil. [α]_D²⁸ +0.27 (c 0.32, MeOH). IR (film) 3332, 3179, 2931, 2862, 2360, 1711, 1538, 1450, 1379, 1246, 1201, 1135 cm⁻¹. ¹H NMR (500 MHz, CD₃OD) δ 0.78 (3H, s), 0.98 (3H, s), 1.17-1.46 (13H, m), 1.55-1.74 (13H, m), 1.83-1.98 (3H, m), 1.85 (3H, CH₃CO), 2.01-2.06 (1H, m), 2.25 (2H, t, *J* = 7.7 Hz), 2.33 (2H, t, *J* = 7.5 Hz), 2.71 (1H, dd, *J* = 15.5, 8.9 Hz, H15a), 2.97 (1H, d, *J* = 8.9 Hz, H17), 3.16-3.26 (2H, m, H5''), 4.32 (1H, t, *J* = 5.8 Hz, H2''), 5.08 (1H, s, H3), 5.51 (1H, dd, *J* = 8.9, 8.9 Hz, H16), 6.21 (1H, d, *J* = 10.0 Hz, H23), 7.44 (1H, s, H21), 8.25 (1H, d, *J* = 10.0 Hz, H22). ¹³C{¹H} NMR (125 MHz, CD₃OD) δ 17.1, 20.9, 22.2, 22.3, 24.3, 26.0, 26.1, 26.2, 26.8, 27.6, 29.9, 30.0, 30.8, 31.5, 31.7, 35.5, 36.2, 36.7, 37.1, 38.5, 41.1, 41.2, 42.0, 43.0, 50.8, 55.1 (deduced from HMQC), 58.2, 72.1, 75.7, 85.0, 113.2, 119.4, 152.1, 152.8, 158.6, 164.4, 171.9, 175.1, 175.6, 178.5 (deduced from HMBC). HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₄₀H₆₀N₄O₁₀Na 779.4202; Found 779.4214.



Vulgarobufotoxin (4) prepared by the Pettit's route.^{S8} A solution of suberic acid (**S6**, 1.04 g, 5.97 mmol) in Ac₂O (2.1 mL) was stirred at 150 °C for 2 h. The reaction mixture was concentrated and azeotroped with toluene (2 mL x3) to afford the crude acid anhydride **S10**

[CAS: 10521-06-9]^{S9} (995 mg) as white solid, which was used in the next reaction without further purification.

The above crude acid anhydride **S10** (3.6 mg) and DMAP (0.7 mg, 6 µmol) were added to a solution of bufotalin (**3**) (5.2 mg, 12 µmol) in CH₂Cl₂ (0.10 mL) at room temperature. After the reaction mixture was stirred at room temperature for 11 h, saturated aqueous NH₄Cl (3 mL) was added to the mixture. The resultant mixture was extracted with EtOAc (3 mL x4). The combined organic layers were washed with brine (3 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by PTLC (CHCl₃/MeOH = 20/1) to afford the crude steroid **S11** (7.1 mg), which was used in the next reaction without further purification: colorless oil. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₃4H₄₈O₉Na 623.3191; Found 623.3193.

N-methyl morpholine (6.0 μ L, 55 μ mol) was added to a suspension of the above crude steroid S11 (7.1 mg) and activated molecular sieves 3A (MS 3A, 60.5 mg) in THF (0.50 mL) at -10 °C. After the reaction mixture was stirred at -10 °C for 10 min, trichloroethyl chloroformate (7.0 μ L, 52 µmol) was added to the mixture. After the reaction mixture was stirred at -10 °C for 30 min, a solution of L-arginine (8.7 mg, 50 µmol) in 1 M aqueous HCl (15 µL, 15 µmol) and MeOH (150 µL) was added to the mixture. After being stirred at room temperature for 2 h, the reaction mixture was concentrated. The residue was purified by flash column chromatography on silica gel (0.5 g, CH₂Cl₂/MeOH/28 wt% aqueous NH₃ = 28/8/1) to afford vulgarobufotoxin (4) (5.2 mg, 6.9 μ mol) in 58% yield over 2 steps: colorless oil. $[\alpha]_D^{28}$ -1.2 (c 0.26, MeOH). IR (film) 3386, 2932, 2861, 1712, 1537, 1378, 1246, 1201, 1134 cm⁻¹. ¹H NMR (500 MHz, CD₃OD) δ 0.78 (3H, s, H18), 0.98 (3H, s, H19), 1.22-1.46 (13H, m), 1.55-1.74 (13H, m), 1.84-1.94 (3H, m), 1.85 (3H, s, CH₃CO), 2.01-2.06 (1H, m), 2.25 (2H, t, J = 7.7 Hz), 2.33 (2H, t, J = 7.5 Hz), 2.71 (1H, dd, J = 15.5, 8.9 Hz, H15a), 2.97 (1H, d, J = 8.9 Hz, H17), 3.17-3.23 (2H, m, H5"), 4.28-4.32 (1H, m, H2"), 4.61 (1H, brs, OH), 5.08 (1H, brs, H3), 5.51 (1H, dd, J = 8.9, 8.9 Hz, H16), 6.21 (1H, d, J = 10.1 Hz, H23), 7.44 (1H, s, H21), 8.11 (1H, s, NH), 8.25 (1H, d, J = 10.1 Hz, H22). ¹³C{¹H} NMR (100 MHz, CD₃OD) δ 17.1, 20.9, 22.2, 22.3, 24.3, 25.9, 26.1, 26.3, 26.8, 27.6, 29.9, 30.0, 30.5, 31.5, 31.7, 35.5, 36.2, 36.7, 36.9, 38.5, 41.1, 41.2, 41.9, 43.0, 50.8, 53.9, 58.2, 72.1, 75.7, 85.0, 113.2, 119.4, 152.1, 152.8, 158.6, 164.4, 171.9, 175.1, 175.9, 176.6. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C40H60N4O10Na 779.4204; Found 779.4224.



Carboxylic acid 29b. Acid anhydride S12 (10.4 mg, 104 µmol) and N-methyl morpholine $(23 \,\mu\text{L}, 0.21 \,\text{mmol})$ were successively added to a solution of amine hydrochloride S8 (54.2 mg, 104 µmol) in DMF (0.20 mL) at room temperature. After the reaction mixture was stirred at room temperature for 4 h, saturated aqueous NH₄Cl (3 mL) was added to the mixture. The resultant mixture was extracted with EtOAc (3 mL x4). The combined organic layers were washed with brine (3 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (5 g, CHCl₃/MeOH = 1/0 to 30/1 to 10/1) to afford carboxylic acid **29b** (47.8 mg, 82.0 μ mol) in 79% yield: colorless oil. [α] $_{D^{18}}$ +16 (c 1.2, CHCl₃). IR (film) 3442, 3336, 2973, 2929, 2871, 1731, 1656, 1551, 1454, 1371, 1253, 1157, 1105 cm⁻¹. ¹H NMR (500 MHz, CDCl₃) δ 1.43 (9H, s, *t*-Bu), 1.45 (6H, s, H4'''), 1.54-1.58 (2H, m, H4"), 1.64-1.71 (1H, m, H3"a), 1.82-1.88 (1H, m, H3"b), 2.08 (3H, s, H3"), 2.43-2.64 (3H, m), 2.48 (3H, s, H1""), 2.54 (3H, s, H2""), 2.76-2.81 (1H, m), 2.94 (2H, s, H5"), 3.18 (2H, brs, H5"), 4.44 (1H, brs, H2"), 6.37 (2H, brs, NH), 6.88 (1H, brs, NHCO). ¹³C{¹H} NMR (125 MHz, CDCl₃) δ 12.4, 17.8, 19.2, 24.8, 27.9, 28.5, 29.4, 29.6, 30.5, 40.6, 43.1, 52.6, 82.4, 86.4, 117.5, 124.6, 132.2 (2C), 138.3, 156.3, 158.7, 171.2, 172.7, 176.0. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₂₇H₄₂N₄O₈SNa 605.2616; Found 605.2594.



3-(*N*-succinyl argininyl) bufotalin (5)

3-(*N*-succinyl argininyl) bufotalin (5) [CAS: 146996-65-8]. Carboxylic acid 29b (8.92 mg, 15.3 µmol), DIC (3.0 µL, 19 µmol), and DMAP (0.54 mg, 4.4 µmol) were added to a solution of bufotalin (3) (5.61 mg, 12.6 µmol) in CH₂Cl₂ (0.25 mL) at room temperature. After the reaction mixture was stirred at room temperature for 12 h, saturated aqueous NaHCO₃ (3 mL) was added to the mixture. The resultant mixture was extracted with EtOAc (3 mL x3). The combined organic layers were washed with brine (3 mL), dried over Na₂SO₄, filtered, and concentrated. The residue was purified by flash column chromatography on silica gel (0.5 g, CHCl₃/MeOH = 1/0 to 50/1) to afford the crude steroid **30b** (13.5 mg), which was used in the next reaction without further purification: colorless oil. HRMS (ESI-TOF) [M+Na]⁺ m/z: Calcd for C₅₃H₇₆N₃O₁₃SNa 1031.5022; Found 1031.5021.

A mixture of CF₃CO₂H (450 µL), PhOMe (10 µL), PhSMe (25 µL), and HSCH₂CH₂SH (15 µL) was added to a solution of the above crude steroid **30b** (13.5 mg) in CH₂Cl₂ (1.5 mL) at room temperature. After the reaction mixture was stirred at room temperature for 4 h, toluene (20 mL) was added to the mixture and concentrated. The residue was purified by reversed-phase HPLC (column: Inertsil ODS-3 10 x 250 mm, eluent A: MeOH+0.05% CF₃CO₂H, eluent B: H₂O+0.05% CF₃CO₂H , A/B = 40/60, flow rate: 2.0 mL/min, detection: UV 254 nm, t_R = 32.4 min) to afford CF₃CO₂H salt of 3-(*N*-succinyl argininyl) bufotalin (**5**) (4.21 mg, 5.17 µmol) in 41% yield over 2 steps. 3-(*N*-succinyl argininyl) bufotalin (**5**) was further purified by flash

column chromatography on silica gel (first time: 0.5 g, EtOAc/EtOH/H₂O = 3/7/1; second time: 2.5 g, EtOAc/EtOH/H₂O = 3/7/1) to compare the NMR data with those in the literature:^{S10} colorless oil. [α]_D²⁵ -9.2 (*c* 0.12, MeOH). IR (film) 3355, 2933, 2869, 1714, 1651, 1594, 1380, 1248, 1025, 854 cm⁻¹. ¹H NMR (500 MHz, DMSO-*d*₆) δ 0.65 (3H, s, H18), 0.88 (3H, s, H19), 1.07-1.19 (2H, m), 1.23-1.78 (19H, m), 1.78 (3H, s, CH₃CO), 1.90-1.96 (1H, m), 2.38-2.46 (4H, m, H2', H3'), 2.67 (1H, dd, *J* = 16.9, 8.6 Hz, H15a), 2.88 (1H, d, *J* = 8.6 Hz, H17), 3.03 (2H, brs, H5''), 3.93 (1H, brs, H2''), 4.41 (1H, s, OH), 4.94 (1H, s, H3), 5.38 (1H, dd, *J* = 8.6, 8.6 Hz, H16), 6.19 (1H, d, *J* = 10.0 Hz, H22). ¹³C{¹H} NMR (125 MHz, DMSO-*d*₆) δ 16.6, 20.66, 20.71, 20.8, 23.5, 24.3, 25.3, 26.1, 29.7, 29.8, 29.9, 30.2, 30.3, 34.6, 34.7, 36.6, 39.2 (deduced from HMQC), 40.0 (deduced from HMQC), 40.4, 41.3, 49.0, 53.6, 55.9, 70.0, 73.8, 82.6, 111.7, 117.3, 150.1, 151.4, 157.3, 161.2, 169.5, 169.7, 171.8, 175.9. HRMS (ESI-TOF) [M+H]⁺ m/z: Calcd for C₃₆H₅₃N₄O₁₀ 701.3756; Found 701.3729.

2. Comparison of 1H and $^{13}C\{^1H\}$ NMR data of reported and synthetic 1–5



Table S1.	¹ H NMR	data of reported ^{S1}	¹ and synthetic	bufalin (1) in CDCl ₃
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reported 1 (500 MHz)		synthetic 1 (400 MHz) ^a	Δ (δ reported - δ synthetic)
NO. —	¹ H [δ , multi, J (Hz)]	¹ H [δ , multi, J (Hz)]	_
1a	1.42 (m)^{b}	1.24 (m)	0.18
1b	1.49 (m)	1.49 (m)	0.00
2a	1.52 (m)	1.52 (m)	0.00
2b	1.58 (m)	1.59 (m)	-0.01
3	4.14 (brs)	4.14 (brs)	0.00
4a	1.35 (m)	1.36 (m)	-0.01
4b	1.88 (m)	1.92 (m)	-0.04
5	1.75 (m)	1.76 (m)	-0.01
ба	1.27 (m)	1.27 (m)	0.00
6b	1.88 (m)	1.91 (m)	-0.03
7a	1.18-1.72 (m)	1.28 (m)	-
7b	1.18-1.72 (m)	1.72 (m)	-
8	1.51 (m)	1.53 (m)	-0.02
9	1.61 (m)	1.61 (m)	0.00
11a	1.18-1.72 (m)	1.17 (m)	-
11b	1.18-1.72 (m)	1.42 (m)	-
12a	1.39 (m)	1.36 (m)	0.03
12b	1.49 (m)	1.50 (m)	-0.01
15a	1.69 (m)	1.69 (m)	0.00
15b	2.06 (m)	2.04 (m)	0.02
16a	1.72 (m)	1.74 (m)	-0.02
16b	2.18 (m)	2.19 (m)	-0.01
17	2.46 (m)	2.46 (dd, 9.6, 6.4)	0.00
18	0.70 (s)	0.70 (s)	0.00
19	0.95 (s)	0.95 (s)	0.00

21	7.24 (d, 2.6)	7.23 (d, 2.8)	0.01
22	7.85 (dd, 9.7, 2.6)	7.84 (dd, 9.8, 2.8)	0.01
23	6.26 (d, 9.7)	6.26 (d, 9.8)	0.00

^{*a*}Chemical shifts were determined by using ¹H-¹³C HMQC spectra. ^{*b*}The δ value of this peak could be misreported.

N	reported 1 (125 MHz)	synthetic 1 (100 MHz)	$\Delta \left(\delta_{\text{reported}} - \delta_{\text{synthetic}} \right)$
No. —	$^{13}C\{^{1}H\}$ (δ)	$^{13}C\{^{1}H\}(\delta)$	-
1	29.62	29.59	0.03
2	27.86	27.87	-0.01
3	66.85	66.81	0.04
4	33.26	33.24	0.02
5	35.96	35.94	0.02
6	26.51	26.48	0.03
7	21.40	21.37	0.03
8	42.33	42.31	0.02
9	35.37	35.35	0.02
10	35.67	35.63	0.04
11	21.40	21.37	0.03
12	40.91	40.88	0.03
13	48.36	48.32	0.04
14	85.39	85.36	0.03
15	32.71	32.70	0.01
16	28.73	28.71	0.02
17	51.21	51.18	0.03
18	16.53	16.52	0.01
19	23.73	23.72	0.01
20	122.78	122.68	0.10
21	148.52	148.46	0.06
22	146.90	146.79	0.11
23	115.27	115.28	-0.01
24	162.49	162.40	0.09

Table S2. ¹³C{¹H} NMR data of reported^{S11} and synthetic bufalin (1) in CDCl₃



Figure S2. ¹H NMR spectra of synthetic 1 (400 MHz) and reported 1 (500 MHz)



Figure S3. ¹³C{¹H} NMR spectra of synthetic 1 (100 MHz) and reported 1 (125 MHz)



No	reported 2^a	synthetic 2 $(500 \text{ MHz})^b$	$\Delta \left(\delta_{\text{reported}} \text{ - } \delta_{\text{synthetic}} ight)$
NO. —	¹ H [δ , multi, J (Hz)]	¹ H [δ , multi, J (Hz)]	_
3	3.89 (brs)	3.89 (brs)	0.00
16	4.43 (m)	4.43 (m)	0.00
17	2.63 (d, 8.1)	2.63 (d, 8.6)	0.00
18	0.64 (s)	0.64 (s)	0.00
19	0.85 (s)	0.85 (s)	0.00
21	7.47 (d, 1.5)	7.47 (d, 2.3)	0.00
22	8.08 (dd, 9.7, 1.5)	8.08 (dd, 9.8, 2.3)	0.00
23	6.12 (d, 9.7)	6.12 (d, 9.8)	0.00

^{*a*}Internal standard was not indicated. ^{*b*}Chemical shifts were determined by using ¹H-¹³C HMQC spectra.



bufotalin (3)

Table S4.	¹ H NMR	data of re	ported ^{S13}	and sy	ynthetic	bufotalin	(3)) in	CDCl	3
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N.	reported 3 $(500 \text{ MHz})^a$	synthetic 3 $(500 \text{ MHz})^b$	$\Delta \left(\delta_{\text{reported}} - \delta_{\text{synthetic}} ight)$
INO.	¹ Η [δ]	¹ H [δ , multi, J (Hz)]	
1a	-	1.25 (m)	-
1b	1.49	1.49 (m)	0.00
2a	1.51	1.52 (m)	-0.01
2b	1.56	1.58 (m)	-0.02
3	4.14	4.14 (brs)	0.00
4a	1.37	1.37 (m)	0.00
4b	1.90	1.89 (m)	0.01
5	1.77	1.77 (brd, 12.6)	0.00
6a	1.29	1.29 (m)	0.00
6b	1.90	1.88 (m)	0.02
7a	1.21	1.20 (m)	0.01
7b	1.77	1.76 (m)	0.01
8	1.54	1.54 (m)	0.00
9	1.55	1.55 (m)	0.00
11a	1.21	1.20 (m)	0.01
11b	1.44	1.44 (m)	0.00
12a	1.28	1.29 (m)	-0.01
12b	1.56	1.56 (m)	0.00
15a	1.84	1.83 (d, 15.8)	0.01
15b	2.64	2.64 (dd, 15.8, 8.9)	0.00
16	5.54	5.53 (dd, 8.9, 8.9)	0.01
17	2.89	2.86 (d, 8.9)	0.03
18	0.78	0.78 (s)	0.00
19	0.98	0.94 (s)	0.04
21	7.25	7.24 (s)	0.01
22	8.03	8.03 (d, 9.8)	0.00

23	6.19	6.19 (d, 9.8)	0.00
Ac	1.87	1.86 (s)	0.01

^{*a*}Internal standard was not indicated. ^{*b*}Chemical shifts were determined by using ¹H-¹³C HMQC spectra.

Table S5. ¹³C{¹H} NMR data of reported^{S13} and synthetic bufotalin (3) in CDCl₃

NT-	reported 3 $(125 \text{ MHz})^a$	synthetic 3 (125 MHz)	$\Delta \left(\delta_{ ext{reported}} - \delta_{ ext{synthetic}} ight)$
INO	$^{13}C\{^{1}H\}(\delta)$	$^{13}C{^{1}H}(\delta)$	-
1	29.56	29.54	0.02
2	27.91	27.88	0.03
3	66.73	66.71	0.02
4	33.27	33.23	0.04
5	35.88	35.85	0.03
6	26.37	26.34	0.03
7	21.14	21.11	0.03
8	42.32	42.27	0.05
9	35.55	35.52	0.03
10	35.29	35.26	0.03
11	21.06	21.04	0.02
12	40.42	40.38	0.04
13	49.39	49.36	0.03
14	84.41	84.39	0.02
15	40.83	40.79	0.04
16	73.57	73.54	0.03
17	57.18	57.13	0.05
18	16.44	16.44	0.00
19	23.71	23.70	0.01
20	116.83	116.84	-0.01
21	150.97	150.94	0.03
22	149.13	149.13	0.00
23	113.12	113.09	0.03
24	161.93	161.95	-0.02
CO of Ac	170.02	170.02	0.00
CH ₃ of Ac	20.94	20.94	0.00

^{*a*}Internal standard was not indicated.



Table S6. ¹H NMR data of vulgarobufotoxin (4) prepared by our route and Pettit's route^{S8} in CD₃OD

No.	synthetic 4 (a) ^{a} (our route, 500 MHz)	synthetic 4 (b) ^{<i>a</i>} (Pettit's route, 500 MHz)	$\Delta \left(\delta_{\text{synthetic (a)}} - \delta_{\text{synthetic (b)}} \right)$		
	¹ H [δ , multi, J (Hz)]	¹ H [δ , multi, J (Hz)]	-		
3	5.08 (s)	5.08 (brs)	0.00		
16	5.51 (dd, 8.9, 8.9)	5.51 (dd, 8.9, 8.9)	0.00		
17	2.97 (d, 8.9)	2.97 (d, 8.9)	0.00		
18	0.78 (s)	0.78 (s)	0.00		
19	0.98 (s)	0.98 (s)	0.00		
21	7.44 (s)	7.44 (s)	0.00		
22	8.25 (d, 10.0)	8.25 (d, 10.1)	0.00		
23	6.21 (d, 10.0)	6.21 (d, 10.1)	0.00		
2"	4.32 (t, 5.8)	4.30 (m)	0.02		
Ac	1.85 (s)	1.85 (s)	0.00		

^{*a*}Chemical shifts were determined by using ¹H-¹³C HMQC spectra.

synthetic 4 (a) (our route, 100 MHz)	synthetic 4 (b) (Pettit's route, 125 MHz)	$\Delta \left(\delta_{synthetic (a)} \text{ - } \delta_{synthetic (b)} \right)$
¹³ C{ ¹ H} (δ)	$^{13}C\{^{1}H\}(\delta)$	_
17.12	17.13	-0.01
20.92	20.93	-0.01
22.21	22.21	0.00
22.28	22.29	-0.01
24.31	24.31	0.00
25.95	25.94	0.01
26.12	26.12	0.00
26.22	26.30	-0.08
26.84	26.82	0.02
27.60	27.60	0.00
29.90	29.90	0.00
30.01	29.97	0.04
30.76	30.47	0.29
31.53	31.53	0.00
31.69	31.69	0.00
35.51	35.50	0.01
36.25	36.25	0.00
36.69	36.67	0.02
37.12	36.92	0.20
38.53	38.52	0.01
41.12	41.11	0.01
41.19	41.18	0.01
42.02	41.93	0.09
43.02	43.01	0.01
50.76	50.75	0.01
55.12	53.92	1.20
58.20	58.18	0.02
72.10	72.09	0.01
75.71	75.70	0.01
84.99	85.00	-0.01
113.15	113.16	-0.01
119.41	119.41	0.00

Table S7. ${}^{13}C{}^{1}H$ NMR data of vulgarobufotoxin (4) prepared by our route and Pettit's route 58 in CD₃OD

152.12	152.11	0.01
152.81	152.80	0.01
158.60	158.60	0.00
164.44	164.42	0.02
171.93	171.92	0.01
175.06	175.06	0.00
175.60	175.94	-0.34
178.50	176.61	1.89


Table S8. ¹H NMR data of reported^{S10} and synthetic 3-(N-succinyl argininyl) bufotalin (5) in DMSO- d_6

No	reported 5 (600 MHz) a,b	synthetic 5 $(500 \text{ MHz})^c$	$\Delta \left(\delta_{\text{reported}} - \delta_{\text{synthetic}} \right)$
	¹ H [δ , multi, J (Hz)]	¹ H [δ , multi, J (Hz)]	_
3	4.95 (brs)	4.94 (s)	0.01
15a	2.67 (m)	2.67 (dd, 16.9, 8.6)	0.00
16	5.38 (dd, 8.4, 8.4)	5.38 (dd, 8.6, 8.6)	0.00
17	2.88 (d, 9.0)	2.88 (d, 8.6)	0.00
18	0.66 (s)	0.65 (s)	0.01
19	0.89 (s)	0.88 (s)	0.01
21	7.51 (s)	7.51 (s)	0.00
22	8.18 (d, 9.6)	8.18 (d, 10.0)	0.00
23	6.19 (d, 9.6)	6.19 (d, 10.0)	0.00
2"	3.92 (d, 7.2)	3.93 (brs)	-0.01
5"	3.03 (brs)	3.03 (brs)	0.00
OH	4.38 (s)	4.41 (s)	-0.03

^{*a*}Internal standard was not indicated. ^{*b*}The ¹H NMR spectrum was provided by Professors Hui-Min Gao and Zhi-Min Wang. ^{*c*}Chemical shifts were assigned by using ¹H-¹³C HMQC spectra.

Supporting Information

reported 5 (150 MHz) ^{a,b}	synthetic 5 (125 MHz)	$\Delta \left(\delta_{reported} - \delta_{synthetic} \right)$
$^{13}C\{^{1}H\}(\delta)$	$^{13}C\{^{1}H\}(\delta)$	
30.20	29.71	0.49
-	29.80	-
30.32	29.85	0.47
30.67	30.17	0.50
-	30.28	-
35.02	34.55	0.47
35.13	34.69	0.44
37.02	36.58	0.44
-	39.30	-
-	40.00	-
40.85	40.39	0.46
41.76	41.30	0.46
49.43	48.98	0.45
54.13	53.61	0.52
56.40	55.92	0.48
70.41	69.96	0.45
74.24	73.81	0.43
83.04	82.59	0.45
112.12	111.68	0.44
117.72	117.29	0.43
150.57	150.14	0.43
151.89	151.44	0.45
157.77	157.31	0.46
161.59	161.18	0.41
169.88	169.47	0.41
170.03	169.70	0.33
172.17	171.75	0.42
176.24	175.91	0.33

Table S9. ¹³C{¹H} NMR data of reported^{S10} and synthetic 3-(*N*-succinyl argininyl) bufotalin (5) in DMSO- d_6

^{*a*}Internal standard was not indicated. ^{*b*}The ¹³C{¹H} NMR spectrum was provided by Professors Hui-Min Gao and Zhi-Min Wang.



3. X-ray structures and crystallographic data of 6 and 18

Figure S4. X-ray crystallographic structure of **6** (CCDC 2031223) with thermal ellipsoids at the 50% probability level (ORTEP)

Supporting Information



Figure S5. X-ray crystallographic structure of **18** (CCDC 2031224) with thermal ellipsoids at the 50% probability level (ORTEP)

Compound	6	18
CCDC number	2031223	2031224
Molecular formula	C30H46O5Si	C31H35O6Br
Formula weight	514.78	583.52
Temperature (K)	93(2)	296(1)
Wavelength (Å)	0.71075	0.71073
Crystal color, habit	colorless, prism	colorless, plate
Crystal size (mm ³)	0.230 x 0.100 x 0.050	0.200 x 0.200 x 0.050
Crystal system	orthorhombic	orthorhombic
Space group	P2 ₁ 2 ₁ 2 ₁ (#19)	P212121 (#19)
Unit cell dimensions		
a (Å)	7.152(3)	7.7359(3)
b (Å)	12.733(5)	12.9937(4)
c (Å)	31.445(12)	26.7557(14)
α (°)	90.0000	90.0000
β (°)	90.0000	90.0000
γ (°)	90.0000	90.0000
Volume (Å ³)	2864(2)	2689.43(19)
Ζ	4	4
Density (calculated) (g/cm ³)	1.194	1.441
$\mu(MoK\alpha)$ (cm ⁻¹)	1.180	15.769
F(000)	1120.00	1216.00
Index ranges	-9<=h<9	-8<=h<10
	-16<=k<13	-17 < = k < 16
	-29<=l<40	-31<=l<35
Reflections collected	22448	18355
Independent reflections	6545	6093
R(int)	0.1066	0.0354
Completeness to theta	27.483°, 100%	29.094°, 100%
Max. and min. transmission	0.994, 0.773	1.000, 0.841
Refinement method	Full-matrix least-squares on F ²	
No. Observations (All reflections)	6545	6093
No. Variables	463	446
Reflection/Parameter Ratio	14.14	13.66
Goodness-of-fit on F ²	1.063	1.025
Residuals: R1 (<i>I</i> >2.00σ(<i>I</i>))	0.0758	0.0416
Residuals: R (All reflections)	0.1104	0.0544
Residuals: wR2 (All reflections)	0.1470	0.1031
Max. and min. peak in Final Diff. Map $(e-/Å^3)$	0.25, -0.30	0.89, -0.49

Table S10. Crystal data and structure refinement for 6 and 18

4. Biological experiments

Cell culture. MCF-7 human breast cancer cells were obtained from the American Type Culture Collection (ATCC). The cells were maintained with growth medium [Dulbecco's Modified Eagle's Medium (DMEM) low glucose (D6046, Sigma) supplemented with 10 v/v% heat-inactivated fetal bovine serum, penicillin G (100 units/mL), and streptomycin (100 μ g/mL)] under atmosphere of 5% CO₂ at 37 °C. The growth medium was refreshed every 2 or 3 days to reach 70-90% cell confluence.

Sulforhodamine B assay. Growth inhibitory activities of the tested compounds were evaluated according to the literature.^{S14} Cell growth (%) was defined as follows:

$$\text{cell growth (\%)} = \frac{FL(sample) - FL(day0)}{FL(control) - FL(day0)} \times 100 \qquad \{FL(sample) \ge FL(day0)\}$$

$$\text{cell growth (\%)} = \frac{FL(sample) - FL(day0)}{FL(day0)} \times 100 \qquad \{FL(day0) > FL(sample)\}$$

FL = mean of the fluorescence intensity (Ex. 485 nm/Em. 585 nm) day0 = time of addition of the tested compound as serial dilutions control = control wells treated by vehicle (DMSO)

Various concentrations of compounds in the growth medium containing 2% DMSO were prepared by serial dilutions. MCF-7 cells were cultured in 6 cm cell culture dishes filled with the growth medium and harvested by trypsinization at 37 °C for 5-10 min. The collected cells were resuspended into the growth medium at 1.25×10^5 cells/mL. The cell suspension (100 µL/well) was seeded into the black polystyrene flat-bottom 96-well plates (sample-plates, 655090, Greiner bio-one). For calculating FL(day0), the same cell suspension (100 μ L/well) was seeded into an independent 96-well plate (day0-plate). The sample-plate and day0-plate were incubated under atmosphere of 5% CO2 at 37 °C for 24 h. Aliquots of the former medium (100 µL) containing compounds were added to each well of the sample-plate. The growth medium containing 2% DMSO (100 µL) was added to the wells for calculating FL(control) of the sample-plate and the wells for calculating FL(day0) of the day0-plate. To the cells in day0-plate was added an ice-cold solution of 30 w/v% trichloroacetic acid (TCA) in H₂O (100 µL/well). The day0-plate was incubated at 4 °C for 60 min, washed with H₂O (x4), dried, and stored at room temperature. The sample-plate was incubated at 37 °C under atmosphere of 5% CO₂ for 48 h. To the cells in the sample-plate was added an ice-cold solution of 30 w/v% TCA in H₂O (100 µL/well). The sample-plate was incubated at 4 °C for 60 min, washed with H₂O (x4), and dried. To each well of the *day*0-plate and sample-plate was added a solution of sulforhodamine B in H₂O (570 µg/mL, 100 µL/well). The fixed cells were stained at room temperature for 30 min in the dark. The cells were washed with AcOH/H₂O (1/99, x4) and To the stained cells was added a solution of 2-amino-2-(hydroxymethyl)-1,3dried. propanediol (Tris) in H₂O (10 mM, 200 µL/well). The plates were vortexed at room temperature for 10 min. The fluorescence (Ex. 485 nm/Em. 585 nm) of each well was measured on Spectra Max Gemini EM microplate reader (Molecular Devices). Growth inhibitory activities of the tested compounds were evaluated as 50% growth inhibitory

concentration (GI₅₀) values (nM) by means of three replicates. Sigmoidal curve fittings were performed on GraphPad Prism (GraphPad Software, Figure S6). The mean GI₅₀ values and standard deviations of ouabain, 1-5, and 28 were determined from three independent experiments (Table S11).

Table S11. Growth inhibitory activity of ouabain and bufadienolides

compound	GI50 [nM] ^a
ouabain (control)	20.5 ± 1.1
ouabain ^{S15}	19.9 ± 3.6
1	13.3 ± 1.8
2	55.0 ± 5.7
3	28.2 ± 3.9
4	301 ± 36
5	1190 ± 340
28	2390 ± 670

^{*a*}Three independent experiments were performed (mean \pm SD).

Supporting Information



Figure S6. Representative concentration-response curves of ouabain, 1–5, and 28. Ouabain was used as a positive control. The cells were incubated for 48 h in the presence of compounds, and the cell growth (%) was evaluated. Each plot is displayed as mean \pm SD of three replicates.

5. References

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