# $\mathrm{C}_{28}$ Terpenoids from Lamiaceous Plant Perovskia scrophulariifolia: Their Structures and Anti-neuroinflammatory Activity 

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## EXPERIMENTAL SECTION

General Experimental Procedures. Specific rotations were obtained by a JASCO P-2200 digital polarimeter. NMR spectra were measured on a Bruker AVANCE-500 spectrometer using the resonances of $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}\left(\delta_{\mathrm{H}} 8.71 ; \delta_{\mathrm{C}} 123.5\right)$ and $\mathrm{CDCl}_{3}\left(\delta_{\mathrm{H}} 7.26 ; \delta_{\mathrm{C}} 77.0\right)$ as internal references for ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR chemical shifts, respectively. IR, UV, and CD spectra were recorded on a JASCO FT/IR-6200, a Hitachi UV-3900H, and a JASCO J-1500 spectrophotometers, respectively. HRESIMS were recorded on a Waters LCT PREMIER 2695.

Extraction and Isolation. The aerial parts of Perovskia scrophulariifolia collected in Uzbekistan were dried and then extracted with MeOH at rt to give the extract ( 180 g ). The extract was partitioned between EtOAc and water. The EtOAc-soluble materials were further partitioned between $n$-hexane and $90 \% \mathrm{MeOH}$ aq. The $90 \% \mathrm{MeOH}$ aq.-soluble materials ( 94 g ) were subjected to Sephadex LH-20 column chromatography ( $\mathrm{MeOH} /$ water, 6:4 to 10:0) to give eight fractions (frs. 1~8). Fr. 5 was applied to a silica gel column $\left(\mathrm{CHCl}_{3} / \mathrm{MeOH}, 99: 1\right.$ to $\left.0: 100\right)$ to give 17 fractions (frs. 5.1~5.17) including methyl rosmarinate ( $3,2.8 \mathrm{~g}$ ) as fr. 5.17. Fr. 5.14 was further separated by Sephadex LH-20 column chromatography ( $\mathrm{MeOH} /$ water, 6:4) to afford 10 fractions (frs. 5.14.1~5.14.10). Fr. 5.14.7 was loaded on an ODS column (MeOH/water, 6:4 to 10:0), and purified by reversed-phase HPLC (COSMOSIL 5C ${ }_{18}$-MS-II, 10 i.d. x 250 mm , $\mathrm{MeCN} /$ water, $58: 42$ with $0.1 \%$ TFA $)$ to furnish perovsfolins $\mathrm{A}(1,3.7 \mathrm{mg})$ and $\mathrm{B}(2,2.7 \mathrm{mg})$.

Perovsfolin A (1): colorless amorphous solid; $[\alpha]_{\mathrm{D}}+251.1$ (c 0.1, MeOH); UV (MeOH) $\lambda_{\max }$ $249(\varepsilon 14,500), 288(7,000)$, and $335(6,200) \mathrm{nm} ; \mathrm{ECD}(\mathrm{MeOH}) \Delta \varepsilon(\mathrm{nm})+15.2(369),-17.8(296)$, +51.1 (220), and -24.9 (203); ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR (Table S1); HRESIMS $m / z 677.2360[\mathrm{M}+\mathrm{Na}]^{+}$ (calcd for $\mathrm{C}_{38} \mathrm{H}_{38} \mathrm{O}_{10} \mathrm{Na}^{+}, 677.2357$ ).

Perovsfolin $B$ (2): colorless amorphous solid; $[\alpha]_{\mathrm{D}}-190.7$ (c 0.1, MeOH); UV (MeOH) $\lambda_{\max }$ $245(\varepsilon 13,600), 289(7,300)$, and $335(6,900) \mathrm{nm}$; ECD (MeOH) $\Delta \varepsilon(\mathrm{nm})-13.4(367),+13.8(296)$, -43.5 (220), and +34.6 (204); ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR (Table S1); HRESIMS $m / z 677.2366[\mathrm{M}+\mathrm{Na}]^{+}$ (calcd for $\mathrm{C}_{38} \mathrm{H}_{38} \mathrm{O}_{10} \mathrm{Na}^{+}, 677.2357$ ).

Chemical Conversions of Perovsfolins A (1) and B (2). A mixture of perovsfolin A (1, 0.4 $\mathrm{mg}), \mathrm{CH}_{3} \mathrm{I}(60 \mu \mathrm{~L})$, and $\mathrm{K}_{2} \mathrm{CO}_{3}(60 \mathrm{mg})$ in dry acetone $(0.4 \mathrm{~mL})$ was heated at $60{ }^{\circ} \mathrm{C}$ (oil bath) in a screw-capped vial for 5 h with stirring. After removal of inorganic salts by filtration, the filtrate was concentrated under the reduced pressure. The residue was purified by silica gel column chromatography (toluene/EtOAc, 1:0 to 9:1) to give permethylperovsfolin A (1a, 0.4 mg ). Permethylpervsfolin A (1a, 0.2 mg ) was treated with NaOH in $\mathrm{MeOH} /$ acetone $(2: 1,0.75 \mathrm{~mL})$ at rt for 12 h with stirring. The reaction mixture was neutralized with 1 M HCl and diluted by water.

The solution was extracted with EtOAc, washed successively with water and brine, dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$, and concentrated to give a residue. Purification of the residue by silica gel column chromatography ( $n$-hexane/acetone, 6:4 to $0: 1$ ) gave a terpenoid ( $\mathbf{1 b}$ ) and a $\mathrm{C}_{6}-\mathrm{C}_{3}(\mathbf{1 c})$ moieties. A solution of $\mathbf{1 c}$ in $\mathrm{MeOH}(0.2 \mathrm{~mL})$ was treated with drops of TMS-CHN $2(0.6 \mathrm{M}$ solution in $n$ hexane), and the mixture was stirred at rt for 0.5 h . The reaction mixture was concentrated to give trimethyltanshinol (1d). The terpenoid moiety (2b) and trimethyltanshinol (2d) were obtained from perovsfolin $B$ (2) according to the same procedure.

Permethylperovsfolin A (1a): colorless amorphous solid; $[\alpha]_{\mathrm{D}}+126.7$ (c 0.02, MeOH); ${ }^{1} \mathrm{H}$ NMR ( $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}, J$ in Hz$) \delta_{\mathrm{H}} 7.45(1 \mathrm{H}, \mathrm{s}), 7.31$ and 7.27 (each $\left.1 \mathrm{H}, \mathrm{d}, 8.0\right), 7.14(1 \mathrm{H}, \mathrm{s}), 7.12(1 \mathrm{H}$, d, 2.0), $7.01(1 \mathrm{H}, \mathrm{dd}, 8.2,2.0), 6.95(1 \mathrm{H}, \mathrm{s}), 6.95(1 \mathrm{H}, \mathrm{d}, 8.2), 6.05(1 \mathrm{H}, \mathrm{d}, 3),. 5.90(1 \mathrm{H}, \mathrm{dd}, 9.3$, 3.8), 4.48 ( $1 \mathrm{H}, \mathrm{dt}, 11.1,6.1$ ), 3.84, 3.75, 3.71, 3.61, and 3.56 (each $3 \mathrm{H}, \mathrm{s}$ ), 3.43 ( $1 \mathrm{H}, \mathrm{dd}, 14.3,3.8$ ), $3.32(1 \mathrm{H}, \mathrm{dd}, 11.1,3.5), 3.25(1 \mathrm{H}, \mathrm{dd}, 14.3,9.3), 3.23(1 \mathrm{H}, \mathrm{m}), 1.86,1.74,1.45$, and 1.34 (each $1 \mathrm{H}, \mathrm{m}$ ), $1.23 \times 2$ (each $3 \mathrm{H}, \mathrm{d}, 7.0$ ), 1.15 and 1.03 (each 3H, s); HRESIMS $m / z 733.2990[\mathrm{M}+\mathrm{Na}]^{+}$ (calcd for $\mathrm{C}_{42} \mathrm{H}_{46} \mathrm{O}_{10} \mathrm{Na}^{+}, 733.2983$ ).

Terpenoid moiety (1b) of 1: colorless amorphous solid; ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}, J\right.$ in Hz$) \delta_{\mathrm{H}} 7.46$ $(1 \mathrm{H}, \mathrm{s}), 7.32$ and 7.27 (each $1 \mathrm{H}, \mathrm{d}, 8.0), 7.12(1 \mathrm{H}, \mathrm{s}), 6.99(1 \mathrm{H}, \mathrm{s}), 6.10(1 \mathrm{H}, \mathrm{d}, 3.0), 4.50(1 \mathrm{H}, \mathrm{dt}$, $11.4,5.7$ ), $3.78,3.58$, and 3.55 (each $3 \mathrm{H}, \mathrm{s}$ ), 3.31 ( 1 H , dd, $11.4,3.0$ ), $3.26(1 \mathrm{H}$, sept, 7.1$), 1.83$, 1.69, 1.61, and 1.40 (each $1 \mathrm{H}, \mathrm{m}$ ), 1.25 and 1.24 (each $3 \mathrm{H}, \mathrm{d}, 7.1$ ), 1.18 and 1.01 (each $3 \mathrm{H}, \mathrm{s}$ ); HRESIMS $m / z 525.2265[\mathrm{M}+\mathrm{Na}]^{+}$(calcd for $\mathrm{C}_{31} \mathrm{H}_{34} \mathrm{O}_{6} \mathrm{Na}^{+}, 525.2248$ ); UV (MeOH) $\lambda_{\text {max }} 250(\varepsilon$ 14,400), $287(5,400)$, and $335(6,200) \mathrm{nm} ; \mathrm{CD}(\mathrm{MeOH}) \Delta \varepsilon+14.4$ (368), -5.6 (322), -18.2 (293), -0.7 (260), -7.1 (249), +54.2 (222), -41.6 (203). UV ( $\varepsilon$ values) and CD ( $\Delta \varepsilon$ values) data for 1b were estimated on the basis of its UV absorption at 335 nm by comparing that of $\mathbf{1}$, since a small amount of $\mathbf{1 b}$ was obtained ( $<0.1 \mathrm{mg}$ ).

Trimethyltanshinol (1d): colorless amorphous solid; ${ }^{1} \mathrm{H} \mathrm{NMR}\left(\mathrm{CDCl}_{3}, J\right.$ in Hz) $\delta_{\mathrm{H}} 6.80(1 \mathrm{H}$, d, 8.5), $6.75(1 \mathrm{H}, \mathrm{brs}), 6.74(1 \mathrm{H}, \mathrm{m}), 4.43(1 \mathrm{H}, \mathrm{dd}, 6.6,4.3), 3.87,3.86$, and 3.76 (each $3 \mathrm{H}, \mathrm{s}$ ), $3.08(1 \mathrm{H}, \mathrm{dd}, 14.1,4.3)$, and $2.92(1 \mathrm{H}, \mathrm{dd}, 14.1,6.6)$; HRESIMS $m / z 263.0872[\mathrm{M}+\mathrm{Na}]^{+}$(calcd for $\mathrm{C}_{12} \mathrm{H}_{16} \mathrm{O}_{5} \mathrm{Na}^{+}, 263.0890$ ).

Permethylperovsfolin B (2a): colorless amorphous solid; $[\alpha]_{\mathrm{D}}-69.2$ (c 0.02, MeOH); ${ }^{1} \mathrm{H}$ NMR ( $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}, J$ in Hz ) $\delta_{\mathrm{H}} 7.45(1 \mathrm{H}, \mathrm{s}), 7.30$ and 7.26 (each $\left.1 \mathrm{H}, \mathrm{d}, 8.0\right), 7.14(1 \mathrm{H}, \mathrm{d}, 2.0), 7.12$ $(1 \mathrm{H}, \mathrm{s}), 7.03(1 \mathrm{H}, \mathrm{dd}, 8.4,2.0), 6.93(1 \mathrm{H}, \mathrm{d}, 8.4), 6.75(1 \mathrm{H}, \mathrm{s}), 6.31(1 \mathrm{H}, \mathrm{d}, 3.2), 5.82(1 \mathrm{H}, \mathrm{dd}$, $10.0,3.9), 4.46(1 \mathrm{H}, \mathrm{dt}, 11.0,5.7), 3.80,3.72,3.71,3.54$, and 3.52 (each $3 \mathrm{H}, \mathrm{s}), 3.46(1 \mathrm{H}, \mathrm{dd}$, $14.0,3.9), 3.31(1 \mathrm{H}, \mathrm{dd}, 11.0,3.2), 3.30(1 \mathrm{H}, \mathrm{dd}, 14.0,10.0), 3.23(1 \mathrm{H}$, sept, 6.9$), 1.23$ and 1.22 (each $3 \mathrm{H}, \mathrm{d}, 6.9$ ), 1.16 and 0.97 (each $3 \mathrm{H}, \mathrm{s}$ ); HRESIMS $m / z 733.2977[\mathrm{M}+\mathrm{Na}]^{+}$(calcd for $\mathrm{C}_{42} \mathrm{H}_{46} \mathrm{O}_{10} \mathrm{Na}^{+}, 733.2983$ ).

Terpenoid moiety (2b) of 2: colorless amorphous solid; ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}\right)$ data was identical to that of 1b; HRESIMS $m / z 525.2260[\mathrm{M}+\mathrm{Na}]^{+}$(calcd for $\mathrm{C}_{31} \mathrm{H}_{34} \mathrm{O}_{6} \mathrm{Na}^{+}, 525.2248$ ); UV (MeOH)
$\lambda_{\max } 250(\varepsilon 16,300), 288(5,700)$, and $335(6,900) \mathrm{nm} ; \mathrm{CD}(\mathrm{MeOH}) \Delta \varepsilon-17.6(368),+6.2(323)$, +5.7 (319), +21.9 (293), +0.7 (259), +8.8 (248), -63.5 (222), and +48.2 (204). UV ( $\varepsilon$ values) and CD ( $\Delta \varepsilon$ values) data for $\mathbf{2 b}$ were estimated on the basis of its UV absorption at 335 nm by comparing that of $\mathbf{2}$, since a small amount of $\mathbf{2 b}$ was obtained ( $<0.1 \mathrm{mg}$ ).

Trimethyltanshinol (2d): colorless amorphous solid; ${ }^{1} \mathrm{H} \mathrm{NMR}\left(\mathrm{CDCl}_{3}\right)$ data was not obtained due to low sample amount; HRESIMS $m / z 263.0894[\mathrm{M}+\mathrm{Na}]^{+}$(calcd for $\mathrm{C}_{12} \mathrm{H}_{16} \mathrm{O}_{5} \mathrm{Na}^{+}, 263.0890$ ).

Chemical Conversion of Methyl Rosmarinate (3) to ( $\pm$ )-Trimethyltanshinol $\{( \pm) \mathbf{3 b}\}$. Methyl rosmarinate $(\mathbf{3}, 690 \mathrm{mg})$ was treated with $\mathrm{CH}_{3} \mathrm{I}(0.44 \mathrm{~mL})$ and $\mathrm{K}_{2} \mathrm{CO}_{3}(1.0 \mathrm{~g})$ in dry acetone $(8 \mathrm{~mL})$ and refluxed in an oil bath for 6 h . After filtration and evaporation, the reaction mixture was purified by silica gel column chromatography (toluene/EtOAc, 1:0 to 9:1) to give rosmarinic acid permethylate ( $\mathbf{3 a}, 680 \mathrm{mg}$ ). To a MeOH solution $(15 \mathrm{~mL})$ of $\mathbf{3 a}(610 \mathrm{mg})$ was added $\mathrm{K}_{2} \mathrm{CO}_{3}(75 \mathrm{mg})$ and stirred at rt for 12 h . The reaction mixture was neutralized with 1 M HCl and diluted by water. The solution was extracted with EtOAc, and the organic layer was washed successively with water and brine, dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$, and concentrated to give a residue. Purification of the residue by silica gel column chromatography ( $n$-hexane/EtOAc, $7: 3$ to $1: 1$ ) gave trimethyltanshinol ( $\mathbf{3 b}, 156 \mathrm{mg}$ ). To a $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ solution $(8 \mathrm{~mL})$ of $\mathbf{3 b}(100 \mathrm{mg})$ were added Dess-Martin periodinane $(185 \mathrm{mg})$ and $\mathrm{NaHCO}_{3}(175 \mathrm{mg})$ and stirred at rt for 4 h . The reaction mixture was quenched with sat. $\mathrm{NaHCO}_{3}$ aq. ( 1 mL ) and sat. $\mathrm{Na}_{2} \mathrm{~S}_{2} \mathrm{O}_{3}$ aq. ( 1 mL ). The aqueous phase was extracted with EtOAc, and the organic layer was washed with brine, dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$, and concentrated under reduced pressure to give the residue. The residue was chromatographed over a silica gel column ( $n$-hexane/EtOAc, $8: 2$ ) to give an oxidized product ( 24 mg ). A mixture of the product $(24 \mathrm{mg})$ and $\mathrm{NaBH}_{4}(4.4 \mathrm{mg})$ in $\mathrm{MeOH}(1 \mathrm{~mL})$ was stirred for 1 h in an ice bath $(0$ ${ }^{\circ} \mathrm{C}$ ). The reaction was quenched with water and evaporated to give a residue. The residue was partitioned with $\mathrm{EtOAc} /$ water, and the organic layer was washed with brine, dried over $\mathrm{Na}_{2} \mathrm{SO}_{4}$, and concentrated under reduced pressure. Purification of the residue on a silica gel column ( $n$ hexane/EtOAc, $8: 2$ to $7: 3$ ) afforded ( $\pm$ )-3b ( 2.5 mg ).

Rosmarinic acid permethylate (3a): colorless amorphous solid; $[\alpha]_{\mathrm{D}}+42.3$ (c 1.1, $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ ); ${ }^{1} \mathrm{H} \mathrm{NMR}\left(\mathrm{CDCl}_{3}, J\right.$ in Hz$) \delta_{\mathrm{H}} 7.65(1 \mathrm{H}, \mathrm{d}, 15.8), 7.09(1 \mathrm{H}, \mathrm{dd}, 8.4,1.8), 7.04(1 \mathrm{H}, \mathrm{d}, 1.8), 6.86$ $(1 \mathrm{H}, \mathrm{d}, 8.4), 6.80(3 \mathrm{H}, \mathrm{m}), 6.33(1 \mathrm{H}, \mathrm{d}, 15.8), 5.37(1 \mathrm{H}, \mathrm{dd}, 8.0,4.8), 3.92,3.91,3.86 \times 2$, and 3.75 (each $3 \mathrm{H}, \mathrm{s}$ ), 3.19 ( 1 H , dd, 14.3, 4.8), and 3.14 ( $1 \mathrm{H}, \mathrm{dd}, 14.3,8.0$ ); HRESIMS $m / z 453.1526$ $[\mathrm{M}+\mathrm{Na}]^{+}$(calcd for $\mathrm{C}_{23} \mathrm{H}_{26} \mathrm{O}_{8} \mathrm{Na}^{+}, 453.1520$ ).
$(+)$-Trimethyltanshinol (3b): colorless amorphous solid; $[\alpha]_{\mathrm{D}}+11.7\left(c 0.3, \mathrm{CH}_{2} \mathrm{Cl}_{2}\right)\left\{\right.$ lit. $[\alpha]_{\mathrm{D}}$ $\left.+10.6\left(c 0.67, \mathrm{CH}_{2} \mathrm{Cl}_{2}\right)\right\}{ }^{15} ;{ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right)$ data was identical to that of $\mathbf{1 d}$ : HRESIMS $\mathrm{m} / \mathrm{z}$ $263.0899[\mathrm{M}+\mathrm{Na}]^{+}$(calcd for $\mathrm{C}_{12} \mathrm{H}_{16} \mathrm{O}_{5} \mathrm{Na}^{+}, 263.0890$ ).
( $\pm$ )-Trimethyltanshinol [( $\pm$ )-3b]: colorless amorphous solid; ${ }^{1} \mathrm{H}$ NMR $\left(\mathrm{CDCl}_{3}\right)$ data was identical to that of 1d: HRESIMS $m / z 263.0886[\mathrm{M}+\mathrm{Na}]^{+}$(calcd for $\mathrm{C}_{12} \mathrm{H}_{16} \mathrm{O}_{5} \mathrm{Na}^{+}, 263.0890$ ).

Chiral HPLC Analyses of Trimethyltanshinols (1d, 2d, and 3b). (+)-Trimethyltanshinol (3b) was subjected to HPLC with a chiral column (YMC Chiral Art Cellulose-SB, 5 mm , i.d. $4.6 \times 250$ mm ; $n$-hexane $/ i$-PrOH, $85: 15$; flow $1.0 \mathrm{~mL} / \mathrm{min}$; UV 254 nm ; temp. $40{ }^{\circ} \mathrm{C}$ ) to give a peak at $t_{\mathrm{R}}$ 14.7 min . The chiral resolution of $( \pm)$ - $\mathbf{3 b}$ with the same condition gave a pair of peaks at $t_{\mathrm{R}} 12.1$ $\min \{(-)-\mathbf{3 b}\}$ and $t_{\mathrm{R}} 14.7 \mathrm{~min}\{(+)-\mathbf{3 b}\}$ in the ratio of ca. 1:1. Similarly, the chiral HPLC analyses of $\mathbf{1 d}$ and $\mathbf{2 d}$ were carried out to show a single peak at $t_{\mathrm{R}} 14.7 \mathrm{~min}$ in each case.

Calculation of ECD Spectrum. A conformational search for the possible stereoisomer ( $\mathbf{1 b}$, $1 R, 11 R, 7^{\prime} R, 8^{\prime} R$ ) of the terpenoid moiety (1b) of perovsfolin $\mathrm{A}(\mathbf{1})$ with the Molecular Mechanics gave stable conformers. Further optimization of the initial conformers (Boltzmann distributions over $1 \%$ ) by DFT calculations $\{\mathrm{B} 3 \mathrm{LYP} / 6-31 \mathrm{G}(\mathrm{d})$, in the presence of MeOH with a polarizable continuum model (PCM) \} gave three stable conformers. The absence of imaginary frequencies of the stable conformers were confirmed by calculations of harmonic vibrational frequencies at the B3LYP/6-31G(d) level in the presence of MeOH with PCM. The stable conformers were subjected to TDDFT calculations \{CAM-B3LYP/6-31+G(d), in the presence of MeOH with a $\operatorname{PCM}\}$. Conversion of the resultant rotatory strengths of the lowest 30 excited states for each conformer with half-bands $(0.25 \mathrm{eV})$ by $\operatorname{SpecDis}(\mathrm{v} 1.61)^{\mathrm{S} 1}$ gave Gaussian-type curves. Finally, the calculated ECD spectra were composed after correction based on the Boltzmann distribution, and red-shifted by 15 nm . The conformational search was run on Spartan 18 program (Wavefunction Inc. Irvine, CA.), while DFT calculations were carried out on Gaussian 09 program, ${ }^{\text {S2 }}$ respectively.

Evaluation of Biological Activity. Perovsfolins A (1) and B (2) were evaluated for their inhibitory effect of IL-1 $\beta$ production from LPS stimulated microglial cells and their antiproliferative activity against human cancer cell lines (A549, Hela, and MCF7) by identical procedures as described in our previous report. ${ }^{11}$

## References

(S1) Bruhn, T.; Schaumlöffel, A.; Hemberger, Y.; Bringmann, G. SpecDis, Version 1.61, University of Wuerzburg, Germany, 2013.
(S2) Frisch, M. J. et al., Gaussian 09, Revision C.01, Gaussian, Inc., Wallingford, CT, 2010.

Scheme S1. Possible biogenetic pathway of perovsfolin A (1).









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Figure S1. ECD spectra of perovsfolins A (1) and B (2).


Figure S2. ${ }^{1} \mathrm{H}$ NMR spectrum of perovsfolin $\mathrm{A}(1)$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(500 \mathrm{MHz})$.


Figure S3. ${ }^{13} \mathrm{C}$ NMR spectrum of perovsfolin $\mathrm{A}(\mathbf{1})$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(125 \mathrm{MHz})$.


Figure S4. ${ }^{1} \mathrm{H}-{ }^{1} \mathrm{H}$ COSY spectrum of perovsfolin $\mathrm{A}(1)$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(500 \mathrm{MHz})$.


Figure $\mathrm{S} 5 . \quad \mathrm{HSQC}$ spectrum of perovsfolin $\mathrm{A}(1)$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(500 \mathrm{MHz})$.


Figure S6. HMBC spectrum of perovsfolin $\mathrm{A}(\mathbf{1})$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(500 \mathrm{MHz})$.


Figure S7. ROESY spectrum of perovsfolin $A(1)$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(500 \mathrm{MHz})$.


Figure S8. ${ }^{1} \mathrm{H}$ NMR spectrum of perovsfolin $\mathrm{B}(2)$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(500 \mathrm{MHz})$.


Figure S9. ${ }^{13} \mathrm{C}$ NMR spectrum of perovsfolin $\mathrm{B}(\mathbf{2})$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(125 \mathrm{MHz})$.


Figure S10. ${ }^{1} \mathrm{H}-{ }^{1} \mathrm{H}$ COSY spectrum of perovsfolin $\mathrm{B}(2)$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(500 \mathrm{MHz})$.


Figure S11. HSQC spectrum of perovsfolin $\mathrm{B}(2)$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(500 \mathrm{MHz})$.


Figure S 12 . HMBC spectrum of perovsfolin $\mathrm{B}(2)$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(500 \mathrm{MHz})$.


Figure S13. ROESY spectrum of perovsfolin $\mathrm{B}(2)$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(500 \mathrm{MHz})$.


Figure S14. ${ }^{1} \mathrm{H}$ NMR spectrum of permethylperovsfolin $\mathrm{A}(\mathbf{1 a})$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(500 \mathrm{MHz})$.


Figure $\mathrm{S} 15 . \quad{ }^{1} \mathrm{H}$ NMR spectrum of terpenoid moiety $(\mathbf{1 b})$ of $\mathbf{1}$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(500 \mathrm{MHz})$.


Figure S16. ${ }^{1} \mathrm{H}$ NMR spectrum of trimethyltanshinol (1d) in $\mathrm{CDCl}_{3}(500 \mathrm{MHz})$.


Figure S17. ${ }^{1} \mathrm{H}$ NMR spectrum of permethylperovsfolin $\mathrm{B}(\mathbf{2 a})$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(500 \mathrm{MHz})$.


Figure S18. ${ }^{1} \mathrm{H}$ NMR spectrum of terpenoid moiety (2b) of 2 in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}(500 \mathrm{MHz})$.


Figure S19. ${ }^{1} \mathrm{H}$ NMR spectrum of rosmarinic acid permethylate (3a) in $\mathrm{CDCl}_{3}(500 \mathrm{MHz})$.


Figure S20. ${ }^{1} \mathrm{H}$ NMR spectrum of $(+)$-trimethyltanshinol ( $\mathbf{3 b}$ ) in $\mathrm{CDCl}_{3}(500 \mathrm{MHz})$.


Figure S21. ${ }^{1} \mathrm{H}$ NMR spectrum of $( \pm)$-trimethyltanshinol ( $\mathbf{3 b}$ ) in $\mathrm{CDCl}_{3}(500 \mathrm{MHz})$.


Table S1. ${ }^{1} \mathrm{H}$ and ${ }^{13} \mathrm{C}$ NMR data for perovsfolins $\mathrm{A}(\mathbf{1})$ and $\mathrm{B}(2)$ in $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}$.

| position | 1 |  | 2 |  |
| :---: | :---: | :---: | :---: | :---: |
|  | $\delta_{\text {c }}$ | $\delta_{\mathrm{H}}(J$ in Hz) | $\delta_{\text {c }}$ | $\delta_{\mathrm{H}}(J$ in Hz) |
| 1 | 37.2 | 4.41, dt (11.2, 6.5) | 37.1 | 4.45 , dt (10.9, 6.5) |
| 2 | 24.3 | 1.82, m | 24.2 | 1.68, m |
|  |  | 1.70, m |  | 1.68, m |
| 3 | 35.6 | 1.52, brt (10.8) | 35.6 | 1.51, m |
|  |  | 1.35, brt (10.8) |  | 1.44, m |
| 4 | 35.2 | - | 35.1 | - |
| 5 | $149.6{ }^{\text {a }}$ | - | 149.4 | - |
| 6 | 127.7 | $7.23{ }^{\text {d }}$ | 127.7 | $7.23{ }^{\text {d }}$ |
| 7 | 128.8 | $7.24{ }^{\text {d }}$ | 128.8 | $7.23{ }^{\text {d }}$ |
| 8 | 127.7 | - | $127.9^{\text {b }}$ | - |
| 9 | 144.9 | - | 145.0 | - |
| 10 | 134.9 | - | 134.9 | - |
| 11 | 96.7 | - | 96.7 | - |
| 12 | 197.8 | - | 197.6 | - |
| 13 | 141.1 | - | 141.2 | - |
| 14 | 139.9 | 7.27, s | 139.8 | 7.25, s |
| 15 | 27.4 | 3.10, sept (6.9) | 27.4 | 3.10, sept (6.7) |
| 16 | 21.4 | 1.07, d (6.9) | 21.4 | 1.05, d (6.7) |
| 17 | 21.2 | 1.13, d (6.9) | 21.3 | 1.13, d (6.7) |
| 18 | 31.5 | 1.05 , s | 31.5 | 0.94, s |
| 19 | 31.9 | 1.11, s | 31.8 | 1.11, s |
| $1^{\prime}$ | $135.3{ }^{\text {a }}$ | - | 135.1 | - |
| $2^{\prime}$ | 108.8 | 7.33, s | 108.7 | 7.19, s |
| $3^{\prime}$ | 148.5 | - | 148.4 | - |
| $4^{\prime}$ | 147.1 | - | $147.1^{\text {c }}$ | - |
| $5^{\prime}$ | 107.8 | 7.43, s | 107.8 | 7.39, s |
| $6^{\prime}$ | 133.8 | - | 134.0 | - |
| $7{ }^{\prime}$ | 82.9 | 6.08, d (3.4) | 82.3 | 6.25, d (2.4) |
| $8^{\prime}$ | 59.9 | $3.32{ }^{\text {d }}$ | 60.5 | 3.28, dd (10.9, 2.4) |
| $9^{\prime}$ | 174.4 | - | 173.6 | - |
| $10^{\prime}$ | 128.0 | - | $128.0^{\text {b }}$ | - |
| $11^{\prime}$ | 117.7 | 7.34, d (1.4) | 117.8 | 7.35, brs |
| $12^{\prime}$ | 147.1 | - | $147.2^{\text {c }}$ | - |
| $13^{\prime}$ | 146.3 | - | 146.4 | - |
| $14^{\prime}$ | 116.6 | 7.22, d (8.1) | 116.6 | $7.23{ }^{\text {d }}$ |
| $15^{\prime}$ | 121.0 | 6.88, dd (8.1, 1.4) | 120.9 | 6.91, brd (7.5) |
| $16^{\prime}$ | 37.0 | $3.32{ }^{\text {d }}$ | 37.0 | 3.36, dd (14.0, 3.2) |
|  |  | 3.12, dd (14.5, 9.9) |  | 3.21, dd (14.0, 9.5) |
| $17^{\prime}$ | 73.9 | 5.74, dd (9.9, 3.7) | 74.2 | 5.71, dd (9.5, 3.2) |
| $18^{\prime}$ | 170.4 | - | 170.4 | - |
| OMe | 52.2 | 3.62, s | 52.1 | 3.61, s |

[^0]Table S2. Cartesian coordinates, total energies (E), relative energies ( $\Delta \mathrm{E}$ ), and Boltzmann populations for the stable conformers of terpenoid moiety $\left(\mathbf{1 b}: 1 R, 11 R, 7^{\prime} R, 8^{\prime} R\right)$ of perovsforin A (1).

(Table S2 continued)

| C | 0.529643 | 2.952839 | -0.11502 | C | 1.072744 | 2.974929 | -0.08491 |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| O | -3.31933 | 1.27783 | 2.757701 | O | -3.11565 | 2.033765 | 2.456572 |
| C | -3.20011 | 1.704698 | 1.489011 | C | -2.8085 | 2.356606 | 1.188233 |
| C | -2.09119 | 0.980048 | 0.742643 | C | -1.82709 | 1.369056 | 0.56763 |
| O | -3.89545 | 2.58675 | 1.010978 | O | -3.23977 | 3.350485 | 0.627107 |
| C | -4.33181 | 1.911764 | 3.572469 | C | -4.00815 | 2.924578 | 3.163975 |
| H | -2.20067 | -0.9502 | 1.643033 | H | -2.49461 | -0.45116 | 1.50758 |
| O | 1.855273 | 4.269935 | -1.69124 | O | 2.625574 | 4.107461 | -1.59572 |
| O | 3.455668 | 2.150278 | -2.23032 | O | 3.832101 | 1.763198 | -2.22974 |
| C | 1.01135 | 5.402995 | -1.46546 | C | 1.996775 | 5.361523 | -1.31501 |
| C | 4.62096 | 2.869937 | -1.7868 | C | 5.098547 | 2.260492 | -1.75945 |
| H | -4.5175 | -0.23492 | 0.848651 | H | -3.89903 | 0.789515 | -0.88091 |
| H | -4.05356 | -1.89807 | 0.586251 | H | -4.48772 | 0.533439 | 0.752909 |
| H | -5.15337 | -0.80934 | -1.45013 | H | -4.50044 | -1.93371 | 0.378057 |
| H | -3.78178 | 0.284487 | -1.54875 | H | -5.54943 | -1.07911 | -0.75223 |
| H | -1.39607 | -3.36671 | -3.03224 | H | -1.94407 | -3.43381 | -2.82278 |
| H | 0.876758 | -3.84109 | -2.23276 | H | 0.391159 | -3.90744 | -2.24416 |
| H | 2.63934 | -3.50455 | -0.66389 | H | 2.228743 | -3.66146 | -0.79245 |
| H | -2.78438 | -2.06342 | -4.15556 | H | -3.4435 | -1.52355 | -3.78988 |
| H | -4.23894 | -1.09929 | -3.86808 | H | -5.01826 | -0.97517 | -3.18415 |
| H | -2.63696 | -0.39063 | -3.58314 | H | -3.55311 | -0.02342 | -2.86451 |
| H | -5.07819 | -2.99908 | -2.41246 | H | -5.42993 | -3.18748 | -1.95131 |
| H | -3.56331 | -3.90576 | -2.55436 | H | -3.9832 | -3.74938 | -2.78573 |
| H | -4.1953 | -3.49718 | -0.95544 | H | -4.11505 | -3.93587 | -1.02533 |
| H | 3.850848 | -2.75152 | 2.683925 | H | 4.187307 | -2.2018 | 2.0013 |
| H | 5.404593 | -4.30899 | 1.612544 | H | 4.281395 | -4.45825 | 3.010963 |
| H | 3.745946 | -4.80934 | 1.234724 | H | 2.712123 | -3.69954 | 3.344114 |
| H | 4.735117 | -3.98615 | 0.010582 | H | 2.84366 | -4.95452 | 2.093388 |
| H | 6.008056 | -1.8589 | 1.873281 | H | 5.562139 | -3.87547 | 0.872156 |
| H | 4.756918 | -0.60662 | 1.781597 | H | 4.826476 | -2.83788 | -0.36449 |
| H | 5.280238 | -1.44858 | 0.307011 | H | 4.196321 | -4.47223 | -0.07372 |
| H | -0.8483 | 1.928721 | 2.299703 | H | -0.57038 | 2.102364 | 2.229432 |
| H | 2.927481 | -0.02574 | -0.92652 | H | 2.92264 | -0.33828 | -1.02525 |
| H | -0.14859 | 3.770681 | 0.101997 | H | 0.542863 | 3.887203 | 0.165381 |
| H | -2.07116 | 1.424987 | -0.25519 | H | -1.66016 | 1.732099 | -0.45028 |
| H | -4.26136 | 1.427281 | 4.545575 | H | -4.12741 | 2.484564 | 4.153343 |
| H | -4.12915 | 2.981764 | 3.658952 | H | -3.56539 | 3.920731 | 3.235743 |
| H | -5.32002 | 1.75415 | 3.133986 | H | -4.96985 | 2.981011 | 2.64865 |
| H | 1.410448 | 6.19543 | -2.09935 | H | 2.540827 | 6.099667 | -1.90512 |
| H | -0.02347 | 5.186366 | -1.75457 | H | 0.943997 | 5.350387 | -1.61965 |
| H | 1.0474 | 5.717198 | -0.41596 | H | 2.07459 | 5.610622 | -0.2504 |
| H | 5.309927 | 2.880237 | -2.63371 | H | 5.786908 | 2.177994 | -2.60289 |
| H | 4.364192 | 3.895872 | -1.5052 | H | 5.017636 | 3.306274 | -1.44761 |
| H | 5.085976 | 2.35293 | -0.93867 | H | 5.460946 | 1.646838 | -0.92575 |

(Table S 2 continued)

(Table S2 continued)

| O | -3.08807 | 3.500004 | 0.445557 |
| :---: | :---: | :---: | :---: |
| C | -3.91868 | 3.242353 | 2.985744 |
| H | -2.56708 | -0.28277 | 1.533285 |
| O | 2.792565 | 3.861148 | -1.74733 |
| O | 3.872837 | 1.430808 | -2.2792 |
| C | 2.23072 | 5.157415 | -1.52185 |
| C | 5.164984 | 1.887575 | -1.83835 |
| H | -3.85309 | 0.873589 | -0.96686 |
| H | -4.48159 | 0.793471 | 0.670836 |
| H | -4.63362 | -1.69096 | 0.509718 |
| H | -5.62714 | -0.87609 | -0.69717 |
| H | -2.14198 | -3.52945 | -2.61212 |
| H | 0.175456 | -4.06915 | -2.01768 |
| H | 2.018361 | -3.8388 | -0.57375 |
| H | -3.57942 | -1.71447 | -3.67809 |
| H | -5.12972 | -1.06097 | -3.11555 |
| H | -3.63225 | -0.13117 | -2.89625 |
| H | -5.60776 | -3.12263 | -1.66235 |
| H | -4.19325 | -3.81796 | -2.4522 |
| H | -4.30743 | -3.82358 | -0.68101 |
| H | 3.342648 | -3.18798 | 2.739823 |
| H | 4.698398 | -4.93235 | 1.681969 |
| H | 2.992583 | -5.22235 | 1.295849 |
| H | 4.0835 | -4.53167 | 0.075578 |
| H | 5.601278 | -2.5766 | 1.950943 |
| H | 4.519034 | -1.17743 | 1.837248 |
| H | 4.949769 | -2.08786 | 0.373846 |
| H | -0.51222 | 2.192957 | 2.153431 |
| H | 2.860557 | -0.56423 | -0.97955 |
| H | 0.706645 | 3.827142 | 0.023842 |
| H | -1.58667 | 1.744129 | -0.52382 |
| H | -4.06604 | 2.865831 | 3.997279 |
| H | -3.43194 | 4.220147 | 3.005179 |
| H | -4.87276 | 3.311911 | 2.45787 |
| H | 2.806027 | 5.83833 | -2.1499 |
| H | 1.175891 | 5.186142 | -1.81812 |
| H | 2.331143 | 5.451737 | -0.47079 |
| H | 5.843706 | 1.728638 | -2.67866 |
| H | 5.13739 | 2.950401 | -1.57956 |
| H | 5.50116 | 1.299517 | -0.97583 |
| C | -3.89163 | 0.242563 | -0.06972 |


[^0]:    ${ }^{\text {a }}$ overlapped with the signals of $\mathrm{C}_{5} \mathrm{D}_{5} \mathrm{~N}$; ${ }^{\text {b.c }}$ signals maybe interchangeable; ${ }^{\mathrm{d}}$ overlapped signal

