

Direct Conversion of *N*-Alkylamines to *N*-Propargylamines Through C–H Activation Promoted by Lewis Acid/Organocopper Catalysis: Application to Late-Stage Functionalization of Bioactive Molecules

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1. Procedures, Materials and Instrumentation

General experimental procedures. All reactions were performed in standard, dry glassware fitted with rubber septa under an inert atmosphere of nitrogen unless otherwise described. Stainless steel syringes or cannula were used to transfer air- and moisture-sensitive liquids. Reported concentrations refer to solution volumes at room temperature. Evaporation and concentration *in vacuo* were performed using house vacuum (ca. 40 mm Hg). Column chromatography was performed with SiliaFlash® 60 (40–63 micron) silica gel from Silicycle. Thin layer chromatography (TLC) was used for reaction monitoring and product detection using pre-coated glass plates covered with 0.25 mm silica gel with fluorescent indicator; visualization by UV light ($\lambda_{\text{ex}} = 254 \text{ nm}$) or KMnO_4 stain.

Materials. Reagents were purchased in reagent grade from commercial suppliers and used without further purification, unless otherwise described. Amines and trimethylsilyl propiolate compounds were prepared according to the procedures reported previously.^{1–4} H_2O , in synthetic procedures, refers to distilled water. Tris(pentafluorophenyl)borane, $\text{Cu}(\text{MeCN})_4\text{PF}_6$, Xantphos, and 1,2-bis(diphenylphosphino)ethane were purchased from TCI and used without further purification. Chiral ligands **L4-7**, **L10**, and **L14-19** were prepared according to the literature procedures.^{5–8}

Instrumentation. Proton nuclear magnetic resonance (^1H NMR) spectra and proton-decoupled carbon nuclear magnetic resonance (^{13}C { $^1\text{H}}$ NMR) spectra were recorded at 25 °C (unless stated otherwise) on Inova 600 (600 MHz), Varian Unity/Inova 500 (500 MHz) or Oxford AS400 (400 MHz) spectrometers at the Boston College nuclear magnetic resonance facility. Chemical shifts for protons are reported in parts per million downfield from tetramethylsilane and are referenced to 0 ppm. Chemical shifts for carbon are reported in parts per million downfield from tetramethylsilane and are referenced to the carbon resonances of the solvent. The peak positions are quoted to one decimal place unless they are indistinguishable. The solvent peak was referenced to 77.0 ppm for ^{13}C for CDCl_3 . Benzotrifluoride was used as an external standard for ^{19}F NMR and referenced to –63.0 ppm. $\text{BF}_3 \cdot \text{OEt}_2$ was used as an external standard for ^{11}B NMR and referenced to 0 ppm. Data are represented as follows: chemical shift, integration, multiplicity (br = broad, s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constants in Hertz (Hz).

Infrared spectra were recorded on a Bruker FT-IR Alpha (ATR mode) spectrophotometer. Data are represented as follows: frequency of absorption (cm^{-1}).

High-resolution mass spectrometry was performed on a JEOL AccuTOF-DART (positive mode) at the Mass Spectrometry Facility, Boston College. Chiral HPLC analyses were carried using Agilent 1200 series instruments and Shimadzu chromatograph with Daicel CHIRALPAK® columns or Daicel CHIRALCEL® columns (internal diameter 4.6 mm, column length 250 mm, particle size 5 μ m).

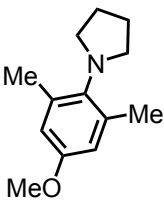
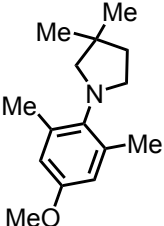
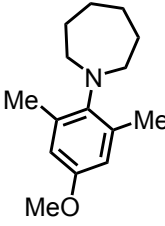
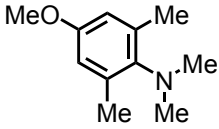
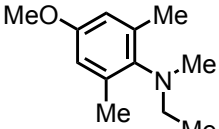
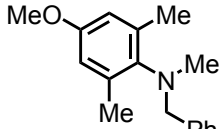
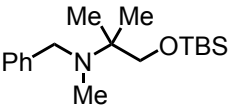
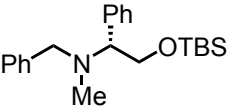
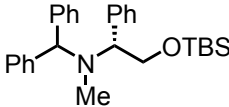
Abbreviations used. Bn = benzyl, COSY = correlated spectroscopy, DART = direct analysis in real time, ESI = electrospray ionization, Et₃N = trimethylamine, EtOAc = ethyl acetate, Et₂O = ethyl ether, HR = high-resolution, HSQC = heteronuclear single quantum coherence, LC = liquid chromatography, MS = mass spectrometry, NOESY = nuclear Overhauser effect spectroscopy, OTf = triflate, PTLC = preparatory thin-layer chromatography, THF = tetrahydrofuran, TLC = thin-layer chromatography, TMS = trimethylsilyl, TBS = *tert*-butyldimethylsilyl, TOF = time-of-flight.

2. Experimental Section

2.1 Substrate Preparation

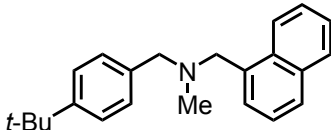
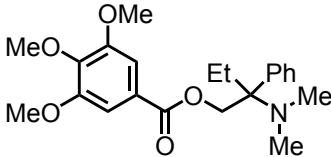
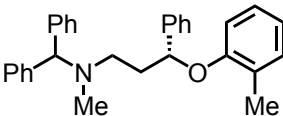
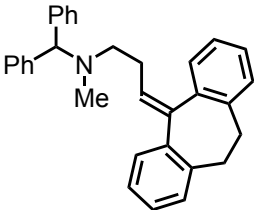
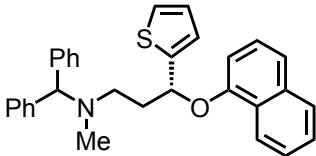
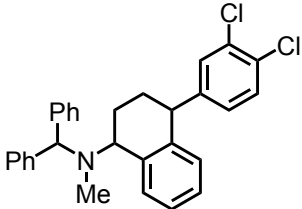
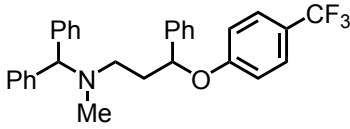
2.1.1 Preparation of Amine Substrates

Table S1-1. List of Amine Substrates (Part 1)

		
1b	1c	1d
		
1e	1f	1g
		
1h	1i	1j

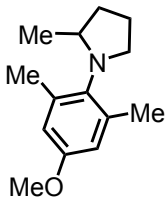
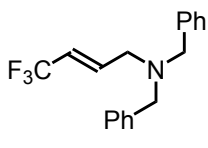
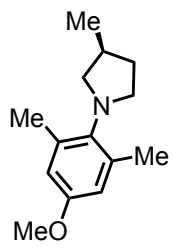
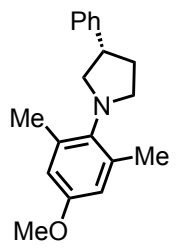
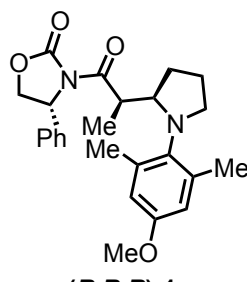
Amines **1b-1g** and **1h-1j** were prepared according to literature procedures.¹ The spectroscopic data for the amine substrates (**1h-1j**) are provided in SI-Section 2.1.

Table S1-2. List of Amine Substrates (Part 2)

 <p>1k</p>	 <p>1l</p>	
 <p>1m</p>	 <p>1n</p>	
 <p>1o</p>	 <p>1p</p>	 <p>1q</p>

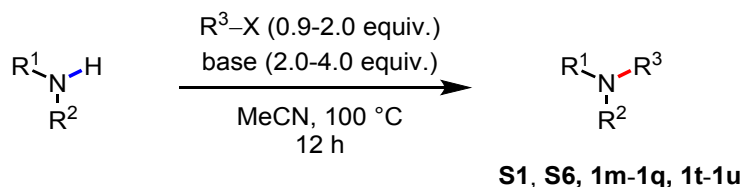
Amines **1m-1q** were prepared according to literature procedures.² Amines **1k** and **1l** were obtained by reacting the commercially available amine hydrochloride salt with NaOH (1.0 M aq.). The spectroscopic data for the amine substrates (**1m-1q**) are provided in SI-Section 2.1.

Table S1-3. List of Amine Substrates (Part 3)

 <p>rac-1r</p>	 <p>1s</p>	 <p>(S)-1t</p>	 <p>(S)-1u</p>	 <p>(R,R,R)-1v</p>
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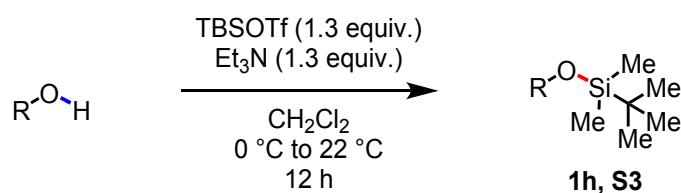
Amines listed above were prepared according to literature procedures.¹ The spectroscopic data for the amine substrates (**1s-1u**) are provided in SI-Section 2.1.

General Procedure for Preparation of Tertiary or Secondary Amines



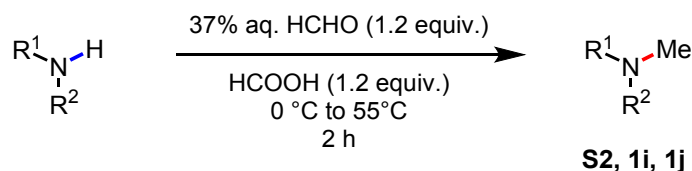
Amines **S1**, **S6**, **1m-1q** and **1t-1u** were prepared by alkylation of the corresponding primary or secondary amines. To a solution of primary or secondary amine (1.0 equiv.) and K_2CO_3 or Et_3N (2.0-4.0 equiv.) in MeCN was added alkyl halide ($\text{R}^3\text{-X}$; 0.9-2.0 equiv.). The reaction mixture was allowed to stir at 100 °C for 12 h. Upon completion (determined by TLC), H_2O was added and the organic material was extracted with EtOAc (3 x 20 mL). The combined organic layers were dried over MgSO_4 , filtered, and concentrated *in vacuo*. The desired amine products were obtained after purification by flash silica gel column chromatography.

General Procedure for TBS Protection of Alcohols



Substrates **1h** and **S3** were prepared by TBS protection of alcohols. To a solution of alcohol in CH_2Cl_2 at 0 °C, Et_3N (1.3 equiv.) and TBSOTf (1.3 equiv.) were added in a dropwise manner. After the addition, the reaction mixture was allowed to warm to 22 °C and stirred for 12 h. Upon completion (determined by TLC), H_2O was added and the organic material was extracted with CH_2Cl_2 (3 x 20 mL). The combined organic layers were dried over MgSO_4 , filtered, and concentrated *in vacuo*. The desired silyl ether products were obtained after purification by flash silica gel column chromatography.

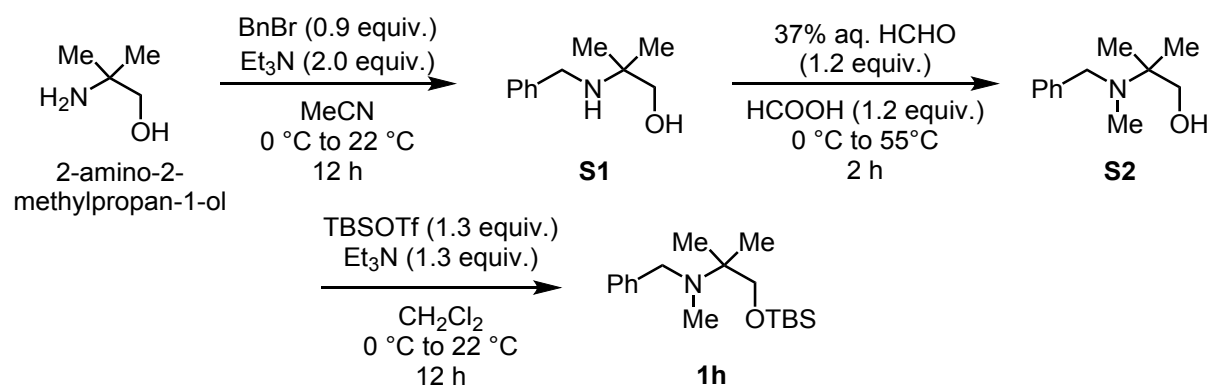
General Procedure for N-Methylation of Secondary Amines



Substrates **S2**, **1i**, and **1j** were prepared by *N*-methylation of secondary amines. A solution of amine and formaldehyde (37% aq. solution, 1.2 equiv.) was cooled to 0 °C. To the reaction mixture was added formic acid (1.2 equiv.) in a dropwise manner. The reaction mixture was

allowed to warm to 55 °C and stirred for 2 h. Upon completion (determined by TLC), the reaction mixture was cooled to 0 °C, NaOH (1.0 M aq. solution) was added until the aqueous layer was alkaline. The organic material was extracted with Et₂O (3 x 20 mL). The combined organic layers were dried (MgSO₄), filtered, and concentrated *in vacuo*. The desired amine products were obtained after flash silica gel column chromatography.

Procedure for Preparation of *N*-Benzyl-1-((*tert*-butyldimethylsilyl)oxy)-*N*,2-dimethylpropan-2-amine (1h)



2-(Benzylamino)-2-methylpropan-1-ol (S1)

2-(Benzylamino)-2-methylpropan-1-ol was prepared following **General Procedure for Preparation of Secondary Amines** using 2-amino-2-methylpropan-1-ol (69 mmol). The amine product **S1** was obtained after purification by flash silica gel column chromatography (EtOAc:hexanes = 1:1) as a colorless oil (9.0 g, 73% yield).

¹H NMR (600 MHz, CDCl₃) δ 7.36 – 7.30 (m, 3H), 7.26 (d, *J* = 1.0 Hz, 2H), 3.69 (s, 2H), 3.35 (s, 2H), 1.15 (d, *J* = 1.0 Hz, 6H).

2-(Benzyl(methyl)amino)-2-methylpropan-1-ol (S2)

2-(Benzyl(methyl)amino)-2-methylpropan-1-ol was prepared following **General Procedure for *N*-Methylation of Secondary Amines** using 2-(benzylamino)-2-methylpropan-1-ol (20 mmol). The amine product **S2** was obtained after purification by flash silica gel column chromatography (EtOAc:Et₃N:hexanes = 20:1:79) as a colorless oil (2.0 g, 50% yield).

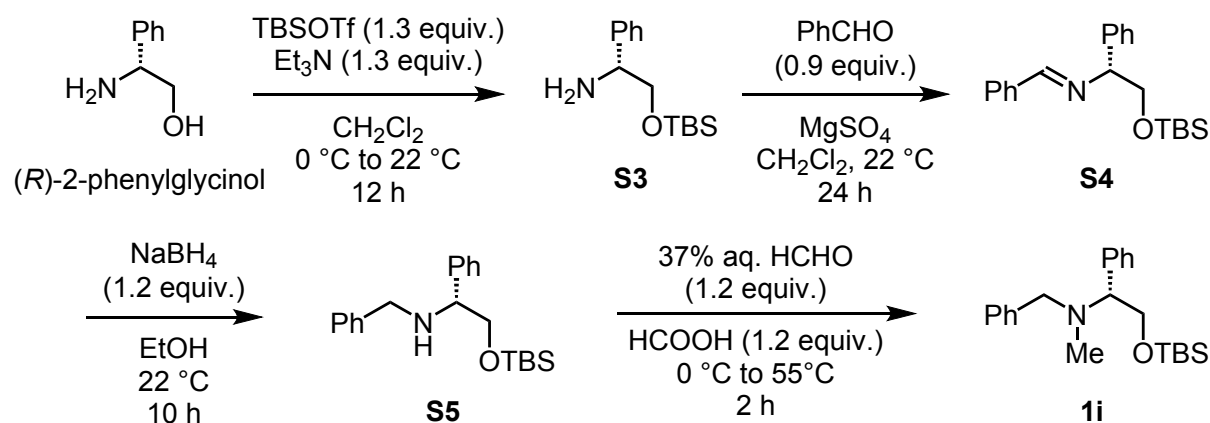
¹H NMR (400 MHz, CDCl₃) δ 7.30 (d, *J* = 1.8 Hz, 1H), 7.26 – 7.20 (m, 4H), 3.52 (s, 2H), 3.44 (s, 2H), 2.09 (s, 3H), 1.13 (s, 6H).

***N*-Benzyl-1-((*tert*-butyldimethylsilyl)oxy)-*N*,2-dimethylpropan-2-amine (**1h**)**

N-Benzyl-1-((*tert*-butyldimethylsilyl)oxy)-*N*,2-dimethylpropan-2-amine was prepared following **General Procedure for TBS Protection of Alcohols** using 2-(benzyl(methyl)amino)-2-methylpropan-1-ol (10 mmol). The amine product **1h** was obtained after purification by flash silica gel column chromatography (Et₂O:hexanes = 1:9) as a colorless oil (3.0 g, 95% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.36 – 7.24 (m, 4H), 7.24 – 7.17 (m, 1H), 3.63 (s, 2H), 3.56 (s, 2H), 2.15 (s, 3H), 1.11 (s, 6H), 0.91 (s, 9H), 0.06 (s, 6H).

Procedure for Preparation of (*R*)-*N*-Benzyl-2-((*tert*-butyldimethylsilyl)oxy)-*N*-methyl-1-phenylethan-1-amine (1i**)**



(*R*)-2-((*tert*-Butyldimethylsilyl)oxy)-1-phenylethan-1-amine (S3**)**

(*R*)-2-((*tert*-Butyldimethylsilyl)oxy)-1-phenylethan-1-amine was prepared following **General Procedure for TBS Protection of Alcohols** using (*R*)-2-amino-2-phenylethan-1-ol (60 mmol). The amine product **S3** was obtained after purification by flash silica gel column chromatography (Et₃N:hexanes = 1:19) as a colorless oil (14.0 g, 93% yield).

¹H NMR (500 MHz, CDCl₃) δ 7.31 – 7.21 (m, 4H), 7.19 – 7.13 (m, 1H), 3.98 (dd, *J* = 8.4, 3.9 Hz, 1H), 3.63 (dd, *J* = 9.8, 3.9 Hz, 1H), 3.43 (dd, *J* = 9.8, 8.3 Hz, 1H), 0.81 (s, 9H), -0.07 (d, *J* = 1.6 Hz, 6H).

(*R,E*)-*N*-(2-((*tert*-Butyldimethylsilyl)oxy)-1-phenylethyl)-1-phenylmethanimine (S4**)**

To a solution of amine **S1** (33 mmol, 1.1 equiv.) and benzaldehyde (30 mmol, 1.0 equiv.) in CH₂Cl₂, was added MgSO₄. The reaction mixture was allowed to stir for 24 h at 22 °C. Upon completion (determined by TLC), the unpurified mixture was filtered over a pad of Celite and rinsed with CH₂Cl₂. The organic layer was concentrated *in vacuo*, and the product obtained

was directly used without further purification.

¹H NMR (400 MHz, CDCl₃) δ 8.26 (s, 1H), 7.76 – 7.65 (m, 2H), 7.49 – 7.37 (m, 2H), 7.37 – 7.28 (m, 3H), 7.28 – 7.21 (m, 2H), 7.20 – 7.12 (m, 1H), 4.33 (dd, *J* = 8.6, 4.4 Hz, 1H), 3.86 – 3.68 (m, 2H), 0.73 (s, 9H), -0.10 (s, 3H), -0.16 (s, 3H).

(*R*)-*N*-Benzyl-2-((*tert*-butyldimethylsilyl)oxy)-1-phenylethan-1-amine (S5)

To a solution of imine **S4** (30 mmol, 1.0 equiv.) in EtOH, was added NaBH₄ (36 mmol, 1.2 equiv.) at 0 °C. The reaction mixture was allowed to stir for 10 h. Upon completion (monitored by TLC), the reaction mixture was diluted with H₂O, and extracted with EtOAc (3 x 20 mL). The combined organic layers were then dried (MgSO₄), filtered, and concentrated *in vacuo*. The amine product **S5** was obtained after purification by flash silica gel column chromatography (Et₃N:hexanes = 1:50) as a colorless oil (10 g, 98% yield).

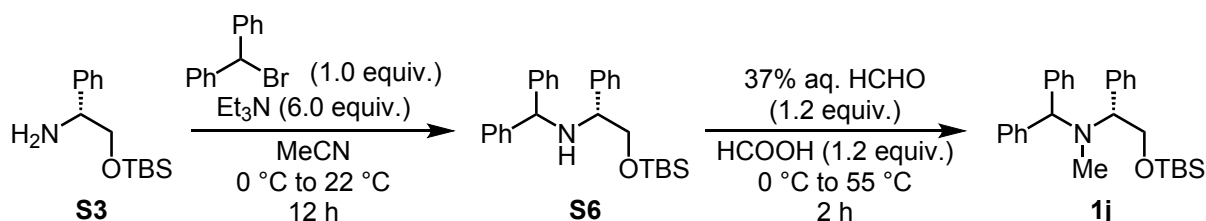
¹H NMR (500 MHz, CDCl₃) δ 7.34 – 7.27 (m, 2H), 7.24 (ddd, *J* = 7.9, 6.7, 1.6 Hz, 2H), 7.22 – 7.15 (m, 5H), 7.13 (td, *J* = 6.9, 1.7 Hz, 1H), 3.71 (dd, *J* = 9.2, 4.0 Hz, 1H), 3.65 (d, *J* = 13.5 Hz, 1H), 3.56 (dd, *J* = 10.0, 4.0 Hz, 1H), 3.51 – 3.42 (m, 2H), 0.78 (d, *J* = 1.8 Hz, 9H), -0.06 – -0.12 (m, 6H).

(*R*)-*N*-Benzyl-2-((*tert*-butyldimethylsilyl)oxy)-*N*-methyl-1-phenylethan-1-amine (1i)

(*R*)-*N*-Benzyl-2-((*tert*-butyldimethylsilyl)oxy)-*N*-methyl-1-phenylethan-1-amine was prepared using **General Procedure for *N*-Methylation of Secondary Amines** using (*R*)-*N*-benzyl-2-((*tert*-butyldimethylsilyl)oxy)-1-phenylethan-1-amine (21 mmol). The amine product **1i** was obtained after purification by flash silica gel column chromatography (EtOAc: Et₃N: hexanes 20:1:79) as a colorless oil (6.8 g, 93% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.41 – 7.34 (m, 3H), 7.34 – 7.28 (m, 5H), 7.28 – 7.24 (m, 1H), 7.24 – 7.18 (m, 1H), 4.07 (dd, *J* = 10.4, 6.1 Hz, 1H), 3.94 – 3.85 (m, 1H), 3.70 – 3.56 (m, 2H), 3.45 (d, *J* = 13.5 Hz, 1H), 2.19 (s, 3H), 0.83 (s, 9H), -0.06 (d, *J* = 7.9 Hz, 6H); [α]_D²⁵ = 1.6° (*c* 1.0, CH₂Cl₂).

Procedure for Preparation of (*R*)-*N*-Benzyl-2-((*tert*-butyldimethylsilyl)oxy)-*N*-methyl-1-phenylethan-1-amine (1j**)**



(*R*)-*N*-Benzhydryl-2-((*tert*-butyldimethylsilyl)oxy)-1-phenylethan-1-amine (S6**)**

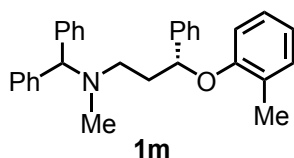
(*R*)-*N*-Benzhydryl-2-((*tert*-butyldimethylsilyl)oxy)-1-phenylethan-1-amine was prepared using **General Procedure for Preparation of Secondary Amines** using (*R*)-2-((*tert*-butyldimethylsilyl)oxy)-1-phenylethan-1-amine (46 mmol). The amine product **S6** was obtained after purification by flash silica gel column chromatography (Et₃N:hexanes = 1:19) as a colorless oil (9.5 g, 49% yield).

¹H NMR (400 MHz, CDCl₃) δ 7.38 – 7.25 (m, 7H), 7.22 (t, *J* = 6.7 Hz, 6H), 7.19 – 7.10 (m, 2H), 4.62 (s, 1H), 3.68 (dd, *J* = 8.5, 3.7 Hz, 1H), 3.61 (dd, *J* = 9.2, 2.3 Hz, 2H), 0.85 (s, 9H), -0.05 (d, *J* = 11.4 Hz, 6H); [α]_D²⁵ = -6.8° (*c* 1.0, CH₂Cl₂).

(*R*)-*N*-Benzyl-2-((*tert*-butyldimethylsilyl)oxy)-*N*-methyl-1-phenylethan-1-amine (1j**)**

(*R*)-*N*-Benzyl-2-((*tert*-butyldimethylsilyl)oxy)-*N*-methyl-1-phenylethan-1-amine was prepared using **General Procedure for *N*-Methylation of Secondary Amines** using (*R*)-*N*-benzhydryl-2-((*tert*-butyldimethylsilyl)oxy)-1-phenylethan-1-amine (24 mmol). The amine product **1j** was obtained after purification by flash silica gel column chromatography (EtOAc:Et₃N: hexanes 20:1:79) as a colorless oil (10.1 g, 98% yield).

¹H NMR (600 MHz, CDCl₃) δ 7.42 (ddd, *J* = 11.6, 8.2, 1.3 Hz, 4H), 7.34 – 7.29 (m, 6H), 7.25 – 7.19 (m, 4H), 7.17 – 7.12 (m, 1H), 4.80 (s, 1H), 4.06 (dd, *J* = 9.7, 5.9 Hz, 1H), 3.98 – 3.89 (m, 2H), 2.13 (s, 3H), 0.85 (s, 9H), -0.03 (d, *J* = 6.5 Hz, 6H).

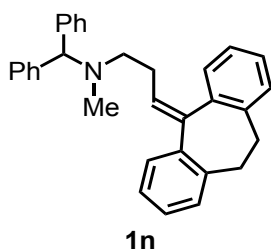


(*R*)-*N*-Benzhydryl-*N*-methyl-3-phenyl-3-(*o*-tolyloxy)propan-1-amine (1m**)**

(*R*)-*N*-Benzhydryl-*N*-methyl-3-phenyl-3-(*o*-tolyloxy)propan-1-amine was prepared following **General Procedure for Preparation of Tertiary Amines** using (*R*)-*N*-methyl-3-phenyl-3-(*o*-

tolyloxy)propan-1-amine (1.5 g, 5.9 mmol), (2-bromoethyl)benzene (1.2 equiv.) and K₂CO₃ (1.2 equiv.). The amine product **1m** was obtained after purification by flash silica gel column chromatography (EtOAc:hexanes = 1:19) as a colorless oil (2.0 g, 82%).

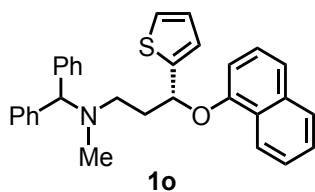
¹H NMR (600 MHz, CDCl₃) δ 7.37 (d, *J* = 7.6 Hz, 2H), 7.29 (d, *J* = 4.4 Hz, 6H), 7.23 (t, *J* = 7.3 Hz, 3H), 7.15 (t, *J* = 6.7 Hz, 1H), 7.10 – 7.05 (m, 4H), 6.95 (t, *J* = 7.8 Hz, 1H), 6.75 (t, *J* = 7.5 Hz, 1H), 6.58 (d, *J* = 8.2 Hz, 1H), 5.25 (dd, *J* = 8.9, 3.7 Hz, 1H), 4.33 (s, 1H), 2.72 – 2.65 (m, 1H), 2.48 (ddd, *J* = 12.6, 8.1, 4.5 Hz, 1H), 2.18 (s, 3H), 2.17 – 2.11 (m, 1H), 2.08 – 2.00 (m, 4H).



***N*-Benzhydryl-3-(10,11-dihydro-5*H*-dibenzo[*a,d*][7]annulen-5-ylidene)-*N*-methylpropan-1-amine (**1n**)**

N-Benzhydryl-3-(10,11-dihydro-5*H*-dibenzo[*a,d*][7]annulen-5-ylidene)-*N*-methylpropan-1-amine was prepared following **General Procedure for Preparation of Tertiary Amines** using 3-(10,11-dihydro-5*H*-dibenzo[*a,d*][7]annulen-5-ylidene)-*N*-methylpropan-1-amine (4.3 g, 16 mmol), (bromomethylene)dibenzene (1.2 equiv.) and K₂CO₃ (2.0 equiv.). The amine product **1n** was obtained after purification by flash silica gel column chromatography (Et₂O:hexanes = 1:19) as a colorless oil (6.5 g, 91%).

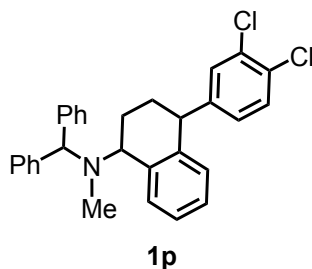
¹H NMR (600 MHz, CDCl₃) δ 7.36 (d, *J* = 7.5 Hz, 5H), 7.23 (s, 4H), 7.19 (d, *J* = 1.0 Hz, 2H), 7.17 – 7.10 (m, 5H), 7.07 (d, *J* = 7.4 Hz, 1H), 7.03 (d, *J* = 4.1 Hz, 1H), 5.82 (t, *J* = 7.5 Hz, 1H), 4.31 (s, 1H), 3.30 (s, 2H), 2.94 (s, 1H), 2.72 (s, 1H), 2.46 (s, 2H), 2.31 (d, *J* = 7.4 Hz, 2H), 2.07 (s, 3H).



(*R*)-*N*-Benzhydryl-*N*-methyl-3-(naphthalen-1-yloxy)-3-(thiophen-2-yl)propan-1-amine (1o)

(*R*)-*N*-Benzhydryl-*N*-methyl-3-(naphthalen-1-yloxy)-3-(thiophen-2-yl)propan-1-amine was prepared following **General Procedure for Preparation of Tertiary Amines** using (*R*)-*N*-methyl-3-(naphthalen-1-yloxy)-3-(thiophen-2-yl)propan-1-amine (0.9 g, 2.9 mmol), (bromomethylene)dibenzene (1.2 equiv.) and K₂CO₃ (2.0 equiv.). The amine product **1o** was obtained after purification by flash silica gel column chromatography (Et₂O:hexanes = 1:4) as a colorless oil (1.2 g, 90%).

¹H NMR (500 MHz, CDCl₃) δ 8.09 (s, 1H), 7.76 (d, *J* = 7.9 Hz, 1H), 7.51 – 7.42 (m, 1H), 7.42 – 7.30 (m, 4H), 7.30 – 7.09 (m, 7H), 7.04 – 6.94 (m, 4H), 6.94 – 6.88 (m, 1H), 6.88 – 6.81 (m, 1H), 5.75 (s, 1H), 4.35 (s, 1H), 2.76 – 2.65 (m, 1H), 2.61 – 2.51 (m, 1H), 2.51 – 2.38 (m, 1H), 2.32 – 2.16 (m, 4H).

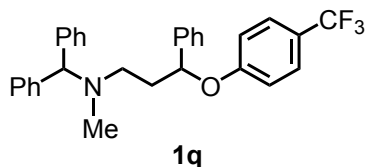


***N*-Benzhydryl-4-(3,4-dichlorophenyl)-*N*-methyl-1,2,3,4-tetrahydronaphthalen-1-amine (1p)**

N-Benzhydryl-4-(3,4-dichlorophenyl)-*N*-methyl-1,2,3,4-tetrahydronaphthalen-1-amine was prepared following **General Procedure for Preparation of Tertiary Amines** using 4-(3,4-dichlorophenyl)-*N*-methyl-1,2,3,4-tetrahydronaphthalen-1-amine (2.9 g, 6.0 mmol), (bromomethylene)dibenzene (1.2 equiv.) and K₂CO₃ (2.0 equiv.). The amine product **1p** was obtained after purification by flash silica gel column chromatography (Et₂O:hexanes = 1:4) as a colorless oil (1.4 g, 49%).

¹H NMR (600 MHz, CDCl₃) δ 8.19 – 8.13 (m, 1H), 7.56 (td, *J* = 7.1, 6.3, 1.5 Hz, 4H), 7.38 – 7.33 (m, 1H), 7.29 (ddt, *J* = 7.9, 4.3, 2.0 Hz, 3H), 7.26 – 7.23 (m, 2H), 7.16 (dtd, *J* = 17.5, 7.2, 1.4 Hz, 3H), 7.08 (d, *J* = 1.9 Hz, 1H), 6.87 (d, *J* = 7.6 Hz, 1H), 6.79 (dt, *J* = 8.2, 1.9 Hz, 1H),

4.73 (d, $J = 1.6$ Hz, 1H), 4.08 – 3.99 (m, 2H), 2.04 (d, $J = 1.7$ Hz, 3H), 1.91 (q, $J = 5.0, 4.5$ Hz, 2H), 1.76 – 1.65 (m, 1H), 1.65 – 1.58 (m, 1H).

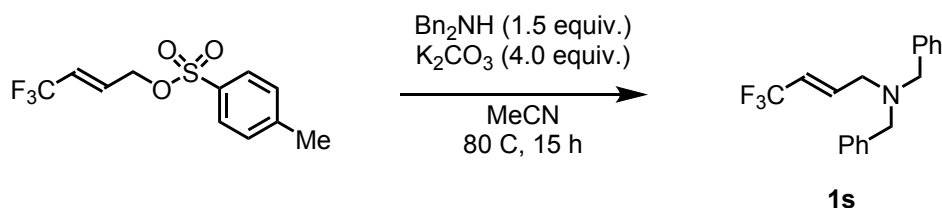


***N*-Benzhydryl-*N*-methyl-3-phenyl-3-(4-(trifluoromethyl)phenoxy)propan-1-amine (1q)**

N-Benzhydryl-*N*-methyl-3-phenyl-3-(4-(trifluoromethyl)phenoxy)propan-1-amine was prepared following **General Procedure for Preparation of Tertiary Amines** using *N*-methyl-3-phenyl-3-(4-(trifluoromethyl)phenoxy)propan-1-amine (2.8 g, 9.0 mmol), (bromomethylene)dibenzene (1.2 equiv.) and K_2CO_3 (2.0 equiv.). The amine product **1q** was obtained after purification by flash silica gel column chromatography (Et₂O:hexanes = 1:9) as a colorless oil (3.0 g, 70%).

¹H NMR (500 MHz, CDCl₃) δ 7.43 (d, $J = 8.4$ Hz, 2H), 7.38 – 7.33 (m, 2H), 7.29 (dtd, $J = 7.3, 5.7, 4.7, 1.6$ Hz, 6H), 7.23 (td, $J = 7.5, 1.7$ Hz, 3H), 7.19 – 7.14 (m, 1H), 7.12 – 7.07 (m, 3H), 6.84 – 6.79 (m, 2H), 5.34 – 5.29 (m, 1H), 4.35 (s, 1H), 2.71 (dt, $J = 13.9, 7.5$ Hz, 1H), 2.39 (dt, $J = 12.2, 5.9$ Hz, 1H), 2.18 (d, $J = 1.6$ Hz, 3H), 2.13 (dd, $J = 13.8, 7.2$ Hz, 1H), 2.08 – 1.99 (m, 1H); ¹⁹F NMR (564 MHz, CDCl₃) δ -61.51.

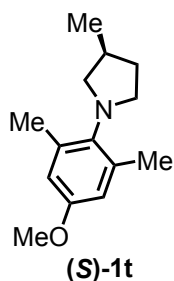
Procedure for Preparation of (*E*)-*N,N*-Dibenzyl-4,4,4-trifluorobut-2-en-1-amine (1s)



(*E*)-*N,N*-Dibenzyl-4,4,4-trifluorobut-2-en-1-amine (1s)

(*E*)-*N,N*-Dibenzyl-4,4,4-trifluorobut-2-en-1-amine was prepared following the literature previously reported.² To a solution of (*E*)-4,4,4-trifluorobut-2-en-1-yl 4-methylbenzenesulfonate (7.0 g, 25 mmol) and K_2CO_3 (13.8 g, 100 mmol) in MeCN (100 mL) was added dibenzylamine (7.4 g, 37.5 mmol). The reaction mixture was then allowed to stir at 80 °C for 15 hours. Upon completion (determined by TLC), the reaction mixture was filtered and concentrated *in vacuo*. The amine product **1s** was obtained after purification by flash silica gel column chromatography (EtOAc:hexanes = 1:20) as a colorless oil (5.9 g, 77%).

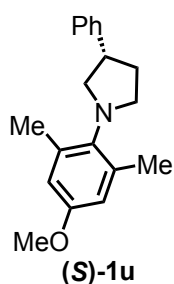
¹H NMR (500 MHz, CDCl₃) δ 7.39 – 7.29 (m, 8H), 7.28 – 7.22 (m, 2H), 6.51 – 6.30 (m, 1H), 5.85 (ddt, *J* = 15.8, 6.5, 1.7 Hz, 1H), 3.59 (s, 4H), 3.26 – 3.03 (m, 2H); **¹⁹F NMR** (470 MHz, CDCl₃) δ -63.97 (d, *J* = 6.1 Hz).



(S)-1-(4-Methoxy-2,6-dimethylphenyl)-3-methylpyrrolidine ((S)-1t)

(S)-1-(4-Methoxy-2,6-dimethylphenyl)-3-methylpyrrolidine was prepared following **General Procedure for Preparation of Tertiary Amines** using 4-methoxy-2,6-dimethylaniline (1.65 g, 10.9 mmol), (S)-1,4-dibromo-2-methylbutane (0.9 equiv.) and K₂CO₃ (2.0 equiv.). The amine product **(S)-1t** was obtained after purification by flash silica gel column chromatography (Et₂O:hexanes = 1:19) as a colorless oil (1.8 g, 82%).

¹H NMR (600 MHz, CDCl₃) δ 6.57 (s, 2H), 3.75 (s, 3H), 3.26 – 3.21 (m, 1H), 3.21 – 3.17 (m, 2H), 2.79 (t, *J* = 7.5 Hz, 1H), 2.39 (dt, *J* = 8.5, 6.8 Hz, 1H), 2.23 (s, 6H), 2.12 – 2.02 (m, 1H), 1.59 (dd, *J* = 11.9, 8.1 Hz, 1H), 1.11 (d, *J* = 6.7 Hz, 3H); [α]_D²⁵ = -19.5° (*c* 0.25, CH₂Cl₂).



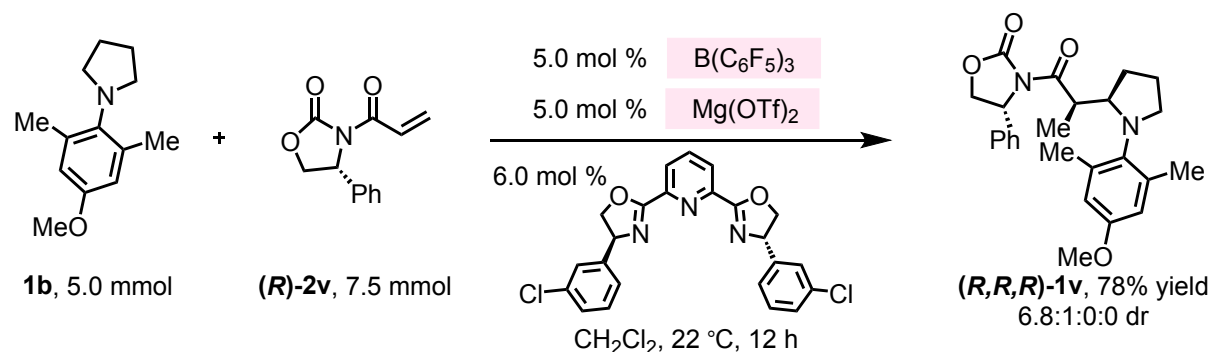
(S)-1-(4-Methoxy-2,6-dimethylphenyl)-3-phenylpyrrolidine ((S)-1u)

(S)-1-(4-Methoxy-2,6-dimethylphenyl)-3-phenylpyrrolidine was prepared following **General Procedure for Preparation of Tertiary Amines** using 4-methoxy-2,6-dimethylaniline (382 mg, 2.5 mmol), (S)-1,4-dibromobutan-2-ylbenzene (0.9 equiv.) and K₂CO₃ (2.0 equiv.). The amine product **(S)-1u** was obtained after purification by flash silica gel column chromatography (Et₂O:hexanes = 1:19) as a colorless oil (549 mg, 78%).

¹H NMR (400 MHz, CDCl₃) δ 7.45 – 7.29 (m, 4H), 7.24 – 7.17 (m, 1H), 6.60 (s, 2H), 3.76 (s, 3H), 3.63 – 3.45 (m, 2H), 3.40 (t, *J* = 8.0 Hz, 1H), 3.33 (dd, *J* = 13.9, 7.2 Hz, 2H), 2.49 – 2.33

(m, 1H), 2.30 (s, 6H), 2.24 – 2.06 (m, 1H); $[\alpha]_D^{25} = -20.5^\circ$ (*c* 0.25, CH₂Cl₂).

Procedure for Preparation of (*R*)-3-((*R*)-2-((*R*)-1-(4-Methoxy-2,6-dimethylphenyl)pyrrolidin-2-yl)propanoyl)-4-phenyloxazolidin-2-one ((*R,R,R*)-1v)

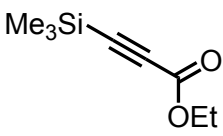
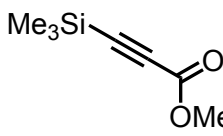
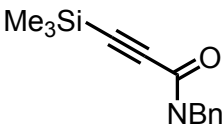
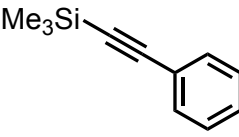
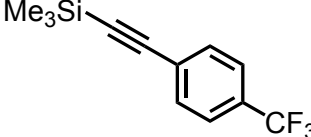
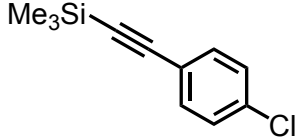
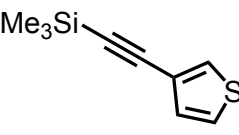
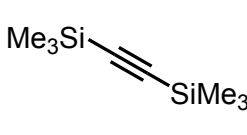
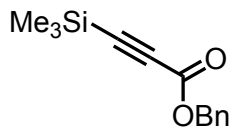


(*R*)-3-((*R*)-2-((*R*)-1-(4-Methoxy-2,6-dimethylphenyl)pyrrolidin-2-yl)propanoyl)-4-phenyloxazolidin-2-one ((*R,R,R*)-1v)

(*R*)-3-((*R*)-2-((*R*)-1-(4-Methoxy-2,6-dimethylphenyl)pyrrolidin-2-yl)propanoyl)-4-phenyloxazolidin-2-one was prepared according to a literature procedure.¹ To a 35 mL oven-dried sealed tube was added Mg(OTf)₂ (0.25 mmol), 2,6-bis((*S*)-4-(3-chlorophenyl)-4,5-dihydrooxazol-2-yl)pyridine (0.30 mmol), CH₂Cl₂ (10 mL) under nitrogen atmosphere. The mixture was allowed to stir for 30 min at 22 °C, then (*R*)-3-acryloyl-4-phenyloxazolidin-2-one (*R*)-2v (1.3 g, 6.0 mmol), 1-(4-methoxy-2,6-dimethylphenyl)pyrrolidine **1b** (1.02 g, 5.0 mmol), B(C₆F₅)₃ (0.25 mmol), and CH₂Cl₂ (5.0 mL) were added to the vessel. The reaction mixture was stirred at 22 °C for 48 h. Upon completion, the solvent was removed *in vacuo*. The diastereomeric ratio was determined to be 6.8:1:0:0 by ¹H NMR analysis of the unpurified product mixture. Purification by silica gel column chromatography (Et₂O:hexanes = 1:4) gave the product as a colorless solid as a mixture of diastereomers (1.63 g, 78% yield). Further purification was carried out by silica gel column chromatography (Et₂O:hexanes = 1:4) to obtain the major diastereomer (*R,R,R*)-1v in 1.47 g as a colorless oil.

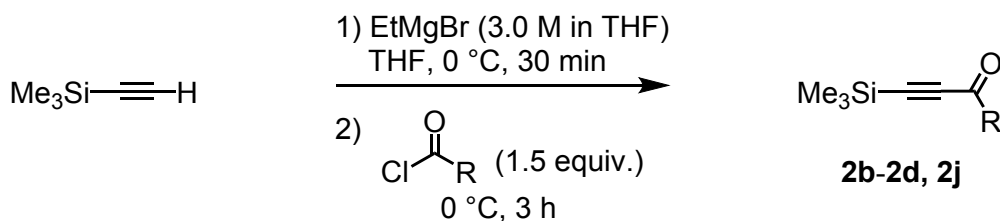
2.1.2. Preparation of Trimethylsilyl Propiolate Substrates

Table S2. List of Trimethylsilyl Propiolate Compounds

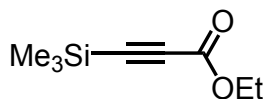
		
2b	2c	2d
		
2e	2f	2g
		
2h	2i	2j

Alkynes **2b-2d** and **2j** were prepared according to a literature procedure.⁴ Alkyne compounds **2e-2i** were obtained from commercial sources and used without further purification.

General Procedure for Preparation of 3-(Trimethylsilyl)propiolates



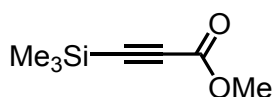
3-(Trimethylsilyl)propiolates **2b-2d**, **2j** were prepared according to a literature procedure.⁴ To a solution of ethynyltrimethylsilane (20 mmol) in THF (20 mL) was added ethylmagnesium bromide (3.0 M solution in THF) in a dropwise manner at 0 °C. The reaction mixture was allowed to stir for 30 min. The corresponding chloroformate (30 mmol) in THF (30 mL) was added dropwise and the reaction mixture was allowed to stir at 0 °C for 3 h. Upon completion (determined by TLC), H₂O (50 mL) was added and the organic material was extracted using Et₂O (3 x 20 mL). The combined organic layers were dried over MgSO₄, filtered, and concentrated *in vacuo*. The desired alkyne products were obtained after purification by flash silica gel column chromatography.



2b

Ethyl 3-(trimethylsilyl)propiolate (**2b**) was prepared according to **General Procedure for Preparation of 3-(Trimethylsilyl)propiolates** using ethyl chloroformate. The propiolate **2b** was obtained after purification by flash silica gel column chromatography (Et₂O:hexanes = 1:99) as a colorless oil (2.7 g, 80%).

¹H NMR (400 MHz, CDCl₃) δ 4.23 (q, *J* = 7.2 Hz, 2H), 1.31 (td, *J* = 7.1, 0.9 Hz, 3H), 0.25 (s, 9H).

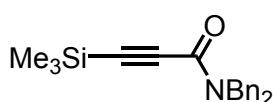


2c

Methyl 3-(trimethylsilyl)propiolate (2c)

Methyl 3-(trimethylsilyl)propiolate was prepared according to **General Procedure for Preparation of 3-(Trimethylsilyl)propiolates** using methyl chloroformate. The propiolate **2c** was obtained after purification by flash silica gel column chromatography (Et₂O:hexanes = 1:99) as a colorless oil (2.6 g, 83%).

¹H NMR (600 MHz, CDCl₃) δ 3.77 (s, 3H), 0.25 (s, 9H).

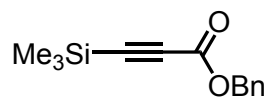


2d

***N,N*-Dibenzyl-3-(trimethylsilyl)propiolamide (2d)**

N,N-Dibenzyl-3-(trimethylsilyl)propiolamide was prepared according to **General Procedure for Preparation of 3-(Trimethylsilyl)propiolates** using *N,N*-dibenzylcarbonyl chloride. The propiolate **2d** was obtained after purification by flash silica gel column chromatography (Et₂O:hexanes = 1:99) as a colorless oil (3.3 g, 51%).

¹H NMR (500 MHz, CDCl₃) δ 7.41 – 7.19 (m, 10H), 4.67 (s, 2H), 4.50 (s, 2H), 0.21 (s, 9H).



2j

Benzyl 3-(trimethylsilyl)propiolate (2j)

Benzyl 3-(trimethylsilyl)propiolate was prepared according to **General Procedure for Preparation of 3-(Trimethylsilyl)propiolates** using benzyl chloroformate. The propiolate **2j** was obtained after purification by flash silica gel column chromatography (Et₂O:hexanes = 1:99) as a colorless oil (3.9 g, 84%).

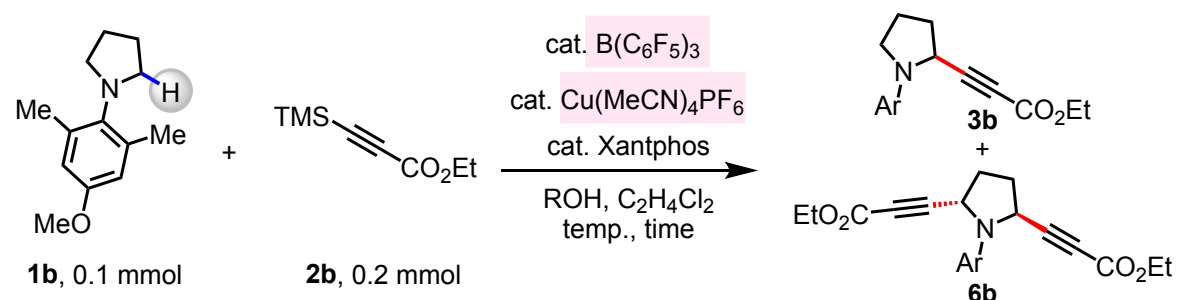
¹H NMR (500 MHz, CDCl₃) δ 6.81 (s, 2H), 5.11 (hept, *J* = 6.2 Hz, 2H), 1.29 (d, *J* = 6.3 Hz, 12H).

2.2 Optimization Studies for α -Alkynylation of *N*-Alkylamines Catalyzed by $B(C_6F_5)_3$ and Organocopper Catalysts

Experimental Procedure for Evaluation of Reaction Conditions (see Table S3)

An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added $Cu(MeCN)_4PF_6$ (0.01 mmol), Xantphos (0.01 mmol), and $C_2H_4Cl_2$ (0.2 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then ethyl 3-(trimethylsilyl)propiolate **2b** (0.2 mmol), alcohol (0.2 mmol), amine **1b** (0.1 mmol), $B(C_6F_5)_3$ (0.01 mmol, 10 mol%), and $C_2H_4Cl_2$ (0.2 mL) were added to the vessel. The reaction mixture was allowed to stir at 80 or 60 °C for 24 or 12 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH_2Cl_2 . The combined organic material was then concentrated *in vacuo*. Yield values were determined by 1H NMR analysis of unpurified reaction mixtures using mesitylene as the internal standard.

Table S3. Evaluation of Alcohol Additive and Reaction Conditions



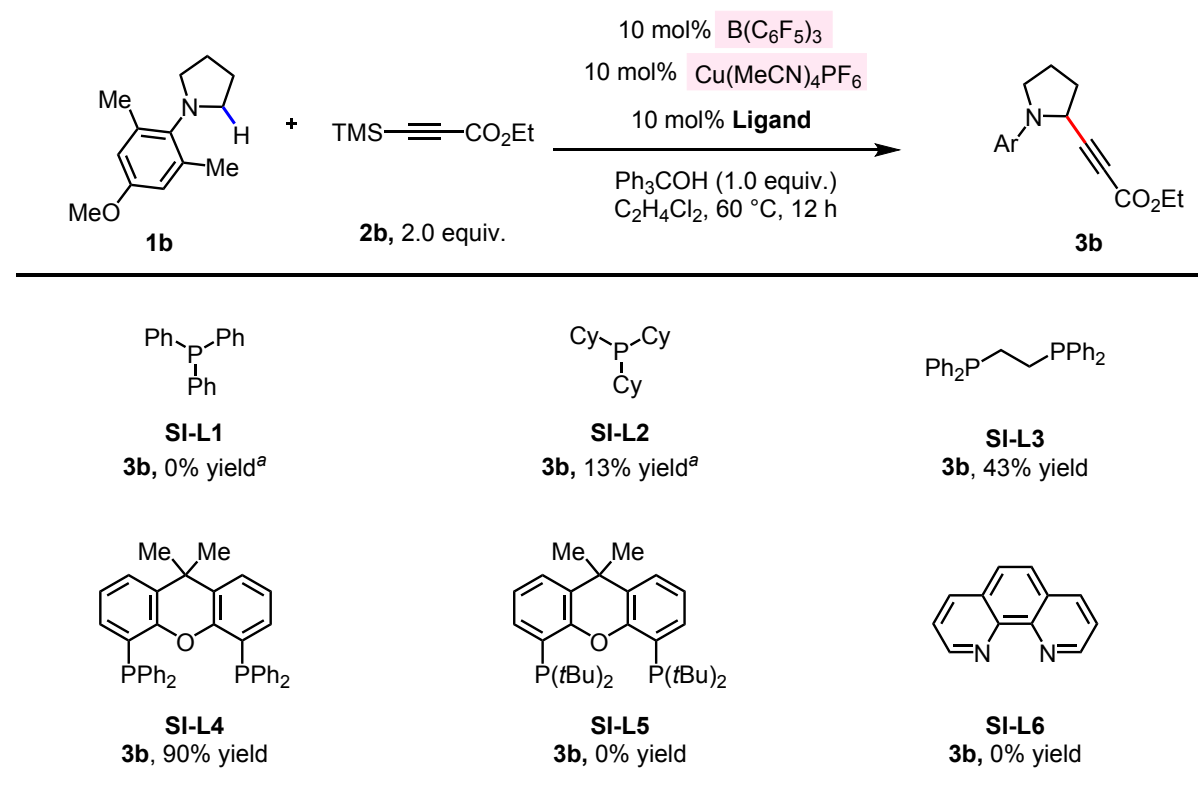
entry	alcohol (equiv.)	Temp. °C	time (h)	yield (%) 3b	yield (%) 6b
1	none	80	24	15	0
2	<i>i</i> -PrOH (2.0)	80	24	26	0
3	<i>t</i> -BuOH (2.0)	80	24	64	5
4	BnOH (2.0)	80	24	34	0
5	Ph_3COH (2.0)	80	24	52	34
6	1-adamantol (2.0)	80	24	93	<5
7	1-adamantol (2.0)	60	24	6	0
8	Ph_3COH (2.0)	60	24	79	19
9	Ph_3COH (1.0)	60	24	83	15
10	Ph_3COH (1.0)	60	12	90	<5

Conditions: *N*-arylpyrrolidine (**1b**, 0.1 mmol), 3-(trimethylsilyl)propiolate (**2b**, 0.2 mmol), $B(C_6F_5)_3$ (10 mol%), $Cu(MeCN)_4PF_6$ (10 mol%), Xantphos (10 mol%), alcohol (0.2 or 0.1 mmol), $C_2H_4Cl_2$ (0.4 mL), under N_2 . Yield values were determined by 1H NMR analysis of unpurified reaction mixtures using mesitylene as the internal standard.

Experimental Procedure for Evaluation of Ligand for 1-(4-Methoxy-2,6-dimethylphenyl)pyrrolidine **1b** (see Table S4)

An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added $\text{Cu}(\text{MeCN})_4\text{PF}_6$ (0.01 mmol), ligand (0.01 or 0.02 mmol), and $\text{C}_2\text{H}_4\text{Cl}_2$ (0.2 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then ethyl 3-(trimethylsilyl)propiolate **2b** (0.2 mmol), triphenylmethanol (0.1 mmol), amine **1b** (0.1 mmol), $\text{B}(\text{C}_6\text{F}_5)_3$ (0.01 mmol, 10 mol%), and $\text{C}_2\text{H}_4\text{Cl}_2$ (0.2 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 12 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH_2Cl_2 . The combined organic material was then concentrated *in vacuo*. Yield values were determined by ^1H NMR analysis of unpurified reaction mixtures using mesitylene as the internal standard.

Table S4. Evaluation of Ligand for the Reaction of 1-(4-Methoxy-2,6-dimethylphenyl)pyrrolidine **1b** and **2b**



Conditions: *N*-arylpyrrolidine (**1b**, 0.1 mmol), 3-(trimethylsilyl)propiolate (**2b**, 0.2 mmol), $\text{B}(\text{C}_6\text{F}_5)_3$ (10 mol%), $\text{Cu}(\text{MeCN})_4\text{PF}_6$ (10 mol%), ligand (10 mol%), triphenylmethanol (0.1 mmol), $\text{C}_2\text{H}_4\text{Cl}_2$ (0.4 mL), under N_2 , 60 °C, 12 h. Yield values were determined by ^1H NMR analysis of unpurified reaction mixtures using mesitylene as the internal standard. ^a20 mol% ligand was used.

Experimental Procedure for Evaluation of Ligand for (*R*)-*N*-Benzhydryl-2-((*tert*-butyldimethylsilyl)oxy)-*N*-methyl-1-phenylethan-1-amine (**1j**) (see Table S5)

An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added Cu(MeCN)₄PF₆ (0.01 mmol), ligand (0.01 or 0.02 mmol), and C₂H₄Cl₂ (0.2 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then ethyl 3-(trimethylsilyl)propiolate **2b** (0.2 mmol), triphenylmethanol (0.1 mmol), amine **1j** (0.1 mmol), B(C₆F₅)₃ (0.01 mmol, 10 mol%), and C₂H₄Cl₂ (0.2 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 12 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH₂Cl₂. The combined organic material was then concentrated *in vacuo*. Yield values were determined by ¹H NMR analysis of unpurified reaction mixtures using mesitylene as the internal standard.

Table S5. Evaluation of Ligand for the Reaction of (*R*)-*N*-Benzhydryl-2-((*tert*-butyldimethylsilyl)oxy)-*N*-methyl-1-phenylethan-1-amine (**1j**) and **2b**

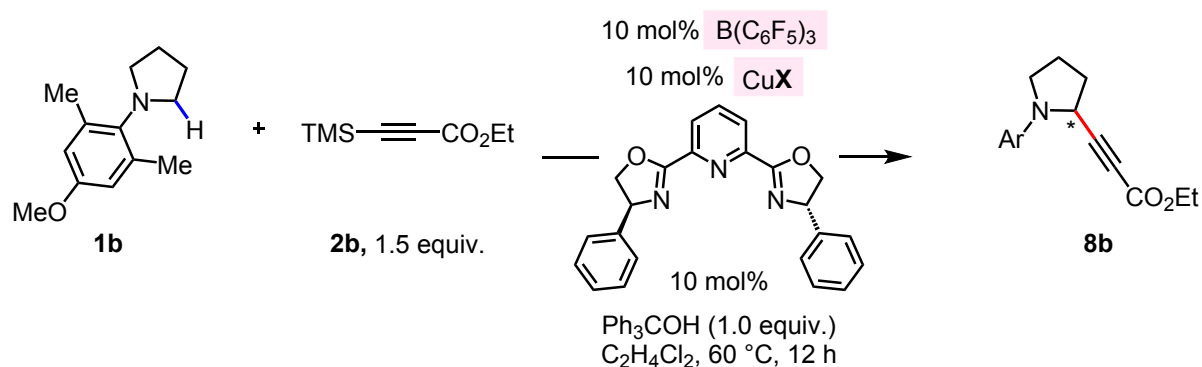
<p>SI-L1 3j, 45% yield^a</p>	<p>SI-L2 3j, 41% yield^a</p>	<p>SI-L3 3j, 88% yield</p>
<p>SI-L7 3j, 45% yield</p>	<p>SI-L4 3j, 45% yield</p>	<p>SI-L5 3j, 9% yield</p>
<p>L1 3j, 45% yield</p>	<p>SI-L8 3j, 38% yield</p>	<p>L3 3j, 64% yield</p>

Conditions: *N*-arypyrrolidine (**1b**, 0.1 mmol), 3-(trimethylsilyl)propiolate (**2b**, 0.2 mmol), B(C₆F₅)₃ (10 mol%), Cu(MeCN)₄PF₆ (10 mol%), ligand (10 mol%), triphenylmethanol (0.2 mmol), C₂H₄Cl₂ (0.4 mL), under N₂, 80 °C, 24 h. Yield values were determined by ¹H NMR analysis of unpurified reaction mixtures with mesitylene as the internal standard. ^a20 mol% ligand was used.

Experimental Procedure for Evaluation of Copper Salt (see Table S6)

An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added CuX (0.01 mmol), (*S*)-Ph-PyBOX (0.01 mmol), and C₂H₄Cl₂ (0.2 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then ethyl 3-(trimethylsilyl)propiolate **2b** (0.15 mmol), triphenylmethanol (0.1 mmol), amine **1b** (0.1 mmol), B(C₆F₅)₃ (0.01 mmol, 10 mol%), and C₂H₄Cl₂ (0.2 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 12 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH₂Cl₂. The combined organic material was then concentrated *in vacuo*. Yield values were determined by ¹H NMR analysis of unpurified reaction mixtures using mesitylene as the internal standard. Enantiomeric ratio (er) was determined by HPLC of the purified product.

Table S6. Evaluation of Copper Salt



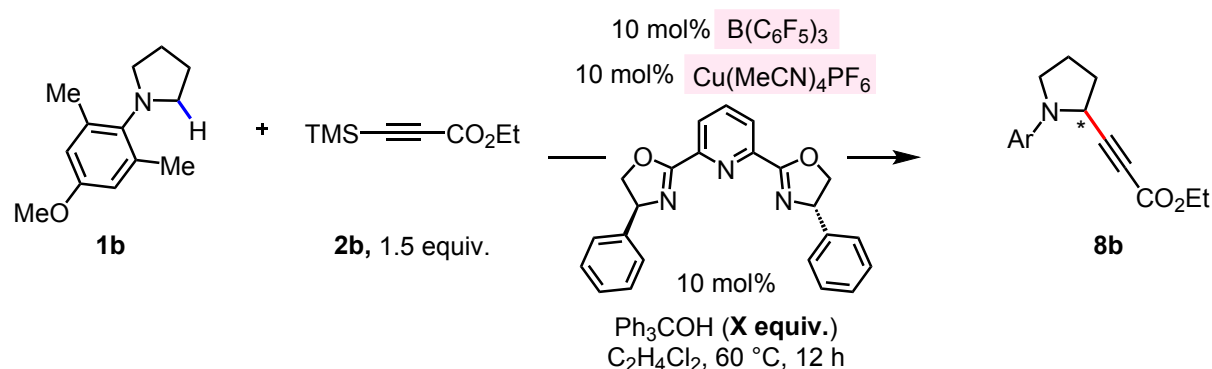
entry	CuX	Yield of 8b (%)	er of 8b
1	CuBr	0	nd
2	CuCl	<3	65:35
3	CuI	0	nd
4	CuOTf·Benzene	13	71:29
5	Cu(OTf) ₂	14	76:24
6	CuOAc	15	62:38
7	Cu(OAc) ₂	4	62:38
8	Cu(MeCN) ₄ PF ₆	69	81:19

Conditions: *N*-arylpyrrolidine (**1b**, 0.1 mmol), 3-(trimethylsilyl)propiolate (**2b**, 0.15 mmol), B(C₆F₅)₃ (10 mol%), CuX (10 mol%), (*S*)-PhPyBOX (10 mol%), triphenylmethanol (0.1 mmol), C₂H₄Cl₂ (0.4 mL), under N₂, 60 °C, 12 h. Yield values were determined by ¹H NMR analysis of unpurified reaction mixtures using mesitylene as the internal standard. Enantiomeric ratio (er) was determined by HPLC of the purified product.

Experimental Procedure for Optimization of Alcohol Additive for the Enantioselective Transformation (see Table S7)

An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added $\text{Cu}(\text{MeCN})_4\text{PF}_6$ (0.01 mmol), (*S*)-Ph-PyBOX (0.01 mmol), and $\text{C}_2\text{H}_4\text{Cl}_2$ (0.2 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then ethyl 3-(trimethylsilyl)propiolate **2b** (0.15 mmol), triphenylmethanol (0.1 or 0.2 mmol), amine **1b** (0.1 mmol), $\text{B}(\text{C}_6\text{F}_5)_3$ (0.01 mmol, 10 mol%), and $\text{C}_2\text{H}_4\text{Cl}_2$ (0.2 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 12 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH_2Cl_2 . The combined organic material was then concentrated *in vacuo*. Yield values were determined by ^1H NMR analysis of unpurified reaction mixtures using mesitylene as the internal standard. Enantiomeric ratio (er) was determined by HPLC of the purified product.

Table S7. Effect of Alcohol Additive for the Enantioselective Transformation



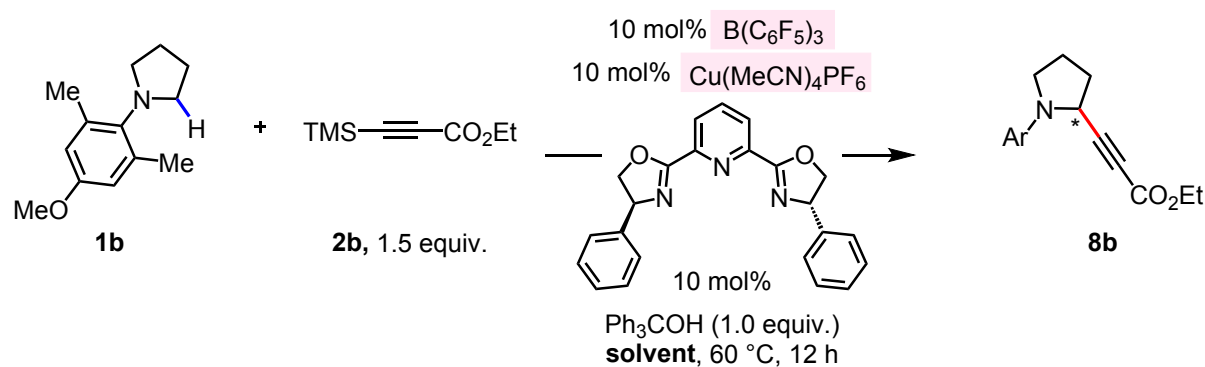
entry	Ph_3COH (X equiv.)	Yield of 8b (%)	er of 8b
1	1.0	69	81:19
2	2.0	98	77:23

Conditions: *N*-arylpyrrolidine (**1b**, 0.1 mmol), 3-(trimethylsilyl)propiolate (**2b**, 0.15 mmol), $\text{B}(\text{C}_6\text{F}_5)_3$ (10 mol%), $\text{Cu}(\text{MeCN})_4\text{PF}_6$ (10 mol%), (*S*)-PhPyBOX (10 mol%), triphenylmethanol (0.1 or 0.2 mmol), $\text{C}_2\text{H}_4\text{Cl}_2$ (0.4 mL), under N_2 , 60 °C, 12 h. Yield values were determined by ^1H NMR analysis of unpurified reaction mixtures with mesitylene as the internal standard. Enantiomeric ratio (er) was determined by HPLC of the purified product.

Experimental Procedure for Evaluation of Solvent (see Table S8)

An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added $\text{Cu}(\text{MeCN})_4\text{PF}_6$ (0.01 mmol), (*S*)-Ph-PyBOX (0.01 mmol), and solvent (0.2 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then ethyl 3-(trimethylsilyl)propiolate **2b** (0.15 mmol), triphenylmethanol (0.1 mmol), amine **1b** (0.1 mmol), $\text{B}(\text{C}_6\text{F}_5)_3$ (0.01 mmol, 10 mol%), and solvent (0.2 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 12 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH_2Cl_2 . The combined organic material was then concentrated *in vacuo*. Yield values were determined by ^1H NMR analysis of unpurified reaction mixtures using mesitylene as the internal standard. Enantiomeric ratio (er) was determined by HPLC of the purified product.

Table S8. Evaluation of Solvent



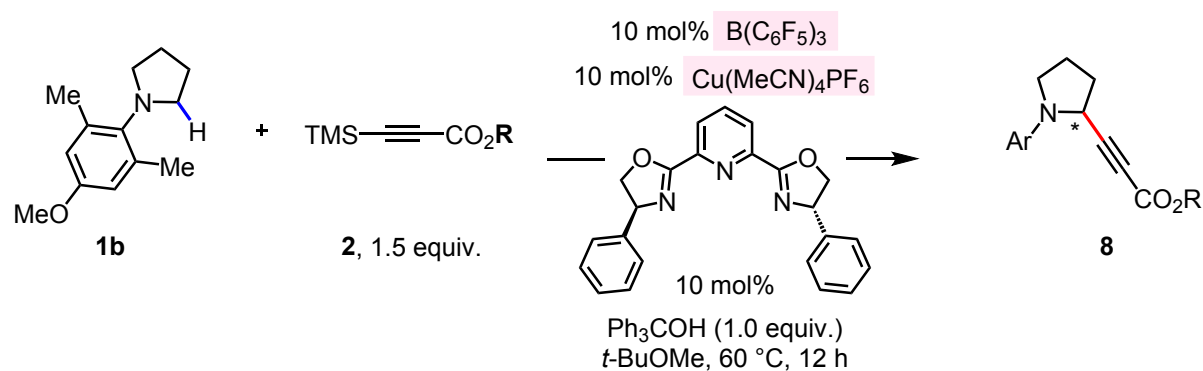
entry	solvent	Yield of 8b (%)	er of 8b
1	CHCl_3	48	59:14
2	CH_2Cl_2	62	78:22
3	$\text{C}_2\text{H}_4\text{Cl}_2$	69	81:19
4	Benzene	43	80:20
5	Toluene	54	71:29
6	THF	63	83:17
7	Et_2O	28	83:17
8	<i>t</i> -BuOMe	84	82:18

Conditions: *N*-arylpyrrolidine (**1b**, 0.1 mmol), 3-(trimethylsilyl)propiolate (**2b**, 0.15 mmol), $\text{B}(\text{C}_6\text{F}_5)_3$ (10 mol%), $\text{Cu}(\text{MeCN})_4\text{PF}_6$ (10 mol%), (*S*)-PhPyBOX (10 mol%), triphenylmethanol (0.1 mmol), solvent (0.4 mL), under N_2 , 60 °C, 12 h. Yield values were determined by ^1H NMR analysis of unpurified reaction mixtures with mesitylene as the internal standard. Enantiomeric ratio (er) was determined by HPLC of the purified product.

Experimental Procedure for Evaluation of Alkyne Substrate for the Stereoselective Transformation (see Table S9)

An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added $\text{Cu}(\text{MeCN})_4\text{PF}_6$ (0.01 mmol), (*S*)-Ph-PyBOX (0.01 mmol), and *t*-BuOMe (0.2 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then 3-(trimethylsilyl)propiolate **2** (0.15 mmol), triphenylmethanol (0.1 mmol), amine **1b** (0.1 mmol), $\text{B}(\text{C}_6\text{F}_5)_3$ (0.01 mmol, 10 mol%), and *t*-BuOMe (0.2 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 12 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH_2Cl_2 . The combined organic material was then concentrated *in vacuo*. Yield values were determined by ^1H NMR analysis of unpurified reaction mixtures using mesitylene as the internal standard. Enantiomeric ratio (er) was determined by HPLC of the purified product.

Table S9. Evaluation of Alkyne Substrates



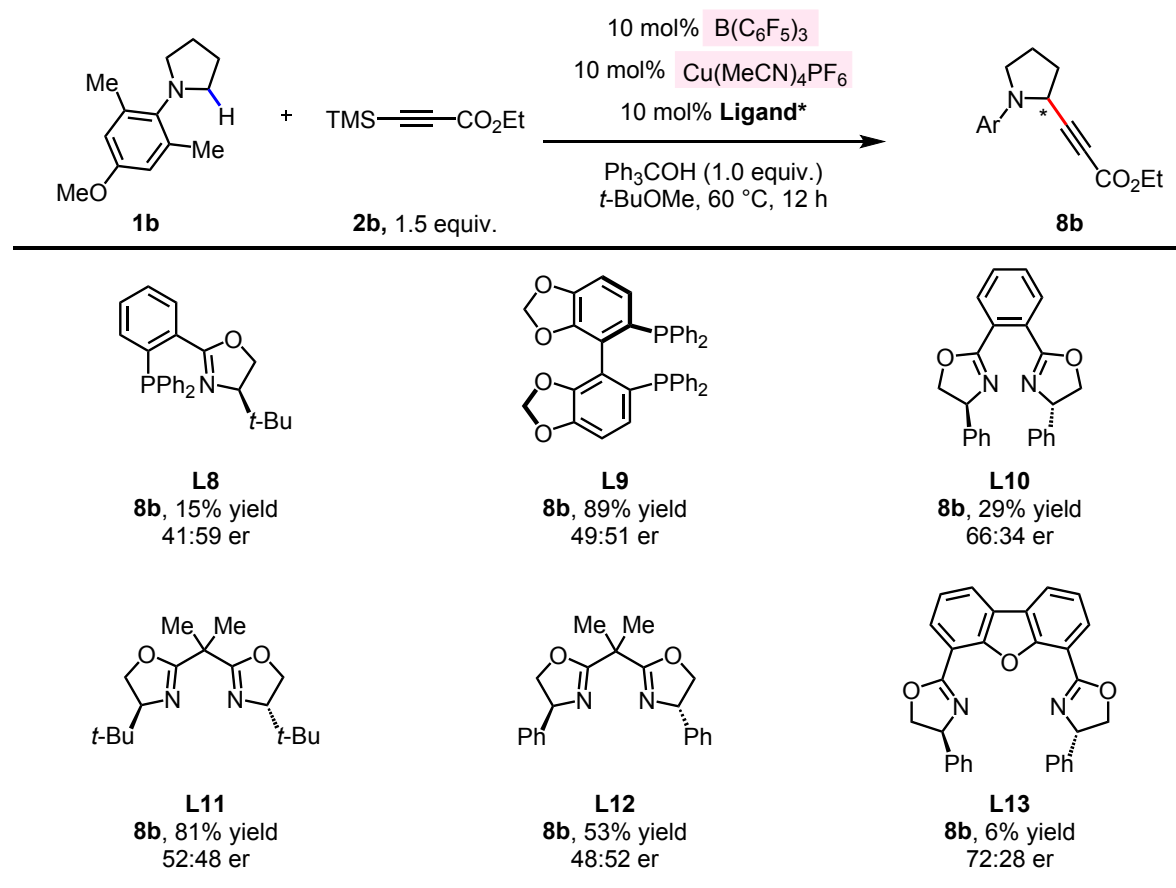
entry	CO_2R	Yield of 8 (%)	er of 8
1	Me	35	82:18
2	Et	84	82:18
3	<i>i</i> -Pr	0	nd
4	<i>i</i> -Bu	30	62:38
5	<i>t</i> -Bu	0	nd
6	Bn	59	64:36

Conditions: *N*-arylpyrrolidine (**1b**, 0.1 mmol), 3-(trimethylsilyl)propiolate (**2**, 0.15 mmol), $\text{B}(\text{C}_6\text{F}_5)_3$ (10 mol%), $\text{Cu}(\text{MeCN})_4\text{PF}_6$ (10 mol%), (*S*)-PhPyBOX (10 mol%), triphenylmethanol (0.1 mmol), *t*-BuOMe (0.4 mL), under N_2 , 60 °C, 12 h. Yield values were determined by ^1H NMR analysis of unpurified reaction mixtures with mesitylene as the internal standard. Enantiomeric ratio (er) was determined by HPLC of the purified product.

Experimental Procedure for Evaluation of Chiral Ligands (see Table S10)

An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added Cu(MeCN)₄PF₆ (0.01 mmol), ligand (0.01 mmol), and *t*-BuOMe (0.2 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then ethyl 3-(trimethylsilyl)propiolate **2b** (0.15 mmol), triphenylmethanol (0.1 mmol), amine **1b** (0.1 mmol), B(C₆F₅)₃ (0.01 mmol, 10 mol%), and *t*-BuOMe (0.2 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 12 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH₂Cl₂. The combined organic material was then concentrated *in vacuo*. Yield values were determined by ¹H NMR analysis of unpurified reaction mixtures using mesitylene as the internal standard. Enantiomeric ratio (er) was determined by HPLC of the purified product.

Table S10. Evaluation of Chiral Ligands

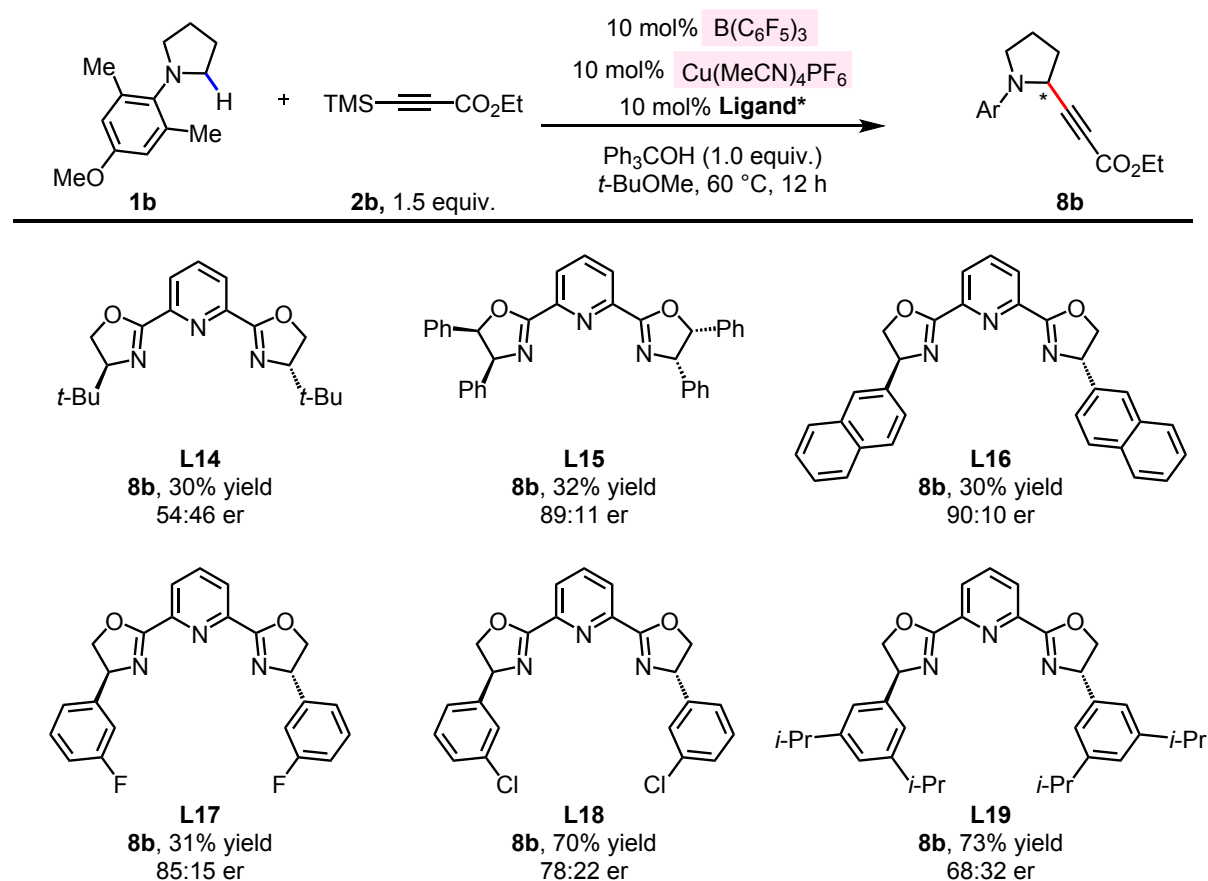


Conditions: *N*-arylpyrrolidine (**1b**, 0.1 mmol), 3-(trimethylsilyl)propiolate (**2b**, 0.15 mmol), B(C₆F₅)₃ (10 mol%), Cu(MeCN)₄PF₆ (10 mol%), Ligand (10 mol%), triphenylmethanol (0.1 mmol), *t*-BuOMe (0.4 mL), under N₂, 60 °C, 12 h. Yield values were determined by ¹H NMR analysis of unpurified reaction mixtures with mesitylene as the internal standard. Enantiomeric ratio (er) was determined by HPLC of the purified product.

Experimental Procedure for Evaluation of PyBOX Ligands (see Table S11)

An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added $\text{Cu}(\text{MeCN})_4\text{PF}_6$ (0.01 mmol), ligand (0.01 mmol), and *t*-BuOMe (0.2 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then ethyl 3-(trimethylsilyl)propiolate **2b** (0.15 mmol), triphenylmethanol (0.1 mmol), amine **1b** (0.1 mmol), $\text{B}(\text{C}_6\text{F}_5)_3$ (0.01 mmol, 10 mol%), and *t*-BuOMe (0.2 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 12 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH_2Cl_2 . The combined organic material was then concentrated *in vacuo*. Yield values were determined by ^1H NMR analysis of unpurified reaction mixtures using mesitylene as the internal standard. Enantiomeric ratio (er) was determined by HPLC of the purified product.

Table S11. Evaluation of PyBOX Ligands

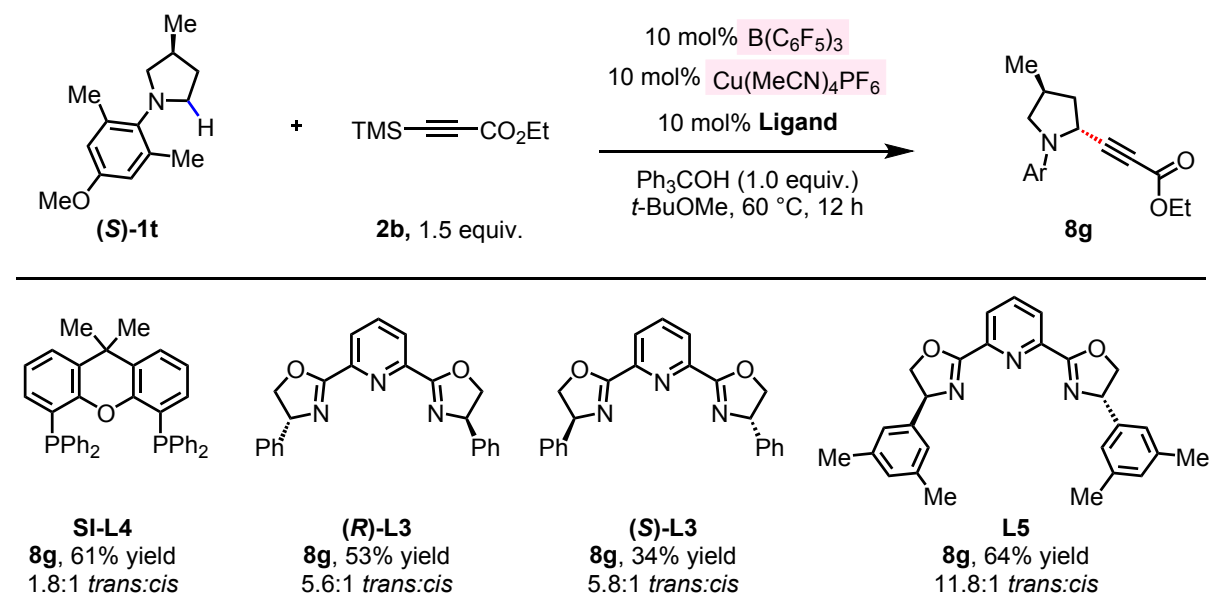


Conditions: *N*-arylpyrrolidine (**1b**, 0.1 mmol), 3-(trimethylsilyl)propiolate (**2b**, 0.15 mmol), $\text{B}(\text{C}_6\text{F}_5)_3$ (10 mol%), $\text{Cu}(\text{MeCN})_4\text{PF}_6$ (10 mol%), Ligand (10 mol%), triphenylmethanol (0.1 mmol), *t*-BuOMe (0.4 mL), under N_2 , 60 °C, 12 h. Yield values were determined by ^1H NMR analysis of unpurified reaction mixtures with mesitylene as the internal standard. Enantiomeric ratio (er) was determined by HPLC of the purified product.

Experimental Procedure for Evaluation of Ligands for Amine (**S**)-**1t** (see Table S12)

An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added Cu(MeCN)₄PF₆ (0.01 mmol), ligand (0.01 mmol), and *t*-BuOMe (0.2 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then 3-(trimethylsilyl)propiolate **2b** (0.15 mmol), triphenylmethanol (0.1 mmol), amine (**S**)-**1t** (0.1 mmol), B(C₆F₅)₃ (0.01 mmol, 10 mol%), and *t*-BuOMe (0.2 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 12 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH₂Cl₂. The combined organic material was then concentrated *in vacuo*. Yield values and *trans*:*cis* ratios were determined by ¹H NMR analysis of unpurified reaction mixtures using mesitylene as the internal standard.

Table S12. Evaluation of Ligands for Amine (**S**)-**1t**

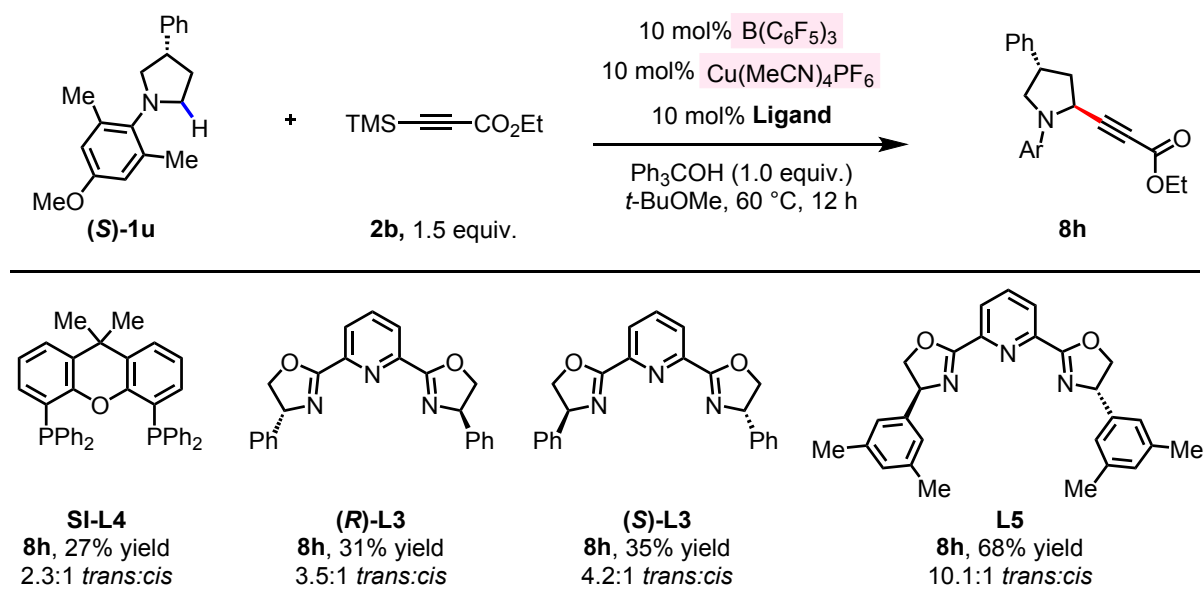


Conditions: *N*-arylpyrrolidine ((**S**)-**1t**, 0.1 mmol), 3-(trimethylsilyl)propiolate (**2b**, 0.15 mmol), B(C₆F₅)₃ (10 mol%), Cu(MeCN)₄PF₆ (10 mol%), ligand (10 mol%), triphenylmethanol (0.1 mmol), *t*-BuOMe (0.4 mL), under N₂, 60 °C, 12 h. Yield values and *trans*:*cis* ratio were determined by ¹H NMR analysis of the unpurified reaction mixtures with mesitylene as the internal standard.

Experimental Procedure for Evaluation of Ligands for the Reaction of Amine (**S**)-**1u** and **2b** (see Table S13)

An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added $\text{Cu}(\text{MeCN})_4\text{PF}_6$ (0.01 mmol), ligand (0.01 mmol), and *t*-BuOMe (0.2 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then 3-(trimethylsilyl)propiolate **2** (0.15 mmol), triphenylmethanol (0.1 mmol), amine (**S**)-**1u** (0.1 mmol), $\text{B}(\text{C}_6\text{F}_5)_3$ (0.01 mmol, 10 mol%), and *t*-BuOMe (0.2 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 12 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH_2Cl_2 . The combined organic material was then concentrated *in vacuo*. Yield values and *trans*:*cis* ratios were determined by ^1H NMR analysis of unpurified reaction mixtures using mesitylene as the internal standard.

Table S13. Evaluation of Ligands for Amine (**S**)-**1u**

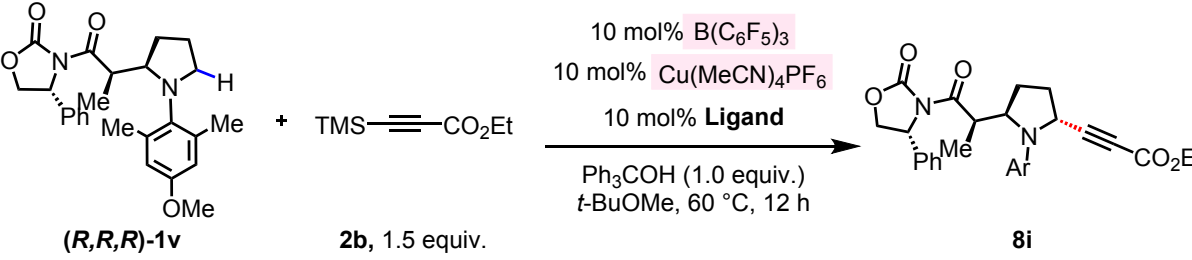
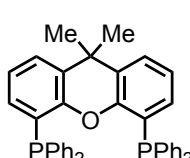
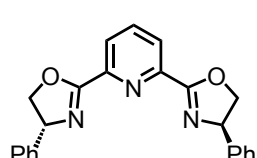
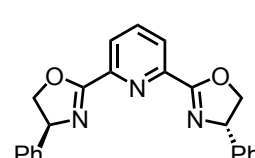
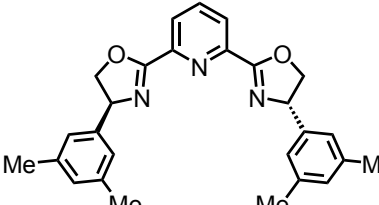


Conditions: *N*-arylpyrrolidine ((**S**)-**1u**, 0.1 mmol), 3-(trimethylsilyl)propiolate (**2b**, 0.15 mmol), $\text{B}(\text{C}_6\text{F}_5)_3$ (10 mol%), $\text{Cu}(\text{MeCN})_4\text{PF}_6$ (10 mol%), ligand (10 mol%), triphenylmethanol (0.1 mmol), *t*-BuOMe (0.4 mL), under N_2 , 60 °C, 12 h. Yield values and *trans*:*cis* ratio were determined by ^1H NMR analysis of the unpurified reaction mixtures with mesitylene as the internal standard.

Experimental Procedure for Evaluation of Ligands for the Reaction of Amine (*R,R,R*)-**1v** and **2b** (see Table S14)

An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added Cu(MeCN)₄PF₆ (0.01 mmol), ligand (0.01 mmol), and *t*-BuOMe (0.2 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then 3-(trimethylsilyl)propiolate **2b** (0.15 mmol), triphenylmethanol (0.1 mmol), amine (*R,R,R*)-**1v** (0.1 mmol), B(C₆F₅)₃ (0.01 mmol, 10 mol%), and *t*-BuOMe (0.2 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 12 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH₂Cl₂. The combined organic material was then concentrated *in vacuo*. Yield values and *trans*:*cis* ratios were determined by ¹H NMR analysis of unpurified reaction mixtures using mesitylene as the internal standard.

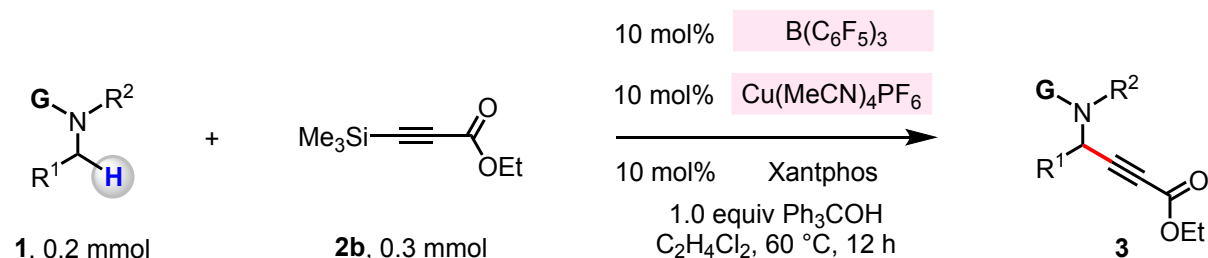
Table S14. Evaluation of Ligands for (*R,R,R*)-**1v**

 <p>(<i>R,R,R</i>)-1v + 2b, 1.5 equiv.</p> <p>10 mol% $\text{B}(\text{C}_6\text{F}_5)_3$ 10 mol% $\text{Cu}(\text{MeCN})_4\text{PF}_6$ 10 mol% Ligand Ph_3COH (1.0 equiv.) $t\text{-BuOMe}$, 60 °C, 12 h</p> <p>8i</p>			
 <p>SI-L4 8i, 54% yield 5.8:1 <i>trans</i>:<i>cis</i></p>	 <p>(R)-L3 8i, 81% yield 1.9:1 <i>trans</i>:<i>cis</i></p>	 <p>(S)-L3 8i, 49% yield 7.2:1 <i>trans</i>:<i>cis</i></p>	 <p>L5 8i, 93% yield 7.7:1 <i>trans</i>:<i>cis</i></p>

Conditions: *N*-arylpyrrolidine ((*R,R,R*)-**1v**, 0.1 mmol), 3-(trimethylsilyl)propiolate (**2**, 0.15 mmol), B(C₆F₅)₃ (10 mol%), Cu(MeCN)₄PF₆ (10 mol%), ligand (10 mol%), triphenylmethanol (0.1 mmol), *t*-BuOMe (0.4 mL), under N₂, 60 °C, 12 h. Yield values and *trans*:*cis* ratio were determined by ¹H NMR analysis of the unpurified reaction mixtures with mesitylene as the internal standard.

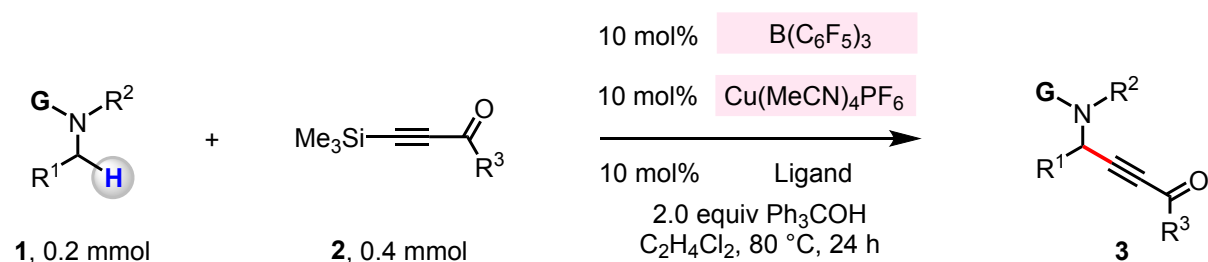
2.3 General Procedures for α -C–H Alkynylation of *N*-Alkylamines by $B(C_6F_5)_3$ and Organocopper Complex

General Procedure A for α -C–H Alkynylation of *N*-Alkylamines Catalyzed by $B(C_6F_5)_3$ and Organocopper Complex (See Table 1 and Figure 2 in the Manuscript)



An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added $Cu(MeCN)_4PF_6$ (0.02 mmol), Xantphos (0.02 mmol), and $C_2H_4Cl_2$ (0.4 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then ethyl 3-(trimethylsilyl)propiolate **2b** (0.3 mmol), triphenylmethanol (0.2 mmol), amine **1** (0.2 mmol), $B(C_6F_5)_3$ (0.02 mmol, 10 mol%), and $C_2H_4Cl_2$ (0.4 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 12 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH_2Cl_2 . The combined organic material was then concentrated *in vacuo*. The propargylamine product was purified and isolated by silica gel column chromatography.

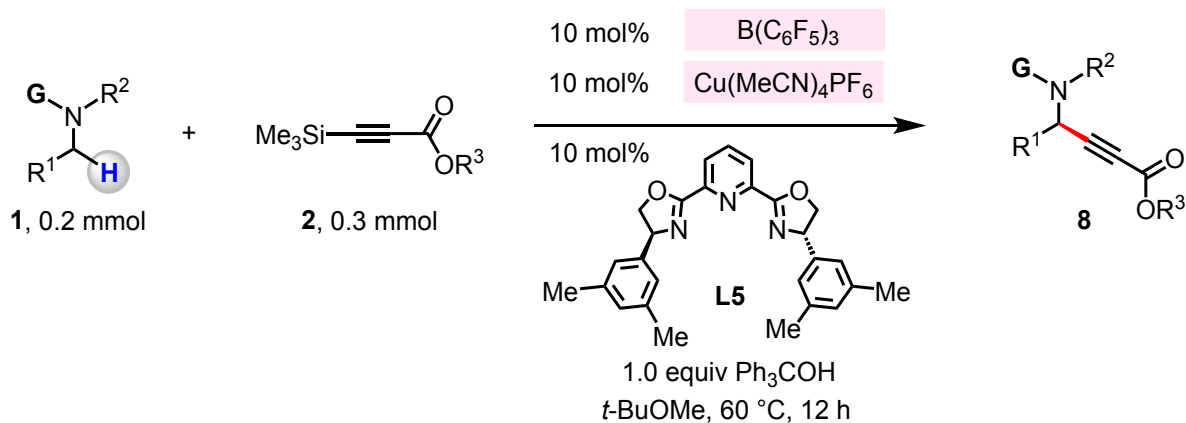
General Procedure B for α -C–H Alkynylation of *N*-Alkylamines Catalyzed by $B(C_6F_5)_3$ and Organocopper Complex (See Figures 2-3 in the Manuscript)



An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added $Cu(MeCN)_4PF_6$ (0.02 mmol), ligand (1,2-bis(diphenylphosphino)ethane or (*S*)-PhPyBOX, 0.02 mmol), and $C_2H_4Cl_2$ (0.4 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then (trimethylsilyl)propiolate **2** (0.4 mmol), triphenylmethanol (0.4 mmol), amine **1** (0.2 mmol), $B(C_6F_5)_3$ (0.02, 10 mol%), and $C_2H_4Cl_2$ (0.4 mL) were added to the vessel. The reaction mixture was allowed to stir at 80 °C for 24 h.

Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH₂Cl₂. The combined organic material was then concentrated *in vacuo*. The propargylamine product was purified and isolated by silica gel column chromatography.

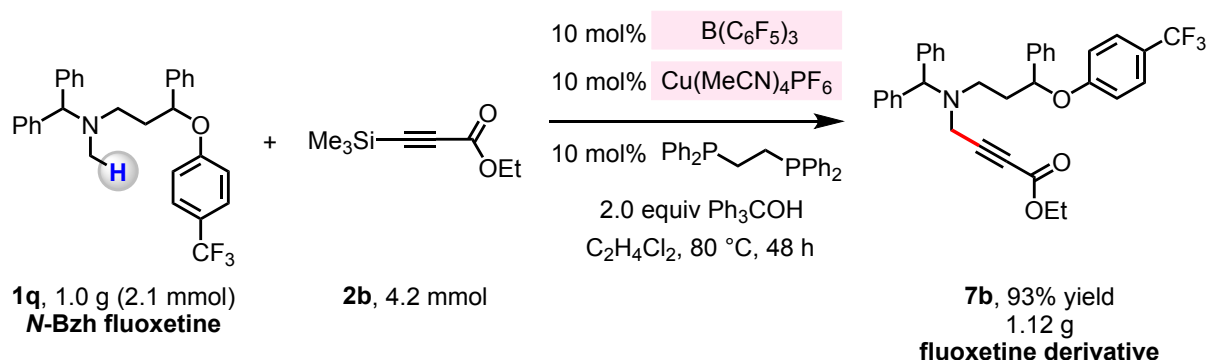
General Procedure C for Stereoselective α -C–H Alkynylation of *N*-Alkylamines Catalyzed by B(C₆F₅)₃ and Organocopper Complex (See Figure 5 in the Manuscript)



An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added Cu(MeCN)₄PF₆ (0.02 mmol), (*S*)-(3,5-dimethylphenyl)PyBOX **L5** (0.02 mmol), and *t*-BuOMe (0.4 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then (trimethylsilyl)propiolate **2** (0.3 mmol), triphenylmethanol (0.2 mmol), amine **1** (0.2 mmol), B(C₆F₅)₃ (0.02 mmol, 10 mol%), and solvent (0.4 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 12 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with Et₂O. The combined organic material was then concentrated *in vacuo*. The propargylamine product was purified isolated by silica gel column chromatography. The er values were determined by HPLC analysis of the isolated product.

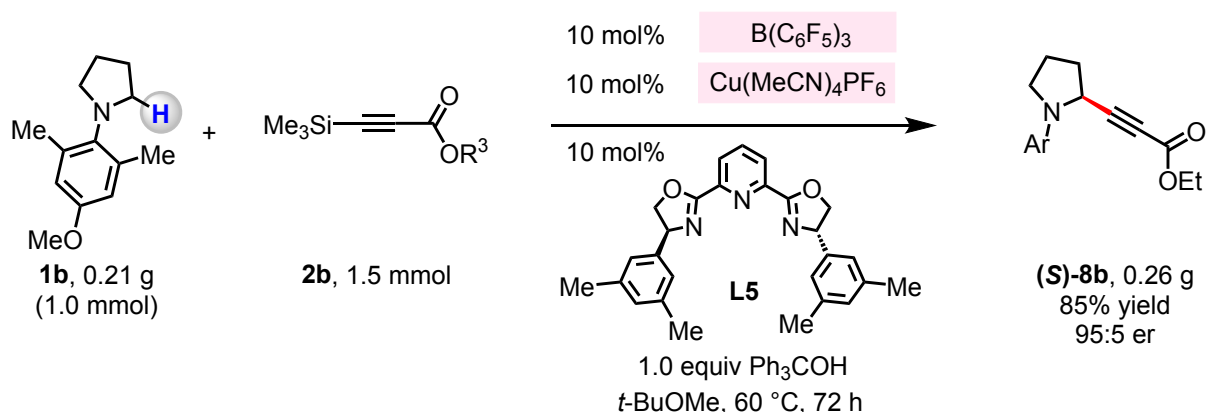
2.4 Procedures for Large Scale Reactions

Procedure for Gram-Scale Synthesis of Alkynylated Fluoxetine Derivative **7b**



An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added $\text{Cu}(\text{MeCN})_4\text{PF}_6$ (78 mg, 0.21 mmol), 1,2-bis(diphenylphosphino)ethane (84 mg, 0.21 mmol), and $\text{C}_2\text{H}_4\text{Cl}_2$ (4.0 mL) under nitrogen atmosphere. The mixture was allowed to stir for 30 minutes at 22 °C, then (trimethylsilyl)propiolate **2b** (715 mg, 4.2 mmol), triphenylmethanol (1.1 g, 4.2 mmol), amine **1q** (1.0 g, 2.1 mmol), $\text{B}(\text{C}_6\text{F}_5)_3$ (108 mg, 0.21 mmol), and $\text{C}_2\text{H}_4\text{Cl}_2$ (4.0 mL) were added to the vessel. The reaction mixture was allowed to stir at 80 °C for 48 h. Upon completion, the unpurified reaction mixture was concentrated *in vacuo*. The amine product **7b** was obtained after purification by flash silica gel column chromatography (Et_2O :hexanes = 1:19) as a colorless solid (1.12 g, 93%).

Procedure for Enantioselective α -Alkynylation Reaction in 1.0 mmol Scale

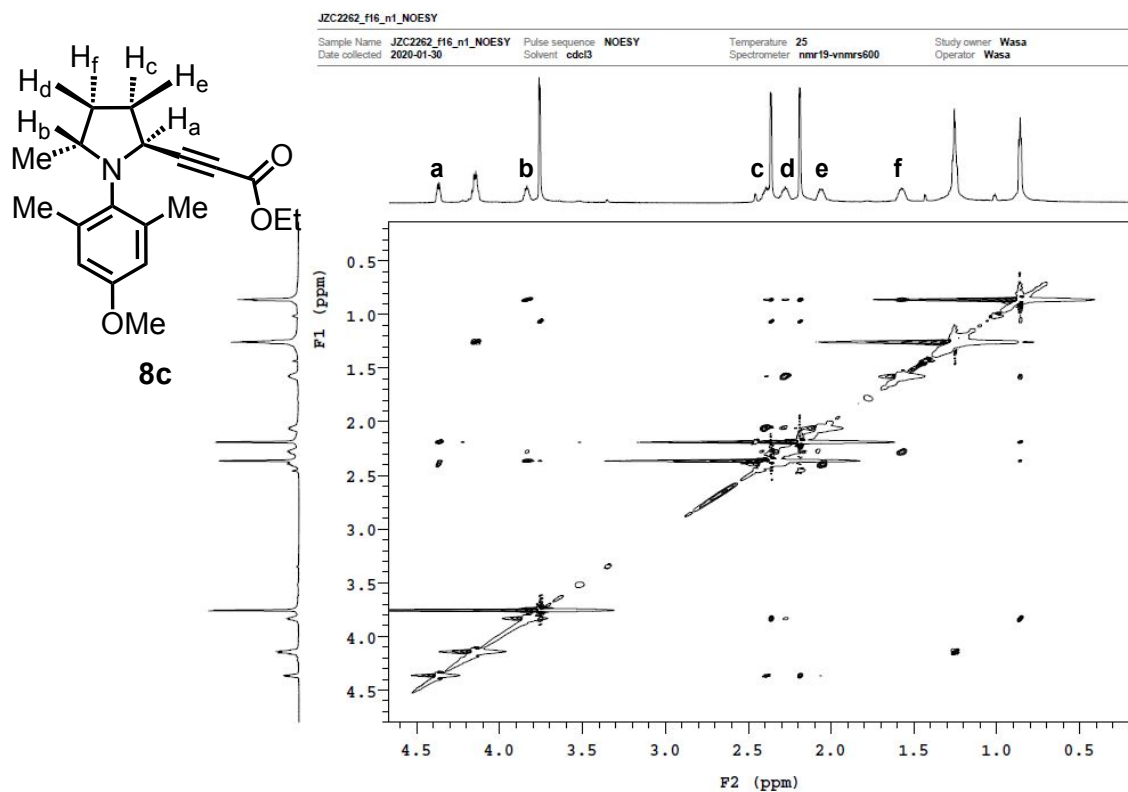


An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added $\text{Cu}(\text{MeCN})_4\text{PF}_6$ (19 mg, 0.05 mmol), (*S*)-(3,5-dimethylphenyl)PyBOX **L5** (23 mg, 0.05 mmol), and *t*-BuOMe (2.0 mL) under nitrogen atmosphere. The mixture was allowed to stir for 30 minutes at 22 °C, then (trimethylsilyl)propiolate **2b** (255 mg, 1.5 mmol), triphenylmethanol (260 mg, 1.0 mmol), amine **1b** (205 mg, 1.0 mmol), $\text{B}(\text{C}_6\text{F}_5)_3$ (26 mg, 0.05 mmol), and *t*-

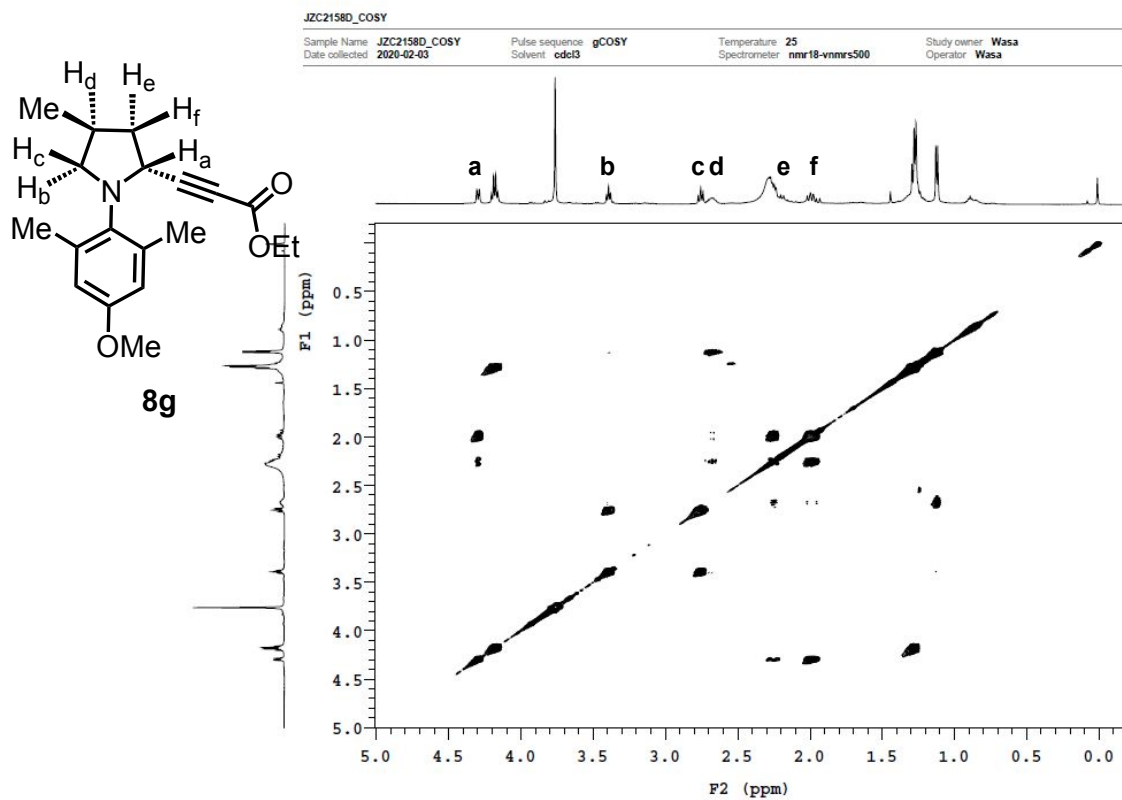
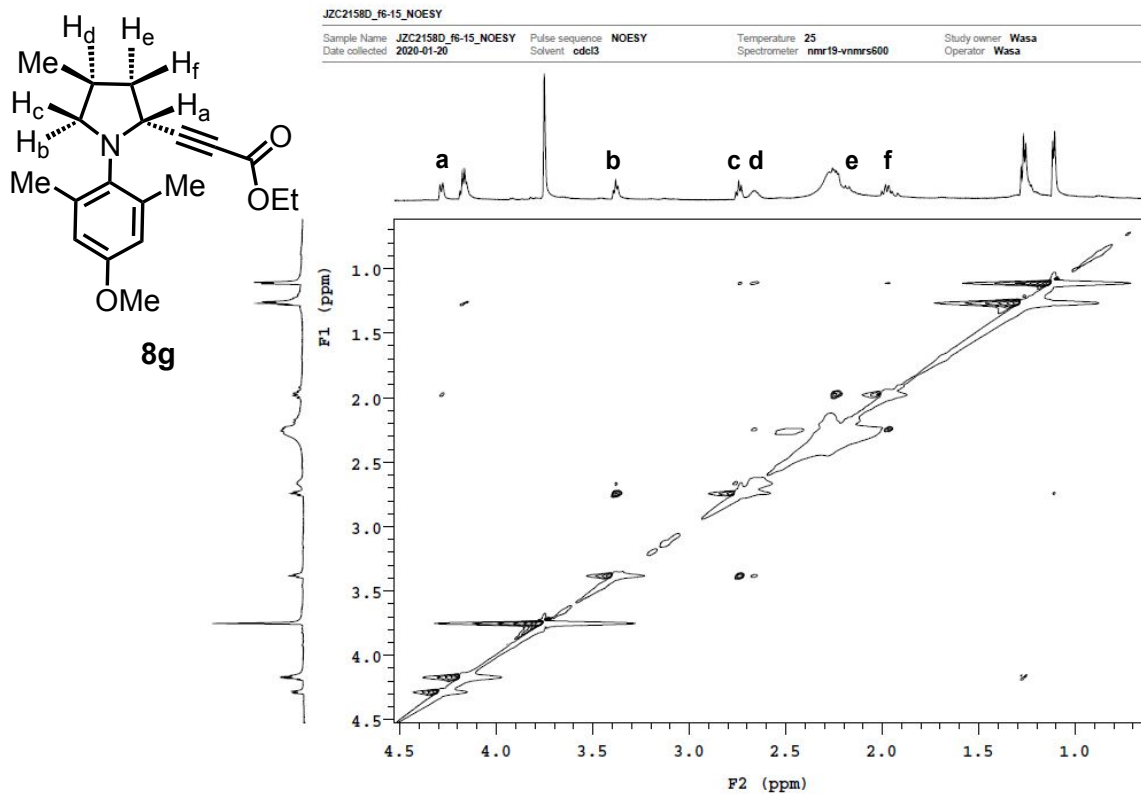
BuOMe (2.0 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 72 h. Upon completion, the unpurified reaction mixture was concentrated *in vacuo*. The amine product **(S)-8b** was obtained after purification by flash silica gel column chromatography (Et₂O:hexanes = 1:19) as a colorless solid (260 mg, 85%).

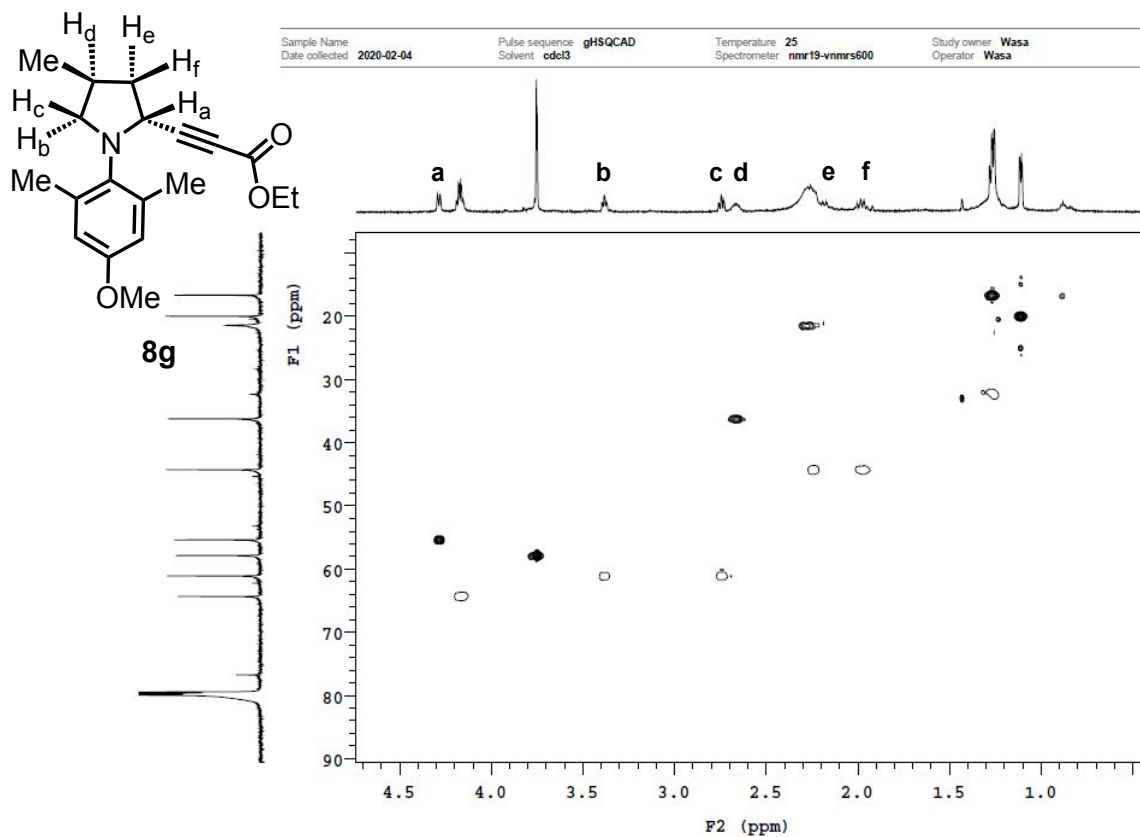
3. Determination of Relative Configuration

We carried out the following 2D NMR studies in order to determine relative configuration of enantioenriched products **8c**, **8g**, **8h**, **8i-trans**, and **8i-cis**.



The relative configuration of the major diastereomer of **8c** was assigned to be *trans*.





The relative configuration of the major diastereomer of **8g** was assigned as *trans*. The relative configuration of **8h** was assigned in analogy.

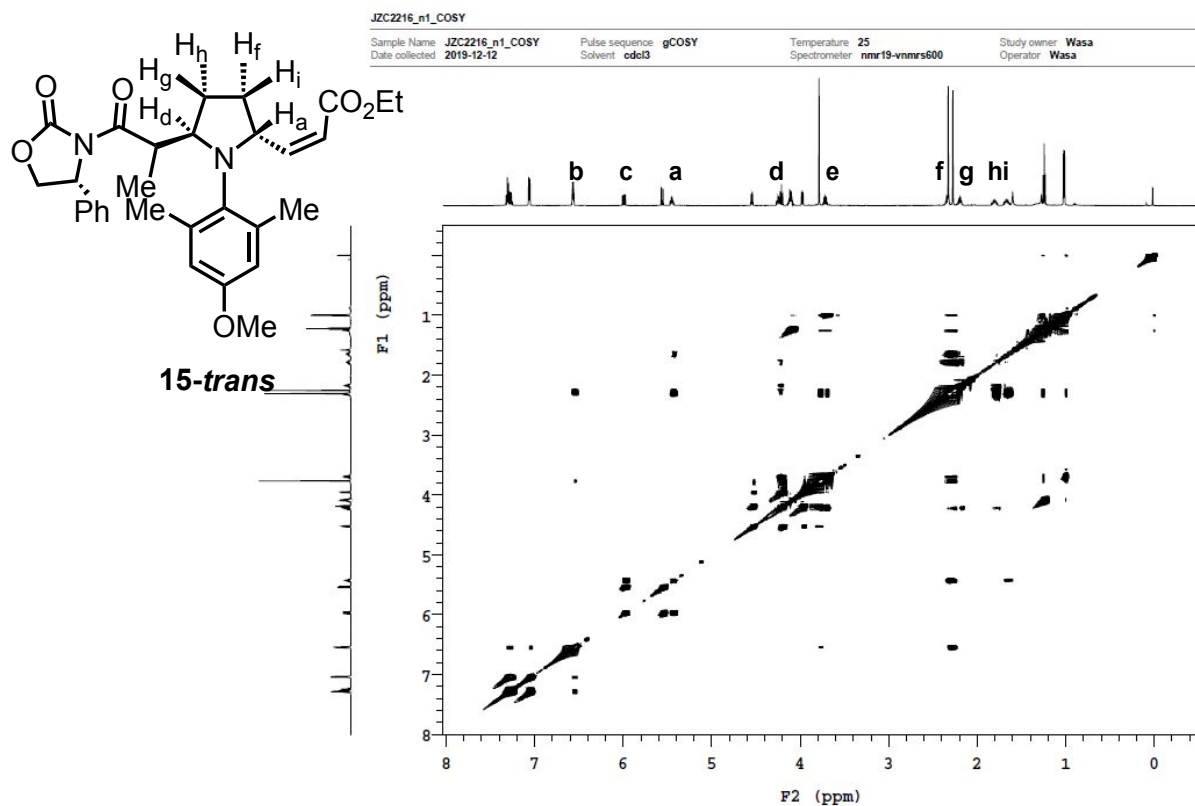
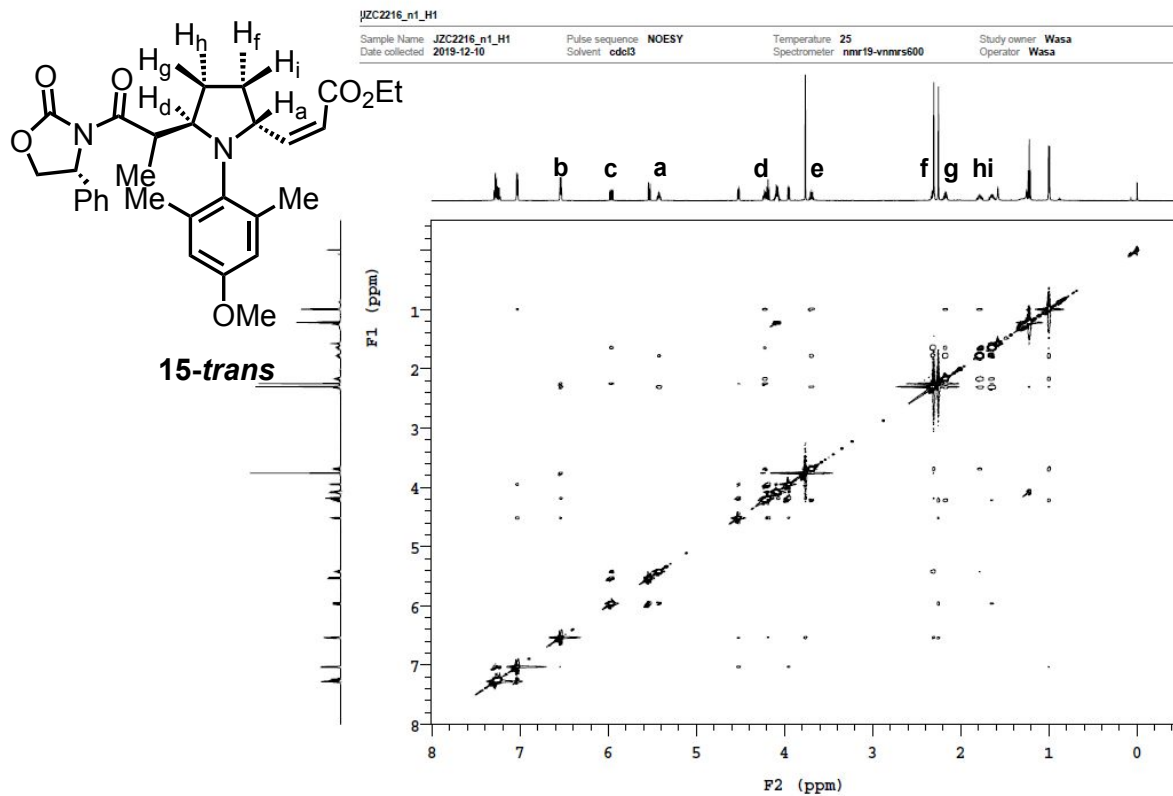
8i-major

10% Pd/CaCO₃
 H₂ (1 atm)
 pyridine
 22 °C, 3 h

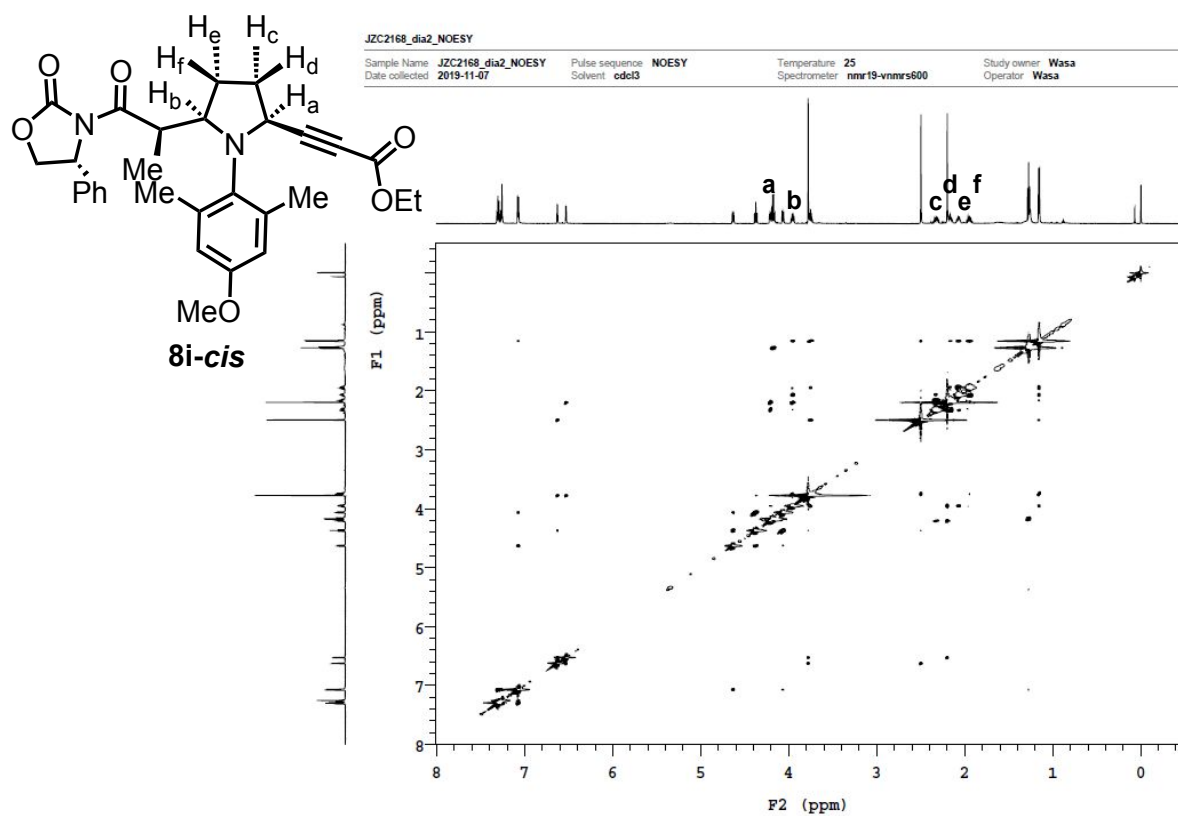
15-major, 98% yield

To a solution of **8i-major** (26 mg, 0.05 mmol) in pyridine (0.5 mL) was added Pd/CaCO₃ (2.6 mg, 10% wt). The reaction mixture was evacuated and filled with hydrogen gas three times. The reaction mixture was allowed to stir under hydrogen atmosphere at 22 °C for 3 h. Upon completion (determined by TLC), the reaction was filtered through a plug of Celite using EtOAc as the eluent. The amine product **15-major** was obtained after purification by flash silica gel column chromatography (Et₂O:hexanes = 1:4) as a colorless oil (25.5 mg, 98%).

¹H NMR (600 MHz, CDCl₃) δ 7.31 – 7.22 (m, 3H), 7.06 – 7.02 (m, 2H), 6.57 – 6.50 (m, 2H), 5.97 (ddd, *J* = 11.6, 8.7, 1.0 Hz, 1H), 5.54 (dt, *J* = 11.7, 1.1 Hz, 1H), 5.43 (td, *J* = 8.9, 5.7 Hz, 1H), 4.52 (dd, *J* = 8.5, 2.2 Hz, 1H), 4.23 (dt, *J* = 9.5, 4.7 Hz, 1H), 4.19 (td, *J* = 8.4, 1.1 Hz, 1H), 4.09 (qdd, *J* = 7.1, 3.8, 1.0 Hz, 2H), 3.95 (ddd, *J* = 8.4, 2.4, 1.1 Hz, 1H), 3.77 (d, *J* = 1.0 Hz, 3H), 3.73 – 3.66 (m, 1H), 2.34 – 2.28 (m, 4H), 2.26 (s, 3H), 2.18 (dt, *J* = 11.8, 6.1 Hz, 1H), 1.83 – 1.75 (m, 1H), 1.65 (tdd, *J* = 12.2, 9.2, 6.5 Hz, 1H), 1.58 (s, 1H), 1.22 (td, *J* = 7.1, 1.0 Hz, 3H), 1.00 (dd, *J* = 6.8, 1.0 Hz, 3H); **IR** (neat) 2957, 2926, 2855, 1778, 1713, 1480, 1411, 1182, 1155, 760, 699 cm⁻¹; [α]_D²⁵ = -289.9° (*c* 0.5, CH₂Cl₂).

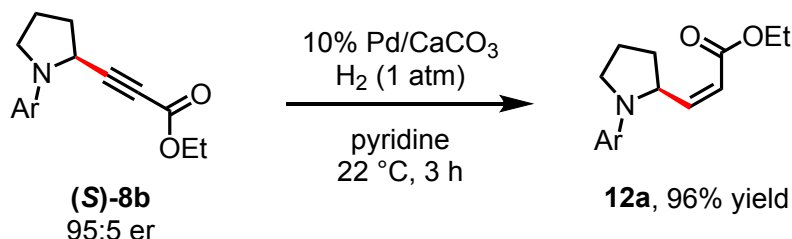


The relative configuration of **15-major** (derivative of **8i-major**) was assigned as *trans*.



The relative configuration of the minor diastereomer of **8i** was assigned as *cis*.

4. Determination of Absolute Configuration and Derivatization Experiments



Ethyl (*S,Z*)-3-(1-(4-methoxy-2,6-dimethylphenyl)pyrrolidin-2-yl)acrylate (**12a**)

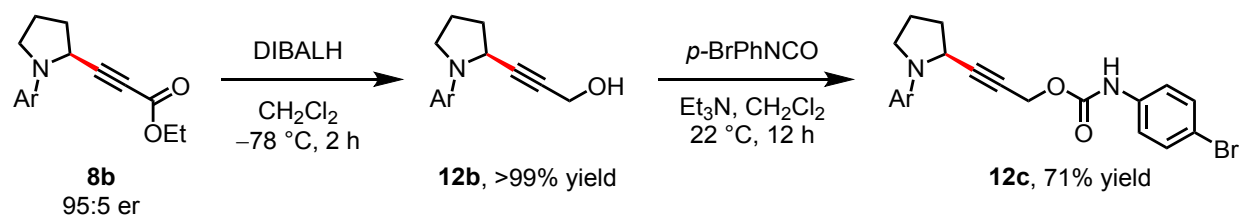
Ethyl (*S,Z*)-3-(1-(4-methoxy-2,6-dimethylphenyl)pyrrolidin-2-yl)acrylate (**12a**) was prepared according to the literature procedure.⁹

To a solution of (**S**)-**8b** (30.1 mg, 0.1 mmol) in pyridine (0.5 mL) was added Pd/CaCO₃ (3.0 mg, 10% wt). The reaction mixture was evacuated and filled with hydrogen gas three times. The reaction was allowed to stir under hydrogen atmosphere at 22 °C for 3 h. Upon completion (determined by TLC), the reaction was filtered through a plug of Celite using EtOAc as the eluent. The amine product **12a** was obtained after purification by flash silica gel column chromatography (Et₂O:hexanes = 1:19) as a colorless oil (29 mg, 96%).

¹H NMR (600 MHz, CDCl₃) δ 6.54 (s, 2H), 6.19 (ddd, *J* = 11.7, 8.7, 1.2 Hz, 1H), 5.56 (dt, *J* = 11.6, 1.3 Hz, 1H), 5.26 – 5.16 (m, 1H), 4.06 (qdd, *J* = 7.0, 5.0, 2.3 Hz, 2H), 3.73 (s, 3H), 3.41 – 3.30 (m, 1H), 3.07 (q, *J* = 7.6 Hz, 1H), 2.37 – 2.30 (m, 1H), 2.28 (s, 6H), 2.06 – 1.96 (m, 2H), 1.72 (dt, *J* = 12.1, 6.9 Hz, 1H), 1.20 (td, *J* = 7.2, 1.2 Hz, 3H); **¹³C NMR** (151 MHz, CDCl₃) δ 166.2, 156.6, 154.2, 137.3, 118.8, 59.8, 58.3, 55.1, 51.8, 33.5, 25.5, 19.3, 14.2; **IR** (neat) 2952, 2835, 1715, 1601, 1482, 1317, 1266, 1189, 1154, 1067, 1030, 827 cm⁻¹; **HRMS** (DART) *m/z* Calcd for C₁₈H₂₆NO₃ (MH⁺): 304.1907; found: 304.1906; [α]_D²⁵ = -97.5° (*c* 0.6, CH₂Cl₂).

Determination of Absolute Configuration for **8b**

We carried out the following studies in order to determine the absolute configuration of enantioenriched products **8b**, **8d**, and **8e**. The absolute configuration of (*S*)-**8b** was determined by X-ray crystallographic analysis of **12c**. The absolute configuration of **8d** and **8e** were assigned in analogy.



(*S*)-3-(1-(4-Methoxy-2,6-dimethylphenyl)pyrrolidin-2-yl)prop-2-yn-1-ol (**12b**)

(*S*)-3-(1-(4-Methoxy-2,6-dimethylphenyl)pyrrolidin-2-yl)prop-2-yn-1-ol (**12b**) was prepared according to the literature procedure.¹⁰ A solution of **8b** (100 mg, 0.3 mmol) in CH₂Cl₂ (5.0 mL) was cooled to -78 °C. DIBAL-H (0.1 mL, 0.5 mmol) was added dropwise. The reaction mixture was allowed to stir for 2 h at -78 °C. Then, potassium sodium tartarate (saturated aqueous solution, 3.0 mL) was added and stirred. The aqueous layer was extracted with CH₂Cl₂ (3 x 5 mL). The combined organic layers were dried (MgSO₄), filtered, and concentrated *in vacuo*. The amine product **12b** was obtained after purification by flash silica gel column chromatography (Et₂O:hexanes = 1:1) as a colorless oil (85.7 mg, >99%).

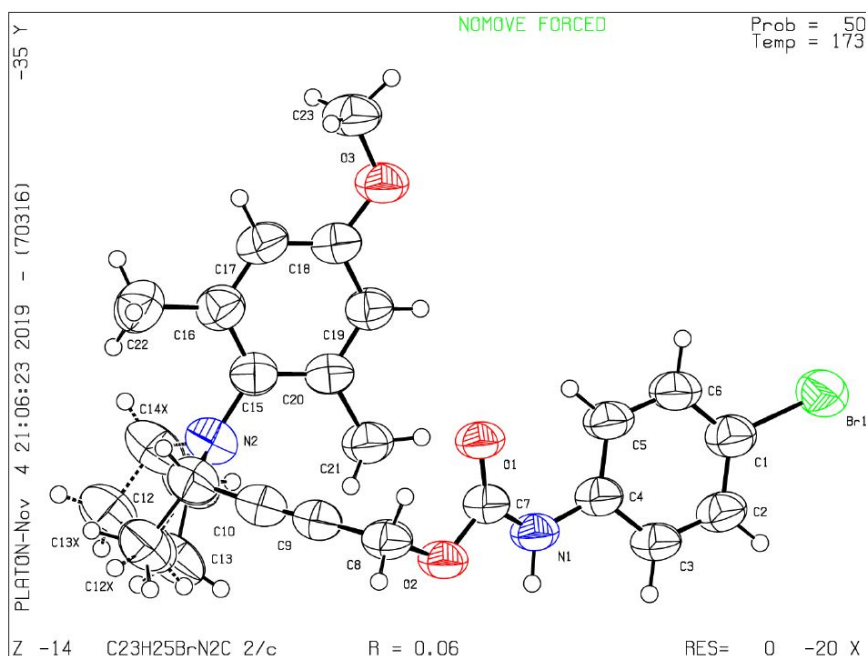
¹H NMR (500 MHz, CDCl₃) δ 6.57 (s, 2H), 4.22 (ddd, *J* = 6.7, 4.8, 1.7 Hz, 1H), 4.13 (s, 2H), 3.73 (s, 3H), 3.34 (td, *J* = 7.6, 4.4 Hz, 1H), 3.06 (q, *J* = 7.4 Hz, 1H), 2.42 – 2.17 (m, 7H), 2.17 – 2.01 (m, 2H), 1.96 (dtd, *J* = 11.7, 6.2, 5.8, 3.1 Hz, 1H), 1.70 (d, *J* = 4.6 Hz, 1H); ¹³C NMR (126 MHz, CDCl₃) δ 156.8, 136.4, 113.5, 88.6, 79.5, 55.1, 52.1, 51.2, 50.7, 34.4, 25.3, 19.0; IR (neat) 3379, 2939, 2834, 1600, 1482, 1273, 1153, 1066, 925, 835, 571 cm⁻¹; HRMS (DART) *m/z* Calcd for C₁₆H₂₂NO₂ (MH⁺): 260.1645; found: 260.1645.

(*S*)-3-(1-(4-Methoxy-2,6-dimethylphenyl)pyrrolidin-2-yl)prop-2-yn-1-yl (4-bromophenyl)carbamate (**12c**)

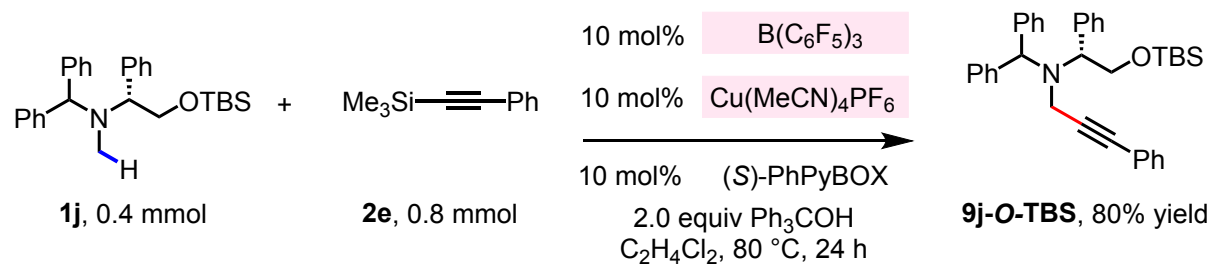
(*S*)-3-(1-(4-Methoxy-2,6-dimethylphenyl)pyrrolidin-2-yl)prop-2-yn-1-yl (4-bromophenyl)carbamate was prepared according to the literature procedure.¹⁰ To a solution of **12b** (50 mg, 0.2 mmol) in CH₂Cl₂ (1.0 mL), *p*-bromophenyl isocyanate (113 mg, 0.6 mmol) and Et₃N (96 mg, 1.0 mmol) were added. The reaction mixture was allowed to stir at 22 °C for

12 h. Upon completion (determined by TLC), the reaction mixture was filtered through a short plug of Celite using CH_2Cl_2 as the eluent. The organic layer was concentrated *in vacuo*. The amine product **12c** was obtained after purification by flash silica gel column chromatography (Et_2O :hexanes = 1:1) as a colorless solid (61.3 mg, 71%).

Amine **12c** was recrystallized using the vapor-vapor diffusion method, using Et_2O to dissolve the product in an inner vial, and pentane as the precipitant placed in the outer vial in order for slow diffusion to occur into the inner vial. The solution was cooled to 0 °C, whereupon a crystal was obtained for X-ray crystallography. The X-ray crystallographic analysis revealed that the absolute configuration of **12c** is (*S*), see SI Section 9 for X-ray crystallographic data. The absolute configuration of **8b** is (*S*). The absolute configuration of **8d** and **8e** were assigned in analogy to **8b**.



Procedure for Preparation of Propargylamine **9j-O-TBS**

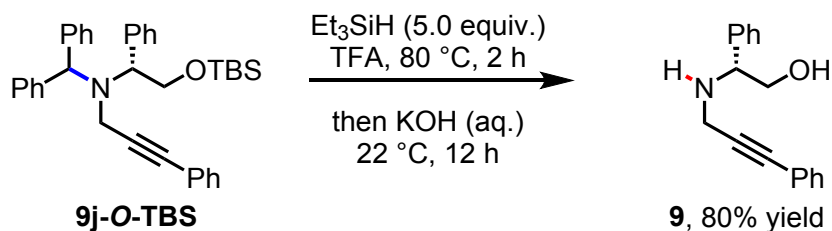


(*R*)-*N*-Benzhydryl-*N*-(2-((*tert*-butyldimethylsilyl)oxy)-1-phenylethyl)-3-phenylprop-2-yn-1-amine (**9j-O-TBS**)

An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added $\text{Cu(MeCN)}_4\text{PF}_6$ (0.04 mmol), (*S*)-PhPyBOX, 0.04 mmol, and $\text{C}_2\text{H}_4\text{Cl}_2$ (0.4 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then (trimethylsilyl)propiolate **2e** (0.8 mmol), triphenylmethanol (0.8 mmol), amine **1j** (0.4 mmol), $\text{B(C}_6\text{F}_5)_3$ (0.04 mmol, 10 mol%), and $\text{C}_2\text{H}_4\text{Cl}_2$ (0.4 mL) were added to the vessel. The reaction mixture was allowed to stir at 80 °C for 24 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH_2Cl_2 . The combined organic material was then concentrated *in vacuo*. The propargylamine product was purified and isolated by silica gel column chromatography (1:9 CH_2Cl_2 :hexane) to afford **9j-O-TBS** as a colorless oil (170.2 mg, 80% yield).

^1H NMR (600 MHz, CDCl_3) δ 7.69 – 7.58 (m, 6H), 7.47 – 7.33 (m, 12H), 7.33 – 7.24 (m, 2H), 5.40 (s, 1H), 4.38 (dd, J = 10.2, 5.1 Hz, 1H), 4.34 (dd, J = 10.2, 7.3 Hz, 1H), 4.30 (dd, J = 7.2, 5.1 Hz, 1H), 3.75 (d, J = 1.4 Hz, 2H), 0.92 (s, 9H), 0.08 (s, 3H), 0.02 (s, 3H); **^{13}C NMR** (151 MHz, CDCl_3) δ 142.6, 142.4, 140.5, 131.5, 128.9, 128.6, 128.5, 128.38, 128.35, 128.07, 128.06, 127.9, 127.7, 127.03, 126.95, 126.9, 123.7, 87.7, 84.4, 69.2, 63.4, 62.8, 37.0, 25.9, 18.2, -5.45, -5.47; **IR** (neat) 3057, 3025, 2924, 2852, 1597, 1488, 1451, 1251, 1095, 835, 813, 744, 699 cm^{-1} ; **HRMS** (DART) m/z Calcd for $\text{C}_{36}\text{H}_{42}\text{NOSi}$ (MH^+): 532.3030; found: 532.3023; $[\alpha]_D^{25} = -25.1^\circ$ (c 1.0, CH_2Cl_2).

Procedure for Deprotection of *N*-Benzhydryl Group

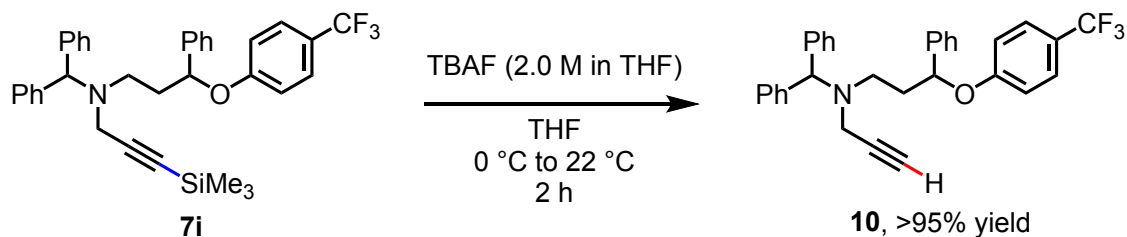


(*R*)-2-Phenyl-2-((3-phenylprop-2-yn-1-yl)amino)ethan-1-ol (**9**)

(*R*)-2-Phenyl-2-((3-phenylprop-2-yn-1-yl)amino)ethan-1-ol was prepared according to the literature procedure.¹¹ To a solution of **9j-O-TBS** (100 mg, 0.2 mmol) in TFA (1.5 mL) was added triethylsilane (0.14 mL, 0.9 mmol). The mixture was allowed to stir at 80 °C for 2 h. The mixture was cooled to 22 °C and KOH (1.0 M, aq.) was added in a dropwise manner until the solution was alkaline. The reaction mixture was allowed to stir at 22 °C for 12 h. CH₂Cl₂ (5 mL) was added and the organic material was extracted with CH₂Cl₂ (3 x 5 mL). The combined organic layers were combined, dried (MgSO₄), filtered, and concentrated *in vacuo*. The amine product **9** was obtained after purification by flash silica gel column chromatography (EtOAc:hexanes = 8:2) as a colorless solid (36.2 mg, 80%).

¹H NMR (600 MHz, CDCl₃) δ 7.43 – 7.38 (m, 2H), 7.38 – 7.33 (m, 4H), 7.32 – 7.26 (m, 4H), 4.06 (dd, *J* = 8.3, 4.4 Hz, 1H), 3.78 (dd, *J* = 10.9, 4.4 Hz, 1H), 3.68 – 3.60 (m, 2H), 3.45 (d, *J* = 17.0 Hz, 1H), 2.42 – 2.12 (m, 2H); **¹³C NMR** (151 MHz, CDCl₃) δ 139.70, 131.63, 128.66, 128.21, 128.03, 127.81, 127.57, 123.09, 87.29, 83.71, 66.79, 63.22, 36.72; **IR** (neat) 3288, 3056, 2914, 2847, 1488, 1451, 1329, 1026, 754, 690, 526 cm⁻¹; **HRMS** (DART) *m/z* Calcd for C₁₇H₁₈NO (MH⁺): 252.1383; found: 252.1379; [α]_D²⁵ = 199.1° (*c* 1.0, CH₂Cl₂).

Procedure for Removal of Trimethylsilyl Group

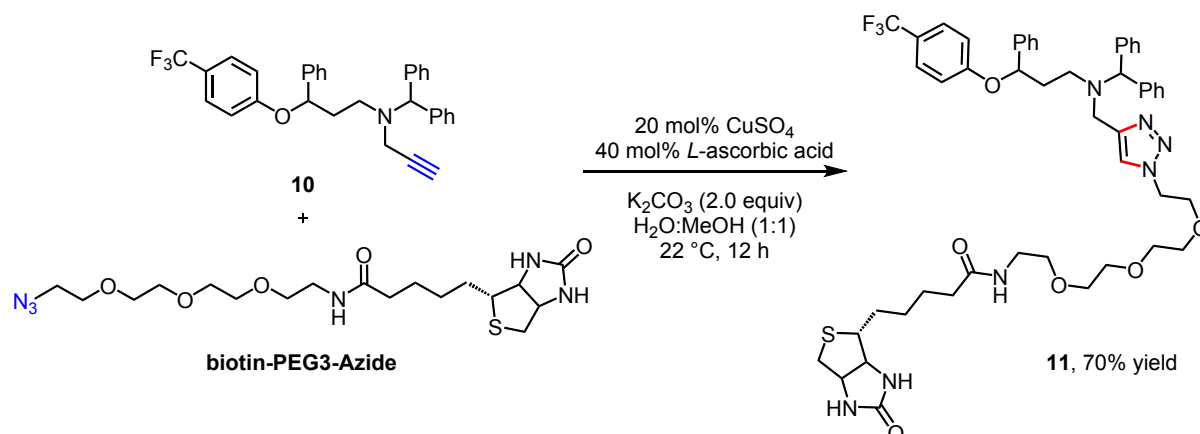


***N*-Benzhydryl-*N*-(3-phenyl-3-(4-(trifluoromethyl)phenoxy)propyl)prop-2-yn-1-amine (10)**

To a solution of **7i** (228 mg, 0.4 mmol) in THF (10 mL) was added TBAF (1.0 mL, 2.0 M in THF) at 0 °C. The reaction mixture was allowed to stir for 2 h at 22 °C. Upon completion (determined by TLC), the reaction mixture was concentrated *in vacuo*. The amine product **10** was obtained after purification by flash silica gel column chromatography (Et₂O:hexane = 1:99) as a colorless solid (200 mg, 99% yield).

¹H NMR (600 MHz, CDCl₃) δ 7.42 (dd, *J* = 8.4, 3.5 Hz, 4H), 7.34 – 7.30 (m, 2H), 7.28 (d, *J* = 5.9 Hz, 4H), 7.23 (t, *J* = 7.7 Hz, 3H), 7.20 – 7.10 (m, 2H), 7.06 (q, *J* = 7.0, 6.4 Hz, 3H), 6.80 (d, *J* = 8.4 Hz, 2H), 5.26 (d, *J* = 3.8 Hz, 1H), 4.68 (s, 1H), 3.44 (qd, *J* = 17.7, 2.4 Hz, 2H), 2.86 – 2.65 (m, 2H), 2.17 (s, 1H), 2.16 – 1.94 (m, 2H); **¹³C NMR** (151 MHz, CDCl₃) δ 160.6, 143.9, 142.6, 142.5, 141.4, 130.0, 129.4, 128.7, 128.6, 128.52, 128.48, 128.44, 128.38, 128.28, 128.25, 128.00, 127.96, 127.93, 127.92, 127.83, 127.80, 127.7, 127.4, 127.24, 127.15, 127.1, 126.96, 126.92, 126.68, 126.65, 126.63, 126.60, 126.3, 125.8, 125.6, 125.5, 125.4, 123.6, 122.6 (q, *J* = 32.7 Hz), 121.8, 115.7, 78.2, 77.8, 73.5, 72.2, 56.8, 46.4, 39.1, 36.8; **¹⁹F NMR** (470 MHz, CDCl₃) δ -61.34; **IR** (neat) ν 3298, 3060, 3025, 2924, 2831, 1700, 1612, 1515, 1491, 1326, 1250, 1110, 1066, 834, 700 cm⁻¹; **HRMS** (DART) Calcd for C₃₂H₂₉NOF₃ (MH⁺): 500.2196; found: 500.2183.

Procedure for Organocopper-Catalyzed Alkyne Azide Click Reaction



N-(2-(2-(2-(2-(4-((Benzhydryl(3-phenyl-3-(4-(trifluoromethyl)phenoxy)propyl)amino)methyl)-1H-1,2,3-triazol-1-yl)ethoxy)ethoxy)ethoxy)ethyl)-5-((4R)-2-oxohexahydro-1H-thieno[3,4-d]imidazol-4-yl)pentanamide (11)

N-(2-(2-(2-(2-(4-((Benzhydryl(3-phenyl-3-(4-(trifluoromethyl)phenoxy)propyl)amino)methyl)-1H-1,2,3-triazol-1-yl)ethoxy)ethoxy)ethoxy)ethyl)-5-((4*R*)-2-oxohexahydro-1H-thieno[3,4-*d*]imidazol-4-yl)pentanamide was prepared according to the literature procedure.¹²

To a solution of alkyne **10** (100 mg, 0.2 mmol) in MeOH (2.0 mL) was added K₂CO₃ (55 mg, 0.4 mmol), CuSO₄ (6.4 mg, 0.04 mmol), Biotin-PEG3-azide (98 mg, 0.22 mmol), L-ascorbic acid (14.1 mg, 0.08 mmol), and H₂O (2.0 mL). The reaction mixture was then allowed to stir for 12 h. Upon completion (determined by TLC), the reaction mixture was concentrated *in vacuo* to remove the organic solvent. EtOAc (3 x 5 mL) was used to extract the organic material. The combined organic layers were dried (MgSO₄), filtered, and concentrated *in vacuo*. The amine product **11** was obtained after purification by flash silica gel column chromatography (MeOH:CH₂Cl₂ = 1:99) as a colorless solid (132 mg, 70%).

¹H NMR (600 MHz, CDCl₃) δ 7.46 – 7.36 (m, 4H), 7.36 – 7.30 (m, 2H), 7.30 – 7.26 (m, 3H), 7.24 (t, *J* = 7.8 Hz, 4H), 7.19 (q, *J* = 7.2, 6.1 Hz, 4H), 6.77 (d, *J* = 8.5 Hz, 2H), 6.64 (s, 1H), 6.57 (s, 1H), 5.63 (s, 1H), 5.23 (dd, *J* = 8.5, 4.2 Hz, 1H), 4.78 (s, 1H), 4.42 (dtt, *J* = 14.3, 10.5, 5.2 Hz, 3H), 4.26 (dd, *J* = 7.8, 4.7 Hz, 1H), 3.94 – 3.74 (m, 4H), 3.54 (d, *J* = 7.2 Hz, 8H), 3.51 (t, *J* = 5.2 Hz, 2H), 3.40 (t, *J* = 5.2 Hz, 2H), 3.10 (td, *J* = 7.3, 4.5 Hz, 1H), 2.85 (dd, *J* = 12.7, 4.9 Hz, 1H), 2.77 – 2.68 (m, 2H), 2.63 (ddd, *J* = 12.9, 7.7, 4.6 Hz, 1H), 2.18 (t, *J* = 7.5 Hz, 2H), 2.12 (dtd, *J* = 14.5, 7.6, 7.0, 3.8 Hz, 2H), 1.78 – 1.56 (m, 4H), 1.46 – 1.34 (m, 2H); **¹³C NMR**

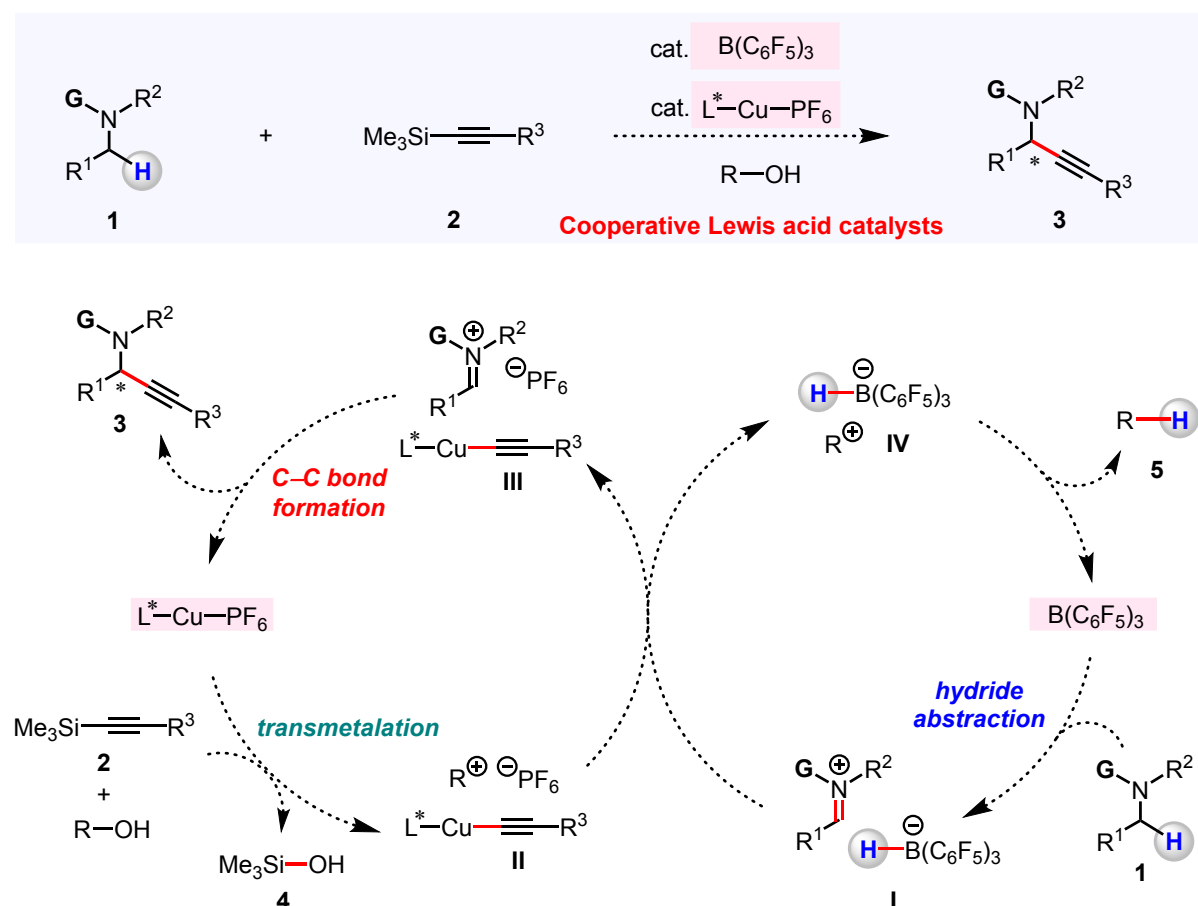
(151 MHz, CDCl₃) δ 173.2, 164.0, 160.5, 144.8, 142.2, 141.6, 141.30, 128.6, 128.5, 128.4, 128.34, 128.31, 128.26, 128.1, 127.6, 127.07, 127.00, 126.96, 126.62, 126.60, 126.57, 126.55, 125.7, 125.5, 125.3, 123.5, 122.4 (q, $J = 32.5$ Hz), 121.9, 115.6, 77.8, 77.2, 77.0, 76.9, 76.8, 70.6, 70.5, 70.4, 70.3, 70.0, 69.9, 69.8, 69.5, 65.8, 61.7, 60.1, 55.6, 50.0, 46.1, 44.9, 40.4, 39.1, 36.1, 35.9, 35.9, 30.3, 29.6, 29.6, 28.2, 28.0, 25.5, 15.2; **¹⁹F NMR** (564 MHz, CDCl₃) δ -61.44; **IR** (neat) 3287, 2921, 2863, 1698, 1612, 1451, 1325, 1250, 1109, 1066, 835, 734, 701 cm⁻¹; **HRMS** (DART) m/z Calcd for C₅₀H₆₁F₃N₇O₆S (MH⁺): 944.4278; found: 944.4342.

5. Mechanistic Studies for α -C–H Alkynylation of *N*-Alkylamines Catalyzed by $\text{B}(\text{C}_6\text{F}_5)_3$ and Organocopper Complex

5.1 Kinetic Experiments for the Coupling of 4-Methoxy-*N,N*,2,6-tetramethylaniline and Ethyl 3-(trimethylsilyl)propiolate

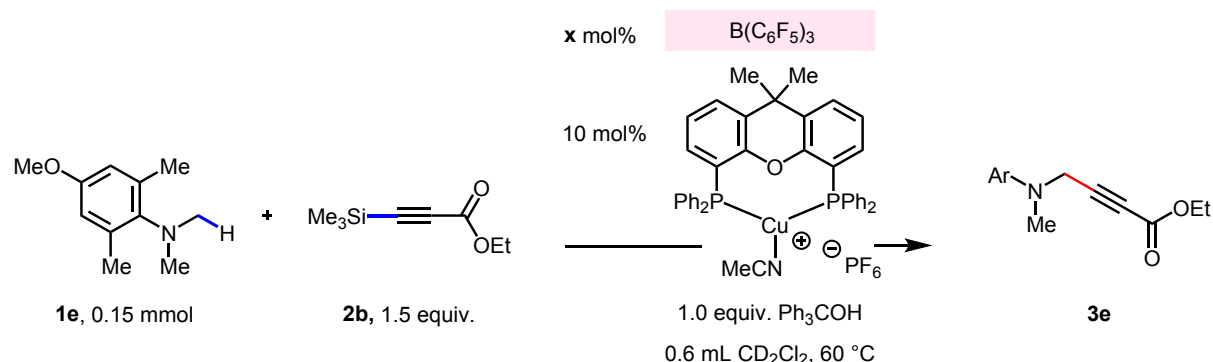
In order to provide evidence for the proposed reaction mechanism, we carried out the following kinetic experiments. We originally proposed (as shown in Figure 1c of the manuscript) that α -alkynylation process is proposed to proceed through $\text{B}(\text{C}_6\text{F}_5)_3$ -catalyzed hydride abstraction of the *N*-alkylamine substrate (**1**) to afford an iminium ion (**I**, Scheme S1). Subsequently, the organocopper catalyst can activate the alkyne substrate (**2**) with the aid of alcohol additive to promote transmetalation (**II**), which generates a nucleophilic alkyne that can attack the iminium ion (**III**) to afford the desired propargylamine **3**.

Scheme S1. Proposed Catalytic Cycle



5.1.1 Determination of Reaction Order of B(C₆F₅)₃

A kinetic study was conducted following the procedure for time course reaction monitoring by ¹H NMR (using internal standard) while varying the concentration of B(C₆F₅)₃ (Figure S1). Initial-rate kinetic analysis, which was determined from the data points in the first 400 seconds, demonstrates half-order kinetics of B(C₆F₅)₃ in the reaction between 4-methoxy-*N,N*,2,6-tetramethylaniline **1e** and ethyl 3-(trimethylsilyl)propiolate **2b** (Figure S2).¹³



Procedure for Time Course Reaction Monitoring by *in situ* ¹H NMR

In a nitrogen-filled glove box, previously prepared (Xantphos)Cu(MeCN)PF₆ (41 mg, 0.11 mmol),¹⁴ 4-methoxy-*N,N*,2,6-tetramethylaniline (197 mg, 1.1 mmol), ethyl 3-(trimethylsilyl)propiolate (281 mg, 1.65 mmol) and mesitylene (132 mg, 1.1 mmol) were weighed in an oven-dried 7.0 mL vial and diluted to 2.2 mL with CD₂Cl₂ (**Stock Solution A**). In another oven-dried 7.0 mL vial, B(C₆F₅)₃ (63.8 mg, 0.125 mmol) was weighed and diluted to 1.0 mL with CD₂Cl₂ (**Stock Solution B**). In 4 oven-dried 7.0 mL vials, were added triphenylmethanol (39 mg, 0.15 mmol). To each oven-dried vial containing triphenylmethanol was added **Stock Solution A** (0.3 mL), **Stock Solution B** (0.06, 0.12, 0.15 and 0.18 mL) and neat CD₂Cl₂ (0.24, 0.18, 0.15 and 0.12 mL) to prepare reaction mixtures (total 0.6 mL) with 0.150 mmol of 4-methoxy-*N,N*,2,6-tetramethylaniline, 0.225 mmol ethyl 3-(trimethylsilyl)propiolate, 0.150 mmol triphenylmethanol, 10.0 mol% (Xantphos)Cu(MeCN)PF₆, and the following amounts of B(C₆F₅)₃ (5.00, 10.0, 12.5 and 15.0 mol%). The reaction mixture was then transferred to a J-Young tube. After the J-Young tube was tightly capped with the Teflon plug, it was taken out of the glove box and ¹H NMR spectra were acquired at 60 °C (preheated) using a pre-acquisition delay in array mode with a spectrum taken every 30 seconds for the length of the experiment. The data were processed using MestReNova and peak integrations were normalized using mesitylene as the internal standard.

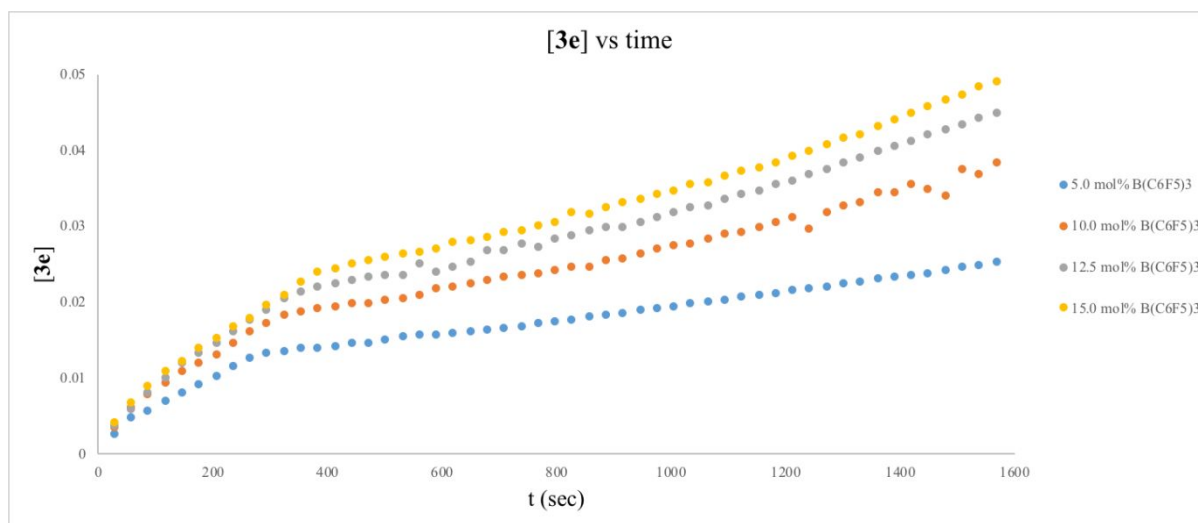


Figure S1. Monitoring the formation of **3e** using different concentrations of $\text{B}(\text{C}_6\text{F}_5)_3$.

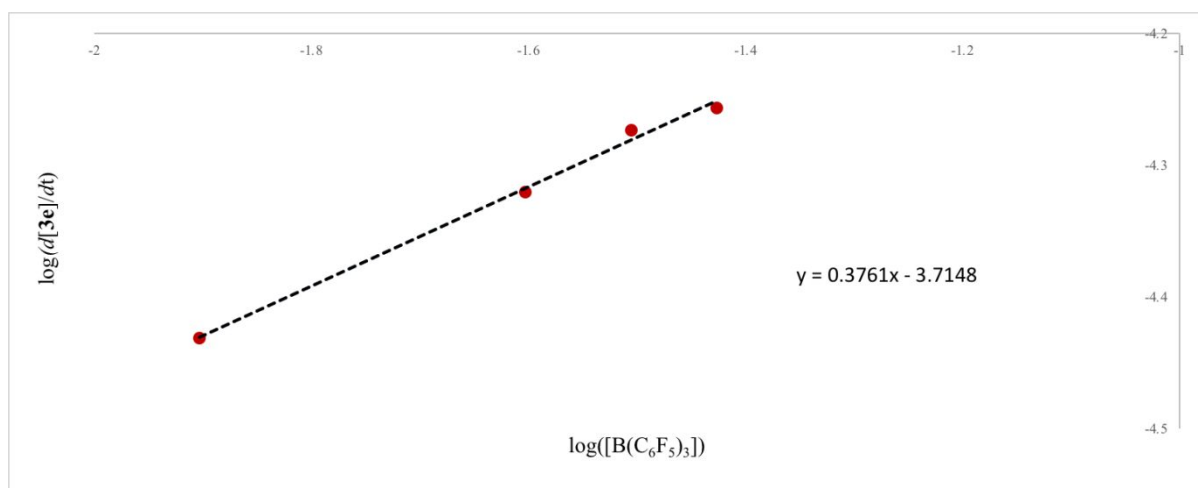
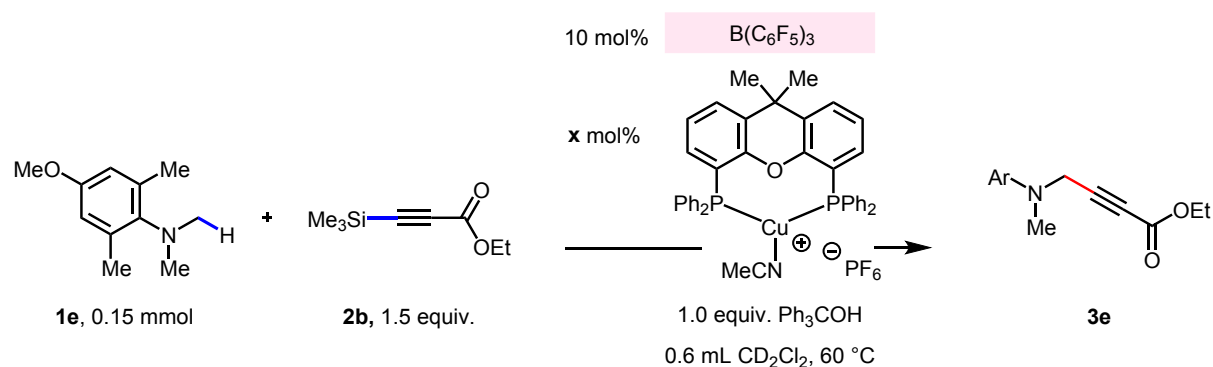


Figure S2. Log(rate) vs Log $[\text{B}(\text{C}_6\text{F}_5)_3]$ plot is employed to determine the reaction order for $\text{B}(\text{C}_6\text{F}_5)_3$. The result suggests that there is approximately 0.5-order dependency on the concentration of $\text{B}(\text{C}_6\text{F}_5)_3$.

5.1.2 Determination of Reaction Order of (Xantphos)Cu(MeCN)PF₆

A kinetic study was conducted following the procedure for time course reaction monitoring by ¹H NMR (using internal standard) while varying the concentration of (Xantphos)Cu(MeCN)PF₆ (Figure S3). Initial-rate kinetic analysis, which was determined from the data points in the first 400 seconds, demonstrates zero-order kinetics of (Xantphos)Cu(MeCN)PF₆ in the reaction between 4-methoxy-*N,N*,2,6-tetramethylaniline **1e** and ethyl 3-(trimethylsilyl)propiolate **2b** (Figure S4).¹³



In a nitrogen-filled glove box, B(C₆F₅)₃ (56.3 mg, 0.11 mmol), 4-methoxy-*N,N*,2,6-tetramethylaniline (197 mg, 1.10 mmol), ethyl 3-(trimethylsilyl)propiolate (281 mg, 1.65 mmol) and mesitylene (132 mg, 1.10 mmol) were weighed in an oven-dried 7.0 mL vial and diluted to 2.2 mL with CD₂Cl₂ (**Stock Solution A**). In another oven-dried 7.0 mL vial, (Xantphos)Cu(MeCN)PF₆ (103.5 mg, 0.125 mmol)¹⁴ was weighed and diluted to 2.00 mL with CD₂Cl₂ (**Stock Solution B**). In 5 oven-dried 7.0 mL vials, were added triphenylmethanol (39 mg, 0.15 mmol). To each oven-dried vial containing triphenylmethanol was added **Stock Solution A** (0.30 mL), **Stock Solution B** (0.06, 0.12, 0.24, 0.30 and 0.36 mL) and neat CD₂Cl₂ (0.30, 0.24, 0.12, 0.06 and 0.00 mL) to prepare reaction mixtures (total 0.62 mL) with 0.150 mmol of 4-methoxy-*N,N*,2,6-tetramethylaniline, 0.225 mmol ethyl 3-(trimethylsilyl)propiolate, 0.15 mmol triphenylmethanol, 10.0 mol% B(C₆F₅)₃, and the following amounts of (Xantphos)Cu(MeCN)PF₆ (2.5, 5.0, 10.0, 12.5 and 15.0 mol%). The reaction mixture was then transferred to a J-Young tube. After the J-Young tube was tightly capped with the Teflon plug, it was taken out of the glove box and ¹H NMR spectra were acquired at 60 °C (preheated) using a pre-acquisition delay in array mode with a spectrum taken every 30 seconds for the length of the experiment. The data were processed using MestReNova and peak integrations were normalized using mesitylene as the internal standard.

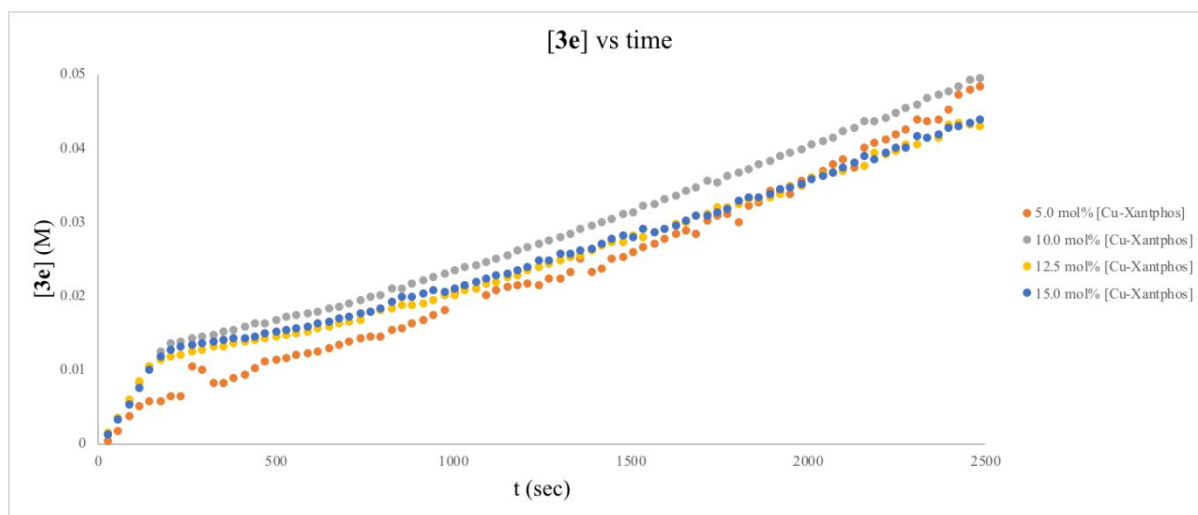


Figure S3. Monitoring the formation of **3e** using different concentrations of (Xantphos)Cu(MeCN)(PF₆).

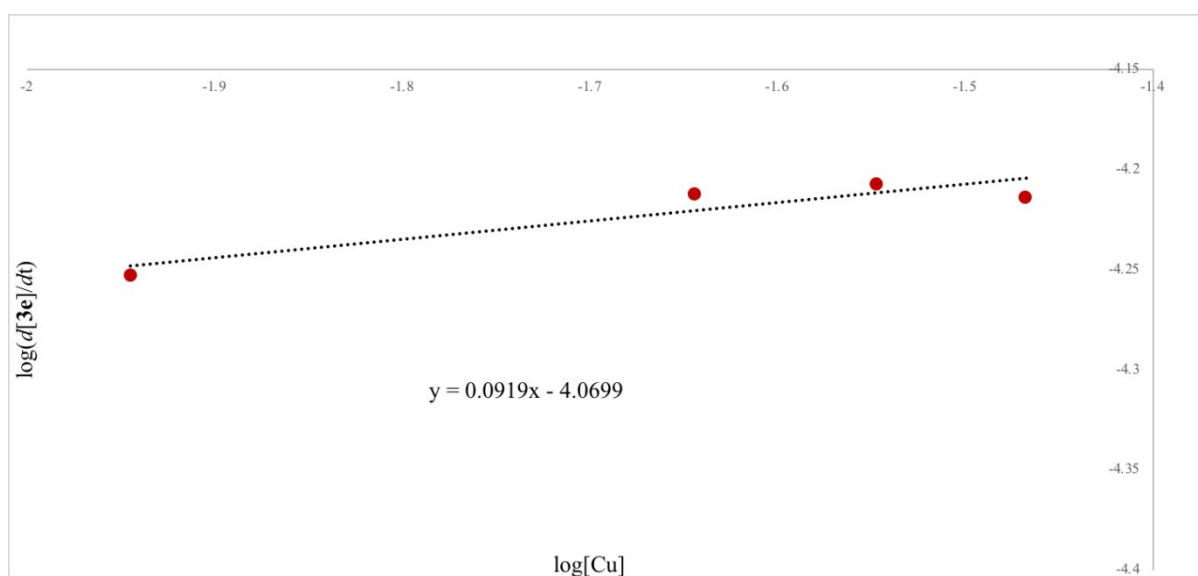
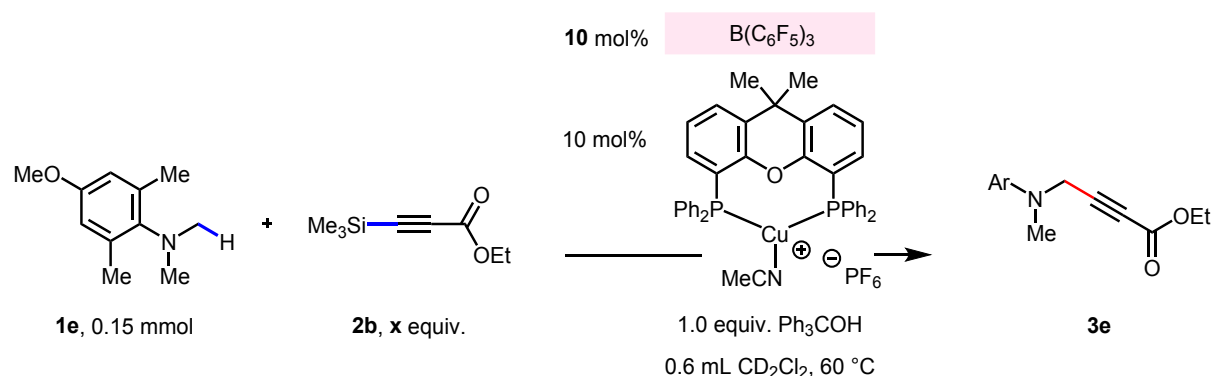


Figure S4. Log(rate) vs Log[(Xantphos)Cu(MeCN)(PF₆)] plot is employed to determine the initial reaction order for (Xantphos)Cu(MeCN)(PF₆). The result suggests that there is 0-order dependency on the concentration of (Xantphos)Cu(MeCN)(PF₆).

5.1.3 Determination of Reaction Order of Ethyl 3-(trimethylsilyl)propiolate **2b**

A kinetic study was conducted following the procedure for time course reaction monitoring by ¹H NMR (using internal standard) while varying the concentration of ethyl 3-(trimethylsilyl)propiolate **2b** (Figure S5). Initial-rate kinetic analysis, which was determined from the data points in the first 400 seconds, demonstrates zero-order kinetics of ethyl 3-

(trimethylsilyl)propiolate in the reaction between 4-methoxy-*N,N*,2,6-tetramethylaniline **1e** and ethyl 3-(trimethylsilyl)propiolate **2b** (Figure S6).¹³



Procedure for Time Course Reaction Monitoring by *in situ* ¹H NMR

In a nitrogen-filled glove box, (Xantphos)Cu(MeCN)PF₆ (103.5 mg, 0.11 mmol),¹⁴ B(C₆F₅)₃ (56.3 mg, 0.11 mmol), 4-methoxy-*N,N*,2,6-tetramethylaniline (197 mg, 1.10 mmol), and mesitylene (132 mg, 1.10 mmol) were weighed in an oven-dried 7.0 mL vial and diluted to 2.20 mL with CD₂Cl₂ (**Stock Solution A**). In another oven-dried 7.0 mL vial, (ethyl 3-(trimethylsilyl)propiolate (623 mg, 3.66 mmol) was weighed and diluted to 1.30 mL with CD₂Cl₂ (**Stock Solution B**). In 5 oven-dried 7.0 mL vials, were added triphenylmethanol (39 mg, 0.15 mmol). To each oven-dried vial containing triphenylmethanol was added **Stock Solution A** (0.30 mL), **Stock Solution B** (0.04, 0.08, 0.12, 0.16 and 0.24 mL) and neat CD₂Cl₂ (0.28, 0.24, 0.18, 0.14 and 0.08 mL) to prepare reaction mixtures (total 0.60 mL) with 0.15 mmol of 4-methoxy-*N,N*,2,6-tetramethylaniline, 0.15 mmol triphenylmethanol, 10.0 mol% B(C₆F₅)₃, 10.0 mol% (Xantphos)Cu(MeCN)PF₆ and the following amounts of ethyl 3-(trimethylsilyl)propiolate (0.75, 1.50, 2.25, 3.00 and 4.50 equiv.). The reaction mixture was then transferred to a J-Young tube. After the J-Young tube was tightly capped with the Teflon plug, it was taken out of the glove box and ¹H NMR spectra were acquired at 60 °C (preheated) using a pre-acquisition delay in array mode with a spectrum taken every 30 seconds for the length of the experiment. The data were processed using MestReNova and peak integrations were normalized using mesitylene as the internal standard.

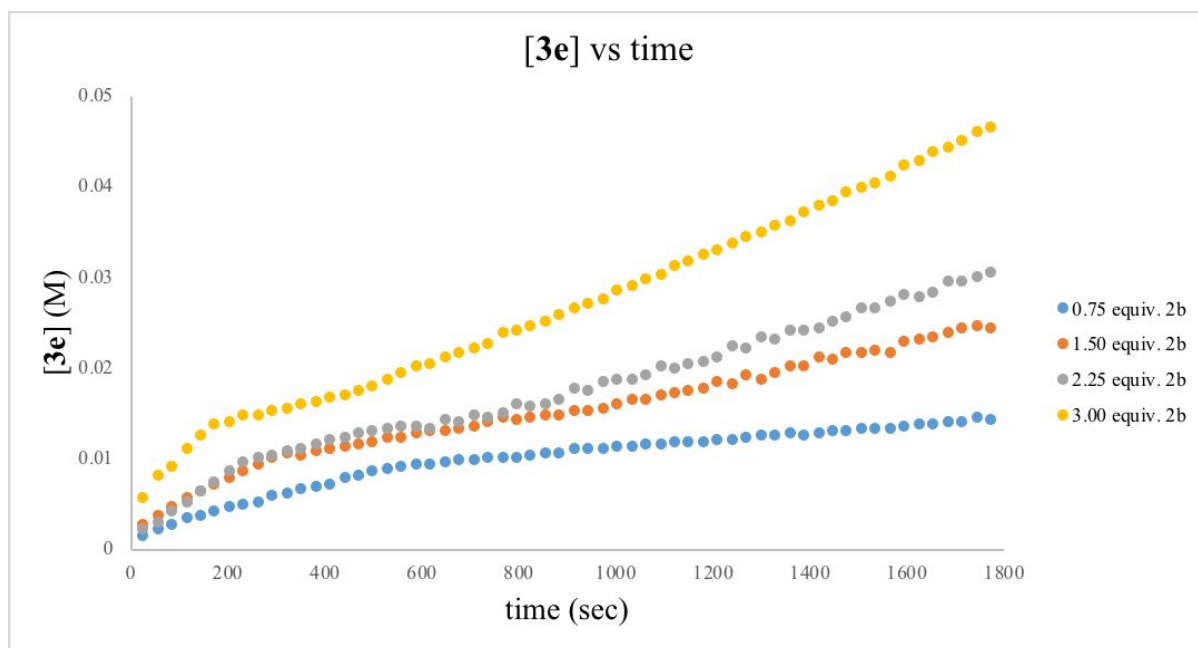


Figure S5. Monitoring the formation of **3e** using different concentrations of **2b**.

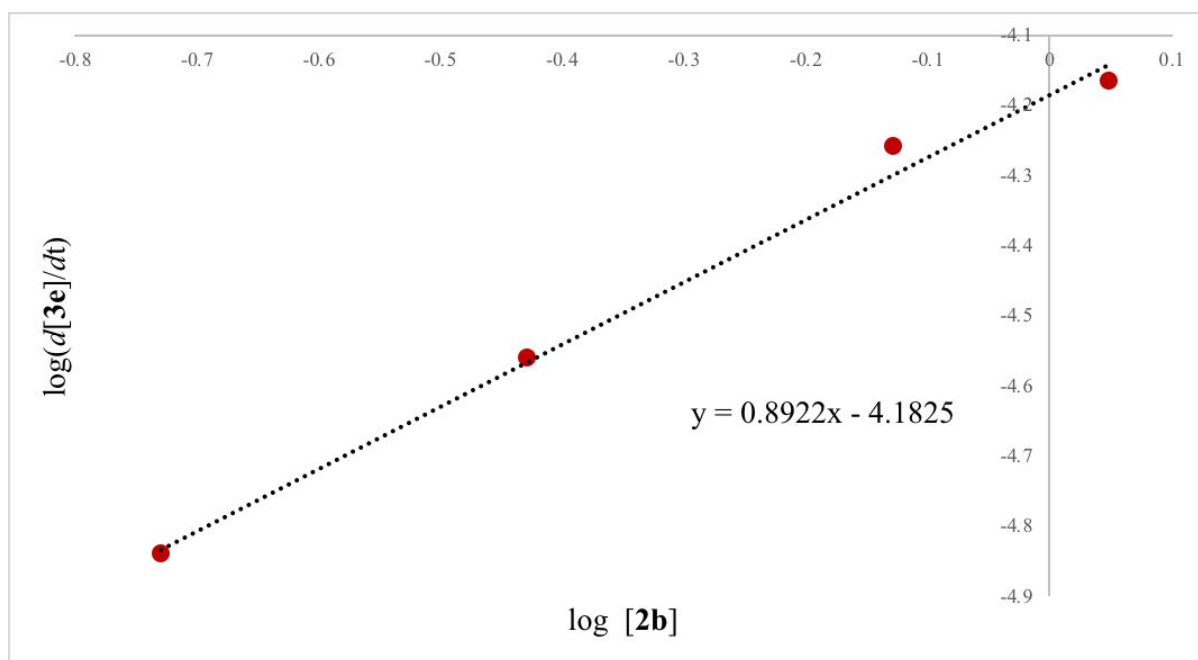
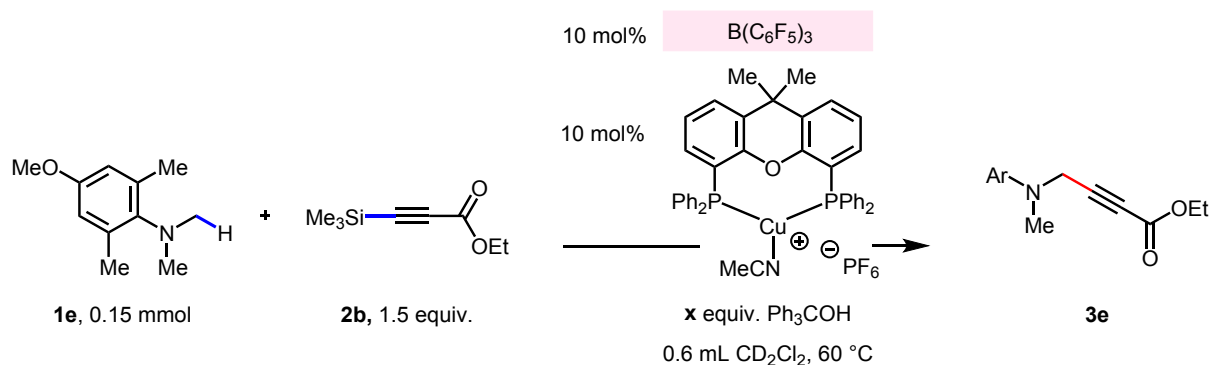


Figure S6. Log(rate) vs Log[**2b**] plot is employed to determine the initial reaction order for **2b**. The result suggests that there is 1.0-order dependency on the concentration of **2b**.

5.1.4 Determination of Reaction Order of Ph_3COH

A kinetic study was conducted following the procedure for time course reaction monitoring by ^1H NMR (using internal standard) while varying the concentration of trityl alcohol (Figure S7). Initial-rate kinetic analysis, which was determined from the data points in

the first 400 seconds, demonstrates zero-order kinetics of trityl alcohol in the reaction between 4-methoxy-*N,N*,2,6-tetramethylaniline **1e** and ethyl 3-(trimethylsilyl)propiolate **2b** (Figure S8).¹³



In a nitrogen-filled glove box, $(\text{Xantphos})\text{Cu}(\text{MeCN})\text{PF}_6$ (91.2 mg, 0.11 mmol),¹⁴ $\text{B}(\text{C}_6\text{F}_5)_3$ (56.3 mg, 0.11 mmol), 4-methoxy-*N,N*,2,6-tetramethylaniline (197 mg, 1.1 mmol), ethyl 3-(trimethylsilyl)propiolate (281 mg, 1.65 mmol) and mesitylene (132 mg, 1.1 mmol) were weighed in an oven-dried 7.0 mL vial and diluted to 2.2 mL with CD_2Cl_2 (**Stock Solution A**). In 5 oven-dried 7.0 mL vials, were added triphenylmethanol (19.5 mg, 39.0 mg, 52.0 mg, 64.7 mg and 78.0 mg). To each oven-dried vial containing triphenylmethanol was added **Stock Solution A** (0.3 mL) and neat CD_2Cl_2 (0.3 mL) to prepare reaction mixtures (total 0.6 mL) with 0.15 mmol of 4-methoxy-*N,N*,2,6-tetramethylaniline, 0.22 mmol ethyl 3-(trimethylsilyl)propiolate, 10.0 mol% $\text{B}(\text{C}_6\text{F}_5)_3$, and the following amounts of $(\text{Xantphos})\text{Cu}(\text{MeCN})\text{PF}_6$ (10.0 mol%) and triphenylmethanol (0.5, 1.0, 1.3, 1.7 and 2.0 equiv.). The reaction mixture was then transferred to a J-Young tube. After the J-Young tube was tightly capped with the Teflon plug, it was taken out of the glove box and ^1H NMR spectra were acquired at 60 °C (preheated) using a pre-acquisition delay in array mode with a spectrum taken every 30 seconds for the length of the experiment. The data were processed using MestReNova and peak integrations were normalized using mesitylene as the internal standard.

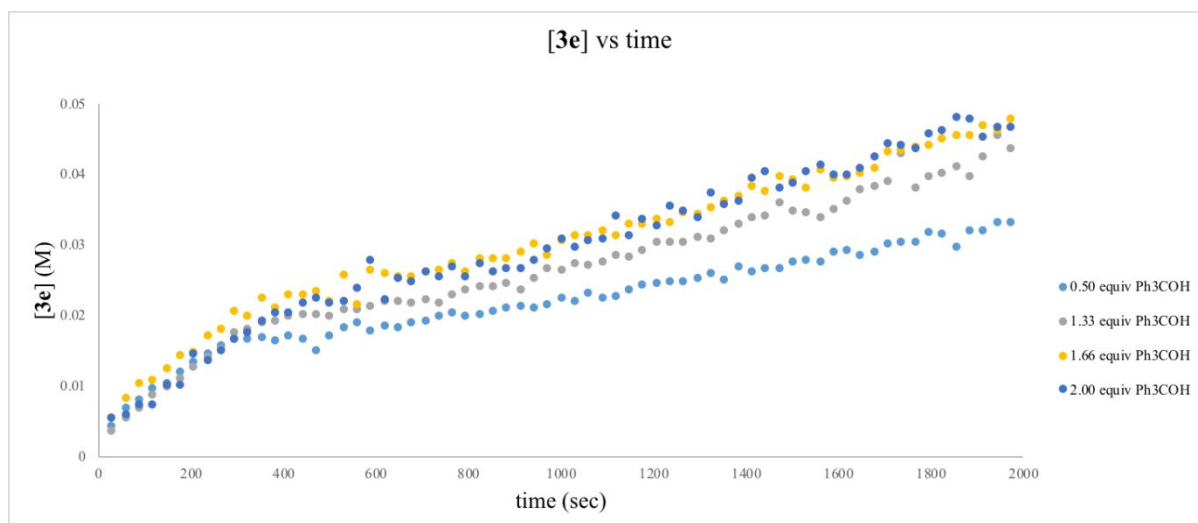


Figure S7. Monitoring the formation of **3e** using different concentrations of Ph₃COH.

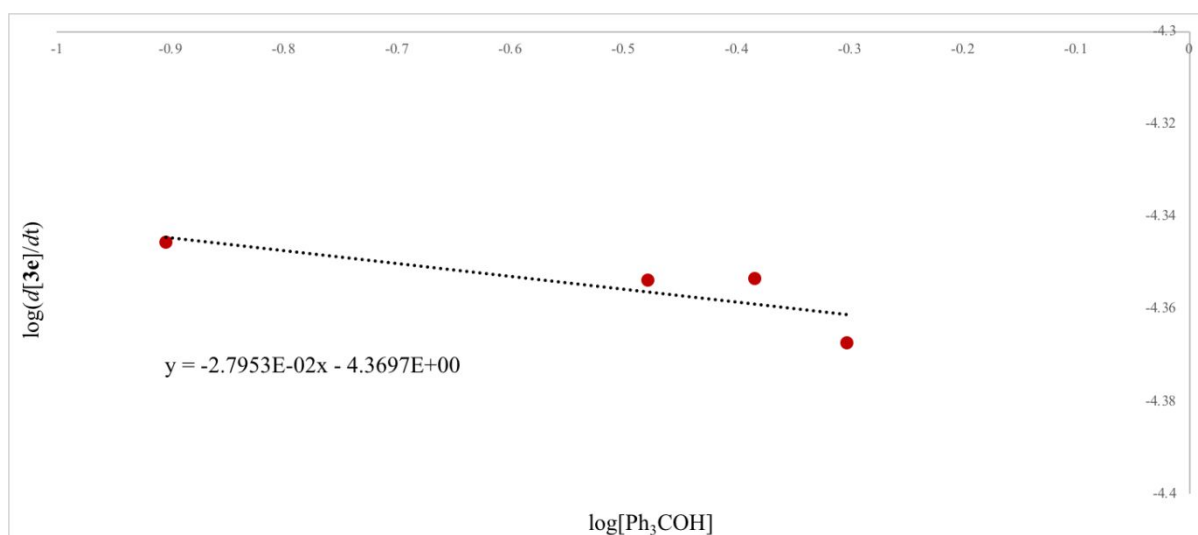
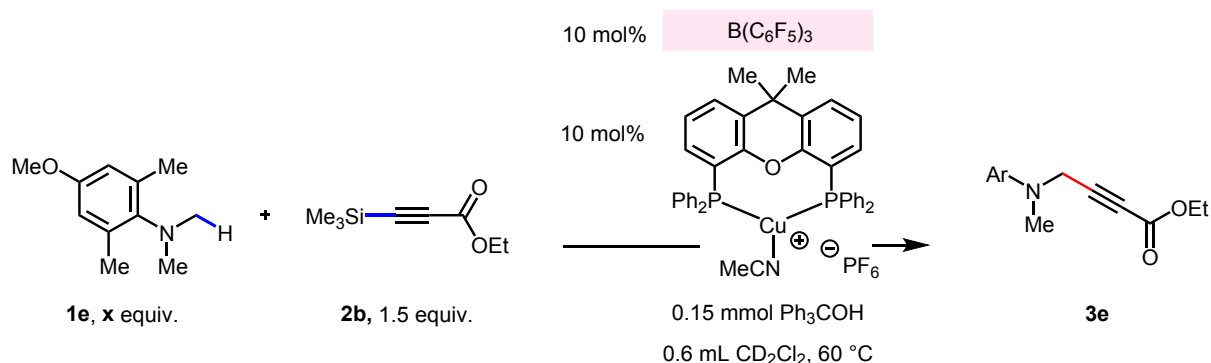


Figure S8. Log(rate) vs Log[Ph₃COH] plot is employed to determine the initial reaction order for Ph₃COH. The result suggests that there is 0-order dependency on the concentration of Ph₃COH.

5.1.5 Determination of Reaction Order in Ethyl 4-methoxy-*N,N*,2,6-tetramethylaniline **1e**

A kinetic study was conducted following the procedure for time course reaction monitoring by ¹H NMR (using internal standard) while varying the concentration of 4-methoxy-*N,N*,2,6-tetramethylaniline **1e** (Figure S9). Initial-rate kinetic analysis, which was determined from the data points in the first 400 seconds, demonstrates zero-order kinetics of 4-methoxy-*N,N*,2,6-tetramethylaniline **1e** (Figure S10).¹³



In a nitrogen-filled glove box, $\text{B}(\text{C}_6\text{F}_5)_3$ (56.3 mg, 0.11 mmol), $(\text{Xantphos})\text{Cu}(\text{MeCN})\text{PF}_6$ (91.2 mg, 0.11 mmol),¹⁴ ethyl 3-(trimethylsilyl)propiolate (281.0 mg, 1.65 mmol) and mesitylene (132 mg, 1.10 mmol) were weighed in an oven-dried 7.0 mL vial and diluted to 2.20 mL with CD_2Cl_2 (**Stock Solution A**). In another oven-dried 7.0 mL vial, 4-methoxy-*N,N*,2,6-tetramethylaniline (436.9 mg, 2.44 mmol) was weighed and diluted to 1.30 mL with CD_2Cl_2 (**Stock Solution B**). In 5 oven-dried 7.0 mL vials, were added triphenylmethanol (39 mg, 0.15 mmol). To each oven-dried vial containing triphenylmethanol was added **Stock Solution A** (0.3 mL), **Stock Solution B** (0.04, 0.06, 0.08, 0.16 and 0.24 mL) and neat CD_2Cl_2 (0.28, 0.26, 0.24, 0.16 and 0.08 mL) to prepare reaction mixtures (total 0.62 mL) with 0.225 mmol ethyl 3-(trimethylsilyl)propiolate, 0.15 mmol triphenylmethanol, 10.0 mol% $\text{B}(\text{C}_6\text{F}_5)_3$, 10.0 mol% $(\text{Xantphos})\text{Cu}(\text{MeCN})\text{PF}_6$ and the following amounts of 4-methoxy-*N,N*,2,6-tetramethylaniline (0.50, 0.75, 1.00, 2.00 and 3.00 equiv). The reaction mixture was then transferred to a J-Young tube. After the J-Young tube was tightly capped with the Teflon plug, it was taken out of the glove box and ^1H NMR spectra were acquired at 60 °C (preheated) using a pre-acquisition delay in array mode with a spectrum taken every 30 seconds for the length of the experiment. The data were processed using MestReNova and peak integrations were normalized using mesitylene as the internal standard.

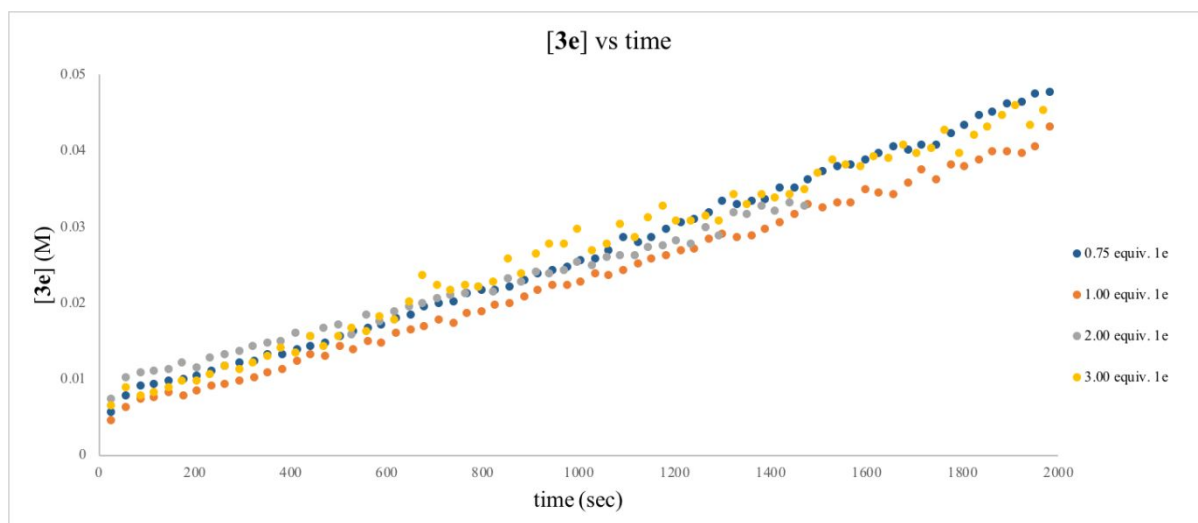


Figure S9. Monitoring the formation of **3e** using different concentrations of amine **1e**.

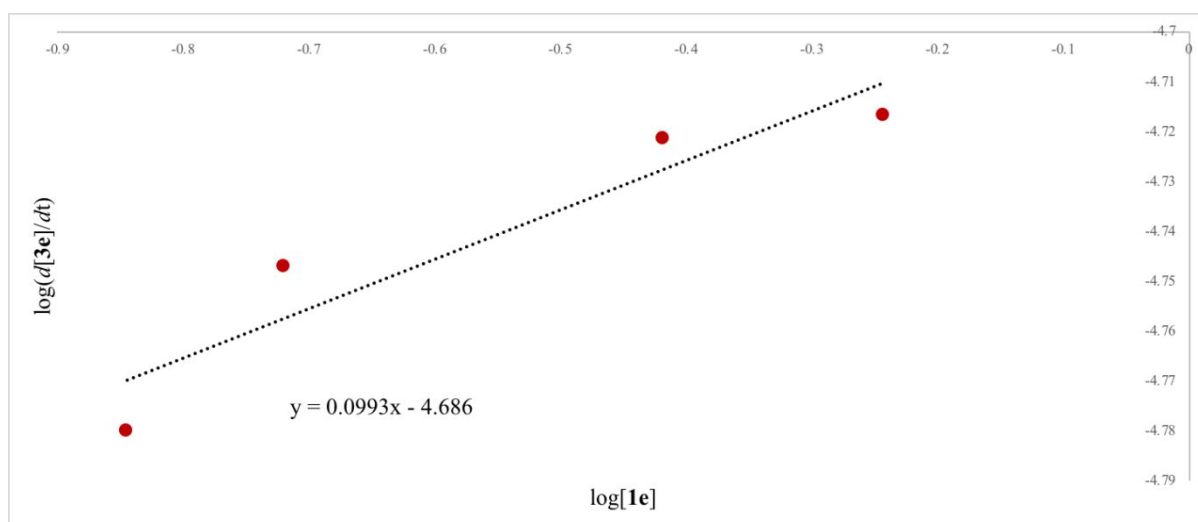


Figure S10. Log(rate) vs Log[**1e**] plot is employed to determine the initial reaction order for amine **1e**. The result suggests that there is 0-order dependency on the concentration of **1e**.

5.2 Parallel and Intermolecular Competition Kinetic Isotope Effect Experiments

5.2.1 Measurements of the Parallel Kinetic Isotope Effect

A parallel kinetic isotope effect study was conducted following the procedure for time course reaction monitoring by ^1H NMR (using internal standard). Kinetic analysis based on the initial rates of the product formation (Figure S11) demonstrates no kinetic isotope effect ($k_{\text{H}}/k_{\text{D}} = 1.02 \pm 0.02$, average of 2 reactions) in the reaction between *N*-benzyl-4-methoxy-*N*,2,6-trimethylaniline **1g** or *N*-benzyl-4-methoxy-2,6-dimethyl-*N*-(methyl- d_3)aniline **1g-d** and ethyl 3-(trimethylsilyl)propiolate **2b**.^{13,15}

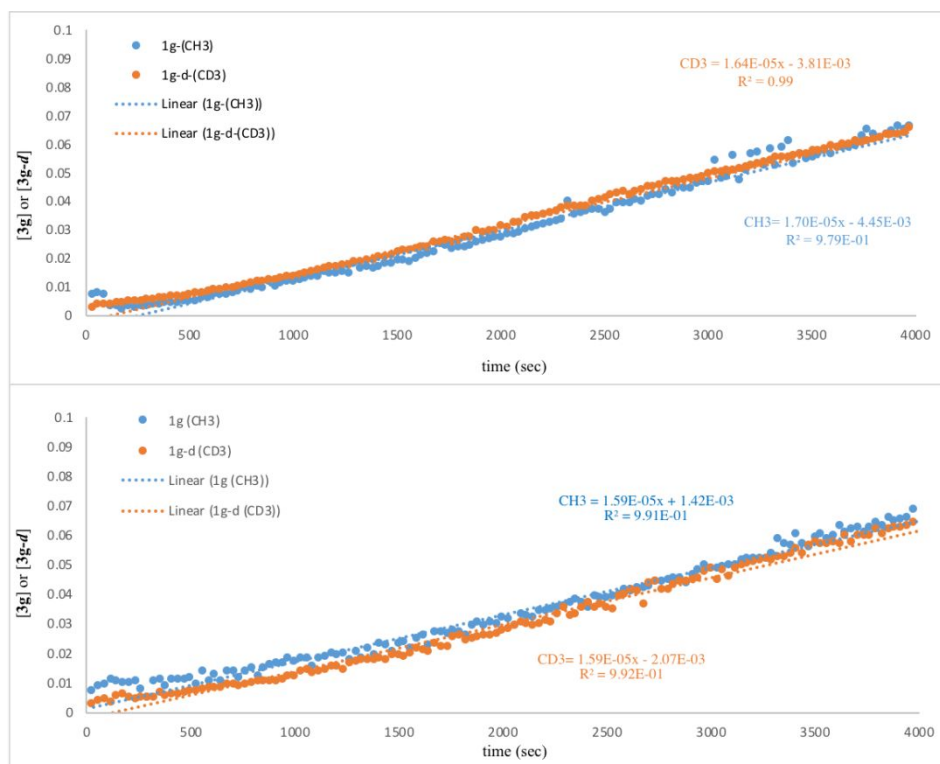
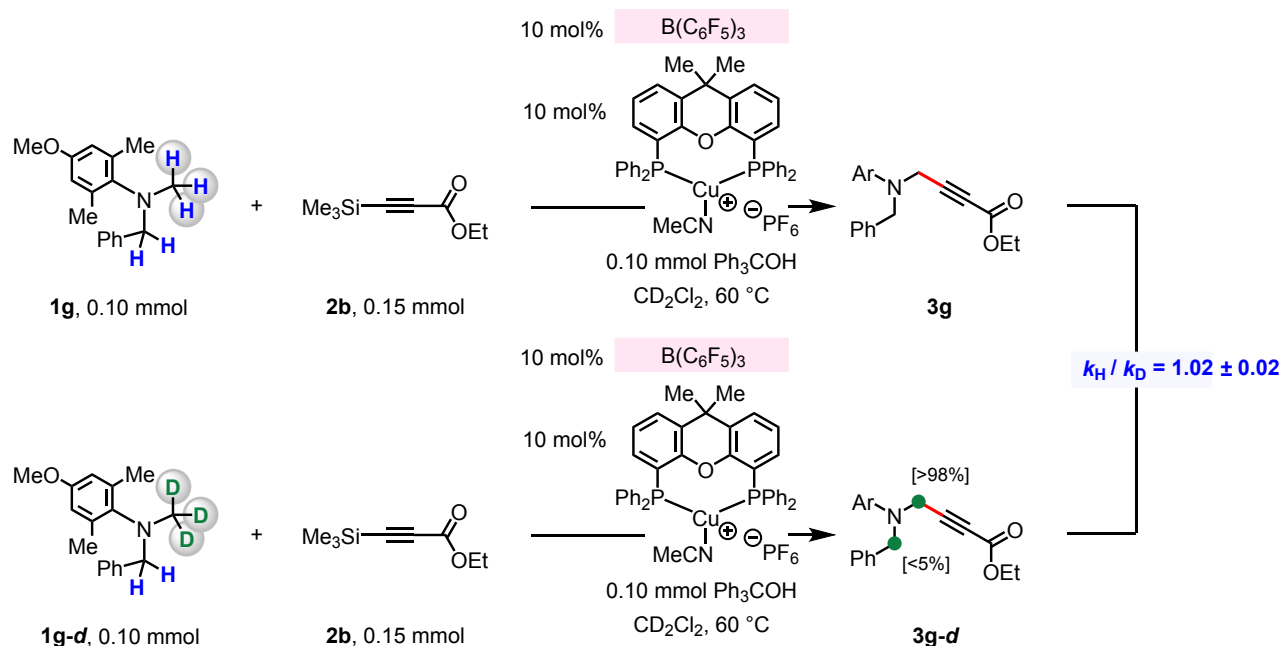


Figure S11. Parallel kinetic isotope effect experiments. The rate of the reaction is unaffected when amine **1g** or **1g-d** are employed as both have similar reaction time course plots. From the set of two parallel KIE experiments, the KIE value of 1.02 ± 0.02 was found.

Experimental Procedure for Measuring the Parallel Kinetic Isotope Effect

To an oven-dried 7.0 mL vial were added (Xantphos)Cu(MeCN)PF₆ (0.01 mmol), ethyl 3-(trimethylsilyl)propiolate **2b** (0.15 mmol), triphenylmethanol (0.10 mmol), amine **1g** (0.10 mmol) or amine **1g-d** (0.10 mmol), B(C₆F₅)₃ (0.01 mmol, 10 mol%), mesitylene (0.10 mmol). This mixture was then diluted to 0.40 mL with CD₂Cl₂ and transferred into a J-Young tube. After the J-Young tube was tightly capped with the Teflon plug, it was taken out of the glove box and ¹H NMR spectra were acquired at 60 °C (preheated) using a pre-acquisition delay in array mode with a spectrum taken every 30 seconds for the length of the experiment. The data were processed using MestReNova and peak integrations were normalized using mesitylene as the internal standard.

5.2.2 Intermolecular Competition Kinetic Isotope Effect Experiment

An intermolecular competition kinetic isotope effect study was conducted between *N*-benzyl-4-methoxy-*N*,2,6-trimethylaniline **1g** and *N*-benzyl-4-methoxy-2,6-dimethyl-*N*-(methyl-*d*₃)aniline **1g-d**. Upon analysis of the unpurified reaction mixture by ¹H NMR spectroscopy, 27% conversion to both **3g** and **3g-d** was detected where 22% conversion of **3g** was detected. This ratio was further verified by studying the ²H NMR spectrum; it revealed that approximately 5% of **3g-d** is formed.

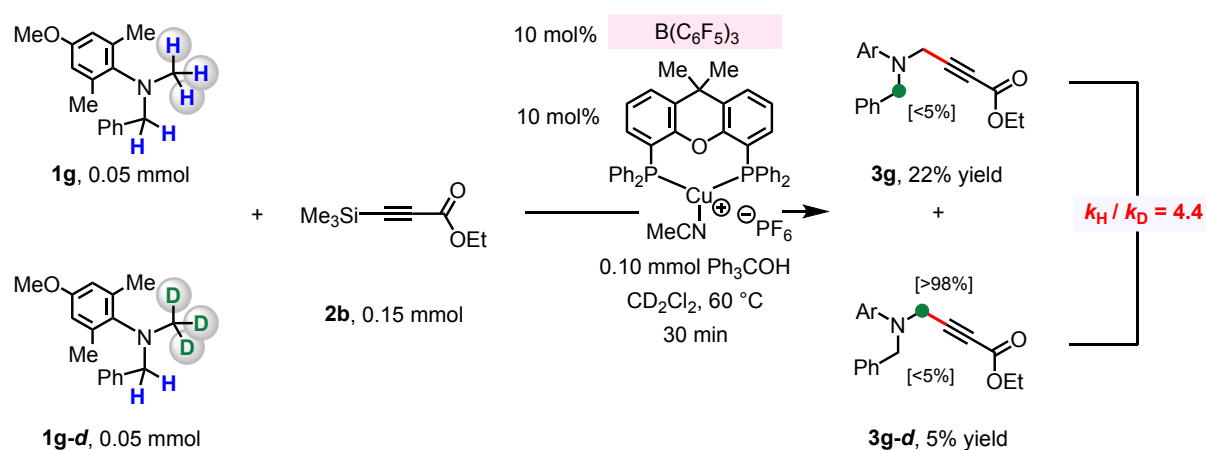


Figure S12. Intermolecular Competition Kinetic Isotope Effect Experiment. The result suggests hydride abstraction step is irreversible.

Experimental Procedure for Measuring the Intermolecular Competition Kinetic Isotope Effect

An oven-dried 7.0 mL vial equipped with a magnetic stir bar was used. To the vial were added (Xantphos)Cu(MeCN)PF₆ (0.01 mmol), ethyl 3-(trimethylsilyl)propiolate **2b** (0.15 mmol), triphenylmethanol (0.10 mmol), amine **1g** (0.05 mmol), amine **1g-d** (0.05 mmol), B(C₆F₅)₃ (0.01 mmol, 10 mol%), mesitylene (0.10 mmol), benzene-*d*₆ (0.05 mmol), and CD₂Cl₂ (0.40 mL). The reaction mixture was then transferred to a J-Young tube and was allowed to heat at 60 °C for 2 h. After 2 h, the reaction mixture was allowed to cool and NMR spectroscopy was obtained. The conversion values were determined by ¹H and ²H NMR (Figure S13) analysis of the unpurified reaction mixture using mesitylene and benzene-*d*₆ as internal standards.

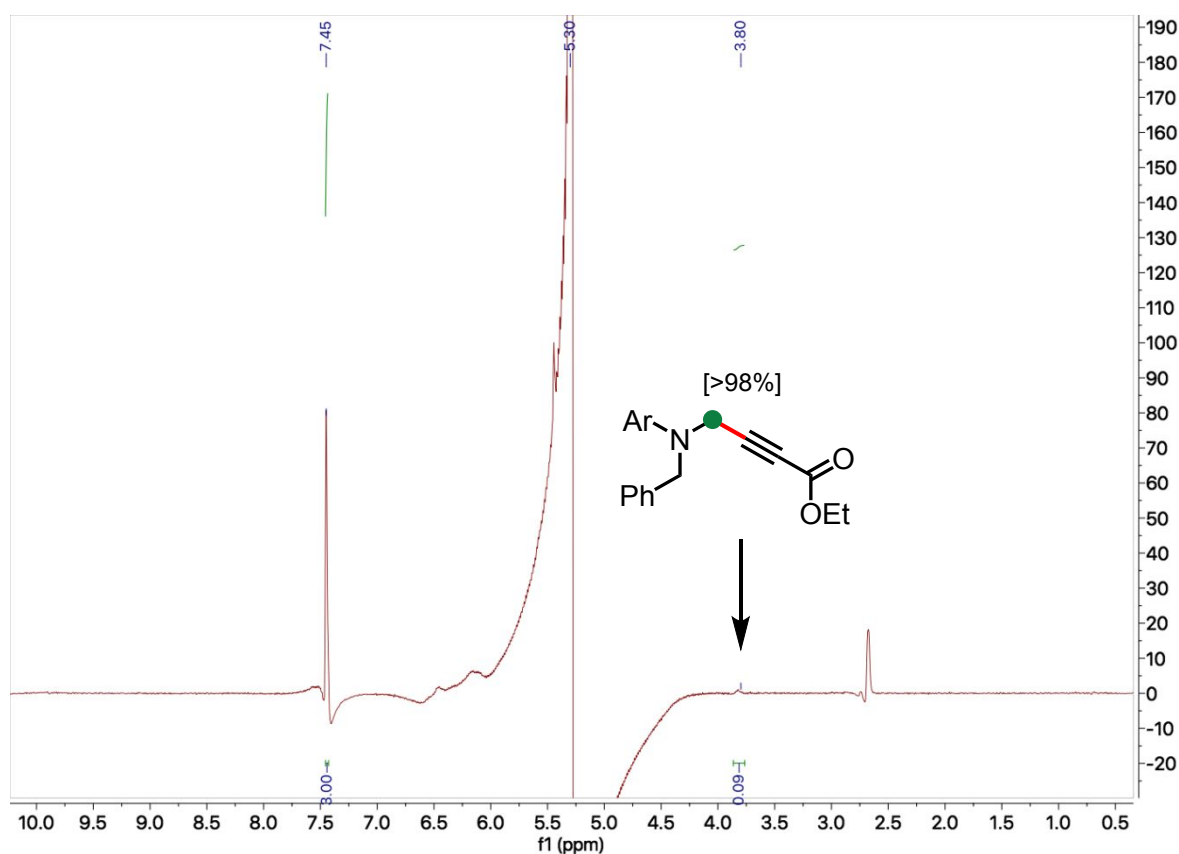


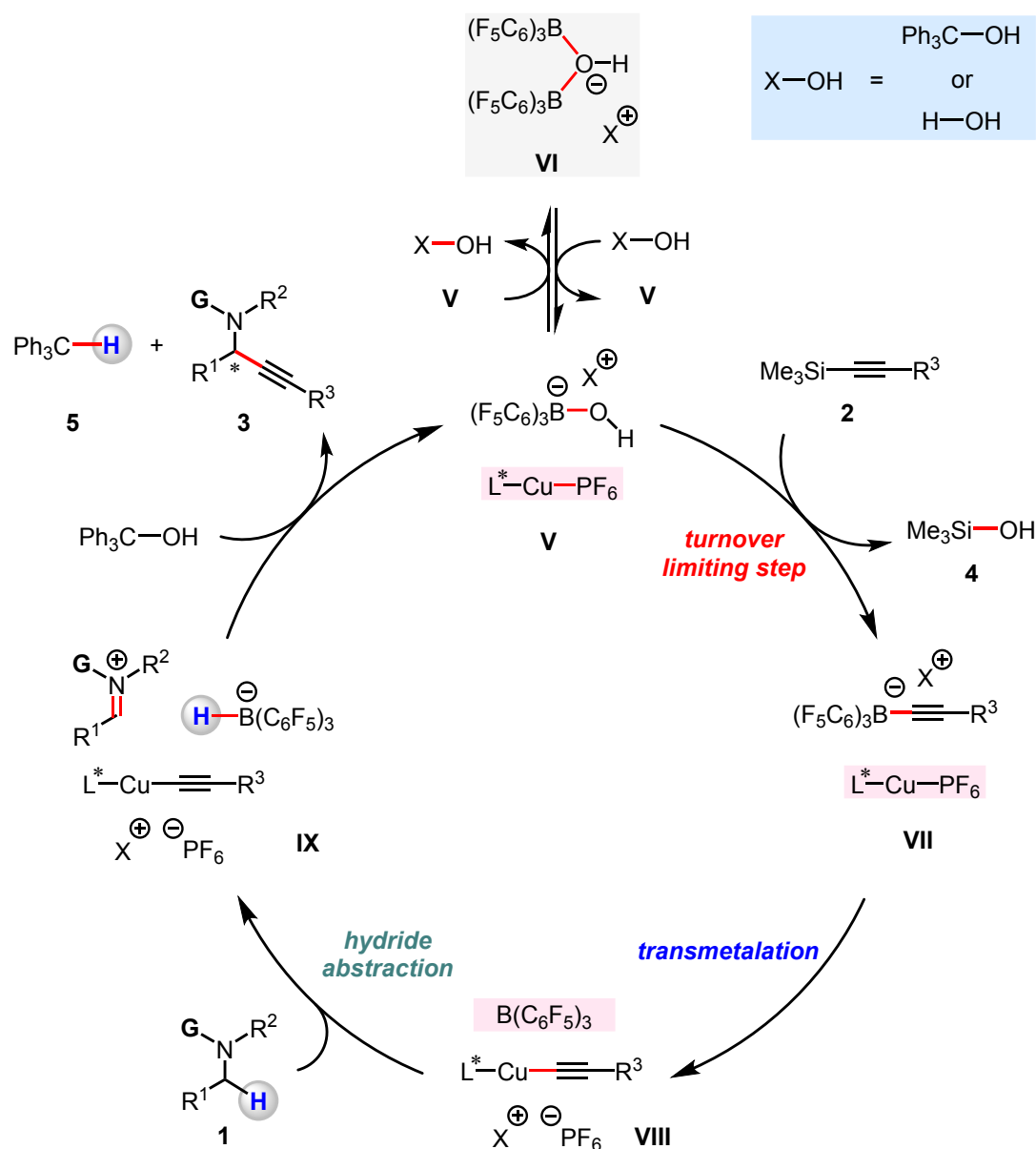
Figure S13: ²H NMR spectrum for intermolecular competition kinetic isotope effect study.

5.3 NMR Experiments for Detection of Proposed Intermediates and the Resting State

5.3.1 Demonstration of Chemical Competency of the Proposed Intermediate VII

Based on the kinetic studies as described above, we propose that borate anion **V** (Scheme S2) reacts with trimethylsilylacetylene **2** to afford $[(F_5C_6)_3B\text{-alkyne}]^- [X]^+$ (**VII**). To demonstrate the competency of **VII** as an intermediate to afford the propargylamine product **3**, we prepared a sample of $[(F_5C_6)_3B\text{-C}\equiv\text{C-CO}_2\text{Et}]^- [\text{H-NR}_3]^+$ ($\text{NR}_3 = \mathbf{1e}$) following a procedure reported previously in the literature.¹⁶

Scheme S2. Proposed Mechanism



Experimental Procedure for the Preparation of $[(F_5C_6)_3B-C\equiv C-CO_2Et]^- [H-1e]^+$

An oven-dried 7.0 mL vial equipped with a magnetic stir bar was used. To the vial were added ethyl propiolate (0.10 mmol), amine **1e** (0.15 mmol), $B(C_6F_5)_3$ (0.10 mmol, 100 mol%), mesitylene (0.10 mmol), trifluorotoluene (0.10 mmol) and CD_2Cl_2 (0.60 mL) under nitrogen atmosphere. The mixture was then transferred to a J-Young tube and NMR spectroscopy was obtained. Analysis of the 1H NMR spectra of unpurified mixture with mesitylene as the internal standard revealed that ethyl propiolate was fully consumed (Figure S14) and $[(F_5C_6)_3B-C\equiv C-CO_2Et]^- [H-1e]^+$ (**VII**) was formed (Figure S15) which was indicated by a characteristic sharp singlet at -21.5 ppm on ^{11}B NMR (Figure S16).¹⁶

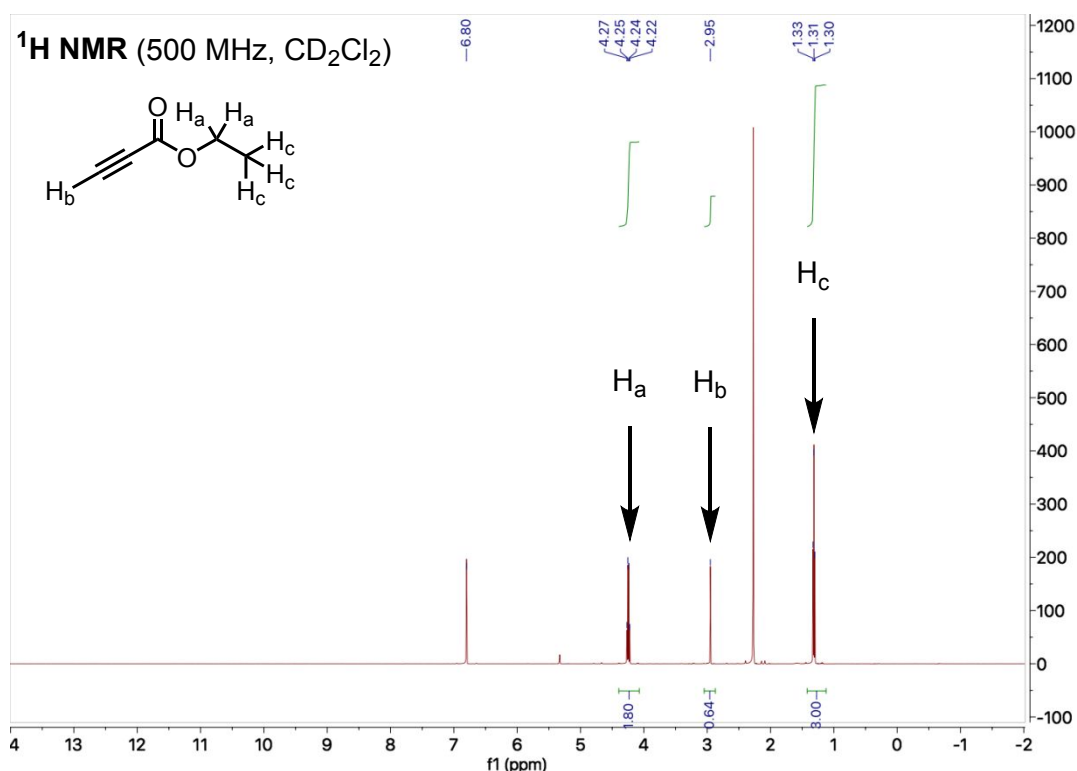


Figure S14. 1H NMR spectrum of ethyl propiolate.

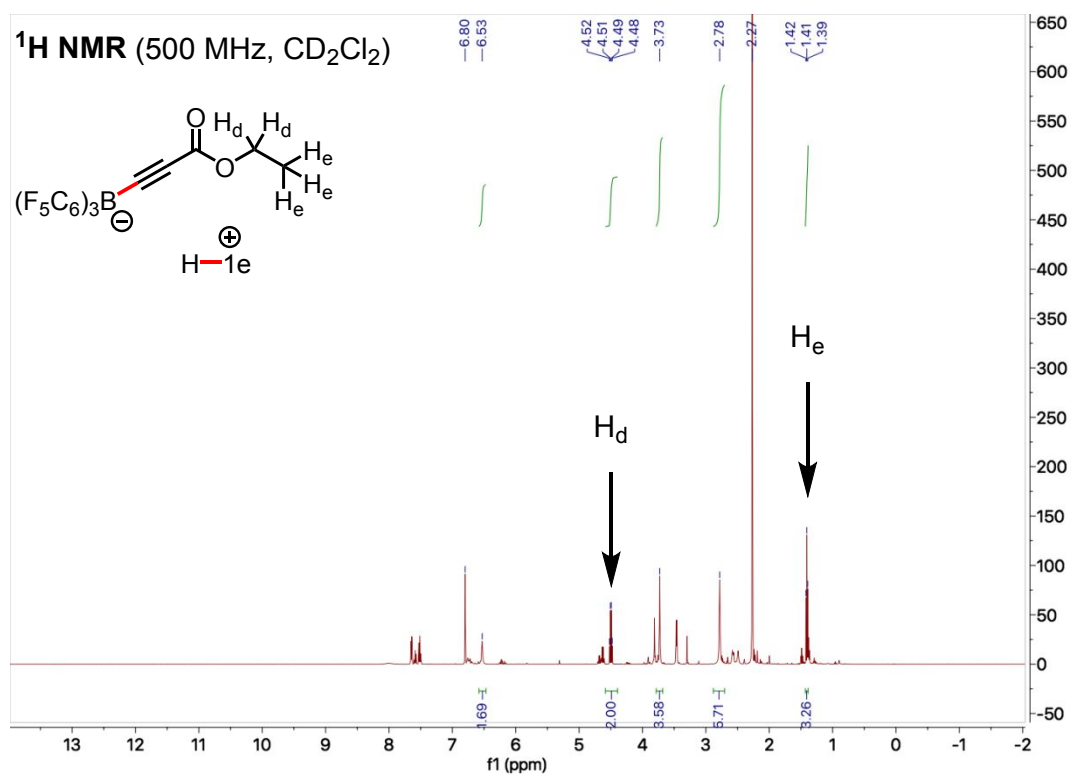


Figure S15. ^1H NMR spectrum of unpurified reaction mixture of $[(\text{F}_5\text{C}_6)_3\text{B}-\text{C}\equiv\text{C}-\text{CO}_2\text{Et}][\text{H}-1\text{e}]^+$.

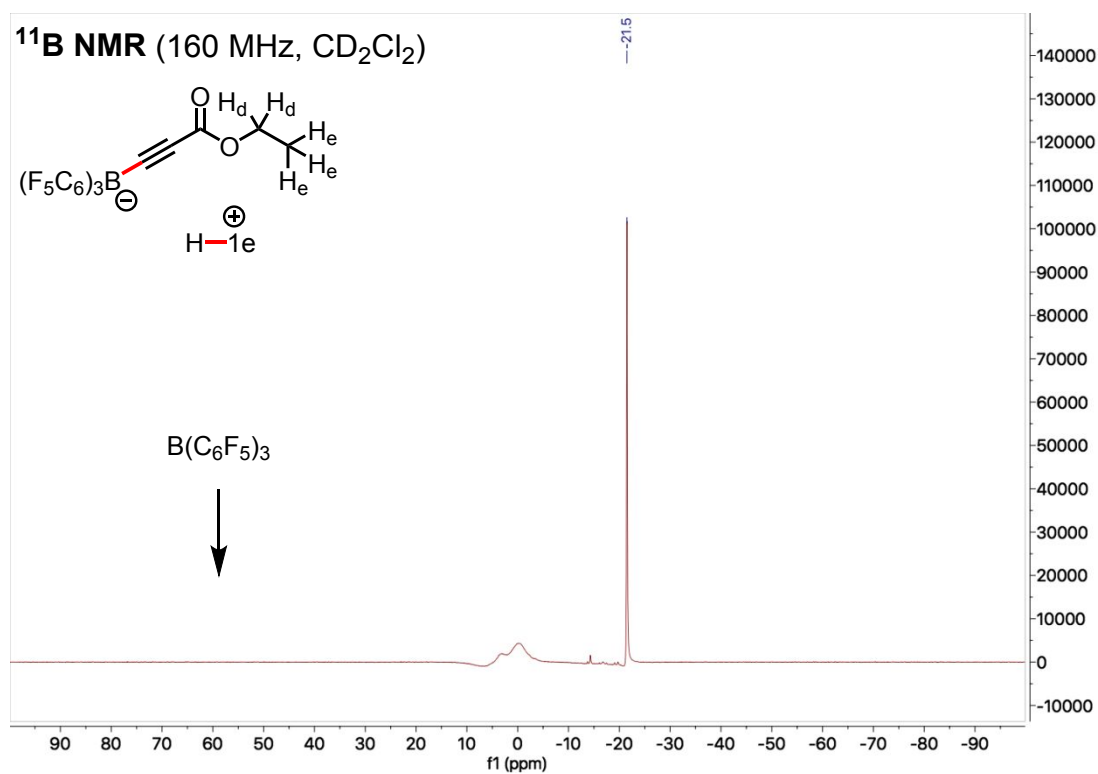


Figure S16. ^{11}B NMR spectrum of unpurified reaction mixture of $[(\text{F}_5\text{C}_6)_3\text{B}-\text{C}\equiv\text{C}-\text{CO}_2\text{Et}][\text{H}-1\text{e}]^+$.

To this unpurified reaction mixture was added (Xantphos)Cu(MeCN)PF₆ (0.10 mmol). The reaction mixture was allowed to stir at 60 °C for 1 h. Then, the reaction mixture was cooled down to 22 °C and a ¹H NMR spectrum was obtained (Figure 17). Analysis of the ¹H spectrum of unpurified mixtures with mesitylene as the internal standard revealed that **3e** was formed in 24% under this reaction conditions, thereby demonstrating competency of the proposed intermediate **VII** in the alkyne incorporation process.

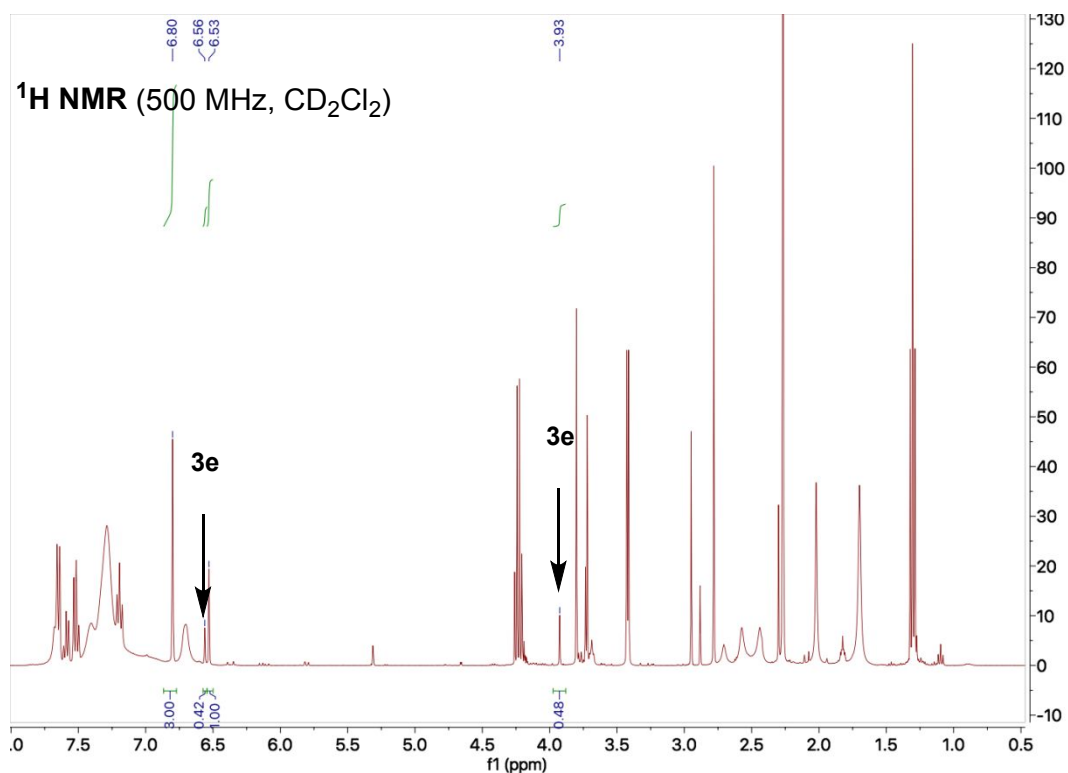
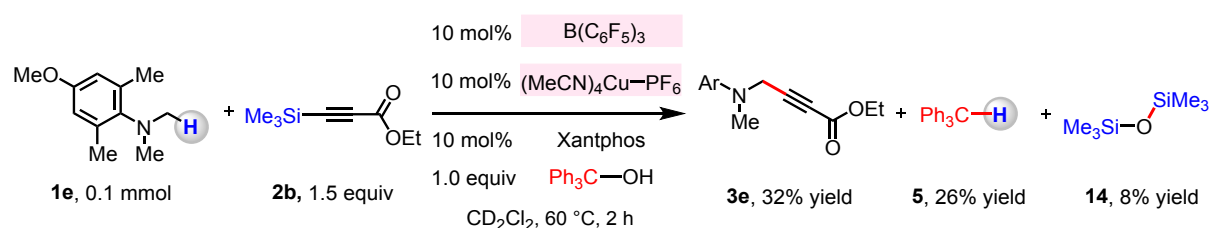


Figure S17. ¹H NMR spectrum of unpurified reaction mixture of **VII** and 100 mol % of (Xantphos)Cu(MeCN)PF₆.

5.3.2 NMR Experiments for Detection of Byproducts

The reaction between 4-methoxy-*N,N*,2,6-tetramethylaniline **1e** and ethyl 3-(trimethylsilyl)propiolate **2b** was monitored by ¹H NMR spectroscopy (Figure S18–S20). This study revealed that Ph₃C–H **5** and Me₃Si–O–SiMe₃ **14** are the stable byproducts of this α -alkynylation reaction (Scheme S2 and S3).

Scheme S3. Formation of Stable Byproducts **5** and **14**



Experimental Procedure for the α -Alkynylation of 4-Methoxy-*N,N*,2,6-tetramethylaniline and Ethyl 3-(trimethylsilyl)propiolate

An oven-dried 7.0 mL vial equipped with a magnetic stir bar was used. To the vial were added Cu(MeCN)₄PF₆ (0.010 mmol), Xantphos (0.01 mmol), and CD₂Cl₂ (0.2 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then ethyl 3-(trimethylsilyl)propiolate **2b** (0.15 mmol), triphenylmethanol (0.1 mmol), amine **1e** (0.10 mmol), mesitylene (0.10 mmol), B(C₆F₅)₃ (0.01 mmol, 10 mol%), and CD₂Cl₂ (0.20 mL) were added to the vial. The reaction mixture was then transferred to a J-Young tube and was allowed to heat at 60 °C for 1 h. After 1 h, the reaction mixture was allowed to cool and NMR spectroscopy was obtained. Upon analysis of the ¹H NMR of the unpurified reaction mixture, triphenylmethane **5** and 1,1,1,3,3,3-hexamethyldisiloxane **14** was detected (Figure S18).

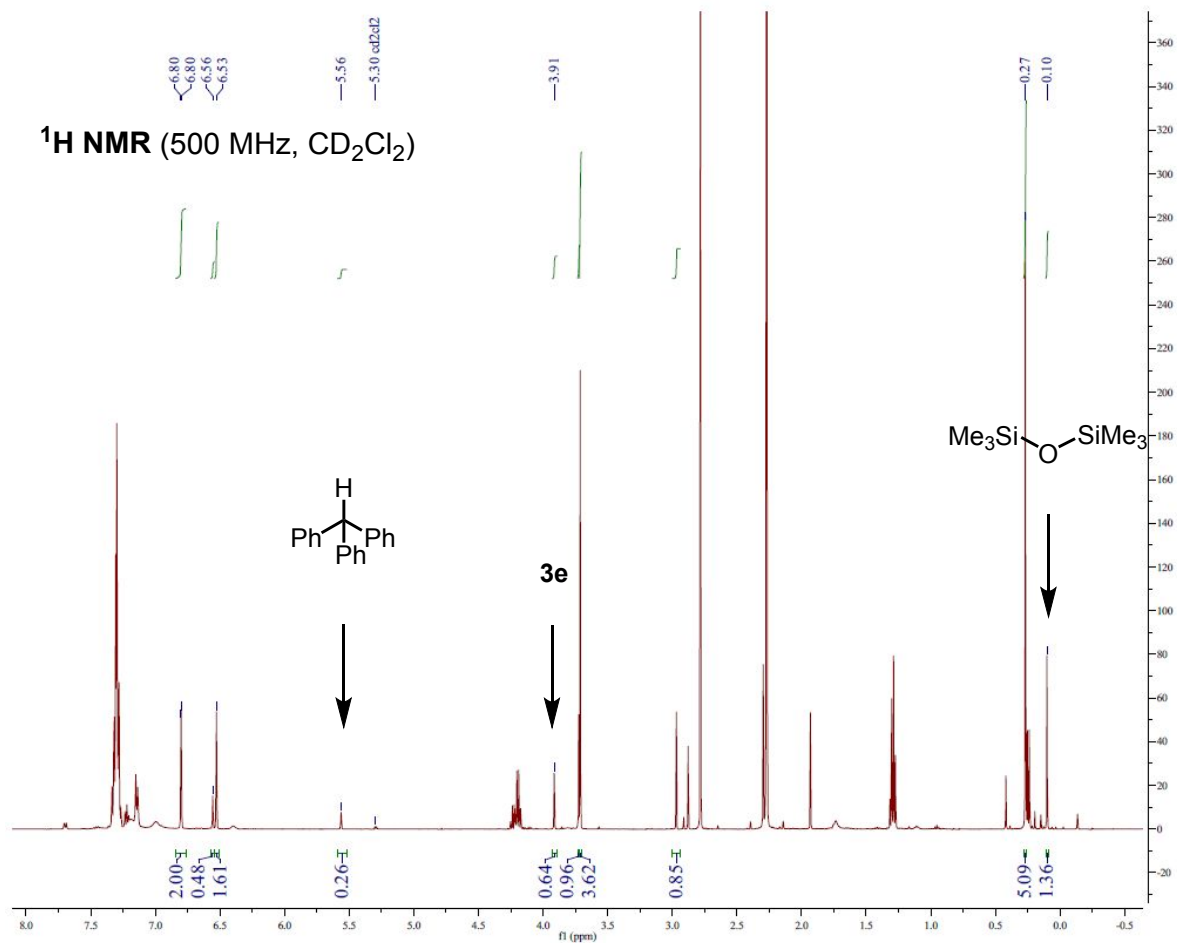


Figure S18. ¹H NMR spectrum of unpurified reaction mixture of **1e** and **2b** under α -alkynylation condition.

To this unpurified reaction mixture was added Me₃Si–O–SiMe₃ **14** obtained from a commercial source (Figure S19).

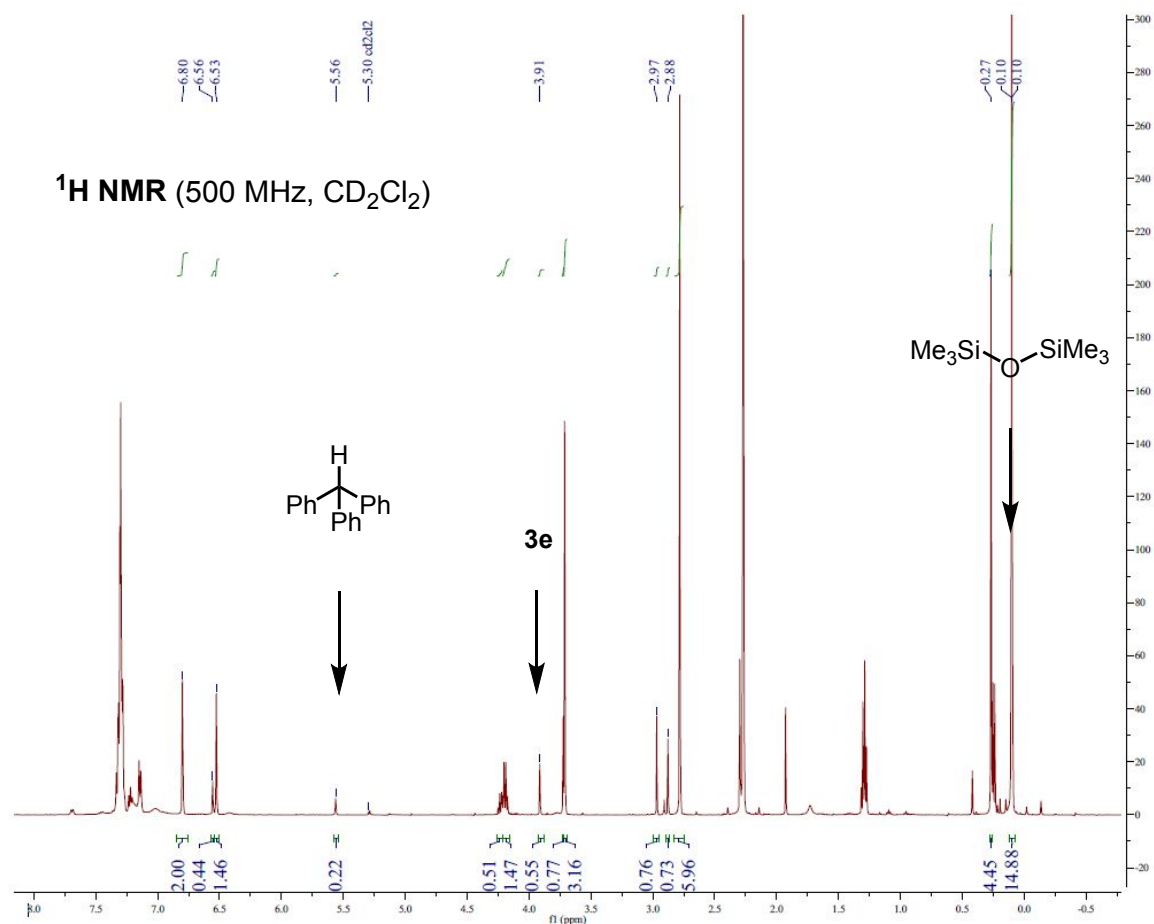


Figure S19. ¹H NMR spectrum of the unpurified reaction mixture of **1e** and **2b** under α -alkynylation conditions, where Me₃Si–O–SiMe₃ **14** was added to the cooled reaction mixture after 1 h.

In order to determine the fate of the hydride abstracted from the amine substrates by B(C₆F₅)₃, we carried out the transformation between 4-methoxy-2,6-dimethyl-*N,N*-bis(methyl-*d*₃)aniline **1e-d** and ethyl 3-(trimethylsilyl)propiolate **2b** in the presence of Ph₃C–OH (Figure 20). Upon analysis of the ¹H NMR of the unpurified product mixture, it was found that deuteride is trapped by in situ generated Ph₃C⁺ to produce (methanetriyl-*d*)tribenzene **5-d** (Figure S20). The yield of **3e-d**, **5-d** and **14** were determined based on the amount of mesitylene added as an internal standard.

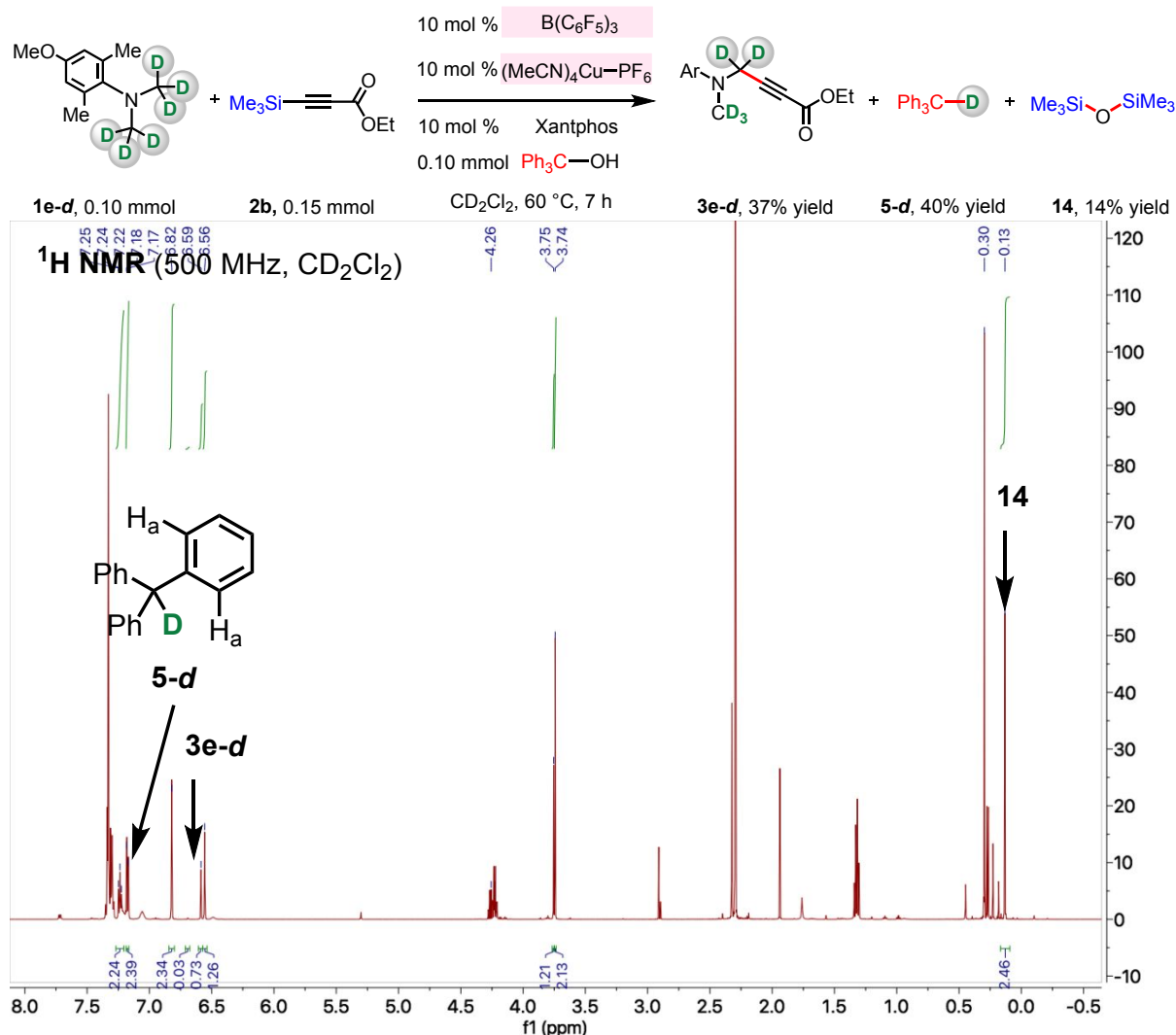


Figure S20. ^1H NMR spectrum of unpurified reaction mixture of **1e-d** and **2b** under α -alkynylation conditions.

5.3.3 NMR Experiments for the Detection of the Resting State Complex Containing $\text{B}(\text{C}_6\text{F}_5)_3$

We embarked on a study to identify the structure of a resting state complex which contains $\text{B}(\text{C}_6\text{F}_5)_3$. Previously, the groups of Pinkas and Resconi have independently reported that a borate anion $[(\text{C}_6\text{F}_5)_3\text{B}(\mu\text{-OH})\text{B}(\text{C}_6\text{F}_5)_3]^-$ can be formed through the reaction of $\text{B}(\text{C}_6\text{F}_5)_3$ and Ph_3COH ,¹⁷ and by reacting $\text{B}(\text{C}_6\text{F}_5)_3$, H_2O and an amine.¹⁸ Since the $\text{B}(\text{C}_6\text{F}_5)_3$ /organocopper co-catalyzed transformation of C–H bonds was found to have a 0.5 order dependency with respect to the concentration of $\text{B}(\text{C}_6\text{F}_5)_3$, we surmised that $[(\text{C}_6\text{F}_5)_3\text{B}(\mu\text{-OH})\text{B}(\text{C}_6\text{F}_5)_3]^-$ containing two molecules of $\text{B}(\text{C}_6\text{F}_5)_3$ could be its resting state.

We first acquired a ^1H NMR spectrum of a sample containing $\text{B}(\text{C}_6\text{F}_5)_3$ and Ph_3COH in CD_2Cl_2 (Figure S21) and compared that to the spectra reported by Pinkas.¹⁷ Based on this analysis, we concluded that $[(\text{F}_5\text{C}_6)_3\text{B}(\mu\text{-OH})\text{B}(\text{C}_6\text{F}_5)_3]^-[\text{CPh}_3]^+$ is generated.

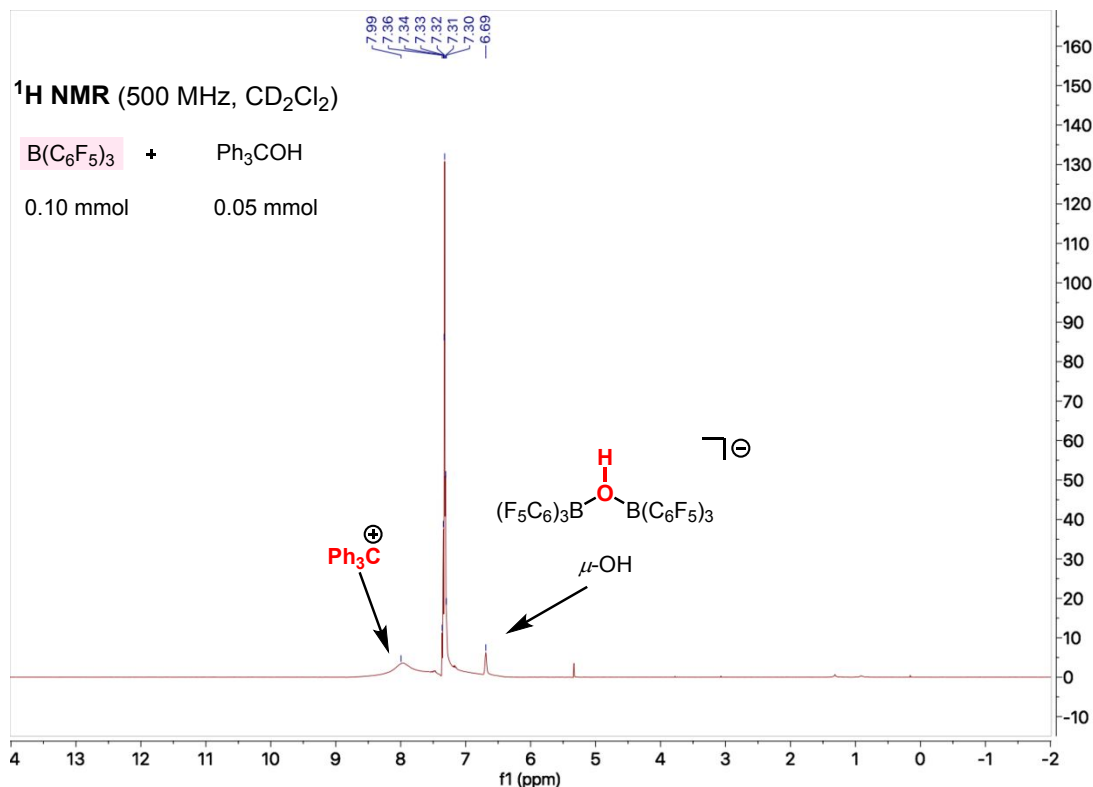


Figure S21. ^1H NMR of $\text{B}(\text{C}_6\text{F}_5)_3$ and Ph_3COH in CD_2Cl_2 .

Next, we obtained the ^{11}B and/or ^{19}F NMR spectra of:

Figures S22–S23: the sample containing $[(\text{F}_5\text{C}_6)_3\text{B}(\mu\text{-OH})\text{B}(\text{C}_6\text{F}_5)_3]^-[\text{CPh}_3]^+$ prepared as described above,

and compared those to the spectra we obtained for:

Figures S24–25: $\text{B}(\text{C}_6\text{F}_5)_3$ only

Figures S26–27: a mixture of $\text{B}(\text{C}_6\text{F}_5)_3$, $[\text{Cu}(\text{Xantphos})(\text{MeCN})][\text{PF}_6]$, 4-methoxy-*N,N*,2,6-tetramethylaniline **1e**, 3-(triethylsilyl)propiolate **2b**, Ph_3COH in CD_2Cl_2 at 22 °C, and

Figure S28: a mixture of $\text{B}(\text{C}_6\text{F}_5)_3$, $[\text{Cu}(\text{Xantphos})(\text{MeCN})][\text{PF}_6]$, 4-methoxy-*N,N*,2,6-tetramethylaniline **1e**, 3-(triethylsilyl)propiolate **2b**, Ph_3COH in CD_2Cl_2 at 60 °C.

The analyses of Figures S26–S28 revealed that there is no free $\text{B}(\text{C}_6\text{F}_5)_3$ remaining in the reaction mixture (characteristic peak at 58.7 ppm corresponding to $\text{B}(\text{C}_6\text{F}_5)_3$ in Figure 24 is not observed in Figure S26).

The comparison of ^{19}F NMR spectra for the authentic sample of $[(\text{F}_5\text{C}_6)_3\text{B}(\mu\text{-OH})\text{B}(\text{C}_6\text{F}_5)_3]^-$

$[\text{CPh}_3]^+$ (Figure 23) with those acquired for the actual reaction mixture (Figures S27 and S28) revealed that there are consistent peaks at -135.94 , -160.88 and -165.53 ppm.

Furthermore, the comparison of ^{11}B NMR spectrum we acquired for the sample of $[(\text{F}_5\text{C}_6)_3\text{B}(\mu\text{-OH})\text{B}(\text{C}_6\text{F}_5)_3][\text{CPh}_3]^+$ (Figure S22) with the one for the reaction mixture (Figure 26) also showed that there are common peaks at 0.8 and -3.7 ppm.

These results serve as evidences to support the formation of $[(\text{F}_5\text{C}_6)_3\text{B}(\mu\text{-OH})\text{B}(\text{C}_6\text{F}_5)_3]^-$ anion under the reaction conditions for $\text{B}(\text{C}_6\text{F}_5)_3/[\text{Cu}(\text{Xantphos})(\text{MeCN})][\text{PF}_6]$ co-catalyzed C–H functionalization.

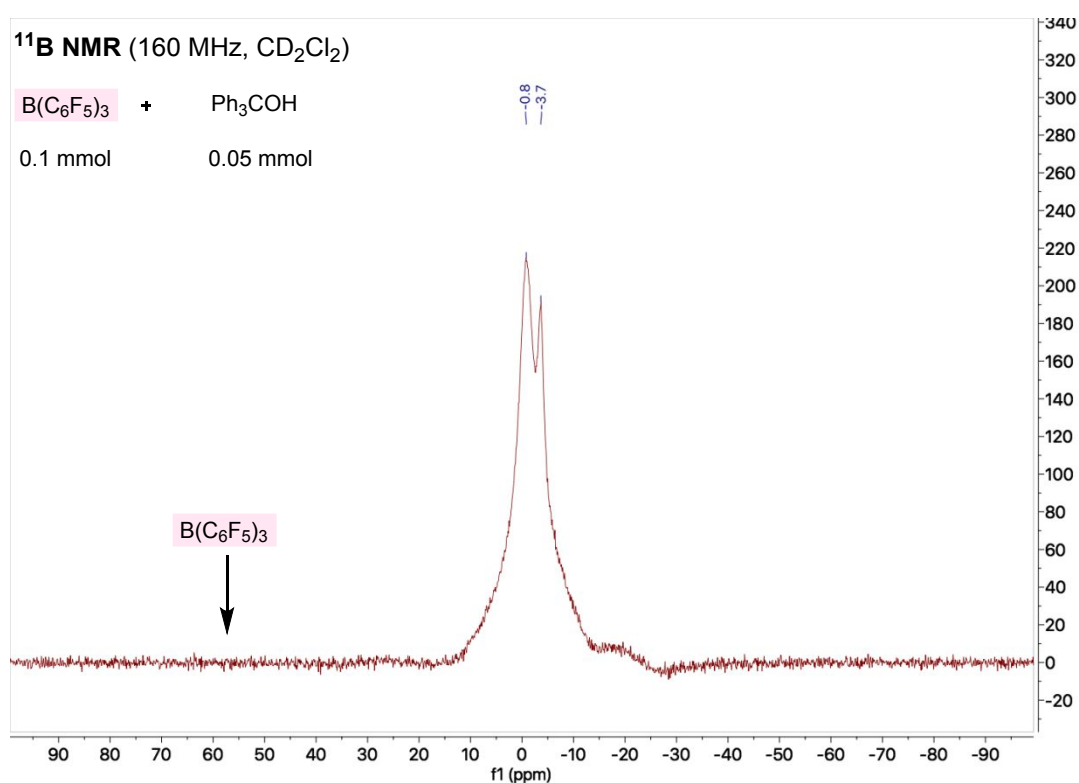


Figure S22. ^{11}B NMR spectrum of a sample containing $\text{B}(\text{C}_6\text{F}_5)_3$ and Ph_3COH in CD_2Cl_2

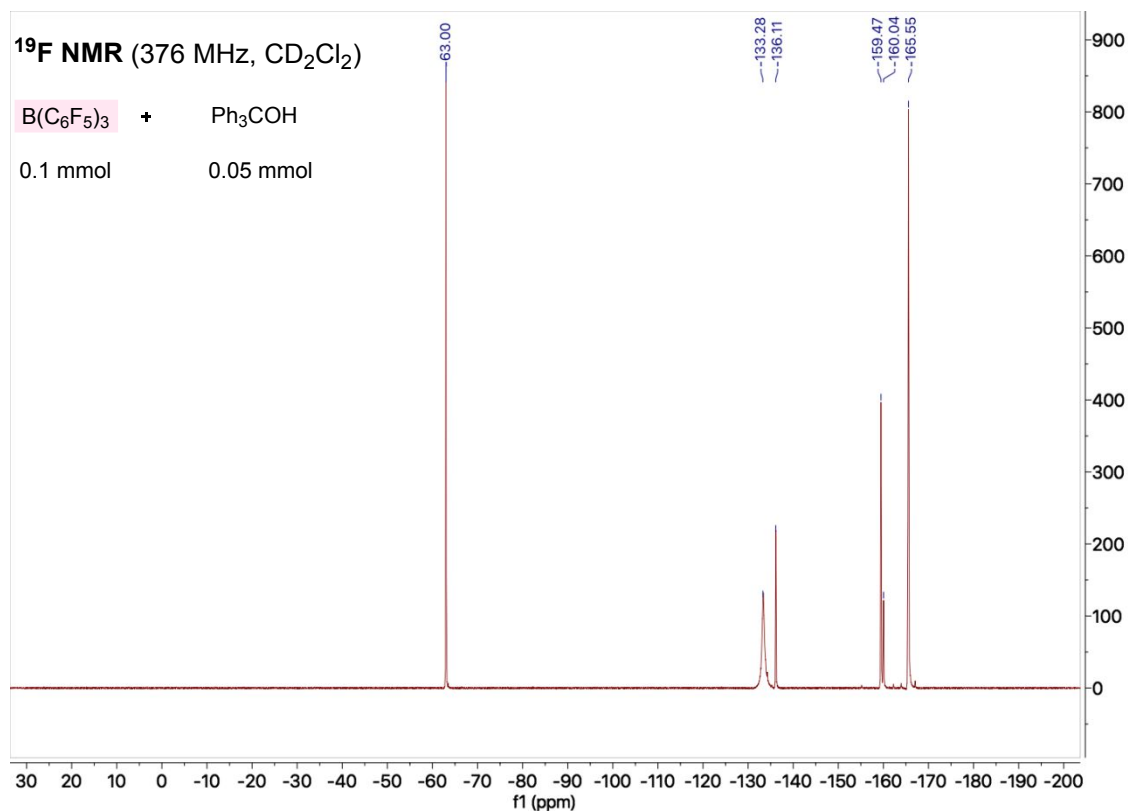


Figure S23. ^{19}F NMR spectrum of a sample containing $\text{B}(\text{C}_6\text{F}_5)_3$ and Ph_3COH in CD_2Cl_2 .

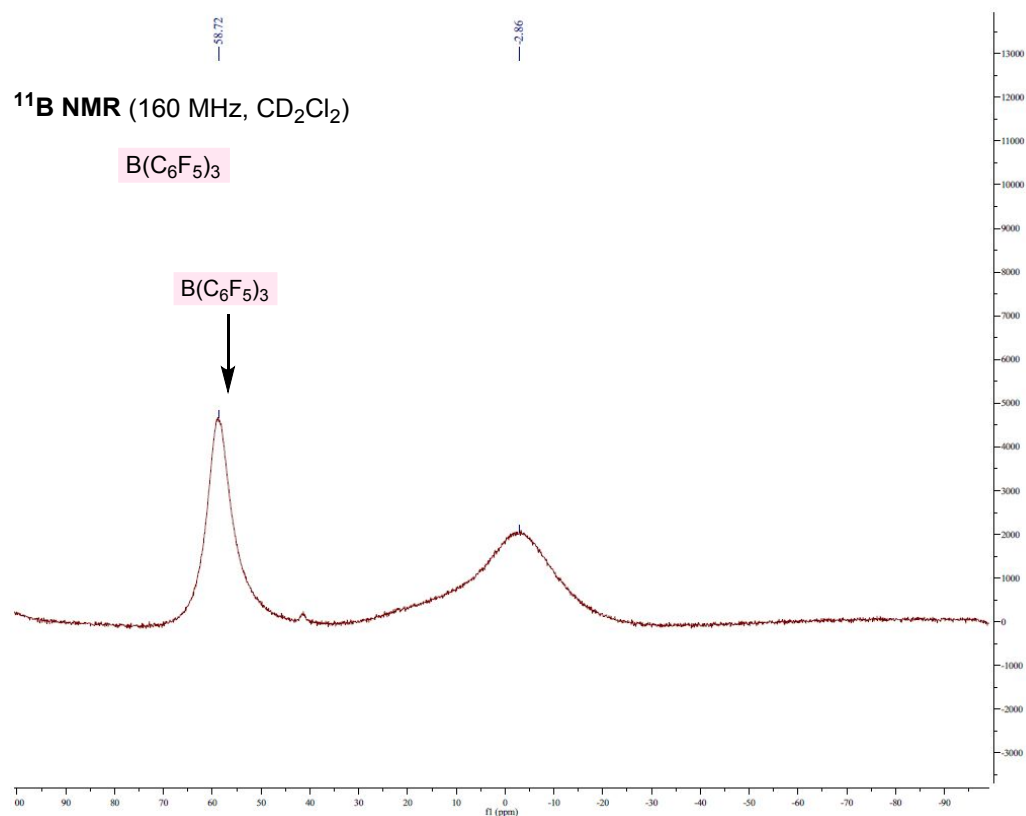


Figure S24. ^{11}B NMR spectrum of $\text{B}(\text{C}_6\text{F}_5)_3$ in CD_2Cl_2 .

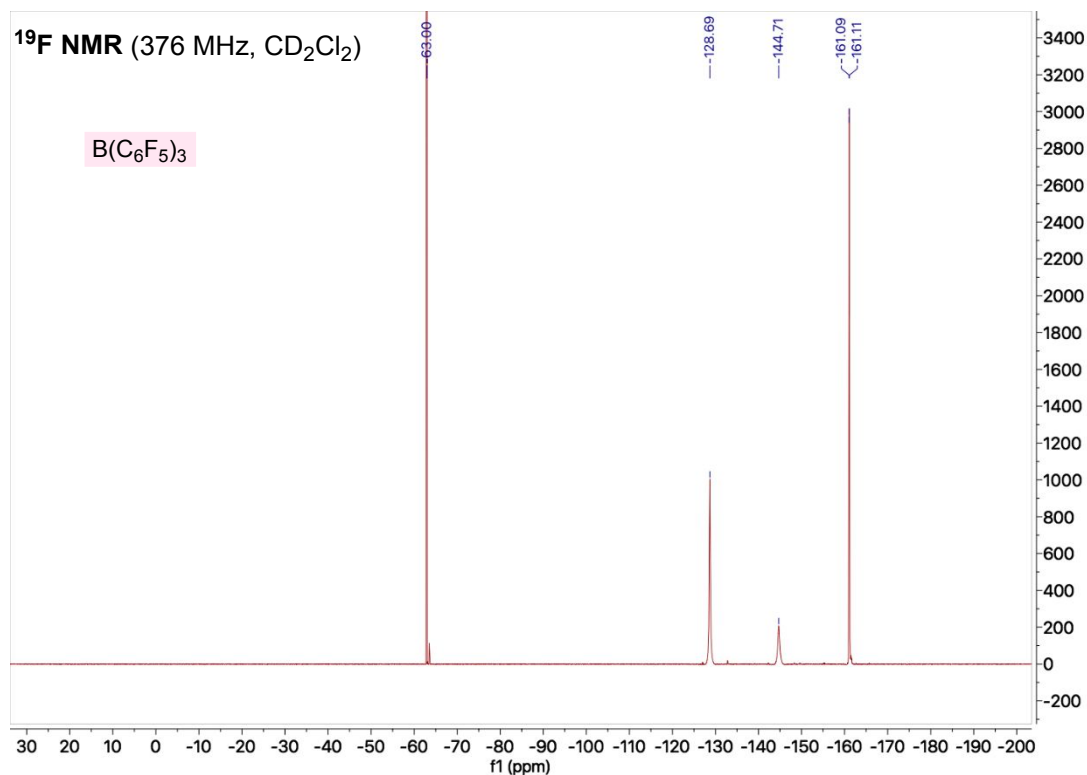


Figure S25. ^{19}F NMR spectrum of $\text{B}(\text{C}_6\text{F}_5)_3$ in CD_2Cl_2 .

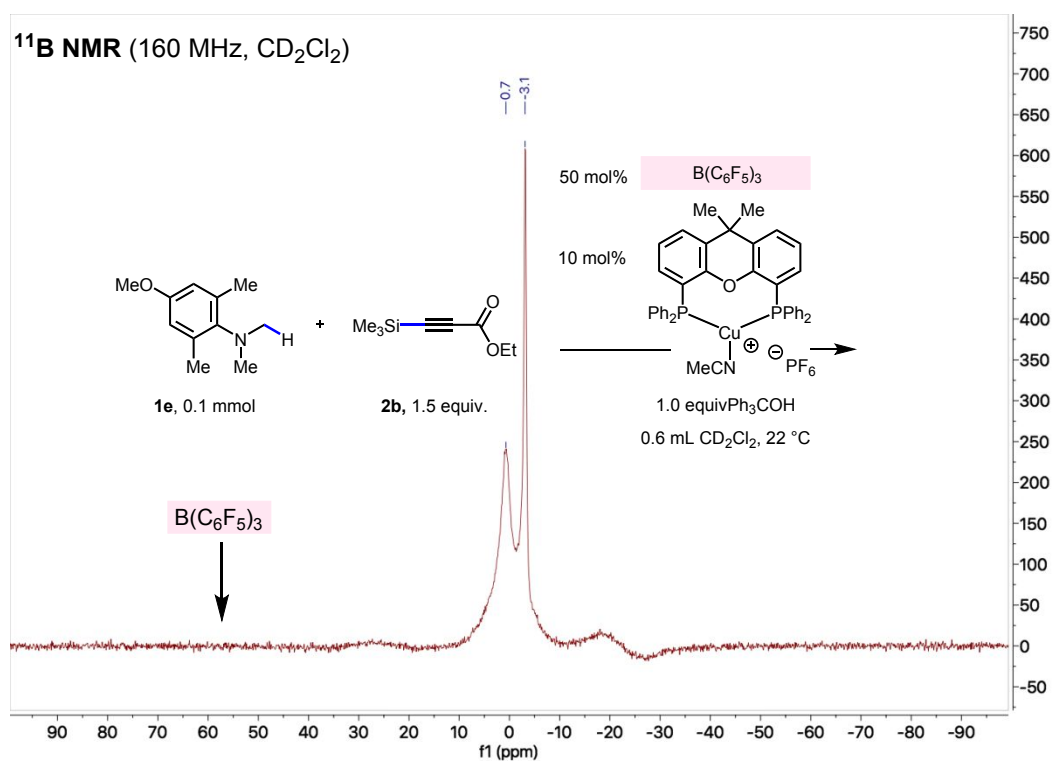


Figure S26. ^{11}B NMR spectrum of α -alkynylation reaction between **1e** and **2b** with 50 mol% $\text{B}(\text{C}_6\text{F}_5)_3$ at 22 °C.

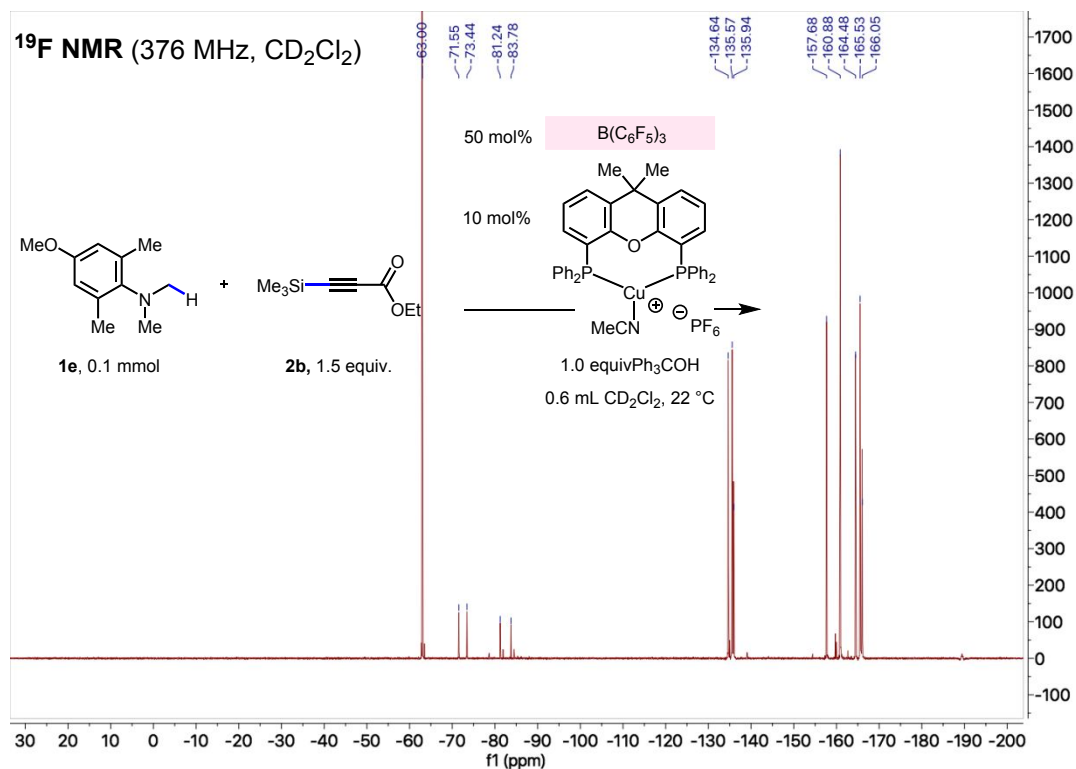


Figure S27. ¹⁹F NMR spectrum of α -alkynylation reaction of amine **1e** with 50 mol% B(C₆F₅)₃ at 22 °C.

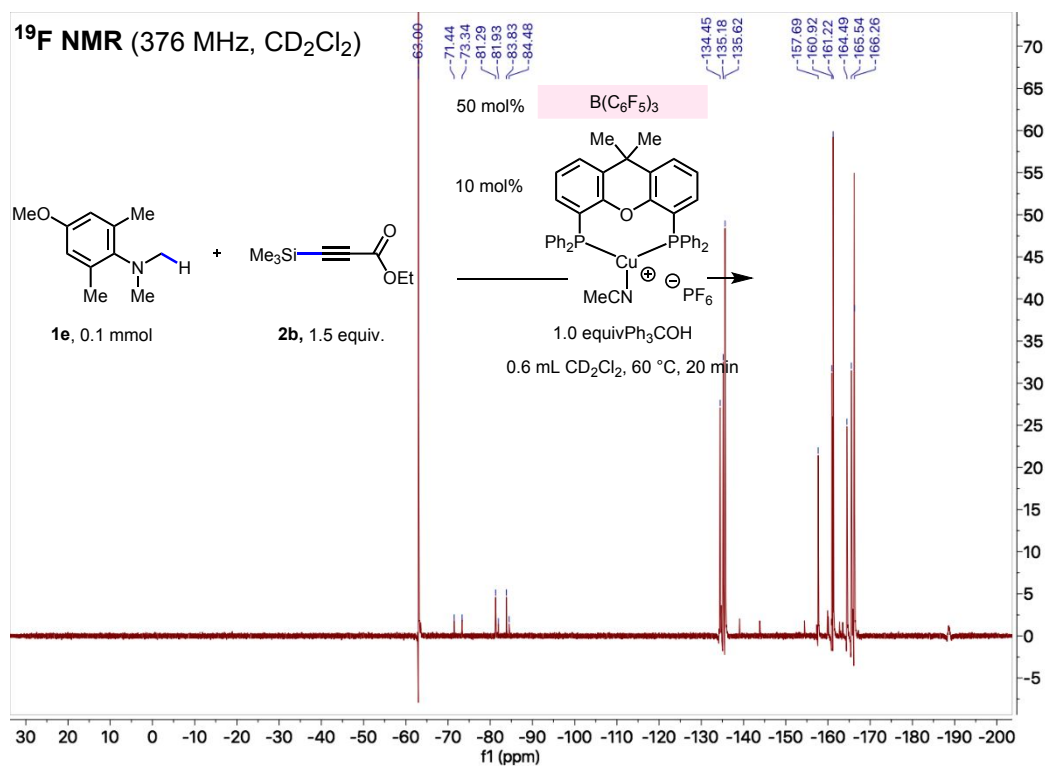


Figure S28. ¹⁹F NMR spectrum of α -alkynylation reaction between **1e** and **2b** using 50 mol% B(C₆F₅)₃ at 22 °C.

As reported by the group of Resconi, the reaction of $\text{B}(\text{C}_6\text{F}_5)_3$ and H_2O may result in the formation of $[(\text{F}_5\text{C}_6)_3\text{B}(\mu\text{-OH})\text{B}(\text{C}_6\text{F}_5)_3]^- [\text{H-NR}_3]^+$.¹⁸ To probe if this can occur under our reaction conditions, we acquired the following spectra:

Figures S29: ^{11}B spectrum for a mixture of $\text{B}(\text{C}_6\text{F}_5)_3$, H_2O and **1e**, and

Figures S26: ^{11}B NMR spectra for a mixture of $\text{B}(\text{C}_6\text{F}_5)_3$, $[\text{Cu}(\text{Xantphos})(\text{MeCN})][\text{PF}_6]$, 4-methoxy-*N,N*,2,6-tetramethylaniline **1e**, 3-(triethylsilyl)propionate **2b**, Ph_3COH in CD_2Cl_2 at 22 °C.

Resconi *et al.*¹⁸ reported that ^{11}B NMR spectrum for $[(\text{F}_5\text{C}_6)_3\text{B}(\mu\text{-OH})\text{B}(\text{C}_6\text{F}_5)_3]^-$ contains a characteristic singlet peak at -1 ppm (298 K). The ^{11}B NMR spectra we acquired (Figures S26 and S29) both possess a singlet at -3.7 ppm, therefore suggesting that the formation of $[(\text{F}_5\text{C}_6)_3\text{B}(\mu\text{-OH})\text{B}(\text{C}_6\text{F}_5)_3]^-$ anion is possible.

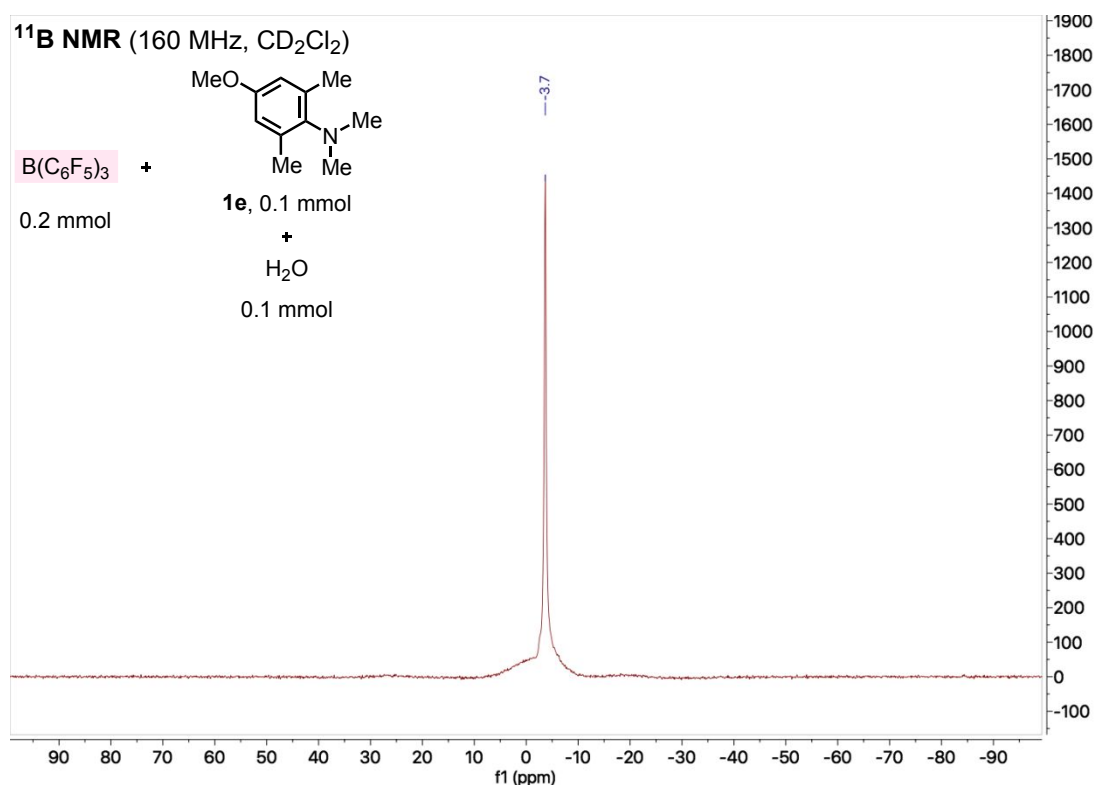


Figure S29. ^{11}B NMR spectrum of a sample containing $\text{B}(\text{C}_6\text{F}_5)_3$, H_2O and amine **1e** in CD_2Cl_2 .

As reported by the group of Basset,¹⁹ a tertiary amine and $\text{B}(\text{C}_6\text{F}_5)_3$ could form an ionic complex containing an iminium ion and $[(\text{F}_5\text{C}_6)_3\text{B-H}]^-$. In the ^{11}B NMR spectrum, $[(\text{F}_5\text{C}_6)_3\text{B-H}]^-$ has been reported to have a characteristic peak at -23.6 ppm.¹⁹ To probe if the related ion pairs are formed in our system, we prepared the following sample and obtained their ^1H , ^{11}B and ^{19}F NMR spectra:

Figures S30-32: a reaction mixture containing 0.1 mmol $\text{B}(\text{C}_6\text{F}_5)_3$ and 0.1 mmol of 4-methoxy-*N,N*,2,6-tetramethylaniline **1e** in CD_2Cl_2 .

Figure S33: 4-methoxy-*N,N*,2,6-tetramethylaniline **1e** in CD_2Cl_2 only.

The comparison of the ^1H NMR spectra (Figure S30 versus Figure 33) suggests that, even in the presence of a stoichiometric quantity of $\text{B}(\text{C}_6\text{F}_5)_3$, amine **1e** is recovered in full and that the formation of corresponding iminium ion cannot be detected.

The analysis of the ^{11}B NMR spectrum (Figure S31) and its comparison to S24 (standard spectrum for $\text{B}(\text{C}_6\text{F}_5)_3$ only) indicates that there is free $\text{B}(\text{C}_6\text{F}_5)_3$ (characteristic peaks at 59.5 ppm was observed in both Figures S24 and S31). In addition, generation of a small quantity of $\text{B}(\text{C}_6\text{F}_5)_3 \cdot \mathbf{1e}$ adduct was observed (characteristic peak at -1.2 ppm was found in S31).

In agreement with the observations mentioned above, the ^{19}F NMR spectrum (Figure S32) contains peaks corresponding to both free $\text{B}(\text{C}_6\text{F}_5)_3$ and the adduct $\text{B}(\text{C}_6\text{F}_5)_3 \cdot \mathbf{1e}$.

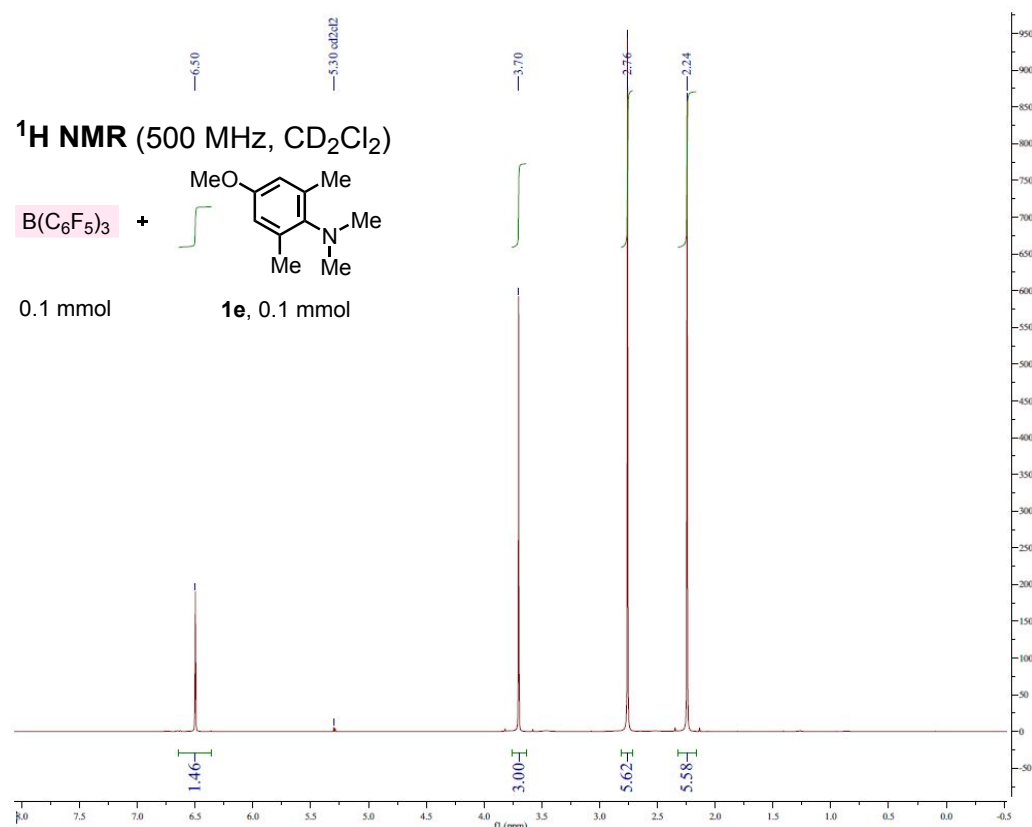


Figure S30. ^1H NMR of spectrum of a sample containing $\text{B}(\text{C}_6\text{F}_5)_3$ and amine **1e** in CD_2Cl_2 .

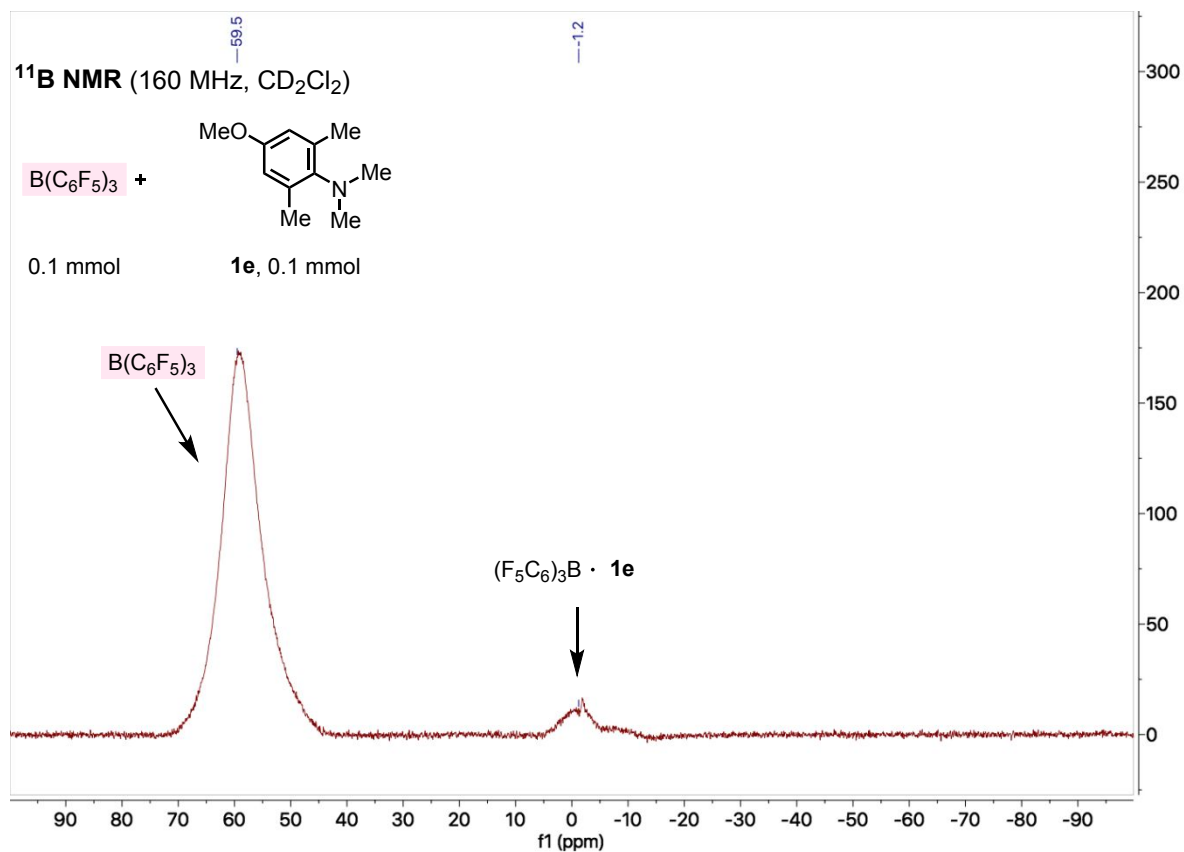


Figure S31. ^{11}B NMR spectrum of a sample containing $\text{B}(\text{C}_6\text{F}_5)_3$ and amine **1e** in CD_2Cl_2 .

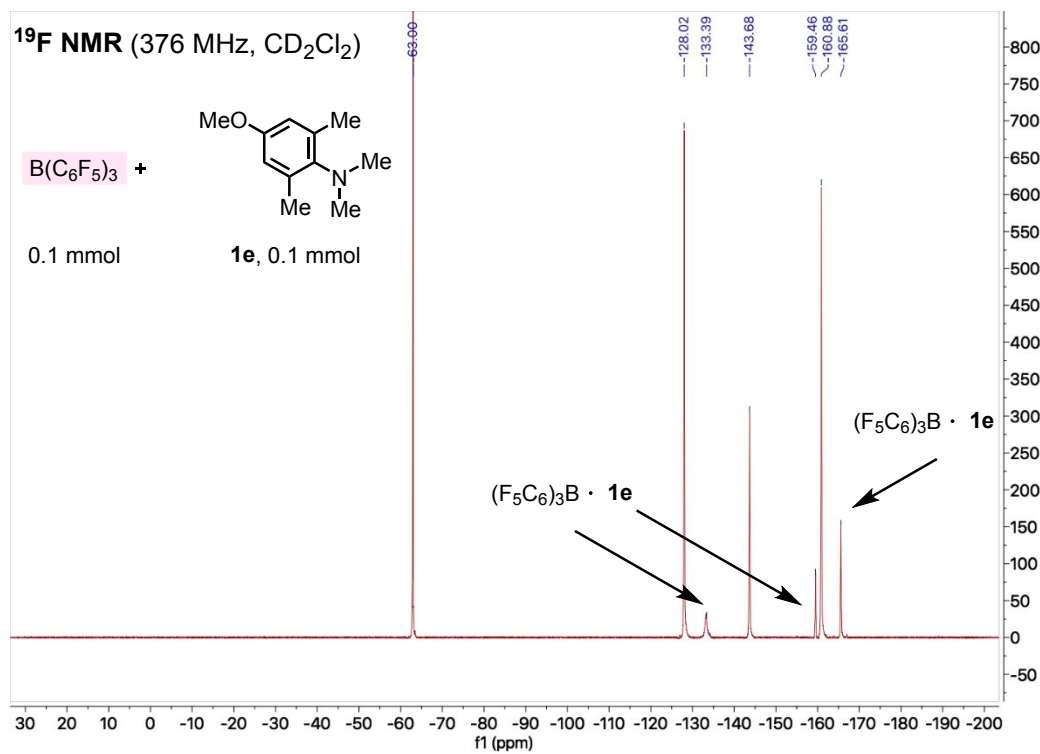


Figure S32. ^{19}F NMR spectrum of a sample containing $\text{B}(\text{C}_6\text{F}_5)_3$ and amine **1e** in CD_2Cl_2 .

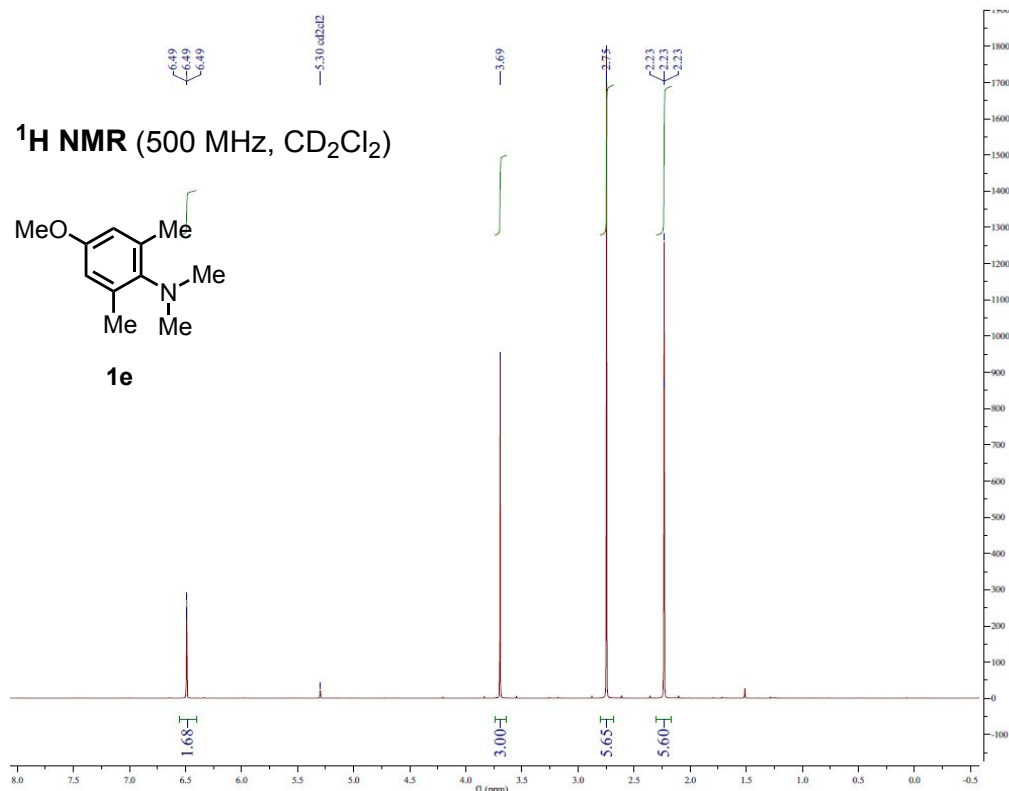


Figure S33. ¹H NMR spectrum of amine **1e** in CD₂Cl₂.

Next, we attempted to detect a Lewis acid/Lewis base adduct that may form between the carbonyl unit of alkynylsilane **2b** and B(C₆F₅)₃.²⁰ The ¹¹B and ¹⁹F NMR spectra were obtained for:

Figures S34 – S35: the sample containing 0.1 mmol of B(C₆F₅)₃ and 0.1 mmol of **2b**.

It was found through studying the ¹¹B NMR spectrum of the sample (Figure S34) that B(C₆F₅)₃ forms the adduct with **2b** (a characteristic peak was observed at –2.1 ppm), and that B(C₆F₅)₃ is consumed.

Furthermore, we detected peaks at –134.14, –157.49 and –164.44 ppm in the ¹⁹F NMR spectrum (Figure S35) that may correspond to the adduct formed while B(C₆F₅)₃ was fully consumed.

In order to determine if the adduct may also be generated in under the standard reaction conditions for catalytic C–H functionalization, ¹⁹F NMR spectrum (Figure S35) was compared to those obtained for the unpurified reaction mixtures (Figures S27–S28). In Figures S27–S28, there were peaks at –134.64, –157.68 and –164.48 ppm (vs those at –134.14, –157.49 and –164.44 ppm found in S35), thereby suggesting that the adduct may be present in the unpurified reaction mixture.

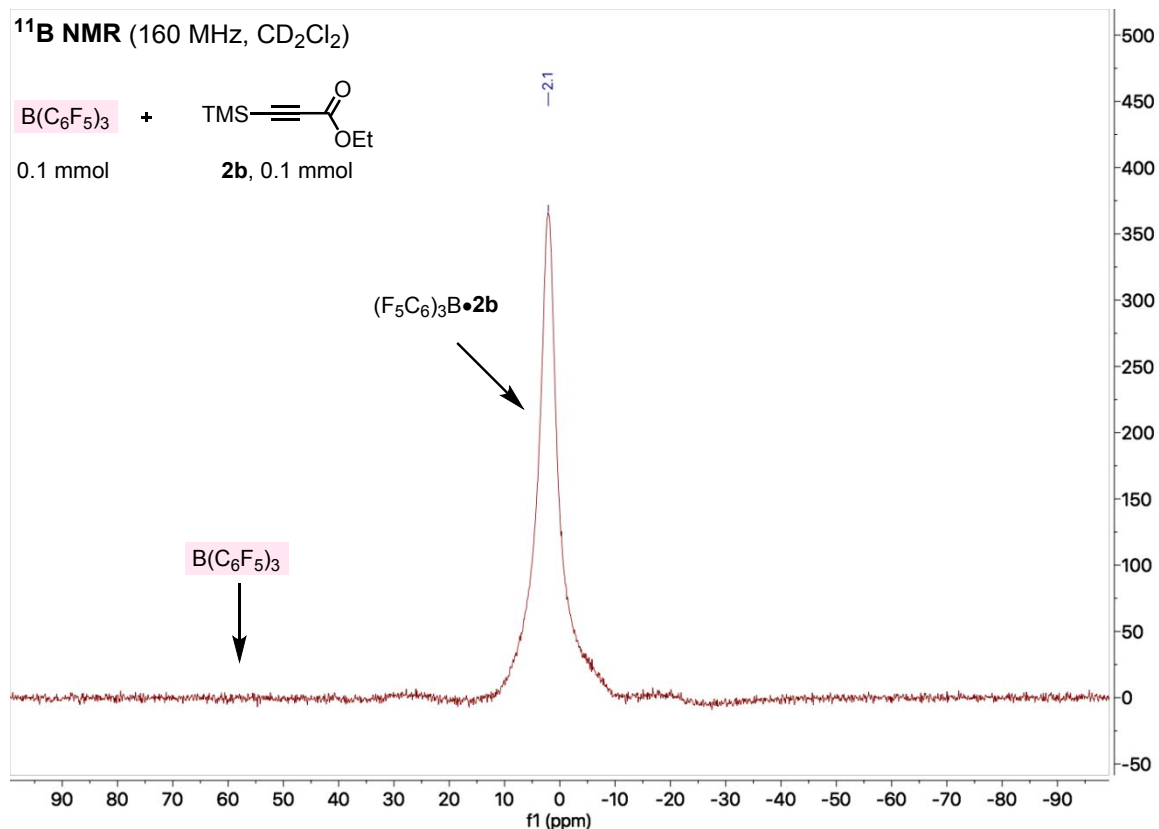


Figure S34. ^{11}B NMR spectrum of a sample containing $\text{B}(\text{C}_6\text{F}_5)_3$ and alkyne **2b** in CD_2Cl_2 .

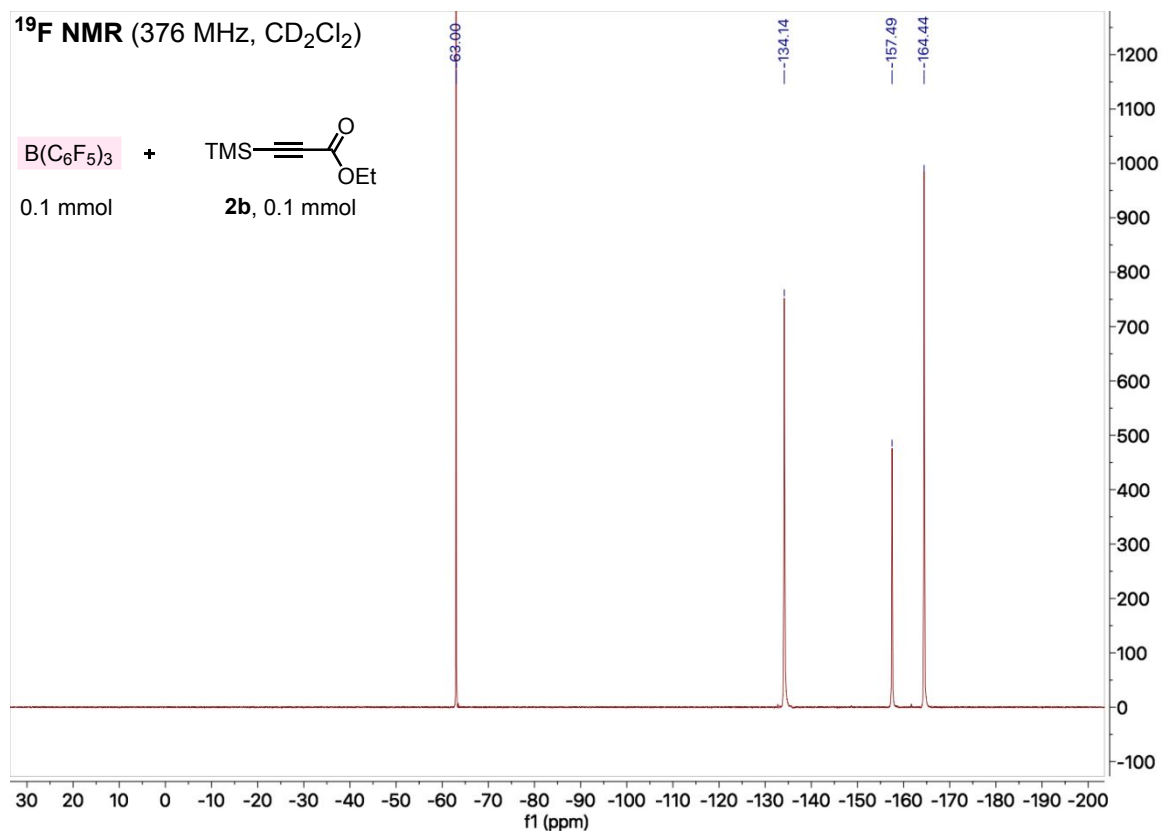
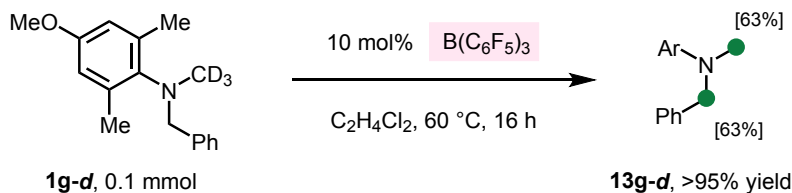


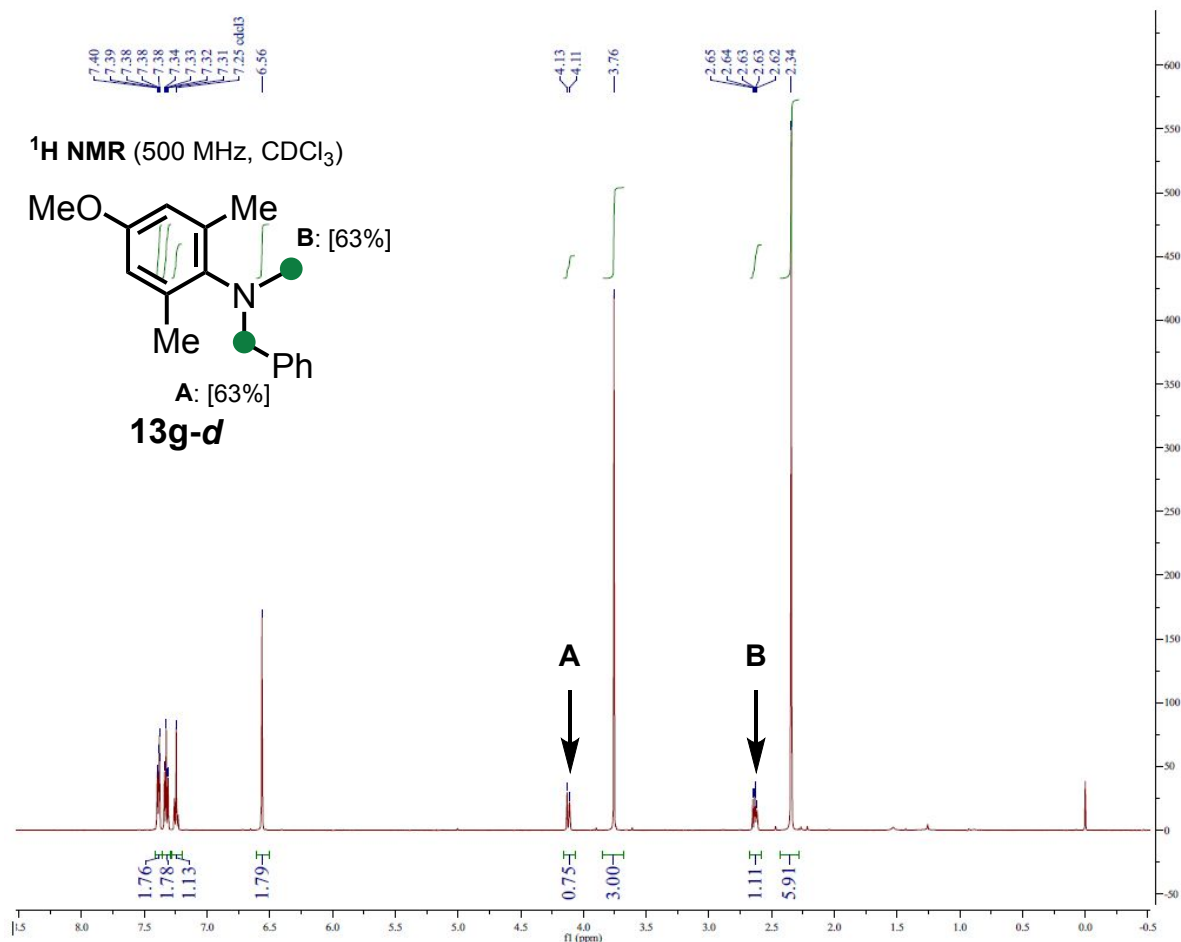
Figure S35. ^{19}F NMR spectrum of a sample containing $\text{B}(\text{C}_6\text{F}_5)_3$ and alkyne **2b** in CD_2Cl_2 .

5.4. Experiments Involving *N*-Benzyl-4-methoxy-2,6-dimethyl-*N*-(methyl-*d*₃)aniline

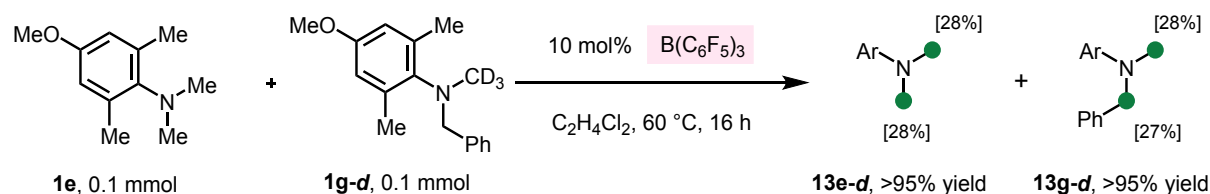


Experimental Procedure for the Isotope Exchange of *N*-Benzyl-4-methoxy-2,6-dimethyl-*N*-(methyl-*d*₃)aniline

An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added amine **1g-d** (0.1 mmol), B(C₆F₅)₃ (0.01 mmol, 10 mol%), and C₂H₄Cl₂ (0.4 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 16 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH₂Cl₂. The combined organic material was then concentrated *in vacuo*. The unpurified mixture was purified by silica gel column chromatography (Et₂O:hexanes = 1:19), to afford **13g-d** as a colorless oil (>95% yield). Deuterium or proton incorporation values were determined based on analysis of the ¹H NMR spectrum of the purified product.

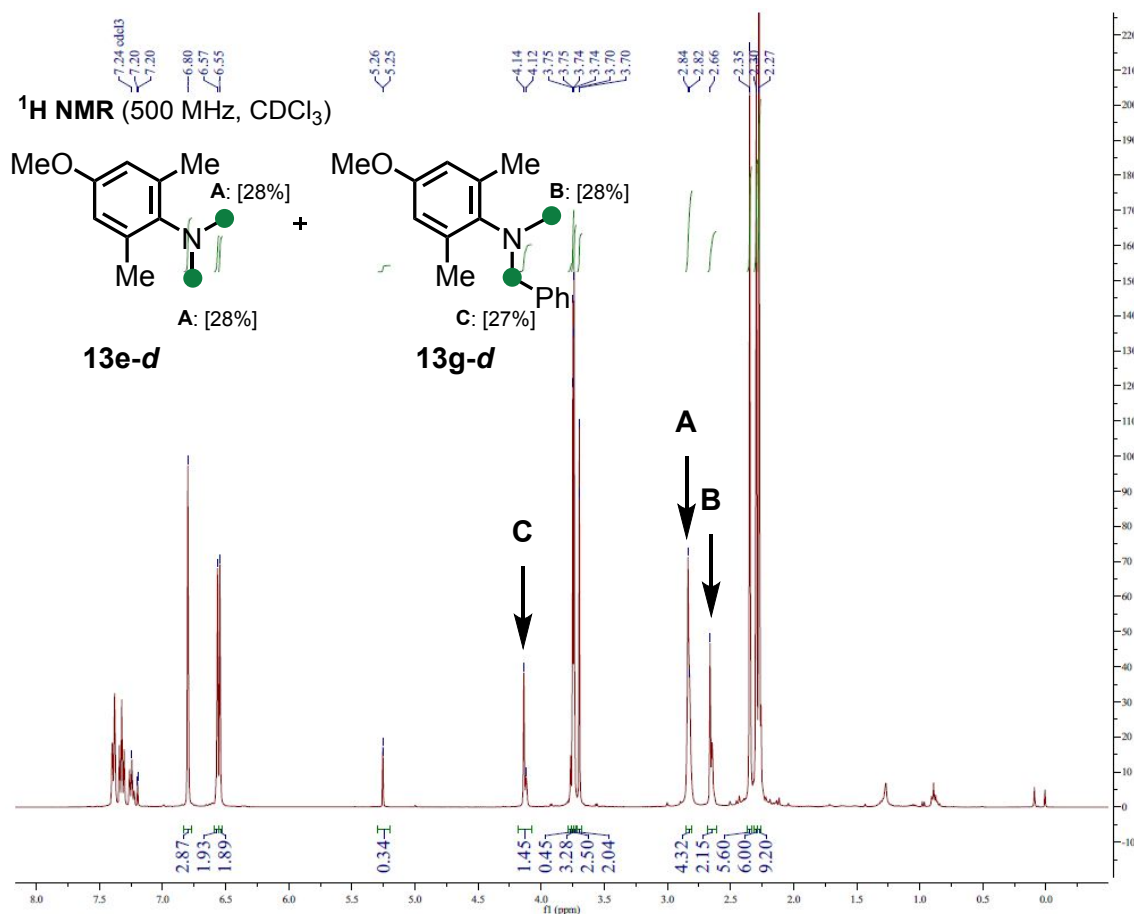


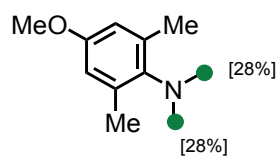
Intermolecular Isotope Exchange Experiment Between *N*-Benzyl-4-methoxy-2,6-dimethyl-*N*-(methyl-*d*₃)aniline and 4-Methoxy-*N,N*,2,6-tetramethylaniline



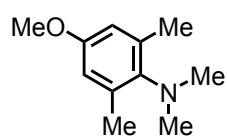
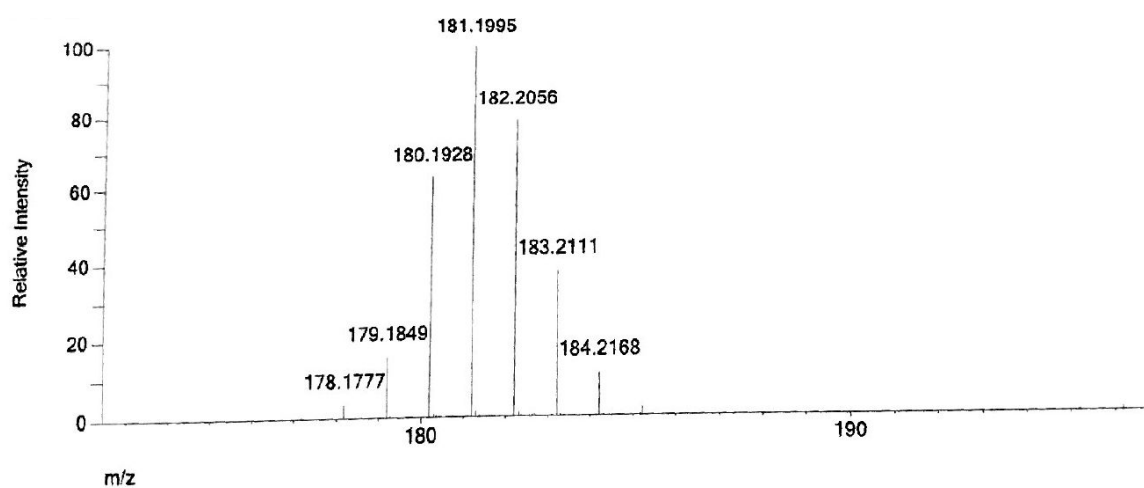
Experimental Procedure for the Isotope Exchange Between of *N*-Benzyl-4-methoxy-2,6-dimethyl-*N*-(methyl-*d*₃)aniline and 4-Methoxy-*N,N*,2,6-tetramethylaniline

An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added amine **1e** (0.1 mmol), amine **1g-d** (0.1 mmol), $\text{B(C}_6\text{F}_5)_3$ (0.02 mmol, 10 mol%), and $\text{C}_2\text{H}_4\text{Cl}_2$ (0.4 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 16 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH_2Cl_2 . The combined organic material was then concentrated *in vacuo*. Deuterium incorporation values were determined based on analysis of the ^1H NMR and HRMS spectra of the purified product.

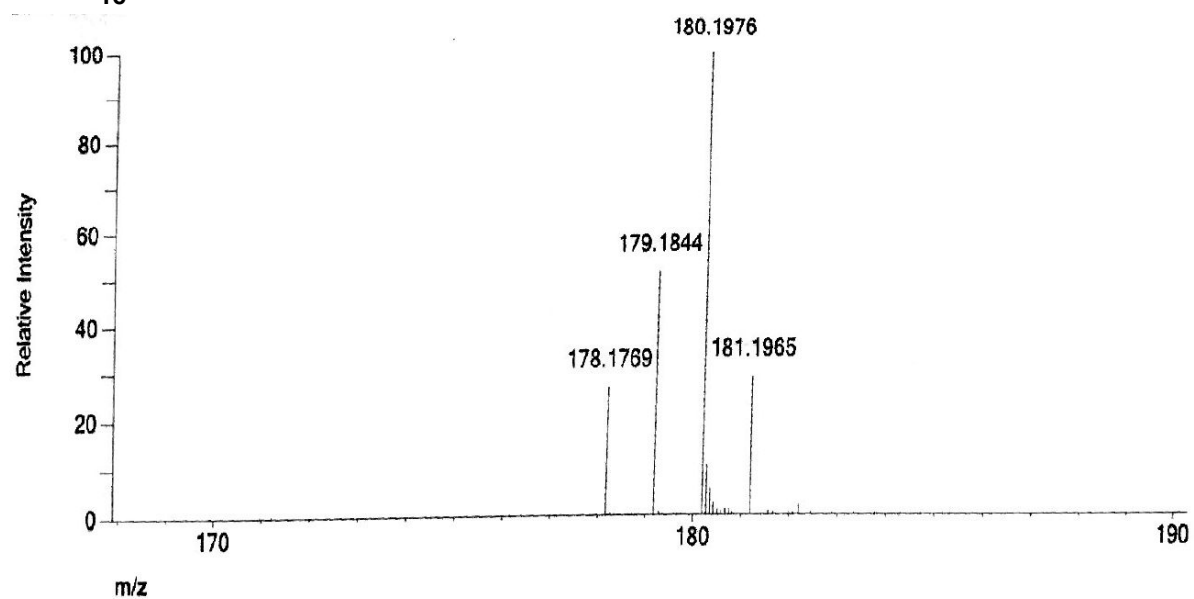


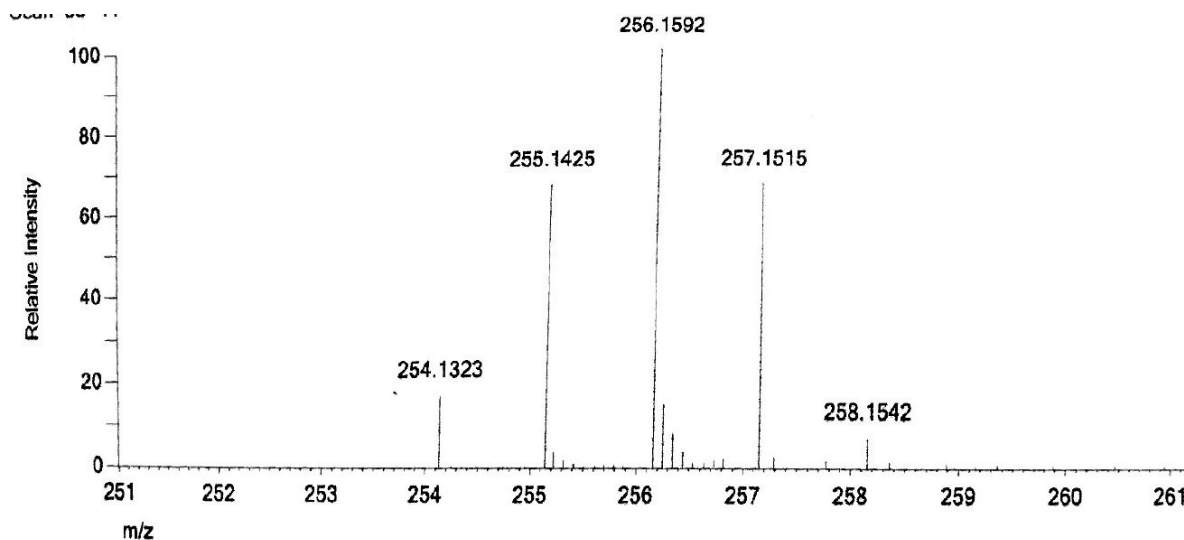
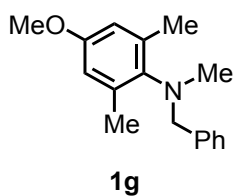
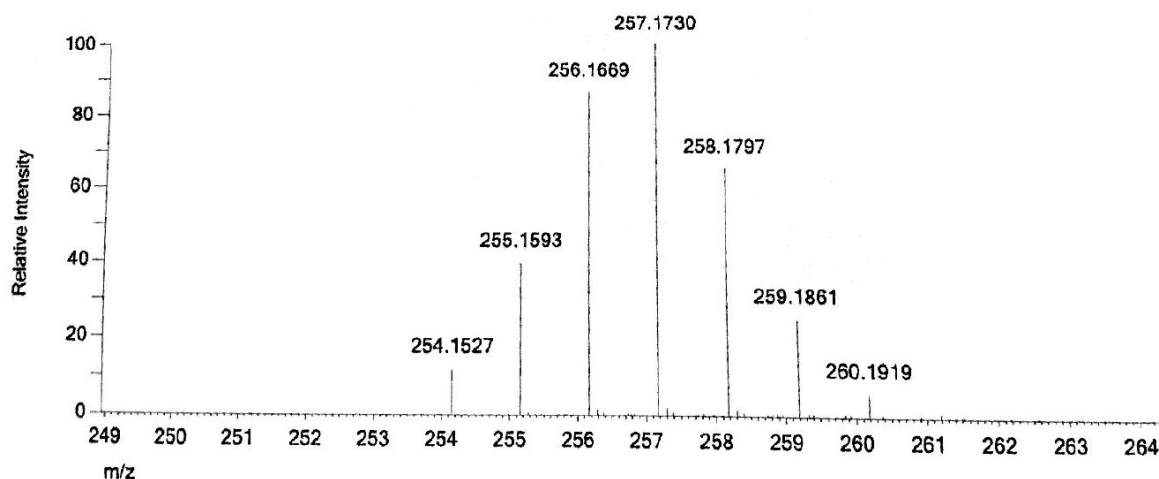
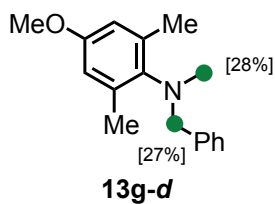


13e-d



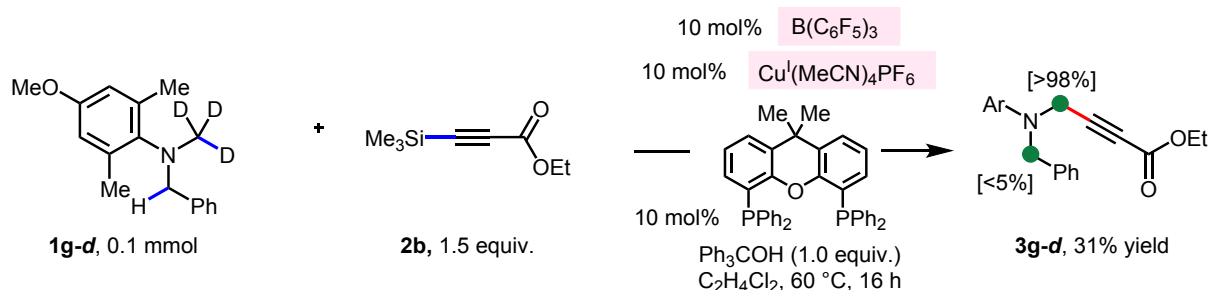
1e





Experiments Involving *N*-Benzyl-4-methoxy-2,6-dimethyl-*N*-(methyl-*d*₃)aniline

We investigated if the H/D exchange reaction may take place under the standard conditions for B(C₆F₅)₃ and organocopper co-catalyzed α -amino C–H alkynylation reaction with 0.10 mmol of *N*-benzyl-4-methoxy-2,6-dimethyl-*N*-(methyl-*d*₃)aniline **1g-d**, 0.15 mmol of 3-(trimethylsilyl)propiolate **2b** and 0.10 mmol of Ph₃COH.

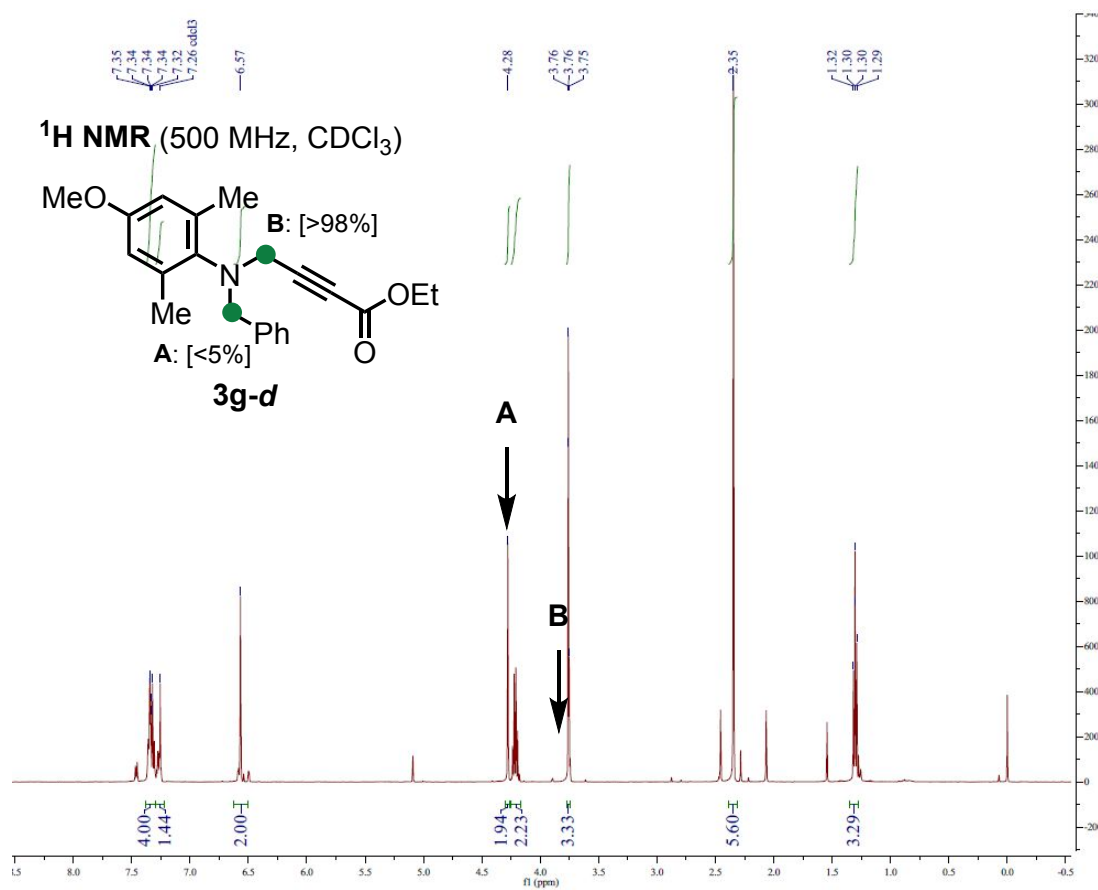


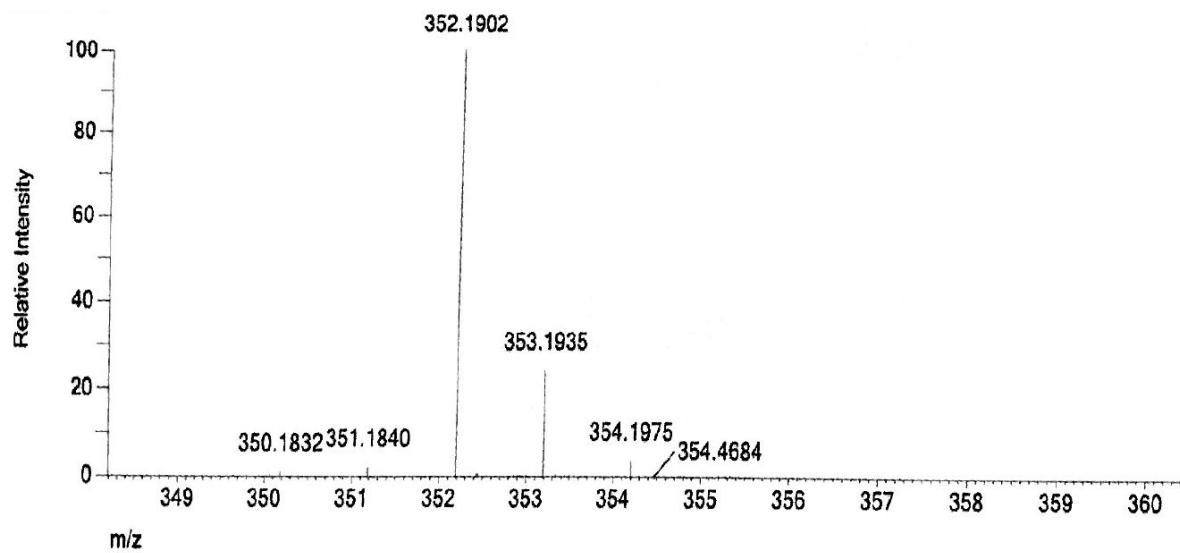
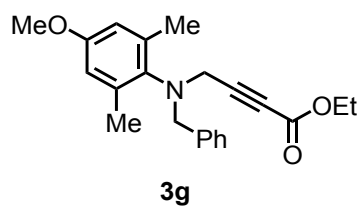
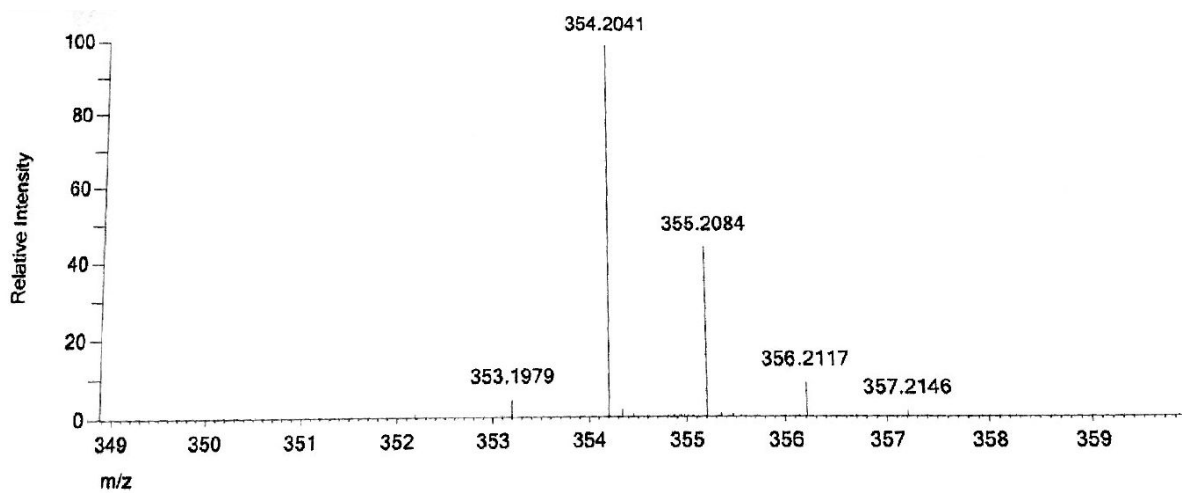
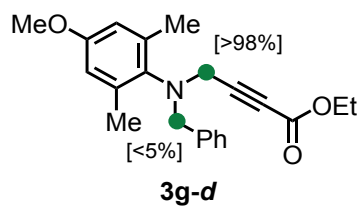
The ^1H NMR and HRMS spectrum of the isolated and purified product **3g-d** showed that there was 19% deuterium incorporation at the benzylic position of **3g-d**.

Experimental Procedure for the α -Alkynylation of *N*-Benzyl-4-methoxy-2,6-dimethyl-*N*-(methyl- d_3)aniline

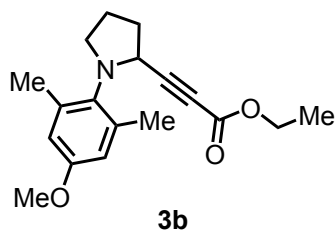
An oven-dried sealed tube equipped with a magnetic stir bar was used. To the sealed tube were added $\text{Cu}(\text{MeCN})_4\text{PF}_6$ (0.01 mmol), Xantphos (0.01 mmol), and $\text{C}_2\text{H}_4\text{Cl}_2$ (0.2 mL) under nitrogen atmosphere. The mixture was allowed to stir for 20 minutes at 22 °C, then ethyl 3-(trimethylsilyl)propiolate **2b** (0.15 mmol), triphenylmethanol (0.1 mmol), amine **1g-d** (0.1 mmol), $\text{B(C}_6\text{F}_5)_3$ (0.01 mmol, 10 mol%), and $\text{C}_2\text{H}_4\text{Cl}_2$ (0.2 mL) were added to the vessel. The reaction mixture was allowed to stir at 60 °C for 16 h. Upon completion, the unpurified reaction mixture was filtered through a short plug of Celite and rinsed with CH_2Cl_2 . The combined organic material was then concentrated *in vacuo*. The propargylamine product was purified and isolated by silica gel column chromatography (Et_2O :hexanes = 1:9), **3g-d** was obtained as a colorless liquid (12.7 mg, 36%). Deuterium incorporation values were determined based on analysis of the ^1H NMR and HRMS spectra of the purified product.

^1H NMR (500 MHz, CDCl_3) δ 7.38 – 7.29 (m, 4H), 7.29 – 7.23 (m, 1H), 6.57 (s, 2H), 4.28 (s, 2H), 4.22 (qd, J = 7.1, 0.9 Hz, 2H), 3.76 (s, 3H), 2.35 (s, 6H), 1.30 (dd, J = 7.8, 6.9 Hz, 3H).





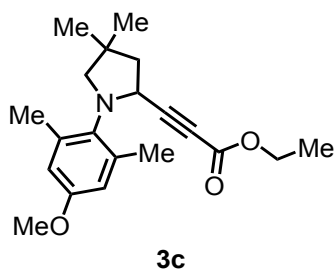
6. Analytical Data



Ethyl 3-(1-(4-methoxy-2,6-dimethylphenyl)pyrrolidin-2-yl)propiolate (**3b**)

1-(4-Methoxy-2,6-dimethylphenyl)pyrrolidine **1b** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure A**. After purification by column chromatography (Et₂O:hexanes = 1:19), **3b** was obtained as a colorless liquid (54 mg, 90%).

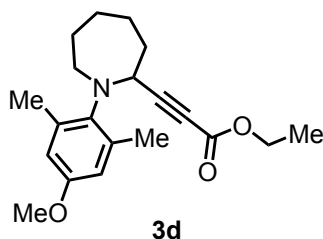
¹H NMR (500 MHz, CDCl₃) δ 6.58 (s, 2H), 4.27 (ddd, *J* = 7.8, 3.5, 1.3 Hz, 1H), 4.17 (qd, *J* = 7.2, 1.3 Hz, 2H), 3.75 (s, 3H), 3.43 – 3.31 (m, 1H), 3.09 (q, *J* = 6.9 Hz, 1H), 2.47 – 2.22 (m, 6H), 2.22 – 2.10 (m, 3H), 2.09 – 1.94 (m, 1H), 1.27 (td, *J* = 7.2, 1.4 Hz, 3H); **¹³C NMR** (151 MHz, CDCl₃) δ 157.2, 153.3, 136.6, 113.2, 91.5, 73.9, 63.3, 55.9, 52.3, 51.0, 35.8, 26.7, 18.4, 15.1; **IR** (neat) ν 2967, 2837, 2224, 1707, 1599, 1476, 1366, 1244, 1153, 1067, cm⁻¹; **HRMS** (DART) Calcd for C₁₈H₂₄NO₃ (MH⁺): 302.1751; found: 302.1755.



Ethyl 3-(1-(4-methoxy-2,6-dimethylphenyl)-3,3-dimethylpyrrolidin-2-yl)propiolate (3c)

1-(4-Methoxy-2,6-dimethylphenyl)-3,3-dimethylpyrrolidine **1c** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure A**. After purification by column chromatography (Et₂O:hexanes = 1:19), **3c** was obtained as a colorless liquid (51 mg, 77%).

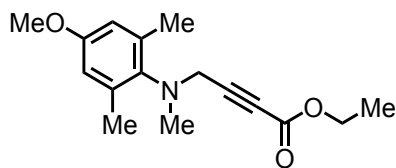
¹H NMR (600 MHz, CDCl₃) δ 6.57 (s, 2H), 4.41 (dd, *J* = 9.0, 3.8 Hz, 1H), 4.15 (q, *J* = 7.1 Hz, 2H), 3.75 (s, 3H), 3.07 (d, *J* = 7.9 Hz, 1H), 2.95 (d, *J* = 7.8 Hz, 1H), 2.37 (s, 5H), 2.19 (dd, *J* = 12.5, 9.0 Hz, 2H), 1.98 (dd, *J* = 12.5, 3.9 Hz, 1H), 1.37 (s, 3H), 1.26 (t, *J* = 7.2 Hz, 3H), 1.15 (d, *J* = 1.0 Hz, 3H); **¹³C NMR** (151 MHz, CDCl₃) δ 157.2, 153.8, 135.4, 113.7, 91.0, 74.3, 64.5, 61.6, 55.2, 51.9, 47.0, 39.1, 27.8, 27.2, 14.0; **IR** (neat) ν 2954, 2864, 2228, 1707, 1601, 1465, 1243, 1094, 1023, 751 cm⁻¹; **HRMS** (DART) Calcd for C₂₀H₂₈NO₃ (MH⁺): 330.2063; found: 330.2069.



Ethyl 3-(1-(4-methoxy-2,6-dimethylphenyl)azepan-2-yl)propiolate (3d)

1-(4-Methoxy-2,6-dimethylphenyl)azepane **1d** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure A**. After purification by column chromatography (Et₂O:hexanes = 1:19), **3d** was obtained as a colorless liquid (51 mg, 77%).

¹H NMR (500 MHz, CDCl₃) δ 6.57 (d, *J* = 13.4 Hz, 2H), 4.18 (d, *J* = 7.1 Hz, 2H), 4.06 (d, *J* = 2.7 Hz, 1H), 3.75 (s, 3H), 3.32 (d, *J* = 9.1 Hz, 1H), 3.08 (d, *J* = 11.1 Hz, 1H), 2.42 (s, 3H), 2.29 (s, 3H), 2.19 (d, *J* = 1.5 Hz, 1H), 1.95 – 1.85 (m, 2H), 1.85 – 1.79 (m, 1H), 1.79 – 1.70 (m, 2H), 1.57 (s, 2H), 1.28 (s, 3H); **¹³C NMR** (151 MHz, CDCl₃) δ 157.4, 153.8, 140.9, 139.7, 137.9, 114.1, 113.6, 90.3, 75.6, 59.4, 55.6, 53.0, 51.3, 34.8, 31.4, 28.9, 25.2, 20.1, 19.6, 14.0; **IR** (neat) ν 2926, 2845, 2221, 1707, 1598, 1474, 1309, 1240, 1065, 853 cm⁻¹; **HRMS** (DART) Calcd for C₂₀H₂₈NO₃ (MH⁺): 330.2063; found: 330.2061.

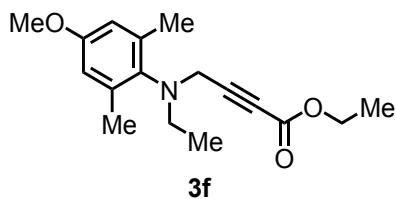


3e

Ethyl 4-((4-methoxy-2,6-dimethylphenyl)(methyl)amino)but-2-ynoate (3e)

4-Methoxy-*N,N*,2,6-tetramethylaniline **1e** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure A**. After purification by column chromatography (Et₂O:hexanes = 1:19), **3e** was obtained as a colorless liquid (50 mg, 90%).

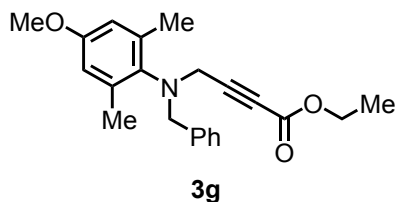
¹H NMR (500 MHz, CDCl₃) δ 6.54 (s, 2H), 4.23 (q, *J* = 7.1, 1.5 Hz, 2H), 3.89 (s, 2H), 3.74 (s, 3H), 2.88 (s, 3H), 2.29 (s, 6H), 1.31 (t, *J* = 7.1, 1.5 Hz, 3H); **¹³C NMR** (151 MHz, CDCl₃) δ 156.3, 153.6, 140.9, 138.7, 112.8, 86.1, 75.6, 61.8, 55.2, 44.8, 40.1, 19.4, 13.9; **IR** (neat) ν 2933, 2228, 1707, 1598, 1480, 1309, 1244, 1155, 1060, 855 cm⁻¹; **HRMS** (DART) Calcd for C₁₆H₂₂NO₃ (MH⁺): 276.1594; found: 276.1609.



Ethyl 4-(ethyl(4-methoxy-2,6-dimethylphenyl)amino)but-2-ynoate (3f)

N-Ethyl-4-methoxy-*N*,2,6-trimethylaniline **1f** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure A**. After purification by column chromatography (Et₂O:hexanes = 1:19), **3f** was obtained as a colorless liquid (24 mg, 42%).

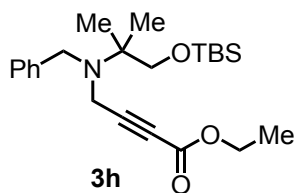
¹H NMR (500 MHz, CDCl₃) δ 6.56 (s, 2H), 4.23 (q, *J* = 7.1 Hz, 2H), 3.92 (s, 2H), 3.76 (s, 3H), 3.21 (q, *J* = 7.2 Hz, 2H), 2.30 (s, 6H), 1.31 (t, *J* = 7.1 Hz, 3H), 1.04 (t, *J* = 7.2 Hz, 3H); **¹³C NMR** (126 MHz, CDCl₃) δ 156.8, 153.7, 139.5, 139.3, 113.6, 86.7, 75.5, 61.8, 55.2, 47.2, 42.9, 19.6, 14.3, 14.0; **IR** (neat) ν 2933, 2230, 1707, 1598, 1490, 1309, 1254, 1120, 1060, 840 cm⁻¹; **HRMS** (DART) Calcd for C₁₇H₂₄NO₃ (MH⁺): 290.1751; found: 290.1755.



Ethyl 4-(benzyl(4-methoxy-2,6-dimethylphenyl)amino)but-2-ynoate (3g)

N-Benzyl-4-methoxy-*N*,2,6-trimethylaniline **1g** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure A**. After purification by column chromatography (Et₂O:hexanes = 1:19), **3g** was obtained as a colorless liquid (51 mg, 72%).

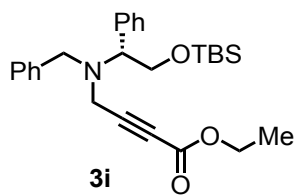
¹H NMR (500 MHz, CDCl₃) δ 7.38 – 7.29 (m, 4H), 7.28 – 7.23 (m, 1H), 6.57 (s, 2H), 4.28 (s, 2H), 4.21 (q, *J* = 7.1 Hz, 2H), 3.78 (s, 2H), 3.76 (s, 3H), 2.34 (s, 6H), 1.30 (t, *J* = 7.1 Hz, 3H); **¹³C NMR** (151 MHz, CDCl₃) δ 156.8, 153.5, 140.6, 139.0, 138.5, 128.9, 128.4, 127.2, 113.4, 85.4, 75.3, 61.8, 58.5, 55.2, 41.5, 19.9, 13.2; **IR** (neat) ν 2927, 2841, 2230, 1708, 1598, 1479, 1312, 1245, 1065, 855 cm⁻¹; **HRMS** (DART) Calcd for C₂₂H₂₆NO₃ (MH⁺): 352.1907; found: 352.1895.



Ethyl 4-(benzyl(1-((*tert*-butyldimethylsilyl)oxy)-2-methylpropan-2-yl)amino)but-2-ynoate (3h)

N-Benzyl-1-((*tert*-butyldimethylsilyl)oxy)-*N*,2-dimethylpropan-2-amine **1h** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure B** using 1,2-bis(diphenylphosphino)ethane as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:19), **3h** was obtained as a colorless liquid (61 mg, 76%).

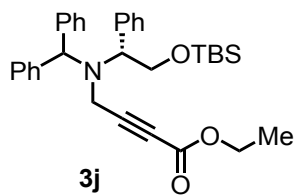
¹H NMR (500 MHz, CDCl₃) δ 7.35 (dt, *J* = 6.6, 1.1 Hz, 2H), 7.32 – 7.27 (m, 2H), 7.24 – 7.20 (m, 1H), 4.23 (q, *J* = 7.2 Hz, 2H), 3.91 (s, 2H), 3.60 (s, 2H), 3.51 (s, 2H), 1.31 (t, *J* = 7.1, 0.8 Hz, 3H), 1.23 (s, 6H), 0.91 (s, 9H), 0.06 (s, 6H); **¹³C NMR** (126 MHz, CDCl₃) δ 153.6, 140.3, 128.4, 128.2, 126.9, 87.6, 76.7, 69.7, 61.8, 58.6, 51.4, 36.7, 25.9, 22.9, 18.2, 14.1, -5.6; **IR** (neat) ν 2949, 2855, 2221, 1708, 1463, 1364, 1237, 1092, 840, 774 cm⁻¹; **HRMS** (DART) Calcd for C₂₃H₃₈NO₃Si (MH⁺): 404.2615; found: 404.2610.



Ethyl (R)-4-(benzyl(2-((*tert*-butyldimethylsilyl)oxy)-1-phenylethyl)amino)but-2-ynoate (3i)

(*R*)-*N*-Benzyl-2-((*tert*-butyldimethylsilyl)oxy)-*N*-methyl-1-phenylethan-1-amine **1i** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure B** using 1,2-bis(diphenylphosphino)ethane as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:19), **3i** was obtained as a colorless liquid (78 mg, 86%).

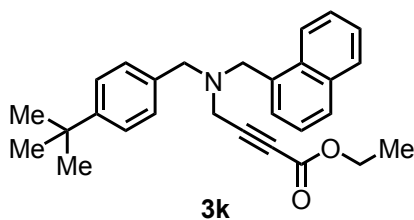
¹H NMR (500 MHz, CDCl₃) δ 7.45 (d, *J* = 7.5 Hz, 2H), 7.37 – 7.20 (m, 8H), 4.26 (q, *J* = 7.1, 2.1 Hz, 2H), 3.94 (dd, *J* = 7.3, 4.9 Hz, 2H), 3.82 (d, *J* = 2.0 Hz, 1H), 3.74 (d, *J* = 2.1 Hz, 1H), 3.70 – 3.60 (m, 2H), 3.37 – 3.28 (m, 1H), 1.34 (t, *J* = 2.0 Hz, 3H), 0.83 (s, 9H), -0.09 (d, *J* = 5.7, 2.1 Hz, 6H); **¹³C NMR** (126 MHz, CDCl₃) δ 153.6, 141.1, 138.8, 128.9, 128.5, 128.30, 128.26, 127.4, 127.1, 84.5, 77.84, 67.6, 65.9, 61.9, 55.1, 39.1, 25.8, 18.2, 14.1, -0.01, -5.67, -5.69; **IR** (neat) ν 2923, 2853, 2222, 1709, 1458, 1365, 1239, 1095, 870, 698 cm⁻¹; **HRMS** (DART) Calcd for C₂₇H₃₈NO₃Si (MH⁺): 452.2615; found: 452.2612; [α]_D²⁵ = 36.7° (*c* 0.2, CH₂Cl₂).



Ethyl (R)-4-(benzhydryl(2-((*tert*-butyldimethylsilyl)oxy)-1-phenylethyl)amino)but-2-ynoate (3j)

(*R*)-*N*-Benzhydryl-2-((*tert*-butyldimethylsilyl)oxy)-*N*-methyl-1-phenylethan-1-amine **1j** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure B** using 1,2-bis(diphenylphosphino)ethane as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:19), **3j** was obtained as a colorless liquid (102 mg, 97%).

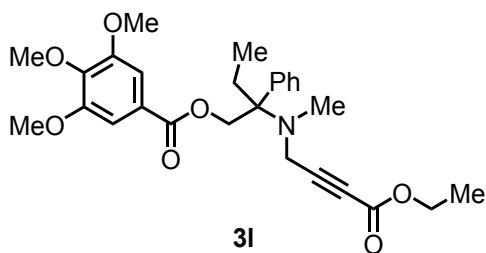
¹H NMR (600 MHz, CDCl₃) δ 7.52 – 7.46 (m, 4H), 7.42 (dt, *J* = 8.1, 1.1 Hz, 2H), 7.35 – 7.14 (m, 9H), 5.23 (s, 1H), 4.24 – 4.06 (m, 5H), 3.55 (s, 2H), 1.31 (t, *J* = 7.1, 3H), 0.82 (s, 9H), -0.04 (d, *J* = 20.0, 6H); **¹³C NMR** (151 MHz, CDCl₃) δ 153.4, 142.0, 141.9, 139.8, 128.7, 128.6, 128.49, 128.47, 128.39, 128.1, 127.24, 127.15, 86.5, 76.6, 69.2, 63.1, 62.9, 61.6, 36.6, 25.9, 18.2, 14.1, -5.55, -5.57; **IR** (neat) ν 3026, 2930, 2855, 2224, 1708, 1456, 1362, 1243, 1095, 837 cm⁻¹; **HRMS** (DART) Calcd for C₃₃H₄₂NO₃Si (MH⁺): 528.2928; found: 528.2922; [α]_D²⁵ = -16.2° (*c* 0.8, CH₂Cl₂).



Ethyl 4-((4-(*tert*-butyl)phenyl)(naphthalen-1-ylmethyl)amino)but-2-ynoate (3k**)**

4-(*tert*-Butyl)-*N*-methyl-*N*-(naphthalen-1-ylmethyl)aniline **1k** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure B** using 1,2-bis(diphenylphosphino)ethane as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:19), **3k** was obtained as a colorless liquid (63 mg, 76%).

¹H NMR (500 MHz, CDCl₃) δ 8.21 (d, *J* = 7.4 Hz, 1H), 7.86 – 7.80 (m, 1H), 7.77 (d, *J* = 8.2 Hz, 1H), 7.55 – 7.45 (m, 3H), 7.40 (dd, *J* = 8.6, 6.6 Hz, 1H), 7.36 – 7.27 (m, 4H), 4.30 (q, *J* = 7.2 Hz, 2H), 4.15 (s, 2H), 3.78 (s, 2H), 3.35 (s, 2H), 1.37 (t, *J* = 7.1 Hz, 4H), 1.30 (s, 9H); **¹³C NMR** (126 MHz, CDCl₃) δ 153.6, 150.3, 135.1, 133.9, 133.7, 132.5, 128.9, 128.4, 128.3, 127.8, 125.8, 125.7, 125.3, 125.2, 124.9, 83.6, 78.3, 62.0, 57.7, 56.2, 40.9, 34.5, 31.4, 14.1; **IR** (neat) ν 2957, 2825, 2220, 1707, 1597, 1509, 1239, 1107, 1050, 791 cm⁻¹; **HRMS** (DART) Calcd for C₂₈H₃₂NO₂ (MH⁺): 414.2428; found: 414.2429.

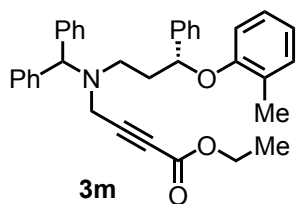


**2-((4-Ethoxy-4-oxobut-2-yn-1-yl)(methyl)amino)-2-phenylbutyl
trimethoxybenzoate (3I)**

3,4,5-

2-(Dimethylamino)-2-phenylbutyl 3,4,5-trimethoxybenzoate **1I** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure B** using 1,2-bis(diphenylphosphino)ethane as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:2), **3I** was obtained as a colorless liquid (69 mg, 71%).

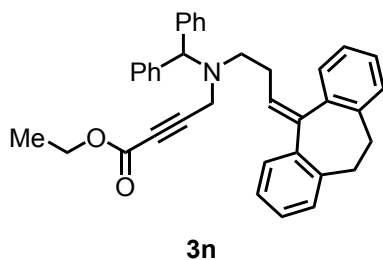
¹H NMR (500 MHz, CDCl₃) δ 7.52 – 7.46 (m, 2H), 7.40 – 7.33 (m, 2H), 7.29 – 7.22 (m, 1H), 7.18 (d, *J* = 1.4 Hz, 2H), 4.89 (dd, *J* = 12.1, 1.3 Hz, 1H), 4.74 (dd, *J* = 12.0, 1.3 Hz, 1H), 4.18 (q, *J* = 7.1 Hz, 2H), 3.89 (s, 3H), 3.82 (s, 6H), 3.63 (d, *J* = 17.6 Hz, 1H), 3.48 (d, *J* = 17.6 Hz, 1H), 2.63 (s, 3H), 1.95 – 1.78 (m, 2H), 1.26 (t, *J* = 7.1 Hz, 2H), 0.69 (t, *J* = 7.3 Hz, 4H); **¹³C NMR** (126 MHz, CDCl₃) δ 165.8, 153.5, 152.9, 142.3, 141.7, 128.2, 127.3, 126.9, 124.8, 106.8, 86.2, 75.5, 65.7, 64.8, 61.8, 60.8, 56.1, 42.1, 35.8, 30.1, 13.9, 8.5; **IR** (neat) ν 2939, 2231, 1709, 1586, 1498, 1331, 1239, 1123, 1006, 756 cm⁻¹; **HRMS** (DART) Calcd for C₂₇H₃₄NO₇ (MH⁺): 484.2330; found: 484.2322.



Ethyl (*R*)-4-(benzhydryl(3-phenyl-3-(*o*-tolylloxy)propyl)amino)but-2-ynoate (3m**)**

(*R*)-*N*-Benzhydryl-*N*-methyl-3-phenyl-3-(*o*-tolylloxy)propan-1-amine **1m** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure B** using 1,2-bis(diphenylphosphino)ethane as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:19), **3m** was obtained as a colorless liquid (58 mg, 56%).

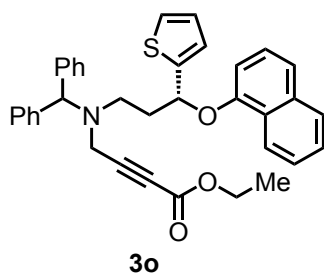
¹H NMR (500 MHz, CDCl₃) δ 7.45 – 7.40 (m, 2H), 7.35 – 7.13 (m, 10H), 7.07 (qt, *J* = 5.7, 1.3 Hz, 4H), 6.94 (td, *J* = 7.8, 1.8 Hz, 1H), 6.75 (td, *J* = 7.4, 1.1 Hz, 1H), 6.57 – 6.52 (m, 1H), 5.19 (dd, *J* = 9.2, 3.4 Hz, 1H), 4.66 (s, 1H), 4.25 (q, *J* = 7.1 Hz, 2H), 3.63 – 3.50 (m, 2H), 2.86 – 2.77 (m, 2H), 2.13 (dddd, *J* = 19.3, 9.2, 7.5, 5.6 Hz, 1H), 2.02 (s, 3H), 2.02 – 1.95 (m, 1H), 1.34 (t, *J* = 7.1 Hz, 3H); **¹³C NMR** (126 MHz, CDCl₃) δ 155.9, 153.4, 142.33, 142.26, 142.24, 130.5, 128.64, 128.57, 128.54, 127.91, 127.88, 127.72, 127.4, 127.20, 127.15, 127.1, 126.4, 125.8, 120.0, 112.3, 83.5, 77.9, 76.9, 72.6, 61.9, 47.3, 39.5, 37.2, 16.3, 14.1; **IR** (neat) ν 3025, 2933, 2831, 2220, 1707, 1594, 1490, 1240, 1049, 749 cm⁻¹; **HRMS** (DART) Calcd for C₃₅H₃₆NO₃ (MH⁺): 518.2689; found: 518.2691.



Ethyl 4-(benzhydryl(3-(10,11-dihydro-5*H*-dibenzo[*a,d*][7]annulen-5-ylidene)propyl)amino)but-2-ynoate (3n)

N-Benzhydryl-3-(10,11-dihydro-5*H*-dibenzo[*a,d*][7]annulen-5-ylidene)-*N*-methylpropan-1-amine **1n** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure B** using 1,2-bis(diphenylphosphino)ethane as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:4), **3n** was obtained as a colorless liquid (79 mg, 74%).

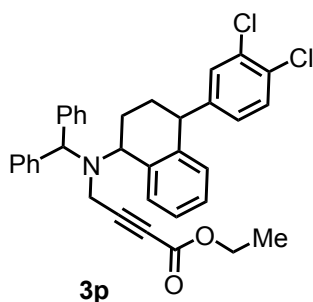
¹H NMR (500 MHz, CDCl₃) δ 7.45 – 7.39 (m, 4H), 7.30 – 7.22 (m, 5H), 7.22 – 7.12 (m, 7H), 7.10 – 7.02 (m, 2H), 5.84 (t, *J* = 7.5, 1.5 Hz, 1H), 4.67 (s, 1H), 4.27 (q, *J* = 7.1, 1.5 Hz, 2H), 3.47 – 3.19 (m, 4H), 2.97 (s, 1H), 2.67 (t, *J* = 7.2 Hz, 3H), 2.30 (q, *J* = 7.2 Hz, 2H), 1.36 (t, *J* = 7.1, 1.4 Hz, 3H); **¹³C NMR** (126 MHz, CDCl₃) δ 153.4, 143.8, 142.4, 141.2, 139.9, 139.3, 137.0, 129.9, 128.9, 128.6, 128.51, 128.48, 128.2, 128.0, 127.9, 127.4, 127.1, 126.9, 125.9, 125.8, 83.7, 77.8, 72.3, 61.9, 50.3, 39.3, 33.7, 31.9, 27.6, 14.1; **IR** (neat) ν 3020, 2922, 2836, 2224, 1707, 1484, 1448, 1365, 11243, 751 cm⁻¹; **HRMS** (DART) Calcd for C₃₇H₃₆NO₂ (MH⁺): 526.2740; found: 526.2751.



Ethyl (R)-4-(benzhydryl(3-(naphthalen-1-yloxy)-3-(thiophen-2-yl)propyl)amino)but-2-ynoate (3o)

(R)-N-Benzhydryl-N-methyl-3-(naphthalen-1-yloxy)-3-(thiophen-2-yl)propan-1-amine **1o** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure B** using 1,2-bis(diphenylphosphino)ethane as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:19), **3o** was obtained as a colorless liquid (76 mg, 68%).

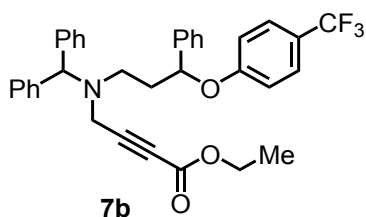
¹H NMR (600 MHz, CDCl₃) δ 8.04 (d, *J* = 8.3 Hz, 1H), 7.76 (d, *J* = 8.0 Hz, 1H), 7.46 (t, *J* = 7.3 Hz, 1H), 7.40 (dd, *J* = 19.1, 7.8 Hz, 4H), 7.27 (dt, *J* = 25.2, 7.3 Hz, 5H), 7.17 (q, *J* = 5.5 Hz, 2H), 7.01 – 6.91 (m, 4H), 6.89 (t, *J* = 4.4 Hz, 1H), 6.81 (d, *J* = 7.6 Hz, 1H), 5.69 (dd, *J* = 8.5, 4.3 Hz, 1H), 4.67 (s, 1H), 4.24 (q, *J* = 7.1 Hz, 2H), 3.59 (d, *J* = 4.6 Hz, 2H), 2.86 (d, *J* = 7.7 Hz, 2H), 2.49 – 2.11 (m, 2H), 1.32 (t, *J* = 7.1 Hz, 3H); **¹³C NMR** (126 MHz, CDCl₃) δ 153.4, 153.3, 145.3, 142.3, 141.9, 134.5, 128.7, 128.5, 127.9, 127.7, 127.3, 127.2, 126.9, 126.5, 126.2, 126.1, 125.6, 125.1, 124.6, 124.5, 122.2, 120.5, 106.4, 83.4, 78.1, 73.9, 72.5, 61.9, 53.4, 47.1, 39.7, 37.2, 14.0; **IR** (neat) ν 3055, 2926, 2837, 2220, 1707, 1579, 1453, 1243, 1092, 702 cm⁻¹; **HRMS** (DART) Calcd for C₃₆H₃₄NO₃S (MH⁺): 560.2254; found: 560.2239.



Ethyl 4-(benzhydryl(4-(3,4-dichlorophenyl)-1,2,3,4-tetrahydronaphthalen-1-yl)amino)but-2-ynoate (3p)

N-Benzhydryl-4-(3,4-dichlorophenyl)-*N*-methyl-1,2,3,4-tetrahydronaphthalen-1-amine **1p** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure B** using 1,2-bis(diphenylphosphino)ethane as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:19), **3p** was obtained as a colorless liquid (76 mg, 67%).

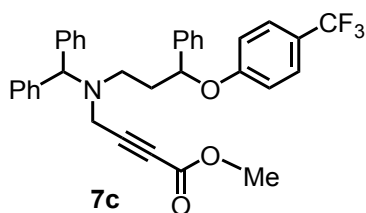
¹H NMR (600 MHz, CDCl₃) δ 8.05 (d, *J* = 8.1 Hz, 1H), 7.72 – 7.56 (m, 4H), 7.42 – 7.07 (m, 10H), 6.85 (dd, *J* = 69.9, 8.1 Hz, 2H), 5.31 – 5.17 (m, 1H), 4.28 – 4.11 (m, 3H), 4.05 (s, 1H), 3.53 (s, 2H), 2.06 (t, *J* = 12.1 Hz, 1H), 2.00 – 1.75 (m, 3H), 1.27 (t, *J* = 7.1, 3.2 Hz, 3H); **¹³C NMR** (126 MHz, CDCl₃) δ 153.3, 147.2, 142.24, 142.18, 139.1, 138.6, 132.2, 130.7, 130.04, 129.99, 129.87, 128.8, 128.74, 128.70, 128.5, 128.3, 128.2, 128.1, 127.93, 127.91, 127.58, 127.56, 127.45, 127.38, 127.3, 127.2, 127.0, 86.3, 77.5, 69.1, 61.7, 57.8, 43.4, 36.2, 30.3, 17.5, 13.9; **IR** (neat) ν 3023, 2931, 2860, 2221, 1706, 1592, 1459, 1241, 1117, 1052 cm⁻¹; **HRMS** (DART) Calcd for C₃₅H₃₂NO₂Cl₂ (MH⁺): 568.1805; found: 568.1799.



Ethyl 4-(benzhydryl(3-phenyl-3-(4-(trifluoromethyl)phenoxy)propyl)amino)but-2-ynoate (7b)

N-Benzhydryl-*N*-methyl-3-phenyl-3-(4-(trifluoromethyl)phenoxy)propan-1-amine **1q** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure B** using 1,2-bis(diphenylphosphino)ethane as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:19), **7b** was obtained as a colorless liquid (94 mg, 82%).

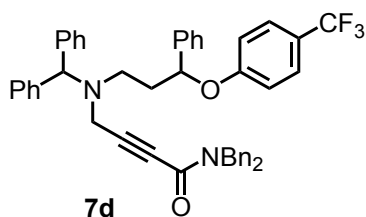
¹H NMR (500 MHz, CDCl₃) δ 7.46 – 7.39 (m, 4H), 7.35 – 7.22 (m, 10H), 7.18 (tt, *J* = 7.4, 1.5 Hz, 1H), 7.14 – 7.04 (m, 3H), 6.83 – 6.77 (m, 2H), 5.28 – 5.19 (m, 1H), 4.67 (s, 1H), 4.26 (q, *J* = 7.2, 2.4 Hz, 2H), 3.64 – 3.50 (m, 2H), 2.88 – 2.78 (m, 1H), 2.78 – 2.68 (m, 1H), 2.19 – 2.08 (m, 1H), 2.04 – 1.94 (m, 1H), 1.34 (t, *J* = 7.2, 2.4 Hz, 3H); **¹³C NMR** (126 MHz, CDCl₃) δ 160.5, 153.4, 142.13, 142.08, 141.2, 128.8, 128.6, 128.5, 127.9, 127.8, 127.3, 127.1, 126.7 (d, *J*_{CF} = 3.3 Hz), 125.6, 124.4 (d, *J*_{CF} = 271.1 Hz), 122.7 (q, *J*_{CF} = 32.7 Hz), 115.7, 83.3, 78.0, 77.7, 72.5, 61.9, 46.9, 39.5, 36.8, 14.0; **¹⁹F NMR** (470 MHz, CDCl₃) δ -61.53; **IR** (neat) ν 3028, 2930, 2834, 2221, 1708, 1612, 1513, 1451, 1324, 1246, 1114 cm⁻¹; **HRMS** (DART) Calcd for C₃₅H₃₃NO₃F₃(MH⁺): 572.2407; found: 572.2402.



Methyl 4-(benzhydryl(3-phenyl-3-(4-(trifluoromethyl)phenoxy)propyl)amino)but-2-ynoate (7c)

N-Benzhydryl-*N*-methyl-3-phenyl-3-(4-(trifluoromethyl)phenoxy)propan-1-amine **1q** was reacted with methyl 3-(trimethylsilyl)propiolate **2c** following **General Procedure B** using 1,2-bis(diphenylphosphino)ethane as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:19), **7c** was obtained as a colorless liquid (76 mg, 68%).

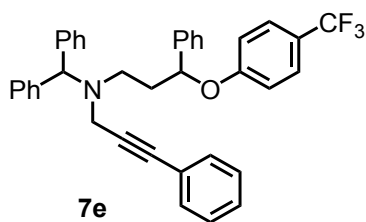
¹H NMR (500 MHz, CDCl₃) δ 7.46 – 7.37 (m, 4H), 7.36 – 7.21 (m, 9H), 7.18 (tt, *J* = 7.1, 1.5 Hz, 1H), 7.12 – 7.05 (m, 3H), 6.80 (d, *J* = 8.4 Hz, 2H), 5.23 (dd, *J* = 9.1, 3.6 Hz, 1H), 4.66 (s, 1H), 3.80 (s, 3H), 3.57 (d, *J* = 10.6 Hz, 2H), 2.88 – 2.67 (m, 2H), 2.20 – 1.95 (m, 2H); **¹³C NMR** (126 MHz, CDCl₃) δ 160.5, 153.8, 142.11, 142.05, 141.2, 128.8, 128.7, 128.6, 127.9, 127.79, 127.75, 127.3, 127.2, 126.7 (d, *J*_{CF} = 3.8 Hz), 125.6, 124.4 (d, *J*_{CF} = 271.3 Hz), 122.7 (q, *J*_{CF} = 32.5 Hz), 115.7, 83.7, 77.7, 72.5, 53.4, 52.7, 46.9, 39.5, 36.8; **¹⁹F NMR** (470 MHz, CDCl₃) δ -61.50; **IR** (neat) ν 3028, 2946, 2832, 2226, 1713, 1513, 1445, 1324, 1249, 1114 cm⁻¹; **HRMS** (DART) Calcd for C₃₄H₃₁NO₃F₃ (MH⁺): 558.2251; found: 558.2246.



4-(Benzhydryl(3-phenyl-3-(4-(trifluoromethyl)phenoxy)propyl)amino)-*N,N*-dibenzylbut-2-ynamide (7d)

N-Benzhydryl-*N*-methyl-3-phenyl-3-(4-(trifluoromethyl)phenoxy)propan-1-amine **1q** was reacted with *N,N*-dibenzyl-3-(trimethylsilyl)propiolamide **2d** following **General Procedure B** using (*S*)-PhPyBOX as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:4), **7d** was obtained as a colorless liquid (116 mg, 80%).

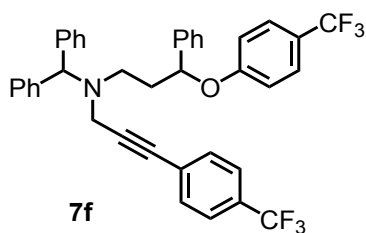
¹H NMR (600 MHz, CDCl₃) δ 7.40 (d, *J* = 8.0 Hz, 4H), 7.35 (t, *J* = 7.5 Hz, 3H), 7.31 (d, *J* = 6.8 Hz, 1H), 7.27 (d, *J* = 7.4 Hz, 4H), 7.23 (t, *J* = 9.3 Hz, 7H), 7.17 (t, *J* = 7.5 Hz, 2H), 7.15 – 7.10 (m, 3H), 7.06 (t, *J* = 7.3 Hz, 1H), 7.01 (t, *J* = 7.4 Hz, 2H), 6.75 (d, *J* = 8.4 Hz, 2H), 5.16 (dd, *J* = 9.0, 3.7 Hz, 1H), 4.78 – 4.65 (m, 2H), 4.55 (q, *J* = 14.8 Hz, 2H), 4.46 (s, 1H), 3.61 (q, *J* = 18.3 Hz, 2H), 2.75 – 2.57 (m, 2H), 2.13 – 1.90 (m, 2H); **¹³C NMR** (151 MHz, CDCl₃) δ 160.5, 154.7, 142.13, 142.06, 141.1, 136.2, 135.9, 129.0, 128.8, 128.7, 128.6, 128.5, 128.0, 127.8, 127.7, 127.6, 127.24, 127.20, 127.1, 126.6 (d, *J*_{CF} = 3.0 Hz), 125.6, 124.4 (d, *J*_{CF} = 271.1 Hz), 122.7 (q, *J*_{CF} = 32.7 Hz), 115.7, 87.7, 78.9, 77.7, 72.6, 51.3, 46.9, 46.5, 39.6, 36.9; **¹⁹F NMR** (470 MHz, CDCl₃) δ -61.48; **IR** (neat) ν 3028, 2925, 2830, 2221, 1628, 1324, 1245, 1162, 1113, 700 cm⁻¹; **HRMS** (DART) Calcd for C₄₇H₄₂N₂O₂F₃ (MH⁺): 723.3193; found: 723.3167.



***N*-Benzhydryl-3-phenyl-*N*-(3-phenyl-3-(4-(trifluoromethyl)phenoxy)propyl)prop-2-yn-1-amine (**7e**)**

N-Benzhydryl-*N*-methyl-3-phenyl-3-(4-(trifluoromethyl)phenoxy)propan-1-amine **1q** was reacted with trimethyl(phenylethynyl)silane **2e** following **General Procedure B** using (*S*)-PhPyBOX as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:49), **7e** was obtained as a colorless liquid (86 mg, 75%).

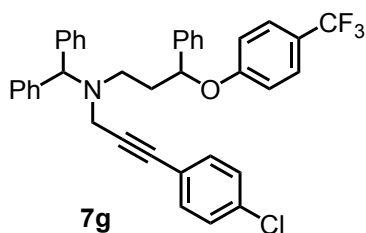
¹H NMR (500 MHz, CDCl₃) δ 7.53 – 7.38 (m, 6H), 7.37 – 7.14 (m, 13H), 7.08 (d, *J* = 6.6 Hz, 3H), 6.81 (d, *J* = 8.4 Hz, 2H), 5.30 (dd, *J* = 9.0, 3.8 Hz, 1H), 4.74 (s, 1H), 3.72 – 3.56 (m, 2H), 2.91 – 2.70 (m, 2H), 2.24 – 2.00 (m, 2H); **¹³C NMR** (126 MHz, CDCl₃) δ 160.6, 142.8, 142.7, 141.4, 131.7, 128.7, 128.5, 128.4, 128.3, 128.1, 127.9, 127.7, 127.1, 126.9, 126.6 (d, *J*_{CF} = 3.1 Hz), 125.7, 124.5 (d, *J*_{CF} = 271.5 Hz), 123.3, 122.6 (q, *J*_{CF} = 32.6 Hz), 115.7, 85.9, 84.2, 77.9, 72.6, 46.7, 39.9, 36.9; **¹⁹F NMR** (470 MHz, CDCl₃) δ -61.49; **IR** (neat) ν 3060, 2927, 2829, 2096, 1513, 1491, 1324, 1249, 1162, 1114, 698 cm⁻¹; **HRMS** (DART) Calcd for C₃₈H₃₃NOF₃ (MH⁺): 576.2509; found: 576.2504.



***N*-Benzhydryl-*N*-(3-phenyl-3-(4-(trifluoromethyl)phenoxy)propyl)-3-(4-(trifluoromethyl)phenyl)prop-2-yn-1-amine (**7f**)**

N-Benzhydryl-*N*-methyl-3-phenyl-3-(4-(trifluoromethyl)phenoxy)propan-1-amine **1q** was reacted with trimethyl((4-(trifluoromethyl)phenyl)ethynyl)silane **2f** following **General Procedure B** using (*S*)-PhPyBOX as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:19), **7f** was obtained as a colorless liquid (106 mg, 82%).

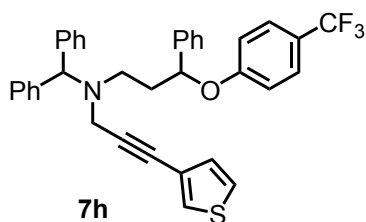
¹H NMR (500 MHz, CDCl₃) δ 7.57 (dd, *J* = 8.4, 4.1 Hz, 2H), 7.48 (dd, *J* = 8.1, 4.1 Hz, 2H), 7.43 (ddd, *J* = 13.0, 8.0, 4.1 Hz, 4H), 7.35 (tt, *J* = 4.9, 2.2 Hz, 2H), 7.29 (d, *J* = 4.8 Hz, 4H), 7.28 – 7.22 (m, 3H), 7.19 (td, *J* = 7.1, 4.4 Hz, 1H), 7.10 (p, *J* = 5.2 Hz, 3H), 6.81 (dd, *J* = 9.0, 4.0 Hz, 2H), 5.30 (dt, *J* = 8.5, 4.0 Hz, 1H), 4.73 (d, *J* = 4.2 Hz, 1H), 3.73 – 3.60 (m, 2H), 2.97 – 2.70 (m, 2H), 2.25 – 2.00 (m, 2H); **¹³C NMR** (126 MHz, CDCl₃) δ 160.6, 142.6, 142.5, 141.3, 132.4, 132.2 (d, *J*_{CF} = 293.4 Hz), 129.8 (q, *J*_{CF} = 32.6 Hz), 128.8, 128.6, 128.5, 128.0, 127.9, 127.8, 127.2, 127.1, 126.7 (d, *J*_{CF} = 2.9 Hz), 125.7, 125.2 (d, *J*_{CF} = 2.8 Hz), 124.2 (d, *J*_{CF} = 211.0 Hz), 122.7 (q, *J*_{CF} = 32.5 Hz), 115.7, 87.1, 84.6, 77.9, 72.6, 46.8, 40.0, 36.9; **¹⁹F NMR** (470 MHz, CDCl₃) δ -61.45, -62.75; **IR** (neat) ν 3027, 2931, 2829, 1612, 1513, 1450, 1322, 1249, 1116, 701 cm⁻¹; **HRMS** (DART) Calcd for C₃₉H₃₂NOF₆ (MH⁺): 644.2383; found: 644.2365.



***N*-Benzhydryl-3-(4-chlorophenyl)-*N*-(3-phenyl-3-(4-(trifluoromethyl)phenoxy)propyl)prop-2-yn-1-amine (**7g**)**

N-Benzhydryl-*N*-methyl-3-phenyl-3-(4-(trifluoromethyl)phenoxy)propan-1-amine **1q** was reacted with ((4-chlorophenyl)ethynyl)trimethylsilane **2g** following **General Procedure B** using (*S*)-PhPyBOX as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:19), **7g** was obtained as a colorless liquid (98 mg, 80%).

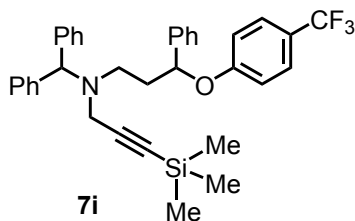
¹H NMR (600 MHz, CDCl₃) δ 7.46 – 7.37 (m, 4H), 7.29 (dddd, *J* = 22.3, 20.6, 9.1, 7.4 Hz, 13H), 7.18 (ddd, *J* = 7.4, 6.1, 1.4 Hz, 1H), 7.13 – 7.03 (m, 3H), 6.81 (d, *J* = 8.4 Hz, 2H), 5.29 (dd, *J* = 9.0, 3.8 Hz, 1H), 4.71 (s, 1H), 3.63 (q, *J* = 17.7 Hz, 2H), 2.93 – 2.65 (m, 2H), 2.25 – 1.97 (m, 2H); **¹³C NMR** (126 MHz, CDCl₃) δ 160.6, 142.7, 142.5, 141.4, 134.0, 132.9, 128.7, 128.6, 128.53, 128.45, 128.0, 127.9, 127.7, 127.1, 127.0, 126.7 (d, *J*_{CF} = 2.6 Hz), 125.7, 124.4 (d, *J*_{CF} = 271.1 Hz), 122.6 (q, *J*_{CF} = 32.8 Hz), 121.8, 115.7, 85.3, 84.7, 77.9, 72.6, 46.7, 40.0, 36.9; **¹⁹F NMR** (470 MHz, CDCl₃) δ -61.50; **IR** (neat) ν 3055, 2928, 2832, 2223, 1707, 1608, 1488, 1245, 1113, 701 cm⁻¹; **HRMS** (DART) Calcd for C₃₈H₃₂NOF₃Cl (MH⁺): 610.2119; found: 610.2125.



***N*-Benzhydryl-*N*-(3-phenyl-3-(4-(trifluoromethyl)phenoxy)propyl)-3-(thiophen-3-yl)prop-2-yn-1-amine (**7h**)**

N-Benzhydryl-*N*-methyl-3-phenyl-3-(4-(trifluoromethyl)phenoxy)propan-1-amine **1q** was reacted with trimethyl(thiophen-3-ylethynyl)silane **2h** following **General Procedure B** using (*S*)-PhPyBOX as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:49), **7h** was obtained as a colorless liquid (86 mg, 74%).

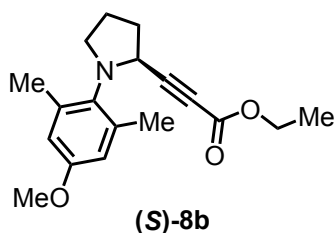
¹H NMR (600 MHz, CDCl₃) δ 7.43 (dt, *J* = 8.9, 5.0 Hz, 4H), 7.38 (d, *J* = 2.9 Hz, 1H), 7.34 (dd, *J* = 6.2, 3.4 Hz, 2H), 7.29 (t, *J* = 3.4 Hz, 4H), 7.25 (ddq, *J* = 9.9, 7.1, 4.3, 3.5 Hz, 4H), 7.18 (dt, *J* = 7.8, 3.8 Hz, 1H), 7.08 (dt, *J* = 8.8, 4.2 Hz, 4H), 6.81 (dd, *J* = 8.8, 2.5 Hz, 2H), 5.30 (dd, *J* = 9.1, 3.9 Hz, 1H), 4.72 (s, 1H), 3.62 (q, *J* = 17.8 Hz, 2H), 2.80 (ddt, *J* = 50.2, 12.0, 6.5 Hz, 2H), 2.12 (dt, *J* = 55.1, 12.1 Hz, 2H); **¹³C NMR** (151 MHz, CDCl₃) δ 160.6, 142.7, 142.6, 141.4, 130.0, 128.7, 128.5, 128.4, 128.3, 128.0, 127.9, 127.7, 127.1, 126.9, 126.7 (d, *J*_{CF} = 3.6 Hz), 124.7 (d, *J*_{CF} = 267.9 Hz), 125.7, 122.6 (d, *J*_{CF} = 32.9 Hz), 122.3, 115.7, 83.8, 80.8, 77.8, 72.5, 46.6, 39.9, 36.9; **¹⁹F NMR** (470 MHz, CDCl₃) δ -61.41; **IR** (neat) ν 3026, 2929, 2829, 2166, 1514, 1450, 1324, 1249, 1114, 700 cm⁻¹; **HRMS** (DART) Calcd for C₃₆H₃₁NOF₃S (MH⁺): 582.2073; found: 582.2074.



***N*-Benzhydryl-*N*-(3-phenyl-3-(4-(trifluoromethyl)phenoxy)propyl)-3-(trimethylsilyl)prop-2-yn-1-amine (**7i**)²¹**

N-Benzhydryl-*N*-methyl-3-phenyl-3-(4-(trifluoromethyl)phenoxy)propan-1-amine **1q** was reacted with 1,2-bis(trimethylsilyl)ethyne **2i** following **General Procedure B** using (*S*)-PhPyBOX as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:19), **7i** was obtained as a colorless liquid (99 mg, 87%).

¹H NMR (600 MHz, CDCl₃) δ 7.41 (dd, *J* = 20.0, 8.2 Hz, 4H), 7.32 – 7.27 (m, 6H), 7.23 (d, *J* = 8.0 Hz, 2H), 7.17 (d, *J* = 7.5 Hz, 2H), 7.10 – 7.05 (m, 3H), 6.80 (d, *J* = 8.4 Hz, 2H), 5.25 (dd, *J* = 9.2, 3.4 Hz, 1H), 4.64 (s, 1H), 3.42 (q, *J* = 17.9 Hz, 2H), 2.80 – 2.66 (m, 2H), 2.13 – 1.98 (m, 2H), 0.20 (s, 9H); **¹³C NMR**; ²¹ **¹⁹F NMR** (470 MHz, CDCl₃) δ -61.50; **IR** (neat) ν 3059, 2951, 2848, 2160, 1612, 1324, 1250, 1116, 840, 700 cm⁻¹; **HRMS** (DART) Calcd for C₃₅H₃₇NOF₃Si(MH⁺): 572.2591; found: 572.2579.



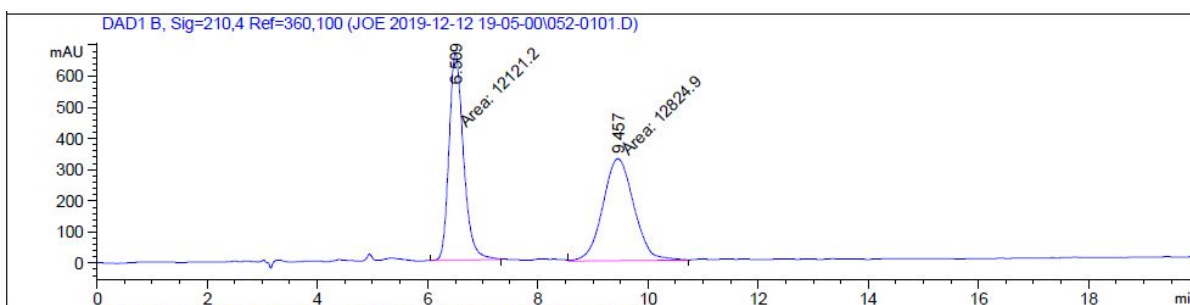
Ethyl (*S*)-3-(1-(4-methoxy-2,6-dimethylphenyl)pyrrolidin-2-yl)propiolate ((*S*)-8b)

1-(4-Methoxy-2,6-dimethylphenyl)pyrrolidine **1b** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure C**. After purification by column chromatography (Et₂O:hexanes = 1:19), (**S**)-**8b** was obtained as a colorless liquid (45 mg, 75%). The absolute configuration of (**S**)-**8b** was assigned as (*S*) as described in SI Section 4.

¹H NMR (500 MHz, CDCl₃) δ 6.58 (s, 2H), 4.27 (ddd, *J* = 7.8, 3.5, 1.3 Hz, 1H), 4.17 (qd, *J* = 7.2, 1.3 Hz, 2H), 3.75 (s, 3H), 3.43 – 3.31 (m, 1H), 3.09 (q, *J* = 6.9 Hz, 1H), 2.47 – 2.22 (m, 6H), 2.22 – 2.10 (m, 3H), 2.09 – 1.94 (m, 1H), 1.27 (td, *J* = 7.2, 1.4 Hz, 3H); **¹³C NMR** (151 MHz, CDCl₃) δ 157.2, 153.3, 136.6, 113.2, 91.5, 73.9, 63.3, 55.9, 52.3, 51.0, 35.8, 26.7, 18.4, 15.1; **IR** (neat) ν 2967, 2837, 2224, 1707, 1599, 1476, 1366, 1244, 1153, 1067, cm⁻¹; **HRMS** (DART) Calcd for C₁₈H₂₄NO₃ (MH⁺): 302.1751; found: 302.1755; [α]_D²⁵ = 76.2° (*c* 1.0, CH₂Cl₂); **HPLC** (Chiralcel OJ-H; 95:5 hexane:isopropanol, 1.0 mL/min; (**S**)-**8b**: tr = 6.5 min (major), 9.5 min (minor); 95:5 er.

```

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                                           Inj Volume: 4.000 µl
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                  0mL.M (Sequence Method)
Last changed    : 12/12/2019 7:05:05 PM by SYSTEM
Method Info     : Column4 60min-1% iPrOH 99% hexane-1.0mL
  
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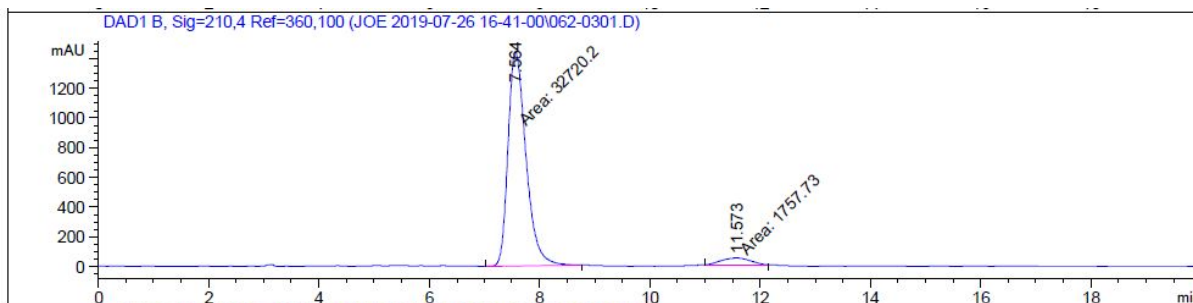


Signal 2: DAD1 B, Sig=210,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
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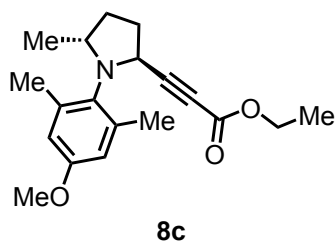
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=====
Acq. Operator   : SYSTEM                      Seq. Line :    3
Acq. Instrument : Wasa_LC1                    Location  :    62
Injection Date  : 7/26/2019 5:44:53 PM        Inj       :    1
                                           Inj Volume: 4.000 µl
Method          : C:\Chem32\1\Data\JOE 2019-07-26 16-41-00\column4 5%IPA 95% hexane 30min-1.
                                           0mL.M (Sequence Method)
Last changed    : 7/26/2019 4:41:04 PM by SYSTEM
Method Info     : Column4 60min-1% iPrOH 99% hexane-1.0mL
  
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Signal 2: DAD1 B, Sig=210,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	7.564	MM	0.3774	3.27202e4	1445.16772	94.9019
2	11.573	MM	0.6270	1757.73438	46.72317	5.0981



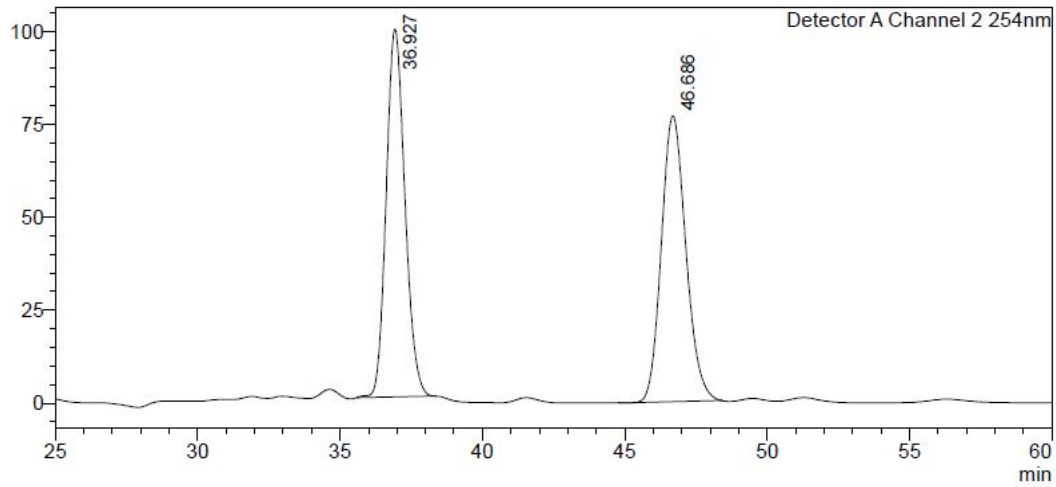
Ethyl 3-((2*S*,5*R*)-1-(4-methoxy-2,6-dimethylphenyl)-5-methylpyrrolidin-2-yl)propionate (8c)

1-(4-Methoxy-2,6-dimethylphenyl)-2-methylpyrrolidine *rac*-**1r** was reacted with ethyl 3-(trimethylsilyl)propionate **2b** following **General Procedure C**. The *trans*:*cis* ratio was determined to be 6.3:1 by ¹H NMR analysis of the unpurified reaction mixtures. After purification by column chromatography (Et₂O:hexanes = 1:19), **8c** was obtained as a yellow oil (42 mg, 66%). The relative configuration of **8c** was assigned by NOESY analysis as described in SI Section 3.

¹H NMR (600 MHz, CDCl₃) δ 6.58 (d, *J* = 1.2 Hz, 2H), 4.37 (dd, *J* = 7.7, 4.1 Hz, 1H), 4.15 (qd, *J* = 7.1, 0.8 Hz, 2H), 3.84 (q, *J* = 6.1 Hz, 1H), 3.76 (s, 3H), 2.43 – 2.37 (m, 1H), 2.36 (s, 3H), 2.32 – 2.25 (m, 1H), 2.19 (s, 3H), 2.09 – 2.02 (m, 1H), 1.61 – 1.55 (m, 1H), 1.25 (d, *J* = 0.9 Hz, 3H), 0.86 (d, *J* = 6.2 Hz, 3H); **¹³C NMR** (151 MHz, CDCl₃) δ 156.8, 153.8, 140.2, 139.7, 133.6, 113.9, 113.2, 91.0, 74.6, 61.7, 55.2, 55.1, 52.2, 33.2, 31.4, 20.3, 19.9, 19.0, 14.0; **IR** (neat) ν 2956, 2222, 1705, 1597, 1471, 1368, 1234, 1152, 1065, 852 cm⁻¹; **HRMS** (DART) Calcd for C₁₉H₂₆NO₃ (MH⁺): 316.1907; found: 316.1905; [α]_D²⁵ = 12.1° (*c* 0.3, CH₂Cl₂); **HPLC** (Chiralpak AD-H; 99.7:0.3 hexane:isopropanol, 0.3 mL/min; **8c**: tr = 36.9 min (minor), 46.7 min (major); 83:17 er.

Sample Name	: 8b_rac1_aMe_f6_AD-H_inst4_0.3mL_3	Sample Type	: Unknown
Sample ID	: 8b_rac1_aMe_f6_AD-H_inst4_0.3mL_3	Acquired by	: System Administrator
Data Filename	: 8b_rac1_aMe_f6_AD-H_inst4_0.3mL_3.lcd	Processed by	: System Administrator
Method Filename	: 99.7_0.3_60min_0.3mL.lcm		
Batch Filename	: batch9.lcb		
Vial #	: 1-20		
Injection Volume	: 30 uL		
Date Acquired	: 2/13/2020 10:43:40 PM		
Date Processed	: 2/13/2020 11:43:41 PM		

mV

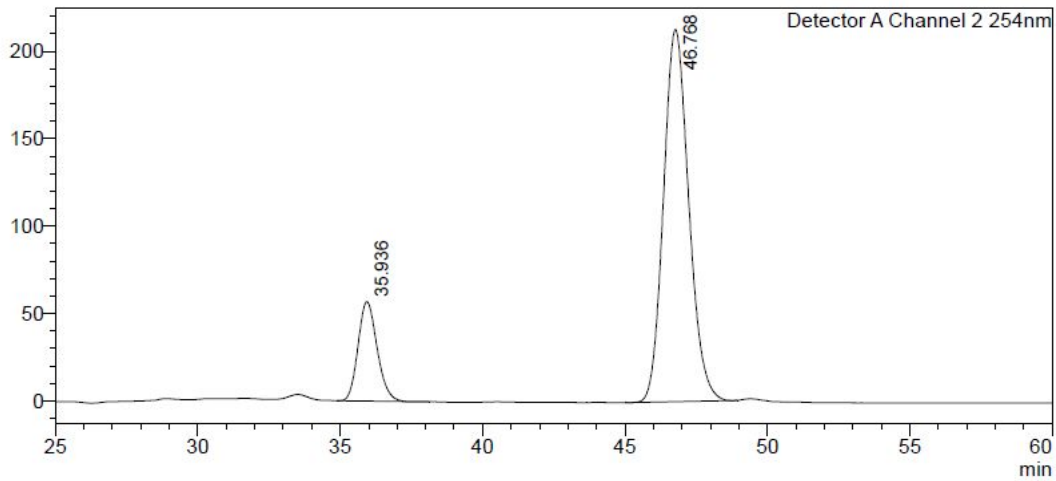


Detector A Channel 2 254nm			
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2	46.686	4589671	50.359
Total		9113925	100.000

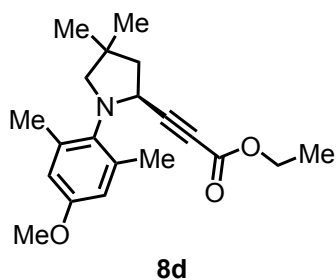
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 Sample ID : 8b_ee2_JZC2264_f9_AD-H_inst4_0.
 Data Filename : 8b_ee2_JZC2264_f9_AD-H_inst4_0.3mL.lcd
 Method Filename : 99.7_0.3_60min_0.3mL.lcm
 Batch Filename : batch9.lcb
 Vial # : 1-21
 Injection Volume : 20 uL
 Date Acquired : 2/13/2020 7:34:16 PM
 Date Processed : 2/13/2020 8:34:17 PM

Sample Type : Unknown
 Acquired by : System Administrator
 Processed by : System Administrator

mV



Detector A Channel 2 254nm			
Peak#	Ret. Time	Area	Area%
1	35.936	2651131	16.814
2	46.768	13116228	83.186
Total		15767359	100.000



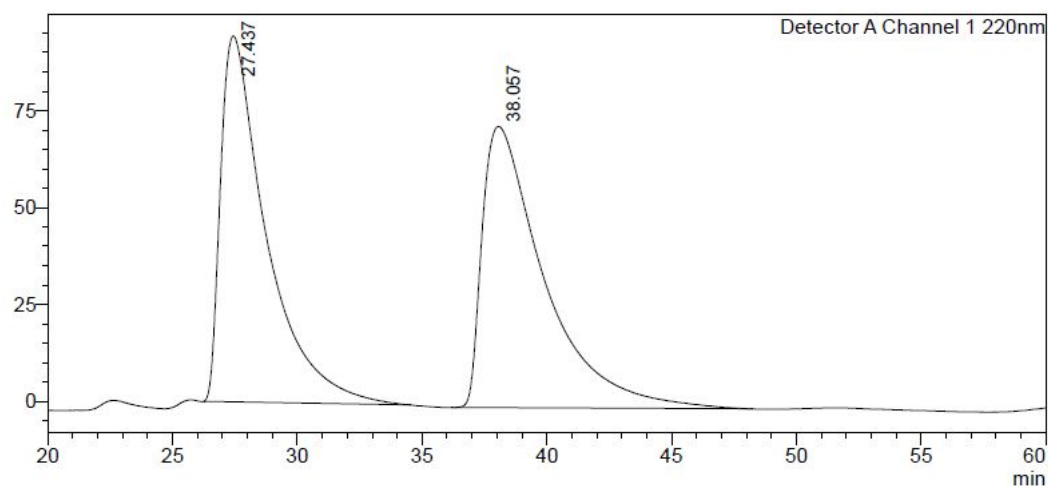
Ethyl (S)-3-(1-(4-methoxy-2,6-dimethylphenyl)-4,4-dimethylpyrrolidin-2-yl)propiolate (8d)

1-(4-Methoxy-2,6-dimethylphenyl)-3,3-dimethylpyrrolidine **1c** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure C**. After purification by column chromatography (Et₂O:hexanes = 1:19), **8d** was obtained as a yellow oil (45 mg, 69%). The absolute configuration of **8d** was assigned in analogy to (**S**)-**8b** (see SI Section 4).

¹H NMR (600 MHz, CDCl₃) δ 6.57 (s, 2H), 4.41 (dd, *J* = 9.0, 3.8 Hz, 1H), 4.15 (q, *J* = 7.1 Hz, 2H), 3.75 (s, 3H), 3.07 (d, *J* = 7.9 Hz, 1H), 2.95 (d, *J* = 7.8 Hz, 1H), 2.37 (s, 5H), 2.19 (dd, *J* = 12.5, 9.0 Hz, 2H), 1.98 (dd, *J* = 12.5, 3.9 Hz, 1H), 1.37 (s, 3H), 1.26 (t, *J* = 7.2 Hz, 3H), 1.15 (d, *J* = 1.0 Hz, 3H); **¹³C NMR** (151 MHz, CDCl₃) δ 157.2, 153.8, 135.4, 113.7, 91.0, 74.3, 64.5, 61.6, 55.2, 51.9, 47.0, 39.1, 27.8, 27.2, 14.0; **IR** (neat) ν 2954, 2864, 2228, 1707, 1601, 1465, 1243, 1094, 1023, 751 cm⁻¹; **HRMS** (DART) Calcd for C₂₀H₂₈NO₃ (MH⁺): 330.2063; found: 330.2069; [α]_D²⁵ = 20.5° (*c* 0.3, CH₂Cl₂); **HPLC** (Chiralpak AY-3; 99.9:0.1 hexane:isopropanol, 0.3 mL/min; **8d**: tr = 27.4 min (major), 38.1 min (minor); 93:7 er.

Sample Name	: 8c_rac5_JZC2152A_n2n1_AY-3_2-10	Sample Type	: Unknown
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Data Filename	: 8c_rac5_JZC2152A_n2n1_AY-3_2-10.lcd	Processed by	: System Administrator
Method Filename	: 99.9_0.1_60min_0.3mL.lcm		
Batch Filename	: batch8.lcb		
Vial #	: 1-24		
Injection Volume	: 10 uL		
Date Acquired	: 2/11/2020 2:39:47 AM		
Date Processed	: 2/11/2020 10:17:04 AM		

mV



Detector A Channel 1 220nm

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Total		24511111	100.000

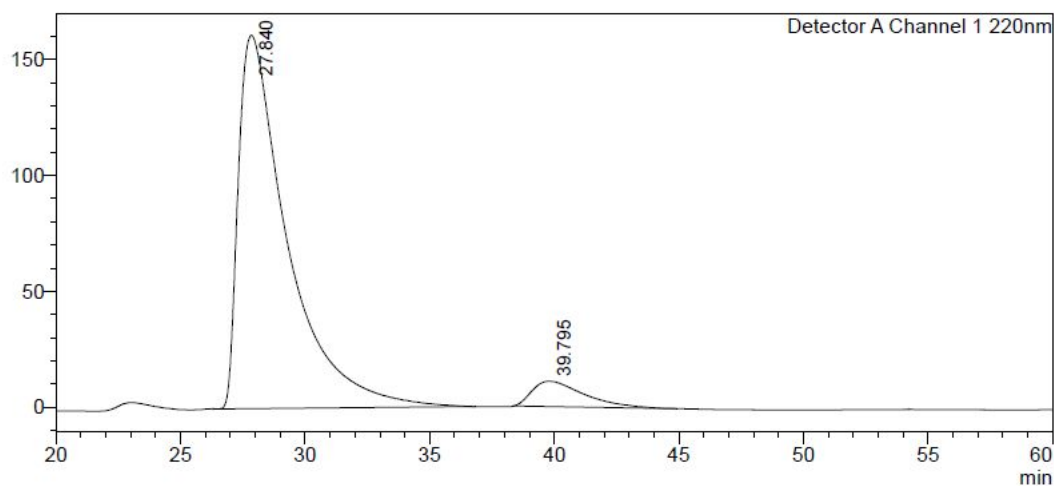
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 Sample ID : 8c_ee4_JZC2101A_AY-3_2-10
 Data Filename : 8c_ee4_JZC2101A_AY-3_2-10.lcd
 Method Filename : 99.9_0.1_60min_0.3mL.lcm
 Batch Filename : batch8.lcb
 Vial # : 1-25
 Injection Volume : 4 uL
 Date Acquired : 2/11/2020 3:40:02 AM
 Date Processed : 2/12/2020 4:33:05 PM

Sample Type : Unknown

Acquired by : System Administrator

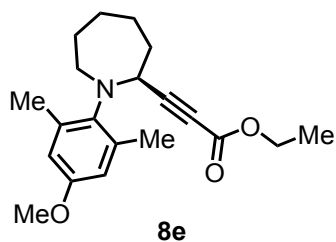
Processed by : System Administrator

mV



Detector A Channel 1 220nm

Peak#	Ret. Time	Area	Area%
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Total		23599165	100.000



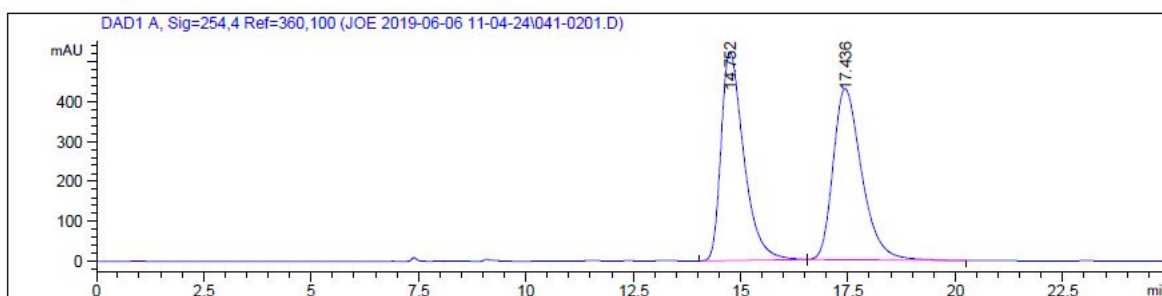
Ethyl (*S*)-3-(1-(4-methoxy-2,6-dimethylphenyl)azepan-2-yl)propiolate (8e**)**

1-(4-Methoxy-2,6-dimethylphenyl)azepane **1d** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure C**. After purification by column chromatography (Et₂O:hexanes = 1:19), **8e** was obtained as a colorless liquid (42 mg, 64%). The absolute configuration of **8e** was assigned in analogy to (*S*)-**8b** (see SI Section 4).

¹H NMR (500 MHz, CDCl₃) δ 6.57 (d, *J* = 13.4 Hz, 2H), 4.18 (d, *J* = 7.1 Hz, 2H), 4.06 (d, *J* = 2.7 Hz, 1H), 3.75 (s, 3H), 3.32 (d, *J* = 9.1 Hz, 1H), 3.08 (d, *J* = 11.1 Hz, 1H), 2.42 (s, 3H), 2.29 (s, 3H), 2.19 (d, *J* = 1.5 Hz, 1H), 1.95 – 1.85 (m, 2H), 1.85 – 1.79 (m, 1H), 1.79 – 1.70 (m, 2H), 1.57 (s, 2H), 1.28 (s, 3H); **¹³C NMR** (151 MHz, CDCl₃) δ 157.4, 153.8, 140.9, 139.7, 137.9, 114.1, 113.6, 90.3, 75.6, 59.4, 55.6, 53.0, 51.3, 34.8, 31.4, 28.9, 25.2, 20.1, 19.6, 14.0; **IR** (neat) ν 2926, 2845, 2221, 1707, 1598, 1474, 1309, 1240, 1065, 853 cm⁻¹; **HRMS** (DART) Calcd for C₂₀H₂₈NO₃ (MH⁺): 330.2063; found: 330.2061; [α]_D²⁵ = 30.1° (c 1.0, CH₂Cl₂); **HPLC** (Chiralcel OJ-H; 98.5:1.5 hexane:isopropanol, 0.5 mL/min; **8e**: tr = 14.8 min (major), 17.4 min (minor); 95:5 er.

```

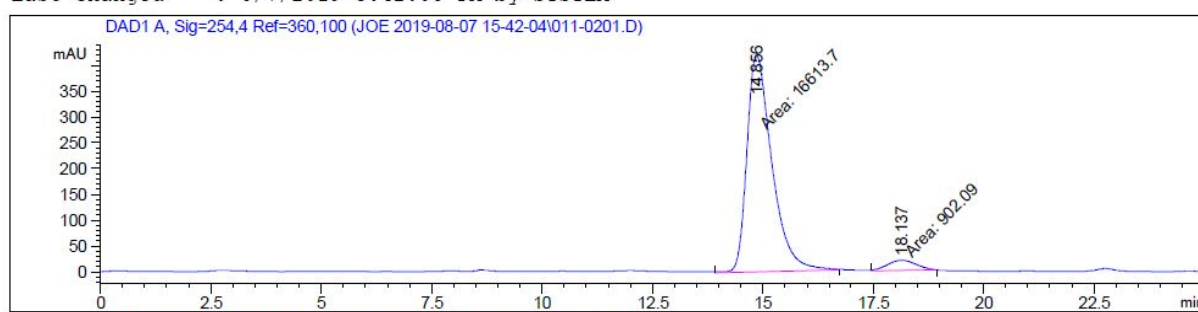
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Acq. Instrument : Wasa_LC1                             Location  :   41
Injection Date  : 6/6/2019 11:37:23 AM                 Inj       :    1
                                                    Inj Volume : 4.000 µl
Acq. Method     : C:\Chem32\1\Data\JOE 2019-06-06 11-04-24\column4 1.5%IPA 98.5% hexane 60min
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Last changed    : 6/6/2019 11:04:28 AM by SYSTEM
  
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Signal 1: DAD1 A, Sig=254,4 Ref=360,100

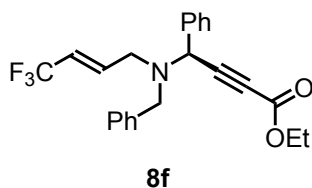
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
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2	17.436	BB	0.7122	1.98270e4	428.21890	50.7942

Acq. Operator : SYSTEM Seq. Line : 2
 Acq. Instrument : Wasa_LC1 Location : 11
 Injection Date : 8/7/2019 4:14:59 PM Inj : 1
 Inj Volume : 4.000 µl
 Acq. Method : C:\Chem32\1\Data\JOE 2019-08-07 15-42-04\column4 1.5%IPA 98.5% hexane 30min
 -0.5mL.M
 Last changed : 8/7/2019 3:42:08 PM by SYSTEM



Signal 2: DAD1 B, Sig=210,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	14.856	MM	0.6745	9.39466e4	2321.40063	95.4662
2	18.137	MM	0.6770	4461.63379	109.84474	4.5338



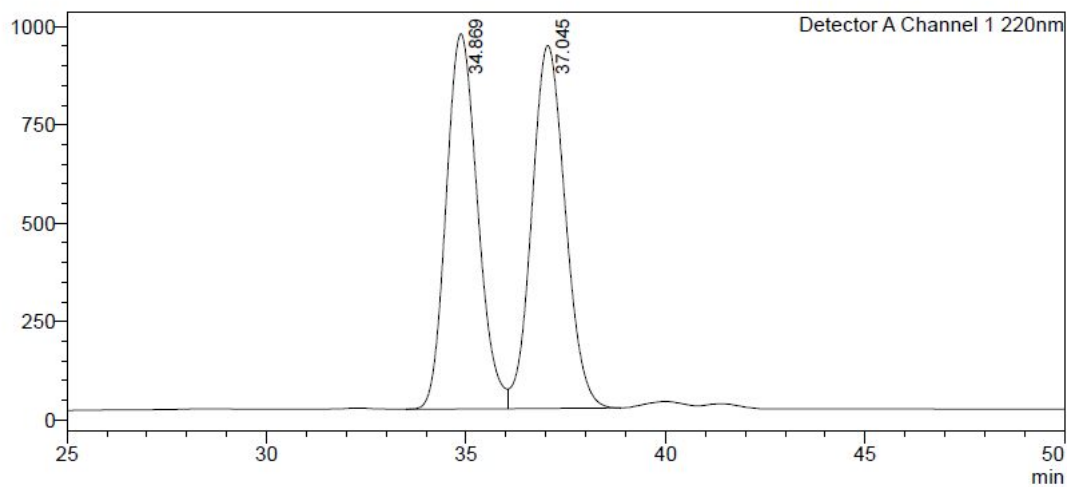
Ethyl (*S,E*)-4-(benzyl(4,4,4-trifluorobut-2-en-1-yl)amino)-4-phenylbut-2-ynoate (8f**)**

(*E*)-*N,N*-Dibenzyl-4,4,4-trifluorobut-2-en-1-amine **1s** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure C** using (*S*)-PhPyBOX as the ligand. After purification by column chromatography (Et₂O:hexanes = 1:19), **8f** was obtained as a colorless liquid (36 mg, 45%).

¹H NMR (600 MHz, CDCl₃) δ 7.56 (dt, *J* = 8.1, 1.1 Hz, 2H), 7.40 – 7.31 (m, 7H), 7.31 – 7.25 (m, 1H), 6.33 (dddd, *J* = 15.8, 7.5, 4.2, 2.1 Hz, 1H), 5.94 – 5.84 (m, 1H), 4.87 (s, 1H), 4.32 (q, *J* = 7.1 Hz, 2H), 3.83 (d, *J* = 13.5 Hz, 1H), 3.54 (d, *J* = 13.5 Hz, 1H), 3.28 (dq, *J* = 15.6, 3.2 Hz, 1H), 3.21 – 3.14 (m, 1H), 1.38 (t, *J* = 7.1 Hz, 3H); **¹³C NMR** (151 MHz, CDCl₃) δ 153.4, 138.1, 138.03, 137.99, 137.95, 137.93, 136.5, 128.84, 128.81, 128.68, 128.65, 128.60, 128.57, 128.5, 128.42, 128.36, 128.3, 128.2, 128.0, 127.9, 127.6, 127.21, 127.16, 123.7, 121.9, 120.7, 120.5, 120.3, 120.0, 82.8, 80.5, 62.3, 58.3, 56.3, 55.4, 53.5, 50.6, 14.1; **¹⁹F NMR** (564 MHz, CDCl₃) δ -64.07, -64.08, -64.08, -64.09, -64.09, -64.10; **IR** (neat) ν 2925, 2222, 1711, 1492, 1450, 1242, 1119, 749, 698 cm⁻¹; **HRMS** (DART) Calcd for C₂₃H₂₃NO₂F₃ (MH⁺): 402.1675; found: 402.1667; [α]_D²⁵ = -45.3° (*c* 1.0, CH₂Cl₂); **HPLC** (Chiralcel AD-H; 99.5:0.5 hexane:isopropanol, 0.3 mL/min; **8f**: tr = 34.9 min (minor), 37.0 min (major); 84:16 er.

Sample Name	: 8e_rac3_JZC2017H2_AD-H_0.3mL	Sample Type	: Unknown
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Method Filename	: 995_05_60min_03mL.lcm		
Batch Filename	: batch9.lcb		
Vial #	: 1-4		
Injection Volume	: 4 uL		
Date Acquired	: 2/9/2020 5:43:00 PM		
Date Processed	: 2/10/2020 9:51:53 AM		

mV

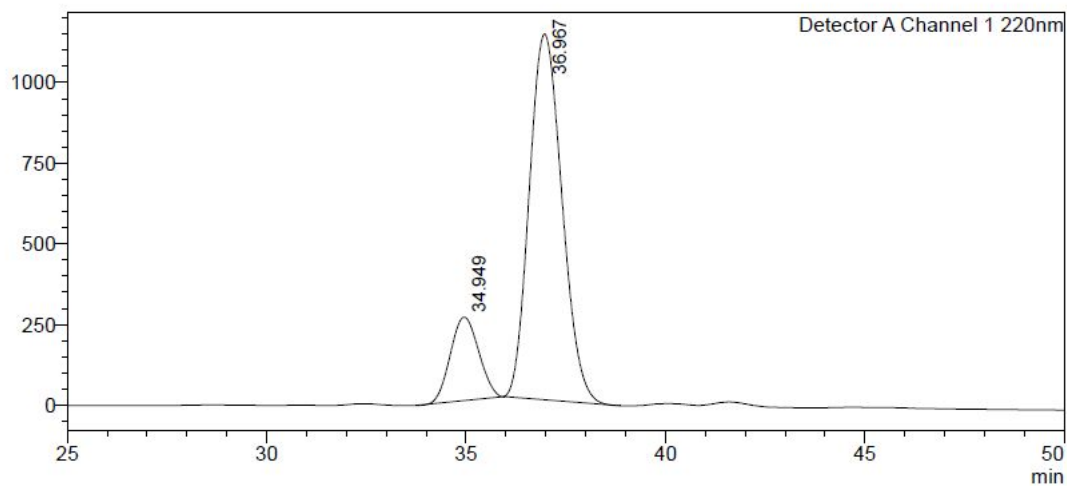


Detector A Channel 1 220nm			
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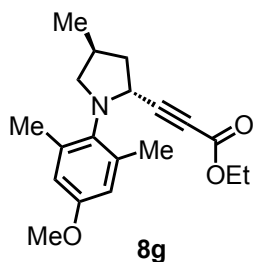
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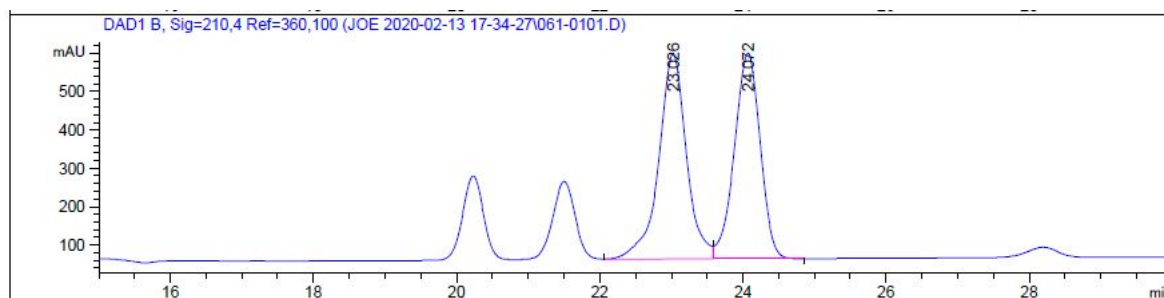


Ethyl 3-((2R,4S)-1-(4-methoxy-2,6-dimethylphenyl)-4-methylpyrrolidin-2-yl)propionate (8g)

(S)-1-(4-Methoxy-2,6-dimethylphenyl)-3-methylpyrrolidine (**S**)-**1t** was reacted with ethyl 3-(trimethylsilyl)propionate **2b** following **General Procedure C**. The *trans:cis* ratio was determined to be 11.8:1 by ^1H NMR analysis of the unpurified reaction mixtures. After purification by column chromatography (Et_2O :hexanes = 1:19), **8g** was obtained as a colorless liquid (40 mg, 64%). The relative configuration of **8g** was assigned by NOESY, COSY, and HSQC analysis (see SI Section 3).

^1H NMR (600 MHz, CDCl_3) δ 6.58 (s, 2H), 4.29 (dd, J = 8.6, 1.8 Hz, 1H), 4.17 (qd, J = 7.1, 1.0 Hz, 2H), 3.75 (d, J = 1.0 Hz, 3H), 3.38 (t, J = 7.2 Hz, 1H), 2.74 (dd, J = 8.8, 7.8 Hz, 1H), 2.71 – 2.62 (m, 1H), 2.42 – 2.18 (m, 7H), 2.03 – 1.94 (m, 1H), 1.27 (td, J = 7.1, 0.9 Hz, 3H), 1.11 (dd, J = 6.5, 0.9 Hz, 3H); ^{13}C NMR (151 MHz, CDCl_3) δ 157.1, 153.9, 135.8, 113.6, 91.2, 74.1, 61.7, 58.4, 55.2, 52.7, 41.6, 33.6, 18.8, 17.3, 14.0; IR (neat) ν 2954, 2924, 1708, 1601, 1484, 1465, 1244, 1154, 1066 cm^{-1} ; HRMS (DART) Calcd for $\text{C}_{19}\text{H}_{26}\text{NO}_3$ (MH^+): 316.1907; found: 316.1904; $[\alpha]^{25}_D$ = 63.0° (c 0.2, CH_2Cl_2); HPLC (Chiralpak IA; 97.5:2.5 hexane:isopropanol, 0.2 mL/min; **8g**: tr = 23.0 min (major), 24.0 min (minor); 97:3 er.

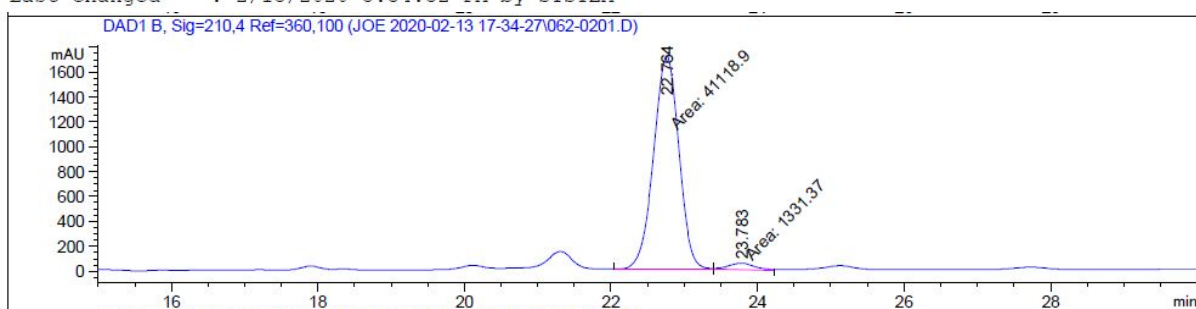
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                                           Inj Volume: 4.000 µl
Acq. Method     : C:\Chem32\1\Data\JOE 2020-02-13 17-34-27\column6 2.5%IPA 97.5% hexane 40min
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Signal 2: DAD1 B, Sig=210,4 Ref=360,100

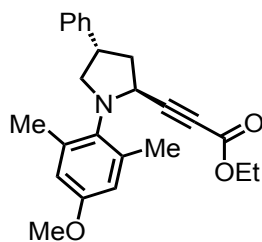
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2	24.072	VB	0.3885	1.34559e4	536.65869	48.2618

Acq. Operator : SYSTEM Seq. Line : 2
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Signal 2: DAD1 B, Sig=210,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	22.764	MM	0.3995	4.11189e4	1715.43152	96.8637
2	23.783	MM	0.4273	1331.37109	51.93531	3.1363



8h

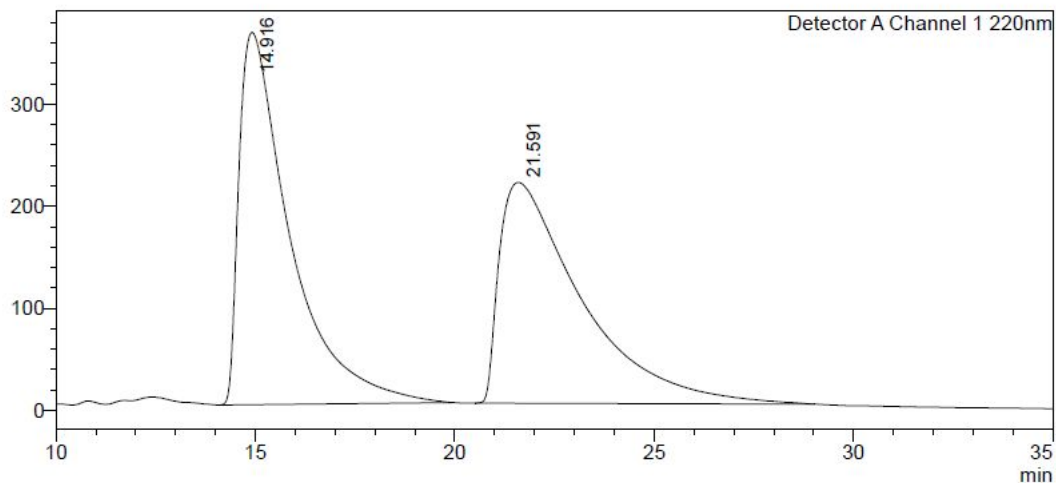
Ethyl 3-((2*S*,4*S*)-1-(4-methoxy-2,6-dimethylphenyl)-4-phenylpyrrolidin-2-yl)propionate (8h)

(*S*)-1-(4-Methoxy-2,6-dimethylphenyl)-3-phenylpyrrolidine (**S**)-**1u** was reacted with ethyl 3-(trimethylsilyl)propionate **2b** following **General Procedure C**. The *trans:cis* ratio was determined to be 10.1:1 by ¹H NMR analysis of the unpurified product mixtures. After purification by column chromatography (Et₂O:hexanes = 1:19), **8h** was obtained as a colorless liquid (51 mg, 68%). The absolute and relative configuration of **8h** was assigned in analogy to **8g** (see SI Section 3).

¹H NMR (600 MHz, CDCl₃) δ 7.33 (t, *J* = 7.6 Hz, 2H), 7.31 – 7.27 (m, 2H), 7.24 (dt, *J* = 6.8, 1.7 Hz, 1H), 6.61 (s, 2H), 4.44 (dd, *J* = 7.3, 2.8 Hz, 1H), 4.20 (q, *J* = 7.1 Hz, 2H), 3.84 (ddd, *J* = 17.7, 10.2, 7.5 Hz, 1H), 3.76 (s, 3H), 3.64 (t, *J* = 7.6 Hz, 1H), 3.25 (dd, *J* = 10.0, 8.0 Hz, 1H), 2.56 – 2.50 (m, 2H), 2.50 – 2.14 (m, 6H), 1.29 (t, *J* = 7.2 Hz, 3H); **¹³C NMR** (151 MHz, CDCl₃) δ 157.4, 153.9, 141.1, 135.5, 128.6, 127.9, 127.2, 126.7, 113.7, 90.6, 74.4, 61.8, 57.7, 55.2, 52.9, 44.1, 40.5, 18.9, 14.0; **IR** (neat) ν 2196, 2847, 2226, 1701, 1601, 1485, 1243, 1153, 1037, 699 cm⁻¹; **HRMS** (DART) Calcd for C₂₄H₂₈NO₃ (MH⁺): 378.2064; found: 378.2063; [α]²⁵_D = –25.5° (*c* 0.6, CH₂Cl₂); **HPLC** (Chiralpak AY-3; 99.5:0.5 hexane:isopropanol, 0.5 mL/min; **8h**: tr = 14.9 min (minor), 21.6 min (major); 88:12 er.

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Method Filename	: 995_05_40min_05mL.lcm		
Batch Filename	: batch8.lcb		
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mV

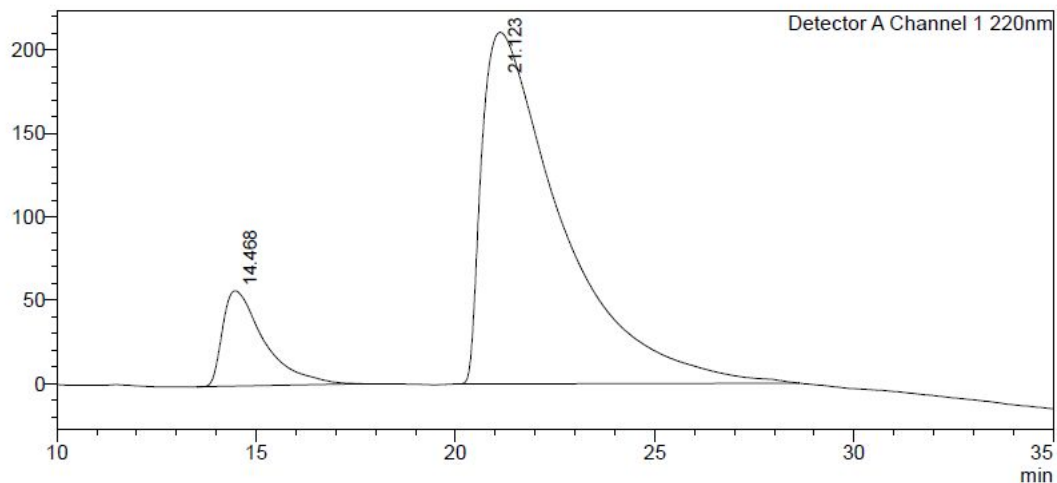


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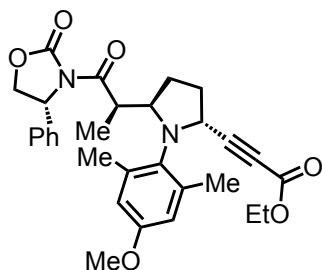
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Sample Type : Unknown
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 Processed by : System Administrator

mV



Detector A Channel 1 220nm			
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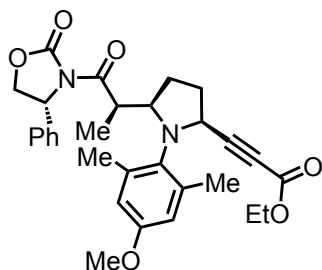


8i-trans

Ethyl 3-((2*R*,5*R*)-1-(4-methoxy-2,6-dimethylphenyl)-5-((*R*)-1-oxo-1-((*R*)-2-oxo-4-phenyloxazolidin-3-yl)propan-2-yl)pyrrolidin-2-yl)propanoate (8i-trans)

(*R*)-3-((*R*)-2-((*R*)-1-(4-Methoxy-2,6-dimethylphenyl)pyrrolidin-2-yl)propanoyl)-4-phenyloxazolidin-2-one (***R,R,R***)-**1v** was reacted with ethyl 3-(trimethylsilyl)propiolate **2b** following **General Procedure C**. The *trans*:*cis* ratio was determined to be 7.7:1 by ^1H NMR analysis of the unpurified reaction mixtures. After purification by column chromatography (Et_2O :hexanes = 1:4), **8i-trans** was obtained as a colorless liquid (96 mg, 93%). The relative configuration of **8i-trans** was assigned by NOESY and COSY analysis (see SI Section 3).

^1H NMR (500 MHz, CDCl_3) δ 7.32 – 7.21 (m, 3H), 7.07 – 7.02 (m, 2H), 6.59 (q, J = 3.1 Hz, 2H), 4.56 (dd, J = 8.4, 2.4 Hz, 1H), 4.47 (t, J = 7.0 Hz, 1H), 4.21 (dt, J = 12.7, 8.5 Hz, 2H), 4.13 (q, J = 7.1 Hz, 2H), 3.98 (dd, J = 8.4, 2.4 Hz, 1H), 3.79 (s, 3H), 3.71 (dq, J = 9.0, 6.9 Hz, 1H), 2.42 (dtd, J = 12.3, 7.0, 3.8 Hz, 1H), 2.34 (s, 3H), 2.26 (s, 3H), 2.21 (dtd, J = 12.2, 6.3, 3.8 Hz, 1H), 2.11 (ddt, J = 12.3, 10.1, 7.0 Hz, 1H), 1.76 (ddt, J = 11.9, 9.8, 7.4 Hz, 1H), 1.24 (t, J = 7.1 Hz, 3H), 1.02 (d, J = 6.9 Hz, 3H); **^{13}C NMR** (151 MHz, CDCl_3) δ 174.2, 157.2, 153.6, 153.1, 143.7, 140.6, 139.5, 133.2, 128.9, 128.3, 125.1, 113.8, 112.4, 89.6, 74.8, 69.7, 62.4, 61.7, 57.4, 55.21, 55.20, 52.5, 42.1, 32.2, 30.3, 20.1, 19.5, 15.9, 13.9; **IR** (neat) ν 2960, 2847, 2226, 2172, 1775, 1699, 1597, 1380, 1240, 1039, 699 cm^{-1} ; **HRMS** (DART) Calcd for $\text{C}_{30}\text{H}_{35}\text{N}_2\text{O}_6$ (MH^+): 519.2489; found: 519. 2475; $[\alpha]^{25}_D$ = -62.2° (c 0.5, CH_2Cl_2).



8i-cis

Ethyl 3-((2*S*,5*R*)-1-(4-methoxy-2,6-dimethylphenyl)-5-((*R*)-1-oxo-1-((*R*)-2-oxo-4-phenyloxazolidin-3-yl)propan-2-yl)pyrrolidin-2-yl)propionate (8i-cis)

The relative configuration of **8i-cis** was assigned by NOESY analysis (see SI Section 3).

¹H NMR (600 MHz, CDCl₃) δ 7.39 – 7.21 (m, 3H), 7.08 (dt, *J* = 7.2, 2.3 Hz, 2H), 6.62 (d, *J* = 3.1 Hz, 1H), 6.53 (d, *J* = 3.2 Hz, 1H), 4.63 (dt, *J* = 8.7, 2.9 Hz, 1H), 4.37 (td, *J* = 8.5, 3.0 Hz, 1H), 4.19 (dt, *J* = 17.6, 7.4, 3.7 Hz, 3H), 4.07 (dt, *J* = 10.5, 2.8 Hz, 1H), 3.95 (q, *J* = 8.7 Hz, 1H), 3.83 – 3.67 (m, 4H), 2.50 (d, *J* = 3.0 Hz, 3H), 2.32 (tt, *J* = 12.3, 4.6 Hz, 1H), 2.25 – 2.12 (m, 4H), 2.07 (dh, *J* = 13.2, 3.5 Hz, 1H), 1.94 (ddd, *J* = 11.9, 7.8, 2.8 Hz, 1H), 1.27 (tt, *J* = 9.0, 4.2 Hz, 4H), 1.15 (dd, *J* = 7.2, 3.0 Hz, 2H); **¹³C NMR** (151 MHz, CDCl₃) δ 174.8, 157.5, 153.7, 153.1, 142.8, 140.8, 139.3, 134.7, 129.1, 129.0, 128.4, 125.2, 114.5, 112.8, 90.0, 74.4, 69.9, 63.8, 61.7, 57.4, 55.3, 53.9, 42.0, 32.2, 29.0, 19.9, 19.4, 14.9, 14.0; **IR** (neat) ν 2876, 2848, 2232, 1779, 1704, 1600, 1465, 1382, 1246, 1194, 1066, 1041, 702 cm⁻¹; **HRMS** (DART) Calcd for C₃₀H₃₅N₂O₆ (MH⁺): 519.2489; found: 519.2483; [α]_D²⁵ = −108.6° (*c* 0.4, CH₂Cl₂).

7. References

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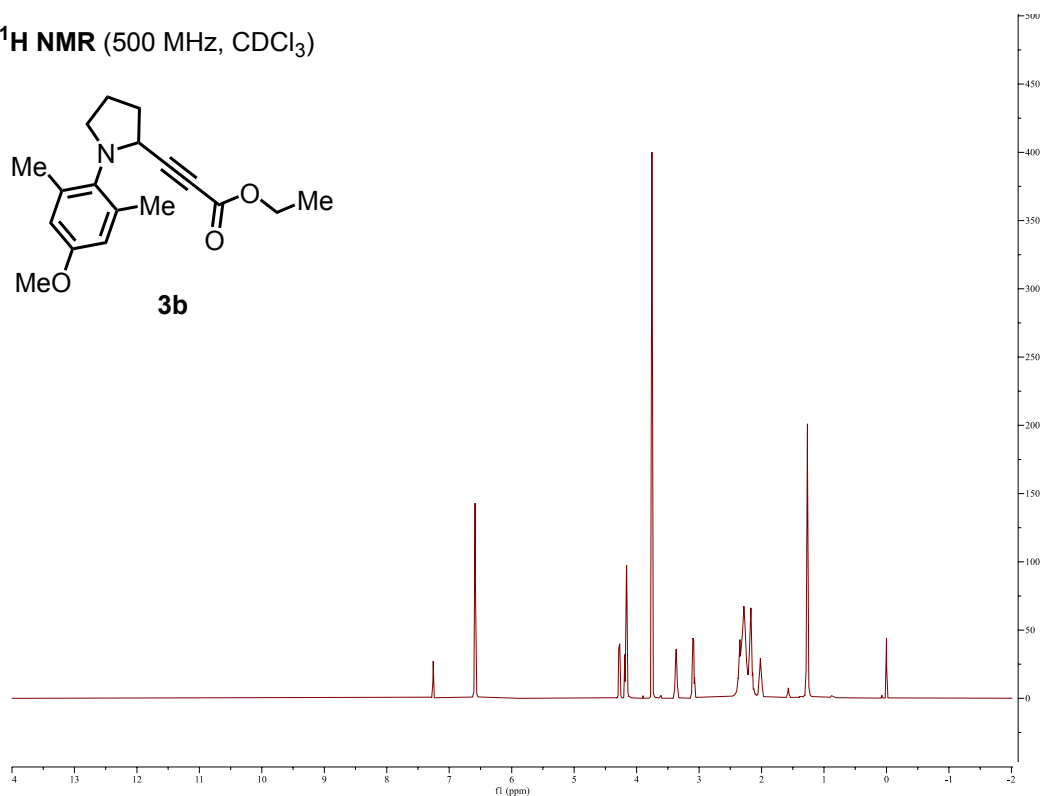
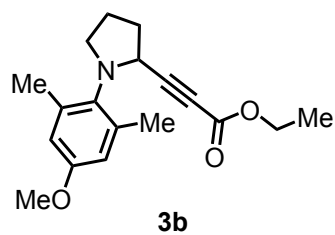
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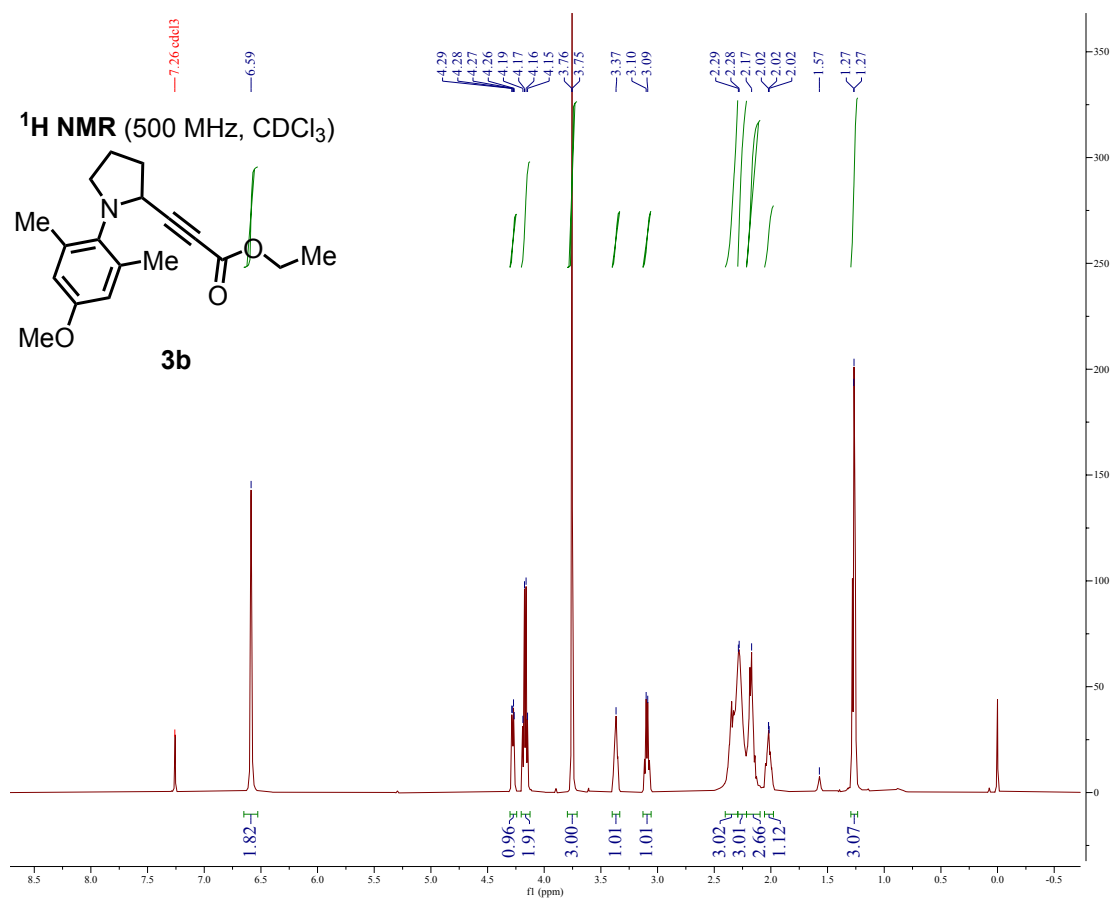
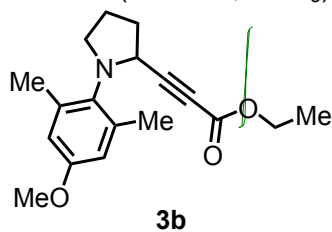
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8. NMR Spectra

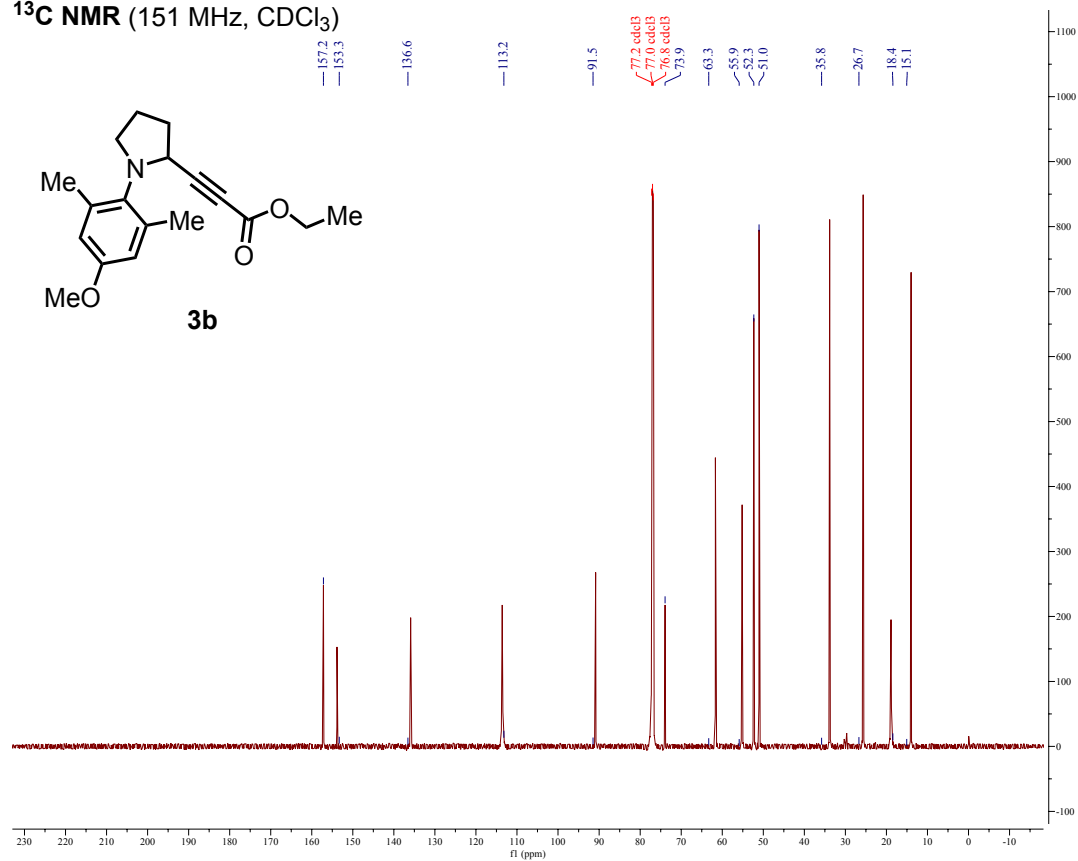
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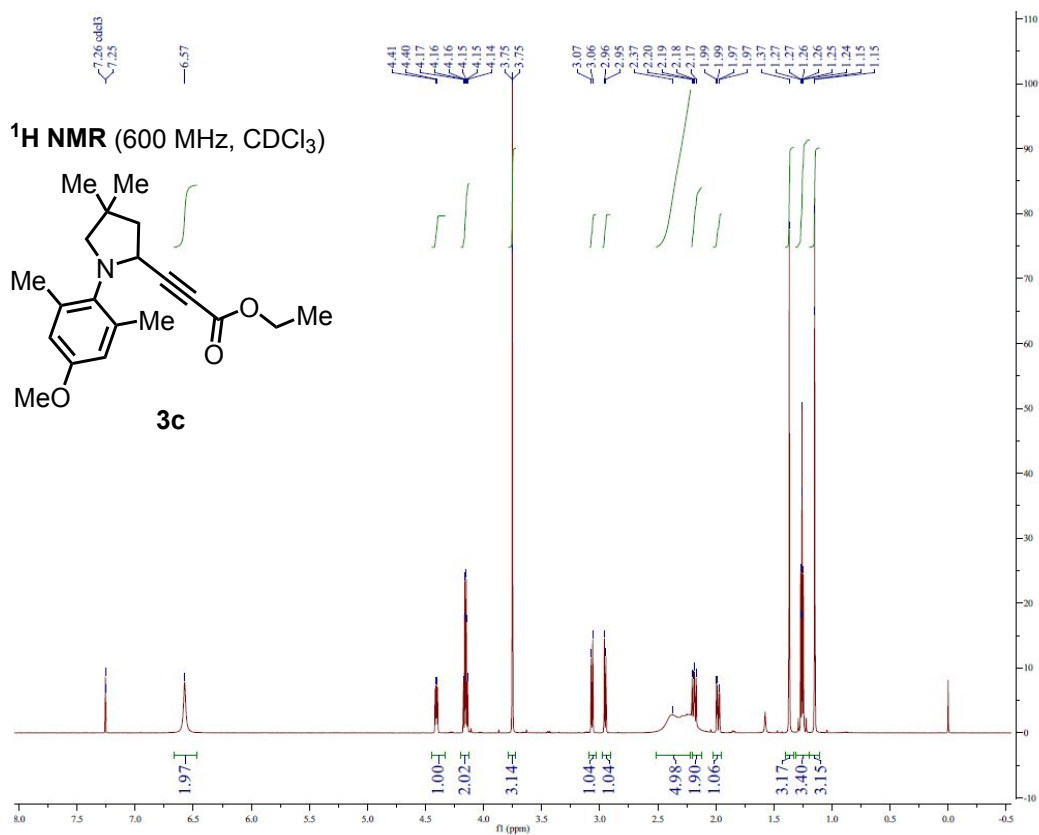
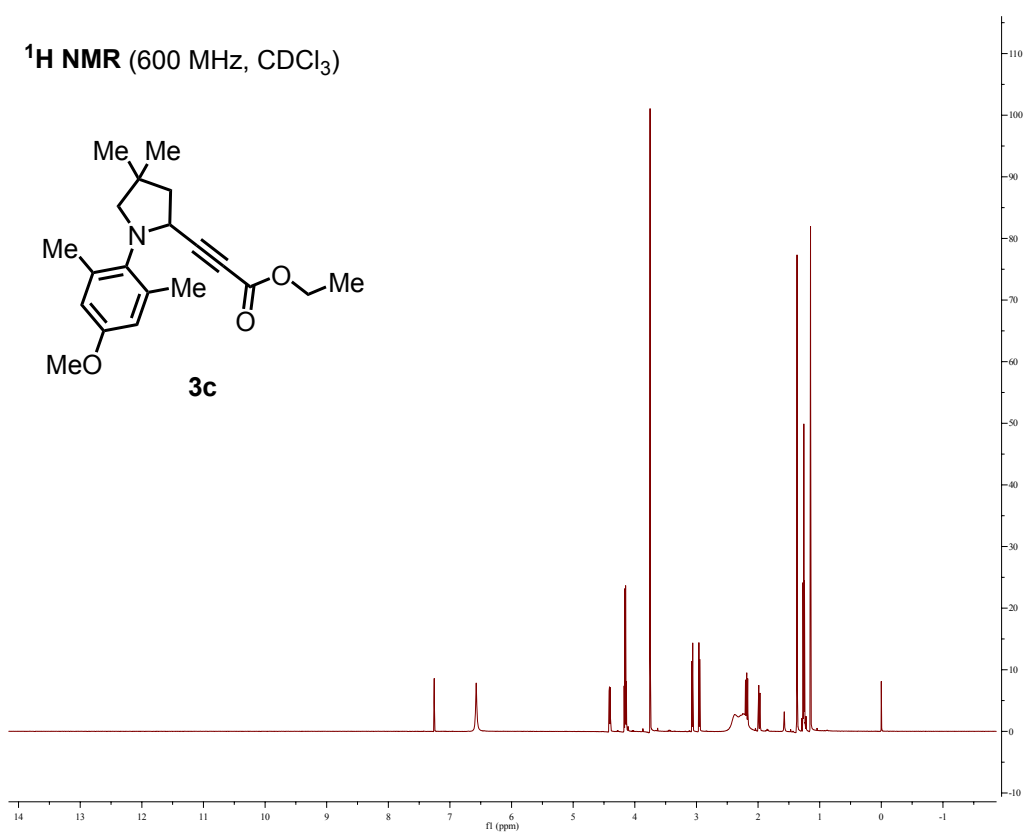
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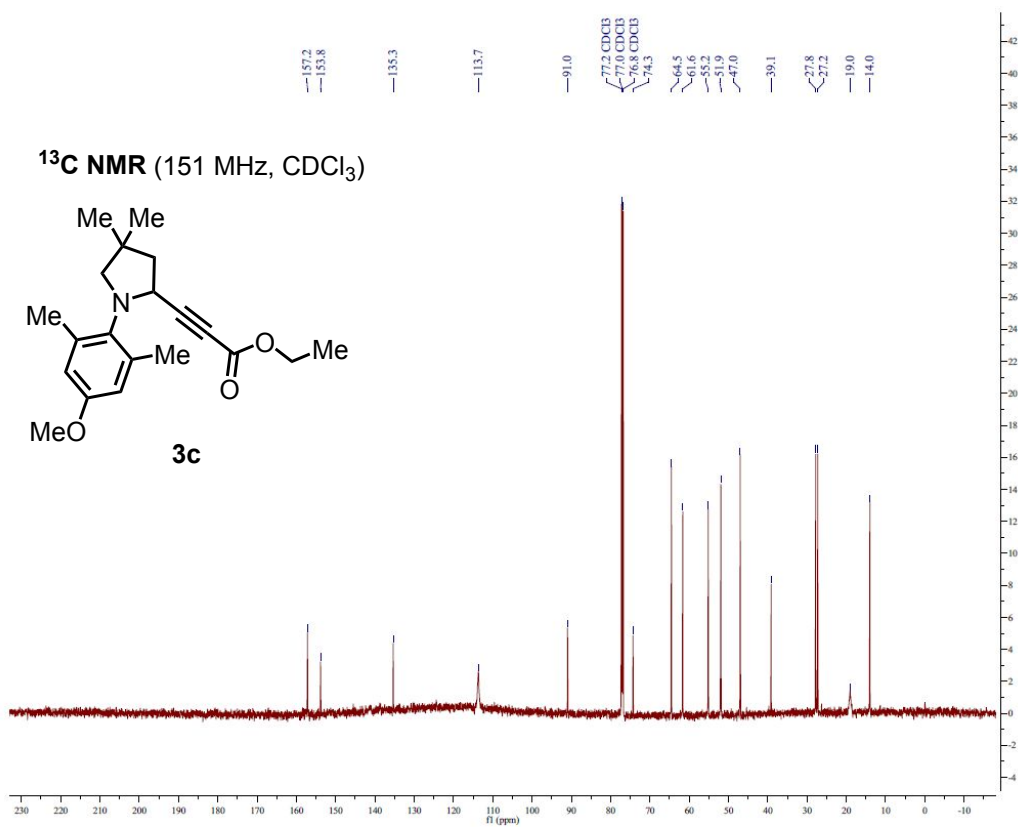


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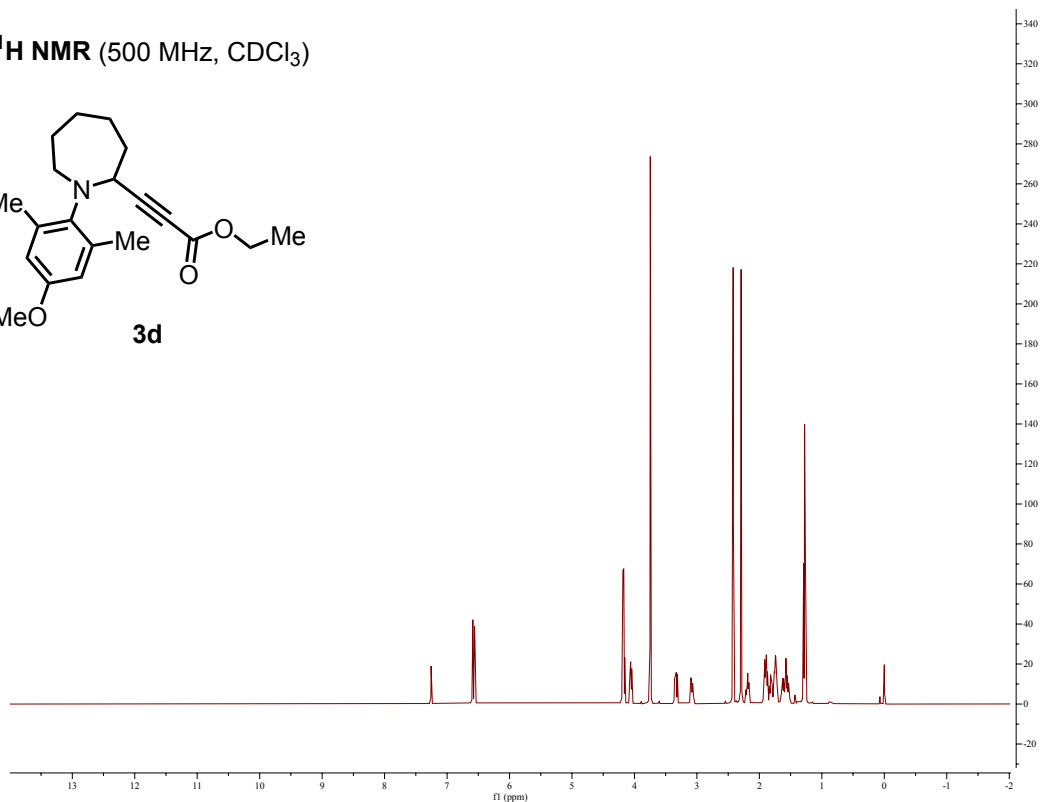
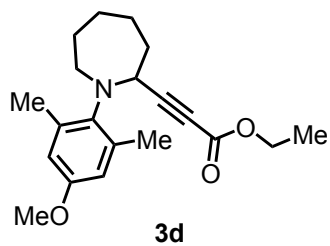


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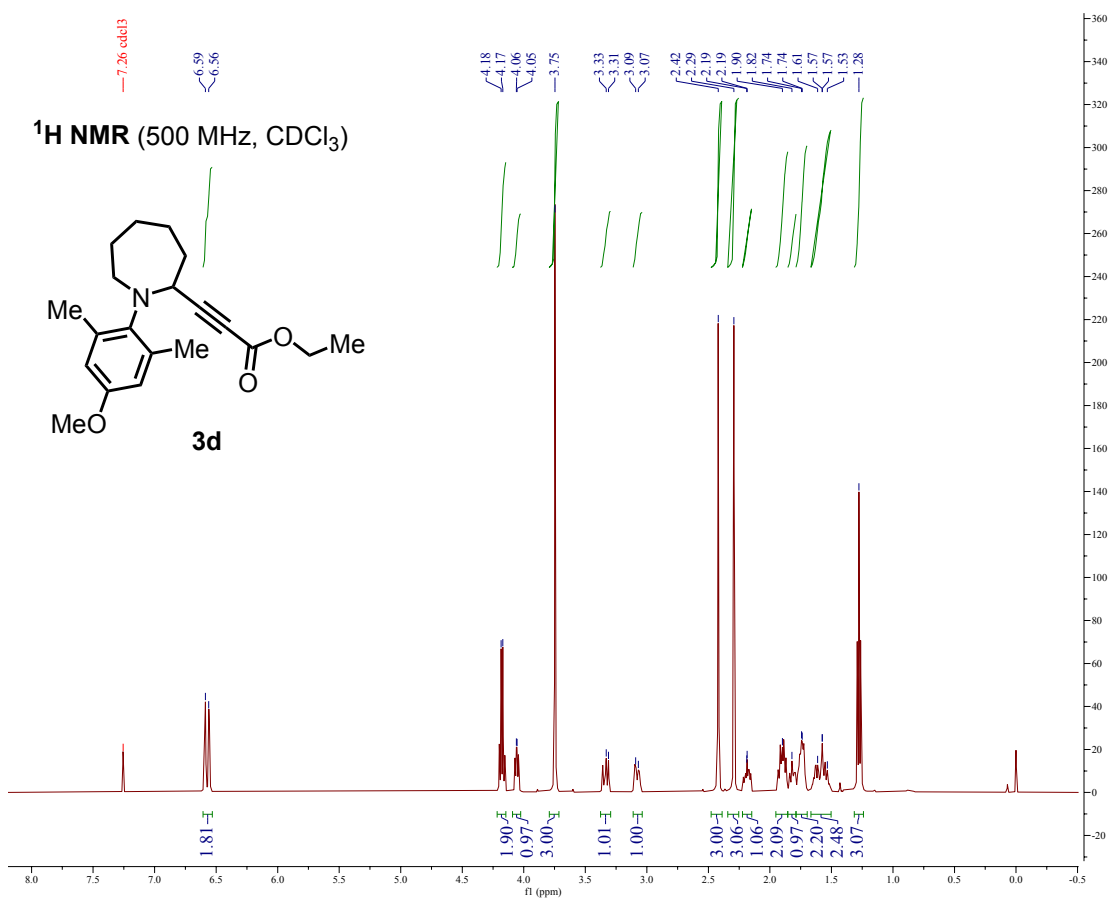
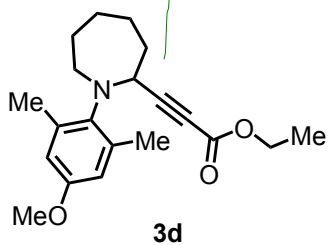


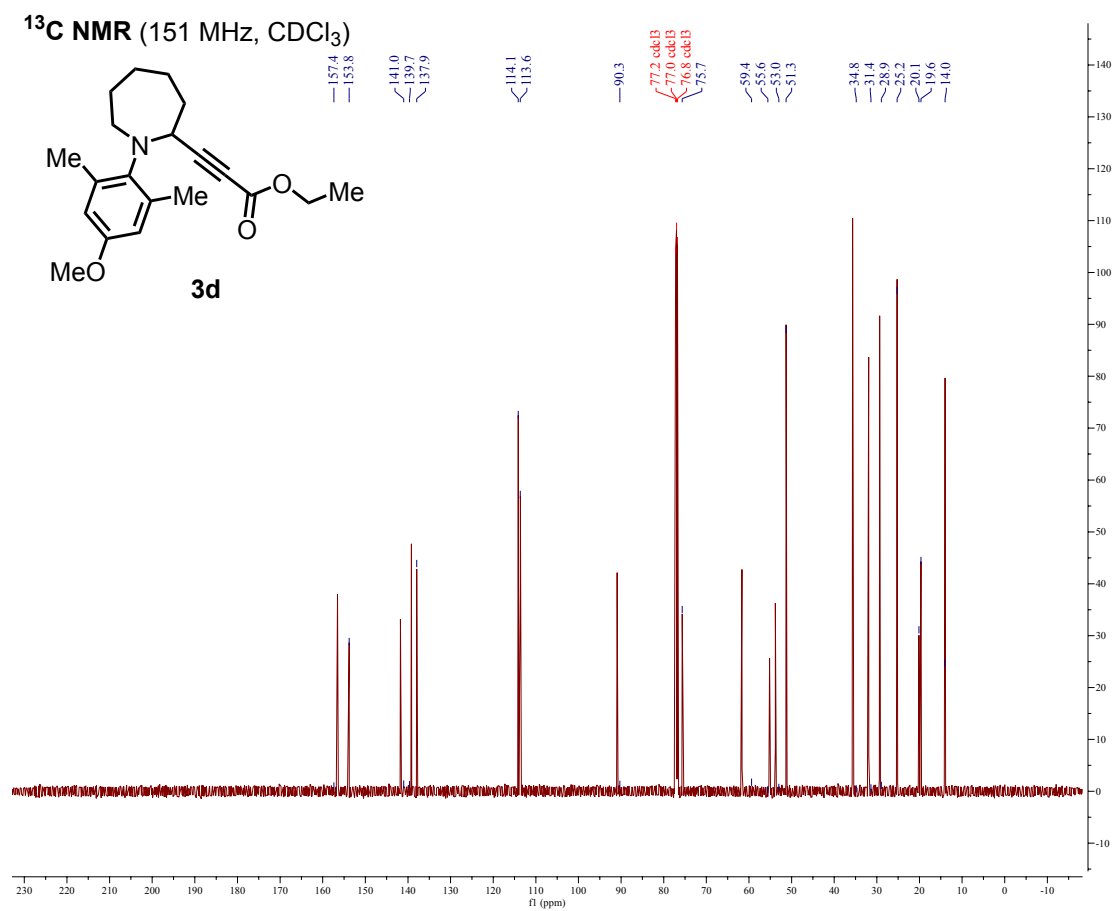


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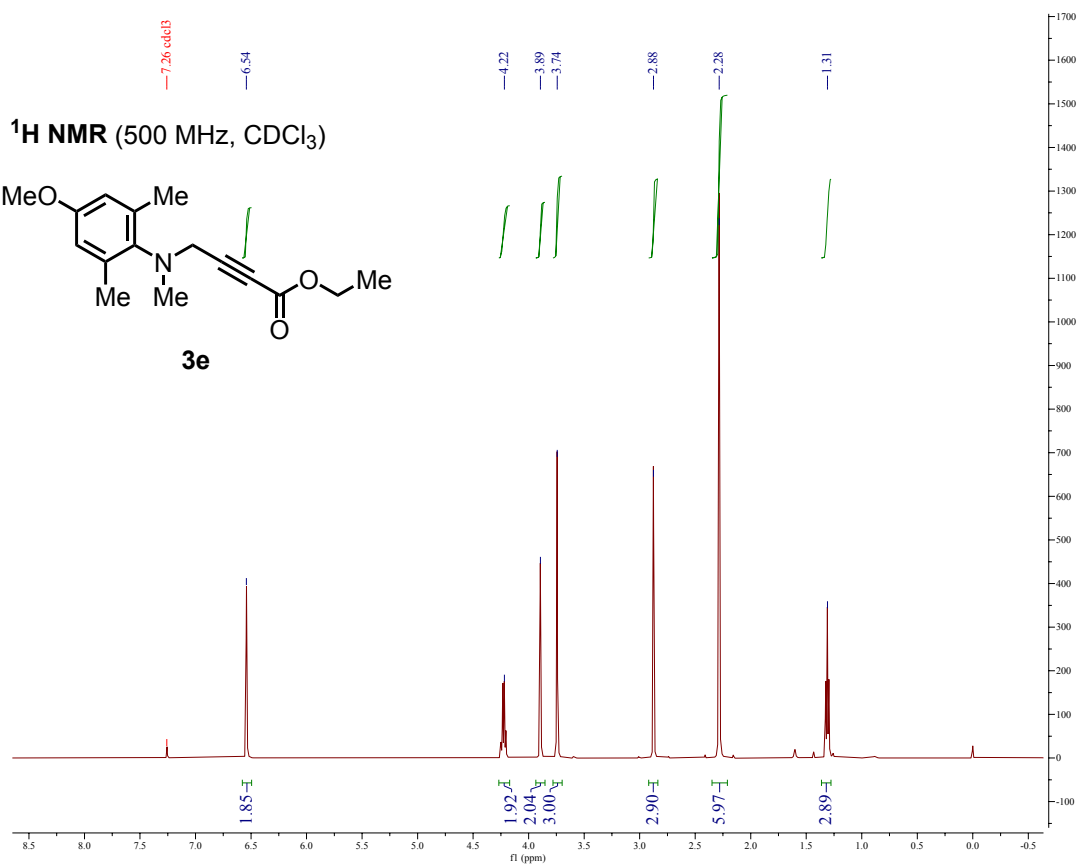
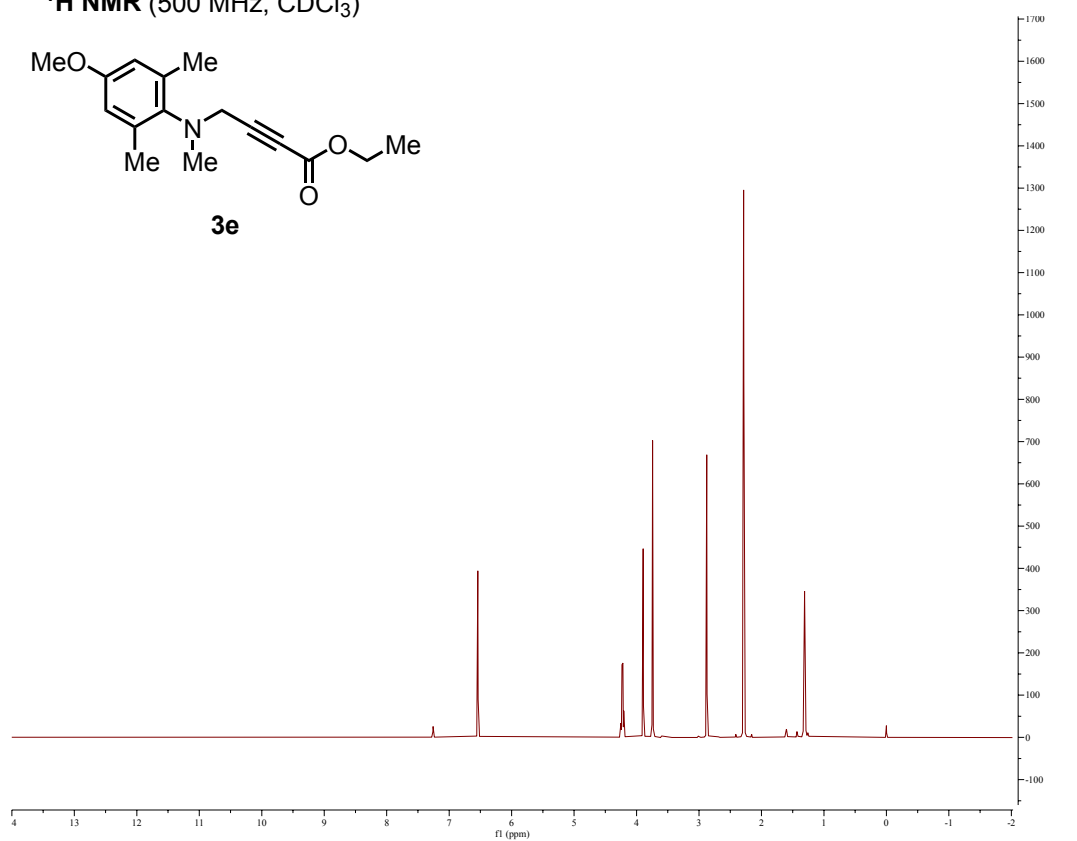


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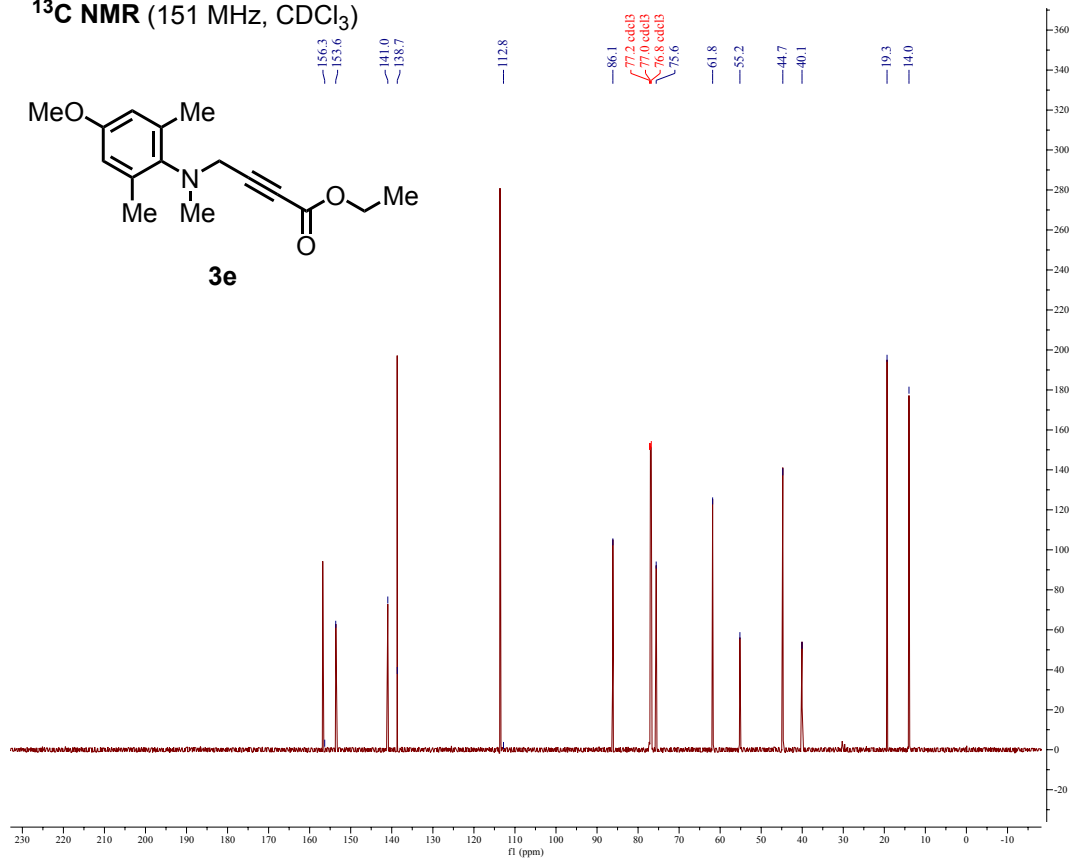




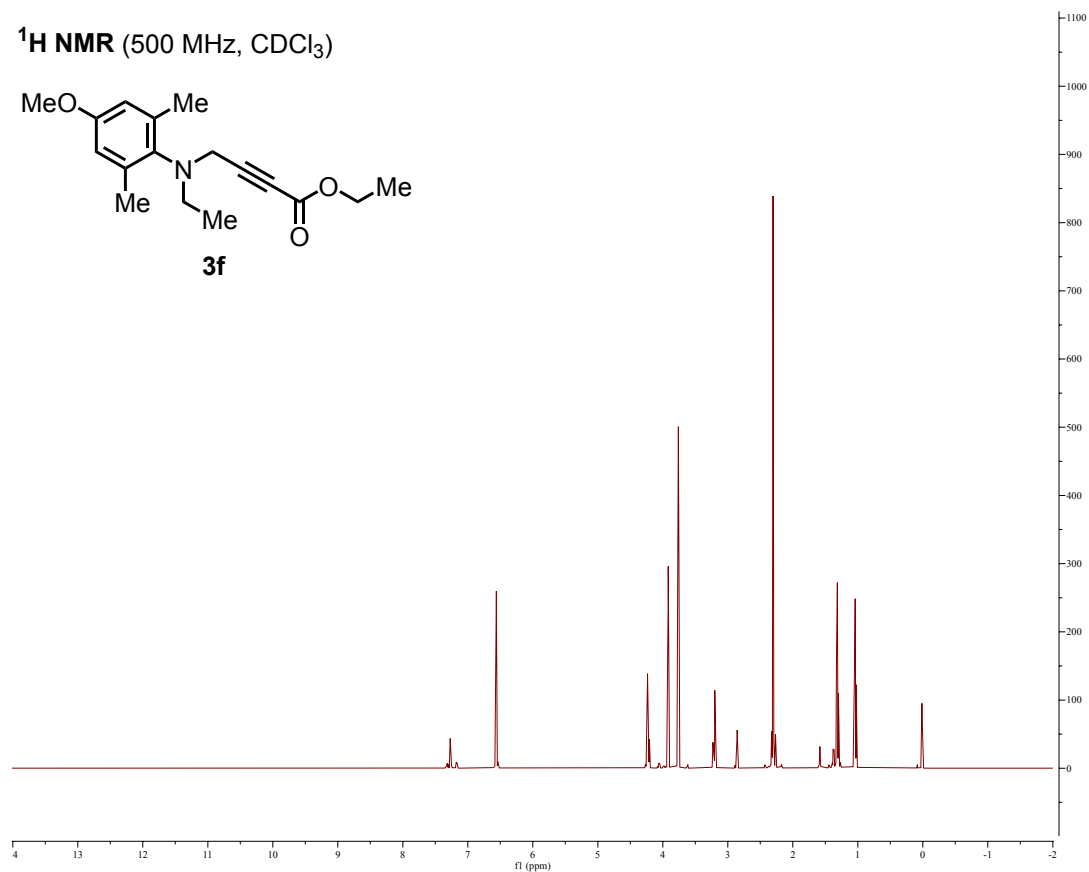
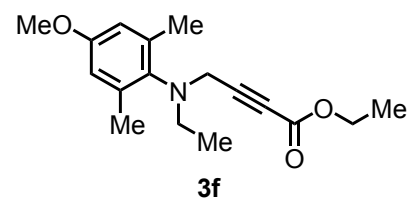
¹H NMR (500 MHz, CDCl₃)



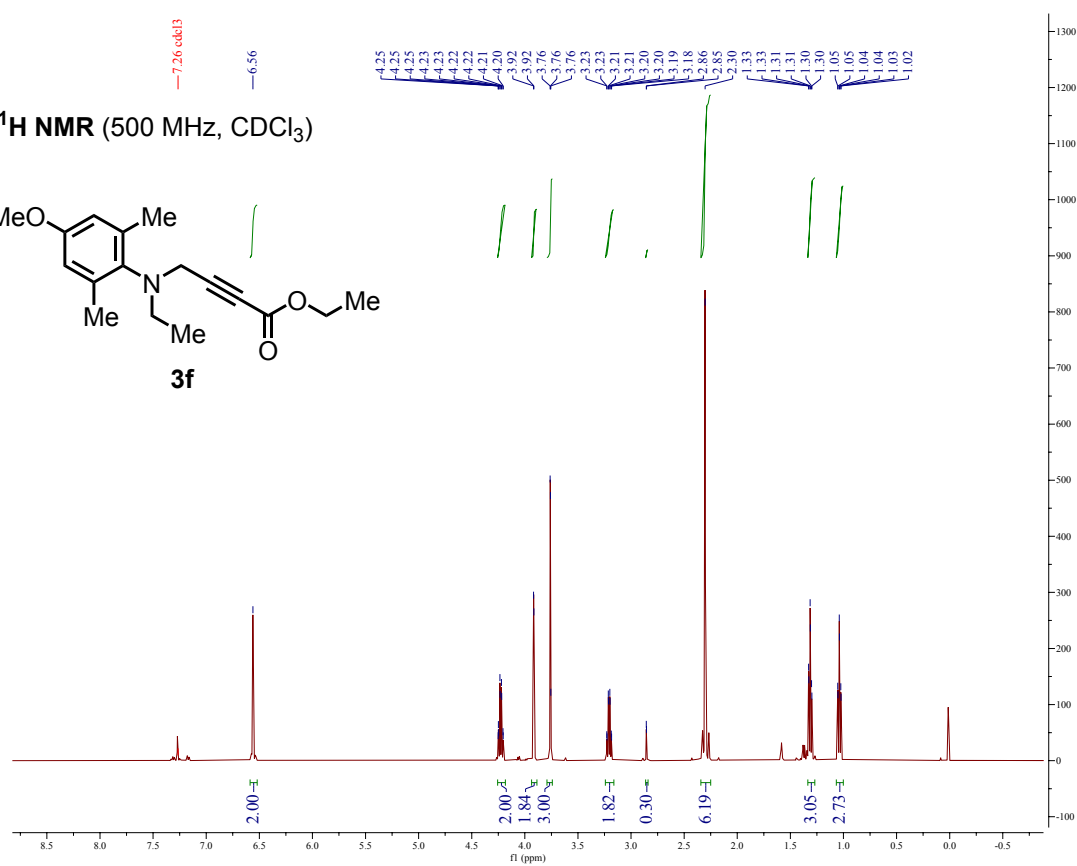
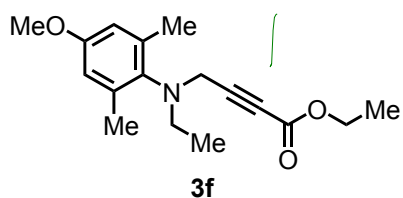
^{13}C NMR (151 MHz, CDCl_3)

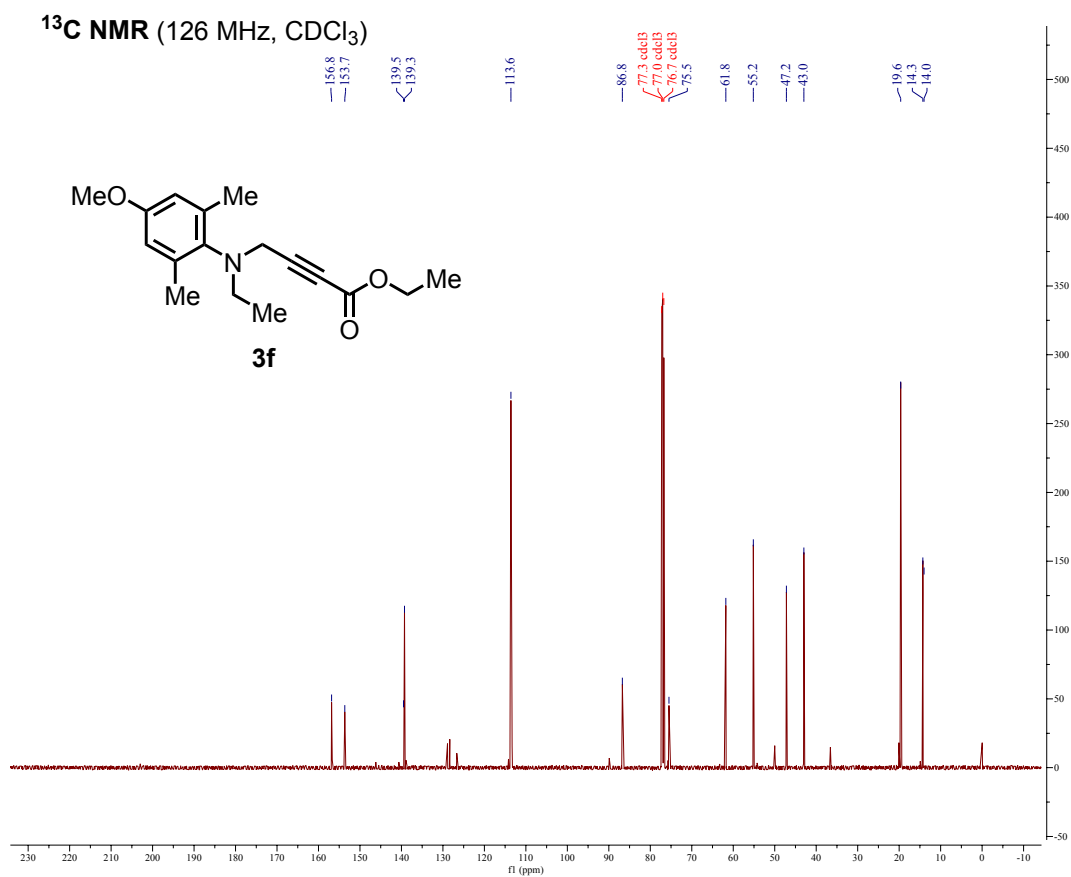


¹H NMR (500 MHz, CDCl₃)

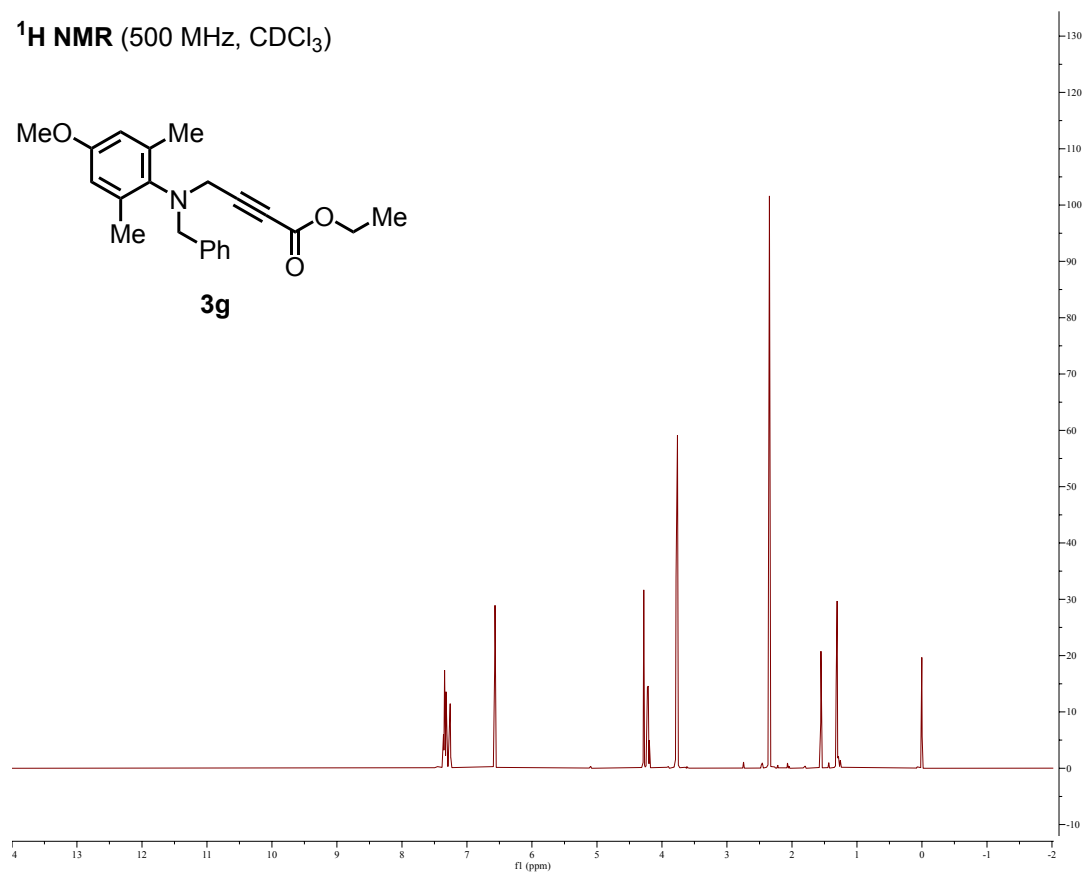
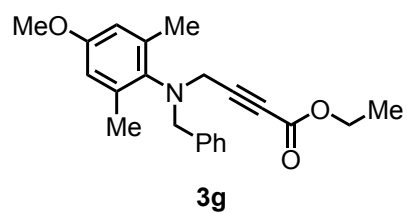


¹H NMR (500 MHz, CDCl₃)

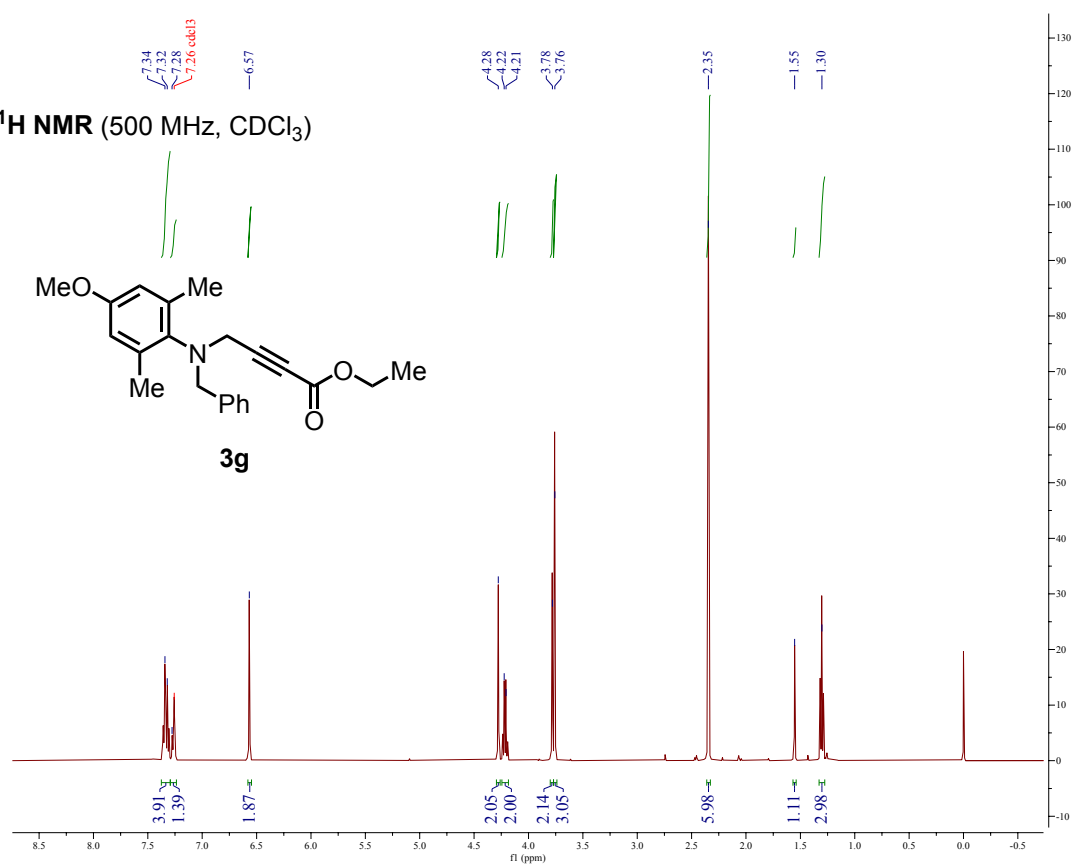
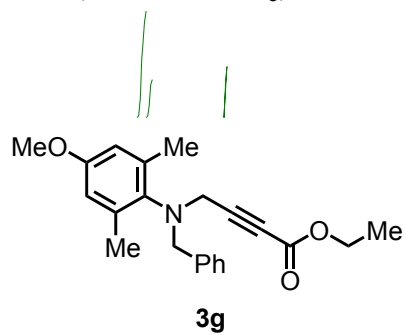




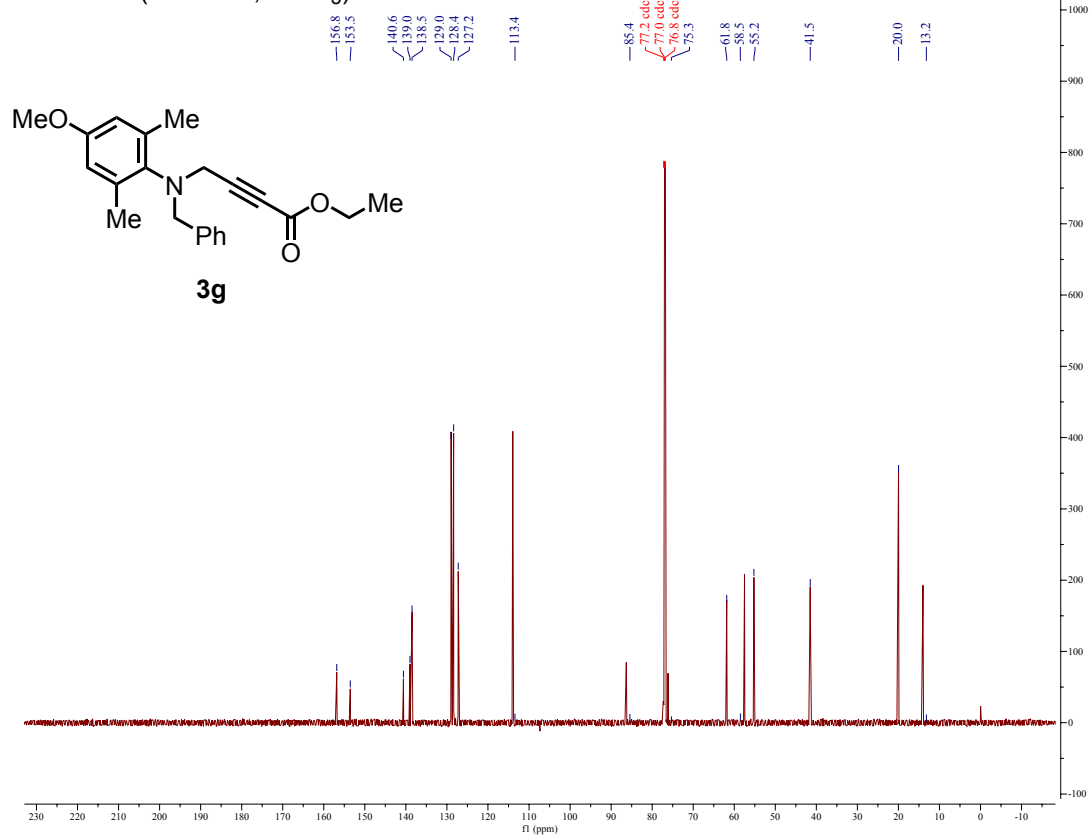
¹H NMR (500 MHz, CDCl₃)



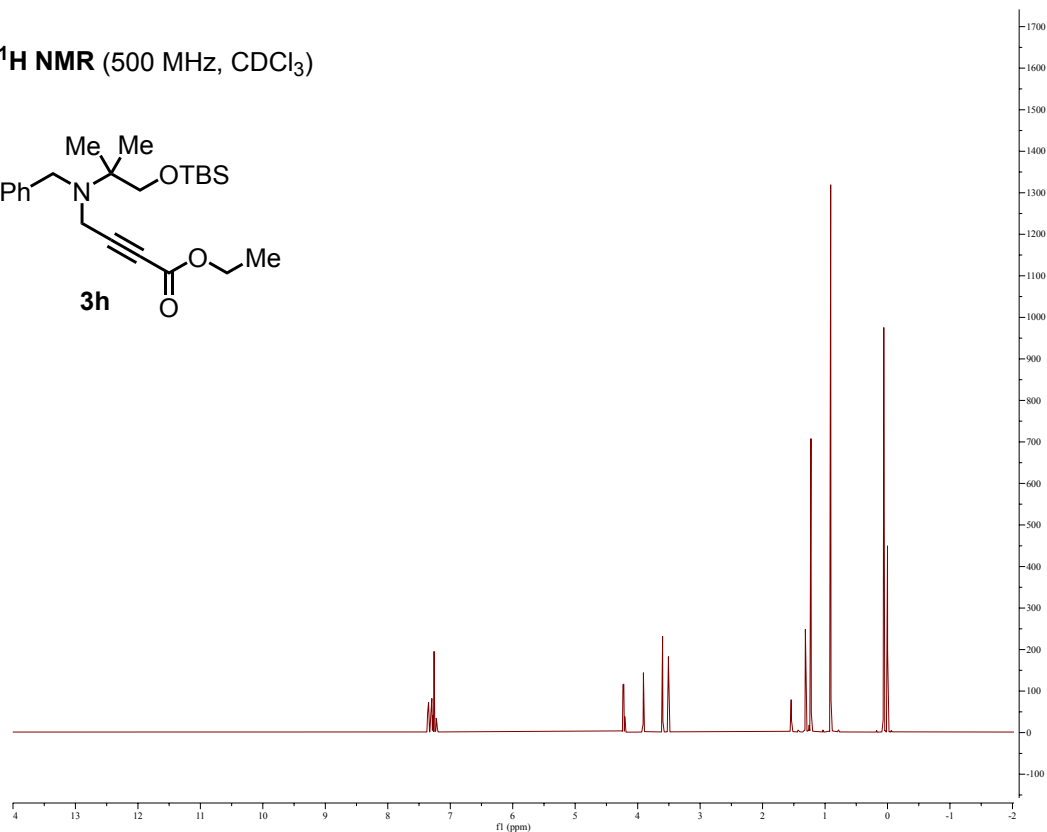
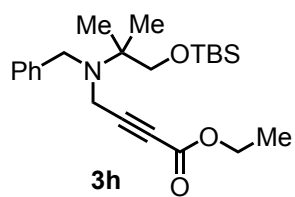
¹H NMR (500 MHz, CDCl₃)



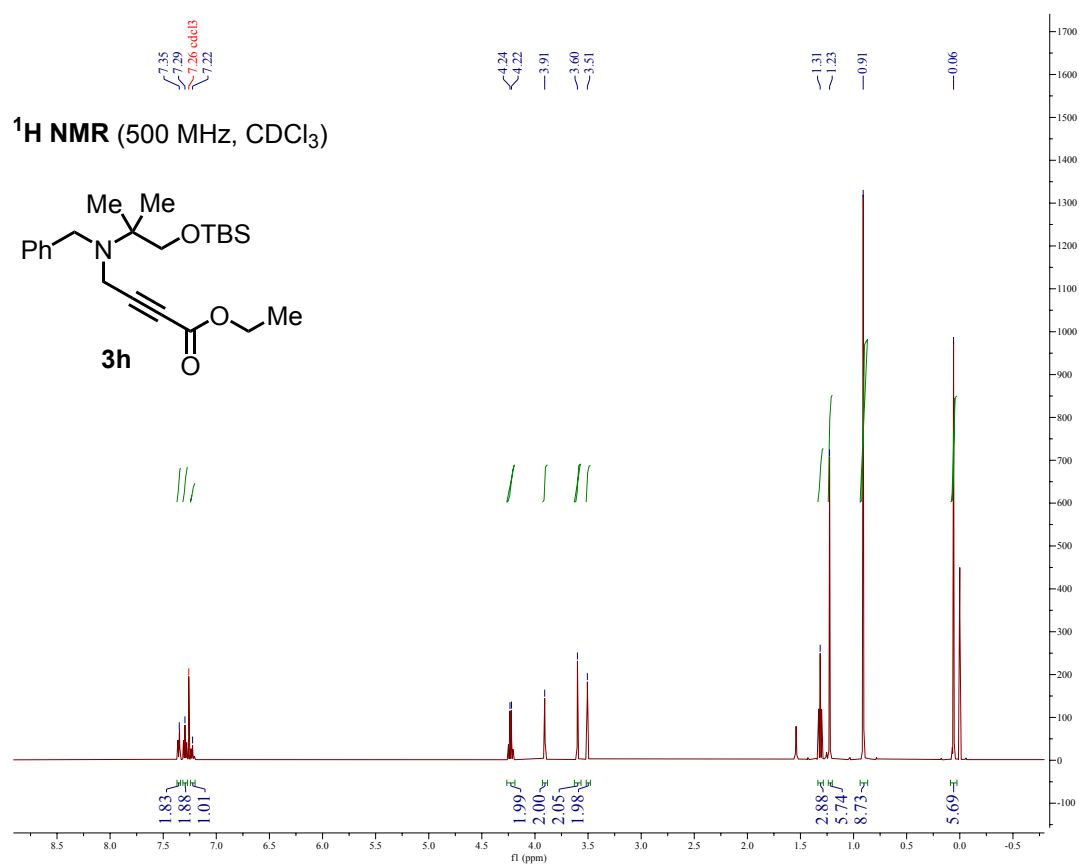
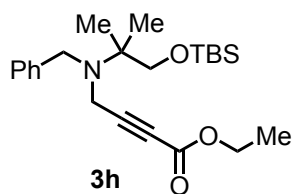
¹³C NMR (151 MHz, CDCl₃)



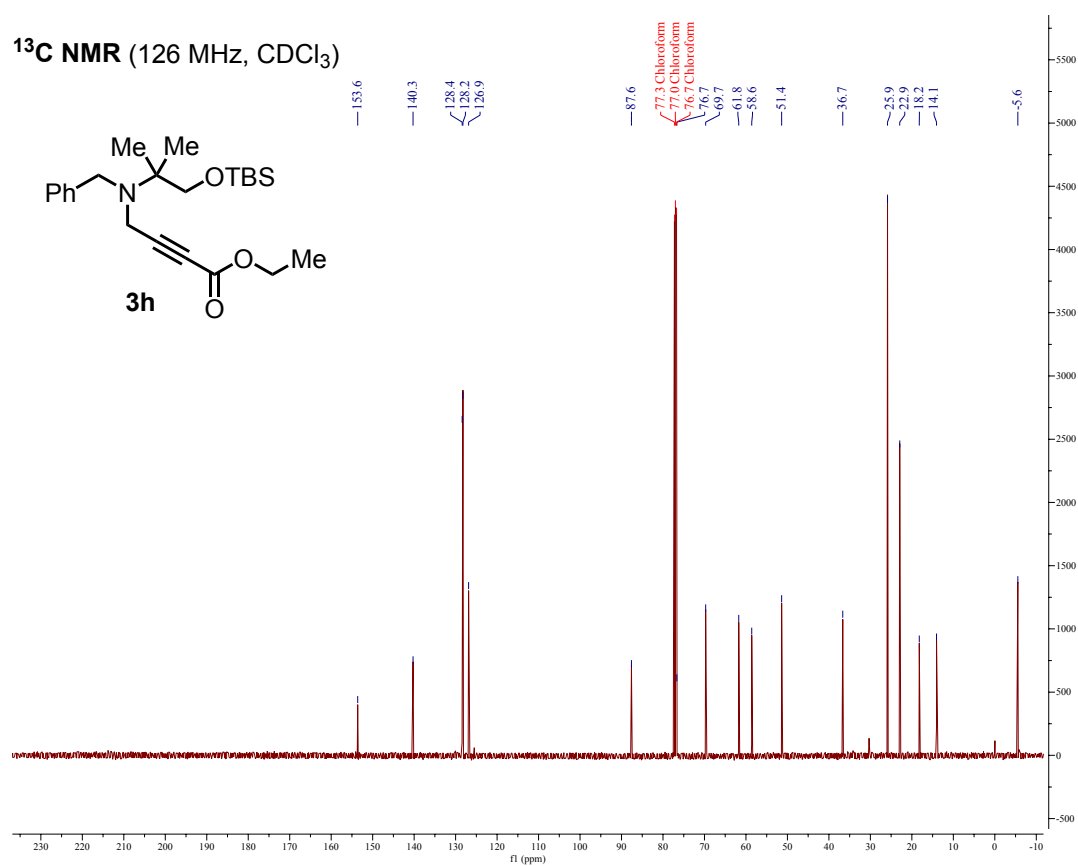
¹H NMR (500 MHz, CDCl₃)



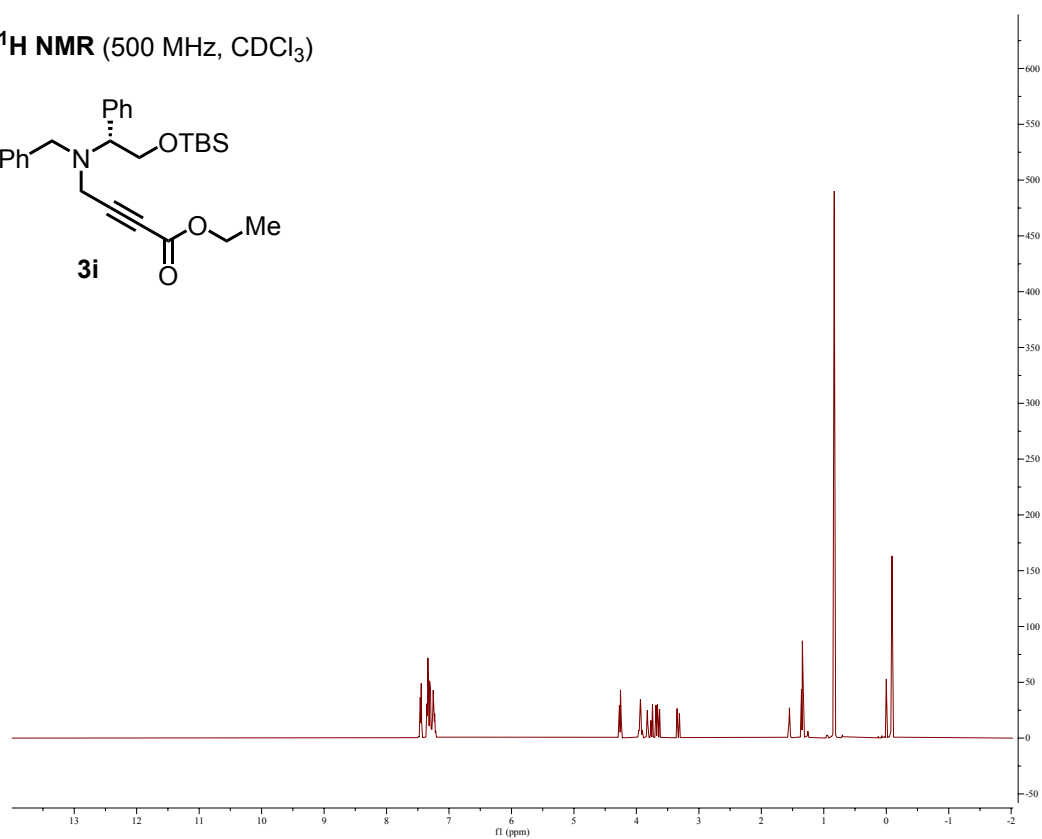
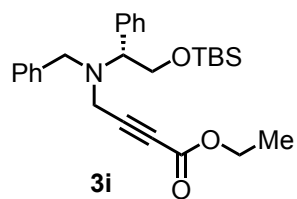
¹H NMR (500 MHz, CDCl₃)



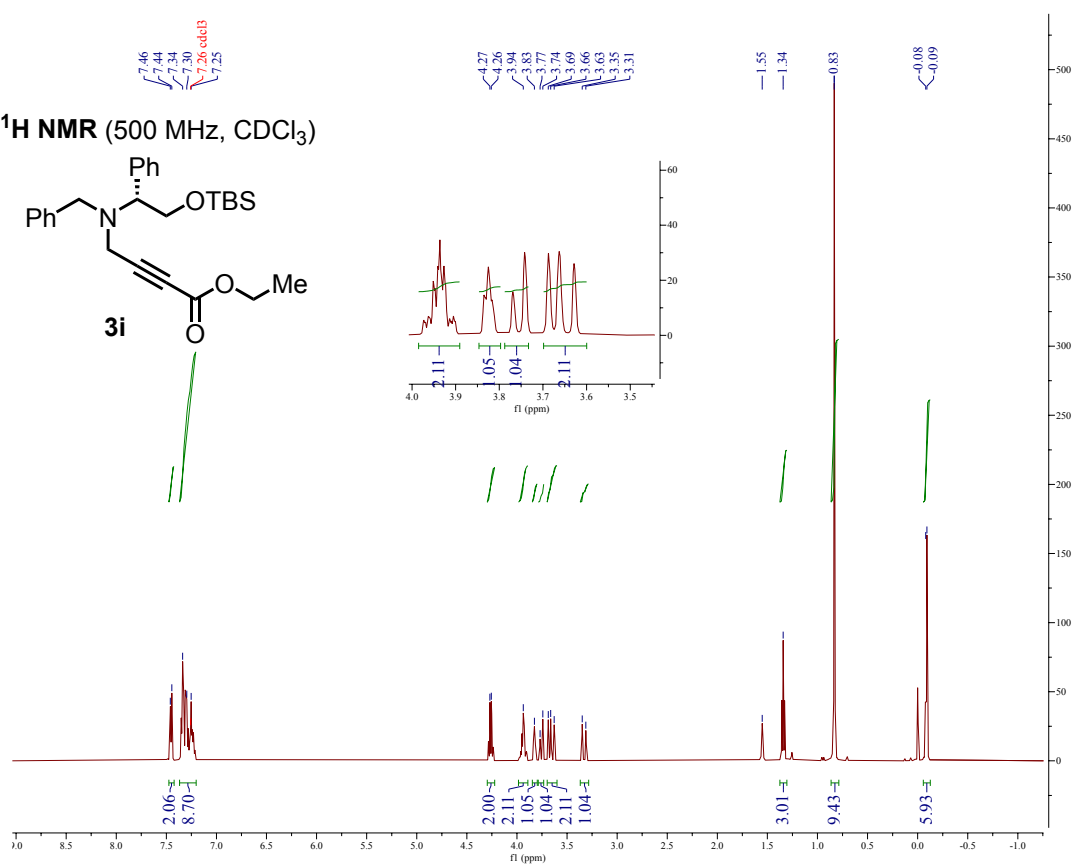
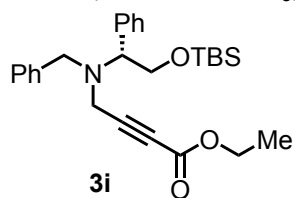
¹³C NMR (126 MHz, CDCl₃)



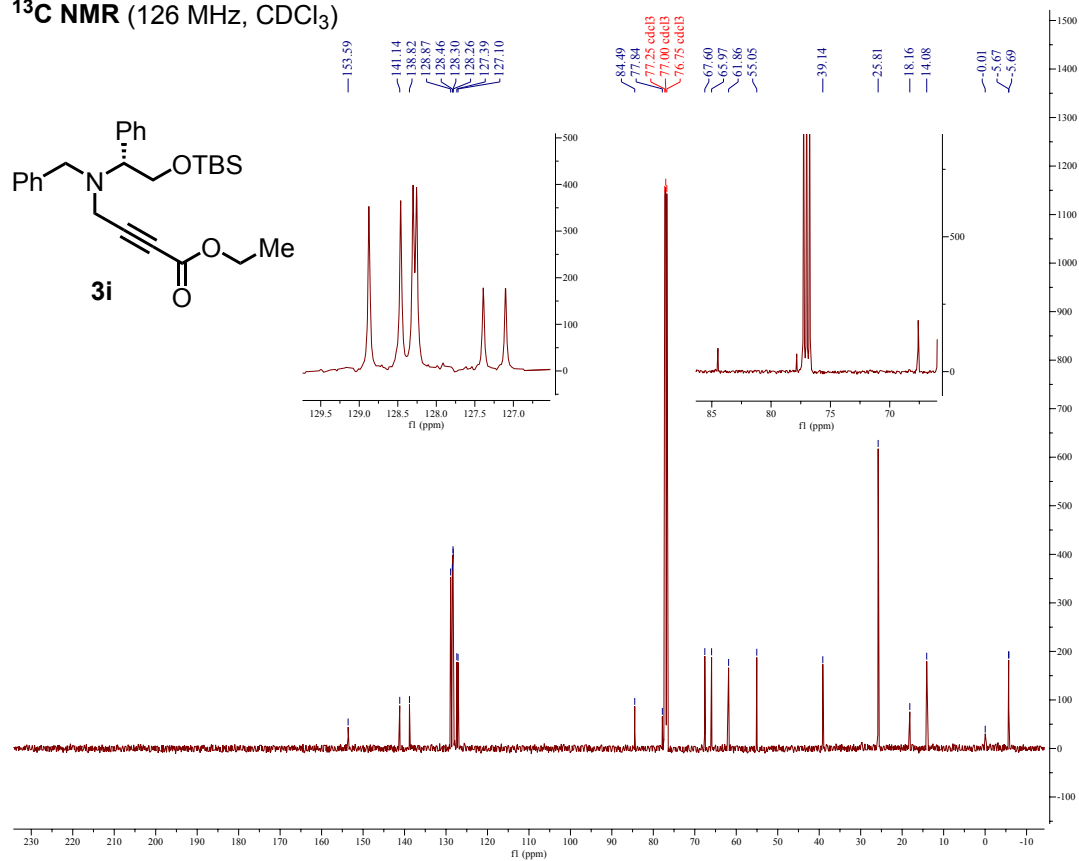
¹H NMR (500 MHz, CDCl₃)



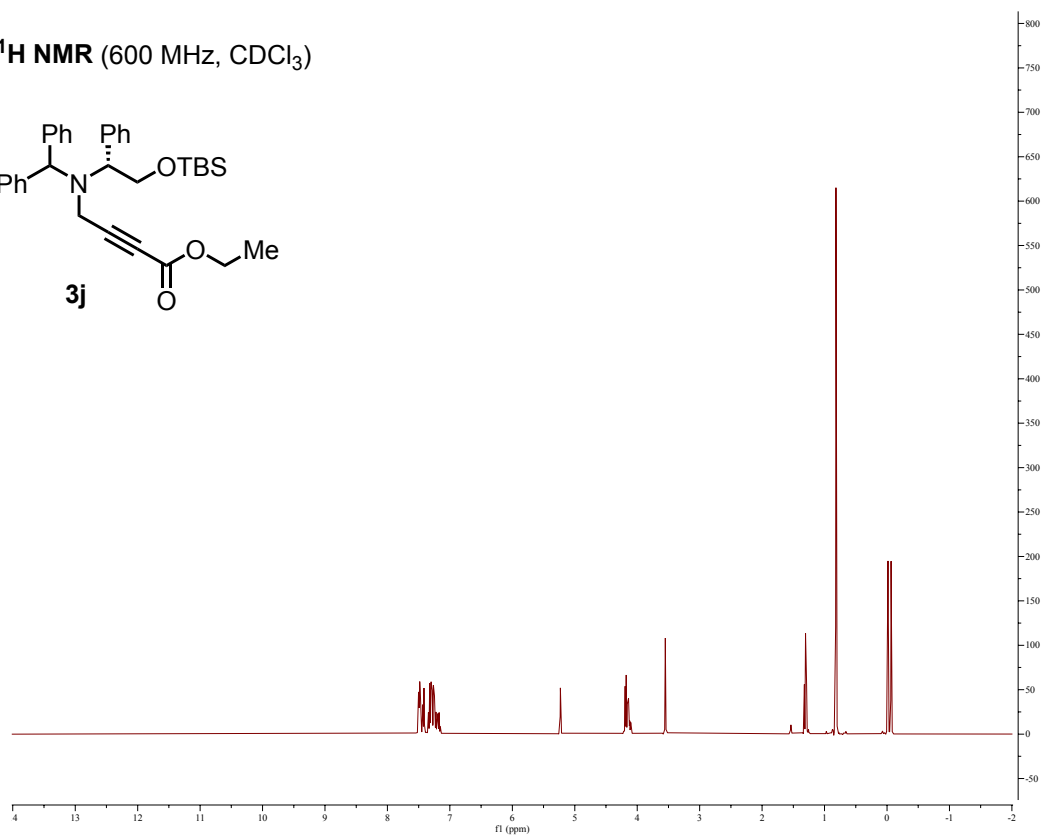
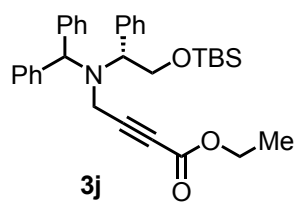
¹H NMR (500 MHz, CDCl₃)



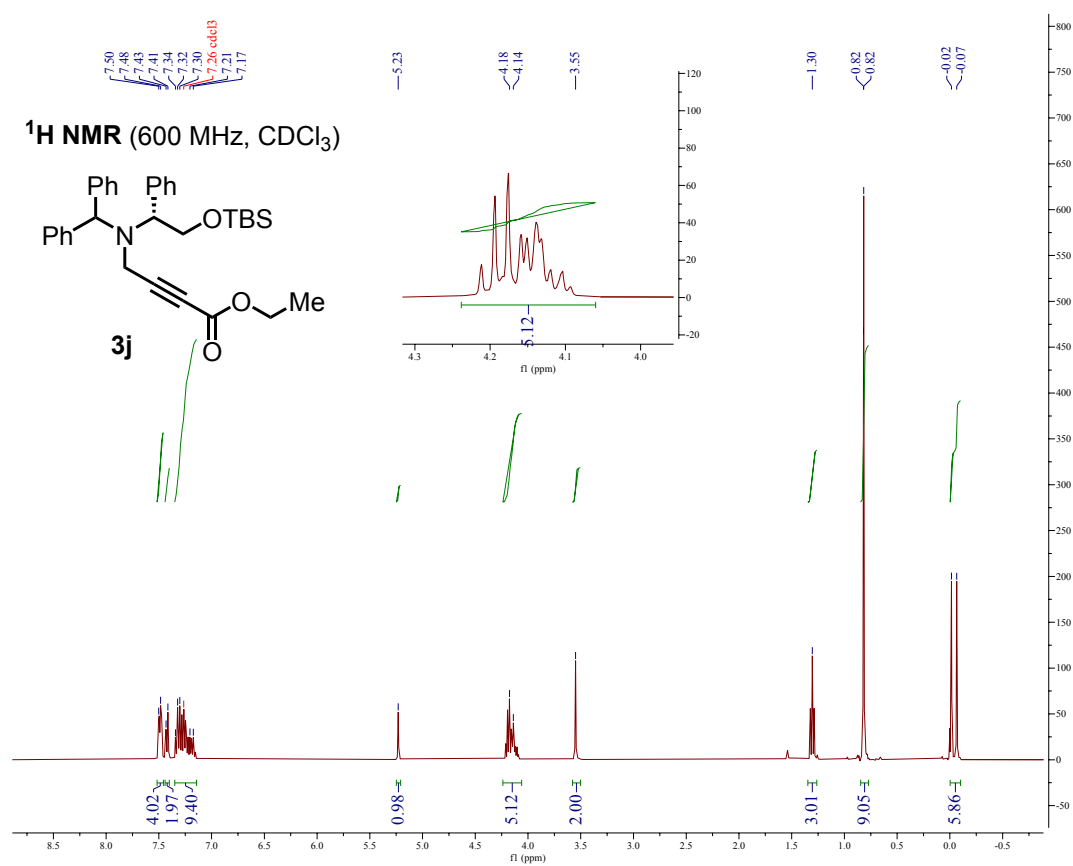
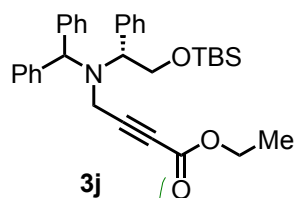
¹³C NMR (126 MHz, CDCl₃)

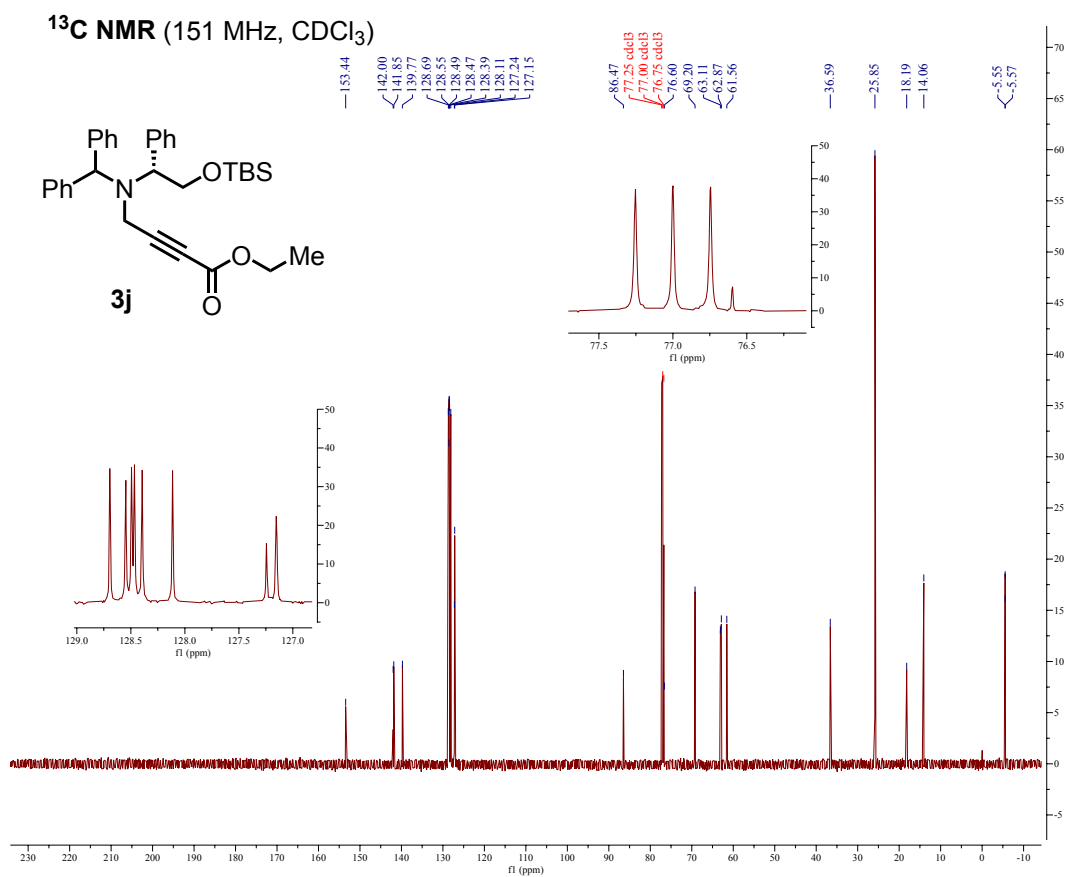


¹H NMR (600 MHz, CDCl₃)

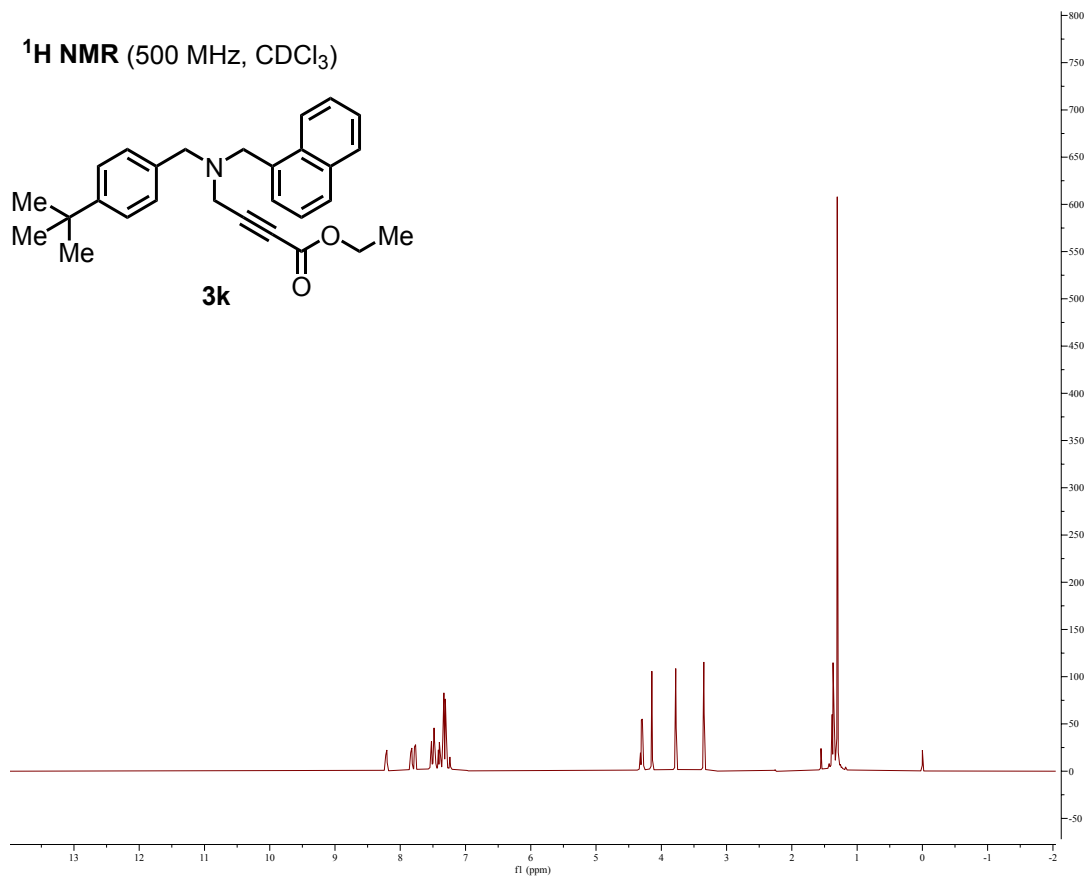
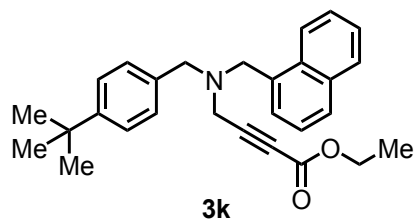


¹H NMR (600 MHz, CDCl₃)

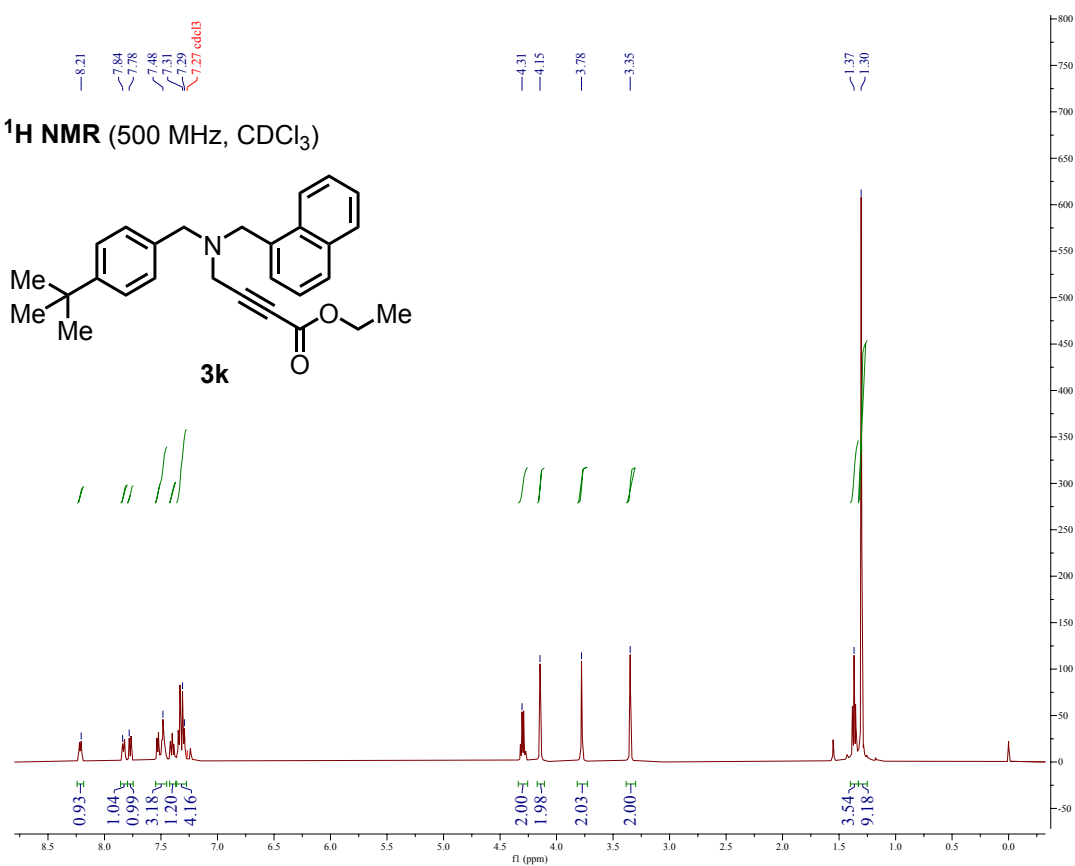
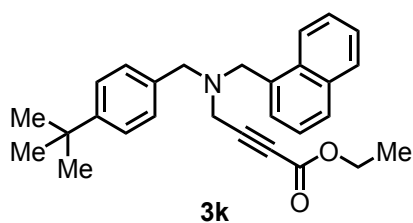


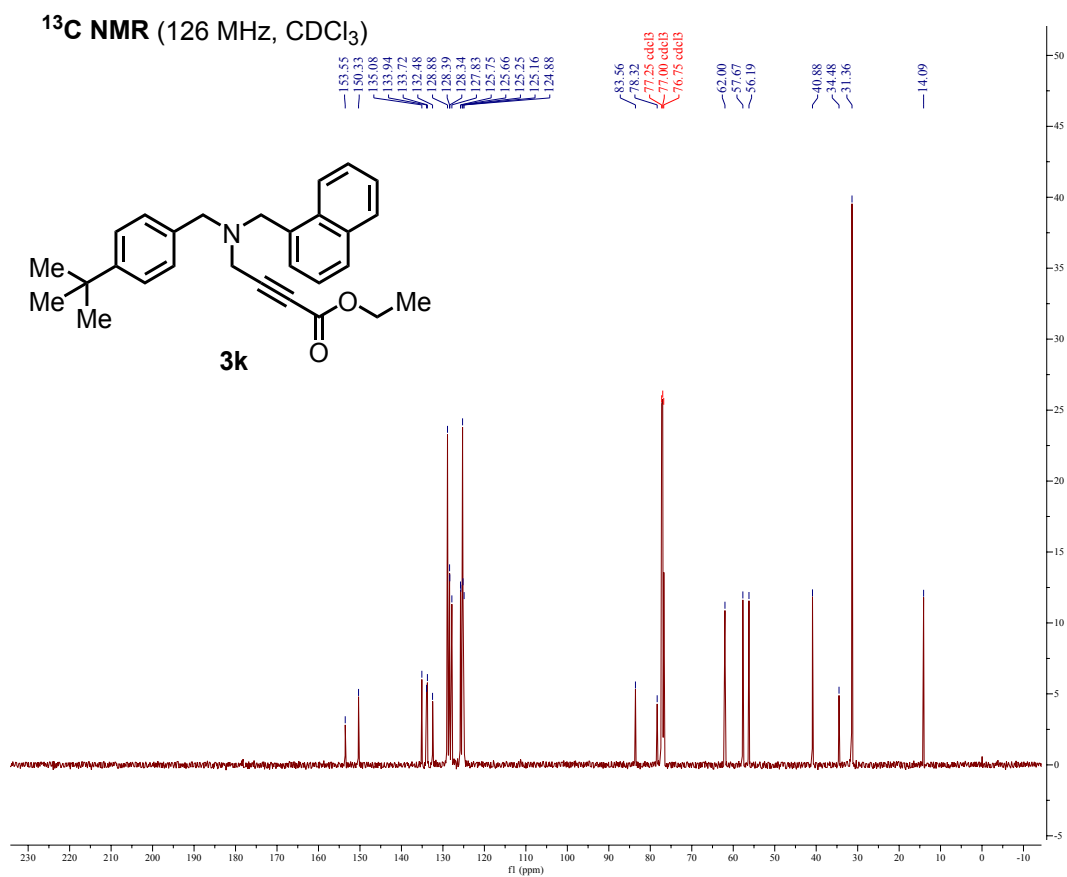


¹H NMR (500 MHz, CDCl₃)

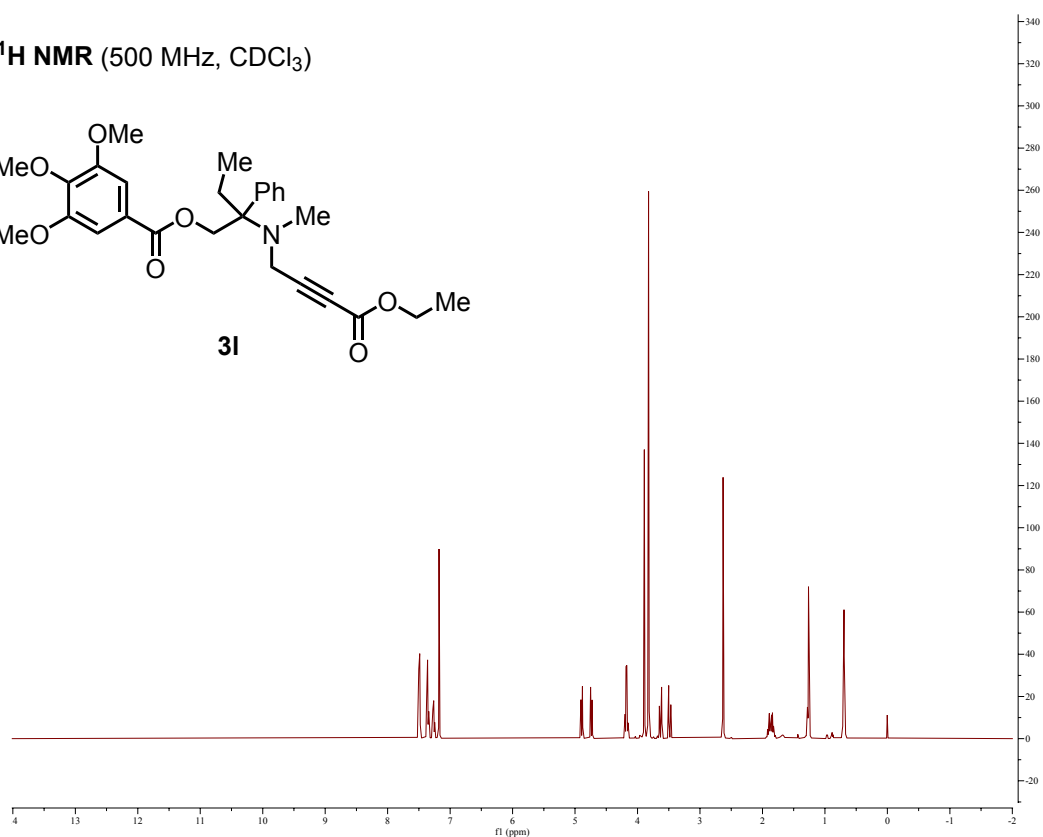
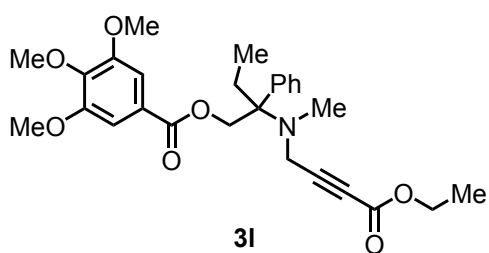


¹H NMR (500 MHz, CDCl₃)

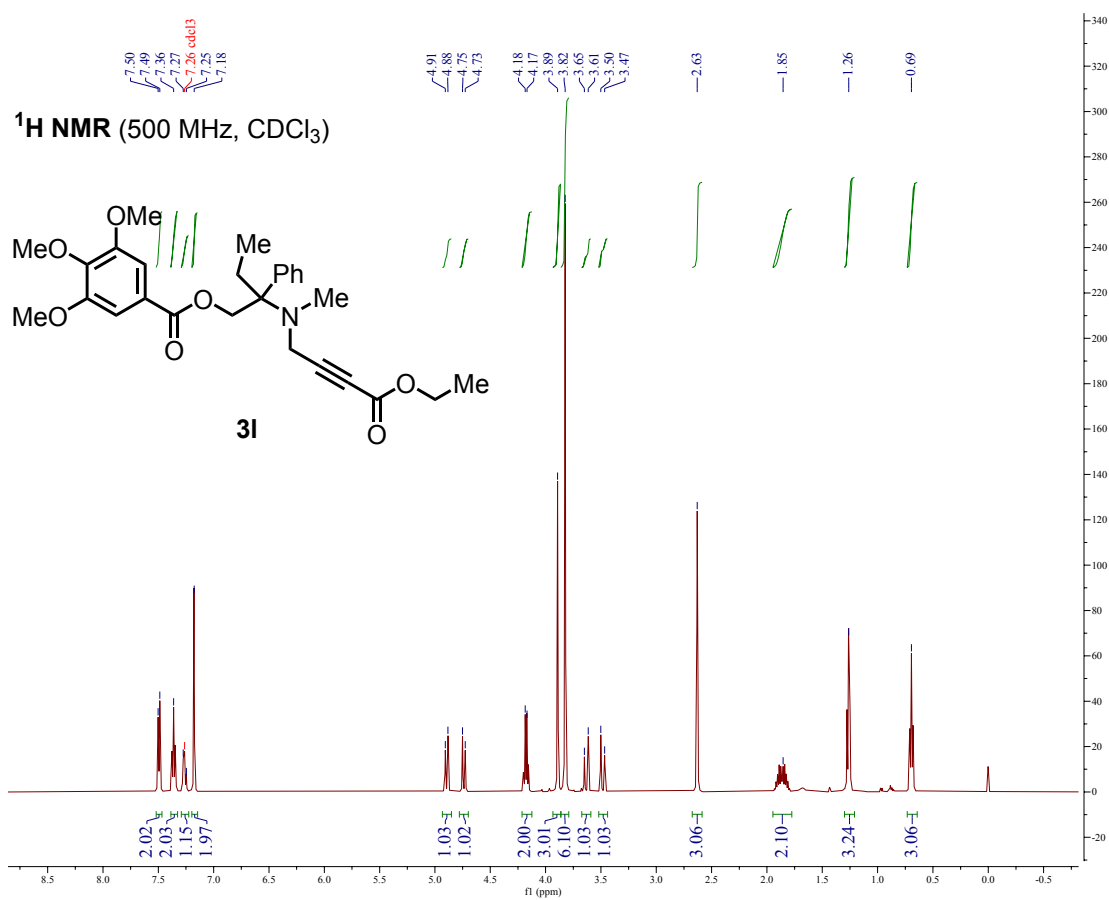
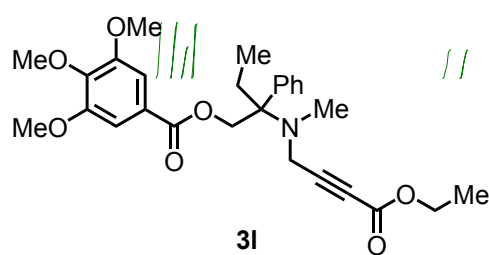




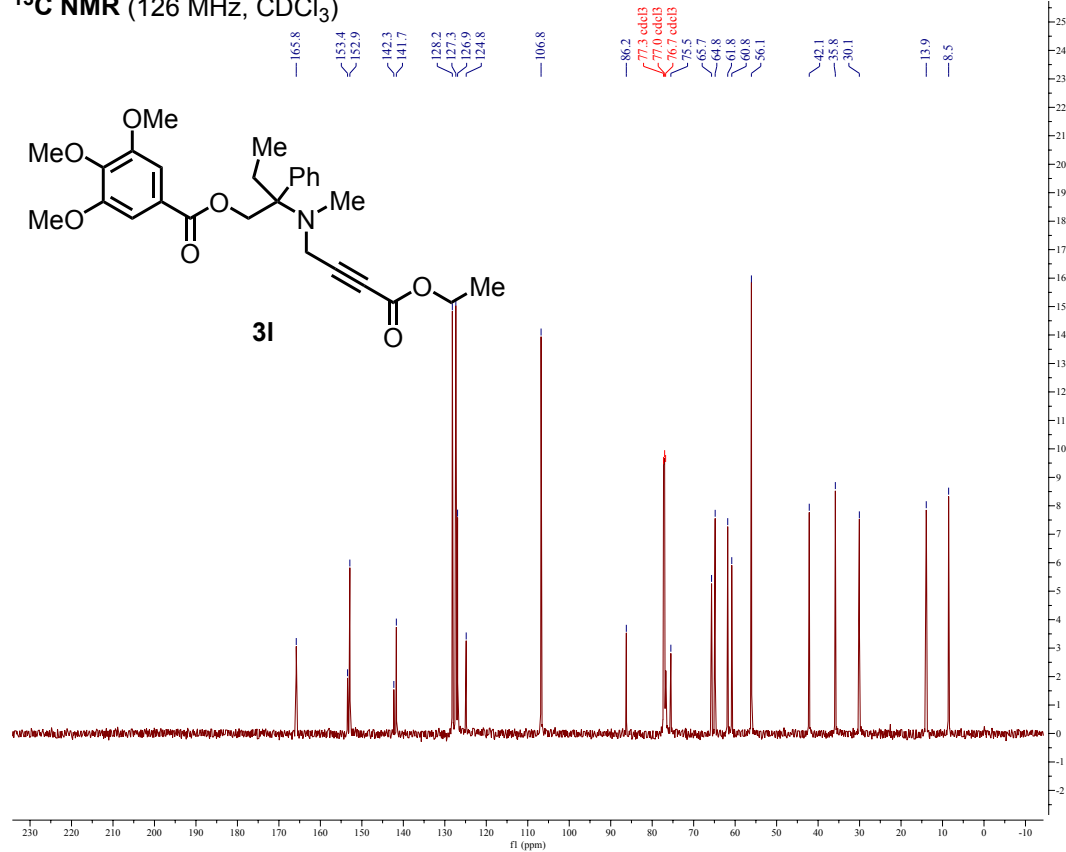
¹H NMR (500 MHz, CDCl₃)



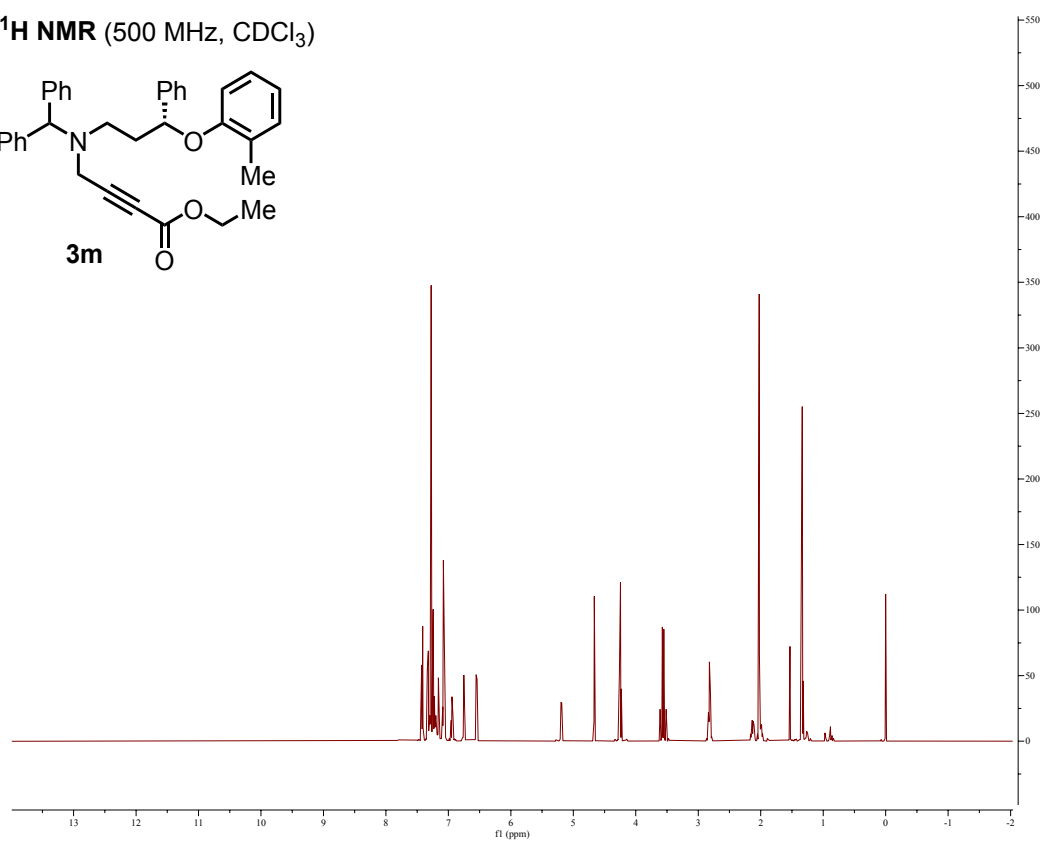
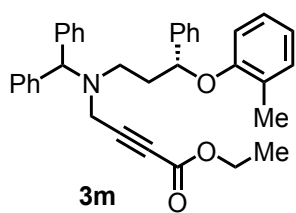
¹H NMR (500 MHz, CDCl₃)



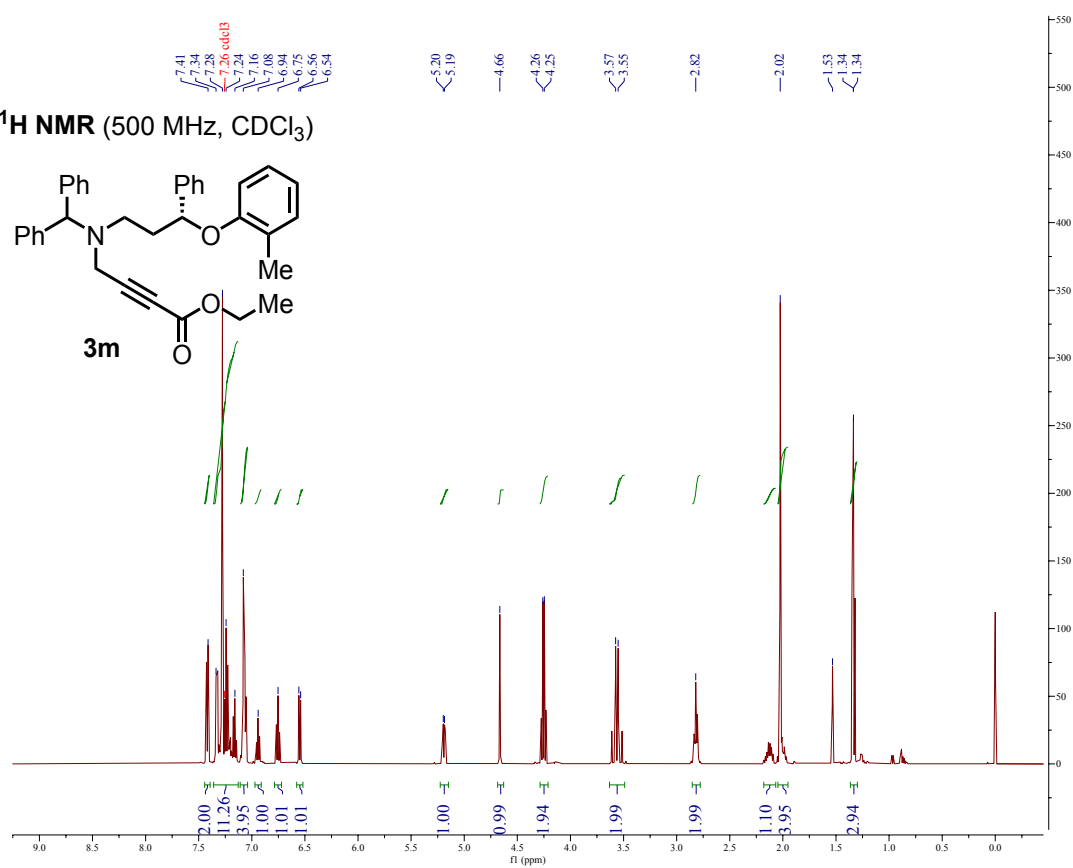
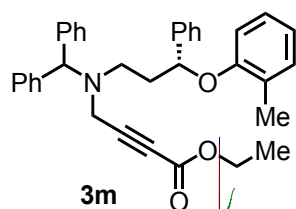
^{13}C NMR (126 MHz, CDCl_3)



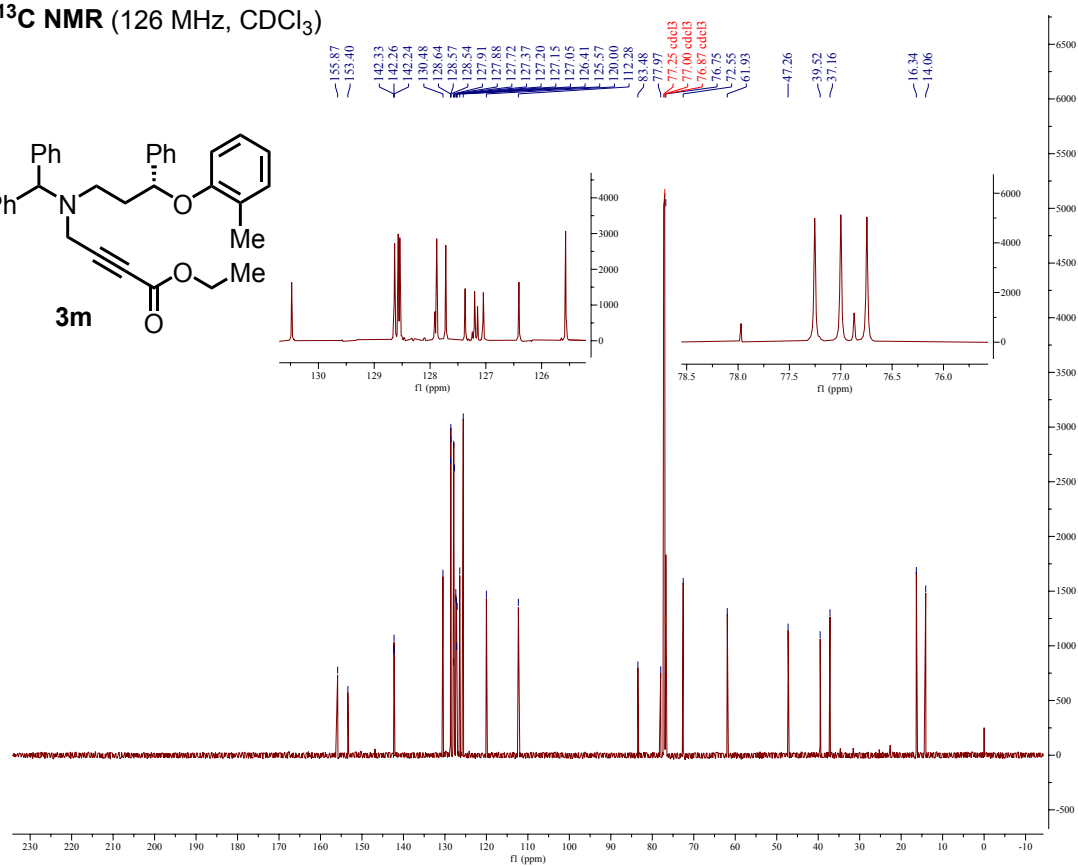
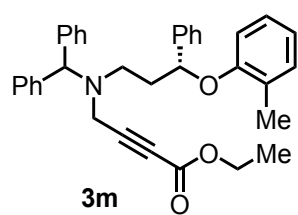
¹H NMR (500 MHz, CDCl₃)

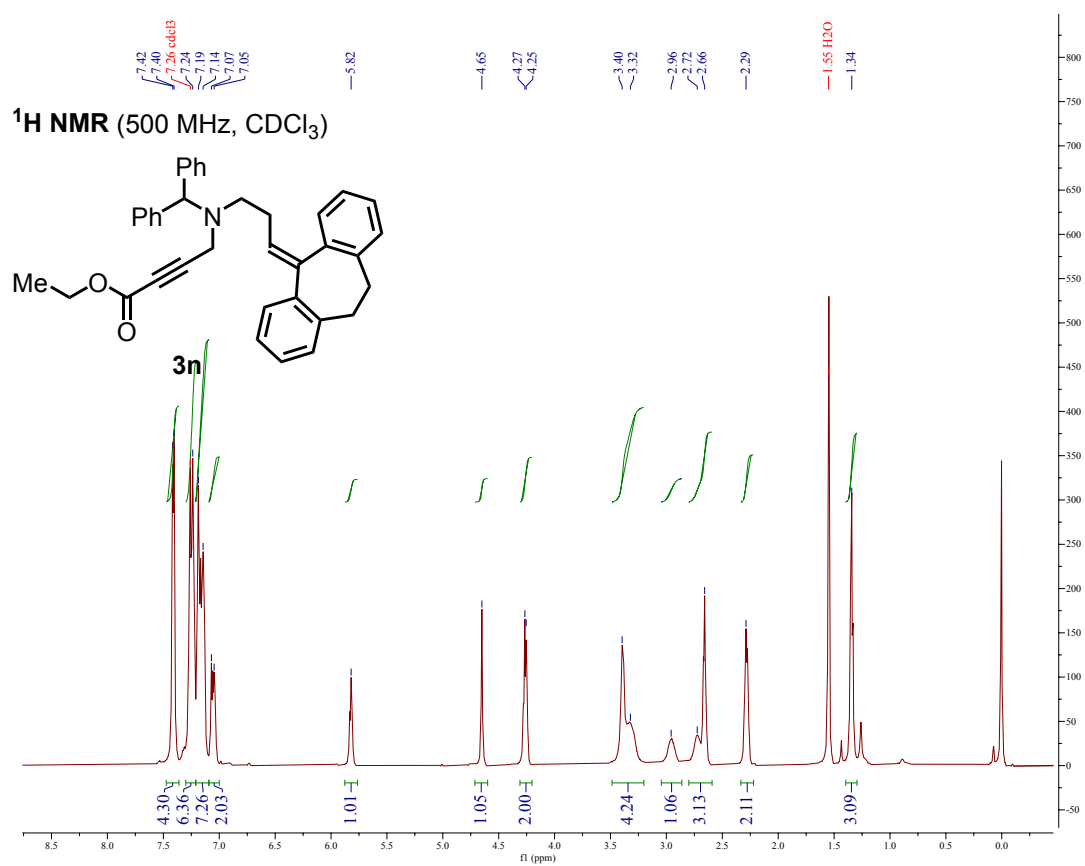
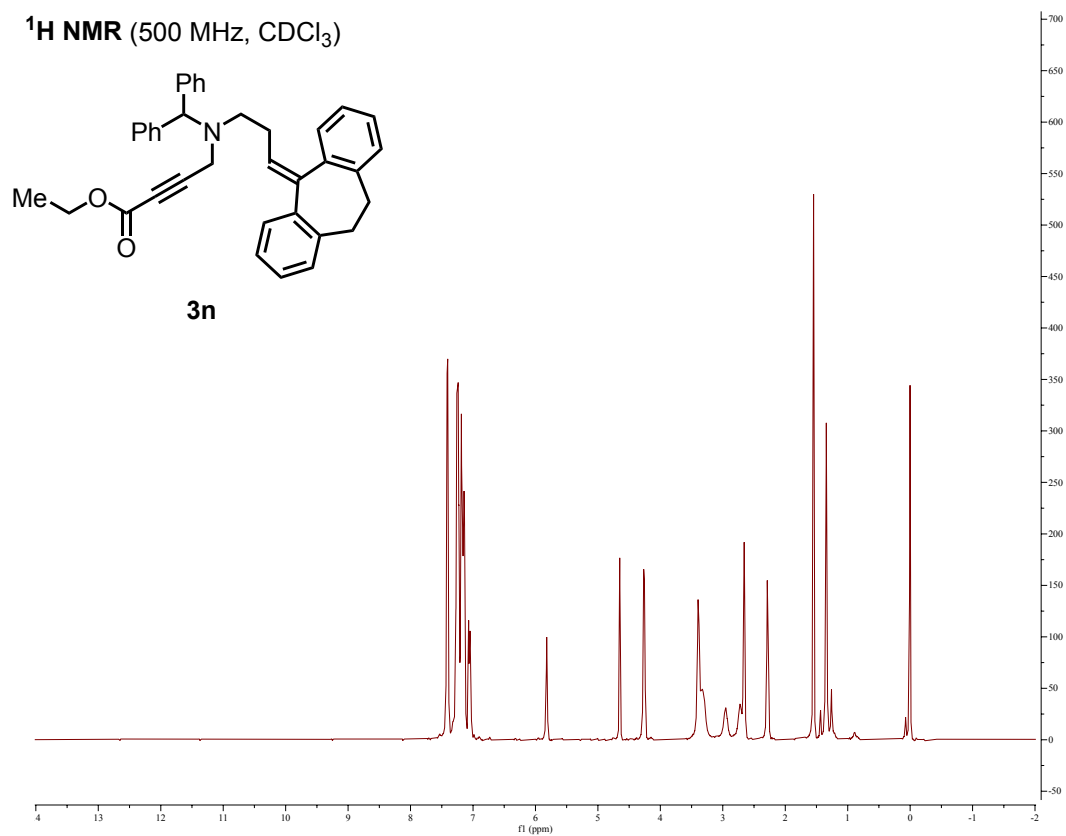


¹H NMR (500 MHz, CDCl₃)

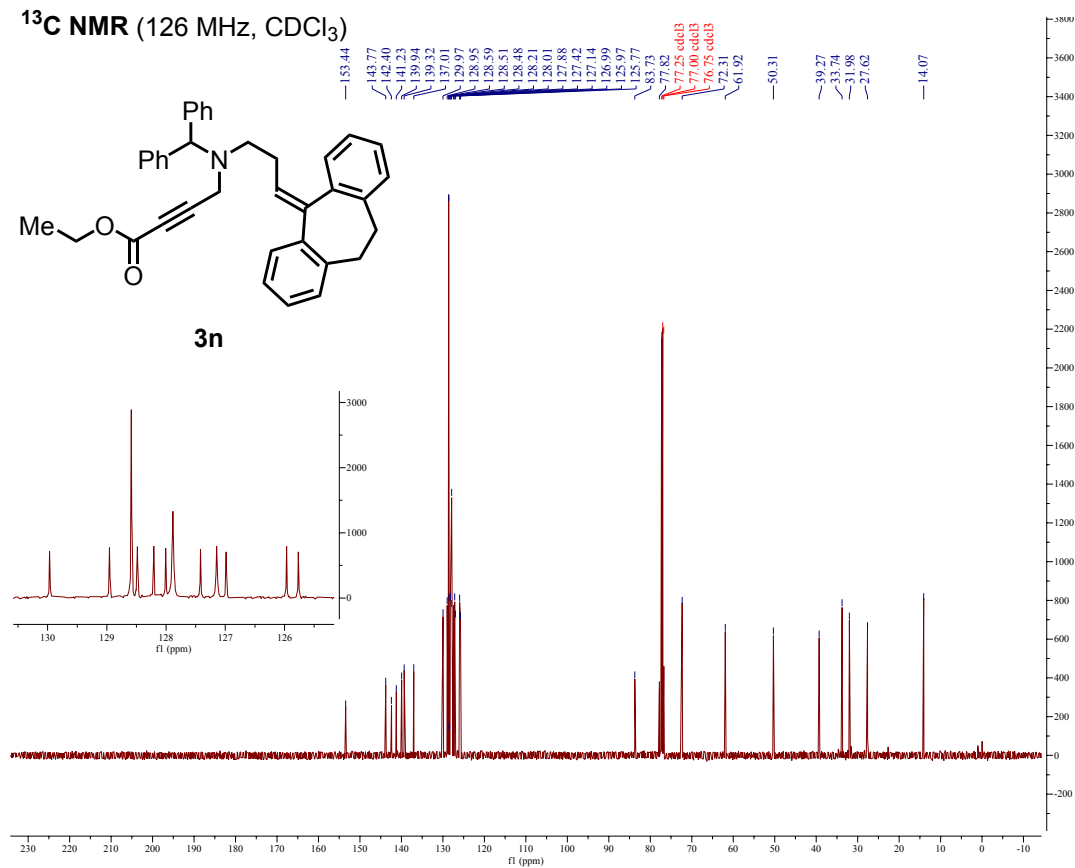
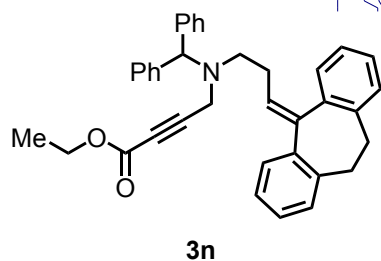


^{13}C NMR (126 MHz, CDCl_3)

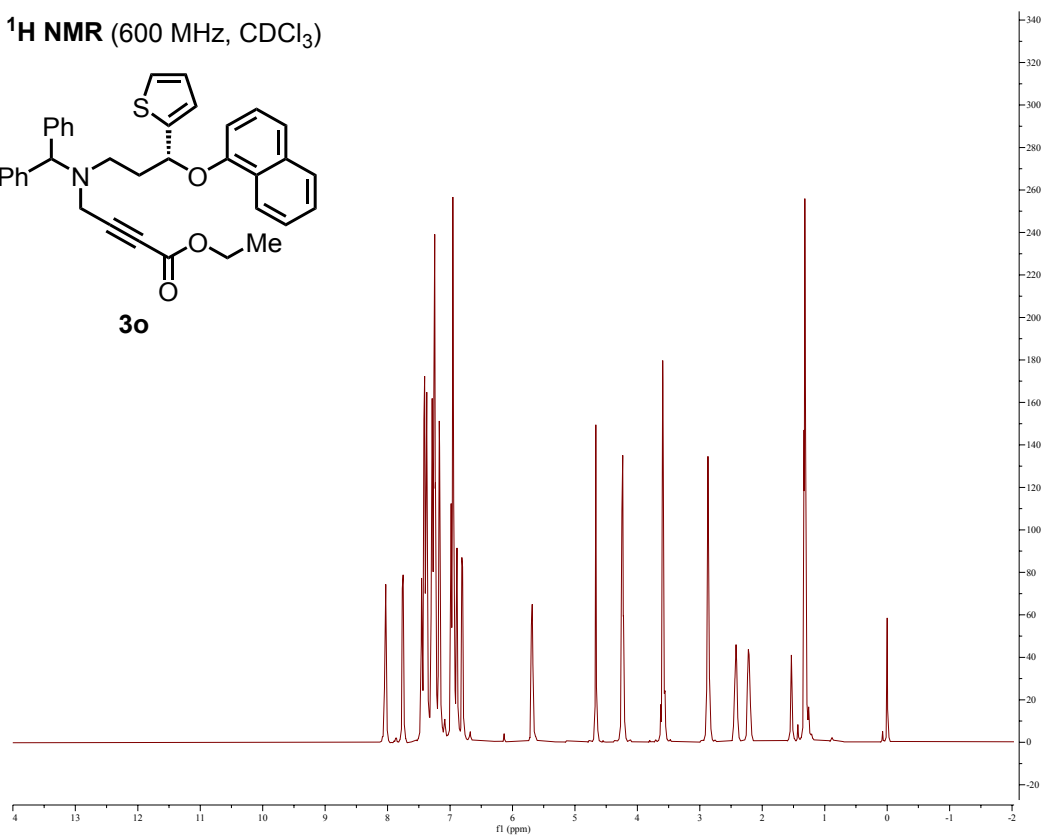
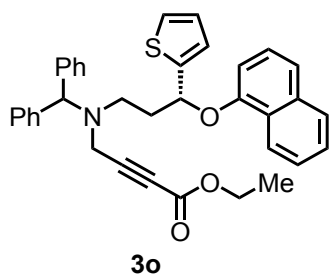




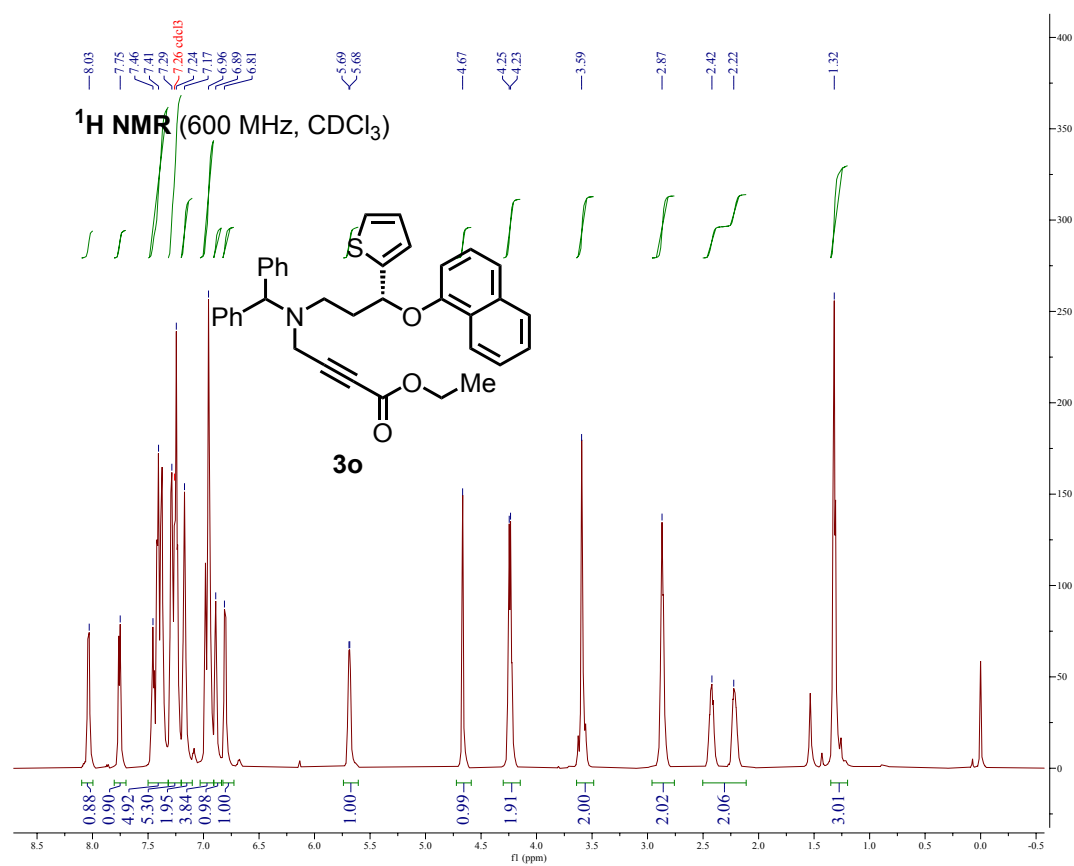
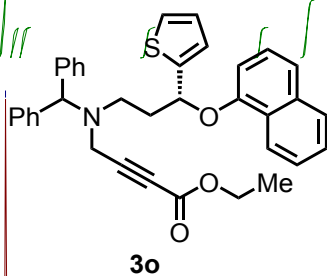
^{13}C NMR (126 MHz, CDCl_3)



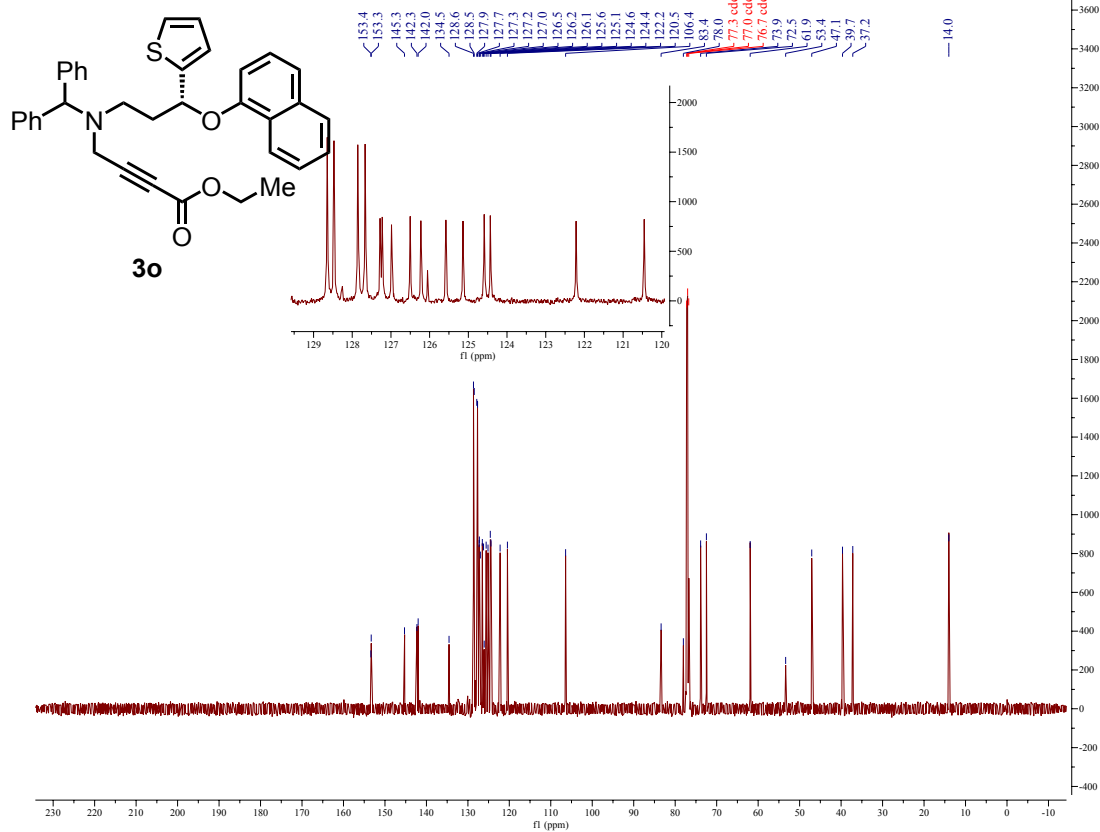
¹H NMR (600 MHz, CDCl₃)



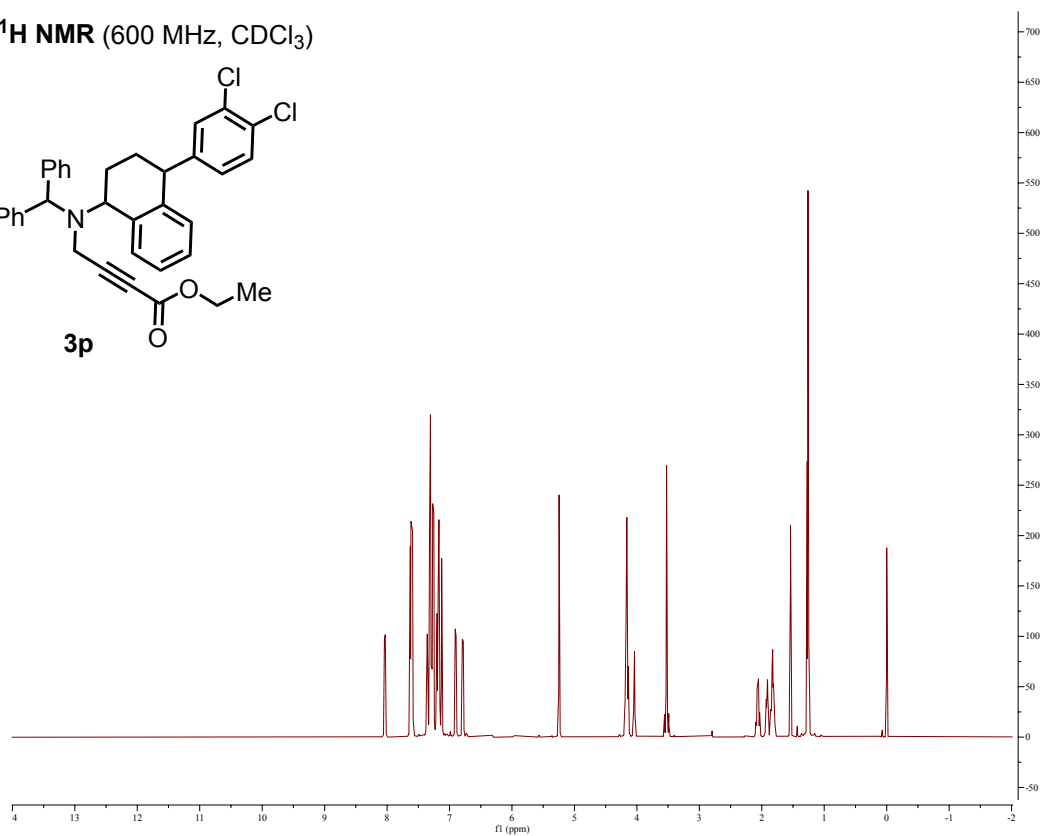
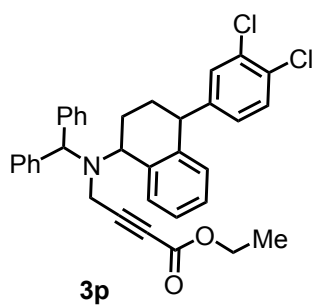
¹H NMR (600 MHz, CDCl₃)



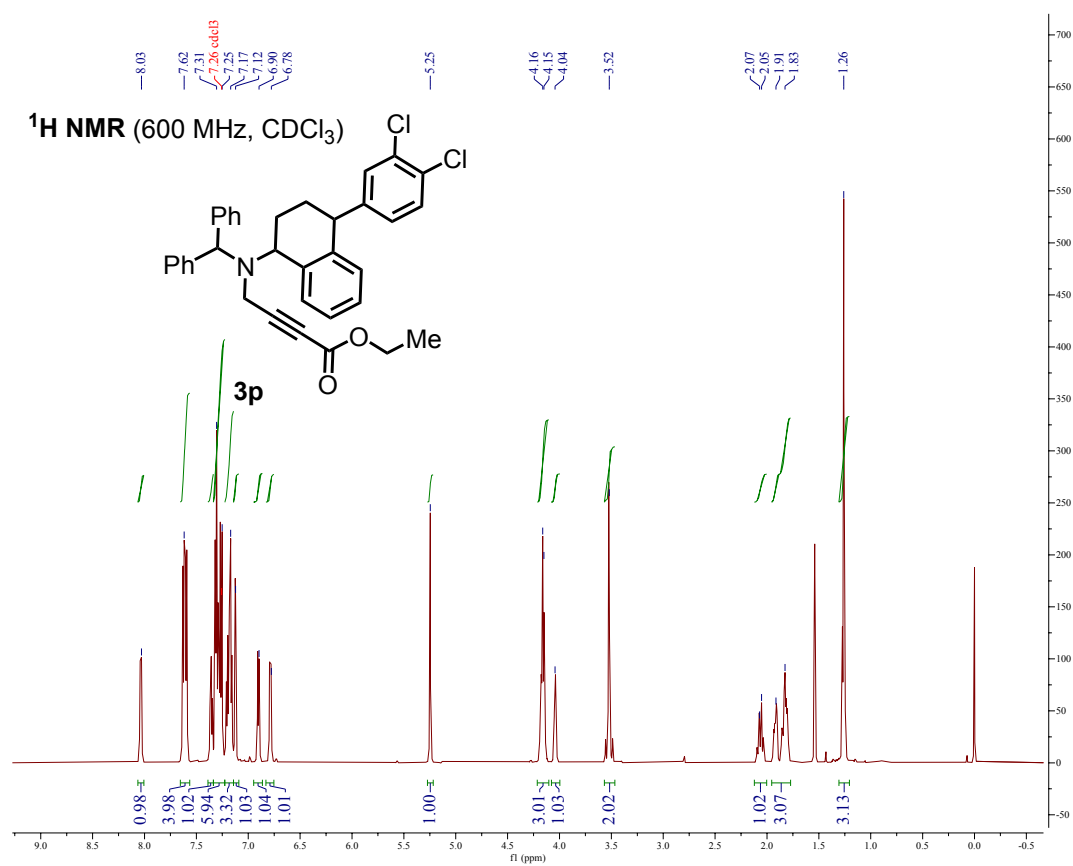
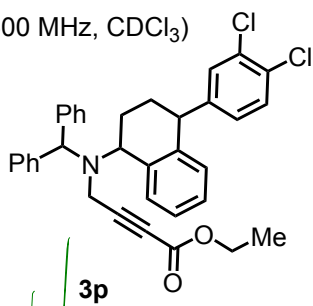
^{13}C NMR (126 MHz, CDCl_3)



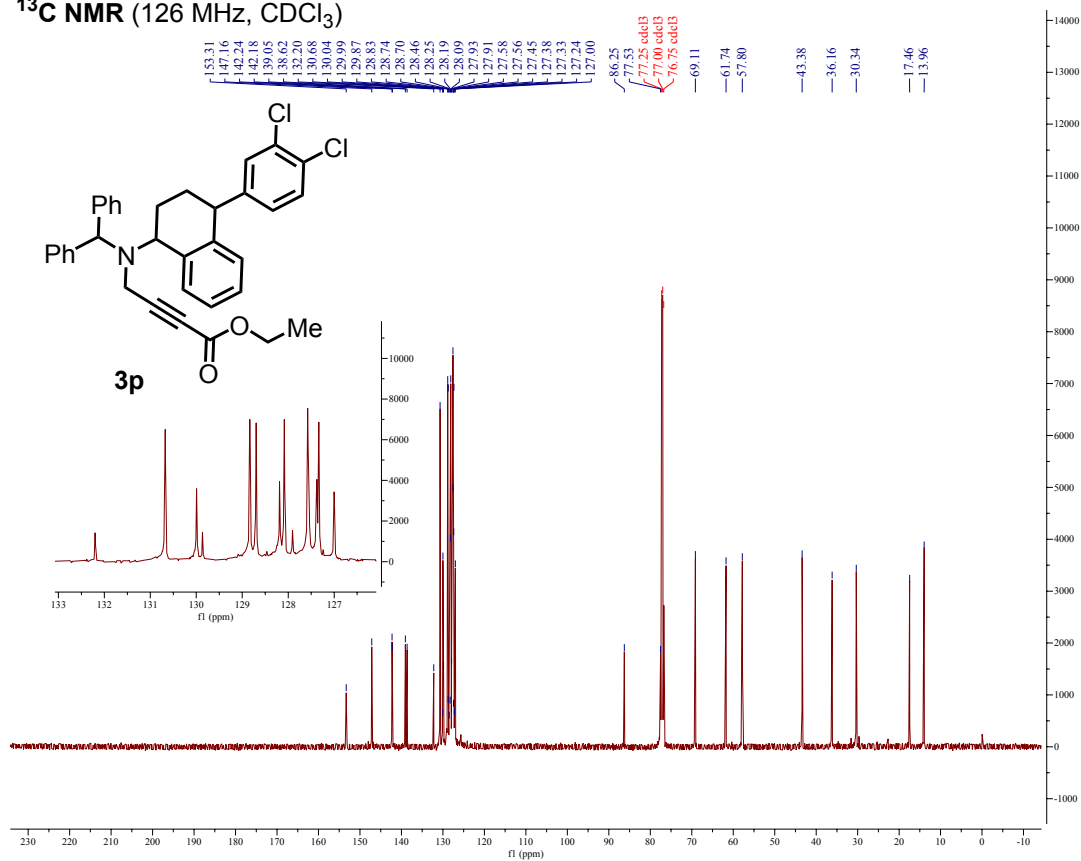
¹H NMR (600 MHz, CDCl₃)



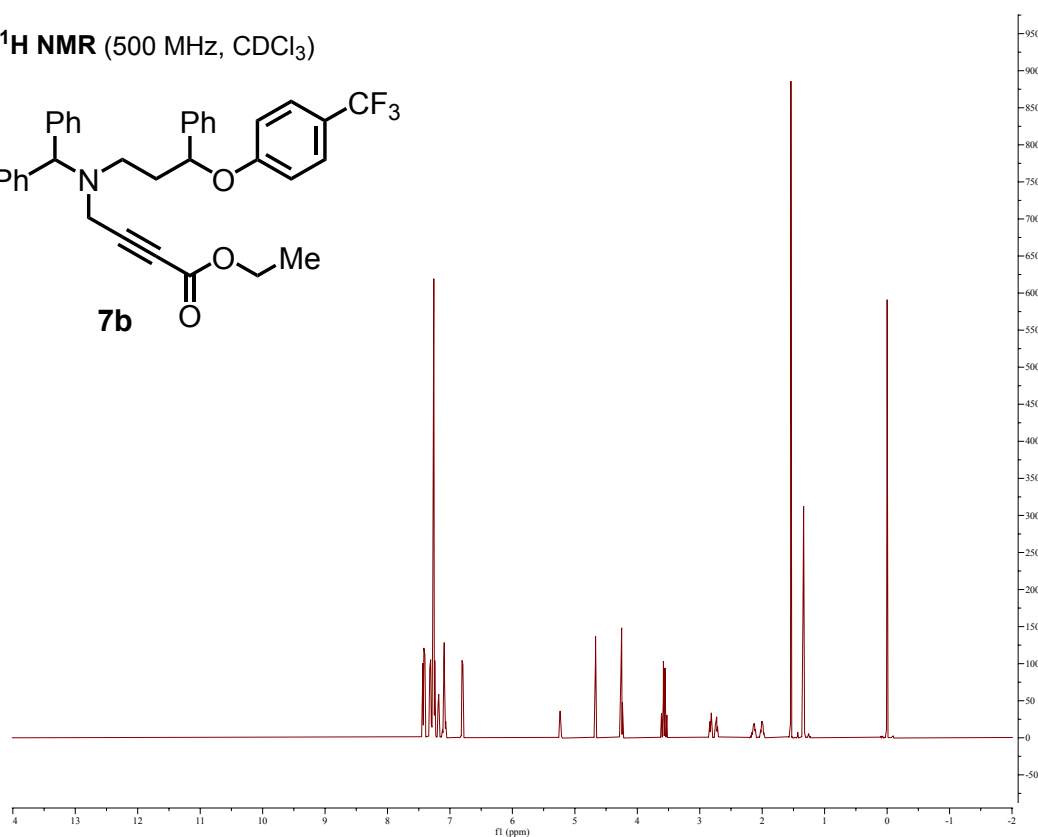
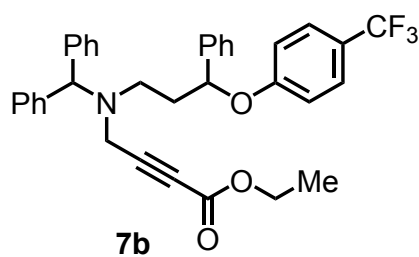
¹H NMR (600 MHz, CDCl₃)



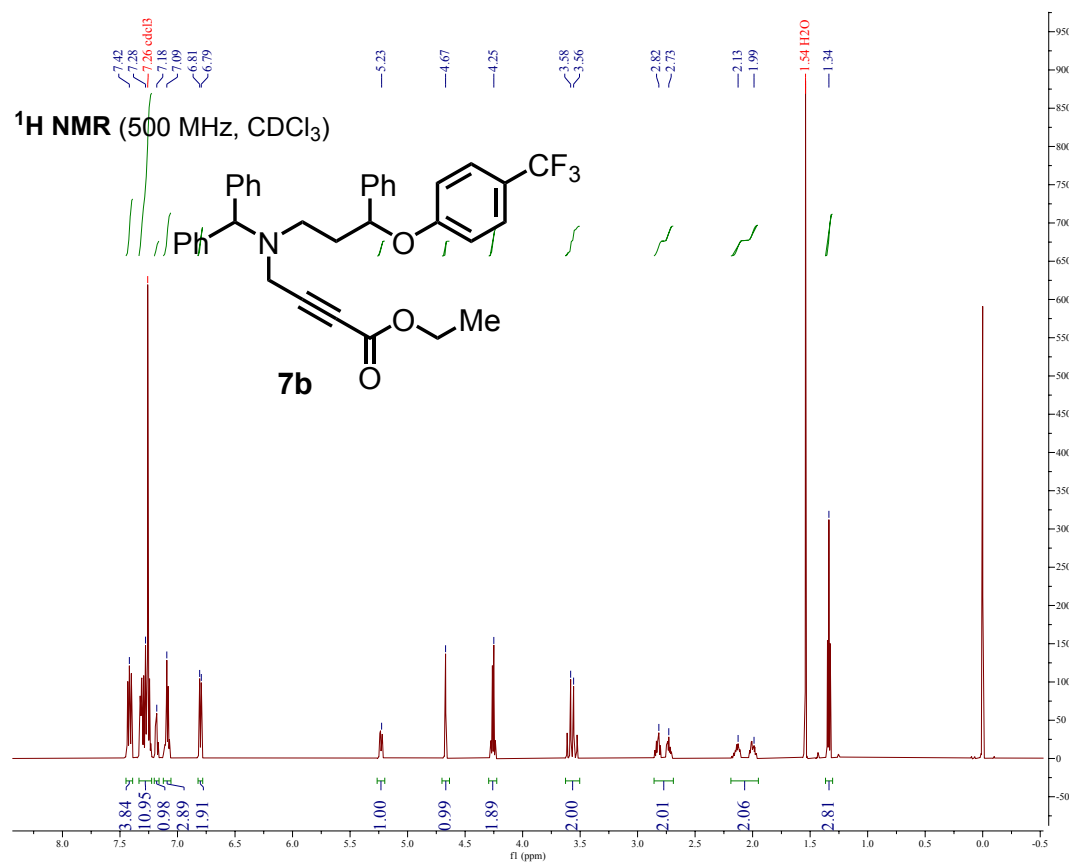
^{13}C NMR (126 MHz, CDCl_3)



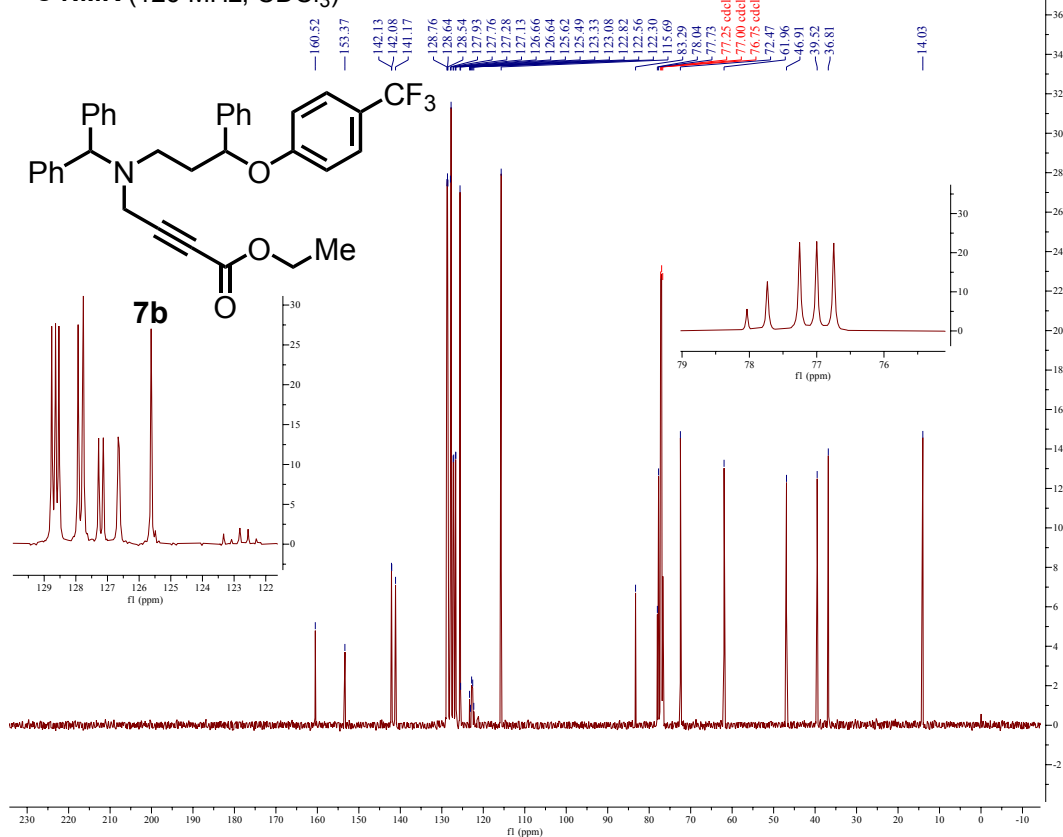
¹H NMR (500 MHz, CDCl₃)



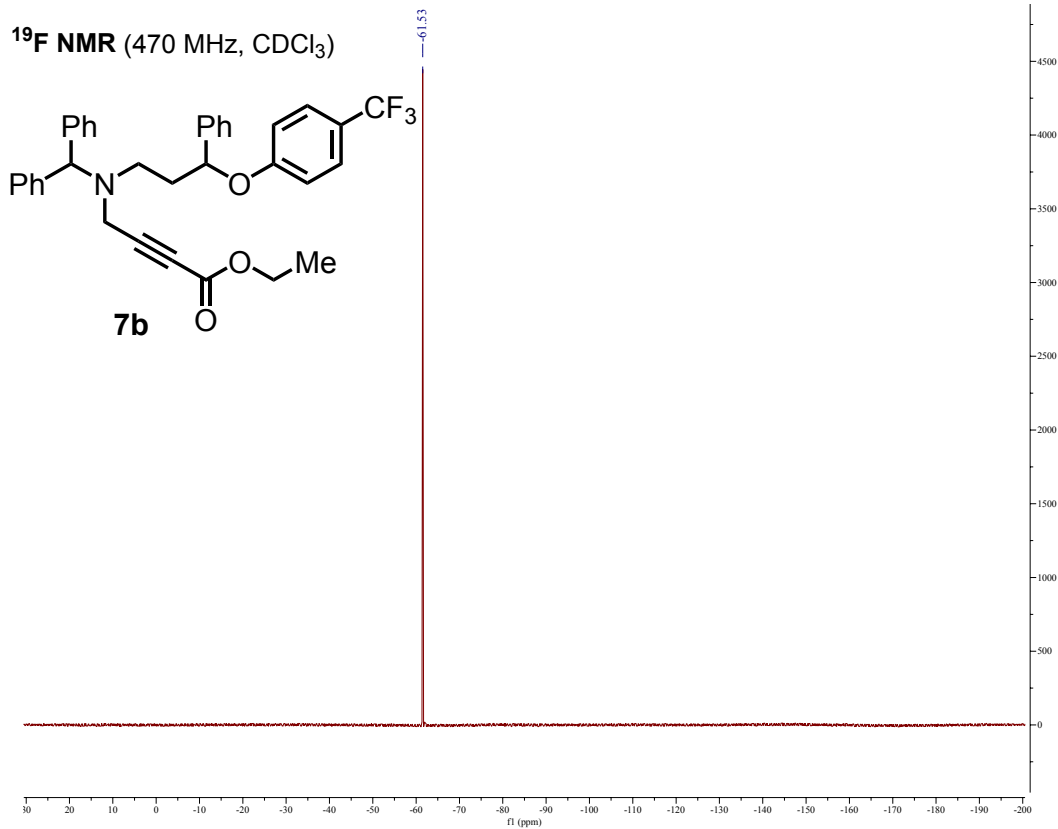
¹H NMR (500 MHz, CDCl₃)



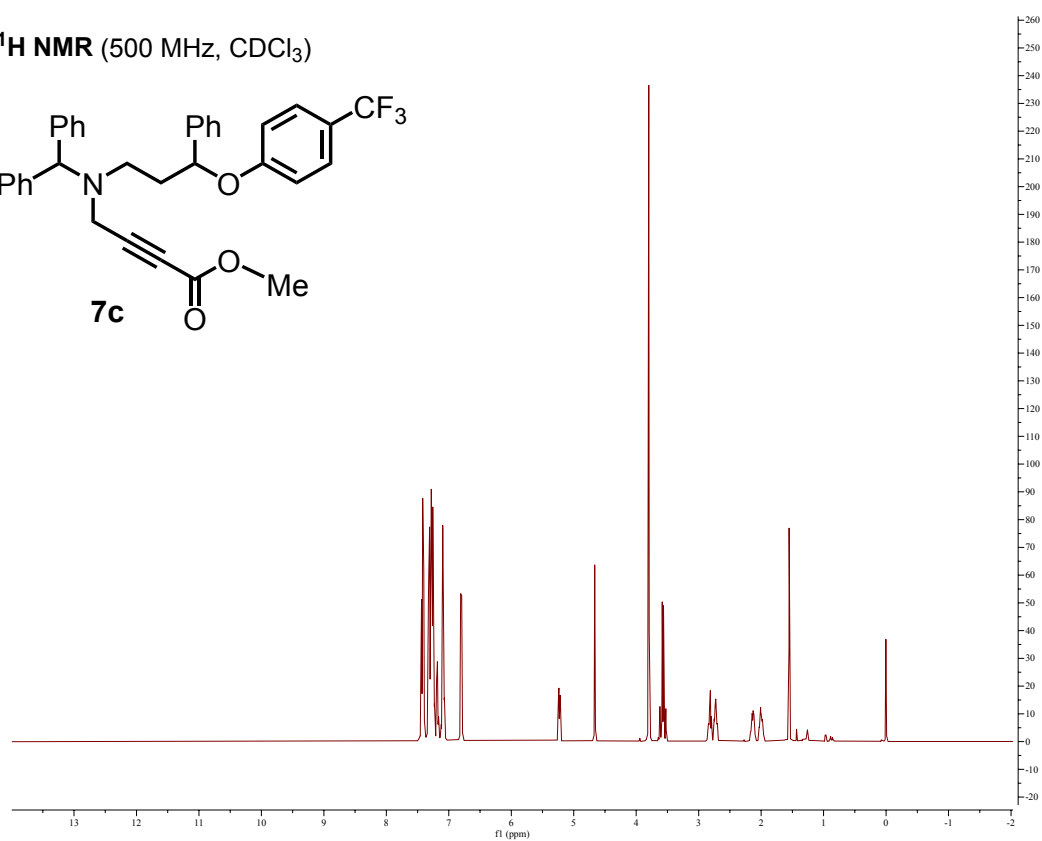
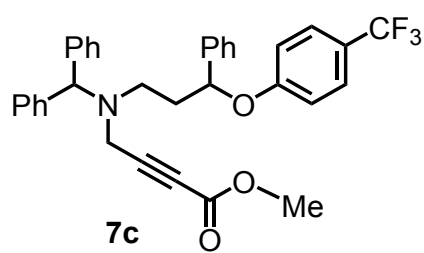
^{13}C NMR (126 MHz, CDCl_3)



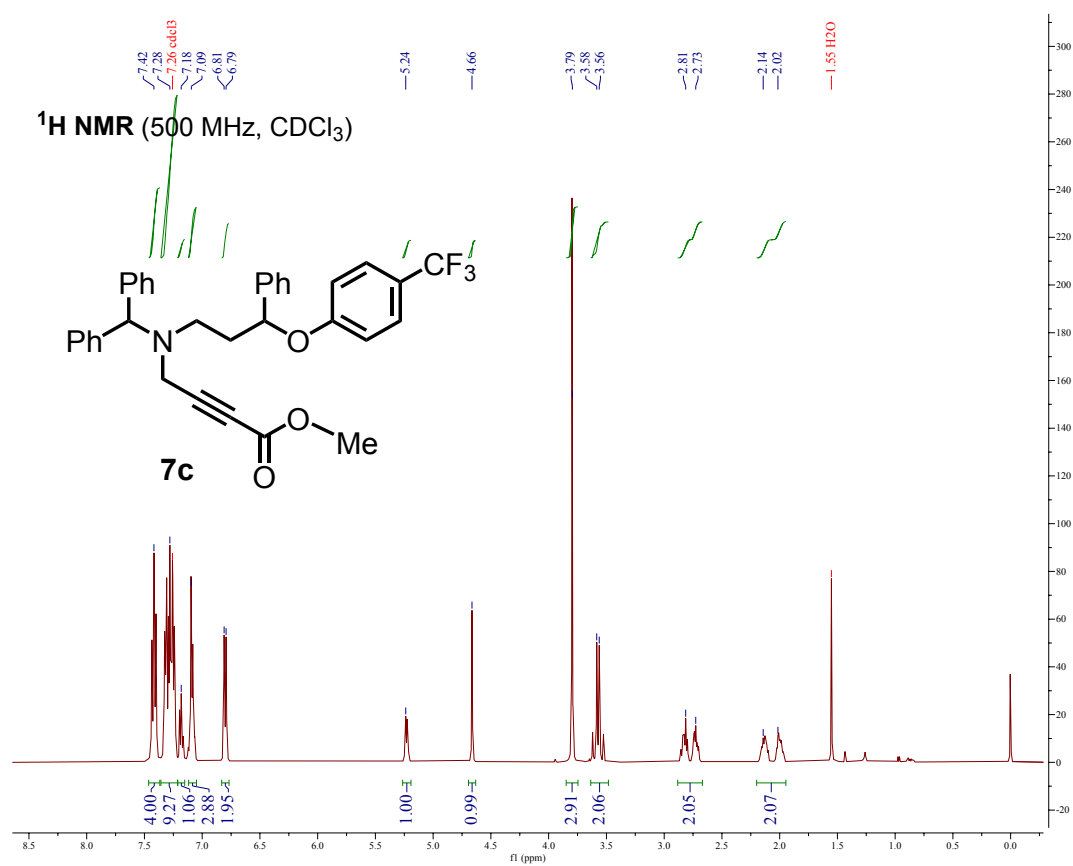
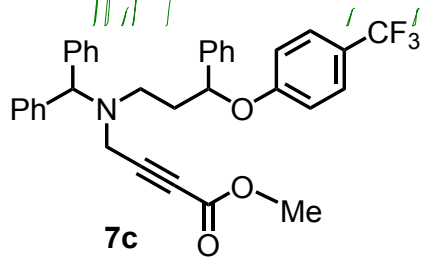
^{19}F NMR (470 MHz, CDCl_3)



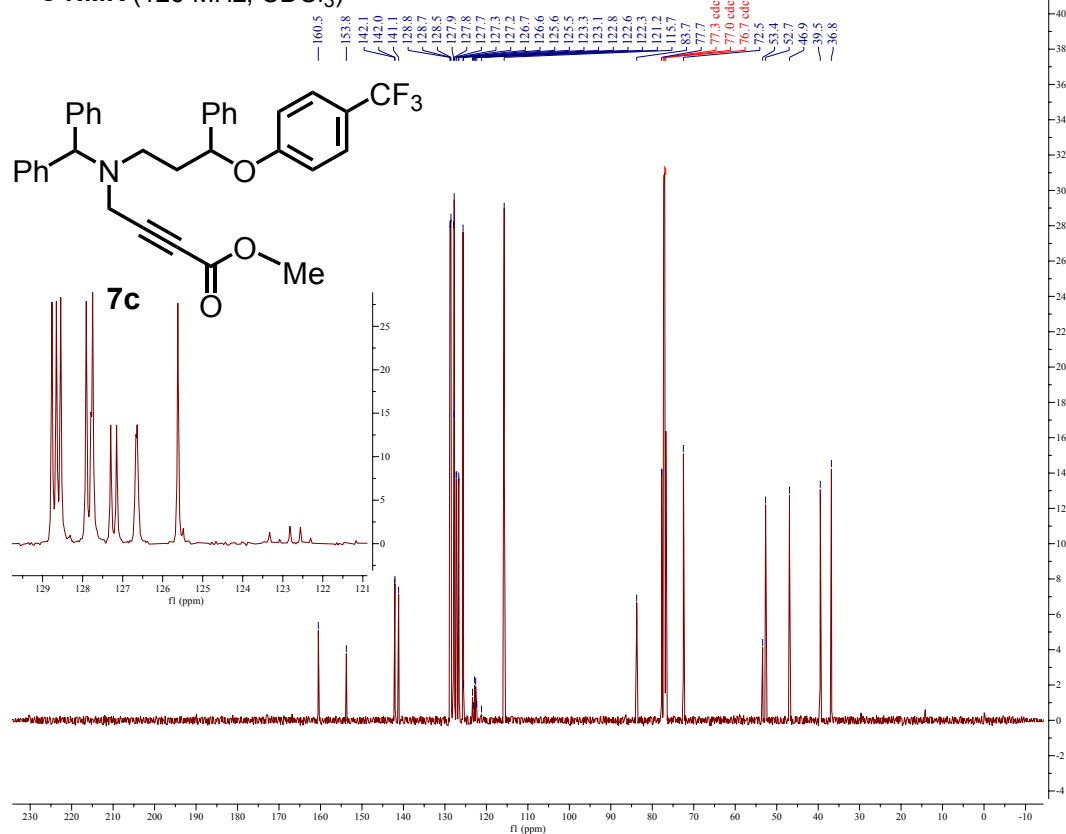
¹H NMR (500 MHz, CDCl₃)



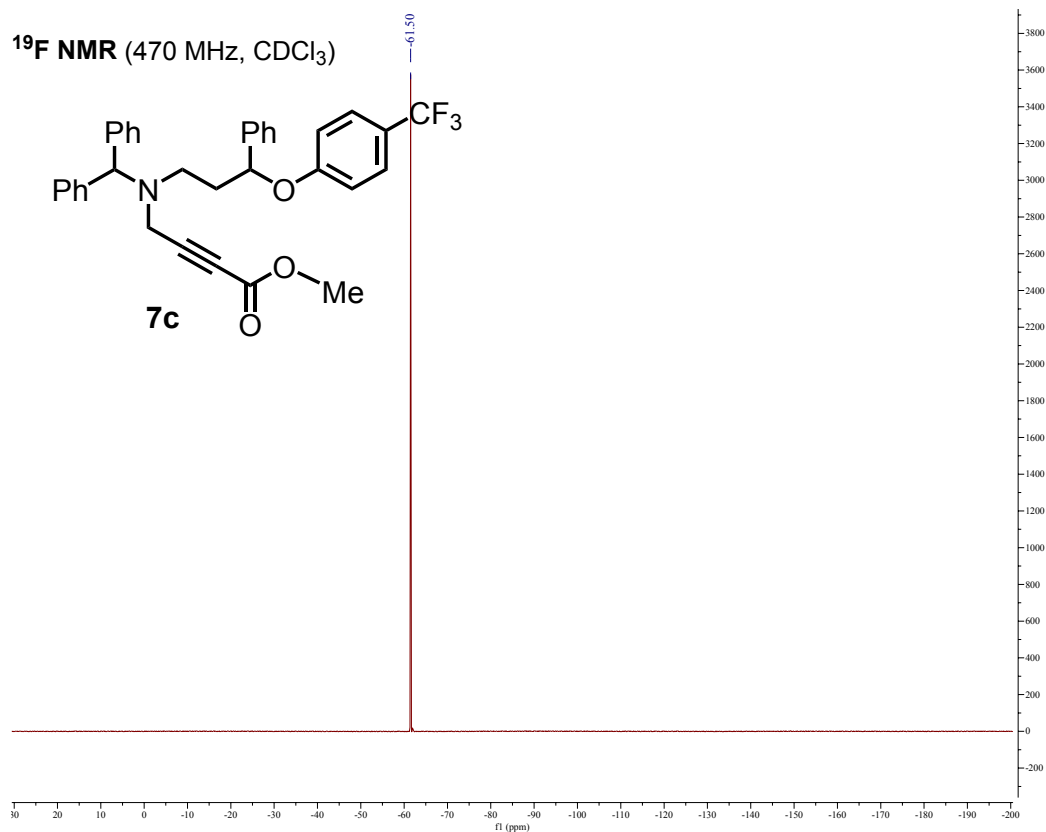
¹H NMR (500 MHz, CDCl₃)



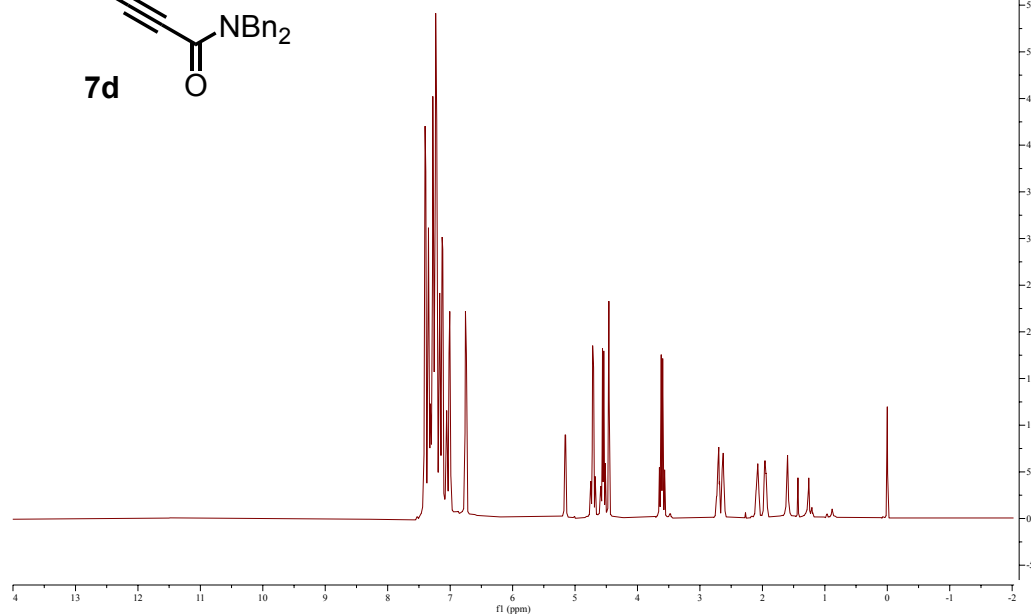
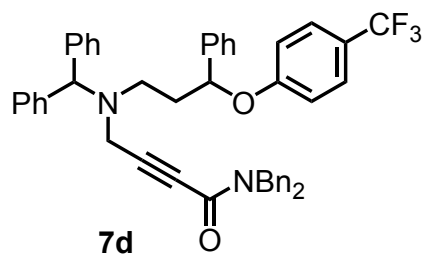
¹³C NMR (126 MHz, CDCl₃)



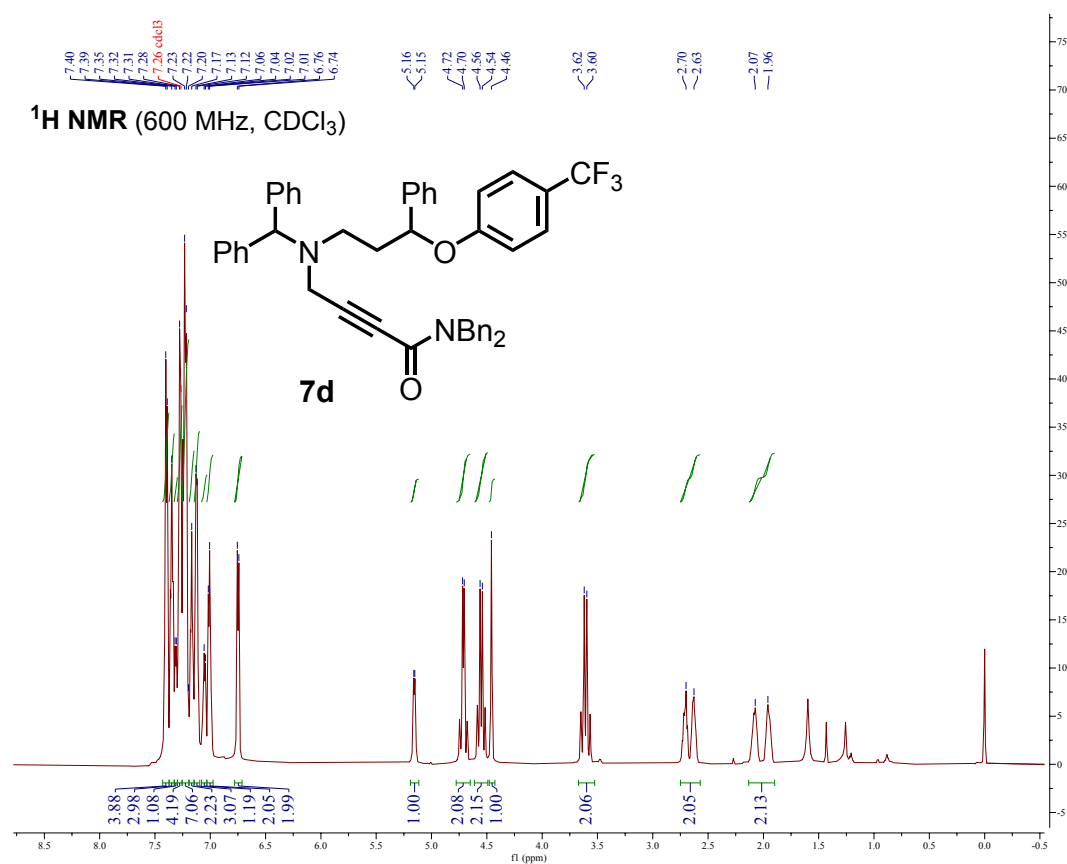
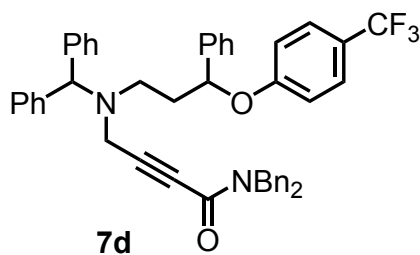
¹⁹F NMR (470 MHz, CDCl₃)



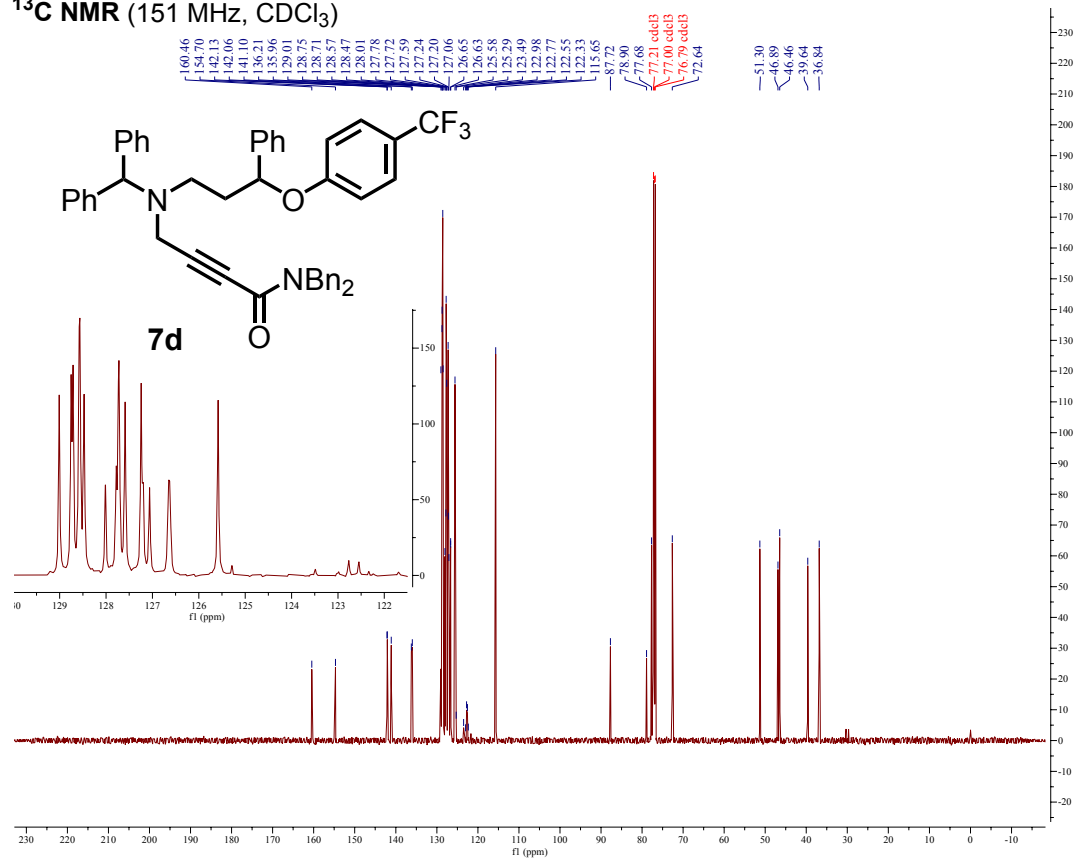
¹H NMR (600 MHz, CDCl₃)



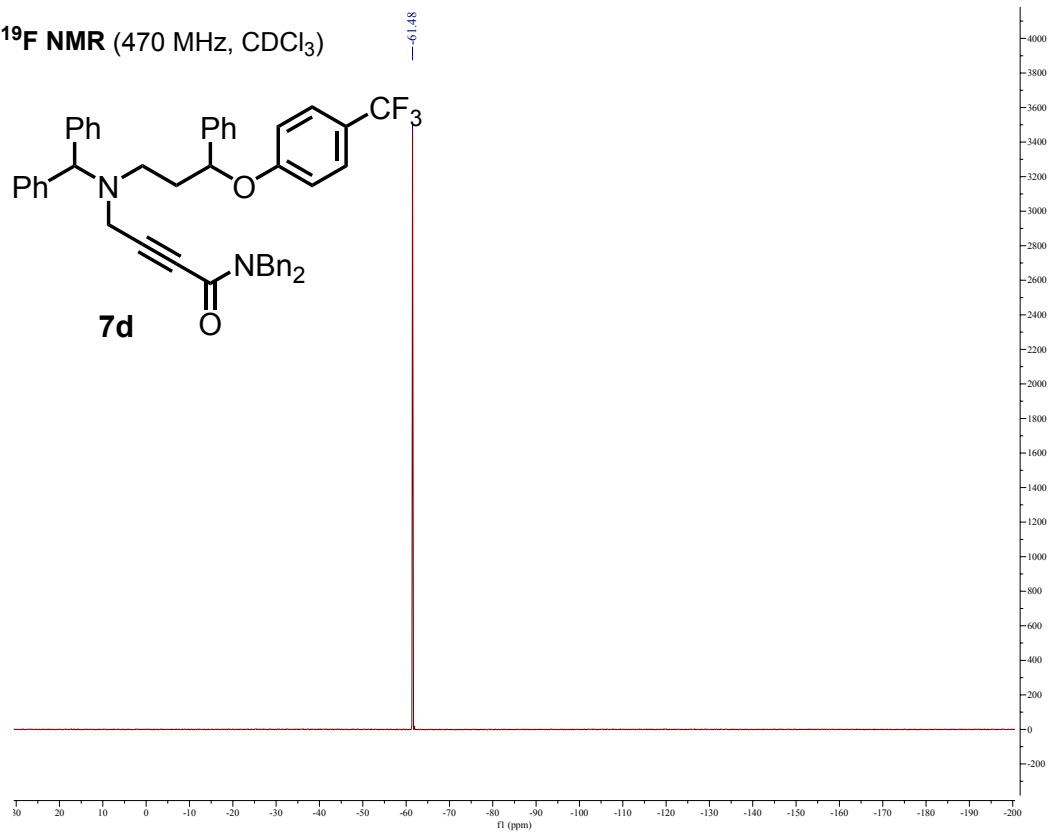
¹H NMR (600 MHz, CDCl₃)



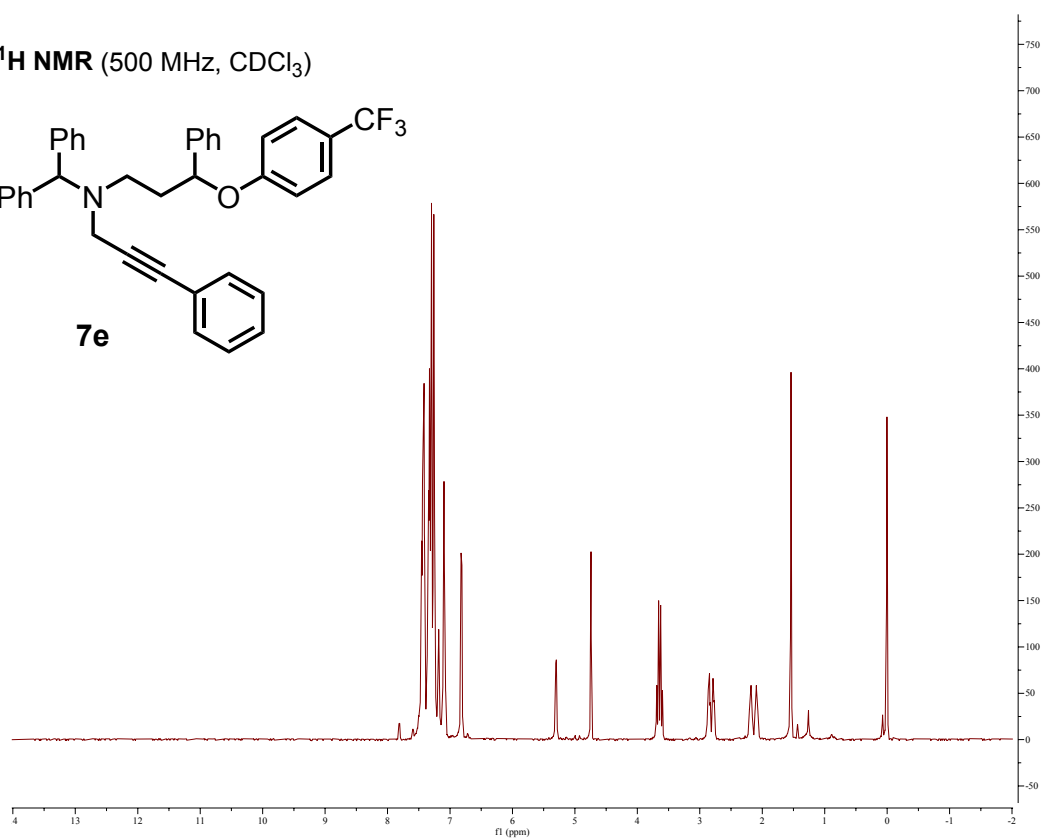
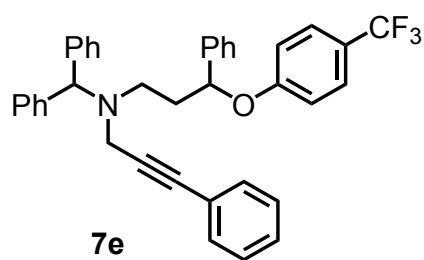
^{13}C NMR (151 MHz, CDCl_3)



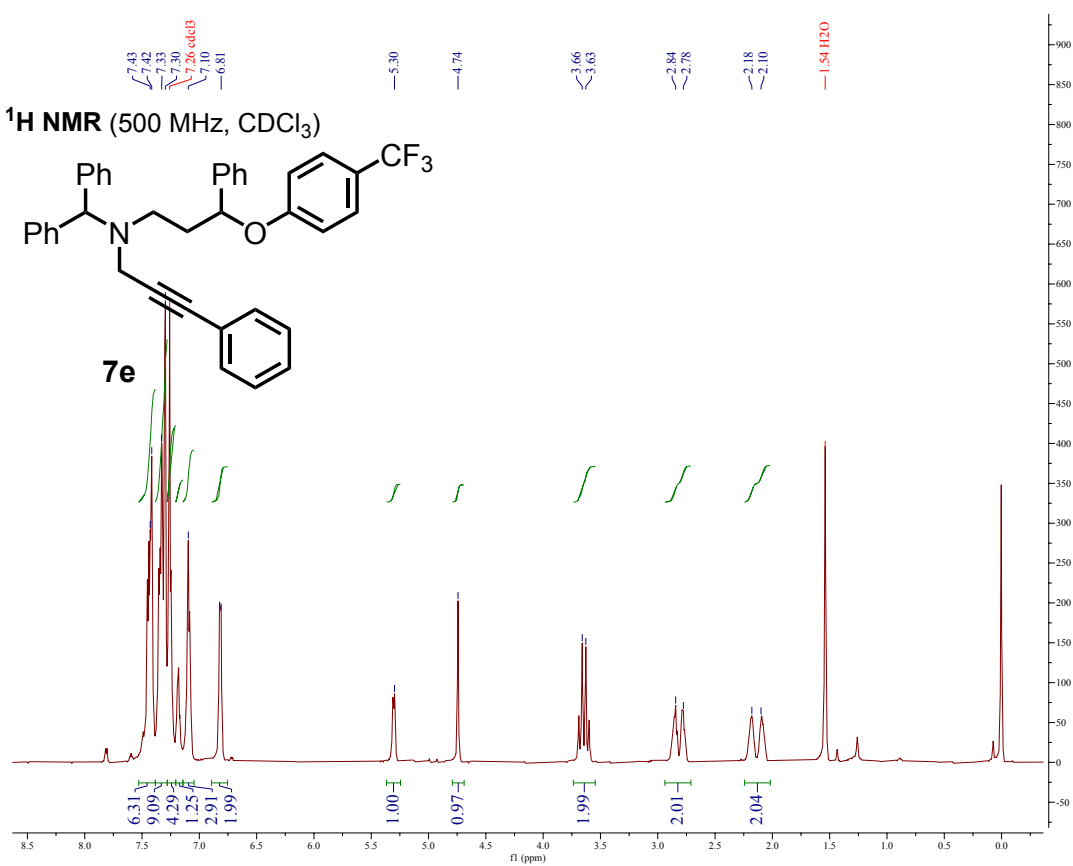
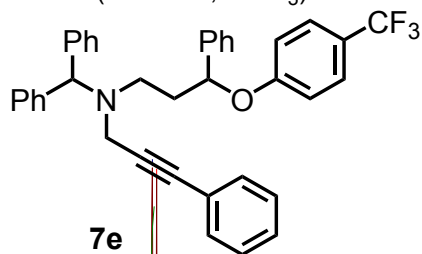
^{19}F NMR (470 MHz, CDCl_3)

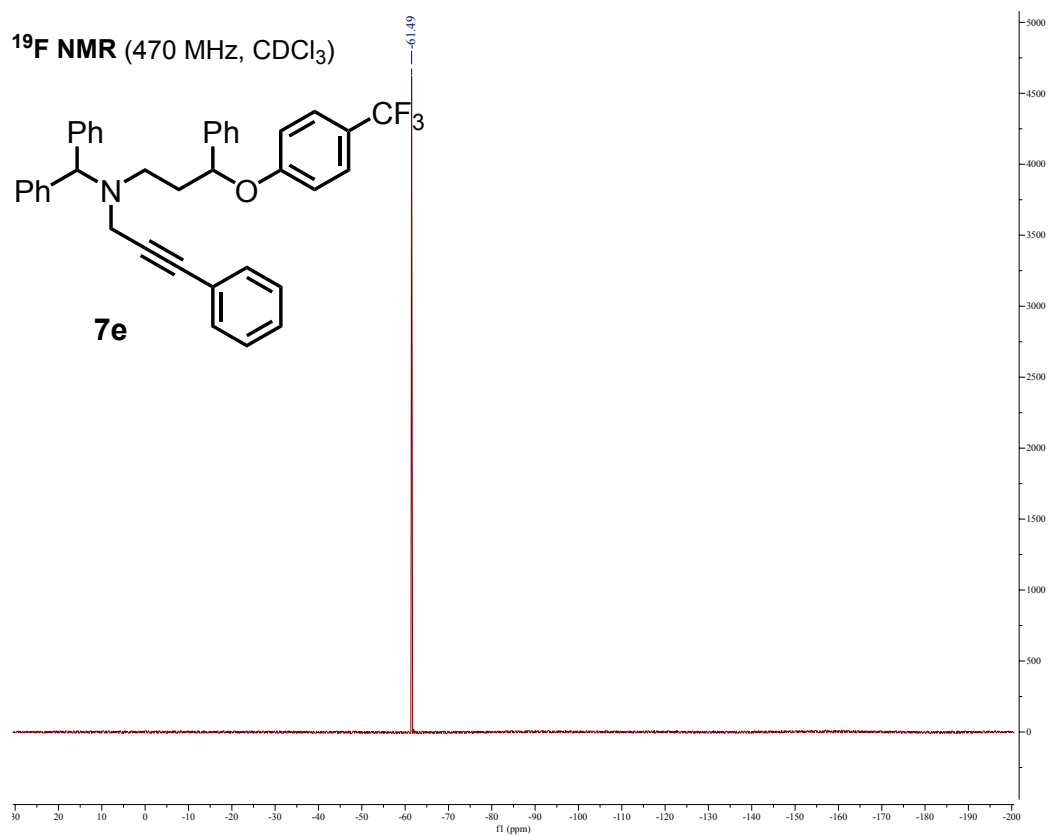
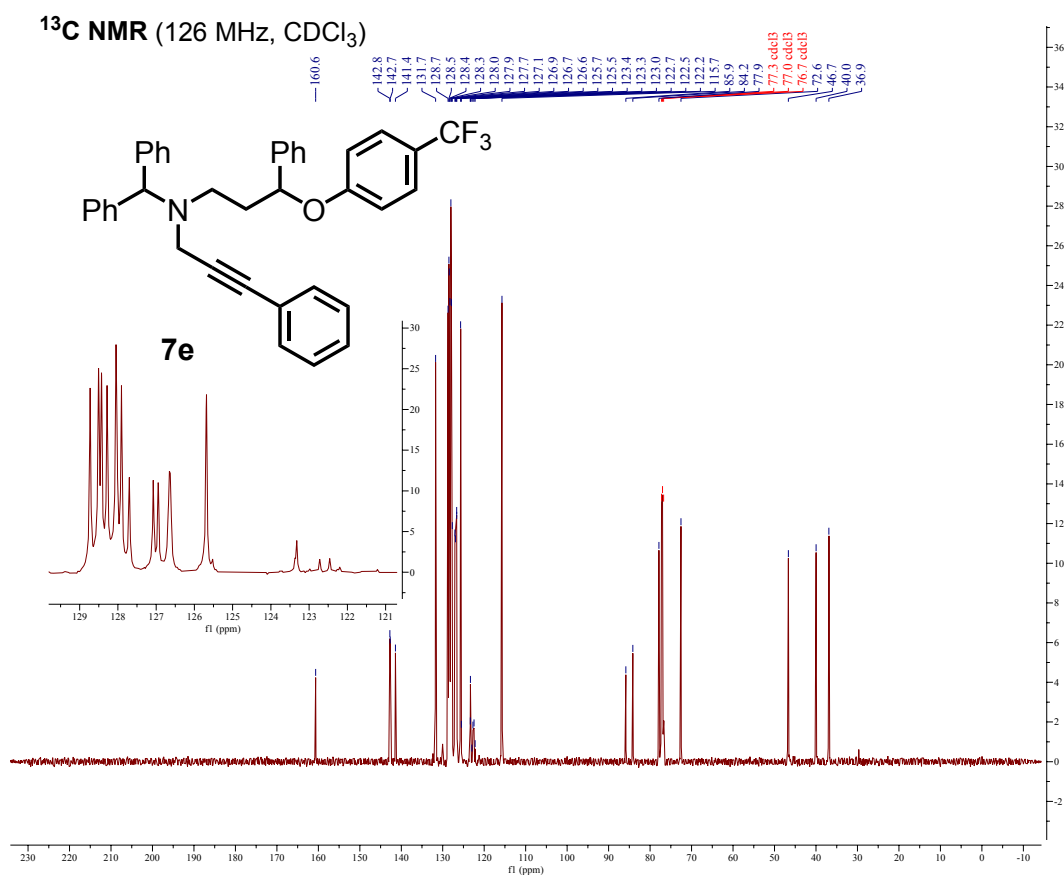


¹H NMR (500 MHz, CDCl₃)



¹H NMR (500 MHz, CDCl₃)





^1H NMR (500 MHz, CDCl_3)

7f

Chemical structure of **7f**: c1ccc(cc1)C#CCN(Cc2ccccc2)C(c3ccccc3)(c4ccccc4)CCOC(c5ccc(cc5)C(F)(F)F)C6=CC=CC=C6

^1H NMR spectrum (500 MHz, CDCl_3) showing peaks in the aromatic region (7.2-7.4 ppm), aliphatic region (4.8 ppm), and alkyne region (3.6 ppm, 3.4 ppm, 2.3 ppm, 2.1 ppm, 1.9 ppm, 1.7 ppm, 1.5 ppm, 1.3 ppm, 1.1 ppm, 0.9 ppm, 0.7 ppm, 0.5 ppm, 0.3 ppm, 0.1 ppm).

¹H NMR (500 MHz, CDCl₃)

7f

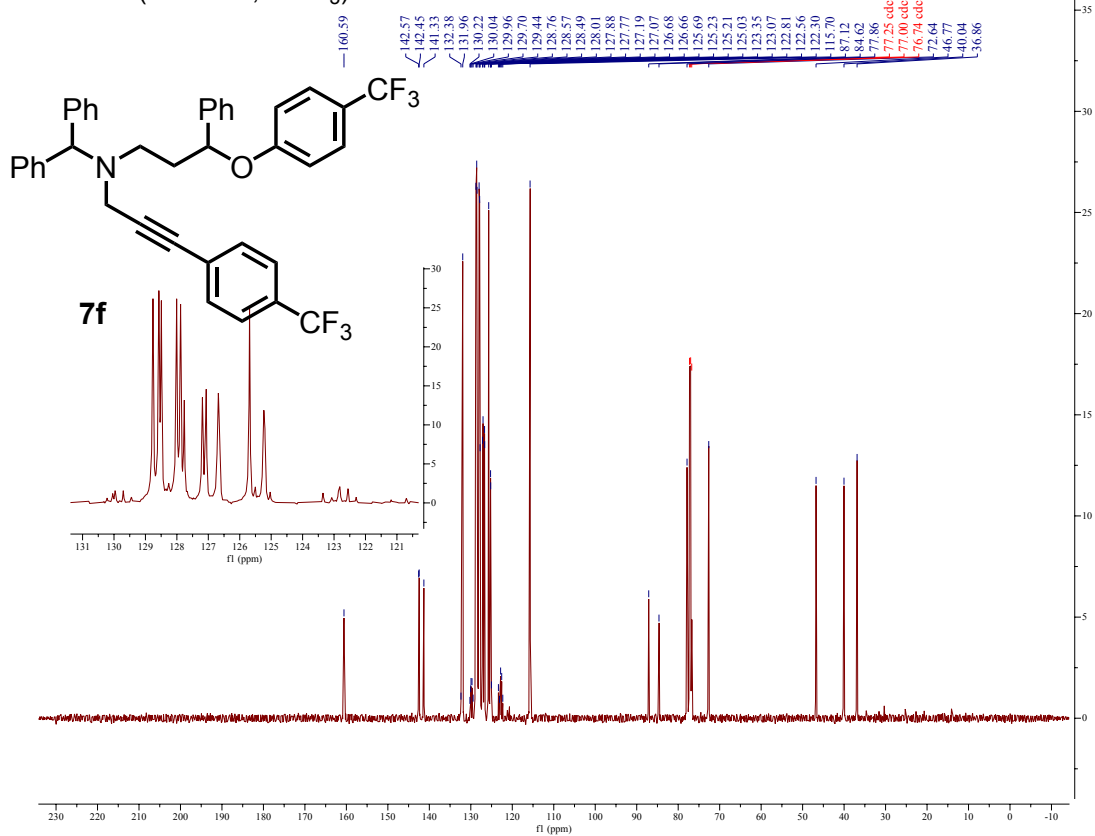
c1ccc(cc1)C#CCN(Cc1ccccc1)C(c2ccccc2)CCOc3ccc(cc3)C(F)(F)F

Chemical structure of compound **7f** is shown above the spectrum. The structure is a tertiary amine with a 4-(trifluoromethyl)phenyl group, a 4-(trifluoromethyl)phenoxy group, and a 4-(trifluoromethyl)phenyl group attached to the nitrogen atom. The spectrum displays peaks corresponding to the protons in the molecule, with chemical shifts (ppm) and integrations (area) indicated below the baseline.

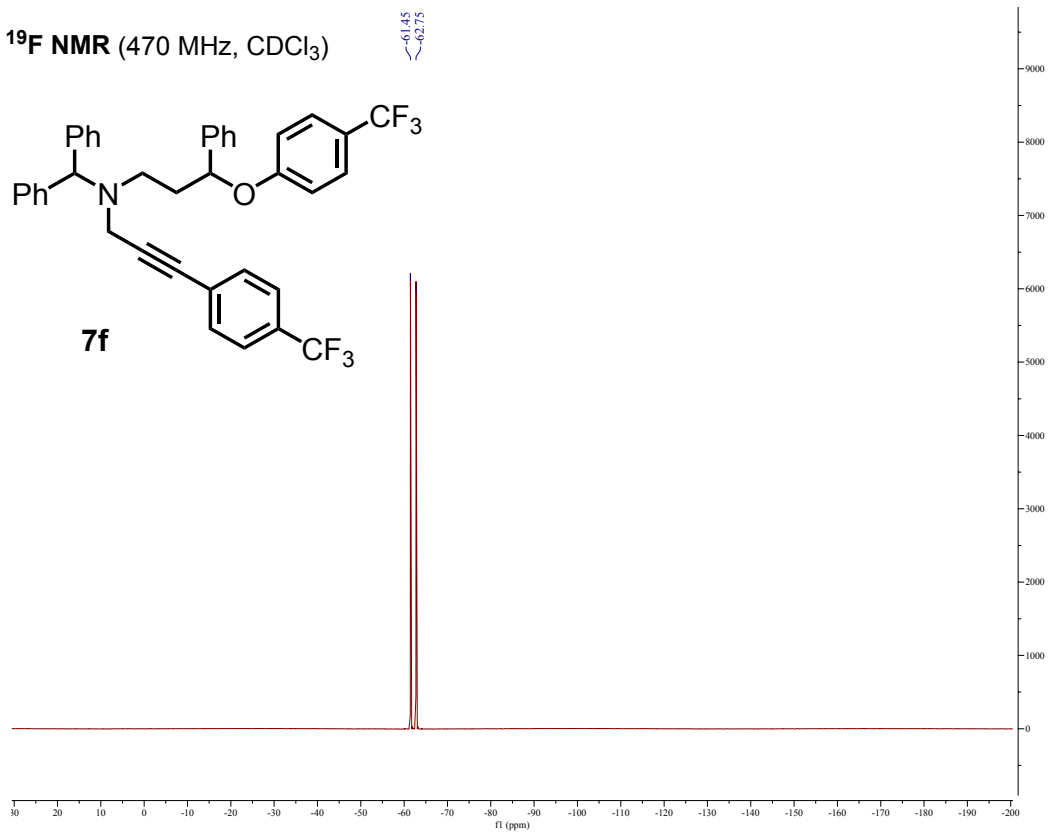
Chemical shifts (ppm) and integrations (area) are listed below the spectrum:

- 7.58, 7.51, 7.46, 7.45, 7.42, 7.36, 7.32, 7.29, 7.26 (cds), 7.21, 7.13, 6.84, 6.83 (aromatic protons, integration 1.85, 1.91, 3.87, 1.98, 6.94, 1.03, 2.93, 1.94)
- 5.31, 5.31 (m, integration 1.00)
- 4.75 (m, integration 0.98)
- 3.68 (m, integration 1.97)
- 2.88, 2.80 (m, integration 2.00)
- 2.21, 2.12 (m, integration 2.07)

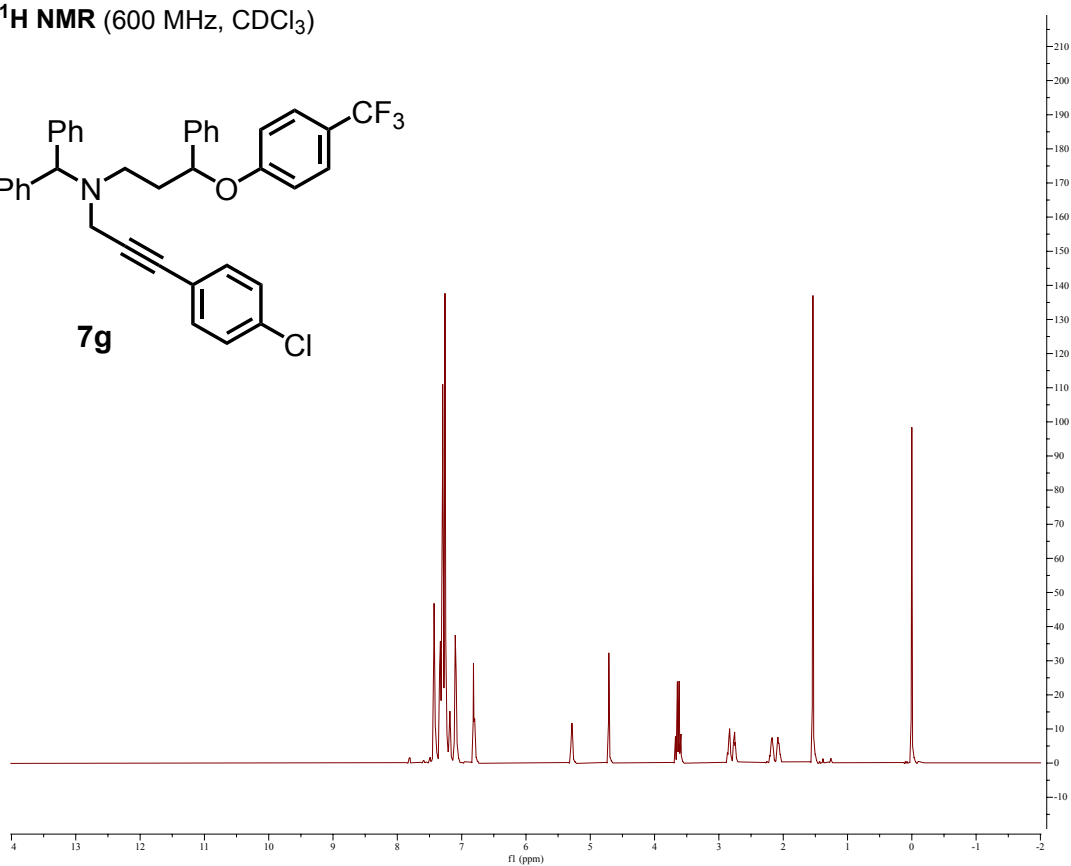
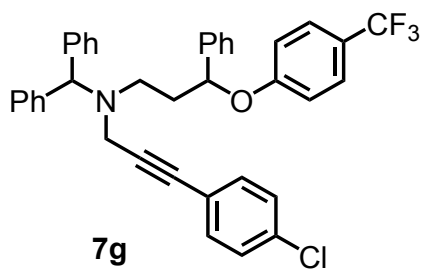
^{13}C NMR (126 MHz, CDCl_3)



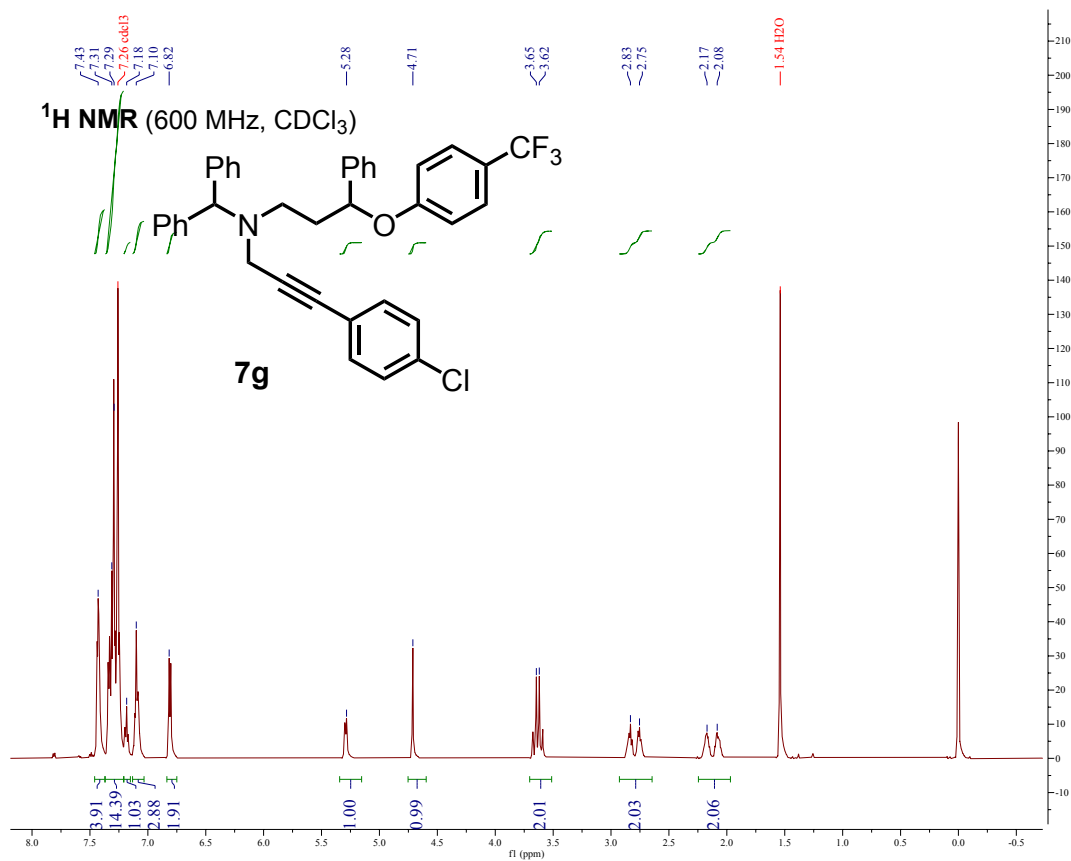
^{19}F NMR (470 MHz, CDCl_3)



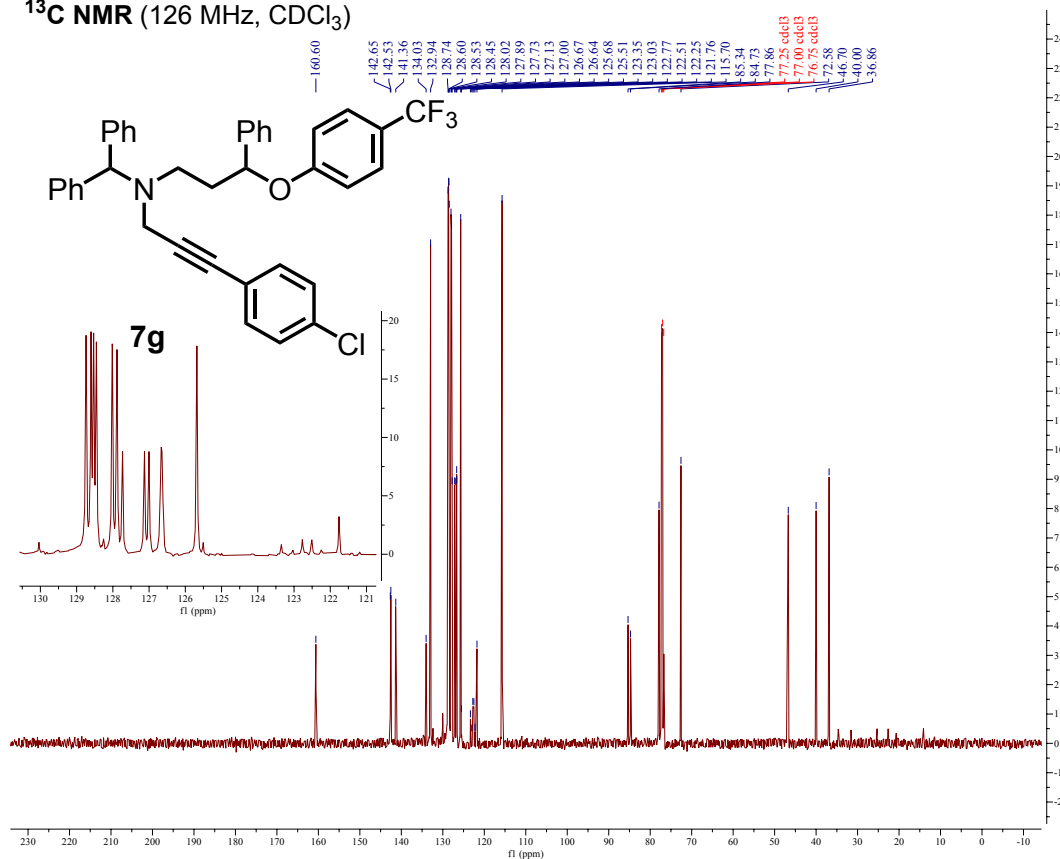
^1H NMR (600 MHz, CDCl_3)



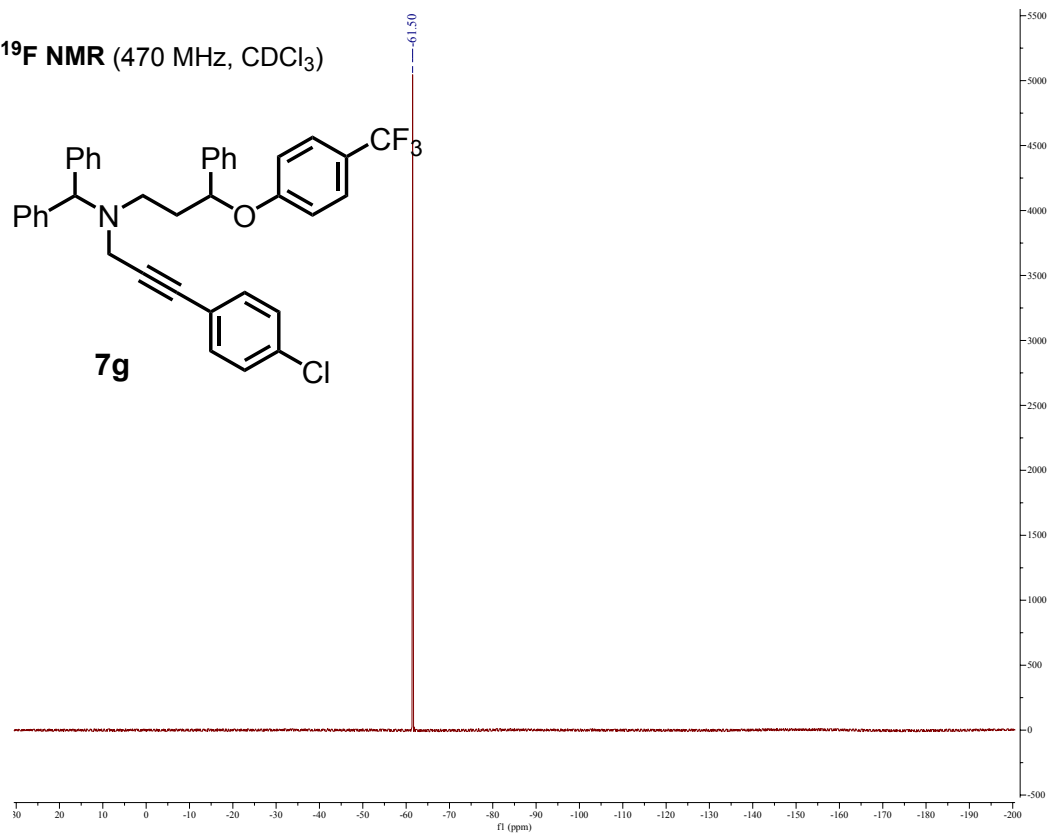
^1H NMR (600 MHz, CDCl_3)



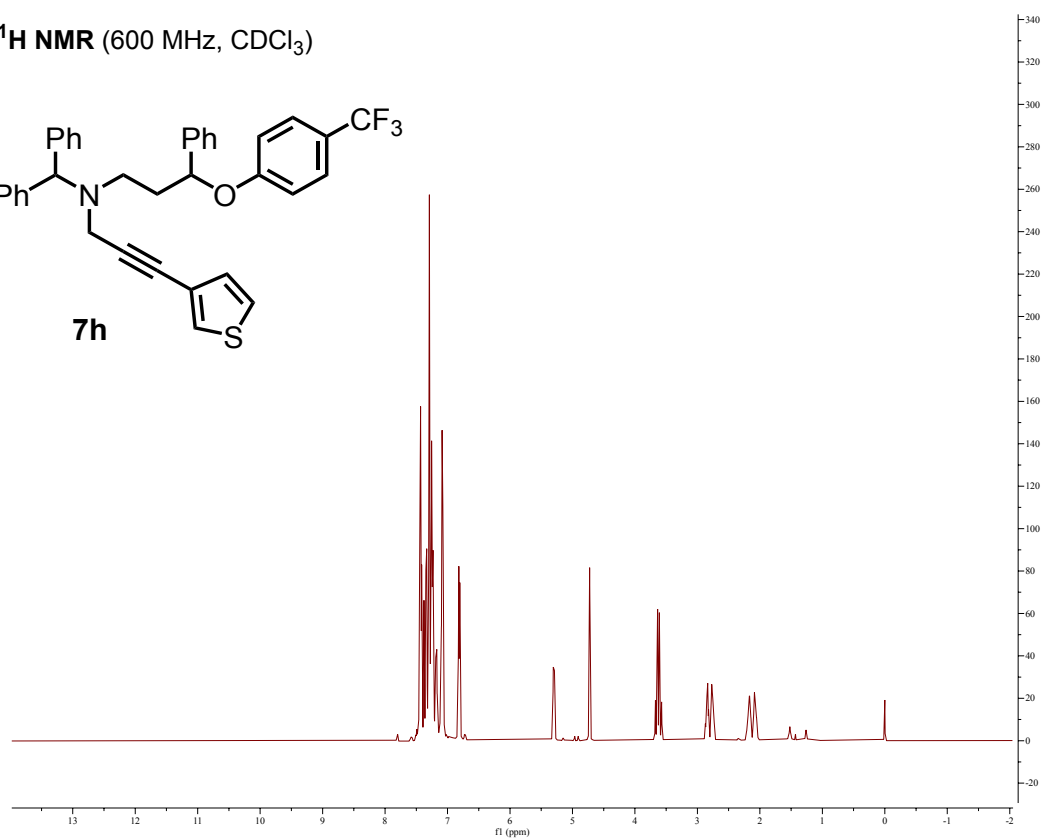
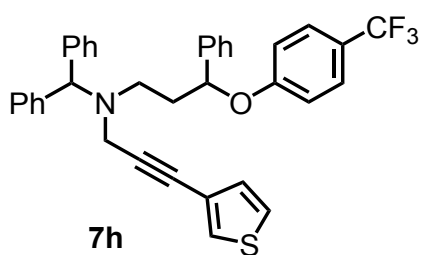
¹³C NMR (126 MHz, CDCl₃)



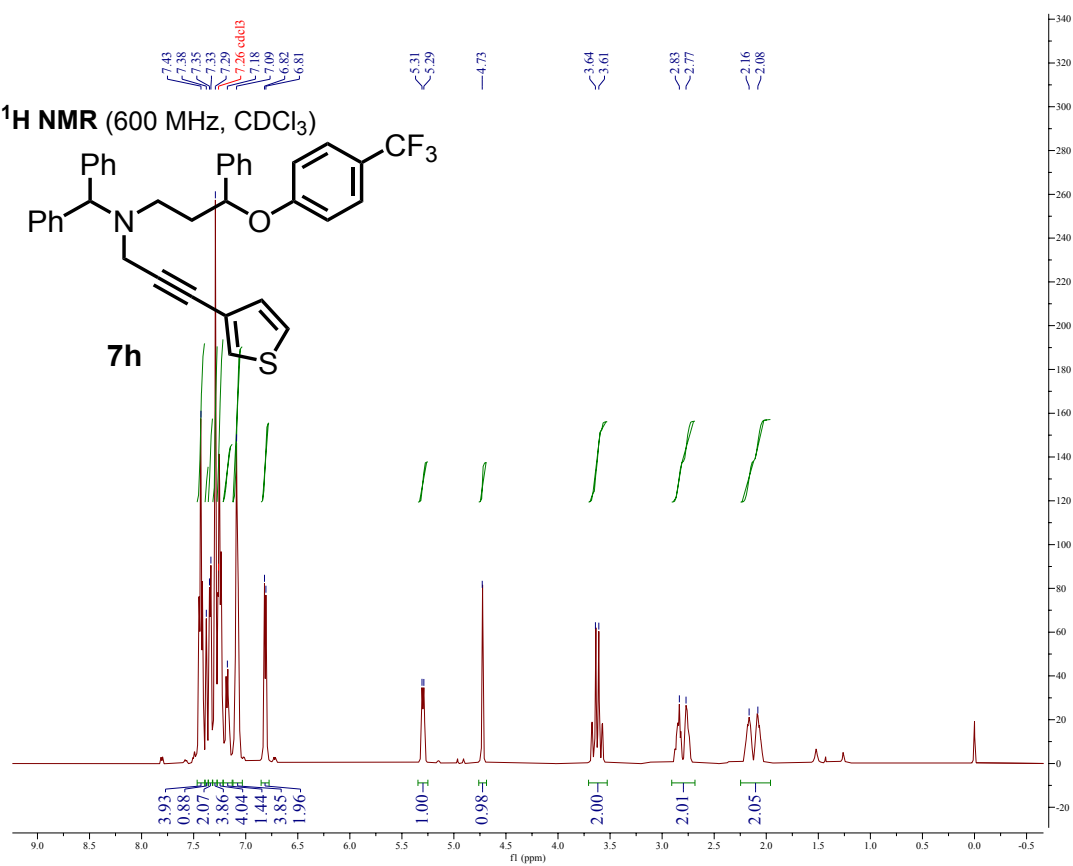
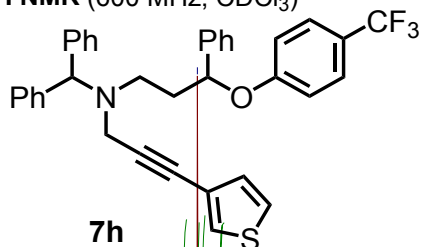
¹⁹F NMR (470 MHz, CDCl₃)



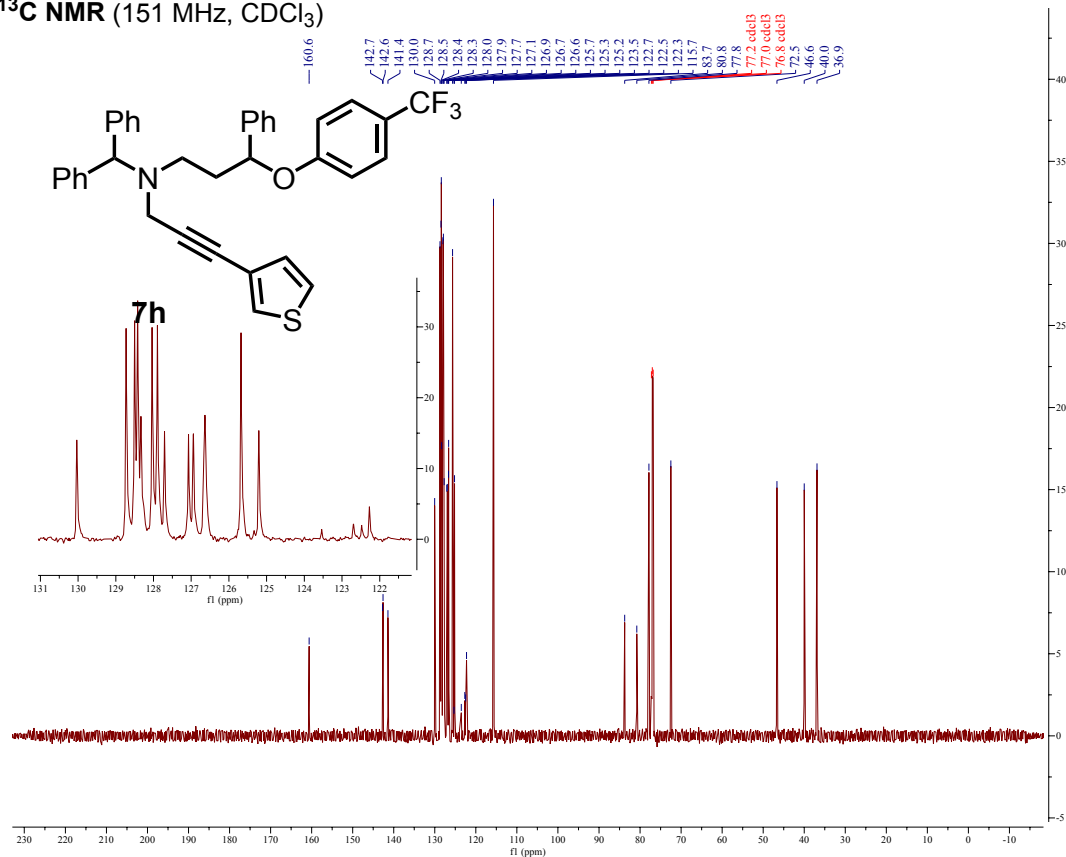
^1H NMR (600 MHz, CDCl_3)



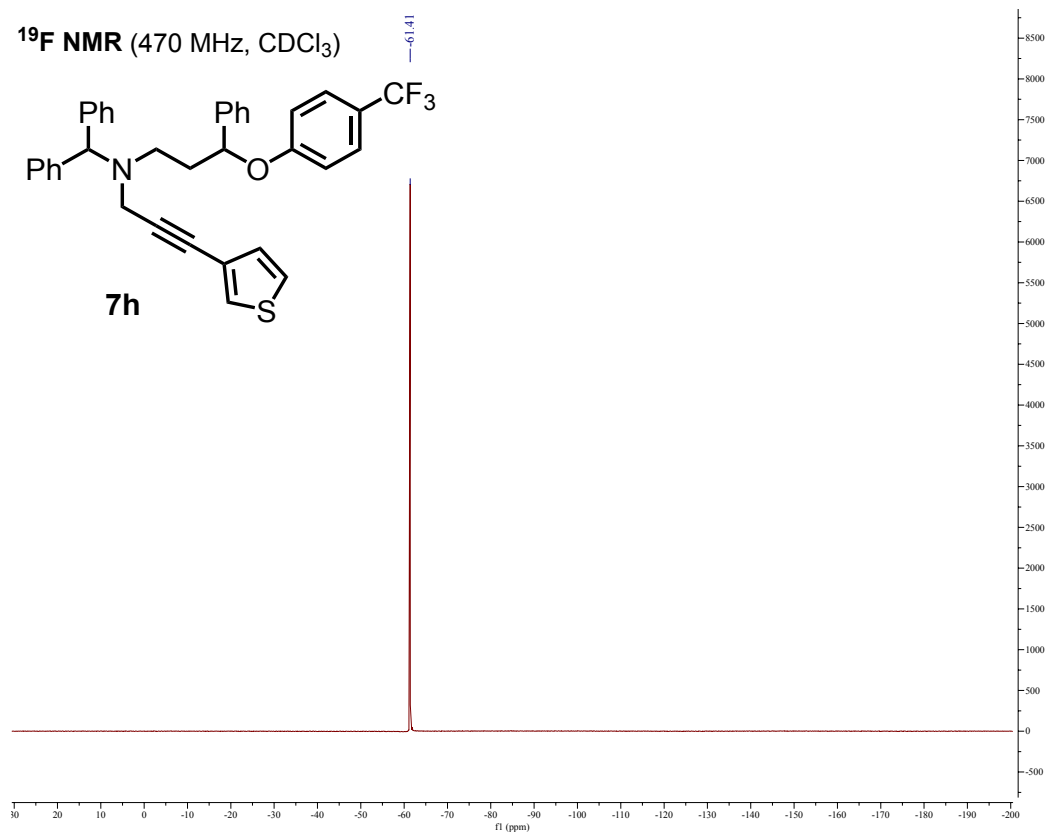
^1H NMR (600 MHz, CDCl_3)



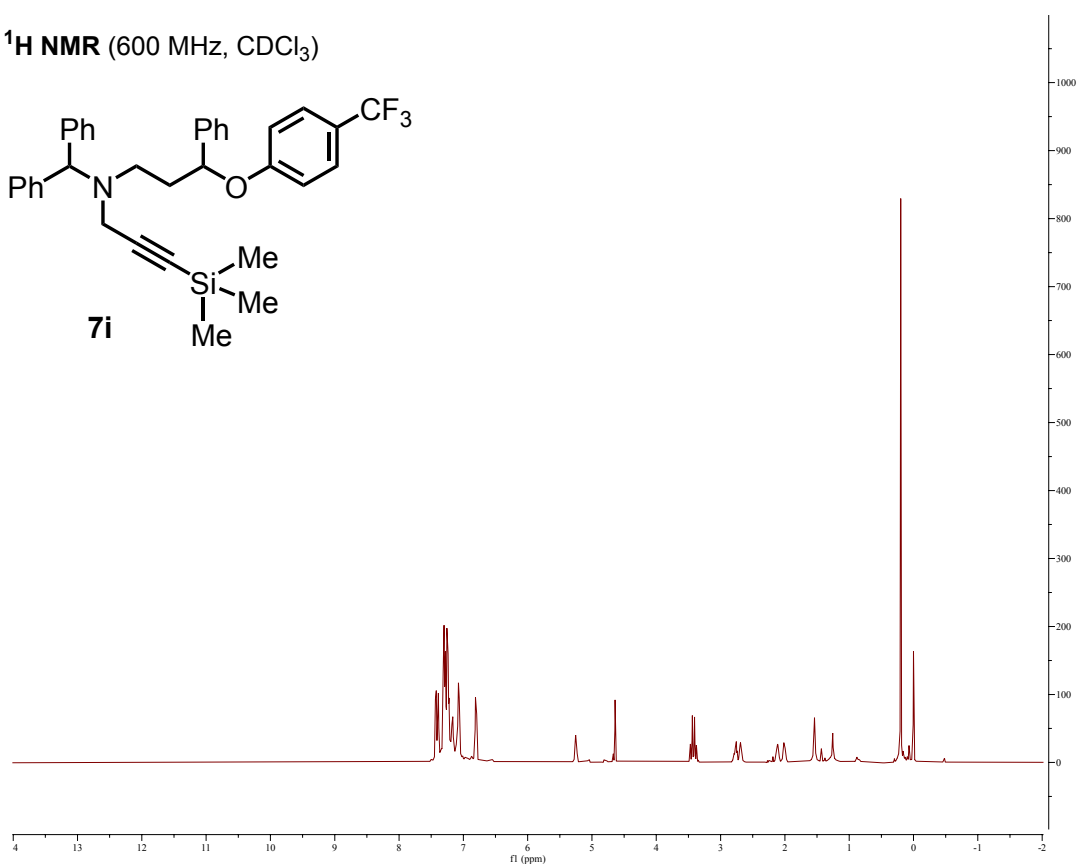
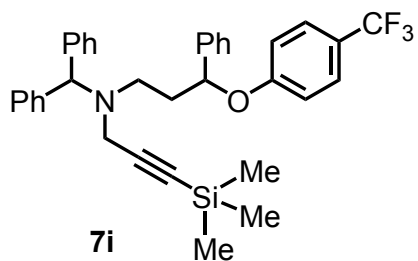
^{13}C NMR (151 MHz, CDCl_3)



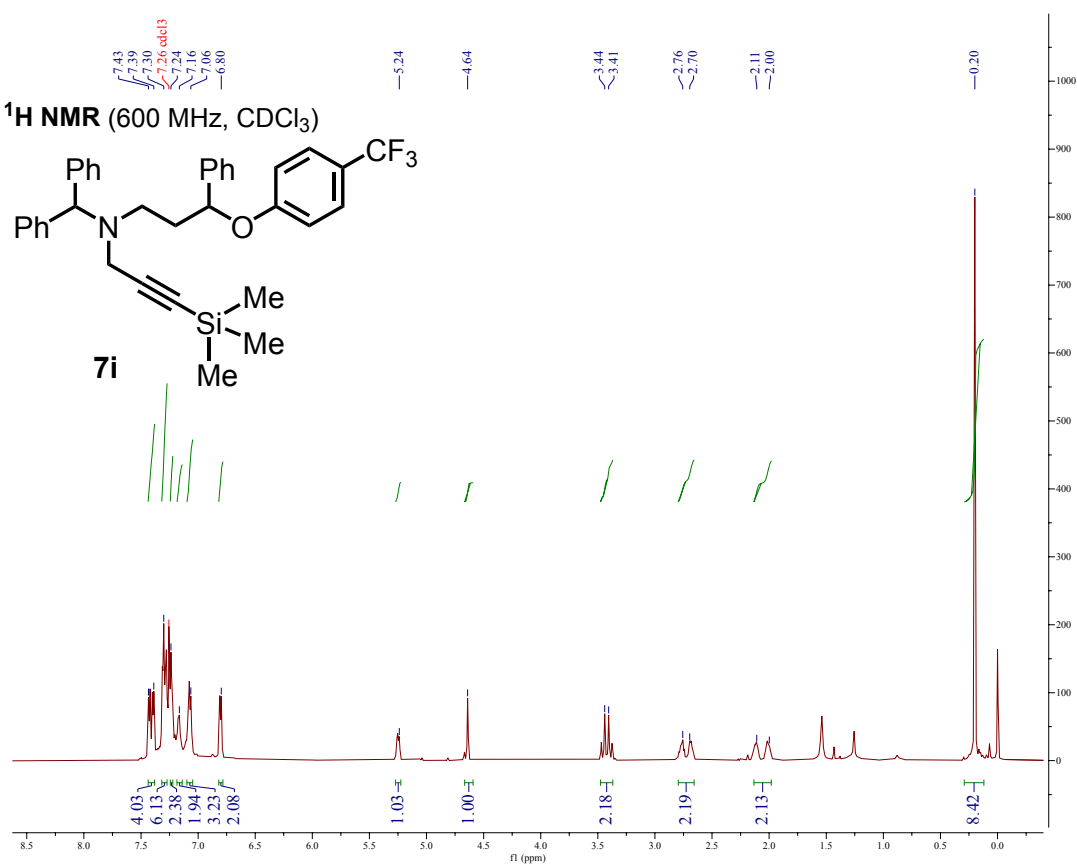
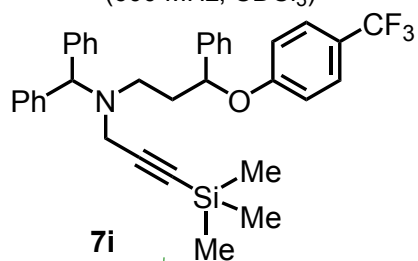
^{19}F NMR (470 MHz, CDCl_3)



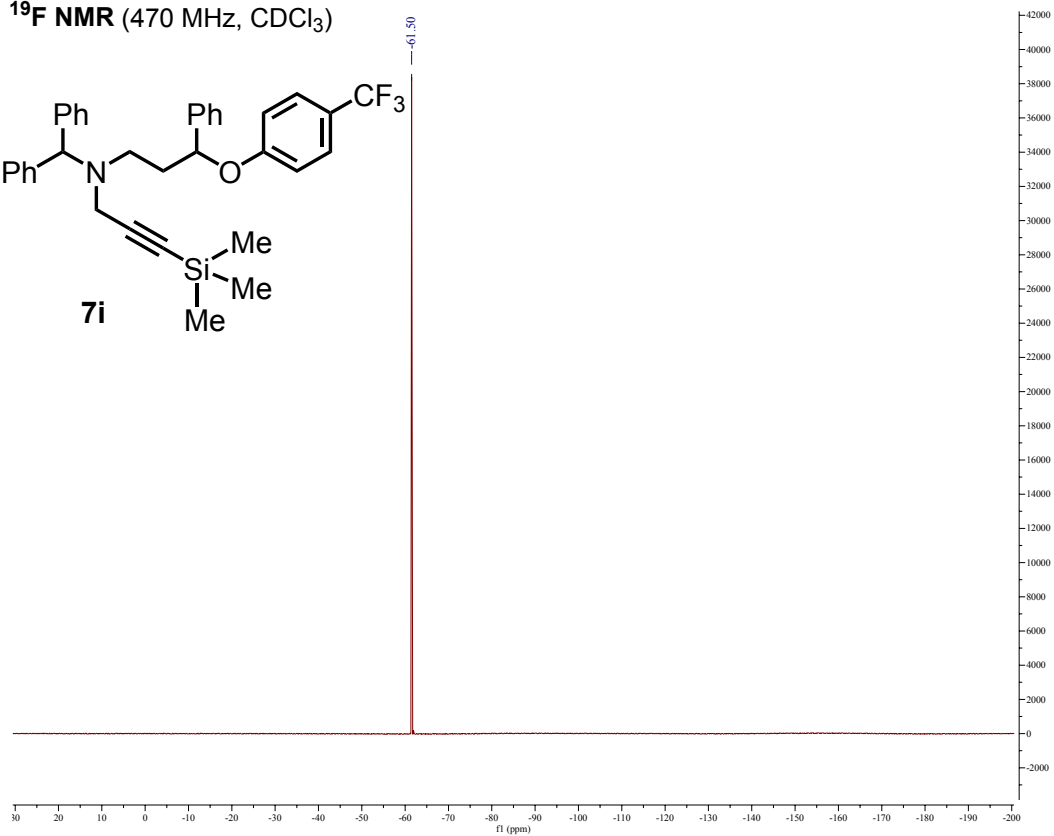
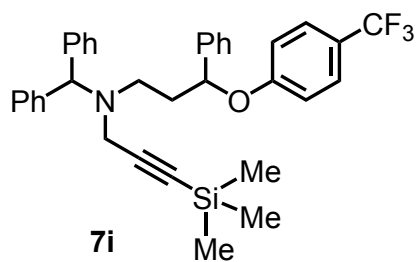
¹H NMR (600 MHz, CDCl₃)



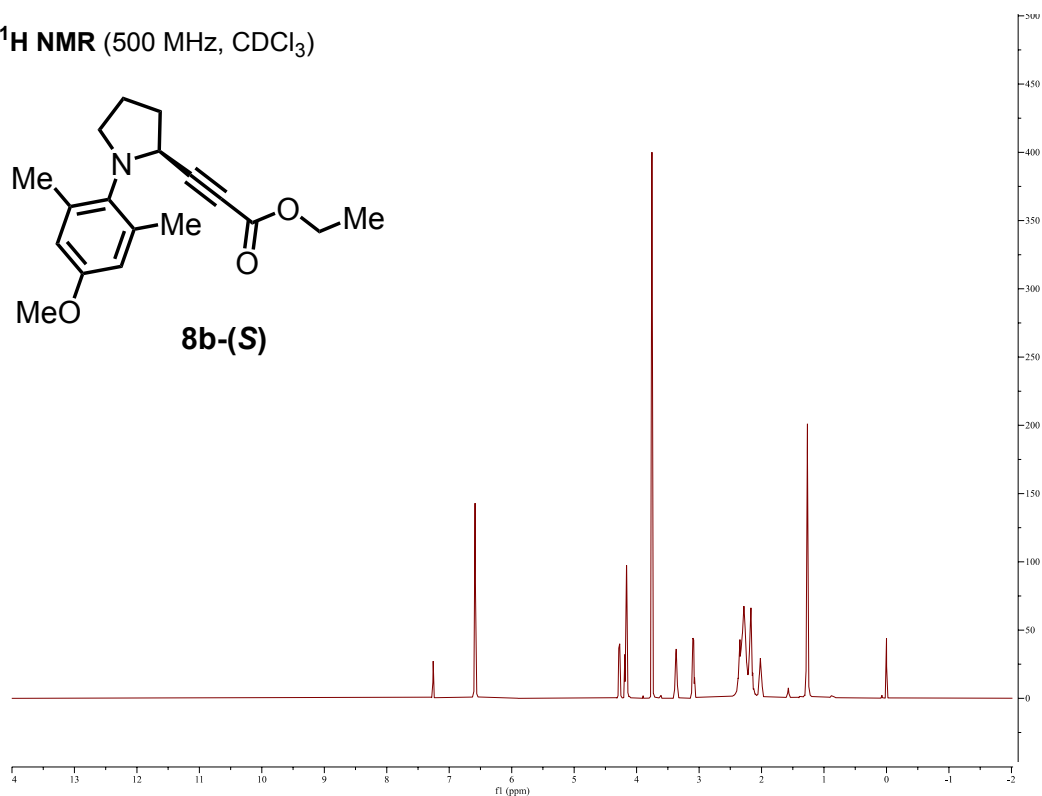
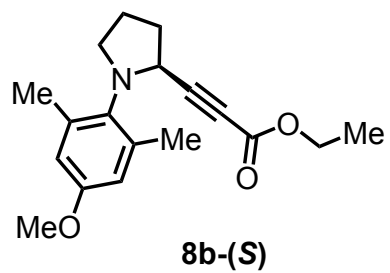
¹H NMR (600 MHz, CDCl₃)



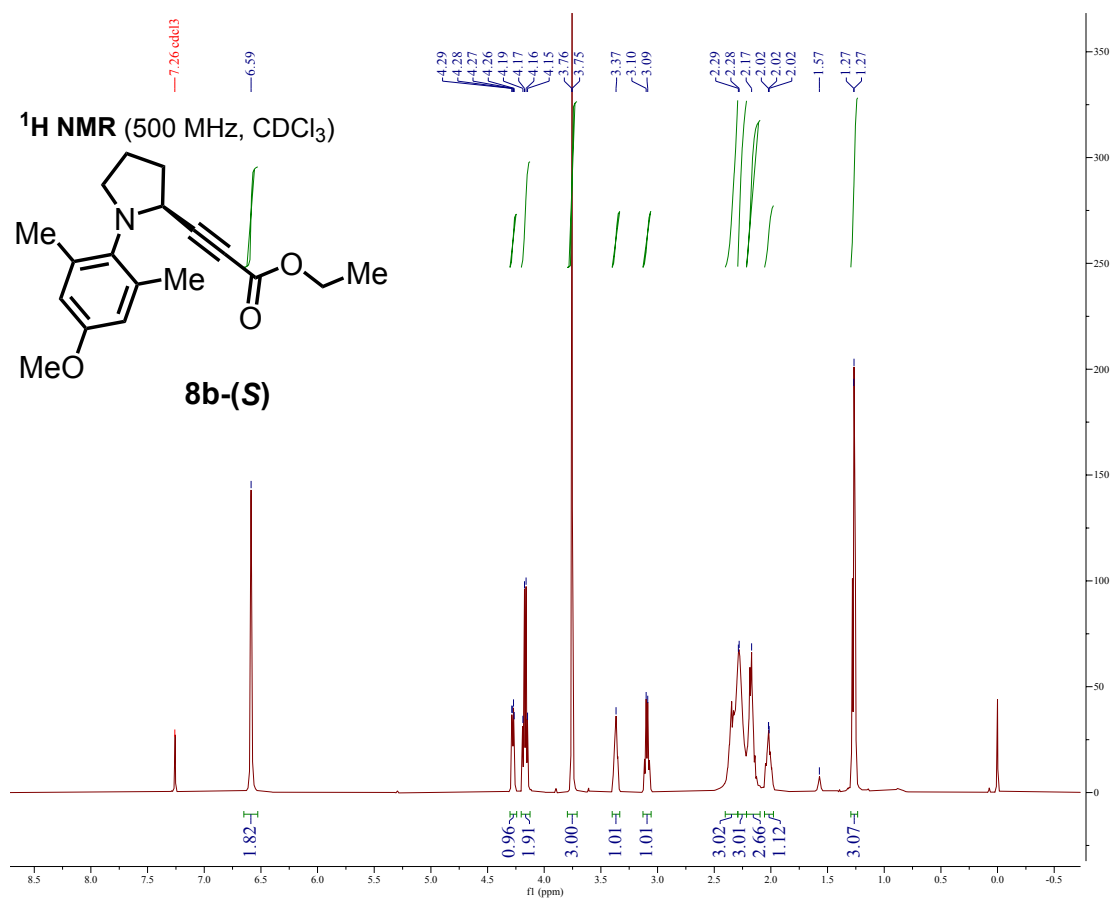
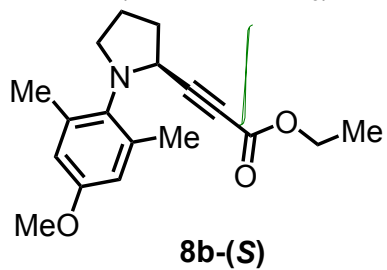
^{19}F NMR (470 MHz, CDCl_3)



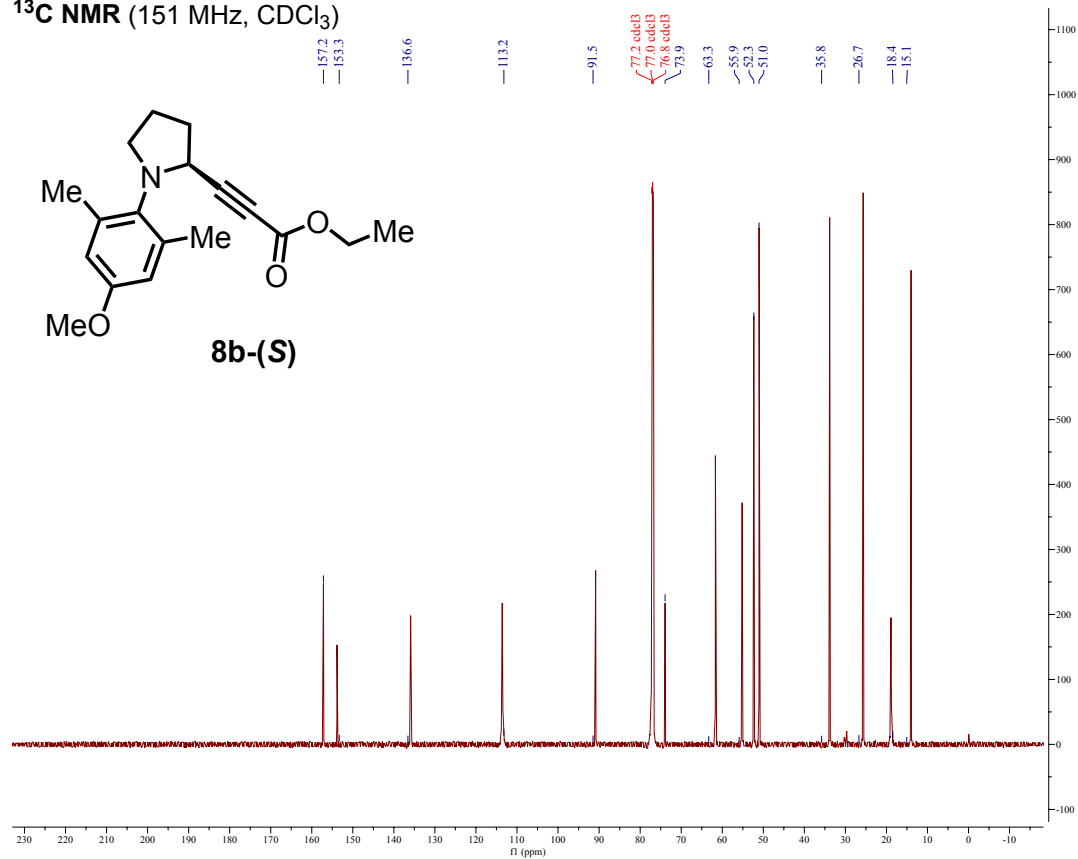
¹H NMR (500 MHz, CDCl₃)



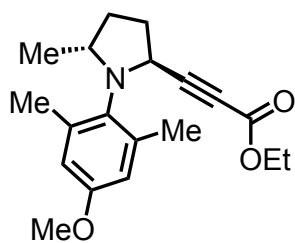
¹H NMR (500 MHz, CDCl₃)



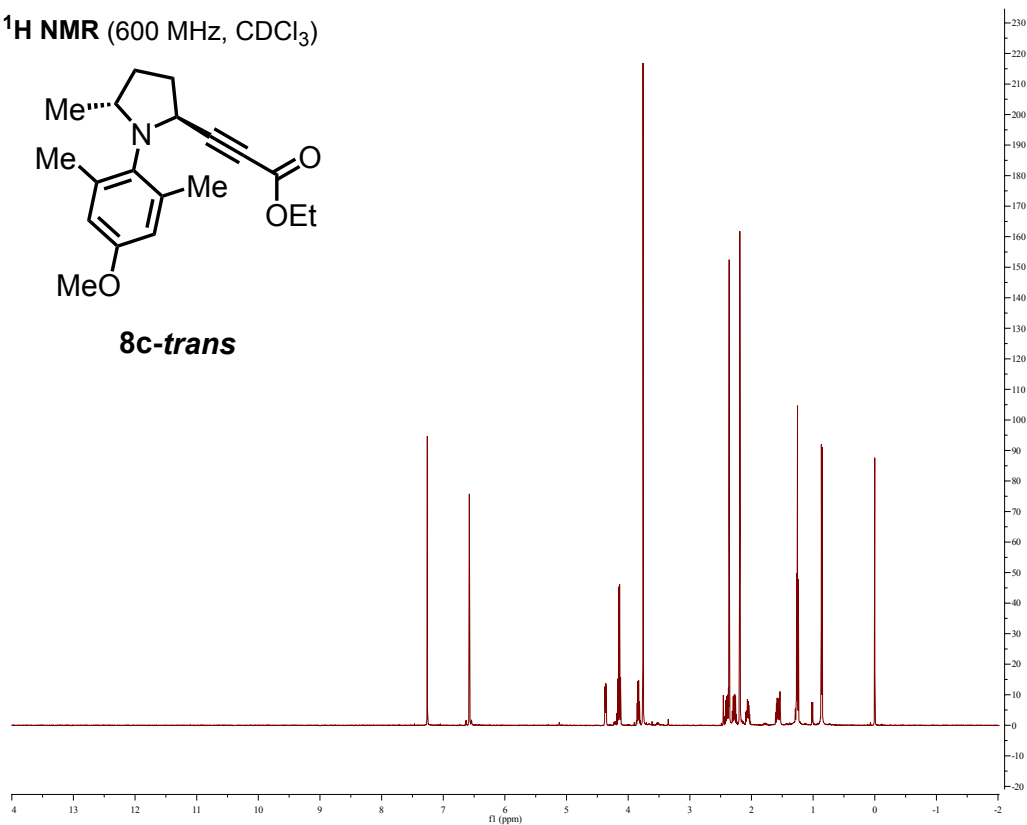
¹³C NMR (151 MHz, CDCl₃)



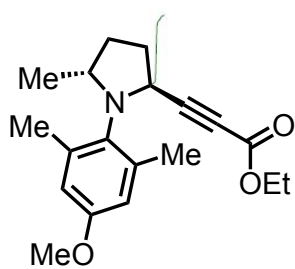
¹H NMR (600 MHz, CDCl₃)



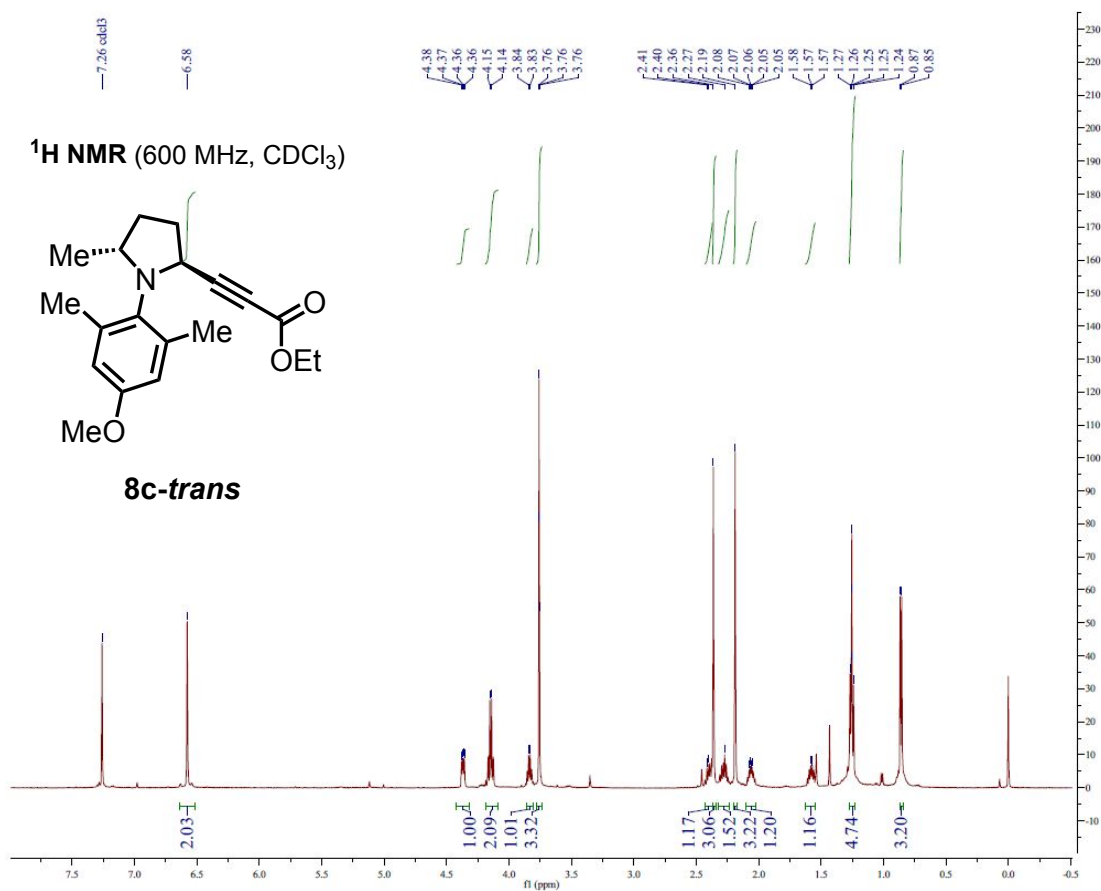
8c-trans



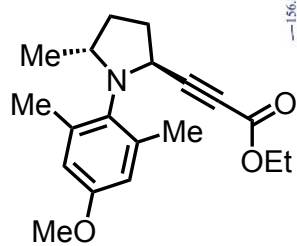
¹H NMR (600 MHz, CDCl₃)



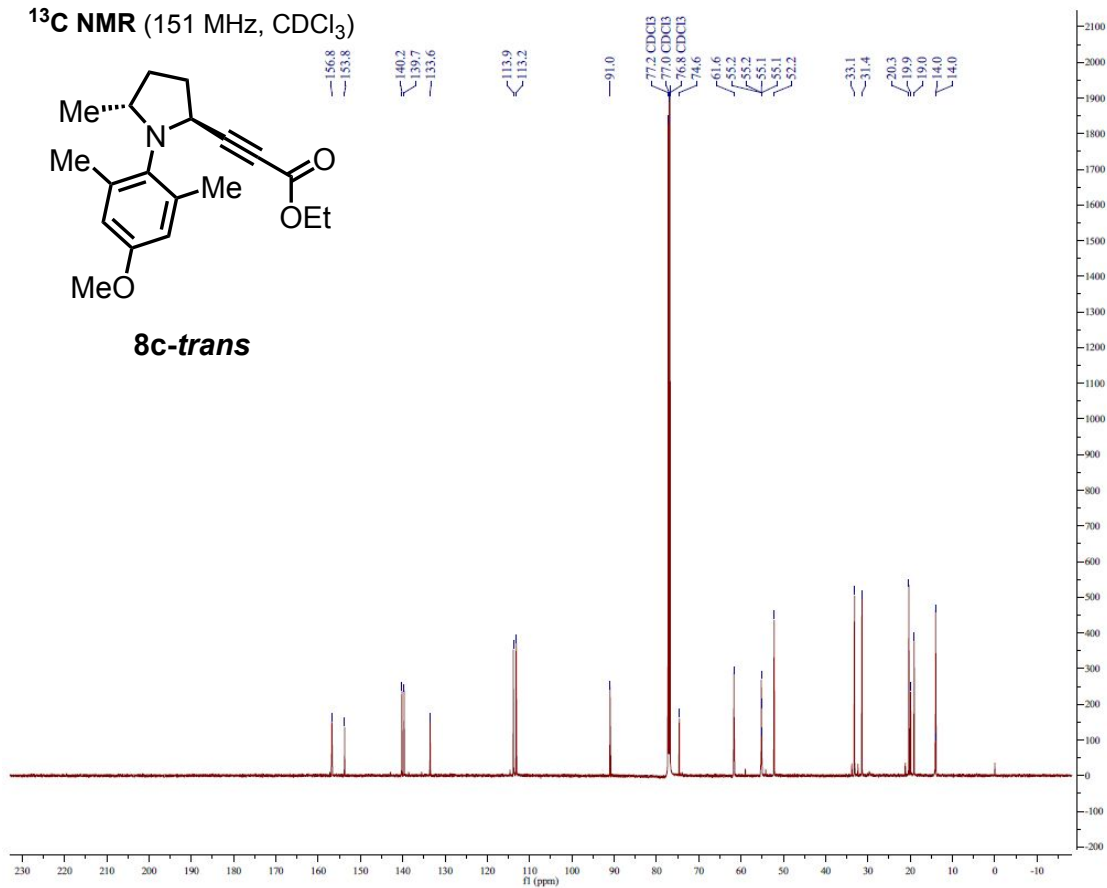
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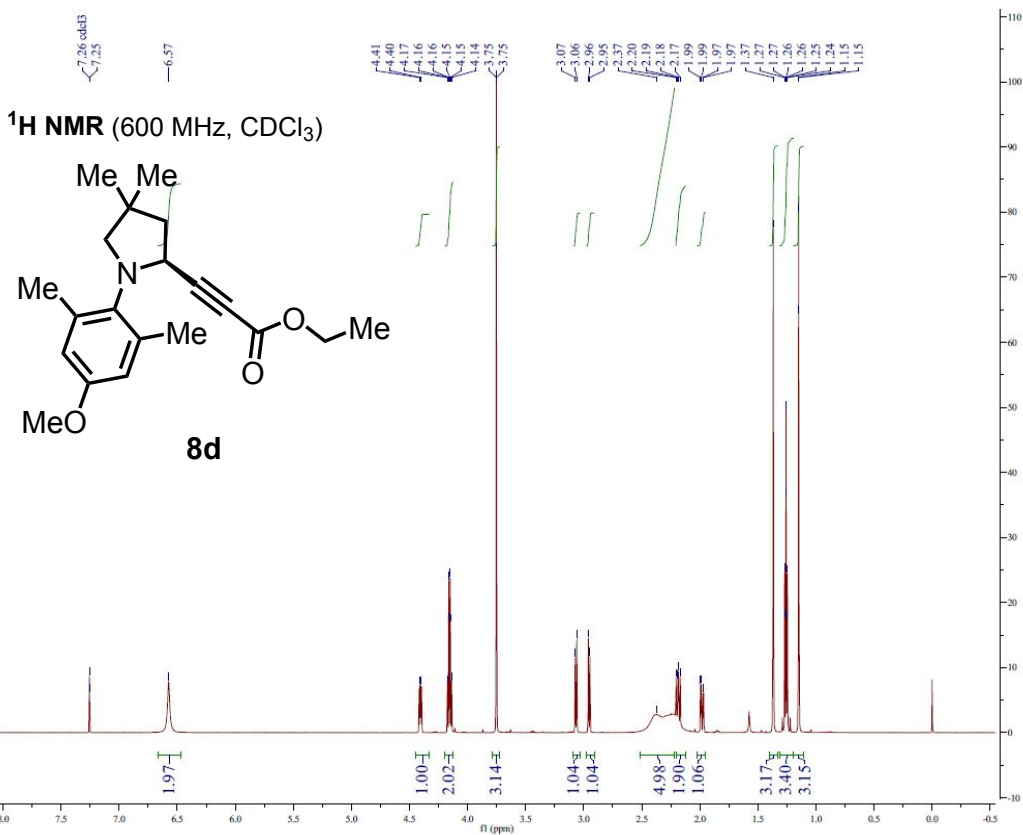
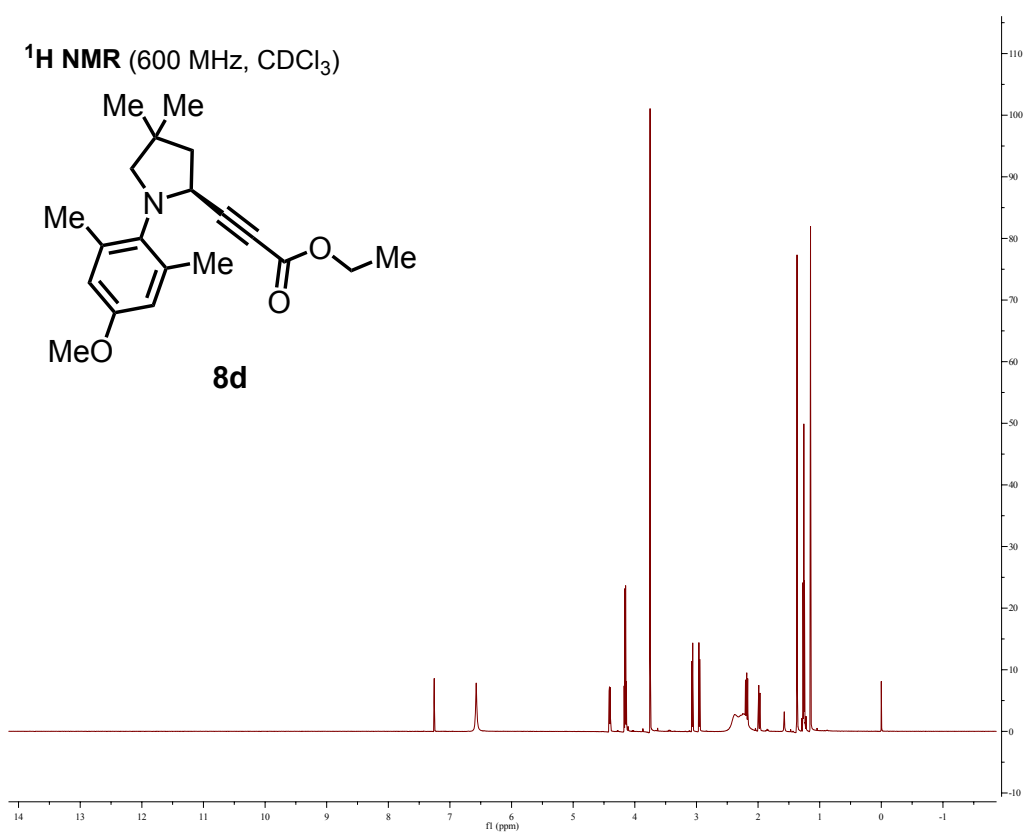
¹³C NMR (151 MHz, CDCl₃)

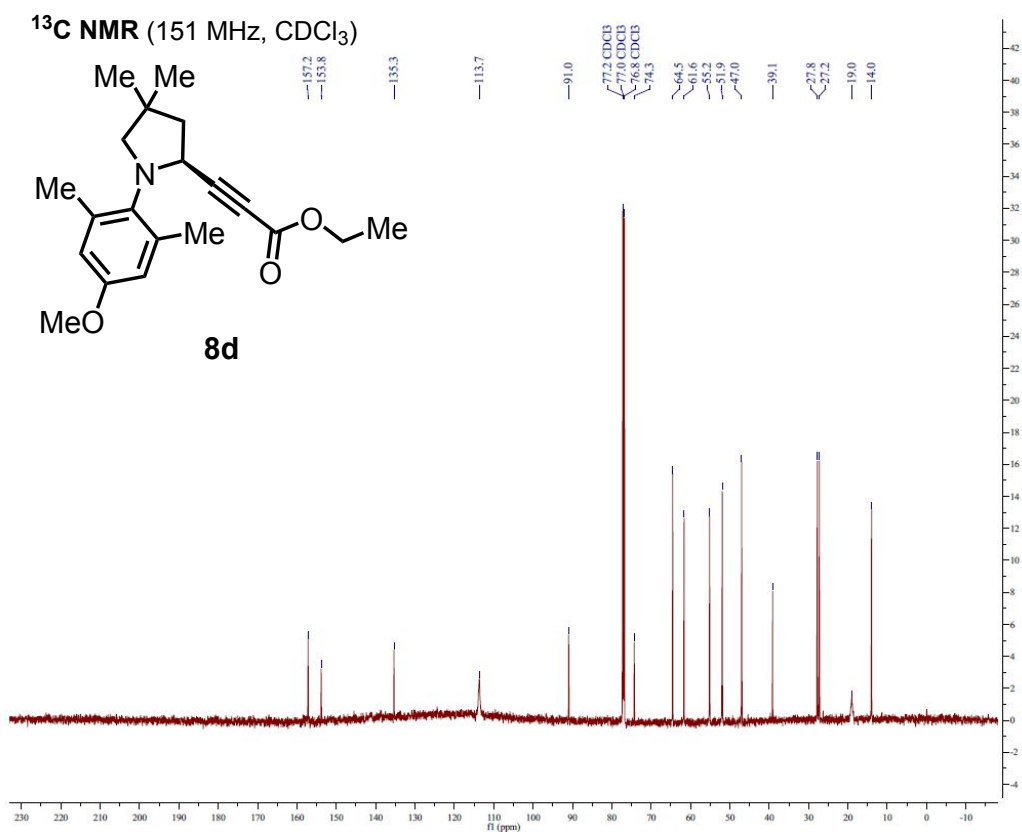


8c-trans

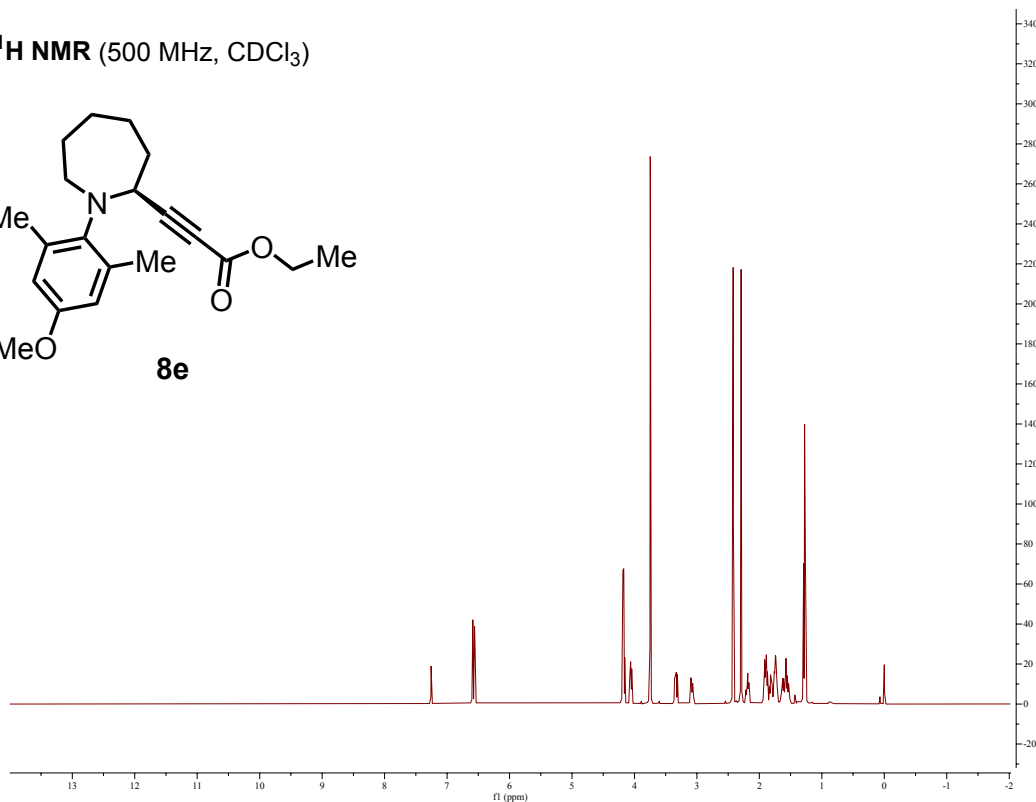
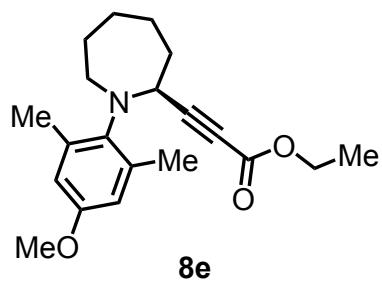


¹H NMR (600 MHz, CDCl₃)

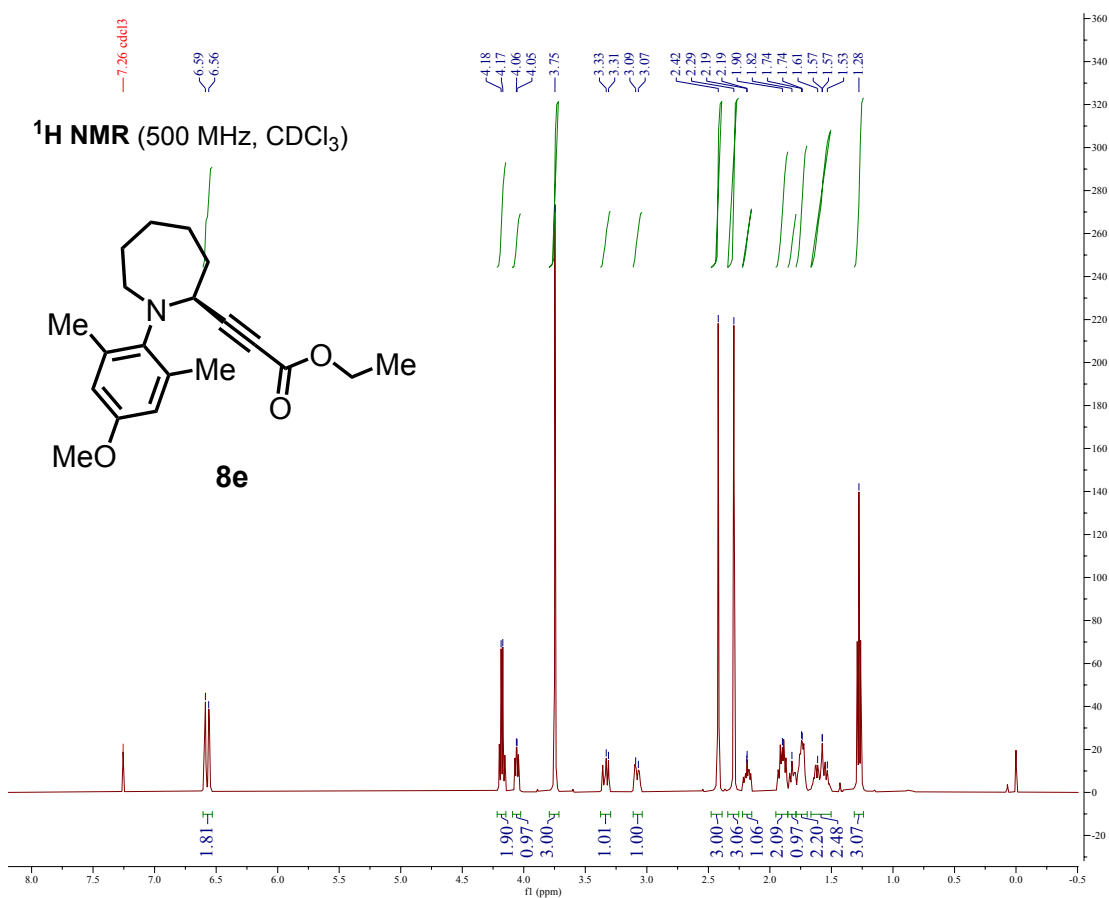
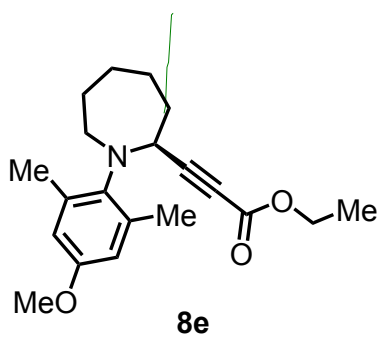


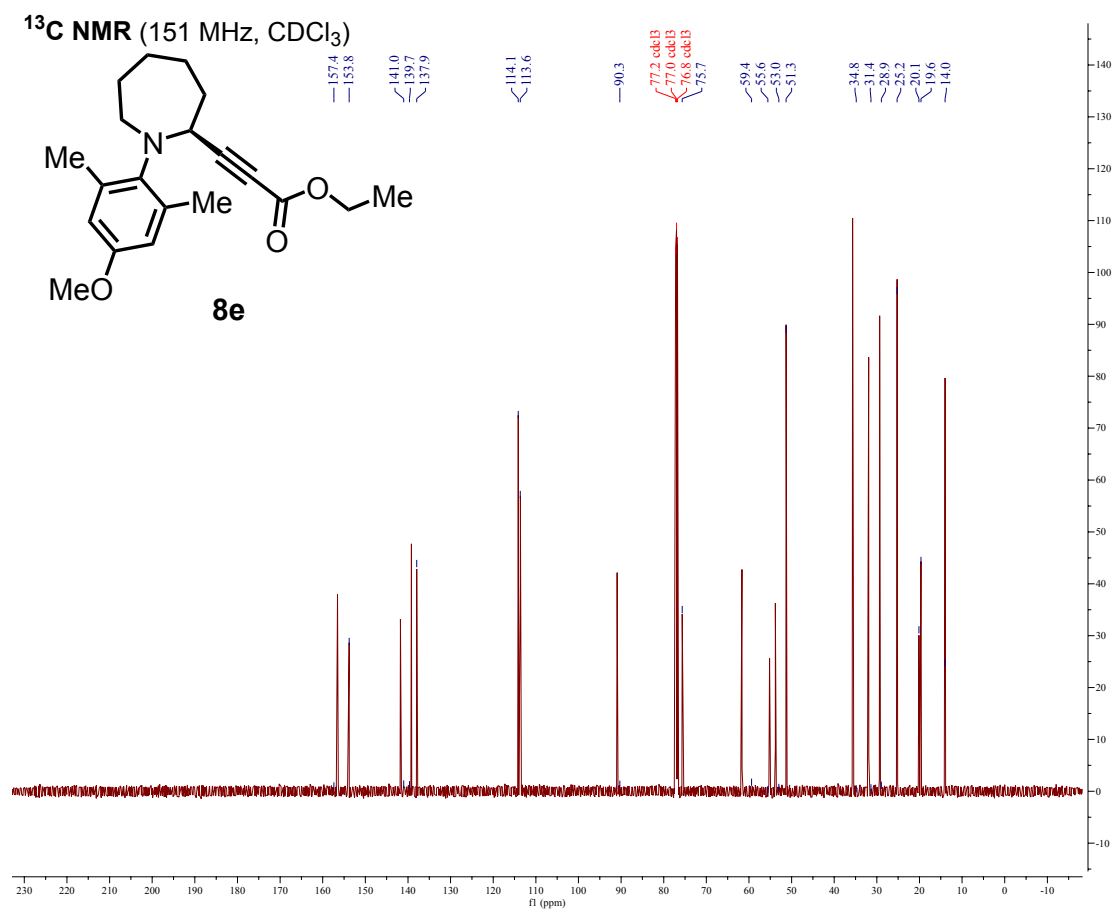


¹H NMR (500 MHz, CDCl₃)

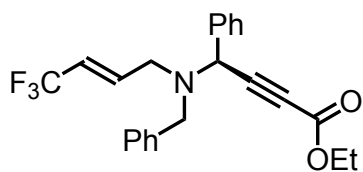


¹H NMR (500 MHz, CDCl₃)

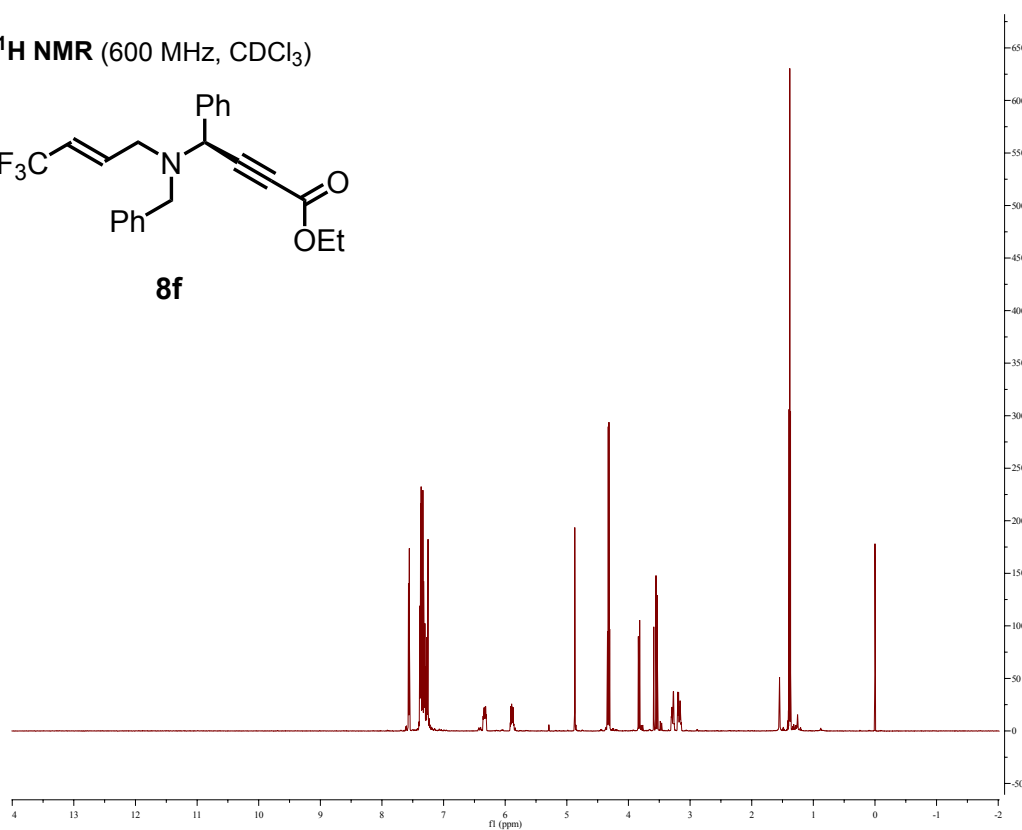




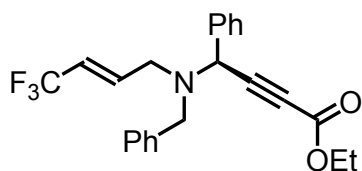
¹H NMR (600 MHz, CDCl₃)



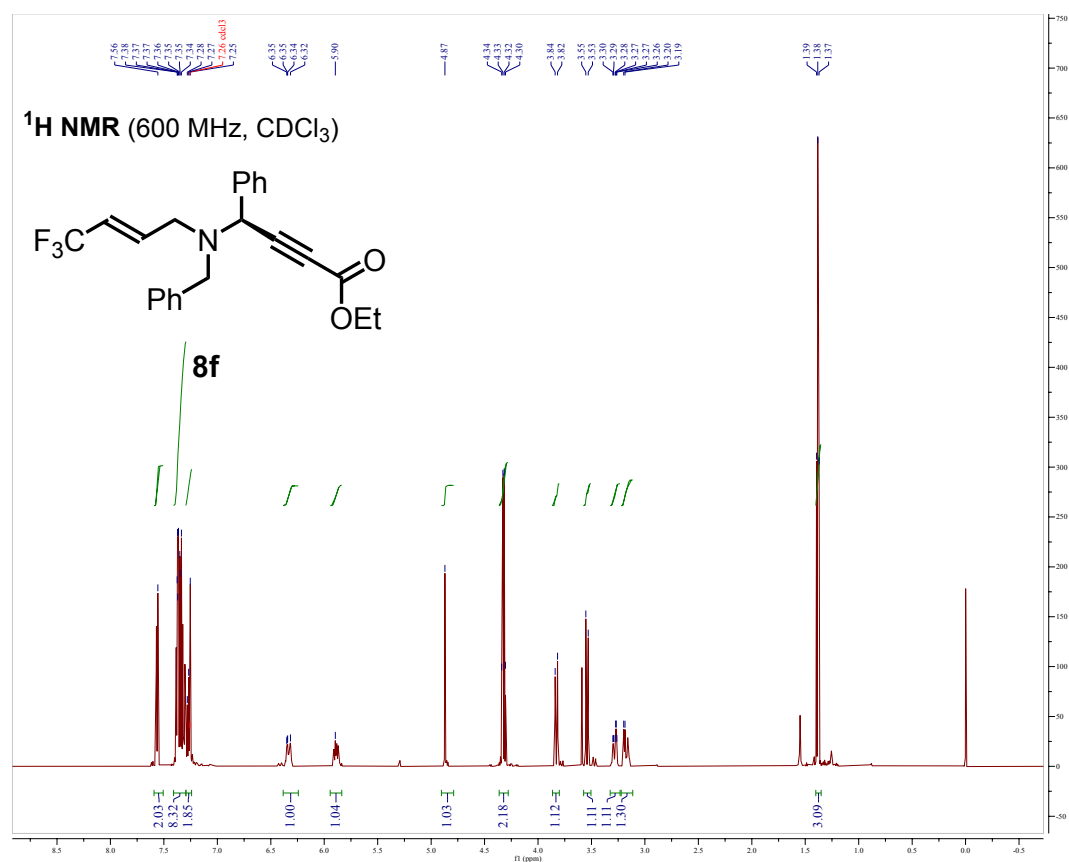
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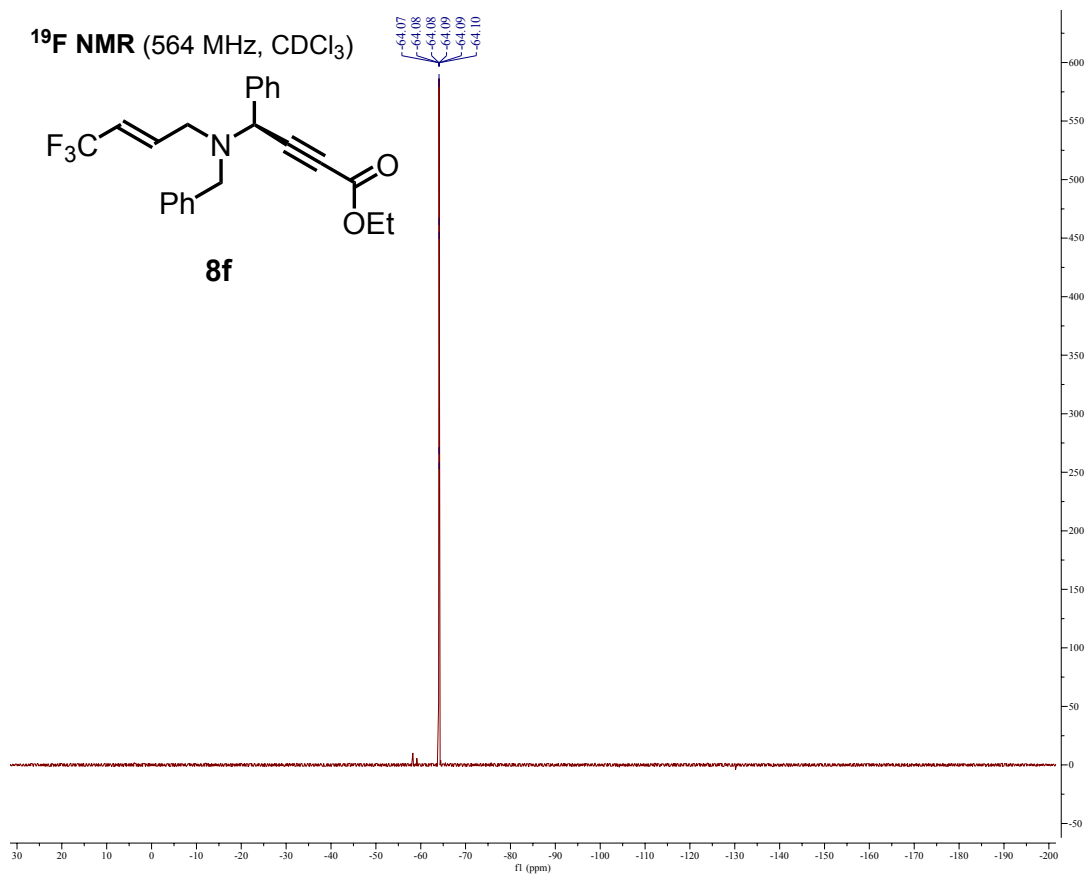
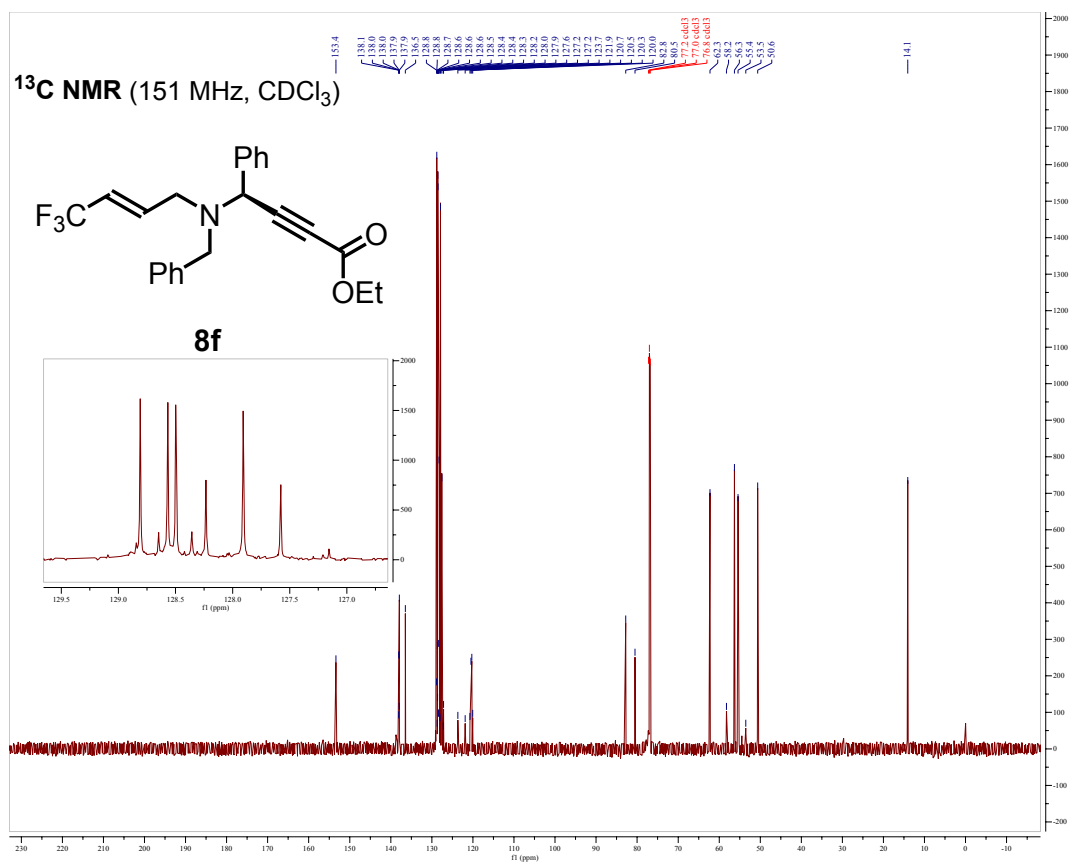


¹H NMR (600 MHz, CDCl₃)

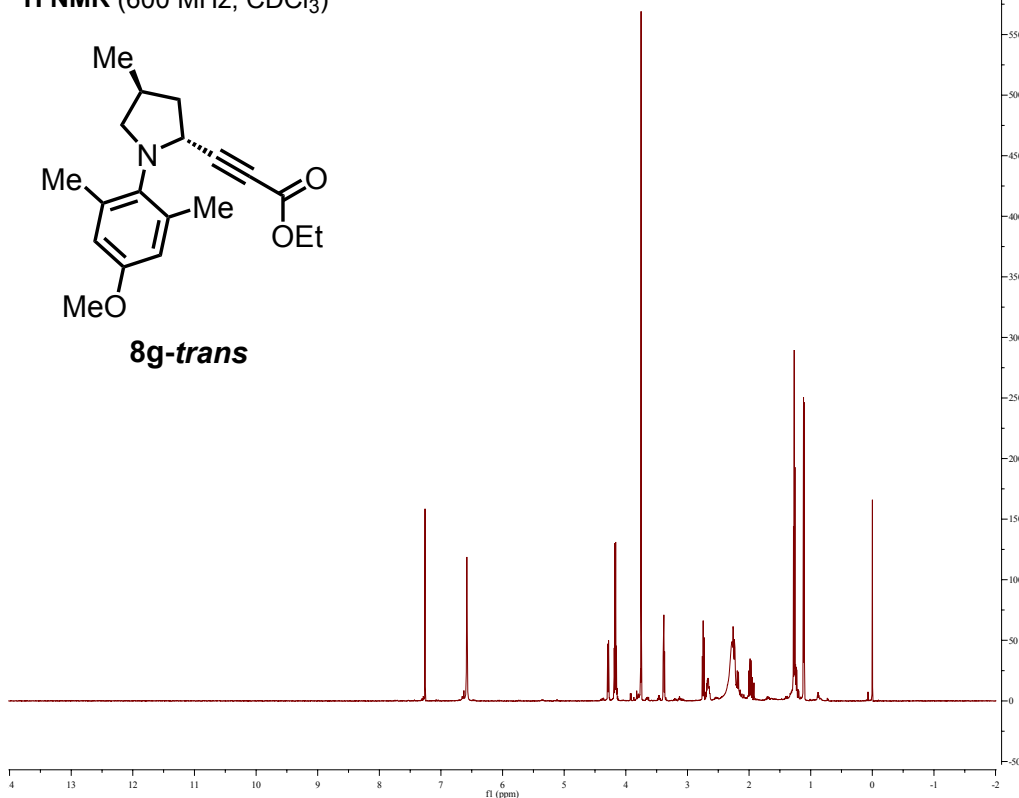


8f

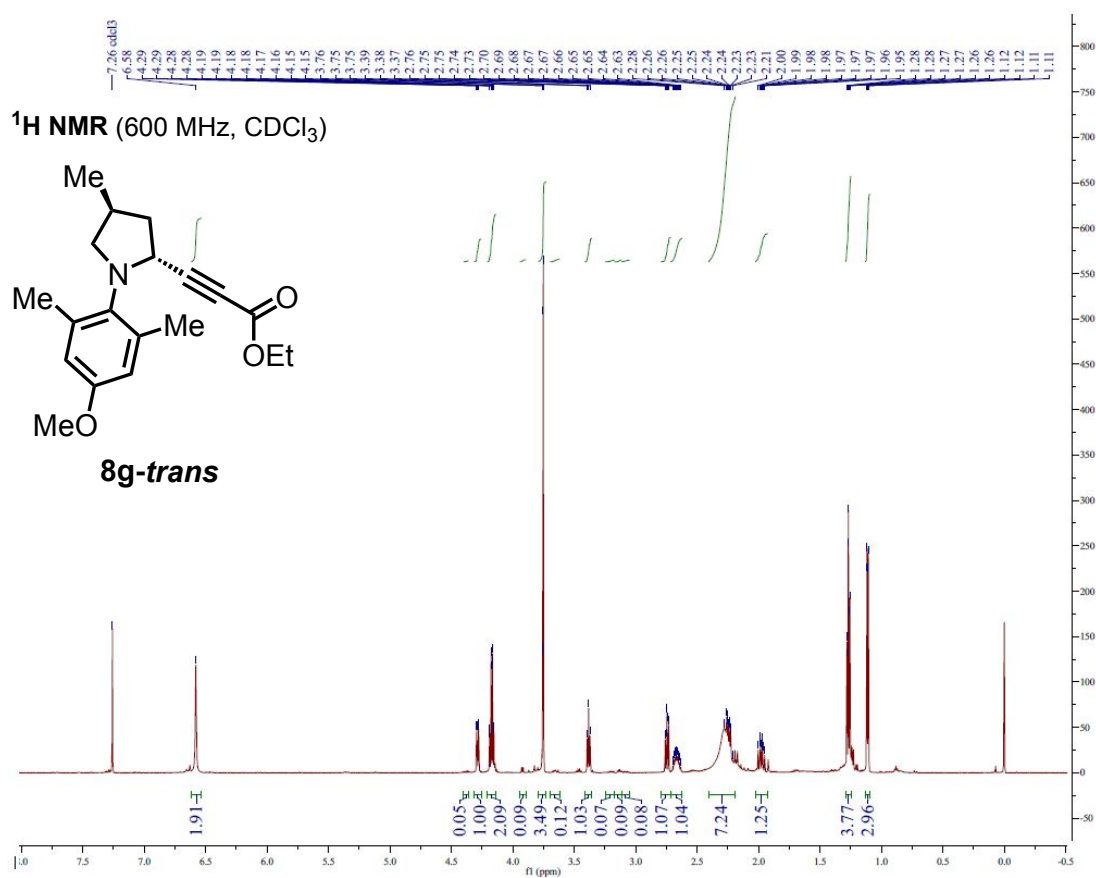


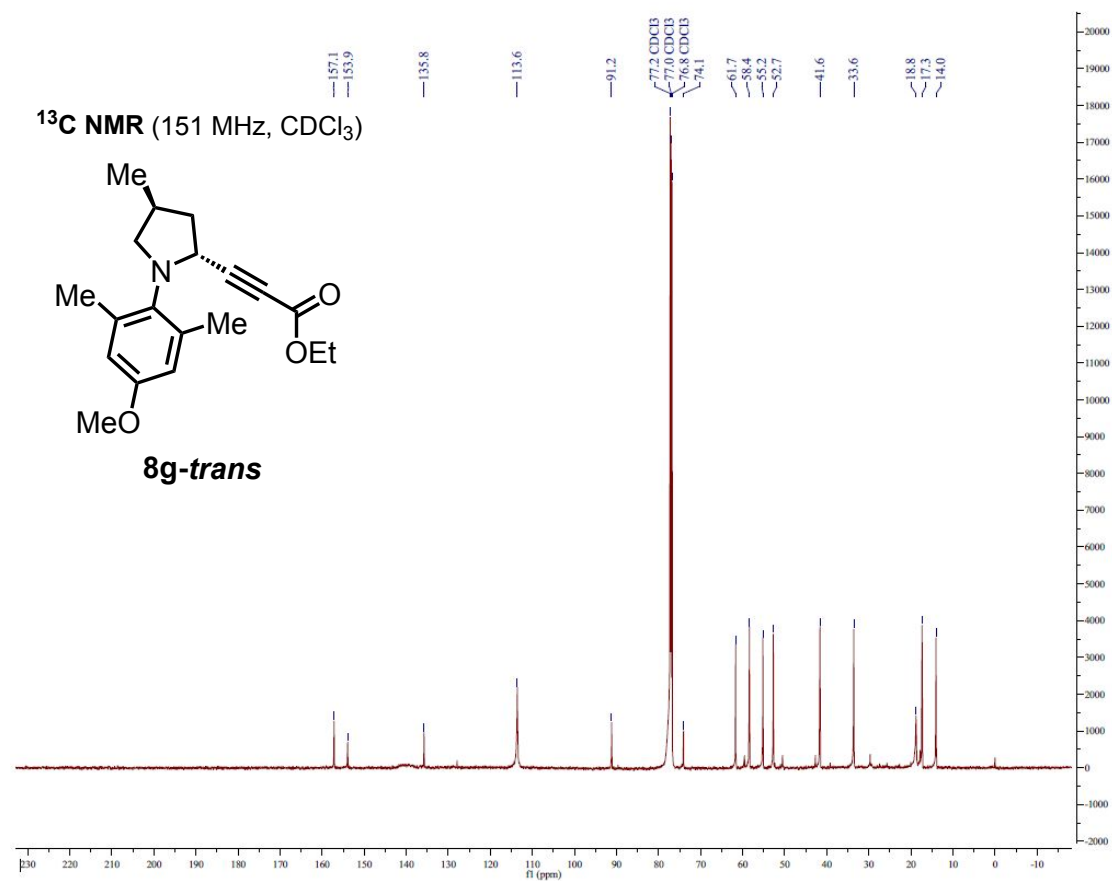


¹H NMR (600 MHz, CDCl₃)

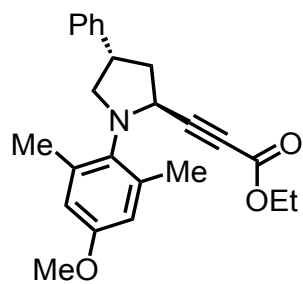


¹H NMR (600 MHz, CDCl₃)

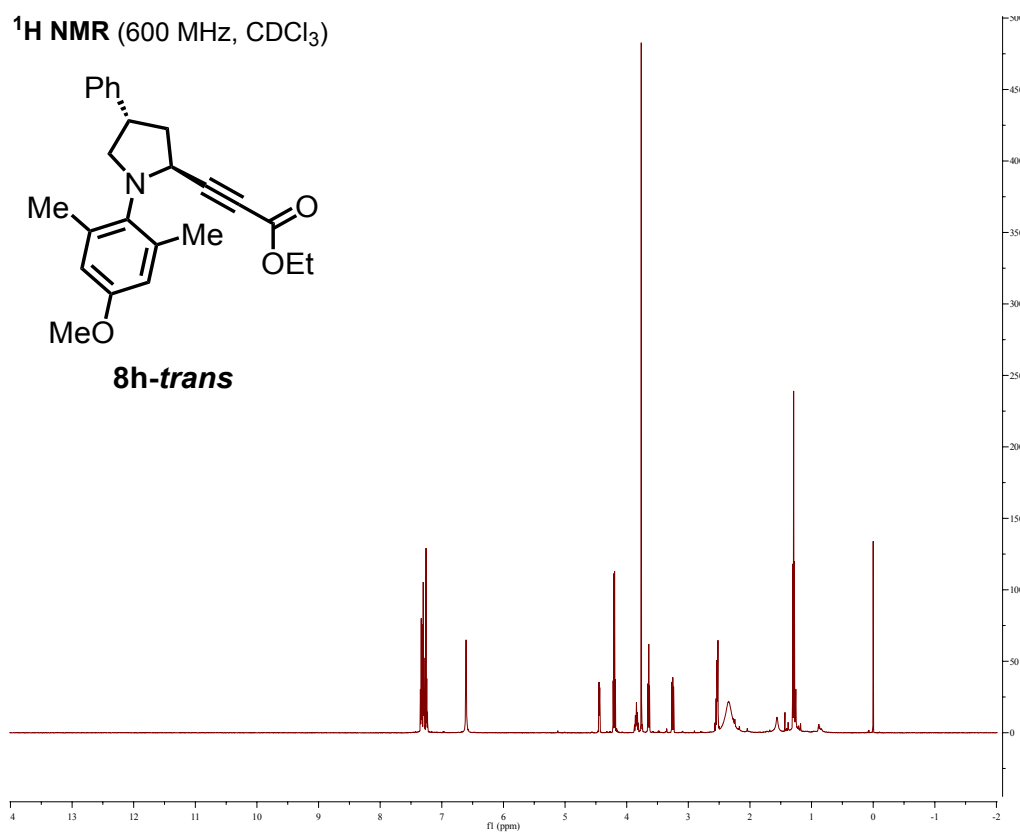




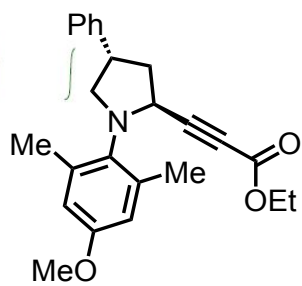
¹H NMR (600 MHz, CDCl₃)



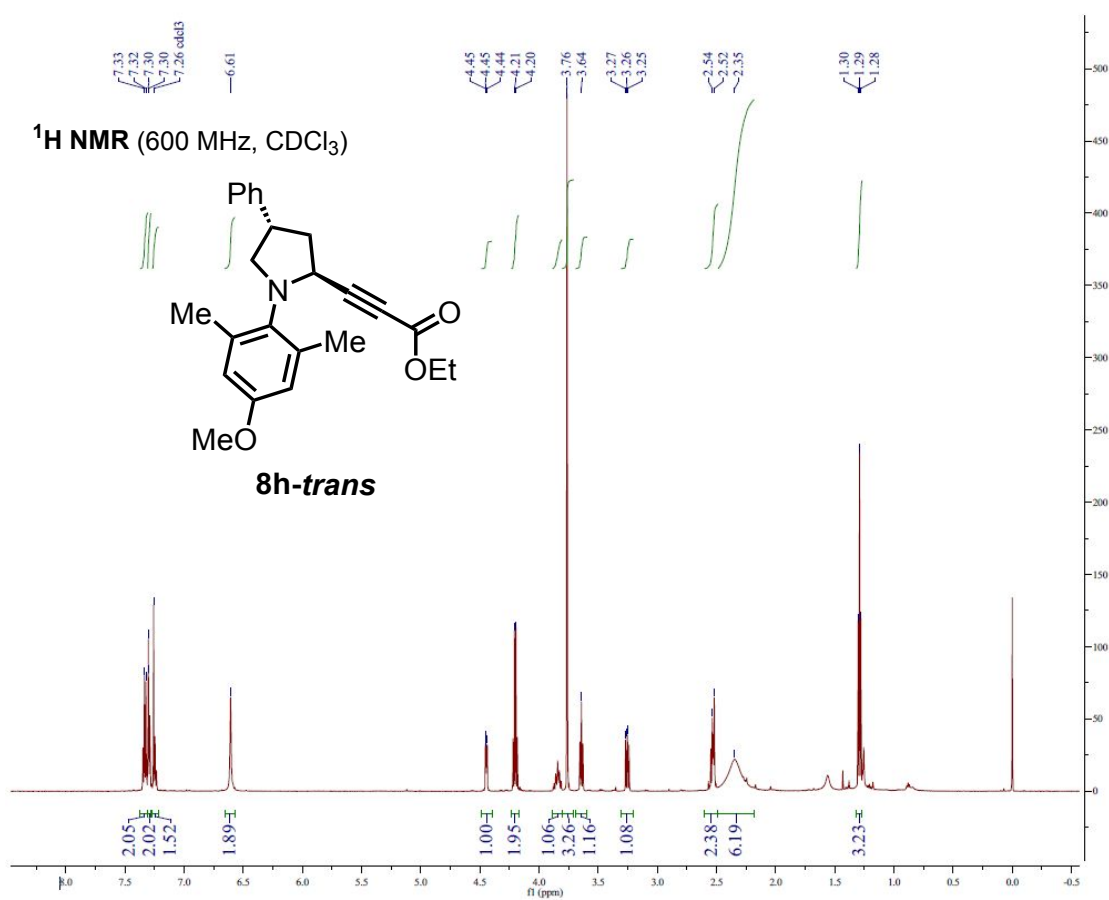
8h-trans

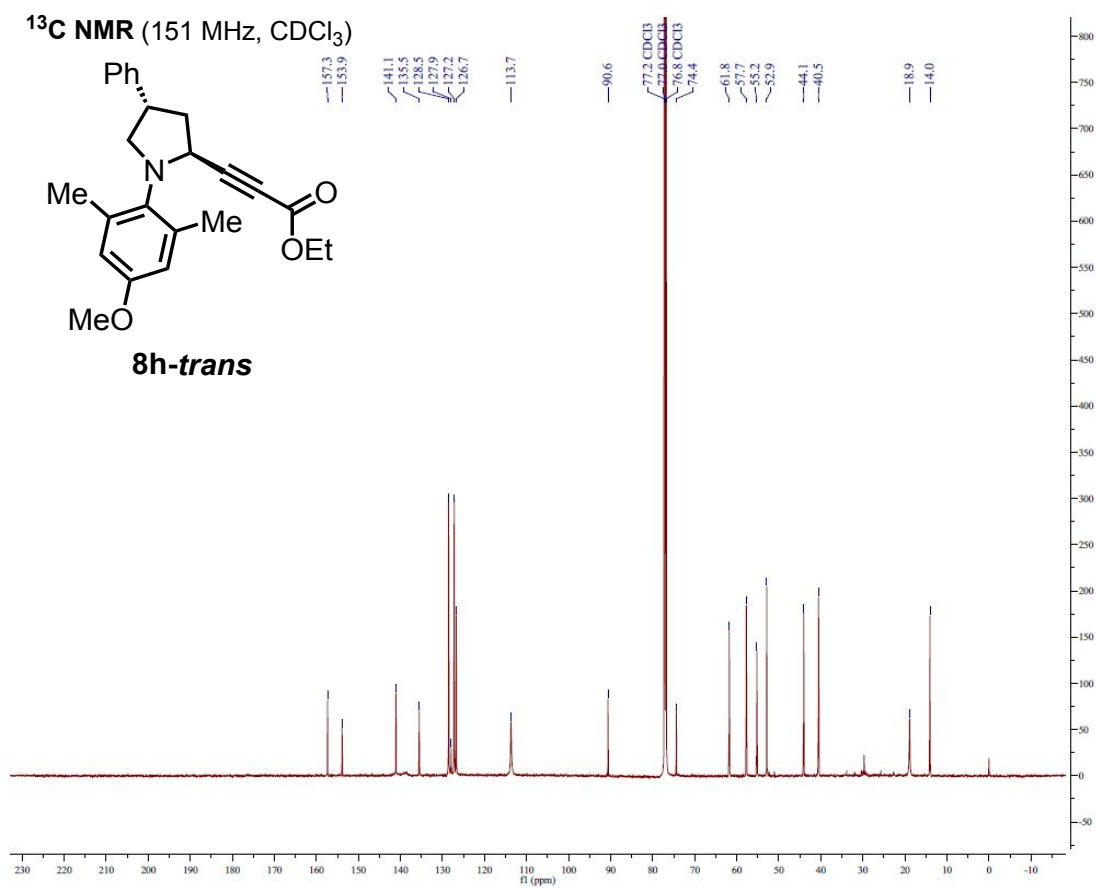


¹H NMR (600 MHz, CDCl₃)

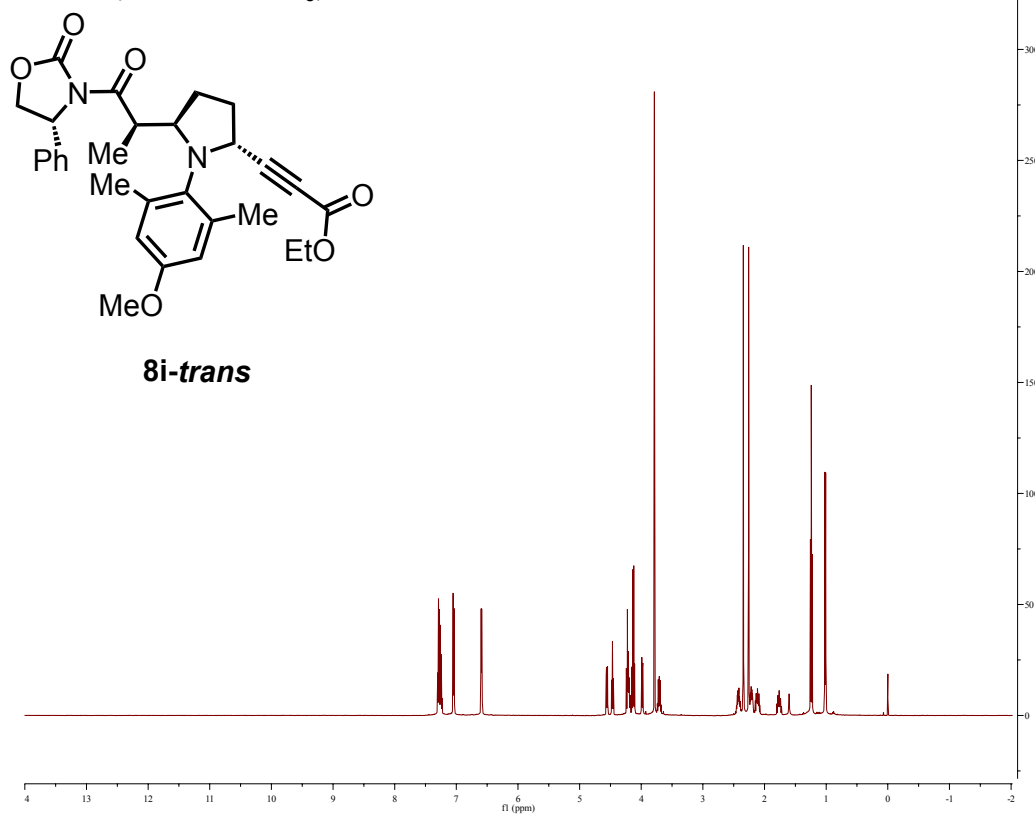


8h-trans

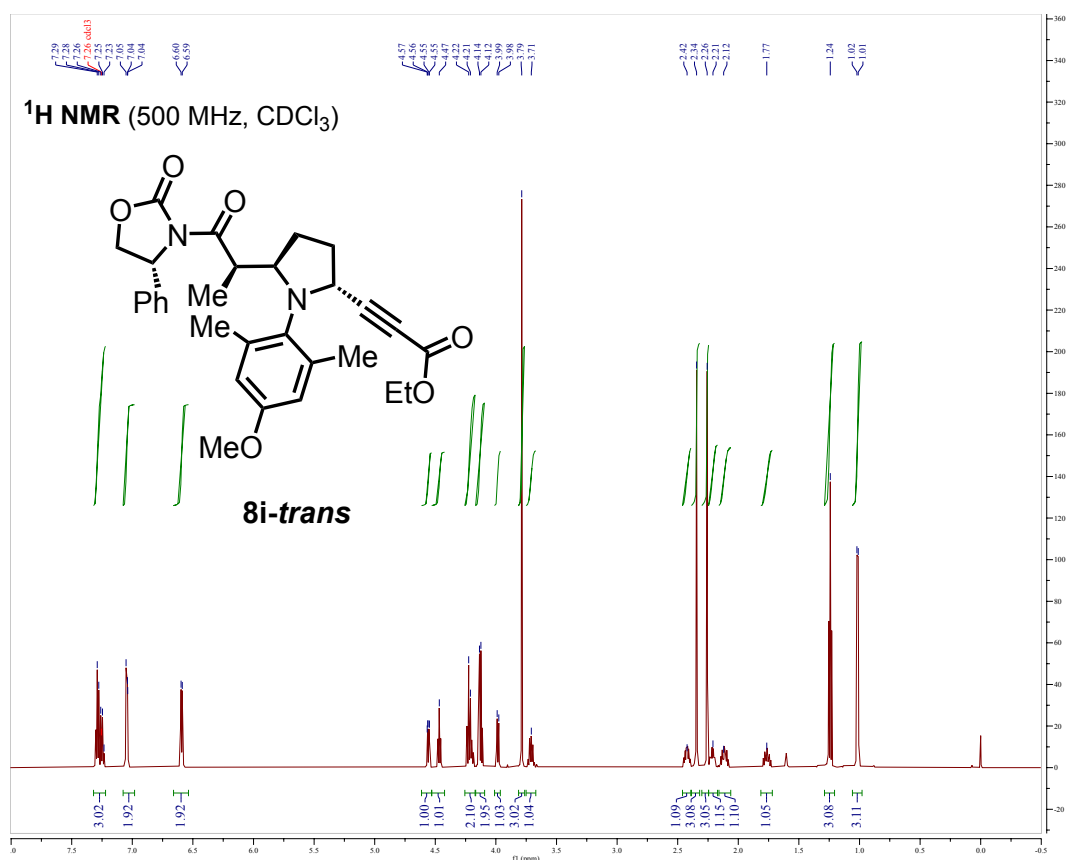


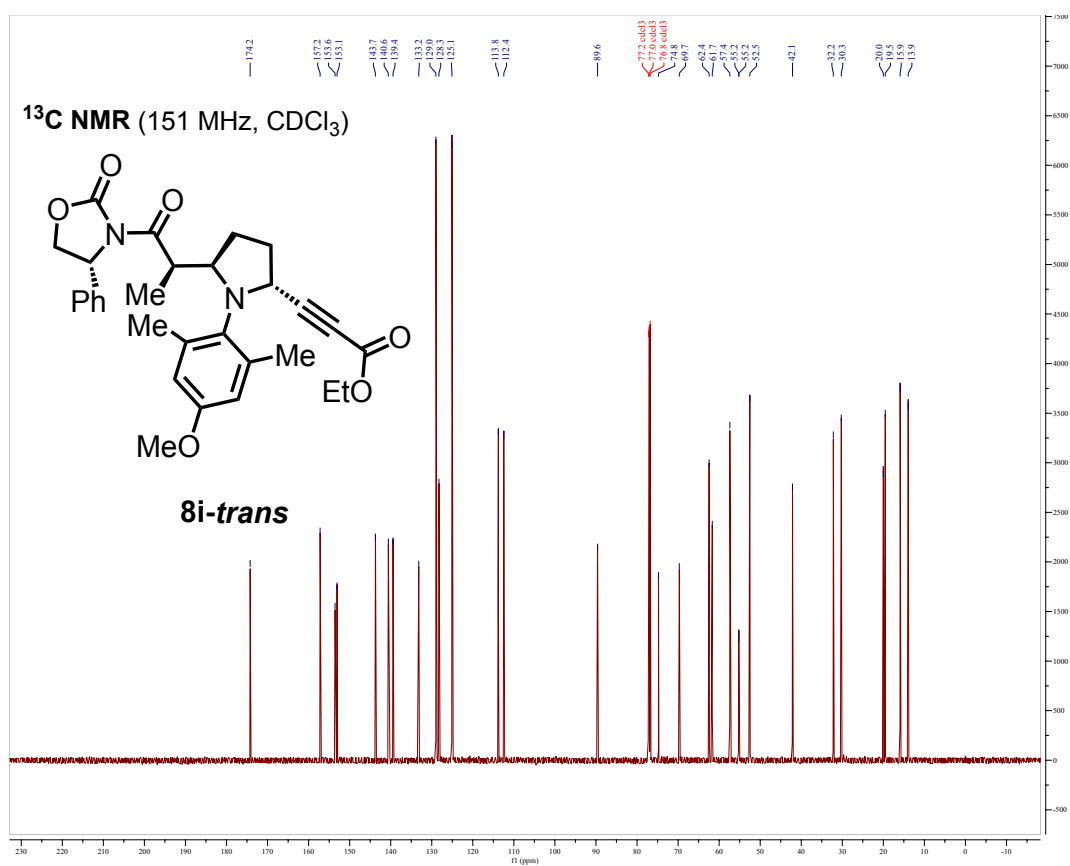


¹H NMR (500 MHz, CDCl₃)

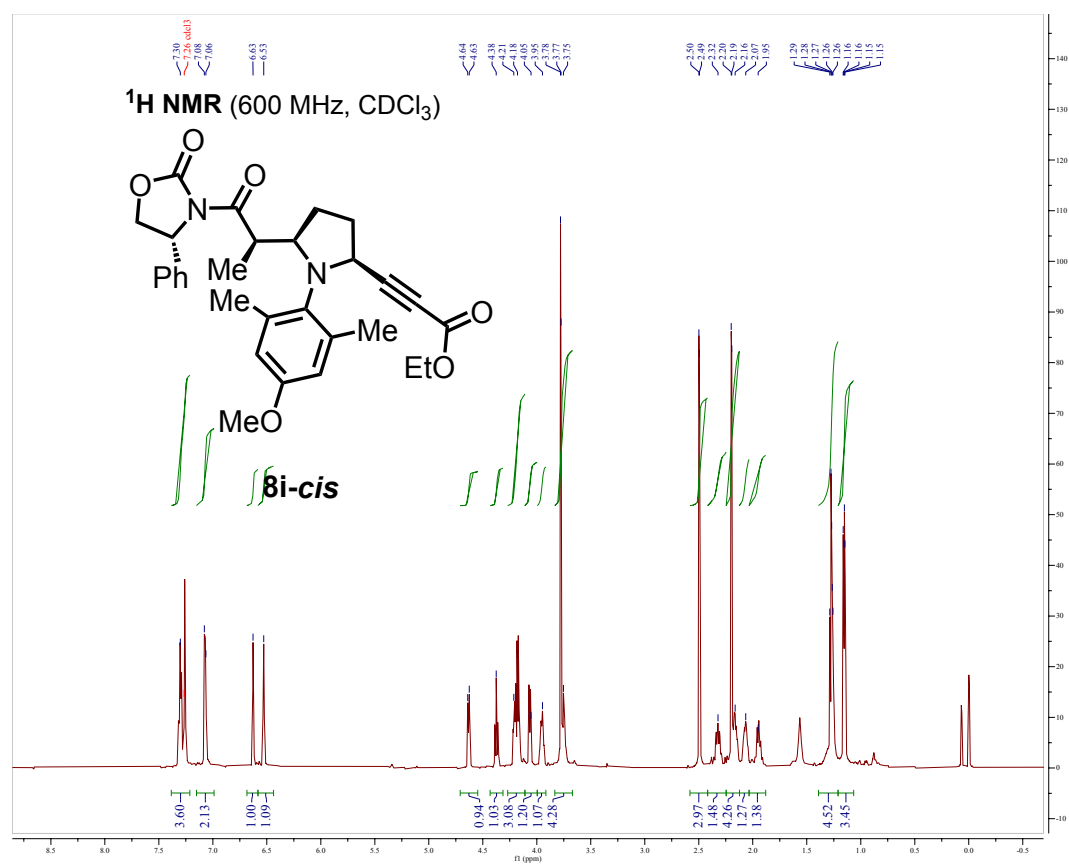
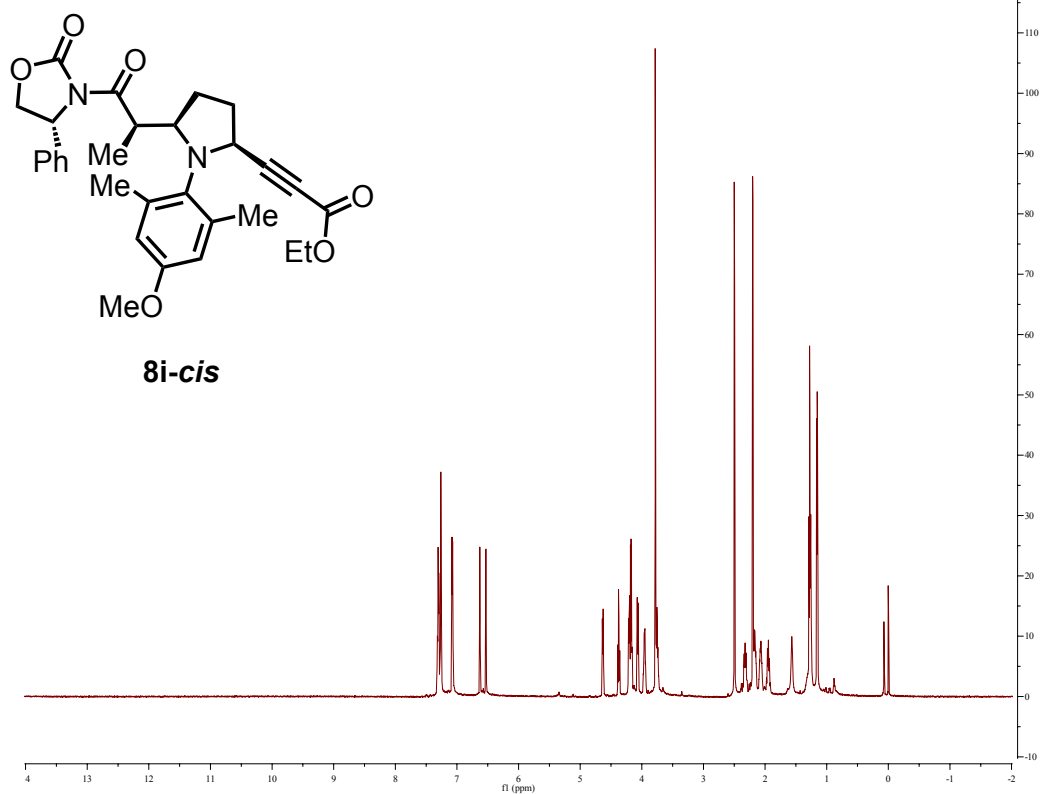


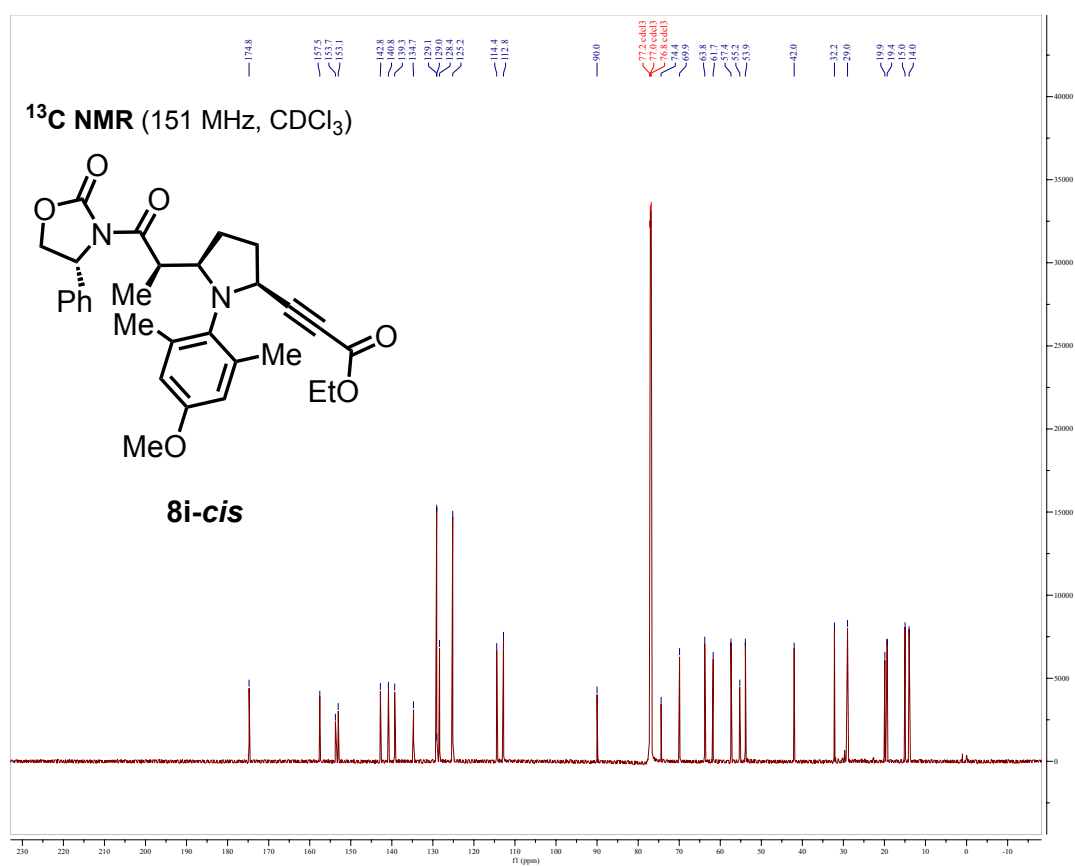
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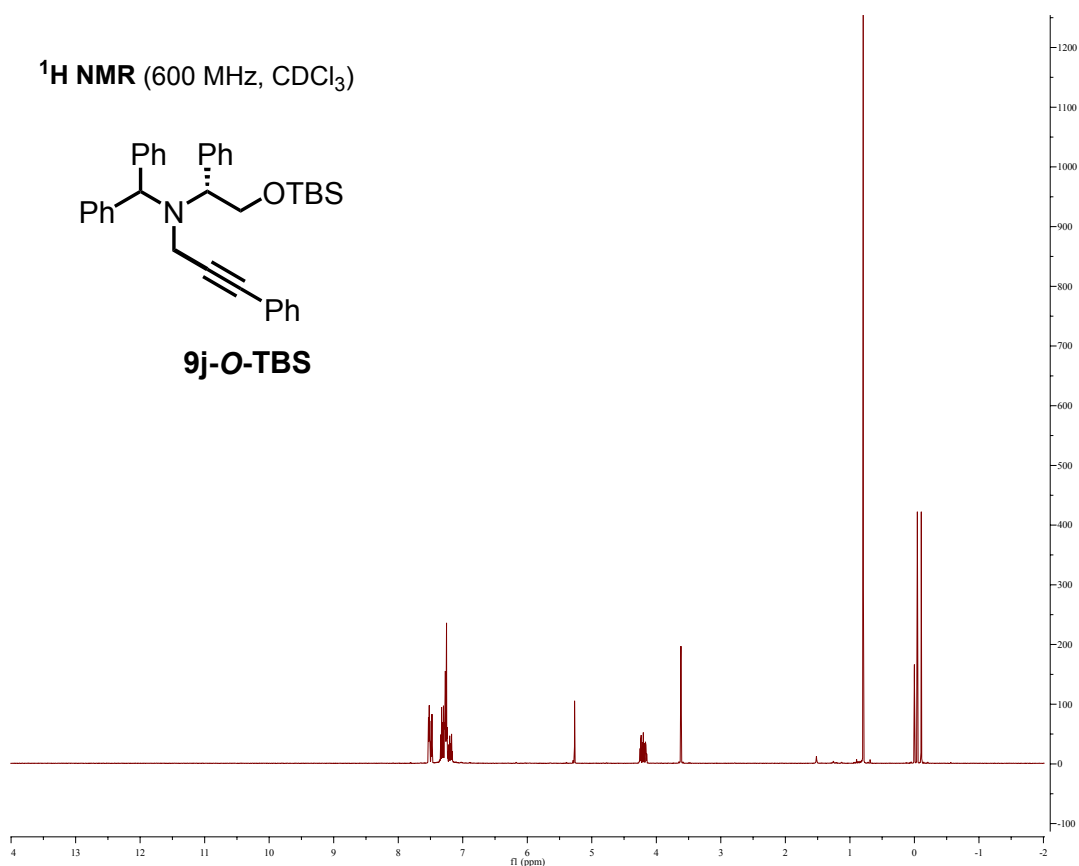
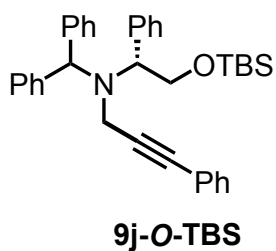


¹H NMR (600 MHz, CDCl₃)

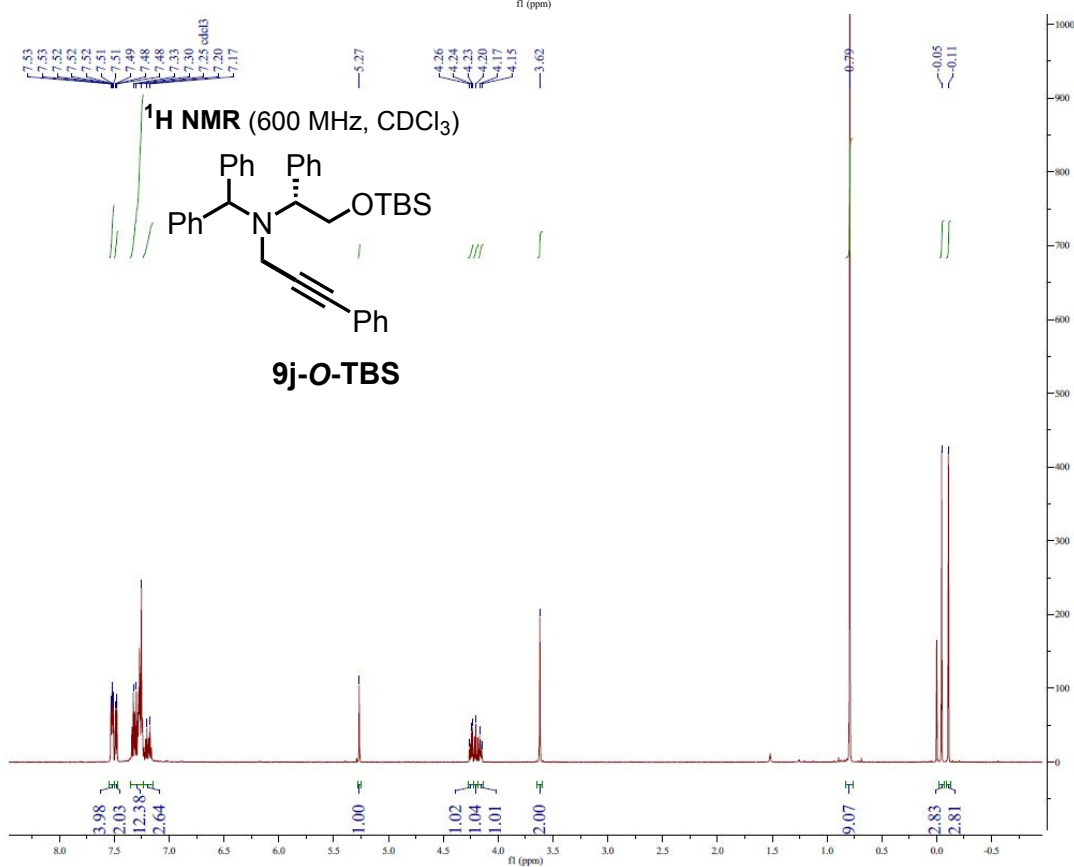
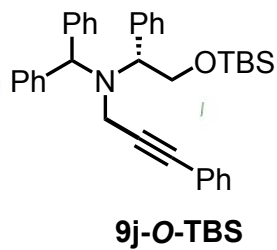




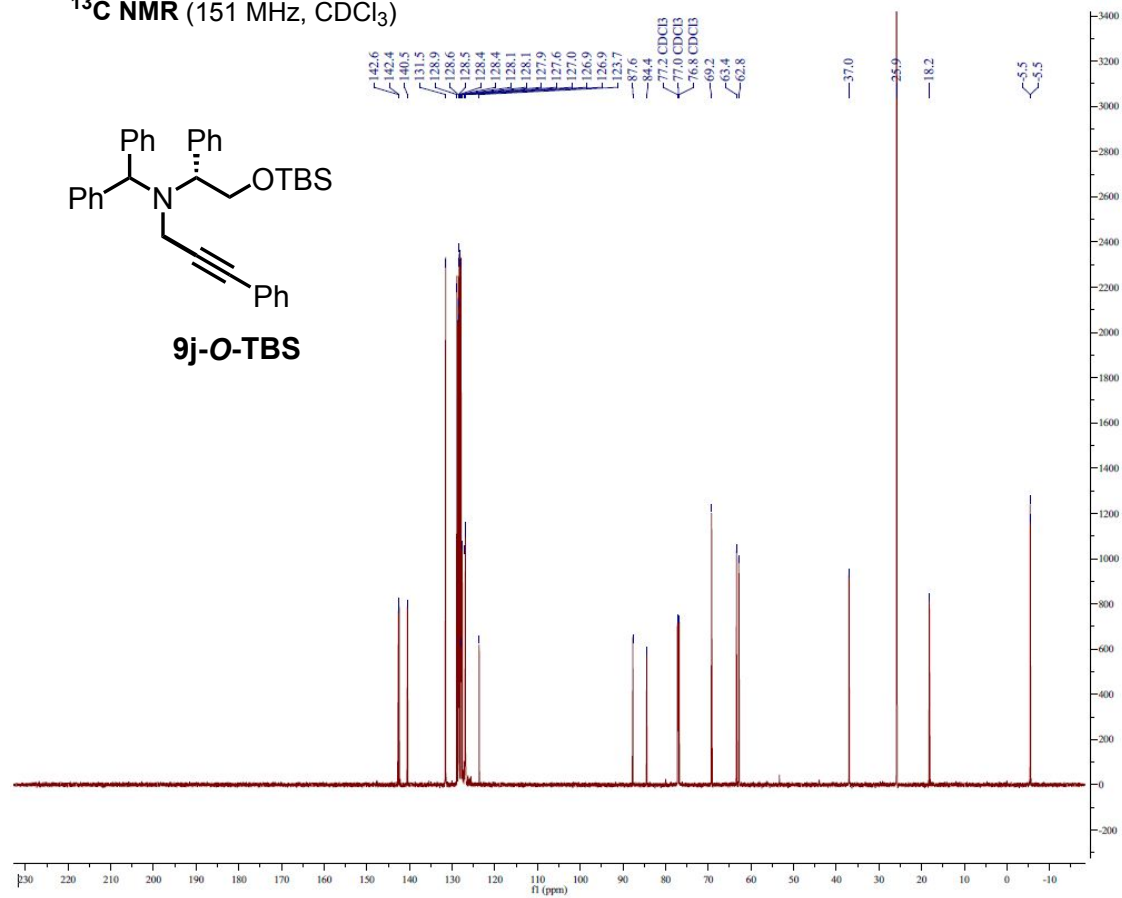
¹H NMR (600 MHz, CDCl₃)



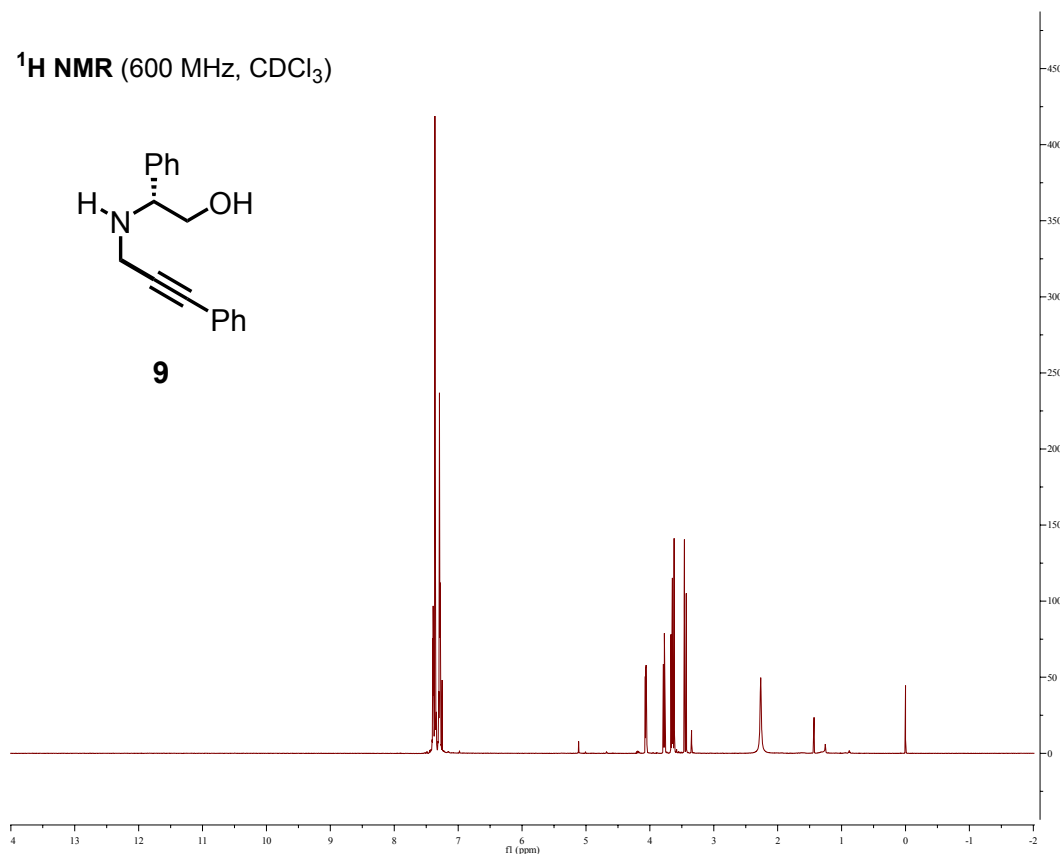
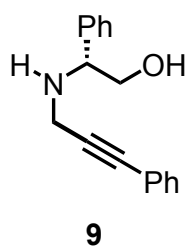
¹H NMR (600 MHz, CDCl₃)



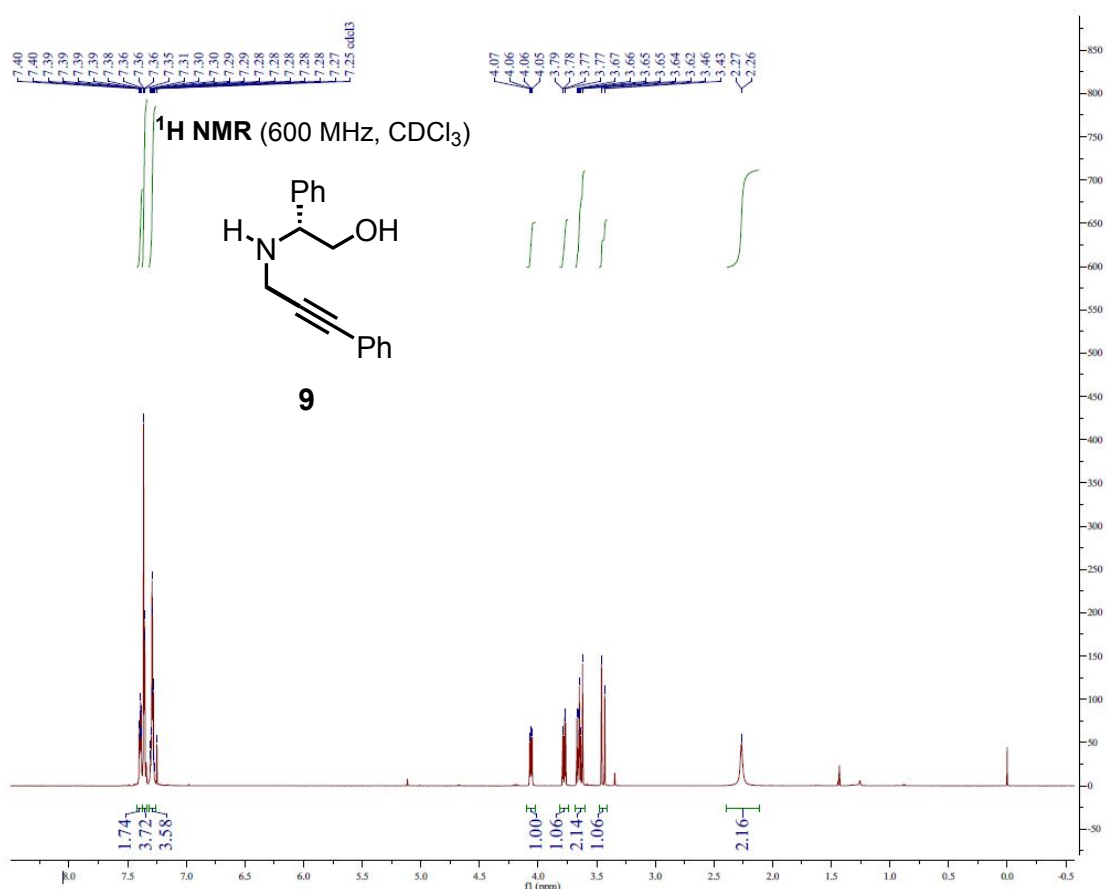
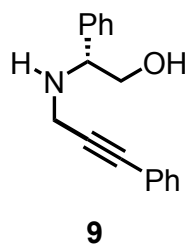
^{13}C NMR (151 MHz, CDCl_3)



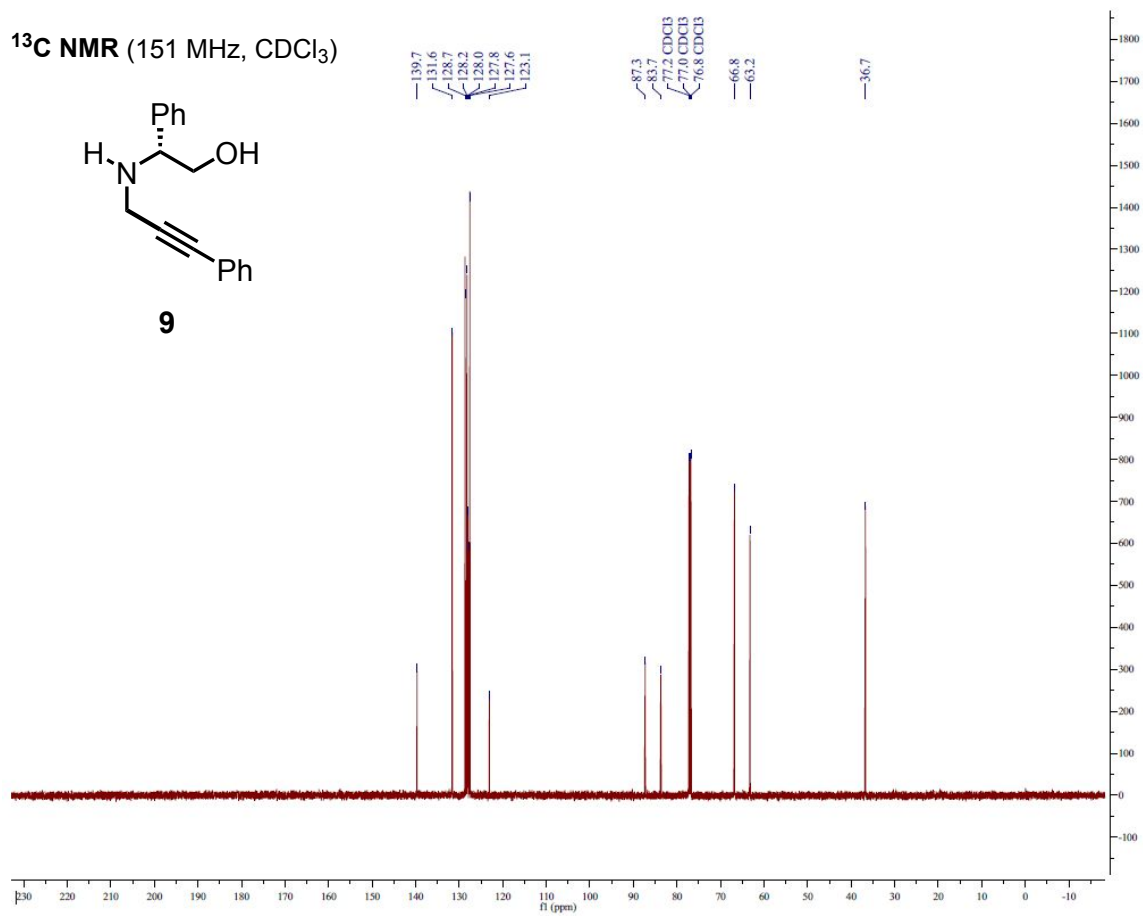
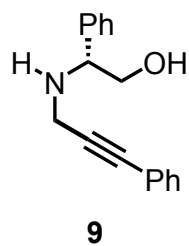
¹H NMR (600 MHz, CDCl₃)

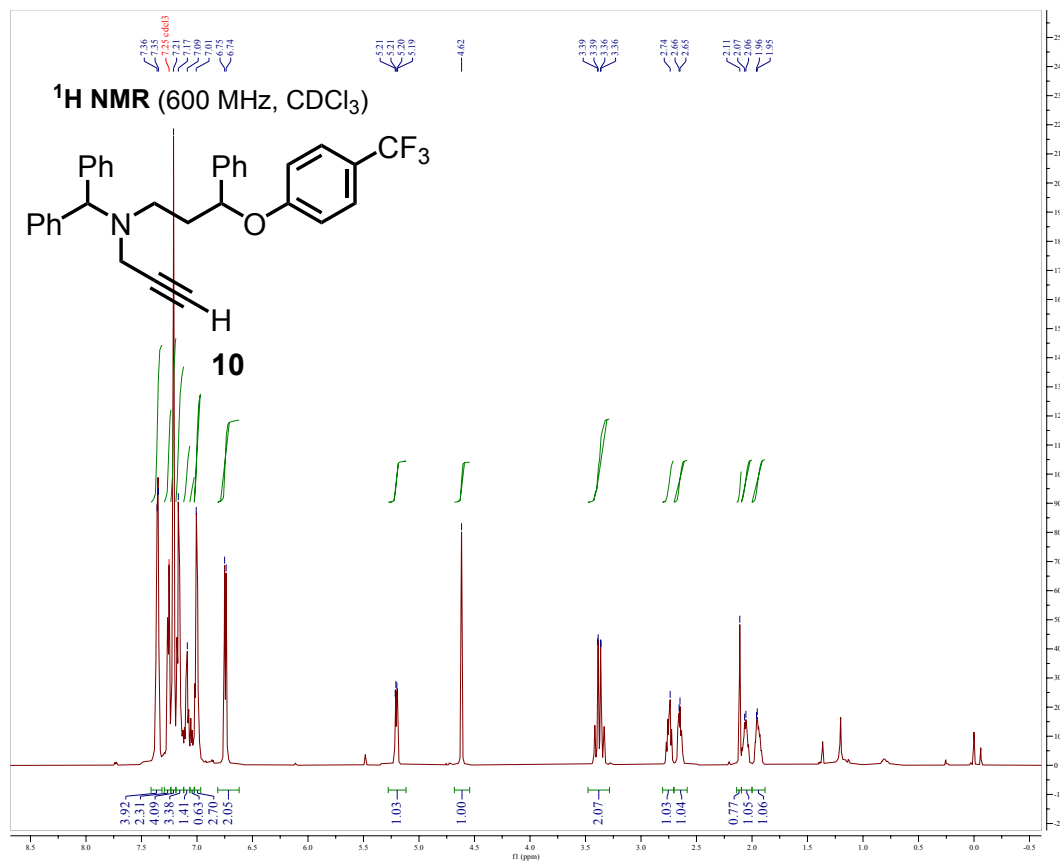
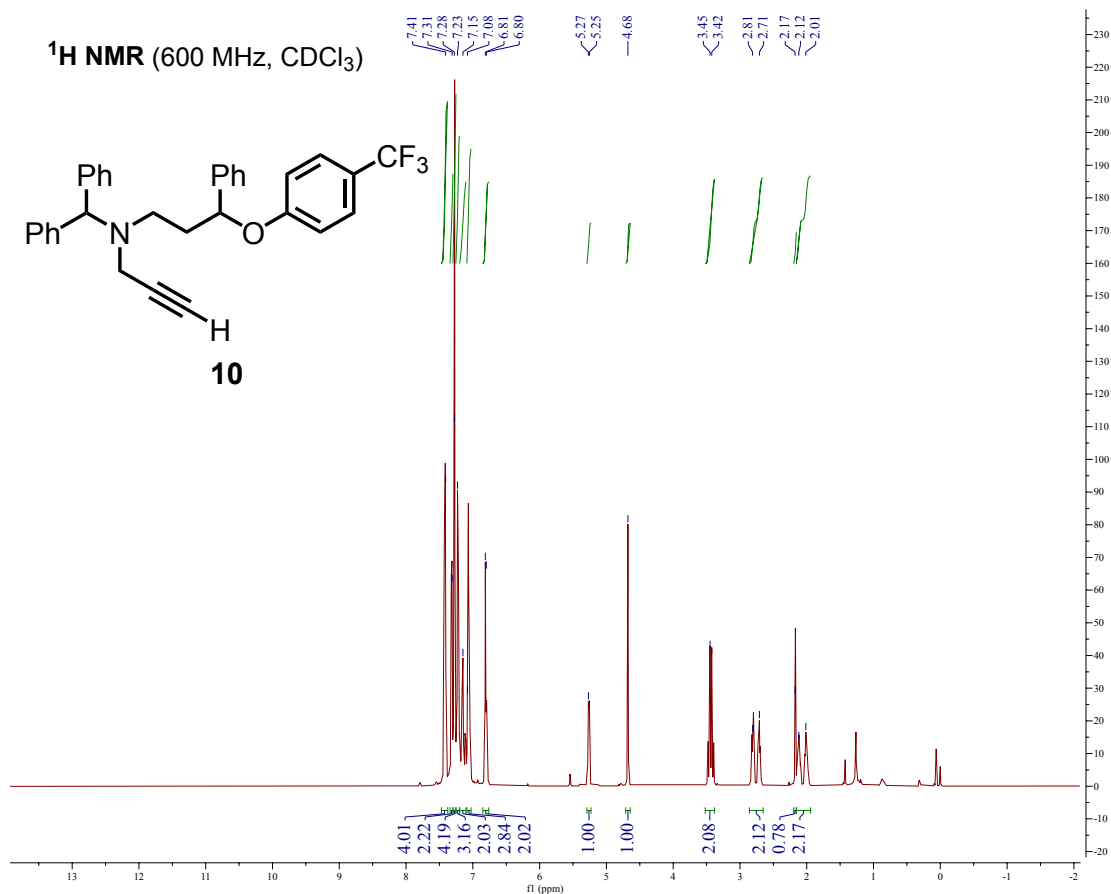


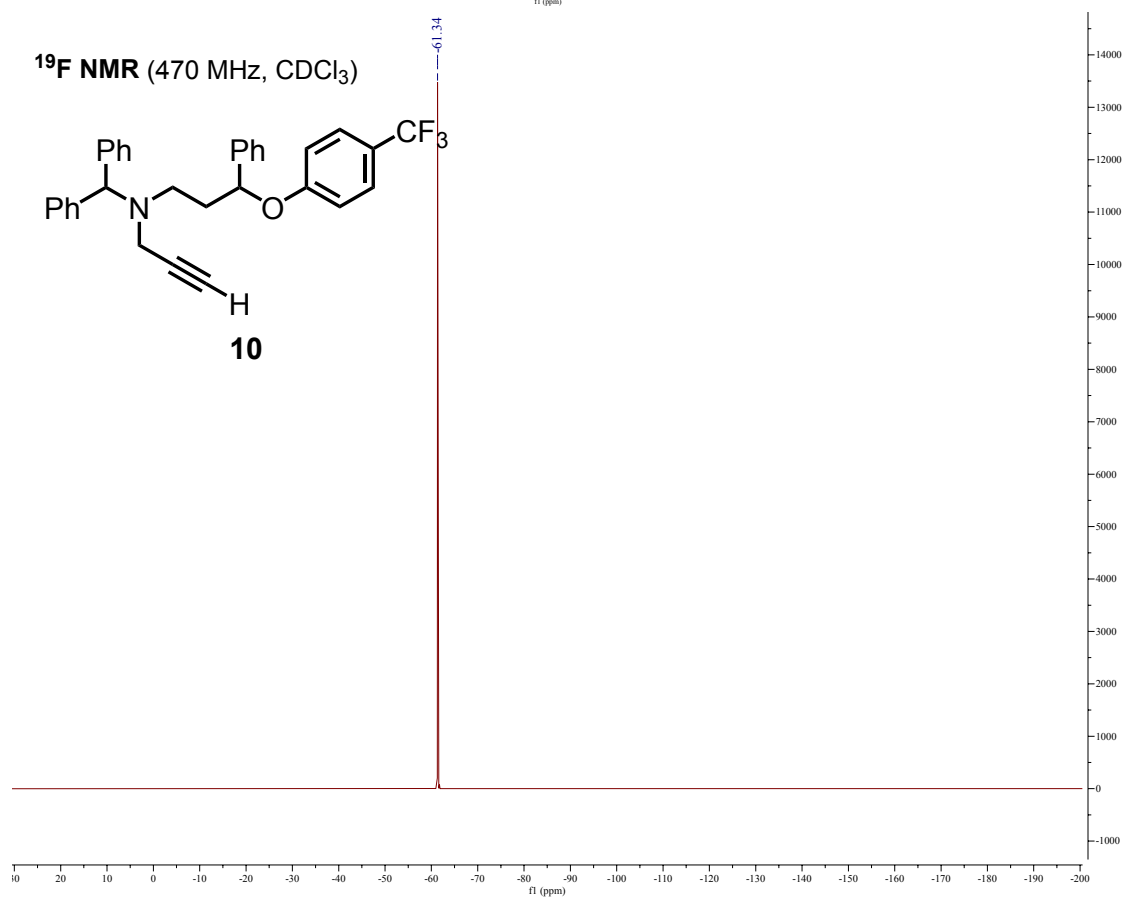
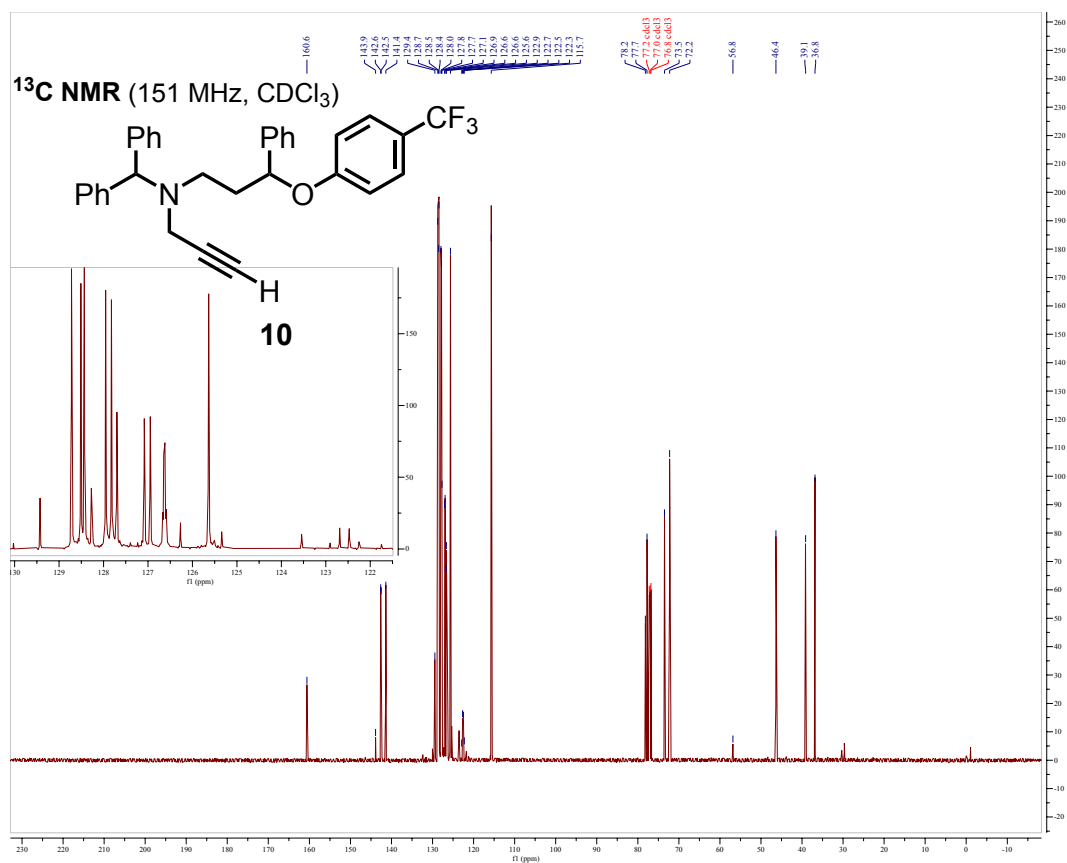
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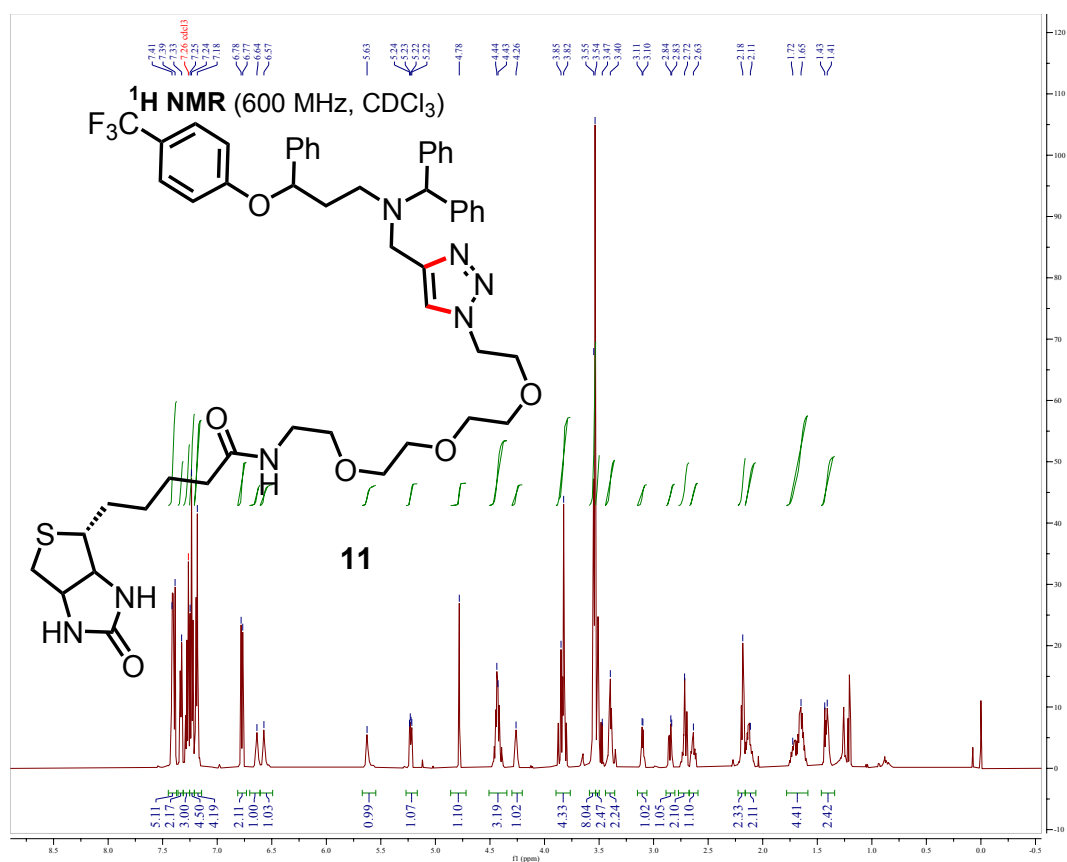
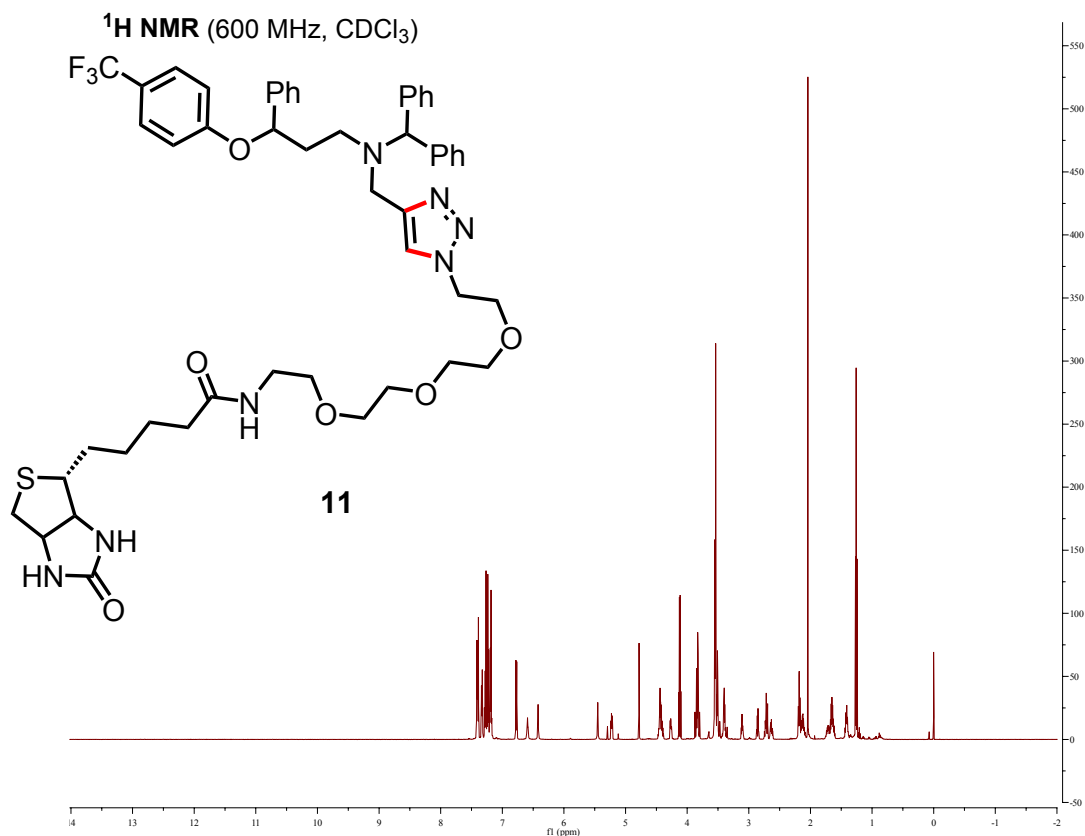


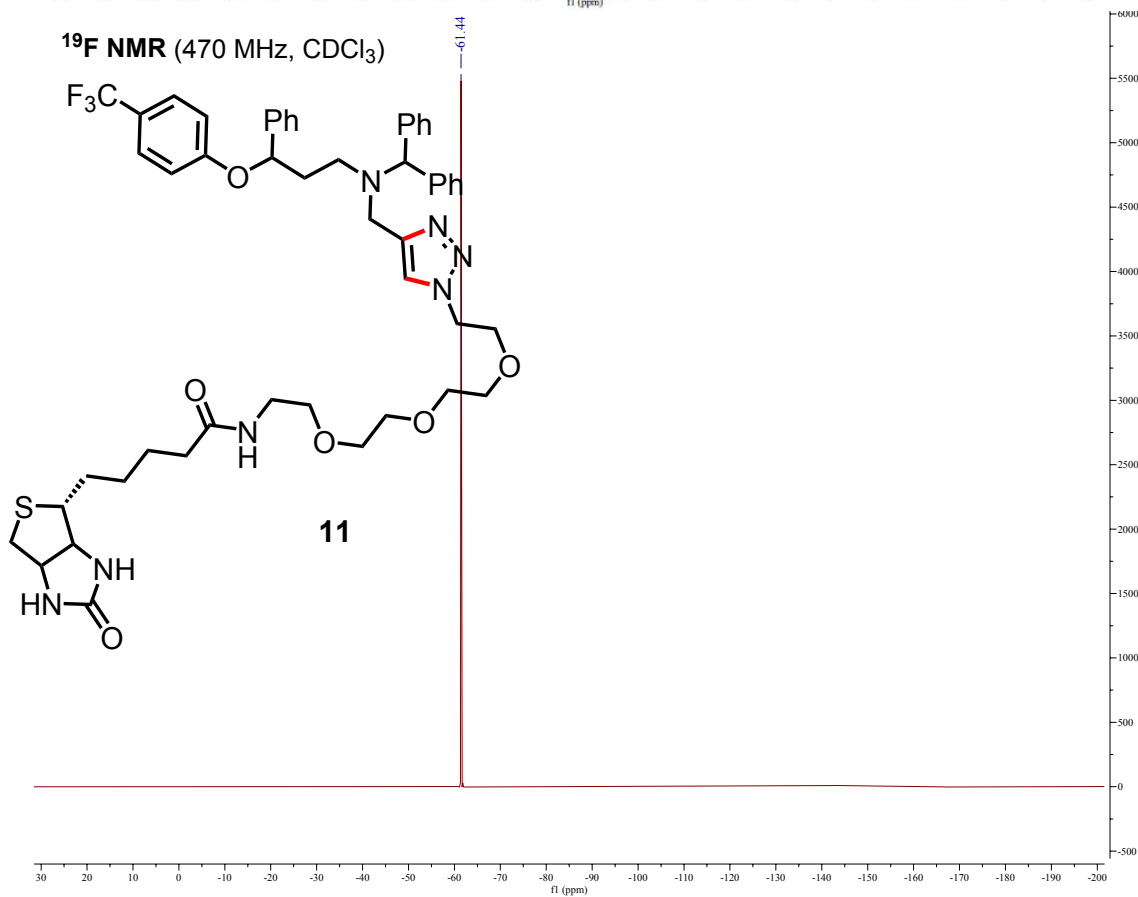
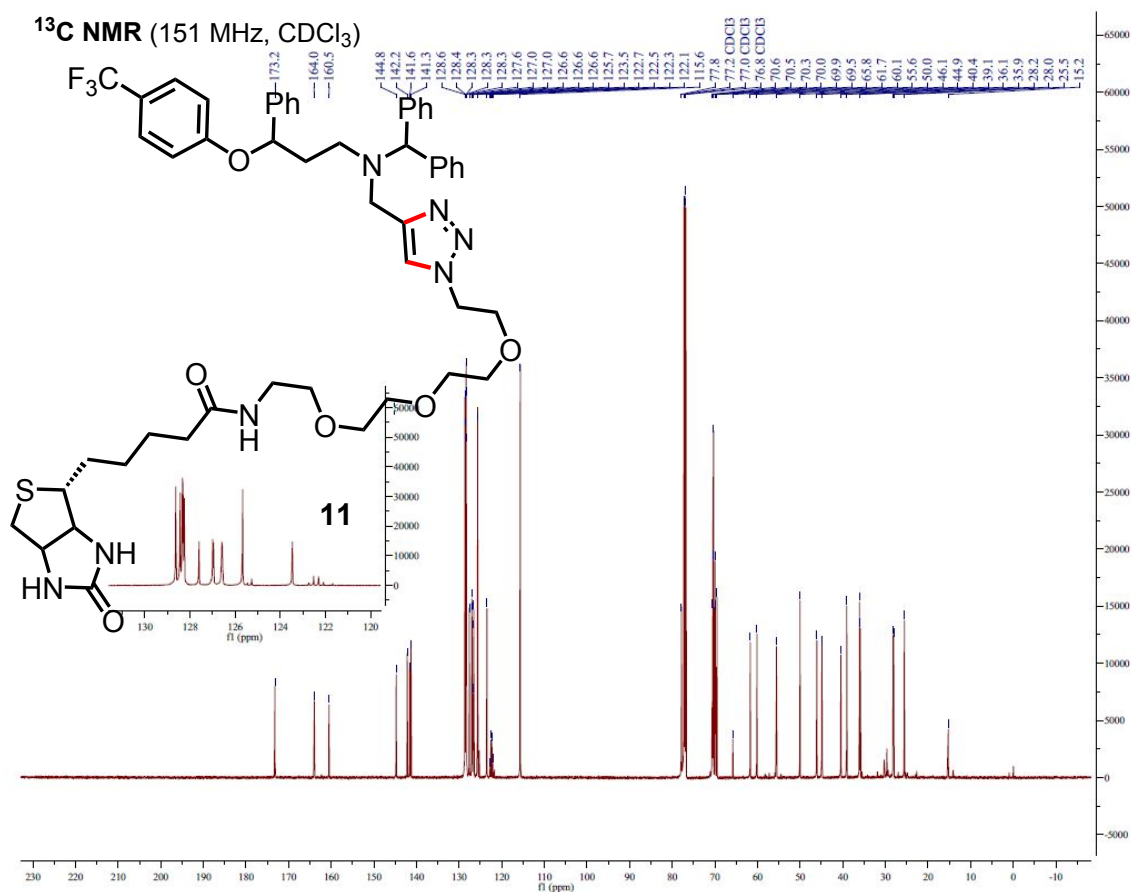
^{13}C NMR (151 MHz, CDCl_3)



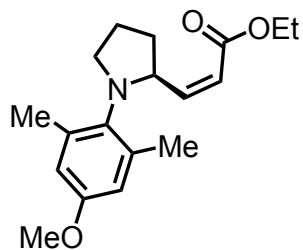




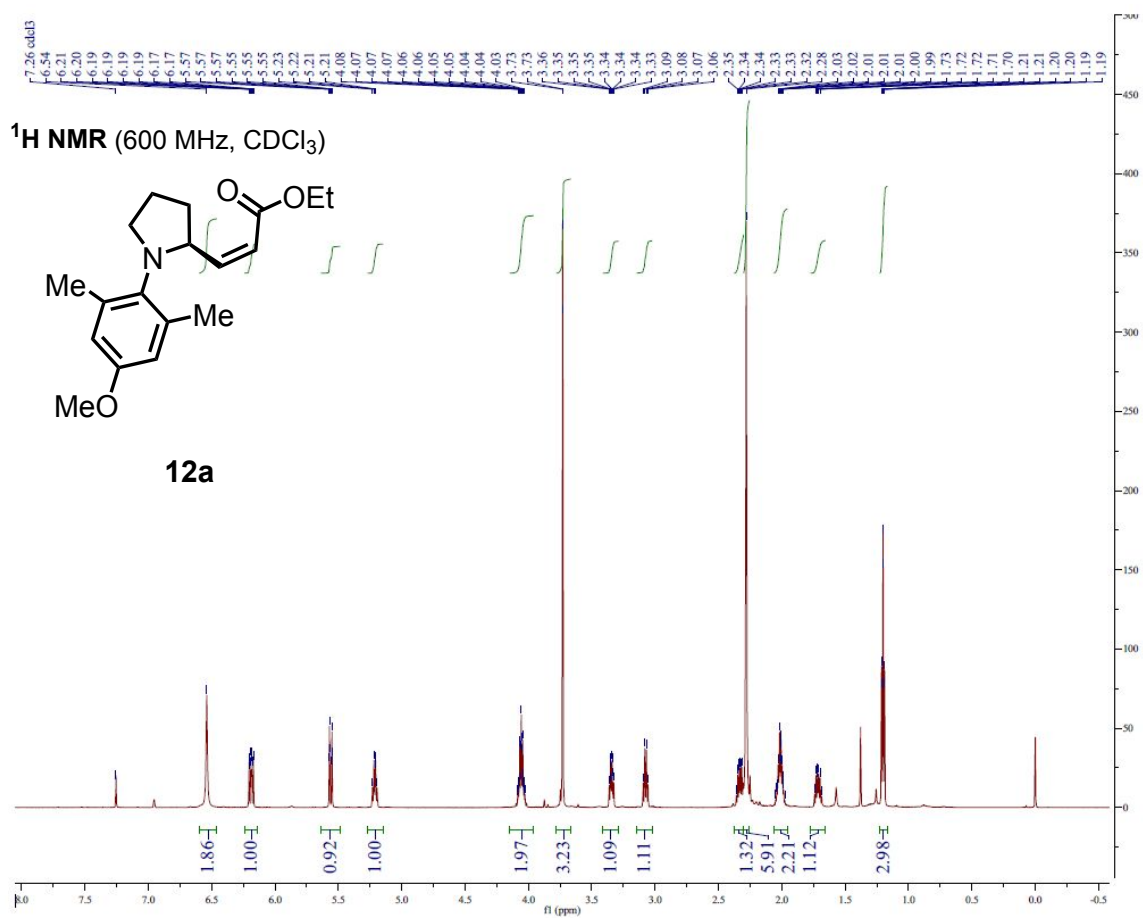
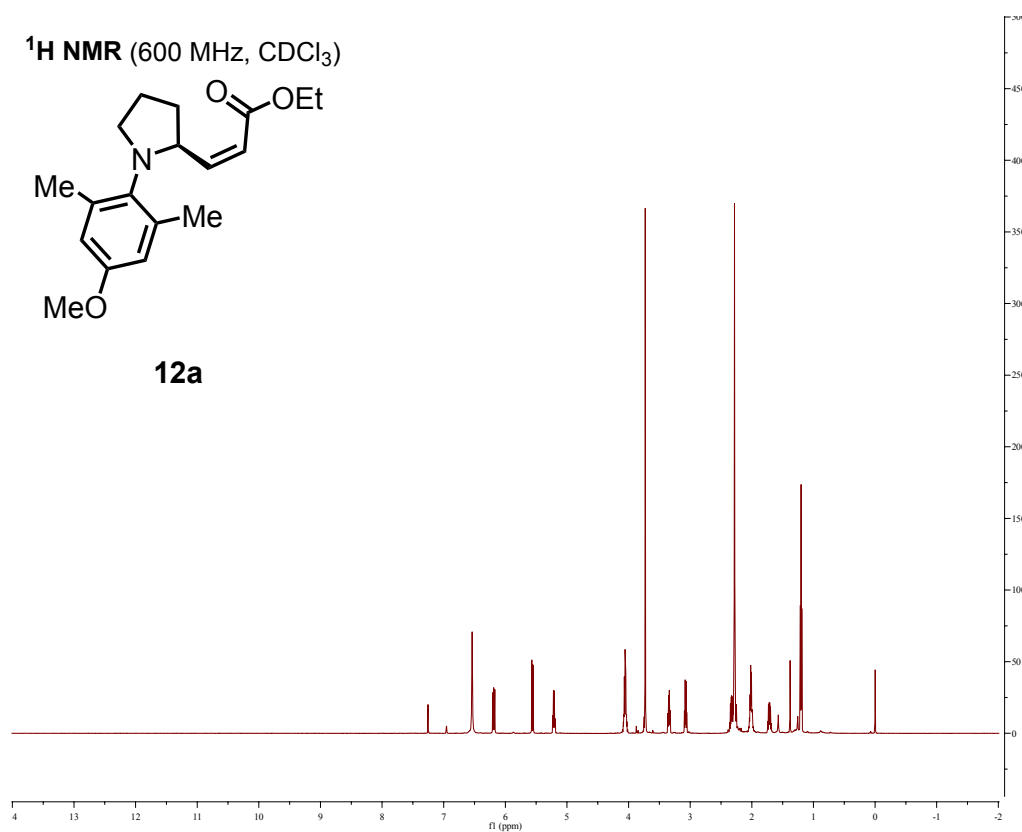




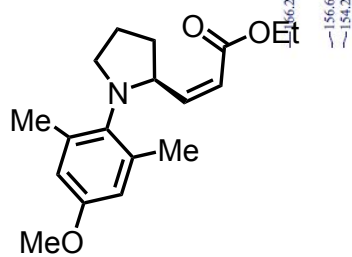
¹H NMR (600 MHz, CDCl₃)



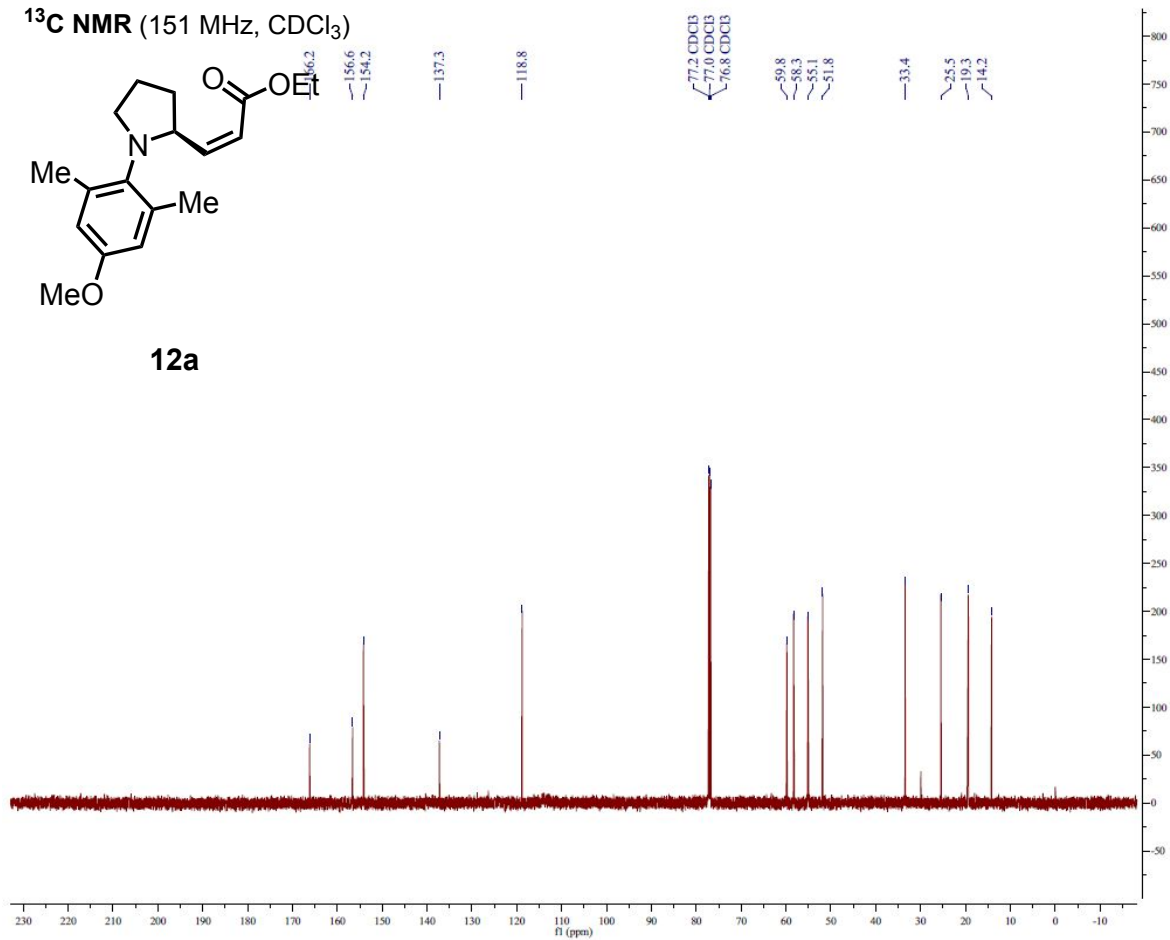
12a



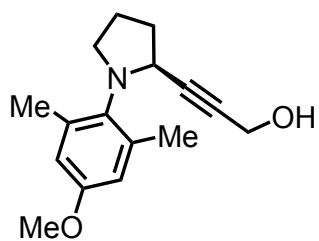
¹³C NMR (151 MHz, CDCl₃)



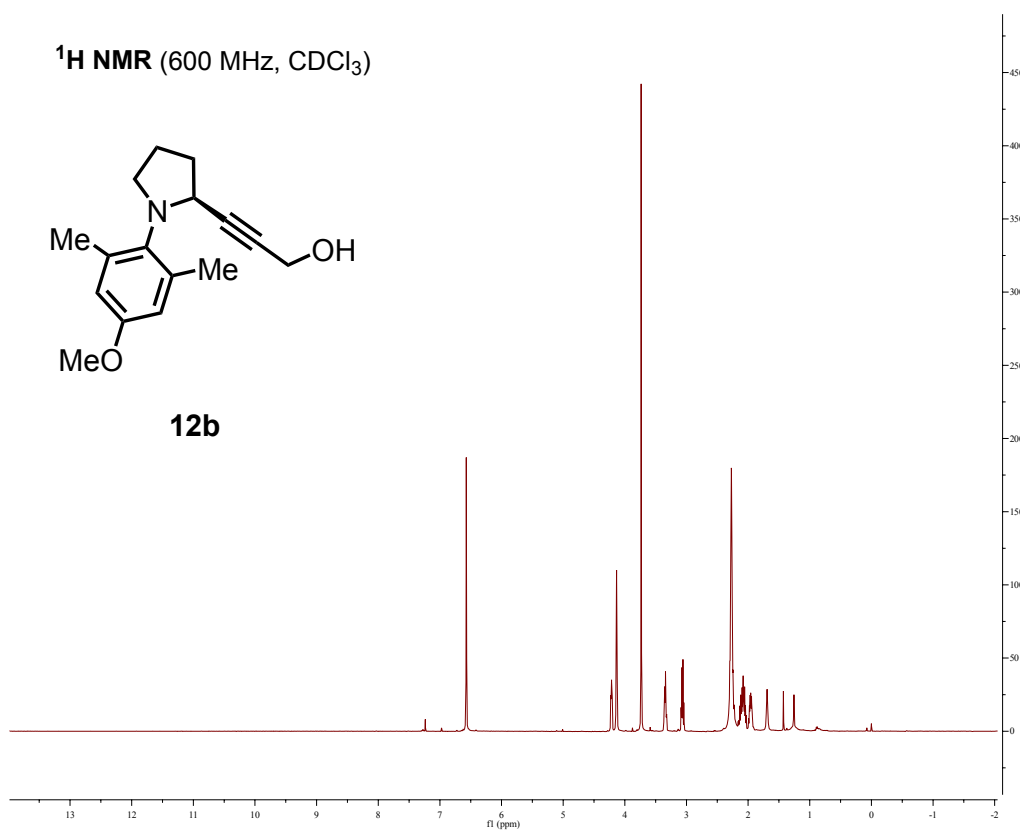
12a



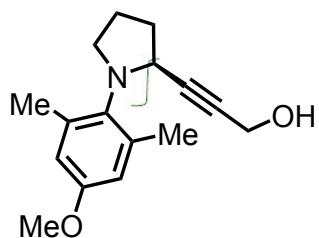
¹H NMR (600 MHz, CDCl₃)



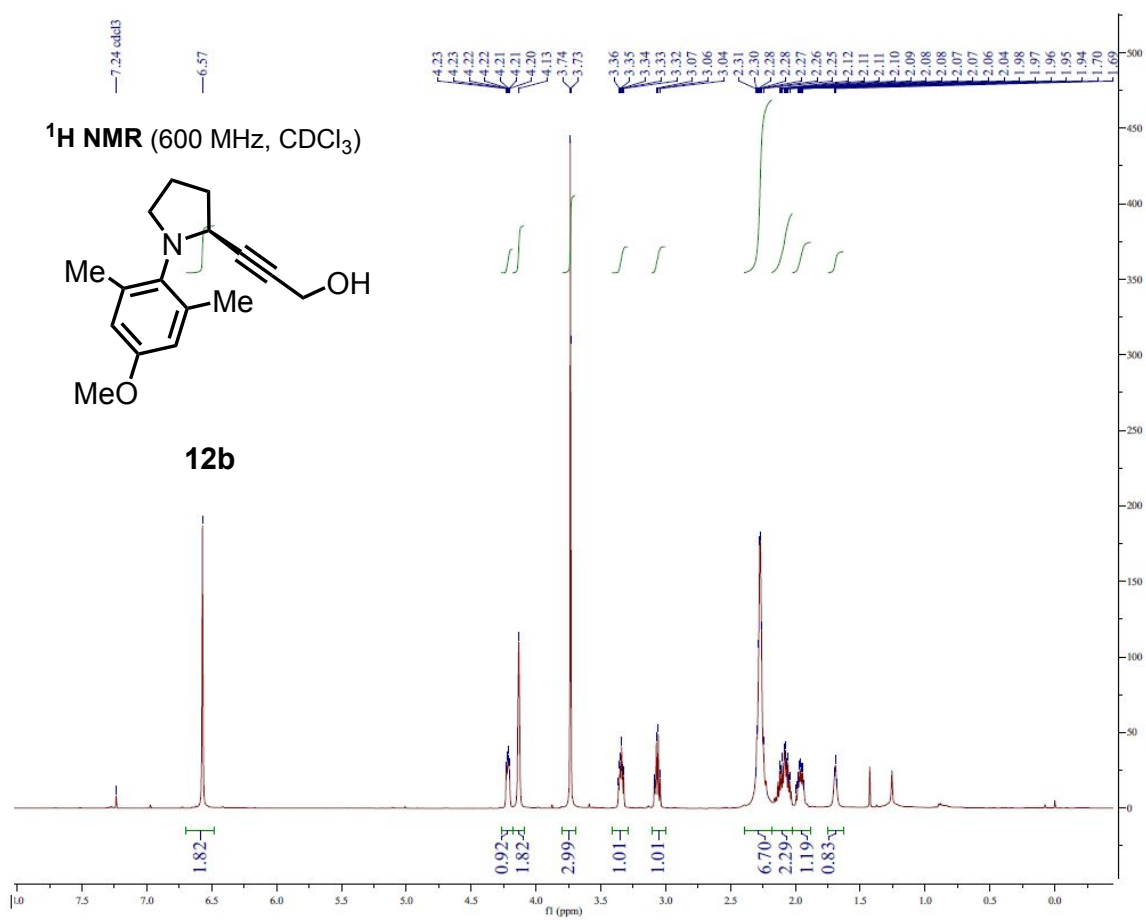
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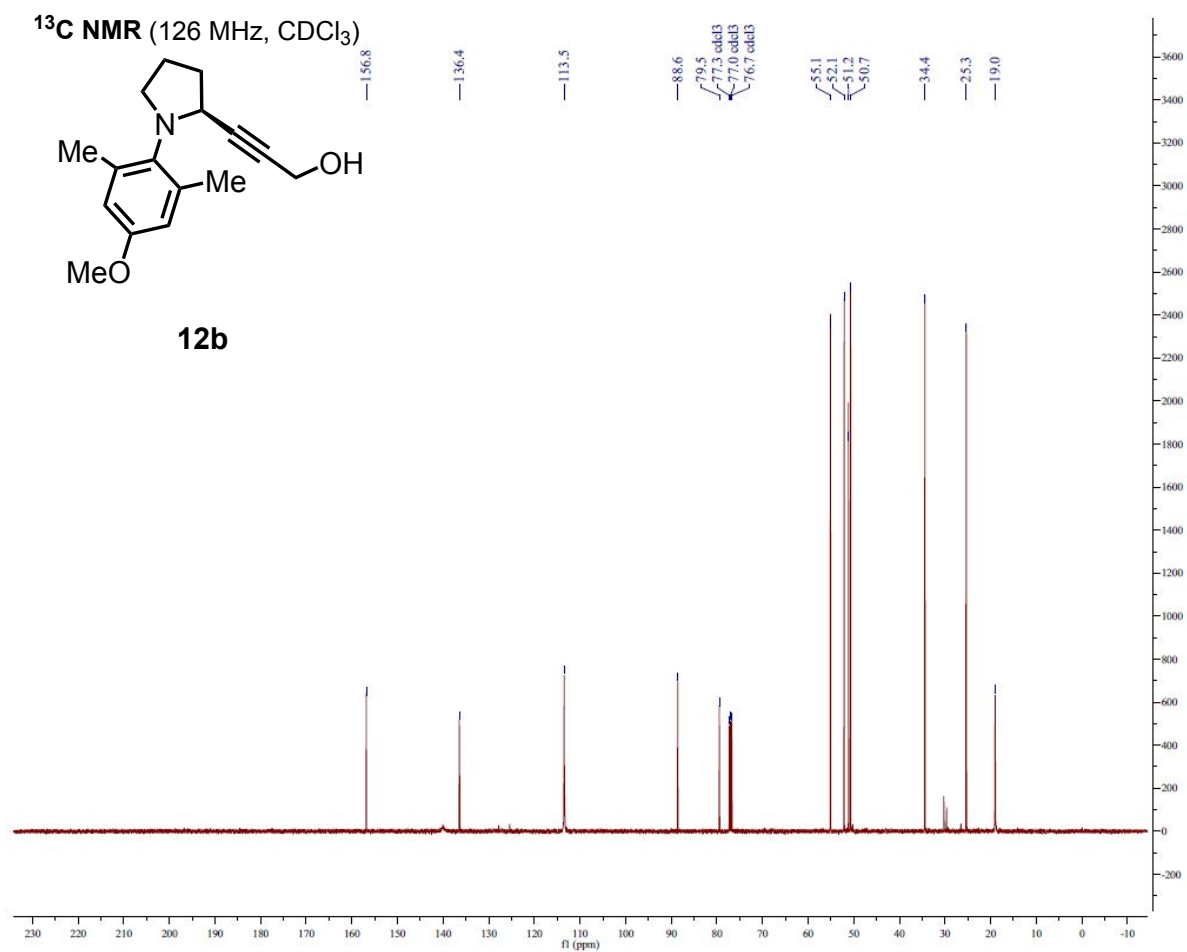


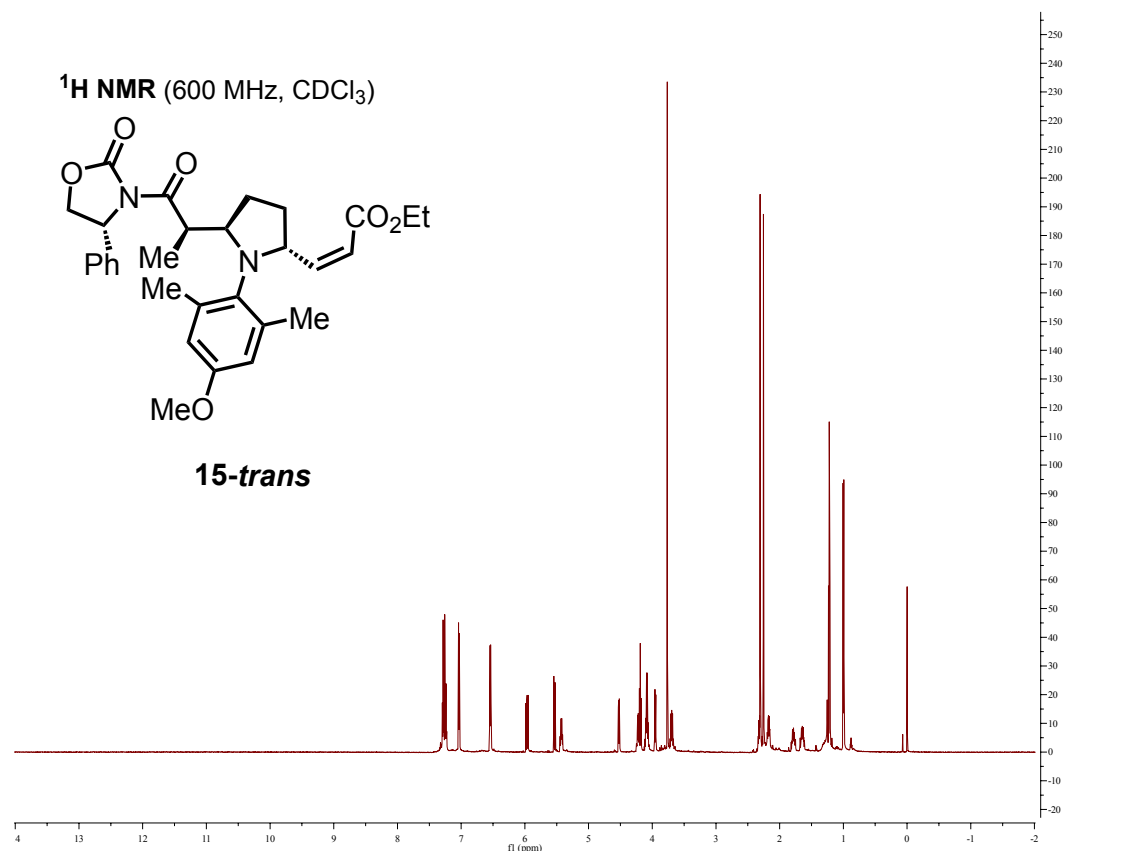
¹H NMR (600 MHz, CDCl₃)

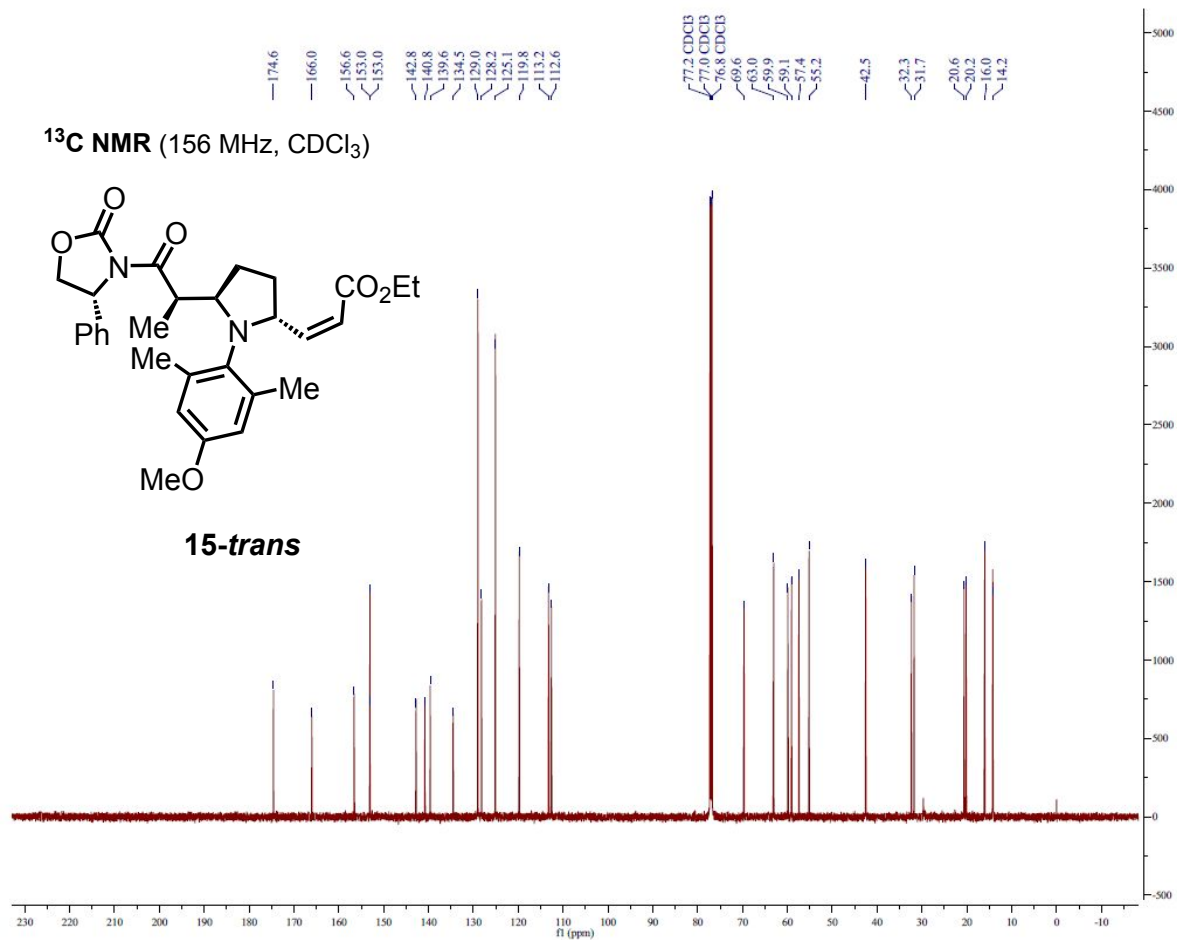


12b









9. X-Ray Crystallography Data

Table S15. Crystal data and structure refinement for $C_{23}H_{25}BrN_2O_3$.

Identification code	$C_{23}H_{25}BrN_2O_3$	
Empirical formula	C23 H25 Br N2 O3	
Formula weight	457.36	
Temperature	173(2) K	
Wavelength	1.54178 Å	
Crystal system	Monoclinic	
Space group	C2/c	
Unit cell dimensions	$a = 31.9630(17)$ Å	$\alpha = 90^\circ$.
	$b = 9.8050(5)$ Å	$\beta = 112.711(3)^\circ$.
	$c = 15.0555(8)$ Å	$\gamma = 90^\circ$.
Volume	$4352.5(4)$ Å ³	
Z	8	
Density (calculated)	1.396 Mg/m ³	
Absorption coefficient	2.783 mm ⁻¹	
F(000)	1888	
Crystal size	0.480 x 0.180 x 0.100 mm ³	
Theta range for data collection	2.997 to 66.526°.	
Index ranges	-37 ≤ h ≤ 34, 0 ≤ k ≤ 11, 0 ≤ l ≤ 17	
Reflections collected	3790	
Independent reflections	3790 [R(int) = 0.1009]	
Completeness to theta = 66.526°	98.8 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.7528 and 0.4332	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	3790 / 258 / 278	
Goodness-of-fit on F ²	1.071	
Final R indices [I > 2σ(I)]	R1 = 0.0611, wR2 = 0.1753	
R indices (all data)	R1 = 0.0826, wR2 = 0.1919	
Extinction coefficient	n/a	
Largest diff. peak and hole	0.704 and -0.647 e.Å ⁻³	

Table S16. Atomic coordinates ($\times 10^4$) and equivalent isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for $\text{C}_{23}\text{H}_{25}\text{BrN}_2\text{O}_3$. U(eq) is defined as one third of the trace of the orthogonalized U_{ij} tensor.

	x	y	z	U(eq)
Br(1)	4645(1)	4818(1)	8410(1)	102(1)
O(1)	2346(1)	5189(2)	7608(2)	72(1)
O(2)	2035(1)	3146(2)	7683(2)	72(1)
O(3)	2085(1)	9312(3)	5196(2)	83(1)
N(1)	2703(1)	3164(3)	7568(3)	70(1)
C(1)	4033(2)	4296(4)	8101(3)	78(1)
C(2)	3927(2)	2965(4)	8208(3)	74(1)
C(3)	3484(1)	2620(4)	8017(3)	69(1)
C(4)	3143(1)	3597(4)	7716(3)	67(1)
C(5)	3255(2)	4934(4)	7597(4)	79(1)
C(6)	3703(2)	5272(4)	7804(4)	85(1)
C(7)	2366(1)	3966(4)	7629(3)	66(1)
C(8)	1644(1)	3829(4)	7738(3)	70(1)
C(9)	1294(2)	4050(4)	6781(3)	70(1)
C(10)	1015(2)	4204(4)	5999(4)	80(1)
N(2)	914(2)	4846(4)	4377(3)	81(1)
C(11)	686(2)	4496(5)	5009(4)	87(1)
C(12)	391(8)	3269(17)	4511(10)	96(2)
C(13)	668(3)	2606(8)	4006(6)	109(2)
C(14)	859(3)	3808(9)	3636(6)	99(2)
C(12X)	430(30)	3200(50)	4500(20)	96(2)
C(13X)	297(7)	3530(20)	3435(13)	99(2)
C(14X)	656(9)	4490(30)	3374(12)	109(2)
C(15)	1209(1)	5996(4)	4559(3)	69(1)
C(16)	1027(2)	7312(5)	4359(3)	76(1)
C(17)	1316(2)	8433(4)	4562(3)	77(1)
C(18)	1776(2)	8273(4)	4965(3)	72(1)
C(19)	1959(1)	6967(4)	5150(3)	68(1)
C(20)	1682(1)	5831(4)	4961(3)	66(1)
C(21)	1900(2)	4441(4)	5148(4)	79(1)
C(22)	518(2)	7512(6)	3924(5)	105(2)

C(23)	1917(2)	10679(5)	5077(4)	93(1)
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Table S17. Bond lengths [Å] and angles [°] for $\text{C}_{23}\text{H}_{25}\text{BrN}_2\text{O}_3$.

Br(1)-C(1)	1.898(5)
O(1)-C(7)	1.200(4)
O(2)-C(7)	1.356(4)
O(2)-C(8)	1.449(5)
O(3)-C(18)	1.367(5)
O(3)-C(23)	1.430(5)
N(1)-C(7)	1.364(5)
N(1)-C(4)	1.404(5)
N(1)-H(1N)	0.93(5)
C(1)-C(6)	1.366(6)
C(1)-C(2)	1.373(6)
C(2)-C(3)	1.373(6)
C(2)-H(2)	0.9500
C(3)-C(4)	1.389(5)
C(3)-H(3)	0.9500
C(4)-C(5)	1.387(5)
C(5)-C(6)	1.384(7)
C(5)-H(5)	0.9500
C(6)-H(6)	0.9500
C(8)-C(9)	1.460(7)
C(8)-H(8A)	0.9900
C(8)-H(8B)	0.9900
C(9)-C(10)	1.181(6)
C(10)-C(11)	1.482(8)
N(2)-C(15)	1.428(5)
N(2)-C(11)	1.445(6)
N(2)-C(14)	1.470(7)
C(11)-C(12)	1.534(8)
C(11)-H(11)	1.0000
C(12)-C(13)	1.52(2)
C(12)-H(12A)	0.9900

C(12)-H(12B)	0.9900
C(13)-C(14)	1.527(10)
C(13)-H(13A)	0.9900
C(13)-H(13B)	0.9900
C(14)-H(14A)	0.9900
C(14)-H(14B)	0.9900
C(15)-C(16)	1.400(6)
C(15)-C(20)	1.403(6)
C(16)-C(17)	1.391(6)
C(16)-C(22)	1.514(7)
C(17)-C(18)	1.366(6)
C(17)-H(17)	0.9500
C(18)-C(19)	1.390(5)
C(19)-C(20)	1.383(5)
C(19)-H(19)	0.9500
C(20)-C(21)	1.508(6)
C(21)-H(21A)	0.9800
C(21)-H(21B)	0.9800
C(21)-H(21C)	0.9800
C(22)-H(22A)	0.9800
C(22)-H(22B)	0.9800
C(22)-H(22C)	0.9800
C(23)-H(23A)	0.9800
C(23)-H(23B)	0.9800
C(23)-H(23C)	0.9800

C(7)-O(2)-C(8)	116.1(3)
C(18)-O(3)-C(23)	117.8(4)
C(7)-N(1)-C(4)	125.8(3)
C(7)-N(1)-H(1N)	114(3)
C(4)-N(1)-H(1N)	117(3)
C(6)-C(1)-C(2)	120.7(4)
C(6)-C(1)-Br(1)	118.9(3)
C(2)-C(1)-Br(1)	120.4(3)
C(3)-C(2)-C(1)	119.4(4)
C(3)-C(2)-H(2)	120.3
C(1)-C(2)-H(2)	120.3

C(2)-C(3)-C(4)	120.9(4)
C(2)-C(3)-H(3)	119.6
C(4)-C(3)-H(3)	119.6
C(5)-C(4)-C(3)	119.1(4)
C(5)-C(4)-N(1)	123.8(4)
C(3)-C(4)-N(1)	117.1(3)
C(6)-C(5)-C(4)	119.5(4)
C(6)-C(5)-H(5)	120.2
C(4)-C(5)-H(5)	120.2
C(1)-C(6)-C(5)	120.4(4)
C(1)-C(6)-H(6)	119.8
C(5)-C(6)-H(6)	119.8
O(1)-C(7)-O(2)	124.1(3)
O(1)-C(7)-N(1)	127.5(4)
O(2)-C(7)-N(1)	108.4(3)
O(2)-C(8)-C(9)	111.1(3)
O(2)-C(8)-H(8A)	109.4
C(9)-C(8)-H(8A)	109.4
O(2)-C(8)-H(8B)	109.4
C(9)-C(8)-H(8B)	109.4
H(8A)-C(8)-H(8B)	108.0
C(10)-C(9)-C(8)	178.4(5)
C(9)-C(10)-C(11)	175.3(5)
C(15)-N(2)-C(11)	121.7(4)
C(15)-N(2)-C(14)	124.7(4)
C(11)-N(2)-C(14)	113.1(4)
N(2)-C(11)-C(10)	111.4(4)
N(2)-C(11)-C(12)	103.7(5)
C(10)-C(11)-C(12)	113.8(9)
N(2)-C(11)-H(11)	109.3
C(10)-C(11)-H(11)	109.3
C(12)-C(11)-H(11)	109.3
C(13)-C(12)-C(11)	102.2(10)
C(13)-C(12)-H(12A)	111.3
C(11)-C(12)-H(12A)	111.3
C(13)-C(12)-H(12B)	111.3
C(11)-C(12)-H(12B)	111.3

H(12A)-C(12)-H(12B)	109.2
C(12)-C(13)-C(14)	104.2(7)
C(12)-C(13)-H(13A)	110.9
C(14)-C(13)-H(13A)	110.9
C(12)-C(13)-H(13B)	110.9
C(14)-C(13)-H(13B)	110.9
H(13A)-C(13)-H(13B)	108.9
N(2)-C(14)-C(13)	101.9(5)
N(2)-C(14)-H(14A)	111.4
C(13)-C(14)-H(14A)	111.4
N(2)-C(14)-H(14B)	111.4
C(13)-C(14)-H(14B)	111.4
H(14A)-C(14)-H(14B)	109.3
C(16)-C(15)-C(20)	119.2(4)
C(16)-C(15)-N(2)	119.9(4)
C(20)-C(15)-N(2)	120.9(4)
C(17)-C(16)-C(15)	119.7(4)
C(17)-C(16)-C(22)	120.3(4)
C(15)-C(16)-C(22)	120.0(4)
C(18)-C(17)-C(16)	121.0(4)
C(18)-C(17)-H(17)	119.5
C(16)-C(17)-H(17)	119.5
C(17)-C(18)-O(3)	125.1(4)
C(17)-C(18)-C(19)	119.5(4)
O(3)-C(18)-C(19)	115.4(4)
C(20)-C(19)-C(18)	120.9(4)
C(20)-C(19)-H(19)	119.6
C(18)-C(19)-H(19)	119.6
C(19)-C(20)-C(15)	119.6(4)
C(19)-C(20)-C(21)	118.4(4)
C(15)-C(20)-C(21)	121.9(4)
C(20)-C(21)-H(21A)	109.5
C(20)-C(21)-H(21B)	109.5
H(21A)-C(21)-H(21B)	109.5
C(20)-C(21)-H(21C)	109.5
H(21A)-C(21)-H(21C)	109.5
H(21B)-C(21)-H(21C)	109.5

C(16)-C(22)-H(22A)	109.5
C(16)-C(22)-H(22B)	109.5
H(22A)-C(22)-H(22B)	109.5
C(16)-C(22)-H(22C)	109.5
H(22A)-C(22)-H(22C)	109.5
H(22B)-C(22)-H(22C)	109.5
O(3)-C(23)-H(23A)	109.5
O(3)-C(23)-H(23B)	109.5
H(23A)-C(23)-H(23B)	109.5
O(3)-C(23)-H(23C)	109.5
H(23A)-C(23)-H(23C)	109.5
H(23B)-C(23)-H(23C)	109.5

Symmetry transformations used to generate equivalent atoms:

Table S18. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for $\text{C}_{23}\text{H}_{25}\text{BrN}_2\text{O}_3$. The anisotropic displacement factor exponent takes the form: $-2\pi^2 [h^2 a^{*2} U^{11} + \dots + 2 h k a^* b^* U^{12}]$

	U11	U22	U33	U23	U13	U12
Br(1)	80(1)	77(1)	152(1)	7(1)	50(1)	-1(1)
O(1)	79(2)	45(1)	97(2)	6(1)	39(2)	3(1)
O(2)	78(2)	47(1)	96(2)	1(1)	41(2)	-1(1)
O(3)	93(2)	52(1)	112(2)	1(1)	47(2)	-1(1)
N(1)	79(2)	45(2)	87(2)	3(1)	35(2)	2(1)
C(1)	83(3)	61(2)	99(3)	5(2)	46(2)	4(2)
C(2)	84(3)	53(2)	93(3)	7(2)	42(2)	11(2)
C(3)	85(2)	46(2)	84(3)	1(2)	41(2)	3(2)
C(4)	77(2)	51(2)	78(3)	3(2)	35(2)	1(2)
C(5)	82(3)	50(2)	111(4)	11(2)	44(3)	6(2)
C(6)	86(3)	52(2)	123(4)	10(2)	46(3)	1(2)
C(7)	74(2)	52(2)	73(2)	2(2)	30(2)	-1(2)
C(8)	84(2)	52(2)	84(3)	-3(2)	44(2)	0(2)
C(9)	79(2)	56(2)	82(3)	-2(2)	39(2)	-6(2)
C(10)	88(3)	63(2)	91(3)	-1(2)	38(2)	-12(2)

N(2)	90(2)	75(2)	83(2)	-8(2)	40(2)	-14(2)
C(11)	85(3)	84(3)	95(3)	1(2)	39(2)	-11(2)
C(12)	89(5)	98(4)	101(3)	-7(3)	37(3)	-25(3)
C(13)	111(5)	95(4)	131(5)	-37(4)	56(4)	-40(4)
C(14)	107(5)	99(4)	101(4)	-33(3)	52(4)	-28(4)
C(12X)	89(5)	98(4)	101(3)	-7(3)	37(3)	-25(3)
C(13X)	107(5)	99(4)	101(4)	-33(3)	52(4)	-28(4)
C(14X)	111(5)	95(4)	131(5)	-37(4)	56(4)	-40(4)
C(15)	77(2)	65(2)	69(2)	2(2)	32(2)	-1(2)
C(16)	78(2)	73(2)	83(3)	8(2)	39(2)	5(2)
C(17)	90(3)	60(2)	90(3)	11(2)	44(2)	14(2)
C(18)	86(2)	57(2)	81(3)	5(2)	39(2)	2(2)
C(19)	77(2)	59(2)	74(3)	0(2)	35(2)	3(2)
C(20)	81(2)	57(2)	67(2)	2(2)	37(2)	1(2)
C(21)	88(3)	58(2)	95(3)	5(2)	39(2)	2(2)
C(22)	81(3)	100(4)	133(4)	26(3)	39(3)	17(3)
C(23)	114(4)	55(2)	120(4)	1(2)	56(3)	2(2)

Table S19. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for $\text{C}_{23}\text{H}_{25}\text{BrN}_2\text{O}_3$.

	x	y	z	U(eq)
H(1N)	2658(15)	2230(50)	7620(30)	84
H(2)	4157	2289	8411	89
H(3)	3410	1701	8092	83
H(5)	3025	5611	7376	95
H(6)	3782	6190	7738	102
H(8A)	1738	4718	8067	84
H(8B)	1518	3270	8124	84
H(11)	486	5270	5029	104
H(12A)	349	2642	4986	115
H(12B)	90	3564	4045	115
H(13A)	475	2024	3467	131
H(13B)	916	2044	4460	131
H(14A)	1153	3575	3598	118
H(14B)	644	4114	2995	118
H(12C)	626	2389	4691	115
H(12D)	157	3049	4649	115
H(13C)	287	2681	3068	118
H(13D)	-6	3962	3166	118
H(14C)	850	4033	3087	131
H(14D)	516	5309	2989	131
H(17)	1191	9324	4419	93
H(19)	2279	6854	5409	82
H(21A)	1899	4062	4544	119
H(21B)	1731	3836	5406	119
H(21C)	2214	4521	5614	119
H(22A)	409	7687	4438	158
H(22B)	373	6688	3571	158
H(22C)	443	8290	3481	158
H(23A)	1743	10860	4393	140
H(23B)	2172	11317	5324	140
H(23C)	1720	10798	5434	140

Table S20. Torsion angles [°] for C₂₃H₂₅BrN₂O₃.

C(6)-C(1)-C(2)-C(3)	-0.3(7)
Br(1)-C(1)-C(2)-C(3)	177.4(3)
C(1)-C(2)-C(3)-C(4)	0.2(6)
C(2)-C(3)-C(4)-C(5)	0.8(6)
C(2)-C(3)-C(4)-N(1)	-177.9(4)
C(7)-N(1)-C(4)-C(5)	-24.6(6)
C(7)-N(1)-C(4)-C(3)	154.0(4)
C(3)-C(4)-C(5)-C(6)	-1.6(7)
N(1)-C(4)-C(5)-C(6)	177.0(4)
C(2)-C(1)-C(6)-C(5)	-0.5(8)
Br(1)-C(1)-C(6)-C(5)	-178.2(4)
C(4)-C(5)-C(6)-C(1)	1.5(8)
C(8)-O(2)-C(7)-O(1)	-1.3(5)
C(8)-O(2)-C(7)-N(1)	-179.2(3)
C(4)-N(1)-C(7)-O(1)	17.4(7)
C(4)-N(1)-C(7)-O(2)	-164.8(4)
C(7)-O(2)-C(8)-C(9)	90.8(4)
C(15)-N(2)-C(11)-C(10)	-60.4(6)
C(14)-N(2)-C(11)-C(10)	111.7(6)
C(15)-N(2)-C(11)-C(12)	176.9(11)
C(14)-N(2)-C(11)-C(12)	-11.1(13)
N(2)-C(11)-C(12)-C(13)	30.5(14)
C(10)-C(11)-C(12)-C(13)	-90.7(10)
C(11)-C(12)-C(13)-C(14)	-39.1(14)
C(15)-N(2)-C(14)-C(13)	158.8(6)
C(11)-N(2)-C(14)-C(13)	-13.0(9)
C(12)-C(13)-C(14)-N(2)	32.0(11)
C(11)-N(2)-C(15)-C(16)	-76.4(6)
C(14)-N(2)-C(15)-C(16)	112.5(7)
C(11)-N(2)-C(15)-C(20)	102.0(5)
C(14)-N(2)-C(15)-C(20)	-69.1(7)
C(20)-C(15)-C(16)-C(17)	-0.2(6)

N(2)-C(15)-C(16)-C(17)	178.2(4)
C(20)-C(15)-C(16)-C(22)	179.9(4)
N(2)-C(15)-C(16)-C(22)	-1.7(6)
C(15)-C(16)-C(17)-C(18)	-0.6(7)
C(22)-C(16)-C(17)-C(18)	179.2(5)
C(16)-C(17)-C(18)-O(3)	-179.5(4)
C(16)-C(17)-C(18)-C(19)	1.8(7)
C(23)-O(3)-C(18)-C(17)	5.1(6)
C(23)-O(3)-C(18)-C(19)	-176.2(4)
C(17)-C(18)-C(19)-C(20)	-2.2(6)
O(3)-C(18)-C(19)-C(20)	179.0(4)
C(18)-C(19)-C(20)-C(15)	1.3(6)
C(18)-C(19)-C(20)-C(21)	178.2(4)
C(16)-C(15)-C(20)-C(19)	-0.1(6)
N(2)-C(15)-C(20)-C(19)	-178.5(4)
C(16)-C(15)-C(20)-C(21)	-176.8(4)
N(2)-C(15)-C(20)-C(21)	4.8(6)

Symmetry transformations used to generate equivalent atoms:

Table S21. Hydrogen bonds for C₂₃H₂₅BrN₂O₃ [Å and °].

D-H...A	d(D-H)	d(H...A)	d(D...A)	<(DHA)
N(1)-H(1N)...O(1)#1	0.93(5)	2.03(5)	2.928(4)	160(4)

Symmetry transformations used to generate equivalent atoms:

#1 -x+1/2,y-1/2,-z+3/2