DNA-Compatible Copper-Catalyzed Oxidative Amidation of Aldehydes with Non-Nucleophilic Arylamines

Ke Li^{†§}, Yi Qu^{†§}, Yulong An[†], Eric Breinlinger[‡], Matthew P. Webster^ψ, Huanan Wen[†], Duanchen Ding[†], Meng Zhao[†], Xiaodong Shi[†], Jiangong Wang[†], Wenji Su[†], Weiren Cui[†], Alexander L. Satz[†], Hongfang Yang[†], Letian Kuai[†], Andrew Little^{‡*} and Xuanjia Peng^{†*}

[†]WuXiAppTec (Shanghai) Co., Ltd. 288 Middle Fu Te Road, Shanghai 200131, China
 [‡]AbbVie Bioresearch Center, 381 Plantation Street, Worcester, MA 01605, United States
 ^ψResearch and Development, AbbVie, 1 North Waukegan Road, North Chicago, Illinois 60064, United States

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SI-1 General Experimental

Dimethylsulfoxide (DMSO), 1-methyl-2-pyrrolidinone (NMP), *N*,*N*-dimethylacetamide (DMAc), acetonitrile (CH₃CN), and EtOH were purchased from Sigma-Aldrich. EDCI (CAS: 25952-53-8), *N*-hydroxysulfosuccinimide sodium salt (*s*-NHS, CAS: 106627-54-7), *tert*-butyl-hydroperoxide (TBHP, 70% in water, 75-91-2), and NaCl were purchased from TCI. The H₂O used was obtained by passing through the Milli-Q Direct. The reaction buffers were purchased from Vazyme and the ligase and ligation buffer were purchased from Thermo. The other reagents were purchased from domestic vendors unless mentioned otherwise. On-DNA reaction conversions were determined by UV traces of LC/MS analysis. The centrifuge instruments were Allegra X-15R, eppendorf-5424R. The peak at 0.27 min sometimes seen in the provided LC/MS traces is attributed to inorganic salts and small molecules contaminants, according to our experience.

SI-2 S-HP (Spacer-headpiece) Material



S-HP is composed of headpiece and PEG4 linker.

SI-3 General Procedure

SI-3-1 Ethanol Precipitation for DNA Compatible Reactions

To the reaction mixture was added 10% (v/v) 5 M NaCl aqueous solutions and 2.5-3 fold the volume of cold ethanol (stored at -20 $^{\circ}$ C before use). The colloidal solution was then allowed to sit at -80 $^{\circ}$ C for more than 2 hours. The solutions were centrifuged at 4 $^{\circ}$ C for 30 min at 4000 g. The supernatants were discarded, and the DNA pellet was dried at 30 $^{\circ}$ C for 1 hour in vacuo. Generally, ethanol precipitation was performed after each chemical reaction.

SI-3-2 General Procedure 1 for DNA conjugated aryl aldehydes



To a 15 mL tube was added a solution of EDCI (200 mM in DMSO, 125 μ L, 50 equiv), *s*-NHS (200 mM in DMSO/H₂O=1/1, 75 μ L, 30 equiv) and **S1** (200 mM in DMAc, 150 μ L, 60 equiv). This solution was mixed and left to stand at 20 °C for 15 min to make the active ester.

Next, S-HP solution (1 mM in pH 9.5 sodium borate buffer (250 mM), 500 μ L, 1 equiv) was added to the freshly prepared active ester solution. The solution was mixed and left to stand at 25 °C for 16 h (pH 9), with reaction conversion monitored by LCMS. The reaction mixture was then precipitated with ethanol (see General Procedure **SI-3-1**).





The DNA-linked material **S-HP-1** (500 nmol) was dissolved in sodium borate buffer (pH 9.45, 250 mM, 500 μ L) to make a 1 mM solution. To 4,4-dimethoxybutanoic acid methyl ester (200 mM in DMAc, 106.25 μ L, 42.5 equiv) was added NaOH (200 mM in H₂O, 106.25 μ L, 42.5 equiv); the solution was mixed and then stood at 60 °C for 2 h. To the DNA material **S-HP-1** was added the above solution and DMT-MM (200 mM in H₂O, 100 μ L, 40 equiv). The solution was mixed and left to stand at 20 °C for another 3 h (pH 9) to make the precursor for **1q** in situ. The reaction conversion monitored by LCMS. The reaction mixture was then precipitated with ethanol (see General Procedure **SI-3-1**).

Next, the material was redissoved in H₂O (500 μ L) and freshly prepared acetic acid solution (300 Mm in H₂O, 166 μ L) was added. The solution was mixed and left to stand at 50 °C for 1 h (pH 9), with reaction conversion monitored by LCMS. The reaction mixture was then precipitated with ethanol (see General Procedure **SI-3-1**).

SI-3-4 General Procedure 3 for Oxidative Amidation.



To the DNA-linked material **1** (10 nmol, 1 mM in H₂O, 10 μ L, 1 equiv) was added aniline **2** (400.0 mM in CH₃CN, 5 μ L, 200 equiv), CuI (50.0 mM in CH₃CN, 16 μ L, 80 equiv) and TBHP (200 mM in H₂O, 5 μ L, 100 equiv). The solution was mixed at 25 °C for 16 h in a shaker (pH 6~7). Next, to the solution was added ethyldithiocarbamate trihydrate (200.0 mM in H₂O, 12 μ L, 240.0 equiv). The solution was mixed at 80 °C for 30 min, with reaction conversion monitored by LCMS. The reaction mixture was then precipitated with ethanol (see General Procedure **SI-3-1**).

SI-3-5 Procedure for Compound 11.



To the DNA starting material **10** (946 nmol, 1 mM in H₂O, 946 μ L, 1 equiv) was added FeSO₄.7H₂O (200.0 mM in H₂O, 378.4 μ L, 80.0 equiv), and NaOH (2500.0 mM in H₂O, 189.2 μ L, 500.0 equiv). The solutions was mixed at 80 °C for 1 h in an ultrasonic unit (pH 12). Next, to the solution was added ethyl-dithiocarbamate trihydrate (100.0 mM in H₂O, 567.6 μ L, 60 equiv). The solution was mixed and left to stand at 80 °C for 30 min, with reaction conversion monitored by LCMS. The reaction mixture was then precipitated with ethanol (see General Procedure **SI-3-1**). The crude product was purified by reverse-phase HPLC to give 167.8 nmol of desired product **11**. Yield (17.7%).



The DNA starting material **11** (5 nmol) was dissolved in MOPS buffer (pH 8.2, 50 mM, 5 μ L). 4-(*N*,*N*-Dimethylcarbamimidoyl)benzoic acid (400.0 mM in H₂O, 2.5 μ L, 200.0 equiv) and DMT-MM.BF₄ (400.0 mM in DMSO, 2.5 μ L, 200 equiv) were then added. After the addition, the tube was centrifuged, eddied and re-centrifuged. The reaction mixture was kept at 25 °C for 16 h (pH 8). After EtOH precipitation, H₂O (5 μ L) and 20% piperidine (5 μ L) were added. The tube was centrifuged, eddied and re-centrifuged. The solution was mixed and left to stand at 25 °C for 2 h, with reaction conversion monitored by LCMS. The reaction mixture was then precipitated with ethanol (see General Procedure **SI-3-1**).





The DNA starting material **11** (5 nmol) was dissolved in phosphoric buffer (pH 5.5, 250 mM, 5 μ L). 4-(1*H*-Tetrazol-5-yl) benzaldehyde (200.0 mM in DMAc, 3 μ L, 20.0 equiv) and NaBH₃CN (400.0 mM in DMAc, 1.5 μ L, 120 equiv) were then added. After the addition, the tube was centrifuged, eddied and recentrifuged. The solution was mixed and left to stand at 20 °C for 16 h (pH 6), with reaction conversion monitored by LCMS. The reaction mixture was then precipitated with ethanol (see General Procedure **SI-3-1**).

SI-3-8 General Procedure 7 for Enzymatic Tagging

To DNA-linked starting materials (100 nmol, 1 mM in H₂O, 100 μ L, 1 equiv) were added DNA tags (160 nmol, 1 mM in H₂O, 160 μ L, 1.6 equiv), 10x T4 DNA ligation buffer (40 μ L), T4 DNA ligase (4 μ L, 30U/ μ L) and H₂O (96 μ L). The solutions were mixed and stirred at 16 °C for 16 h. The reaction mixtures were precipitated with ethanol, according to General Procedure **SI-3-1**, and the samples were analyzed by 4% gel picture.

SI-4 DNA Damage Evaluation

Copper-mediated oxidative amidation of aldehyde reactions (abbreviated as "the oxidative amidation reaction" below) were performed with DNA conjugated compound with a double stranded DNA coding region to mimic the library component. The DNA conjugated compound was subjected to the oxidative amidation reaction under different conditions (Table S1) to determine DNA damage.

| ID | Condition No. | Condition Info. | Theoretical In- put |
|----|---------------|---------------------|------------------------|
| A1 | 1 | Normal reaction | 5 nmol |
| A2 | 2 | No CuI/TBHP | 5 nmol |
| A3 | 3 | No CuI | 5 nmol |
| A4 | 4 | No TBHP | 5 nmol |
| A5 | 5^a | formaldehyde(PC1) | 5 nmol |
| A6 | 6^b | No reaction(NC) | 5 nmol |
| A7 | 7^c | No reaction (Ab NC) | 5 nmol |

Table S1, sampling logic of the comparison.

Note: ^a Compounds in this group act as positive control.^{[1] b} Compounds in this group act as negative control and go through the basic liquid transfer processes in the oxidative amidation reaction, but no reagent or material was added. ^c Compounds in this group don't go through the oxidative amidation reaction and are kept as an absolute negative control.

[1] Huang, H., Hopkins, P. B. (1993) DNA interstrand cross-linking by formaldehyde: nucleotide sequence preference and covalent struc-ture of the predominant cross-link formed in synthetic oligonucleo-tides. *J. Am. Chem. Soc. 115*, 9402-9408.

The products of each reaction were visualized by an Agilent 2100 Bioanalyzer (Fig S1), then ligated to an oligonucleotide to generate a full-length DNA fragment, similar to DEL molecules. If there is DNA damage in the oxidative amidation reaction, the band of the experimental group may become vague or turn into a smear compared with the negative control. The shape of the band from the experimental group A1 is similar to the negative group A7, while the band of positive control of the formaldehyde treated group A5 is disappearing. These results indicate that normal reaction conditions will not affect the DNA as observed in condition A5. As the band of A2 to A4 are quite similar, it is clear that the different reagents in the oxidative amidation reaction will not affect the DNA. For subsequent qPCR analysis, only the groups of A1 (oxidative amidation reaction group), A5 (formaldehyde treated, positive control) and A7 (negative control) were used.



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Figure S1. Gel image of different comparison groups. Lane 1 to 7 is the condition A1 to A7.

SI-4-1 QPCR Test

After EtOH Precipitation, according to General Procedure **SI-3-1**, the ligation products of A1 (oxidative amidation), A5 (formaldehyde treated) and A7 (negative control) were step-wise diluted 10 fold to a final 1e4 fold dilution. The serial dilutions were used as templates for qPCR testing using a SYBR Green Master Mix kit (Thermo) and a Real-Time PCR System (QuantStudio 7 Flex) (Table S2). All samples were run in triplicate and subjected to PCR cycles as follows: 95 °C heat activation for 5 min followed by 40 cycles of 95 °C denaturation for 10 seconds, 55 °C annealing for 15 seconds and extension at 72 °C for 30 seconds.



Table S2. Summary of qPCR Data Analysis.

The results showed that no obvious concentration difference was observed between oxidative amidation and no reaction, but the concentration of formaldehyde at the same dilution ratio is significantly lower. The R^2 value are all above 0.99 in these three groups. This indicates that the oxidative amidation reaction did not affect the detectability of DNA tags.

SI-4-2 Next-generation sequencing.

1e7 copies of the three groups mentioned above were used as template for PCR amplification. To a PCR tube was added diluted sample (2 μ L), 10x high fidelity PCR buffer (5 μ L), 50 mM MgSO₄ (2 μ L), 10 mM dNTP mix (1 μ L), PlatinumTM Taq DNA Polymerase (0.2 μ L), 10 μ M forward primer (2 μ L), 10 μ M reverse primer (2 μ L), and nuclease-free water (35.8 μ L). After 20 cycles, the PCR products (204 bp, forward primer brought in additional 20 bp at 5' terminal, reverse primer brought in additional 66 bp at 3' terminal) were purified by the Agencourt AMPure XP Beads method. The purified samples were quantified by Qubit 4.0 to appear at 1.2 ng/μL. PCR products were diluted to 3nM and pooled together according to the ratio of PCR input copies, then provided for next-generation sequencing (Illumina HiSeq XTen). Bowtie2 were used to mapping the sequenced reads to reference by local alignment. The detailed mapping identity were extracted from CIGAR string and XM flag in the SAM format. The results of NGS showed that all samples retained the right sequence as expected (Fig. S2), indicating that the chemical reactions did not affect the encodability of DNA tags.



Figure S2. Statistics of next-generation sequencing results. The left Y-axis is the ratio of perfect match read count normalized by the negative control, while the right Y-axis is the fraction of 1bp mismatch.

In conclusion, our data revealed that the oxidative amidation reaction used in this paper caused no damage to DNA, and thus could be used for the encoded library construction.

SI-5 Selected examples of the active molecule





To investigate the molecules that could potentially be synthesized with the oxidative amidation reaction, the library of CHEMBL_25.sdf was searched through. 11921 (0.64% of the total) pyridin-2-yl analogous amides and 28517 (1.52% of the total) pyrrol-2-yl analogous amides were found, among which 283 molecules were counted in both of the categories. The total coverage of molecules that could be synthesized with the newly developed method is 2.15%. (CHEMBL database (CHEMBL 25) accessed on 2020. Feb. 26th under https://www.ebi.ac.uk/chembl/)

SI-6 LC Trace and Mass

10-

0-

700

[M-7]/7

776.3

800

850.8

900

1000

SI-6-1 LC Trace and Mass of 3a

Following **General Procedure 3** Yield: 92.31% Exact mass: 5438.56

Triply charged mass [M-3]/3, calculated: 1811.9; observed: 1812.1.





1300

m/z

1400

1500

1700

1800

1900

1600

1119.6

1100

1200

S14

2000

S15

SI-6-2 LC Trace and Mass of 3b

Yield: >99% Exact mass: 5623.77 Triply charged mass [M-3]/3, calculated: 1873.6; observed: 1873.4.



Figure S5. LC trace and mass of 3b

SI-6-3 LC Trace and Mass of 3c

Yield: 77.13% Exact mass: 5675.77 Triply charged mass [M-3]/3, calculated: 1890.9; observed: 1891.2.



Figure S6. LC trace and mass of 3c

SI-6-4 LC Trace and Mass of 3d

Following **General Procedure 3** Yield: 69.87% Exact mass: 5597.71 Triply charged mass [M-3]/3, calculated: 1864.9; observed: 1865.2.



Figure S7. LC trace and mass of 3d

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SI-6-5 LC Trace and Mass of 3e

Following **General Procedure 3** Yield: 99.12% Exact mass: 5593.8 Triply charged mass [M-3]/3, calculated: 1863.6; observed: 1863.8.







Figure S8. LC trace and mass of 3e

SI-6-6 LC Trace and Mass of 3f

Following General Procedure 3

Yield: 88.75% Exact mass: 5579.72

Triply charged mass [M-3]/3, calculated: 1858.9; observed: 1858.5.



Figure S9. LC trace and mass of 3f

SI-6-7 LC Trace and Mass of 3g

Following **General Procedure 3** Yield: 80.94% Exact mass: 5514.85 Triply charged mass [M-3]/3, calculated: 1837.3; observed: 1837.4.



Figure S10. LC trace and mass of 3g

SI-6-8 LC Trace and Mass of 3h

Following **General Procedure 3** Yield: 66.01% Exact mass: 5496.83 Triply charged mass [M-3]/3, calculated: 1831.3; observed: 1831.2.





SI-6-9 LC Trace and Mass of 3i

Following **General Procedure 3** Yield: 74.54% Exact mass: 5438.75

Triply charged mass [M-3]/3, calculated: 1811.9; observed: 1812.2.



Figure S12. LC trace and mass of 3i

SI-6-10 LC Trace and Mass of 3j

Following **General Procedure 3** Yield: 79.33% Exact mass: 5408.72 Triply charged mass [M-3]/3, calculated: 1801.9; observed: 1802.1.



Figure S13. LC trace and mass of 3j

SI-6-11 LC Trace and Mass of 3k

Following **General Procedure 3** Yield: 74.7% Exact mass: 5422.75 Triply charged mass [M-3]/3, calculated: 1806.6; observed: 1806.9.





Figure S14. LC trace and mass of 3k

SI-6-12 LC Trace and Mass of 31

Following **General Procedure 3** Yield: 48.70% Exact mass: 5498.85 Triply charged mass [M-3]/3, calculated: 1832.0; observed: 1832.2.



Figure S15. LC trace and mass of 31

SI-6-13 LC Trace and Mass of 3m

Following **General Procedure 3** Yield: 51.06% Exact mass: 5485.81 Triply charged mass [M-3]/3, calculated: 1827.6; observed: 1827.4.





SI-6-14 LC Trace and Mass of 3n

Following **General Procedure 3** Yield: 85.95% Exact mass: 5555.7 Triply charged mass [M-3]/3, calculated: 1850.9; observed: 1851.3.



Figure S17. LC trace and mass of 3n

SI-6-15 LC Trace and Mass of 3o

Following **General Procedure 3** Yield: 17.14% Exact mass: 5448.75 Triply charged mass [M-3]/3, calculated: 1815.3; observed: 1815.0.



Figure S18. LC trace and mass of 30

SI-6-16 LC Trace and Mass of 3p

Following **General Procedure 3** Yield: 76.37% Exact mass: 5448.74 Triply charged mass [M-3]/3, calculated: 1815.3; observed: 1815.6.



Figure S19. LC trace and mass of 3p

m/z

SI-6-17 LC Trace and Mass of 4b

Following **General Procedure 3** Yield: 92.75% Exact mass: 5547.89 Triply charged mass [M-3]/3, calculated: 1848.3; observed: 1848.3.





Figure S20. LC trace and mass of 4b

SI-6-18 LC Trace and Mass of 4c

Following **General Procedure 3** Yield: 42.26% Exact mass: 5506.75 Triply charged mass [M-3]/3, calculated: 1834.6; observed: 1834.5.





Figure S21. LC trace and mass of 4c

SI-6-19 LC Trace and Mass of 4d

Yield: 90.67% Exact mass: 5456.75 Triply charged mass [M-3]/3, calculated: 1817.9; observed: 1818.2.



Figure S22. LC trace and mass of the mixture of 4d

SI-6-20 LC Trace and Mass of 4e

Following **General Procedure 3** Yield: 80.83% Exact mass: 5452.78 Triply charged mass [M-3]/3, calculated: 1816.6; observed: 1816.3.



Figure S23. LC trace and mass of 4e

SI-6-21 LC Trace and Mass of 4f

Following **General Procedure 3** Yield: 51.46% Exact mass: 5517.65 Triply charged mass [M-3]/3, calculated: 1838.2; observed: 1838.4.



Figure S24. LC trace and mass of 4f

SI-6-22 LC Trace and Mass of 4g

Following **General Procedure 3** Yield: 88.49% Exact mass: 5509.83 Triply charged mass [M-3]/3, calculated: 1835.6; observed: 1835.9.



Figure S25. LC trace and mass of 4g

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SI-6-23 LC Trace and Mass of 4h

Following **General Procedure 3** Yield: 77.95% Exact mass: 5622.98 Triply charged mass [M-3]/3, calculated: 1873.3; observed: 1873.7.





Figure S26. LC trace and mass of 4h

SI-6-24 LC Trace and Mass of 4i

RT: 0.00 - 3.00

100-

90

Following General Procedure 3 Yield: 39.03% Exact mass: 5622.01 Triply charged mass [M-3]/3, calculated: 1873; observed: 1873.3.







Figure S27. LC trace and mass of 4i

SI-6-25 LC Trace and Mass of 4j

Following **General Procedure 3** Yield: 80.82% Exact mass: 5497.82 Triply charged mass [M-3]/3, calculated: 1831.6; observed: 1832.0.



Figure S28. LC trace and mass of 4j

SI-6-26 LC Trace and Mass of 4k

Following **General Procedure 3** Yield: 71.01% Exact mass: 5439.74 Triply charged mass [M-3]/3, calculated: 1812.3; observed: 1811.9.





Figure S29. LC trace and mass of 4k

SI-6-27 LC Trace and Mass of 41

Following **General Procedure 3** Yield: 67.64% Exact mass: 5489.8 Triply charged mass [M-3]/3, calculated: 1828.9; observed: 1829.1.



Figure S30. LC trace and mass of 41

SI-6-28 LC Trace and Mass of 4m

Following **General Procedure 3** Yield: 34.39% Exact mass: 5483.8 Triply charged mass [M-3]/3, calculated: 1826.9; observed: 1826.8.





SI-6-29 LC Trace and Mass of 4n

Following **General Procedure 3** Yield: 51.95% Exact mass: 5570.72 Triply charged mass [M-3]/3, calculated: 1855.9; observed: 1855.5.



Figure S32. LC trace and mass of 4n

SI-6-30 LC Trace and Mass of 40

Following **General Procedure 3** Yield: 35.27% Exact mass: 5529.89 Triply charged mass [M-3]/3, calculated: 1842.3; observed: 1842.4.



Figure S33. LC trace and mass of 40

SI-6-31 LC Trace and Mass of 4p

Following **General Procedure 3** Yield: 76.37% Exact mass: 5596.76 Triply charged mass [M-3]/3, calculated: 1864.6; observed: 1864.8.



Figure S34. LC trace and mass of 4p

1300

m/z

1400

1151.4

1200

1100

958.8

1000

900

0-

700

800

2000

1700

1600

1500

1800

1900

SI-6-32 LC Trace and Mass of 4q

Following **General Procedure 3** Yield: 64.7% Exact mass: 5570.72

Triply charged mass [M-3]/3, calculated: 1855.9; observed: 1856.0.



Figure S35. LC trace and mass of 4q

SI-6-33 LC Trace and Mass of 4r

Following **General Procedure 3** Yield: 46.72% Exact mass: 5557.68 Triply charged mass [M-3]/3, calculated: 1851.6; observed: 1851.7.





SI-6-34 LC Trace and Mass of 1q

Following **General Procedure 2** Yield: 89.23% Exact mass: 5282.6 Triply charged mass [M-3]/3, calculated: 1759.9; observed: 1760.3.



Figure S37. LC trace and mass of 1q

SI-6-35 LC Trace and Mass of 5a

Following **General Procedure 3** Yield: 55.91 % Exact mass: 5374.67 Triply charged mass [M-3]/3, calculated: 1790.6; observed: 1790.9.





SI-6-36 LC Trace and Mass of 5p

Following **General Procedure 3** Yield: 80.97% Exact mass: 5532.69 Triply charged mass [M-3]/3, calculated: 1843.2; observed: 1843.9.



Figure S39. LC trace and mass of 5p

SI-6-37 LC Trace and Mass of 5q

Following **General Procedure 3** Yield: 59.97% Exact mass: 5506.65

Triply charged mass [M-3]/3, calculated: 1834.6; observed: 1834.6.





Figure S40. LC trace and mass of 5q

SI-6-38 LC Trace and Mass of 5s

Following **General Procedure 3** Yield: 54.07% Exact mass: 5520.67 Triply charged mass [M-3]/3, calculated: 1839.2; observed: 1839.9.



Figure S41. LC trace and mass of 5s

SI-6-39 LC Trace and Mass of 5t

Following **General Procedure 3** Yield: 53.04% Exact mass: 5456.59 Triply charged mass [M-3]/3, calculated: 1817.9; observed: 1818.0.



Figure S42. LC trace and mass of 5t

SI-6-40 LC Trace and Mass of 5u

Following **General Procedure 3** Yield: 42.77% Exact mass: 5492.62 Triply charged mass [M-3]/3, calculated: 1829.87; observed: 1830.2.



Figure S43. LC trace and mass of 5u

SI-6-41 LC Trace and Mass of 5v

Following **General Procedure 3** Yield: 38.71% Exact mass: 5539.62 Triply charged mass [M-3]/3, calculated: 1845.5; observed: 1845.6.



Figure S44. LC trace and mass of 5v

SI-6-42 LC Trace and Mass of 5w

Following **General Procedure 3** Yield: 34.93% Exact mass: 5493.6 Triply charged mass [M-3]/3, calculated: 1830.2; observed: 1829.9.



Figure S45. LC trace and mass of 5w

SI-6-43 LC Trace and Mass of 5x

Following **General Procedure 3** Yield: 32.22% Exact mass: 5483.61 Triply charged mass [M-3]/3, calculated: 1826.8; observed: 1826.2.



Figure S46. LC trace and mass of 5x

SI-6-44 LC Trace and Mass of 5y

Following **General Procedure 3** Yield: 43.69% Exact mass: 5467.61 Triply charged mass [M-3]/3, calculated: 1821.5; observed: 1821.8.



Figure S47. LC trace and mass of 5y

SI-6-45 LC Trace and Mass of 5z

Following **General Procedure 3** Yield: 67.24% Exact mass: 5503.64 Triply charged mass [M-3]/3, calculated: 1833.6; observed: 1833.3.







SI-6-46 LC Trace and Mass of 5aa

Following **General Procedure 3** Yield: 65.03% Exact mass: 5503.64 Triply charged mass [M-3]/3, calculated: 1833.6; observed: 1833.7.





SI-6-47 LC Trace and Mass of 9

Following **General Procedure 1** Yield: 89.75% Exact mass: 5419.69 Triply charged mass [M-3]/3, calculated: 1805.6; observed: 1805.5.





Figure S50. LC trace and mass of 9

SI-6-48 LC Trace and Mass of 10

Following **General Procedure 3** Yield: 79.02% Exact mass: 5546.25 Triply charged mass [M-3]/3, calculated: 1847.8; observed: 1848.1.



Figure S51. LC trace and mass of 10

SI-6-49 LC Trace and Mass of 11

Yield: 51.36% Exact mass: 5516.25 Triply charged mass [M-3]/3, calculated: 1837.8; observed: 1837.5.



Figure S52. LC trace and mass of crude 11



Figure S53. LC trace and mass of 11 after prep-HPLC

SI-6-50 LC Trace and Mass of 12

Yield: 83.16% Exact mass: 5690.46 Triply charged mass [M-3]/3, calculated: 1895.8; observed: 1895.9.





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SI-6-51 LC Trace and Mass of 13

Yield: 70.44% Exact mass: 5674.41 Triply charged mass [M-3]/3, calculated: 1890.5; observed: 1890.5.



Figure S55. LC trace and mass of 13