# Elemental Sulfur-Incorporated Cyclizations of Pyrrolidines 

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Table S1. Optimization of the reaction conditions. ${ }^{a}$


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## 2. EPR experiment procedure for interaction of 4-benzyl-2-phenyl-5,

 6-dihydro-4H-thieno[3,2-b]pyrrole (1a) with elemental sulfur.1-benzyl-2-(phenylethynyl)pyrrolidine ( $78.4 \mathrm{mg}, 0.30 \mathrm{mmol}$ ) and elemental sulfur $\mathbf{S}_{8}(115.2 \mathrm{mg}, 0.45 \mathrm{mmol})$ were combined in a 50 mL flame-dried Young-type tube equipped with a stir bar, and then the tube was sealed. Next, the Schlenk tube was purged three times with $\mathrm{N}_{2}$. Then, 1,4 -dioxane ( $(.0 \mathrm{~mL}$ ) was injected into the Schlenk tube with a syringe under $\mathrm{N}_{2}$ atmosphere. The contents of the Schlenk tube were then allowed to stir at $120^{\circ} \mathrm{C}$ by using a heating mantle for 1.0 h . Then, DMPO was added to the mixture and preserved in liquid nitrogen for EPR examination. No organic radical was observed.

## 3. ${ }^{1} \mathrm{H}$ NMR and ${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR copies of substrates

${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ )


${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$


${ }^{1} \mathrm{H} \operatorname{NMR}\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$





${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$


${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$


${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR ( $101 \mathrm{MHz}, \mathrm{CDCl}_{3}$ )







## ${ }^{1} \mathrm{H}$ NMR $\left(600 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$






${ }^{1} \mathrm{H}$ NMR $\left(600 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$


${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}\right.$, DMSO- $d_{6}$ )


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${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$


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## ${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ )



${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR ( $101 \mathrm{MHz}, \mathrm{CDCl}_{3}$ )



${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$






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${ }^{1} \mathrm{H} \operatorname{NMR}\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$

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${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ )




${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$



${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$




${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$



${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$




${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$


${ }^{1} \mathrm{H}$ NMR $\left(600 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$


${ }^{1} \mathrm{H} \operatorname{NMR}\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$




${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$










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| 80 | 170 | 160 | 150 | 140 | ${ }_{130}$ | ${ }_{120}$ | ${ }_{110}$ | ${ }_{100}$ | ${ }_{90}$ | 80 | 70 | 60 | 50 | 40 | 30 | 20 | 10 | ${ }_{0}$ |

${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$




| 80 | 170 | 160 | 150 | 140 | 130 | 120 | 110 | 100 | 90 | 80 | 70 | 60 | 50 | 10 | 30 | 20 | 10 |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |

${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ )


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${ }^{1} \mathrm{H} \operatorname{NMR}\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$




${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ )



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${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR ( $101 \mathrm{MHz}, \mathrm{CDCl}_{3}$ )







${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$





${ }^{1} \mathrm{H}$ NMR $\left(600 \mathrm{MHz}\right.$, DMSO- $\left.d_{6}\right)$




${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR $\left(101 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$



${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$


${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR $\left(101 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$



## 4. ${ }^{1} \mathrm{H}$ NMR and ${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR copies of products



${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$








${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$


${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR ( $101 \mathrm{MHz}, \mathrm{CDCl}_{3}$ )



${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR $\left(101 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$







${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$



##  

${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$


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${ }^{3} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR $\left(101 \mathrm{MHz} \mathrm{CDCl}_{3}\right)$

(376 MHz, $\left.\mathrm{CDCl}_{3}\right)$
${ }^{1} \mathrm{H} \operatorname{NMR}\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$


${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\} \mathrm{NMR}\left(101 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$


















${ }^{1} \mathrm{H} \operatorname{NMR}\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$





${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$














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${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$


$\left.{ }^{13} \mathrm{C}^{1}{ }^{1} \mathrm{H}\right\} \mathrm{NMR}\left(101 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$





${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$







${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR $\left(101 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$

${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$



${ }^{1} \mathrm{H}$ NMR ( $400 \mathrm{MHz}, \mathrm{CDCl}_{3}$ )


${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR (101 MHz, $\left.\mathrm{CDCl}_{3}\right)$ (





${ }^{1} \mathrm{H}$ NMR $\left(400 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$



${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR $\left(151 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$

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${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR $\left(151 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$



${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR $\left(151 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$






${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\}$ NMR ( $151 \mathrm{MHz}, \mathrm{CDCl}_{3}$ )


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| ${ }^{1} \mathrm{H}$ NMR（ $600 \mathrm{MHz}, \mathrm{CDCl}_{3}$ ） |  |  |  |



${ }^{1} \mathrm{H}$ NMR $\left(600 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$

${ }^{13} \mathrm{C}\left\{{ }^{1} \mathrm{H}\right\} \mathrm{NMR}\left(151 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$

${ }^{1} \mathrm{H}$ NMR ( $600 \mathrm{MHz}, \mathrm{CDCl}_{3}$ )


$\left.{ }^{13} \mathrm{C}_{\{ }{ }^{1} \mathrm{H}\right\} \operatorname{NMR}\left(101 \mathrm{MHz}, \mathrm{CDCl}_{3}\right)$


## 5. Single-Crystal X-ray diffraction.

Single-crystal XRD studies on compounds $\mathbf{2 b}$ and $\mathbf{2 h}$ were performed on a Supernova CCD diffractometer at 293(2) K. Determination of unit cell parameters and data collection were performed with Mo-Ka radiation at a wavelength of $0.71073 \AA$ using the x -scan technique. The structures were solved by direct methods and refined by full matrix least-squares on $F^{2}$ using SHELXS-97 and SHELXL-97 programs. ${ }^{1}$ The metal atoms in each compound were located from the E-maps, and other non-hydrogen atoms were located in successive difference Fourier syntheses and refined with anisotropic thermal parameters on $F^{2}$. The hydrogen atoms were added theoretically, riding on the concerned atoms and refined with fixed thermal factors. The SQUEEZE function in PLATON was utilized during the refinement of $\mathbf{2 b}$ and $\mathbf{2 h}$ owing to the disordered solvents. ${ }^{2}$

The structure was then refined again using the data generated. Crystal data and details of the data collection are given in Table S1-S2. CCDC 1946767 (2b) and 1946766 (2h) contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre.
(1) M. Sheldrick, G. SHELXS-97. Program for X-ray crystal structure determination, Gottingen University, Germany, 1997.
(2) L. Spek, A. Single-crystal structure validation with the program PLATON. J. Appl. Crystallogr. 2003, 36, 7-13.

Single crystals (2b and 2h) suitable for X-ray analysis were obtained by slow evaporation of $\mathrm{CH}_{2} \mathrm{Cl}_{2}$ solvent.
Table S1. Crystal data and structure refinement for 2b

| Identification code | shs-20180927 |
| :--- | :--- |
| Empirical formula | $\mathrm{C}_{20} \mathrm{H}_{17} \mathrm{NS}$ |
| Formula weight | 303.41 |
| Temperature/K | $293(2) \mathrm{K}$ |
| Crystal system | monoclinic |
| Space group | $\mathrm{P} 121 / \mathrm{c} 1$ |
| $\mathrm{a} / \AA$ | $13.4137(6) \mathrm{A}$ |


| b/Å | 5.9329(2) A |
| :---: | :---: |
| c/Å | 20.1416(8) A |
| $\alpha /{ }^{\circ}$ | 90 |
| $\beta /{ }^{\circ}$ | 90.210(4) |
| $\gamma^{/ 0}$ | 90 |
| Volume/ $/{ }^{\text { }}$ 3 | 1602.90(11) |
| Z | 4 |
| $\rho_{\text {calc }} / \mathrm{cm}^{3}$ | 1.257 |
| $\mu / \mathrm{mm}^{-1}$ | 0.198 |
| $\mathrm{F}(000)$ | 640 |
| Crystal size/ $\mathrm{mm}^{3}$ | $0.45 \times 0.28 \times 0.11$ |
| Radiation | $\operatorname{MoK} \alpha(\lambda=0.71073)$ |
| $2 \Theta$ range for data collection/ ${ }^{\circ}$ | 3.58 to 29.21 |
| Index ranges | $-17<=\mathrm{h}<=16,-8<=\mathrm{k}<=6,-26<=1<=25$ |
| Reflections collected | 8879 / 3702 [ $\mathrm{R}(\mathrm{int})=0.0224]$ |
| Independent reflections | unique 3702 |
| Data/restraints/parameters | 3702 / 0 / 200 |
| Goodness-of-fit on $\mathrm{F}^{2}$ | 1.038 |
| Final R indexes [ $\mathrm{I}>=2 \sigma$ ( I$)$ ] | $\mathrm{R} 1=0.0439, \mathrm{wR} 2=0.0992$ |
| Final R indexes [all data] | $\mathrm{R} 1=0.0604, \mathrm{wR} 2=0.1081$ |
| Largest diff. peak/hole / e $\AA^{-3}$ | 0.218 and -0.236 |



Figure S1. Additional X-ray crystallographic structures 2b with $30 \%$ probability ellipsoid.

Table S2. Crystal data and structure refinement for 2h

| Identification code | shs-2-20180919 |
| :--- | :--- |
| Empirical formula | $\mathrm{C}_{21} \mathrm{H}_{19} \mathrm{NOS}$ |
| Formula weight | 333.43 |
| Temperature/K | $293(2) \mathrm{K}$ |


| Crystal system | monoclinic |
| :---: | :---: |
| Space group | C1c1 |
| a/Å | 28.9068(5) A |
| b/Å | 5.75310(10) A |
| c/Å | 10.5484(2) A |
| $\alpha /{ }^{\circ}$ | 90 |
| $\beta /{ }^{\circ}$ | 98.531(2) |
| $\gamma /{ }^{\circ}$ | 90 |
| Volume/A ${ }^{3}$ | 1734.83(5) |
| Z | 4 |
| $\rho_{\text {calc }} / \mathrm{cm}^{3}$ | 1.277 |
| $\mu / \mathrm{mm}^{-1}$ | 1.694 |
| $\mathrm{F}(000)$ | 704 |
| Crystal size $/ \mathrm{mm}^{3}$ | $0.20 \times 0.20 \times 0.20$ |
| Radiation | $\operatorname{MoK} \alpha(\lambda=0.71073)$ |
| $2 \Theta$ range for data collection/ ${ }^{\circ}$ | 6.19 to 71.18 |
| Index ranges | $-34<=\mathrm{h}<=35,-5<=\mathrm{k}<=6,-12<=\mathrm{l}<=9$ |
| Reflections collected | $5226 / 2646$ [ $\mathrm{R}(\mathrm{int})=0.0166]$ |
| Independent reflections | unique 2646 |
| Data/restraints/parameters | 2646 / 2 / 218 |
| Goodness-of-fit on $\mathrm{F}^{2}$ | 1.079 |
| Final R indexes [ $\mathrm{l}>=2 \sigma$ (I)] | $\mathrm{R} 1=0.0363, \mathrm{wR} 2=0.1052$ |
| Final R indexes [all data] | $\mathrm{R} 1=0.0369, w R 2=0.1060$ |
| Largest diff. peak/hole / e $\AA^{-3}$ | 0.108 and -0.191 |



Figure S2. Additional X-ray crystallographic structures 2h with $30 \%$ probability ellipsoid.


[^0]:    ${ }^{a}$ Standard condition: 1a $(0.30 \mathrm{mmol}), \mathbf{S}_{\mathbf{8}}\left(1.5\right.$ equiv), solvent $(2.0 \mathrm{~mL}), \mathrm{N}_{2}, 120{ }^{\circ} \mathrm{C}, 24 \mathrm{~h}$.

