## **Supporting Information**

A versatile adeno-associated viral vector cross-linking platform capable of tuning cellular tropisms and simultaneously inducing solid-phase gene delivery *Seungju Yoo<sup>§,‡</sup>, Byunguk Kang<sup>§,†,‡</sup>, Seokmin Oh<sup>§</sup>, Yunha Kim<sup>§</sup>, Jae-Hyung Jang<sup>§,\*</sup>* <sup>§</sup>Department of Chemical and Biomolecular Engineering, Yonsei University, 50 Yonsei-ro, Seodaemun-gu, Seoul 03722, Korea <sup>†</sup>Department of Bioengineering, Rice University, 6100 Main Street, Houston, Texas, United States of America <sup>\*</sup> Corresponding Author (Email: j-jang@yonsei.ac.kr)

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**Figure S1**. The degree of the conjugation of A488 to AAV8 (A) and i488 to AAV8-DTSSP complexes (B). The A488 and i488 were utilized as alternative substances containing the same functional groups (i.e., sulfo-NHS and amine group) to those on the DTSSP and aminated-L-fucose molecules, respectively. The negative control (i.e., Neg.) indicates the condition without including fluorescence molecules (i.e., POROS beads only, beads+AAV, beads+AAV+DTSSP). The symbol \* indicates significant differences (P<0.05).



**Figure S2**. Investigation of delivery mechanisms. Cells (MIA PaCa-2) were treated with trypsin-EDTA (0.25%) and transduced by naïve AAV8 or AAV8-D-Fc vectors at an MOI of 20,000. The media was replaced with fresh media at 6 hours post-transduction (i.e., prior to the full recovery of the membrane-bound receptors), and the number of GFP expressing cells was quantified using flow cytometry at 48 hours post-infection. Symbols \* and # indicate the significant differences in the percentages of GFP-expressing cells (P<0.05).



**Figure S3**. Cellular transduction (HEK293T cells) by **(A)** AAV2-DTSSP complexes and **(B)** AAV8-DTSSP complexes displaying Tris molecules (i.e., AAV2-D-Tris and AAV8-D-Tris). Symbols \* and # indicate the significant differences in the percentages of GFP-expressing cells compared to the ones by the naïve AAV2 or 8 vectors.



**Figure S4.** Comparison of the morphologies of the AAV-DTSSP pellets and the AAV precipitates formed by ammonium sulfate.



Figure S5. Reversibility of AAV-DTSSP pellet formation. The AAV-DTSSP pellets were

resuspended by mild sonication and then centrifuged to reform the pellets (arrow).



**Figure S6** Cell viability of HEK293T cells after transducing with PBS, AAV2, AAV2-DTSSP-Fucose (AAV2-D-Fc) and AAV2-DTSSP (at final concentrations of 8.2, 16.4 and 32.8 mM DTSSP) at a MOI of 5000.