Supporting Information

Placental Trophoblast-Inspired Lipid Bilayers for Cell-Free Investigation of Molecular Interactions

Christina M. Bailey-Hytholt, ¹ Tun-Li Shen, ² Bonnee Nie, ³ Anubhav Tripathi, ¹ Anita Shukla ^{1*}

¹School of Engineering, Center for Biomedical Engineering, Brown University, Providence, RI 02912

²Department of Chemistry, Brown University, Providence, RI 02912

³Department of Biochemistry and Molecular Biology, Brown University, Providence, RI 02912

*Corresponding author: anita_shukla@brown.edu

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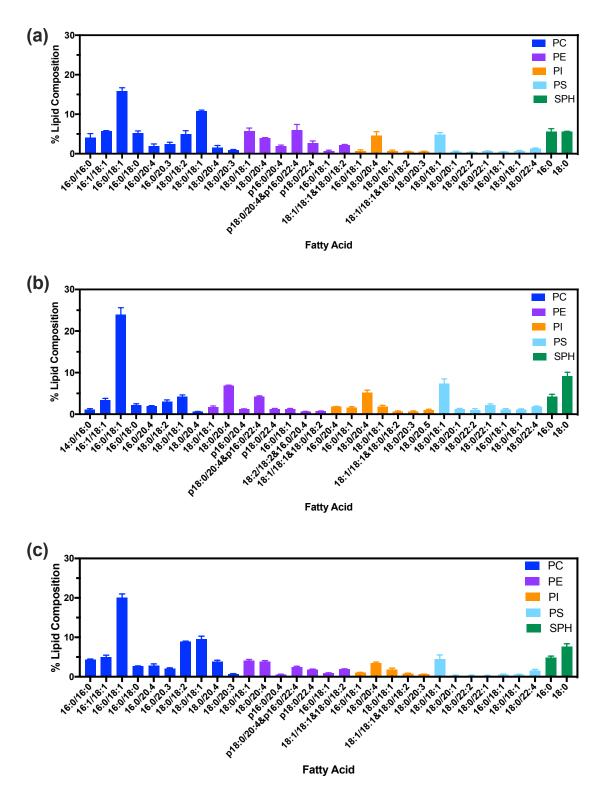


Figure S1. Lipid composition with all fatty acid distributions for (a) HTR-8, (b) TCL-1, and (c) primary trophoblast cells. Results are shown as mean \pm standard deviation; n=3.

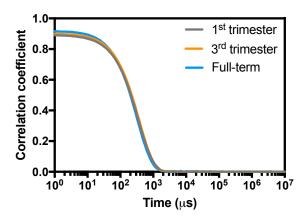


Figure S2. Representative dynamic light scattering correlation curves for 1st trimester, 3rd trimester, and full-term lipid vesicles.

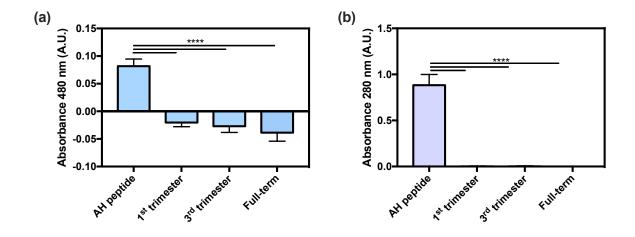


Figure S3. Confirmation of AH peptide removal after rupture of supported placental lipid bilayers. (a) Absorbance measurements for AH peptide using a pierce quantitative colorimetric peptide assay. The control AH peptide concentration was 13 μ M, which is the experimental concentration. After bilayer formation, values were below the detection limit (1 μ M). (b) Absorbance measurements after 1st trimester, 3rd trimester, and full-term bilayer formation, when QCM-D silica coated quartz crystals were rinsed with 2% (w/v) SDS to remove the adsorbed layer. The control

AH peptide concentration was 13 μ M, which is the concentration used in bilayer rupture. After bilayer formation, absorbance values were below the detection limit (0.3 μ M). Statistical analysis was performed using one-way ANOVA with Tukey's post hoc analysis; n=4; α =0.05; ****p<0.0001.

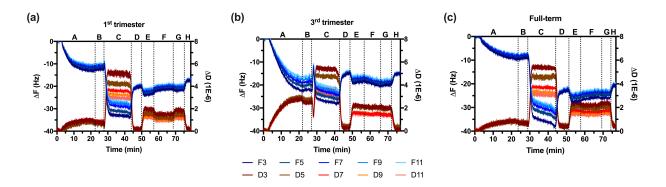


Figure S4. Amphotericin B interactions with (a) 1^{st} trimester, (b) 3^{rd} trimester, and (c) full-term supported placental lipid bilayers investigated via QCM-D. Frequency changes (ΔF) and dissipation changes (ΔD) are shown in blue and red, respectively for overtones 3, 5, 7, 9, and 11. Solution changes are indicated by letters indicating for A: Vesicles, B: Tris NaCl, C: AH peptide, D: Tris NaCl, E: 1% (v/v) DMSO, F: amphotericin B, G: 1% (v/v) DMSO, H: Tris NaCl. Representative QCM-D traces are shown for $n \geq 3$.

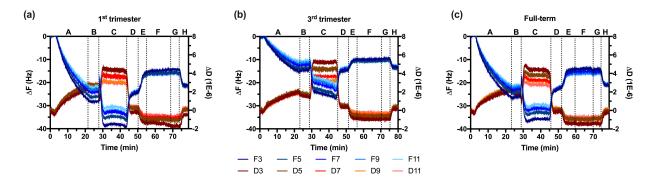


Figure S5. AmBisome interactions with (a) 1st trimester, (b) 3rd trimester, and (c) full-term supported placental lipid bilayers investigated via QCM-D. Frequency changes (ΔF) and dissipation changes (ΔD) are shown in blue and red, respectively for overtones 3, 5, 7, 9, and 11. Solution changes are indicated by letters indicating for A: 1st trimester vesicles, B: Tris NaCl, C: AH peptide, D: Tris NaCl, E: 67.8% (w/w) sucrose and 2% (w/w) succinic acid buffer, which is the clinically utilized suspension buffer, F: AmBisome in the suspension buffer, G: 67.8% (w/w) sucrose and 2% (w/w) succinic acid buffer, H: Tris NaCl. Representative QCM-D traces are shown for $n \ge 3$.

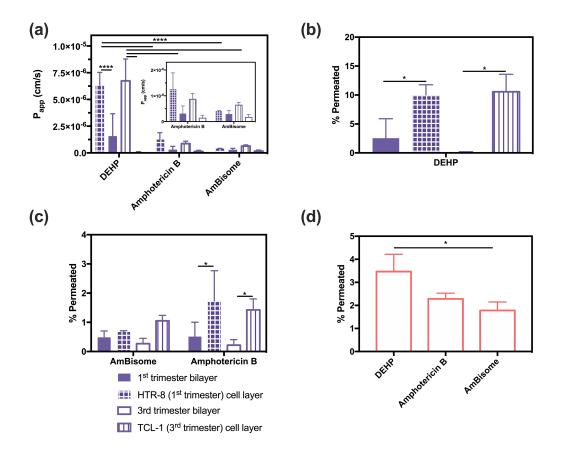


Figure S6. Examining transport across 1st and 3rd trimester suspended placental lipid bilayers and corresponding cell monolayers (HTR-8 and TCL-1, respectively). (a) Apparent permeability (P_{app}) of DEHP, amphotericin B, and AmBisome for placental lipid bilayer and cell monolayers. (b) Percent of 200 μM DEHP that transports across the placental lipid bilayer after 2 h at 25°C or cell monolayer after 2 h at 37 °C. (c) Percent of AmBisome and amphotericin B that transports across the placental lipid bilayer after 2 h at 25°C or cell monolayer after 2 h at 37 °C. (d) Percent of DEHP, amphotericin B, and AmBisome that transports across a placental collagen coated transwell plate. Statistical analysis was performed using one- or two-way ANOVA with Tukey's post hoc analysis; n≥3; α=0.05; *p<0.05; ****p<0.0001.