# **Supporting Information**

# An Endoplasmic Reticulum Targeted Ratiometric Fluorescent Probe for Sensing of Hydrogen Sulfide in Living Cells and Zebrafish

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#### 1. Experimental reagents and instruments

All other chemicals were obtained from commercial suppliers and used without further purification. Silica gel (200-300 mesh, Qingdao Haiyang Chemical Co.) was used for column chromatography. <sup>1</sup>H NMR and <sup>13</sup>C NMR spectra were recorded on a Bruker Advance at 400MHz or at 100 MHz,  $\delta$  values are in parts per million relatives to TMS in DMSO-*d*<sub>6</sub>. Mass spectra (MS) were measured with Bruker Apex IV FTMS using electrospray ionization (ESI). Absorption spectra were recorded on a Purkinje TU-1901 spectrophotometer. Fluorescence measurements were taken on a Hitachi F-7000 fluorescence spectrometer with a 10mm quartz cuvette. pH measurements were carried out with a pH acidometer (Mettler Toledo FE-30). Fluorescence imaging was observed under an Olympus IX81 confocal fluorescence microscope.

HeLa cells were cultured in high-glucose DMEM supplemented with 10% fetal bovine serum, 1% penicillin, and 1% streptomycin with 5% CO<sub>2</sub> at 37°C.

4-day-old zebrafish were obtained from Shanghai Feixi Biotechnology Co., Ltd.

#### 2. Normalized spectra of probe M-H<sub>2</sub>S to H<sub>2</sub>S



Fig. S1. (A) Normalized UV–*vis* absorption spectra and fluorescence spectra of probe M-H<sub>2</sub>S.
(B) Normalized UV–*vis* absorption spectra and fluorescence spectra of M-H<sub>2</sub>S recognized after H<sub>2</sub>S.

 $\lambda_{ex}$  = 480 nm, slit width:  $d_{ex} = d_{em} = 10$  nm, 10 mM PBS, pH 7.4, containing 10% DMSO, v/v at

room temperature.

#### 3. The stability of M-H<sub>2</sub>S at different pH values



Fig. S2. Fluorescent intensity ( $F_{650}/F_{560}$ ) responses of M-H<sub>2</sub>S (10  $\mu$ M) at different pH values (pH=5.0, 6.0, 7.4, 8.0). Spacing interval is 1 min.

#### 4. Time response spectra of probe M-H<sub>2</sub>S to H<sub>2</sub>S



Fig. S3. UV-vis absorption A) and ratio intensity changes B) of M-H<sub>2</sub>S (10 µM) upon addition

of  $H_2S$  (50  $\mu$ M). Fluorescence spectra C) and ratio intensity changes D) of **M-H<sub>2</sub>S** (10  $\mu$ M) upon addition of  $H_2S$  (50  $\mu$ M).

#### 5. Determination of the Detection Limit

According to IUPAC, the detection limits were determined based on the fluorescence titrations, carried out in PBS / DMSO (9:1, v/v), pH 7.4, using the following equation:

Detection limit = 
$$3\sigma/k$$

where  $\sigma$  is the standard deviation of blank measurements and *k* is the slope of the plot of fluorescence intensity vs H<sub>2</sub>S concentration. The standard deviations  $\sigma = 0.0011$ .

6. The capabilities of M-H<sub>2</sub>S for detecting H<sub>2</sub>S at different pH



Fig. S4. Fluorescence responses of  $M-H_2S$  (10  $\mu$ M) in the absence and presence of  $H_2S$  (50

μM) at different pH.

#### 7. High-resolution mass spectra of reaction system



Fig. S5. The HRMS of the product after the reaction between M-H<sub>2</sub>S and H<sub>2</sub>S.

## 8. Cytotoxicity assay



Fig. S6. MTT assay for estimating cell viability (%) of HeLa cells. The concentrations of probe

**M-H<sub>2</sub>S** were used: 1: blank, 2: 5 μM, 3: 10μM, 4: 15 μM, 5: 20 μM, respectively.

## 9. Characterization data



Fig. S7. <sup>1</sup>H NMR spectral of compound M-1.



Fig. S8. HRMS spectral of compound M-1.







Fig. S10. HRMS spectral of compound M-OH.



Fig. S11. <sup>1</sup>H NMR spectral of compound M-H<sub>2</sub>S.



Figure. S12 <sup>13</sup>C NMR spectral of compound M-H<sub>2</sub>S.



Fig. S13. HRMS spectral of compound M-H<sub>2</sub>S.