Discovery of novel selective and orally bioavailable phosphodiesterase-1

inhibitors for the efficient treatment of idiopathic pulmonary fibrosis

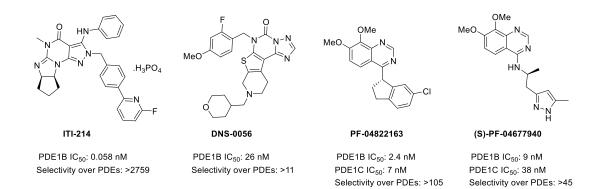
Yinuo Wu[#], Yi-Jing Tian[#], Mei-Ling Le[#], Si-Rui Zhang, Chen Zhang, Meng-Xing Huang, Mei-Yan Jiang, Bei Zhang, and Hai-Bin Luo*

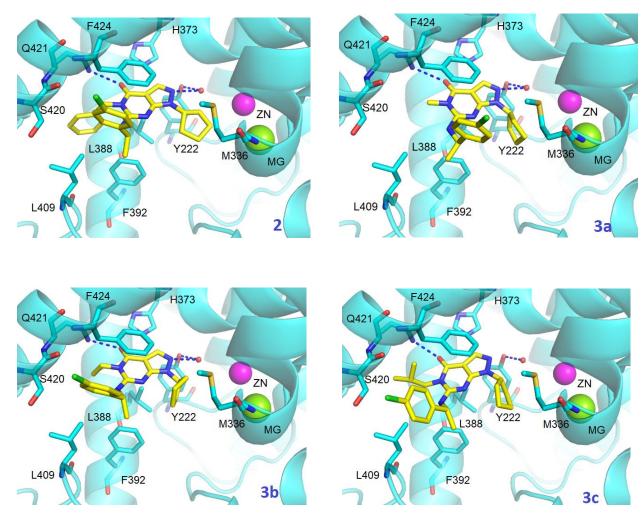
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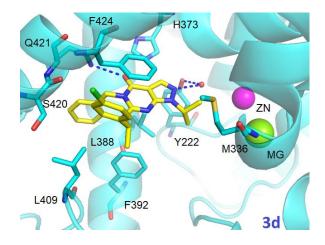
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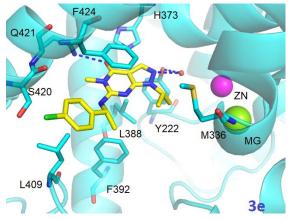
1. Figure S1. Chemical structures of PDE1 inhibitors.

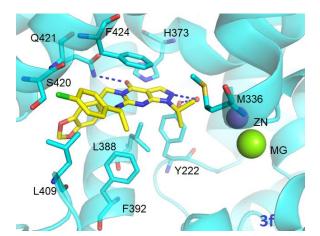


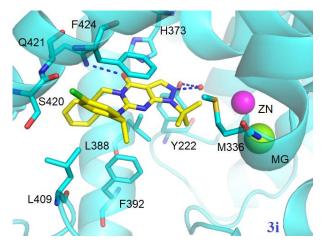


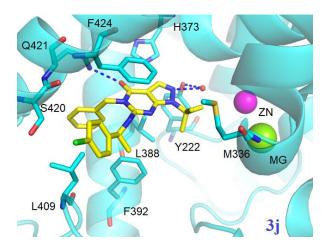
2. Figure S2: The binding modes of representative compounds

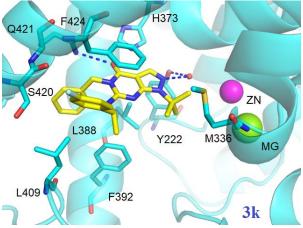


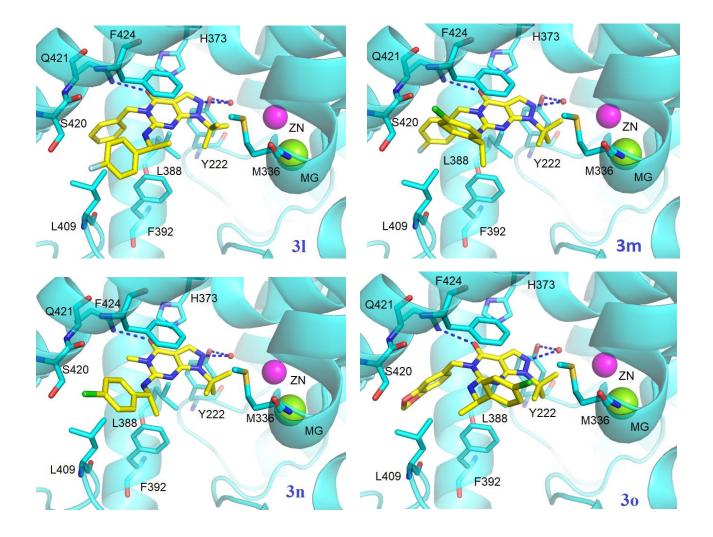












3. Stability of 3m in the rat liver microsomes.

The assays were performed at the Shanghai institute of Materia Medica, Chinese Academy of Sciences, Shanghai, China. **3m** was dissolved in 100% DMSO to prepare a 0.5mM stock solution and diluted to a final concentration of 1.5µM for the experiments. Ketanserin (NPGAH-CO from TCI company) was used as the positive control.

4. In vivo pharmacokinetics analysis of 3m.

The pharmacokinetic properties of **3m** were analyzed by Shanghai institute of Materia Medica, Chinese Academy of Sciences, Shanghai, China. Six male SD rats with a body weight of 230-260 g were purchased from Shanghai SIPPR-BK LAB Animal Ltd., Shanghai, China, and used for the pharmacokinetic analysis of **3m**. It was dissolved/suspended in 5% DMSO, 10% Solutol, and 85% water for intravenous administration (iv) and oral administration (po). A final dosage of 2.5 and 5.0 mg/kg rat of the formulated compounds was administered for iv and po purposes, respectively, and the blood samples were taken at various time points during a 24 h period. The concentration of the compounds in the blood was analyzed by LC-MS/MS (Shimadzu liquid chromatographic system and API4000 mass spectrometer, Applied Biosystems, Ontario, Canada).

5. hERG inhibition and humanCYP450 inhibition of 3m.

The assays were performed by the Medicilon Company, Shanghai, China. hERG inhibition was performed using an automated patch clamp electrophysiology measurement in CHO-hERG cells. TheCYP450 inhibition assay was performed by incubating compound **3m** with human liver microsomes and NADPH in the presence of theCYP450-isoform specific probe substrate. The IC₅₀ values of compound **3m** for five CYP isoenzymes (i.e., CYP1A2, CYP2C9, CYP2D6, CYP3A4-M

and CYP3A4-T) were determined.

6. Acute toxicity of compound 3m.

The acute toxicity was tested according to similar protocols that were described in our previous study.¹ Thirty KM mice (22 days, 18-20 g), which were purchased from the Laboratory Animal Center of Sun Yat-Sen University (Guangzhou, China), were used to evaluate the acute toxicity of **3m**. Mice were randomly divided into three groups, and each group was given in single oral dose of 0, 1000, or 1500 mg/kg **3m** on the first day of the experiment. Mice were maintained on a 12 h light/dark cycle (light from 7:00 to 19:00) at room temperature and 60-70% relative humidity. Sterile food and water were provided according the institutional guidelines. Prior to each experiment, mice were fasted overnight and allowed free access to water. Compound **3m** was dissolved in 5% DMSO/10% Solutol/85% water solution and orally administered. Mice were observed for any abnormal behavior and mortality and weighed 4 h after **3m** was administered and then every 24h for 14 days. Animals were sacrificed on the 14th day, and tissue samples of the heart, liver, and kidney were macroscopically examined for possible damage.

7. Molecular Docking and Molecular Dynamics Simulations.

The X-ray crystal structure of PDE1 (PDB code: 5UP0) and **C33**-PDE9A complex (PDB ID: 4Y874) was selected for molecular modeling, and Surflex-dock embedded in the software Tripos Sybyl 2.0 was used. ^{2,3} Two metal ions crucial for the PDE's catalytic activity in the catalytic domain and water molecules coordinate these two metal ions were retained. Hydrogen atoms were added, and the ionizable residues were protonated at the neutral pH. The protomol, was generated using the

parameters by default. The parameters of proto_thresh and proto_bloat were assigned 0.5 and 0, respectively. After the protomol was the prepared, molecular docking was performed for test molecules.

After molecular docking completed, similar MD simulation procedures as previous studies were used to equilibrate the whole system. Here 8 ns MD simulations were carried out in the NPT ensemble with a constant pressure of 1 atm and a constant temperature of 300 K. The periodic boundary conditions were adopted, along with an 8 Å cutoff for long-range electrostatic interactions with the partial mesh Ewald (PME) method. The SHAKE algorithm was utilized to deal with all bonds involving hydrogen atoms, and hence the time step was set to 2 fs. An Intel Xeon E5620 CPU and an NVIDIA Tesla C2050 GPU, which are available in performing floating-point calculations, were applied to accelerate the process of MD simulations for each system. Subsequently, the 100 snapshots were isolated from the final 1.0 ns period of the MD simulation trajectories, and then used for bindingfree-energy calculations by the MM-PBSA approach. For the electrostatic contribution to the solvation-free energy, the PBSA program in the Amber 16 suite was used, which could numerically solves the Poissone Boltzmann equations.

In light of the MM-PBSA method, the binding free energy (ΔG_{bind}) can be calculated by the following equation 1. G_{complex} , G_{rec} and G_{lig} represent the free energies of complex, receptor and ligand, respectively.

$$\Delta G_{\text{bind}} = G_{\text{complex}} - G_{\text{rec}} - G_{\text{lig}} \tag{1}$$

Each free energy was evaluated by the sum of the MM energy E_{MM} , the solvation free energy G_{solv} , and the entropy contribution *S*, respectively, leading to equation 2.

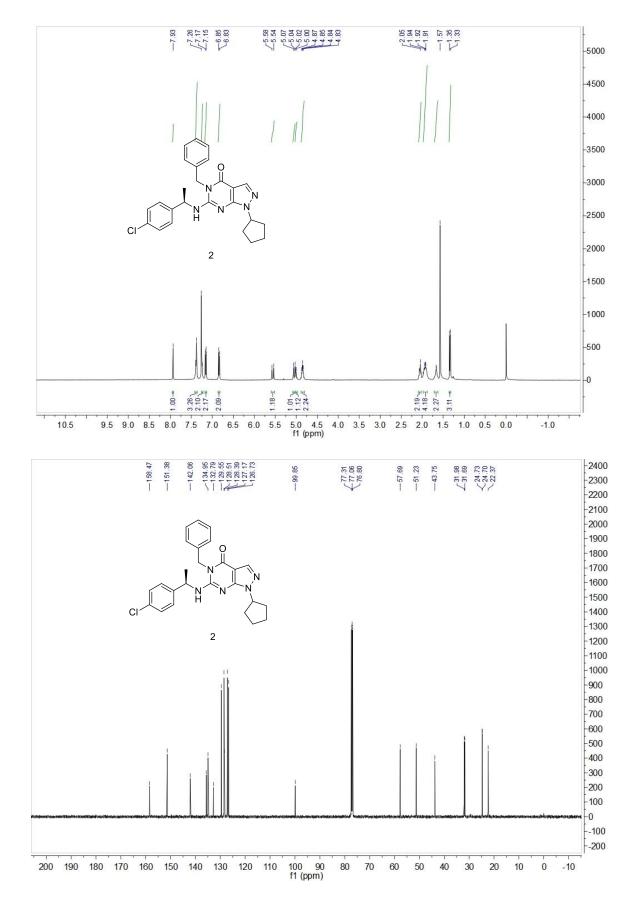
 $\Delta G_{bind} = \Delta E_{\rm MM} + \Delta G_{\rm solv} - T\Delta S \tag{2}$

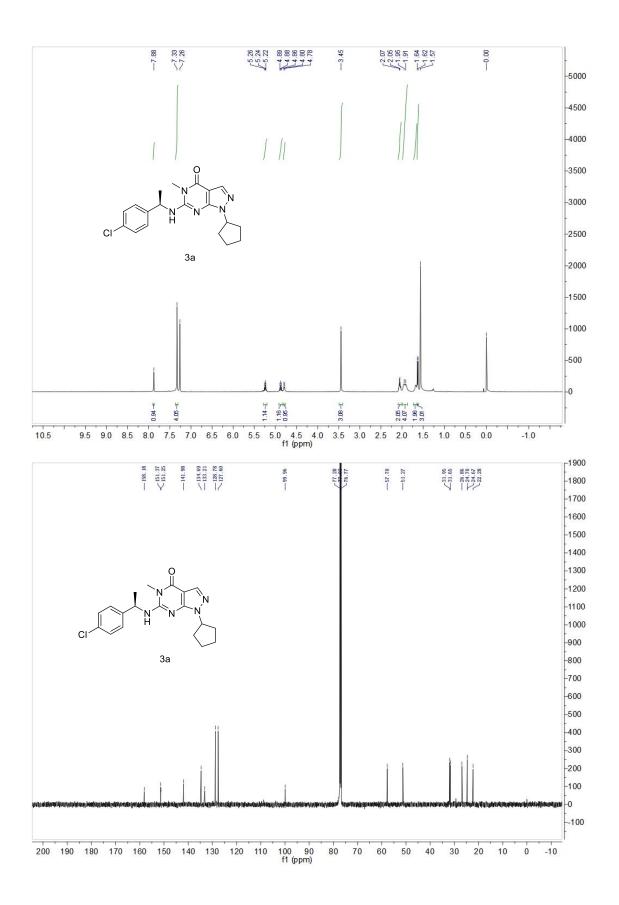
 ΔE_{MM} is the gas phase interaction energy and can be decomposed into $E_{\text{MM, comp}}$, $E_{\text{MM, rec}}$ and $E_{\text{MM, rec}}$ lig. Solvation free energy is evaluated by the sum of the electrostatic solvation free energy and nonpolar solvation free energy. The electrostatic solvation free energy, ΔG_{PB} , was solved by the Poisson Boltzmann (PB) equation, while the nonpolar solvation free energy varies proportionally with the solvation accessible surface area (SASA), leading to by equation 3 and 4.

$$\Delta G_{\text{solv}} = \Delta G_{\text{PB}} + \Delta G_{\text{np}}$$
(3)
$$\Delta G_{\text{np}} = \gamma \text{ SASA} + b$$
(4)

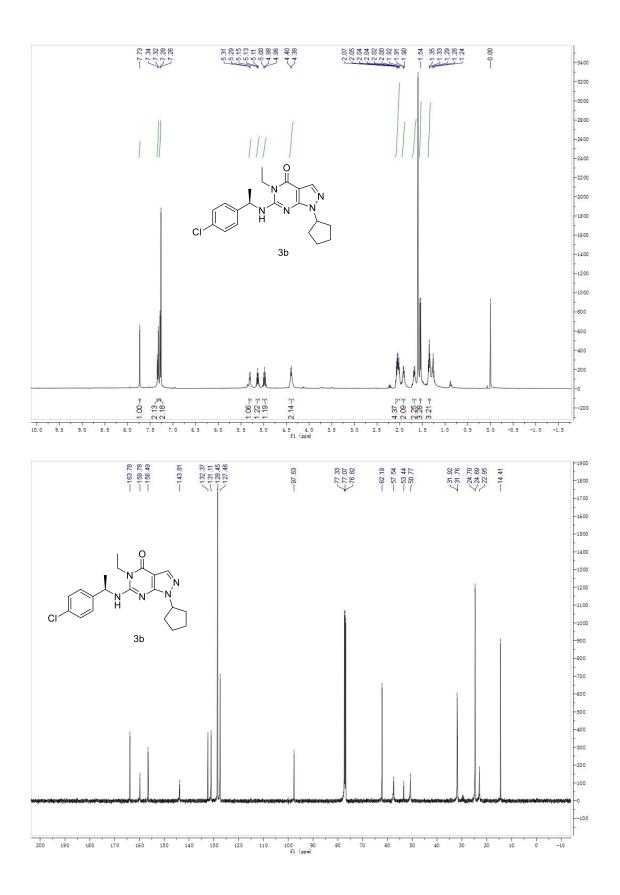
In order to get the compromise between efficiency and accuracy, entropy contribution term was omitted for ΔG_{bind} in equation 2, since the calculations of the entropy contribution is extremely time-consuming for large protein-ligand systems. The default parameters were adopted, with $\gamma = 0.0072$ kcal/(Å²) and b = 0 kcal/mol.The charges of Mg²⁺ and Zn²⁺ were assigned 2.0 for PB calculations, and their bond raddi were used for SA calculations.

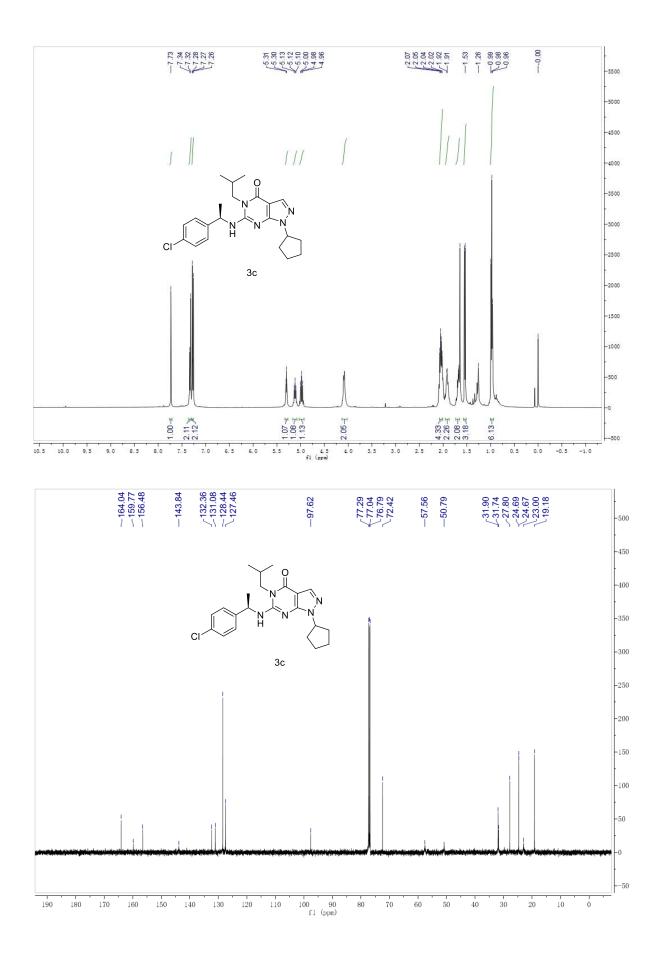


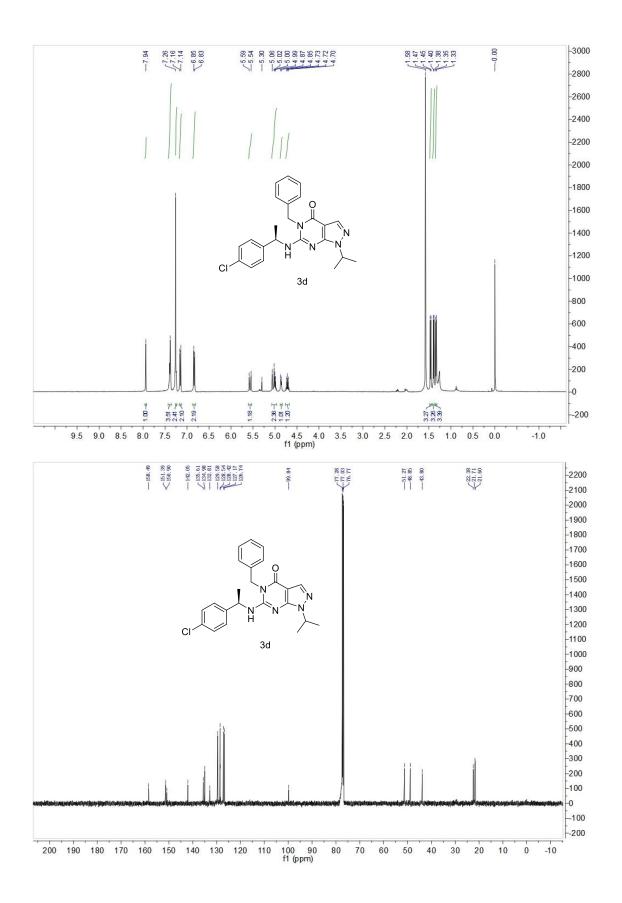


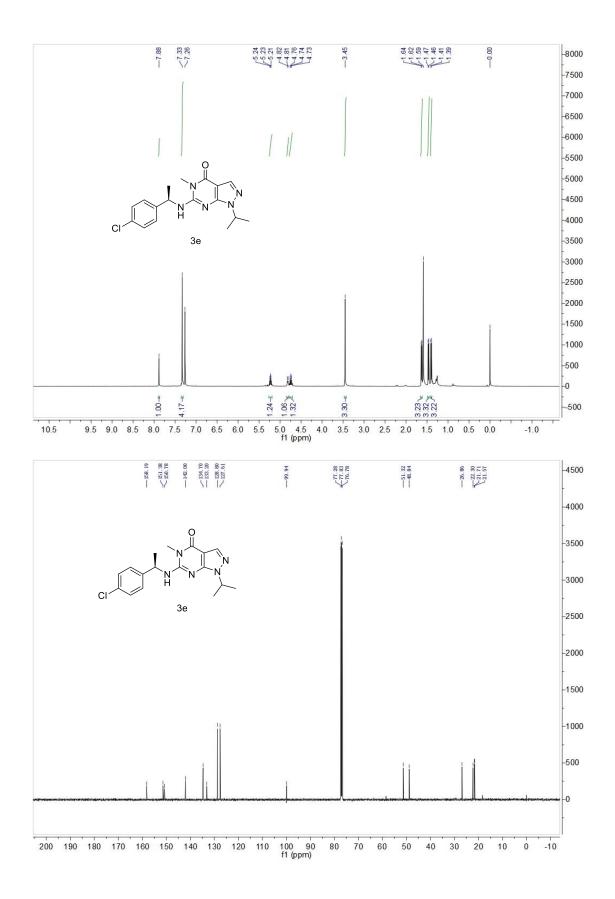


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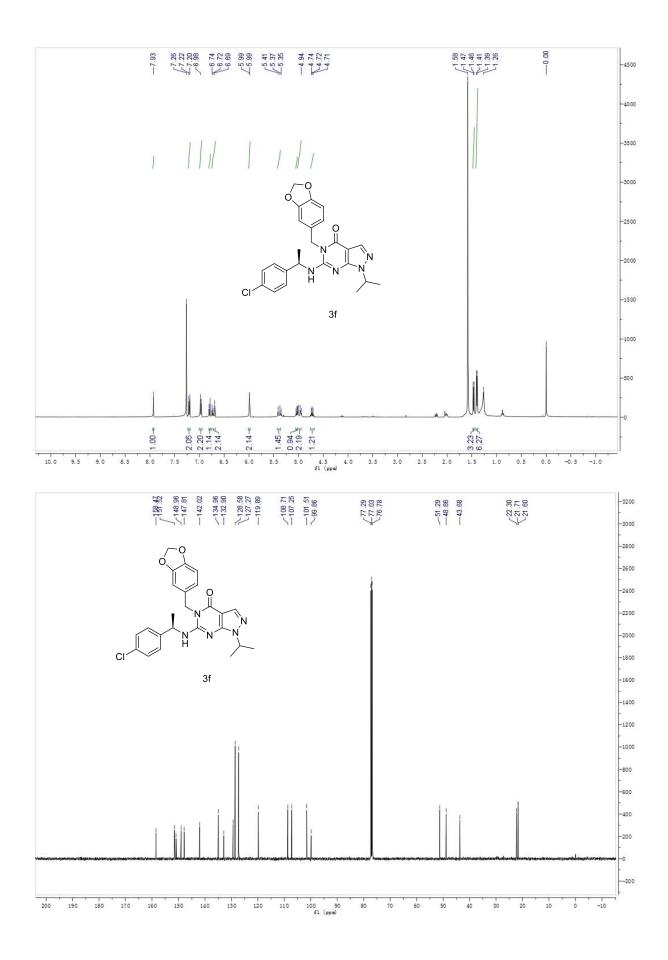


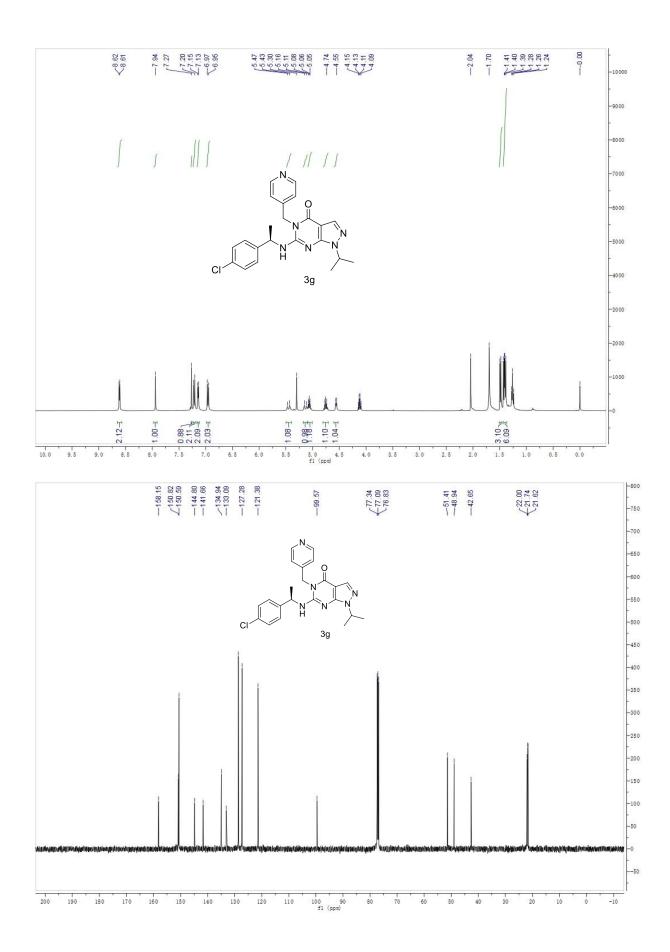


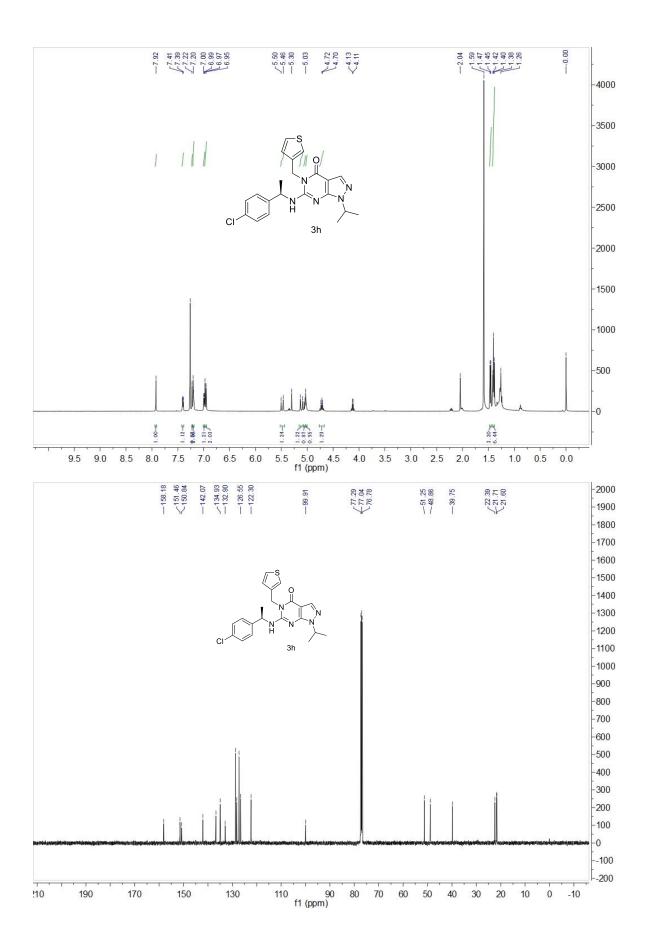


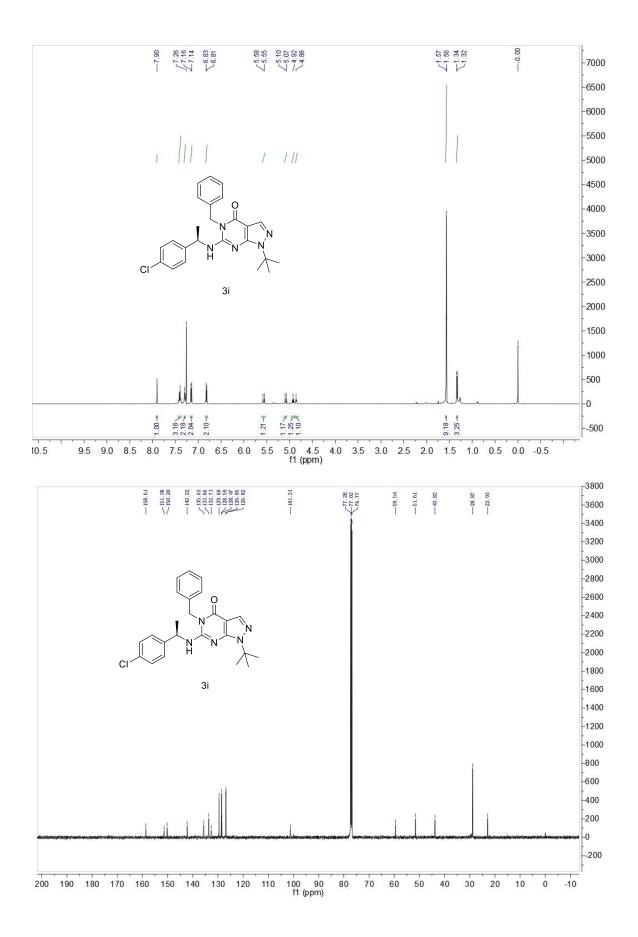


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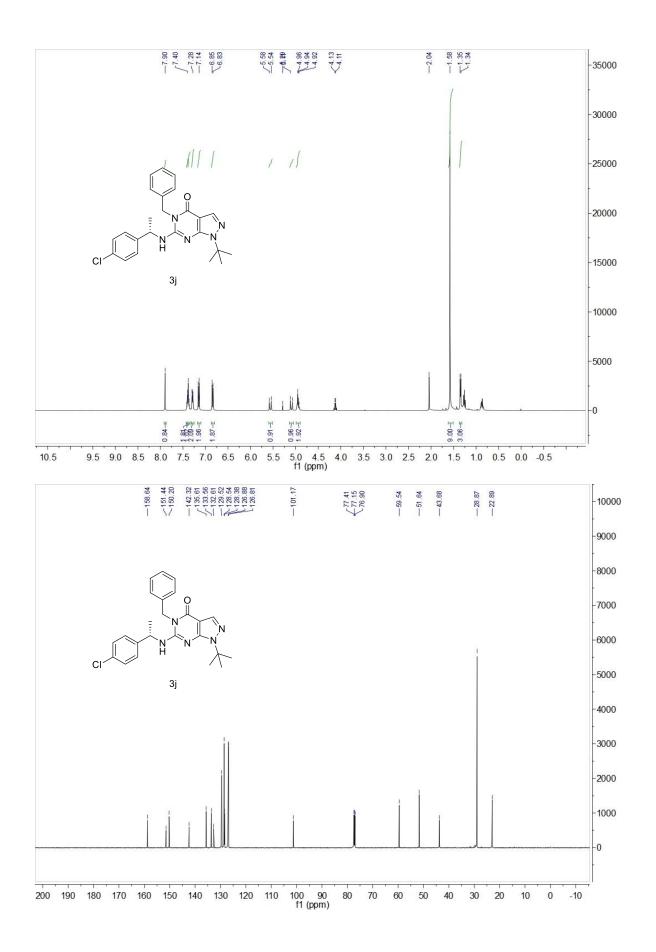


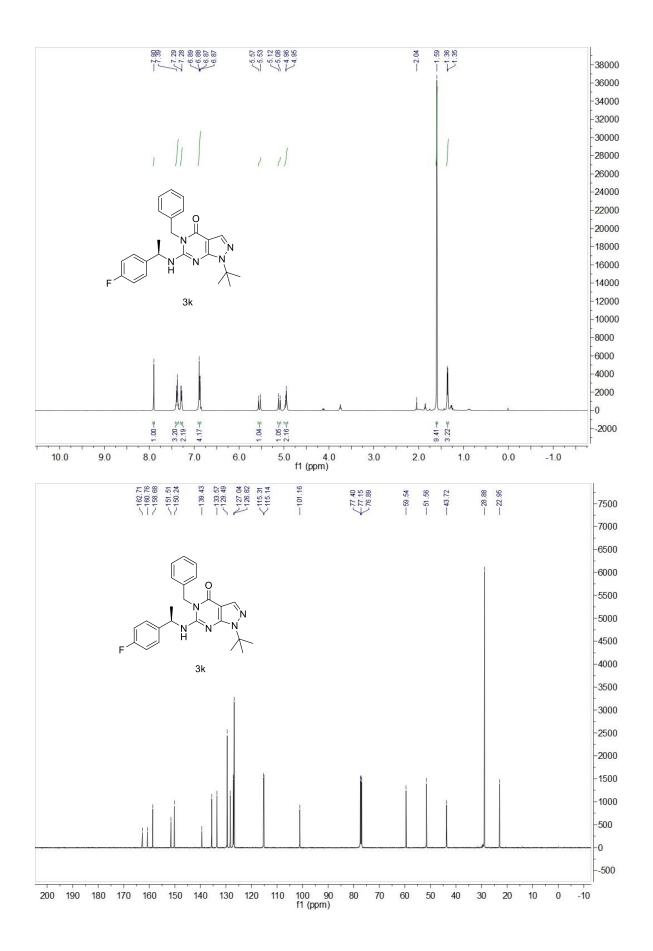


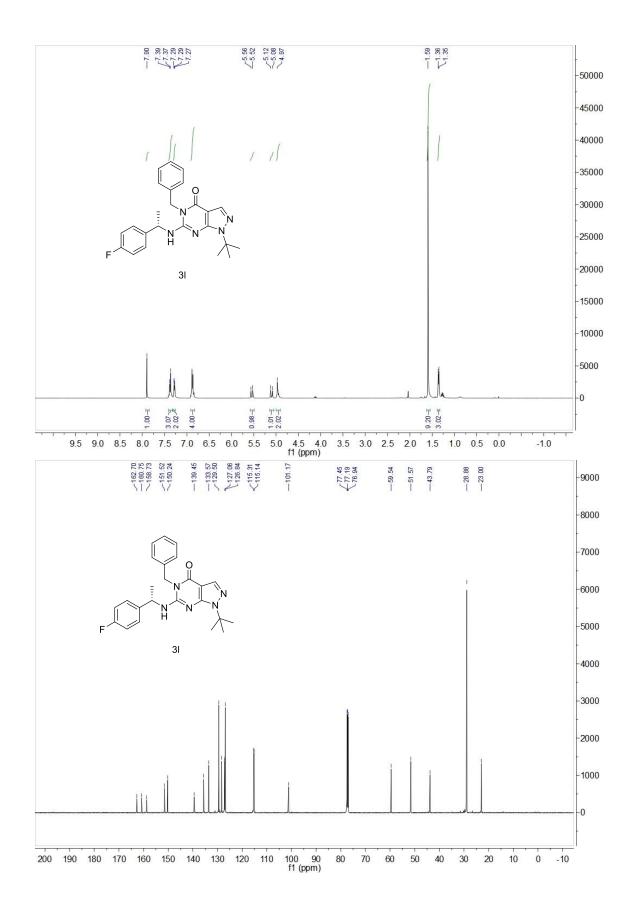


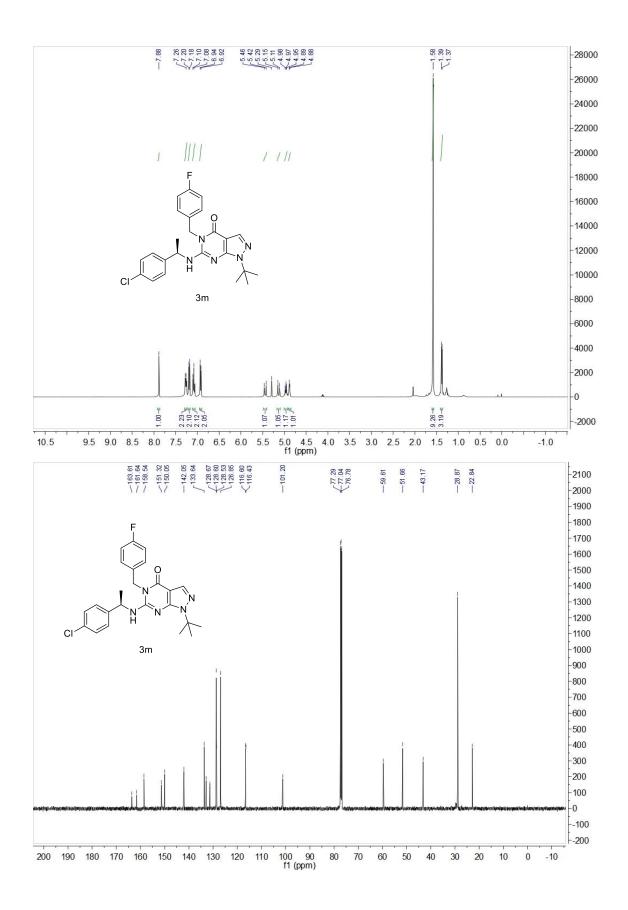


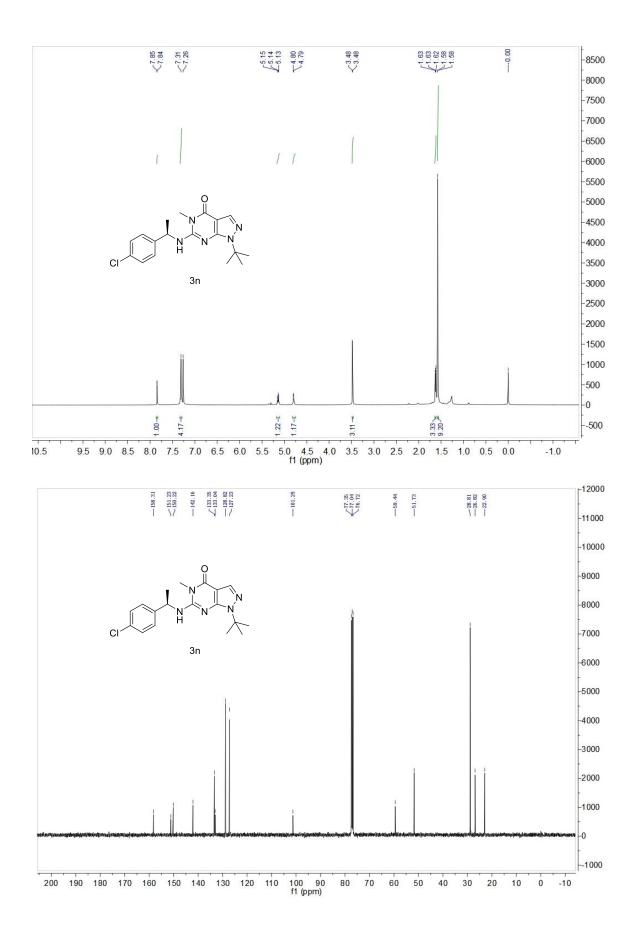
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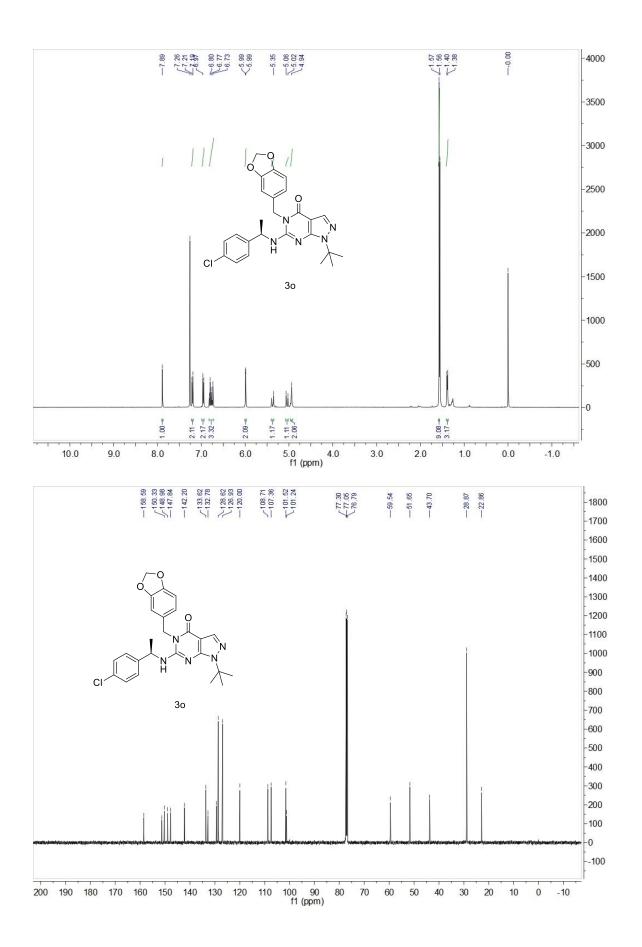


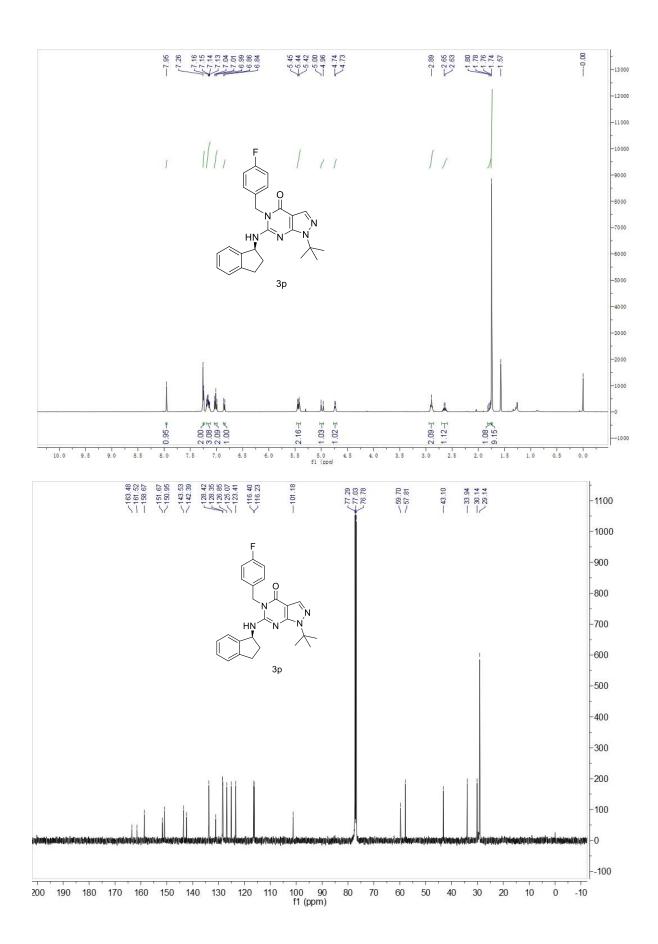


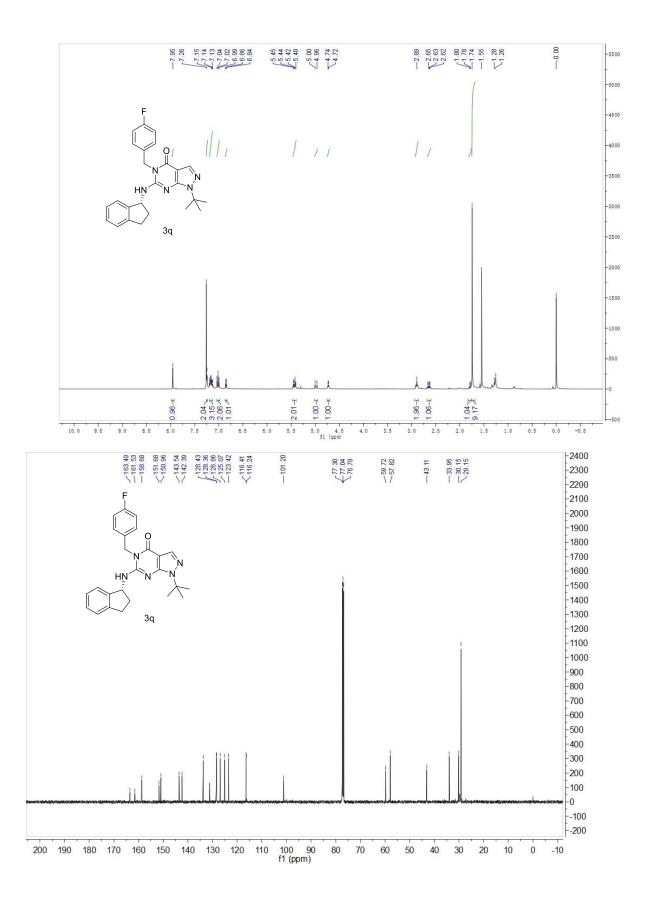


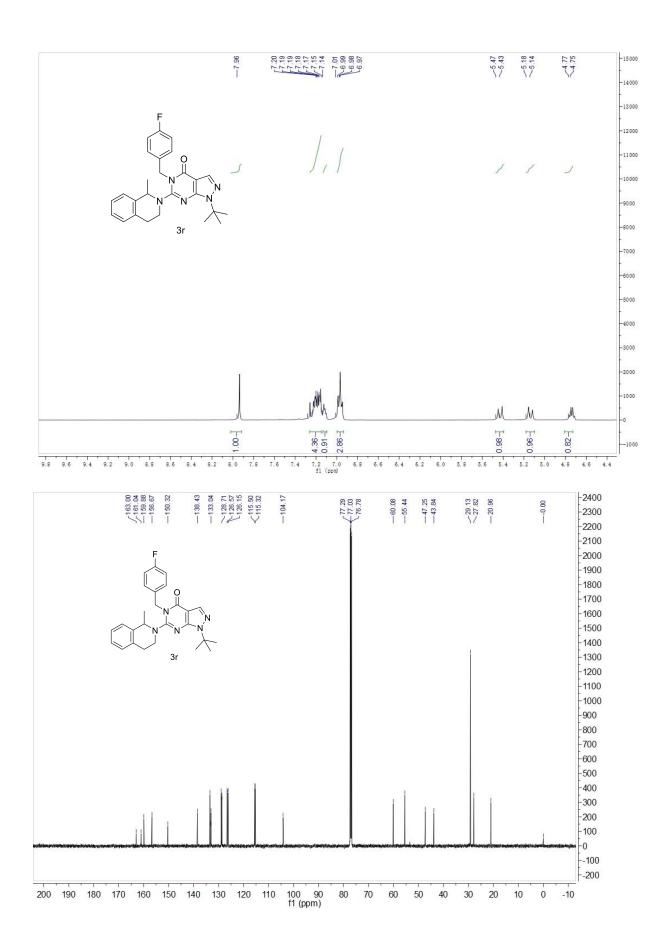




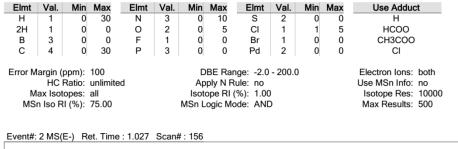


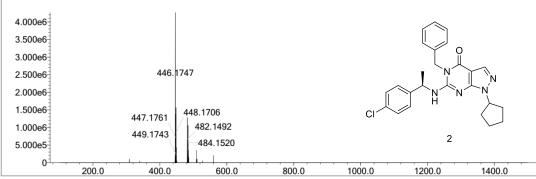




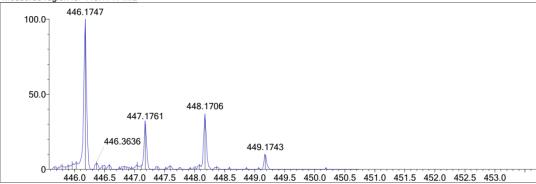


9. The HRMS spectrums of the target compounds.

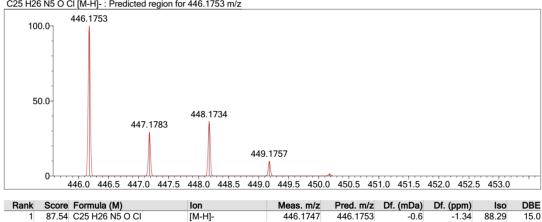


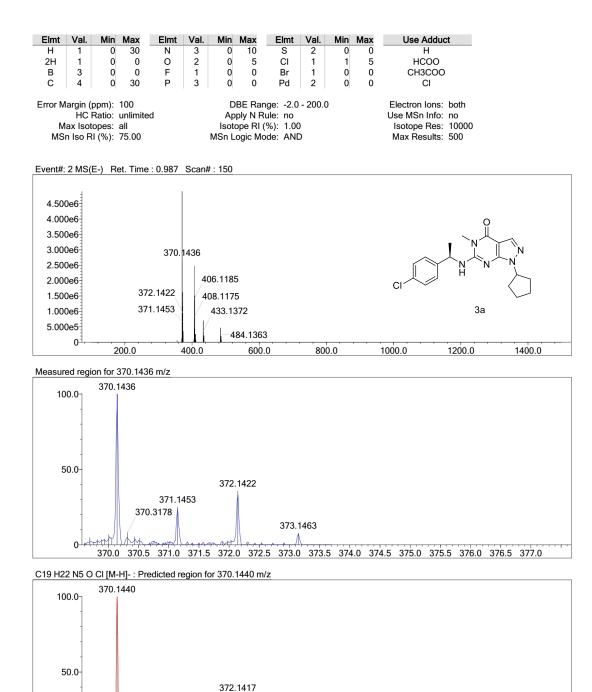


Measured region for 446.1747 m/z



C25 H26 N5 O CI [M-H]- : Predicted region for 446.1753 m/z





373.1442

370.0 370.5 371.0 371.5 372.0 372.5 373.0 373.5 374.0 374.5 375.0 375.5 376.0 376.5 377.0

Meas. m/z

370.1436

Pred. m/z Df. (mDa) Df. (ppm)

-0.4

370.1440

DBE

11.0

lso

93.45

-1.08

371.1469

lon

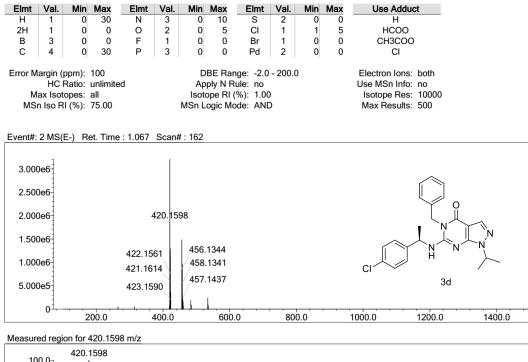
[M-H]-

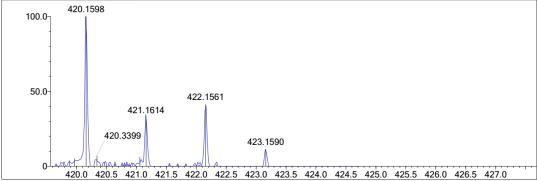
0-

1

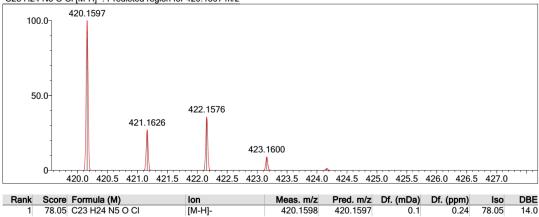
Rank Score Formula (M)

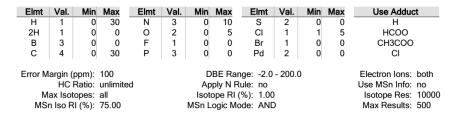
93.26 C19 H22 N5 O CI



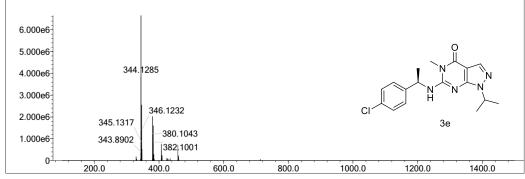


C23 H24 N5 O CI [M-H]- : Predicted region for 420.1597 m/z

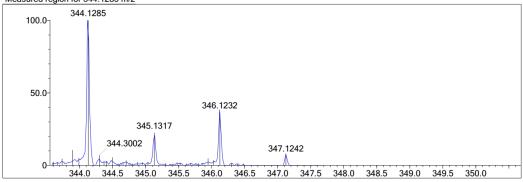




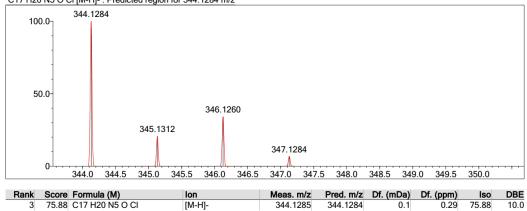


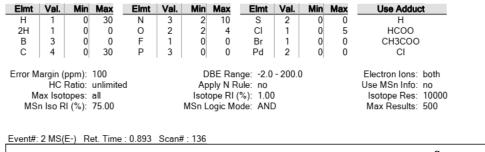


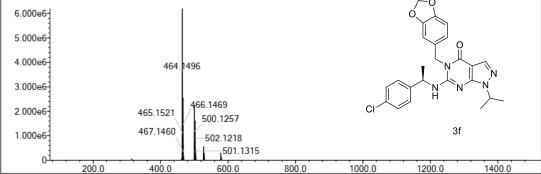
Measured region for 344.1285 m/z

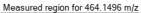


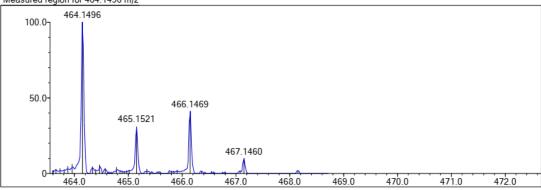




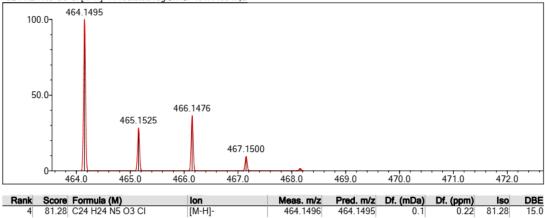


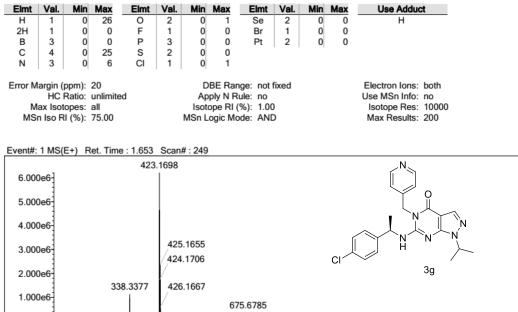




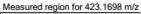


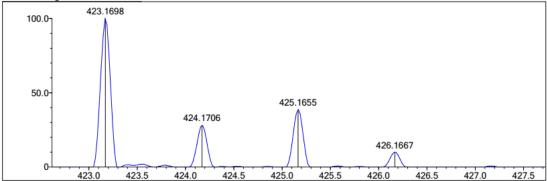




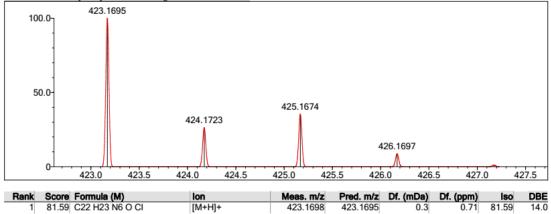


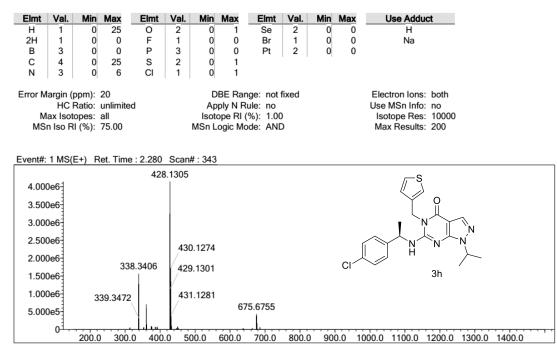


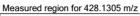


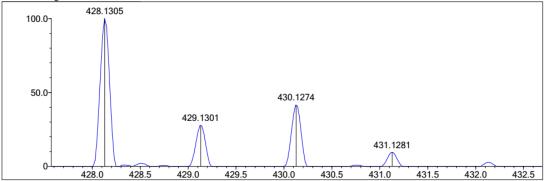


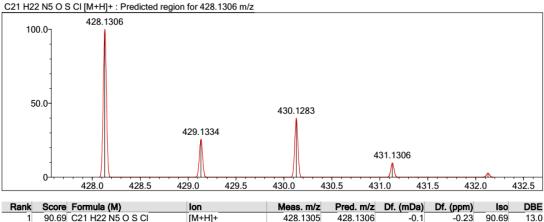












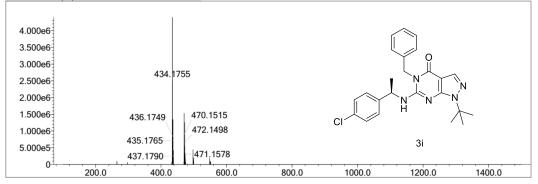


90.69 C21 H22 N5 O S CI 428.1305 -0.23 90.69 [M+H]+ 428.1306 -0.1 1

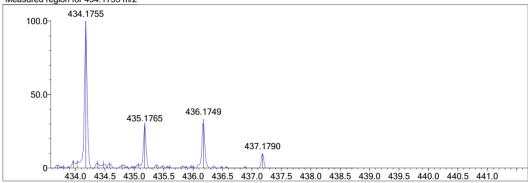
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2H	1	0	0	0	2	0	5	CI	1	1	5	HCOO
В	3	0	0	F	1	0	0	Br	1	0	0	CH3COO
С	4	0	30	Р	3	0	0	Pd	2	0	0	CI
	HC ax Isot		unlimit all	ed	DBE Range: -2.0 - 200.0 Apply N Rule: no Isotope RI (%): 1.00 MSn Logic Mode: AND							Electron lons: both Use MSn Info: no Isotope Res: 10000 Max Results: 500

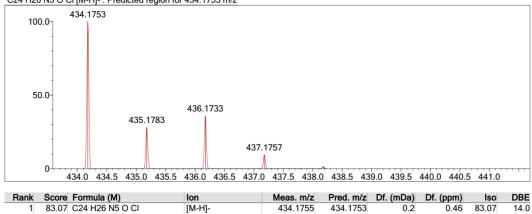
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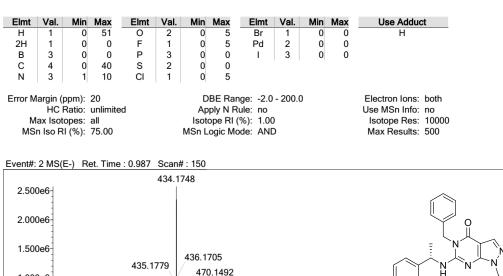


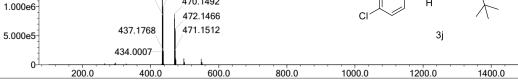
Measured region for 434.1755 m/z



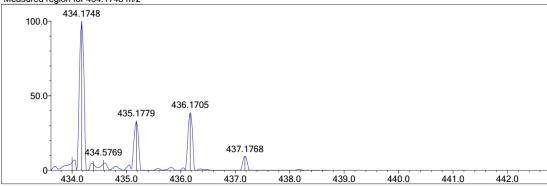
C24 H26 N5 O CI [M-H]- : Predicted region for 434.1753 m/z



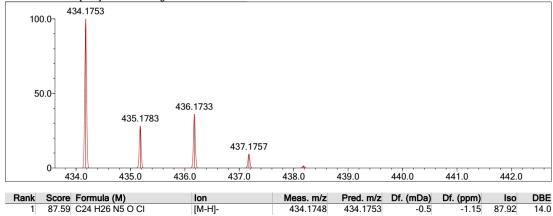


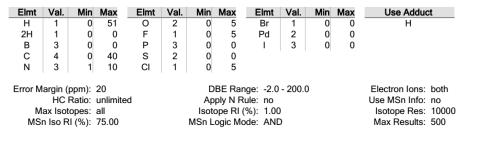


Measured region for 434.1748 m/z

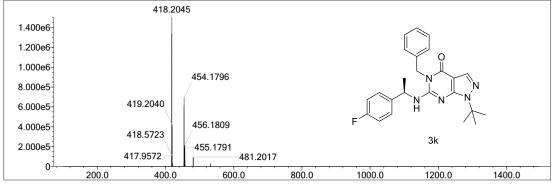


C24 H26 N5 O CI [M-H]- : Predicted region for 434.1753 m/z

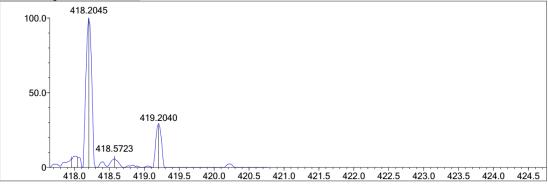




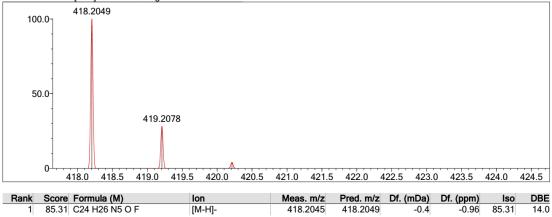
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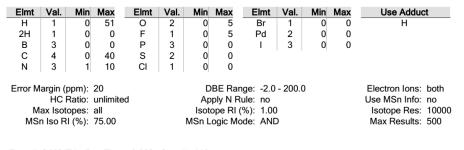


Measured region for 418.2045 m/z

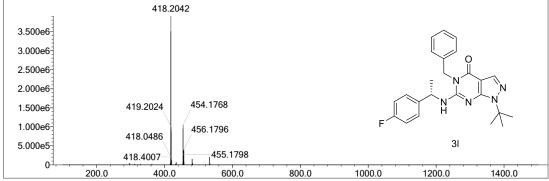


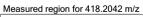
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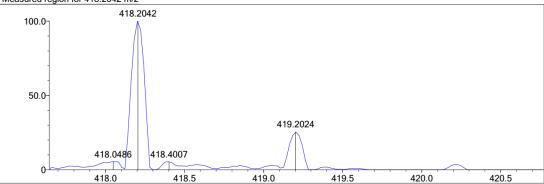




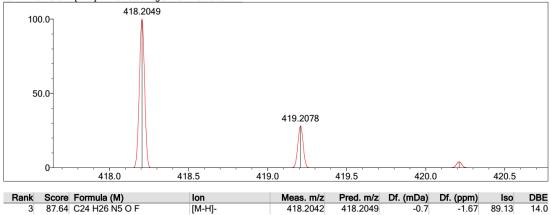
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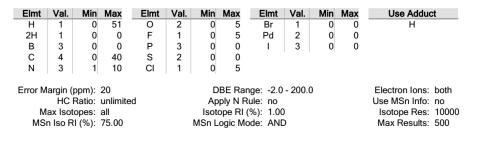




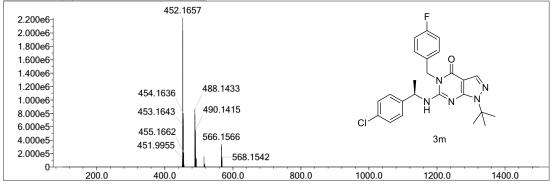




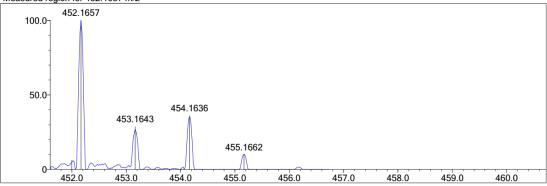
S38



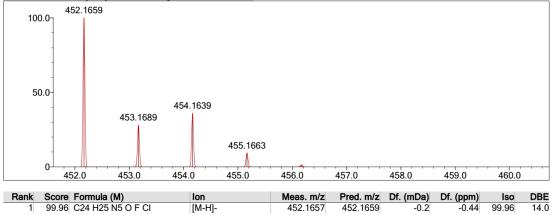
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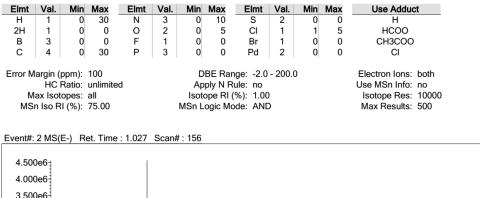


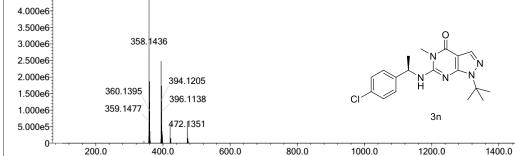




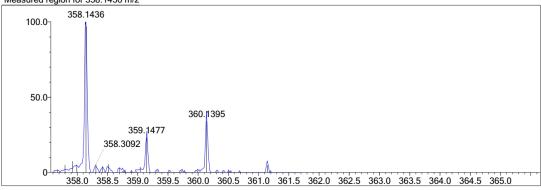
C24 H25 N5 O F CI [M-H]- : Predicted region for 452.1659 m/z



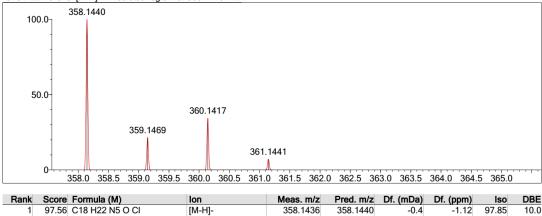


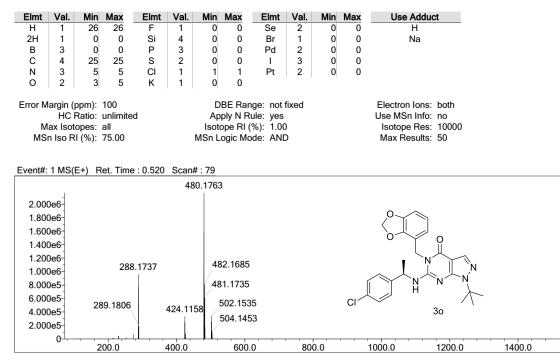


Measured region for 358.1436 m/z

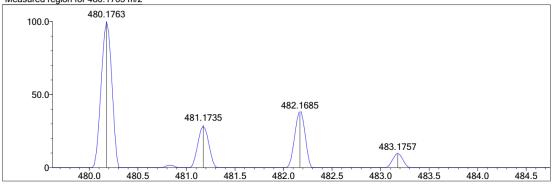


C18 H22 N5 O CI [M-H]- : Predicted region for 358.1440 m/z

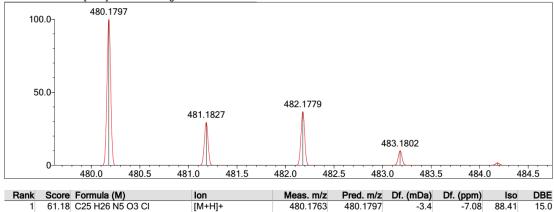


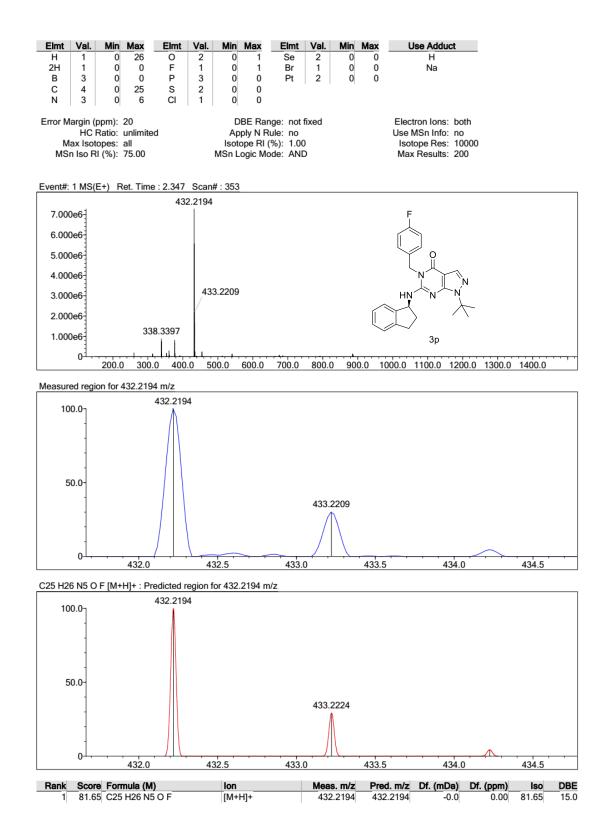


Measured region for 480.1763 m/z

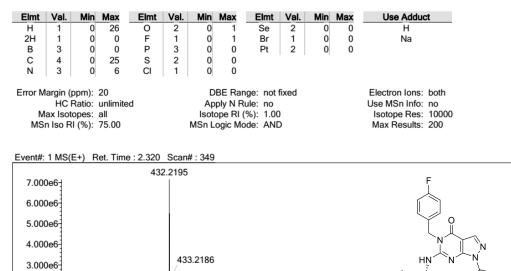


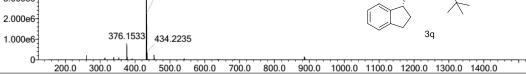
C25 H26 N5 O3 CI [M+H]+ : Predicted region for 480.1797 m/z



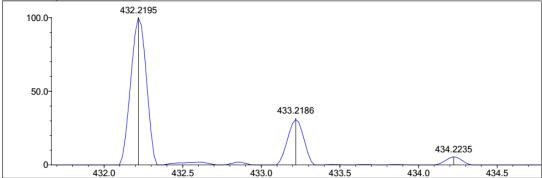


S42

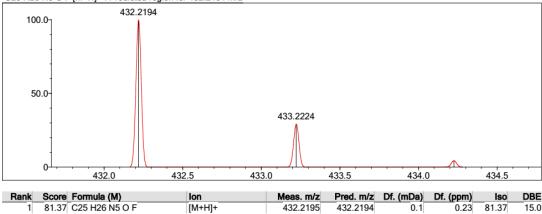


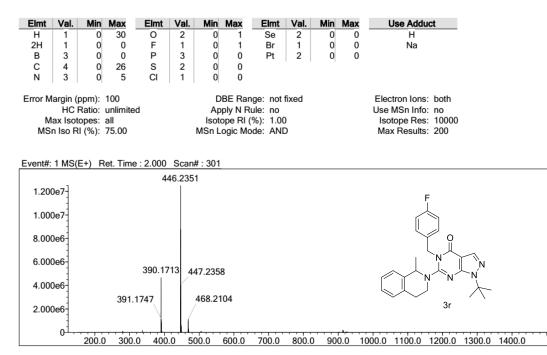


Measured region for 432.2195 m/z

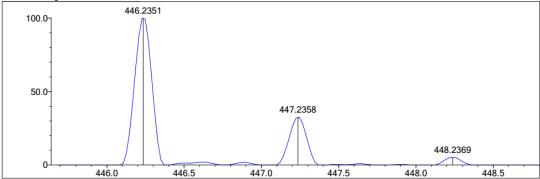


C25 H26 N5 O F [M+H]+ : Predicted region for 432.2194 m/z

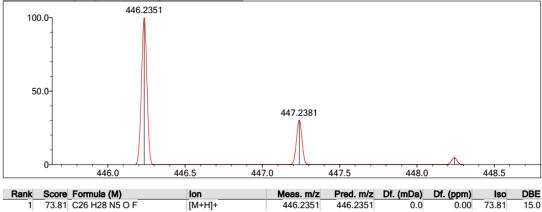




Measured region for 446.2351 m/z

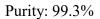


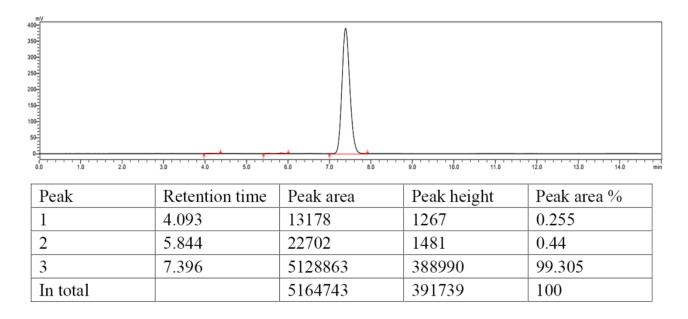




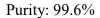
10. The purity spectrums of the target compounds.

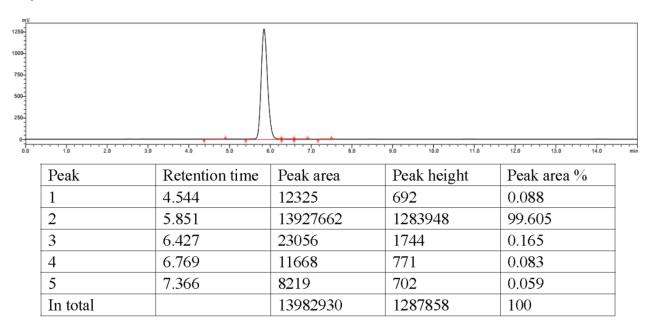
Compound 2: UV detection at 254nm; elution, MeOH (100%); T = 25°C; Flow rate = 1.0 mL/min.





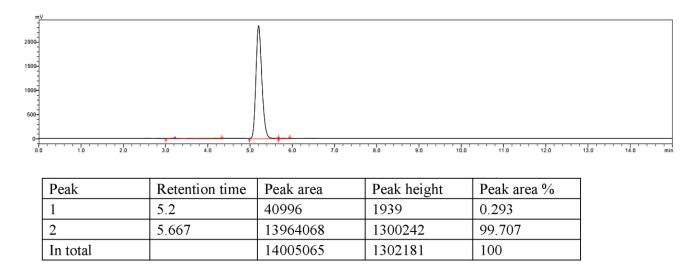
Compound **3a**: UV detection at 254nm; elution, MeOH (100%); T = 25°C; Flow rate = 1.0 mL/min.





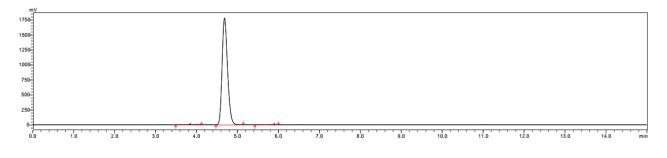
Compound 3d: UV detection at 254 nm; elution, MeOH (100%); T = 25°C; Flow rate = 1.0 mL/min.

Purity: 99.7%



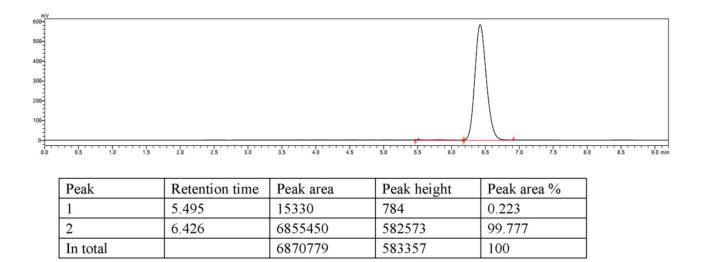
Compound 3e: UV detection at 254 nm; elution, MeOH (100%); T = 25°C; Flow rate = 1.0 mL/min.

Purity: 99.7%



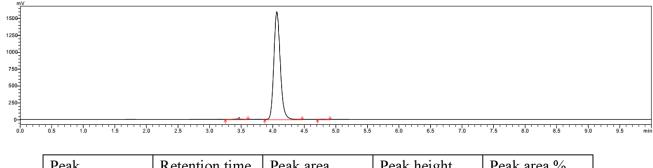
Peak	Retention time	Peak area	Peak height	Peak area %
1	3.813	38101	2109	0.231
2	4.689	16406837	1778960	99.677
3	5.868	15122	1360	0.092
In total		16460061	1782428	100

Compound **3g** : UV detection at 254 nm; elution, MeOH (100%); T = 25°C; Flow rate = 1.0 mL/min. Purity: 99.8%



Compound **3h** : UV detection at 254 nm; elution, MeOH (100%); $T = 25^{\circ}C$; Flow rate = 1.0 mL/min.

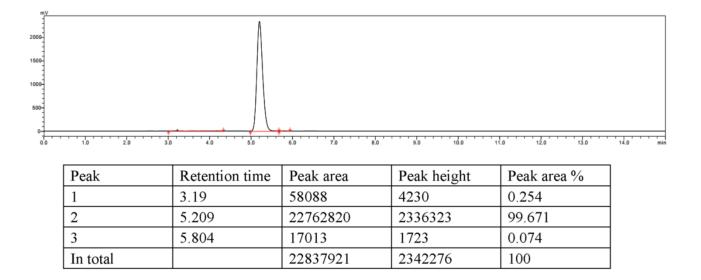
Purity: 99.4%



Peak	Retention time	Peak area	Peak height	Peak area %
1	3.445	52224	7289	0.46
2	4.07	11295035	1595970	99.403
3	4.795	15577	2264	0.137
In total		11362836	1605523	100

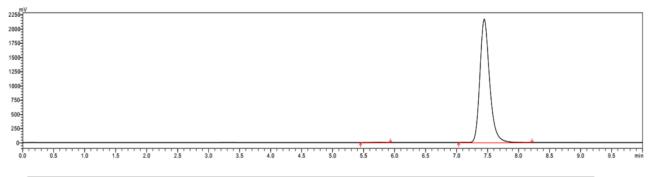
Compound 3k: UV detection at 254 nm; elution, MeOH (100%); T = 25°C; Flow rate = 1.0 mL/min.

Purity: 99.7%



Compound **31** : UV detection at 254 nm; elution, MeOH (100%); $T = 25^{\circ}C$; Flow rate = 1.0 mL/min.

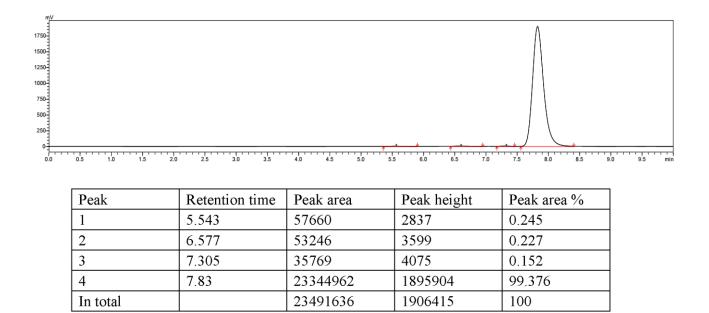
Purity: 99.8%



Peak	Retention time	Peak area	Peak height	Peak area %
1	5.757	48562	3721	0.205
2	7.45	23665138	2170088	99.795
In total		23713700	2173809	100

Compound 3m: UV detection at 254 nm; elution, MeOH (100%); T = 25°C; Flow rate = 1.0 mL/min.

Purity: 99.4%



11. Reference

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