# Tryptic shaving of Staphylococcus aureus unveils immunodominant epitopes on the bacterial cell surface 

Annette Dreisbach ${ }^{1, \#}$, Min Wang ${ }^{1, \#}$, Magdalena M. van der Kooi-Pol ${ }^{1, \#}$, Ewoud Reilman ${ }^{1, \#,}$ Dennis G.A.M. Koedijk ${ }^{1}$, Ruben A.T. Mars ${ }^{1,2}$, José Duipmans ${ }^{3}$, Marcel Jonkman ${ }^{3, \dagger}$, Joris J. Benschop ${ }^{4}$, Hendrik P.J. Bonarius ${ }^{5}$, Herman Groen ${ }^{5}$, Michael Hecker ${ }^{6}$, Andreas Otto ${ }^{6}$, Katrin Bäsell ${ }^{6}$, Jörg Bernhardt ${ }^{6}$, Jaap Willem Back ${ }^{4}$, Dörte Becher ${ }^{6}$, Girbe Buist ${ }^{1}$ and Jan Maarten van Dij1 $1^{1, *}$<br>${ }^{1}$ Department of Medical Microbiology, University of Groningen, University Medical Center Groningen, Hanzeplein 1, P.O. box 30001, 9700 RB Groningen, the Netherlands<br>${ }^{2}$ current address: Division of Gastroenterology and Hepatology, Mayo Clinic, Rochester, MN, 55902, USA<br>${ }^{3}$ Department of Dermatology, University of Groningen, University Medical Center Groningen, Hanzeplein 1, Groningen, the Netherlands<br>${ }^{4}$ Pepscan Therapeutics BV, P.O. Box 2098, 8203 AB Lelystad, the Netherlands<br>${ }^{5}$ IQ Therapeutics, Rozenburglaan 13a, Groningen, the Netherlands<br>${ }^{6}$ Institut für Mikrobiologie, Ernst-Moritz-Arndt Universität Greifswald, Friedrich-Ludwig-Jahn-Str. 15, D-17489 Greifswald, Germany<br>\#These authors contributed equally to this work<br>${ }^{\dagger}$ Deceased<br>*To whom correspondence should be addressed: J.M.van.Dij101@umcg.nl

## Table of Contents

Supplementary Figure S1. Original Western blots ............................................................... 2
Supplementary Figure S2. Comparison of the $\mathrm{p} I$ values of identified proteins and peptides in the different sub-proteome fractions 3
Supplementary Figure S3. Predicted subcellular localization of the proteins identified with
the different approaches for strains USA300 and Newman........................................... 4
Supplementary Figure S4. Comparative overview of the results from the Pepscan analysis. 5
Supplementary Figure S5. Comparison of the results from the proteomic studies and epitope mapping.
Supplementary Table S1. Results from the mass spectrometric analyses for the samples derived from S. aureus strain USA300.
Supplementary Table S2. Results from the mass spectrometric analyses for the samples derived from $S$. aureus strain Newman.
Supplementary Table S3. Comparison of the peptide identifications for $S$. aureus strains USA300 and Newman.
Supplementary Table S4. Results from the Pepscan analysis.
Supplementary Table S5. Comparison of epitope mapping with the peptide identifications from the shaving and exoproteome analyses.

## Supplementary Figure S1.



Supplementary Figure S1. Original Western blots. Panel A shows the full blot used to generate Figures 3A, panel B shows the full blot used to generate Figure 3B, and panel C shows the full blot used to generate Figure 4B.

## Supplementary Figure S2.



Supplementary Figure $S 2$. Comparison of the $\mathrm{p} I$ values of identified proteins and peptides in the different sub-proteome fractions. The percentages of protein ( $\mathbf{A}, \mathbf{B}$ ) and peptide ( $\mathbf{C}$, D) identifications with a certain $\mathrm{p} I$ are displayed for the different sub-proteome fractions of strains Newman (A, C) and USA300 (B, D).
$\square$ shaving $1, \mathbb{*}$ shaving 2 , $\mathbb{Z}$ cell wall, $\square$ exoproteome.

## Supplementary Figure S3.



Supplementary Figure S3. Predicted subcellular localization of the proteins identified with the different approaches for strains USA300 and Newman. The overlapping as well as unique proteins identified for the four different sub-proteome fractions from strains USA300 and Newman were analyzed regarding their predicted subcellular localization. CW, covalently cell wall-bound proteins; M, transmembrane- and lipoproteins; Sec, secreted proteins; and No, proteins with no predicted motif for subcellular localization.


Supplementary Figure S4. Comparative overview of the results from the Pepscan analysis. The binding of antibodies from seven Epidermolysis bullosa patients to immobilized overlapping 15 -mer peptide sequences from 54 different staphylococcal surface-exposed proteins were analyzed. For most of the proteins the overlap of consecutive peptides was 11 amino acids. For LytM and IsaA the overlap was 14 amino acids (high resolution, HR). Peptides were synthesizes linearly and with the CLIPS ${ }^{\text {TM }}$ technology. The quantified signals were normalized to the signal average per chip and are presented in a bee swarm plot, where the respective proteins are indicated on the $y$-axis and their normalized signal intensities on the x -axis. EB01, •EB09, • EB51, • EB11, • EB15, • EB53, • EB02.

## Supplementary Figure S5.

## Legend



Figure S5. Comparison of the results from the proteomic studies and epitope mapping. The locations of peptides in particular proteins as identified by the analyses of four different sub-proteome fractions of $S$. aureus strains USA300 and Newman are highlighted in the respective linearly depicted protein sequences. In addition, known protein domains are indicated. The graphs display the signals from the epitope mapping normalized to the median signal of the respective protein and serum for the seven different EB patients.

## Cell Wall-Anchored Proteins

## Clumping factor B , ClfB (Q2FDM9, CLFB)



Iigand_A_binding_region (e)


Fibronectin binding protein A, FnbA (Q2FE03, A6QJY9)


Newman
(truncated)


Iron-regulated surface protein B, IsdB (Q2FHV2, ISDB)


Serine-aspartate repeat-containing protein D, SdrD (Q2FJ78)


## Lipoproteins

Cell wall binding lipoprotein (YkyA-like) (Q2FHY8, A6QFU8)

shaving 1
USA300
shaving 2

shaving 1
Newman



## Foldase protein, PrsA (Q2FFQ5, PRSA)


culture supernatant
USA300
shaving 2
culture supernatant
Newman


Oligopeptide permease peptide binding protein (Q2FE32, A6QJV4)



Peptide ABC transporter, peptide binding protein (Q2FKI7)


Putative Zinc-binding metalloprotease (Q2FJP2, A6QE54)



## Zn-binding lipoprotein AdcA-like protein (Q2FE92, A6qjp6)


culture supernatant
USA300
shaving 1
shaving 2

culture supernatant
Newman shaving 2



## Membrane Proteins

Cell divison protein FtsL (Q2FHQ7, A6QG80)


## Elastin binding protein, EbpS (Q2FGW 1, EBPS)





Glycerol phosphate lipoteichoic acid synthase, LtaS (Q2FIS2, A6QF27)



## Am



Penicillin binding protein 2, Pbp2 (Q2FGZ0)



## Penicillin binding protein 2', Pbp2A(Q2FKM6)



Uncharacterized protein (Q2FE65)


## Secreted Proteins

Bifunctional autolysin (Q2FI25, A6QFR2)

culture supernatant
shaving 1
shaving 2

cell wall
Newman
shaving 1
shaving 2
domains


## Chemotaxis inhibitory protein, CHIPS (Q2FFF7, CHIPS)



## Extracellular matrix binding protein, Emp( EMP)



Fibrinogen binding related protein (Q2FHS8, A6QG56)


## FPRL1 inhibitory protein, FLIPR (FLIPR)



## signal



IgG binding protein, Sbi (Q2FE79, SBI)


culture supernatant
USA300
shaving 1
shaving 2
culture supernatant
cell wall
shaving 1
shaving 2
domains

Lipase, Lip (Q2FDJ1)





N-acetylmuramoyl-L-alanine amidase, family 4 (A6QJE7)
Newman
domains
100

10

0.1


Peptidoglycan hydrolase, LytM (Q2FJZ4, A6QDQ0)



## Staphylocoagulase, Coa (Q2FK40, A6QDK6)



Staphylococcal enterotoxin-like toxin (A6QE90)



Staphylococcal secretory antigen, SsaA (Q2FEJ4, A6QJD9)

culture supernatant
USA300
cell wall
shaving 1
shaving 2

culture supernatant
Newman
cell wall
shaving 1
shaving 2

Truncated putative cell wall surface anchor protein, truncated SasG (Q2FE08)



## uncharacterized protein (Q2FJ23, A6QES5)



## "Cytosolic" Proteins

## CsbD-like protein (A6QHG7)

 shaving 2

Newman


CsbD-like superfamily protein (Q2FIG2, A6QFC3)


## DNA-binding protein HU (Q2FGW9, A6QH22)



Elongation factor, Ef-G (Q2FJ93, EFG)

culture supernatant USA300
cell wall
shaving 1
shaving 2
cell wall Newman
shaving 1
shaving 2


## Elongation factor Ts, Ef-Ts (Q2FHI1, EFTS)


culture supernatant
USA300

shaving 1
Newman


## Elongation factor Tu, Ef-Tu (Q2FJ92, EFTU)



## Enolase, Eno (Q2FIL7, ENO)


culture supernatant
USA300
cell wall
shaving 1
shaving 2

cell wall
Newman
shaving 1
shaving 2


Fructose-bisphosphate-aldolase, Alf1 (Q2FF03, A6QIW9)

shaving 2 USA300

shaving 2
Newman


Glyceraldehyde 3-phosphate dehydrogenase GapDH (Q2FIM1, A6QF81)

culture supernatant USA300
cell wall
shaving 1
shaving 2
shedding

cell wall
Newman

shaving 1

shaving 2


Glycine cleavage system H protein, GcsH (Q2FII7)

culture supernatant
USA300


## Mqo malate:quinone oxidoreductase (Q2FDQ3, A6QK94)


cell wall
USA300

shaving 1
shaving 2

shaving 2
Newman


Phosphoglycerate kinase, Pgk (Q2FIM0)


## Thioredoxin like (Q2FII9, A6QFB4)


culture supernatant USA300
shaving 1

shaving 2
Newman


Triosephosphate isomerase, TpiS (Q2FIL9, TPIS)
 culture supernatant USA300 cell wall shaving 2

cell wall
shaving 2


## universal stress protein, Y1819 (Q2FG28, A6QHP4)



UPF0337protein (A6QHG6)


## Virulence factor, EsxA (Q2FJY6)


culture supernatant
USA300

cell wall

shaving 1



## 30S ribosomal protein S5, RS5 (Q2FEQ6, RS5)



30S ribosomal protein S7, RS7 (Q2FJ94, RS7)

culture supernatant
USA300
 shaving 2
shaving 2
Newman

## 30S ribosomal protein S13, RS13 (Q2FER3)



culture supernatant
USA300
cell wall
shaving 1
shaving 2
shaving 1
Newman
shaving 2


