Supporting Information

Synthesis and Biological Activity of 2,22-Dimethylene Analogues of 19-Norcalcitriol and Related Compounds

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Purity criteria for the synthesized vitamin D compounds

All vitamin D analogues synthesized by us gave single sharp peaks on HPLC and they were judged at least 99% pure. Two HPLC systems (straight- and reversed-phase) were employed as indicated in the Table 2. The purity and identity of the synthesized vitamins were additionally confirmed by inspection of their ¹H NMR and high-resolution mass spectra.

	Compd. No.	HPLC Retention Volumes	
Compound		Straight-phase ^a	Reversed-phase ^b
		(hexane/2-propanol)	(methanol/water)
(20 <i>R</i>)-1a-Hydroxy-2,22-dimethylene-	Q	h/p (95:5)	m/w (94:6)
19-norvitamin D ₃	o	35 mL	37 mL
(20 <i>R</i>)-1α,25-Dihydroxy-2,22-dimethylene-	9	h/p (95:5)	m/w (87:13)
19-norvitamin D ₃		36 mL	29 mL
(20 <i>R</i>)-1α,25-Dihydroxy-2,22-dimethylene-	10	h/p (92:8)	m/w (85:15)
24-homo-19-norvitamin D ₃		45 mL	52 mL
(20 <i>R</i>)-1α,25-Dihydroxy-2,22-dimethylene-	11	h/p (95:5)	m/w (90:10)
26,27-dihomo-19-norvitamin D ₃		27 mL	29 mL
(20 <i>R</i>)-1α,25-Dihydroxy-2,22-dimethylene-	12	h/p (92:8)	m/w (90:10)
24,26,27-trihomo-19-norvitamin D_3		25 mL	38 mL
(20 <i>R</i>)-1α-Hydroxy-2,22-dimethylene-	13	h/p (90:10)	m/w (85:15)
24-(1'-hydroxycyclopentyl)-19,25,26,27- tetranorvitamin D_3		40 mL	50 mL

Table 2. Purity Criteria for Target Vitamin D Compounds

^aZorbax-Sil; 9.4 mm \times 25 cm column; ^bZorbax Eclipse XDB-C18; 9.4 mm \times 25 cm column.

Conformational analysis of model 8-methylene des-A,B-compounds BM-FM

Structural analysis of model 8-methylene *des*-A,B-compounds **BM-FM**, performed according to a protocol described by us previously in detail,¹ in all cases gave similar results. They are exemplified by Figure 1S showing the "dot map" of model compound **BM** characterized by the presence of the side chain identical as in the planned 2MD analog **9**. Thus, in the low-energy side chain conformers of the examined compound, falling into the 1 kcal/mol energy window, the 25-oxygen resides in the spatial regions EG and EA; the former rear region (EG) is somewhat less active but the latter front region EA is considered as the most active one.²



Figure 1S. The structures of the model 8-methylene compounds **BM-FM** possessing side chains identical to those of the synthesized analogs **9-13**. Stereo-view illustrating side-chain conformational analysis of compound **BM**. Energy-minimized conformations of this compound (energy window 1 kcal/mol) were overlaid; the circles show the location of 25-oxygen atoms in the corresponding conformers. Side-chain carbons and hydrogen atoms are omitted for clarity. There are also indicated spatial regions EG, G, EA, and A as defined by Yamada.²

Preparation of (20*R*)-8β-[(triethylsilyl)oxy]-*des*-A,B-23,24-dinorcholan-22-al (16)

For the preparation of **16** we used slightly modified literature procedures.^{3,4} Thus, the Inhoffen-Lythgoe diol **i**, easily obtained from vitamin D_2 ,⁵ was oxidized with TEMPO/NCS system⁶ and the formed hydroxy aldehyde **ii** was subjected to equilibration with piperidine.⁷ The resulting crude mixture of epimeric aldehydes (**iv** and unchanged **ii** at the ratio of 3:1) was directly reduced with NaBH₄ and the formed epimeric alcohols were separated by column chromatography. The minor isomer **i** was recycled and the main 20-epi-Ihoffen-Lythgoe diol (**iii**) oxidized to the hydroxy aldehyde **iv** which after silvlation provided the desired synthon **16**.



Scheme 1S. (a) TEMPO, TBACl, NSC, NaHCO₃/K₂CO₃ buffer pH 8.6, CH₂Cl₂, (ii: 93%, iv: 88%);
(b) piperidine, benzene, 93%; (c) NaBH₄, THF, EtOH, (iii: 75%); (d) TESOTf, 2,6-lutidine, CH₂Cl₂, 88%.

(20*S*)-8β-Hydroxy-*des*-A,B-23,24-dinorcholan-22-al (ii). To the vigorously stirred solution of Inhoffen-Lythgoe diol i [1.37 g, 6.48 mmol; prepared as described by Grzywacz et al.⁵ in CH₂Cl₂ (50 mL) and NaHCO₃/K₂CO₃ buffer (pH 8.6; 50 mL), TEMPO (101 g, 647 μ mol) was added followed by TBACl (180 mg, 1.64 mmol) and NCS (1.731 g, 12.9 mmol). The reaction mixture was stirred overnight at the room temperature, poured into water and extracted with CH₂Cl₂. The combined extracts were dried over Na₂SO₄ and concentrated. Purification by column chromatography on silica with hexane/ethyl acetate (7:3) yielded aldehyde ii (1.27 g, 93%) as a colorless oil which spectral data were identical with these reported in the literature.⁴

(20*R*)-*des*-A,B-23,24-Dinorcholano-8 β ,22-diol (iii). Solution of aldehyde ii (378 mg, 1.8 mmol) in anhydrous benzene (9 mL) and piperidine (213 μ L, 2.15 mmol) was refluxed for 8 h using Bidwell-Sterling trap. The solvents were removed on the rotary evaporator. Spectral analysis (¹H NMR, 500 MHz, CDCl₃) of the crude products obtained in 93% yield confirmed a presence of two isomeric aldehydes: iv [9.542 ppm (1H, d, *J* = 5.1 Hz, 22-H)]⁴ and unchanged ii [9.575 ppm (1H, d, *J* = 3.1 Hz, 22-H)]⁴ at the ratio of 3:1. This crude mixture of aldehydes was then dissolved in dry THF (4 mL), treated with NaBH₄ (123 mg) and EtOH (4 mL). After 2 h reaction was quenched with water and materials were extracted with ethyl acetate. Combined organics were dried over Na₂SO₄ and concentrated. 20-*Epi*-Inhoffen- Lythgoe diol iii (261 mg, 75%) was isolated by column chromatography on silica with hexane/ethyl acetate (7:3) as white crystals which spectral data were identical with these reported in the literature.⁴

(20*R*)-8 β -[(Triethylsilyl)oxy]-*des*-A,B-23,24-dinorcholan-22-al (16). Diol iii (273 mg, 1.28 mmol) was then subjected to selective oxidation of the primary alcohol according to the procedure described above for the alcohol i. The obtained hydroxy aldehyde iv (242 mg, 88%), which spectral data were identical with these reported in the literature,⁴ was dissolved in anhydrous CH₂Cl₂ (30

mL) containing 2,6-lutidine (395 μ L, 3.41 mmol). To this solution triethylsilyl trifluoromethanesulfonate (715 μ L, 3.16 mmol) was added dropwise at -40 °C and the mixture was stirred for 30 min. After addition of saturated NaHCO₃, the mixture was allowed to warm up to room temperature, and then it was extracted with CH₂Cl₂, dried (Na₂SO₄) and concentrated. The residue was purified by column chromatography on silica with hexane/ethyl acetate (95:5) to give silylated compound **16** (329 mg, 88%) as a colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 0.536 (6[H, q, *J* = 7.8 Hz, 3 × SiCH₂), 0.909 (3H, s, 18-H₃), 0.926 (9H, t, *J* = 7.8 Hz, 3 × SiCH₂C<u>H₃</u>), 0.997 (3H, d, *J* = 6.8 Hz, 21-H₃), 4.03 (1H, narr m, 8α-H), 9.53 (1H, d, *J* = 10 Hz, C<u>H</u>O); ¹³C NMR (100 MHz, CDCl₃) δ 5.2, 7.1, 13.6, 14.8, 17.6, 22.6, 25.8, 34.7, 39.7, 41.9, 48.5, 52.8, 52.9, 69.1, 206.1; HRMS (ESI) exact mass calculated for C₁₉H₃₇O₂Si (M⁺ + H) 325.2562, found 325.2574.

Preparation of bromides (A-G) and the corresponding Grignard reagents



Figure 2S

1-Bromo-3-methyl-3-[(triethylsilyl)oxy]butane (B) and 1-bromo-3-ethyl-3-[(triethylsilyl)oxy]pentane (D).⁸ Commercially available ethyl-3-bromopropionate was treated with methyl magnesium bromide or ethyl magnesium bromide, respectively, according to the standard procedures described in the literature.⁹ The resultant tertiary alcohols were next silylated by treatment with TESOTf and 2,6-lutidine.⁹

1-Bromo-4-methyl-4-[(triethylsilyl)oxy]pentane (C) and 1-bromo-4-ethyl-4-[(triethylsilyl)oxy]hexane (E). Analogously, as described above for **B** and **D**, these bromides were obtained from commercially available ethyl-4-bromobutyrate in the reaction with the corresponding Grignard reagents followed by protection of the formed tertiary alcohols with TESOTf and 2,6-lutidine.

1-(2'-Bromoethyl)-1-[(triethylsilyl)oxy]cyclopentane (F) was obtained from commercially available β -propiolactone by the four-step procedure involving: (a) the reaction with the Grignard reagent derived from 1,4-dibromobutane,^{9,10} (b) tosylation of the primary alcohol in the formed product using *p*-toluenesulfonyl chloride in pyridine, (c) conversion of the obtained tosylate to the bromide¹¹ (Adv. Synth. Catal. 2012, 354, 1519-28), and (d) protection of tertiary

alcohol with TESOTf and 2,6-lutidine.

(a) 1,4-Dibromobutane (3.82 mL, 32 mmol) was slowly added to the flask containing magnesium turnings (1.75 g, 73 mmol) in anhydrous THF (72 mL) at 0 °C under argon. After addition was complete, the mixture was stirred at room temperature for 30 min. Freshly generated Grignard reagent was then cooled to -40 °C and β -propiolactone (1.81 mL, 28.78 mmol) was slowly added with stirring. Cooling bath was removed after 20 min, the mixture was allowed to reach room temperature and the stirring was continued overnight. Reaction was quenched by addition of sat. aq. NH₄Cl solution, then it was extracted with ethyl acetate, dried over Na₂SO₄ and concentrated. Purification by column chromatography on silica (0 \rightarrow 100% ethyl acetate/hexane) afforded 1-(2-hydroxyethyl)cyclopentanol (2.34 g, 64%).

(b) *p*-Toluenesulphonyl chloride (2.21 g, 11.6 mmol) was added to a stirred solution of the prepared diol (1.171 g, 9 mmol) and DMAP (76 mg, 0.62 mmol) in anh. CH₂Cl₂ (70 mL) containing TEA (4.7 mL) at 0 °C under argon. Reaction mixture was allowed to reach room temperature and stirring was continued for 2 h. Reaction was quenched by addition of water followed by 2% HCl aq. sol.. The mixture was extracted with CH₂Cl₂, dried over Na₂SO₄, and concentrated. Purification on silica column yielded 2-(1-hydroxycyclopentyl)ethyl 4-methylbenzenesulfonate (2.36 g, 92%): ¹H NMR (400 MHz, CDCl₃) δ 1.353 (1H, s), 1.97 (2H, t, *J* = 6.8 Hz), 2.45 (3H, s), 4.24 (2H, t, *J* = 6.8 Hz), 7.34 and 7.79 (2H and 2H, each d, each *J* = 8.2 Hz, Ar-H); ¹³C NMR (100 MHz, CDCl₃) δ 21.6, 23.4, 39.6, 40.0, 68.0, 80.5, 127.9, 129.8, 132.9, 144.8; HRMS (ESI) exact mass calculated for C₁₄H₂₁O₄S (M⁺ + H) 285.1160, found 285.1171.

(c) Mixture of the tosylate (50 mg, 0.176 mmol) and anhydrous LiBr in anhydrous THF (1 mL)

was refluxed for 1 h. Oil bath was removed, mixture was allowed to reach room temperature and water was added. Materials were extracted with ethyl acetate, dried over Na₂SO₄, and concentrated. Purification on silica column yielded 1-(2-bromoethyl)cyclopentanol (32 mg, 95%); ¹H NMR (400 MHz, CDCl₃) δ 1.64 (6H, m), 1.71 (2H, m), 2.20 (2H, t, *J* = 8.1 Hz), 3.54 (2H, t, *J* = 8.1 Hz); ¹³C NMR (100 MHz, CDCl₃) δ 23.4, 28.9, 39.7, 44.7, 82.3; HRMS (ESI) exact mass calculated for C₇H₁₃BrONa (M⁺ + Na) 215.0047, found 215.0053.

(d) 2,6-Lutidine (940 µL, 8.135 mmol) was added to the solution of the prepared cyclopentanol (1.41 g, 7.33 mmol) in dry CH₂Cl₂ (20 mL) at -40 °C under argon followed by TESOTf (1.86 mL, 8.24 mmol). Reaction was stirred for 4 h, brine was added and the mixture was extracted with CH₂Cl₂. The combined organics were washed with 2% HCl aq. sol., dried over Na₂SO₄, and concentrated. Purification by column chromatography on silica eluted with hexane/ethyl acetate (95:5) afforded 1-(2'-bromoethyl)-1-[(triethylsilyl)oxy]cyclopentane (**F**; 2.19 g, 97%) as a colorless oil: ¹H NMR (500 MHz, CDCl₃) δ 0.590 (6H, q, *J* = 6.7 Hz), 0.949 (9H, t, *J* = 6.7 Hz), 1.56 (6H, m), 1.72 (2H, m), 2.12 (2H, m), 3.48 (2H, m); ¹³C NMR (125 MHz, CDCl₃) δ 6.5, 7.1, 23.3, 29.3, 39.7, 45.8, 84.5; HRMS (ESI) exact mass calculated for C₁₁H₂₂BrOSi (M⁺ - Et) 277.0618, found 277.0628.

The ethereal solutions of the corresponding Grignard reagents were prepared from the bromides **A-F** and magnesium powder using standard procedure, analogous to that described below for the isopentyl magnesium bromide. Although this Grignard reagent was commercially available (Aldrich), it was freshly prepared by us from the bromide **A**.

1-Bromo-3-methylbutane (A; 393 μ L, 3.28 mmol) was added dropwise to a stirred mixture of magnesium powder (78 mg, 3.21 mmol) in anhydrous THF (6 mL) under argon at room

temperature. After addition was complete the mixture was stirred at room temperature for 1 h. Then

the prepared solution of isopentyl magnesium bromide was added to the solution of the aldehyde

16 as described in Experimental Section.

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Figure 38. ¹H NMR spectrum of the vitamin D analogue 8



Figure 4S. ¹³C NMR spectrum of the vitamin D analogue 8



Figure 5S. ¹H NMR spectrum of the vitamin D analogue 9



Figure 6S. ¹³C NMR spectrum of the vitamin D analogue 9



Figure 78. ¹H NMR spectrum of the vitamin D analogue 10



Figure 8S. ¹³C NMR spectrum of the vitamin D analogue 10



Figure 9S. ¹H NMR spectrum of the vitamin D analogue 11



Figure 10S. ¹³C NMR spectrum of the vitamin D analogue 11



Figure 11S. ¹H NMR spectrum of the vitamin D analogue 12



Figure 12S. ¹³C NMR spectrum of the vitamin D analogue 12



Figure 13S. ¹H NMR spectrum of the vitamin D analogue 13



Figure 14S. ¹³C NMR spectrum of the vitamin D analogue 13








































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