Supporting Information

Effects of microstripe geometry on guided cell migration

Xiang Yao, Jiandong Ding*

State Key Laboratory of Molecular Engineering of Polymers, Department of Macromolecular Science, Fudan University, Shanghai 200438, China

*Correspondence should be addressed to J.D. DING. E-mail: <u>jdding1@fudan.edu.cn</u>

Supplementary Materials include 17 figures, 13 tables and 3 movies. In this file, the figures and tables are presented in the sequence of their citations in the main manuscript. The movies are shown as three separate video files.

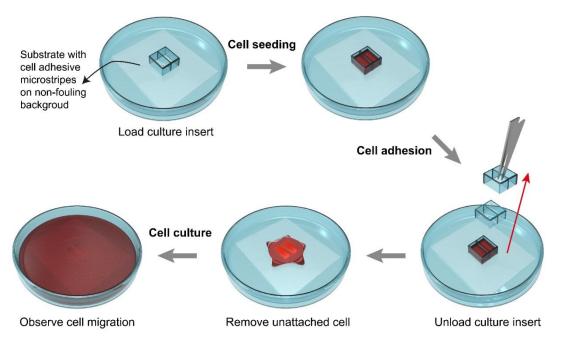


Figure S1. Schematic presentation of the procedure to observe the microstripe-guided cell migration by using a surface patterning technique and a cell culture insert.

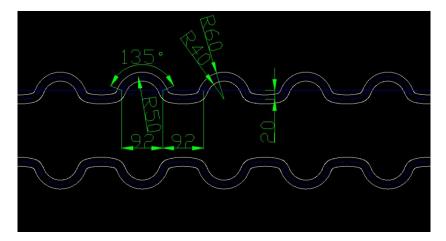


Figure S2. Original design drawing of the combinatory microstripe R50. The white lines here are the corresponding edges of combinatory microstripe R50, the green lines here are the auxiliary lines for the pattern designing with the indicated radiuses (R) and lengths in units of micrometer.

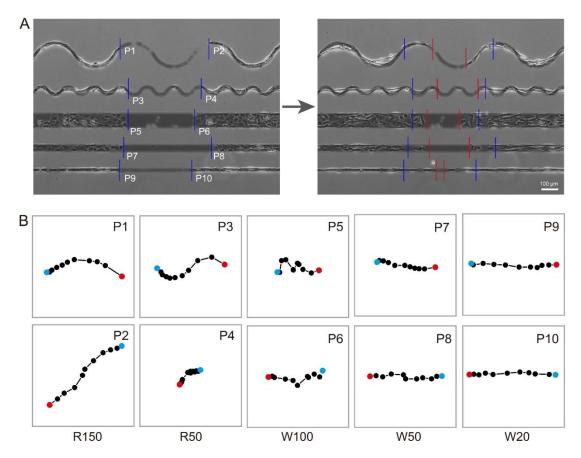


Figure S3. Cell migration along the typical straight and wavy microstripes in one view field in an optical microscope. (A) Phase contrast micrograph to show the relative positions of cell clusters (P1 – P10) at the initial and final states for Movie S1. The blue and red vertical lines indicate the corresponding initial and final positions of the fronts of the cell clusters, respectively. (B) Trajectories of the fronts of the indicated cell clusters during 5.8 h. Each site represents a time-dependent position of the cell fronts; the blue and red dots indicate the corresponding initial and final site, respectively. Each square in (B) has side length of about 280 μ m.

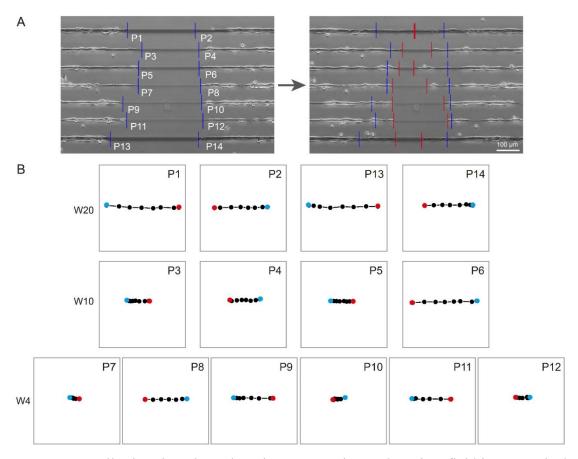


Figure S4. Cell migration along the micropatterns in another view field in our optical microscope to demonstrate typical cases of straight microstripes of widths equal to or less than 20 μ m. (A) Phase contrast micrograph to show the relative positions of cell clusters (P1 – P14) at the initial and final states for Movie S2. The blue and red vertical lines indicate the corresponding initial and final positions of the fronts of the cell clusters, respectively. In this independent experiment, the gap distance (about 300 μ m) is smaller than that in a regular experiment (about 500 μ m). So, we only show the 4 h data for the gap region here, while the statistical results of migration velocity in other figures are still from the 6 h data from other ROI (side regions), as shown at the top-left corner of Figure 2. (B) Trajectories of the fronts of the indicated cell clusters during 4.1 h. Each square in (B) has side length of about 200 μ m.

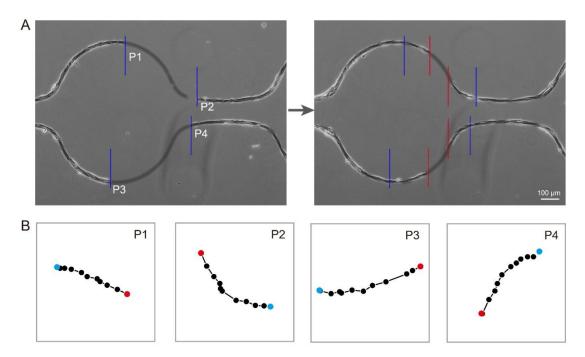


Figure S5. Cell migration along the combinatory microstripes of R400 in one view field in our optical microscope. (A) Phase contrast micrograph to show the relative positions of cell clusters (P1 – P4) at the initial and final states for Movie S3. The blue and red vertical lines indicate the corresponding initial and final positions of the fronts of the cell clusters, respectively. (B) Trajectories of the fronts of the indicated cell clusters during 5.8 h. Each square in (B) has side length of about 300 μ m.

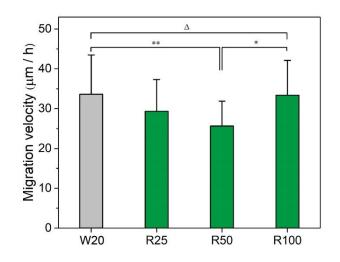


Figure S6. Statistical results of migration velocities of rMSCs on wavy microstripes with arc radiuses of 25 μ m (R25), 50 μ m (R50) and 100 μ m (R100) during the initial 6 h. Here the group of the straight microstripe with width of 20 μ m (W20) is as control. The widths of wavy microstripes are all 20 μ m and the indicated R's are actually the arc radiuses of the middle arcs of the corresponding wavy stripes. The symbols indicate the significance or insignificance of difference between two indicated groups. " Δ ": p > 0.05; "*": p < 0.05; "**": p < 0.01.

Groups	W4	W10	W20	W50	W100
W4		9.7E-7	3.3E-10	0.85	0.02
		***	***	Δ	*
W10	9.7E-7		0.34	0.003	4.0E-8
	***		Δ	**	***
W20	3.3E-10	0.34		2.1E-4	3.2E-10
	***	Δ		***	***
W50	0.85	0.003	2.1E-4		0.04
	Δ	**	***		*
W100	0.02	4.0E-8	3.3E-10	0.04	
	*	***	***	*	

Table S1. The *p* values of one-way ANOVA of the data in Figure 3A for migration of rMSCs on straight microstripes with different widths.

The *p* value of a global test among W4, W10 and W20 is 1.2E-9 (***).

The p value of a global test among W20, W50 and W100 is 1.8E-12 (***).

" Δ ": p > 0.05, no significant difference; "*": p < 0.05, significant difference;

Groups	W20	R50	R150
W20		0.008	0.20
		**	Δ
R50	0.008		0.003
	**		**
R150	0.20	0.003	
	Δ	**	

Table S2. The *p* values of one-way ANOVA of the data in Figure 3B for migration of rMSCs on W20 and wavy microstripes with different arc radiuses.

" Δ ": p > 0.05, no significant difference; "*": p < 0.05, significant difference;

"**": p < 0.01, significant difference; "***": p < 0.001, very significant difference.

Table S3. The *p* values of one-way ANOVA of the data in Figure 3C for migration of rMSCs on W20 and combinatory microstripes with different arc radiuses.

W20	R50	R150	R400
L-R vs R-L	CCW vs CW	CCW vs CW	CCW vs CW
0.98	0.44	0.15	0.04
Δ	Δ	Δ	*

" Δ ": p > 0.05, no significant difference; "*": p < 0.05, significant difference;

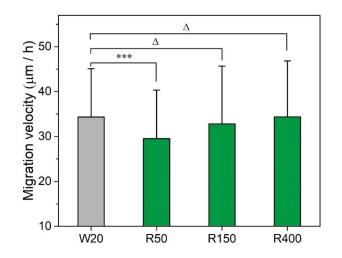


Figure S7. Statistical results of migration velocities of rMSCs on combinatory microstripes of varied arc radiuses during the initial 6 h. CW migration and CCW migration were not distinguished here. The symbols indicate the significance or insignificance of difference between two indicated groups. " Δ ": p > 0.05; "***": p < 0.001.

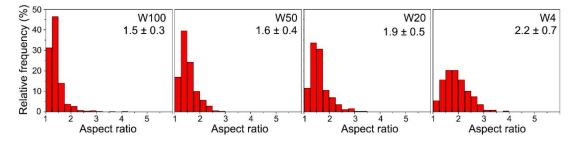


Figure S8. Statistical results of nuclear aspect ratios of rMSCs after 5 h of cell seeding on the indicated straight microstripes. Data on the top-right corner are mean \pm SD of the corresponding aspect ratios of cell nuclei.

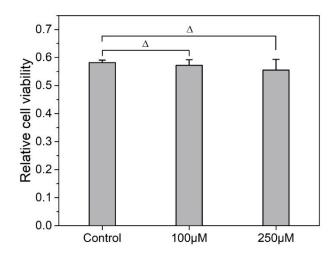


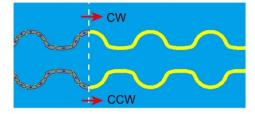
Figure S9. Relative cell viability of rMSCs cultured on tissue culture plates (TCPs) without or with a gap-junction inhibitor Gap-27 of the indicated concentrations. The symbols indicate the significance or insignificance of difference between two indicated groups. " Δ ": p > 0.05.

W20	R50	R150	R400
L-R vs R-L	CCW vs CW	CCW vs CW	CCW vs CW
0.72	0.31	0.94	0.91
Δ	Δ	Δ	Δ

Table S4. The *p* values of one-way ANOVA of the data in Figure 5B for migration of rMSCs on combinatory microstripes and W20 in presence of Gap-27.

" Δ ": p > 0.05, no significant difference

One independent event (CCW vs CW)



Without Gap-27

Percentage of indicated independent events

	CCW>CW	CCW <cw< th=""><th>CCW=CW</th></cw<>	CCW=CW
R50	48.3%	43.1%	8.6%
R150	51.7%	33.3%	15.0%
R400	61.4%	33.3%	5.3%

With Gap-27

Percentage of indicated independent events

	CCW>CW	CCW <cw< th=""><th>CCW=CW</th></cw<>	CCW=CW
R50	44.6%	47.3%	8.1%
R150	48.6%	45.8%	5.6%
R400	46.8%	46.8%	6.4%

Figure S10. Comparison between CW and CCW events of migration of rMSCs on paired combinatory microstripes during the initial 6 h trying a modified range of criteria. Schematic presentation of one independent event for the comparison of CW and CCW migrations (two adjacent combinatory microstripes with the same arc radius) and corresponding statistical results of fractions of independent events under the conditions without or with Gap-27. Here, we have modified the evaluation criteria of "CCW = CW" as $95\% \times CW \le CCW \le 105\% \times CW$, different from $99\% \times CW \le$ $CCW < 101\% \times CW$ in Figure 3D in the main manuscript. On each pair of microstripes as seen in the above image, an independent cellular event was defined via comparison between a CW migration on one microstripe and a CCW migration on its adjacent microstripe: a cellular event was regarded as "CCW > CW" if the migration velocity of CCW be 1.05 times higher than that of CW; if the migration velocity of CCW be 0.95 times lower than that of CW, this independent event was marked as "CCW < CW". The modification of the criteria range from (99%, 101%) to (95%, 105%) was confirmed not to alter the conclusion about migration chirality under the same experimental condition.

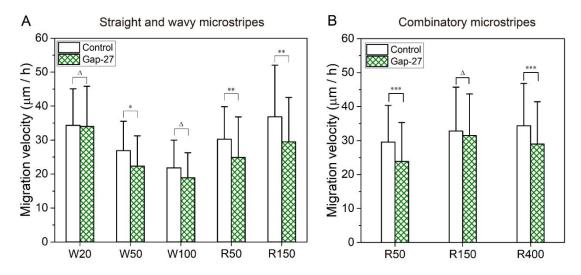


Figure S11. Migration velocities of rMSCs on typical microstripes with or without Gap-27 during the initial 6 h. (A) Statistical results of migration velocities on typical straight (W20, W50, W100) and wavy (R50, R150) microstripes. (B) Statistical results of migration velocities on combinatory microstripes (R50, R150, R400). CW migration and CCW migration on the combinatory microstripes were not distinguished here. The symbols indicate the significance or insignificance of difference between two indicated groups. " Δ ": p > 0.05; "*": p < 0.05; "**": p < 0.01; "***": p < 0.001.

Groups	W4	W10	W20	W50	W100
W4		1.6E-4	5.4E-8	0.15	0.07
		***	***	Δ	Δ
W10	1.6E-4		0.07	7.8E-5	2.3E-5
	***		Δ	***	***
W20	5.4E-8	0.07		2.5E-7	6.7E-8
	***	Δ		***	***
W50	0.15	7.8E-5	2.5E-7		0.74
	Δ	***	***		Δ
W100	0.07	2.3E-5	6.7E-8	0.74	
	Δ	***	***	Δ	

Table S5. The *p* values of one-way ANOVA of the data in Figure 6A for migration of NIH3T3 cells on straight microstripes with different widths.

The *p* value of global test among W4, W10 and W20 is 6.125E-7 (***).

The *p* value of global test among W20, W50 and W100 is 1.743E-11 (***).

"

"p > 0.05, no significant difference; "*":
 p < 0.05, significant difference;

Groups	W4	W10	W20	W50	W100
W4		1.0E-4	1.2E-4	0.33	0.67
		***	***	Δ	Δ
W10	1.0E-4		0.89	0.02	0.004
	***		Δ	*	**
W20	1.2E-4	0.89		0.03	0.005
	***	Δ		*	**
W50	0.33	0.02	0.03		0.61
	Δ	*	*		Δ
W100	0.69	0.004	0.005	0.61	
	Δ	**	**	Δ	

Table S6. The *p* values of one-way ANOVA of the data in Figure 6A for migration of Hela cells on straight microstripes with different widths.

The *p* value of global test among W4, W10 and W20 is 1.2E-4 (***).

The *p* value of global test among W20, W50 and W100 is 0.005 (**).

" Δ ": p > 0.05, no significant difference; "*": p < 0.05, significant difference;

Groups	W20	R50	R150
W20		0.007	0.17
		**	Δ
R50	0.007		0.26
	**		Δ
R150	0.17	0.26	
	Δ	Δ	

Table S7. The *p* values of one-way ANOVA of the data in Figure 6A for migration of NIH3T3 cells on W20 and wavy microstripes with different arc radiuses.

" Δ ": p > 0.05, no significant difference; "*": p < 0.05, significant difference;

"**": p < 0.01, significant difference; "***": p < 0.001, very significant difference.

Table S8. The *p* values of one-way ANOVA of the data in Figure 6A for migration of Hela cells on W20 and wavy microstripes with different arc radius.

Groups	W20	R50	R150
W20		0.002	1.0
		**	Δ
R50	0.002		0.01
	**		*
R150	1.0	0.01	
	Δ	*	

" Δ ": p > 0.05, no significant difference; "*": p < 0.05, significant difference; "**": p < 0.01, significant difference; "***": p < 0.001, very significant difference.

Table S9. The <i>p</i> values of one-way ANOVA of the data in Figure 6B for migration of
NIH3T3 cells on W20 and combinatory microstripes with different arc radiuses.

W20	R50	R150	R400
L-R vs R-L	CCW vs CW	CCW vs CW	CCW vs CW
0.99	0.83	0.88	0.30
Δ	Δ	Δ	Δ

" Δ ": p > 0.05, no significant difference.

Table S10. The *p* values of one-way ANOVA of the data in Figure 6B for migration of Hela cells on W20 and combinatory microstripes with different arc radiuses.

W20	R50	R150	R400
L-R vs R-L	CCW vs CW	CCW vs CW	CCW vs CW
0.75	0.17	0.36	0.87
Δ	Δ	Δ	Δ

" Δ ": p > 0.05, no significant difference.

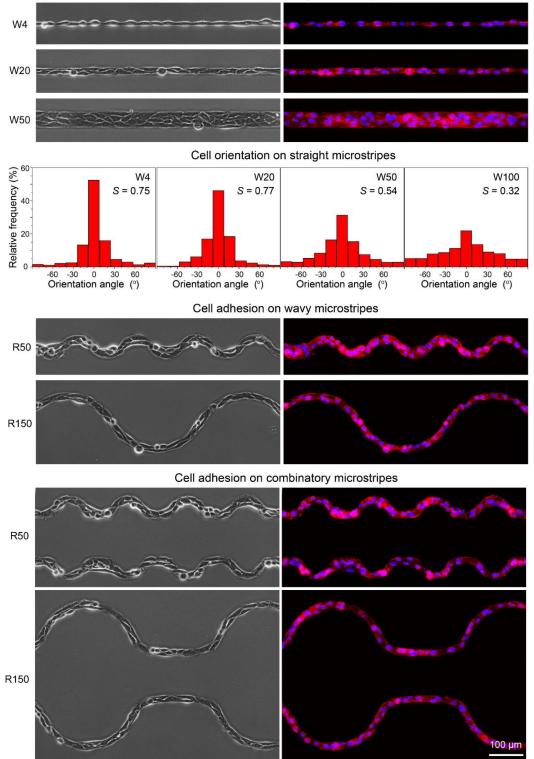


Figure S12. Adhesion of Hela cells on microstripes with varied geometric features. Phase contrast (left column) and corresponding fluorescence (right column) micrographs of cells on typical microstripes were captured 5 h after cell seeding. Color scheme: red, F-actins; blue, nuclei. The data of the histograms came from statistics of nuclear orientation of Hela cells on the indicated straight microstripes.

Cell adhesion on straight microstripes

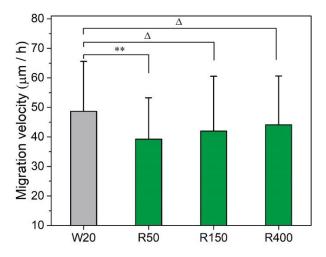


Figure S13. Statistical results of migration velocities of NIH3T3 cells on combinatory microstripes during the initial 6 h. CW migration and CCW migration were not distinguished here. The symbols indicate the significance or insignificance of difference between the two indicated groups. " Δ ": p > 0.05; "**": p < 0.01.

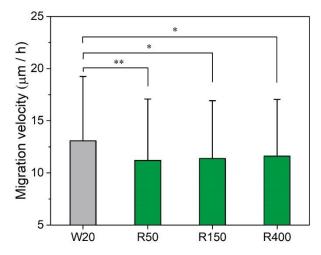


Figure S14. Statistical results of migration velocity of Hela cells on combinatory microstripes during the initial 6 h. CW migration and CCW migration were not distinguished here. The symbols indicate the significance of difference between the two indicated groups. "*": p < 0.05; "**": p < 0.01.

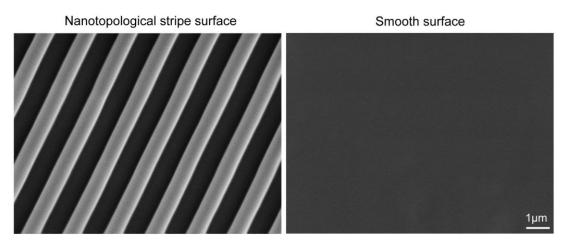


Figure S15. SEM micrographs of petri dishes with a nanotopological stripe surface and a pattern-free smooth surface.

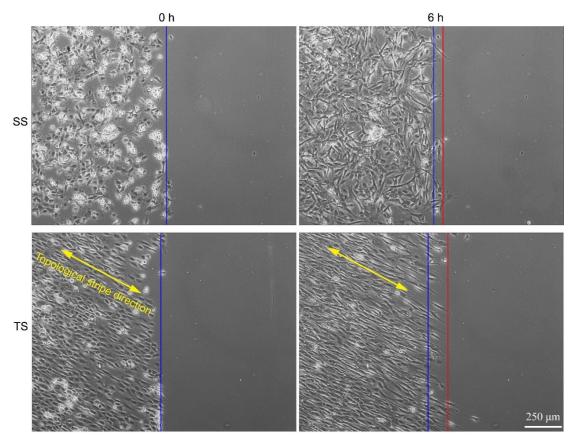


Figure S16. Phase contrast micrographs of migration of rMSCs on nanotopological stripe surface (TS) and smooth surface (SS). "0 h": initial state of the observation of cell migration (about 4 h after cell seeding). "6 h": 6 h after the initial observation. The blue and red lines indicate roughly the positions of the initial and migration fronts, respectively.

Groups	W20	TS	SS
W20		4.0E-5	3.3E-25
		***	***
TS	4.0E-5		1.1E-7
	* * *		***
SS	3.3E-25	1.1E - 7	
	***	***	

Table S11. The *p* values of one-way ANOVA of the data in Figure 7B for migration of rMSCs on W20, nanotopological stripe and smooth surfaces.

"***": p < 0.001, very significant difference.

Table S12. The *p* values of one-way ANOVA of the data in Figure 7B for migration of NIH3T3 cells on W20, nanotopological stripe and smooth surfaces.

Groups	W20	TS	SS
W20		5.0E-7	1.1E-20
		***	***
TS	5.0E-7		7.7E-8
	***		***
SS	1.1E-20	7.7E-8	
	***	***	

"***": p < 0.001, very significant difference.

Table S13. The *p* values of one-way ANOVA of the data in Figure 7B for migration of Hela cells on W20, nanotopological stripe and smooth surfaces.

Groups	W20	TS	SS
W20		0.01	3.7E-8
		*	***
TS	0.01		0.02
	*		*
SS	3.7E-8	0.02	
	***	*	

" Δ ": p > 0.05, no significant difference; "*": p < 0.05, significant difference;

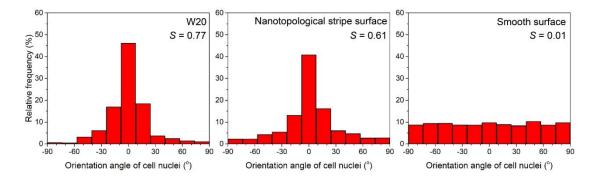


Figure S17. Statistical results of nuclear orientation of Hela cells after 5 h of cell seeding on the indicated surfaces. W20: straight microstripe with width of 20 μ m. *S* refers to order parameter.