

Supporting Information

Citric Acid-Modified Cellulose-Based Tough and Self-Healable Composite Formed by Two Kinds of Non-Covalent Bonding

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1. Experimental details

Materials: The HEA, 12-aminolauric acid, and acryloyl chloride were purchased from Tokyo Chemical Industry Co., Ltd. The γ CD was obtained from Junsei Chemical Co. Ltd. The IRGACURE 184 was purchased from BASF Japan Co., Ltd. The cellulose powder, 1,4-dioxane, ethyl acetate, sodium hydroxide (NaOH), hydrochloric acid (HCl), and sodium sulfate (Na_2SO_4) were purchased from Nacalai Tesque Inc. 1-Butyl-3-methylimidazolium chloride was purchased from Sigma-Aldrich Corporation. The 6-acrylamido methylether- γ CD ($\gamma\text{CDAAmMe}$)¹ and PAc γ CD² were prepared according to our previous report. The citric acid (hydrate) was purchased from Fujifilm Wako Pure Chemical Corporation. The CAC was prepared according to previous report.³ The dimethyl sulfoxide (DMSO)- d_6 was obtained from Merck & Co., Inc. The water used for the preparation of the aqueous solutions was purified with a Millipore Elix 5 system. Other reagents were used without further purification.

Measurements:

NMR spectroscopy: The ^1H and ^{13}C spectra were recorded at 500 MHz with a JEOL JNM-ECA 500 NMR spectrometer at 25°C. In all NMR measurements, the chemical shifts were referenced to the solvent values [^1H NMR : $\delta = 0$ ppm for tetramethylsilane (TMS) and 2.49 ppm for DMSO- d_6 , ^{13}C NMR : $\delta = 0$ ppm for TMS and 39.5 ppm for DMSO- d_6). The ^1H field gradient magic angle spinning (FGMAS) NMR spectra were recorded at 400 MHz with a JEOL JNM-ECA 400WB NMR spectrometer. The sample spinning rate was 7 kHz with a relaxation delay of 10 s at 30°C. Chemical shifts were referenced to hydrogen oxide deuterium (HOD) as a standard ($\delta = 4.79$ ppm). The ^{13}C cross-polarization magic angle spinning (CPMAS) NMR was recorded at 75 MHz in the solid state on a Chemagnetics JNM-CMX300W spectrometer with a sample spinning rate of 4.0 kHz at 30°C.

IR spectrometry: The attenuated total reflectance Fourier-transform infrared spectroscopy (ATR-FTIR) were recorded using JASCO FT/IR-6100 spectrometer in the wavenumber range from 4000 to 400 cm^{-1} in ATR method.

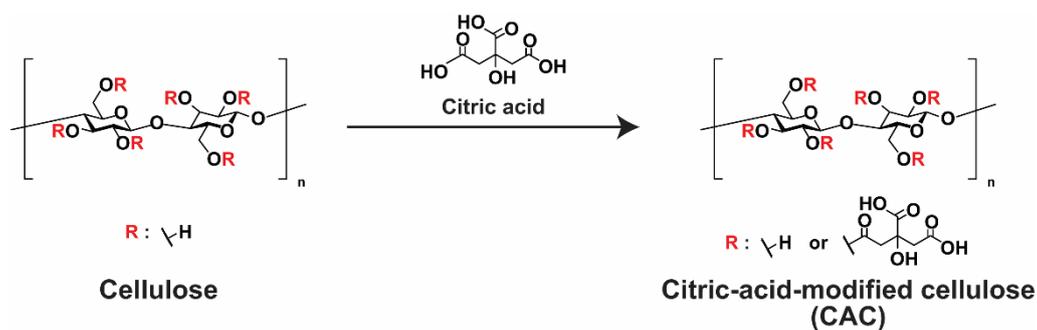
Scanning electron microscope (SEM): SEM images of surfaces were examined with a JEOL JSM-7600F instrument with an accelerating voltage of 2 kV.

Mechanical property measurement: The mechanical properties (fracture energy, Young's modulus, and self-healing) of the CAC-based composites were measured using Autograph AG-X plus (Shimadzu Co.) equipped with a 50 N load cell, at a specific deformation rate ($1 \text{ mm}\cdot\text{s}^{-1}$).

Laser scanning confocal microscope: The laser scanning confocal microscope images were examined using a Keyence Co. VK-X250.

Differential scanning calorimeter (DSC) measurements: Glass transition temperatures (T_g) of the samples were determined by Hitachi High-Technologies Corporation DSC7020 System with a heating rate $10^\circ\text{C}/\text{min}$.

2. Preparation of citric acid-modified cellulose (CAC)



Scheme S1. Preparation of citric-acid-modified cellulose (CAC).

Citric acid-modified cellulose (CAC) were prepared by dispersing 30 g of cellulose in 200 mL deionized water with addition of 1.4 mL 1 M NaOH aqueous solution. The mixture was stirred for several minutes and 90 g citric acid was added. The mixture was heated to 130 °C for 12 h until all the component completely reacted. After the reaction, the water was evaporated. The supernatant was washed with water until unreacted citric acid was completely removed (pH 7) then followed washing with methanol and acetone. CAC was obtained by evaporating under pressure. CAC was characterized by FTIR measurement. The content of carboxylic acid is 1.8 mmol·g⁻¹ (determined by conductivity titration from CAC aqueous dispersion).

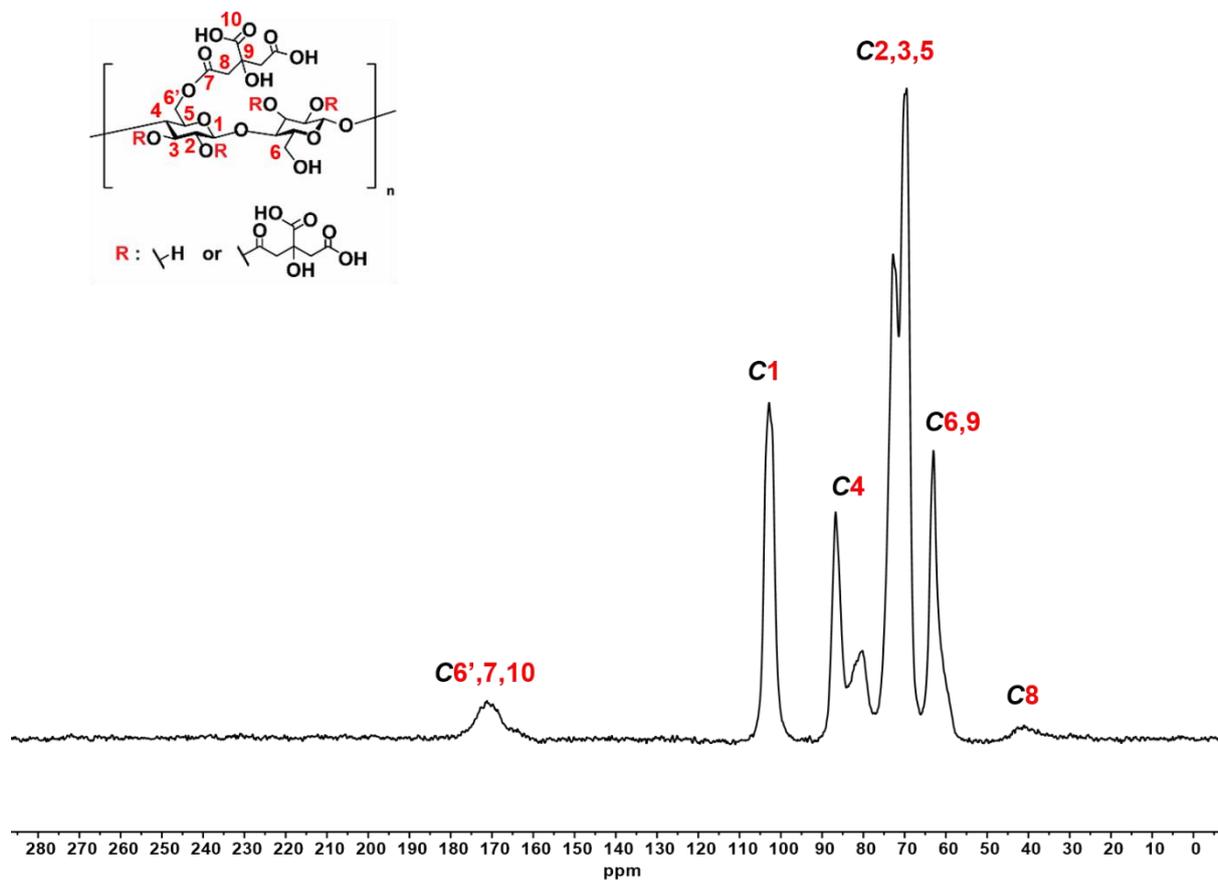


Figure S1. Solid-state ^{13}C CPMAS NMR spectrum of CAC (TMS for standard, 75 MHz, 25 °C, rotation frequency = 7 kHz).

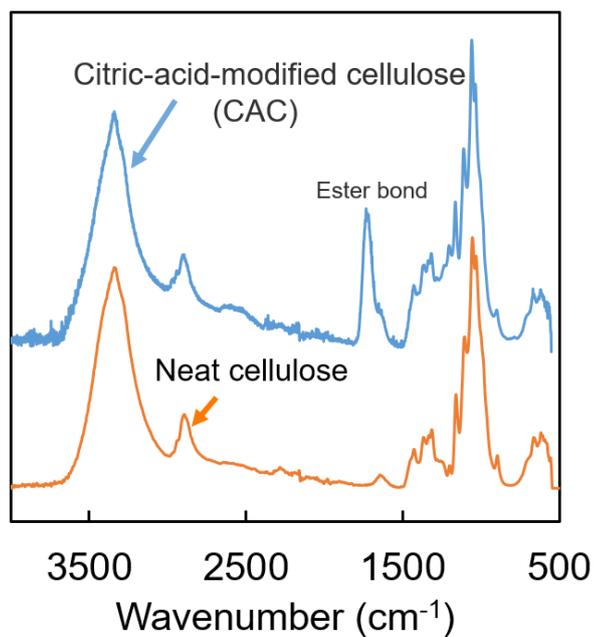
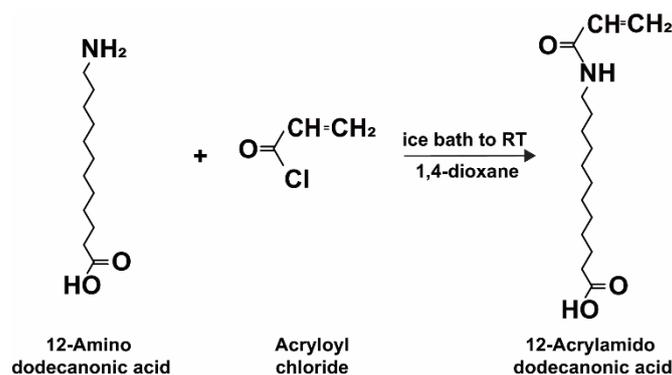


Figure S2. ATR-FTIR spectra of citric acid-modified cellulose (CAC) compared with neat cellulose

3. Preparation of 12-acrylamido dodecanoic acid (Dod)



Scheme S2. Preparation of 12-acrylamido dodecanoic acid (Dod).

12-Aminododecanoic acid (5 g, 323 mmol) and 2M NaOH (2.32 g NaOH in 29 mL water) were dissolved in 1,4-dioxane (0.10 L) and put in ice bath. After the solution was cooled, acryloyl chloride (3.15 g, 35 mmol) was added dropwise. The solution was stirred overnight in room temperature. Next day, HCl was added to solution until pH 2-3 and then the solution was extracted with ethyl acetate. After extraction, the organic phase was filtrated to remove the salt and then extracted again with ethyl acetate. Then, the solution was evaporated and dried at 40 °C for 24 hours. Yield: 90%.

¹H NMR (500 MHz, DMSO-*d*₆) of Dod: δ = 11.92 (s, 1H, -NH), 7.99 (t, J = 5.6 Hz, 1H, -COOH), 6.16 (dd, J = 17.1, 10.1 Hz, 1H, CH₂CH-), 6.02 (dd, J = 17.1, 2.3 Hz, 1H, CH₂CH-), 5.51 (dd, J = 10.1, 2.3 Hz, 1H, CH₂CH-), 3.06 (td, J = 7.1, 5.7 Hz, 2H, -NHCH₂CH₂-), 2.14 (t, J = 7.4 Hz, 2H, -CH₂CH₂COOH-), 1.44 (t, J = 7.3 Hz, 2H, -CH₂(CH₂)₁₁CH₂COOH), 1.37 (q, J = 6.8 Hz, 2H, -CH₂(CH₂)₁₁CH₂COOH), 1.21 (t, J = 2.7 Hz, 14H, -CH₂(CH₂)₁₁CH₂COOH).

¹³C NMR (125 MHz, DMSO-*d*₆) of Dod: δ = 175.05, 164.98, 132.44, 125.27, 39.05, 34.20, 29.55, 29.49, 29.46, 29.42, 29.26, 29.07, 26.97, 25.02.

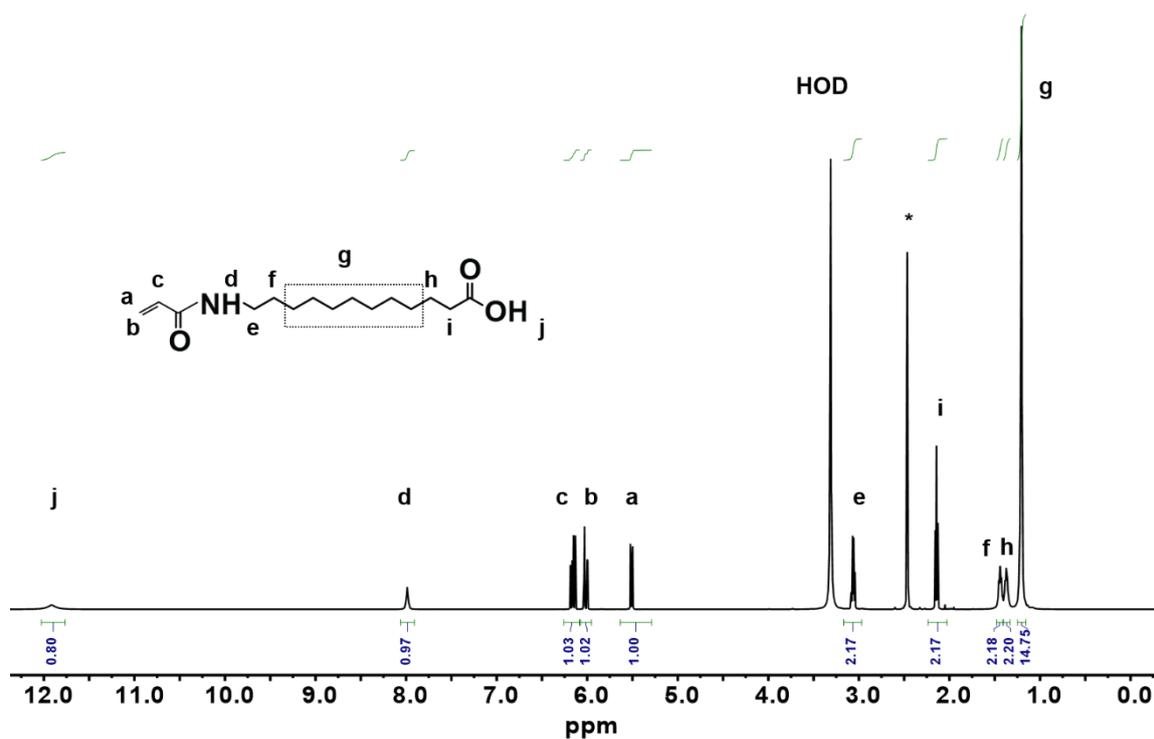


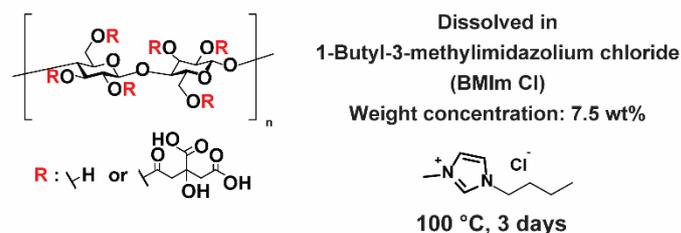
Figure S3. 500 MHz ^1H NMR spectrum of Dod in $\text{DMSO-}d_6$. (TMS for standard, 400 MHz, 25 °C, rotation frequency = 7 kHz).



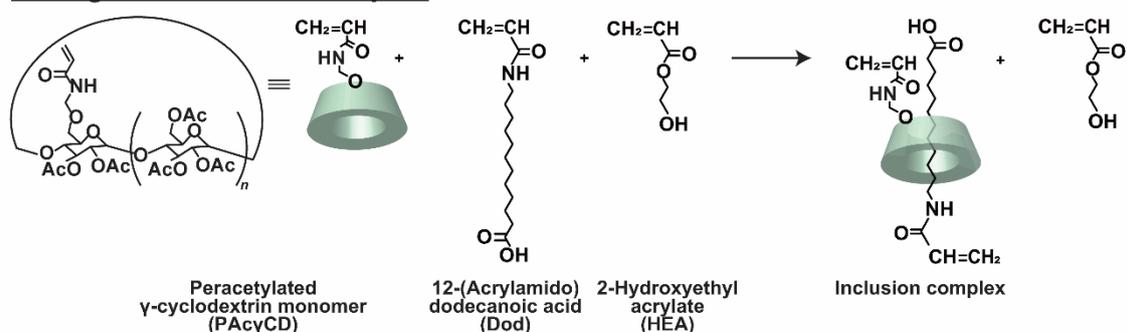
Figure S4. 125 MHz ^{13}C NMR spectrum of Dod in $\text{DMSO-}d_6$. (TMS for standard, 400 MHz, 25 °C, rotation frequency = 7 kHz).

4. Preparation of materials

(a) Citric-acid-modified cellulose (CAC) solution



(b) Host-guest inclusion complex



(c) CAC-based composite formed by host-guest interactions and hydrogen bonding

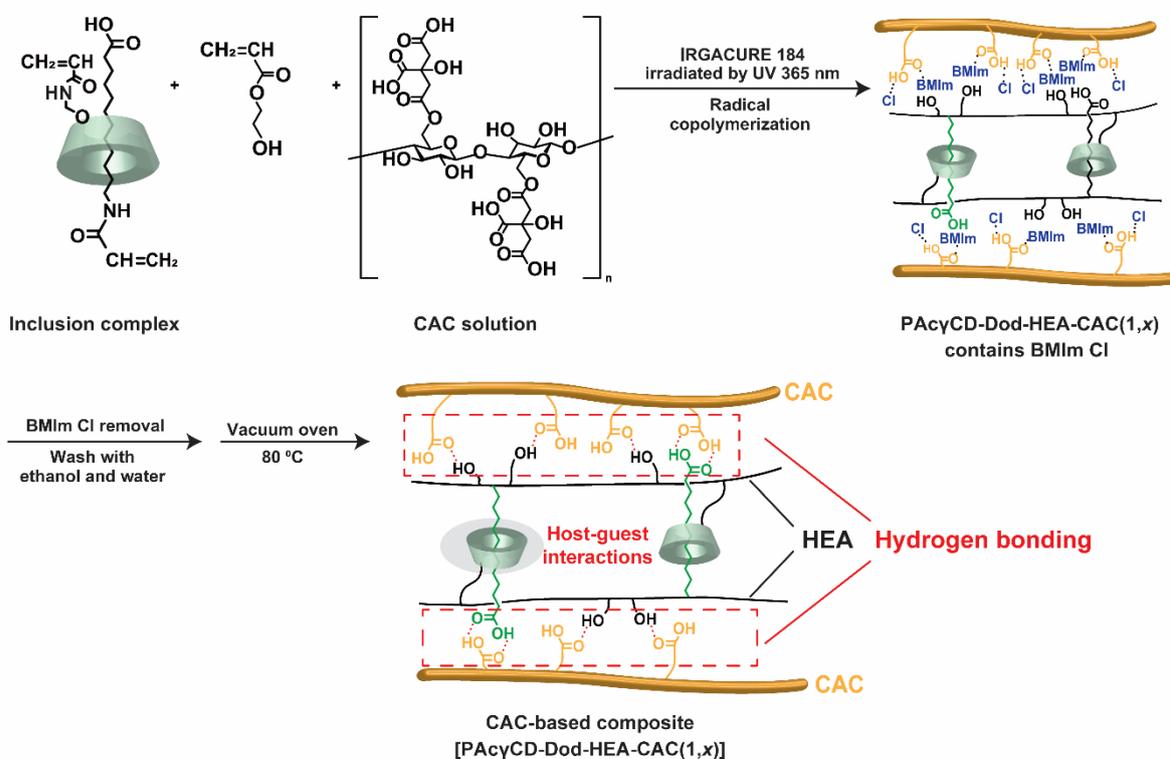


Figure S5. Preparation of CAC-based composite formed by hydrogen bonding and host-guest interactions [PAcyCD-Dod-HEA-CAC(1,x)]

Preparation of CAC-based composite [PAC γ CD-Dod-HEA-CAC(1,x)]

CAC solution was prepared by dissolving CAC (1, 3, or 5 wt%) in BMIm Cl at 7.5wt% concentration at 100 °C for 3 days. PAC γ CD (1 mol%) and Dod (1 mol%) were sonicated in HEA (98 mol%) for 90 minutes. CAC solution was added into host-guest inclusion complexes and radical copolymerized using IRGACURE 184 (0.2 mol%) successfully giving CAC reinforced supramolecular materials [PAC γ CD-Dod-HEA-CAC(1,x)] that still containing BMIm Cl. Then PAC γ CD-Dod-HEA-CAC(1,x) were washed with ethanol and water to remove remaining BMIm Cl and dried in vacuum oven 80 °C for 24 h. The notation “1” at PAC γ CD-Dod-HEA-CAC(1,x) indicates mol% of host guest inclusion complex between PAC γ CD and Dod units and “x” indicates the wt% of CAC.

Table S1. Preparation of PAC γ CD-Dod-HEA-CAC (1,x).

(1,x)	PAC γ CD / g	Dod / mg	HEA / g	CAC / g	IRGACURE 184 / mg
(1,1)	0.41	47	2.0	0.025	7.2
(1,3)	0.41	47	2.0	0.074	7.2
(1,5)	0.41	47	2.0	0.12	7.2

Preparation of polymer without CAC [PAC γ CD-Dod-HEA(1)]

PAC γ CD (1 mol%) and Dod (1 mol%) were sonicated in HEA (98 mol%) for 90 minutes to perform inclusion complex. Then the inclusion complex solution was radical copolymerized using IRGACURE 184 (0.2 mol%) to supramolecular elastomer [PAC γ CD-Dod-HEA(1)]. Then PAC γ CD-Dod-HEA(1) were dried in vacuum oven 80 °C for 24 h. The notation “1” at PAC γ CD-Dod-HEA(1) indicates mol% of host guest inclusion complex between PAC γ CD and Dod units.

Table S2. Preparation of PAC γ CD-Dod-HEA(1).

(1)	PAC γ CD / g	Dod / mg	HEA / g	IRGACURE 184 / mg
(1)	0.41	47	2.0	7.2

Preparation of host polymer with CAC [PAC γ CD-HEA-CAC(1,5)]

CAC solution was prepared by dissolving CAC (5 wt%) in BMIm Cl at 7.5wt% concentration at 100 °C for 3 days. PAC γ CD (1 mol%) was sonicated in HEA (99 mol%) for 10 minutes. CAC solution was added into PAC γ CD-HEA solution and radical copolymerized using IRGACURE 184 (0.2 mol%) successfully giving CAC reinforced materials [PAC γ CD-HEA-CAC(1,5)] that still containing BMIm Cl. Then PAC γ CD-HEA-CAC(1,5) were washed with ethanol and water to remove remaining BMIm Cl and dried in vacuum oven 80 °C for 24 h. The notation “1” at PAC γ CD-HEA-CAC(1,5) indicates mol% of PAC γ CD units and “5” indicates the wt% of CAC.

Table S3. Preparation of PAC γ CD-HEA-CAC(1,5).

x	PAC γ CD / g	HEA / g	CAC / g	IRGACURE 184 / mg
5	0.41	2.0	0.12	7.1

Preparation of guest polymer with CAC [Dod-HEA-CAC(1,5)]

CAC solution was prepared by dissolving CAC (5 wt%) in BMIm Cl at 7.5wt% concentration at 100 °C for 3 days. Dod (1 mol%) was sonicated in HEA (99 mol%) for 10 minutes. CAC solution was added into Dod-HEA solution and radical copolymerized using IRGACURE 184 (0.2 mol%) successfully giving CAC reinforced materials [Dod-HEA-CAC(1,5)] that still containing BMIm Cl. Then Dod-HEA-CAC(1,5) were washed with ethanol and water to remove remaining BMIm Cl and dried in vacuum oven 80 °C for 24 h. The notation “1” at Dod-HEA-CAC(1,5) indicates mol% of Dod units and “5” indicates the wt% of CAC.

Table S4. Preparation of PA γ CD-HEA-CAC(1,5).

x	Dod / mg	HEA / g	CAC / g	IRGACURE 184 / mg
5	47	2.00	0.11	7.1

Preparation of linear polymer with CAC [HEA-CAC(5)]

CAC solution was prepared by dissolving CAC (1 and 5 wt%) in BMIm Cl at 7.5wt% concentration at 100 °C for 3 days. CAC solution was added into HEA and radical copolymerized using IRGACURE 184 (0.2 mol%) successfully giving CAC reinforced materials [HEA-CAC(5)] that still containing BMIm Cl. Then HEA-CAC(5) were washed with ethanol and water to remove remaining BMIm Cl and dried in vacuum oven 80 °C for 24 h. The notation “5” indicates the wt% of CAC.

Table S5. Preparation of HEA-CAC(1,5).

x	HEA / g	CAC / g	IRGACURE 184 / g
5	2.0	0.10	7.0

Preparation of CAC-based composite with guest unit without carboxyl group [PA γ CD-DAA-HEA-CAC(1,5)]

CAC solution was prepared by dissolving CAC (5 wt%) in BMIm Cl at 7.5wt% concentration at 100 °C for 3 days. PA γ CD (1 mol%) and DAA (1 mol%) were sonicated in HEA (98 mol%) for 90 minutes. CAC solution was added into host-guest inclusion complexes and radical copolymerized using IRGACURE 184 (0.2 mol%) successfully giving CAC reinforced supramolecular materials [PA γ CD-Dod-HEA-CAC(1, x)] that still containing BMIm Cl. Then PA γ CD-Dod-HEA-CAC(1, x) were washed with ethanol and water to remove remaining BMIm Cl and dried in vacuum oven 80 °C for 24 h. The notation “1” at PA γ CD-Dod-HEA-CAC(1, x) indicates mol% of host guest inclusion complex between PA γ CD and Dod units and “ x ” indicates the wt% of CAC.

Table S6. Preparation of PA γ CD-DAA-HEA-CAC (1,5).

x	PA γ CD / g	DAA / mg	HEA / g	CAC / g	IRGACURE 184 / mg
5	0.41	42	2.0	0.12	7.2

Preparation of CAC-based composite with main chain without hydroxyl group [PAC γ CD-Dod-EA-CAC(1,5)]

CAC solution was prepared by dissolving CAC (5 wt%) in BMIm Cl at 7.5wt% concentration at 100 °C for 3 days. PAC γ CD (1 mol%) and DAA (1 mol%) were sonicated in HEA (98 mol%) for 90 minutes. CAC solution was added into host-guest inclusion complexes and radical copolymerized using IRGACURE 184 (0.2 mol%) successfully giving CAC reinforced supramolecular materials [PAC γ CD-Dod-HEA-CAC(1, x)] that still containing BMIm Cl. Then PAC γ CD-Dod-HEA-CAC(1, x) were washed with ethanol and water to remove remaining BMIm Cl and dried in vacuum oven 80 °C for 24 h. The notation “1” at PAC γ CD-Dod-HEA-CAC(1, x) indicates mol% of host guest inclusion complex between PAC γ CD and Dod units and “ x ” indicates the wt% of CAC.

Table S7. Preparation of PAC γ CD-Dod-HEA-CAC (1,5).

x	PAC γ CD / g	Dod / mg	EA / g	CAC / g	IRGACURE 184 / mg
5	0.48	55	2.0	0.13	8.3

Preparation of CAC-based composite with no carboxyl and hydroxyl group [PAC γ CD-DAA-EA-CAC(1,5)]

CAC solution was prepared by dissolving CAC (5 wt%) in BMIm Cl at 7.5wt% concentration at 100 °C for 3 days. PAC γ CD (1 mol%) and DAA (1 mol%) were sonicated in HEA (98 mol%) for 90 minutes. CAC solution was added into host-guest inclusion complexes and radical copolymerized using IRGACURE 184 (0.2 mol%) successfully giving CAC reinforced supramolecular materials [PAC γ CD-Dod-HEA-CAC(1, x)] that still containing BMIm Cl. Then PAC γ CD-Dod-HEA-CAC(1, x) were washed with ethanol and water to remove remaining BMIm Cl and dried in vacuum oven 80 °C for 24 h. The notation “1” at PAC γ CD-Dod-HEA-CAC(1, x) indicates mol% of host guest inclusion complex between PAC γ CD and Dod units and “ x ” indicates the wt% of CAC.

Table S8. Preparation of PAC γ CD-DAA-EA-CAC (1,5).

x	PAC γ CD / g	DAA / mg	EA / g	CAC / g	IRGACURE 184 / mg
5	0.48	49	2.0	0.13	8.3

5. NMR characterization of materials

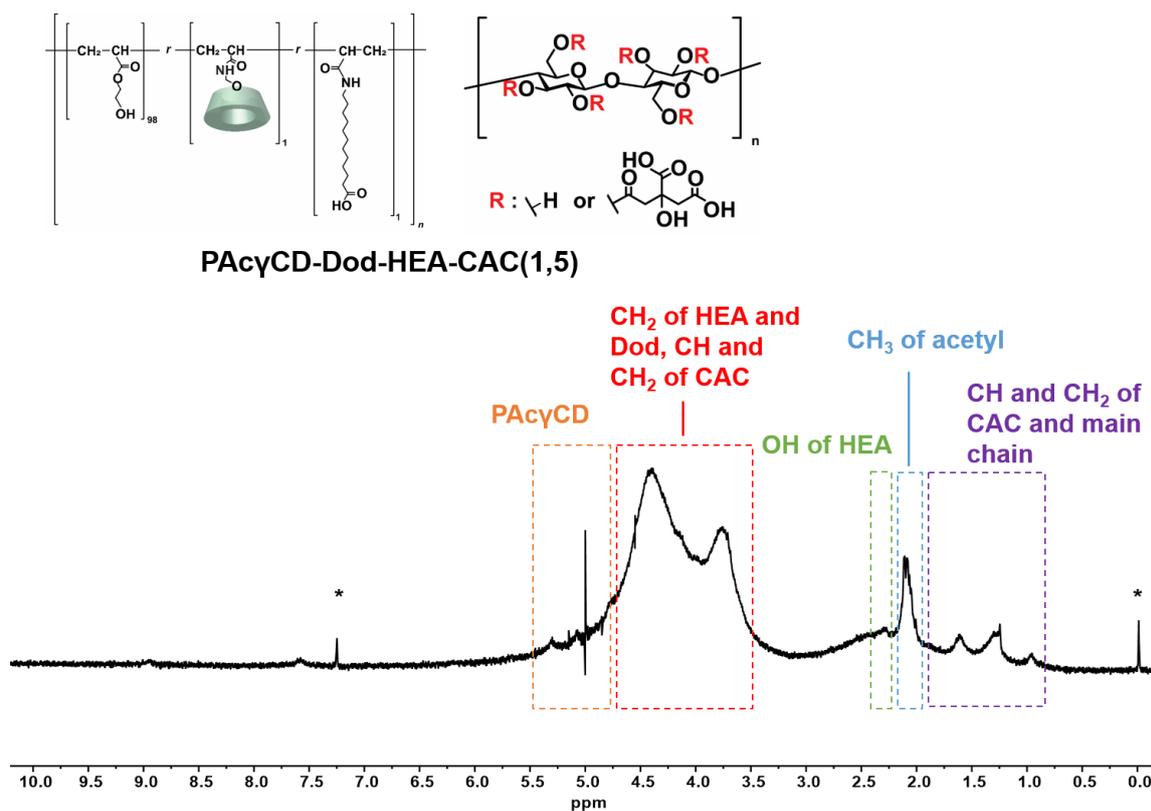


Figure S6. Solid-state ¹H FGMAS NMR spectrum of **PAcyCD-Dod-HEA-CAC(1,5)** (TMS for standard, 400 MHz, 25 °C, rotation frequency = 7 kHz).

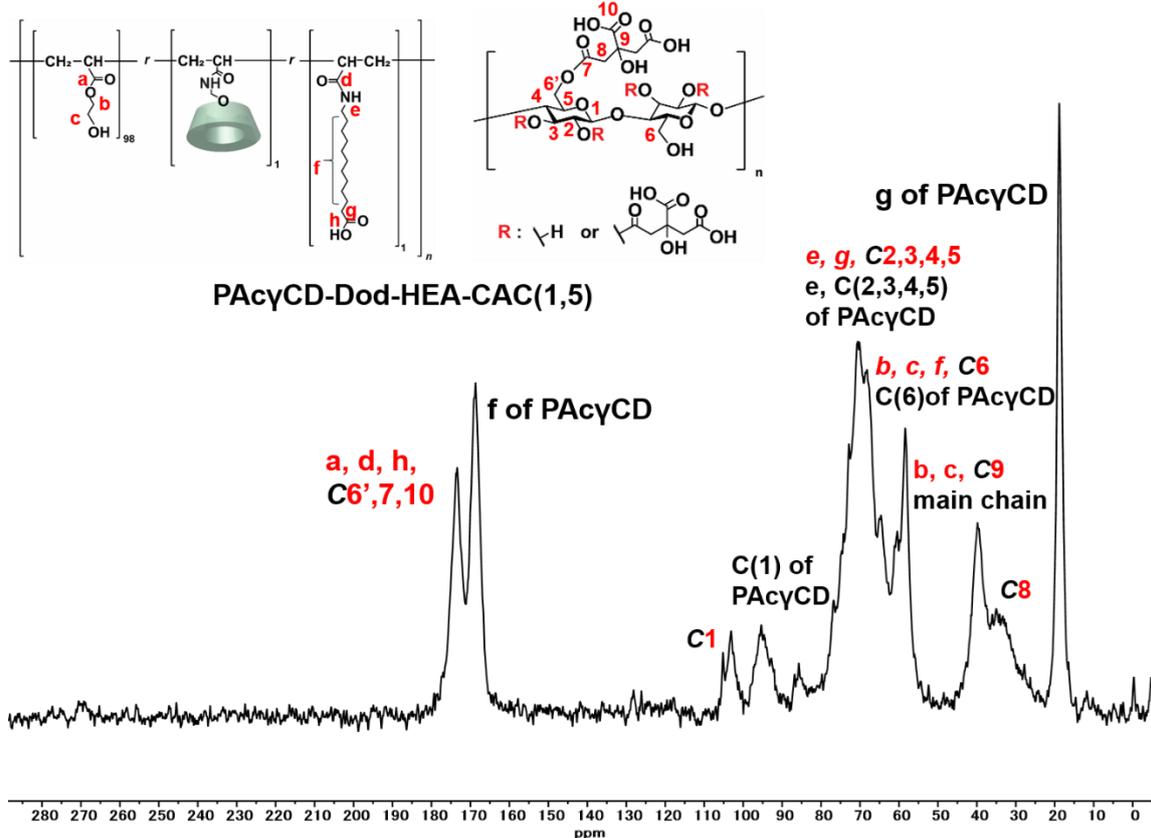


Figure S7. Solid-state ¹³C CPMAS NMR spectrum of **PAcyCD-Dod-HEA-CAC(1,5)** (TMS for standard, 75 MHz, 25 °C, rotation frequency = 7 kHz).

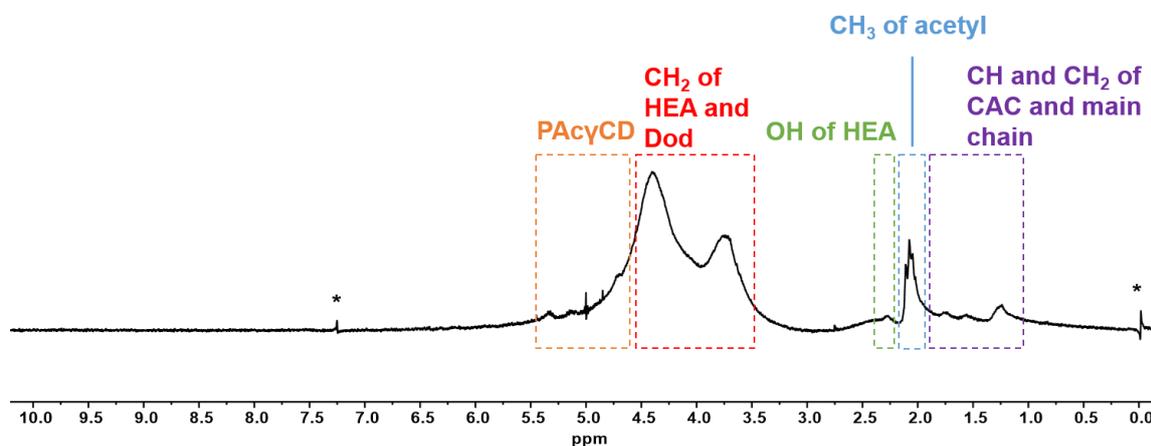
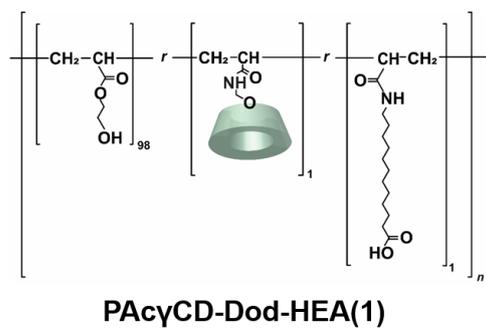


Figure S8. Solid-state ^1H FGMAS NMR spectrum of **PAcyCD-Dod-HEA(1)** (TMS for standard, 400 MHz, 25 °C, rotation frequency = 7 kHz).

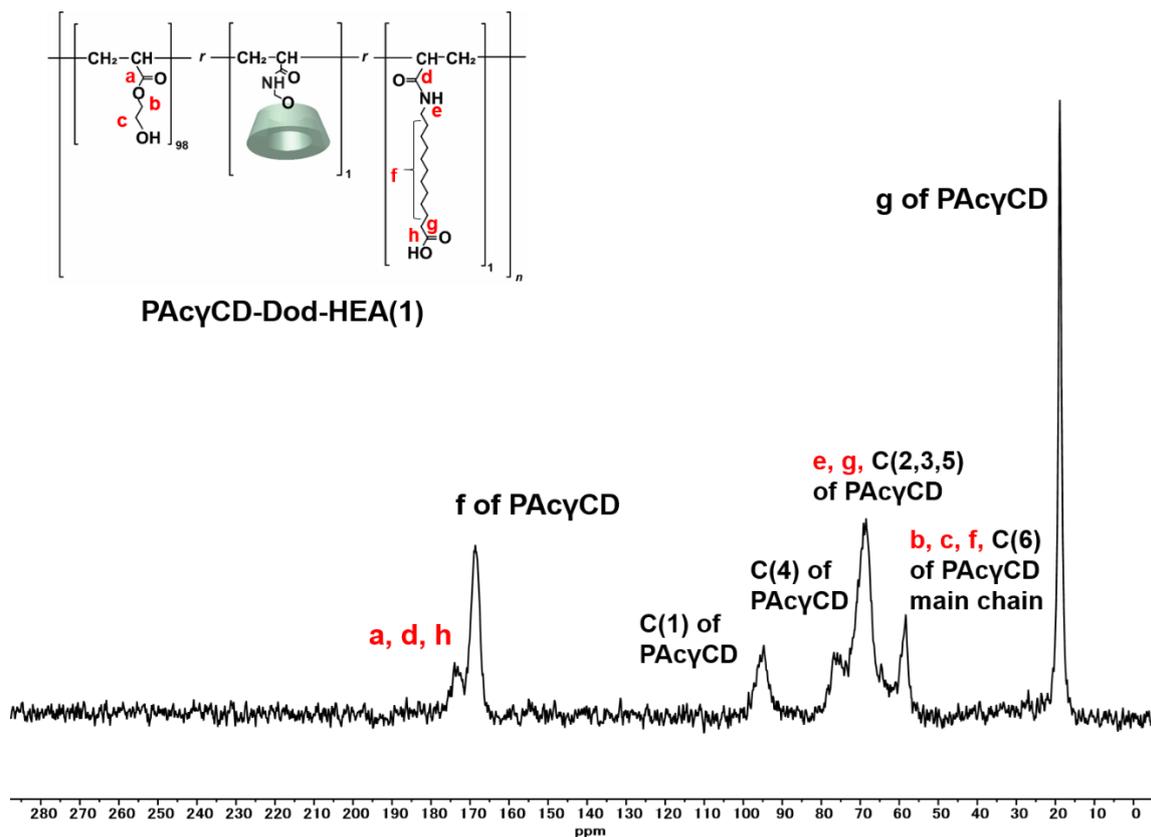


Figure S9. Solid-state ^{13}C FGMAS NMR spectrum of **PAcyCD-Dod-HEA(1)** (TMS for standard, 75 MHz, 25 °C, rotation frequency = 7 kHz).

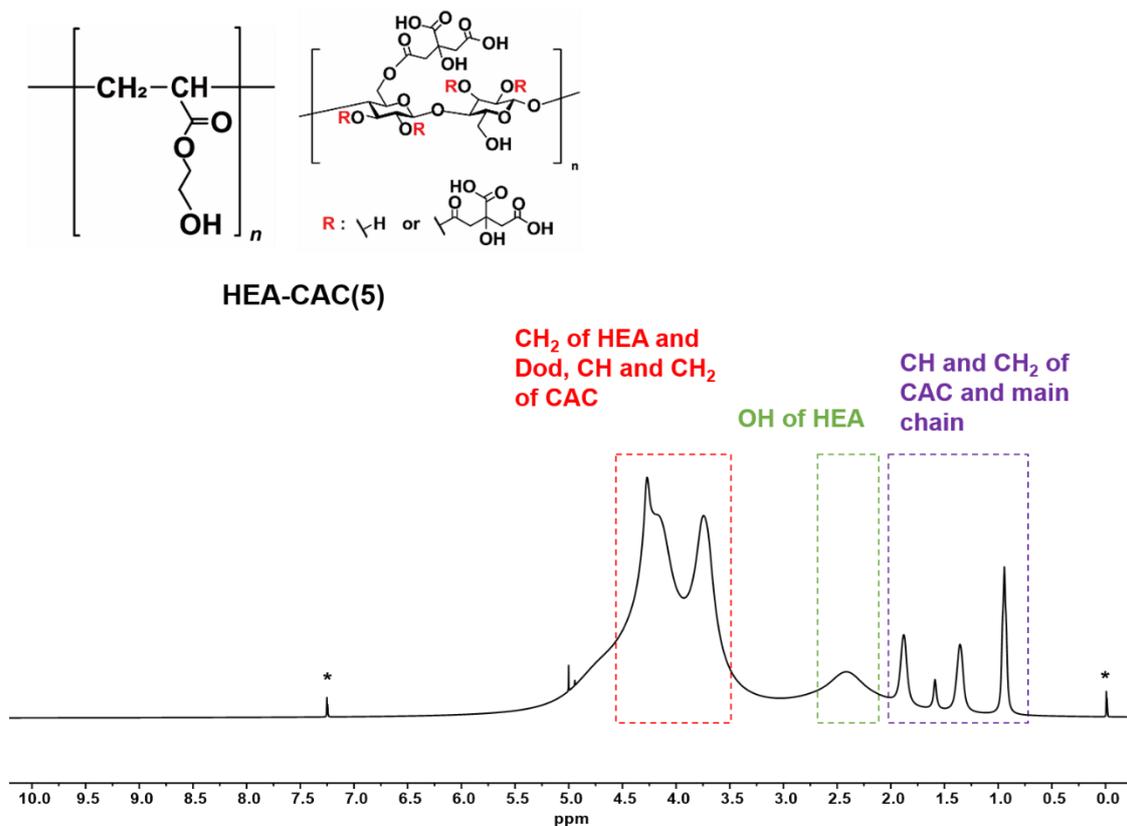


Figure S10. Solid-state ¹H FGMAS NMR spectrum of **PAcyCD-HEA-CAC(1,5)** (TMS for standard, 400 MHz, 25 °C, rotation frequency = 7 kHz).

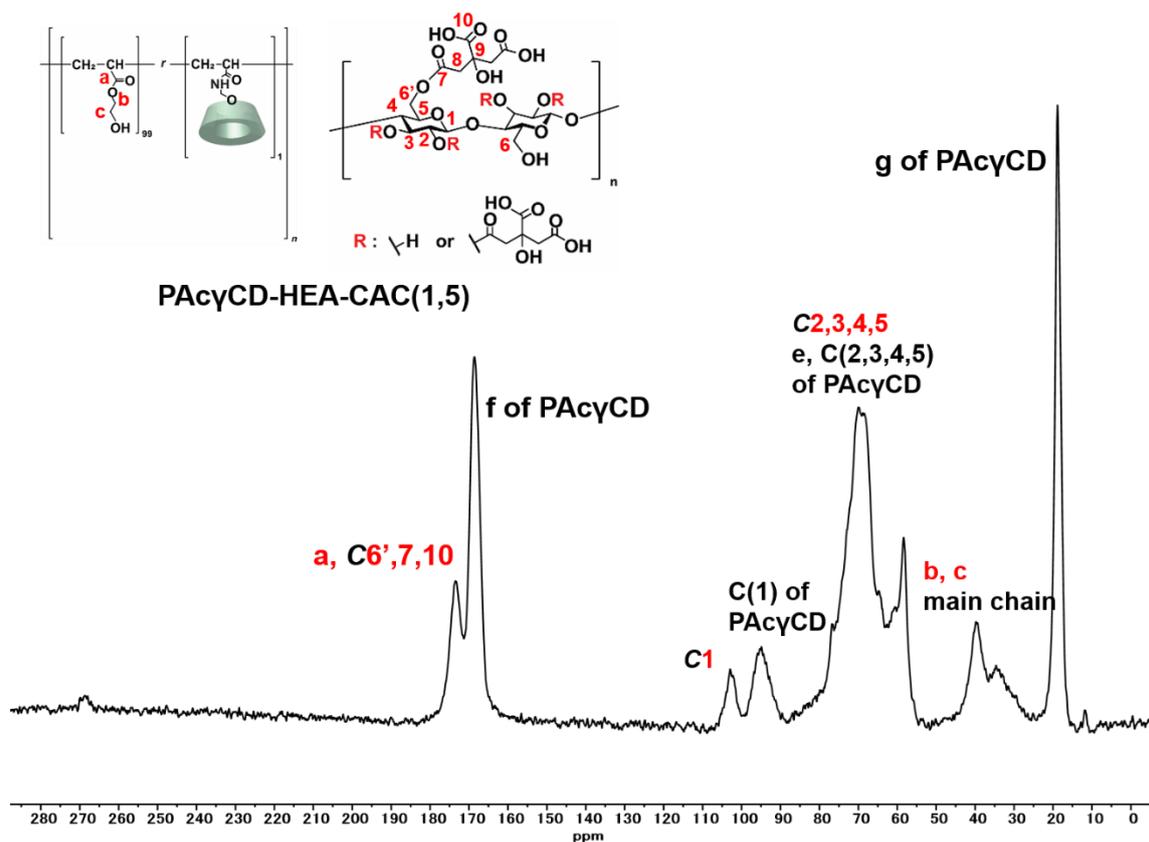


Figure S11. Solid-state ¹³C CPMAS NMR spectrum of **PAcyCD-HEA-CAC(1,5)** (TMS for standard, 75 MHz, 25 °C, rotation frequency = 7 kHz).

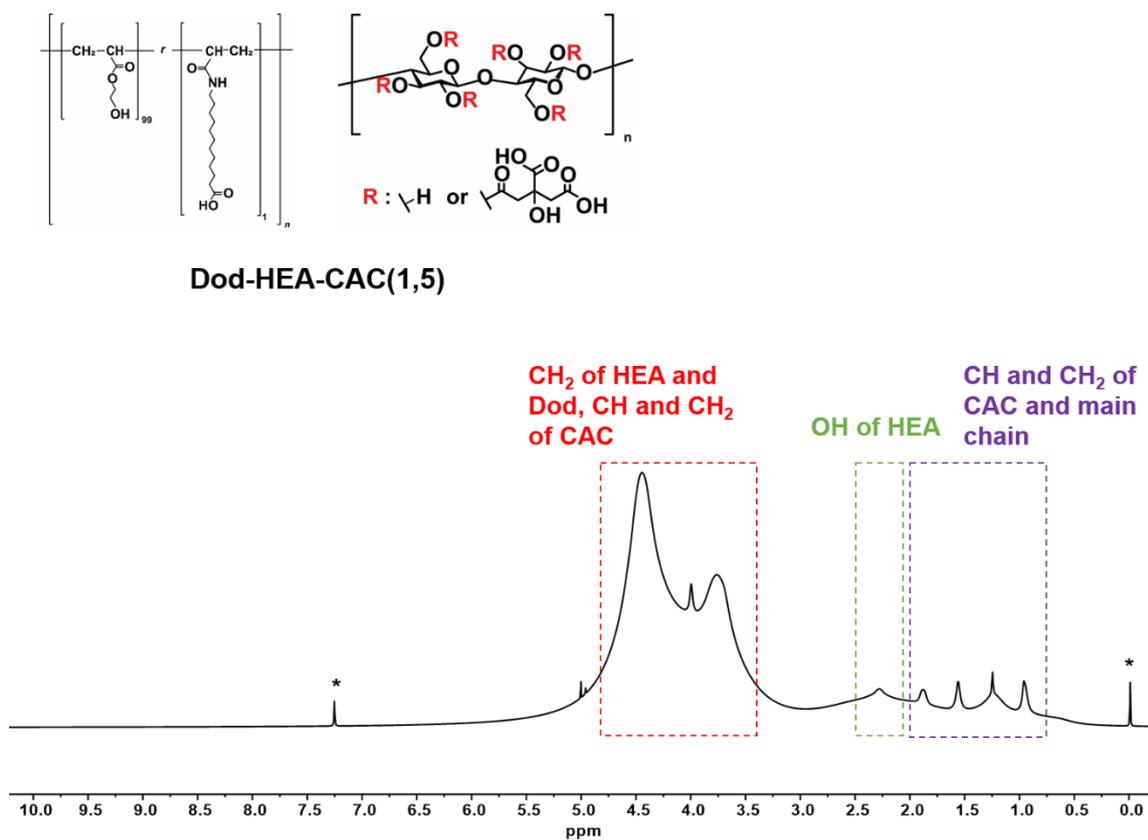


Figure S12. Solid-state ^1H FGMAS NMR spectrum of **Dod-HEA-CAC(1,5)** (TMS for standard, 400 MHz, 25 °C, rotation frequency = 7 kHz).

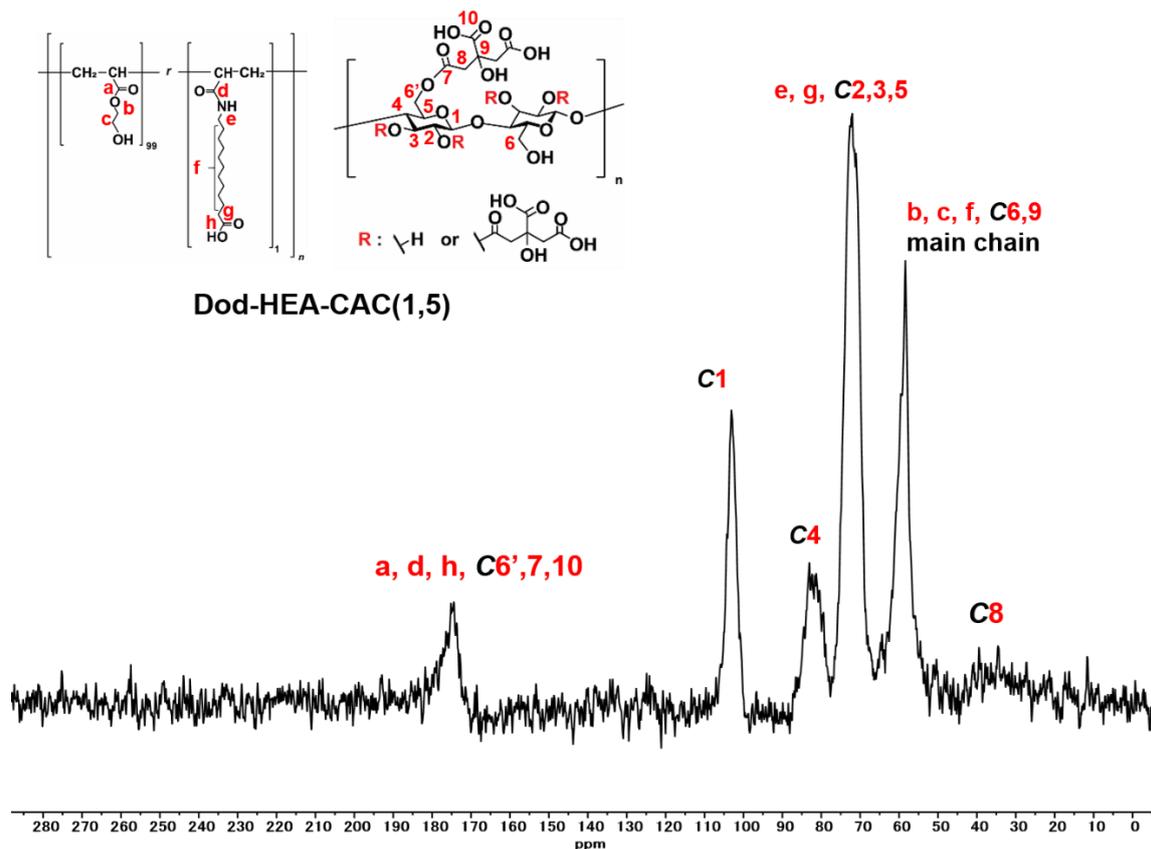


Figure S13. Solid-state ^{13}C CPMAS NMR spectrum of **Dod-HEA-CAC(1,5)** (TMS for standard, 75 MHz, 25 °C, rotation frequency = 7 kHz).

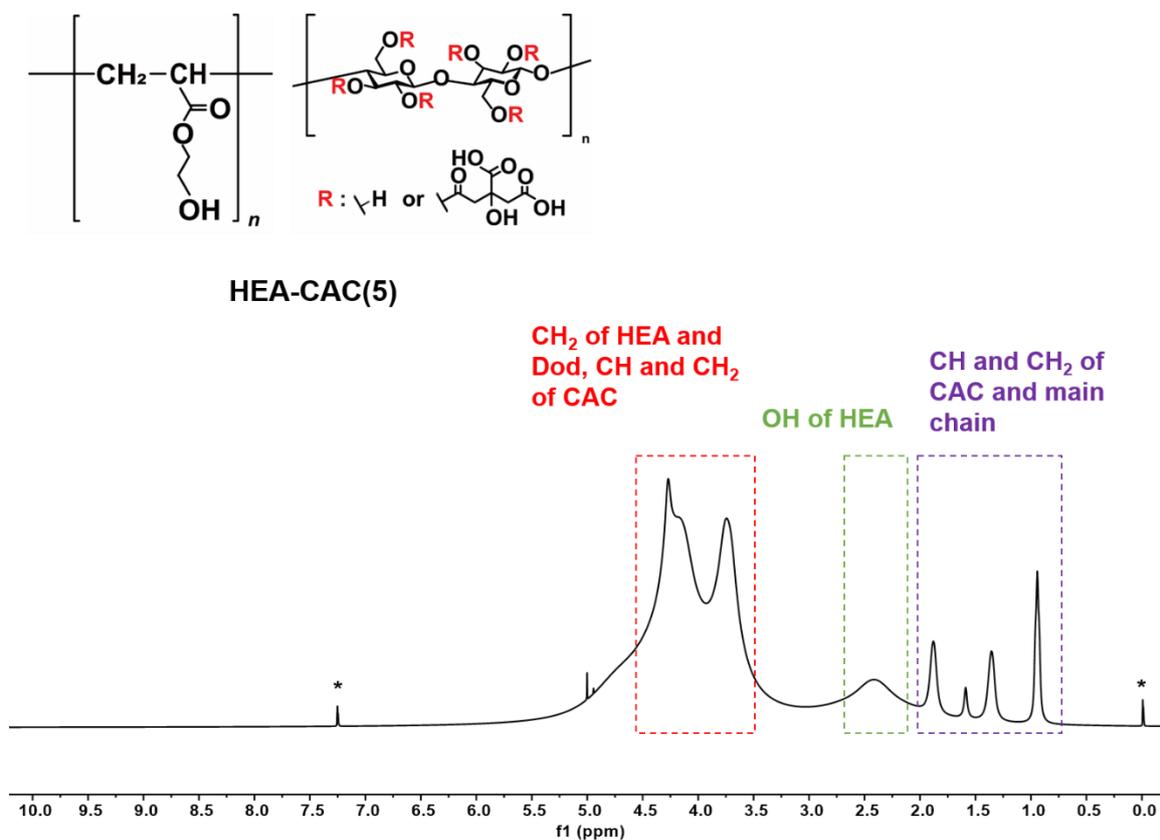


Figure S14. Solid-state ¹H FGMAS NMR spectrum of **HEA-CAC(15)** (TMS for standard, 400 MHz, 25 °C, rotation frequency = 7 kHz).

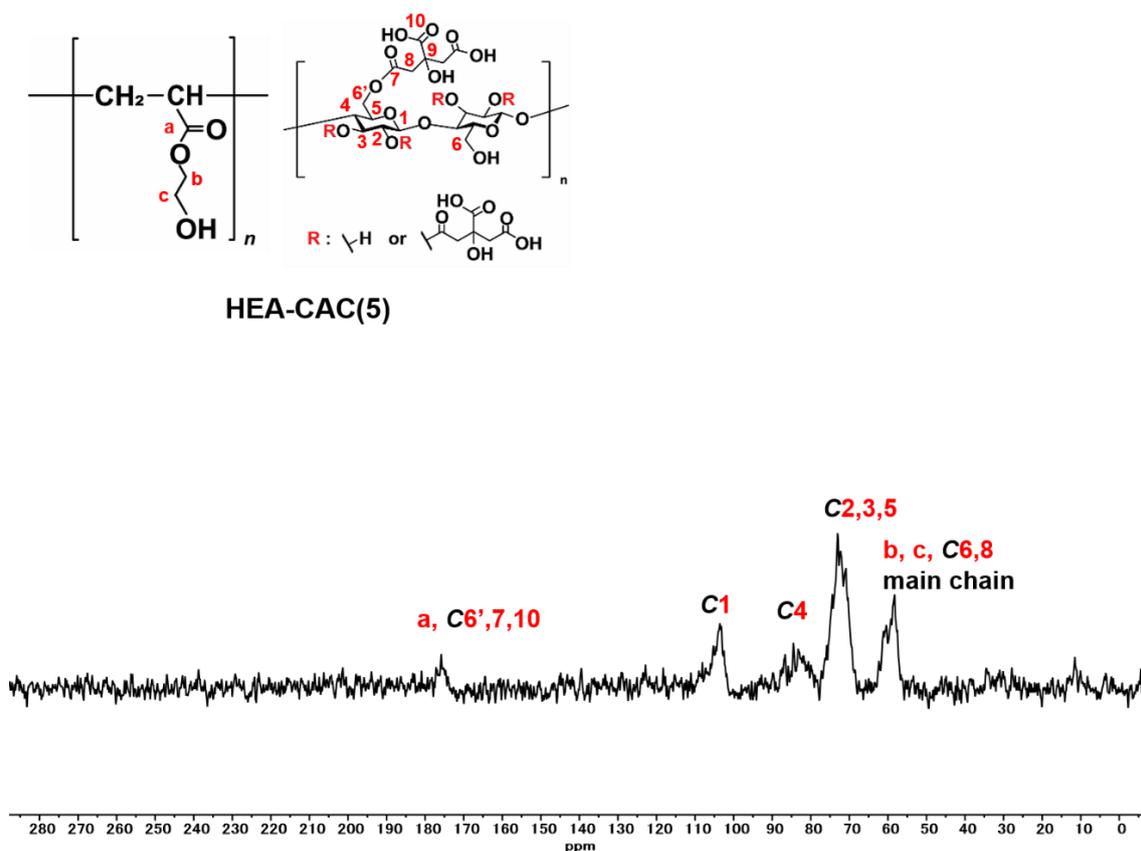


Figure S15. Solid-state ¹³C CPMAS NMR spectrum of **HEA-CAC(15)** (TMS for standard, 75 MHz, 25 °C, rotation frequency = 7 kHz).

6. FTIR of materials

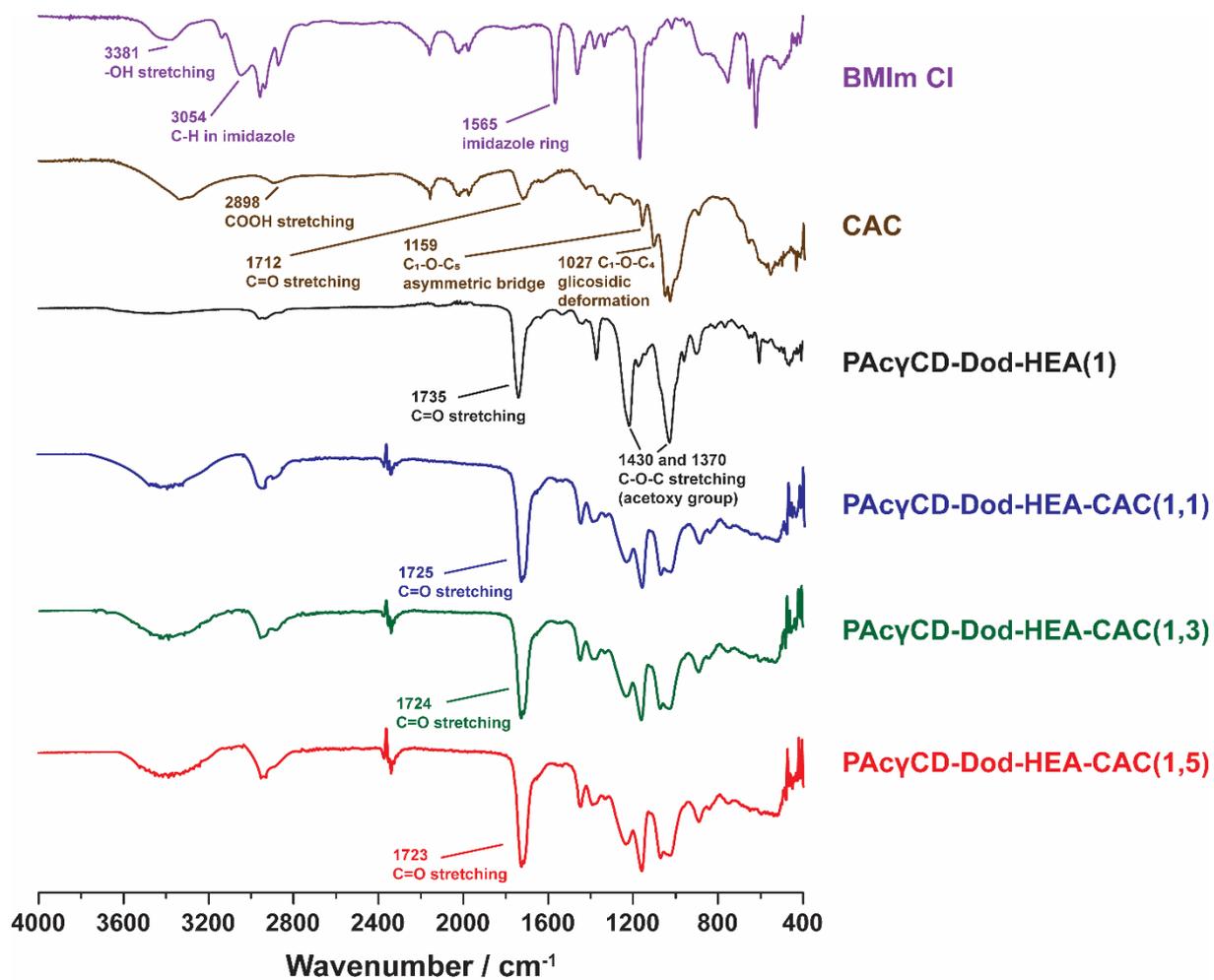


Figure S16. Attenuated Total Reflectance Fourier-transform infrared spectroscopy (ATR-FTIR) of 1-butyl-3-methylimidazolium chloride (BMIm Cl), citric acid-modified cellulose (CAC), supramolecular elastomer [PAcyCD-Dod-HEA(1)], composite with 1 wt% CAC [PAcyCD-Dod-HEA-CAC(1,1)], composite with 3 wt% CAC [PAcyCD-Dod-HEA-CAC(1,3)], and composite with 5 wt% CAC [PAcyCD-Dod-HEA-CAC(1,5)].

7. Cyclic tensile test measurement

Cyclic tensile tests were measured using Autograph AG-X plus (Shimadzu Co.). Test pieces were continuously stretched and recovered without interval. Maximum strains were set to 100%, 200%, 300%, 400%, 500%, 600%, 700%, and 800% at deformation rate of $1 \text{ mm}\cdot\text{s}^{-1}$ in tests for both PAcyCD-Dod-HEA-CAC(1,5) and PAcyCD-Dod-HEA(1).

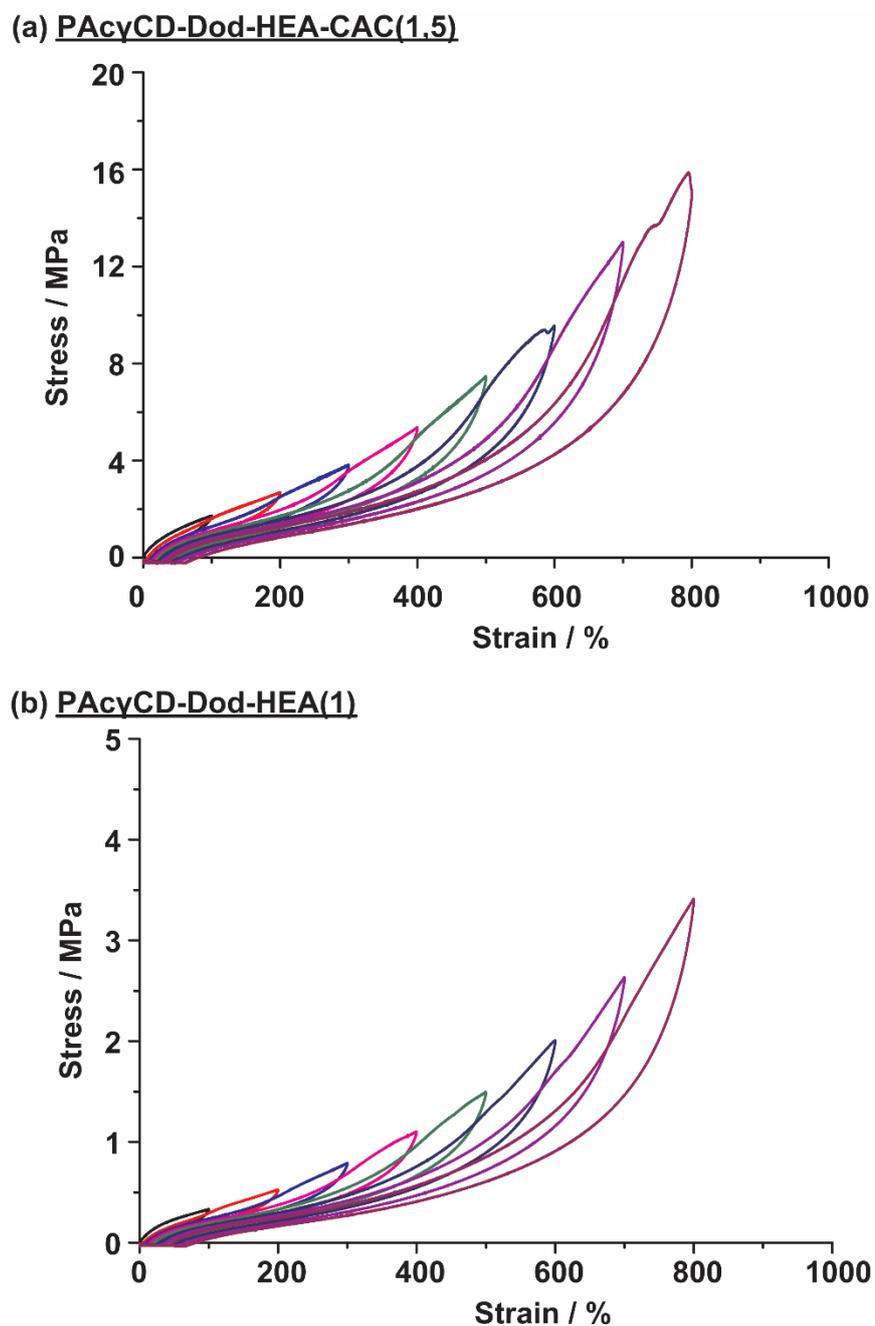


Figure S17. Cyclic tensile test of (a) PAcyCD-Dod-HEA-CAC(1,5) and (b) PAcyCD-Dod-HEA(1).

8. Creep and stretch recovery measurement

Creep recovery measurement was performed using Autograph AG-X plus (Shimadzu Co.). Stress applying to the test pieces was held at 500 kPa for both PA γ CD-Dod-HEA(1) and PA γ CD-Dod-HEA-CAC(1,5) at 1 hour. Then, the stress was released and strain recovery of the test pieces was recorded as a length of test pieces.

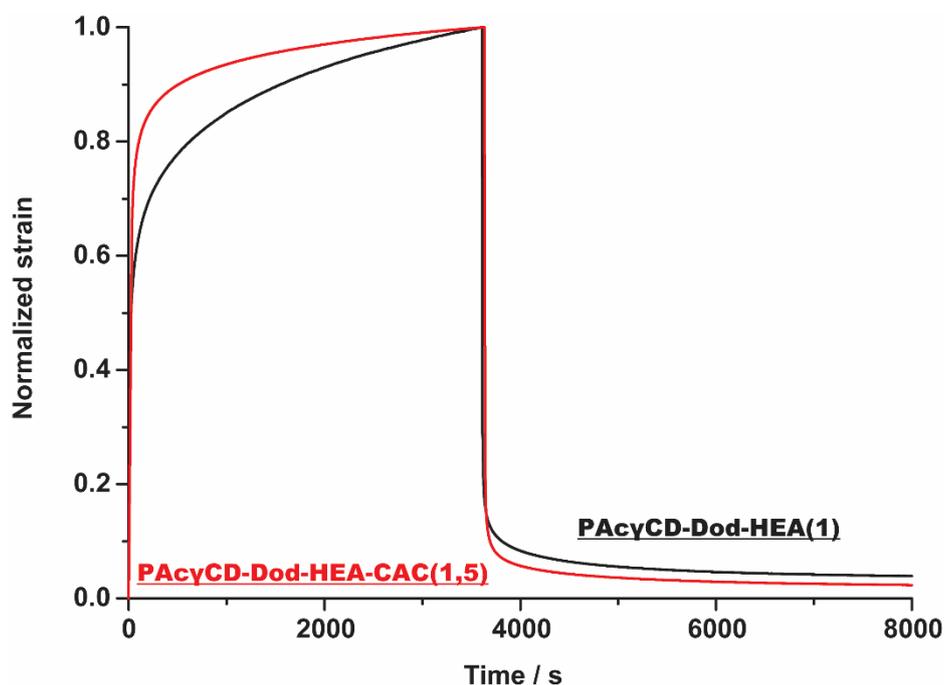


Figure S18. Creep recovery test for PA γ CD-Dod-HEA(1) and PA γ CD-Dod-HEA-CAC(1,5). Stress applying to the test pieces was held at 500 kPa for 1 hour then released.

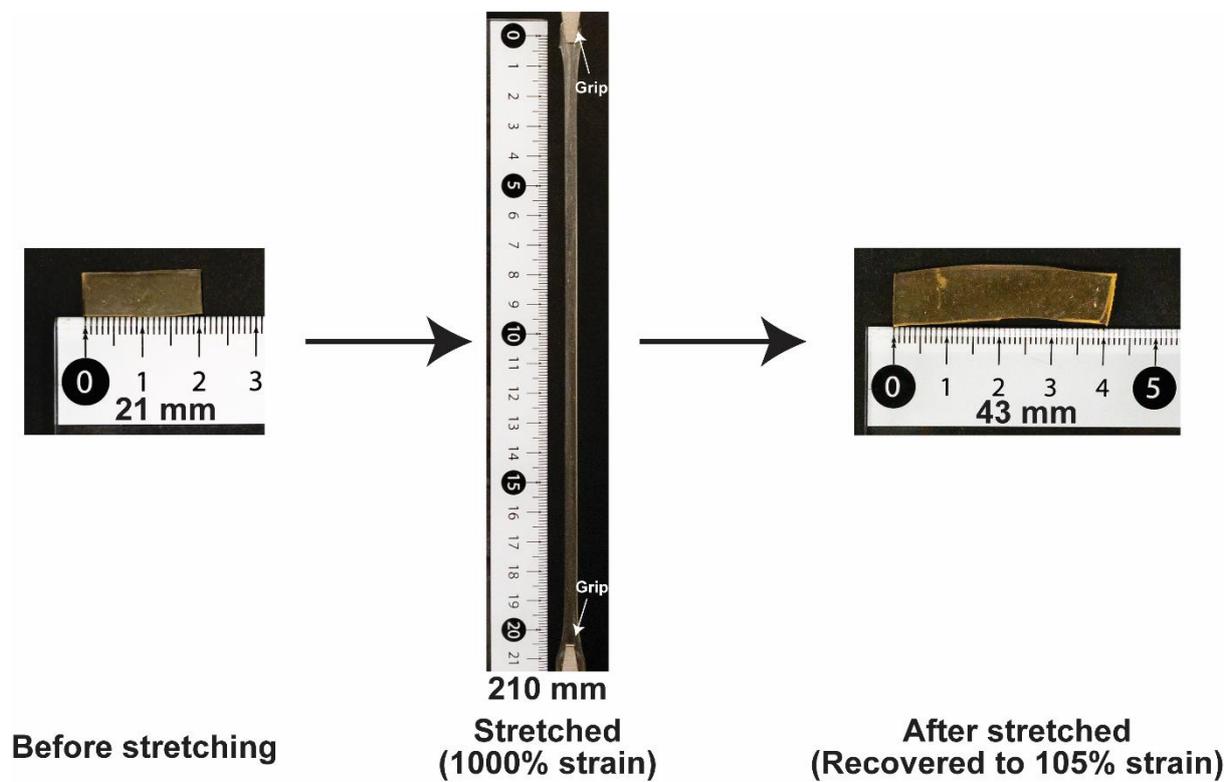


Figure S19. Stretch and recovery test for PAc γ CD-Dod-HEA-CAC(1,5).

9. Time dependency self-healing ratio calculation

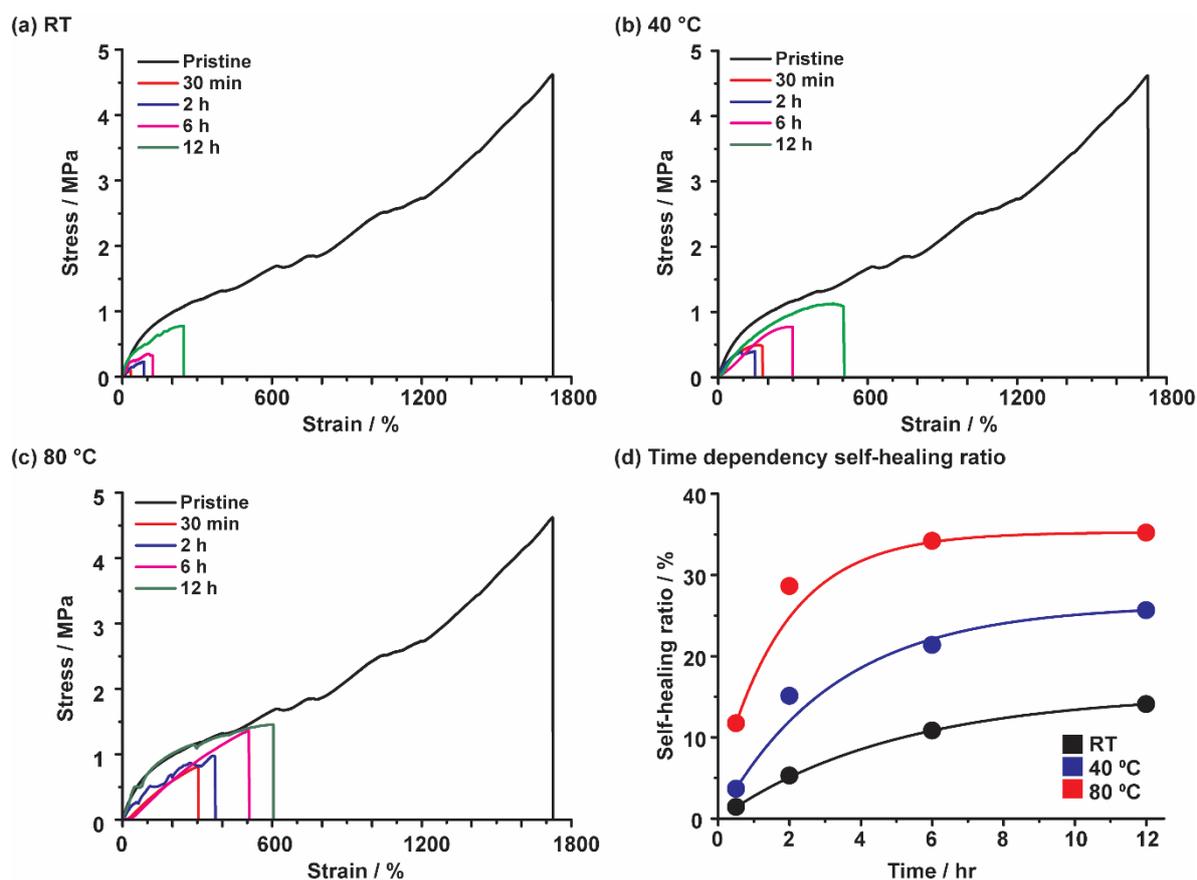


Figure S20. Stress strain curve of PAC γ CD-Dod-HEA(1) for self-healing ratio calculation at (a) RT, (b) 40 °C, and (c) 80 °C. (d) Time dependency self-healing ratio of PAC γ CD-Dod-HEA(1).

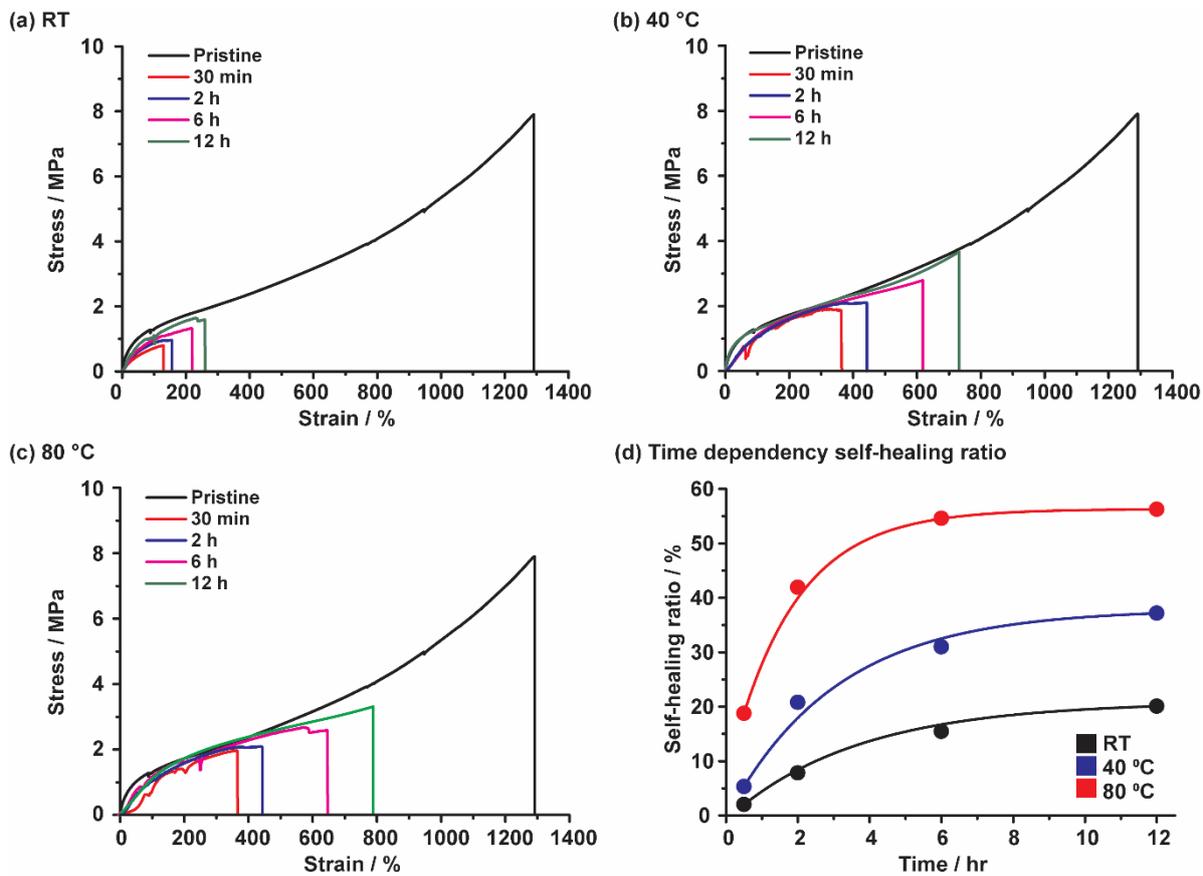


Figure S21. Stress strain curve of PA γ CD-Dod-HEA-CAC(1,1) for self-healing ratio calculation at (a) RT, (b) 40 °C, and (c) 80 °C. (d) Time dependency self-healing ratio of PA γ CD-Dod-HEA-CAC(1,1).

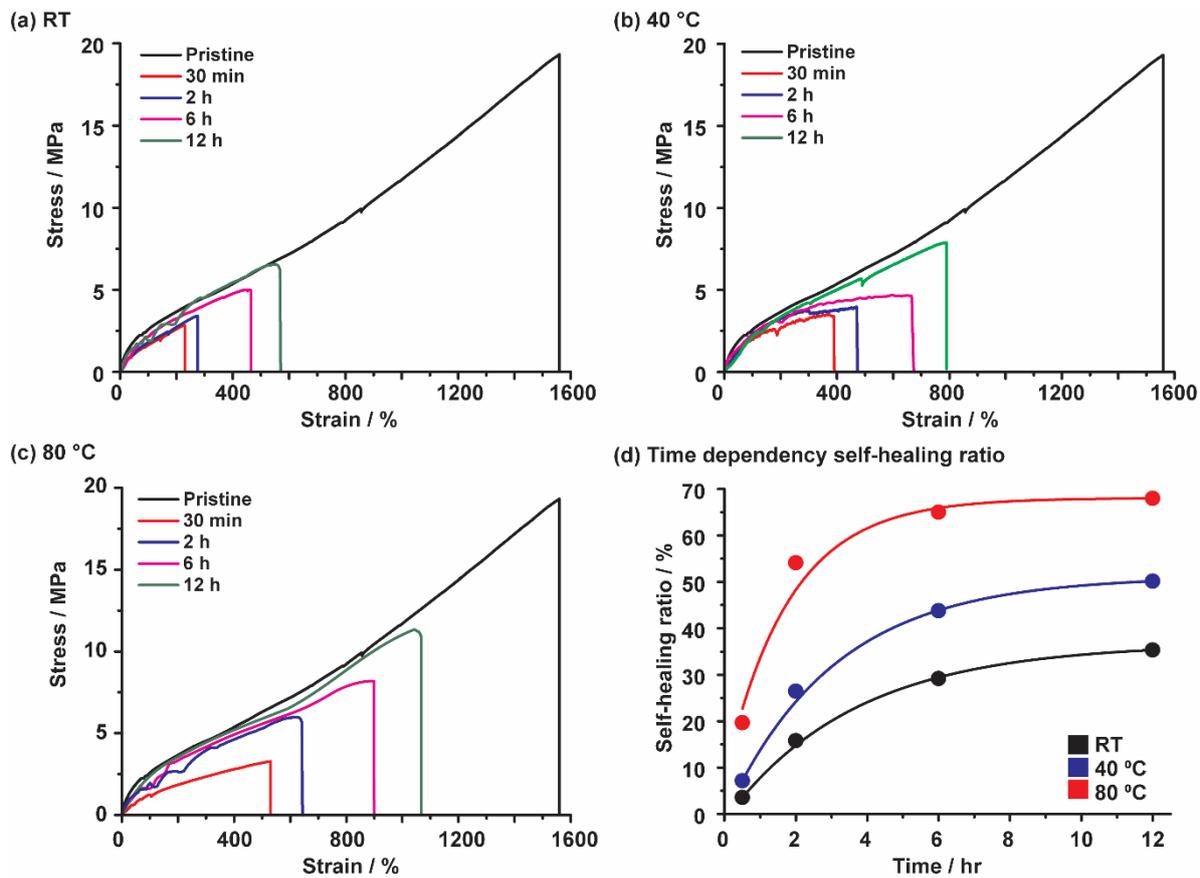


Figure S22. Stress strain curve of PACyCD-Dod-HEA-CAC(1,3) for self-healing ratio calculation at (a) RT, (b) 40 °C, and (c) 80 °C. (d) Time dependency self-healing ratio of PACyCD-Dod-HEA-CAC(1,3).

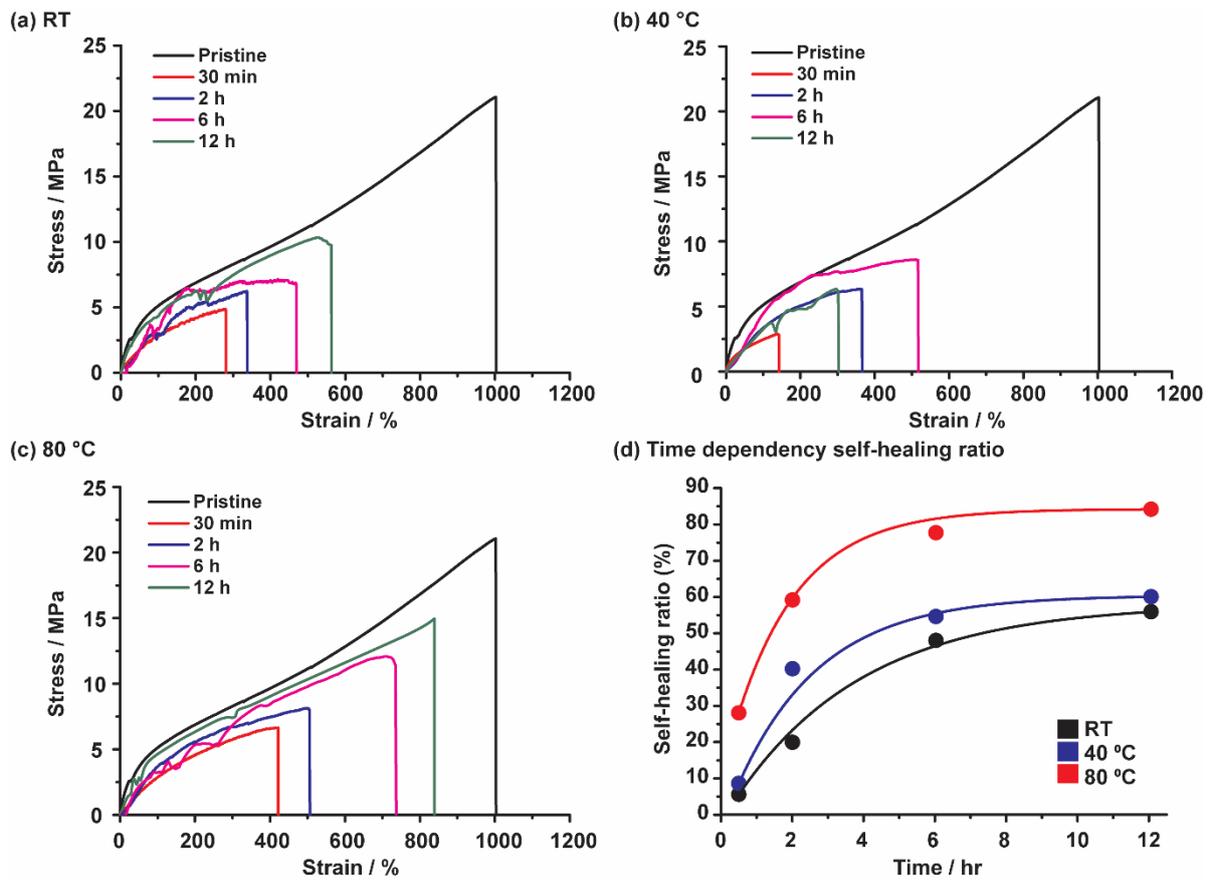


Figure S23. Stress strain curve of PACyCD-Dod-HEA-CAC(1,1) for self-healing ratio calculation at (a) RT, (b) 40 °C, and (c) 80 °C. (d) Time dependency self-healing ratio of PACyCD-Dod-HEA-CAC(1,5).

10. DSC curve of materials

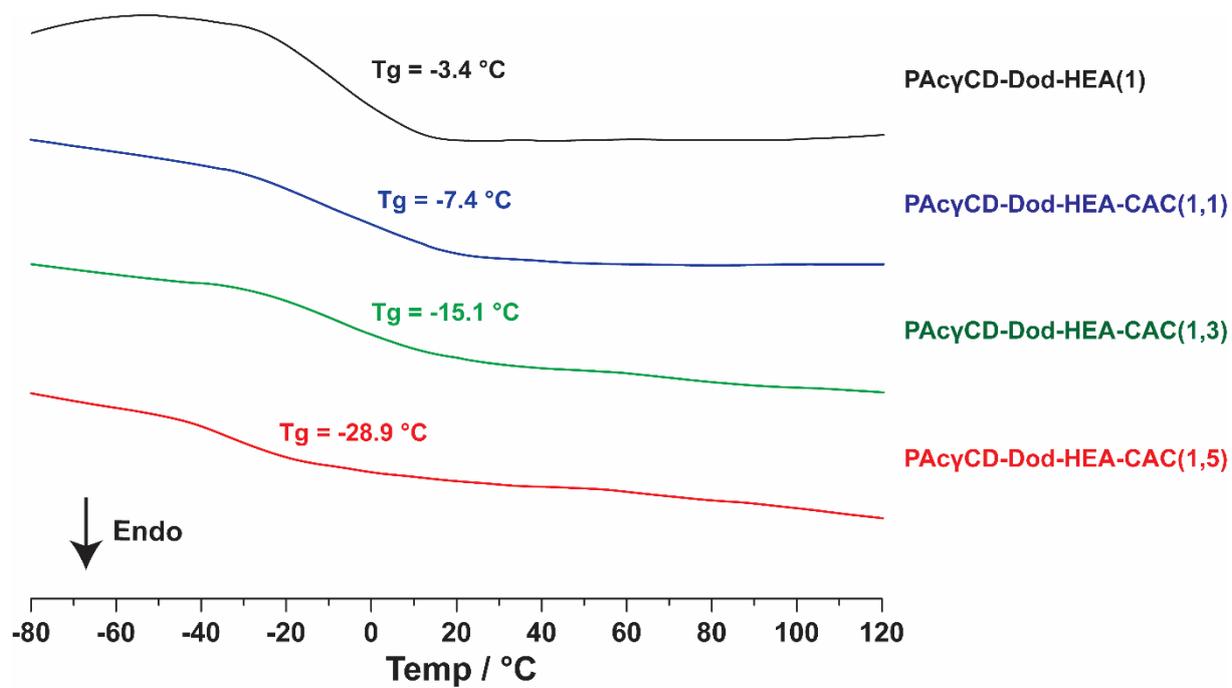


Figure S24. DSC curve for supramolecular elastomer [PAcyCD-Dod-HEA(1)], composite with 1 wt% CAC [PAcyCD-Dod-HEA-CAC(1,1)], composite with 3 wt% CAC [PAcyCD-Dod-HEA-CAC(1,3)], and composite with 5 wt% CAC [PAcyCD-Dod-HEA-CAC(1,5)].

11. Laser scanning confocal microscope

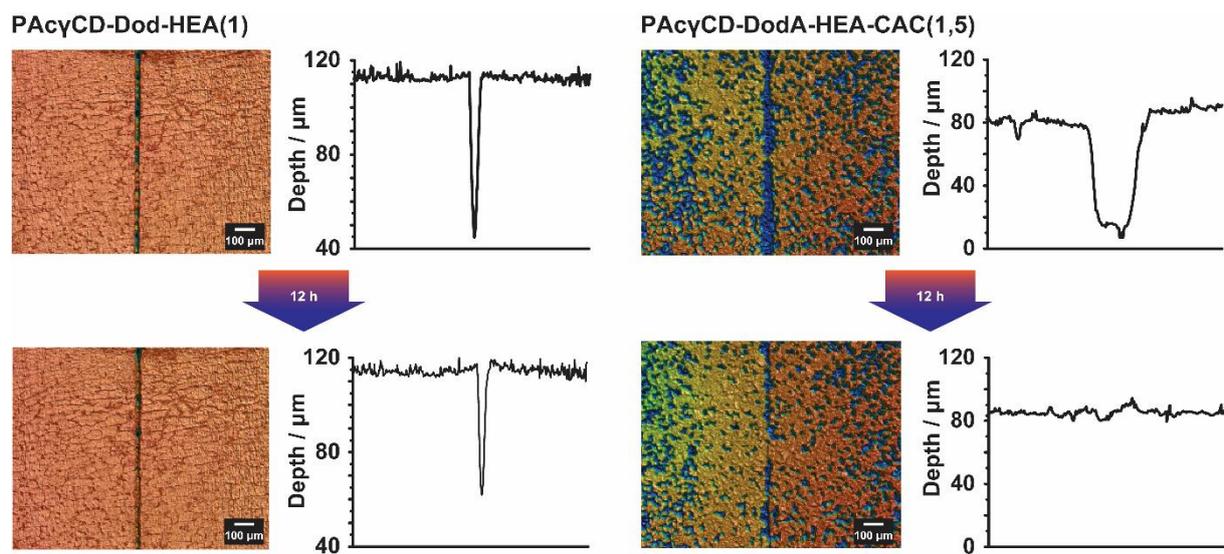


Figure S25. Laser scanning confocal microscope and depth profile of PAcyCD-Dod-HEA(1) and PAcyCD-Dod-HEA-CAC(1,5) at 80°C after reattached for 12 hours.

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