Supporting Information

Modeling Gene Expression Instability by Programmed and

Switchable Polymerization/Nicking DNA Nano-Machineries

Zhixin Zhou, ‡ Daoqing Fan, ‡ and Itamar Willner*

Institute of Chemistry, The Minerva Center for Biohybrid Complex Systems, The Hebrew

University of Jerusalem, Jerusalem 91904, Israel

[‡]These authors contributed equally.

*E-mail: willnea@vms.huji.ac.il.

Fax: 972-2-6527715.

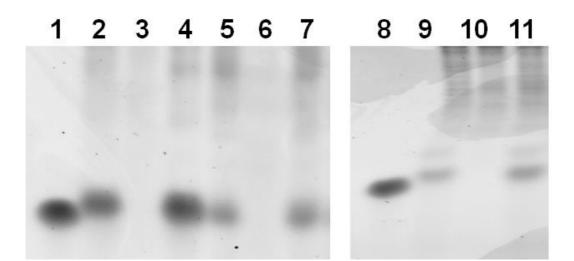


Figure S1. Native polyacrylamide gel electrophoresis analysis of the switchable polymerization/nicking machinery using T-A•T triplex as blocker unit and strand (6) as deblocker: Lane 1-reference lane of "gene" (2); lane 2-the polymerization/nicking machinery, in the absence of T-A duplex; lane 3-upon addition of T-A duplex, the blockage of the polymerization/nicking machinery; lane 4-upon the addition of DNA sequence (6) to the T-A·T triplex blocked machinery. Native polyacrylamide gel electrophoresis analysis of the switchable polymerization/nicking machinery using ATP as blocker unit and ATPase as deblocker: Lane 5-the polymerization/nicking machinery, in the absence of ATP; lane 6-upon addition of ATP, the blockage of the polymerization/nicking machinery; lane 7-upon the addition of ATPase to the ATP blocked machinery. Native polyacrylamide gel electrophoresis analysis of the switchable polymerization/nicking machinery using Sr²⁺-ion-stabilized G-quadruplex as blocker unit and kryptofix [2.2.2] (KP) as deblocker: Lane 8-reference lane of "gene" (2); Lane 9-the polymerization/nicking machinery, in the absence of Sr²⁺-ions; lane 10-upon addition of Sr²⁺-ions, the blockage of the polymerization/nicking machinery; lane 11-upon the addition of KP to the Sr²⁺-ions blocked machinery.