

# Supporting Information

## Molecular mechanisms of chromium(III) immobilization by organo-ferrihydrite coprecipitates: The significant roles of ferrihydrite and carboxyl

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**1 Cr(III) adsorbed on Ferrihydrite (Fh) (Fh-Cr(III)), amorphous Cr(OH)<sub>3</sub> (Cr(OH)<sub>3</sub>(am)) and organo-Cr(III) complexes (OC-Cr(III)) synthesis**

The Fh was synthesized following the method described by Schwertmann and Cornell.<sup>1</sup> Briefly, the pH of 0.1 M FeCl<sub>3</sub> solution was adjusted to ~7 using a 0.1 M NaOH solution. After shaking for 24 h, the suspension was separated by centrifugation and the paste was washed twice to remove residual ions using Mili-Q water, and then freeze-dried for 48 h. The adsorption of Cr(III) to Fh was conducted by mixing 20 mg dried Fh with 20 mL 0.1 mM CrCl<sub>3</sub> solution in a 50 ml plastic tube. To prevent the possible formation of Cr(OH)<sub>3</sub> precipitation on the surface of Fh, a low pH of 4.5 was kept during the adsorption experiment. The sorption result showed that the 80% Cr(III) was removed from solution to Fh. According to the Cr(III) binding sites density of Fh (1.3 sites nm<sup>-2</sup>) and the specific surface area (SSA) of Fh (305.05 m<sup>2</sup> g<sup>-1</sup>) (Table S2, IS), ~11.6% Cr(III) binding sites of Fh were occupied by Cr(III), which suggested no Cr(OH)<sub>3</sub> surface precipitates formation in the product.<sup>2</sup> Therefore, the adsorption product should be theoretically predominant as Fh-Cr(III). However, Cr(III) surface hydroxyl polymers, having a similar structure with Cr(OH)<sub>3</sub>, most probably presented on the Fh surface instead of isolated Cr atoms according to previous study.<sup>2</sup>

The synthesis method of amorphous chromium(III) hydroxide (Cr(OH)<sub>3</sub>(am)) was same as the Fh. The Cr(OH)<sub>3</sub>(am) rather than crystalline Cr(OH)<sub>3</sub> was selected as a standard reference because NaOH solution was used to adjust pH, and thus no crystalline Cr(OH)<sub>3</sub> formed in this study.<sup>3</sup> This was because the synthesis of crystalline Cr(OH)<sub>3</sub> required NH<sub>3</sub>H<sub>2</sub>O or KOH solutions.<sup>4, 5</sup>

The OC-Cr(III) was made by mixing 60 mg L<sup>-1</sup> rice or rape DOC with 0.1 mM CrCl<sub>3</sub> at

pH of 4.5. The paste of OC-Cr(III) was obtained by centrifugation and washed twice with Mili-Q water to remove residual DOC or Cr(III). Then the paste was freeze-dried for XANES measurements. Since there is little difference of Cr K-edge XANES between rice and rape derived OC-Cr(III) (data are not shown). In this study, we chose rice derived OC-Cr(III) as a standard compound for further bulk XANES analysis.

## **2 XRD, SEM-EDS and BET measurements**

All the OFC samples were freeze-dried and ground into powders for further characterization. XRD analysis was conducted at ambient conditions using Cu-K $\alpha$  radiation produced with 40 kV and 30 mA electron beam. SEM-EDS analysis was performed using scanning electron microscope (SEM) (Zeiss Auriga Compact) in conjunction with energy dispersive X-ray spectroscopy (EDS) (X-MAX80). The SSA of the OFC was determined through measurements of N<sub>2</sub> gas adsorption isotherms after degassing.

## **3 C K-edge and Cr L-edge XANES**

Bulk C K-edge XANES spectra were collected at the beamline BL08U at the Shanghai Synchrotron Radiation Facility (SSRF) in China.<sup>6, 7</sup> The two DOC stocking solutions were freeze-dried and the deposited samples were ground into powders. Then the OC powders and oxalic acid reference were attached onto Cu tape fixed by a holder for spectral collection in the total electron yield mode. To improve the signal-to-noise ratio, at least of two scans were conducted for each sample and then all the spectra were processed using Athena software.<sup>8</sup>

## **4 STXM data analysis**

The Stack Analyze, aXis2000 software package were applied for the alignment of image sequence and spectra processing. After alignment, the equation: optical density (OD) =  $\ln(I_0/I)$

was used to convert data to optical density, where  $I$  referred to the flux passed through the sample, and the  $I_0$  (incident flux) was obtained by measuring an empty region without samples. The C, Fe and Cr distribution maps were obtained by subtracting average images at the pre-edge range from the averaged images for each element. The energy ranges were listed in Table S5. The C K-edge, Cr and Fe  $L_{3,2}$ -edge XANES spectra of the selected regions of interest (ROIs) of the sorption and co-precipitation experiments were derived using aXis2000 software, and plotted using origin 8.5 for fingerprint analysis. PCA-Cluster analysis of the sorption and co-precipitation samples were processed using PCA GUI (1.1.1) software.

### 5 Impact of pH increase on Cr(III) precipitation in $\text{CrCl}_3$ solution

The solution of 0.1 mM  $\text{CrCl}_3$  was prepared in two replicates by dissolving  $\text{CrCl}_3$  chemicals in 10 mM NaCl solution, and the pH of 0.1 mM  $\text{CrCl}_3$  was determined. Then NaOH solution was used to adjust the pH of the  $\text{CrCl}_3$  solution to the experimental pH (5.5). After shaking at 200 rpm for 24 hours at 25 °C, the  $\text{CrCl}_3$  solution were centrifuged at 5,000 rpm for 15 mins, and the supernatants of the  $\text{CrCl}_3$  solution were filtrated using 0.45  $\mu\text{m}$  membrane filters to determine the concentrations of soluble Cr using ICP-OES (Agilent 5110). The removal of the Cr in the solution was calculated by the difference between the initial added Cr and the remaining Cr in the supernatants.

### 6 Additional results

Table S1. The contents of interested elements in the rice and rape straw-derived DOC.

	C (mg L <sup>-1</sup> )	N (mg L <sup>-1</sup> )	Al (mg L <sup>-1</sup> )	Ca (mg L <sup>-1</sup> )	Fe (mg L <sup>-1</sup> )	K (mg L <sup>-1</sup> )	Mn (mg L <sup>-1</sup> )	P (mg L <sup>-1</sup> )	S (mg L <sup>-1</sup> )
Rice	2476.5	1364.1	0.9	6.1	3.7	1677.2	44.9	36.2	77.6
Rape	2077.1	1390.5	0.2	214.4	0.5	1834.7	0.4	88.8	282.3

Table S2. Final Fe/C molar ratios and SSA of OFC formed under different initial solution Fe/C molar ratios.

DOC	Initial Fe/C in solution	Fe/C in OFC <sup>a</sup>	SSA (m <sup>2</sup> g <sup>-1</sup> )
-	control	-	305.05
Rice	10	11.31	317.06
	0.5	0.53	0.15
	0.1	0.02	1.08
Rape	10	9.94	294.02
	0.5	0.52	_ <sup>b</sup>
	0.1	0.01	1.01

<sup>a</sup>The Fe/C molar ratio in OFC was calculated through the difference of Fe, C concentrations in solution before and after reactions.

<sup>b</sup> The SSA is under the detection limit with our sample weight.

Table S3. Results of the first four components obtained from PCA analysis on Cr K-edge XANES spectra of the studied OFC samples.

Components	Eigenvalue	Variation	Accumulated variation	IND
Component 1	113.8	0.925	0.925	0.0295
Component 2	7.091	0.057	0.983	0.0058
Component 3	0.901	0.007	0.990	0.0043
Component 4	0.479	0.003	0.994	0.0032

Table S4. Linear combination fitting results of the Cr K-edge XANES spectra of the OFCs from the sorption and co-precipitation experiments under different Fe/C molar ratios. The standard deviation is shown in the bracket

Sample	Goodness of fit		Percentage of Cr(III) species		
	R factor	$\chi^2$	Fh-Cr(III)	OC-Cr(III)	Cr(OH) <sub>3</sub>
Sor_control	0.0003	0.0395	90 (3)		10 (3)
Sor_Ri_0.5	0.0006	0.0815	68 (6)	32 (6)	
Sor_Ra_0.5	0.0005	0.0739	52 (6)	48 (6)	
Sor_Ri_0.1	0.0003	0.0483	27 (5)	73 (5)	

Sor_Ra_0.1	0.0002	0.0335	62 (4)	38 (4)	
Cop_control	0.0014	0.1903	77 (7)		23 (7)
Cop_Ri_0.5	0.0017	0.2254	75 (11)	25 (11)	
Cop_Ra_0.5	0.0050	0.6034	73 (17)	27 (17)	
Cop_Ri_0.1	0.0011	0.1436	70 (8)	30 (8)	
Cop_Ra_0.1	0.0005	0.0779	53 (6)	47 (6)	

Table S5. Pre-edge energy and edge-energy ranges used for the generation of average stack maps using STXM data.

Element	Absorption edge	Pre-edge energy stack range (eV)	Edge energy stack range (eV)
Total C	K	280-282	300-320
Aromatic C		280-282	285.2-285.8
Aliphatic C		280-282	287.1-287.9
Carboxylic C		280-282	288.3-288.9
Fe	L	700-704	709.0-709.8
Cr	L	570-572	577.2-578.1

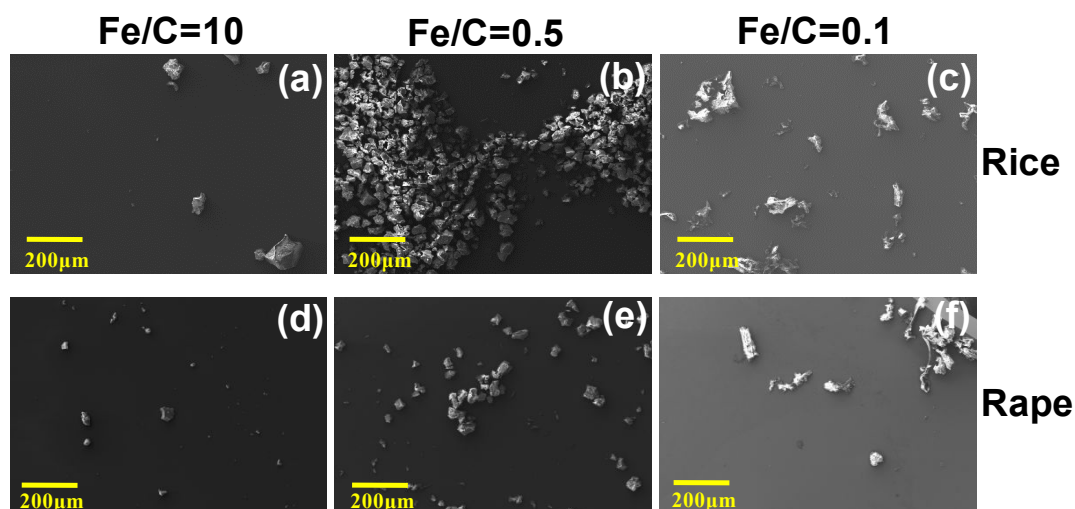


Figure S1. SEM images of the OFC under different Fe/C molar ratios at a relative large scale.

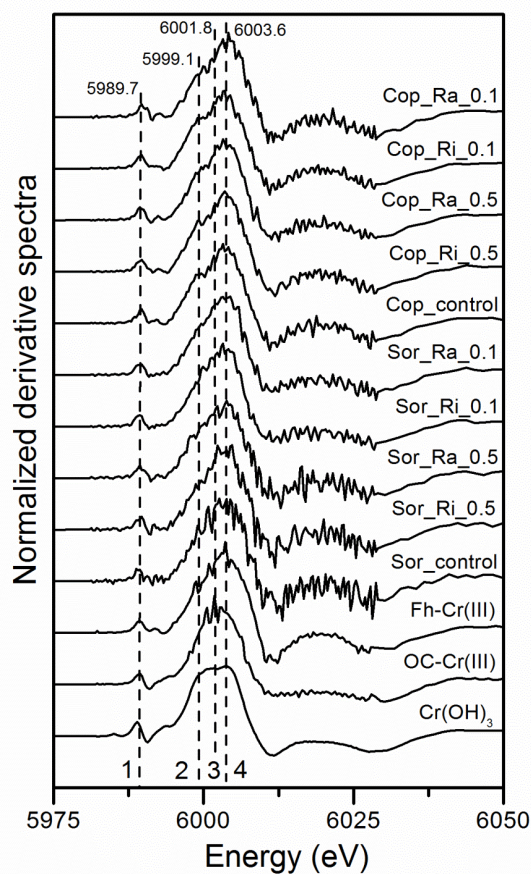


Figure S2. The first derivative spectra of Cr K-edge XANES spectra of OFC and standards (a). Sor\_Ri\_10 or Cop\_Ra\_10 refer to the OFC with Cr produced from adsorption or co-precipitation in the presence of DOCri or DOCra at the Fe/C molar ratio of 10, respectively, the others were named similarly.

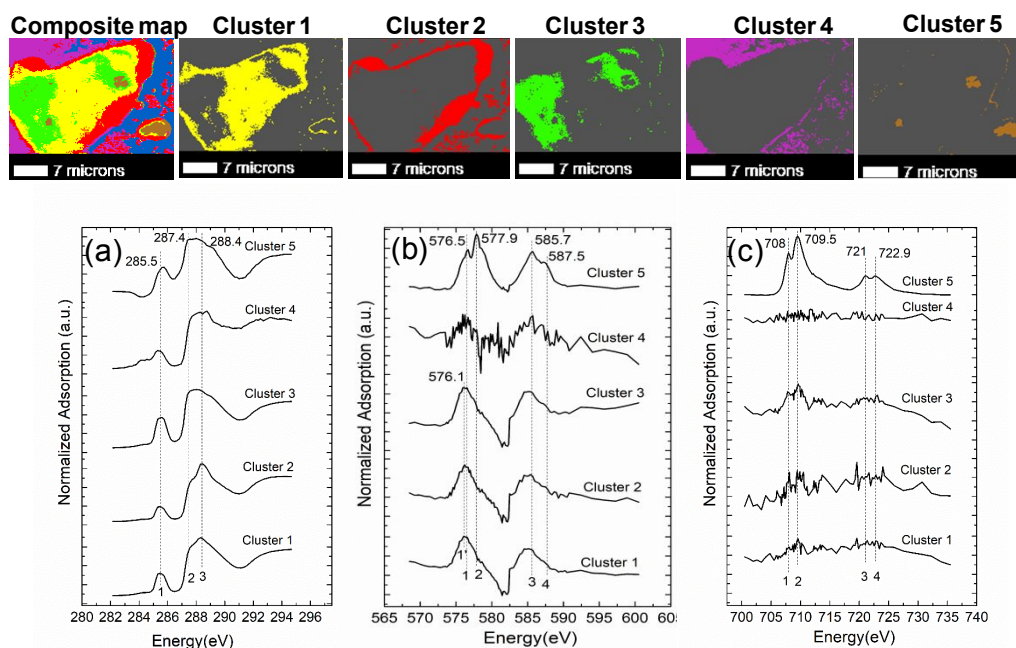


Figure S3. The PCA-cluster results of the sorption OFC sample synthesized using rice-derived DOC under the Fe/C molar ratio of 0.1 (Sor\_Ri\_0.1). Corresponding C K-edge, Cr and Fe L<sub>3,2</sub>-edge XANES spectra of each cluster identified is shown in (a), (b) and (c), respectively.

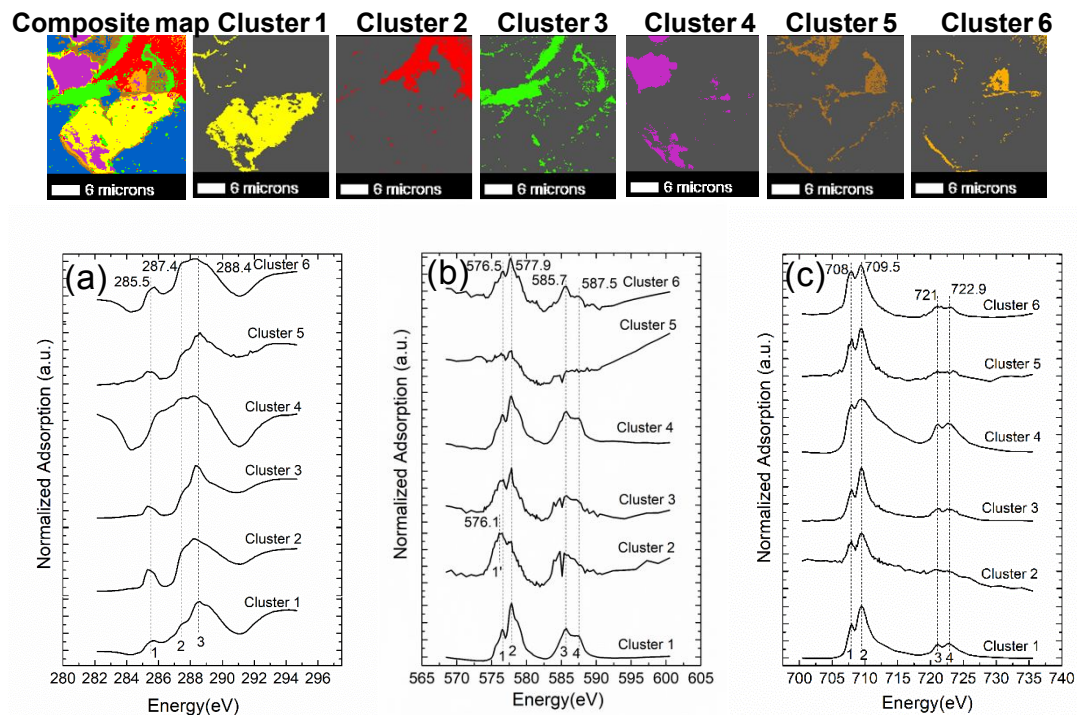
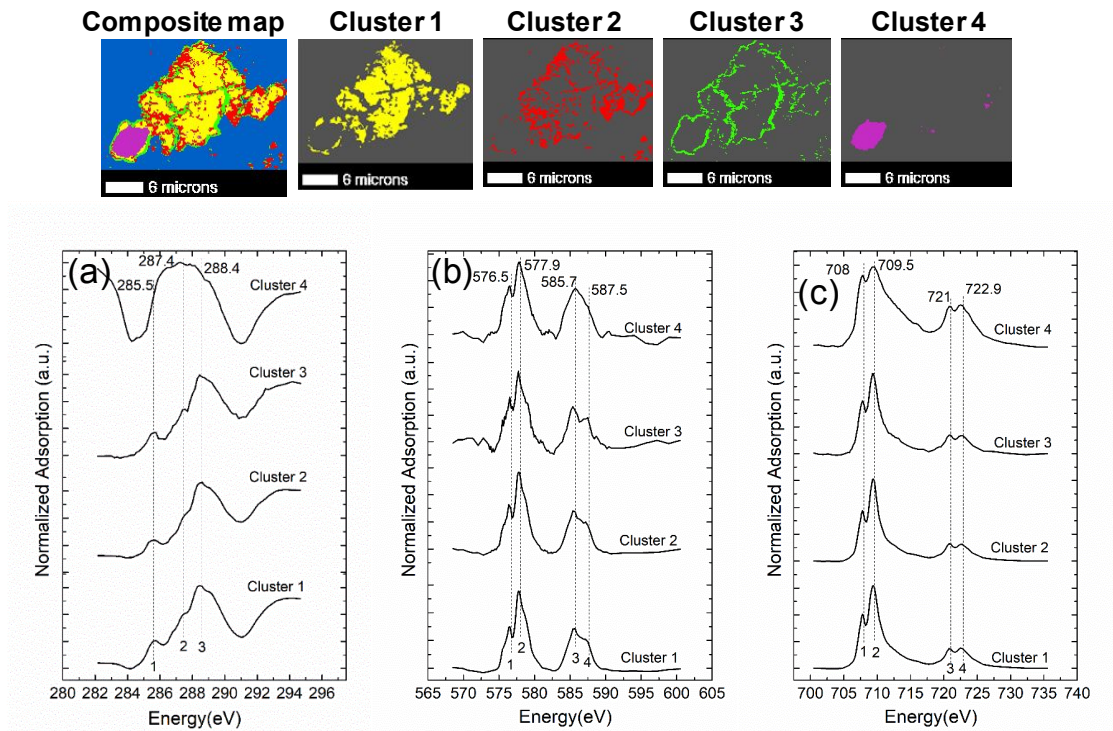


Figure S4. The PCA-cluster results of the sorption OFC sample synthesized using rape-derived DOC under the Fe/C molar ratio of 0.1 (Sor\_Ra\_0.1). Corresponding C K-edge, Cr and Fe L<sub>3,2</sub>-edge XANES spectra of each cluster identified is shown in (a), (b) and (c), respectively.



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147 Figure S5. The PCA-cluster results of the co-precipitation OFC sample synthesized using  
148 rice-derived DOC under the Fe/C molar ratio of 0.1 (Cop\_Ri\_0.1). Corresponding C K-edge, Cr  
149 and Fe L<sub>3,2</sub>-edge XANES spectra of each cluster identified is shown in (a), (b) and (c),  
150 respectively.

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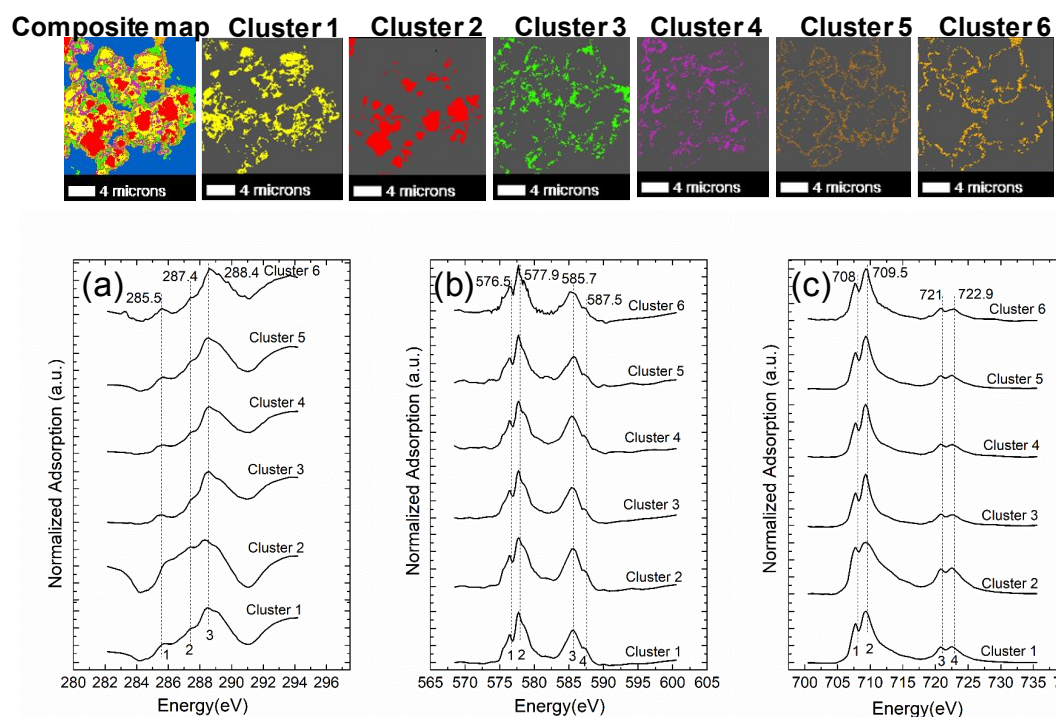
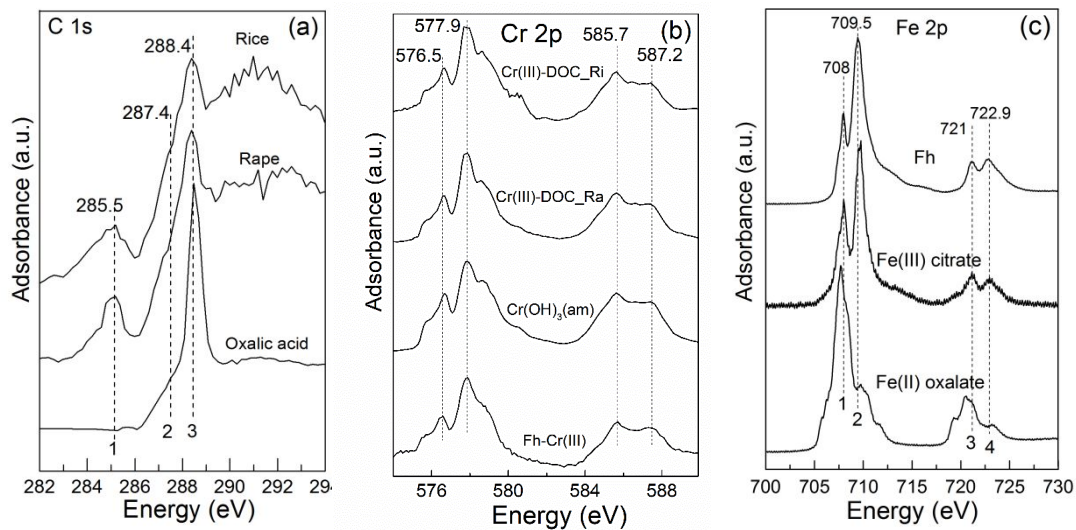


Figure S6. The PCA-cluster results of the co-precipitation OFC sample synthesized using rape-derived DOC under the Fe/C molar ratio of 0.1 (Cop\_Ra\_0.1). Corresponding C K-edge, Cr and Fe L<sub>3,2</sub>-edge XANES spectra of each cluster identified is shown in (a), (b) and (c), respectively.

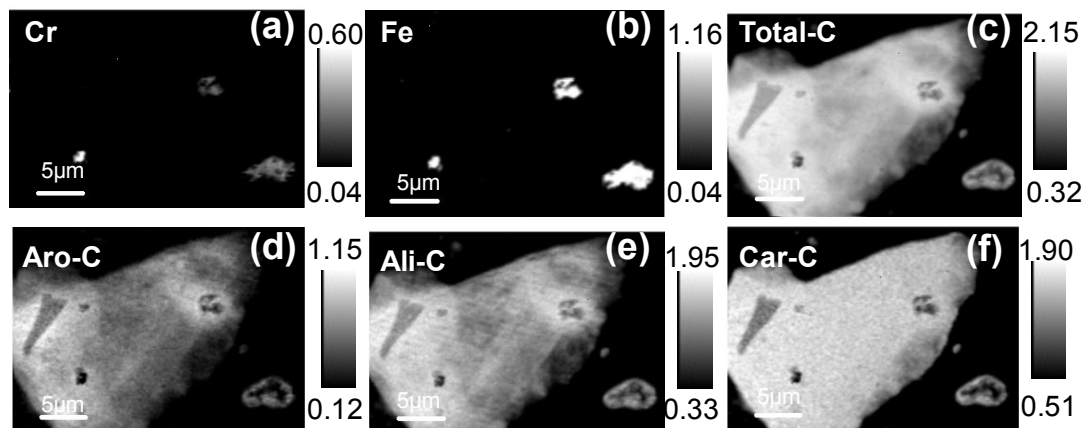


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Figure S7. The XANES spectra at the C K-edge of rice (DOC\_Ri) and rape derived DOC (DOC\_Ra) (a), at the Cr L<sub>3,2</sub>-edge of organo-Cr(III) complexes formed using DOC\_Ri and DOC\_Ra, amorphous Cr(OH)<sub>3</sub>(am) and Cr(III) adsorbed on Fh (Fh-Cr(III)) (b) and Fe L<sub>3,2</sub>-edge of ferrihydrite (Fh), Fe(III) citrate and Fe(II) oxalate (c) from previous studies,<sup>9, 10</sup> respectively.

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Figure S8. Submicro-scale distribution of Cr (a), Fe (b), total C (c, Total-C), aromatic carbon (d, Aro-C), aliphatic carbon (e, Ali-C) and carboxylic carbon (f, Car-C) in the sorption OFC sample, synthesized using rice-derived DOC under the Fe/C molar ratio of 0.1 (Sor\_Ri\_0.1) determined by STXM.

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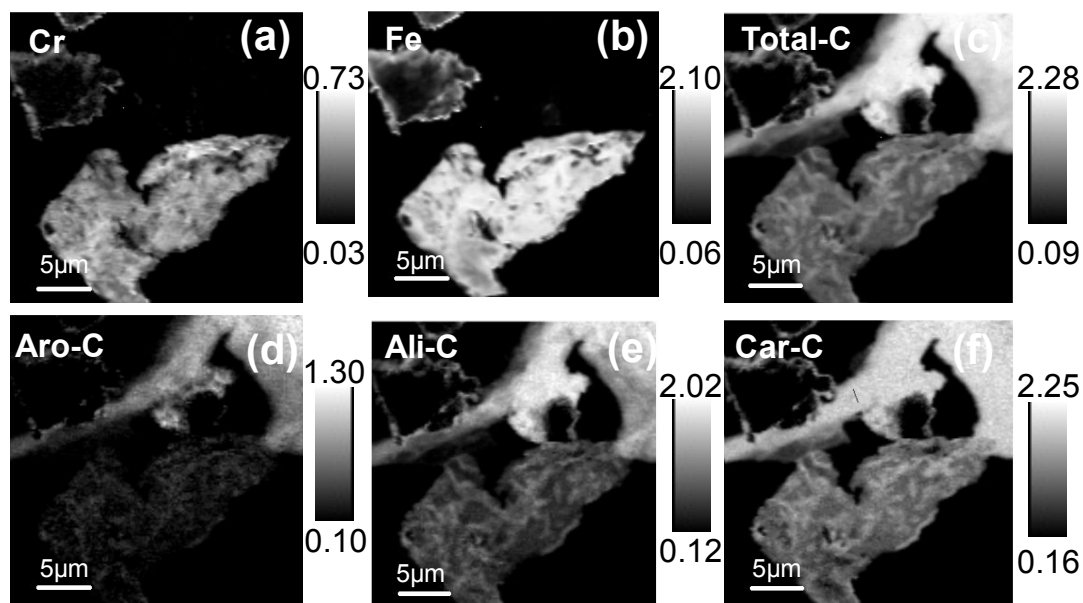


Figure S9. Submicro-scale distribution of Cr (a), Fe (b), total C (c, Total-C), aromatic carbon (d, Aro-C), aliphatic carbon (e, Ali-C) and carboxylic carbon (f, Car-C) in the sorption OFC sample, synthesized using rape-derived DOC under the Fe/C molar ratio of 0.1 (Sor\_Ra\_0.1 ) determined by STXM.

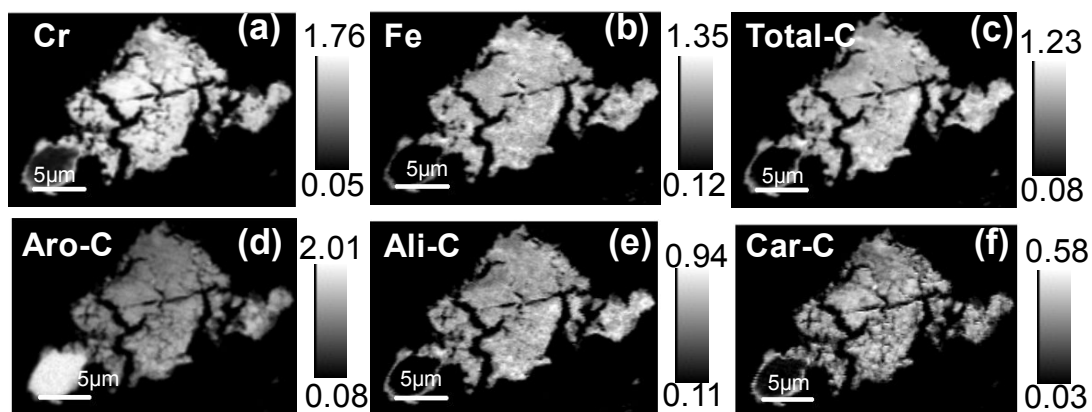


Figure S10. Submicro-scale distribution of Cr (a), Fe (b), total C (c, Total-C), aromatic carbon (d, Aro-C), aliphatic carbon (e, Ali-C) and carboxylic carbon (f, Car-C) in the co-precipitation OFC sample, synthesized using rice-derived DOC under the Fe/C molar ratio of 0.1 (Cop\_Ri\_0.1) determined by STXM.

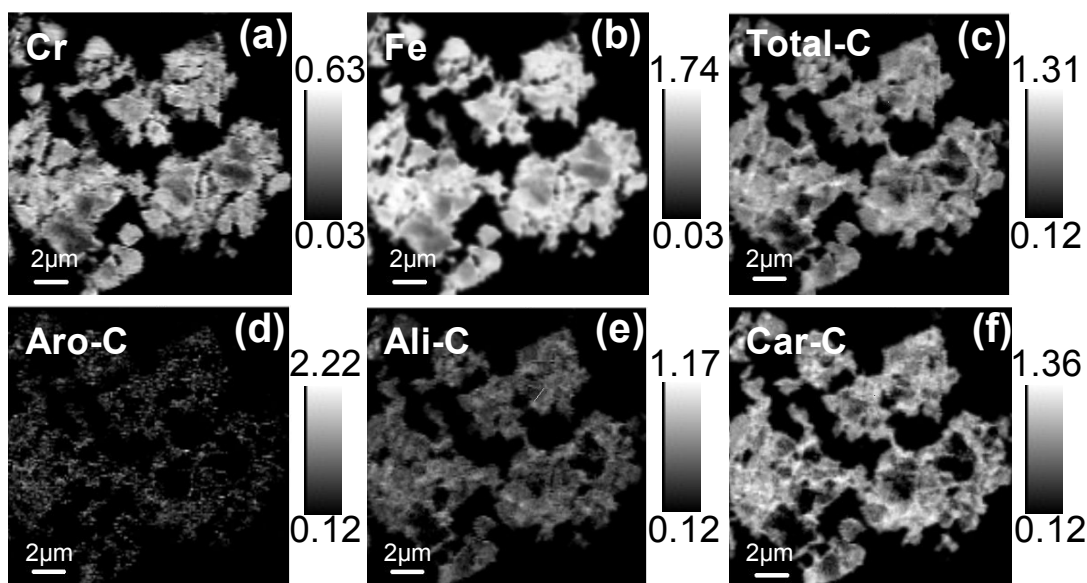


Figure S11. Submicro-scale distribution of Cr (a), Fe (b), total C (c, Total-C), aromatic carbon(d, Aro-C), aliphatic carbon (e, Ali-C) and carboxylic carbon (f, Car-C) in the co-precipitation OFC sample, synthesized using rape-derived DOC under the Fe/C molar ratio of 0.1 (Cop\_Ra\_0.1) determined by STXM.

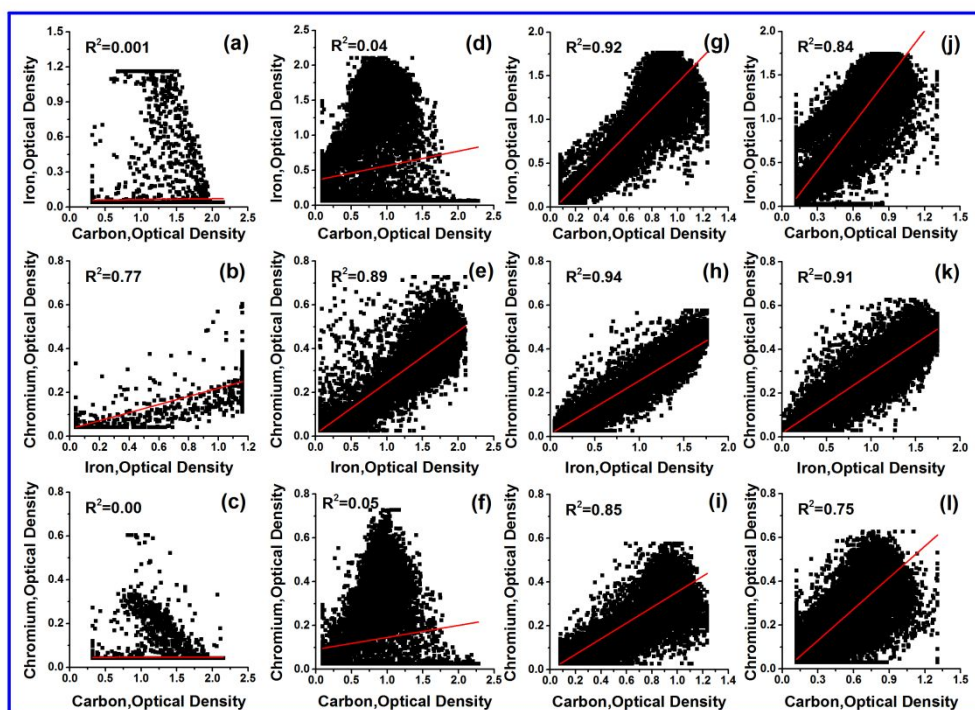


Figure S12. The elemental optical density correlation plots among C, Fe, and Cr for the sorption samples synthesized under the Fe/C molar ratio of 0.1 using rice- (Sor\_Ri\_0.1, a-c)

and rape-derived DOC (Sor\_Ra\_0.1, d-f), as well as the co-precipitation OFC samples under the Fe/C molar ratio of 0.1 using rice- (Cop\_Ri\_0.1, g-h) and rape-derived DOC (Cop\_Ra\_0.1, j-l), respectively.

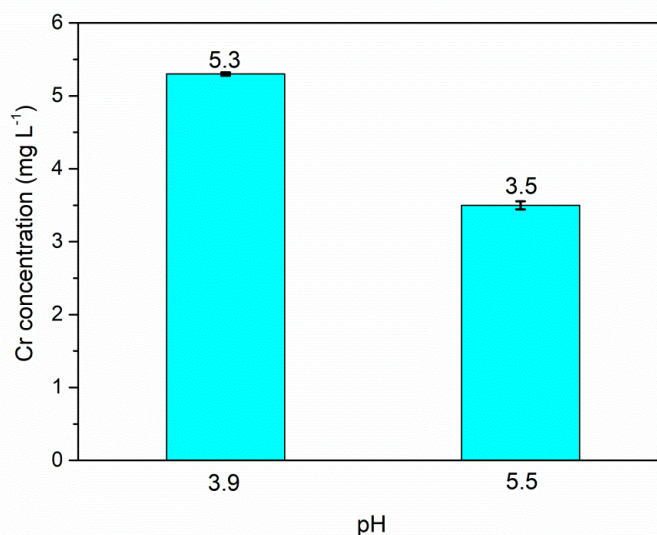


Figure S13. Concentrations of Cr in the supernatants of solutions containing 0.1 mM CrCl<sub>3</sub> and 10 mM NaCl at pH 3.9 (mean value, 5.3) and pH 5.5 (mean value, 3.5) after 24 h shaking.

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