Design of Ru(II)-NHC-Diamine Precatalysts Directed by Ligand Cooperation: Applications and Mechanistic Investigations for Asymmetric Hydrogenation

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(A) General

Unless otherwise noted, all reactions were carried out under an atmosphere of argon in oven-dried glassware. Reaction temperatures are reported as the temperature of the bath surrounding the vessel unless otherwise stated. The solvents used were purified by distillation over the drying agents indicated in parentheses and were transferred under argon: *n*-hexane (CaH₂), THF (Na-benzophenone), toluene (CaH₂), *i*-PrOH (molecular sieves).

All hydrogenation reactions were performed in Berghof High Pressure Reactors using hydrogen gas. Commercially available chemicals were bought from Acros Organics, Aldrich Chemical Co., Strem Chemicals, Alfa Aesar, ABCR, TCI Europe, Combi-Blocks and Chempur and used as received unless otherwise stated. NHC Ligands were synthesized following literature known procedures.¹ Chiral amines for the preparation of NHC ligands (R,R)-INpEt•Cl, (R,R)-SINpEt•HBF4, (S,S)-SINpEt•HBF4 were received from BASF SE. The chiral diamine ligands were purchased from Aldrich Chemical Co., Strem Chemicals, Alfa Aesar or prepared according to the literature.² All 3-substituted isocoumarin substrates **1** and benzothiophene 1,1-dioxides **3** were synthesized according to the literature.^{3,4} All analytical data were in agreement with the reported data.

Analytical thin layer chromatography was performed on Polygram SIL G/UV₂₅₄ plates and alox B. Visualization was accomplished with short wave UV light, and/or KMnO₄ staining solutions followed by heating. Flash chromatography was performed on Merck silica gel (40–63 mesh) by standard technique eluting with solvents as indicated.

¹H and ¹³C-NMR spectra were recorded on a Bruker AV 300, AV 400, Varian 500 MHz INOVA or Varian Unity plus 600 in the indicated solvents. Chemical shifts (δ) are given in ppm relative to TMS. The residual solvent signals were used as references and the chemical shifts converted to the TMS scale (CDCl₃: $\delta_H = 7.26$ ppm,

 $\delta_{\rm C} = 77.16$ ppm; THF- d_8 : $\delta_{\rm H} = 1.72$ ppm, $\delta_{\rm C} = 25.31$ ppm; toluene- d_8 : $\delta_{\rm H} = 2.08$ ppm, $\delta_{\rm C} = 20.43$ ppm). ESI mass spectra were recorded on a Bruker Daltonics MicroTof. Specific rotation was measured on a Perkin Elmer 341 polarimeter at 22 °C using a quartz glass cell (100 mm path length).

(B) Preparation of Ru(II)-NHC-diamine complexes

a. Procedure 1



The silver carbene complex was synthesized following a literature procedure:⁵ Ag₂O (0.55 mmol) was added to a solution of chloro-imidazolinium salt (*R*,*R*)-INpEt⁻HCl (1.0 mmol) in CH₂Cl₂ (15 mL). The mixture was stirred at room temperature for 24 h, filtered through celite and washed with CH₂Cl₂ (15 mL) to give a solution **C0** in CH₂Cl₂ (30 mL). Then [RuCl₂(benzene)]₂ (0.5 mmol) was added and the mixture was stirred at room temperature for 24 h. The mixture was filtered through celite and washed with CH₂Cl₂ (10 mL) to give the ruthenium-NHC complex in CH₂Cl₂ (40 mL). (*R*,*R*)-1,2-diphenylethylenediamine (DPEN) (1.0 mmol) was added to the solution and the mixture was stirred at room temperature for 16 h. The crude mixture was filtered through celite, washed with CH₂Cl₂, concentrated and purified by column chromatography on silica gel (pentane/ethylacetate = 20:1, later 10:1, 4 : 1) to yield the pure complex **C1** (56% yield over 3 steps).



¹**H** NMR (500 MHz, CDCl₃, connectivities were confirmed by gCOSY, gHSQC, and gHMBC experiments) δ 8.18 – 8.08 (m, 2H, *H*C18, *H*C28), 7.49 – 7.41 (m, 3H, *H*C27, *H*C23, *H*C12), 7.42 – 7.33 (m, 3H, *H*C17, *H*C15, *H*C22), 7.35 – 7.28 (m, 1H, *H*C6), 7.29 (d, *J* = 2.2 Hz, 1H, *H*C3), 7.27 – 7.21 (m,

2H, *H*C16, *H*C25), 7.17 (d, J = 2.2 Hz, 1H, *H*C2), 7.08 – 7.01 (m, 1H, *H*C26), 6.99 – 6.90 (m, 5H, *H*C32, *H*C34, *H*C33, *H*C39, *H*C21), 6.90 (d, J = 9.0 Hz, 1H, *H*C13), 6.81 (t, J = 7.7 Hz, 2H, *H*C38, *H*C40), 6.59 – 6.54 (m, 2H, *H*C35, *H*C31), 6.22 – 6.13 (m, 2H, *H*C37, *H*C41), 5.79 (q, J = 6.5 Hz, 1H, *H*C4), 4.99 (d, J = 5.8 Hz, 1H, *H*C11), 3.74 (td, J = 11.7, 4.8 Hz, 1H, *H*C9), 3.66 (dd, J = 10.6, 5.0 Hz, 1H, *H*₂N4), 3.18 – 3.04 (m, 2H, *H*C8, *H*₂N4), 2.00 (d, J = 7.0 Hz, 3H, *H*₃C7), 1.69 (dd, J = 12.2, 10.4 Hz, 1H, *H*₂N3), 1.45 (d, J = 6.5 Hz, 3H, *H*₃C5), 0.34 (dd, J = 10.3, 4.9 Hz, 1H, *H*₂N3). ¹³C **NMR** (126 MHz, 26 °C, CDCl₃, connectivities were confirmed by, gHSQC, gHMBC) δ 186.6 (C1), 140.3 (C20), 140.1 (C30), 138.8 (C19), 138.7 (C36), 136.5 (C12), 133.5 (C24), 131.1 (C14), 129.3 (C25), 128.8 (C29), 128.4 (C32, C34), 128.3 (C38, C40), 128.2 (C23), 127.6 (C40), 127.4 (C27), 127.3 (C39), 127.2 (C15), 126.8 (C31, C35), 126.5 (C37, C41), 126.1 (C18), 125.9 (C26), 125.8 (C16), 125.8 (C22), 125.6 (C17), 123.0 (C21), 122.4 (C13), 121.7 (C28), 118.6 (C3), 118.1 (C2), 92.0 (C10), 72.7 (C11), 65.5 (C9), 60.0 (C8), 57.1 (C4), 54.7 (C6), 22.0 (C7), 21.2 (C5).

ESI-MS: calculated [C₄₁H₄₀Cl₂N₄Ru + Na]⁺: 783.1566, found: 783.1569.





S5





gCOSY



gHMBC



b. Procedure 2



In a glove box, to a flame-dried screw-capped tube equipped with a magnetic stirring bar was added [Ru(2-methylallyl)₂(COD)] (1.5 mmol; COD = cyclooctadiene), (R,R)-SINpEt·HBF₄ (1.5 mmol), (R,R)-1,2-diphenylethylenediamine (1.5 mmol), and dry NaOt-Bu (1.8 mmol). The mixture was suspended in n-hexane (60 mL) and stirred at 40 °C for 4 days to give a black solution. HCl (4M in dioxane) (2.7 mmol) was then added to the reaction mixture at 0 °C. After stirring for 30 min at 0 °C, the mixture was concentrated and purified by column chromatography on silica gel (pentane/ethylacetate = 20:1, later 10:1, 4:1, 2:1) to yield the complexes C2 (3% yield) and C3 (40% yield).



¹**H NMR** (600 MHz, 26 °C, CDCl₃, connectivities were confirmed by gCOSY, gHSQC, and gHMBC experiments) δ 8.14 (d, J = 8.6 Hz, 1H, HC28), 8.11 (d, J = 8.2 Hz, 1H, HC18), 7.52 (d, J = 7.0 Hz, 1H, HC21), 7.48 (d, J = 8.1 Hz, 1H, HC23), 7.46 – 7.40 (m, 3H, HC27, HC22, HC12), 7.41 – 7.35 (m, 2H,

*H*C17, *H*C15), 7.33 (d, *J* = 8.1 Hz, 1H, *H*C25), 7.24 (d, *J* = 7.4 Hz, 1H, *H*C16), 7.10 (t, *J* = 7.5 Hz, 1H, *H*C26), 6.99 – 6.85 (m, 6H, *H*C33, *H*C32, *H*C34, *H*C6, *H*C13, *H*C39), 6.76 (t, J = 7.6 Hz, 2H, HC40, HC38), 6.57 (d, J = 6.7 Hz, 2H, HC31, HC35), 6.17 (d, J = 7.6 Hz, 2H, HC37, HC41), 5.20 (d, J = 5.8 Hz, 1H, HC11), 5.07 (q, J = 6.4 Hz, 1H, *H*C4), 4.03 (dd, *J* = 11.5, 7.2 Hz, 2H, *H*₂C3), 3.87 (dt, *J* = 9.1, 7.2 Hz, 1H, *H*₂C2), 3.77 -3.70 (m, 1H, HC9), 3.72 - 3.63 (m, 2H, H2C2, H₂N4), 3.19 (t, J = 11.0 Hz, 1H, H_2 N4), 3.13 (td, J = 12.0, 4.8 Hz, 1H, HC8), 1.83 (d, J = 7.0 Hz, 3H, H_3 C7), 1.77 (t, J= 11.4 Hz, 1H, H_2 N3), 1.21 (d, J = 6.4 Hz, 3H, H_3 C5), 0.43 (dd, J = 10.5, 4.9 Hz, 1H, H₂N3). ¹³C NMR (151 MHz, 26 °C, CDCl₃, connectivities were confirmed by, gHSQC, gHMBC) & 220.5 (C1), 140.2 (C30), 139.9 (C20), 139.0 (C19), 138.7 (C36), 136.5 (C12), 133.8 (C24), 131.2 (C14), 129.4 (C29), 129.4 (C25), 128.4 (C32, C34), 128.3 (C38, C40), 128.1 (C23), 127.6 (C33), 127.3 (C39), 127.2 (C15), 127.1 (C27), 126.8 (C31, C35), 126.5 (C18), 126.3 (C37, C41), 125.8 (C26), 125.8 (C16), 125.6 (C22), 125.5 (C17), 122.9 (C21), 122.7 (C13), 122.3 (C28), 93.1 (C10), 73.0 (C11), 65.6 (C9), 59.7 (C8), 56.9 (C4), 53.6 (C6), 47.1 (C3), 46.9 (C2), 20.9 (C7), 20.6 (C5). ESI-MS: calculated $[C_{41}H_{42}Cl_2N_4Ru - Cl]^+$: 727.2136, found: 727.2144.

S8



S9

gCOSY:



11.0 10.5 10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5 f2 (ppm)

Here the total

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120

130

-140 -150 -160

gHMBC:





¹H NMR (600 MHz, 26 °C, Toluene- d_8 , connectivities were confirmed by gCOSY, gHSQC, and gHMBC experiments) δ 8.70 (d, J = 8.2 Hz, 1H, HC18), 7.78 (d, J = 7.3 Hz, 1H, HC32), 7.50 – 7.40 (m, 2H, HC28, HC21), 7.38 – 7.34 (m, 2H, HC33, HC25), 7.32 (ddd, J = 8.3, 6.6, 1.8 Hz, 1H, HC17),

7.28 – 7.23 (m, 2H, *H*C23, *H*C27), 7.21 (t, J = 7.3 Hz, 1H, *H*C34), 7.18 (t, J = 7.7 Hz, 1H, *H*C22), 7.14 – 7.07 (m, 4H, *H*C15, *H*C16, *H*C37, *H*C41), 7.07 – 7.04 (m, 1H, *H*C26), 7.02 (d, J = 7.3 Hz, 1H, *H*C35), 6.72 – 6.67 (m, 2H, *H*C38, *H*C40), 6.67 – 6.63 (m, 1H, *H*C39), 6.48 – 6.37 (m, 2H, *H*C6, *H*C13), 6.27 (dd, J = 9.1, 5.7 Hz, 1H, *H*C12), 5.51 (q, J = 6.4 Hz, 1H *H*C4), 4.00 (dd, J = 10.8, 3.6 Hz, 1H, *H*2N4), 3.76 (d, J = 5.7 Hz, 1H, *H*C11), 3.30 – 3.22 (m, 2H, *H*₂C3), 3.21 (s, 1H, *H*C8), 3.14 (td, J = 8.4, 4.9 Hz, 1H, *H*₂C2), 3.09 – 2.99 (m, 1H, *H*₂C2), 2.85 (t, J = 10.2 Hz, 1H, *H*2N4), 2.75 (dd,

J = 9.8, 4.0 Hz, 1H, HC9), 1.43 (d, J = 7.0 Hz, 3H, H_3C7), 1.08 (d, J = 6.4 Hz, 3H, H_3C5), 1.04 (d, J = 9.4 Hz, 1H, H_2N3), -0.02 (d, J = 9.4 Hz, 1H, H_2N3). ¹³C NMR (151 MHz, 26 °C, Toluene- d_8 , connectivities were confirmed by gHSQC and gHMBC experiments) δ 225.3 (C1), 176.5 (C31), 147.7 (C30), 142.1 (C19), 140.3 (C20), 140.2 (C32), 139.7 (C36), 137.0 (C12), 134.3 (C24), 131.3 (C14), 129.9 (C29), 129.6 (C25), 128.8 (C38, C40), 128.3 (C23), 127.8 (C18), 127.5 (C39), 127.1 (C27), 126.9 (C15), 126.6, (C37, C41) 126.3 (C22), 125.9 (C17), 125.8 (C33), 125.7 (C26), 124.6 (C16), 123.8 (C21), 122.6 (C28), 121.8 (C34), 121.4 (C13), 121.4 (C35), 83.4 (C10), 67.5 (C8), 66.8 (C11), 62.8 (C9), 57.8 (C4), 53.4 (C6), 46.8 (C2), 46.6 (C3), 20.3 (C5), 20.1 (C7). **ESI-MS**: calculated [C₄₁H₄₁ClN₄Ru – Cl]⁺: 691.2369, found: 691.2374







gHSQC:







Complex C4 was prepared in 38% yield according to the Procedure 2 using (1R,2R)-1,2-di-ptolylethane-1,2-diamine.

¹H NMR (500 MHz, 26 °C, THF- d_8 , connectivities were confirmed by gCOSY, gHSQC and gHMBC experiments) δ 8.35 (d, J = 8.5 Hz, 1H, HC18),

7.81 – 7.76 (m, 1H, HC28), 7.75 (ddd, J = 8.2, 1.4, 0.7 Hz, 1H, HC25), 7.70 – 7.63 (m, 2H, HC23, HC32), 7.57 (dt, J = 7.2, 1.0 Hz, 1H, HC21), 7.50 – 7.44 (m, 1H, HC27), 7.40 (dd, J = 8.2, 7.2 Hz, 1H, HC22), 7.36 (ddd, J = 8.0, 6.8, 1.1 Hz, 1H, HC26), 7.24 -7.20 (m, 2H, HC37, HC41), 7.20 - 7.17 (m, 1H, HC17), 7.07 (dd, J = 7.7, 1.7 Hz, 1H, HC15), 7.05 – 7.01 (m, 1H, HC16), 6.96 (d, J = 7.4 Hz, 1H, HC35), 6.88 – 6.84 (m, 2H, *H*C38, *H*C40), 6.81 (ddd, *J* = 7.4, 1.8, 0.7 Hz, 1H, *H*C34), 6.51 – 6.43 (m, 1H, HC13), 6.38 (q, J = 7.0 Hz, 1H, HC6), 6.05 (dd, J = 9.0, 5.7 Hz, 1H, HC12), 5.29 (q, J= 6.4 Hz, 1H, HC4), 4.10 – 4.04 (m, 1H, H₂N4), 3.85 – 3.74 (m, 1H, H₂C3), 3.70 (d, J = 5.7 Hz, 1H, HC11), 3.68 (s, 1H, HC8), 3.68 – 3.63 (m, 1H, H₂C2), 3.64 – 3.58 (m, 1H, H_2 C3), 3.55 – 3.45 (m, 1H, H_2 N4), 3.42 – 3.32 (m, 1H, H_2 C2), 3.11 – 3.04 (m, 1H, *H*C9), 2.47 (s, 3H, *H*₃C42), 2.12 (s, 3H, *H*₃C43), 1.72 (d, *J* = 3.4 Hz, 3H, *H*₃C7), 1.16 -1.10 (m, 4H, H_3 C5, H_2 N3), 0.10 (d, J = 9.4 Hz, 1H, H_2 N3). ¹³C NMR (126 MHz, 26 °C, THF- d_8 connectivities were confirmed by gHSQC and gHMBC experiments) δ 225.8 (C1), 176.8 (C31), 145.8 (C30), 142.7 (C19), 141.6 (C32), 140.4 (C20), 138.0 (C36), 137.7 (C12), 137.3 (C39), 134.9 (C24), 134.5 (C33), 131.9 (C14), 131.3 (C29), 129.8 (C38, C40), 129.6 (C25), 128.6 (C23), 128.1 (C18), 127.9 (C27), 127.3 (C37, C41), 126.9 (C15), 126.3 (C22), 126.3 (C26), 125.5 (C17), 124.5 (C21), 124.4 (C16), 124.3 (C28), 122.7 (C34), 121.4 (C13), 121.3 (C35), 82.9 (C10), 68.0 (C8), 67.1 (C11), 63.0 (C9), 58.5 (C4), 54.2, (C6) 47.8 (C2), 46.7 (C3), 21.7 (C42), 20.8 (C43), 20.5 (C5), 20.0 (C7). **ESI-MS**: calculated [C₄₃H₄₅ClN₄Ru – Cl]⁺: 719.2682, found: 719.2692







gHMBC





Complex C5 was isolated in 34% yield according to the Procedure 2 using (1R,2R)-1,2-bis(4-methoxyphenyl)ethane-1,2-diamine.

¹H NMR (500 MHz, 26 °C, THF- $d_{8,}$ connectivities were confirmed by gCOSY, gHSQC and gHMBC experiments) δ 8.34 (d, J

= 8.1 Hz, 1H, *H*C18), 7.89 (d, J = 8.5 Hz, 1H, *H*C28), 7.78 – 7.71 (m, 1H, *H*C25), 7.67 (d, J = 8.2 Hz, 1H, *H*C23), 7.56 (d, J = 7.2 Hz, 1H, *H*C21), 7.49 (ddd, J = 8.5, 6.8, 1.4 Hz, 1H, *H*C27), 7.43 – 7.38 (m, 1H, *H*C22), 7.39 (d, J = 2.4 Hz, 1H, *H*C32), 7.36 (ddd, J = 7.9, 6.9, 0.9 Hz, 1H, *H*C26), 7.26 (d, J = 8.6 Hz, 2H, *H*C37, *H*C41), 7.19 (ddd, J = 8.3, 6.9, 1.6 Hz, 1H, *H*C17), 7.07 (dd, J = 7.6, 1.5 Hz, 1H, *H*C15), 7.03 (ddd, J = 7.7, 7.0, 1.1 Hz, 1H, *H*C16), 6.96 (d, J = 8.1 Hz, 1H, *H*C35), 6.63 – 6.57 (m, 2H, *H*C38, *H*C40), 6.55 (dd, J = 8.0, 2.5 Hz, 1H, *H*C34), 6.47 (d, J = 9.1 Hz, 1H, *H*C13), 6.33 (q, J = 6.9 Hz, 1H, *H*C6), 6.06 (dd, J = 9.0, 5.7 Hz, 1H, *H*C12), 5.26 (q, J = 6.4 Hz, 1H,

*H*C4), 3.96 (dd, J = 10.9, 3.4 Hz, 1H, H_2 N4), 3.89 (s, 3H, H_3 C42), 3.78 (dt, J = 11.7, 9.3 Hz, 1H, H_2 C3), 3.73 (d, J = 5.7 Hz, 1H, HC11), 3.66 (dd, J = 9.0, 2.3 Hz, 1H, H_2 C2), 3.62 (s, 1H, HC8), 3.58 (s, 3H, H_3 C43), 3.58 – 3.54 (m, 2H, H_2 N4, H_2 C3), 3.39 – 3.29 (m, 1H, H_2 C2), 3.08 (d, J = 9.1 Hz, 1H, HC9), 1.74 (d, J = 6.9 Hz, 3H, H_3 C7), 1.13 – 1.08 (m, 4H, H_3 C5, H_2 N3), 0.06 (d, J = 9.5 Hz, 1H, H_2 N3). ¹³C **NMR** (126 MHz, 26 °C, THF- d_8 , connectivities were confirmed by gHSQC and gHMBC experiments) δ 225.4 (C1), 178.5 (C31), 159.6 (C39), 158.3 (C33), 142.5 (C19), 141.1 (C30), 140.0 (C20), 137.4 (C12), 134.7 (C24), 132.6 (C36), 131.7 (C14), 131.1 (C29), 129.4 (C25), 128.5 (C23), 128.3 (C37, C41), 127.9 (C18), 127.7 (C27), 126.8 (C13), 126.1 (C22, C26), 125.7 (C32), 125.3 (C17), 124.4 (C21), 124.3 (C16, C28), 121.7 (C35), 121.4 (C13), 114.3 (C38, C40), 107.5 (C34), 82.9 (C10), 67.5 (C3), 20.4 (C5), 20.0 (C7). **ESI-MS**: calculated [C4₃H₄₅ClN₄O₂Ru – C1]⁺: 751.2581, found: 751.2587.





gHSQC:





Complex C6 was isolated in 32% yield according to the Procedure 2 using (1R,2R)-1,2-bis(4-fluorophenyl)ethane-1,2-diamine.

¹**H NMR** (600 MHz, 26 °C, CDCl₃, connectivities were confirmed by gCOSY, gHSQC and gHMBC experiments) δ 8.36 (d, *J* = 8.0 Hz, 1H, *H*C18), 7.75

-7.73 (m, 1H, HC25), 7.65 (d, J = 8.2 Hz, 1H, HC23), 7.52 (d, J = 7.2 Hz, 1H, HC21), 7.49 – 7.44 (m, 2H, HC32, HC22), 7.39 (ddd, J = 8.4, 5.5, 3.1 Hz, 1H, HC17), 7.35 (ddd, *J* = 8.1, 6.9, 1.1 Hz, 1H, *H*C26), 7.31 (ddd, *J* = 8.3, 6.9, 1.5 Hz, 1H, *H*C27), 7.20 -7.14 (m, 3H, HC28, HC15, HC16), 7.01 (dd, J = 8.2, 5.6 Hz, 1H, HC35), 6.95 (ddt, J = 8.3, 5.3, 2.6 Hz, 2H, HC38, HC40), 6.78 (ddd, J = 9.1, 8.1, 2.6 Hz, 1H, HC34), 6.76 -6.72 (m, 2H, HC37, HC41), 6.57 (dt, J = 9.1, 0.7 Hz, 1H, HC13), 6.43 (q, J = 7.1 Hz, 1H, HC6), 6.17 (dd, J = 9.0, 5.7 Hz, 1H, HC12), 5.44 (q, J = 6.4 Hz, 1H, HC4), 4.08 -4.01 (m, 1H, H₂C3), 4.02 – 3.96 (m, 1H, H₂C3), 3.84 – 3.78 (m, 1H, H₂C2), 3.73 (d, J = 5.5 Hz, 1H, HC11), 3.70 (dt, J = 10.8, 1.8 Hz, 1H, H₂C2), 3.67 (s, 1H, HC8), 3.44 - $3.38 (m, 1H, H_2N4), 2.84 - 2.74 (m, 2H, H_2N4, HC9), 1.68 (d, J = 7.1 Hz, 3H, H_3C7),$ $1.20 (d, J = 6.5 Hz, 3H, H_3C5), 1.10 (d, J = 9.5 Hz, 1H, H_2N3), 0.13 (d, J = 9.9 Hz, 1H, H_2N3)$ H₂N3). ¹³C NMR (151 MHz, 26 °C, CDCl₃, connectivities were confirmed by gHSQC and gHMBC experiments) δ 223.8 (C1), 179.74 (d, J = 2.9 Hz, C31), 162.03 (d, J =247.0 Hz, C39), 161.41 (d, J = 247.1 Hz, C33), 142.85 (d, J = 1.9 Hz, C30), 141.1 (C19), 140.2 (C20), 136.1 (C12), 134.83 (d, J = 3.2 Hz, C36), 134.0 (C24), 130.8 (C14),129.7 (C25), 129.0 (C29), 128.1 (C23), 128.04 (d, *J* = 8.0 Hz, C38, C40), 126.9 (C27), 126.9 (C15), 126.7 (C18), 126.6 (C22), 125.8 (C17), 125.7 (C26), 124.98 (d, J = 15.8 Hz, C32), 124.7 (C16), 123.4 (C21), 121.61 (d, *J* = 8.0 Hz, C35), 121.5 (C13), 121.4 (C28), 115.80 (d, *J* = 21.4 Hz, C37, C41), 108.34 (d, *J* = 22.4 Hz, C34), 85.3 (C10), 68.5 (C11), 66.2 (C8), 62.0 (C9), 57.0 (C4), 53.5 (C6), 47.3 (C3), 46.7 (C2), 20.7 (C7), 20.3 (C5). ¹⁹F NMR (564 MHz, CDCl₃) δ -114.6, -118.4. ESI-MS: calculated $[C_{41}H_{39}ClF_2N_4Ru - Cl]^+$: 727.2181, found: 727.2171.







----118.44

gHSQC:



gHMBC:





Complex C4a was isolated in 12% yield according to the Procedure 2 using (1R,2R)-1,2-bis(2methylphenyl)ethane-1,2-diamine.

¹H NMR (600 MHz, 26 °C, Chloroform-*d*, connectivities were confirmed by gCOSY, gHSQC,

and gHMBC experiments) δ 8.40 (d, J = 8.2 Hz, 1H, HC18), 7.72 (dd, J = 8.2, 1.3 Hz, 1H, HC28), 7.66 – 7.60 (m, 3H, HC22, HC32, HC41), 7.52 (d, J = 7.2 Hz, 1H, HC21), 7.43 – 7.38 (m, 2H, HC16, HC23), 7.34 (ddd, J = 8.0, 6.8, 1.1 Hz, 1H, HC26), 7.25 (ddd, J = 8.2, 6.9, 1.1 Hz, 1H, HC27), 7.20 (t, J = 7.4 Hz, 1H, HC33), 7.18 - 7.15 (m, HC27))2H, HC15, HC17), 7.02 - 6.96 (m, 2H, HC25, HC39), 6.95 - 6.89 (m, 2H, HC34, *H*C40), 6.86 (d, *J* = 7.4 Hz, 1H, *H*C38), 6.70 (q, *J* = 7.0 Hz, 1H, *H*C6), 6.55 (d, *J* = 9.0 Hz, 1H, HC13), 6.22 (dd, J = 9.0, 5.7 Hz, 1H, HC12), 5.50 (q, J = 6.4 Hz, 1H, HC4), 4.03 (m, 1H, H₂C2), 3.99 – 3.92 (m, 2H, H₂C2, HC8), 3.82 – 3.79 (m, 1H, H₂C3), 3.78 $(d, J = 5.7 \text{ Hz}, 1\text{H}, H\text{C}11), 3.73 - 3.66 \text{ (m, 1H, } H_2\text{C}3), 3.51 \text{ (dd, } J = 11.2, 4.1 \text{ Hz}, 1\text{H},$ H_2 N4), 3.02 – 2.97 (m, 1H, HC9), 2.89 (t, J = 10.5 Hz, 1H, H_2 N4), 2.33 (s, 3H, H_3 C42), 1.66 (s, 3H, H_3 C43), 1.59 (d, J = 7.1 Hz, 3H, H_3 C7), 1.22 (d, J = 6.5 Hz, 3H, H_3 C5), 0.89 - 0.83 (m, 1H, H_2N_3), 0.14 (d, J = 9.7 Hz, 1H, H_2N_3). ¹³C NMR (151 MHz, 26 °C, Chloroform-d connectivities were confirmed by gHSQC, and gHMBC experiments) & 225.1 (C1), 176.0 (C31), 144.9 (C35), 141.5 (C14), 140.9 (C20), 138.2 (C36, C32), 136.8 (C12), 134.3 (C37), 134.1 (C24), 130.9 (C19), 130.6 (C38), 129.8 (C30), 129.6 (C28), 129.1 (C29), 127.9 (C22), 127.4 (C39), 127.3 (C40), 126.7 (C15, C18), 126.6 (C27), 126.5 (C23), 126.1 (C41), 125.7 (C16), 125.5 (C26), 125.4 (C33), 124.4 (C17), 123.4 (C34), 123.2 (C21), 122.0 (C25), 120.9 (C13), 84.8 (C10), 68.1 (C11), 61.2 (C8), 57.1 (C4), 56.8 (C9), 53.3 (C6), 47.3 (C3), 46.7 (C2), 21.0 (C7, C42), 20.3 (C5), 19.7 (C43). **ESI-MS**: calculated $[C_{43}H_{45}CIN_4Ru - CI]^+$: 719.2694, found: 719.2690.



S27

gCOSY:



gHSQC:



gHMBC:



(C) X-Ray diffraction of complexes C1, C2, and C3

X-Ray diffraction: Data sets for compound C1 were collected with a Nonius Kappa CCD diffractometer. Programs used: data collection, COLLECT (R. W. W. Hooft, Bruker AXS, 2008, Delft, The Netherlands); data reduction Denzo-SMN (Z. Otwinowski, W. Minor, *Methods Enzymol.* 1997, *276*, 307-326); absorption correction, Denzo (Z. Otwinowski, D. Borek, W. Majewski, W. Minor, *Acta Crystallogr.* 2003, *A59*, 228-234); structure solution SHELXS-97 (G. M. Sheldrick, *Acta Crystallogr.* 1990, *A46*, 467-473); structure refinement SHELXL-97 (G. M. Sheldrick, *Acta Crystallogr.* 2008, *A64*, 112-122). Data sets for compounds C2 and C3 were collected with a D8 Venture CMOS diffractometer. Programs used: data collection: APEX3 V2016.1-0 (Bruker AXS Inc., 2016); cell refinement: SAINT V8.37A (Bruker AXS Inc., 2015); data reduction: SAINT V8.37A (Bruker AXS Inc., 2015); absorption correction, SADABS V2014/7 (Bruker AXS Inc., 2014); structure solution SHELXT-2015 (Sheldrick, 2015); structure refinement SHELXL-2015 (Sheldrick, 2015) and graphics, XP (Bruker AXS Inc., 2015). *R*-values are given for observed reflections, and *w*R² values are given for all reflections.

Exceptions and special features: For compound C1 one phenyl group and one naphthyl group and for compound C3 one phenyl group were found disordered over two positions in the asymmetric unit. Several restraints (SADI, SAME, ISOR and SIMU) were used in order to improve refinement stability. For compound C1 a badly disordered THF molecule and for compound C3 four badly disordered THF molecules were found in the asymmetrical unit and could not be satisfactorily refined. The program SQUEEZE (Spek, A. L. (2015). *Acta Cryst. C71*, 9-18.) was therefore used to remove mathematically the effect of the solvent. The quoted formula and derived parameters are not included the squeezed solvent molecules.

X-ray crystal structure analysis of C1 (glo9197): formula C₄₁H₄₀Cl₂N₄Ru, M = 760.74, orange crystal, 0.38 x 0.04 x 0.02 mm, a = 22.9420(8) Å, b = 10.7497(5) Å, c = 17.5379(8) Å, $\beta = 92.037(2)^{\circ}$, V = 4322.50(3) Å³, $\rho_{calc} = 1.169$ gcm⁻³, $\mu = 0.516$ mm⁻¹, empirical absorption correction (0.828 $\leq T \leq 0.990$), Z = 4, monoclinic, space group C2 (No. 5), $\lambda = 0.71073$ Å, T = 173(2) K, ω and φ scans, 19941 reflections collected ($\pm h, \pm k, \pm l$), 7260 independent ($R_{int} = 0.070$) and 6371 observed reflections [$l > 2\sigma I$)], 575 refined parameters, R = 0.053, $wR^2 = 0.124$, max. (min.) residual electron density 0.69 (-0.50) e.Å⁻³, the hydrogen atoms at N3 and N4 were refined freely, but with N-H distance restraint (DFIX and U-fixed value); others hydrogen atoms were calculated and refined as riding atoms. Flack parameter was refined to 0.04(4). CCDC Nr.: 1879259.



Figure S1. Crystal structure of compound **C1**. (Thermals ellipsoids are shown with 15% probability.)

X-ray crystal structure analysis of C2 (glo8770): A orange prism-like specimen of C41H42Cl2N4Ru, approximate dimensions 0.103 mm x 0.139 mm x 0.188 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured. A total of 1604 frames were collected. The total exposure time was 24.13 hours. The frames were integrated with the Bruker SAINT software package using a wide-frame algorithm. The integration of the data using a monoclinic unit cell yielded a total of 28574 reflections to a maximum θ angle of 72.07° (0.81 Å resolution), of which 6769 were independent (average redundancy 4.221, completeness = 98.8%, R_{int} = 3.63%, R_{sig} = 4.17%) and 6649 (98.23%) were greater than $2\sigma(F^2)$. The final cell constants of <u>a</u> = 12.4948(6) Å, <u>b</u> = 11.9927(5) Å, <u>c</u> = 13.5247(6) Å, β = 116.6000(10)°, volume = 1812.12(14) Å³, are based upon the refinement of the XYZ-centroids of 9189 reflections above 20 $\sigma(I)$ with 7.913° < 2 θ < 144.1°. Data were corrected for absorption effects using the multi-scan method (SADABS). The ratio of minimum to maximum apparent transmission was 0.841. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.4460 and 0.6210. The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group P21, with Z = 2 for the formula unit, $C_{41}H_{42}Cl_2N_4Ru$. The final anisotropic full-matrix leastsquares refinement on F^2 with 455 variables converged at R1 = 2.18%, for the observed data and wR2 = 5.28% for all data. The goodness-of-fit was 1.041. The largest peak in the final difference electron density synthesis was $0.336 \text{ e}^{-1}\text{Å}^{3}$ and the largest hole was -0.591 e⁻/Å³ with an RMS deviation of 0.055 e⁻/Å³. On the basis of the final model, the calculated density was 1.398 g/cm³ and F(000), 788 e⁻. The hydrogens at N3, N4 and C12 atoms were refined freely. Flack parameter was refined to 0.06(1). CCDC Nr.: 1879260.



Figure S2. Crystal structure of compound C2. (Thermals ellipsoids are shown with 30% probability.)

X-ray crystal structure analysis of C3 (glo8794): A yellow needle-like specimen of C₄₁H₄₁ClN₄Ru · 0.5 x C₄H₈O, approximate dimensions 0.051 mm x 0.064 mm x 0.163 mm, was used for the X-ray crystallographic analysis. The X-ray intensity data were measured. A total of 1563 frames were collected. The total exposure time was 22.61 hours. The frames were integrated with the Bruker SAINT software package using a wide-frame algorithm. The integration of the data using a triclinic unit cell yielded a total of 45033 reflections to a maximum θ angle of 68.80° (0.83 Å resolution), of which 15014 were independent (average redundancy 2.999, completeness = 99.2%, R_{int} = 7.48%, R_{sig} = 7.39%) and 13253 (88.27%) were greater than $2\sigma(F^2)$. The final cell constants of <u>a</u> = 12.3170(4) Å, <u>b</u> = 13.9678(4) Å, <u>c</u> = 15.0411(5) Å, α = 63.519(2)°, β = 80.198(2)°, γ = 67.747(2)°, volume = 2143.69(13) Å³, are based upon the refinement of the XYZ-centroids of 249 reflections above 20 $\sigma(I)$ with 7.504° < 20 < 103.1°. Data were corrected for absorption effects using the multi-scan method (SADABS). The

ratio of minimum to maximum apparent transmission was 0.820. The calculated minimum and maximum transmission coefficients (based on crystal size) are 0.5780 and 0.8310. The structure was solved and refined using the Bruker SHELXTL Software Package, using the space group *P*1, with Z = 2 for the formula unit, C₄₁H₄₁ClN₄Ru · 0.5 x C₄H₈O. The final anisotropic full-matrix least-squares refinement on F² with 972 variables converged at R1 = 4.78%, for the observed data and wR2 = 10.55% for all data. The goodness-of-fit was 1.036. The largest peak in the final difference electron density synthesis was 0.695 e⁻/Å³ and the largest hole was -0.726 e⁻/Å³ with an RMS deviation of 0.066 e⁻/Å³. On the basis of the final model, the calculated density was 1.181 g/cm³ and F(000), 792 e⁻. The hydrogens at N3A, N4A, N3B, N4B, C12A and C12B atoms were refined freely, but with N-H and C-H distance restraints (DFIX and U-fixed value). Flack parameter was refined to -0.01(1). CCDC Nr.: 1879261.



Figure S3. Crystal structure of compound **C3**. Only one molecule (molecule "A") of two found in the asymmetric unit is shown. (Thermals ellipsoids are shown with 30% probability.)

- APEX3 (2016), SAINT (2015) and SADABS (2014), Bruker AXS Inc., Madison, Wisconsin, USA.
- 2. SHELX software: Sheldrick, G. M. Acta Cryst., 2015, A71, 3-8.

(D) General procedure for the enantioselective hydrogenations

a. Ru(II)-catalyzed asymmetric hydrogenation of isocoumarins:

To a glass vial, complex C4 (0.004 mmol, 2 mol%), NaOt-Bu (0.02 mmol, 10 mol%), isocoumarin 1 (0.20 mmol), and *n*-hexane (4 mL) were added under an argon atmosphere. The glass vial was placed in a 150 mL stainless steel autoclave under an argon atmosphere. The autoclave was pressurized and depressurized with hydrogen gas five times before the indicated pressure (50 bar or 80 bar) was set. The reaction mixture was stirred at 15–25 °C for the indicated time. After the autoclave was carefully depressurized, the mixture was directly purified by flash column chromatography on silica gel (pentane/ethylacetate = 50:1, later 30:1, 20:1, 10:1) to afford the desired product **2**. The e.r. value of the product was determined by HPLC analysis using chiral column AS-H or AD-H.

Substrate Scope of the Ru(II)-Catalyzed Enantioselective Hydrogenation of Isocoumarins^a



^{*a*}Unless otherwise noted, reactions were carried out with **1** (0.2 mmol), **C4** (0.004 mmol, 2 mol%), and NaO*t*-Bu (0.02 mmol, 10 mol%) under 50 bar H₂ in *n*-hexane (4.0 mL) at 15–25 °C for the indicated time. Isolated yields after column chromatography are reported. E.r. values were determined by HPLC analysis using a chiral stationary phase. Results in parentheses are our previously reported data (*J. Am. Chem. Soc.* **2017**, *139*, 2585) using in-situ prepared catalyst solution. ^{*b*}Using a solvent mixture of *n*-hexane : toluene (3 : 1). ^{*c*}Using 80 bar of H₂.

3-methylisochroman-1-one (2a)



Colorless oil; H₂ (50 bar), 15 °C, 18 h, 83% yield, 98.5:1.5 e.r. $[\alpha]_D^{22} =$ +141.0 (c = 1.00 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 80/20, flow rate = 1 mL/min, λ = 230 nm,

retention time: 9.0 min (major), 10.9 min (minor). ¹**H NMR** (500 MHz, CDCl₃) δ 8.15 - 8.02 (m, *J* = 7.8 Hz, 1H), 7.53 (td, *J* = 7.5, 1.3 Hz, 1H), 7.38 (t, *J* = 7.6 Hz, 1H), 7.23 (d, *J* = 7.6 Hz, 1H), 4.71 - 4.65 (m, 1H), 3.06 - 2.87 (m, 2H), 1.52 (d, *J* = 6.3 Hz, 3H).


Known compound, see: Li, W.; Wiesenfeldt. M. P.; Glorius, F. J. Am. Chem. Soc. 2017,

139, 2585.

3,6-dimethylisochroman-1-one (2b)

Colorless solid; H₂ (50 bar), 25 °C, 22 h, 85% yield, 96:4 e.r. $[\alpha]_D^{22}$ =+120.4 (c = 1.20 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 80/20, flow rate = 1 mL/min, λ = 254 nm, retention time: 10.3 min (major), 12.8 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.96 (d, *J* = 7.9 Hz, 1H), 7.17 (d, *J* = 7.9 Hz, 1H), 7.02 (s, 1H), 4.64 (dqd, *J* = 10.1, 6.3, 3.8 Hz, 1H), 2.88 (qd, *J* = 16.2, 7.3 Hz, 2H), 2.39 (s, 3H), 1.50 (d, *J* = 6.3 Hz, 3H). Known compound see: Li, W.; Wiesenfeldt. M. P.; Glorius, F. *J. Am. Chem. Soc.* 2017, *139*, 2585.



3,7-dimethylisochroman-1-one (2c)



retention time: 7.6 min (major), 11.3 min (minor). ¹**H NMR** (500 MHz, CDCl₃) δ 7.89 (dd, J = 1.2, 0.5 Hz, 1H), 7.37 – 7.29 (m, 1H), 7.11 (d, J = 7.7 Hz, 1H), 4.64 (dqd, J = 10.0, 6.3, 4.7 Hz, 1H), 2.96 – 2.79 (m, 2H), 2.37 (s, 3H), 1.50 (d, J = 6.3 Hz, 3H). Known compound, see: Li, W.; Wiesenfeldt. M. P.; Glorius, F. *J. Am. Chem. Soc.* **2017**, *139*, 2585.



3,8-dimethylisochroman-1-one (2d)



Colorless solid; H₂ (50 bar), 15 °C, 16 h, 78% yield, 99:1 e.r. $[\alpha]_D^{22} =$ +160.7 (c = 0.37 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*hexane/2-propanol = 80/20, flow rate = 1 mL/min, λ = 254 nm, retention time: 5.9 min (major), 7.0 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.36 (t, J = 7.6 Hz, 1H), 7.19 (d, J = 7.7 Hz, 1H), 7.05 (d, J = 7.5 Hz, 1H), 4.62 - 4.51 (m,1H), 2.99 - 2.83 (m, 2H), 2.67 (s, 3H), 1.49 (d, J = 6.3 Hz, 3H). Known compound see: Li, W.; Wiesenfeldt. M. P.; Glorius, F. J. Am. Chem. Soc. 2017, 139, 2585.



3-methyl-7-phenylisochroman-1-one (2e)



mL/min, $\lambda = 230$ nm, retention time: 9.3 min (major), 15.9 min (minor). ¹H NMR (500 MHz, CDCl₃) δ 8.34 (d, J = 1.8 Hz, 1H), 7.76 (dd, J = 7.9, 2.0 Hz, 1H), 7.62 (dd, J = 5.2, 3.3 Hz, 2H), 7.46 (dd, J = 10.4, 4.8 Hz, 2H), 7.38 (dd, J = 10.5, 4.3 Hz, 1H), 7.31 (d, J = 7.9 Hz, 1H), 4.76 – 4.62 (m, 1H), 3.04 – 2.89 (m, 2H), 1.55 (d, J = 6.3 Hz, 3H).



Known compound, see: Li, W.; Wiesenfeldt. M. P.; Glorius, F. J. Am. Chem. Soc. 2017, 139, 2585.

7-methoxy-3-methylisochroman-1-one (2f)



1 mL/min, $\lambda = 230$ nm, retention time: 8.4 min (major), 12.5 min (minor). ¹H NMR (500 MHz, CDCl₃) δ 7.57 (d, J = 2.7 Hz, 1H), 7.13 (dd, J = 8.4, 0.4 Hz, 1H), 7.08 (dd, J = 8.4, 2.7 Hz, 1H), 4.65 (dqd, J = 9.8, 6.3, 5.0 Hz, 1H), 3.83 (s, 3H), 2.94 – 2.76 (m,

2H), 1.50 (d, *J* = 6.3 Hz, 3H). Known compound, see: Li, W.; Wiesenfeldt. M. P.; Glorius, F. *J. Am. Chem. Soc.* **2017**, *139*, 2585.



3,6,7-trimethylisochroman-1-one (2g)



Colorless solid; H₂ (50 bar), 25 °C, 21 h, 80% yield, 96:4 e.r. $[\alpha]_D^{22}$ = +119.9 (c = 1.25 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 80/20, flow rate = 1 mL/min, λ = 254 nm,

retention time: 7.9 min (major), 11.5 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.84 (s, 1H), 6.98 (s, 1H), 4.69 – 4.55 (m, 1H), 2.94 – 2.73 (m, 2H), 2.30 (s, 3H), 2.28 (s,



3H), 1.49 (d, J = 6.3 Hz, 3H). Known compound, see: Li, W.; Wiesenfeldt. M. P.;
Glorius, F. J. Am. Chem. Soc. 2017, 139, 2585.

3-methyl-7-(trifluoromethyl)isochroman-1-one (2h)



Colorless solid; H₂ (50 bar), 15 °C, 16 h, 70% yield, 99.7:0.3 e.r. $[\alpha]_D^{22} = +122.0$ (c = 1.19 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 80/20, flow rate = 1

mL/min, $\lambda = 230$ nm, retention time: 6.3 min (major), 9.4 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 8.37 (d, J = 0.7 Hz, 1H), 7.77 (dd, J = 8.0, 1.4 Hz, 1H), 7.39 (d, J = 8.0 Hz, 1H), 4.80 – 4.62 (m, 1H), 3.02 (d, J = 7.2 Hz, 2H), 1.54 (d, J = 6.3 Hz, 3H). Known



compound, see: Li, W.; Wiesenfeldt. M. P.; Glorius, F. J. Am. Chem. Soc. 2017, 139, 2585.

7-fluoro-3-methylisochroman-1-one (2i)



Colorless solid; H₂ (50 bar), 15 °C, 21 h, 73% yield, 99:1 e.r. $[\alpha]_D^{22}$ = +102.5 (c = 1.10 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 80/20, flow rate = 1 mL/min, λ = 230 nm,

retention time: 7.6 min (major), 9.8 min (minor). ¹**H NMR** (400 MHz, CDCl₃) δ 7.80 - 7.69 (m, 1H), 7.25 - 7.19 (m, 2H), 4.76 - 4.51 (m, 1H), 2.91 (d, *J* = 6.5 Hz, 2H), 1.52 (d, *J* = 6.3 Hz, 3H). Known compound, see: Li, W.; Wiesenfeldt. M. P.; Glorius, F. *J. Am. Chem. Soc.* **2017**, *139*, 2585.



7-chloro-3-methylisochroman-1-one (2j)

Clorless solid; H₂ (50 bar), 15 °C, 16 h, 80% yield, 96:4 e.r. $[\alpha]_D^{22}$ = +119.0 (c = 0.95 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 80/20, flow rate = 1 mL/min, λ = 254

nm, retention time: 10.9 min (major), 13.0 min (minor). ¹**H NMR** (400 MHz, CDCl₃) δ 8.02 (d, *J* = 8.3 Hz, 1H), 7.40 – 7.32 (m, 1H), 7.27 – 7.18 (m, 1H), 4.67 (dqd, *J* = 10.5, 6.3, 4.2 Hz, 1H), 3.01 – 2.84 (m, 2H), 1.52 (d, *J* = 6.3 Hz, 3H). Known compound, see: Li, W.; Wiesenfeldt. M. P.; Glorius, F. *J. Am. Chem. Soc.* **2017**, *139*, 2585.



7-bromo-3-methylisochroman-1-one (2k)

Br (α) (α) (α)

nm, retention time: 8.8 min (major), 14.8 min (minor). ¹**H NMR** (500 MHz, CDCl₃) δ 8.23 – 8.18 (m, 1H), 7.63 (dd, *J* = 8.1, 2.2 Hz, 1H), 7.17 – 7.10 (m, 1H), 4.76 – 4.52 (m, 1H), 2.95 – 2.85 (m, 2H), 1.51 (d, *J* = 6.3 Hz, 3H). Known compound, see: Li, W.; Wiesenfeldt. M. P.; Glorius, F. *J. Am. Chem. Soc.* **2017**, *139*, 2585.



6-methyl-6,7-dihydro-4H-thieno[3,4-c]pyran-4-one (2l)



Colorless oil; H₂ (50 bar), 15 °C, 45 h, 72% yield, 93:7 e.r. $[\alpha]_D^{22} =$ +33.2 (c = 0.35 in CHCl₃). HPLC DAICEL CHIRALCEL AD-H, *n*-hexane/2-propanol = 95/5, flow rate = 0.8 mL/min, $\lambda = 230$ nm,

retention time: 19.9 min (minor), 21.0 min (major). ¹H NMR (400 MHz, CDCl₃) δ 8.22 (d, J = 2.8 Hz, 1H), 7.05 (dd, J = 1.3, 0.8 Hz, 1H), 4.74 – 4.52 (m, 1H), 3.00 (dd, J = 16.0, 3.1 Hz, 1H), 2.89 – 2.65 (m, 1H), 1.50 (d, J = 6.3 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 162.3, 137.3, 134.0, 128.2, 120.2, 76.4, 32.1, 20.8. ESI-MS: calculated [C₈H₈O₂S + Na]⁺: 191.0137, found: 191.0141.



3-butylisochroman-1-one (2m)



Colorless oil; H₂ (80 bar), 15 °C, 40 h, 73% yield, 93.5:6.5 e.r.. $[\alpha]_D^{22} = +90.2$ (c = 1.20 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 90/10, flow rate =

1 mL/min, $\lambda = 230$ nm, retention time: 9.4 min (major), 10.7 min (minor). ¹H NMR (300 MHz, CDCl₃) δ 8.08 (dd, J = 7.7, 0.9 Hz, 1H), 7.52 (td, J = 7.5, 1.4 Hz, 1H), 7.37 (t, J = 7.6 Hz, 1H), 7.23 (d, J = 7.6 Hz, 1H), 4.59 – 4.36 (m, 1H), 3.05 – 2.78 (m, 2H), 1.88 (dddd, J = 12.6, 10.0, 7.3, 5.2 Hz, 1H), 1.79 – 1.64 (m, 1H), 1.63 – 1.27 (m, 4H), 0.93 (t, J = 7.2 Hz, 3H). Known compound, see: Li, W.; Wiesenfeldt. M. P.; Glorius, F. J. Am. Chem. Soc. **2017**, 139, 2585.



3-cyclopropylisochroman-1-one (2n)

Colorless oil; H₂ (50 bar), 15 °C, 40 h, 78% yield, 98:2 e.r. $[\alpha]_D^{22}$ = +92.8 (c = 1.06 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*hexane/2-propanol = 80/20, flow rate = 1 mL/min, λ = 230 nm, retention time: 9.4 min (major), 13.2 min (minor). ¹H NMR (300 MHz, CDCl₃) δ 8.01 (dd, *J* = 7.7, 0.9 Hz, 1H), 7.46 (td, *J* = 7.5, 1.4 Hz, 1H), 7.31 (t, *J* = 7.6 Hz, 1H), 7.18 (d, *J* = 8.4 Hz, 1H), 3.74 (ddd, *J* = 10.4, 8.7, 3.9 Hz, 1H), 3.02 (qd, *J* = 16.3, 7.2 Hz, 2H), 1.27 – 1.03 (m, 1H), 0.71 – 0.43 (m, 3H), 0.35 – 0.16 (m, 1H). Known compound, see: Li, W.; Wiesenfeldt. M. P.; Glorius, F. *J. Am. Chem. Soc.* **2017**, *139*, 2585.



3-(methoxymethyl)isochroman-1-one (20)



230 nm, retention time: 8.5 min (major), 11.0 min (minor). ¹**H NMR** (300 MHz, CDCl₃) δ 8.08 (dd, J = 7.7, 1.0 Hz, 1H), 7.53 (td, J = 7.5, 1.4 Hz, 1H), 7.38 (t, J = 7.6 Hz, 1H), 7.26 (d, J = 7.6 Hz, 1H), 4.67 (dtd, J = 11.6, 4.9, 3.4 Hz, 1H), 3.68 (qd, J = 10.4, 4.9 Hz, 2H), 3.44 (s, 3H), 3.15 (dd, J = 16.4, 11.6 Hz, 1H), 2.95 (dd, J = 16.4, 3.4 Hz, 1H). Known compound, see: Li, W.; Wiesenfeldt. M. P.; Glorius, F. *J. Am. Chem. Soc.* 2017, *139*, 2585.



b. Ruthenium-catalyzed asymmetric hydrogenation of benzothiophene 1,1dioxides:

To a glass vial, complex C3 (0.0015 mmol, 0.5 mol%), NaOt-Bu (0.02 mmol, 6.7 mol%), benzothiophene 1,1-dioxide 3 (0.3 mmol), and toluene (2.0 mL) were added under an argon atmosphere. The glass vial was placed in a 150 mL stainless steel autoclave under an argon atmosphere. The autoclave was pressurized and depressurized with hydrogen gas five times before the indicated pressure (5 bar or 30 bar) was set. The reaction mixture was stirred at 25 or 0 °C for 24 h. After the autoclave was carefully depressurized, the mixture was directly purified by flash column chromatography on silica gel (pentane/ethylacetate = 20:1 to 4:1) to afford the desired product 4.

2-methyl-2,3-dihydrobenzo[b]thiophene 1,1-dioxide (4a)

Colorless solid; H₂ (5 bar), 25 °C, 24 h, 99% yield, 97:3 e.r. $[\alpha]_D^{22} =$ +18.5 (c = 1.00 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 70/30, flow rate = 1 mL/min, λ = 230 nm, retention

time: 14.4 min (major), 21.6 min (minor). ¹**H NMR** (300 MHz, CDCl₃) δ 7.74 (d, J = 7.7 Hz, 1H), 7.56 (dd, J = 10.7, 4.2 Hz, 1H), 7.45 (t, J = 7.4 Hz, 1H), 7.34 (d, J = 7.6 Hz, 1H), 3.61 – 3.37 (m, 2H), 2.95 (dd, J = 15.5, 7.1 Hz, 1H), 1.52 (d, J = 6.7 Hz, 3H). Known compound, see: Tosatti, P.; Pfaltz, A. *Angew. Chem., Int. Ed.* **2017**, *56*, 4579.



2,5-dimethyl-2,3-dihydrobenzo[b]thiophene 1,1-dioxide (4b)



hexane/2-propanol = 70/30, flow rate = 0.8 mL/min, λ = 230 nm, retention time: 25.7 min (major), 45.74 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.63 (d, *J* = 7.9 Hz, 1H), 7.26 (d, *J* = 7.9 Hz, 1H), 7.15 (s, 1H), 3.53 (dt, *J* = 13.6, 6.8 Hz, 1H), 3.42 (dd, *J* = 16.0, 7.5 Hz, 1H), 2.91 (dd, *J* = 15.9, 7.4 Hz, 1H), 2.43 (s, 3H), 1.52 (dd, *J* = 6.8, 0.9 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 144.3, 136.9, 135.9, 129.7, 127.6, 121.8, 56.9, 33.9, 21.8, 12.5. ESI-MS: calculated [C₁₀H₁₂O₂S + Na]⁺: 219.0450, found: 219.0456.



5-methoxy-2-methyl-2,3-dihydrobenzo[b]thiophene 1,1-dioxide (4c)



nm, retention time: 42.1 min (major), 68.1 min (minor). ¹H NMR (300 MHz, CDCl₃) δ 7.65 (d, J = 8.6 Hz, 1H), 6.95 (dd, J = 8.6, 2.3 Hz, 1H), 6.82 – 6.71 (m, 1H), 3.85 (s, 3H), 3.53 (dt, J = 14.2, 7.1 Hz, 1H), 3.40 (dd, J = 16.0, 7.6 Hz, 1H), 2.90 (dd, J = 16.0, 7.5 Hz, 1H), 1.50 (d, J = 6.8 Hz, 3H). ¹³C NMR (75 MHz, CDCl₃) δ 163.7, 139.1, 130.6, 123.5, 115.4, 111.2, 56.9, 55.8, 34.0, 12.5. ESI-MS: calculated [C₁₀H₁₂O₃S + Na]⁺: 235.0399, found: 235.0395.



2-methyl-5-(trifluoromethyl)-2,3-dihydrobenzo[b]thiophene 1,1-dioxide (4d)



Colorless oil; H₂ (5 bar), 25 °C, 24 h, 86% yield, 99:1 e.r. $[\alpha]_D^{22} =$ +13.4 (c = 0.75 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 70/30, flow rate = 0.8 mL/min, $\lambda = 230$ nm, retention time: 12.9 min (major), 19.2 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.88 (d, J = 8.0 Hz, 1H), 7.73 (d, J = 8.1 Hz, 1H), 7.63 (s, 1H), 3.69 – 3.40 (m, 2H), 3.02 (dd, J = 15.9, 7.2 Hz, 1H), 1.55 (dd, J = 6.7, 0.9 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 142.1, 137.6, 135.4 (q, J = 32.8 Hz), 126.2 (q, J = 3.6 Hz), 124.6 (dd, J = 7.7, 3.9 Hz), 123.0, 121.9, 57.0, 34.0, 12.4. ¹⁹F NMR (282 MHz, CDCl₃) δ –62.94. ESI-MS: calculated [C₁₀H₉F₃O₂S + Na]⁺: 273.0168, found: 273.0165.



5-fluoro-2-methyl-2,3-dihydrobenzo[b]thiophene 1,1-dioxide (4e)



Colorless oil; H₂ (5 bar), 25 °C, 24 h, 88% yield, 97.5:2.5 e.r. $[\alpha]_D^{22}$ = +18.0 (c = 1.0 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 70/30, flow rate = 0.8 mL/min, λ = 230 nm,

retention time: 21.5 min (major), 33.6 min (minor). ¹H NMR (400 MHz, CDCl₃) δ 7.74 (dd, J = 8.5, 5.0 Hz, 1H), 7.15 (td, J = 8.5, 2.1 Hz, 1H), 7.03 (dd, J = 8.3, 0.9 Hz, 1H),3.65 - 3.51 (m, 1H), 3.44 (dd, J = 16.2, 7.6 Hz, 1H), 2.94 (dd, J = 16.3, 7.6 Hz, 1H), 1.52 (d, J = 6.9 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 165.7 (d, J = 254.7 Hz), 139.9 (d, J = 9.6 Hz), 134.7 (d, J = 2.8 Hz), 124.5 (d, J = 10.0 Hz), 116.8 (d, J = 23.8 Hz),114.2 (d, J = 23.1 Hz), 57.1, 33.9 (d, J = 1.7 Hz, 14H), 12.5. ¹⁹F NMR (282 MHz, CDCl₃) δ -104.45. **ESI-MS**: calculated [C₉H₉FO₂S + Na]⁺: 223.0199, found: 223.0208.



5-chloro-2-methyl-2,3-dihydrobenzo[b]thiophene 1,1-dioxide (4f)

Colorless solid; H₂ (5 bar), 25 °C, 24 h, 92% yield, 98:2 e.r. $[\alpha]_D^{22}$ CI = +13.0 (c = 1.0 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 70/30, flow rate = 0.8 mL/min, λ = 230 nm,

retention time: 21.6 min (major), 35.1 min (minor). ¹H NMR (400 MHz, CDCl₃) & 7.67 S56

(d, J = 8.2 Hz, 1H), 7.42 (d, J = 8.2 Hz, 1H), 7.34 (s, 1H), 3.68 – 3.50 (m, 1H), 3.43 (dd, J = 16.2, 7.5 Hz, 1H), 2.93 (dd, J = 16.2, 7.6 Hz, 1H), 1.51 (dd, J = 6.8, 0.8 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 139.7, 138.6, 137.2, 129.4, 127.4, 123.4, 56.9, 33.7, 12.4. ESI-MS: calculated [C₉H₉ClO₂S + Na]⁺: 238.9904, found: 238.9913.



5-bromo-2-methyl-2,3-dihydrobenzo[b]thiophene 1,1-dioxide (4g)

Br Colorless solid; H₂ (5 bar), 25 °C, 24 h, 93% yield, 98.5:1.5 e.r. $[\alpha]_D^{22} = +9.3$ (c = 1.0 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 70/30, flow rate = 0.8 mL/min, λ =

230 nm, retention time: 23.3 min (major), 37.7 min (minor). ¹H NMR (400 MHz,

CDCl₃) δ 7.67 – 7.56 (m, 2H), 7.51 (s, 1H), 3.62 – 3.50 (m, 1H), 3.43 (dd, J = 16.1, 7.5 Hz, 1H), 2.93 (dd, J = 16.1, 7.5 Hz, 1H), 1.51 (dd, J = 6.8, 1.0 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 138.6, 137.6, 132.2, 130.3, 128.0, 123.4, 56.8, 33.6, 12.3. ESI-MS: calculated [C₉H₉BrO₂S + Na]⁺: 282.9399, 284.9385, found: 282.9404, 284.9385.



2-propyl-2,3-dihydrobenzo[b]thiophene 1,1-dioxide (4h)



Colorless solid; H₂ (30 bar), 0 °C, 24 h, 96% yield, 94:6 e.r. $[\alpha]_D^{22}$ = +17.7 (c = 1.0 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 70/30, flow rate = 1.0 mL/min, λ = 230 nm,

retention time: 12.9 min (major), 20.8 min (minor). ¹**H NMR** (400 MHz, CDCl₃) δ 7.73 (d, *J* = 7.7 Hz, 1H), 7.54 (td, *J* = 7.5, 1.1 Hz, 1H), 7.44 (t, *J* = 7.5 Hz, 1H), 7.33 (d, *J* =

7.7 Hz, 1H), 3.43 (tt, J = 21.1, 7.6 Hz, 2H), 3.05 – 2.84 (m, 1H), 2.20 – 1.99 (m, 1H), 1.82 – 1.48 (m, 3H), 1.02 (t, J = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 139.2, 136.5, 133.3, 128.7, 127.0, 121.8, 61.3, 32.5, 29.6, 20.3, 13.9. ESI-MS: calculated [C₁₁H₁₄O₂S + Na]⁺: 233.0607, found: 233.0611.



2-(methoxymethyl)-2,3-dihydrobenzo[b]thiophene 1,1-dioxide (4i)



Colorless solid; H₂ (30 bar), 0 °C, 24 h, 90% yield, 96.5:3.5 e.r. $[\alpha]_D^{22} = +14.1$ (c = 1.0 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 70/30, flow rate = 0.8 mL/min, λ =

210 nm, retention time: 31.2 min (major), 39.7 min (minor). ¹H NMR (300 MHz, CDCl₃) δ 7.74 (d, J = 7.7 Hz, 1H), 7.56 (t, J = 7.5 Hz, 1H), 7.45 (t, J = 7.5 Hz, 1H),

7.35 (d, J = 7.7 Hz, 1H), 3.97 (ddd, J = 10.7, 6.7, 3.6 Hz, 1H), 3.82 – 3.64 (m, 2H), 3.51 – 3.34 (m, 4H), 3.11 (dd, J = 16.4, 6.8 Hz, 1H). ¹³C NMR (75 MHz, CDCl₃) δ 139.1, 136.1, 133.6, 128.9, 127.3, 121.9, 69.3, 60.5, 59.5, 29.5. ESI-MS: calculated [C₁₀H₁₂O₃S + Na]⁺: 235.0399, found: 235.0407.



2-(4-chlorobutyl)-2,3-dihydrobenzo[b]thiophene 1,1-dioxide (4j)



Colorless oil; H₂ (30 bar), 0 °C, 24 h, 98% yield, 94:6 e.r. $[\alpha]_D^{22} = +9.6$ (c = 1.0 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 70/30, flow rate

= 1.0 mL/min, λ = 210 nm, retention time: 23.7 min (major), 36.3 min (minor). ¹H

NMR (300 MHz, CDCl₃) δ 7.74 (d, J = 7.7 Hz, 1H), 7.56 (t, J = 7.5 Hz, 1H), 7.45 (t, J = 7.5 Hz, 1H), 7.34 (d, J = 7.6 Hz, 1H), 3.58 (t, J = 6.4 Hz, 2H), 3.52 – 3.34 (m, 2H), 3.11 – 2.89 (m, 1H), 2.26 – 2.03 (m, 1H), 2.03 – 1.64 (m, 5H). ¹³C NMR (75 MHz, CDCl₃) δ 139.1, 136.4, 133.5, 128.9, 127.2, 122.0, 61.4, 44.6, 32.5, 32.3, 27.1, 24.4. **ESI-MS**: calculated [C₁₂H₁₅ClO₂S + Na]⁺: 281.0373, found: 281.0383.



c. Enantioselective hydrogenation of ketones catalyzed by Ru(II)-NHC-diamine complexes:

For an easier handling, a stock solution of complex of C4 (0.006 mmol, 4.7 mg) in THF (4.0 mL) was prepared. To a glass vial, 200 μ L of the C4 solution was added, and the solvent was removed under vacuum. To this vial, NaO*t*-Bu (0.02 mmol, 6.7 mol%),

ketone **5** (1.0 mmol), and *i*-PrOH (1.0 mL) were added under an argon atmosphere. The glass vial was placed in a 150 mL stainless steel autoclave under an argon atmosphere. The autoclave was pressurized and depressurized with hydrogen gas five times before the indicated pressure (5 bar or 10 bar) was set. The reaction mixture was stirred at 22 $^{\circ}$ C for 24 h. After the autoclave was carefully depressurized, the mixture was directly purified by flash column chromatography on silica gel (pentane/ethylacetate = 20:1 to 4:1) to afford the desired product **6**. All the obtained products **6** have been reported previously.

1-phenylethan-1-ol (6a)

OH Colorless oil; H₂ (5 bar), 22 °C, 24 h, 99% yield, 93:7 e.r. $[\alpha]_D^{22} = +48.0$ (c = 0.85 in CHCl₃). HPLC DAICEL CHIRALCEL OJ-H, *n*-hexane/2propanol = 98/2, flow rate = 0.8 mL/min, $\lambda = 254$ nm, retention time: 25.9 min (minor), 28.7 min (major). ¹H NMR (300 MHz, CDCl₃) δ 7.45 – 7.25 (m, 5H), 4.92 (q, J = 6.4 Hz, 1H), 2.01 (s, 1H), 1.53 (d, J = 6.5 Hz, 3H).



Peak	RetTime	Туре	Width	Area	Height	Area
#	[min]		[min]	[mAU*s]	[mAU]	8
1	25.855	MM	0.4778	101.69521	3.54747	7.1326
2	28.742	MM	0.6680	1324.07715	33.03671	92.8674

1-(benzo[d][1,3]dioxol-5-yl)ethan-1-ol (6b)

OH Colorless oil; H₂ (10 bar), 22 °C, 24 h, 99% yield, 92:8 e.r. $[\alpha]_D^{22} =$ +40.3 (c = 0.90 in CHCl₃). HPLC DAICEL CHIRALCEL AD-H, *n*-hexane/2-propanol = 95/5, flow rate = 0.8 mL/min, λ = 254 nm,

retention time: 20.8 min (major), 23.3 min (minor). ¹**H NMR** (300 MHz, CDCl₃) δ 6.88 (d, *J* = 1.1 Hz, 1H), 6.79 (dt, *J* = 14.1, 4.6 Hz, 2H), 5.94 (s, 2H), 4.81 (q, *J* = 6.4 Hz, 1H), 1.92 (s, 1H), 1.45 (d, *J* = 6.4 Hz, 3H).



1-(thiophen-3-yl)ethan-1-ol (6c)

HO
Colorless oil; H₂ (10 bar), 22 °C, 24 h, 99% yield, 91:9 e.r.
$$[\alpha]_D^{22} = +25.5$$

(c = 1.40 in CHCl₃). HPLC DAICEL CHIRALCEL OJ-H, *n*-hexane/2-
propanol = 95/5, flow rate = 1.0 mL/min, λ = 230 nm, retention time: 12.2

min (minor), 14.5 min (major). ¹**H NMR** (300 MHz, CDCl₃) δ 7.30 (dd, *J* = 5.0, 3.0 Hz, 1H), 7.24 – 7.17 (m, 1H), 7.10 (dd, *J* = 5.0, 1.3 Hz, 1H), 4.97 (q, *J* = 6.4 Hz, 1H), 1.90 (s, 1H), 1.53 (d, *J* = 6.5 Hz, 3H).



1,2,3,4-tetrahydronaphthalen-1-ol (6d)



min (minor), 12.9 min (major). ¹**H NMR** (300 MHz, CDCl₃) δ 7.49 – 7.35 (m, 1H), 7.25 – 7.16 (m, 2H), 7.11 (dd, *J* = 5.5, 3.5 Hz, 1H), 4.78 (s, 1H), 2.94 – 2.61 (m, 2H), 2.08 – 1.71 (m, 5H).



2 12.861 BB 0.2935 1.97897e4 1014.93079 96.6781

6-methoxy-1,2,3,4-tetrahydronaphthalen-1-ol (6e)

OH Colorless oil; H₂ (10 bar), 22 °C, 24 h, 96% yield, 93:7 e.r. $[\alpha]_D^{22} =$ -20.9 (c = 1.0 in CHCl₃). HPLC DAICEL CHIRALCEL OD-H, *n*-hexane/2-propanol = 95/5, flow rate = 0.8 mL/min, λ = 230 nm, retention time: 14.3 min (minor), 15.3 min (major). ¹H NMR (400 MHz, CDCl₃) δ 7.32

(d, *J* = 8.5 Hz, 1H), 6.76 (dd, *J* = 8.5, 2.4 Hz, 1H), 6.62 (d, *J* = 1.7 Hz, 1H), 4.72 (d, *J* = 4.8 Hz, 1H), 3.78 (s, 3H), 2.84 – 2.63 (m, 2H), 2.06 (s, 1H), 2.01 – 1.84 (m, 3H), 1.79 – 1.71 (m, 1H).



5,7-dimethyl-1,2,3,4-tetrahydronaphthalen-1-ol (6f)



Colorless oil; H₂ (10 bar), 22 °C, 24 h, 96% yield, 91:9 e.r. $[\alpha]_D^{22} = -$ 28.4 (c = 1.0 in CHCl₃). HPLC DAICEL CHIRALCEL OD-H, *n*-hexane/2-propanol = 95/5, flow rate = 1.0 mL/min, $\lambda = 230$ nm,

retention time: 7.0 min (minor), 8.4 min (major). ¹**H NMR** (300 MHz, CDCl₃) δ 7.13 (s, 1H), 6.94 (s, 1H), 4.75 (s, 1H), 2.76 – 2.59 (m, 1H), 2.59 – 2.42 (m, 1H), 2.31 (s, 3H), 2.21 (s, 3H), 2.07 – 1.68 (m, 5H).



7-fluoro-1,2,3,4-tetrahydronaphthalen-1-ol (6g)



Colorless oil; H₂ (10 bar), 22 °C, 24 h, 99% yield, 96:4 e.r. $[\alpha]_D^{22} = -$ 36.8 (c = 1.0 in CHCl₃). HPLC DAICEL CHIRALCEL AD-H, *n*-hexane/2-propanol = 95/5, flow rate = 1.0 mL/min, $\lambda = 254$ nm,

retention time: 9.8 min (minor), 10.9 min (major). ¹**H NMR** (300 MHz, CDCl₃) δ 7.15 (dd, *J* = 9.6, 2.7 Hz, 1H), 7.05 (dd, *J* = 8.4, 5.8 Hz, 1H), 6.89 (td, *J* = 8.4, 2.7 Hz, 1H), 4.74 (s, 1H), 2.87 – 2.54 (m, 2H), 2.13 – 1.66 (m, 5H).



7-bromo-1,2,3,4-tetrahydronaphthalen-1-ol (6h)



Colorless oil; H₂ (10 bar), 22 °C, 24 h, 98% yield, 95:5 e.r. $[\alpha]_D^{22} = -$ 60.2 (c = 1.0 in CHCl₃). HPLC DAICEL CHIRALCEL AD-H, *n*hexane/2-propanol = 95/5, flow rate = 1.0 mL/min, $\lambda = 230$ nm,

retention time: 9.7 min (minor), 12.1 min (major). ¹**H NMR** (400 MHz, CDCl₃) δ 7.59 (s, 1H), 7.29 (d, *J* = 8.2 Hz, 1H), 6.97 (d, *J* = 8.2 Hz, 1H), 4.73 (s, 1H), 2.93 – 2.58 (m, 2H), 2.05 – 1.72 (m, 5H).



#	[min]		[min]	[mAU*s]	[mAU]	00
1	9.690	BB	0.2246	1503.73853	100.50675	5.0088
2	12.094	BB	0.2987	2.85183e4	1454.51062	94.9912

chroman-4-ol (6i)



Colorless oil; H₂ (10 bar), 22 °C, 24 h, 95% yield, 96.5:3.5 e.r. $[\alpha]_D^{22} = -$ 67.5 (c = 1.0 in CHCl₃). HPLC DAICEL CHIRALCEL AS-H, *n*-hexane/2-propanol = 95/5, flow rate = 1.0 mL/min, $\lambda = 230$ nm, retention

time: 9.3 min (minor), 11.7 min (major). ¹**H NMR** (300 MHz, CDCl₃) δ 7.30 (dd, *J* = 7.6, 1.6 Hz, 1H), 7.21 (ddd, *J* = 8.8, 7.5, 1.7 Hz, 1H), 6.92 (td, *J* = 7.5, 1.2 Hz, 1H), 6.84 (dd, *J* = 8.2, 1.0 Hz, 1H), 4.76 (d, *J* = 3.8 Hz, 1H), 4.31 – 4.16 (m, 2H), 2.19 – 1.94 (m, 3H).



6,7,8,9-tetrahydro-5H-benzo[7]annulen-5-ol (6j)

HQ Colorless oil; H₂ (10 bar), 22 °C, 24 h, 99% yield, 95.5:4.5 e.r. $[\alpha]_D^{22} =$ -67.5 (c = 1.0 in CHCl₃). HPLC DAICEL CHIRALCEL AD-H, *n*-hexane/2-propanol = 95/5, flow rate = 0.8 mL/min, λ = 254 nm, retention time: 12.1 min (major), 13.5 min (minor). ¹H NMR (300 MHz, CDCl₃) δ 7.44 (d, J =

7.2 Hz, 1H), 7.25 - 7.05 (m, 3H), 4.93 (d, J = 7.7 Hz, 1H), 2.93 (dd, J = 14.0, 8.3 Hz, 1H), 2.73 (dd, J = 17.6, 7.1 Hz, 1H), 2.18 - 1.89 (m, 3H), 1.81 (ddd, J = 9.7, 7.4, 4.7 Hz, 3H), 1.58 - 1.37 (m, 1H).



(E) Stoichiometric reactions for mechanistic investigations




C3 (0.1 mmol, 73mg) and toluene- d_8 (0.7 mL) were added to a Young NMR tube in the glove box, and ¹H NMR was measured. NaOt-Bu (0.1 mmol, 1 equiv.) was then added to the mixture in the glove box. The dehydrochlorination reaction immediately occured visuliazed by a color change from yellow to dark red. After 5 minutes, the solvent and the byproduct *t*-BuOH were remove under vaccum. Toluene- d_8 (0.7 mL) was again added to the mixture, and NMR spectra were measured (step1). After quickly removing the gas of the NMR tube in vacumm, H_2 was added to reach 1 atm. After shaking the NMR tube several times, the reaction completed within 5 min. ¹H NMR of step2 was measured immediately at 26 °C. The full NMR spectra of C8 (¹H, ¹³C, gCOSY, gHSQC, and gHMBC) were measured at -30 °C because C8 is not stable in solution at room temperature. After releasing the H₂ gas from the NMR tube, substrate 3a (0.2 mmol, 2 equiv.) was added to the solution of C8. The reaction was completed within 5 mins, and ¹H NMR was measured (step3). Finally, 1.0 equiv. of HCl solution (4M in dioxane) was added to the reaction, and ¹H NMR was measured (step4). The product 4a and C3 were recovered by column chromatography on silica gel (pentane/ethylacetate = 20:1, 10:1, 4:1, 2:1). The e.r. value of **4a** was also measured.



¹H NMR (600 MHz, 26 °C, Toluene- d_8 , connectivities were confirmed by gCOSY, gHSQC, gHMBC, 1D-NOESY and 1D-TOCSY experiments) δ 7.78 (d, J = 8.0 Hz, 1H, HC18), 7.37 (t, J = 6.9 Hz, 2H, HC32, HC25), 7.27 – 7.22 (m, 3H, HC33, HC21, HC23), 7.21 – 7.13 (m, 3H, HC34, HC17, HC27),

7.13 – 7.08 (m, 2H, HC22, HC26), 7.06 (d, J = 7.1 Hz, 1H, HC35), 6.99 – 6.90 (m, 3H, *H*C15, *H*C16, *H*C28), 6.86 (d, *J* = 7.5 Hz, 2H, *H*C37, *H*C41), 6.79 (t, *J* = 7.4 Hz, 2H, *H*C38, *H*C40), 6.72 (q, *J* = 6.7 Hz, 2H, *H*C39, *H*C6), 6.60 (dd, *J* = 9.0, 6.0 Hz, 1H, HC12), 6.17 (d, J = 8.9 Hz, 1H, HC13), 4.85 (q, J = 5.6 Hz, 1H, HC4), 4.20 (s, 1H, HN4), 3.71 (d, J = 6.0 Hz, 1H, HC11), 3.62 (s, 1H, HC8), 3.54 – 3.45 (m, 2H, HC9, H_2 C3), 3.35 (dt, J = 13.0, 9.5 Hz, 1H, H_2 C3), 3.26 (t, J = 8.9 Hz, 1H, H_2 C2), 3.06 (q, J= 10.2 Hz, 1H, H_2 C2), 1.31 (d, J = 7.1 Hz, 3H, H_3 C7), 1.26 (d, J = 6.2 Hz, 3H, H_3 C5), 1.03 (d, J = 8.6 Hz, 1H, H_2 N3), -0.03 (d, J = 8.1 Hz, 1H, H_2 N3). ¹³C NMR (151 MHz, 26 °C, Toluene- d_8 , connectivities were confirmed by gHSQC and gHMBC experiments) & 228.2 (C1), 165.8 (C31), 146.4 (C30), 145.5 (C36), 143.7 (C20), 140.4 (C12), 140.4 (C32), 140.1 (C19), 134.4 (C24), 132.3 (C14), 129.9 (C29), 129.0 (C25), 128.0 (C38, C40), 127.6 (C23), 127.1 (C15), 126.7 (C37, C41), 126.3 (C39), 126.2 (C27), 126.0 (C17), 125.5 (C22), 125.5 (C26), 125.5 (C33), 123.6 (C28), 123.1 (C16), 122.5 (C18), 121.6 (C21), 121.3 (C35, C34), 116.7 (C13), 85.5 (C10), 71.9 (C9), 71.1 (C8), 65.3 (C11), 57.8 (C4), 53.4 (C6), 47.5 (C3), 46.8 (C2), 20.9 (C7), 20.9 (C5). ESI-**MS:** calculated $[C_{41}H_{40}N_4Ru + H^+]^+$: 691.2369, found: 691.2378.





−0.02
 −0.04



gHSQC:



gHMBC:





¹H NMR (600 MHz, -30 °C, Toluene- d_{8} , connectivities were confirmed by gCOSY, gHSQC, gHMBC, 1D-NOESY and 1D-TOCSY experiments) δ 8.54 (d, J = 6.2 Hz, 1H, HC32), 8.39 (d, J = 7.9 Hz, 1H, HC18), 7.58 (t, J = 7.0 Hz, 1H, HC33), 7.43 (t, J= 7.3 Hz, 1H, HC34), 7.39 – 7.33 (m, 2H, HC25,

HC17), 7.33 – 7.28 (m, 2H, HC21, HC35), 7.28 – 7.24 (m, 1H, HC27), 7.24 – 7.20 (m, 1H, HC23), 7.20 - 7.00 (m, 5H, HC37, HC41, HC22, HC26, HC28), 6.97 - 6.89 (m, 2H, HC16, HC15), 6.88 – 6.79 (m, 1H, HC6), 6.79 – 6.72 (m, 1H, HC12), 6.69 – 6.56 (m, 3H, HC38, HC40, HC39), 6.05 (d, J = 9.1 Hz, 1H, HC13), 4.87 (q, J = 6.4 Hz, 1H, *H*C4), 3.31 (d, J = 5.7 Hz, 1H, *H*C11), 3.26 – 2.98 (m, 4H, *H*₂C3, *H*₂C2, *H*C8), 2.90 (t, *J* = 9.8 Hz, 1H, *H*C2), 2.81 (t, *J* = 9.5 Hz, 1H, *H*C9), 2.62 (d, *J* = 10.2 Hz, 1H, *H*₂N4), 2.26 (d, J = 11.1 Hz, 1H, H_2 N4), 1.42 (d, J = 6.9 Hz, 3H, H_3 C7), 1.25 (d, J = 6.4 Hz, 3H, H_3 C5), 0.98 (d, J = 9.3 Hz, 1H, H_2 N3), 0.56 (d, J = 9.3 Hz, 1H, H_2 N3), -2.08 (s, 1H, HRu). ¹³C NMR (151 MHz, -30 °C, Toluene- d_8 connectivities were confirmed by gHSQC and gHMBC experiments) & 231.0 (C1), 190.6 (C31), 148.2 (C30), 147.0 (C19), 142.0 (C20), 141.2 (C12), 140.8 (C32), 139.7 (C36), 134.0 (C24), 131.8 (C14), 130.0 (C30), 129.4 (C25), 128.6 (C38, C40), 128.2 (C28), 127.6 (C23), 127.6 (C39), 127.0 (C15, C27), 125.9 (C17), 125.8 (C18), 125.7 (C22), 125.7 (C26), 125.4 (C37, C41), 125.0 (C33), 122.8 (C16), 122.7 (C35, C21), 122.0 (C34), 117.4 (C13), 68.3 (C10), 66.0 (C8), 61.8 (C4), 61.0 (C9), 53.6 (C6), 50.6 (C11), 46.7 (C2), 46.41 (C3), 20.8 (C7), 19.9 (C5). ESI-MS: calculated $[C_{41}H_{42}N_4Ru - H^-]^+$: 691.2369, found: 691.2377.



gCOSY:



gHSQC:



gHMBC:



(F) Quantum chemical investigation of the reaction mechanism

1. Computational Details

All calculations for this work have been performed with the Turbomole v7.2.1 program package⁶ using DFT with the PBE0 hybrid functional.⁷ In addition, the D3 dispersion correction⁸ with Becke-Johnson damping⁹ is applied to ensure a robust description of dispersive interactions. The solvent effects of toluene have been taken into account using the COSMO implicit solvent model.¹⁰ For the numerical integration, the Turbomole m4 grid was used. SCF convergence was assumed after energy changes of less than 1E-7 a.u., while geometry convergence thresholds were set to 1E-6 a.u. for energy changes and 1E-3 a.u. for the norm of the SCF-energy gradient. Frequency analyses have been performed for all optimized structures to confirm minima and transition states and to calculate Gibbs free energies. For the latter, a correction for small vibrational frequencies¹¹ was applied. For geometry optimizations, transition state search and frequency analysis, the def2-SVP basis set was employed.¹²

2. Validation: Comparison of Structures

To confirm that the chosen method is suitable to describe the given systems, the crystal structure of the catalyst is geometrically optimized and the obtained bond lengths and angles are compared to the experimentally determined values. A selection of the obtained results is given in Table S1 for the bond lengths and Table S2 for the bond angles. The data show that both the angles and the bond lengths are in overall good agreement.





In Figure S4 the results are visualized by overlaying the computationally relaxed structure and the experimental crystal structure. It can be seen that they are in good general agreement. The only exception is the phenyl group, which is not coordinated to Ru and thus can rotate without significant barrier. In conclusion it can be assumed that the chosen method is sufficient to describe the systems investigated.

Bond Length (Å)				
	Crystal Structure	DFT (PBE0/def2-SVP)		
(Ru)-(Cl)	2.53	2.51		
(Ru)-(N:2)	2.16	2.16		
(N:2)-(H)	0.92	1.03		
(Ru)-(C:1)	1.94	1.95		
(Ru)-(C:2)	2.21	2.16		
(C:1)- (N:1)	1.38	1.36		

Table S1. Comparison of bond lengths between the crystal structure and the calculated geometry.

Table S2. Comparison of bond angles between the crystal structure and the calculated geometry.

Bond Angle (deg)			
	Crystal Structure	DFT (PBE0/def2-SVP)	
(N:2)-(Ru)-(Cl)	78.05	76.82	
(H)-(N:2)-(Ru)	98.21	96.43	
(N:2)-(Ru)-(C:1)	97.77	101.48	
(Ru)-(C:1)-(N:1)	135.10	135.51	

3. Conceivable Mechanisms

For the adsorption of the substrate to the catalyst and the transfer of the hydride and the proton to the substrate several reaction pathways and configurations have to be considered. Fortunately, the stoichiometric NMR-experiment conducted allows to identify one of the amino-groups as the proton donor, thus restricting the number of possible mechanisms. In the first set of the remaining possible reaction pathways (Scheme S1) the hydrogen atom bonded to the nitrogen of the catalyst forms a hydrogen-bond to the oxygen on the substrate, and the hydride attacks in β -position of the sulfone group. Second, the proton is transferred to the α -position relative to the same sulfone group. It is conceivable that this pathway can proceed in three different ways: First, a concerted mechanism is possible (1a), where proton and hydride are transferred

at the same time. Then, a stepwise mechanism is possible where the hydride is transferred first, followed by a direct hydrogen transfer (1b). The third possibility (1c) for this mechanism to happen is that after the hydride attacked in β -position (as in 1b) the hydrogen is transferred to the oxygen it was hydrogen-bonded to. The final product is then generated through tautomerization.



Scheme S1. Proposed mechanisms for the hydrogenation in β -position.



Scheme S2. Proposed mechanisms for the hydrogenation in α -position.

The second set of possible reaction pathways (Scheme S2) is characterized by the transfer of the hydride to the α -position of the sulfone group on the substrate. As for the first pathway, three different mechanisms are conceivable: A concerted mechanism (2a), the formation of a hydrogen bond with subsequent transfer of the hydrogen to the β -position (2b) and the transfer of the hydrogen to the oxygen of the substrate followed by tautomerization to yield the final product (2c). For all possible pathways both the attack to the Re- and to the Si-face of the substrate are explored and compared.

4. Results

4.1. Optimized Structures

All reaction pathways discussed in the following sections have some common structures, whose optimized geometries are presented in this section. First, the general structure of the catalyst is shown, both for the hydrogenated and the dehydrogenated from. Then the substrate and one enantiomer of the product are presented, followed by the precatalysts for both enantiomers. Finally, the product associated to the dehydrogenated catalyst is given for both enantiomers.

The Hydrogenated Catalyst



Figure S5. Front view (left) and top view (right) of the structure of the hydrogenated catalyst optimized using PBE0/def2-SVP.

The Dehydrogenated Catalyst



Figure S6. Front view (left) and top view (right) of the structure of the dehydrogenated catalyst optimized using PBE0/def2-SVP.

Starting Material / Substrate



Figure S7. Front view (left) and top view (right) of the structure of the substrate to be hydrogenated in the reaction optimized using PBE0/def2-SVP.

Product [(S)-Enantiomer]



Figure S8. Front view (left) and top view (right) of the structure of the product of the hydrogenation [(S)-enantiomer] optimized using PBE0/def2-SVP.



Figure S9. Front view (left) and top view (right) of the structure of the precatalyst formed leading to (R)-product optimized using PBE0/def2-SVP.

Precatalyst [(R)-Product]

Precatalyst [(S)-Product]



Figure S10. Front view (left) and top view (right) of the structure of the precatalyst formed leading to (*S*)-product optimized using PBE0/def2-SVP.

Product-Catalyst Complex [(R)-Product]



Figure S11. Front view (left) and top view (right) of the structure of the complex between the newly formed product and the dehydrogenated catalyst leading to the (R)-product optimized using PBE0/def2-SVP.

Product-Catalyst Complex [(S)-Product]



Figure S12. Front view (left) and top view (right) of the structure of the complex between the newly formed product and the dehydrogenated catalyst leading to the (*S*)-product optimized using PBE0/def2-SVP.

4.2. Proton Transfer to the Sulfone Group (Pathways 1c, 2a and 2c)

The reaction pathways 1c, 2a and 2c all have in common that the proton is transferred to the oxygen of the sulfone group and then the final product is formed *via* tautomerization. However, during the exploration of these mechanisms it turned out, that intermediates with the proton transferred to the sulfone group are thermodynamically unstable. Since it was not possible to optimize minimum structures for these intermediates, the general thermodynamics of the reaction was investigated by calculating the Gibbs free energy of the optimized isolated molecules (as shown in Scheme S3). As a result of this study it turned out that the isolated products of the proton transfer to the sulfone group are destabilized by 66.4 kcal/mol, ruling out all mechanisms including this step (Pathways 1c, 2a and 2c).



Scheme S3. Model reaction to explore the general thermodynamics of a proton transfer to the oxygen of the sulfone group at the substrate.

4.3. Pathway 1a

Reaction pathway 1a corresponds to a concerted mechanism where the hydride is transferred to the β -position of the sulfone group and at the same time the proton is transferred to the α -position (Scheme S4). For this pathway a mechanism leading to the (*S*)-product and one leading to the (*R*)-product have been optimized and the calculated Gibbs free energy for each optimized structure is given in Table S3 and Figure S13. For both pathways, first a pre-complex (PC / C8…3a) is formed from the starting materials (SMs / C8+3a). Then via a single transition state (TS) the products (P…Cat / C7…4a) are formed, which then dissociate (P+Cat / C7+4a). One can see that the precatalyst leading to the (*R*)-product is 2.4 kcal/mol more stable than that leading to the (*S*)-product. However, the transition state is 3.2 kcal/mol more stable for the (*S*)-product and a barrier of 3.4 kcal/mol to generate the (*S*)-product.



Scheme S4. Schematic mechanism for the reaction pathway 1a leading to the (S)-product.

Table S3. Relative Gibbs free energies for pathway 1a leading to the (R)- and (S)-product. All values are given in kcal/mol (PBE0/def2-SVP).



Figure S13. Relative Gibbs free energies for pathway 1a leading to the (R)- and (S)-product (PBE0/def2-SVP).



Transition-State [(R)-Product]

Figure S14. Front view (left) and top view (right) of the transition state in a concerted mechanism with the hydride transferred to β -position leading to the (*R*)-product optimized using PBE0/def2-SVP.



Transition-State [(S)-Product]

Figure S15. Front view (left) and top view (right) of the transition state in a concerted mechanism with the hydride transferred to β -position leading to the (*S*)-product optimized using PBE0/def2-SVP.

4.4. Pathway 1b

Reaction pathway 1b corresponds to a stepwise mechanism in which the hydride is transferred to the β -position relative to the sulfone group. Then, in a second step the proton is transferred to the α -position (see Scheme S5). The Gibbs free energies for all optimized structures are given in Table S4 and Figure S16. For both, the pathways leading to the (*R*)- and (*S*)-product, the starting materials form a pre-catalyst. Afterwards, the hydride is transferred to the β -position *via* a first transition-state (TS1) yielding the intermediate (IN). For the mechanism leading to the (*S*)-product this barrier is 3.4 kcal/mol. However, the barrier for the mechanism leading to the (*R*)-product is 39.4 kcal/mol, thus ruling out his pathway. For that reason only the pathway leading to the (*S*)-product was further investigated. As a next step in this pathway the proton is transferred *via* a second transition state (TS2) to the α -position with a barrier of 2.8 kcal/mol yielding the (*S*)-product.



Scheme S5. Schematic mechanism for the reaction pathway 1b leading to the (S)-product.

Table S4. Relative Gibbs free energies for pathway 1b leading to the (R)- and (S)-product. All values are given in kcal/mol (PBE0/def2-SVP).

	SM	PC	TS1	IN	TS2	P…Cat	P+Cat
(S)- Product	0.0	-0.8	2.9	-9.6	-6.8	-20.9	-21.0
(<i>R</i>)- Product	0.0	-3.2	36.2	-10.3		-21.0	-21.0



Figure S16. Relative Gibbs free energies for pathway 1b leading to the (R)- and (S)-product (PBE0/def2-SVP).

Transition-State [(R)-Product]



Figure S17. Front view (left) and top view (right) of the transition state for the hydride transfer to β -position in a configuration leading to the (*R*)-product optimized using PBE0/def2-SVP.

Transition-State [(S)-Product]



Figure S18. Front view (left) and top view (right) of the transition state for the hydride transfer to β -position in a configuration leading to the (*S*)-product optimized using PBE0/def2-SVP.

Intermediate [(R)-Product]



Figure S19. Front view (left) and top view (right) of the intermediate where the hydride is transferred to β -position in a configuration leading to the (*R*)-product optimized using PBE0/def2-SVP.

Intermediate [(S)-Product]



Figure S20. Front view (left) and top view (right) of the intermediate where the hydride is transferred to β -position in a configuration leading to the (*S*)-product optimized using PBE0/def2-SVP.

4.5. Pathway 2b

Reaction pathway 2b corresponds to a stepwise mechanism in which in a first step the hydride is transferred to the α -position relative to the sulfone group. Afterwards the proton is transferred to the corresponding β -position (Scheme S6). Both mechanisms [yielding the (*R*)- and (*S*)-product, respectively] were calculated and the Gibbs free energies are given in Table S5 and Figure S21. First, the starting materials form a precatalyst from which the hydride is transferred to the α -position relative to the sulfone group *via* a first transition-state. With a barrier of 10.8 kcal/mol for the mechanism leading to the (*S*)-product and 7.1 kcal/mol for the mechanism leading to the (*R*)-product and 2b. As a result, the transition states for the second step, i.e., the transfer of the proton to the β -position, have not been investigated any further.



Scheme S6. Schematic mechanism for the reaction pathway 2b leading to the (S)-product.

Table S5. Relative Gibbs free energies for pathway 2b leading to the (R)- and (S)-product. All values are given in kcal/mol (PBE0/def2-SVP).



Figure S21. Relative Gibbs free energies for pathway 2b leading to the (R)- and (S)-product (PBE0/def2-SVP).

Transition-State [(R)-Product]



Figure S22. Front view (left) and top view (right) of the transition state for the hydride transfer to α -position in a configuration leading to the (*R*)-product optimized using PBE0/def2-SVP.



Transition-State [(S)-Product]

Figure S23. Front view (left) and top view (right) of the transition state for the hydride transfer to α -position in a configuration leading to the (*S*)-product optimized using PBE0/def2-SVP.

Intermediate [(*R*)-Product]



Figure S24. Front view (left) and top view (right) of the intermediate where the hydride is transferred to α -position in a configuration leading to the (*R*)-product optimized using PBE0/def2-SVP.



Intermediate [(S)-Product]

Figure S25. Front view (left) and top view (right) of the intermediate where the hydride is transferred to α -position in a configuration leading to the (*S*)-product optimized using PBE0/def2-SVP.

5. Discussion

As shown in Section 3 there are 6 conceivable mechanisms which can yield the (S)- and the (R)-product. In Section 4.2 the pathways including a proton transfer to the oxygen of the sulfone group on the substrate (pathways 1c, 2a and 2c) have been further investigated for their general thermodynamics and on that basis could be ruled out. During the exploration of the remaining mechanisms it became evident that the reaction barriers for the hydride transfer tend to be higher than the barriers for the proton transfer. For this reason, only the transition states for the hydride transfer of mechanism 1a, 1b and 2b yielding the (S)- and the (R)-product have been optimized in a first step. The activation energies resulting from these transition states are summarized in Table S6. The two mechanisms with the lowest activation energy for the first reaction step both lead to the (S)-product and transfer the hydride to the β -position of the substrate. Moreover, both mechanisms are very similar with the only difference being that pathway 1a corresponds to a concerted mechanism with an activation energy of 3.4

kcal/mol, while pathway 1b happens in a stepwise fashion with the highest activation energy being the hydride transfer with 3.7 kcal/mol. Due to the similarity of the mechanism and the similar reaction barriers the transition states have been compared in detail. We found that they are identical except for rotations of side groups. In order to clarify to which reaction pathway the transition state actually belongs, an intrinsic reaction coordinate exploration has been performed. It showed that the reaction coordinate from the transition state belongs to the stepwise mechanism 1b. The remaining transition state for the proton transfer in step two of pathway 1b was calculated with a Gibbs free energy of -6.8 kcal/mol, and the associated activation energy is 3.2 kcal/mol. The full reaction pathway is shown in Figure S26. It can be seen that the activation energy for the proton transfer is lower than that for the hydride transfer and consequently, pathway 1b provides the most favorable mechanism for this reaction.

Table S6. Summary of the activation energies for the hydride transfer (PBE0/def2-SVP).

Mechanism	Enantiomer	Barrier
1a	(<i>R</i>)	9.0 kcal/mol
14	(S)	3.4 kcal/mol
1b	(<i>R</i>)	39.4 kcal/mol
	(S)	3.7 kcal/mol
$1c \qquad (R) \qquad H \qquad (S) \qquad H \qquad (R) \qquad (R) \qquad H \qquad (R)$	Excluded due to unfavorable proton transfer	
	(S)	
2a	(<i>R</i>)	Excluded due to unfavorable proton transfer

	(S)	
2b	(<i>R</i>)	7.1 kcal/mol
	(S)	10.8 kcal/mol
2c	(R)	Excluded due to unfavorable proton transfer
	(S)	1



Figure S26. Final reaction pathway (1b) with the overall lowest activation energies (PBE0/def2-SVP).

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(H) Copies of NMR spectra



S102





S104


































7.5 5.5 4.5 4.0 fl (ppm) 3.5 3.0 1.5 9.0 8.5 8.0 7.0 6.5 6.0 2.5 0.5 5.0 2.0 1.0 0.0









S119









S123



S124



S125