

Designing an Artificial Pathway for the Biosynthesis of a Novel Phenazine *N*-oxide in *Pseudomonas chlororaphis* HT66

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Table S1. DNA sequences of *phzS* from *Pseudomonas aeruginosa* PAO1, *phzO* from *Pseudomonas chlororaphis* GP72 and *phzNO1* from *Nocardiopsis* sp. 13-12-13 phenazine biosynthesis operons.

Gene	DNA sequences
<i>phzS</i>	<p>CGACACCGCTGCGCCGGCGTTTCATGGCGGATAACCGCAAGCGGTTATTCGCCCTACGC GGCCTTGGAGCCCATCTAACCGCACGGGTCATGCGTACCGCGGCCTCGGAGCCGGTT CGTAGGGCGAATGACGCCACCGGCGTTATCCGCGCTGCGCCGACGTTTCATCGCGGT AAACGGTCATCCATCCCAGCCGAACCCCATCGATTGAACTCGAGAAAAGGAAGC ACCC</p> <p>ATGAGCGAACCCATCGATATCCTCATCGCCGGCGCCGGCATCGGCGGCCT CAGTTGCGCCCTGGCCCTGCACCAGGCCGGCATCGGCAAGGTCACGCTGC TGGAAAGCAGCAGCGAGATACGCCCCCTTGGCGTCGGCATCAATATCCAG CCGGCGGCGGTGAGGCCCTTGCCGAACTGGGCCTCGGCCCGGCGCTGGC GGCCACCGCCATCCCACCCACGAGCTGCGCTACATCGACCAGAGCGGCG CCACGGTATGGTCCGAGCCGCGCGGGGTGGAAGCCGGCAACGCCTATCCG CAGTACTCGATCCATCGCGGCGAACTGCAGATGATCCTGCTCGCCGCGGT GCGCGAGCGCTCGGCCAACAGGCGGTACGCACCGGTCTCGGCGTGGAGC GTATCGAGGAGCGCGACGGCCGCTGCTGATCGGCGCCCGCGACGGACAC GGCAAGCCCCAGGCGCTCGGTGCCGATGTGCTGGTCGGCGCCGACGGTAT CCATTGCGCGGTCCGCGCGCACCTGCATCCCGACCAGAGGCCGCTGTCCC ACGGTGGGATCACCATGTGGCGCGGCGTACCGAGTTCGACCCTTCCTC GACGGCAAGACCATGATCGTCGCCAACGACGAGCACTGGTCGCGCCTGGT CGCCTATCCGATCTCGGCGCGTCACGCGGCCGAAGGCAAGTCGCTGGTGA ACTGGGTGTGCATGGTGGCGAGCGCCGCGTCCGCGCCAGCTCGACAACGAG GCCGACTGGAACCGCGACGGGCGCCTGGAGGACGTGCTGCCGTTCTTCGC CGACTGGGACCTGGGCTGGTTCGACATCCGCGACCTGCTGACCCGCAACC AGTTGATCCTGCAGTACCCGATGGTAGACCGCGATCCGCTGCCGCACTGG GGCCGGGGACGCATCACCTGCTCGGCGACGCCGCCACCTGATGTATCC GATGGGCGCCAACGGCGCTTCGCAAGCAATCCTCGACGGCATCGAGCTGG CCGCCGCGCTGGCGCGCAACGCCGACGTGGCCGACGCCCTGCGCGAATAC GAAGAAGCGCGGCGGCCGACCGCCAACAAGATCATCCTGGCCAACCGAG AACGGGAAAAAGAGGAATGGGCCGCGGCTTCGCGACCGAAGACCGAGAA GAGCGCGGCGCTGGAAGCGATCACCGGCAGCTACCGCAACCAGGTGGAA CGGCCACGCTAG</p>
<i>phzO</i>	<p>GTACCGAGATAAACATGCTTTGAAGTGCCTGGCTGCTCCAACTTCGAACTC ATTGCGCGAACTTCAACACTTATGACATCCGGTCAACATGAGAAGAGTCC ATATGCGAAAGAACGCGTATTCGAAATACCAAACAGAGAGTCCGGATCAC CAAAGTGTGTAACGACATTAATTCCTATCTGAATCTTATAGTTGCTCTAGA ACGTTGTCCTTGACCCAGCGATAGACATCGGGCCAGAGACGACACAAACA AAGTTAGACATTACTGAGGCTGCTACCATGCTAGATCTTCAAAAACAAGCG TAAATATCTGAAAAGTGCAGAATCCTTCAAAGCTTCACTGCGTGATGACC GCACTGTTATTTATCAAGGCCAAGTTGTTGAGGATGTGACTACACACTTCT</p>

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naphzNOI

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CCCGACTTCAAGGGCCGCGACAGCTTCGGGGCCGCTGGTCCACACCGG
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CAACGCCGAGGTGGACGAGGCCTACTTCACCGACCTGCGCGGCCGCTACG
ACCAGTCTGGGAGCAGGCCAAGAAGTCCCAGTGGCGTGGCGTTCGAG
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TCTTCCAGGAGAACTGGGACCGGGGCAACGGCTTCGCTTCATGTTCCGGC
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AACCCCTCGGTGGTGACCGCCT**TGA**

P_{PHZ}

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GTTGAAAACAGGCCGTTAGACTAGACCAACCTGAACCCCTGTCAACAAGC
AAAACCACAGAGTCATAGACCGCTTCGTACTAGCACTAGATTTCTGCATC
CGCCCCAACTAACTGGCCATAGGACATTGCATCCTGACGATGTTAGTAAG
CCATTAGTGCCTCCCAGCCGACGAATGAACCTGCCTCTTACAATAATGCA
ACAGTTCACCCGGTGTGACTGCAGCCAGCTCGTTCACGCCCTAATCACTAC
AAGATCTGGTAGTTCAGCCCCAAGAAATGCAGGTGTATAACAACACCC
GTTCAGCACCGCCACGAAGGAACAATAGTTAAAACTGTAAATTATCTAC
CCGCACCAAATTAACCTAACCATTTACTTTCAAAAAATACCAGCCCACTAA
GGAGGATGCCGCC

*The red capital letter indicates promoter of the gene *phzS*.

Table S2. ^1H (MeOH) and ^{13}C (MeOH) spectra data for 1-hydroxyphenazine *N'*10-oxide recorded by 600 MHz NMR spectrometer.

Position	δ_{H} (<i>J</i> in Hz)	δ_{C}	HMBC	HSQC (ppm)	COSY
1		153.5	H-3,4		
2	8.7/1.0	120.2	H-4	7.66, dd	H-3
3	8.7/7.8	134.0		7.78, dd	H-4,2
4	7.8/1.0	113.3	H-2	7.07, dd	H-3
4a		147.9	H-3		
5a		146.6	H-9,7		
6	8.9	130.8	H-8,9a	8.16, d	H-7
7		133.5	H-9	7.95, m	H-8,6
8		132.5	H-6	7.88, m	H-9,7
9	8.9	119.1	H-7	8.62, d	H-8
9a		134.2	H-6,8		
10a		126.4	H-4,2		

Table S3. Primers used in this study.

primers	Sequence 5' → 3'	application
<i>phzS</i> -F1	ccggggatcctctagaCGAAGTATCAGGCAATGGCGTCA	
<i>phzS</i> -R1	TCAGGGCTGCAGGCGGGT	
<i>phzS</i> -F	<i>ACCCGCCTGCAGCCCTGACGACACCGCTGCGCCGGCGT</i>	<i>phzS</i> replaced gene
<i>phzS</i> -R	CTAGCGTGGCCGTTCCACCTGGTTG	<i>phzH</i>
<i>phzS</i> -F2	<i>GTGGAACGGCCACGCTAGTACGAGCCTGAGGGAGCCAC</i>	
<i>phzS</i> -R2	ggccagtccaagcttCGCCATATGCTGGTTCGGC	
<i>phzO</i> -F1	acatgattacgaattaGCCGCTGTTGGGTAAAGG	
<i>phzO</i> -R1	GTTTATCTCGGTACCTCAGGGTTGCAAACGCC	
<i>phzO</i> -F	<i>GGTACCGAGATAAACATGCTTTGAAGTGC</i>	<i>phzO</i> replaced gene
<i>phzO</i> -R	CTATTTGGCGTTGAGCCCCACCATA	<i>phzH</i>
<i>phzO</i> -F2	<i>CTCAACGCCAAATAGTACGAGCCTGAGGGAGCCACGGCAG</i>	
<i>phzO</i> -R2	cgactctagaggatcaTGGCCGAACCACCTTGC	
<i>NaphzNO1</i> -F1	ccggggatcctctagaTGCGCGAAGGGTAATGC	
<i>NaphzNO1</i> -R1	GCAAAGACTCCTGAGTTCAAGC	
<i>NaphzNO1</i> -PF	<i>GAACTCAGGAGTCTTTGCTACGAATCCCATCCCAACTGC</i>	<i>NaphzNO1</i> replaced gene <i>pykA</i>
<i>NaphzNO1</i> -PR	GGCGGCATCCTCCTTAGTTG	(PF, PR were used to clone PHZ promoter.
<i>NaphzNO1</i> -F	<i>ACTAAGGAGGATGCCCGCTGACCAACGCGAAGAACACC</i>	
<i>NaphzNO1</i> -R	TCAGGCGGTCACCACCGA	
<i>NaphzNO1</i> -F2	<i>TCGGTGGTGACCGCCTGAGCCACCTGACGCAACAATAAAG</i>	
<i>NaphzNO1</i> -R2	ggccagtccaagcttTGTTGTTGCTGGGTGAGGGTT	
PET- <i>NaphzNO1</i> -F	GGAATTCC CATATG GTGACCAACGCGAAGAACACC	<i>NaphzNO1</i> expression
PET- <i>NaphzNO1</i> -R	CCG CTCGAG TCAGGCGGTCACCACCGA	

*The lowercase indicates that the overlapped fragments were used to construct plasmids by the In-fusion method. The italic capital letter indicates the overlapped fragments between Upstream and downstream homologous arms. The red capital letter indicates enzyme cut site.

Figure S2. The MS/MS result of 1-hydroxyphenazine *N*⁷10-oxide.

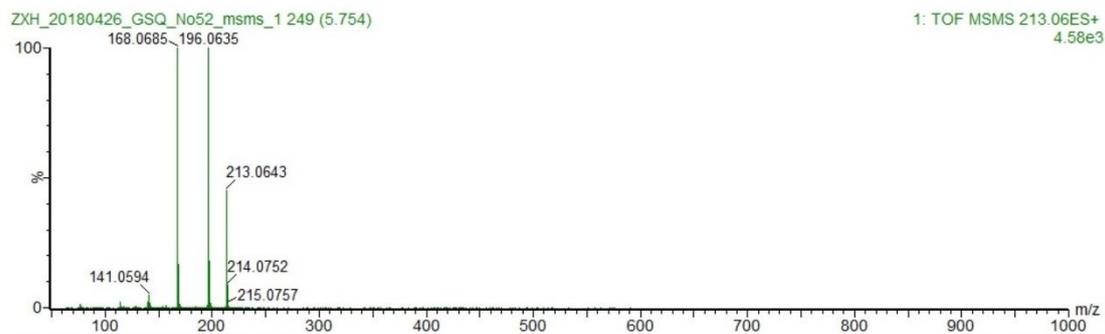
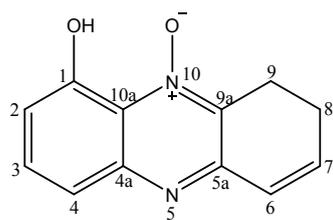
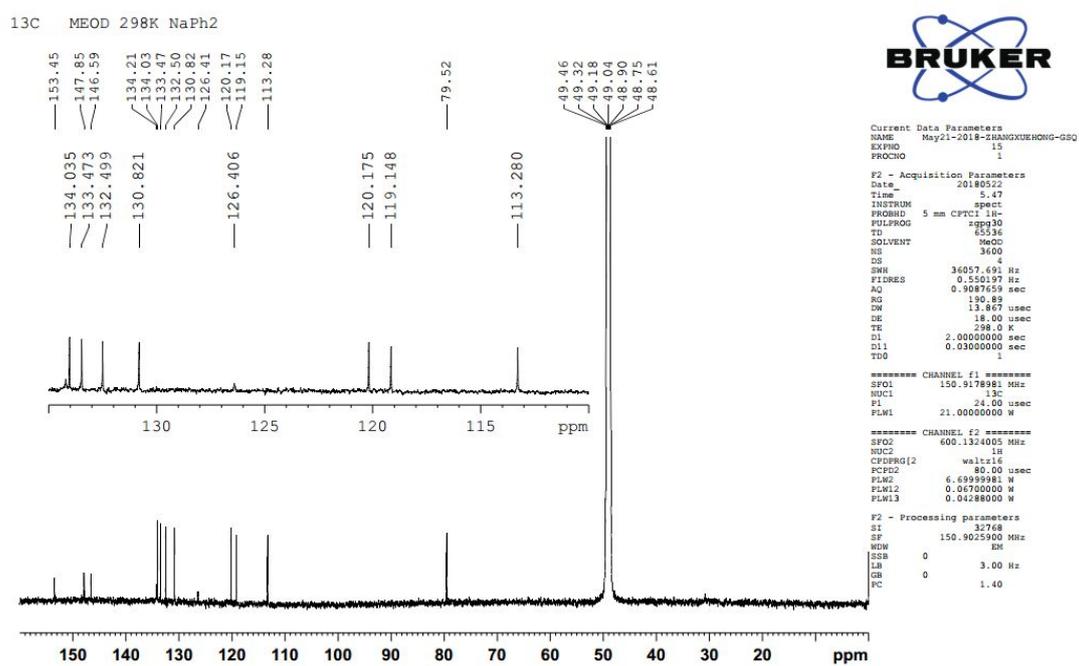


Figure S4. The ^{13}C NMR spectrum of 1-hydroxyphenazine *N'*10-oxide (MeOH, 151 MHz).



*The peak at ~78 ppm in ^{13}C -NMR was impurity signal interference.

Figure S5. The COSY spectrum of 1-hydroxyphenazine *N*'10-oxide (MeOH, 600 MHz).

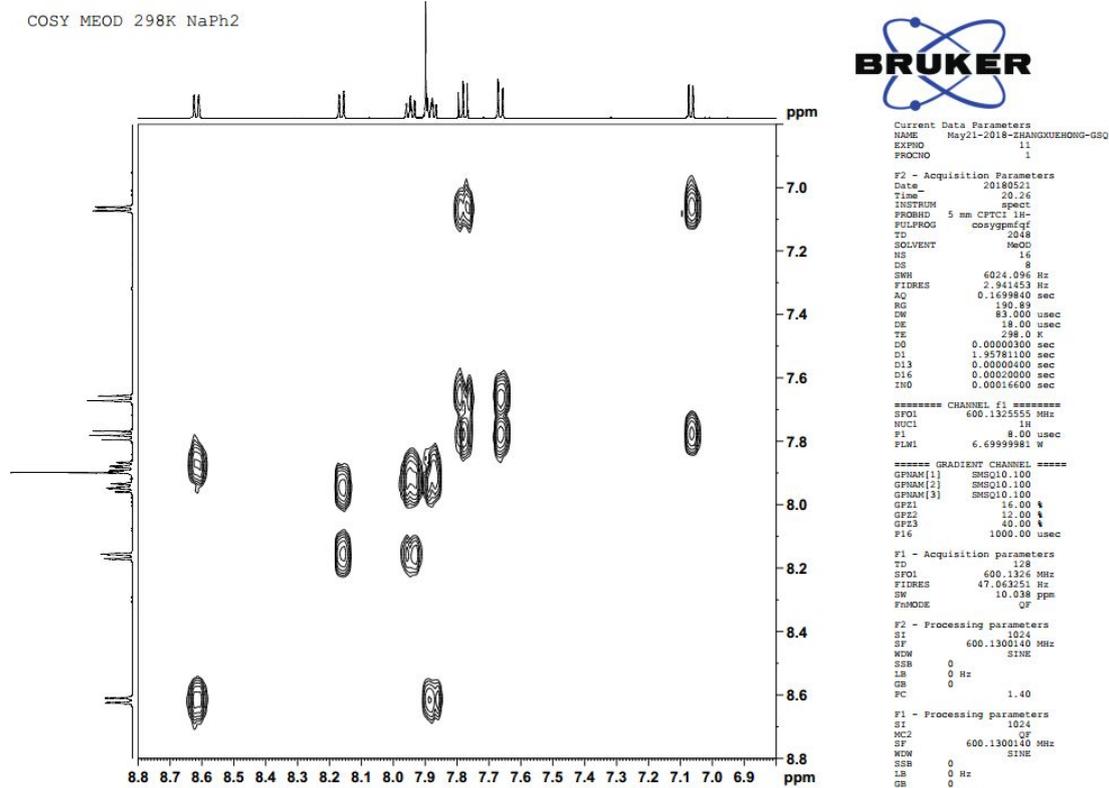
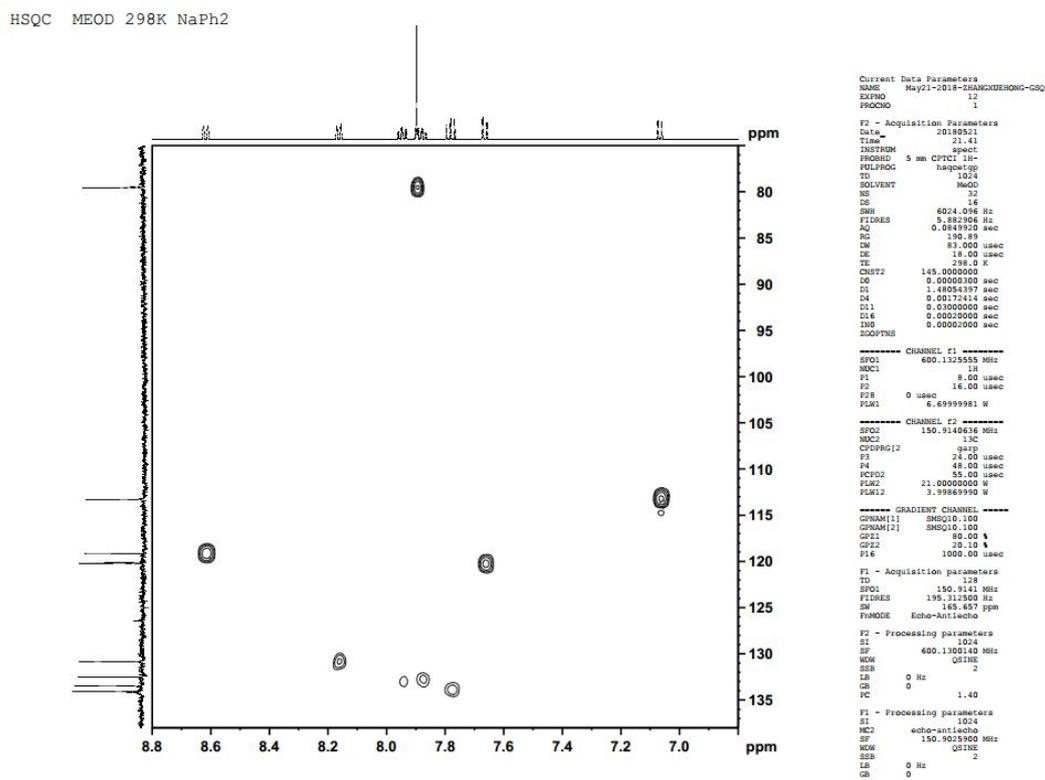


Figure S6. The HSQC spectrum of 1-hydroxyphenazine *N*¹⁰-oxide (MeOH, 151 MHz).



*The peak at ~78 ppm in ¹³C-NMR was impurity signal interference.

Figure S7. The HMBC spectrum of 1-hydroxyphenazine *N*'10-oxide (MeOH, 151 MHz).

HMBC MEOD 298K NaPh2

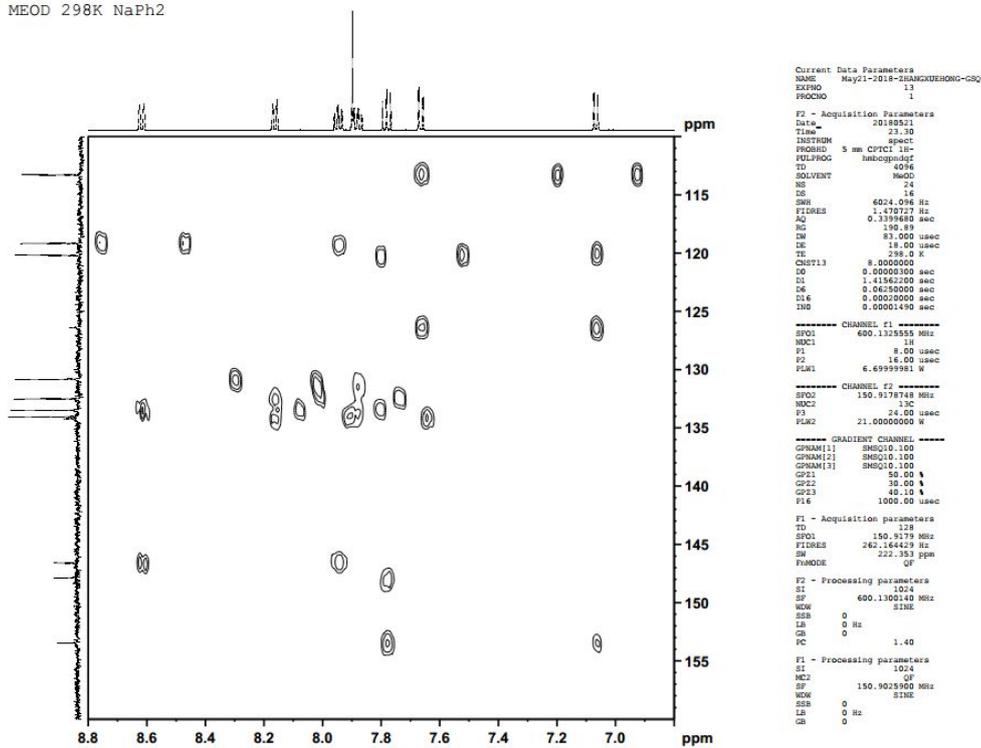


Figure S8. Culture profiles of 1-hydroxyphenazine *N*10-oxide-producing *P. chlororaphis* strains. (A): HT66-SN; (B): P3-SN. (Data represent the mean \pm SD from three independent cultures).

