

# Investigation of Temporal Apparent C4 Sugar Change in Manuka Honey

Anatoly Chernyshev\*, Terry Braggins

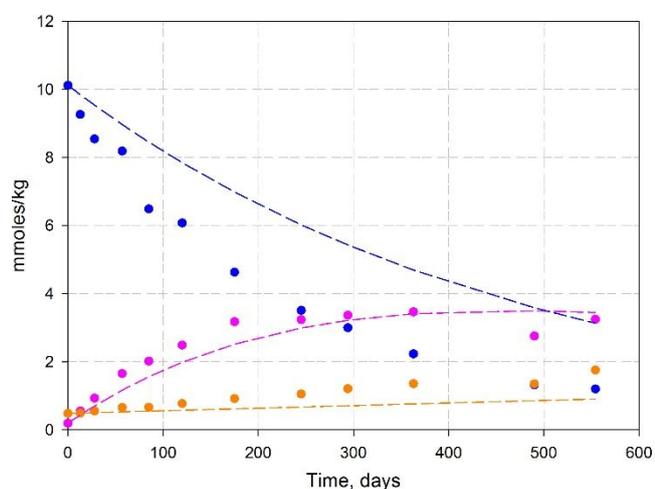
Analytica Laboratories Ltd., Ruakura Research Centre, 10 Bisley Rd, Hamilton 3240, New Zealand

## Supplementary materials

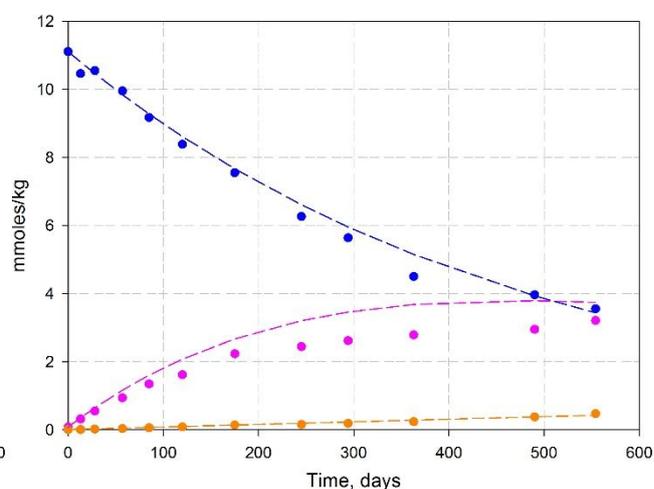
**Table S1.** The results of retesting a set of manuka honeys with Bruker Honey-Profiling™ NMR method (adopted from a conference presentation<sup>11</sup>).

Sample #	A	B	C	D	E	F	G	H
$\delta^{13}C_h$	-24.70	-25.72	-25.16	-24.82	-25.62	-25.36	-25.51	-25.45
$\delta^{13}C_p$	-27.06	-27.61	-26.91	-26.18	-26.61	-26.03	-27.24	-26.10
Apparent C4 sugar %	13.6	10.6	10.2	8.3	5.9	4.1	9.9	4.0
DHA, mmol/kg	4.9	4.2	5.7	20.0	23.1	30.1	5.0	28.0
MG, mmol/kg	8.5	10.6	13.0	5.5	5.0	6.1	11.8	6.2
HMF, mmol/kg	0.9	0.6	0.7	0.1	0.1	0.1	0.7	0.1
<b>Bruker Honey-Profiling™ NMR test results</b>								
Sugar syrups	No	No	Yes	No	No	No	No	No

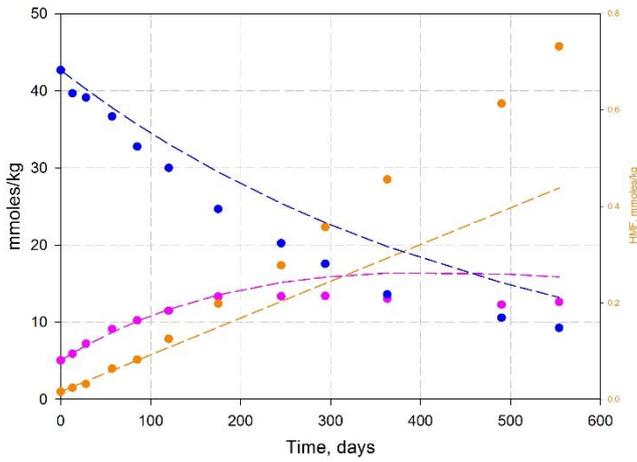
01\_NIUE\_DHA



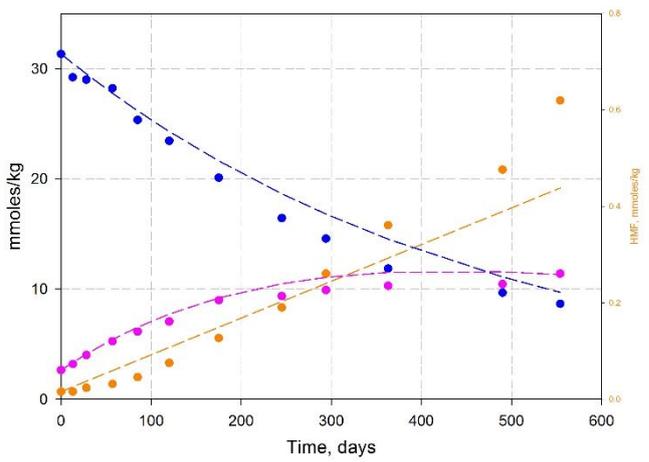
02\_CLOVER\_DHA



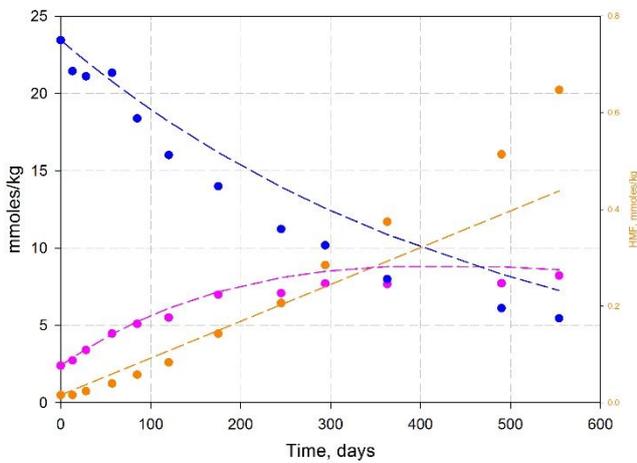
04\_MANUKA



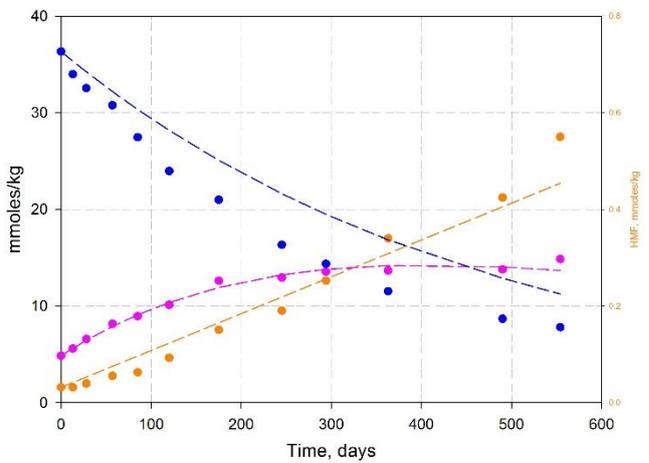
05\_MANUKA



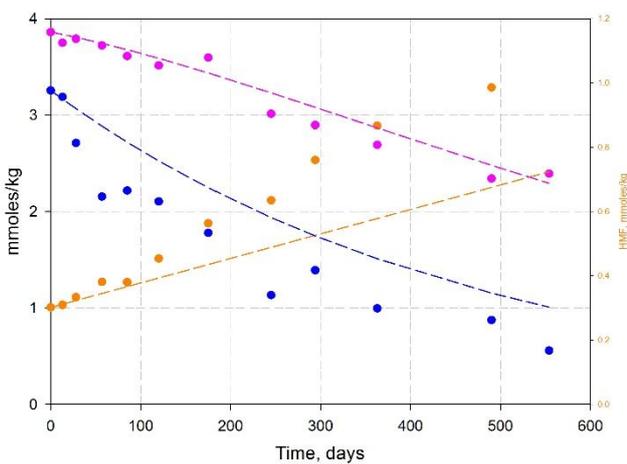
06\_MANUKA



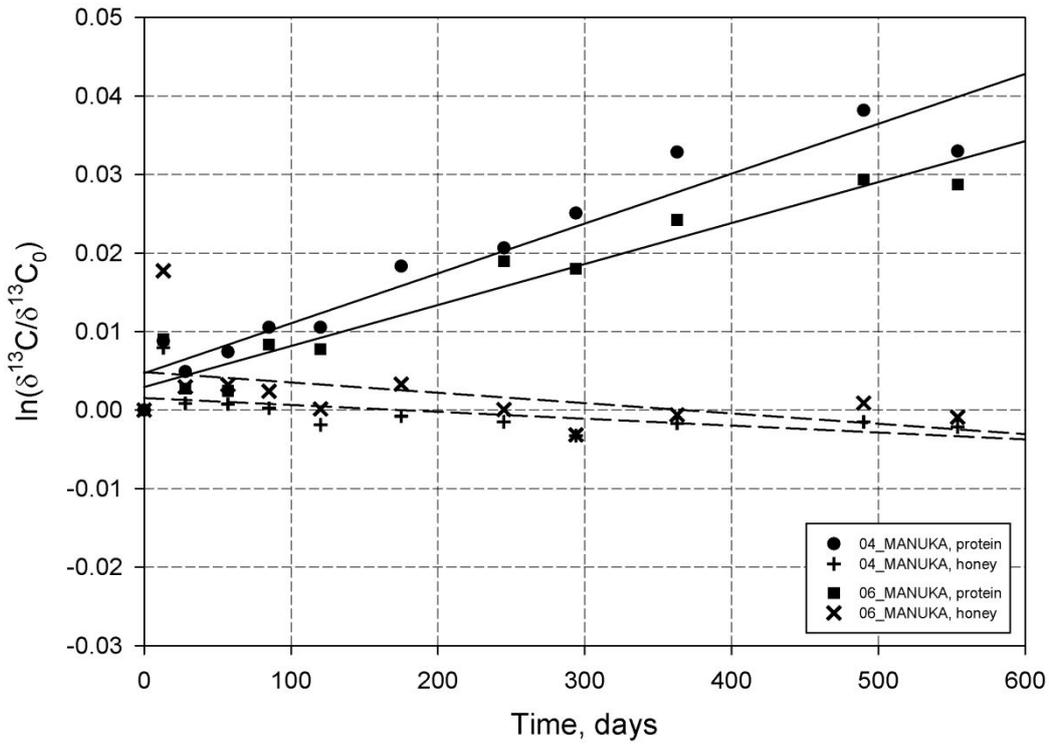
07\_MANUKA



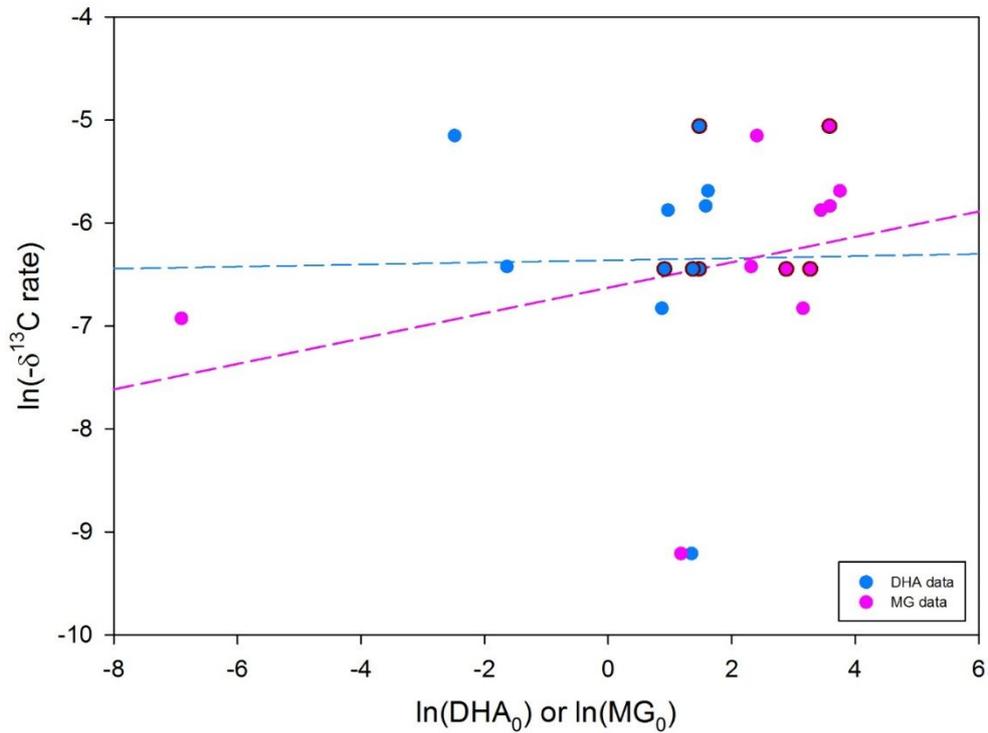
08\_MANUKA\_OLD



**Figure S1.** Change in concentrations of DHA (blue), MG (magenta), HMF (brown) over time in the studied honey samples. Dots — experimental values; dashed lines — forecast by Analytica’s kinetic model. Sample #3 (pure clover honey) is not shown.

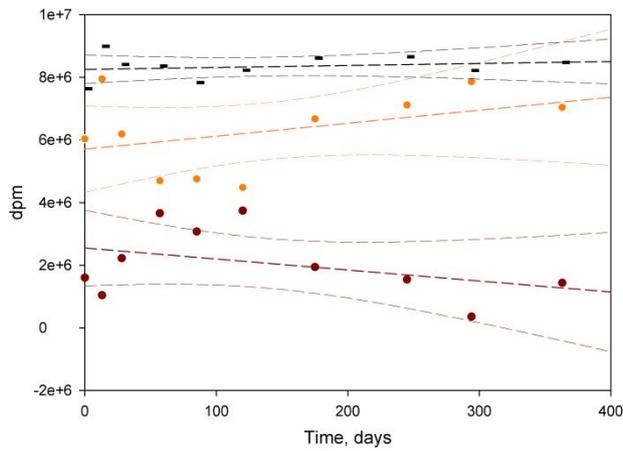


**Figure S2.** The plots of  $\ln(\delta^{13}\text{C}/\delta^{13}\text{C}_0)$  vs. time for the other two high quality manuka honeys (group 3). The grey dashes are 95% confidence intervals.

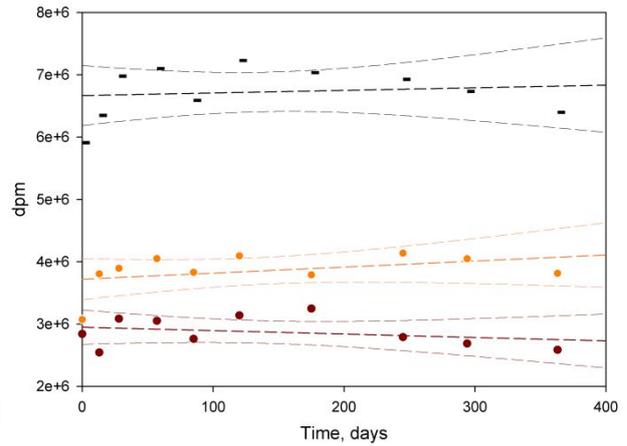


**Figure S3.** The plot of  $\ln(-d\delta^{13}\text{C}_p/dt)$  vs.  $\ln(\text{DHA}_0)$  and  $\ln(\text{MG}_0)$ . The plot includes the data of Rogers<sup>1</sup> (bordered circles).

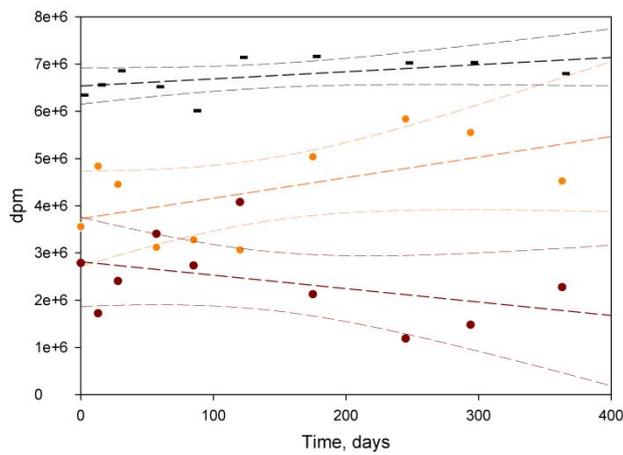
01\_NIUE\_DHA



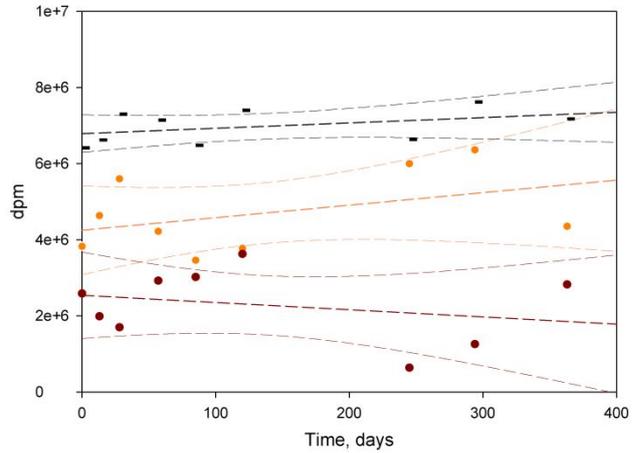
02\_CLOVER\_DHA



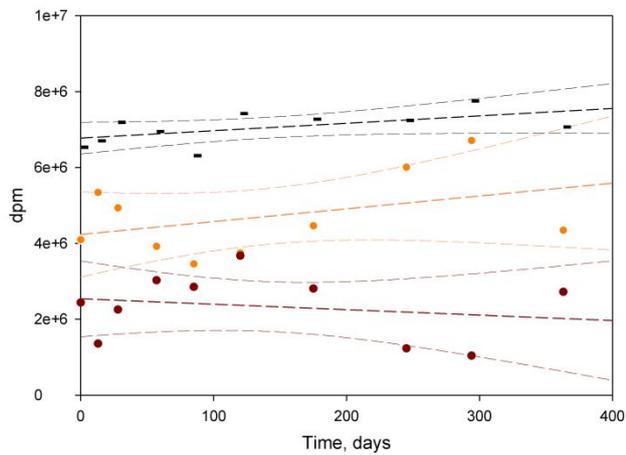
04\_MANUKA



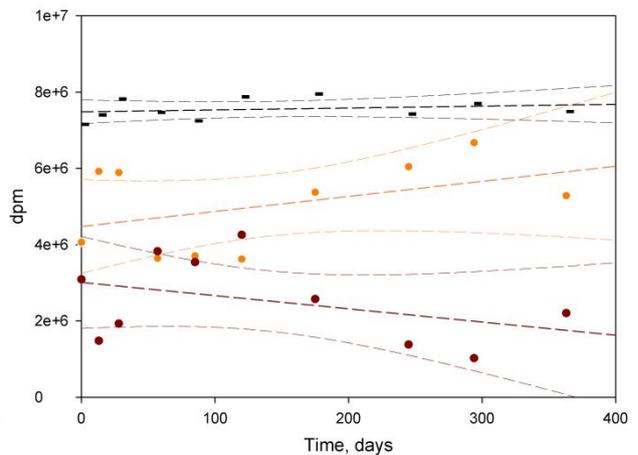
06\_MANUKA



07\_MANUKA



08\_MANUKA\_OLD



**Figure S4.** Change in scintillation counts, with 95% confidence intervals, for honey (light brown), protein (dark brown), and total (black) over time in the studied honey samples. The black dashes show the total amount of  $^{14}\text{C}$  label in the sample, and serve as a control to check in there was any loss of the labelled material during the protein precipitation.

**Table S2.** The results of isotopic headspace analysis of the honey samples.

Sample ID	O <sub>2</sub> , %	CO <sub>2</sub> , ppm	δ <sup>13</sup> C
Laboratory air	20.5	770	-23.4
01_NIUE_DHA	20.3	1260	-27.6
02_CLOVER_DHA	19.2	1500	-29.8
03_CLOVER	19.3	6530	-26.4
04_MANUKA	18.8	12110	-35.2
05_MANUKA	18.4	4840	-35.0
06_MANUKA	18.0	12300	-37.8
07_MANUKA	19.3	1200	-25.7
08_MANUKA_OLD	19.2	6240	-34.9
Fresh, non-incubated manuka honey*	20.3	1540	-27.1

**Table S3.** The results of δ<sup>13</sup>C measurements in artificial honey. Highlighted bold is the component changing its isotopic composition.

Time	Sample	δ <sup>13</sup> C	σ(δ <sup>13</sup> C)
Day 0	Artificial Honey + DHA	-12.28	0.02
	<b>Artificial Protein + DHA</b>	<b>-10.00</b>	0.02
	Artificial Honey - DHA	-12.17	0.07
	Artificial Protein - DHA	-9.99	0.03
Day 7	Artificial Honey + DHA	-12.32	0.04
	<b>Artificial Protein + DHA</b>	<b>-10.71</b>	0.02
	Artificial Honey - DHA	-12.28	0.05
	Artificial Protein - DHA	-10.12	0.08