Supporting Information

Discovery of highly potent benzimidazole derivatives as indoleamine 2,3-dioxygenase-1 (IDO1) inhibitors: from structure-based virtual screening to *in vivo* pharmacodynamic activity

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Molecular modelling



Figure S1. Docking pose of compounds 23 (A) and 35 (B) on IDO1 crystal structure (PDB id:

2D0T). Amino acids of pocket A, pocket B and pocket C are depicted in red, green and blue,

respectively. Heme group, compounds 23 and 35 are depicted as cyan, blue and orange

sticks, respectively.



Figure S2. MST binding curve of compound 10 to recombinant human IDO1.



Figure S3. MST binding curve of epacadostat (1) to recombinant human IDO1.



Figure S4. (**A**) RMSD of the atomic positions for compound **10** (in red, Lig fit Prot) and the protein IDO-1 (C α positions in blue) of the 100 ns molecular dynamics simulations using Desmond package. (**B**) A timeline representation of the interactions and contacts (H-bonds, Hydrophobic, Ionic, Water bridges).



Figure S5. (A) RMSD of the atomic positions for compound 23 (in red, Lig fit Prot) and the

protein IDO-1 (C α positions in blue) of the 100 ns molecular dynamics simulations using

Desmond package. (**B**) A timeline representation of the interactions and contacts (H-bonds, Hydrophobic, Ionic, Water bridges).



Figure S6. (**A**) RMSD of the atomic positions for compound **35** (in red, Lig fit Prot) and the protein IDO-1 (C α positions in blue) of the 100 ns molecular dynamics simulations using Desmond package. (**B**) A timeline representation of the interactions and contacts (H-bonds, Hydrophobic, Ionic, Water bridges).

NMR Spectra of compounds

10: ¹H (DMSO-*d*₆), ¹³C (DMSO-*d*₆) -11.85 -8.63 7,56 7,50 7,50 7,50 7,29 7,29 6,89 -5.49 -4,41 -4.0E+11 -3.5E+11 [[-3.0E+11 -2.5E+11 -2.0E+11 -1.5E+11 -1.0E+11 -5.0E+10 -0.0E+00 P:83-J 0.90 0.96 2.06 4 0.93 로 1.98-1 2.00-I 0.75-I 12.0 11.5 11.0 10.5 10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 fl (ppm) 144.65 144.65 144.05 134.15 134.15 127.22 127.22 127.23 12 160.06 -95.46 -48.13 -42.28 -7.0E+10 -6.0E+10 -5.0E+10 -4.0E+10 -3.0E+10 -2.0E+10 -1.0E+10 -0.0E+00 -1.0E+10 -2.0E+10 -3.0E+10 -4.0E+10 -5.0E+10)0 190 180 170 160 150 140 130 120 110 100 f1 (ppm) 90 80 70 60 50 40 30 10 0 20

23: ¹H (DMSO-*d*₆), ¹³C (DMSO-*d*₆)



35: ¹H (DMSO-*d*₆), ¹³C (DMSO-*d*₆)



S11

Biology

Commence 1	Cell viability (%)	IDO1 cellular assay		
Compound	(a) $10 \ \mu M \pm SD$	inhibition (%) @ 10 µM	IC ₅₀ (μM)	
VS1	95 ± 4.5	76 ± 4.9	4.4	
VS2	87 ± 6.1	68 ± 6	0.9	
VS3	98 ± 3.9	28 ± 4	n.d.	
VS4	97 ± 2.7	78 ± 2	0.6	
VS5	92 ± 4.2	76 ± 3.5	3.7	
VS6	98 ± 2.6	70 ± 5.6	2.6	
VS7	98 ± 1.9	32 ± 6	n.d.	
VS8	96 ± 3.5	86 ± 9	0.8	
VS9	94 ± 3.3	56 ± 7	n.d.	
<i>VS10</i> (10)	100 ± 5.5	99 ± 0.7	0.016	

Table S1. Biological profiles of the 10 active compounds from virtual screening.



Fig. S7. Concentration-response curves for selected compounds (10, 23 and 35) in IDO1 inhibition cell-based assay. A375 cells were treated (48 h) with IFN- γ (1000 U/mL) and increasing concentrations (0.001-3 μ M) of each compound. The L-KYN levels in cell culture supernatants were measured by HPLC method. The data represent mean \pm SEM of at least three independent assays run

in triplicate. Percent inhibition against inhibitor concentration was plotted and IC_{50} value was determined. The data were evaluated by GraphPad 7.0.



Fig. S8. Concentration-response curves (μ M) for selected compounds (10, 23 and 35) in stable transfectant P1.IDO1 and P1.TDO cell lines.

Metabolic stability data

Chromatographic methods

1) LC-UV methods for purity and metabolic stability determination.

A Shimadzu HPLC system (Shimadzu, Kyoto, Japan), consisting of two LC-10AD Vp module pumps, an SLC-10A Vp system controller, an SIL-10AD Vp autosampler, and a DGU-14-A on-line degasser were used for the analysis. The SPD-M10Avp photodiode array detector was used to detect the analytes. LC-Solution 1.24 software was used to process the chromatograms.

- Column: *Phenomenex Synegi Polar*, 150 × 4.6 mm (4 μm d.p.) protected with a SecurityGuard[®] (Torrance, CA, USA).
- Eluent:

A: 0.2% formic acid in water.

B: 0.2% formic acid in acetonitrile.

- Flow rate: 1 mL/min.
- Injection volume: 20 µL.
- Wavelength: 294 nm (23), 268 nm (10), 254 nm (35),
- Gradient program: 0.00 min [B%=20%], 14.50 min [B%=80%], 15.00 min [B%=20%], 20 min [B%=20%].

2) LC-HRMS methods for metabolites characterization.

A Thermo Scientific Q-Exactive Plus system equipped with a Thermo Scientific Vanquish UHPLC system with a binary pump VF-P10, a split sampler VF-A10, and a column compartment VH-C10. Data were acquired and processed using Xcalibur[®] software.

- Column: *Phenomenex Synergi Polar 150 × 2 mm (4μm d.p.)* protected with a SecurityGuard[®] kept at 35 °C (Torrance, CA, USA).
- Eluent:
 - A: 0.1% formic acid in water.
 - B: acetonitrile.
- Flow rate: 0.250 mL/min.
- Injection volume: $5 \mu L$.
- Gradient program: 0.00 min [B%=20%], 14.50 min [B%=80%], 15.00 min [B%=20%], 20.00 min [B%=20%].

The operating conditions of the mass spectrometer were as follows: positive mode; sheath gas flow rate, 45 Auxiliary Units (A.U.), auxiliary gas flow rate, 5 A.U.; sweep gas flow rate, 0 A.U.; spray voltage, 3.2 kV; capillary temperature, 300 °C; auxiliary gas heater temperature, 280 °C. Data were acquired in full-scan/data dependent MS² mode using a mass scan range m/z 110-800. The stepped collision energy was set at: 20, 40, 80% levels.

3) LC-HRMS methods for pharmacokinetic analysis and evaluation of L-Kyn trend levels.

A Thermo Scientific Q-Exactive Plus system equipped with a Thermo Scientific Vanquish UHPLC system with a binary pump VF-P10, a split sampler VF-A10, and a column compartment VH-C10. Data were acquired and processed using Xcalibur[®] software.

- Column: *Phenomenex Synergi Polar 150 × 2 mm (4 μm d.p.)* protected with a SecurityGuard[®] kept at 35 °C (Torrance, CA, USA).
- Eluent:
 - A: 0.1% formic acid in water.
 - B: acetonitrile.
- Flow rate: 0.250 mL/min.
- Injection volume: $5 \mu L$.
- Gradient program: 0.00 min [B%=20%], 1.00 min [B%=20%], 7.50 min [B%=95%], 10.00 min [B%=95%], 10.50 min [B%=20%], 16.00 min [B%=20%].

The operating conditions of the mass spectrometer were as follows: positive mode; sheath gas flow rate, 45 Auxiliary Units (A.U.), auxiliary gas flow rate, 5 A.U.; sweep gas flow rate, 0 A.U.; spray voltage, 3.2 kV; capillary temperature, 300 °C; auxiliary gas heater temperature, 280 °C. Data were acquired in full-scan and parallel reaction monitoring (PRM) using a mass scan range m/z 110-800 with the inclusion list and the collision energy parameters reported in Table S2.

Compound	Mass [m/z]	Formula [M]	(N)CE
10	409.06585	$C_{20}H_{17}BrN_4O$	35
23	381.17099	$C_{24}H_{20}N_4O$	35
L-Kyn	209.09207	$C_{10}H_{12}N_2O_3$	10

Table S2



Figure S9. Representative PRM chromatogram of all the analytes.

Structural characterization of metabolites by HRMS

Scheme S1. Metabolic pathways of compound 10.



	RT (min)	Molecular Formula	Teoretical [M+H] ⁺	Measured [M+H] ⁺	Mass shift (ppm)	Metabolic pathway
10	8.02	$C_{20}H_{17}BrN_4O$	409.06584	409.06607	0.55	
M1	4.40	$C_{15}H_{12}N_2O_2$	253.09715	253.09723	0.30	oxidative N-dealkylation
M2	6.92	$C_{20}H_{17}BrN_4O_2$	425.06076	425.06091	0.36	+ O
M3	7.28	$C_{20}H_{17}BrN_4O_2$	425.06076	425.06090	0.34	+ O
M4	8.14	$C_{13}H_{13}BrN_2O_2$	309.02331	309.02347	0.52	oxidative N-dealkylation + reduction
M5	8.61	$C_{13}H_{11}BrN_2O_3$	323.00257	323.00266	0.27	oxidative N-dealkylation
M6	9.85	$C_{13}H_{11}BrN_2O_2$	307.00760	307.00778	0.59	oxidative N-dealkylation + reduction
M7	10.49	$C_{20}H_{17}BrN_4O_2$	425.06076	425.06088	0.29	+ 0

Br

Ň-H





IL-Pb_002 (F18) #1241, RT=6.889 min, MS2, FTMS (+), (HCD, DDF, 425.0610@30,



IL-P-GSH_002 (F19) #671, RT=4.416 min, MS2, FTMS (+), (HCD, DDF, 253.0972@30, +1

HO

0











IL-Pa 002 (F17) #1333, RT=7.292 min, MS2, FTMS (+), (HCD, DDF, 425.0609@30,







IL-Pb_002 (F18) #1935, RT=9.823 min, MS2, FTMS (+), (HCD, DDF, 307.0078@30,





IL-P-GSH_002 (F19) #1648, RT=8.605 min, MS2, FTMS (+), (HCD, DDF, 323.0027@30, +1)



II -Ph 002 (F18) #2088 RT=10 483 min MS2 FTMS (+) (HCD DDF 425 0608@30,

Scheme S2. Metabolic pathways of compound 23.



M1

	RT (min)	Molecular Formula	Teoretical [M+H] ⁺	Measured [M+H] ⁺	Mass shift (ppm)	Metabolic pathway
23	9.77	$C_{24}H_{20}N_4O$	381.17099	381.17020	2.07	
M1	4.78	$C_{15}H_{12}N_2O_2$	253.09715	253.09671	1.75	oxidative N-dealkylation
M2	7.03	$C_{24}H_{20}N_4O_2$	397.16590	397.16568	0.56	+ O
M3	7.41	$C_{24}H_{20}N_4O_2$	397.16590	397.16565	0.63	+ O
M4	8.24	$C_{24}H_{20}N_4O_2$	397.16590	397.16562	0.71	+ O
M5	8.79	$C_{24}H_{20}N_4O_2$	397.16590	397.16547	1.09	+ O
M6	9.28	$C_{17}H_{14}N_2O_3$	295.10772	295.10742	1.01	oxidative N-dealkylation



P-GSH 002 (F6) #1709, RT=4.758 min, MS2, FTMS (+), (HCD, DDF, 253.0969@(20;40;80),



B0_002 (F1) #4241. RT=9.750 min. MS2. FTMS (+). (HCD. DDF. 381.1703@(20:40;80), +1'



Pa_002 (F4) #2872, RT=7.017 min, MS2, FTMS (+), (HCD, DDF, 397.1656@(20;40;80),

Pa_002 (F4) #3071, RT=7.415 min, MS2, FTMS (+), (HCD, DDF, 397.1656@(20;40;80), +1)





+ 135.04382 C8 H7 O2 [M-e]+1

144.04420 C9 H6 N O [M-e]+1/

150

160.03923 C9 H6 N O2 [M-e]+1

m/z

204.08035

200

1

295.10727 C17 H15 N2 O3 [M+H]+1 (+Oxidation)

296.1106

300

HN-C

252.10005 C16 H14 N O2 [M-e]+1 1 (+Oxidation)

250

- S HN- (

234.09116

206.09651 C16 H12 N O [M-e]+1

Pa 002 (F4) #3481. RT=8.232 min. MS2. FTMS (+). (HCD. DDF. 397.1657@(20:40:80), +1

I[]

107.04902 C7 H7 O [M-e]+1

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95.0490

100

79.05412 91.05413 C6 H7 [M-e]+1 C7 H7 [M-e]+1 118.06493 C8 H8 N [M-e]+1

130.0649

117.057

117.033

M6

• *•*

N

Н

Н

Intensity [counts] (10^A6) 0 00 00

> 0 + 50

Ωİ

65.03876 C5 H5 [M-e]+1

Q,

56.96491 77.0387 •

Scheme S3. Metabolic pathways of compound 35.



M4, M6





35



M5

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	RT (min)	Molecular Formula	Teoretical [M+H] ⁺	Measured [M+H] ⁺	Mass shift (ppm)	Metabolic pathway
35	8.10	$C_{23}H_{18}N_4O$	367.15534	367.15463	1.93	
M1	4.18	$C_{15}H_{14}N_2O$	239.11789	239.11746	1.80	oxidative <i>N</i> -dealkylation + reduction
M2	4.78	$C_{15}H_{12}N_2O_2$	253.09715	253.09677	1.52	oxidative N-dealkylation
M3	5.81	$C_{15}H_{12}N_2O$	237.10224	237.10193	1.30	oxidative N-dealkylation + reduction
M4	6.86	$C_{23}H_{18}N_4O_2$	383.15025	383.14981	1.15	+ O
M5	7.21	$C_{16}H_{14}N_2O_2$	267.11284	267.11243	1.53	oxidative N-dealkylation + reduction
M6	7.30	$C_{23}H_{18}N_4O_2$	383.15025	383.14966	1.55	+ O
M7	7.84	$C_{16}H_{12}N_2O_3$	281.09207	281.09180	0.96	oxidative N-dealkylation

B0_002 (F23) #3421, RT=8.102 min, MS2, FTMS (+), (HCD, DDF, 367.1548@(20;40;80),

NC





IL-P-GSH_002 (F19) #671, RT=4.416 min, MS2, FTMS (+), (HCD, DDF, 253.0972@30,



Pa 002 (F25) #1402, RT=4.170 min, MS2, FTMS (+), (HCD, DDF, 239.1175@(20;40;80),

+

HO

HO

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M2



Pb_002 (F26) #2792, RT=6.848 min, MS2, FTMS (+), (HCD, DDF, 383.1497@(20;40;80), +1



Pa. 002 (F25) #2265. RT=5.812 min. MS2. FTMS (+). (HCD. DDF. 237.1019@(20:40;80),

Pa_002 (F25) #2961, RT=7.181 min, MS2, FTMS (+), (HCD, DDF, 267.1124@(20;40;80), +1











P-GSH_002 (F27) #3289, RT=7.833 min, MS2, FTMS (+), (HCD, DDF, 281.0919@(20;40;80),

In vivo PK evaluation of compound 23.

Parameter	Unit	Value
$t_{1/2}$	h	0.605
T _{max}	h	0.033
C _{max}	μg/L	2684.268
V_d	L/Kg	6.545
Cl	L/h/Kg	7.493

Table S3. PK parameters of single administration of compound 23 (10 mg/kg).

In vivo PD evaluation of compound 23.

Fig. S10. Residual KYN levels after endovenous administration of 23.

