Molecular Networking Reveals Serpentinine-related Bisindole Alkaloids from *Picralima nitida*, a Previously Well-Investigated Species

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Figure S1. Molecular network: "molecular family" A

Figure S1. "Molecular family" **A** of the full molecular network of the EtOH extract and fractions from the aerial parts of *Picralima nitida*, with the dereplicated compounds (the stereochemistry depicted in this figure derives from that of the standards)

Compounds	Score (cosine)	ΔRT^1 (min)	MSI identification level ²
10-hydroxy- geissoschizol	0.91	0.43	1
3,4,5,6-tetradehydro- geissoschizol	0.93	0.88	1
akagerine	0.79	1.57	1
akuammicine	0.69	2.57	2
akuammigine	0.95	0.41	1
alloyohimbine	0.86	1.41	1
cinchonine	0.75	/3	2
corynantheidal	0.91	1.02	1
corynantheol	0.93	2.69	2
iboxygaine	0.67	1.15	1
N-methyl antirhine	0.85	5.95	2
pericine	0.88	5.78	2
pericyclivine	0.74	0.05	1
pseudoyohimbine	0.83	5.26	2
strictosamide	0.91	4.55	2
strychnine-N-oxide	0.61	/ 3	2
raubasine	0.88	3.48	2
vindolinine	0.65	4.99	2
voachalotine	0.73	5.77	2

Table S2. Dereplicated Compounds Against the MIADB-GNPS

¹ Retention times of the monoterpene indole alkaloids contained in the MIADB are accessible from MetaboLights under the identifier:MTBLS142. Delta RTs can be calculated since both acquisitions were realized under the same chromatographic conditions.

² E. L. Schymanski, J. Jeon, R. Gulde, K. Fenner, M. Ruff, H. P. Singer and J. Hollender, Environ. Sci. Technol., **2014**, *48*, 2097–2098.

³ The retention times were not available since these compounds do not belong to the MIADB



Figure S3. Molecular network: molecular families A and B

Figure S3. Molecular families A and B of the full molecular network of the ethanolic extract and fractions from the aerial parts of *Picralima nitida*.

Table S4.HRMS-based dereplication – Selected compounds

Table S4: Dereplication of the selected compounds against DNP and Reaxys®

compounds	species	measured m/z	exact <i>m/z</i>	hits	molecular formula
1	$[M^+]$	631.3628	631.3643	13 hits - Not described in P. nitida	$C_{40}H_{47}N_4O_3{}^+$
2	$[M^+]$	629.3477	629.3486	9 hits - Not described in P. nitida	$C_{40}H_{45}N_4O_3^+$
3	$[M^+]$	685.3410	685.3384	2 hits - serpentinine and 3-oxovoafrine B	$C_{42}H_{45}N_4O_5^+$
4	$[M^+]$	671.3247	671.3228	No hits	$C_{41}H_{43}N_4O_5^+$

databases

5	$[M^+]$	435.1923	435.1914	No hits	$C_{25}H_{27}N_2O_5^+$





Figure S5. Comparison between the experimental ECD spectra of compounds 1 (red) and 2 (black), in methanol

Figure S6. Comparison between the experimental ECD spectra of 3 and 4



Figure S6. Comparison between the experimental ECD spectra of compounds 3 (black) and 4 (red), in methanol



Figure S7. Comparison of 3D models of 1 and moandaensine

S8. Computational methods

Conformations of compound **1**, **3** and **5** were fully optimized *in vacuo* and without constraint using DFT^{1,2} with the hybrid Becke-3-parameter-Lee–Yang–Parr^{3,4} exchange– correlation functional and the 6-31G* basis,⁵ as implemented in the Gaussian 16 revB.01 software package.⁶ Upon geometrical optimization convergence, a frequency calculation within the harmonic approximation was conducted at the same level of theory and local minima were characterized by the absence of imaginary frequency. TDDFT at the same level of theory was

then employed to predict energies as well as rotational strengths of the first 60 electronic transitions of the lowest energy conformer. The UCSF Chimera v1.11⁷ software package was used for the depiction of the most stable conformer of compounds **1**, **3** and **5**. ECD spectrum envelope was then calculated using SpecDis v1.71⁸ (with different sigma/gamma values) and rendered using Gnuplot v5.2.⁹

Coordinates of the lowest energy conformer of 1 found at the B3LYP/6-31G* level

Electronic energy (B3LYP): -1997.04808272 Ha.

Lowest frequency: 9.0584 cm⁻¹.

Free enthalpy: -1996.31721 Ha.

Atomic coordinates:

1 C 7.790196 1.279433 2.416231 2 C 8.370060 2.507594 2.709519 3 C 8.100666 3.645833 1.923094 4 C 7.245048 3.580286 0.828358 5 C 6.662630 2.343479 0.537660 6 C 6.921965 1.180094 1.315717 7 N 5.783038 1.982025 -0.469090 8 C 6.172308 0.104956 0.719029 9 C 5.506699 0.627114 -0.360372 10 C 6.109214 -1.362133 1.022502 11 C 4.545134 -0.109424 -1.251153 12 N 4.624176 -1.564798 -1.012691 13 C 5.795626 -2.081596 -0.305316 14 C 3.114145 0.417284 -0.977606 15 C 1.992257 -0.338900 -1.734528 16 C 2.385070 -1.802820 -1.867058 17 C 3.355991 -2.252759 -0.792438 18 C 2.087901 -2.608376 -2.900109 19 C 1.323204 -2.291771 -4.155064 20 C 0.630792 -0.149324 -0.976908 21 C -0.558349 -0.758178 -1.766000 22 C 0.346781 1.349890 -0.816378 23 0 0.091609 2.036714 -1.783715 24 O 0.350309 1.937816 0.399575 25 C 0.558189 1.220632 1.624892 26 C -1.890233 -0.613758 -1.067901 27 C -2.381609 -1.522061 -0.106487 28 C -3.656022 -1.260827 0.476432 29 C -4.430030 -0.147420 0.155830 30 N -3.907332 0.695454 -0.770570 31 C -2.693359 0.474473 -1.359833 32 N -3.953182 -2.267029 1.364945 33 C -2.910479 -3.181002 1.387119 34 C -1.903092 -2.750182 0.479450 35 C -0.736276 -3.527162 0.333334 36 C -0.612001 -4.688963 1.079425 37 C -1.626313 -5.094685 1.971828 38 C -2.787034 -4.350723 2.140720 39 C -5.782252 0.092691 0.771419 40 C -6.338408 1.515325 0.592082 41 C -6.109784 1.958557 -0.869420 42 C -4.606662 1.961969 -1.151615 43 C -6.699301 3.346552 -1.202328 44 C -6.476733 3.816576 -2.646441 45 C -7.811438 1.599666 1.040235 46 C -8.087796 0.992540 2.416464 47 0 -7.110002 1.489605 3.326211 48 H 4.788937 0.103316 -2.309175 49 H 1.865091 0.110970 -2.725533 50 H -7.406739 1.304074 4.229902 51 H 5.500401 2.579252 -1.231155 52 H -4.798717 -2.340323 1.910559 53 H 8.006172 0.407709 3.028714 54 H 9.043950 2.595432 3.557082 55 H 8.570717 4.592238 2.175078 56 H 7.040442 4.459111 0.222252 57 H 5.347170 -1.600016 1.780325 58 H 7.065969 -1.725237 1.420587 59 H 5.636454 -3.153502 -0.146366 60 H 6.673188 -1.981327 -0.958616 61 H 2.955148 0.336672 0.103986 62 H 3.071080 1.484835 -1.218814 63 H 3.537472 -3.329320 -0.878637 64 H 2.923562 -2.088632 0.218766 65 H 2.496872 -3.619203 -2.866777 66 H 1.929623 -2.550386 -5.032254 67 H 1.047958 -1.237188 -4.240251 68 H 0.403876 -2.888549 -4.231202 69 H 0.709904 -0.647238 -0.005773 70 H -0.352600 -1.812833 -1.958566 71 H -0.608350 -0.251925 -2.733285 72 H 1.556988 0.779587 1.672990 73 H 0.462545 1.970412 2.411098 74 H -0.201375 0.445411 1.770155 75 H -2.374605 1.231103 -2.065562 76 H 0.055625 -3.232271 -0.347072 77 H 0.279569 -5.299184 0.977202 78 H -1.497094 -6.010068 2.541322 79 H -3.564539 -4.666930 2.829318 80 H -5.717633 -0.131777 1.842547 81 H -6.482633 -0.637607 0.335369 82 H -5.761260 2.181489 1.246848 83 H -6.583415 1.219427 -1.535138 84 H -4.117203 2.771576 -0.597146

85H-4.4041762.110059-2.21285886H-6.2905954.088496-0.50161787H-7.7768743.310510-1.01487188H-6.8302793.070448-3.36852789H-7.0323154.741030-2.83130090H-5.4244164.028600-2.86604791H-8.1069882.6527121.06699492H-8.4611041.1005560.30702693H-8.053444-0.1086282.36877894H-9.1048181.2672032.728608

Coordinates of the lowest energy conformer of 3 found at the B3LYP/6-31G* level

Electronic energy (B3LYP): -2222.52864472 Ha.

Lowest frequency: 7.9883 cm⁻¹.

Free enthalpy: -2221.801413 Ha.

Atomic coordinates:

1 C -5.515717 0.713951 -3.866587 2 C -5.718905 0.651657 -2.359976 3 H -5.615427 -0.387324 -2.015353 4 O -7.101757 1.026215 -2.136292 5 C -7.473610 1.370078 -0.889850 6 C -6.639198 1.560073 0.153814 7 C -7.122198 1.863510 1.512062 8 0 -8.454359 2.045489 1.584456 9 C -8.977461 2.332949 2.896043 10 0 -6.374973 1.929664 2.476813 11 C -5.144515 1.466907 -0.034147 12 H -4.678317 2.308733 0.492121 13 C -4.579175 0.166324 0.578579 14 C -3.241722 -0.282613 0.055041 15 C -2.523335 -1.309666 0.659973 16 N -2.856408 -2.010152 1.794085 17 C -1.876867 -2.957258 2.049357 18 C -1.806451 -3.881686 3.094289 19 C -0.712194 -4.736583 3.111935 20 C 0.287383 -4.676549 2.118309 21 C 0.217305 -3.754706 1.085231 22 C -0.879356 -2.870922 1.038419 23 C -1.290764 -1.813826 0.147789 24 C -0.776784 -1.239173 -1.034533 25 C 0.521175 -1.653175 -1.688830 26 C 1.766597 -0.892966 -1.167257

27 H 1.843093 -0.946078 -0.079709 28 C 1.652069 0.579282 -1.544344 29 0 2.188385 1.369937 -0.606088 30 C 2.238423 2.782134 -0.896673 31 0 1.147475 0.994689 -2.573712 32 C 3.063875 -1.540130 -1.784430 33 C 3.402695 -2.847138 -1.092186 34 C 3.947033 -2.681305 0.328260 35 C 3.329188 -4.067278 -1.646818 36 C 2.855879 -4.458928 -3.018950 37 H 2.837137 -1.721763 -2.839739 38 C 4.332758 -0.649342 -1.749797 39 C 5.076623 -0.678169 -0.377647 40 N 4.198483 -1.278309 0.640644 41 C 4.668394 -1.083923 2.017196 42 C 4.621073 0.399421 2.409594 43 C 5.495335 0.684938 0.087563 44 C 5.289696 1.204467 1.336962 45 C 5.824347 2.542133 1.334235 46 C 5.914237 3.567567 2.291434 47 C 6.528830 4.764459 1.942879 48 C 7.058135 4.961895 0.651816 49 C 6.980706 3.968301 -0.319122 50 C 6.359407 2.767525 0.034226 51 N 6.126944 1.624361 -0.714111 52 C -1.523046 -0.222222 -1.604265 53 N -2.702447 0.233217 -1.081059 54 C -3.309123 1.423518 -1.750660 55 C -4.811480 1.582195 -1.531876 56 H -5.046430 2.604419 -1.856650 57 H -5.548365 1.748075 -4.225659 58 H -6.311367 0.154133 -4.365562 59 H -4.560110 0.266579 -4.157859 60 H -8.548710 1.489029 -0.812696 61 H -8.536249 3.251894 3.288939 62 H -8.763463 1.509323 3.581613 63 H -10.052019 2.449923 2.758630 64 H -5.286981 -0.658128 0.409145 65 H -4.527708 0.300366 1.664725 66 H -3.667370 -1.845085 2.372018 67 H -2.574951 -3.932196 3.859420 68 H -0.625618 -5.469076 3.908764 69 H 1.124091 -5.366199 2.165904 70 H 0.996335 -3.724375 0.331508 71 H 0.446269 -1.473310 -2.765701

72 H 0.677864 -2.725944 -1.551733 73 H 1.228495 3.201188 -0.909009 74 H 2.710884 2.955601 -1.865764 75 H 2.831603 3.219100 -0.094723 76 H 4.856893 -3.303866 0.447344 77 H 3.220685 -3.052361 1.064371 78 H 3.681535 -4.904951 -1.040058 79 H 2.502570 -3.614156 -3.616155 80 H 2.039657 -5.190943 -2.956918 81 H 3.664512 -4.948042 -3.577814 82 H 4.083323 0.379920 -2.021098 83 H 5.009442 -1.020608 -2.527601 84 H 5.984581 -1.307629 -0.498530 85 H 5.697363 -1.470290 2.147865 86 H 4.017839 -1.670565 2.675914 87 H 5.120616 0.531965 3.378039 88 H 3.574799 0.708596 2.546428 89 H 5.514875 3.424664 3.292346 90 H 6.608021 5.561694 2.676589 91 H 7.537362 5.906191 0.409689 92 H 7.391254 4.122496 -1.313742 93 H 6.507791 1.450177 -1.631744 94 H -1.174408 0.289985 -2.491624 95 H -3.057343 1.351042 -2.809410 96 H -2.793785 2.301678 -1.347201

Coordinates of the lowest energy conformer of 5 found at the B3LYP/6-31G* level

Electronic energy (B3LYP): -1454.38856820 Ha.

Lowest frequency: 16.8398 cm⁻¹.

Free enthalpy: -1453.959186 Ha.

Atomic coordinates:

1C-3.7966831.956272-0.7669052C-4.1834133.273151-0.9598013C-3.2441424.325679-0.9249864C-1.8938734.091034-0.6982235C-1.5033752.764295-0.5065156C-2.4314301.684483-0.5340367N-0.2330982.257799-0.2723108C-1.6654820.488281-0.2998709C-0.3075450.893367-0.14168210C-1.962555-0.890644-0.221372

11 C 0.735269 -0.000843 0.087407 12 N 0.394771 -1.315015 0.150941 13 C -0.900073 -1.742236 0.005083 14 C 2.166276 0.456394 0.170015 15 C 3.185765 -0.591392 0.668701 16 C 2.845427 -2.002369 0.158328 17 C 1.403557 -2.356673 0.513485 18 C -3.362231 -1.433471 -0.364348 19 C 4.591407 -0.234353 0.250881 20 C 5.210657 -0.924750 -0.729434 21 0 4.658830 -1.911467 -1.459494 22 C 3.235190 -2.147745 -1.325973 23 C -4.222783 -1.231393 0.904170 24 C -5.705075 -1.217507 0.560972 25 0 -6.160498 -0.671653 -0.425314 26 0 -6.437389 -1.843614 1.487726 27 C -7.868142 -1.827923 1.279033 28 C 2.988566 -3.520421 -1.934269 29 C 5.231234 0.887136 0.960934 30 0 6.520048 1.076051 0.623233 31 0 4.631654 1.579926 1.769708 32 C 7.187703 2.170864 1.281728 33 H 3.143503 -0.596798 1.764702 34 H 3.464913 -2.723508 0.707658 35 H -4.539700 1.165458 -0.795365 36 H -5.228759 3.500753 -1.141524 37 H -1.175205 4.904253 -0.672778 38 H 0.605489 2.813856 -0.189491 39 H -1.041040 -2.813262 0.082870 40 H 2.453391 0.800994 -0.834116 41 H 2.238594 1.334123 0.821670 42 H 1.085517 -3.283489 0.033669 43 H 1.327402 -2.498649 1.596912 44 H -3.321792 -2.500225 -0.609737 45 H -3.859491 -0.939761 -1.204805 46 H 6.240303 -0.743274 -1.017161 47 H 2.730777 -1.379427 -1.929511 48 H -4.002598 -0.257517 1.360693 49 H -4.014284 -1.992199 1.660393 50 H -8.116381 -2.317247 0.334690 51 H -8.287609 -2.375452 2.121949 52 H -8.235541 -0.799424 1.261945 53 H 3.451460 -3.569315 -2.923544 54 H 1.918348 -3.714681 -2.057953 55 H 3.424772 -4.310364 -1.313660

56 H8.2046412.1652130.89070857 H7.1890322.0200252.36367458 H6.6902603.1158591.05026559 H-3.5843735.345211-1.079359

Experimental Method

S9. Biological Assays

Antiplasmodial Evaluation. The chloroquine-resistant strain FcB1/Colombia of *Plasmodium falciparum* was obtained from the National Museum Natural History collection, Paris, France (n°MNHN-CEU-224-PfFCB1). Parasites were maintained in vitro in human erythrocytes in RPMI 1640 medium supplemented by 8% (v/v) heatinactivated human serum at 37 °C under an atmosphere of 3% CO₂, 6% O₂, and 91% N₂. Human red blood cells and serum were provided by the Etablissement Français du Sang Ile de France under the C-CPSL-UNT approval n°13/EFS/126. In vitro drug susceptibility was measured by [3H]-hypoxanthine incorporation as described previously.¹⁰ Stock solutions of drugs were prepared in DMSO. Compounds were serially diluted two-fold with 100 µL culture medium in 96-well plates. Asynchronous parasite cultures (100 µL, 1% parasitemia and 1% final hematocrit) were then added to each went and incubated for 24 h at 37 °C prior to the addition of 0.5 µCi of [³H]hypoxanthine (GE Healthcare; 1 to 5 Ci.mmol/mL) per well. After a further incubation of 24 h, plates were frozen and thawed. Cell lysates were then collected onto glassfiber filters and counted in a liquid scintillation spectrometer. The growth inhibition for each drug concentration was determined by comparison of the radioactivity incorporated in the treated culture with that in the control culture maintained on the same plate. The concentration causing 50% growth inhibition (IC_{50}) was obtained from the drug concentration-response curve and the results were expressed as mean values ± standard deviations as determined from three independent experiments. Chloroquine diphosphate, used as a positive control, was purchased from Sigma (purity>99%).

Cell Culture and Proliferation Assay. The MRC-5 cell line was obtained from the American Type Culture Collection (Rockville, MD, USA) and cultured according to the supplier's instructions. Human MRC-5 cells were grown in DMEM supplemented with 10% fetal calf serum (FCS) and 1% glutamine. The cell line was maintained at 37 °C in a humidified atmosphere containing 5% CO₂. Cell growth inhibition was determined by an MTS assay according to the manufacturer's instructions (Promega, Madison, WI, USA). Briefly, the cells were seeded in 96-well plates (2.5×10^3 cells/well) containing 200 µL of growth medium. After 24 h of culture, the cells were treated with the test compounds at different final concentrations. After 72 h of

incubation, 40 μ L of resazurin was added for 2 h before recording absorbance at 490 nm with a spectrophotometric plate reader. The IC₅₀ value corresponded to the concentration of compound inducing a decrease of 50% in absorbance of drug-treated cells compared with untreated cells. Experiments were performed in triplicate. Paclitaxel was used as the reference compound.



Figure S10. ¹H NMR Spectrum (500 MHz, DMSO- d_6) of moandaensine B (1)



Figure S11. ¹³C NMR Spectrum (125 MHz, DMSO- d_6) of moandaensine B (1)



Figure S12. COSY spectrum (500 MHz, DMSO- d_6) of moandaensine B (1)



Figure S13. HSQC spectrum (500 MHz, DMSO- d_6) of moandaensine B (1)



Figure S14. HMBC spectrum (500 MHz, DMSO- d_6) of moandaensine B (1)



Figure S15. ROESY spectrum (500 MHz, DMSO- d_6) of moandaensine B (1)



Figure S16. ¹H NMR spectrum (500 MHz, DMSO- d_6) of moandaensine C (2)



Figure S17. ¹³C NMR spectrum (125 MHz, DMSO- d_6) of moandaensine C (2)





Figure S19. HSQC spectrum (500 MHz, DMSO- d_6) of moandaensine C (2)





Figure S21. ROESY spectrum (500 MHz, DMSO- d_6) of moandaensine C (2)



Figure S22. ¹H NMR spectrum (600 MHz, DMSO-*d*₆) of 20'-*epi*-serpentinine (3)



Figure S23. ¹³C NMR spectrum (150 MHz, DMSO-*d*₆) of 20'-*epi*-serpentinine (3)











Figure S28. ¹H NMR spectrum (500 MHz, DMSO-*d*₆) of 16-demethoxy-20'-*epi*-serpentinine (4)



Figure S29. ¹³C NMR spectrum (125 MHz, DMSO-*d*₆) of 16-demethoxy-20'-*epi*-serpentinine (4)





Figure S31. HSQC spectrum (500 MHz, DMSO-*d*₆) of 16-demethoxy-20'-*epi*-serpentinine (4)









Figure S35. ¹³C NMR spectrum (100 MHz, DMSO- d_6) of alstonine 6-methylpropionate (5)











Figure S40. ECD and UV spectra of $1\,$



Figure S40. ECD and UV spectra of 2



Figure S40. ECD and UV spectra of 3



Figure S40. ECD and UV spectra of 4



Figure S40. ECD and UV spectra of 5

References

- 1. Hohenberg, P.; Kohn, W., Inhomogeneous Electron Gas. *Physical Review* **1964**, *136*, B864-B871.
- 2. Kohn, W.; Sham, L. J., Self-Consistent Equations Including Exchange and Correlation Effects. *Physical Review* **1965**, *140*, A1133-A1138.
- 3. Becke, A. D., Density-functional thermochemistry. III. The role of exact exchange. *The Journal of Chemical Physics* **1993**, *98*, 5648-5652.
- 4. Lee, C.; Yang, W.; Parr, R. G., Development of the Colle-Salvetti correlation-energy formula into a functional of the electron density. *Physical Review B* **1988**, *37*, 785-789.
- Wiberg, K. B., Ab Initio Molecular Orbital Theory by W. J. Hehre, L. Radom, P. v. R. Schleyer, and J. A. Pople, John Wiley, New York, 548pp. Price: \$79.95 (1986). *Journal of Computational Chemistry* 1986, 7, 379-379.
- 6. Frisch, M. J. T., H.B.; Schlegel, G.W.; Scuseria, G.E.; Robb, M.A.; Cheeseman, J.R.; Scalmani, G.; Barone, V.; Petersson, G.A.; Nakatsuji, H.; *et al.* Gaussian 16 Revision B.01; Gaussian Inc.: Wallingford, CT, USA, 2016.
- 7. Pettersen, E. F.; Goddard, T. D.; Huang, C. C.; Couch, G. S.; Greenblatt, D. M.; Meng, E. C.; Ferrin, T. E., UCSF Chimera—A visualization system for exploratory research and analysis. *Journal of Computational Chemistry* **2004**, *25*, 1605-1612.
- 8. Bruhn, T.; Schaumlöffel, A.; Hemberger, Y.; Bringmann, G., SpecDis: Quantifying the Comparison of Calculated and Experimental Electronic Circular Dichroism Spectra. *Chirality* **2013**, *25*, 243-9.
- 9. Williams, T. K., C., and many others., Gnuplot 4.6: an Interactive Plotting Program. http://gnuplot.sourceforge.net/.
- Guillon, J.; Grellier, P.; Labaied, M.; Sonnet, P.; Léger, J.-M.; Déprez-Poulain, R.; Forfar-Bares, I.; Dallemagne, P.; Lemaître, N.; Péhourcq, F.; Rochette, J.; Sergheraert, C.; Jarry, C., Synthesis, Antimalarial Activity, and Molecular Modeling of New Pyrrolo[1,2-a]quinoxalines, Bispyrrolo[1,2-a]quinoxalines, Bispyrido[3,2e]pyrrolo[1,2-a]pyrazines, and Bispyrrolo[1,2-a]thieno[3,2-e]pyrazines. *J. Med. Chem.* **2004**, *47*, 1997-2009.