

Diversity-Oriented Library Synthesis from Steviol and Isosteviol-Derived Scaffolds

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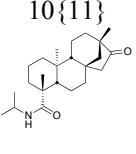
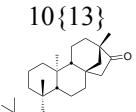
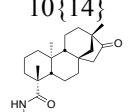
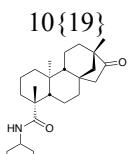
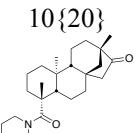
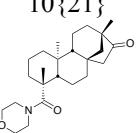
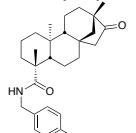
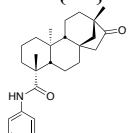
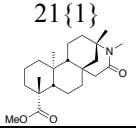
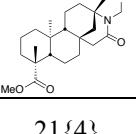
Supporting Information

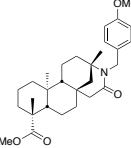
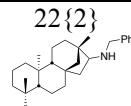
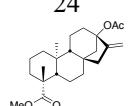
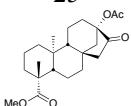
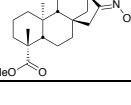
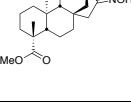
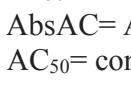
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Table S1. Complete current activity table of isosteviol and steviol analogs from PubChem

Compound	PubChem ID	Assay ID	BioAssay	Activity (if given)	
				Assay Type	Result or Value (μM)
1 	9905087	602438	uHTS identification of modulators of interaction between CendR and NRP-1 using fluorescence polarization assay	Primary screen	active
2 	42601320	652104	qHTS of TDP-43 inhibitors	Confirmatory	$\text{IC}_{50}^{\text{A}}$ 19.953
5 	42601318	720706	HTS for bacterial rRNA inhibitors measured in microorganism-based system	Primary screen	active
10{2} 	42601330	540271	<i>In vivo</i> -based yeast HTS to detect compounds rescuing yeast growth/survival of Plasmodium falciparum HSP40-mediated toxicity measured in whole organism system	Confirmatory	AbsAC _c 1.427
		686978	qHTS for inhibitors of human TDP1: qHTS in cells in absence of CPT	Confirmatory	EC_{50} 18.356
		652041	Cell-based secondary assay to test the inhibitory activity of small molecule on Plasmodium falciparum (HB3 strain) survival in red blood cells measured in cell-based system	Confirmatory	AbsAC 21.72
		652047	Cell-based secondary assay to test the inhibitory activity of small molecule on Plasmodium falciparum (3D7 strain) survival in red blood cells measured in cell-based system	Confirmatory	AbsAC 30.22
		2825	uHTS Luminescent assay for identification of inhibitors of NALP3 in yeast	Primary screen	active
		435006	Single concentration confirmation of uHTS for the identification of inhibitors of NALP3 in yeast using a luminescent assay	Primary screen	active
		504582	<i>In vivo</i> -based yeast HTS to detect compounds rescuing yeast growth/survival of Plasmodium falciparum HSP40-mediated toxicity measured in whole organism system	Primary screen	active
10{4} 	42601335	624417	qHTS of GLP-1 receptor inverse agonists	Confirmatory	EC_{50} 10
		624466	Fluorescence-based cell-based primary HTS assay to identify antagonists of human TAAR1	Primary screen	active
		2825	uHTS Luminescent assay for identification of inhibitors of NALP3 in yeast	Primary screen	active
		504582	<i>In vivo</i> -based yeast HTS to detect compounds rescuing yeast growth/survival of Plasmodium falciparum HSP40-mediated toxicity measured in whole organism system	Primary screen	active
10{10} 	42601331	488784	Single concentration confirmation of inhibitors of NALP3 in yeast using Caspase-1-ASC counter screen	Primary screen	active
		2685	qHTS assay for lipid storage modulators in drosophila S3 cells	Confirmatory	EC_{50} 0.651
		2825	uHTS Luminescent assay for identification of inhibitors of NALP3 in yeast	Primary screen	active
		463195	uHTS identification of small molecule inhibitors of tim10 yeast via a luminescent assay	Primary screen	active
		435006	Single concentration confirmation of uHTS for the identification of inhibitors of NALP3 in yeast using a luminescent assay	Primary screen	active

		488794	Single concentration confirmation of uHTS for the identification of inhibitors of NALP3 in yeast using a luminescent assay-retest	Primary screen	active
	42601329	2825	uHTS luminescent assay for identification of inhibitors of NALP3 in yeast	Primary screen	active
		488784	Single concentration confirmation of inhibitors of NALP3 in yeast using Caspase-1-ASC counter screen	Primary screen	active
		488794	Single concentration confirmation of uHTS for the identification of inhibitors of NALP3 in yeast using a luminescent assay-retest	Primary screen	active
		2825	uHTS luminescent assay for identification of inhibitors of NALP3 in yeast	Primary screen	active
	42601326	686978	qHTS for inhibitors of human TDP1: qHTS in cells in absence of CPT	Confirmatory	EC ₅₀ 23.109
	42601334	686978	qHTS for inhibitors of human TDP1: qHTS in cells in absence of CPT	Confirmatory	EC ₅₀ 23.109
	42601327	686979	qHTS for inhibitors of human TDP1: qHTS in cells in absence of CPT	Confirmatory	EC ₅₀ 18.356
602438		uHTS identification of modulators of interactions between CendR and NRP-1 using fluorescence polarization assay	Primary screen	active	
		602123	Fluorescence polarization-based primary biochemical HTS assay to identify inhibitors of E. coli DNA-binding ATP-dependent protease La (eLon)	Primary screen	active
624169		Luminescence-based cell-based primary HTS assay to identify agonists of the mouse HTR2A	Primary screen	active	
	42601333	652048	qHTS of D3 dopamine receptor agonist	Primary screen	active
652051		qHTS of D3 dopamine receptor potentiators	Primary screen	active	
	42601336	504582	<i>In vivo</i> -based yeast HTS to detect compounds rescuing yeast growth/survival of Plasmodium falciparum HSP40-mediated toxicity measured in whole organism system	Primary screen	active
	42601328	602438	uHTS identification of modulators of interactions between CendR and NRP-1 using fluorescence polarization assay	Primary screen	active
	42601319	-	Inconclusive data in 7 assays and inactive in 417 assays	Primary screen	inactive
	53299290	-	N/A	N/A	N/A
21{4}	42601337	624132	Shn3: Dual-Go Shn3RL cells measured in cell-based system	Confirmatory	AC ₅₀ ^D 10.58

	624133	Schnurri-3 Inhibitors: specific inducers of adult bone formation measured in cell-based system	Confirmatory	AC ₅₀ 8.16
	504832	Primary qHTS for delayed death inhibitors of the malarial parasite plastid, 48 hr incubation	Confirmatory	IC ₅₀ 1.651
	504834	Primary qHTS for delayed death inhibitors of the malarial parasite plastid, 96 hr incubation	Confirmatory	IC ₅₀ 1.472
	504444	Nrf2 qHTS screen for inhibitors	Confirmatory	IC ₅₀ 14.581
	651820	qHTS assay for inhibitors of hepatitis C virus	Confirmatory	IC ₅₀ 12.589
	686978	qHTS for inhibitors of human TDP1: qHTS in cells in absence of CPT	Confirmatory	EC ₅₀ 6.513
	686979	qHTS for inhibitors of human TDP1: qHTS in cells in absence of CPT	Confirmatory	EC ₅₀ 11.582
	743417	Schnurri-3 inhibitors: specific inducers of adult bone formation measured in cell-based system	Confirmatory	AC ₅₀ 8.16
	42601325	686978 qHTS for inhibitors of human TDP1: qHTS in cells in absence of CPT	Confirmatory	EC ₅₀ 20.596
	42601321	588850 uHTS identification of cystic fibrosis induced NFkb inhibitors in a fluorescence assay	Primary screen	active
	42601321	686979 qHTS for inhibitors of human TDP1: qHTS in cells in absence of CPT	Confirmatory	EC ₅₀ 18.356
	42601322	602438 uHTS identification of modulators of interactions between CendR and NRP-1 using fluorescence polarization assay	Primary screen	active
	42601322	686979 qHTS for inhibitors of human TDP1: qHTS in cells in absence of CPT	Confirmatory	EC ₅₀ 20.596
	42601323	624466 Fluorescence-based cell-based primary HTS assay to identify antagonists of human TAAR1	Primary screen	active
	42601323	686978 qHTS for inhibitors of human TDP1: qHTS in cells in absence of CPT	Confirmatory	EC ₅₀ 16.360
	42601324	686979 qHTS for inhibitors of human TDP1: qHTS in cells in absence of CPT	Confirmatory	EC ₅₀ 16.360
	42601324	686978 qHTS of inhibitors of human TDP1: qHTS in cells in absence of CPT	Confirmatory	EC ₅₀ ^B 20.596
	463212	uHTS identification of small molecule inhibitors of tim23-1 yeast via a luminescent assay	Primary screen	active
	463218	Single concentration confirmation of small molecule inhibitors of tim23-1 yeast via a luminescent assay	Primary screen	active

^A IC₅₀= concentration of an inhibitor required for 50% inhibition of maximum control response

^B EC₅₀= concentration of an agonist required to produce 50% maximum (effective) response

^C AbsAC= Absolute active concentration with compounds below 10 μM to be considered active hits

^D AC₅₀= concentration required to elicit a 50% response in an *in vitro* assay

Calculations of Drug-like Properties

We sought to determine the potential drug-like qualities of these compounds by calculating parameters beyond the Lipinski's "rule of five."¹ Walters and coworkers previously analyzed over 415,000 molecules reported in the *Journal of Medicinal Chemistry* from 1959 to 2009.² In their investigation, they identified the eight important properties for all drugs or drug-like molecules to be molecular weight (MW), cLogP, total polar surface area (TPSA), rotatable bonds, hydrogen bond donors (HBD), hydrogen bond acceptors (HBA), complexity, and fraction of sp³ carbons (Fsp3). From this information, Hergenrother and coworkers established the importance for compounds to have higher Fsp3 values, from 0 to 1, and lower cLogP values, lower than 5 units, in order to be developed into drugs.³ The Fsp3 value pertains to the number of sp³-hybridized carbon atoms in a compound divided by the sum of carbon atoms, and this value has been associated with lower melting points as well as enhanced aqueous solubility.⁴ They calculated four main parameters: Fsp3, cLogP, number of stereocenters, and Tanimoto similarity coefficients of their compounds compared to a 150,000-member ChemBridge collection. In our case, we compared our library to the ChemBridge CombiSet and the Maybridge Diversity Set of compounds, which contain 30,000 and about 54,000 compounds, respectively. We expected this would offer insight into the diversity of our libraries relative to two commercially-available libraries.

We employed Pipeline Pilot to calculate ALogP values,⁵ the number of stereocenters, and the Fsp3 ratio.⁶ While our compounds have an average cLogP of 4.63 (Figure S1), the Hergenrother group reported an average cLogP of 2.90.³ The ChemBridge and MayBridge library sets that we chose for comparison had an average cLogP value of 3.11 and 3.04, respectively. With such a lipophilic core scaffold, we endeavored to create less lipophilic compounds by adding various substituents and hydrophilic groups that would lower cLogP values. Our Fsp3 calculations (Figure S2) yielded an average of 0.79, while the ChemBridge and MayBridge libraries have average Fsp3 values of 0.45 and 0.22, respectively.

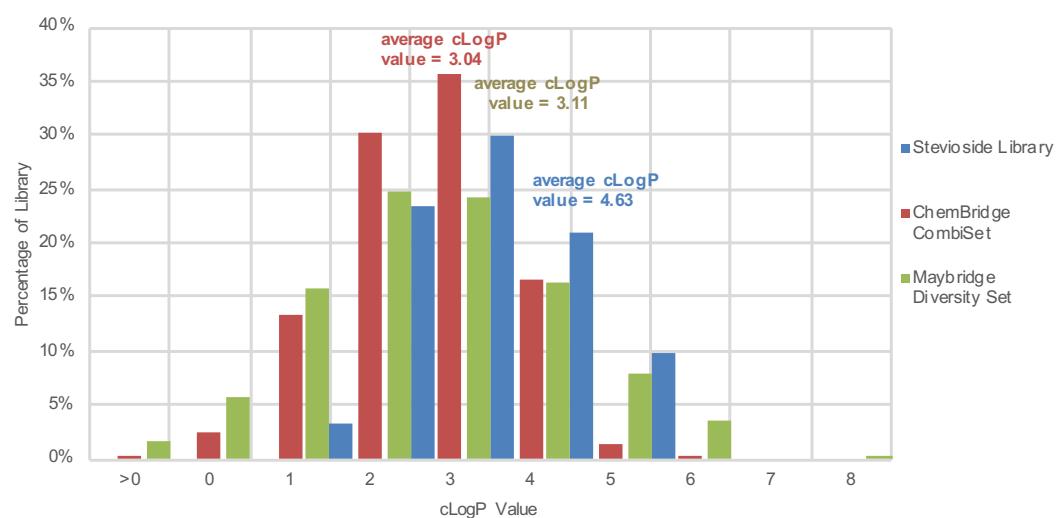


Figure S1. cLogP values of stevioside library vs commercial library sets.

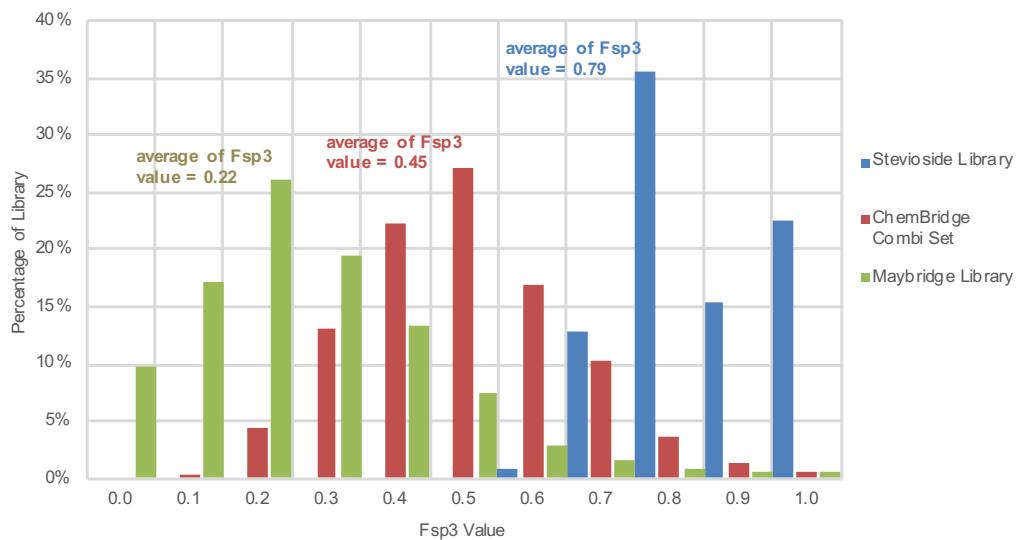


Figure S2. Fsp3 values of stevioside library vs commercial library sets.

The number of stereocenters in a compound can correlate to its structural complexity; on average, the structures in our compound library contained six stereocenters (Figure S3), while the ChemBridge and Maybridge libraries both have zero averages. Interestingly, Hergenrother's library of compounds had an average of 5.17 stereocenters.³ The majority of the compounds in the commercial libraries contain aromatic rings and aryl group substitutions and this largely accounts for their lack of stereocenters. Our compounds as well as those in the compound library of Hergenrother's group appear to be more structurally complex with a greater number of stereocenters because they are natural product analogs. ChemBridge and Maybridge compound libraries contain compounds without or few stereocenters, because these compounds are typically less complex and therefore, relatively easier to synthesize.⁷

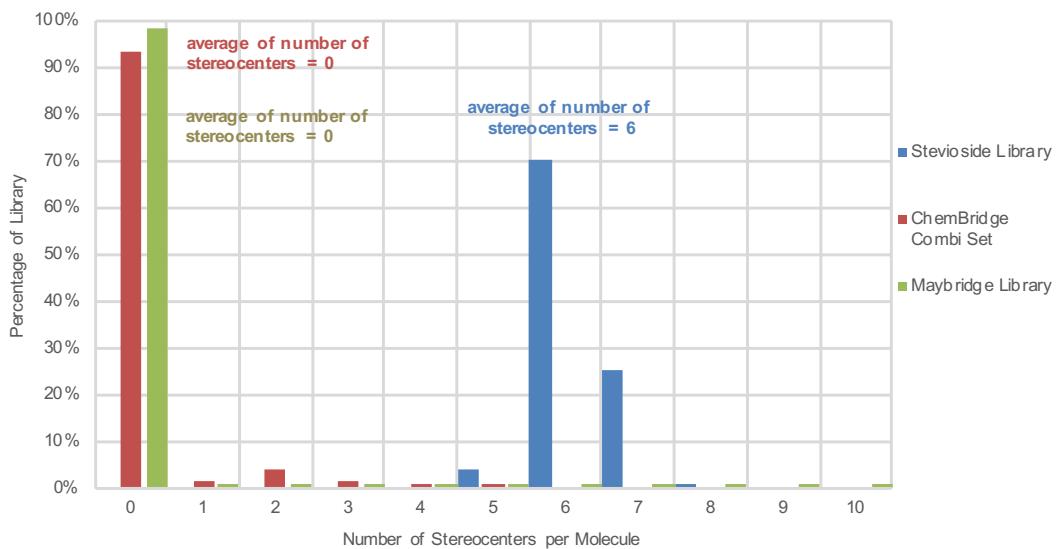


Figure S3. Number of stereocenters in stevioside library vs commercial library sets.

Furthermore, similarity coefficients were generated (Tanimoto, ECFP_4) to determine the structural similarity between the compounds in our libraries. These values are graphed in a matrix format in Chart S1 to allow visualization of the pairwise similarity of the compounds. That is an intersection of a column and a row contains the calculated similarity between the compounds in the row and column. Low scores (blue) indicate a relatively high level of difference between the structures, while high scores (red) indicate very similar ($0.700 < \text{ratio} < 1.00$) or the identical structure (coefficient = 1.00) when a compound is compared to itself. Low (blue) pairwise similarity is desired in a diverse library. The calculated average similarity between compounds in the library was 0.44. Over half, or 63%, of the complete steviol and isosteviol analog library compounds had similarities below 0.44, represented in Chart S1 as indicated by the blue cells in Chart S1. A detailed analysis is shown in Table S2.

Cmpd	24	25	11{1}	10{1}	13{1}	11{5}	10{5}	13{5}	19{2}	17{2}	15{2}	20{3}	20{5}	21{3}	21{5}	22{3}	22{5}	23{1}	26	27	8	5	7	2	1
24	1.000	0.792	0.456	0.406	0.389	0.403	0.346	0.333	0.677	0.329	0.262	0.444	0.463	0.472	0.507	0.438	0.456	0.384	0.737	0.567	0.525	0.576	0.420	0.443	0.525
25	0.792	1.000	0.380	0.455	0.394	0.338	0.405	0.354	0.561	0.333	0.265	0.451	0.470	0.500	0.538	0.444	0.463	0.389	0.750	0.576	0.534	0.614	0.426	0.526	0.435
11{1}	0.456	0.380	1.000	0.695	0.661	0.657	0.440	0.423	0.427	0.425	0.346	0.321	0.329	0.346	0.368	0.333	0.342	0.651	0.378	0.406	0.348	0.391	0.295	0.444	0.691
10{1}	0.406	0.455	0.695	1.000	0.712	0.453	0.631	0.453	0.382	0.437	0.354	0.329	0.338	0.372	0.397	0.342	0.351	0.700	0.370	0.462	0.358	0.424	0.303	0.648	0.444
13{1}	0.389	0.394	0.661	0.712	1.000	0.436	0.453	0.647	0.367	0.419	0.341	0.317	0.325	0.358	0.382	0.329	0.338	0.667	0.355	0.380	0.343	0.644	0.291	0.438	0.424
11{5}	0.403	0.338	0.657	0.453	0.436	1.000	0.721	0.690	0.450	0.487	0.405	0.318	0.294	0.341	0.329	0.345	0.306	0.430	0.337	0.359	0.308	0.346	0.279	0.389	0.594
10{5}	0.346	0.405	0.440	0.631	0.453	0.721	1.000	0.746	0.395	0.507	0.420	0.329	0.305	0.369	0.358	0.357	0.317	0.447	0.333	0.411	0.320	0.378	0.289	0.565	0.375
13{5}	0.333	0.354	0.423	0.453	0.647	0.690	0.746	1.000	0.381	0.487	0.405	0.318	0.294	0.356	0.345	0.345	0.306	0.430	0.321	0.342	0.308	0.567	0.279	0.389	0.360
19{2}	0.677	0.561	0.427	0.382	0.367	0.450	0.395	0.381	1.000	0.434	0.357	0.514	0.432	0.500	0.473	0.468	0.427	0.363	0.529	0.500	0.463	0.507	0.413	0.391	0.463
17{2}	0.329	0.333	0.425	0.437	0.419	0.487	0.507	0.487	0.434	1.000	0.521	0.453	0.411	0.354	0.325	0.507	0.425	0.413	0.316	0.338	0.583	0.342	0.288	0.368	0.357
15{2}	0.262	0.265	0.346	0.354	0.341	0.405	0.420	0.405	0.357	0.521	1.000	0.281	0.241	0.289	0.261	0.322	0.253	0.337	0.253	0.268	0.250	0.272	0.662	0.289	0.282
20{3}	0.444	0.451	0.321	0.329	0.317	0.318	0.329	0.318	0.514	0.453	0.281	1.000	0.656	0.583	0.434	0.712	0.529	0.313	0.427	0.457	0.581	0.464	0.395	0.352	0.342
20{5}	0.463	0.470	0.329	0.338	0.325	0.294	0.305	0.294	0.432	0.411	0.241	0.656	1.000	0.421	0.537	0.529	0.683	0.321	0.443	0.477	0.643	0.484	0.389	0.364	0.353
21{3}	0.472	0.500	0.346	0.372	0.358	0.341	0.369	0.356	0.500	0.354	0.289	0.583	0.421	1.000	0.682	0.575	0.416	0.461	0.453	0.486	0.449	0.515	0.403	0.400	0.370
21{5}	0.507	0.538	0.368	0.397	0.382	0.329	0.358	0.345	0.473	0.325	0.261	0.434	0.537	0.682	1.000	0.429	0.529	0.493	0.486	0.523	0.484	0.556	0.411	0.431	0.397
22{3}	0.438	0.444	0.333	0.342	0.329	0.345	0.357	0.345	0.468	0.507	0.322	0.712	0.529	0.575	0.429	1.000	0.662	0.325	0.421	0.451	0.571	0.457	0.390	0.347	0.338
22{5}	0.456	0.463	0.342	0.351	0.338	0.306	0.317	0.306	0.427	0.425	0.253	0.529	0.683	0.416	0.529	0.662	1.000	0.333	0.437	0.470	0.632	0.477	0.384	0.358	0.348
23{1}	0.384	0.389	0.651	0.700	0.667	0.430	0.447	0.430	0.363	0.413	0.337	0.313	0.321	0.461	0.493	0.325	0.333	1.000	0.351	0.375	0.338	0.400	0.288	0.431	0.418
26	0.737	0.750	0.378	0.370	0.355	0.337	0.333	0.321	0.529	0.316	0.253	0.427	0.443	0.453	0.486	0.421	0.437	0.351	1.000	0.732	0.500	0.548	0.403	0.444	0.431
27	0.567	0.576	0.409	0.462	0.380	0.359	0.411	0.342	0.500	0.338	0.268	0.457	0.477	0.486	0.523	0.451	0.470	0.375	0.732	1.000	0.544	0.596	0.433	0.564	0.467
8	0.525	0.534	0.348	0.358	0.343	0.308	0.320	0.308	0.463	0.583	0.250	0.581	0.643	0.449	0.484	0.571	0.632	0.338	0.500	0.544	1.000	0.554	0.415	0.414	0.400
5	0.576	0.614	0.391	0.424	0.644	0.346	0.378	0.567	0.507	0.342	0.272	0.464	0.484	0.515	0.556	0.457	0.477	0.400	0.548	0.596	0.554	1.000	0.439	0.491	0.450
7	0.420	0.426	0.295	0.303	0.291	0.279	0.289	0.279	0.413	0.288	0.662	0.395	0.389	0.403	0.411	0.390	0.384	0.288	0.403	0.433	0.415	0.439	1.000	0.324	0.314
2	0.443	0.526	0.444	0.648	0.438	0.389	0.565	0.389	0.391	0.368	0.289	0.352	0.364	0.400	0.431	0.347	0.358	0.431	0.444	0.564	0.414	0.491	0.324	1.000	0.640
1	0.525	0.435	0.691	0.444	0.424	0.594	0.375	0.360	0.463	0.357	0.282	0.342	0.353	0.370	0.397	0.338	0.348	0.418	0.431	0.467	0.400	0.450	0.314	0.640	1.000

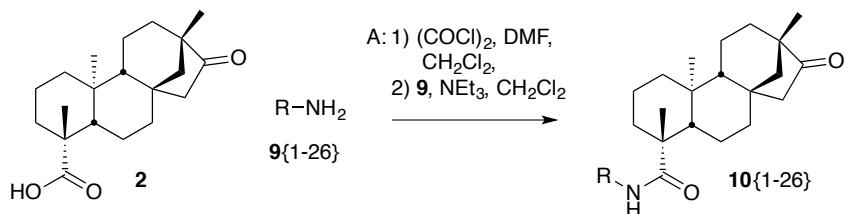
Chart S1. A representative chart of the pairwise similarity for the steviol and isosteviol library compounds described herein. Red indicates similarity coefficient of 1.0 (identical) and Blue indicates pairwise structural comparison with similarities less than the average pairwise similarity for the entire library.

Table S2. Similarity coefficients for Stevioside compound libraries

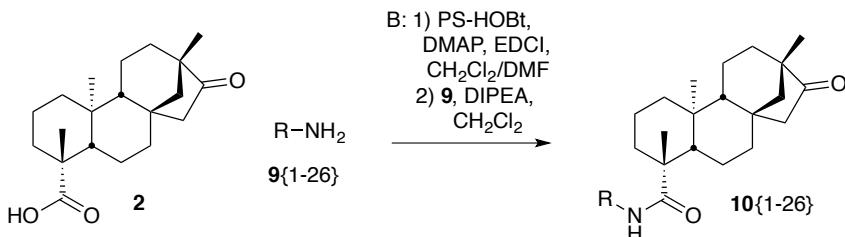
Experimental Details and Compound Characterization.

General Methods. Unless otherwise noted, all reactions were carried out open to air with reagent grade solvents. Purification of reaction products was carried out by flash column chromatography using silica gel 60 (230–400 mesh). Analytical thin layer chromatography (TLC) was performed on 0.25 mm silica gel 60-F plates. Visualization was accomplished with UV light or cerium molybdate stain followed by heating. High-resolution mass spectral data were acquired utilizing the electrospray ionization technique. ¹H nuclear magnetic resonance (NMR) spectra were recorded at ambient temperature at 400 MHz and are reported in ppm using a solvent as an internal standard (CDCl_3 at 7.26 ppm). Proton-decoupled ¹³C NMR spectra were recorded at 100 MHz and are reported in ppm using a solvent as an internal standard (CDCl_3 at 77.16 ppm). The data are reported as follows: chemical shift on the δ scale in ppm, multiplicity (b = broad, s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constants (Hz), and integration. Solvent abbreviations: tetrahydrofuran (THF), dimethylformamide (DMF), Hex (hexanes), methyl tert.-butyl ether (MTBE). Steviol and steviol methyl ester data were in agreement with previous reports.¹

Syntheses of compounds **21{1}**, **21{2}**, and **24–27** were reported by us before.⁸

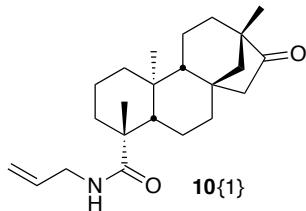


Standard procedure for the solution phase amide bond formation (Scheme 2, A). To an oven-dried reaction flask under nitrogen at 0 °C (ice bath), was added DMF (37 μL , 0.47 mmol, 1.5 equiv) in CH_2Cl_2 (1.0 mL) followed by oxalyl chloride (41 μL , 0.47 mmol, 1.5 equiv). The ice bath was removed and the mixture was allowed to stir at room temperature for 1 h. Then the reaction flask was cooled to 0 °C (ice bath) and isosteviol (**2**, 100 mg, 0.31 mmol) and Et_3N (130 μL , 0.47 mmol, 3 equiv) in CH_2Cl_2 (1.0 mL) were added. After stirring for 5 min, allylamine (**9{1}**, 1 mL, 13.4 mmol) was added and the mixture was stirred for 2 h. (For all reactions 1mL of the amine was added). Then the mixture was loaded onto a 20-mL silica gel plug in a 70-mL column and eluted with dry THF. The solvent was removed under vacuum, the residue was dissolved in CH_2Cl_2 (10 mL), and methylisocyanate polystyrene resin was added. The mixture was shaken for 20 h and was then loaded onto a 20-mL silica gel plug in a 70-mL column and eluted with ethyl acetate (40 mL). The solvent was removed and the residues was purified using column chromatography on silica gel with 10% EtOAc:Hex as the eluent to provide the allyl amide product **10{1}** in 47% yield.

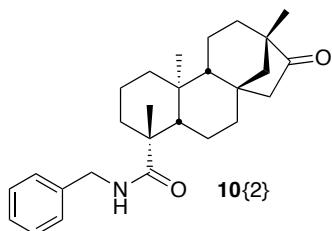


Standard procedure for solid phase amide bond formation (Scheme 2, B). To a flame-dry reaction vial of isosteviol (**2**, 100.0 mg, 0.318 mmol, 1.0 equiv) in $\text{CH}_2\text{Cl}_2/\text{DMF}$ (4:1) was added PS-hydroxybenzotriazole (PS-HOBt, 1.00 mmol/g, 0.67 equiv), 4-dimethylaminopyridine (15.5 mg, 0.127

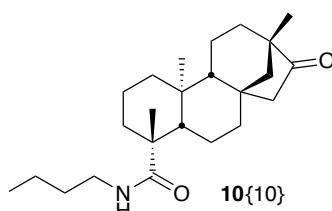
mmol, 0.40 equiv), and then 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (148 mg, 0.954 mmol, 3.0 equiv). The reaction mixture was shaken slowly for 24 h and then filtered. The resin was washed with MeOH (5 mL) and CH₂Cl₂ (5 mL) thrice to remove excess acid. Then, the resin was added to a solution of allylamine (**9{1}**, 15.5 μ L, 0.220 mmol, 0.70 equiv) and diisopropylethylamine (55.4 μ L, 0.318 mmol, 1.0 equiv) in CH₂Cl₂ and shaken slowly for another 24 h. The mixture was filtered again was washed with MeOH (5 mL) and CH₂Cl₂ (5 mL) thrice before the solvent was removed under vacuum. Column chromatography on silica gel with 20% EtOAc:Hex as the eluent gave amide product **10{1}** in 43 mg (38% yield).



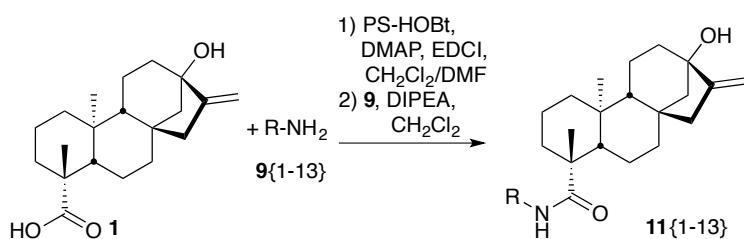
(4*R*,4*aS*,6*aR*,9*S*,11*aR*,11*bS*)-*N*-Allyl-4,9,11b-trimethyl-8-oxotetradecahydro-6*a*,9-methanocyclohepta[*a*]naphthalene-4-carboxamide (10{1}). Oil; ¹H NMR (400 MHz, CDCl₃): δ 5.83 – 5.70 (m, 1H), 5.62 (s, 1H), 5.09 (dd, J = 23.1, 13.7 Hz, 2H), 3.79 (t, J = 5.1 Hz, 2H), 2.58 (dd, J = 18.6, 3.6 Hz, 1H), 2.03 - 1.85 (m, 2H), 1.73 (d, J = 18.5 Hz, 3H), 1.66 – 1.58 (m, 2H), 1.55 – 1.41 (m, 4H), 1.43 – 1.33 (m, 3H), 1.21 – 1.13 (m, 2H), 1.13 (s, 3H), 1.10 – 1.02 (m, 1H), 0.91 (s, 3H), 0.87 (d, J = 4.5 Hz, 1H), 0.71 (s, 3H), 0.66 (d, J = 3.1 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃): δ 222.6, 176.4, 134.4, 116.6, 57.5, 57.2, 54.7, 54.2, 48.7, 48.4, 43.7, 41.9, 41.7, 40.2, 39.5, 38.1, 37.3, 30.2, 22.2, 20.3, 19.9, 19.2, 13.6; LRMS (ESI) (*m/z*): [M+H]⁺ calcd for C₂₃H₃₆NO₂ 358.2668, found 358; $[\alpha]_D^{23}$ –80.7 (*c* 0.950, CHCl₃).



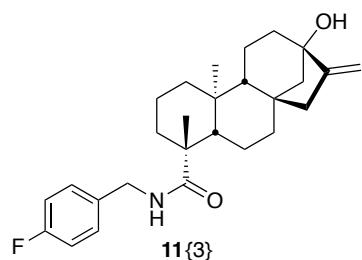
(4*R*,4*aS*,6*aR*,9*S*,11*aR*,11*bS*)-*N*-Benzyl-4,9,11b-trimethyl-8-oxotetradecahydro-6*a*,9-methanocyclohepta[*a*]naphthalene-4-carboxamide (10{2}). Pale yellow solid; mp 68–69 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.36 – 7.25 (m, 5H), 5.86 (t, J = 4.9 Hz, 1H), 4.40 (d, J = 5.4 Hz, 2H), 2.62 (dd, J = 18.6, 3.6 Hz, 1H), 2.06 – 1.90 (m, 2H), 1.83 – 1.63 (m, 6H), 1.59 – 1.32 (m, 7H), 1.27 – 1.22 (m, 1H), 1.21 (s, 3H), 1.18 – 1.12 (m, 2H), 0.96 (s, 3H), 0.95 – 0.79 (m, 1H), 0.74 (s, 3H). ¹³C NMR (125 MHz, CDCl₃): δ 222.4, 176.4, 138.5, 128.9, 128.7, 128.0, 127.5, 127.5, 57.6, 54.7, 54.3, 48.7, 48.4, 43.7, 41.7, 40.2, 39.5, 38.1, 38.1, 37.3, 30.2, 22.2, 20.3, 19.9, 19.2, 13.6; LRMS (ESI) (*m/z*): [M+Na]⁺ calcd for C₂₇H₃₇NO₂Na 430.2824, found 430.4; $[\alpha]_D^{23}$ –44.0 (*c* 1.00, CHCl₃).



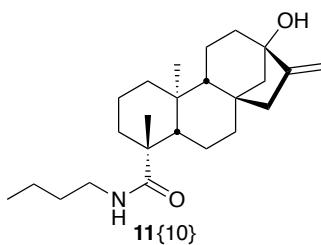
(4*R*,4*aS*,6*aR*,9*S*,11*aR*,11*bS*)-*N*-Butyl-4,9,11*b*-trimethyl-8-oxotetradecahydro-6*a*,9-methanocyclohepta[*a*]naphthalene-4-carboxamide (10{1}**)**. Oil; ^1H NMR (400 MHz, CDCl_3): δ 5.59 (s, 1H), 3.21 (dt, $J = 11.8, 6.7$ Hz, 2H), 2.65 (dd, $J = 18.6, 3.6$ Hz, 1H), 2.01 (d, $J = 14.3$ Hz, 1H), 1.95 (d, $J = 13.3$ Hz, 1H), 1.77 (dd, $J = 19.9, 14.1$ Hz, 4H), 1.71 – 1.65 (m, 2H), 1.63 – 1.57 (m, 1H), 1.56 – 1.52 (m, 1H), 1.51 – 1.45 (m, 4H), 1.40 (dd, $J = 11.8, 3.7$ Hz, 1H), 1.37 – 1.30 (m, 3H), 1.25 – 1.18 (m, 2H), 1.17 (s, 3H), 1.16 – 1.09 (m, 2H), 0.97 (s, 3H), 0.92 (t, $J = 7.2$ Hz, 4H), 0.77 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ 222.4, 176.5, 57.6, 54.8, 54.3, 48.7, 48.4, 43.6, 41.8, 40.2, 39.5, 39.2, 38.2, 38.1, 37.3, 31.5, 30.2, 22.3, 20.3, 20.3, 19.9, 19.2, 13.8, 13.5; LRMS (ESI) (m/z): [M+H] $^+$ calcd for $\text{C}_{24}\text{H}_{40}\text{NO}_2$ 374.2981, found 374.3; $[\alpha]_D^{23} -32.2$ (c 0.500, CHCl_3).



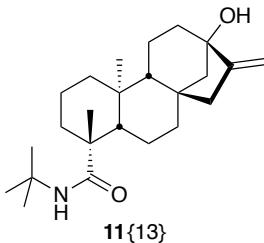
Standard procedure for solid-phase bond formation of steviol amide analogs **11{1-13}** (Scheme 3). To a solution of steviol (**1**, 100 mg, 0.318 mmol, 1.00 equiv) in $\text{CH}_2\text{Cl}_2/\text{DMF}$ (4:1) in a flame-dry reaction vial, was added PS-hydroxybenzotriazole (PS-HOBt, 1.00 mmol/g, 0.213 mmol, 0.670 equiv), 4-dimethylaminopyridine (55.4 μL , 0.318 mmol, 0.40 equiv), and then 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (148 mg, 0.954 mmol, 3.0 equiv). The reaction mixture was shaken slowly for 24 h and then filtered. The resins were washed with MeOH (5 mL) and CH_2Cl_2 (5 mL) thrice to remove excess acid. Then, the resin was added to a solution of 4-fluorobenzylamine (**9{3}**, 27.5 mg, 0.220 mmol, 0.70 equiv) and diisopropylethylamine (55.4 μL , 0.318 mmol, 1.0 equiv) in CH_2Cl_2 and shaken slowly for another 24 h. The mixture was filtered again was washed with MeOH (5 mL) and CH_2Cl_2 (5 mL) thrice before the solvent was removed under vacuum. The crude residue was purified by column chromatography on silica gel with 20% EtOAc:Hex to furnish the amide **11{3}**.



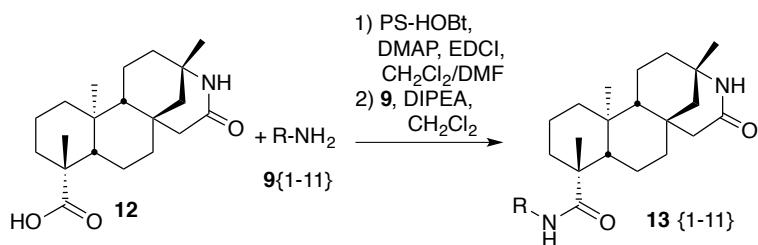
(4*R*,4*aS*,6*aR*,9*S*,11*aR*,11*bS*)-*N*-(4-Fluorobenzyl)-9-hydroxy-4,11*b*-dimethyl-8-methylenetetradecahydro-6*a*,9-methanocyclohepta[*a*]naphthalene-4-carboxamide (11{3}**)**. Oil; ^1H NMR (400 MHz, CDCl_3): δ 8.27 (s, 1H), 7.70 (dd, $J = 8.5, 5.7$ Hz, 2H), 7.03 (t, $J = 8.6$ Hz, 2H), 4.82 (d, $J = 67.5$ Hz, 2H), 4.69 (s, 2H), 2.13 – 1.93 (m, 4H), 1.88 – 1.63 (m, 6H), 1.58 – 1.22 (m, 5H), 1.20 – 1.16 (m, 1H), 1.11 (s, 3H), 1.08 – 0.97 (m, 3H), 0.88 (d, $J = 7.9$ Hz, 1H), 0.79 (s, 3H), 0.75 (dd, $J = 12.7, 3.7$ Hz, 1H). ^{13}C NMR (125 MHz, CDCl_3): δ 176.6, 160.6, 156.0, 130.1, 129.6, 129.4, 115.6, 115.4, 103.0, 80.2, 64.2, 57.4, 53.7, 47.4, 46.9, 43.8, 42.9, 41.7, 40.9, 39.4, 38.2, 30.1, 23.0, 22.4, 20.4, 19.3, 15.7; LRMS (ESI) (m/z): [M+H] $^+$ calcd for $\text{C}_{27}\text{H}_{37}\text{FNO}_2$ 426.2730, found 426; $[\alpha]_D^{23} -84.0$ (c 0.451, CHCl_3).



(4*R*,4*aS*,6*aR*,9*S*,11*aR*,11*bS*)-*N*-Butyl-9-hydroxy-4,11*b*-dimethyl-8-methylenetetradecahydro-6*a*,9-methanocyclohepta[*a*]naphthalene-4-carboxamide (11{10}). Oil; ^1H NMR (400 MHz, CDCl_3): δ 7.99 (s, 1H), 5.58 (s, 1H), 4.88 (d, $J = 68.1$ Hz, 2H), 3.20 (dt, $J = 12.2, 6.8$ Hz, 2H), 2.90 (d, $J = 30.2$ Hz, 1H), 2.18 (d, $J = 17.0$ Hz, 1H), 2.14 – 2.09 (m, 1H), 2.07 – 2.00 (m, 2H), 1.89 (dd, $J = 21.7, 5.2$ Hz, 3H), 1.82 – 1.71 (m, 3H), 1.63 – 1.51 (m, 2H), 1.51 – 1.43 (m, 4H), 1.42 – 1.30 (m, 3H), 1.26 (d, $J = 10.8$ Hz, 1H), 1.13 (s, 3H), 1.10 – 0.99 (m, 2H), 0.94 – 0.90 (m, 3H), 0.89 (s, 3H), 0.86 – 0.77 (m, 1H); ^{13}C NMR (125 MHz, CDCl_3): δ 176.6, 156.1, 102.9, 80.2, 57.3, 53.8, 47.4, 46.9, 43.7, 41.7, 41.6, 41.1, 39.4, 39.3, 39.2, 38.3, 31.5, 30.1, 22.4, 20.5, 20.3, 19.4, 15.6, 13.8; LRMS (m/z): [M+H] $^+$ calcd for $\text{C}_{24}\text{H}_{40}\text{NO}_2$ 374.2981, found 374.3; $[\alpha]_D^{23} -77.5$ (c 1.00, CHCl_3).

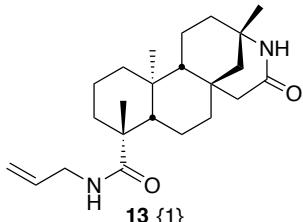


(4*R*,4*aS*,6*aR*,9*S*,11*aR*,11*bS*)-*N*-(tert-Butyl)-9-hydroxy-4,11*b*-dimethyl-8-methylenetetradecahydro-6*a*,9-methanocyclohepta[*a*]naphthalene-4-carboxamide (11{13}). Oil; ^1H NMR (400 MHz, CDCl_3): δ 5.34 (s, 1H), 4.88 (d, $J = 66.6$ Hz, 2H), 2.94 (s, 1H), 2.18 (d, $J = 19.5$ Hz, 1H), 2.10 (d, $J = 17.2$ Hz, 1H), 1.99 (d, $J = 13.6$ Hz, 1H), 1.91 – 1.84 (m, 4H), 1.82 – 1.73 (m, 4H), 1.60 – 1.50 (m, 2H), 1.49 – 1.37 (m, 3H), 1.33 (s, 3H), 1.31 (s, 6H), 1.27 (dd, $J = 14.7, 3.6$ Hz, 1H), 1.12 (s, 3H), 1.08 – 0.98 (m, 2H), 0.95 (s, 3H), 0.82 (d, $J = 4.0$ Hz, 1H); ^{13}C NMR (125 MHz, CDCl_3): δ 175.7, 156.1, 102.9, 80.2, 57.2, 53.8, 50.7, 47.4, 46.9, 44.1, 41.7, 41.6, 41.2, 39.5, 39.3, 38.5, 30.2, 28.8, 28.7, 28.7, 22.5, 20.5, 19.3, 15.9; LRMS (ESI) (m/z): [M+H] $^+$ calcd for $\text{C}_{24}\text{H}_{40}\text{NO}_2$ 374.2981, found 374.3; $[\alpha]_D^{23} -102.1$ (c 0.750, CHCl_3).

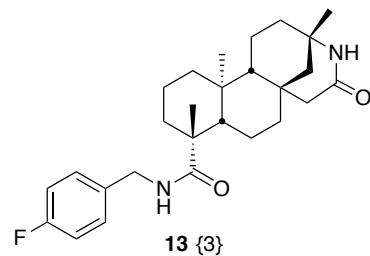


Standard procedure for solid-phase bond formation of isosteviol lactam **13{1-11}** from previously reported **12**² (Scheme 3). To a solution of lactam **12** (100 mg, 0.333 mmol, 1.00 equiv) in $\text{CH}_2\text{Cl}_2/\text{DMF}$ (4:1) in a flame-dry reaction vial, was added PS-hydroxybenzotriazole (PS-HOBt, 1.00 mmol/g, 0.223 mmol, 0.670 equiv), 4-dimethylaminopyridine (58.0 μL , 0.333 mmol, 0.40 equiv), and then 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (155 mg, 1.00 mmol, 3.0 equiv). The reaction mixture was shaken

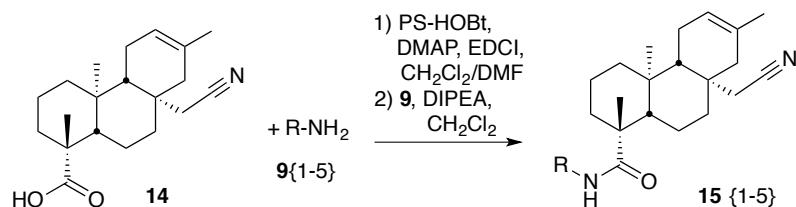
slowly for 24 h and then filtered. The resins were washed with MeOH (5 mL) and CH₂Cl₂ (5 mL) thrice to remove excess acid. Then, the resin was added to a solution of allylamine (**9{1}**, 29.1 mg, 0.220 mmol, 0.70 equiv) and diisopropylethylamine (58.0 μ L, 0.333 mmol, 1.0 equiv) in CH₂Cl₂ and shaken slowly for another 24 h. The mixture was filtered again was washed with MeOH (5 mL) and CH₂Cl₂ (5 mL) thrice before the solvent was removed under vacuum. The crude residue was purified by column chromatography on silica gel with 20% EtOAc:Hex to furnish the amide **13{1}**.



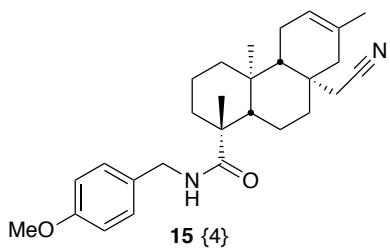
(3*S*,6*aR*,8*a**S*,9*R*,12*a**S*,12*b**R*)-*N*-Allyl-3,9,12*a*-trimethyl-5-oxotetradecahydro-2*H*-3,6*a*-methanonaphtho[2,1-*d*]azocine-9-carboxamide (**13{1}**)**. Oil; ¹H NMR (400 MHz, CDCl₃): δ 5.88 – 5.80 (m, 1H), 5.66 (s, 1H), 5.22 – 5.13 (m, 2H), 3.92 – 3.79 (m, 2H), 2.92 (d, *J* = 29.2 Hz, 1H), 2.52 (ddd, *J* = 52.5, 16.6, 2.1 Hz, 1H), 2.20 – 1.70 (m, 8H), 1.59 (s, 6H), 1.53 – 1.23 (m, 4H), 1.20 (s, 3H), 1.19 – 1.08 (m, 3H), 0.90 (dd, *J* = 13.6, 3.8 Hz, 1H), 0.80 – 0.71 (m, 3H). ¹³C NMR (125 MHz, CDCl₃): δ 176.3, 134.5, 131.4, 120.0, 116.8, 57.9, 52.0, 45.9, 43.8, 42.1, 40.3, 39.5, 38.1, 37.6, 35.4, 33.2, 30.2, 23.4, 22.4, 20.5, 20.1, 19.4, 14.0; HRMS (ESI) (*m/z*): [M+Na]⁺ calcd for C₂₃H₃₆O₂N₂Na 395.2777, found 395.2680; $[\alpha]_D^{23}$ –10.3 (*c* 1.20, CHCl₃).



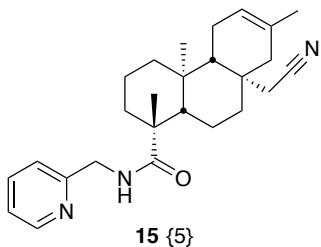
(3*S*,6*aR*,8*a**S*,9*R*,12*a**S*,12*b**R*)-*N*-(4-Fluorobenzyl)-3,9,12*a*-trimethyl-5-oxotetradecahydro-2*H*-3,6*a*-methanonaphtho[2,1-*d*]azocine-9-carboxamide (**13{3}**)**. Oil; ¹H NMR (400 MHz, CDCl₃): δ 7.33 – 7.29 (m, 1H), 7.25 – 7.22 (m, 1H), 7.03 – 6.97 (m, 2H), 6.51 (t, *J* = 6.0 Hz, 1H), 4.56 (dd, *J* = 15.0, 6.5 Hz, 1H), 4.49 – 4.33 (m, 2H), 3.10 (d, *J* = 17.6 Hz, 1H), 2.40 (d, *J* = 12.1 Hz, 1H), 2.31 (d, *J* = 17.8 Hz, 1H), 2.17 (d, *J* = 13.4 Hz, 1H), 1.92 – 1.83 (m, 3H), 1.78 (d, *J* = 13.2 Hz, 1H), 1.73 – 1.67 (m, 2H), 1.64 – 1.56 (m, 2H), 1.51 – 1.42 (m, 1H), 1.39 (s, 3H), 1.36 – 1.28 (m, 2H), 1.25 (s, 3H), 1.22 – 1.15 (m, 1H), 1.13 – 1.04 (m, 2H), 1.02 – 0.93 (m, 2H), 0.86 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 183.0, 176.1, 171.0, 161.1, 129.5, 129.5, 115.7, 115.5, 77.3, 77.0, 76.7, 59.5, 57.3, 56.6, 51.2, 43.7, 43.6, 42.8, 42.0, 39.8, 38.0, 37.7, 37.5, 35.0, 28.8, 26.1, 19.5, 19.1, 18.9, 13.8; LRMS (ESI) (*m/z*): [M+H]⁺ calcd for C₂₇H₃₈FN₂O₂ 441.2339, found 441; $[\alpha]_D^{23}$ 19.2 (*c* 1.00, CHCl₃).



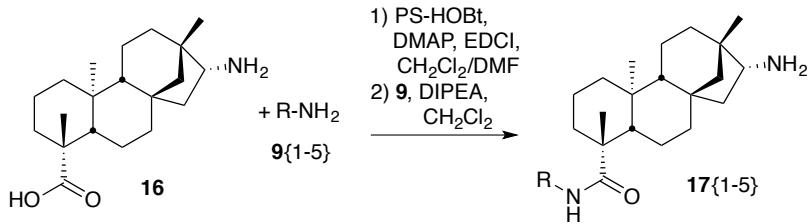
Standard procedure for amide bond formation of **15{1-5}** from **14**. To a flame-dried reaction flask of the nitrile acid **14** (1.0 equiv) in $\text{CH}_2\text{Cl}_2/\text{DMF}$ (4:1) in a flame-dry reaction vial, was added PS-hydroxybenzotriazole (PS-HOBt, 1.00 mmol/g, 0.67 equiv), 4-dimethylaminopyridine (55.4 μL , 0.318 mmol, 0.40 equiv), and then 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (148 mg, 0.954 mmol, 3.0 equiv). The reaction mixture was shaken slowly for 24 h and then filtered. The resins were washed with MeOH (5 mL) and CH_2Cl_2 (5 mL) thrice to remove excess acid. Then, the resin was added to a solution of amine **9** (0.220 mmol, 0.70 equiv) and diisopropylethylamine (55.4 μL , 0.318 mmol, 1.0 equiv) in CH_2Cl_2 and shaken slowly for another 24 h. The mixture was filtered again was washed with MeOH (5 mL) and CH_2Cl_2 (5 mL) thrice before the solvent was removed under vacuum. The crude residue was purified by column chromatography on silica gel with 20% EtOAc:Hex to furnish the amide **15**.



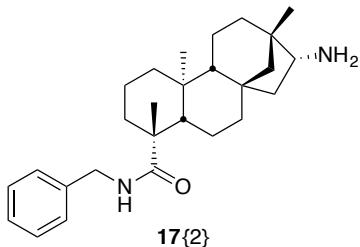
(1*R*,4*aS*,4*bR*,8*aR*,10*aS*)-*N*-Benzyl-8*a*-(cyanomethyl)-1,4*a*,7-trimethyl-1,2,3,4,4*a*,4*b*,5,8,8*a*,9,10,10*a*-dodecahydronaphthalene-1-carboxamide (15{4}). Pale yellow crystals; mp 63 – 64 °C; ^1H NMR (400 MHz, CDCl_3): δ 7.21 (d, J = 8.4 Hz, 2H), 6.86 (d, J = 8.4 Hz, 2H), 5.69 (s, 1H), 4.36 (d, J = 5.6 Hz, 2H), 3.80 (s, 3H), 2.98 (dd, J = 19.0, 2.5 Hz, 1H), 2.17 (d, J = 13.1 Hz, 1H), 2.01 (s, 3H), 2.05 – 1.96 (m, 1H), 1.90 – 1.66 (m, 4H), 1.61 (d, J = 13.6 Hz, 2H), 1.51 – 1.38 (m, 4H), 1.28 (s, 1H), 1.23 (s, 3H), 1.09 (s, 3H), 1.01 (td, J = 13.5, 3.8 Hz, 1H), 0.93 – 0.87 (m, 1H), 0.85 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ 183.0, 170.6, 170.0, 159.2, 130.4, 129.4, 129.4, 114.2, 114.2, 77.5, 77.2, 76.8, 57.3, 56.4, 55.5, 55.1, 44.0, 43.7, 43.4, 40.8, 40.1, 39.6, 38.4, 37.2, 29.2, 23.4, 22.3, 21.7, 20.6, 19.1, 13.6; LRMS (ESI) (m/z): [M+Na] $^+$ calcd for $\text{C}_{28}\text{H}_{38}\text{N}_2\text{O}_2$ 434.2933, found 457.3; $[\alpha]_D^{23}$ 3.60 (c 0.600, CHCl_3).



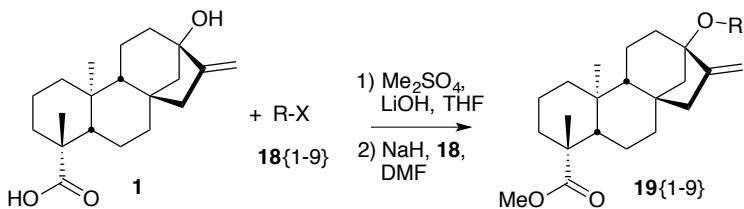
(1*R*,4*aS*,4*bR*,10*aS*)-8*a*-(Cyanomethyl)-1,4*a*,7-trimethyl-*N*-(pyridin-2-ylmethyl)-1,2,3,4,4*a*,4*b*,5,8,8*a*,9,10,10*a*-dodecahydronaphthalene-1-carboxamide (15{5}). Oil; ^1H NMR (400 MHz, CDCl_3): δ 8.57 (d, J = 4.5 Hz, 1H), 7.67 (t, J = 7.6 Hz, 1H), 7.27 (d, J = 7.9 Hz, 1H), 7.25 – 7.17 (m, 1H), 7.11 (s, 1H), 5.35 (s, 1H), 5.31 (s, 1H), 4.53 (tdd, J = 16.5, 11.7, 4.7 Hz, 2H), 2.78 – 2.53 (m, 1H), 2.49 – 2.29 (m, 1H), 2.26 – 2.12 (m, 2H), 2.12 – 1.92 (m, 5H), 1.87 – 1.77 (m, 1H), 1.75 – 1.69 (m, 1H), 1.66 (s, 3H), 1.58 – 1.49 (m, 1H), 1.35 – 1.27 (m, 1H), 1.23 (s, 3H), 1.22 – 1.12 (m, 3H), 0.92 (dt, J = 16.0, 8.2 Hz, 1H), 0.65 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ 176.6, 156.4, 149.0, 136.7, 131.2, 122.4, 122.3, 119.9, 119.0, 57.7, 51.7, 45.7, 44.4, 43.7, 40.2, 39.4, 37.9, 37.4, 35.3, 29.9, 23.3, 22.2, 20.2, 19.9, 19.1, 13.5; LRMS (m/z): [M+Na] $^+$ calcd for $\text{C}_{26}\text{H}_{35}\text{N}_3\text{O}_2\text{Na}$ 428.2780, found 428.3; $[\alpha]_D^{23}$ –34.0 (c 1.00, CHCl_3).



Standard procedure for the amide bond formation for amine **17{1-5}** from previously reported **16**.⁹ To a flame-dried reaction flask of the amine **16** (1.0 equiv) in CH₂Cl₂/DMF (4:1) in a flame-dry reaction vial, was added PS-hydroxybenzotriazole (PS-HOBt, 1.00 mmol/g, 0.67 equiv), 4-dimethylaminopyridine (55.4 μL, 0.318 mmol, 0.40 equiv), and then 1-ethyl-3-(3-dimethylaminopropyl) carbodiimide (148 mg, 0.954 mmol, 3.0 equiv). The reaction mixture was shaken slowly for 24 h and then filtered. The resins were washed with MeOH (5 mL) and CH₂Cl₂ (5 mL) thrice to remove excess acid. Then, the resin was added to a solution of amine **9** (0.220 mmol, 0.70 equiv) and diisopropylethylamine (55.4 μL, 0.318 mmol, 1.0 equiv) in CH₂Cl₂ and shaken slowly for another 24 h. The mixture was filtered again was washed with MeOH (5 mL) and CH₂Cl₂ (5 mL) thrice before the solvent was removed under vacuum. The crude residue was purified by column chromatography on silica gel with 5% MeOH: CH₂Cl₂ to furnish the amide **17**.

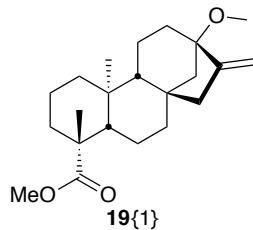


(4*R*,4*aS*,6*aR*,8*R*,9*S*,11*aR*,11*bS*)-8-Amino-*N*-benzyl-4,9,11*b*-trimethyltetradecahydro-6*a*,9-methanocyclohepta[*a*]naphthalene-4-carboxamide (17{2}). Oil; ¹H NMR (400 MHz, CDCl₃): δ 7.96 – 7.83 (m, 1H), 7.36 (d, *J* = 6.8 Hz, 2H), 7.34 – 7.30 (m, 3H), 4.53 (d, *J* = 6.2 Hz, 2H), 2.18 (d, *J* = 13.9 Hz, 1H), 1.94 – 1.78 (m, 4H), 1.76 – 1.71 (m, 3H), 1.69 – 1.58 (m, 3H), 1.49 – 1.41 (m, 3H), 1.40 – 1.33 (m, 2H), 1.30 – 1.26 (m, 1H), 1.25 (s, 3H), 1.17 – 1.12 (m, 1H), 1.11 – 1.01 (m, 3H), 0.98 (t, *J* = 7.4 Hz, 1H), 0.93 (s, 3H), 0.92 – 0.85 (m, 1H), 0.82 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 182.3, 159.9, 136.9, 128.8, 128.8, 127.9, 127.9, 57.6, 56.9, 56.0, 55.6, 43.8, 43.6, 42.6, 42.0, 41.3, 40.5, 39.9, 38.2, 37.8, 33.9, 29.0, 24.8, 21.6, 20.7, 18.8, 13.5; LRMS (*m/z*): [M+H]⁺ calcd for C₂₇H₄₀N₂O 408.3141, found 409; [α]_D²³ –5.25 (*c* 0.750, CHCl₃).

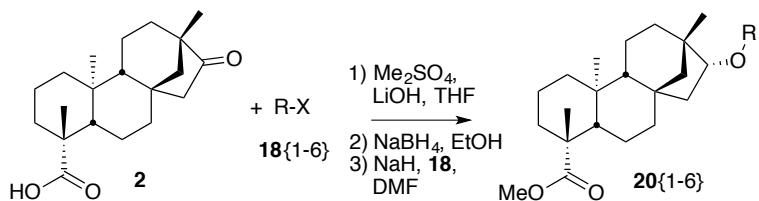


Steviol Methyl Ester.³ To a solution of steviol (**1**, 10 g, 32 mmol) in THF (50 mL) in a 500-mL round-bottomed flask at room temperature was added LiOH•H₂O (1.5 g, 34 mmol). The reaction stirred at room temperature for 1 h under nitrogen, then Me₂SO₄ (3.3 mL, 35 mmol) was added slowly, and the reaction vessel was fitted with a reflux condenser. The reaction was refluxed for 18 h at 80 °C. Once the reaction cooled to room temperature, the off-white precipitate was collected via filtration to furnish steviol methyl

ester (7.6 g, 73%) as off-white crystals. mp 117–118 °C (Lit 113–115 °C); ^1H NMR (400 MHz, CDCl_3): δ 4.92 (d, $J = 64.0$ Hz, 2H), 3.66 (s, 3H), 2.28 – 2.04 (m, 4H), 1.93 – 1.73 (m, 7H), 1.62 – 1.40 (m, 5H), 1.33 – 1.26 (m, 1H), 1.19 (s, 3H), 1.10 – 0.93 (m, 3H), 0.85 (s, 3H), 0.82 – 0.75 (m, 1H); ^{13}C NMR (100 MHz, CDCl_3): δ 178.0, 156.0, 102.9, 80.3, 56.9, 53.8, 51.2, 47.4, 46.9, 43.8, 41.6, 41.3, 40.7, 39.4, 39.2, 38.0, 28.7, 21.9, 20.4, 19.1, 15.3. HRMS (ESI) (m/z): $[\text{M}+\text{Na}]^+$ calcd for $\text{C}_{21}\text{H}_{32}\text{O}_3\text{Na}$ 355.2249, found 355.2080; $[\alpha]_D^{23} -191.3$ (c 1.00, CHCl_3).



Methyl (4*R*,4*aS*,6*aR*,9*S*,11*aR*,11*bS*)-9-Methoxy-4,11*b*-dimethyl-8-methylenetetradecahydro-6*a*,9-methanocyclohepta[*a*]naphthalene-4-carboxylate (19{1}). To a solution of steviol methyl ester (100 mg, 0.318 mmol, 1.0 equiv) in DMF (5 mL) in a flame-dry reaction vial was added slowly via cannula a solution of sodium hydride (38.3 mg in 60% dispersion in oil, 0.954 mmol, 3.0 equiv) in dry DMF (5 mL). Then, methyl iodide (90.3 mg, 0.636 mmol, 2.0 equiv) was added to the reaction flask and the mixture was allowed to stir for 2 h at ambient temperature. The reaction was quenched with MTBE and the organic phase was washed successively with water. The aqueous phase was extracted with MTBE and the combined organic layers were dried over MgSO_4 , filtered, and concentrated *in vacuo*. Purification by flash column chromatography on silica gel with 20% EtOAc:Hex yielded the product **19**. Oil; ^1H NMR (400 MHz, CDCl_3): δ 4.87 (d, $J = 6.1$ Hz, 2H), 3.64 (s, 3H), 3.22 (s, 3H), 2.18 (d, $J = 13.2$ Hz, 1H), 2.14 – 2.00 (m, 2H), 1.89 – 1.80 (m, 4H), 1.79 – 1.71 (m, 3H), 1.69 – 1.58 (m, 1H), 1.57 – 1.49 (m, 2H), 1.48 – 1.40 (m, 3H), 1.17 (s, 3H), 1.07 – 0.94 (m, 3H), 0.82 (s, 3H), 0.85 – 0.76 (m, 1H); ^{13}C NMR (125 MHz, CDCl_3): δ 178.0, 151.0, 103.7, 85.2, 57.0, 54.0, 51.2, 50.1, 48.1, 43.8, 41.7, 41.5, 40.7, 40.0, 39.3, 38.8, 38.0, 28.7, 21.9, 20.2, 19.1, 15.3, LRMS (ES) (m/z): $[\text{M}+\text{CH}_2\text{O}_2]^+$ calcd for $\text{C}_{22}\text{H}_{34}\text{O}_3$ 390.2508, found 390.3; $[\alpha]_D^{23} -93.5$ (c 0.800, CHCl_3).

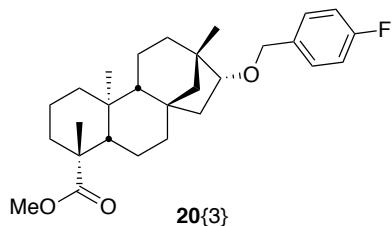


Isosteviol methyl ester.⁸ To a flame-dried 500 mL round-bottomed flask was added isosteviol (**2**, 20 g, 63 mmol) and dry THF (120 mL). Upon dissolution, $\text{LiOH}\cdot\text{H}_2\text{O}$ (2.9 g, 68 mmol) was added and the reaction stirred for 1 h at room temperature under an atmosphere of nitrogen. Me_2SO_4 (6.5 mL, 69 mmol) was slowly added, then a reflux condenser was fitted to the flask and the temperature was raised to 80 °C for 18 h. The colorless precipitate was then recovered through filtration. The cake was washed repeatedly with Et_2O and then concentrated *in vacuo* to furnish isosteviol methyl ester (18.3 g, 88%). The reaction was quenched with 10% NaOH and then washed with brine and dried over MgSO_4 . $R_f = 0.3$ (Hex:EtOAc 9:1); white crystals; mp 200–202 °C (Lit 202–203 °C)¹; ^1H NMR (400 MHz, CDCl_3): δ 3.63 (s, 3H), 2.62 (dd, $J = 18.6, 3.8$ Hz, 1H), 2.18 (d, $J = 13.3$ Hz, 1H), 1.89 (dd, $J = 13.6, 2.9$ Hz, 1H), 1.74–1.85 (m, 2H), 1.57–1.74 (m, 5H), 1.52 (ddd, $J = 17.8, 12.5, 3.3$ Hz, 2H), 1.33–1.45 (m, 3H), 1.19–1.30 (m, 2H), 1.19 (s, 3H), 1.10–1.16 (m, 1H), 0.99–1.06 (m, 1H), 0.97 (s, 3H), 0.92 (dd, $J = 13.2, 4.2$ Hz, 1H), 0.68 (s, 3H); ^{13}C

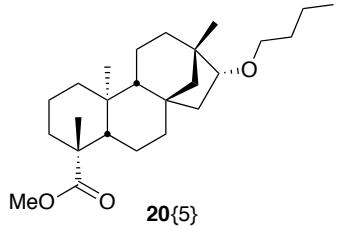
NMR (100 MHz, CDCl₃): δ 222.4, 177.8, 57.0, 54.7, 54.3, 51.2, 48.7, 48.5, 48.4, 43.8, 41.5, 39.8, 39.4, 37.9, 37.3, 28.8, 21.7, 20.3, 19.9, 18.9, 13.2; IR (film) 2952, 1744, 1720, 1452, 1240, 1175, 1153 cm⁻¹; HRMS (ESI) (*m/z*): [M+Na]⁺ calcd for C₂₁H₃₂O₃Na 355.2244; found 355.2234; $[\alpha]_D^{23}$ -82.2 (*c* 1.00, CHCl₃); reported $[\alpha]_D^{25}$ -69.0 (*c* 1.02, CHCl₃).⁸

Reduction of isosteviol methyl ester.¹⁰ To a solution of the isosteviol methyl ester (1.00 g, 3.00 mmol) in MeOH (25 mL) at 0 °C (ice bath), was slowly added NaBH₄ (134 mg, 3.55 mmol). The reaction mixture was stirred for 1 h at 0 °C (ice bath), and then the solvent was evaporated under reduced pressure, followed by dilution with water and extraction with MTBE. Evaporation of the solvent and purification of the product on a silica gel column using 10% EtOAc:Hex gave the hydroxyl methyl ester (450 mg, 45%) as a white crystals. Mp 169-171 °C (Lit 163-166 °C)¹¹; ¹H NMR (400 MHz, CDCl₃): δ 3.85 (dd, *J* = 10.6, 4.6 Hz, 1H), 3.42 (s, 3H), 1.96 (d, *J* = 13.4 Hz, 1H), 1.73 (s, 1H), 1.68 – 1.47 (m, 6H), 1.45 – 1.27 (m, 4H), 1.24 – 0.97 (m, 4H), 0.96 (s, 3H), 0.86 – 0.75 (m, 4H), 0.70 (s, 3H), 0.69 – 0.61 (m, 1H), 0.52 (s, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 178.1, 80.4, 57.1, 55.8, 55.2, 51.1, 43.7, 42.8, 42.0, 42.0, 41.7, 39.9, 38.0, 38.0, 33.7, 28.9, 24.9, 21.7, 20.4, 18.9, 13.1. HRMS (ESI) (*m/z*): [M+Na]⁺ calcd for C₂₁H₃₄O₃Na 357.2406, found 357.2398; $[\alpha]_D^{23}$ -57.1 (*c* 0.833, CHCl₃).

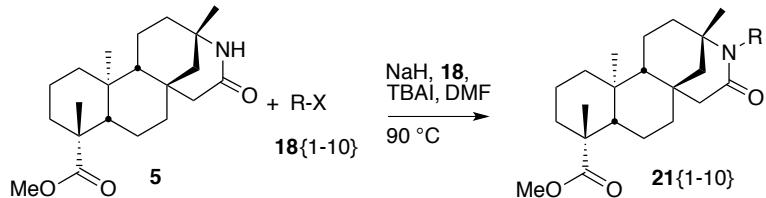
Standard procedure for the synthesis of **20{1-6}**. To a solution of the above hydroxyl methyl ester (100 mg, 0.345 mmol, 1.0 equiv) in DMF (5 mL) in a flame-dry reaction vial was added slowly via cannula a solution of sodium hydride (38.3 mg in 60% dispersion in oil, 0.954 mmol, 3.0 equiv) in dry DMF (5 mL). Then, methyl iodide (90.3 mg, 0.636 mmol, 2.0 equiv) was added to the reaction flask and the mixture was allowed to stir for 2 h at ambient temperature. The reaction was quenched with MTBE and the organic phase was washed successively with water. The aqueous phase was extracted with MTBE and the combined organic layers were dried over MgSO₄, filtered, and concentrated *in vacuo*. Purification by flash column chromatography on silica gel with 20% EtOAc:Hex yielded the product **20**.



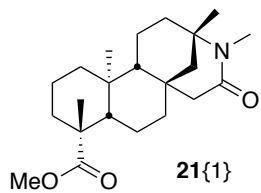
Methyl (4*R*,4a*S*,6a*R*,8*S*,9*S*,11a*R*,11b*S*)-8-((4-Fluorobenzyl)oxy)-4,9,11b-trimethyltetradecahydro-6a,9-methanocyclohepta[*a*]naphthalene-4-carboxylate (20{3}**).** White crystals; mp 151-152 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.29 (dd, *J* = 8.4, 5.7 Hz, 2H), 7.01 (t, *J* = 8.7 Hz, 2H), 4.45 (dd, 2H), 3.63 (s, 3H), 3.54 (dd, *J* = 10.3, 4.1 Hz, 1H), 2.17 (d, *J* = 13.3 Hz, 1H), 1.96 – 1.79 (m, 4H), 1.76 – 1.62 (m, 3H), 1.59 – 1.53 (m, 2H), 1.50 – 1.36 (m, 3H), 1.33 – 1.24 (m, 2H), 1.17 (s, 3H), 1.06 – 0.96 (m, 4H), 0.93 (s, 3H), 0.91 – 0.83 (m, 1H), 0.72 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 178.3, 135.3, 129.0, 128.9, 115.9, 115.2, 115.0, 87.0, 71.3, 57.3, 56.0, 55.5, 51.3, 43.9, 42.4, 42.2, 42.0, 40.3, 40.0, 38.2, 38.1, 34.5, 29.0, 25.6, 21.9, 20.6, 19.1, 13.3; HRMS (ESI) (*m/z*): [M+Na]⁺ calcd for C₂₈H₃₉O₃FNa 465.2883, found 465.2795; $[\alpha]_D^{23}$ -103.7 (*c* 0.655, CHCl₃).



Methyl (4R,4aS,6aR,8R,9S,11aR,11bS)-8-Butoxy-4,9,11b-trimethyltetradecahydro-6a,9-methanocyclohepta[a]naphthalene-4-carboxylate (20{5}). Oil; ^1H NMR (400 MHz, CDCl_3): δ 3.65 (s, 3H), 3.53 – 3.43 (m, 1H), 3.43 – 3.31 (m, 2H), 2.17 (d, $J = 13.0$ Hz, 1H), 1.89 – 1.84 (m, 2H), 1.83 – 1.77 (m, 3H), 1.72 – 1.66 (m, 2H), 1.64 – 1.56 (m, 3H), 1.51 (dd, $J = 12.2, 6.1$ Hz, 3H), 1.44 – 1.37 (m, 4H), 1.32 – 1.24 (m, 2H), 1.21 (d, $J = 4.7$ Hz, 1H), 1.18 (s, 3H), 1.12 – 1.03 (m, 2H), 1.02 – 0.97 (m, 2H), 0.94 (s, 3H), 0.92 – 0.83 (m, 2H), 0.73 (s, 3H); ^{13}C NMR (125 MHz, CDCl_3): δ 178.2, 87.3, 69.9, 57.3, 56.0, 55.5, 51.1, 43.8, 42.2, 42.0, 40.5, 39.9, 38.1, 38.0, 34.4, 32.3, 28.9, 25.6, 21.8, 21.7, 20.4, 19.5, 19.0, 14.0, 13.1; HRMS (ESI) (m/z): [M+Na] $^+$ calcd for $\text{C}_{25}\text{H}_{42}\text{O}_3\text{Na}$ 413.3134, found 413.2998; $[\alpha]_D^{23} -42.8$ (c 0.850, CHCl_3).

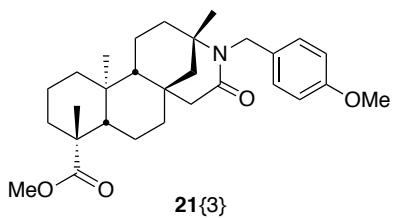


Standard procedure for *N*-alkylation of known lactam **5** to form **21(1-10)**.⁸ To a solution of the lactam methyl ester **5** (100 mg, 0.347 mmol, 1.0 equiv) in DMF (5 mL) in a flame-dried reaction flask, was added sodium hydride (3.0 equiv, pre-washed with hexanes), alkyl halide **18** (5.0 equiv) and a catalytic amount of tetrabutylammonium iodide (TBAI, 10 mol%). The reaction flask was heated to 90 °C, stirred overnight and then the reaction was carefully quenched with water (2 mL). The organic layer was then washed with brine and the aqueous layers were re-extracted with Et₂O (2 x 10 mL). The combined organic layers were then dried over MgSO₄, filtered, and the Et₂O was removed under reduced pressure, while a centrifugal evaporator was used to remove the DMF to furnish alkylated lactam methyl ester **21**. Column chromatography on silica gel with 5% MeOH:CH₂Cl₂ gave the substituted-lactam products.

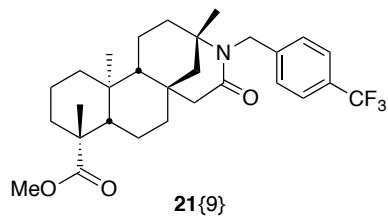


(3S,8aS,9R,12aS,12bR)-Methyl 3,4,9,12a-Tetramethyl-5-oxotetradecahydro-1*H*-3,6a-methanonaphtho[2,1-*d*]azocine-9-carboxylate (21{1}). Yellow solid; mp 168–172 °C; ^1H NMR (400 MHz, CDCl_3): δ 3.62 (s, 3H), 2.98 (dd, $J = 18.3, 2.9$ Hz, 1H), 2.84 (s, 3H), 2.16 (d, $J = 13.3$ Hz, 1H), 2.04 (d, $J = 18.3$ Hz, 1H), 1.95 – 1.22 (m, 2H), 1.83 – 1.66 (m, 3H), 1.65 – 1.54 (m, 2H), 1.51 (dt, $J = 12.9, 3.2, 3.2$ Hz, 1H), 1.48 – 1.37 (m, 1H), 1.30 (dd, $J = 12.8, 2.9$ Hz, 1H), 1.25 (s, 3H), 1.21 (d, $J = 13.0, 3.8$ Hz, 1H), 1.16 (s, 3H), 1.16 – 0.93 (m, 4H), 0.92 – 0.78 (m, 2H), 0.75 (s, 3H); ^{13}C NMR (100 MHz, CDCl_3): δ 177.7, 171.9, 57.4, 56.8, 55.3, 51.1, 51.0, 44.3, 43.7, 41.0, 39.9, 38.0, 37.8, 35.7, 33.9, 28.5,

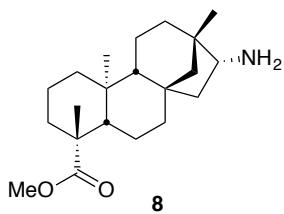
27.4, 26.8, 19.7, 18.8, 18.7, 13.5; HRMS (ESI) (*m/z*): [M+H]⁺ calcd for C₂₂H₃₆NO₃ 362.2695; found 362.2677; [α]_D²³ 18.9 (*c* 0.800, CHCl₃).



Methyl (3*S*,6*aR*,8*aS*,9*R*,12*aS*,12*bR*)-4-(4-Methoxybenzyl)-3,9,12*a*-trimethyl-5-oxotetradecahydro-2*H*-3,6*a*-methanonaphtho[2,1-*d*]azocine-9-carboxylate (21{3}). Pale yellow solid; mp 151–152 °C; ¹H NMR (400 MHz, CDCl₃): 7.16 (d, *J* = 8.7 Hz, 2H), 6.81 (d, *J* = 8.7 Hz, 2H), 4.86 (d, *J* = 15.4 Hz, 1H), 4.20 (d, *J* = 15.5 Hz, 1H), 3.78 (s, 3H), 3.64 (s, 3H), 3.07 (dd, *J* = 18.4, 2.7 Hz, 1H), 2.16 (d, *J* = 18.2 Hz, 2H), 1.93 – 1.82 (m, 1H), 1.81 – 1.72 (m, 4H), 1.67 (dd, *J* = 13.0, 2.8 Hz, 1H), 1.54 (dd, *J* = 12.9, 3.1 Hz, 2H), 1.48 – 1.39 (m, 1H), 1.31 – 1.19 (m, 3H), 1.17 (s, 3H), 1.14 (s, 3H), 1.12 – 0.95 (m, 3H), 0.90 – 0.79 (m, 2H), 0.78 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): 177.8, 172.3, 158.3, 132.1, 128.3 (2C), 113.7 (2C), 57.5, 56.9, 56.4, 55.2, 51.7, 51.2, 44.3, 44.0, 43.8, 41.2, 40.0, 38.0, 37.8, 37.1, 34.1, 28.6, 28.1, 19.8, 18.9, 18.6, 13.7; LRMS (ESI) (*m/z*): [M+H]⁺ calcd for C₂₉H₄₂NO₄ 468.3036, found 468.3; [α]_D²³ –15.6 (*c* 1.00, CHCl₃).

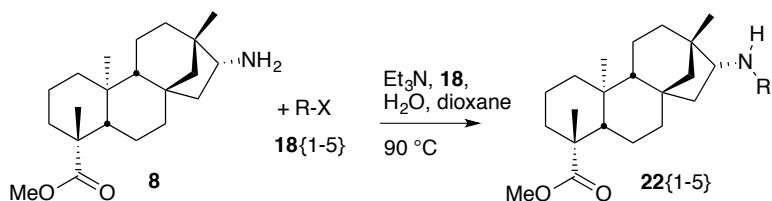


Methyl (3*S*,6*aR*,8*aS*,9*R*,12*aS*,12*bR*)-3,9,12*a*-Trimethyl-5-oxo-4-(4-(trifluoromethyl)benzyl)tetradecahydro-2*H*-3,6*a*-methanonaphtho[2,1-*d*]azocine-9-carboxylate (21{9}). Lightly yellow solid; mp 151–152 °C; ¹H NMR (400 MHz, CDCl₃): δ 7.53 (d, *J* = 8.1 Hz, 2H), 7.32 (d, *J* = 8.1 Hz, 2H), 5.08 (d, *J* = 16.2 Hz, 1H), 4.15 (d, *J* = 16.1 Hz, 1H), 3.64 (s, 3H), 3.09 (dd, *J* = 18.2, 2.5 Hz, 1H), 2.23 – 2.15 (m, 2H), 1.91 – 1.75 (m, 5H), 1.71 (dd, *J* = 13.1, 2.5 Hz, 1H), 1.64 – 1.55 (m, 2H), 1.45 (d, *J* = 13.7 Hz, 1H), 1.36 – 1.29 (m, 1H), 1.27 – 1.20 (m, 2H), 1.18 (s, 3H), 1.17 – 1.11 (m, 1H), 1.10 (s, 3H), 1.08 – 0.96 (m, 2H), 0.94 – 0.84 (m, 2H), 0.79 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 177.7, 172.7, 144.0, 127.1, 127.1, 127.1, 125.4, 125.3, 125.3, 57.4, 56.8, 56.6, 51.5, 51.2, 44.5, 44.2, 43.8, 41.1, 40.0, 38.0, 37.8, 36.9, 34.1, 28.6, 28.0, 19.7, 18.9, 18.6, 13.7; LRMS (ESI) (*m/z*): [M]⁺ calcd for C₂₉H₃₈F₃NO₃ 505.2814, found 506.4; [α]_D²³ –36.6 (*c* 0.905, CHCl₃).

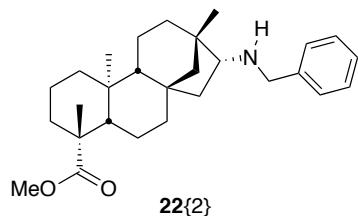


Methyl (4*R*,4*aS*,6*aR*,9*S*,11*aR*,11*bS*)-8-Amino-4,9,11*b*-trimethyltetradecahydro-6*a*,9-methanocyclohepta[a]naphthalene-4-carboxylate (8).¹² To a solution of methyl

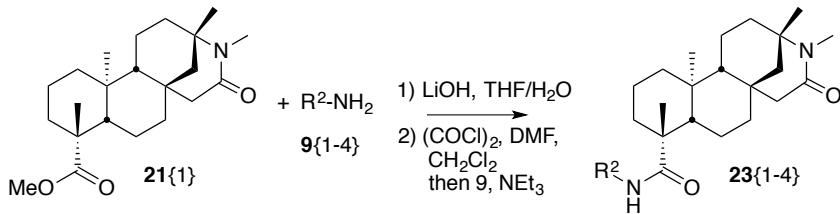
(*4R,4aS,6aR,9S,11aR,11bS*)-8-(hydroxyimino)-4,9,11b-trimethyltetradecahydro-6a,9-methanocyclohepta[*a*]naphthalene-4-carboxylate¹ (348 mg, 1.00 mmol, 1.0 equiv) in MeOH, cooled to 0 °C (ice bath), was added sodium borohydride (189 mg, 5.0 equiv) then molybdenum (VI) oxide (216 mg, 1.5 equiv). The reaction was stirred at room temperature overnight. The next day the reaction was filtered, and then the solvent was evaporated. The resulting residue was treated with aqueous KOH (20%) and extracted with CH₂Cl₂. The combined organic layers were washed with brine, dried with MgSO₄, and then concentrated *in vacuo*. Column chromatography on silica gel with 5% MeOH/ CH₂Cl₂ provided amine **8** in 20% yield as an off-white solid. mp: 255–258 °C; ¹H NMR (400 MHz, CDCl₃): δ 3.60 (s, 3H), 3.12 (dd, *J* = 10.6, 6.7 Hz, 1H), 2.13 (d, *J* = 13.2 Hz, 1H), 1.86 – 1.49 (m, 10H), 1.42 – 1.25 (m, 4H), 1.26 – 1.21 (m, 1H), 1.14 (s, 3H), 1.11 – 1.01 (m, 2H), 1.00 (s, 3H), 0.99 – 0.92 (m, 2H), 0.88 – 0.78 (m, 2H), 0.71 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 178.1, 77.0, 76.7, 59.6, 57.0, 56.0, 55.6, 51.1, 43.7, 42.4, 41.2, 41.1, 39.7, 39.0, 38.0, 38.0, 33.0, 28.8, 24.6, 21.6, 20.2, 18.8, 12.9; HRMS (ESI) (*m/z*): [M+H]⁺ calcd for C₂₁H₃₆NO₂ 334.2688, found 334.2747; [α]_D²³ –153.1 (*c* 1.00, CHCl₃).



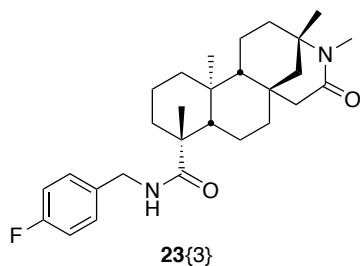
Standard procedure for *N*-alkylation of amine **8** for the synthesis of **22{1-5}**. To a flame-dried reaction flask of the amine **8** (100 mg, 0.333 mmol, 1.0 equiv) and triethylamine (1.5 equiv) in a 1,4-dioxane-water mixed solvent (1:1, 6 mL), the alkyl halide (**18**, 1.0 equiv) was added. The reaction flask was heated to 90 °C and allowed to stir for one day. The reaction mixture was quenched with EtOAc and the organic layer was washed with saturated aqueous NH₄Cl. The organic layer was then washed with brine and the aqueous layers were extracted with EtOAc (2 x 10 mL). The combined organic layers were then dried over MgSO₄, filtered, and concentrated *in vacuo* to give the crude alkylated amine product **22**. Purification was done by column chromatography on silica gel with 5% MeOH: CH₂Cl₂.



Methyl (4*R*,4*aS*,6*a**R*,8*R*,9*S*,11*a**R*,11*b**S*)-8-(Benzylamino)-4,9,11*b*-trimethyltetradecahydro-6*a*,9-methanocyclohepta[*a*]naphthalene-4-carboxylate (22{2}).** Oil; ¹H NMR (400 MHz, CDCl₃): δ 7.35 (d, *J* = 7.6 Hz, 4H), 7.29 – 7.25 (m, 1H), 3.80 (q, *J* = 46.0, 13.4, 12.4 Hz, 2H), 3.66 (s, 3H), 3.64 (s, 1H), 2.77 (dd, *J* = 10.6, 5.4 Hz, 1H), 2.19 (d, *J* = 13.3 Hz, 1H), 1.86 – 1.80 (m, 2H), 1.74 (d, *J* = 13.5 Hz, 3H), 1.64 – 1.59 (m, 3H), 1.57 – 1.51 (m, 2H), 1.46 – 1.40 (m, 1H), 1.38 – 1.31 (m, 2H), 1.19 (s, 3H), 1.10 – 1.04 (m, 2H), 1.03 – 0.98 (m, 3H), 0.92 (s, 3H), 0.90 – 0.86 (m, 1H), 0.74 (s, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 178.2, 141.4, 128.3, 128.2, 128.0, 128.0, 126.7, 66.2, 57.3, 57.0, 55.9, 53.2, 51.1, 43.8, 42.4, 41.8, 41.6, 41.2, 40.0, 38.1, 38.1, 34.3, 28.9, 25.6, 21.8, 20.7, 19.0, 13.3; HRMS (ESI) (*m/z*): [M]⁺ calcd for C₂₈H₄₁NO₂ 423.3137, found 424.3; [α]_D²³ –58.5 (*c* 1.00, CHCl₃).



Standard procedure for amide bond formation lactam ester **23{1-4}** from **21{1}**. To a solution of *N*-methyl lactam ester **21{1}** (1.00 g, 2.77 mmol) in THF/H₂O (1:1, 10 mL) in a round-bottom flask was added LiOH (1M aq. 5 mL) at room temperature. The reaction was heated to reflux and stirred overnight. Then, it was cooled to room temperature and quenched with HCl (1M aq.) to pH 4. The resultant mixture was extracted with EtOAc and the combined organic layers were washed with brined and dried over MgSO₄. The acid was carried on without further purification. To an oven-dried reaction flask under nitrogen at 0 °C (ice bath), was added DMF (37 μL, 0.47 mmol, 1.5 equiv) in CH₂Cl₂ (1.0 mL) followed by oxalyl chloride (41 μL, 0.47 mmol, 1.5 equiv). The ice bath was removed and the mixture was allowed to stir at room temperature for 1 h. Then the reaction flask was cooled to 0 °C (ice bath) and *N*-methyl lactam acid (108 mg, 0.31 mmol) and Et₃N (130 μL, 0.47 mmol, 3 equiv) in CH₂Cl₂ (1.0 mL) were added. After stirring for 5 min, amine **9** (1 mL) was added and the mixture was stirred for 2 h. Then, the solvent was removed and the reaction was purified using column chromatography on silica gel with 10%>20% EtOAc:Hex as the eluent to provide the substituted lactam amide product **23**.

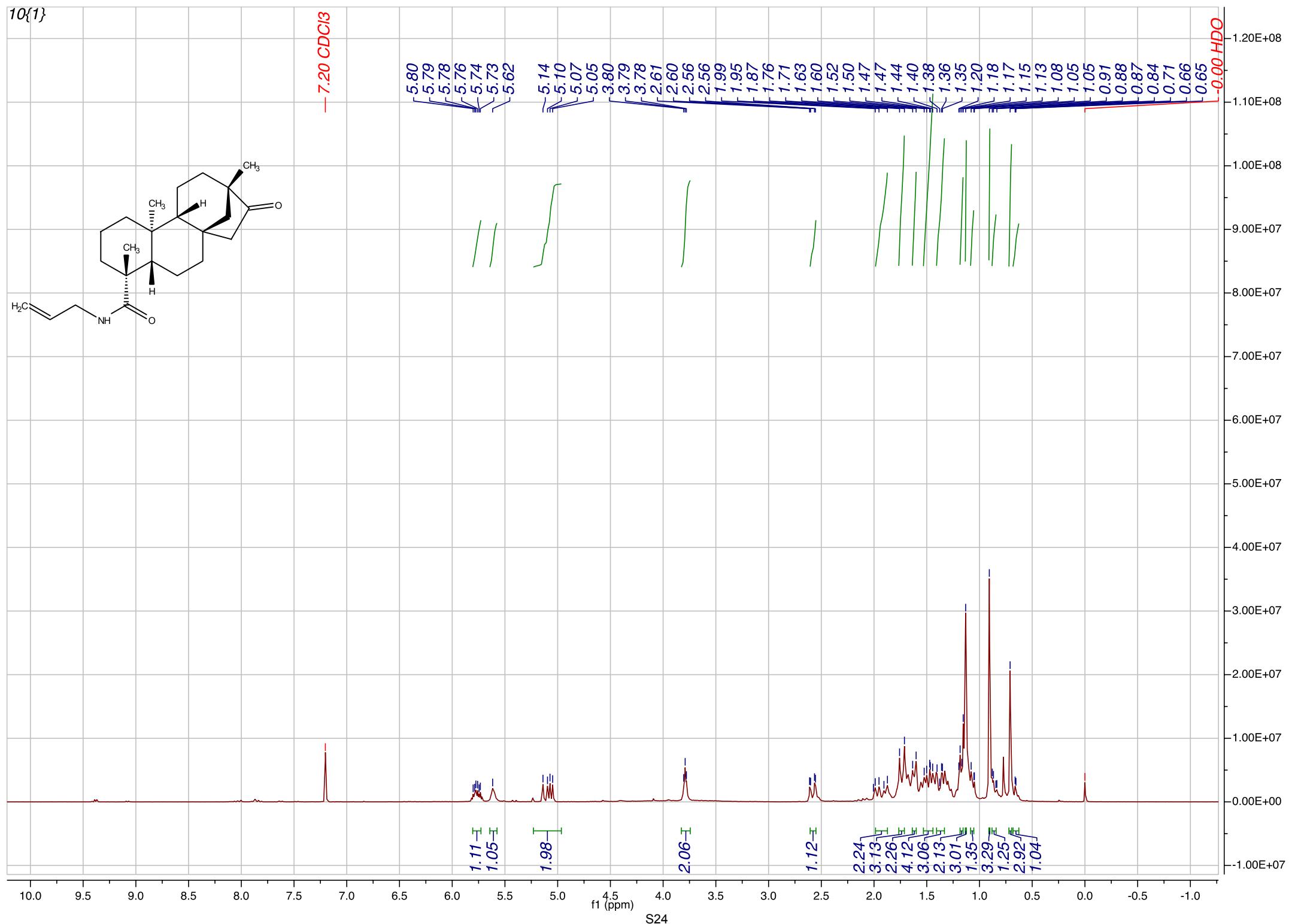


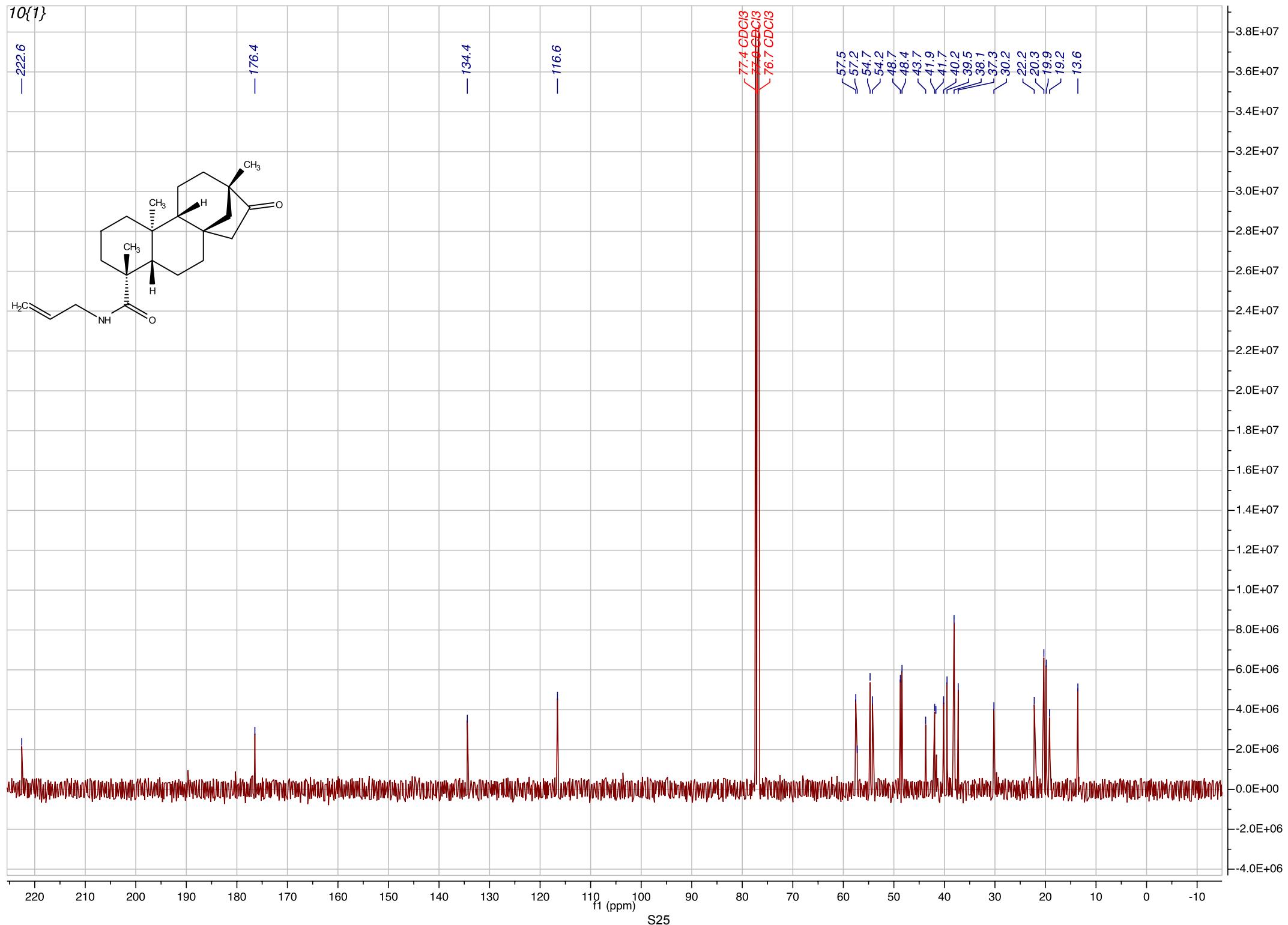
(3*S*,6*aR*,8*aS*,9*R*,12*aS*,12*bR*)-*N*-(4-Fluorobenzyl)-3,4,9,12*a*-tetramethyl-5-oxotetradecahydro-2*H*-3,6*a*-methanonaphtho[2,1-*d*]azocine-9-carboxamide(23{3}). Oil; ¹H NMR (400 MHz, CDCl₃): δ 7.23 (dd, *J* = 8.3, 5.4 Hz, 2H), 7.00 (t, *J* = 8.5 Hz, 2H), 5.20 (d, *J* = 15.4 Hz, 1H), 4.67 (s, 1H), 4.44 (d, *J* = 15.4 Hz, 1H), 3.63 (s, 3H), 2.15 (dd, *J* = 23.4, 13.3 Hz, 2H), 1.87 – 1.72 (m, 2H), 1.70 (s, 3H), 1.67 – 1.56 (m, 3H), 1.55 – 1.44 (m, 2H), 1.42 – 1.32 (m, 3H), 1.30 (s, 3H), 1.28 – 1.18 (m, 2H), 1.15 (s, 3H), 1.11 – 0.89 (m, 4H), 0.87 (dd, *J* = 13.0, 3.8 Hz, 1H), 0.78 (td, *J* = 13.4, 3.9 Hz, 1H); ¹³C NMR (125 MHz, CDCl₃): δ 188.2, 177.4, 133.8, 129.1, 129.0, 116.1, 115.9, 96.4, 58.7, 56.9, 56.2, 51.3, 51.2, 43.7, 43.1, 39.5, 39.3, 37.7, 37.3, 37.2, 35.4, 28.9, 25.2, 19.2, 19.2, 19.0, 14.2, 12.9; LRMS (ESI) (*m/z*): [M+H]⁺ calcd for C₂₈H₄₀FN₂O₂ 455.2996, found 455; [α]_D²³ −74.3 (*c* 0.450, CHCl₃).

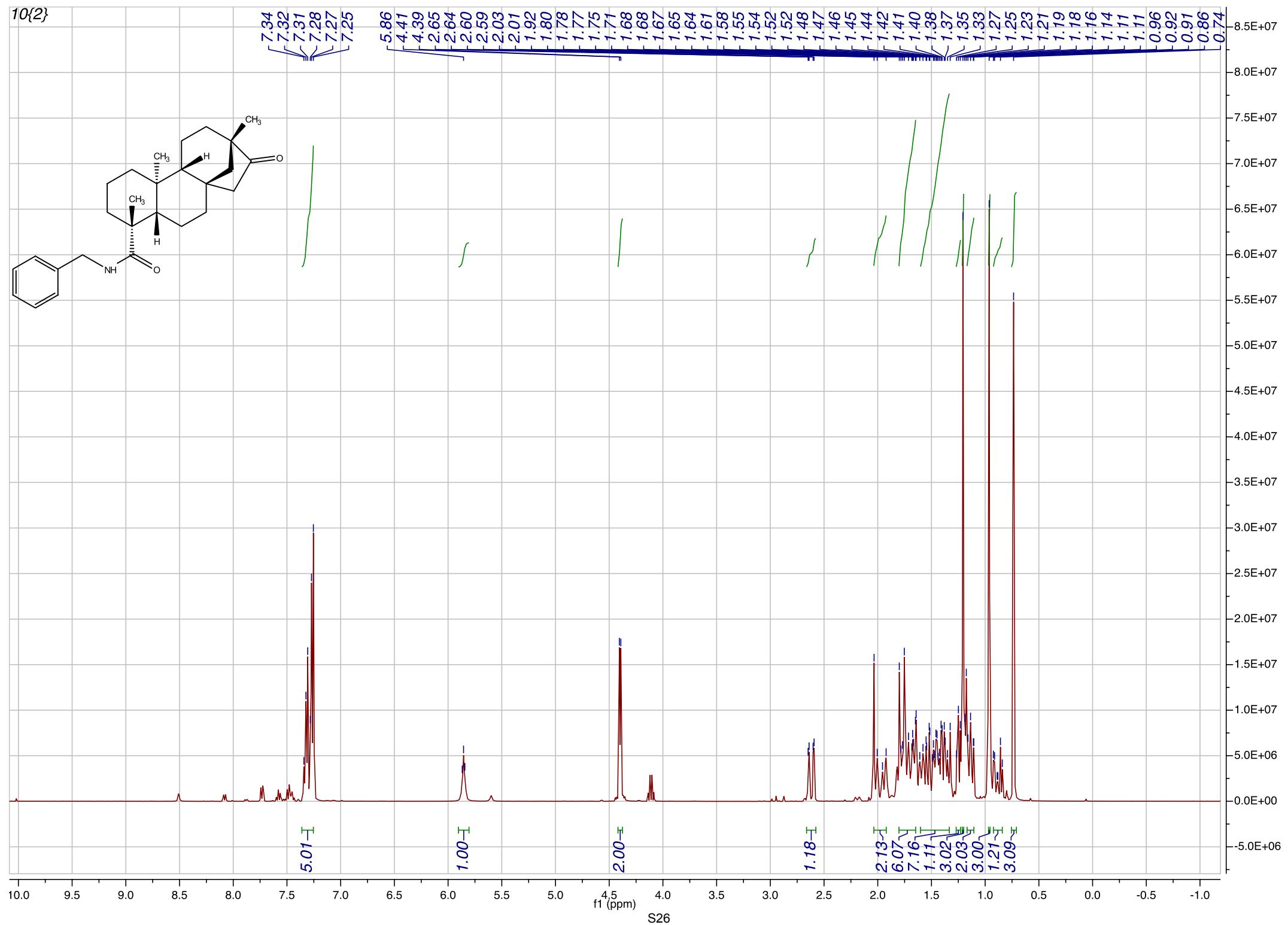
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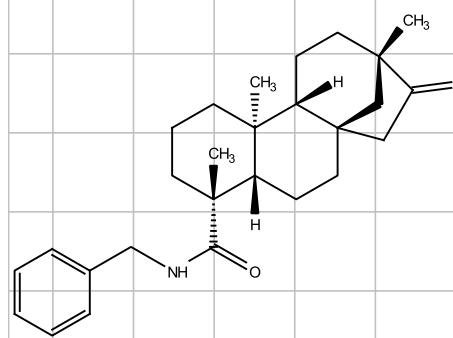






10{2}

-222.4



-176.4

-138.5

 $\begin{cases} 128.7 \\ 128.0 \\ 127.5 \end{cases}$
 $\begin{cases} 77.3 \text{ CDCl}_3 \\ 77.0 \text{ CDCl}_3 \\ 76.7 \text{ CDCl}_3 \end{cases}$

57.6

54.7

54.3

48.7

48.4

43.7

41.7

40.2

39.5

38.1

38.1

37.3

30.2

22.2

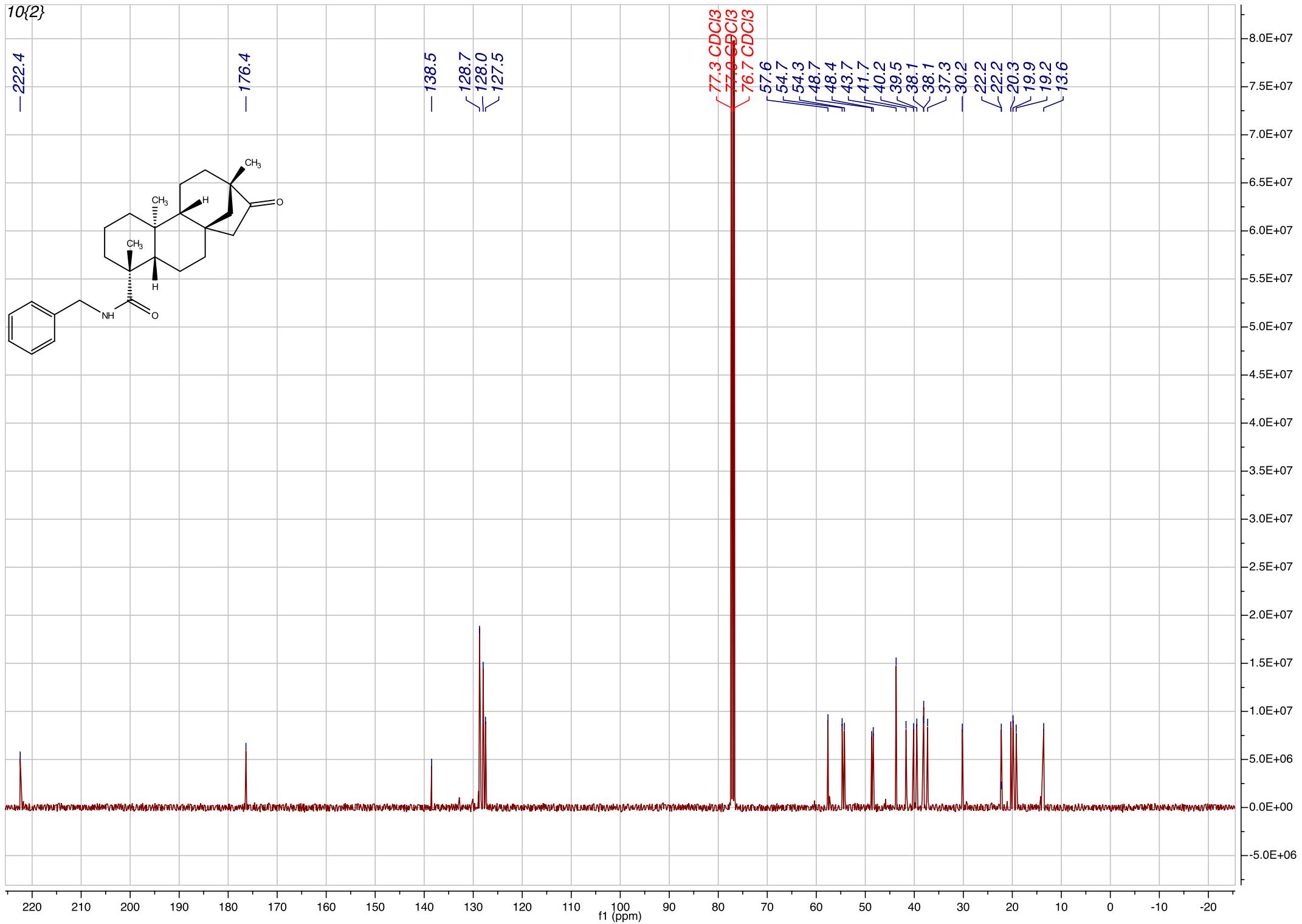
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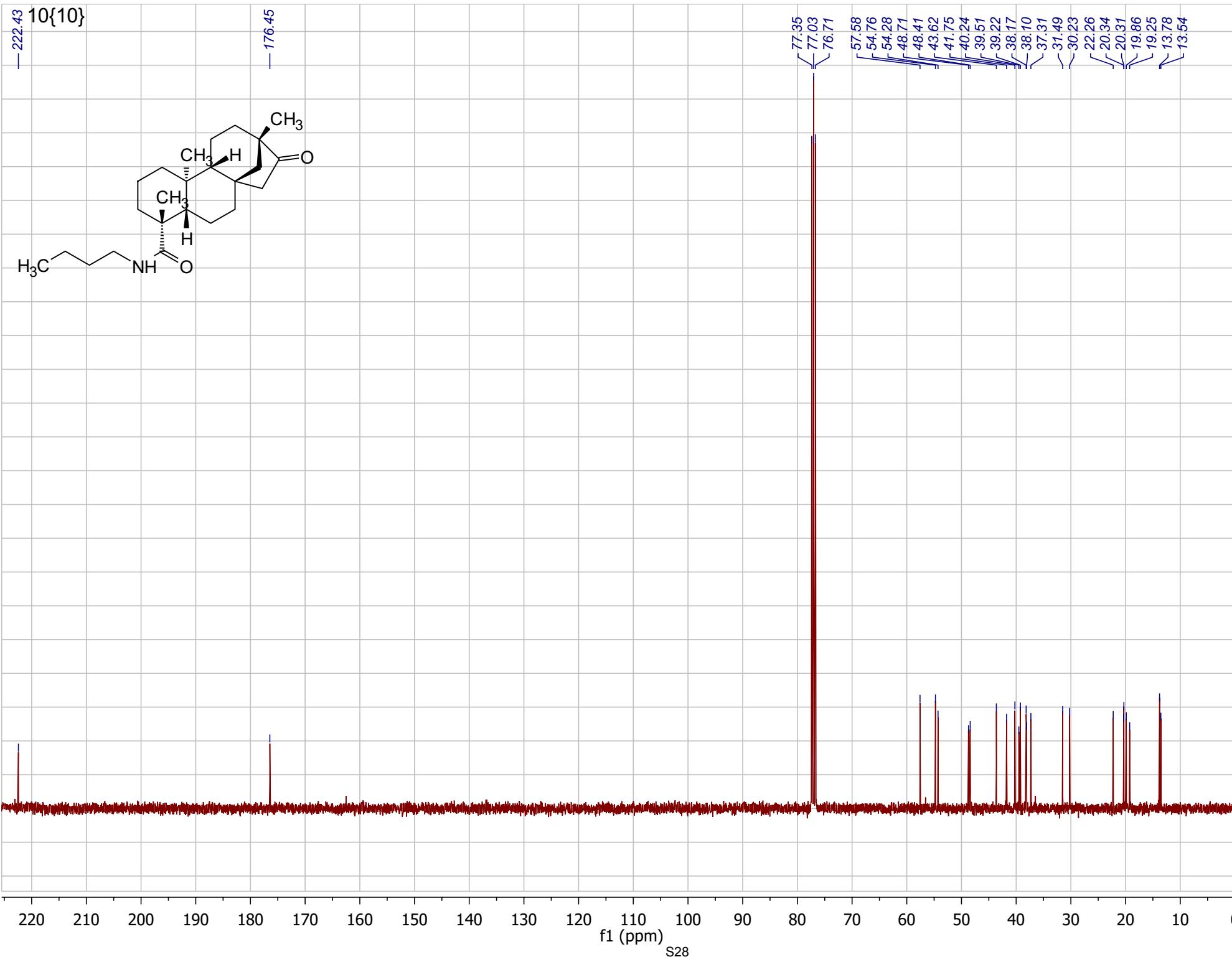
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19.9

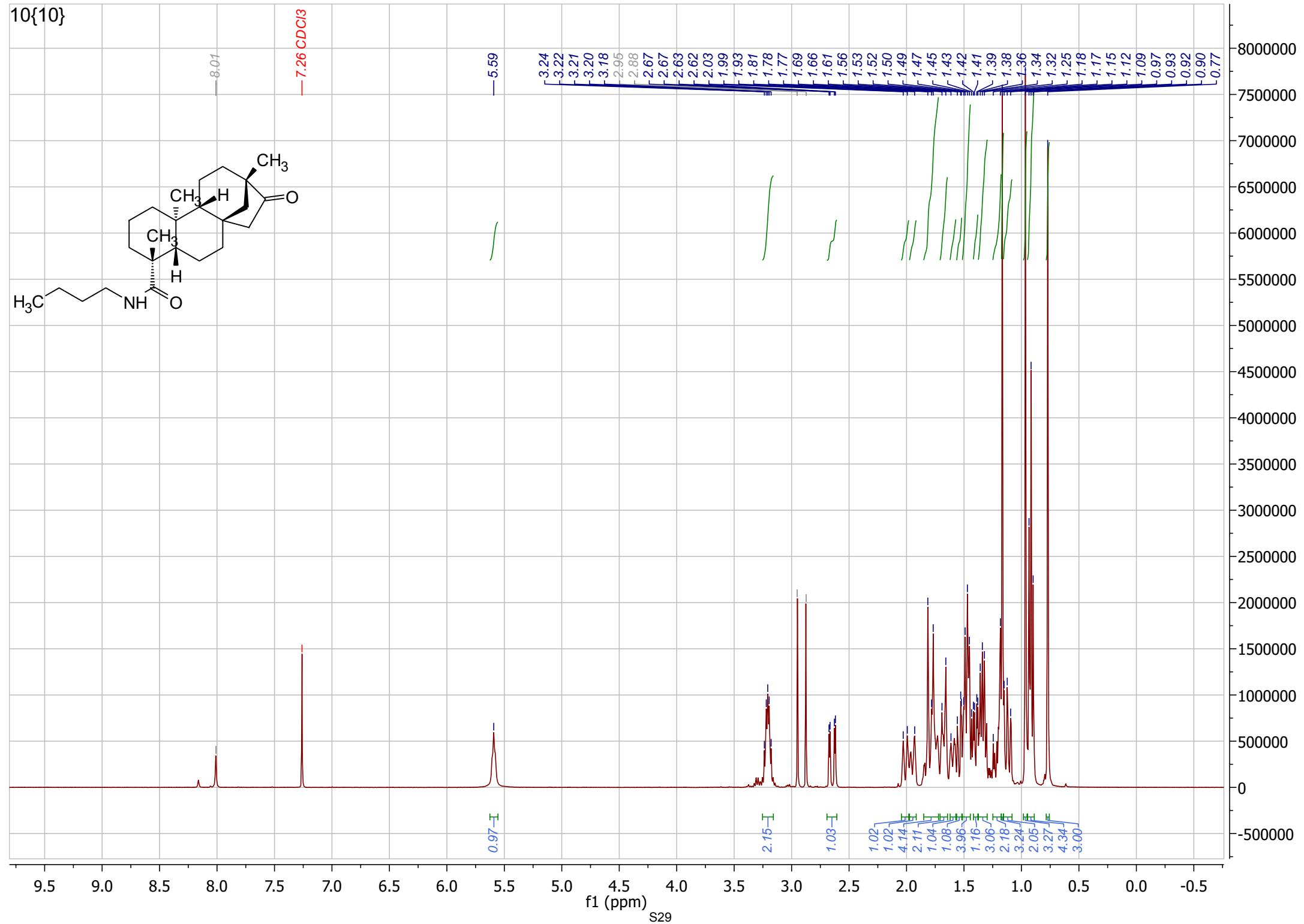
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13.6

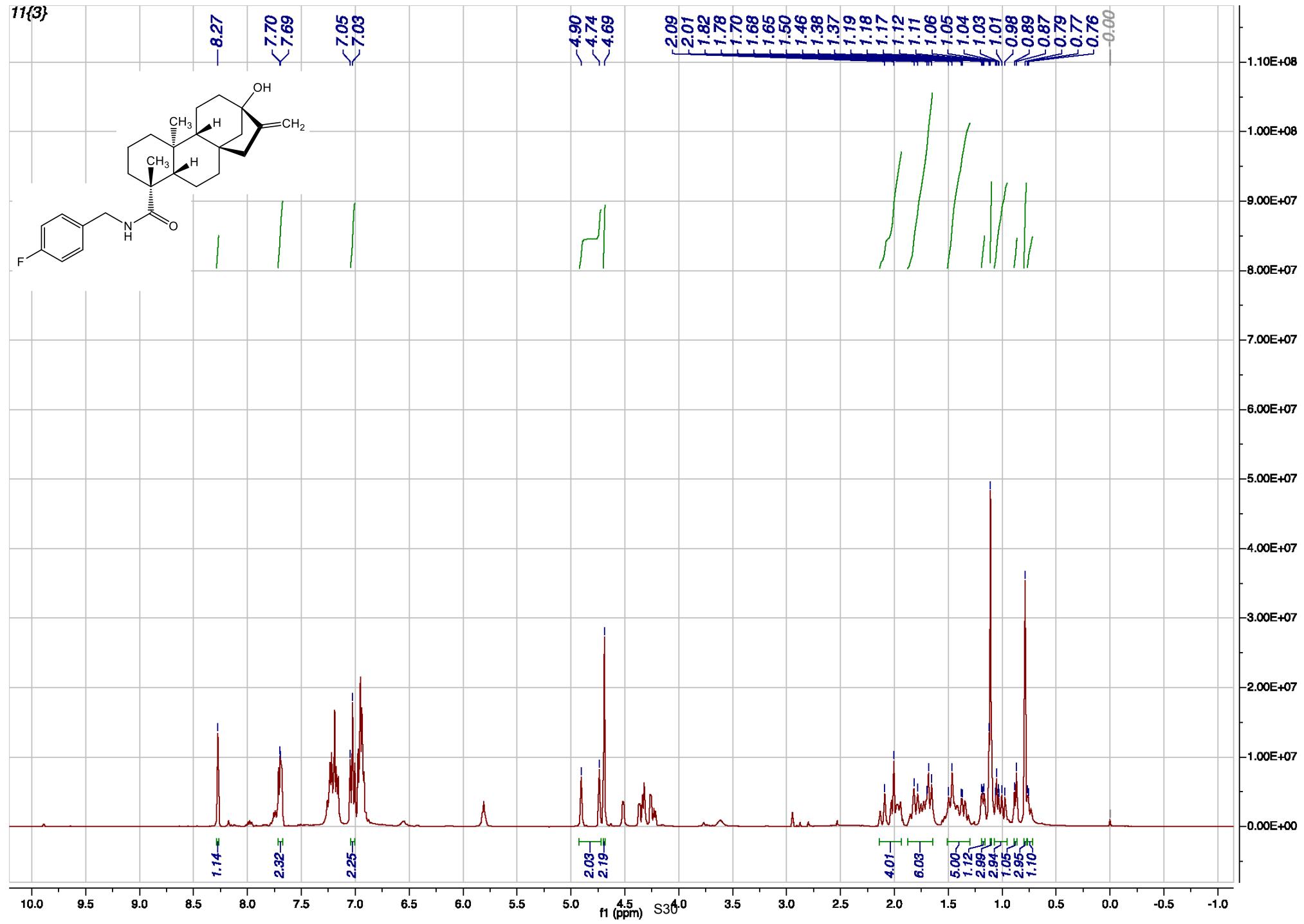




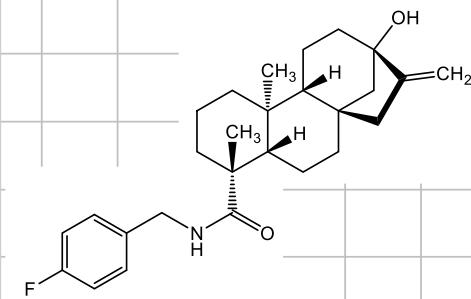
10{10}



11{3}



11{3}



- 176.6

- 160.6

- 158.0

< 130.1

< 115.6

< 115.4

- 103.0

80.2

77.4 CDCl₃77.1 CDCl₃76.7 CDCl₃

64.2

57.4

53.7

47.4

46.9

43.8

42.9

41.7

40.9

39.4

38.2

30.1

23.0

22.4

20.4

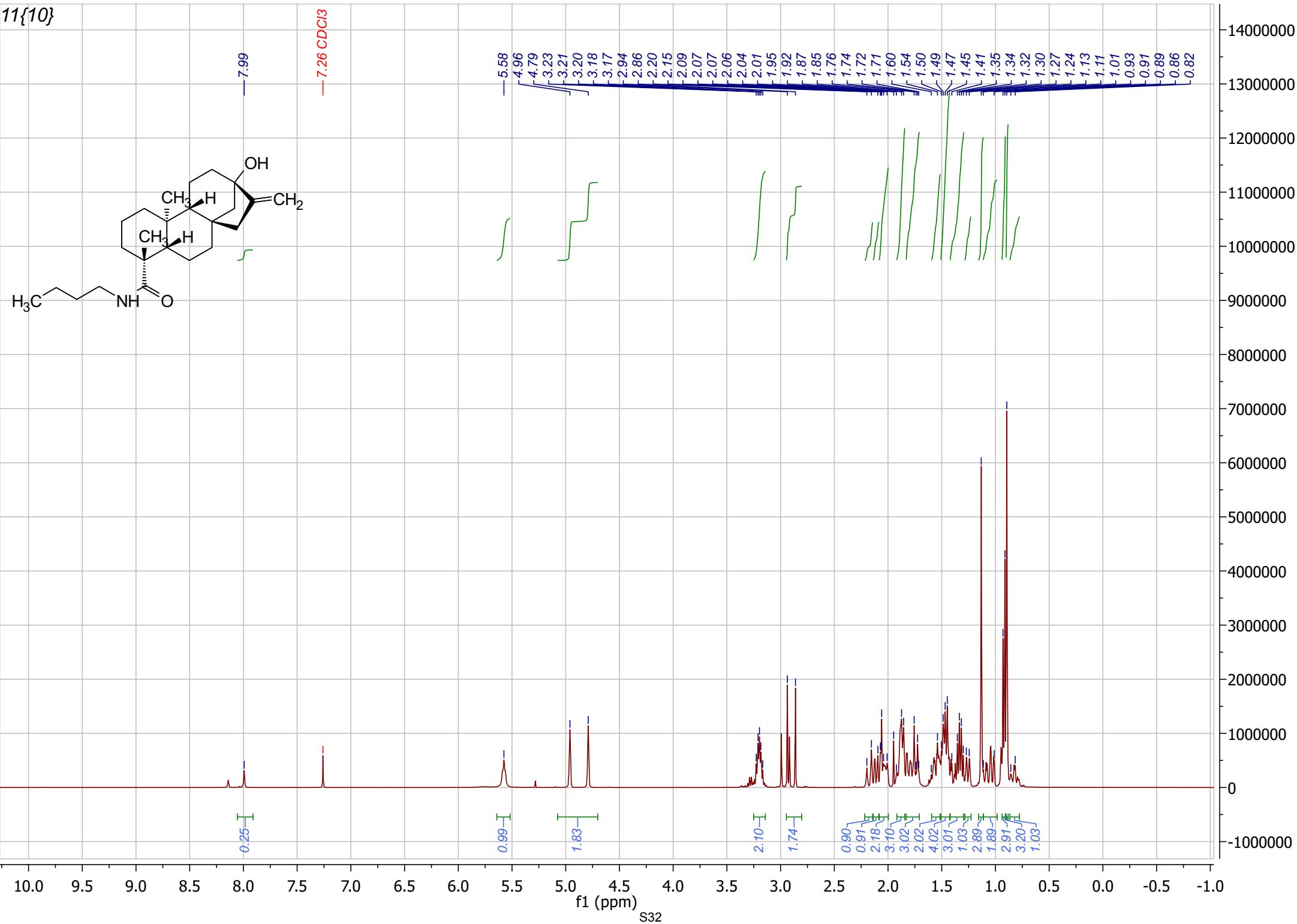
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15.7

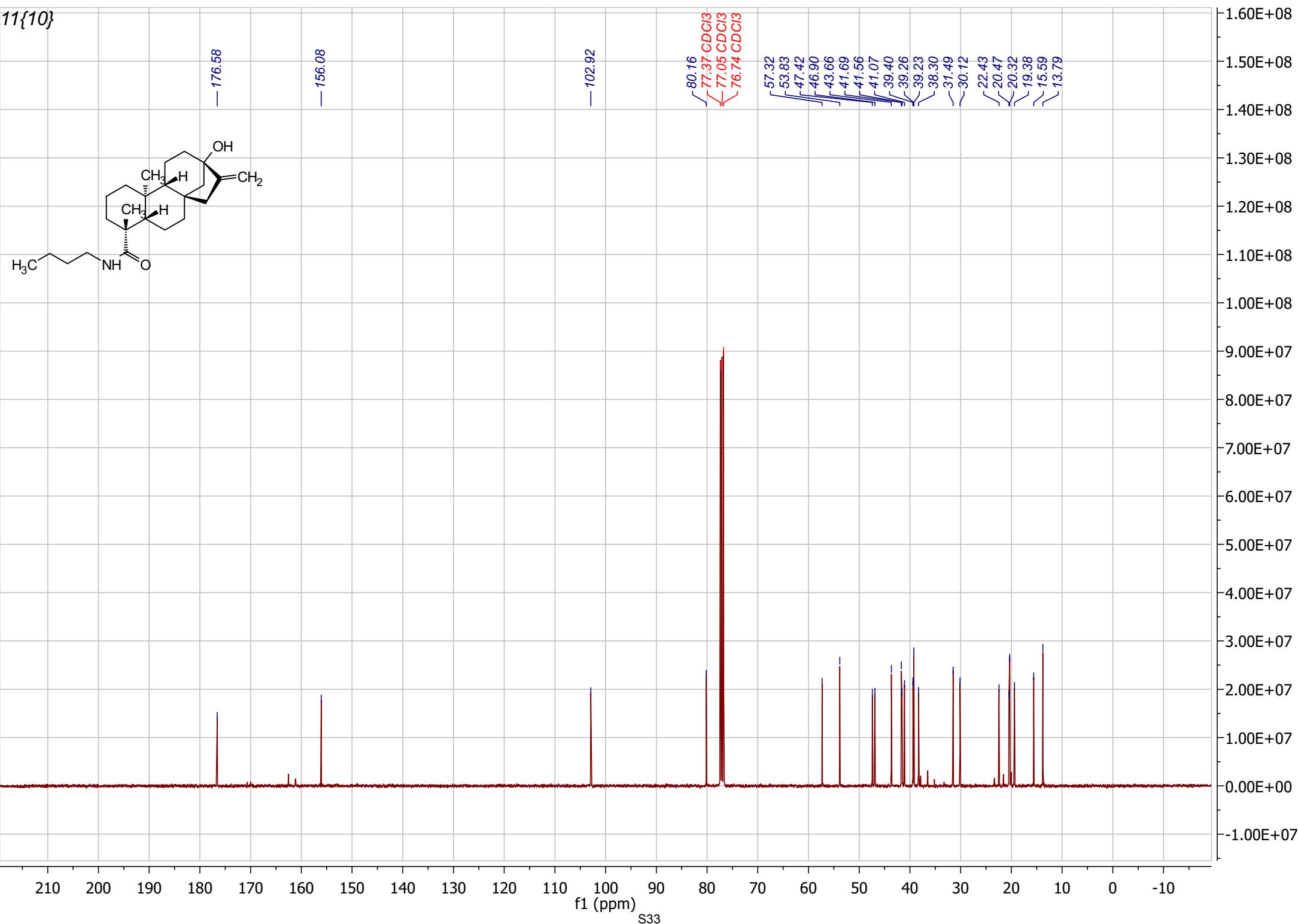
220 210 200 190 180 170 160 150 140 130 120 110 f1 (ppm) S31

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1.00E+08
9.00E+07
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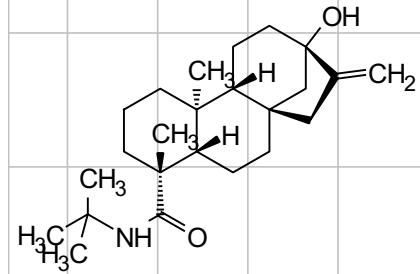
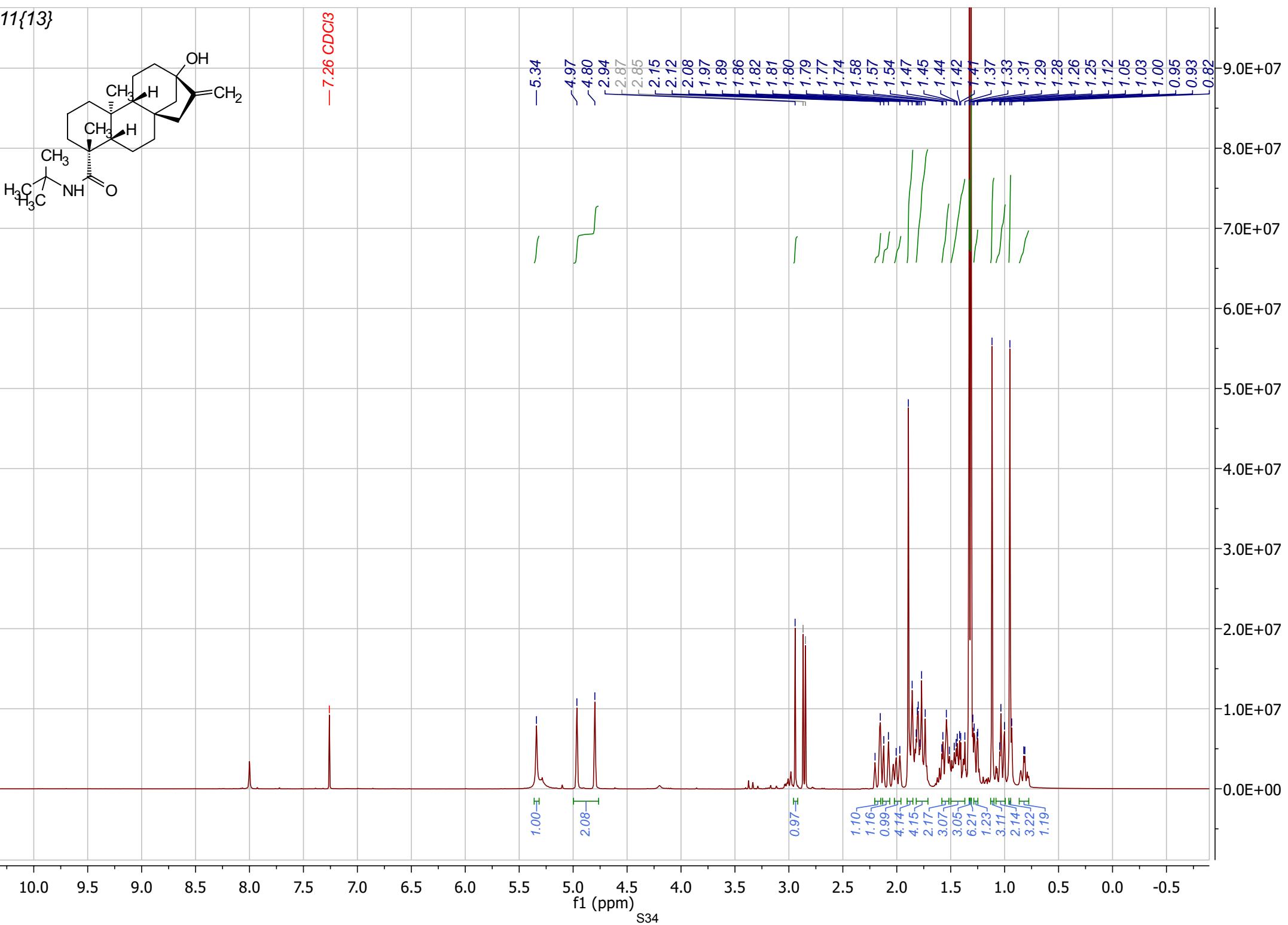
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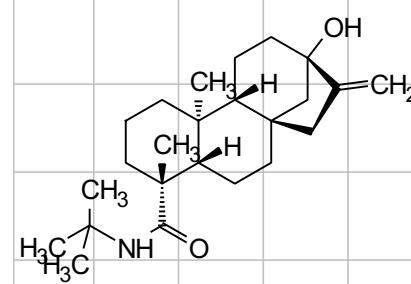
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11{13}

—7.26 CDCl₃

11{13}



-175.72

-156.09

-102.91

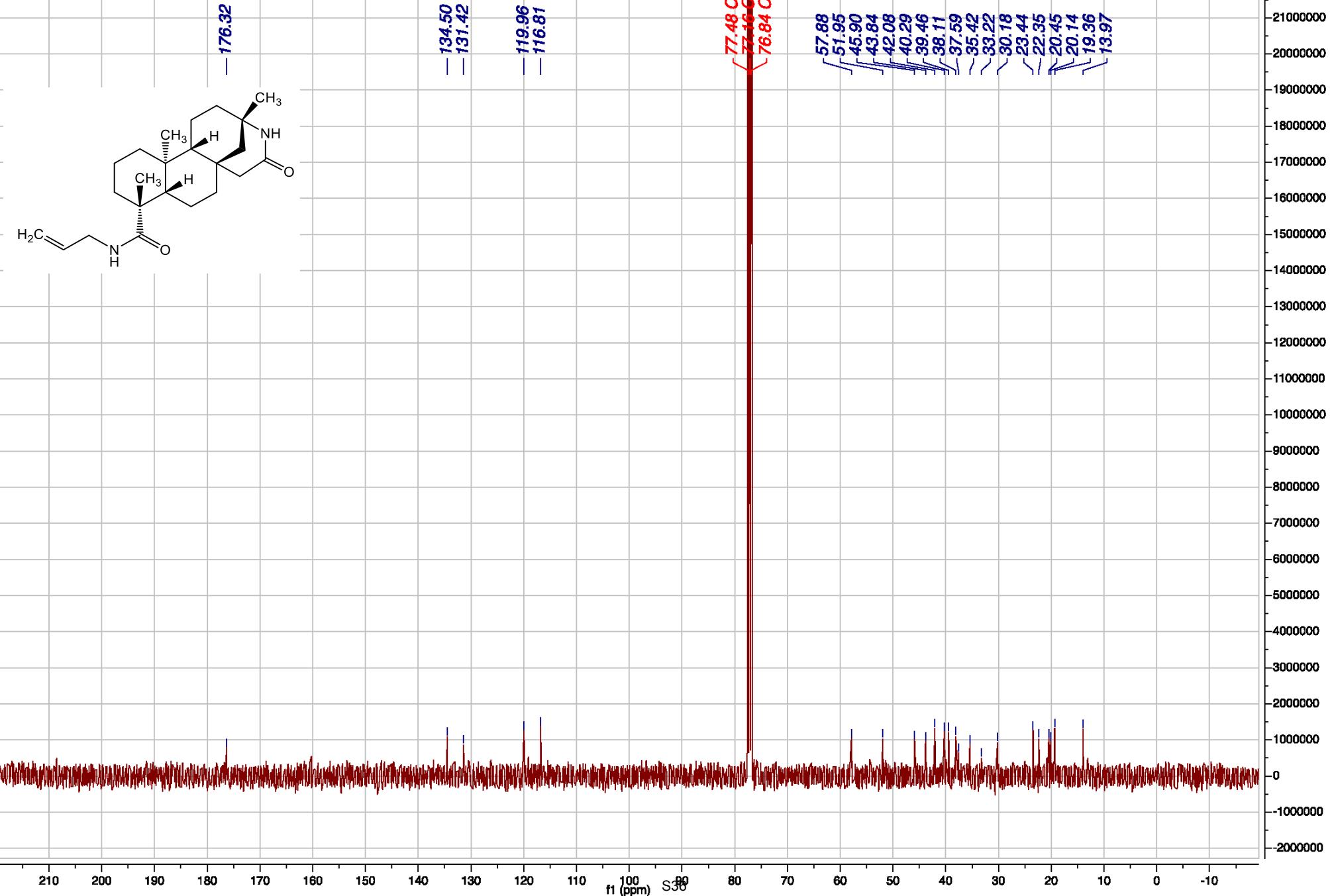
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76.72 CDCl₃
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41.71
41.17
39.50
39.26
38.53
30.21
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-22.45
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-15.94

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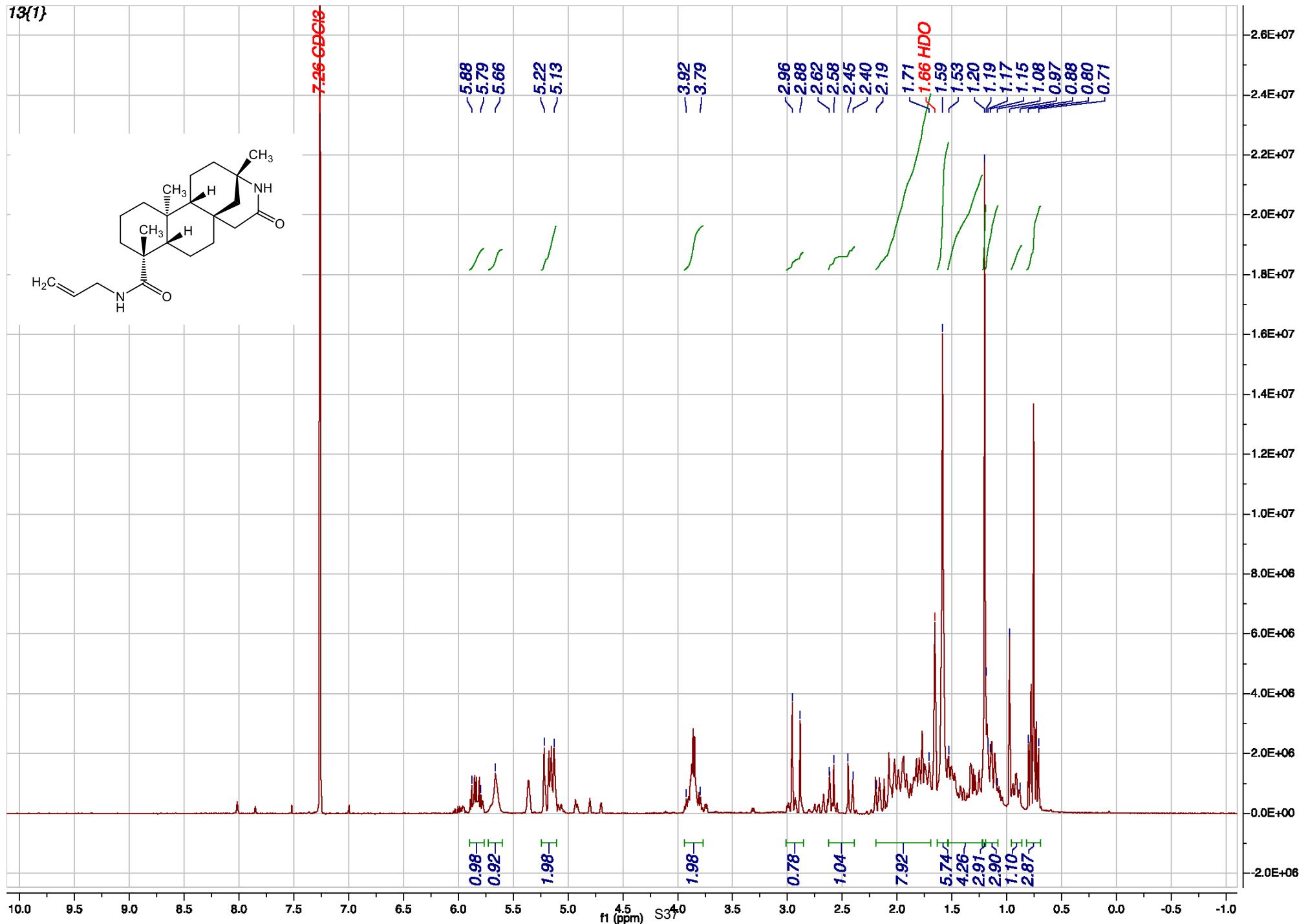
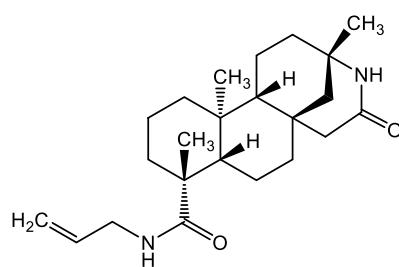
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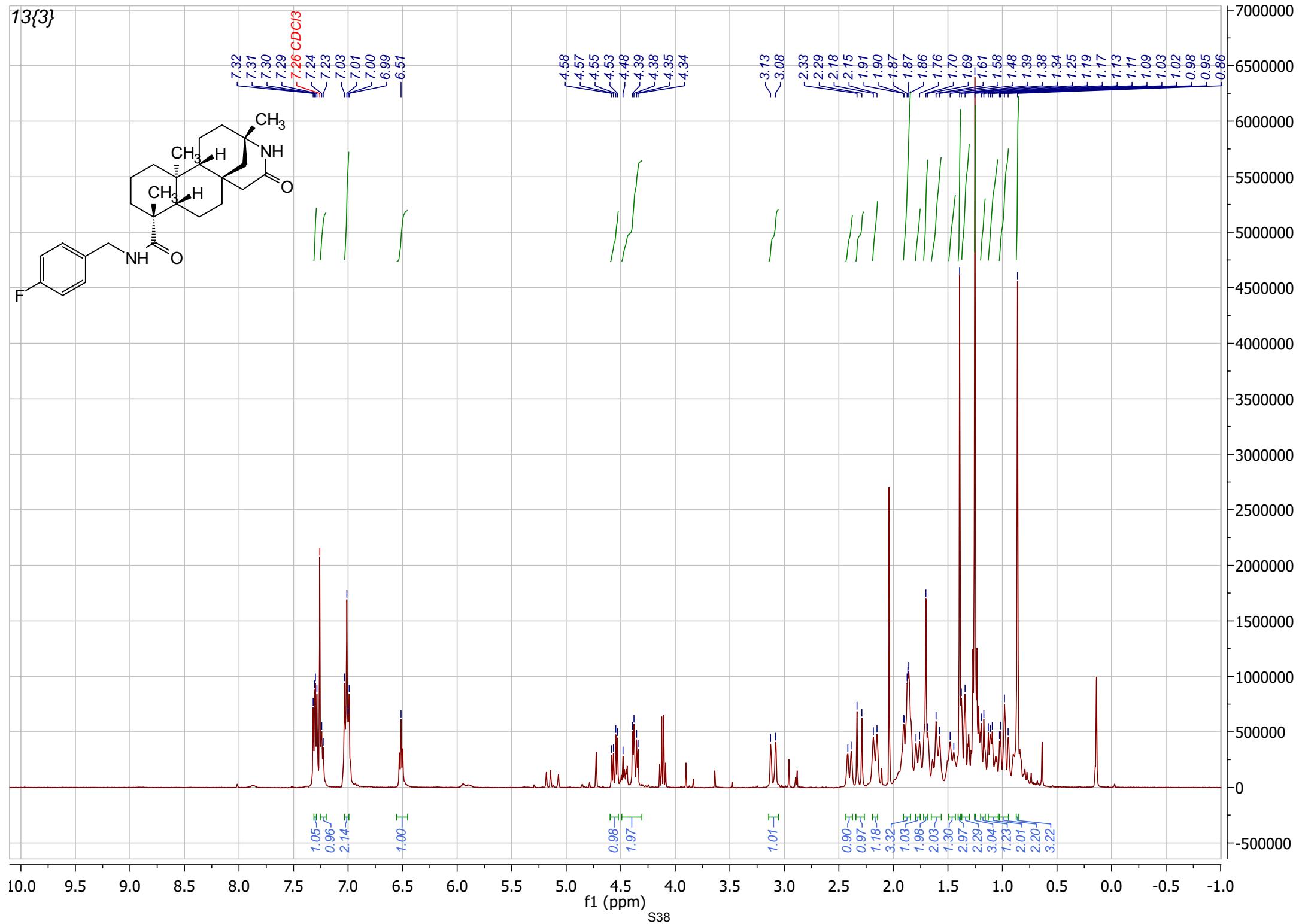
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13{1}

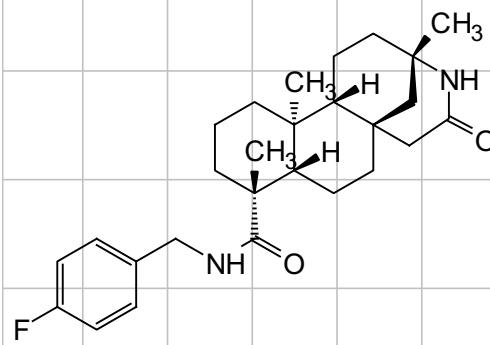


13{1}





13{3}

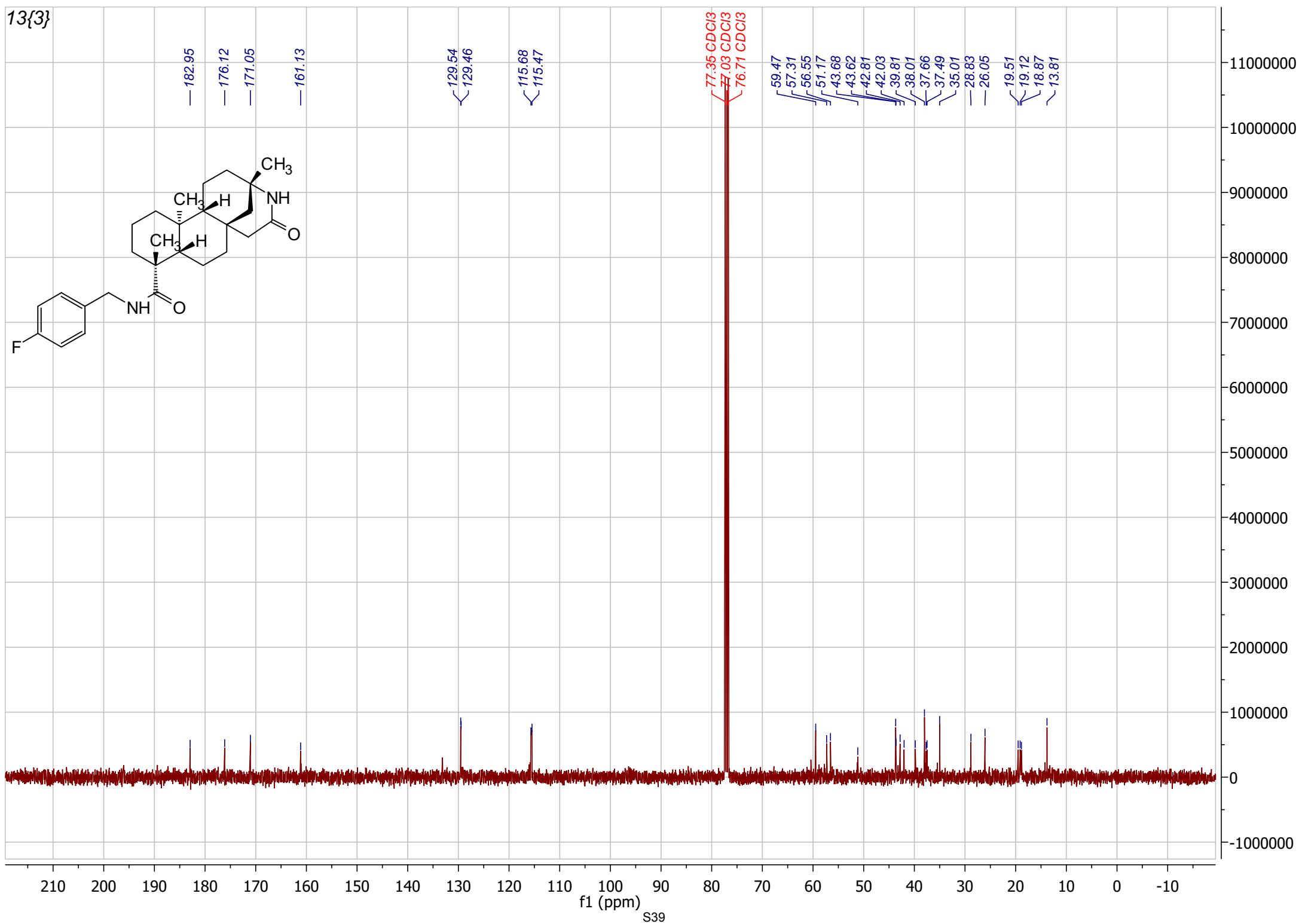


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— 171.05
— 161.13

— 129.54
— 129.46
— 115.68
— 115.47

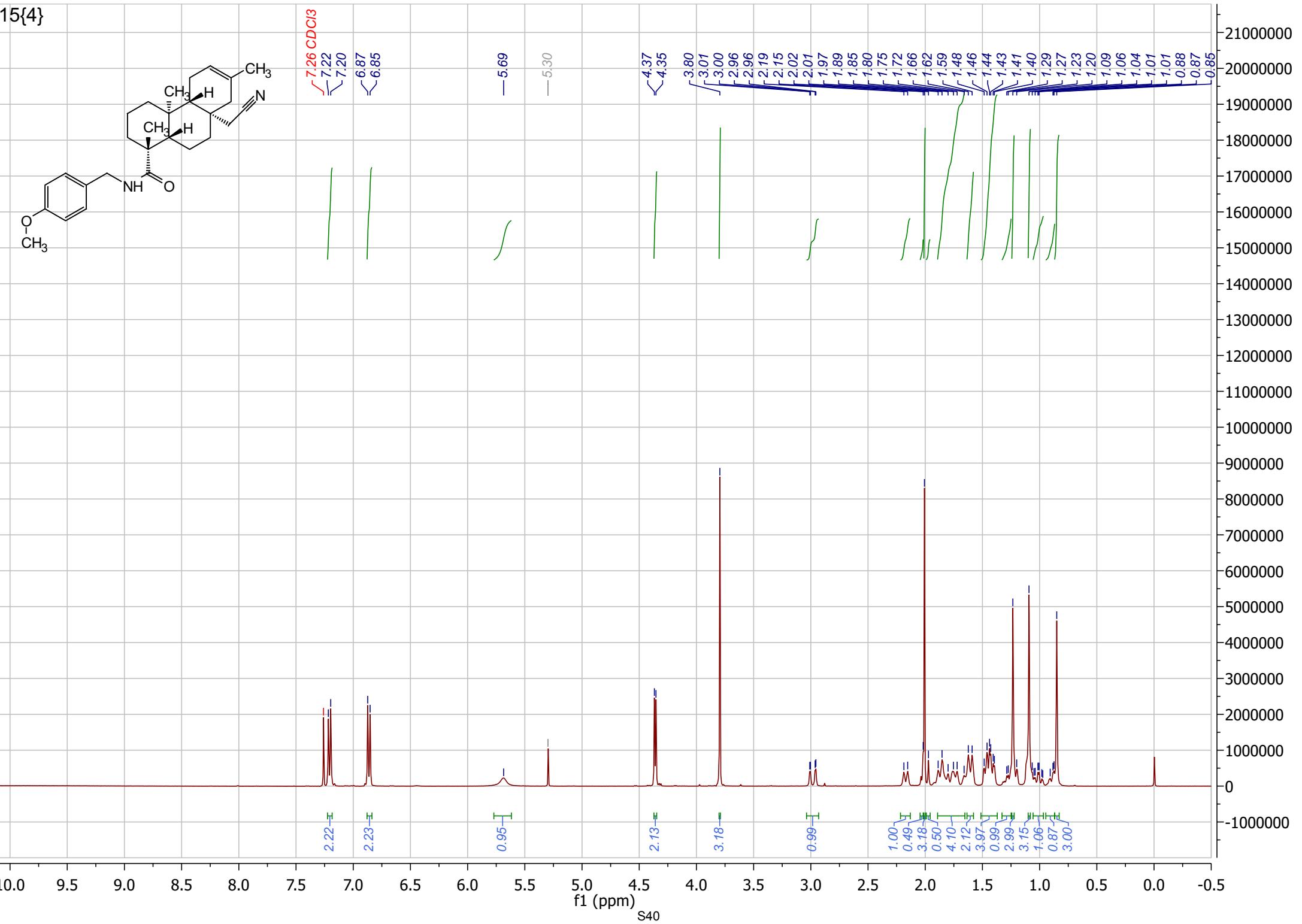
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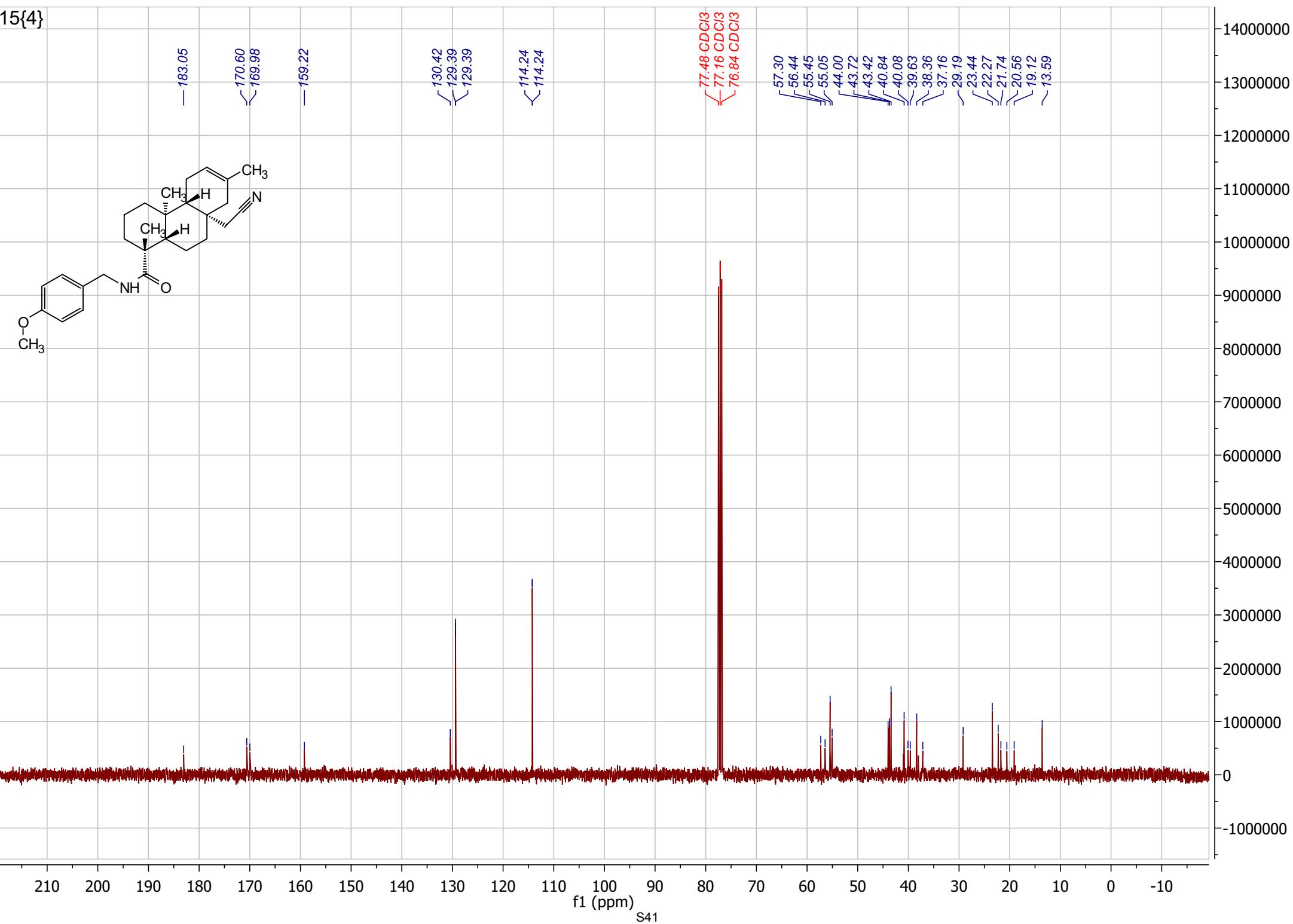
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— 13.81



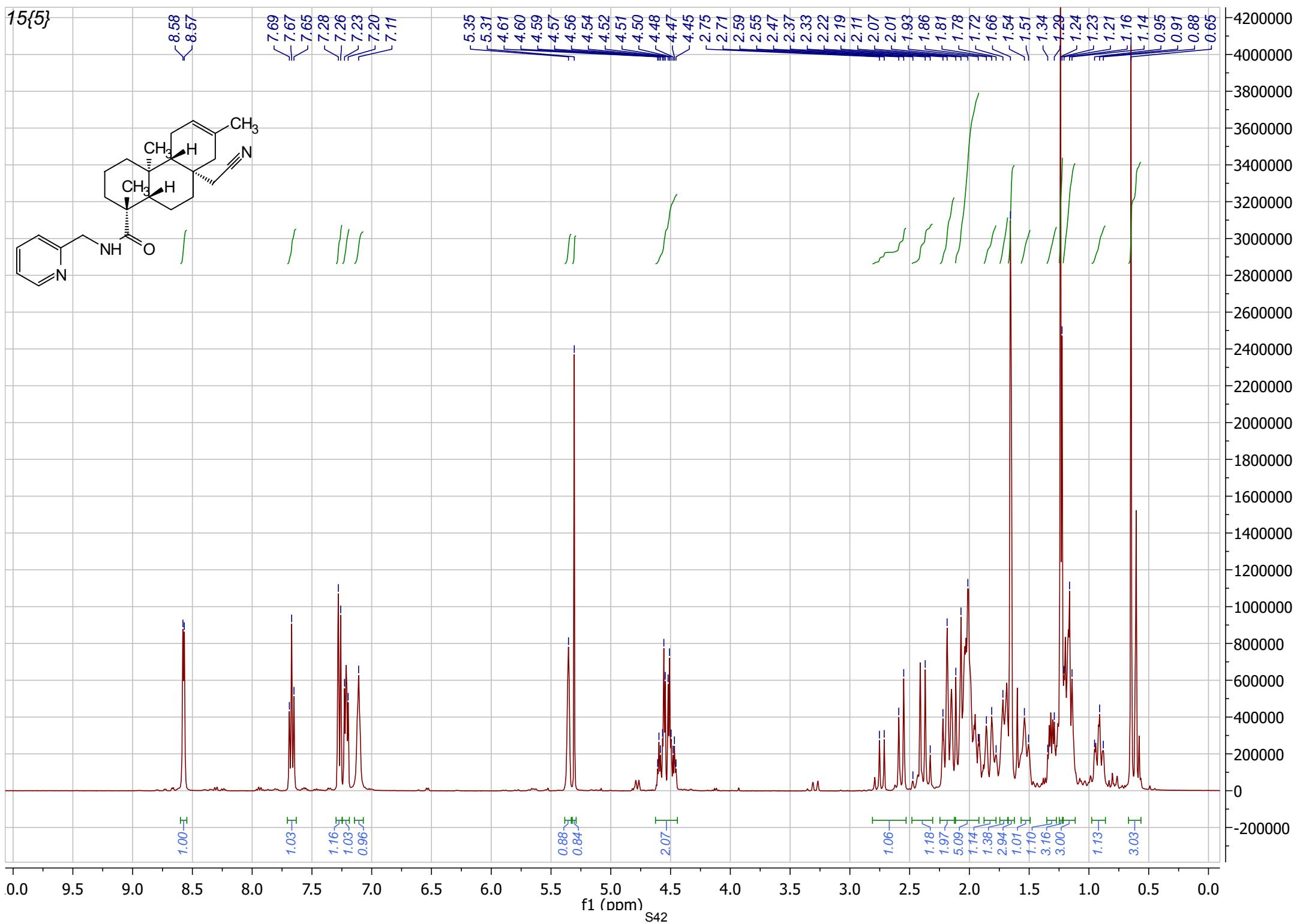
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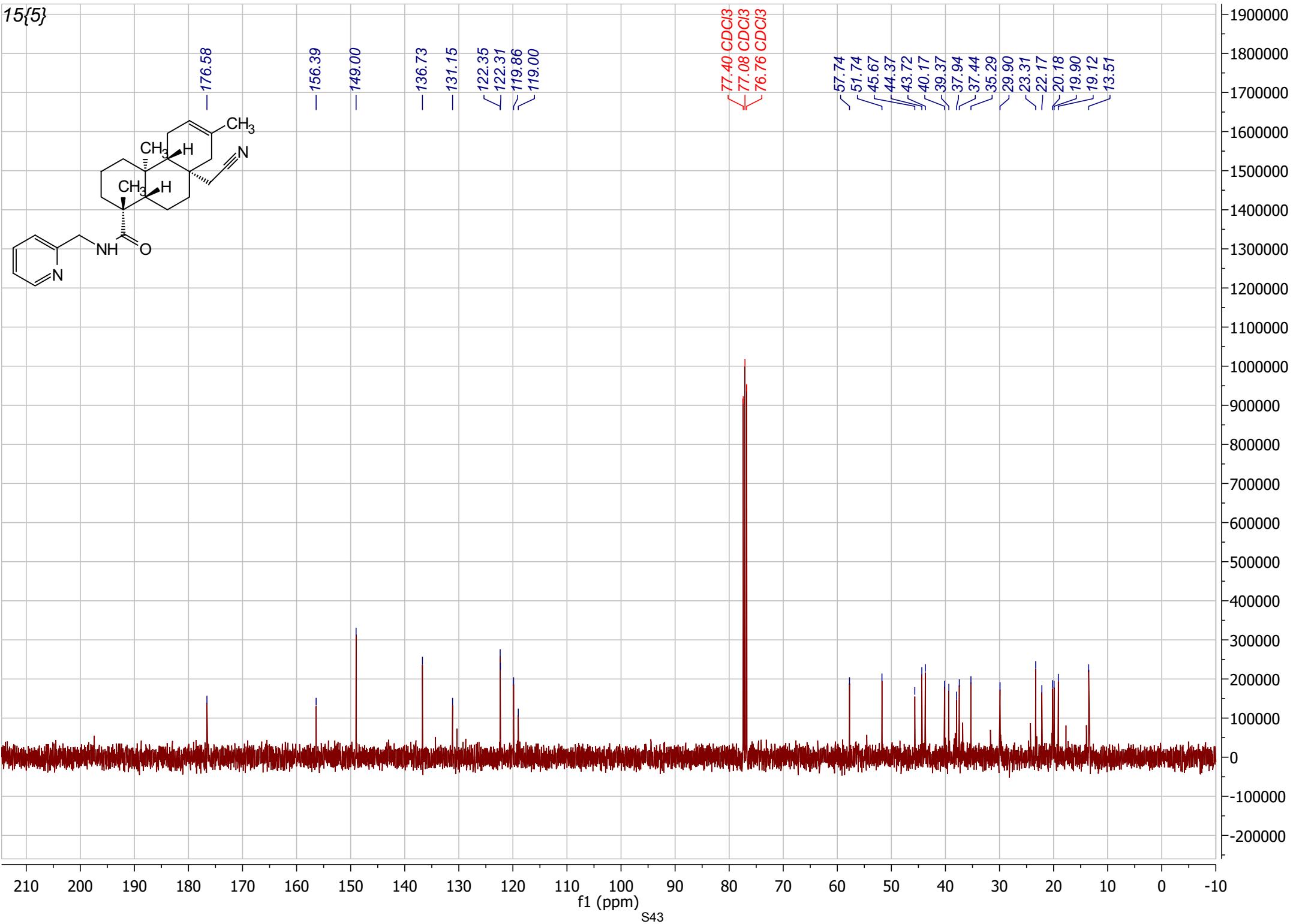
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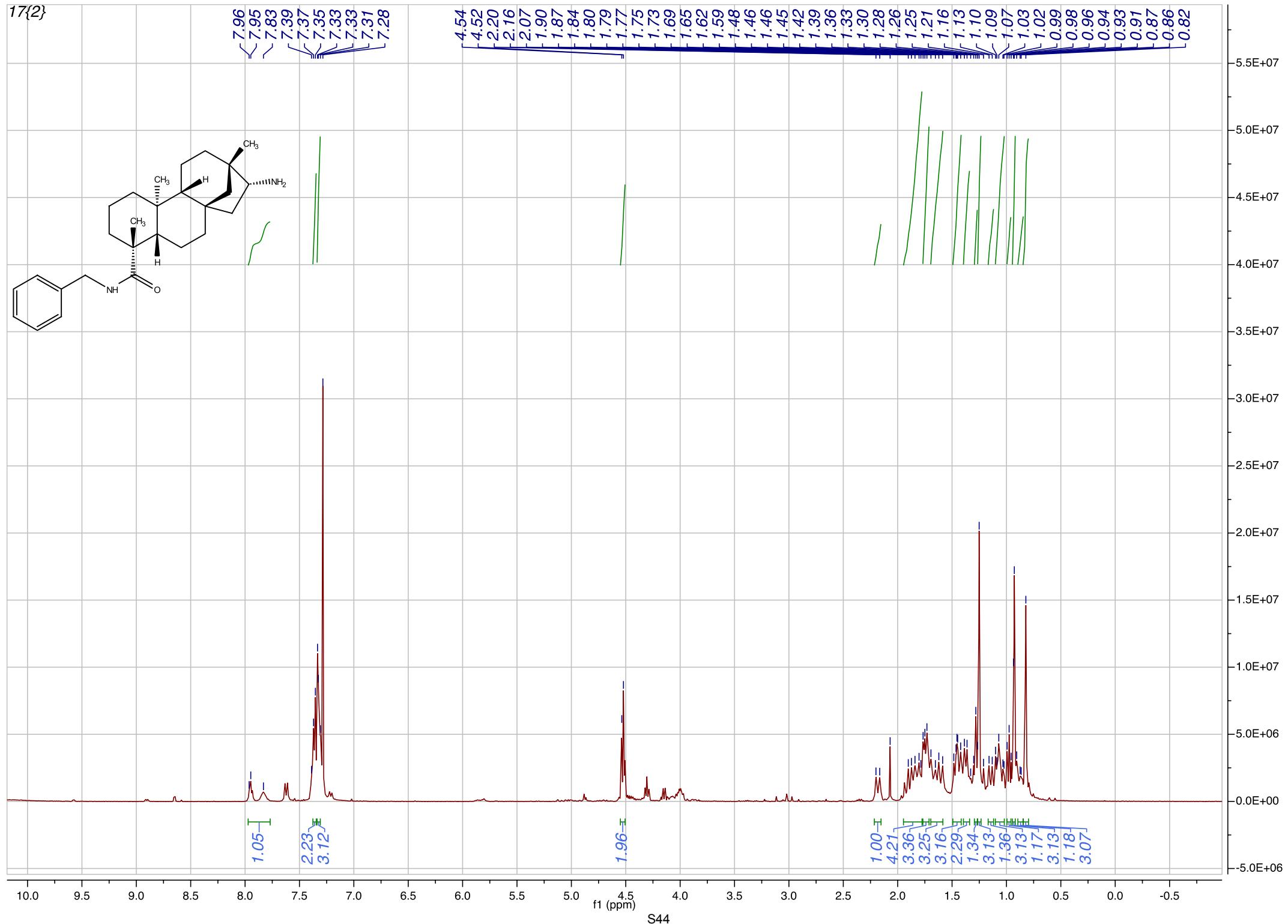


15{5}





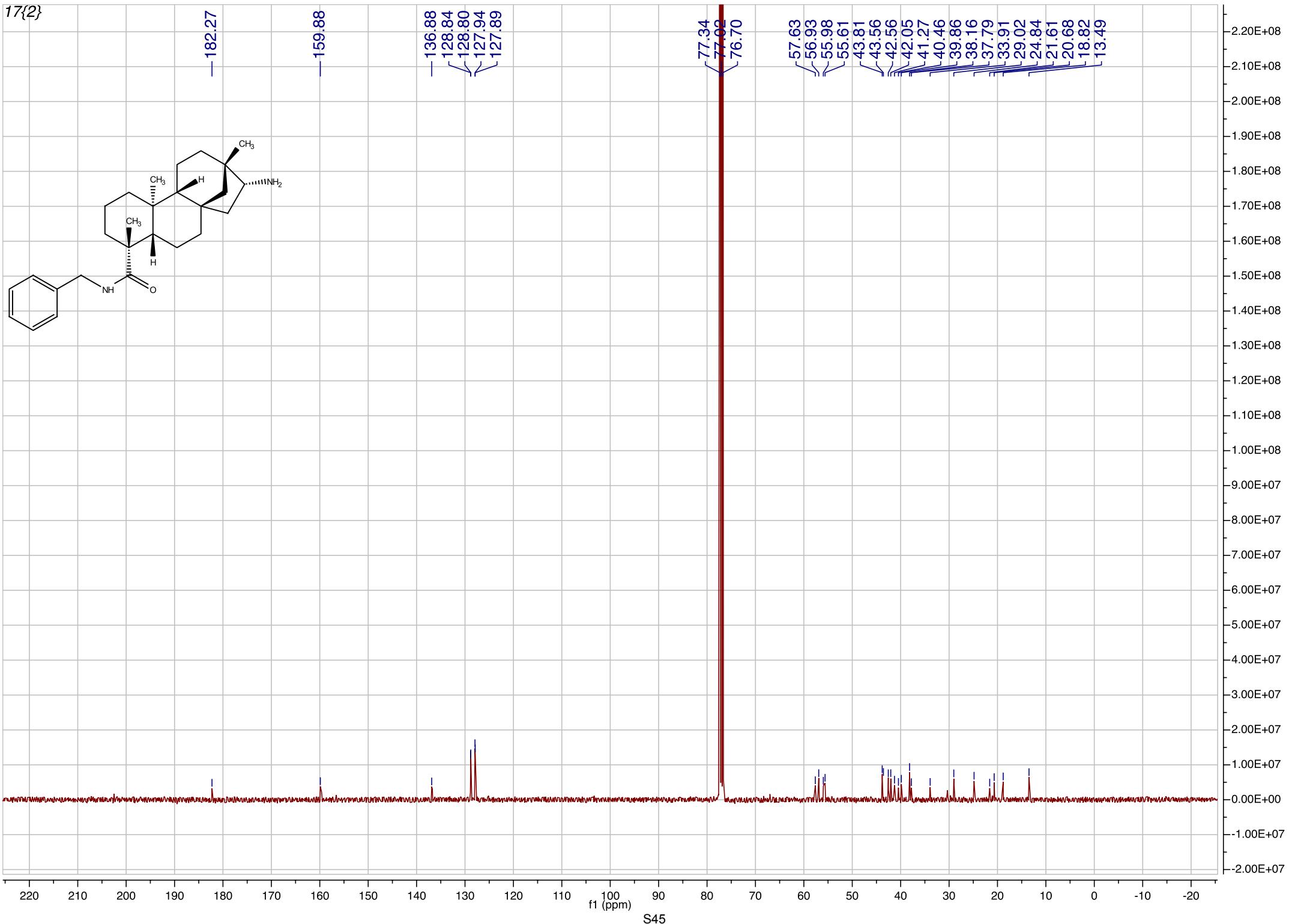
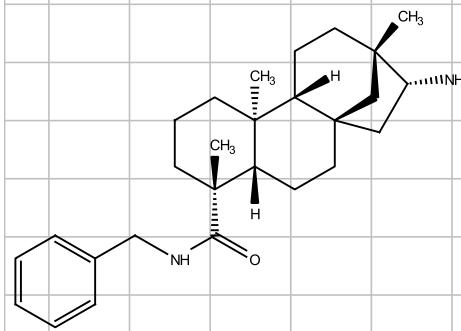
17{2}



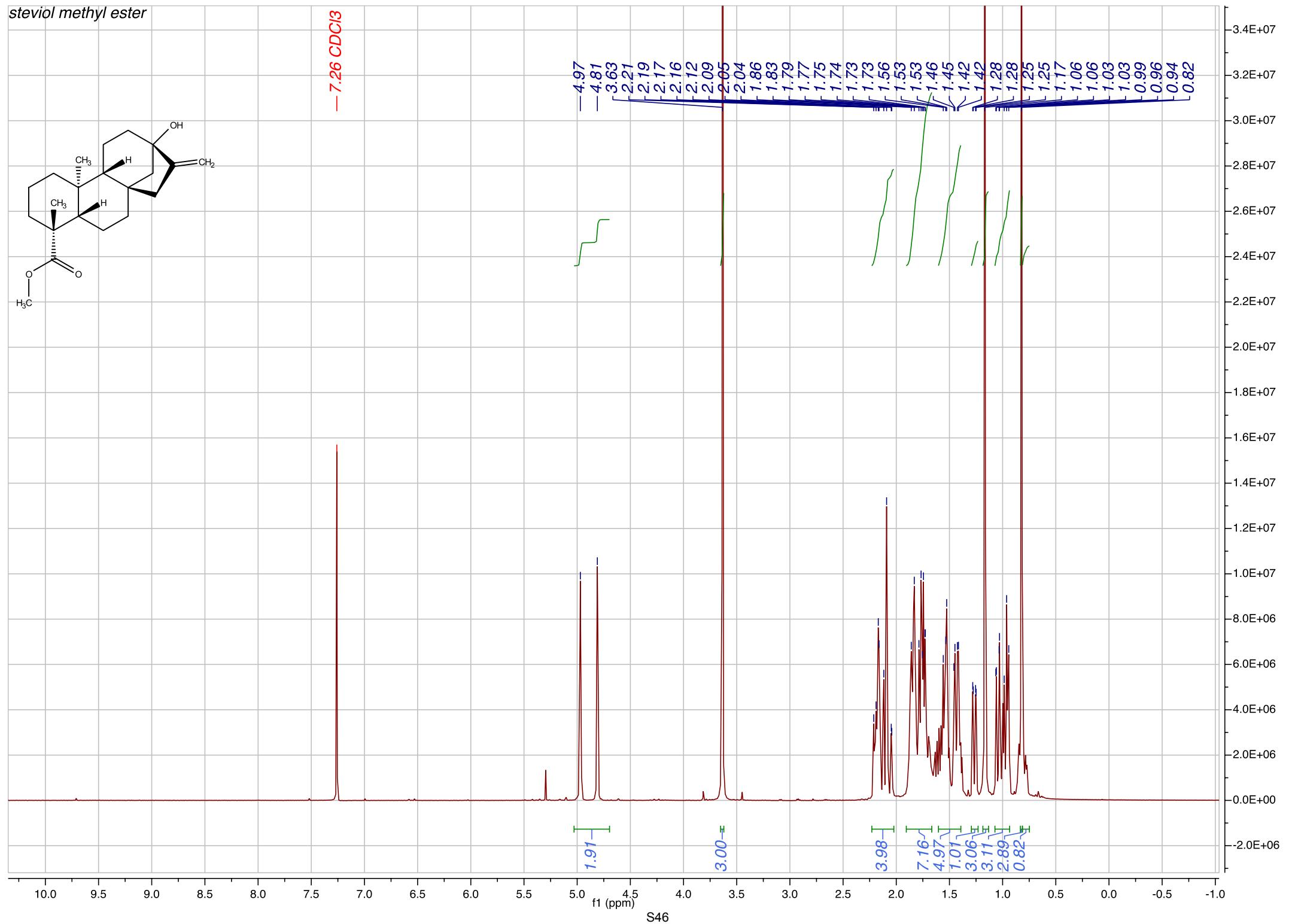
10.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 5.5 5.0 4.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 0.0 -0.5

S44

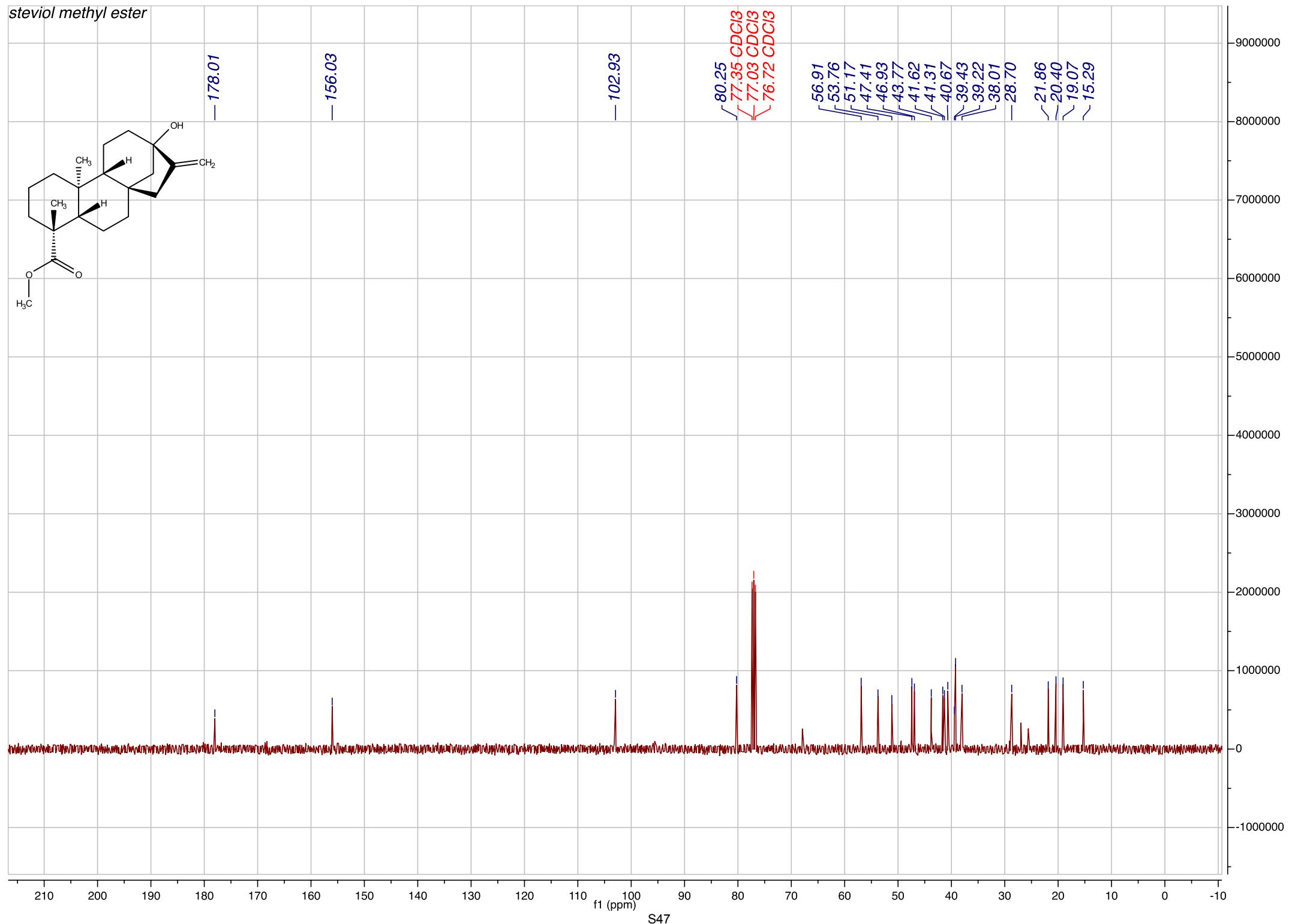
17{2}



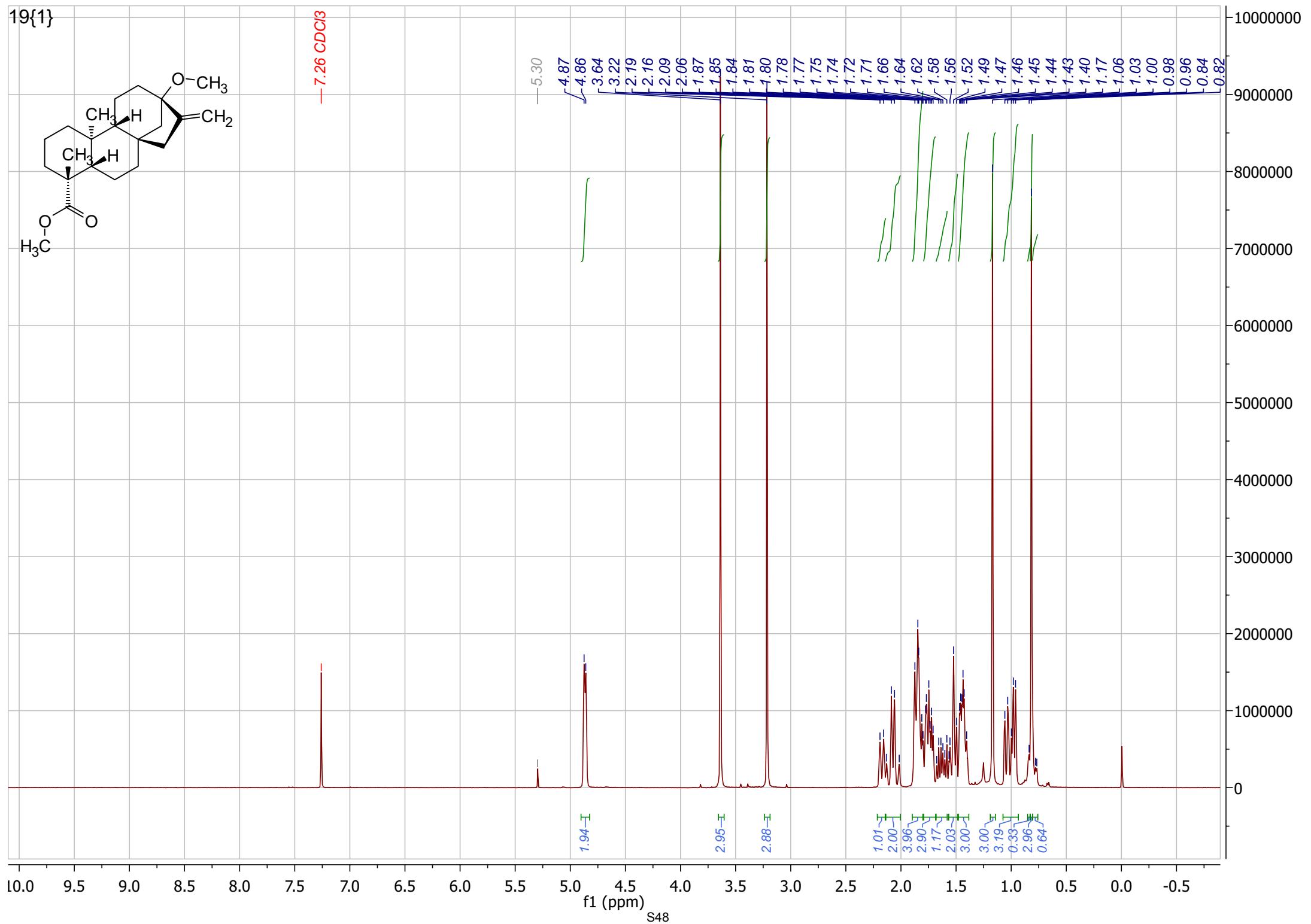
steviol methyl ester

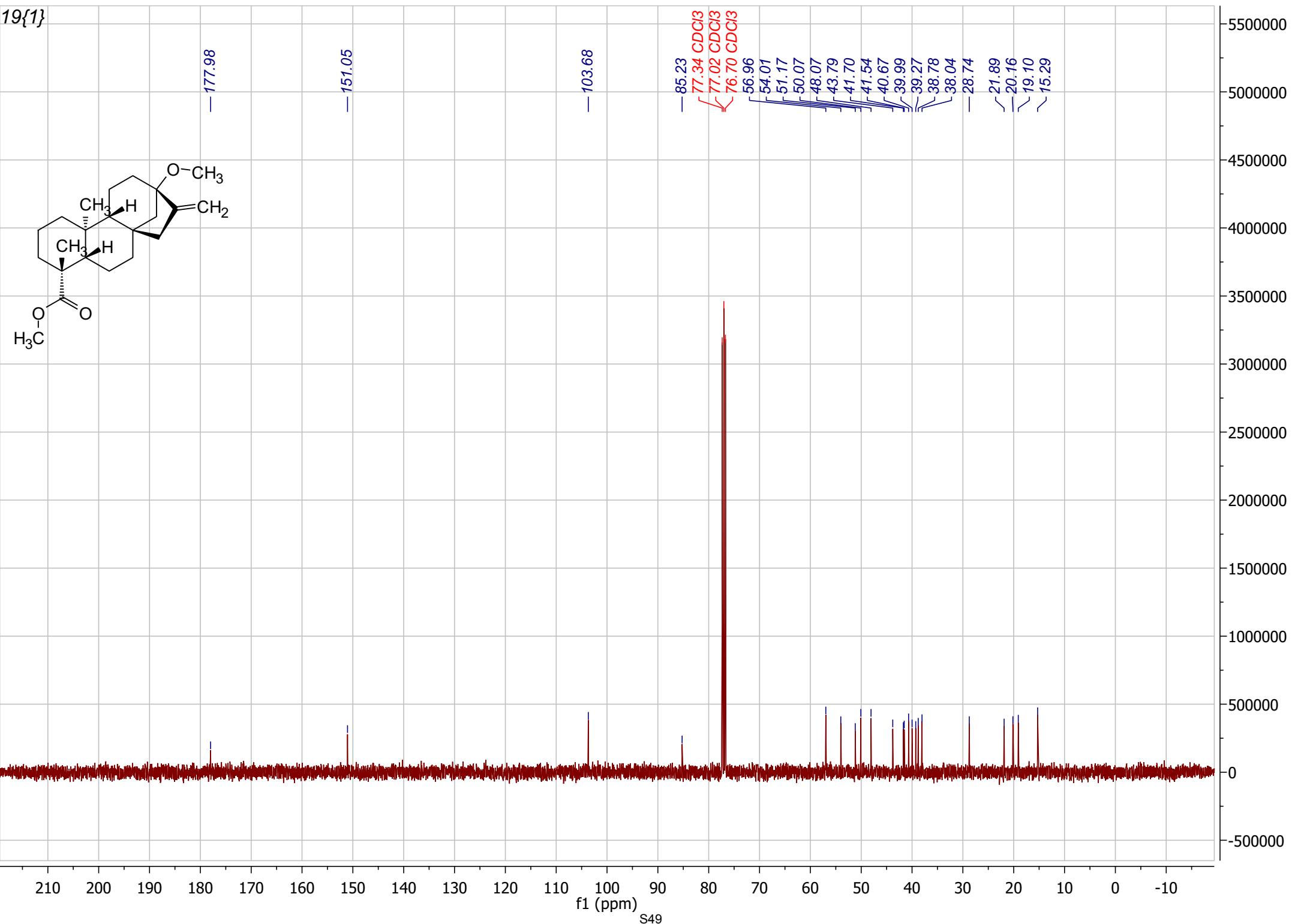


steviol methyl ester

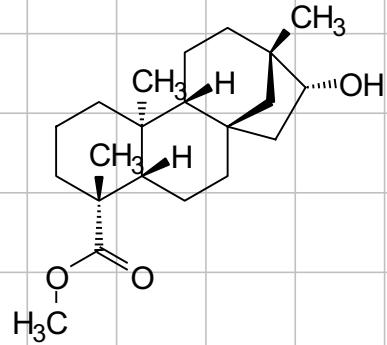


19{1}

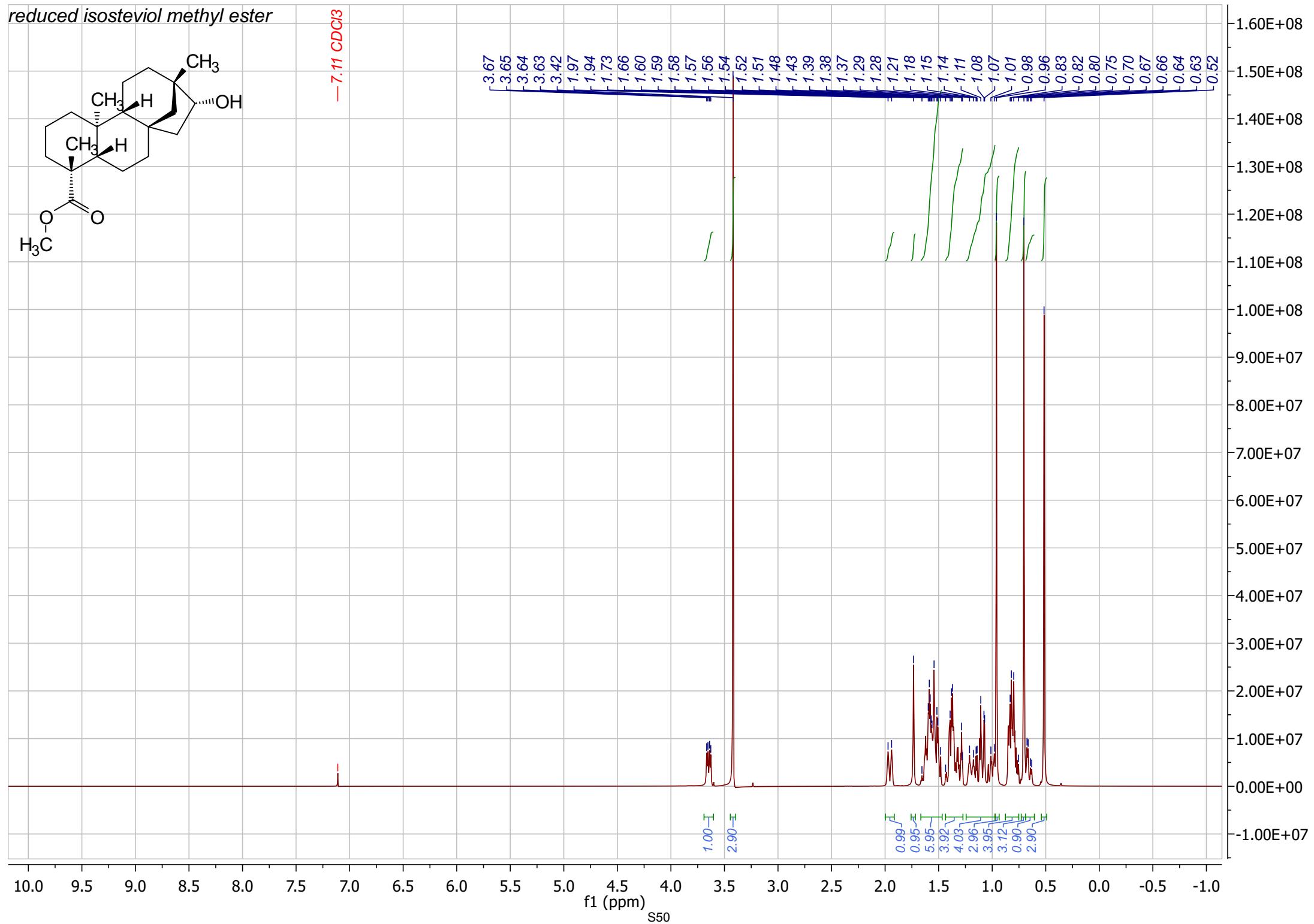




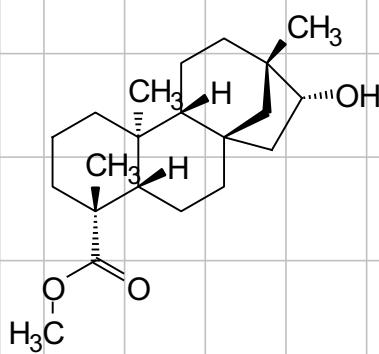
reduced isosteviol methyl ester



—7.11 CDCl₃



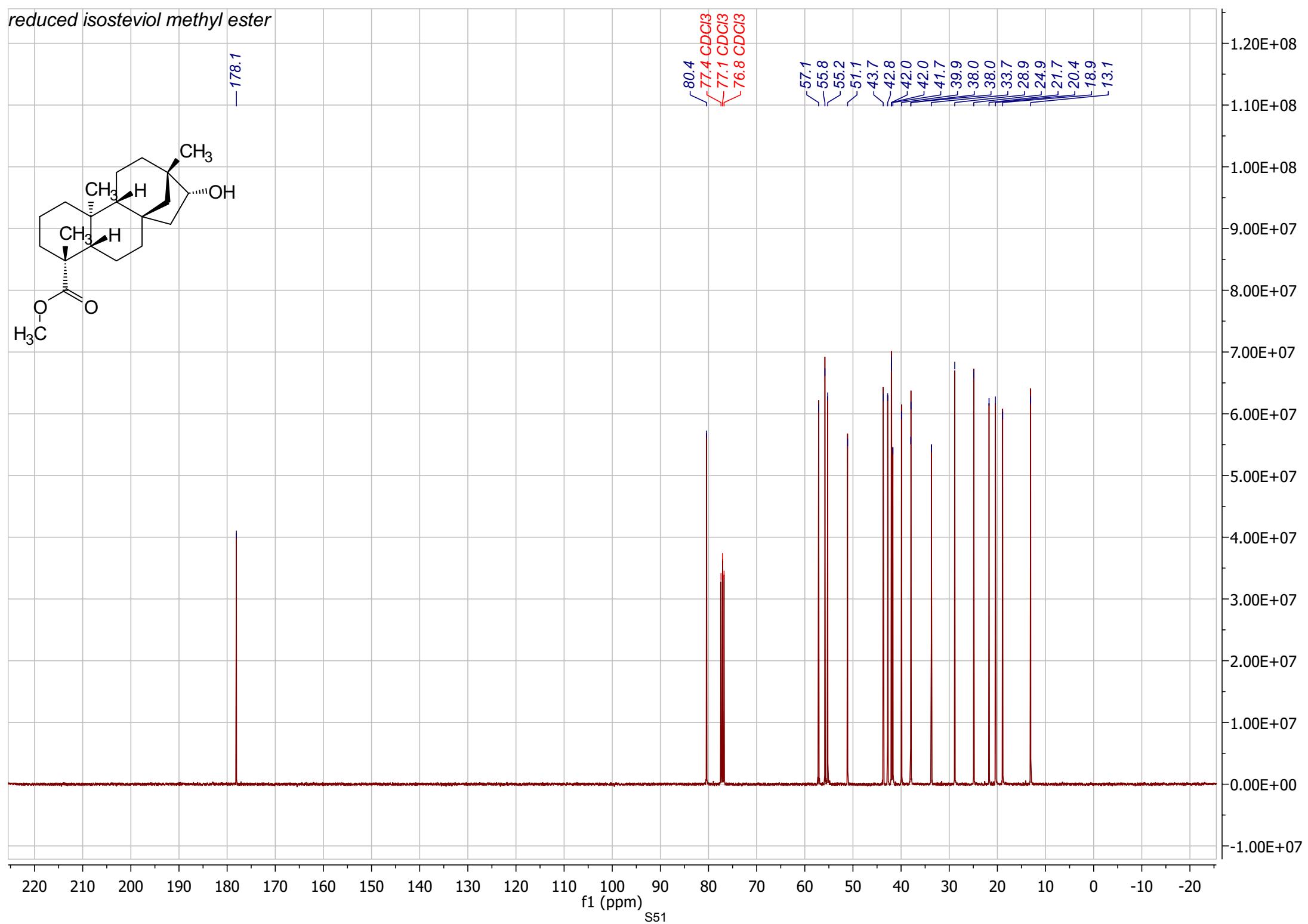
reduced isosteviol methyl ester

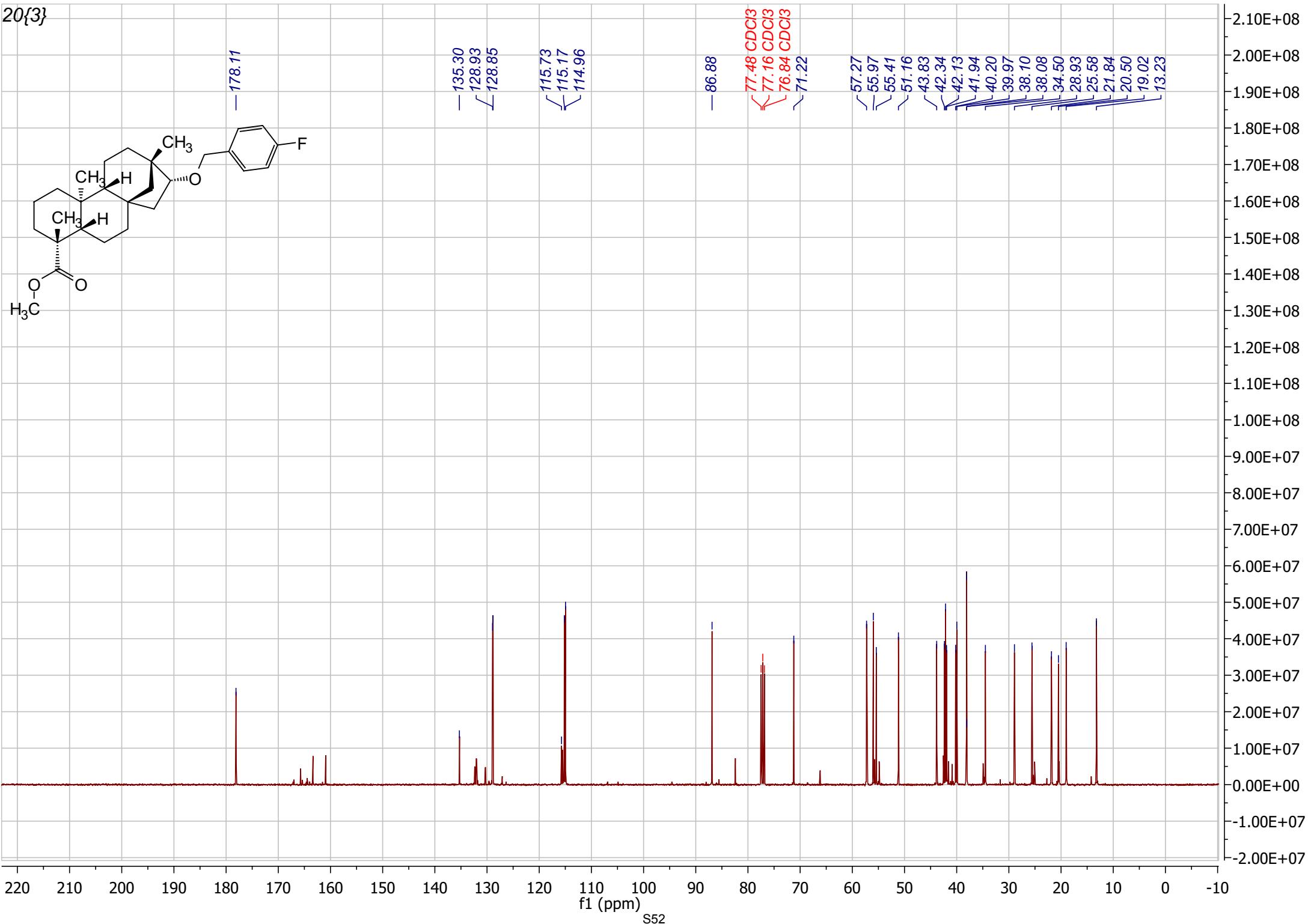


— 178.1

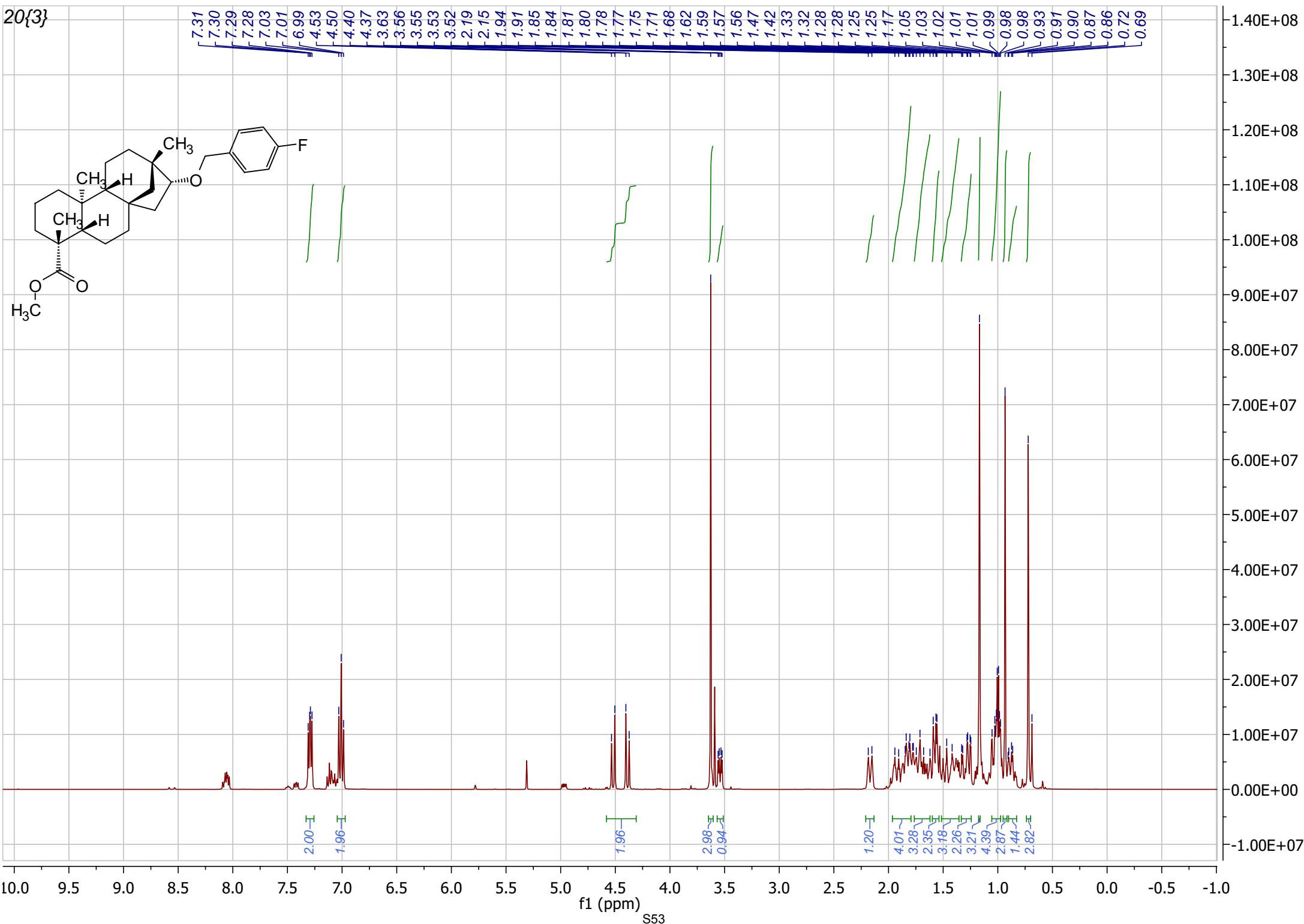
80.4
77.4 CDCl₃
77.1 CDCl₃
76.8 CDCl₃

57.1
55.8
55.2
51.1
43.7
42.8
42.0
41.7
39.9
38.0
38.0
33.7
28.9
24.9
21.7
20.4
18.9
13.1

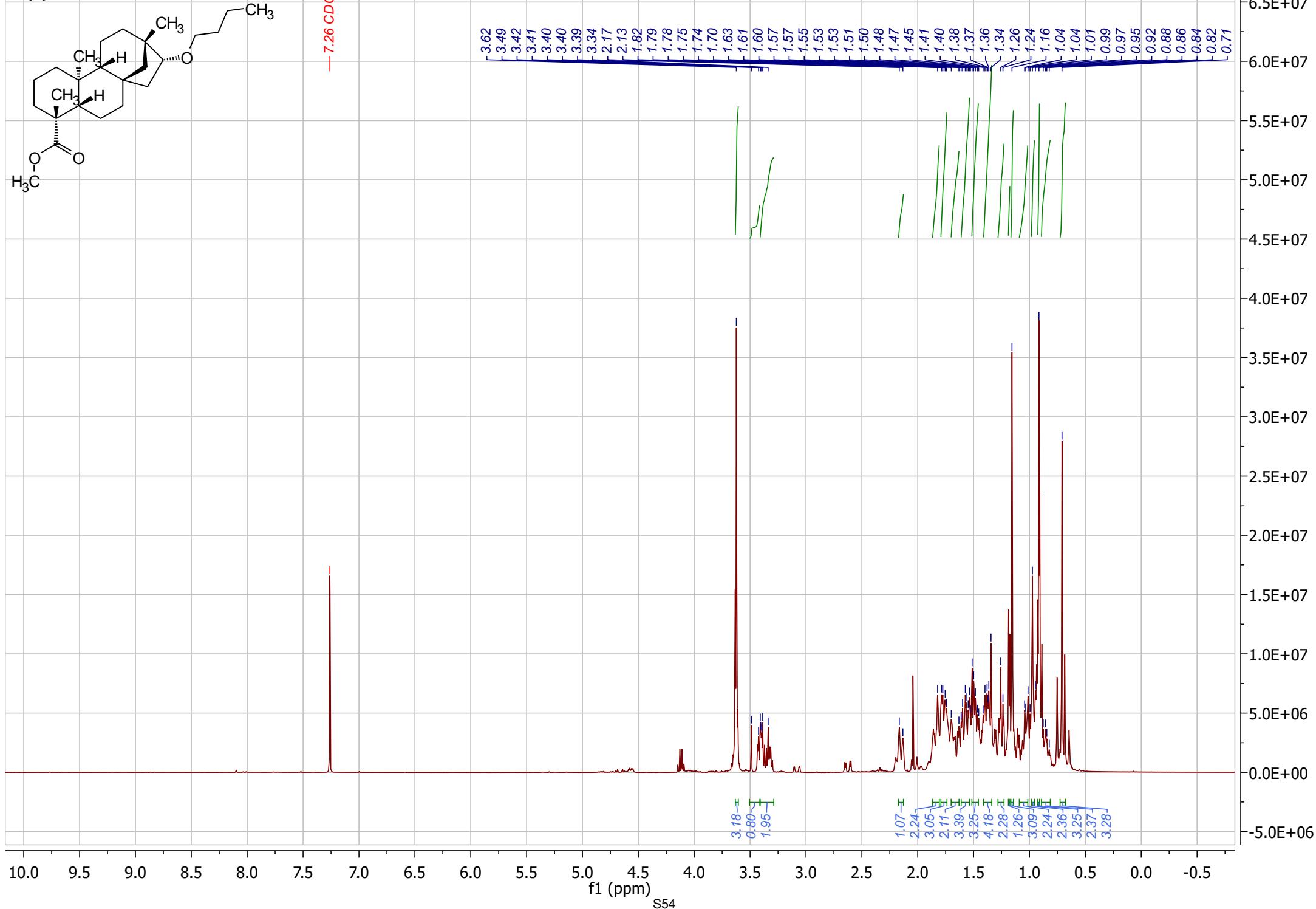




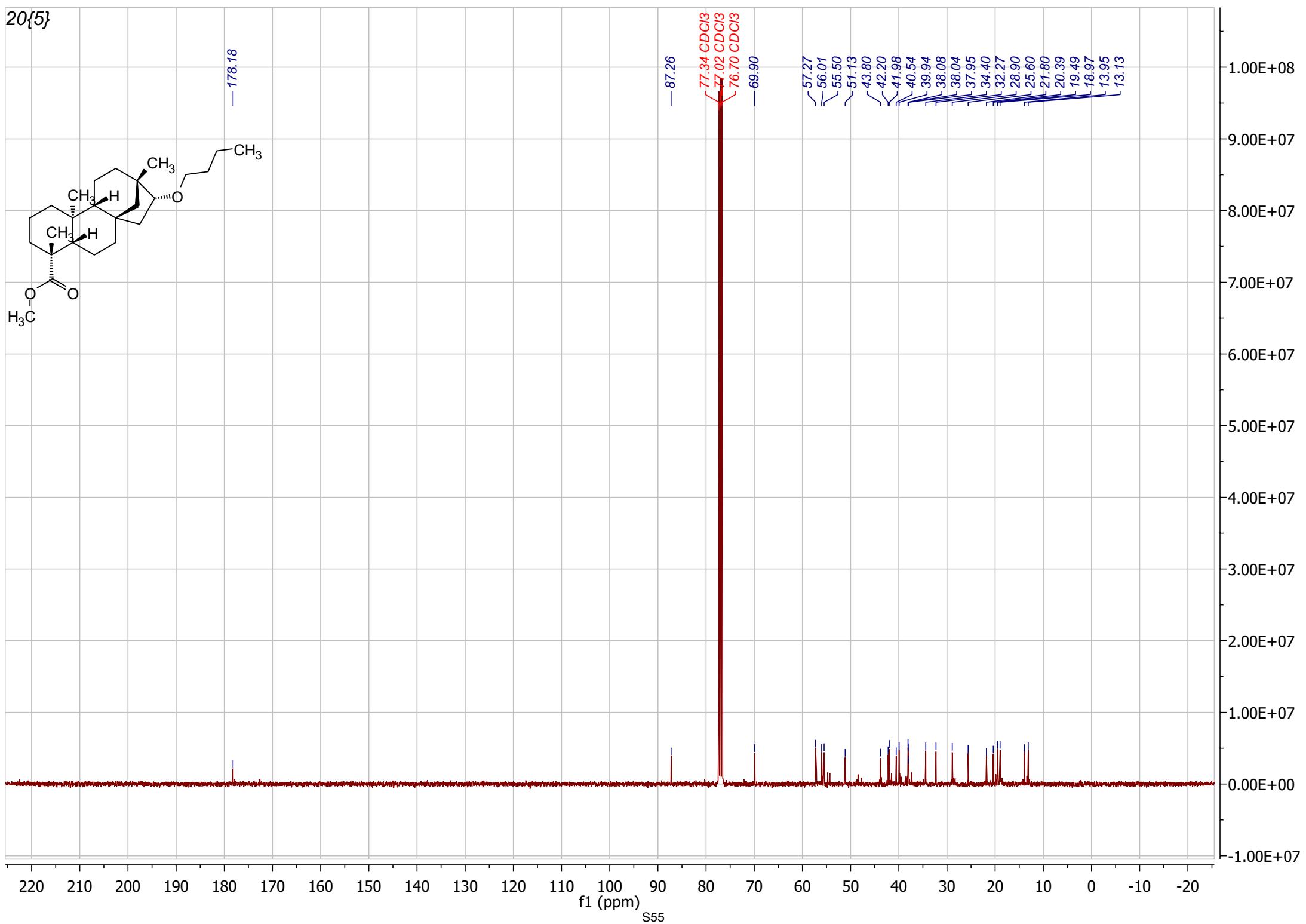
20{3}



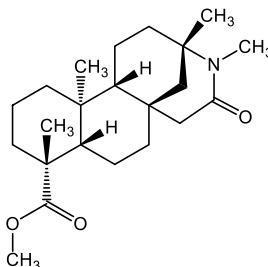
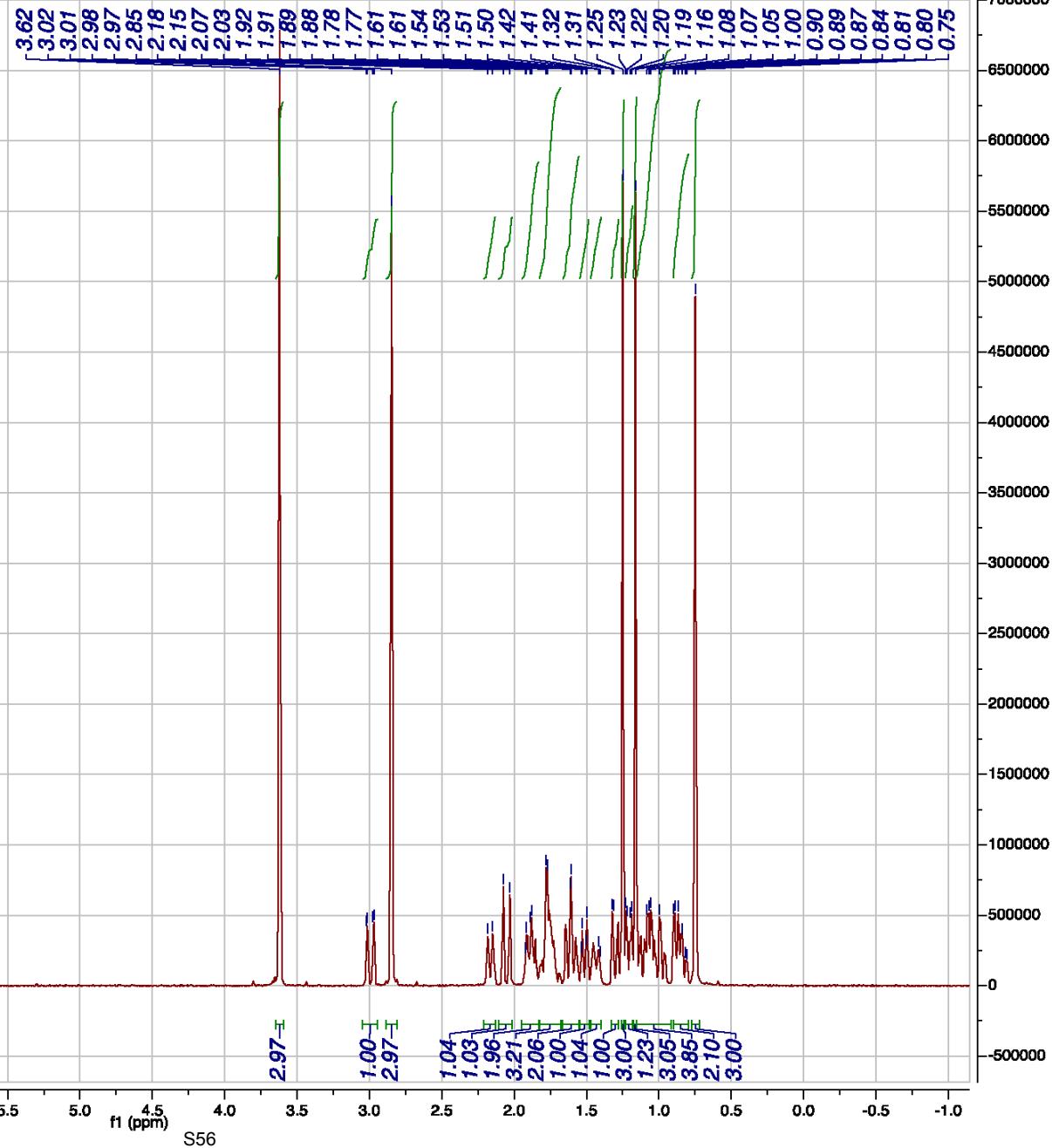
20{5}



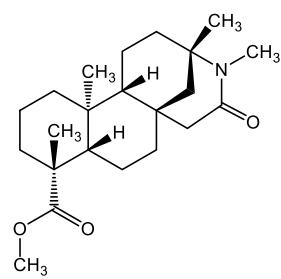
20{5}



21{1}

-7.26 CDCl₃

21{1}



-177.59

-171.79

77.16 CDCl₃76.94 CDCl₃76.52 CDCl₃

57.30
56.69
55.25
51.04
50.84
44.20
43.62
40.82
39.84
37.90
37.68
35.55
33.77
28.36
27.32
26.76
19.59
18.71
18.57
13.39

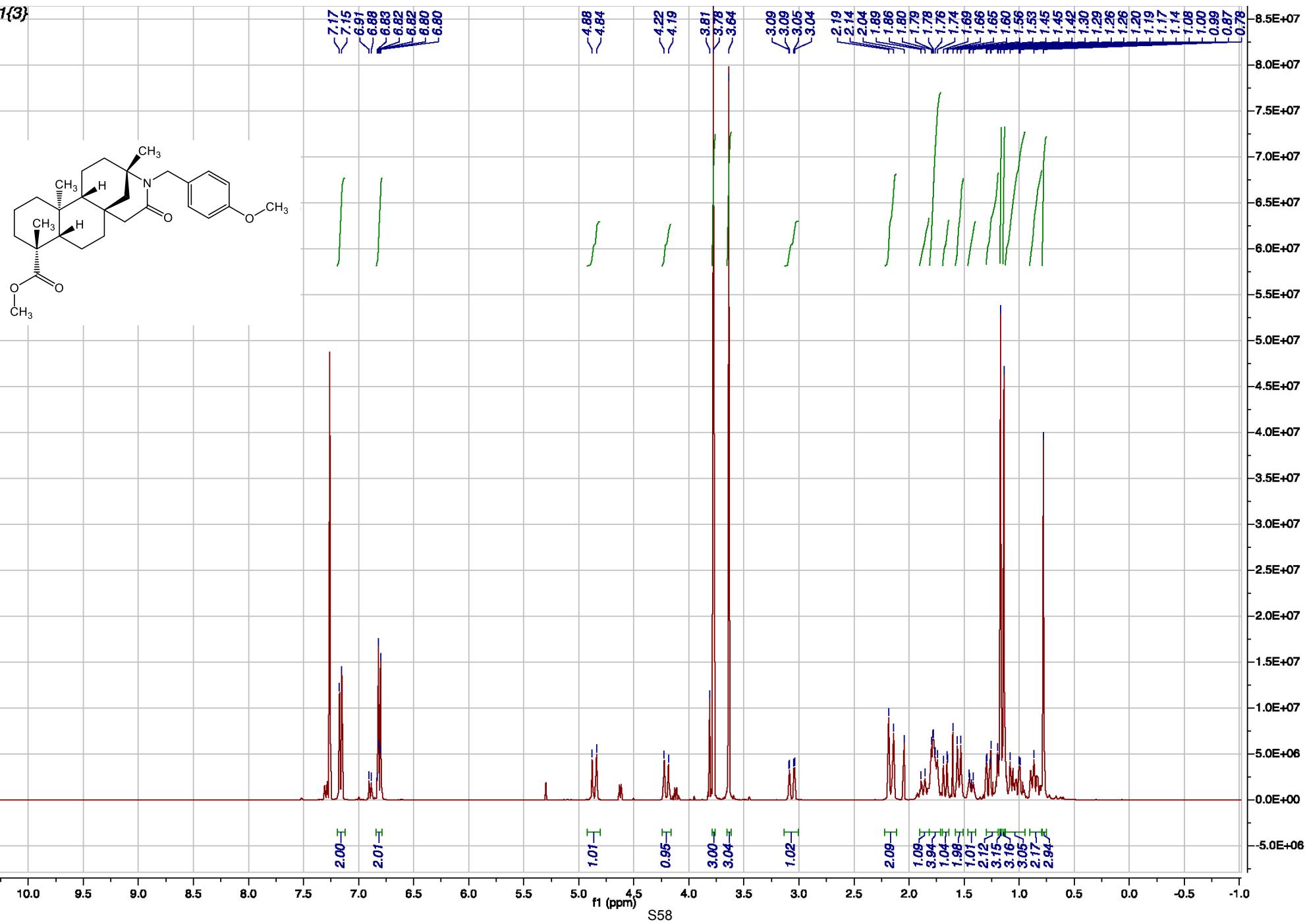
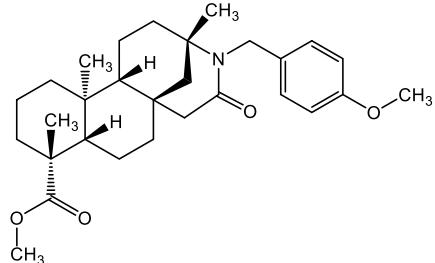
210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0 -10

f1 (ppm)

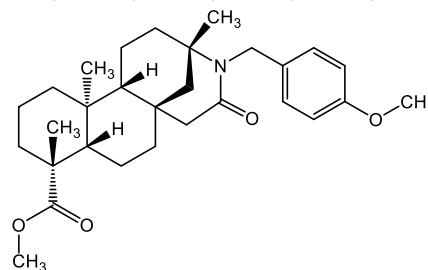
S57

1.30E+08
1.20E+08
1.10E+08
1.00E+08
9.00E+07
8.00E+07
7.00E+07
6.00E+07
5.00E+07
4.00E+07
3.00E+07
2.00E+07
1.00E+07
0.00E+00
-1.00E+07

21{3}



21{3}



-177.8
 -172.3

-158.3

-132.1
 -128.3

-113.7

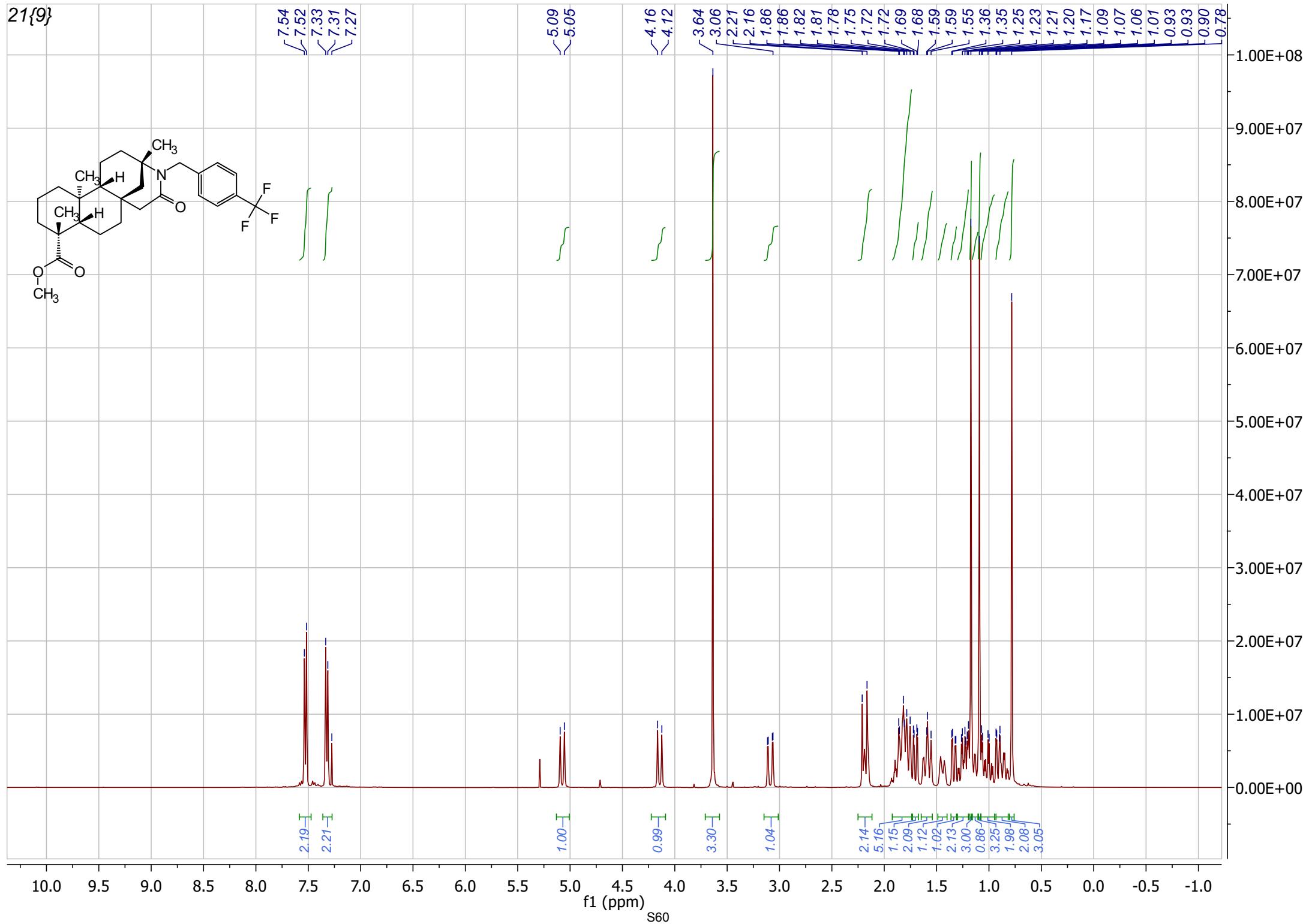
$77.3 \text{ } ^\text{13}^\text{C}\text{DCD}_3$
 $77.0 \text{ } ^\text{13}^\text{C}\text{DCD}_3$
 $76.7 \text{ } ^\text{13}^\text{C}\text{DCD}_3$

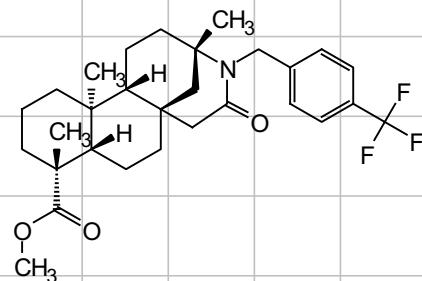
57.5
 56.9
 56.4
 55.2
 51.7
 51.2
 44.3
 44.0
 43.8
 41.2
 40.0
 38.0
 37.8
 37.1
 34.1
 28.6
 28.1
 19.8
 18.9
 18.6
 13.7

210 200 190 180 170 160 150 140 130 120 110 f_1 (ppm) 100 90 80 70 60 50 40 30 20 10 0 -10

S59

21{9}





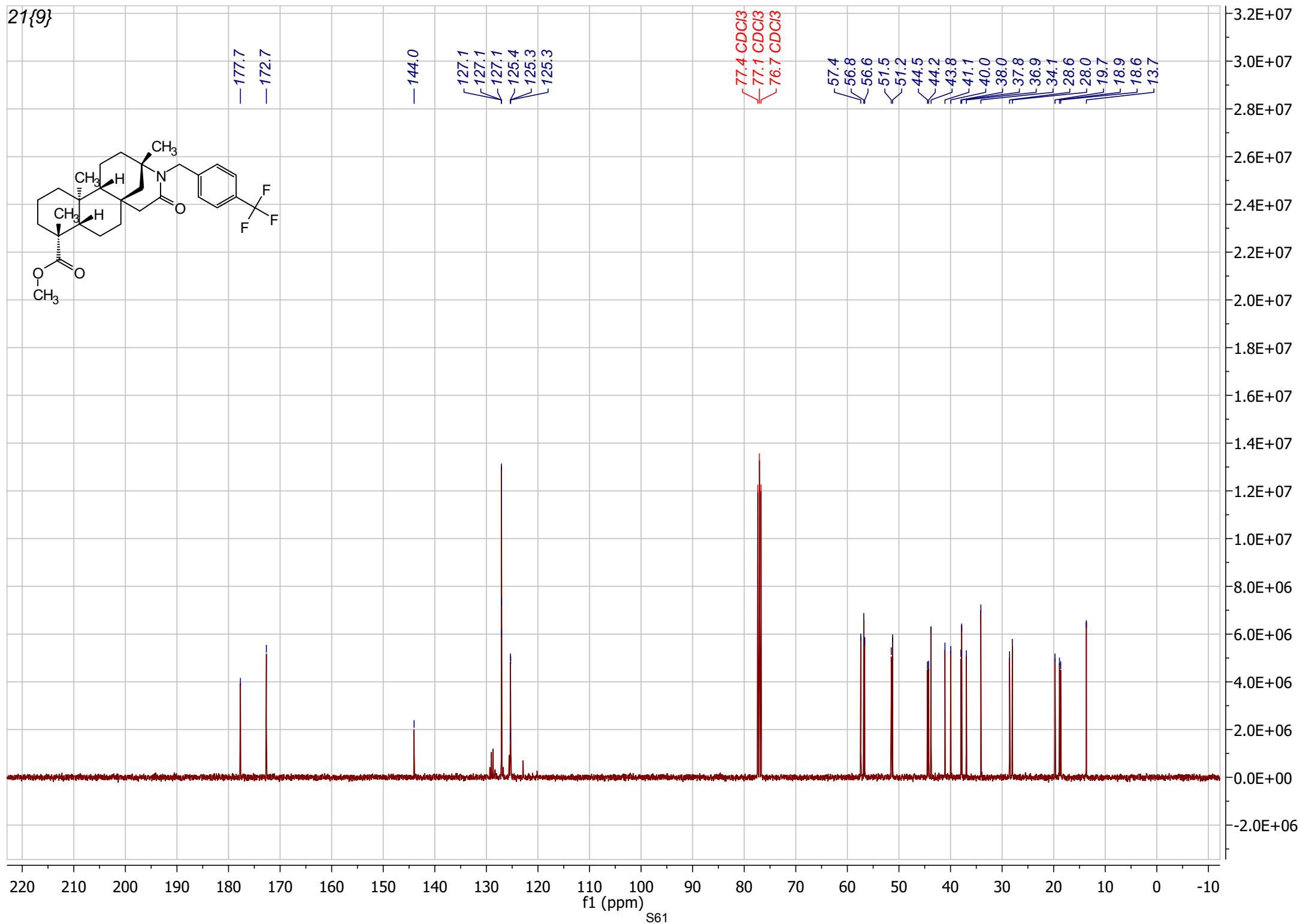
-177.7
-172.7

-144.0

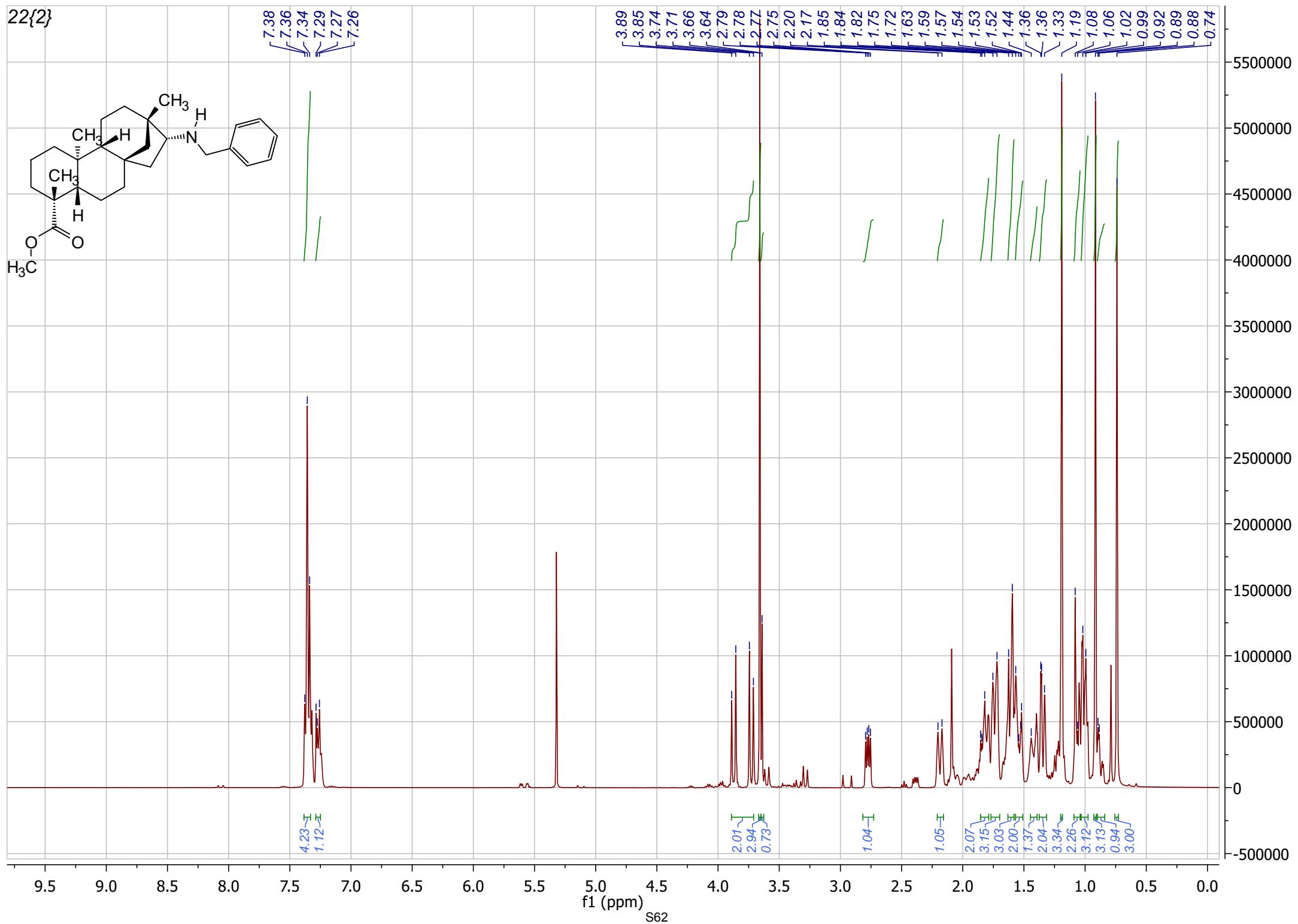
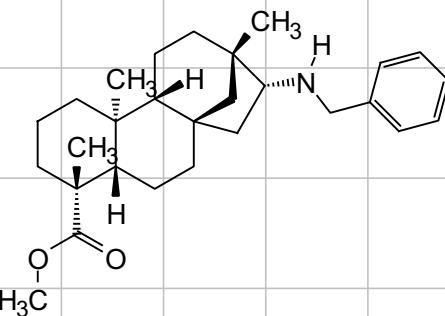
127.1
127.1
127.1
125.4
125.3
125.3

77.4 CDCl₃
77.1 CDCl₃
76.7 CDCl₃

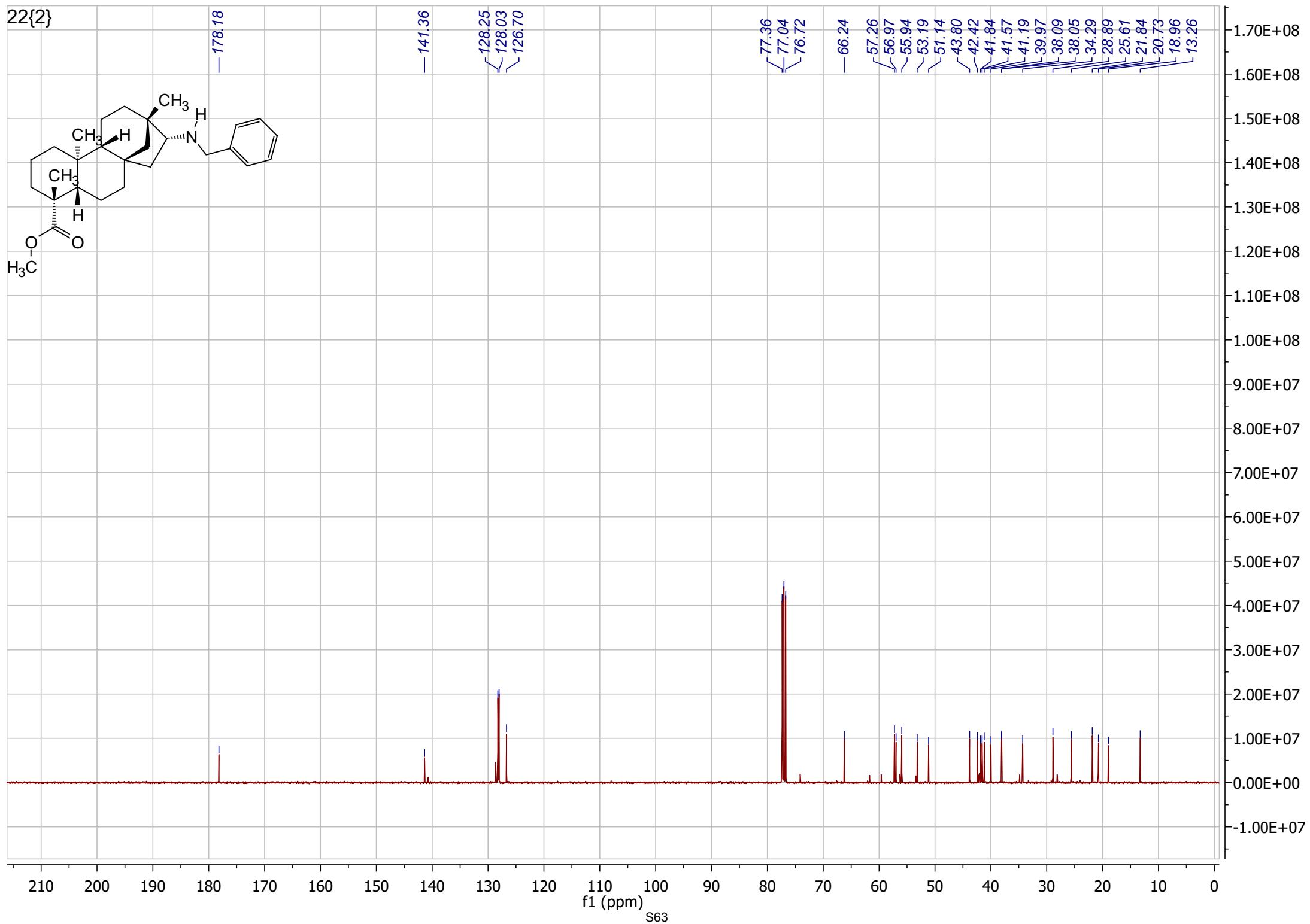
57.4
56.8
56.6
51.5
51.2
44.5
44.2
43.8
41.1
40.0
38.0
37.8
36.9
34.1
28.6
28.0
19.7
18.9
13.7

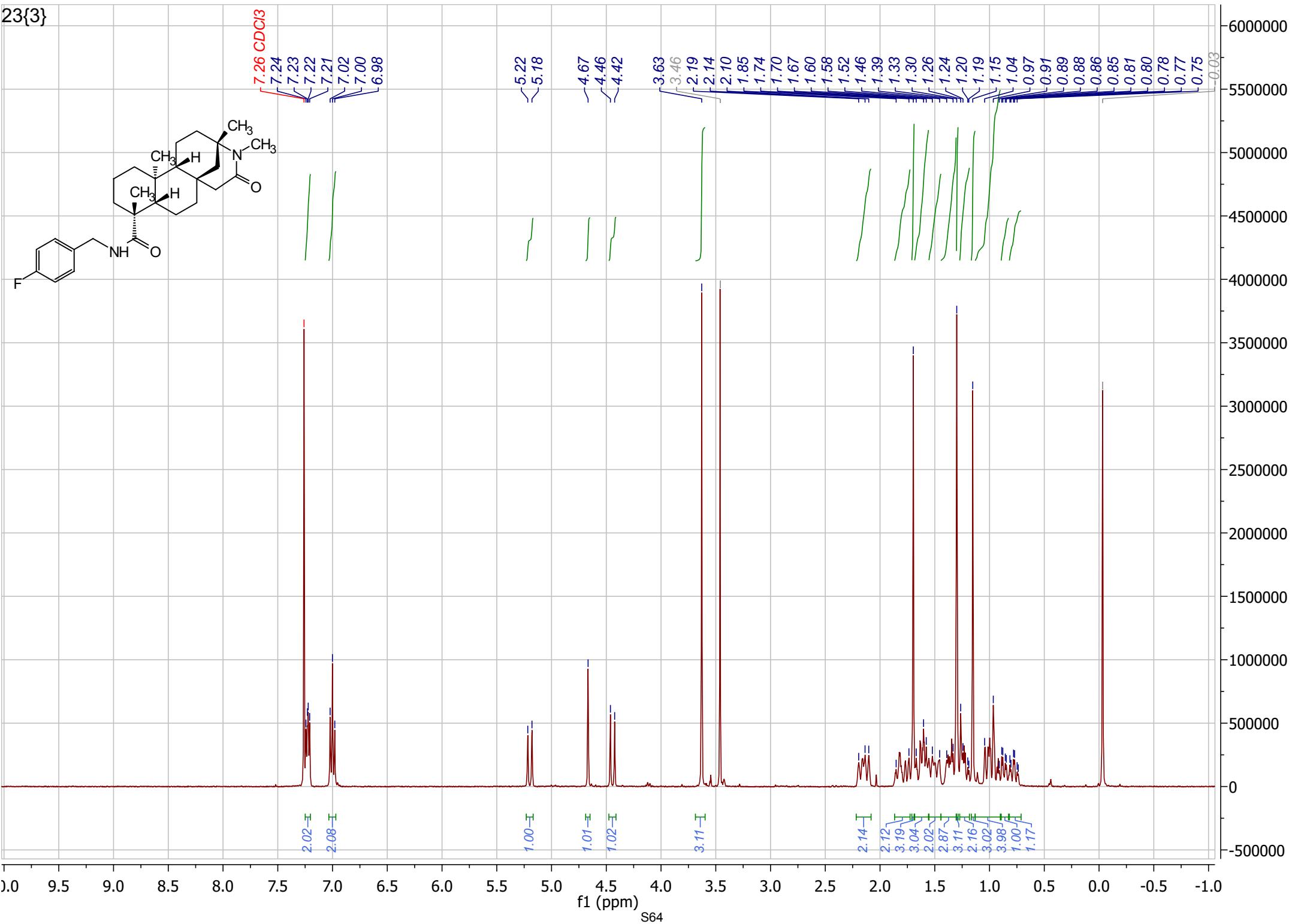


22{2}



22{2}





23{3}

