

# Radioiodinated Nitroxide Derivative for the Detection of Lipid Radicals

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## Table of Contents

1. Experimental Procedure	S2
2. NMR spectra	S4
3. ESR spectra	S5
4. HPLC chromatograms	S6

## 1. Synthetic Experimental Procedure

### General Methods

Reagents and solvents from commercial suppliers were used without further purification. The reaction was monitored by thin layer chromatography on silica gel 60 plates. The compounds were detected by examination under UV light and by staining with iodine vapor or 5% phosphomolybdic acid EtOH solution. Flash column chromatography was performed with prepacked silica gel column. HRMS were measured on a Thermo Fisher Scientific Exactive spectrometer. NMR data for **2** and **3** were obtained by *in situ* reduction with phenylhydrazine (PhNHNH<sub>2</sub>) to reduce the broadening effect derived from the paramagnetism. ESR spectra were obtained by X-band ESR spectrometer in DMSO solution at rt. The purity was checked by HPLC measurement. The measurement condition was described in Figure S5 and S6 legend.

**4-(4-Iodobenzamido)-2,2,6,6-tetramethylpiperidine-1-oxyl (2).** To a solution of **1** (0.50 g, 2.9 mmol) in THF (3 mL), 4-iodobenzoic acid (0.72 g, 2.9 mmol) and DMT-MM (1.22 g, 4.4 mmol) were added and stirred for 1.5 hours at rt. After extraction with EtOAc and drying over Na<sub>2</sub>SO<sub>4</sub>, the solvent was evaporated under reduced pressure, and purification was performed by flash column chromatography using hexane: EtOAc = 2: 1 (R<sub>f</sub>: 0.23) as an elution solvent to afford orange solid **2** (0.92 g, 79%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub> w/ PhNHNH<sub>2</sub>) δ ppm 1.45 (s, 6 H), 1.56 (s, 6 H), 2.02 - 2.14 (m, 2 H), 2.14 - 2.33 (m, 2 H), 4.41 - 4.74 (m, 1 H), 6.43 (d, *J*=7.6 Hz, 1 H), 7.50 (d, *J*=8.4 Hz, 2 H), 7.78 (d, *J*=8.4 Hz, 2 H). ESR (DMSO): *g* = 2.0060, *a<sub>N</sub>* = 1.549 mT (triplet). HRMS (ESI/Q-TOF) *m/z*: [M+Na]<sup>+</sup> Calcd for C<sub>16</sub>H<sub>22</sub>IN<sub>2</sub>NaO<sub>2</sub><sup>+</sup> 424.0618; Found 424.0622.

**2,2,6,6-Tetramethyl-4-(4-(tributylstannyl)benzamido)piperidine-1-oxyl (3).** To a solution of **2** (0.20 g, 0.5 mmol) in anhydrous toluene (5 mL), Pd (PPh<sub>3</sub>)<sub>4</sub> (29 mg, 25 μmol) is added, and bubbling is carried out for 5 minutes while stirring, then (SnBu<sub>3</sub>)<sub>2</sub> (0.50 mL, 1.0 mmol) was added and refluxed at 100°C in an oil bath for 24 hours. After the reaction, the solution was evaporated under reduced pressure, and purification was performed by flash column chromatography by using hexane: Et<sub>2</sub>O = 3: 1 (R<sub>f</sub>: 0.38) as an elution solvent to afford orange solid **3** (145 mg, 51%). <sup>1</sup>H NMR (300 MHz, CDCl<sub>3</sub> w/ PhNHNH<sub>2</sub>) δ ppm 0.88 (t, *J*=7.2 Hz, 9 H), 1.02 - 1.13 (m, 6 H), 1.27 - 1.39 (m, 6 H), 1.47 - 1.59 (m, 6 H), 1.49 (s, 6 H), 1.63 (s, 6 H), 2.05 - 2.18 (m, 2 H), 2.20 - 2.43 (m, 2 H), 4.50 - 4.69 (m, 1 H), 6.25 (d, *J*=8.1 Hz, 1 H), 7.54 (d, *J*=8.0 Hz, 2 H), 7.67 (d, *J*=8.0 Hz, 2 H). ESR (DMSO): *g* = 2.0060, *a<sub>N</sub>* = 1.572 mT (triplet). HRMS (ESI/Q-TOF) *m/z*: [M+Na]<sup>+</sup> Calcd for C<sub>28</sub>H<sub>49</sub>N<sub>2</sub>NaO<sub>2</sub>Sn<sup>+</sup> 588.2708; Found 588.2711.

**Radiolabeling for 4-(4-[<sup>125</sup>I]Iodobenzamido)-2,2,6,6-tetramethylpiperidine-1-oxyl ([<sup>125</sup>I]**2**).** Compound **3** and NCS were each dissolved in MeOH: AcOH (100: 1) to 1 mg/mL. The solutions of **3** (50 μl) and NCS (50 μl) were added to Na<sup>125</sup>I, vortexed and allowed to stand at rt for 15 minutes. The reaction mixture was injected into HPLC (mobile phase: A: H<sub>2</sub>O, B: MeCN; 10% B (t = 0 min), 100% B (t = 5 to 30 mins); column: COSMOSIL 5C<sub>18</sub>-AR-II, 4.6 mm I.D. x 250 mm; flow rate: 1 mL/min), to purify the desired compound. Collected fraction was concentrated using N<sub>2</sub> gas flow.

**Reactivity assay.** The conditions of each reaction are as follows. (i) Lipid alkyl radical: Arachidonic acid, 0.8 mM; soybean lipoxidase (LOX), 0.05 mg/mL; **2**, 50  $\mu$ M. (ii)  $\text{H}_2\text{O}_2$ :  $\text{H}_2\text{O}_2$ , 1%; **2**, 50  $\mu$ M. (iii) Hydroxyl radical ( $\cdot\text{OH}$ ):  $\text{H}_2\text{O}_2$ , 1%;  $\text{FeSO}_4$ , 0.2 mM; **2**, 50  $\mu$ M; (iv) Superoxide anion radical ( $\text{O}_2^{\cdot-}$ ): hypoxanthine, 1.25 mM; xanthine oxidase, 78 mU/mL; **2**, 50  $\mu$ M. (v) Hypochlorite ion ( $\text{ClO}^-$ ):  $\text{NaClO}$ , 0.1%; **2**, 50  $\mu$ M. Each solution was mixed in 10 mM phosphate buffer (pH 7.4) in equal volumes (40  $\mu$ L), then the mixed solution was transferred to a capillary tube and set into the ESR cavity. The ESR spectra were recorded on an X-band ESR spectrometry. The amounts of the nitroxide were calculated from the double-integrated ESR signal intensity, using  $\text{Mn}^{2+}$  as an external standard. The conditions for the ESR measurements were as follows: power, 10 mW; frequency, 9.4 GHz; magnetic field, 337 mT; modulation amplitude, about one-third of the line width; time constant, 0.3 s.

**Cell culture.** HepG2 cells were grown in Dulbecco's Modified Eagle Medium (DMEM) supplemented with 10% fetal bovine serum, streptomycin (100  $\mu\text{g}/\text{mL}$ ) and penicillin (100 U/mL). Cells were maintained at 37  $^\circ\text{C}$  in 5%  $\text{CO}_2$ /95% air.

**Cellular uptake assay.** HepG2 cells were plated at a density of  $2 \times 10^5$  cells per well in a 24-well plate for 1 day prior to the assay. The medium was removed and medium containing each additive, such as 500  $\mu\text{M}$  arachidonic acid and  $[\text{}^{125}\text{I}]\textbf{2}$  (0.5  $\mu\text{Ci}$  with 1% EtOH and 0.1% Tween 80 as co-solvents), was added to each well and allowed to stand in a  $\text{CO}_2$  incubator at 37  $^\circ\text{C}$  for 5, 10, 15 minutes. Then the medium was removed, cells were washed with PBS, and dissolved in 0.2 M NaOH. The radioactivity was measured with a  $\gamma$  counter. In addition, the amount of protein was measured by BCA protein assay kit (Thermo Fisher Scientific).

**Statistical Analysis.** All results are expressed as the mean  $\pm$  standard deviation of at least three independent experiments. Statistical analyses were performed using Student's t test. The minimum level of significance was set at  $p < 0.05$ .

## 2. NMR spectra

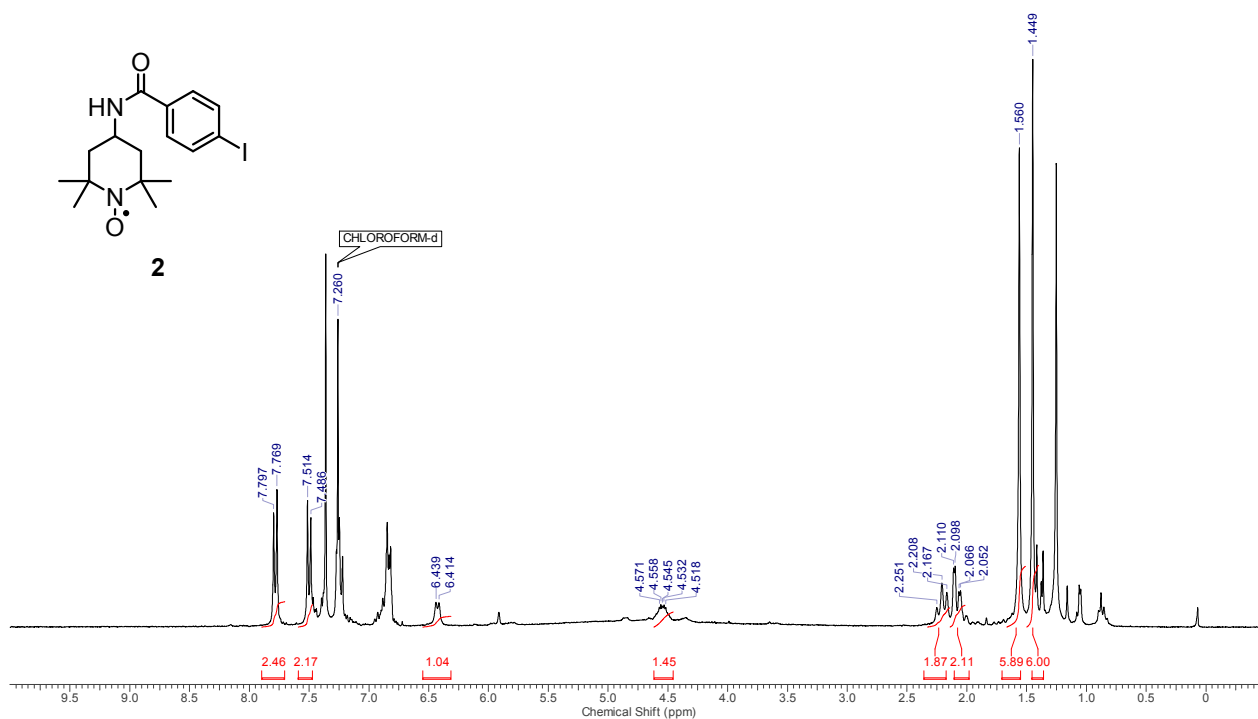


Figure S1. NMR spectrum of **2** w/ PhNHNH<sub>2</sub>.

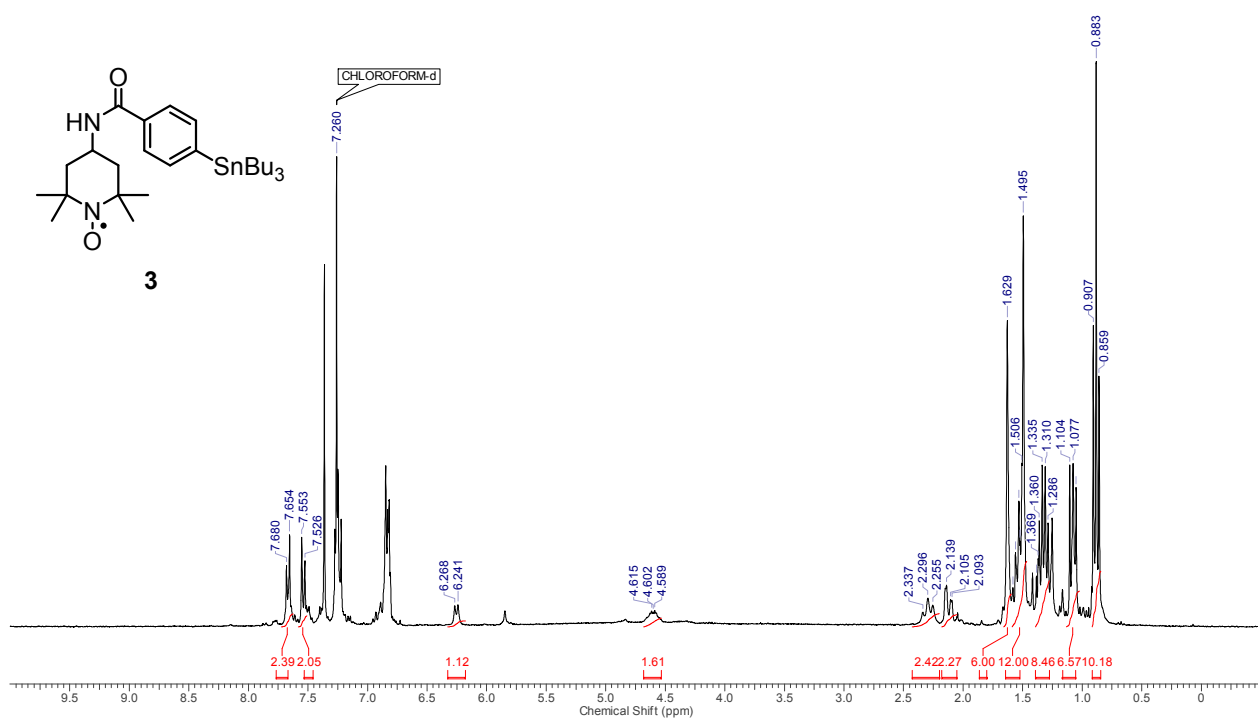
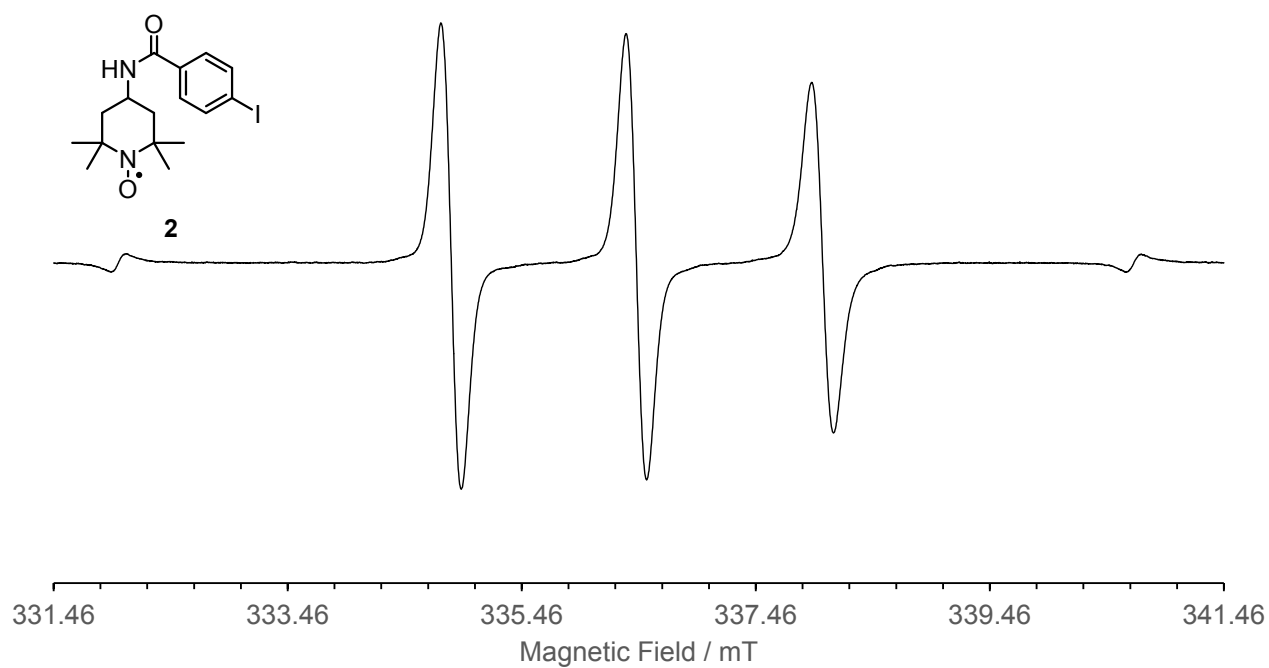
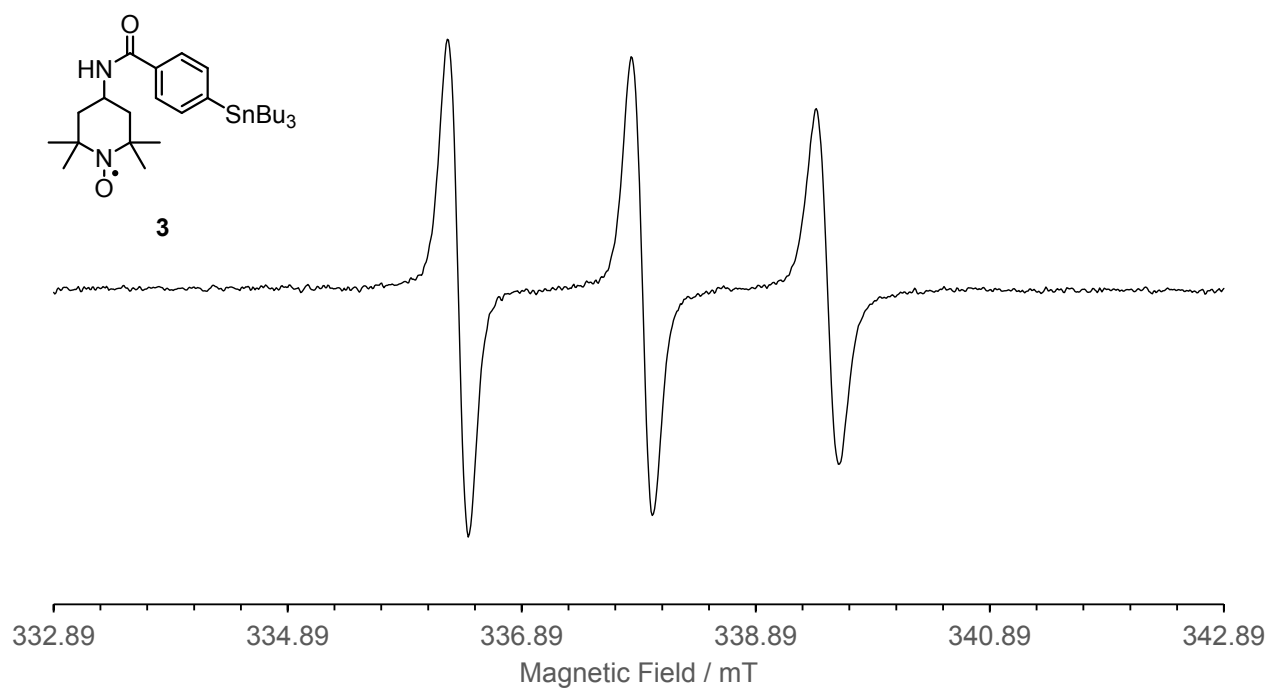


Figure S2. NMR spectrum of **3** w/ PhNHNH<sub>2</sub>.

### 3. ESR spectra

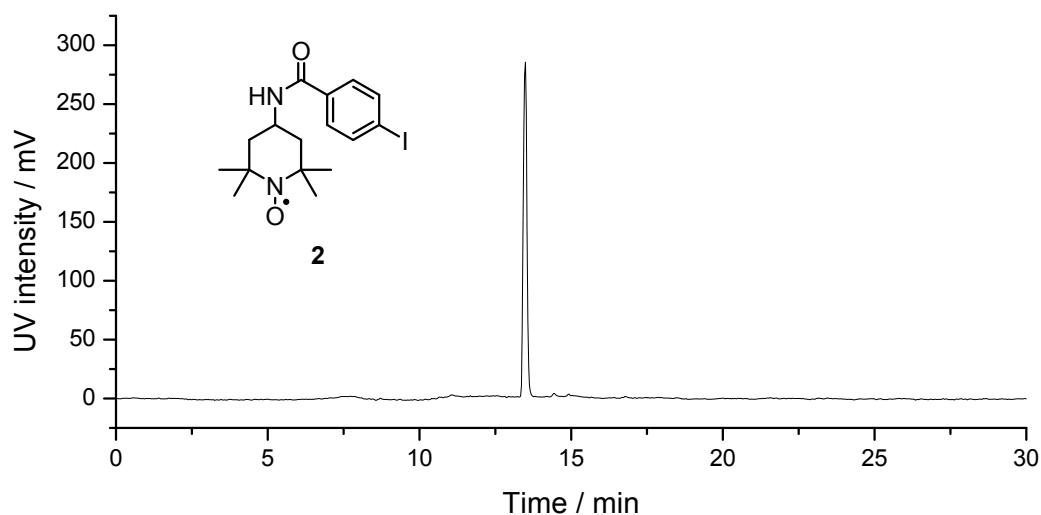


**Figure S3.** ESR spectrum of **2** in DMSO.

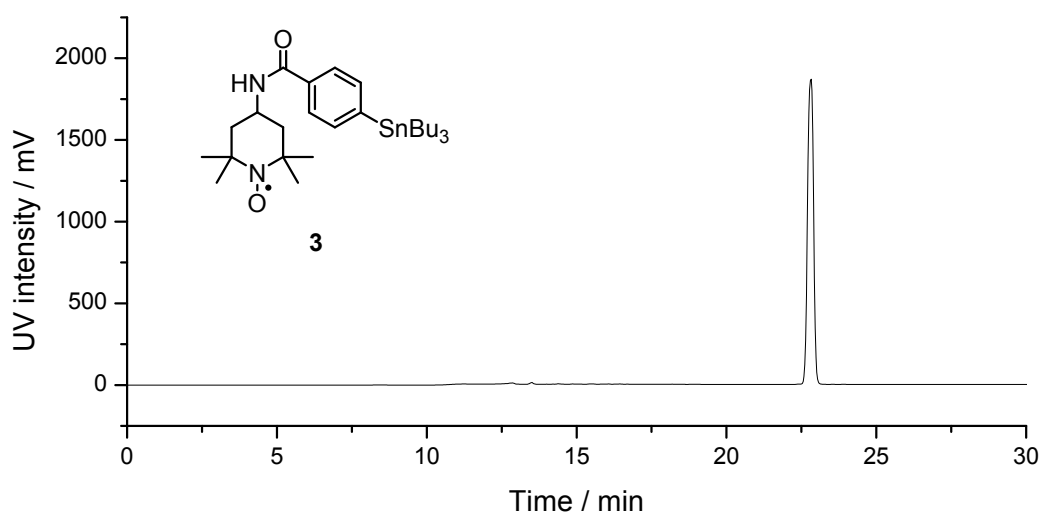


**Figure S4.** ESR spectrum of **3** in DMSO.

#### 4. HPLC chromatograms



**Figure S5.** HPLC chromatogram of **2**. Mobile phase: A: H<sub>2</sub>O, B: MeCN; 10% B (t = 0 min), 100% B (t = 5 to 30 mins); column: COSMOSIL 5C<sub>18</sub>-AR-II, 4.6 mmI.D. x 250 mm; UV: 254 nm; flow rate: 1 mL/min.



**Figure S6.** HPLC chromatogram of **3**. Mobile phase: A: H<sub>2</sub>O, B: MeCN; 10% B (t = 0 min), 100% B (t = 5 to 30 mins); column: COSMOSIL 5C<sub>18</sub>-AR-II, 4.6 mmI.D. x 250 mm; UV: 254 nm; flow rate: 1 mL/min.