Supporting Information

Enzymatic Preparation of Plasmonic-Fluorescent Quantum Dot-Gold Hybrid Nanoprobes for Sensitive Detection of Glucose and Alkaline Phosphatase and Dual-Modality Cell Imaging

Jie zhang, + Guohua Qi, + ‡Chen Xu, +§ and Yongdong Jin*+ ‡§ † State Key Laboratory of Electroanalytical Chemistry, Changchun Institute of Applied Chemistry, Chinese Academy of Sciences, Changchun, Jilin 130022, P. R. China. ‡University of Chinese Academy of Sciences, Beijing 100049, P. R. China §University of Science and Technology of China, Hefei 230026, P. R. China *e-mail: ydjin@ciac.ac.cn

Table of Contents:

Figure S1. The PL intensities of polymer-coated QDs against H ₂ O ₂ chemical oxidation and L-ascorbic
acid etching
Figure S2. The time evolution fluorescence emission spectra of the QD- enzyme solution after adding
HAuCl4 solution
Figure S3. The time evolution fluorescence emission spectra of the QD- polymer-GOx solution in the
presence of HAuCl ₄ after adding 20 μ L of glucose (2×10 ⁻³ M)S-3
Figure S4. Selectivity analysis for glucose detection by the PL intensity of the QD-GOx
NSs
Figure S5. The fluorescence emission spectra of the QD-polymer-ALP solution after adding different
concentrations of AAP in PBSS-4
Figure S6. The fluorescence intensity of the ALP sensing system as a function of the pH
valuesS-4
Figure S7. The fluorescence intensity of the ALP sensing system as a function of the enzymatic reaction
time in the presence and absence of ALP
Figure S8. Representative TEM images and the core-shell separation distribution of the QD-gold core-
shell nanoparticles
Figure S9. The representative XPS spectra of the QD-Au NPs prepared by ALP enzymatic
reactionS-6
Figure S10. The photostability of the polymer-coated QD and QD–gold hybrid
nanoparticles



Figure S1. The PL intensities of polymer-coated QDs against chemical oxidation and L-ascorbic acid etching, in the presence of increasing concentrations of H₂O₂ and L-ascorbic acid.



Figure S2. The time evolution fluorescence emission spectra of the QD- polymer-GOx and QD-polymer-AAP solution after adding HAuCl₄ solution(10 µL, w/w 1%).



Figure S3. The time evolution fluorescence emission spectra of the QD- polymer-GOx solution in the presence of HAuCl₄ after adding 20 μ L of glucose (2 × 10⁻³ M).



Figure S4. Selectivity analysis for glucose detection by the PL intensity of the QD-GOx NSs after 2 h incubation with glucose $(2 \times 10^{-3} \text{ M})$, lactose $(4 \times 10^{-3} \text{ M})$, sucrose $(4 \times 10^{-3} \text{ M})$, and fructose $(4 \times 10^{-3} \text{ M})$, respectively.



Figure S5. The fluorescence emission spectra of the QD-polymer-ALP solution after adding different concentrations of AAP in PBS (pH 7.4, 10 mM).



Figure S6. The fluorescence intensity of the ALP sensing system as a function of the pH values (10 mM Tris-HCl). ALP, 0.05 mU/mL; incubation at room temperature (25-30°C) for 30 min. λ ex = 450 nm.



Figure S7. The fluorescence intensity of the ALP sensing system as a function of the enzymatic reaction time in the presence and absence of ALP, 0.05 mU/mL; pH 9.0 (10 mM Tris-HCl); incubation at room temperature (25-30 °C). λ ex = 450 nm.



Figure S8. a) Representative TEM images of the QD–gold core–shell nanoparticles with layers of polyelectrolyte spacers. b) Histograms of the core–shell separation distribution of particles with polyelectrolytes layers. Each histogram was plotted based on >100 nanoparticles. Insets show the TEM measurements of representative particles.



Figure S9. The representative XPS spectra of the QD-Au NPs prepared by ALP enzymatic reaction.



Figure S10. The fluorescence intensities of water-soluble polymer-coated QDs and QD-Au core-shell NPs. Under identical illumination conditions (450 nm laser).