## Supporting Information

## Graph Convolutional Neural Networks for Predicting Drug-Target Interactions

Wen Torng ${ }^{1} \&$ Russ B. Altman ${ }^{1,2}$
${ }^{1}$ Deparment of Bioengineering, Stanford University, Stanford, CA 94305, ${ }^{2}$ Department of Genetics, Stanford University, Stanford, CA, 94305

Wen Torng:
Shriram Room 213
443 Via Ortega Drive
Stanford, CA 94305-4145
Email: wtorng@stanford.edu

Corresponding Author:
Russ B. Altman MD, PhD
Shriram Room 209, MC: 4245
443 Via Ortega Drive
Stanford, CA 94305-4145
Email: russ.altman@stanford.edu
Phone: 650-725-3394

Contact: rbaltman@stanford.edu
Key words: Graph convolutional neural networks, Protein-ligand binding, Pocket representations, Structural Bioinformatics, Drug discovery

## Note S1. Stanford XStream and Sherlock Servers

The Stanford XStream GPU cluster is made of 65 compute nodes for a total of 520 Nvidia K80 GPU cards. The Stanford Sherlock cluster includes 6 GPU nodes with dual socket Intel(R) Xeon(R) CPU E5-2640 v2 @ 2.00GHz; 256 GB RAM; 200 GB local storage.

## Note S2. Error analysis of MUV - Case Analysis plot

(1) Extent of separation of the actives from the negatives in simple molecular descriptor space

For a given MUV target dataset, to quantify the extent of similarity among the actives and the separation between the actives and negatives, we conceptually view the actives and the negatives as two different clusters and calculate the average silhouette scores of the actives. Specifically, for the active $\mathrm{i}^{\text {th }}$ active, we calculate the $\mathrm{s}(\mathrm{i})$ by the following equations:

- $a(i)=$ average distance between the $i^{\text {th }}$ active and all other actives
- $\quad b(i)=$ average distance between the $\mathrm{i}^{\text {th }}$ active and all negatives

$$
s(i)=\frac{b(i)-a(i)}{\max \{a(i), b(i)\}}
$$

The final score for the MUV target is then calculated by

$$
\mathrm{S}=\text { average(s(i)) for all actives }
$$

Distances between the molecules are calculated by Euclidean distances of the molecules, represented using the Morgan (ECFP) fingerprints.
(2) Average binding site similarity of the MUV target to the DUD-E targets

We quantify pairwise pocket similarities between each MUV target and the DUD-E targets using the PocketFEATURE program. For each MUV target, we then average its pocket similarity score (negative PocketFEATURE score) for all the DUD-E targets to obtain a single final score.

## Note S3. Alternative MUV structures to examine model sensitivity to pocket choice

To examine the model sensitivity to pocket conformational changes, we selected alternative structures of the MUV targets --- unbound structures, or bound structures co-crystalized with ligands that are dis-similar to test ligands as below.
(1) For each MUV target, we retrieved PDB structures that map to the same target by Uniprot ID.
(2) For each retrieved structure, if the structure does not have a bound ligand at the known pocket site, we include it as an apo structure. Otherwise, we compute pairwise Dice similarity of the bound ligand to the actives of the corresponding MUV target, and select structures that have maximum bound-ligand Dice similarity lower than a 0.5 threshold to the test actives. The ligands are represented by Morgan Fingerprints with radius of 2 and the Dice similarity was computed using the rdkit.DataStructs.DiceSimilarity module.

Dice similarity between the bound ligands of the alternative structures and the corresponding MUV actives are summarized in Table S5. For Human cAMP-dependent protein kinase (MUV target 548), we included an apo structure, 6BYS, to evaluate our model. We were not able to find alternative structure for MUV target 466 based on the above criteria.

## Note S4. Derivation of pocket and ligand importance scores

We calculate the importance scores by following three stages:

- Saliency of classification score to hidden nodes in the Interaction Layer

We first calculate the derivative of the true class score of the pocket-molecule pair with respect to the interaction nodes $\mathrm{H}_{\text {inter }}$ at $\mathrm{H}_{\text {inter0 }}$, where $\mathrm{H}_{\text {inter0 }}$ denotes the interaction node values of the given pocket and molecule pair. The derivative is then multiplied by $H_{\text {inter0 }}$ to obtain the saliency score for each interaction node. By first order Taylor approximation, the saliency score of each node approximates the effect on the true class score when removing the corresponding interaction node.

$$
\begin{array}{rl}
\operatorname{grad}_{H_{\text {inter }}} & =\frac{\partial s c o r e_{\text {class }}}{\partial H_{\text {inter }}} \\
H_{H_{\text {inter } 0}} & \mathrm{Eq}(1) \\
\operatorname{sali}_{H_{\text {inter }}} & =\operatorname{grad}_{H_{\text {inter }}} * H_{\text {inter } 0} \mathrm{Eq}(2)
\end{array}
$$

Interaction nodes that have positive saliency scores $\left(\operatorname{sali}_{H_{\text {inter }}}\right)$ are then identified and sorted according to their saliency scores, where a higher saliency score indicate larger contribution. For each pocket-ligand pair, we visualize the top 5 (out of 100)
interaction nodes that have the highest saliency scores.

## - Saliency of key interaction node to pocket and molecule fingerprint attributes

For each identified key interaction node (with index $h_{i d x}$ ), we derive the contribution of each molecule fingerprint and pocket fingerprint attribute to the interaction node value by similarly calculate the saliency score of each fingerprint attribute to the interaction node value.

$$
\begin{gathered}
\operatorname{sali}_{h_{i d x}}=\operatorname{sali}_{H_{\text {inter }}}\left[h_{i d x}\right] \quad \mathrm{Eq}(3) \\
\operatorname{grad}_{F P_{p o c}}=\left.\frac{\partial H_{\text {inter }}\left[h_{i d x}\right]}{\partial F P_{p o c}}\right|_{F P_{p o c 0}} \\
\mathrm{Eq}(4) \\
\operatorname{sali}_{F P_{p o c}}=\operatorname{grad}_{F P_{p o c}} * F P_{p o c 0} \\
\operatorname{grad}_{F P_{\text {mol }}}=\left.\frac{\mathrm{Eq}(5)}{} \frac{\partial H_{\text {inter }}\left[h_{i d x}\right]}{\partial F P_{\text {mol }}}\right|_{F P_{\text {molo }}} \\
\operatorname{sali}_{F P_{\text {mol }}}=\operatorname{grad}_{F P_{\text {mol }}} * F P_{\text {mol0 }}
\end{gathered} \mathrm{Eq} \mathrm{(7)} \text { (7) }
$$

Where $F P_{p o c 0}$ denotes the pocket fingerprint values for the given input pocket. $F P_{\text {mol0 }}$ denotes the molecular fingerprint values for the given input ligand.

Pocket and molecular fingerprint attributes that have positive saliency scores to the key interaction node are then identified, with their saliency scores recorded.

- Saliency of key pocket / molecule fingerprint to pocket residues and ligand atoms

For each identified key pocket and molecular fingerprint attribute, we identify the key contributing pocket residues and atoms. Below we describe the procedure to derive contribution of each pocket residue to the pocket fingerprint attribute indexed by $f p_{i d x}$. Let $x_{v}$ be the node feature of residue node $v$, and $X_{p o c}$ be a matrix containing the node features of all nodes within a pocket graph arranged as columns. Different from the previous procedures, we cannot directly take derivatives of $F P_{p o c}$ with respect to $X_{p o c}$ to compute the saliency scores. This is because as the Softmax function in Equation (9) reaches saturation, the gradients vanish to 0 , prohibiting the gradient to flow freely from $F P_{p o c}$ to the input features $X_{p o c}$.

$$
\begin{array}{r}
S_{v}=W_{F P p o c} x_{v}+b_{F P_{p o c}} \mathrm{Eq}(8) \\
F P_{p o c}=\sum_{v \in \text { pocket }} F P_{v}=\sum_{v \in \text { pocket }} \text { Softmax }\left(S_{v}\right) \tag{9}
\end{array}
$$

Instead, for each residue in the pocket, we compute

$$
\begin{gathered}
\operatorname{sali}_{F P_{\left(f p_{i d x}, v\right)}}=\operatorname{grad}_{F P_{p o c}}\left[f p_{i d x}\right] * F P_{\mathrm{v}}\left[f p_{i d x}\right]_{0} \\
\operatorname{grad}_{X_{p o c}}=\left.\frac{\partial s_{v}\left[f p_{i d x}\right]}{\partial X_{p o c}}\right|_{\mathrm{x}_{\mathrm{poco} 0}} \quad \mathrm{Eq}(11) \\
\operatorname{sali}_{X_{p o c}}=\operatorname{grad}_{X_{p o c}} * X_{p o c 0} \\
\mathrm{Eq}(12) \\
\text { Importance }\left[h_{i d x}, f p_{i d x}, v\right]=\operatorname{sali}_{h_{i d x}} * \operatorname{sali}_{F P_{\left(f p_{\left.i d x^{\prime}, v\right)}\right.} * \operatorname{sali}_{X_{p o c}}} \quad \mathrm{Eq}(13)
\end{gathered}
$$

Where $F P_{\mathrm{v}}\left[f p_{i d x}\right]_{0}$ denotes the value of the $f p_{i d x}{ }^{\text {th }}$ attribute of residue fingerprint of node $v$ in the given pocket graph. $X_{p o c 0}$ denotes the values of the node features of the given pocket graph.

Contribution of each pocket residue to the $h_{i d x}{ }^{\text {th }}$ interaction node are then computed and integrated. Specifically,

Importance $\left[h_{i d x}\right]=\sum_{f p_{i d x} \in \operatorname{pos}\left(h_{i d x}\right)} \sum_{\mathrm{v} \in \text { pocket }}$ Importance $\left[h_{i d x}, f p_{i d x}, \mathrm{v}\right]$
Where $\operatorname{pos}\left(h_{i d x}\right)$ denotes all the pocket fingerprint indexes which have positive saliency scores for interaction node $h_{i d x}$.

The resulting importance scores of residues in the pocket are then normalized by the maximum score so that all scores have values between 0 to 1 . Importance scores of atoms in the ligand are calculated similarly.

Table S1. Dice similarity between bound ligands of DUD-E targets and DUD-E actives

| Target | Average | Standard Deviation | Max |
| :---: | :---: | :---: | :---: |
| AA2AR | 0.410 | 0.131 | 1.000 |
| ABL1 | 0.384 | 0.079 | 0.859 |
| ACE | 0.408 | 0.149 | 0.882 |
| ACES | 0.266 | 0.075 | 0.889 |
| ADA | 0.406 | 0.168 | 1.000 |
| ADA17 | 0.349 | 0.128 | 0.821 |
| ADRB1 | 0.309 | 0.091 | 0.729 |
| ADRB2 | 0.244 | 0.079 | 1.000 |
| AKT1 | 0.212 | 0.080 | 0.929 |
| AKT2 | 0.273 | 0.170 | 1.000 |
| ALDR | 0.435 | 0.162 | 0.933 |
| AMPC | 0.469 | 0.232 | 1.000 |
| ANDR | 0.250 | 0.177 | 0.898 |
| AOFB | 0.270 | 0.087 | 0.718 |
| BACE1 | 0.265 | 0.054 | 0.600 |
| BRAF | 0.410 | 0.176 | 1.000 |
| CAH2 | 0.168 | 0.044 | 0.354 |
| CASP3 | 0.320 | 0.107 | 0.651 |
| CDK2 | 0.310 | 0.067 | 0.719 |
| COMT | 0.519 | 0.148 | 1.000 |
| CP2C9 | 0.349 | 0.056 | 0.523 |
| CP3A4 | 0.260 | 0.078 | 0.664 |
| CSF1R | 0.339 | 0.147 | 0.845 |
| CXCR4 | 0.401 | 0.315 | 1.000 |
| DEF | 0.325 | 0.136 | 1.000 |
| DHI1 | 0.311 | 0.110 | 0.882 |
| DPP4 | 0.289 | 0.120 | 0.894 |
| DRD3 | 0.268 | 0.059 | 0.794 |
| DYR | 0.395 | 0.096 | 0.884 |
| EGFR | 0.430 | 0.104 | 0.856 |
| ESR1 | 0.519 | 0.191 | 0.948 |
| ESR2 | 0.387 | 0.115 | 1.000 |
| FA10 | 0.335 | 0.098 | 1.000 |
| FA7 | 0.479 | 0.118 | 1.000 |
| FABP4 | 0.499 | 0.242 | 1.000 |
| FAK1 | 0.467 | 0.162 | 1.000 |
| FGFR1 | 0.297 | 0.050 | 0.420 |
| FKB1A | 0.388 | 0.104 | 0.694 |
| FNTA | 0.379 | 0.075 | 0.712 |
| FPPS | 0.478 | 0.118 | 1.000 |
| GCR | 0.324 | 0.066 | 0.526 |
| GLCM | 0.137 | 0.081 | 0.261 |


| GRIA2 | 0.245 | 0.112 | 1.000 |
| :---: | :---: | :---: | :---: |
| GRIK1 | 0.272 | 0.111 | 0.758 |
| HDAC2 | 0.398 | 0.151 | 0.859 |
| HDAC8 | 0.432 | 0.119 | 1.000 |
| HIVINT | 0.310 | 0.044 | 0.422 |
| HIVPR | 0.390 | 0.090 | 0.688 |
| HIVRT | 0.243 | 0.074 | 0.637 |
| HMDH | 0.508 | 0.185 | 0.901 |
| HS90A | 0.315 | 0.074 | 0.529 |
| HXK4 | 0.366 | 0.126 | 1.000 |
| IGF1R | 0.371 | 0.099 | 0.909 |
| INHA | 0.353 | 0.235 | 1.000 |
| ITAL | 0.377 | 0.181 | 0.835 |
| JAK2 | 0.381 | 0.140 | 1.000 |
| KIF11 | 0.427 | 0.116 | 0.741 |
| KIT | 0.267 | 0.079 | 1.000 |
| KITH | 0.404 | 0.207 | 1.000 |
| KPCB | 0.444 | 0.149 | 0.695 |
| LCK | 0.360 | 0.105 | 0.804 |
| LKHA4 | 0.467 | 0.094 | 0.763 |
| MAPK2 | 0.349 | 0.054 | 0.497 |
| MCR | 0.192 | 0.153 | 1.000 |
| MET | 0.458 | 0.127 | 0.854 |
| MK01 | 0.407 | 0.108 | 0.610 |
| MK10 | 0.344 | 0.082 | 0.736 |
| MK14 | 0.315 | 0.067 | 1.000 |
| MMP13 | 0.449 | 0.104 | 1.000 |
| MP2K1 | 0.299 | 0.155 | 1.000 |
| NOS1 | 0.091 | 0.087 | 0.625 |
| NRAM | 0.261 | 0.087 | 1.000 |
| PA2GA | 0.373 | 0.132 | 1.000 |
| PARP1 | 0.375 | 0.071 | 0.736 |
| PDE5A | 0.343 | 0.140 | 0.913 |
| PGH1 | 0.303 | 0.094 | 0.855 |
| PGH2 | 0.419 | 0.151 | 0.915 |
| PLK1 | 0.313 | 0.077 | 0.839 |
| PNPH | 0.305 | 0.113 | 0.699 |
| PPARA | 0.423 | 0.101 | 0.940 |
| PPARD | 0.425 | 0.081 | 0.783 |
| PPARG | 0.436 | 0.084 | 0.783 |
| PRGR | 0.335 | 0.083 | 0.768 |
| PTN1 | 0.289 | 0.114 | 0.595 |
| PUR2 | 0.534 | 0.063 | 0.660 |
| PYGM | 0.325 | 0.047 | 0.421 |
| PYRD | 0.547 | 0.214 | 0.922 |


| RENI | 0.298 | 0.095 | 0.613 |
| :---: | :---: | :---: | :---: |
| ROCK1 | 0.261 | 0.066 | 0.416 |
| RXRA | 0.111 | 0.034 | 0.204 |
| SAHH | 0.668 | 0.118 | 1.000 |
| SRC | 0.330 | 0.086 | 0.591 |
| TGFR1 | 0.493 | 0.136 | 1.000 |
| THB | 0.412 | 0.124 | 0.764 |
| THRB | 0.308 | 0.062 | 0.534 |
| TRY1 | 0.316 | 0.076 | 0.488 |
| TRYB1 | 0.331 | 0.071 | 0.512 |
| TYSY | 0.409 | 0.113 | 0.894 |
| UROK | 0.330 | 0.082 | 0.904 |
| VGFR2 | 0.362 | 0.119 | 1.000 |
| WEE1 | 0.620 | 0.113 | 0.859 |
| XIAP | 0.446 | 0.091 | 0.683 |

We computed pairwise Dice similarity between the bound-ligand of each DUD-E target to the corresponding DUD-E actives using the rdkit.DataStructs.DiceSimilarity module. The ligands are represented by Morgan Fingerprints with radius of 2.

Table S2. Dice similarity between bound ligands in MUV targets and MUV actives

| Target | Average | Standard Deviation | Max |
| :---: | :---: | :---: | :---: |
| 846 | 0.329 | 0.060 | 0.447 |
| 600 | 0.068 | 0.045 | 0.155 |
| 692 | 0.064 | 0.032 | 0.147 |
| 859 | 0.233 | 0.055 | 0.478 |
| 852 | 0.087 | 0.020 | 0.122 |
| 548 | 0.257 | 0.055 | 0.343 |
| 713 | 0.245 | 0.065 | 0.365 |
| 466 | 0.215 | 0.046 | 0.321 |
| 689 | 0.275 | 0.045 | 0.351 |
| 832 | 0.108 | 0.038 | 0.185 |

We computed pairwise Dice similarity score of the bound-ligand of each MUV target to the corresponding MUV actives using the rdkit.DataStructs.DiceSimilarity module. The ligands are represented by Morgan Fingerprints with radius of 2.

Table S3. List of functional atoms used to determine the functional centers representing each residue type

| Residue Type | Site 1 | Site 2 |
| :--- | :--- | :--- |
| Glycine (G) | CA | - |
| Cysteine (C) | SG | - |
| Arginine (R) | CZ | - |
| Serine (S) | OG | - |
| Threonine (T) | OG1 | - |
| Lysine (K) | NZ | - |
| Methionine(M) | SD | - |
| Alanine (A) | CB | - |
| Leucine (L) | CB | - |
| Isoleucine (I) | OD1, CG, OD2 | - |
| Valine (V) | OE1, CD, OE2 | - |
| Aspartic acid (D) | NE2, ND1 | - |
| glutamic acid (E) | OD1, CG, ND2 | - |
| histidine (H) | N, CA, CB, CD, CG | - |
| Asparagine (N) | OE1, CD, NE2 | - |
| Proline (P) | CG, CD1, CD2, CE1, CE2, CZ | - |
| Glutamine (Q) | CD2, CE2, CE3, CZ2, CZ3, CH2 | NE1 |
| Phenylalanine (F) | CG, CD1, CD2, CE1, CE2, CZ | OH |
| Tryptophan (W) | Tyrosine (Y) |  |

For each residue type, the average location of the listed functional atoms for a given site is used to represent the corresponding amino acid environment. Tyrosine and Tryptophan are each represented by two sites due to their larger sizes.

Table S4. Network architecture and parameters of unsupervised pocket graph autoencoders.

| Pocket <br> Graph-Autoencoder | Input | Parameters | Output |
| :---: | :---: | :---: | :---: |
| Layer1 <br> Autoencoder I | Pocket Graph $G_{\text {pocket }}$ With residue embedding from FEATURE program $\in\left[\mathrm{N}_{\mathrm{res}}, 480\right]$ | $\begin{aligned} & W_{\text {self }_{L 1}} \in \mathrm{R}[480,200], \\ & W_{\text {deg }_{y_{L 1}}} \in \mathrm{R}[480,200], \\ & \text { max degree }=20 \end{aligned}$ | Residue embedding $\in \mathrm{R}\left[\mathrm{N}_{\mathrm{res}}, 200\right]$ |
| Layer1 <br> Autoencoder II |  | $W_{F P_{L 1}} \in \mathrm{R}[480,512]$ | $\begin{aligned} & F P_{\text {res }_{L 1}} \\ & \in \mathrm{R}\left[\mathrm{~N}_{\mathrm{res}}, 512\right] \end{aligned}$ |
| Layer2 <br> Autoencoder I | Pocket Graph $G_{\text {pocket }}$ With residue embedding from Layer $1 \in\left[\mathrm{~N}_{\mathrm{res}}, 200\right]$ | $\begin{aligned} & W_{\text {self }_{L 2}} \in \mathrm{R}[200,100], \\ & W_{\text {deg }_{y_{L 2}}} \in \mathrm{R}[200,100], \\ & \max \text { degree }=20 \end{aligned}$ | Residue embedding $\in \mathrm{R}\left[\mathrm{N}_{\mathrm{res}}, 100\right]$ |
| Layer2 <br> Autoencoder II |  | $W_{F P_{L 2}} \in \mathrm{R}[200,512]$ | $\begin{aligned} & F P_{r e s_{L 2}} \\ & \in \mathrm{R}\left[\mathrm{~N}_{\mathrm{res}}, 512\right] \end{aligned}$ |
| Average Layer | $\begin{aligned} & F P_{\text {po }_{L 1}}=\frac{1}{N_{\text {res }}} \sum_{\text {res }} F P_{\text {res }_{L 1}}, \in[1,512] \\ & F P_{\text {poc }}^{L 2} \\ & =\frac{1}{N_{\text {res }}} \sum_{\text {res }} F P_{\text {res }}^{L 2} \end{aligned}, \in[1,512]$ |  |  |
| Output Layer | $F P_{\text {oc }}=F P_{\text {poc }}{ }_{L 1}+F P_{\text {poc }}{ }_{L 2}, \in[1,512]$ |  |  |

Our pocket graph-autoencoder comprises two graph-autoencoder layers, each including two autocoders. In each layer, Autoencoder I takes in the pocket graph, and compresses local graph neighborhood information in the previous layer into new residue embeddings using convolutional filters. Autoencoder II takes in embeddings of residue nodes within the same layer, and integrates them into a fixed-size pocket fingerprint. The residue embeddings in Layer 1 are the FEATURE vectors which describe the amino acid environment for each key residue, whereas the residue embeddings in Layer 2 are the output from Autoencoder I in Layer 1. The input and output columns describe the input and output of each module respectively. The parameter column describes the learnable parameters in each module. The bias terms are omitted here for simplicity.

Table S5. Dice similarity between bound ligands in alternative MUV structures and MUV actives

| MUV Target | Alternative <br> PDB of the <br> MUV target | Bound <br> ligand | Dice similarity to MUV actives |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  | Standard <br> Deviation | Max |  |
|  | 1KYN | KTP | 0.303 | 0.079 | 0.444 |
| 859 | 6OIJ | IXO | 0.111 | 0.029 | 0.165 |
| 692 | 4QJR | PIZ | 0.054 | 0.027 | 0.130 |
| 600 | 4QJR | PIZ | 0.058 | 0.037 | 0.130 |
| 689 | 2XYU | Q9G | 0.228 | 0.056 | 0.422 |
| 548 | 6BYR | ATP | 0.168 | 0.039 | 0.262 |
| 548 | 5BX7 | 4W1 | 0.266 | 0.068 | 0.392 |
| 548 | 5BX6 | 495 | 0.287 | 0.059 | 0.396 |
| 713 | 5U2D | OBH | 0.248 | 0.061 | 0.343 |
| 713 | 5T92 | 77W | 0.279 | 0.068 | 0.414 |

For each MUV alternative structure, we compute pairwise Dice similarity between its bound ligand and the actives of the corresponding MUV target and summarized the average, maximum and standard deviation of the similarity scores.

Table S6. Reconstruction errors of the pocket graph-autoencoder layers

| Pocket graph-autoencoder |  | Input Attribute <br> Size | Reconstruction <br> Error | Percentage of <br> Error $^{2}$ |
| :--- | :--- | :---: | :---: | :---: |
| Layer 1 | Autoencoder I | 480 | 1.623 | $0.169 \%$ |
|  | Autoencoder II | 480 | 4.345 | $0.905 \%$ |
| Layer 2 | Autoencoder I | 200 | 2.817 | $0.705 \%$ |
|  | Autoencoder II | 200 | 6.020 | $3.010 \%$ |

1: The reconstruction error for Autoencoder I is defined as Error $_{\text {reconstruct }}=\frac{1}{N} \sum_{p} \frac{1}{R_{p}} \sum_{r} \sum_{i}\left[\left(v_{x(p, r, i)}^{\prime}-v_{x(p, r, i)}\right)^{2}+\left(v_{n(p, r, i)}^{\prime}-v_{n(p, r, i)}\right)^{2}\right]$, where $v_{x(p, r, i)}$ denotes the true value of attribute $i$ of the node embedding of residue $r$ in pocket $p$, $v_{x(p, r, i)}^{\prime}$ denotes the reconstructed value of attribute $i$ of the node embedding of residue $r$ in pocket $p$ by Autoencoder I. $v_{n(p, r, i)}$ denotes the value of attribute $i$ of the neighborhood vector of residue $r$ in pocket $p, v_{n(p, r, i)}^{\prime}$ denotes the reconstructed value of the neighborhood vector of residue $r$ in pocket $p$ by Autoencoder I. $N$ is the total number of pockets, and $R_{p}$ is the number of residues in pocket $p$.

The reconstruction error for Autoencoder II is defined as Error $_{\text {reconstruct }}=\frac{1}{N} \sum_{p} \frac{1}{R_{p}} \sum_{r} \sum_{i}\left(v_{x(p, r, i)}^{\prime \prime}-v_{x(p, r, i)}\right)^{2}$, where $v_{x(p, r, i)}$ denotes the true value of attribute $i$ of the node embedding of residue $r$ in pocket $p, v_{x(p, r, i)}^{\prime \prime}$ denotes the reconstructed value of attribute $i$ of the node embedding of residue $r$ in pocket $p$ by Autoencoder II. $N$ is the total number of pockets, and $R_{p}$ is the number of residues in pocket $p$.

2: The percentage of error is defined as $\frac{\text { Error }_{\text {reconstruct }}}{\text { Error }_{\text {max }}}$, where Error $_{\text {max }}$ of Autoencoder I is defined as $\sum_{i} 2 * v_{\max _{-} i}{ }^{2}$ and Error $\max$ of Autoencoder II is defined as $\sum_{i} v_{\max -i}{ }^{2}$, where $v_{\text {max } \_i}$ denotes the maximum possible value of attribute $i$ in vector $v$.

Table S7. AUC Performance on MUV dataset using alternative structures

| Target | Original MUV PDB | Alternative PDB | AUC Performance |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
|  |  |  | Original <br> Pocket | Alternative <br> Pocket | Dummy <br> Pocket |
| 832 | $1 \mathrm{AU8}$ | 1KYN | 0.530 | 0.549 | 0.552 |
| 859 | 5CXV | 6OIJ | 0.619 | 0.629 | 0.536 |
| 692 | 1YOW | 4QJR | 0.536 | 0.538 | 0.446 |
| 600 | 1YOW | 4QJR | 0.583 | 0.577 | 0.372 |
| 689 | 2 Y 60 | 2XYU | 0.717 | 0.699 | 0.6 |
| 548 | 3 POO | 6BYR | 0.698 | 0.669 | 0.390 |
|  | 3 POO | 5BX7 | 0.698 | 0.758 | 0.390 |
|  | 3 POO | 5BX6 | 0.698 | 0.766 | 0.390 |
|  | 3 POO | $\begin{gathered} \text { 6BYS } \\ \text { (Apo structure) } \end{gathered}$ | 0.698 | 0.733 | 0.390 |
| 713 | 5TN7 | 5U2D | 0.591 | 0.612 | 0.532 |
|  | 5TN7 | 5 T 92 | 0.591 | 0.597 | 0.532 |

For all MUV targets, our model showed comparable performance using the original and alternative structures of the same targets as input, suggesting that the model is generally robust to the choice of input pocket. Both original and alternative pockets generally performed significantly better than the dummy pockets.

