

Supporting information

***N*-Amino-7-Azaindole as *N,N'*- Bidentate Directing Group: Ruthenium-Catalyzed Oxidative Annulation of *N*-(7-Azaindole) Benzamides with Alkynes *via* C-H Bond Activation**

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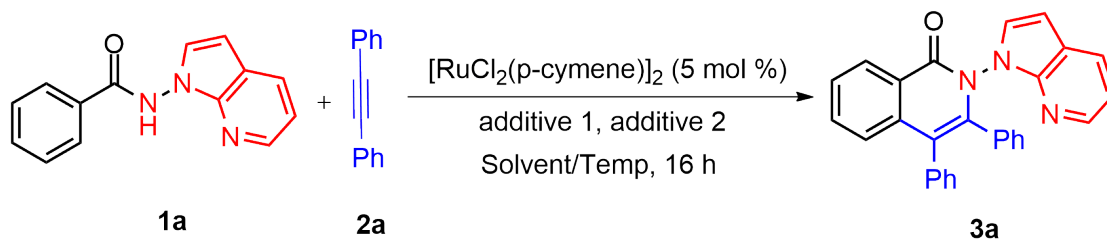
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Table S1. Optimization of Reaction Conditions:

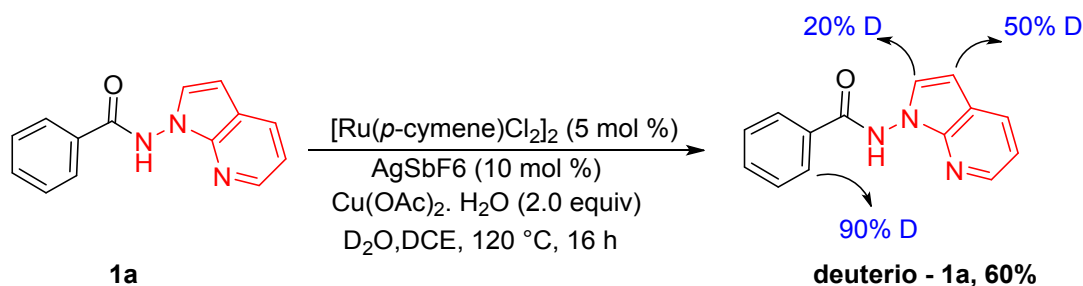


Entry	Additive 1	Additive 2	Solvent/Temp (°C)	Yield ^b (%)
1	-	NaOAc	MeOH/100	30
2	PivOH	CsOAc	MeOH/120	30
3	MesCOOH	KOAc	H ₂ O /100	50
4	MesCOOH	KOAc	TFE/100	60
5	MesCOOH	Cu(OAc) ₂ ·H ₂ O	TFE/100	65
6	MesCOOH	KOAc	PEG-400/100	15
7	MesCOOH	KOAc	DCE/120	45
8	-	CsOAc	DCE/120	40
9	AgSbF ₆	-	DCE/120	25
10	-	Cu(OAc) ₂ ·H ₂ O	DCE/120	50
11 ^c	AgSbF ₆	Cu(OAc) ₂ ·H ₂ O	DCE/120	28
12 ^d	AgSbF ₆	Cu(OAc) ₂ ·H ₂ O	DCE/120	34
13 ^e	AgSbF ₆	Cu(OAc) ₂ ·H ₂ O	DCE/120	42
14 ^f	AgSbF ₆	Cu(OAc) ₂ ·H ₂ O	DCE/120	60
15 ^g	AgSbF₆	Cu(OAc)₂·H₂O	DCE/120	85
16 ^h	-	-	DCE/120	trace

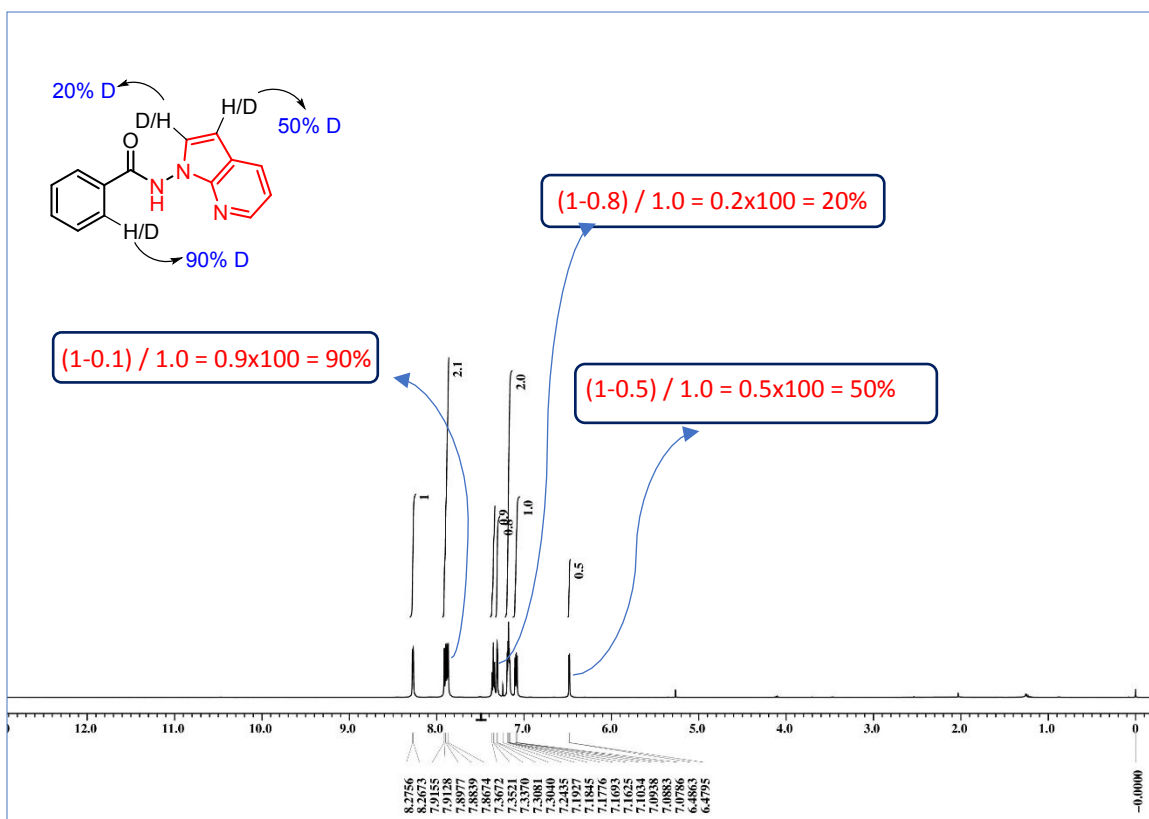
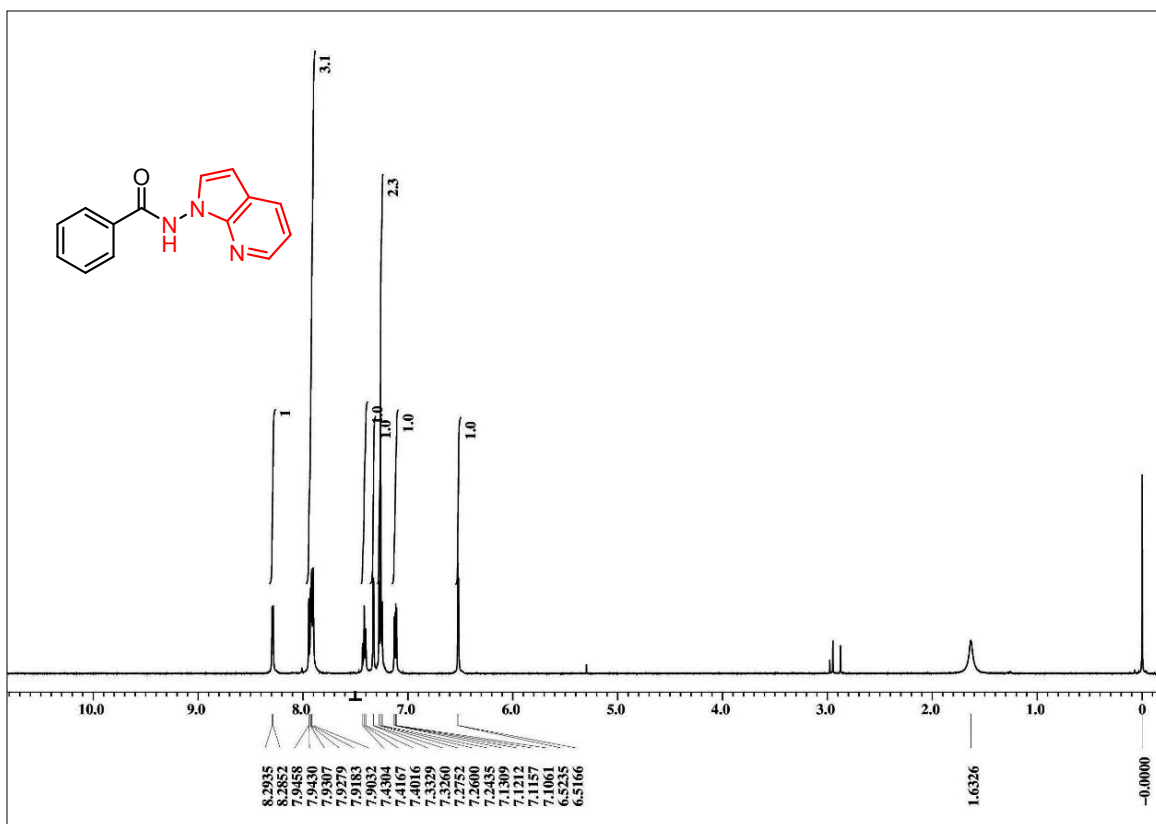
Reaction conditions: [a] 1a (0.5 mmol), 2a (1.2 equiv), [Ru(p-cymene)Cl₂]₂ (5.0 mol %), Additive 1 (10 mol %), Additive 2 (2.0 equiv), solvent (3.0 mL), 100 – 120 °C, 16h. [b] Isolated Yield. ^c Cu(OAc)₂·H₂O (10 mol %), ^d Cu(OAc)₂·H₂O (30 mol %), ^e Cu(OAc)₂·H₂O (50 mol %), ^f Cu(OAc)₂·H₂O (100 mol %), ^g Cu(OAc)₂·H₂O (200 mol %), ^h without additives.

2. Mechanistic Studies:

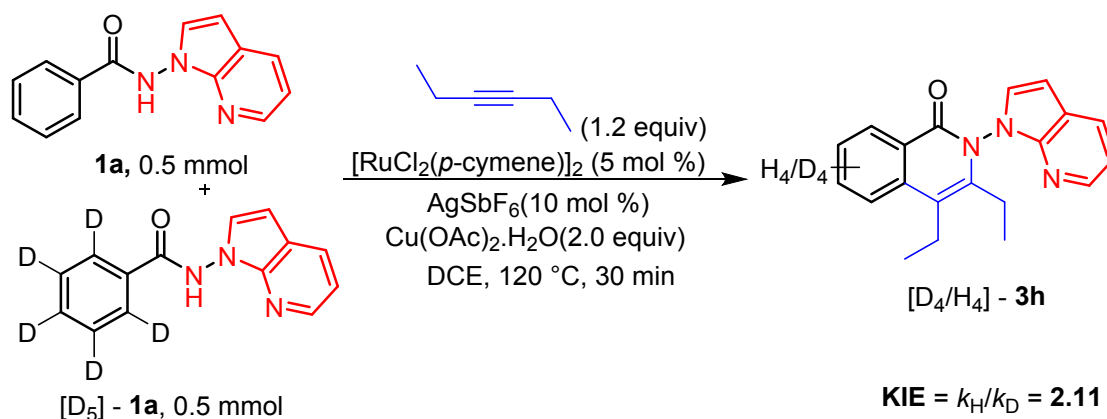
Scheme S1. Deuterium labeling experiments:



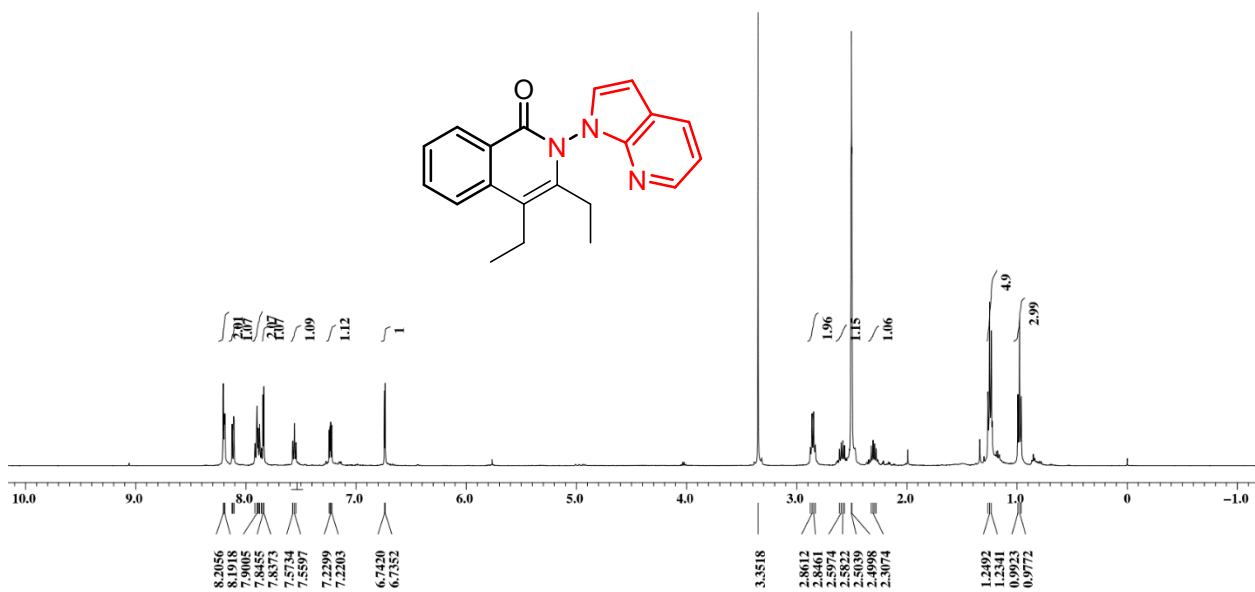
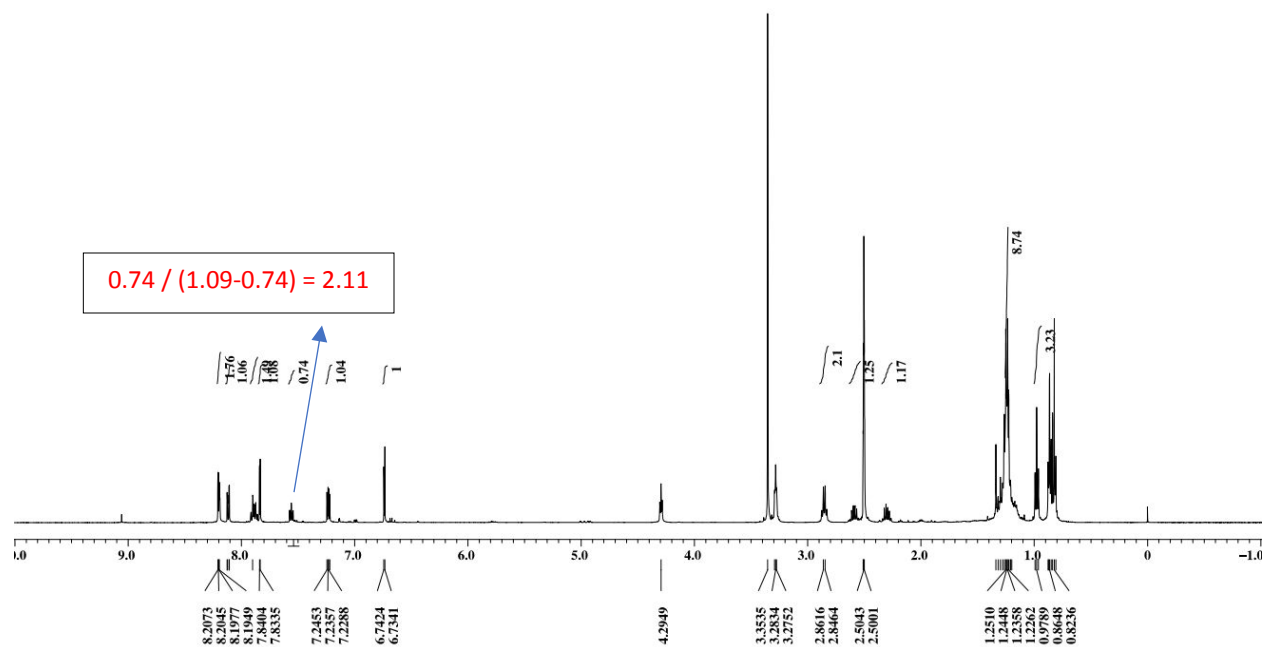
A 15 mL screw cap vial was equipped with magnetic stir bar and charged with starting compound **1a** (0.21 mmol, 50 mg), $[\text{Ru}(p\text{-cymene})\text{Cl}_2]_2$ (5 mol %), AgSbF_6 (10 mol %), $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ (2.0 equiv), D_2O (1.0 mL) and DCE (1.0 mL). The vessel was heated at 120 °C for 16h, and cooled down to room temperature. Later, the reaction mixture was diluted with 10 mL of dichloromethane and filtered through a Celite pad. The filtered solution was concentrated under reduced pressure. Then, the residue was purified by column chromatography with EtOAc/Hexane as eluent. The H/D exchange was calculated by ^1H NMR.



Scheme S2. Intermolecular competition KIE for the reaction



A 15 mL screw cap vial was equipped with magnetic stir bar and charged with starting compound **1a** (0.5 mmol, 50 mg), **[D₅]-1a** (0.5 mmol), 3-hexyne (1.2 mmol), $[\text{Ru}(p\text{-cymene})\text{Cl}_2]_2$ (5 mol %), AgSbF_6 (10 mol %), $\text{Cu}(\text{OAc})_2 \cdot \text{H}_2\text{O}$ (2.0 equiv), and DCE (3.0 mL). The reaction mixture was stirred at 120 °C for 16h, and cooled down to room temperature. Later, the reaction mixture was diluted with 15 mL of dichloromethane and filtered through a Celite pad. The filtered solution was concentrated under reduced pressure. Then, the residue was purified by column chromatography with EtOAc/Hexane as eluent to afford the desired product **[D₄/H₄]-3h**. The crude ^1H NMR spectrum of the mixture **[D₄/H₄]-3h**, revealed the different integration related to the hydrogen resonances, the kinetic isotopic calculated to be $k_{\text{H}}/k_{\text{D}} \approx 2.11$.



3. Copies of ^1H & ^{13}C NMR Data:

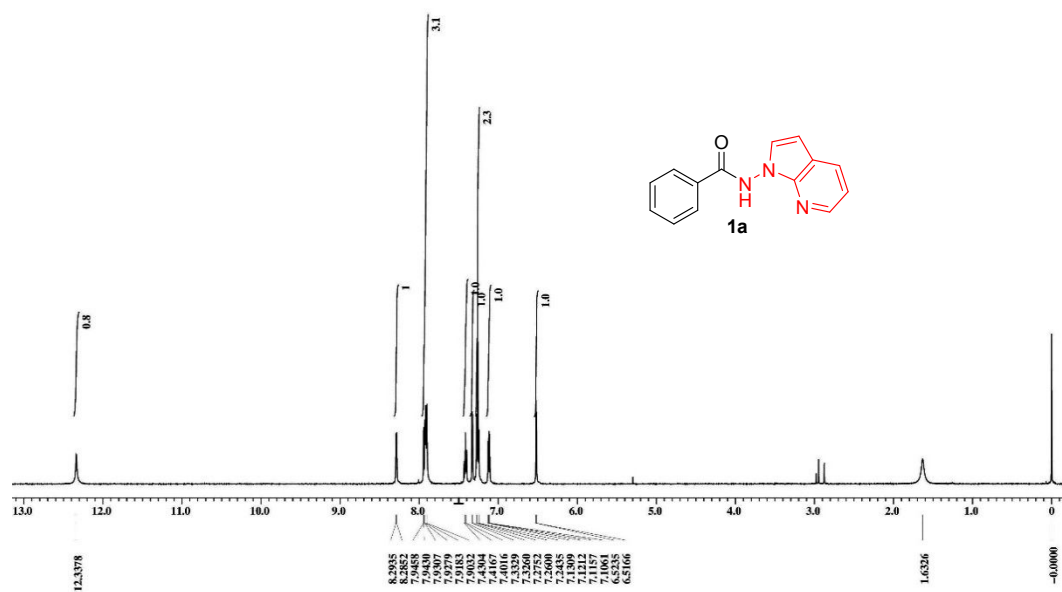


Fig. S1. ^1H NMR spectrum of **1a** (500 MHz, CDCl_3)

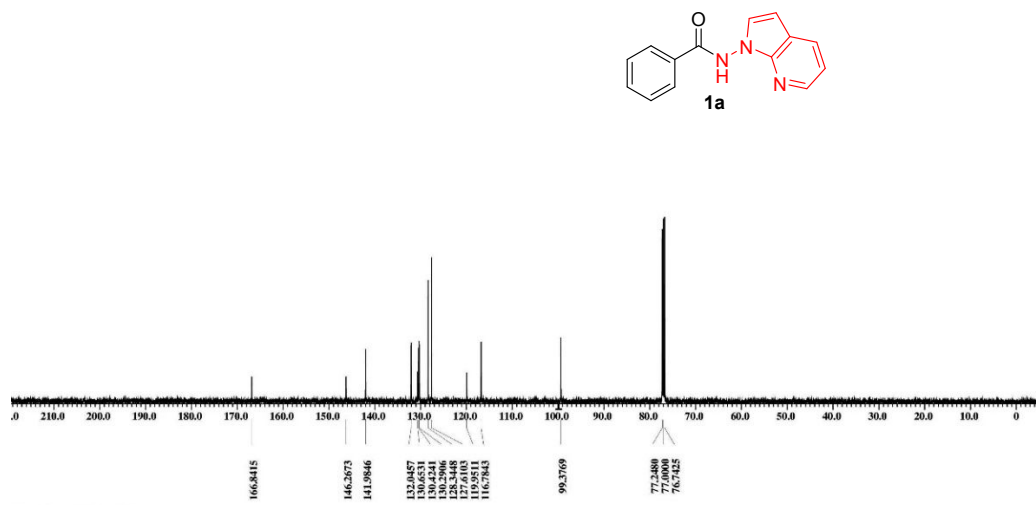


Fig. S2. ^{13}C NMR spectrum of **1a** (125 MHz, CDCl_3)

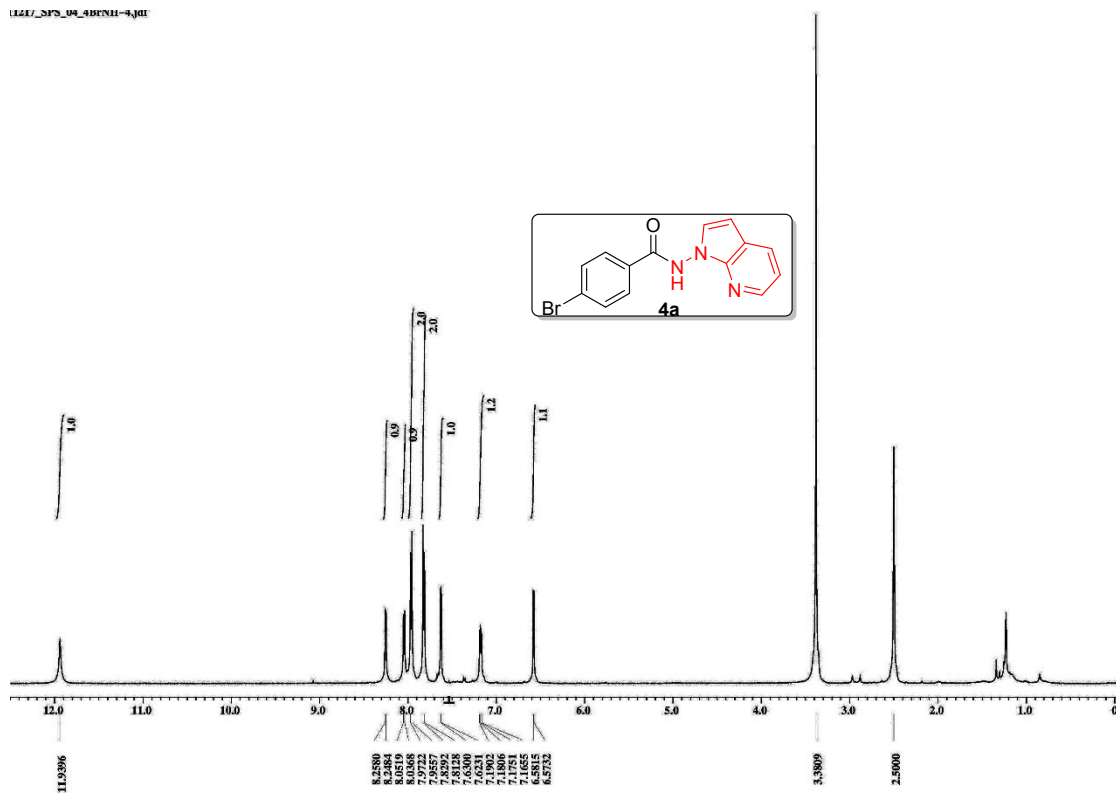


Fig. S3. ¹H NMR spectrum of **4a** (500 MHz, DMSO-d₆)

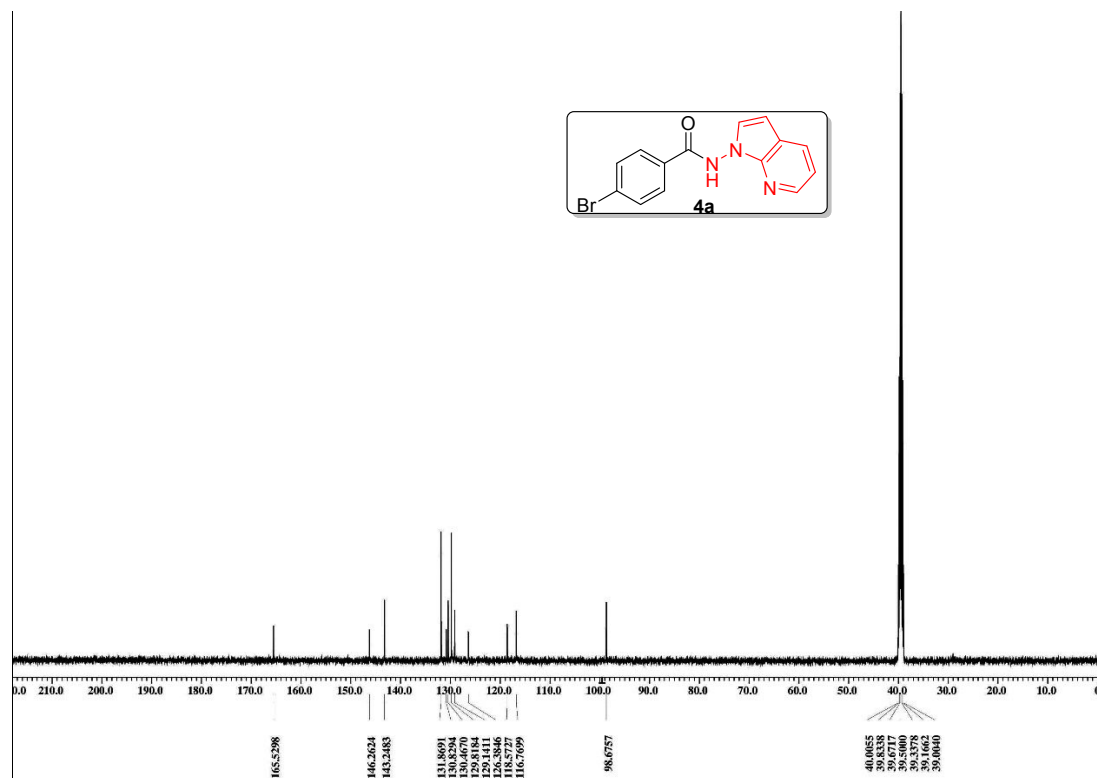


Fig. S4. ¹³C NMR spectrum of **4a** (125 MHz, DMSO-d₆)

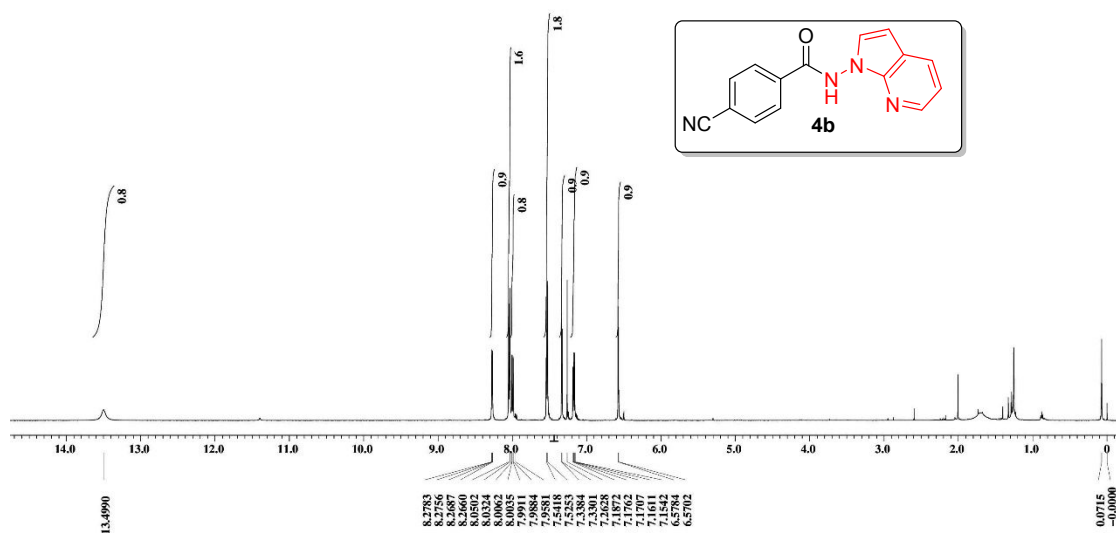


Fig. S5. ¹H NMR spectrum of **4b** (500 MHz, CDCl₃)

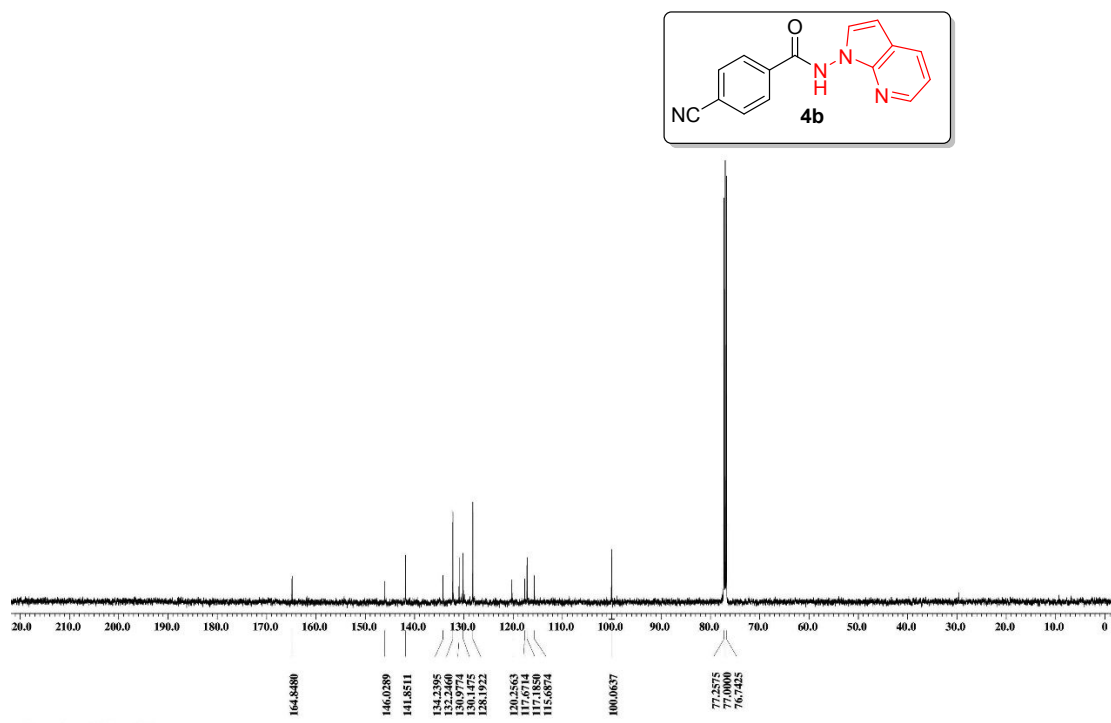


Fig. S6. ¹³C NMR spectrum of **4b** (125 MHz, CDCl₃)

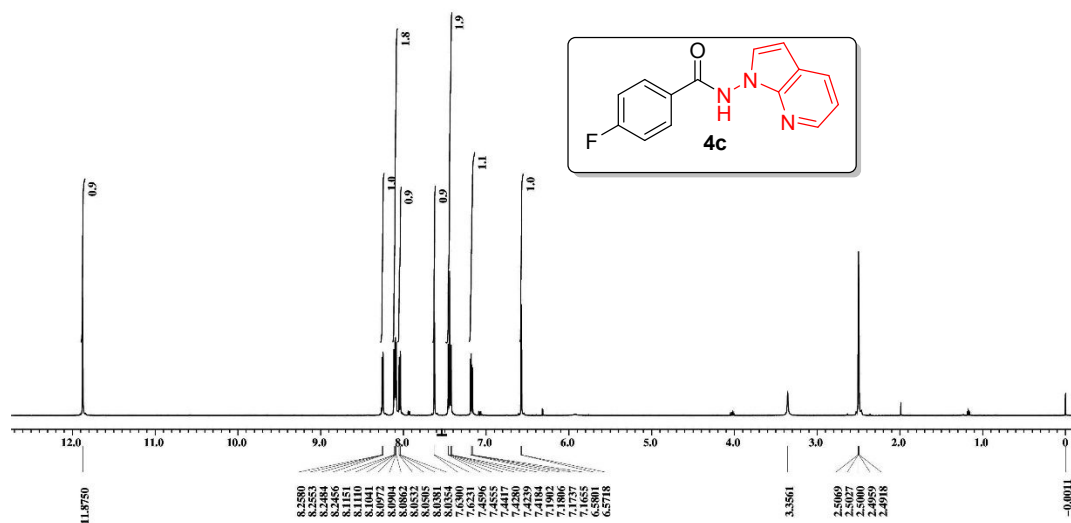


Fig. S7. ¹H NMR spectrum of **4c** (500 MHz, DMSO-d₆)

250719_SPS_05_4FAZA_CARBON-4.jdf

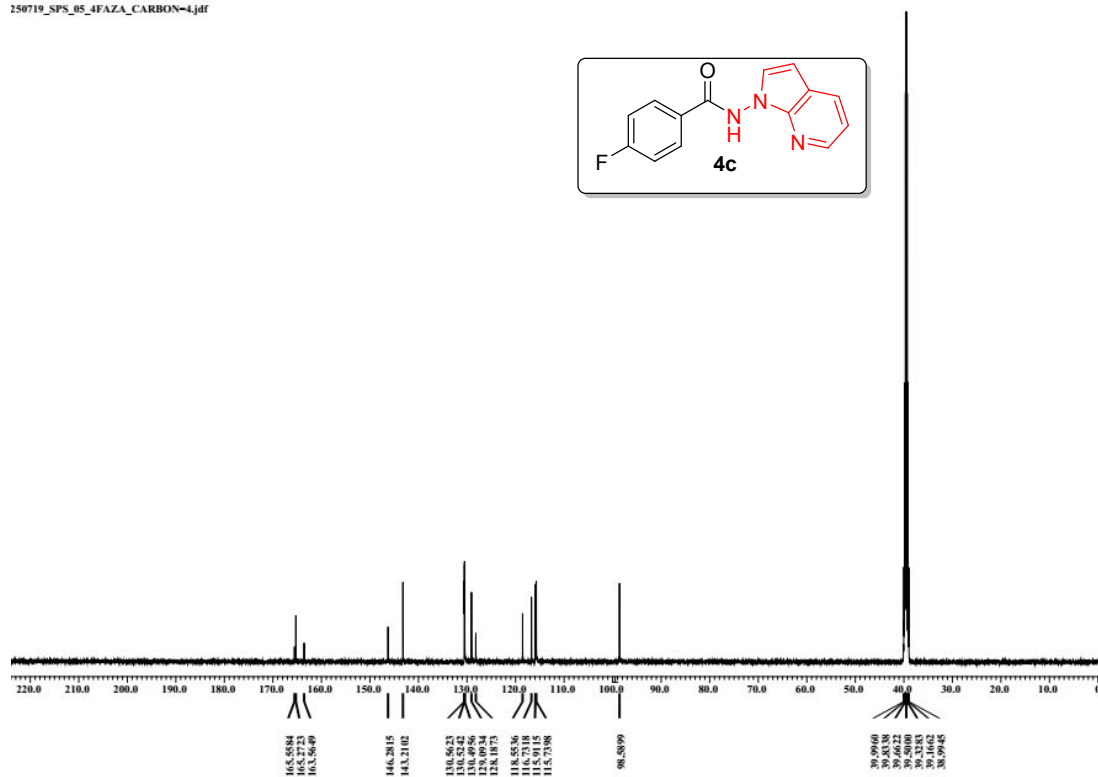


Fig. S8. ¹³C NMR spectrum of **4c** (125 MHz, DMSO-d₆)

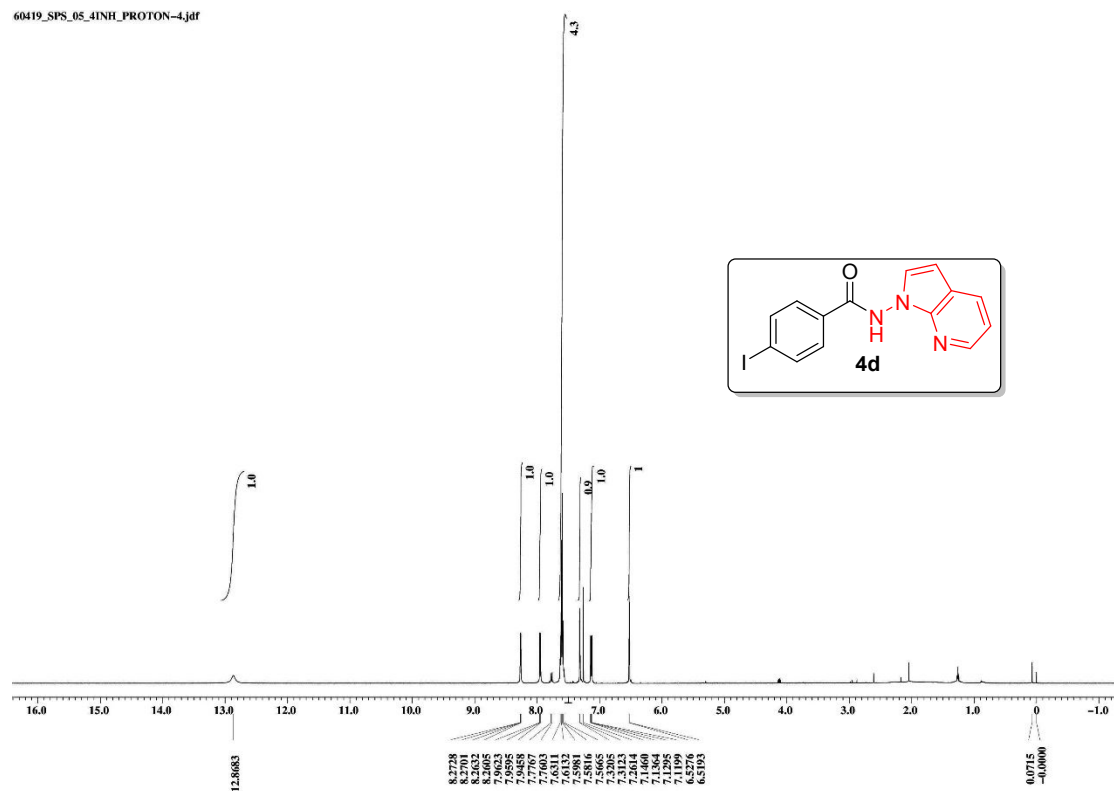


Fig. S9. ¹H NMR spectrum of **4d** (500 MHz, CDCl₃)

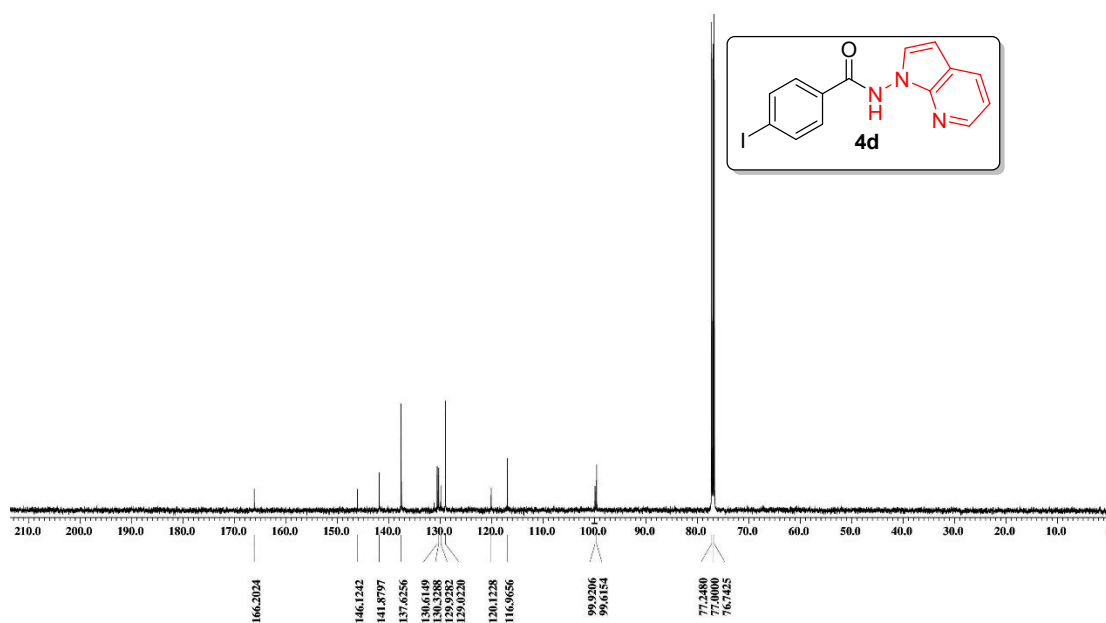


Fig. S10. ¹³C NMR spectrum of **4d** (125 MHz, CDCl₃)

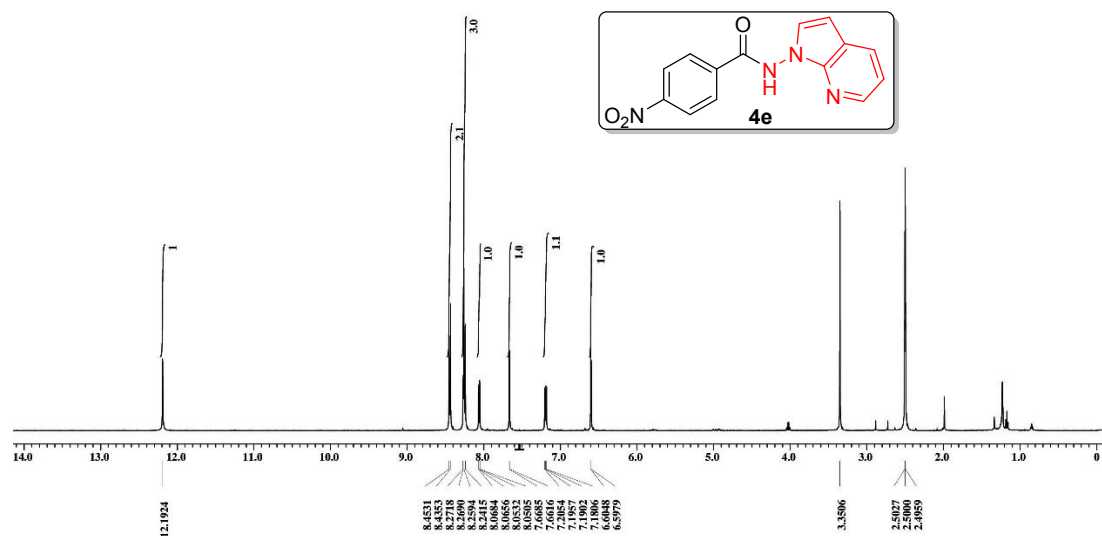


Fig. S11. ^1H NMR spectrum of **4e** (500 MHz, DMSO-d_6)

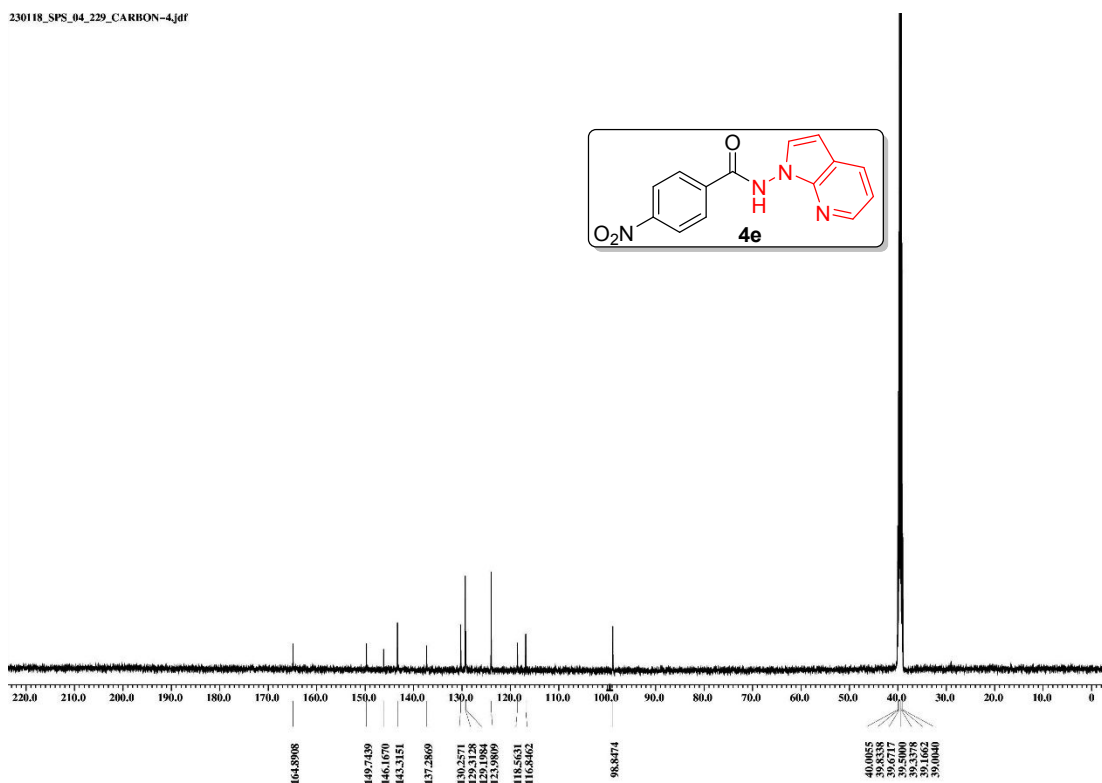


Fig. S12. ^{13}C NMR spectrum of **4e** (125 MHz, DMSO-d_6)

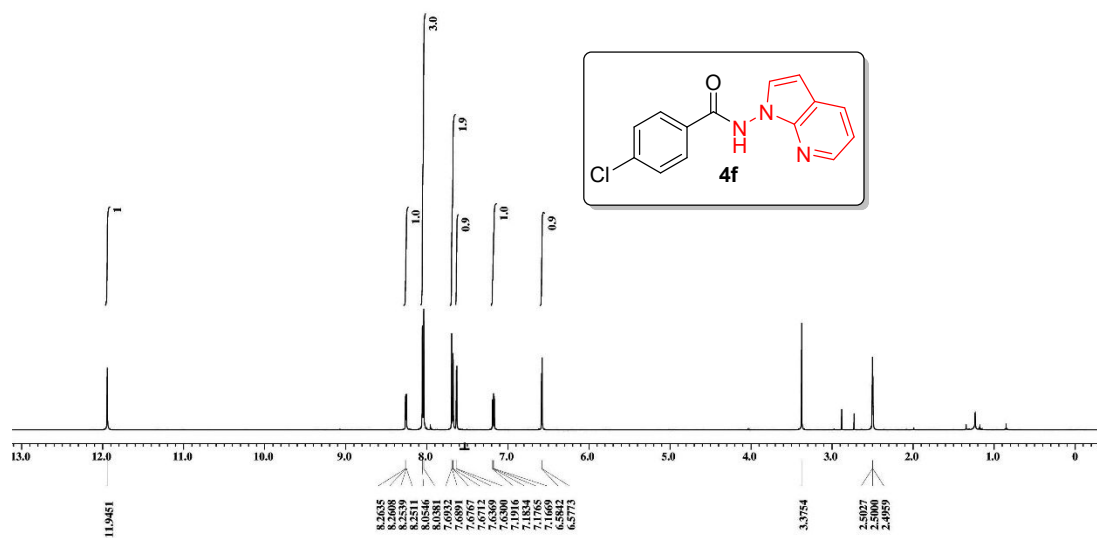


Fig. S13. ¹H NMR spectrum of **4f** (500 MHz, DMSO-d₆)

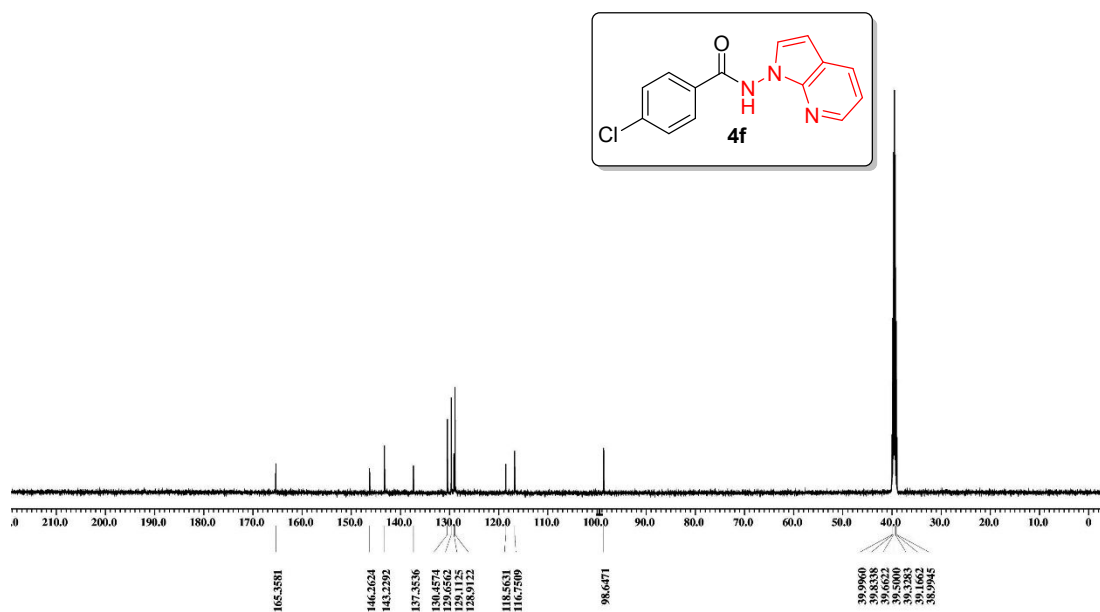


Fig. S14. ¹³C NMR spectrum of **4f** (125 MHz, DMSO-d₆)

4g

166.9113
155.4463
146.4803
142.1785
130.6466
127.5085
127.8710
126.5851
120.0782
116.9019
99.4886
77.3751
77.1176
76.8900
34.9868
31.1619
29.7979

S15

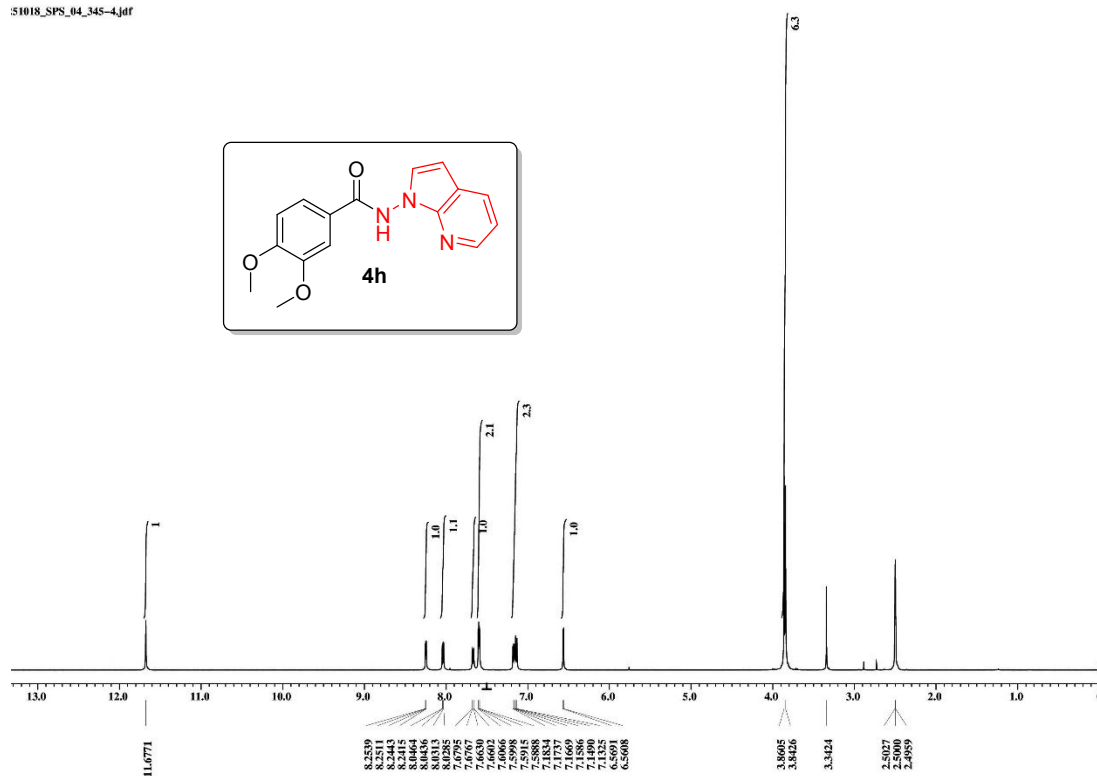


Fig. S17. ^1H NMR spectrum of **4h** (500 MHz, DMSO-d_6)

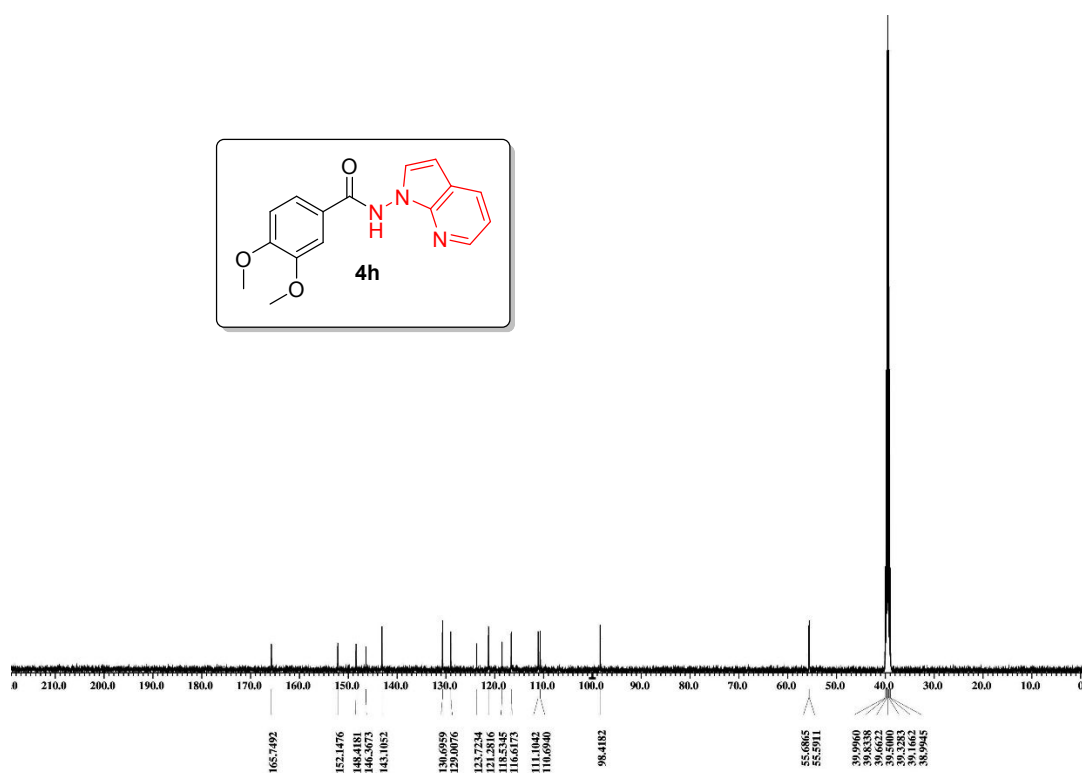


Fig. S18. ^{13}C NMR spectrum of **4h** (125 MHz, DMSO-d_6)

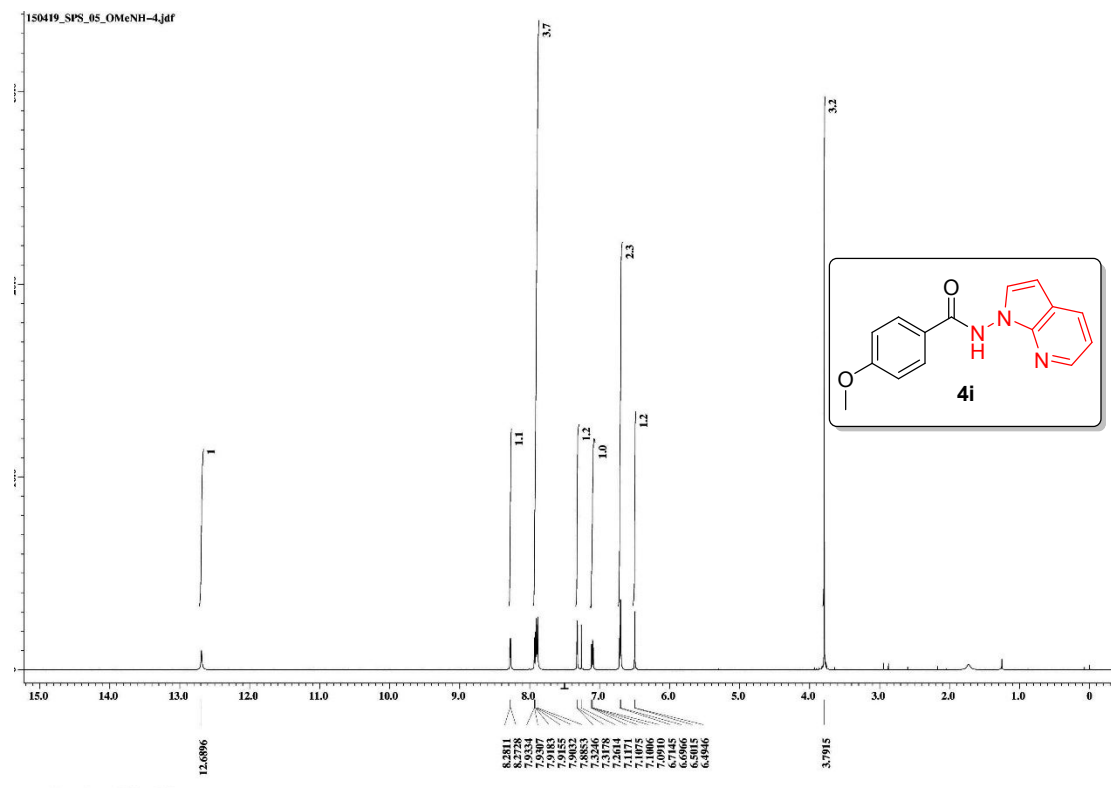


Fig. S19. ^1H NMR spectrum of **4i** (500 MHz, CDCl_3)

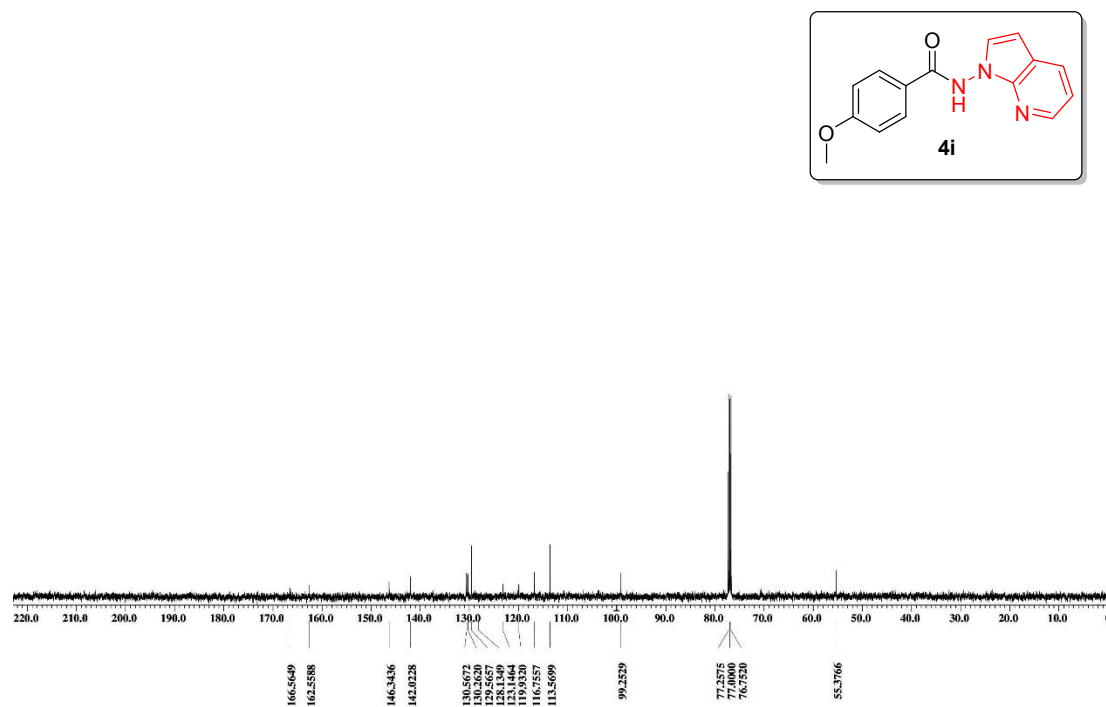


Fig. S20. ^1H NMR spectrum of **4i** (125 MHz, CDCl_3)

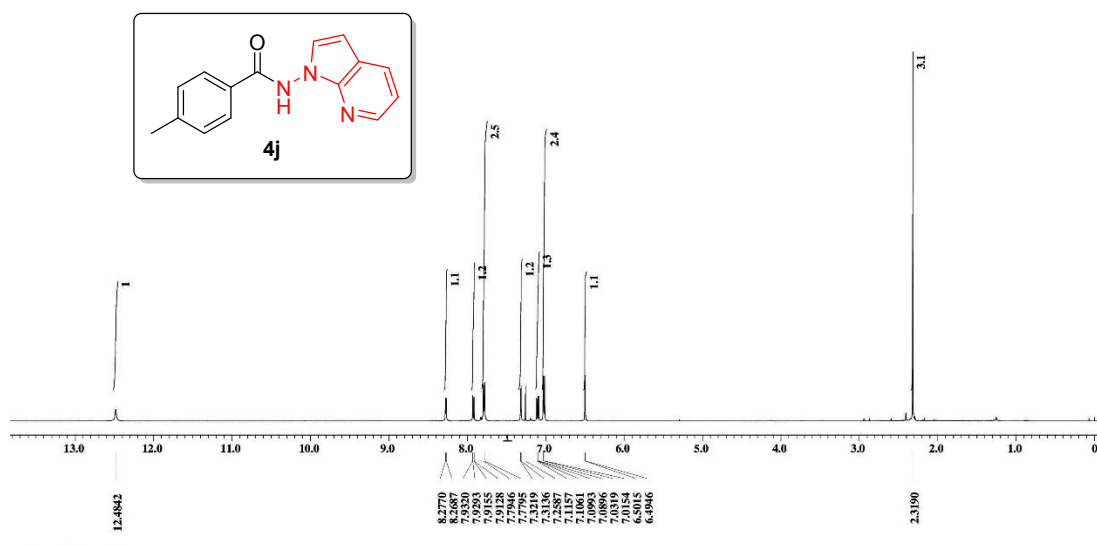


Fig. S21. ¹H NMR spectrum of **4j** (500 MHz, CDCl₃)

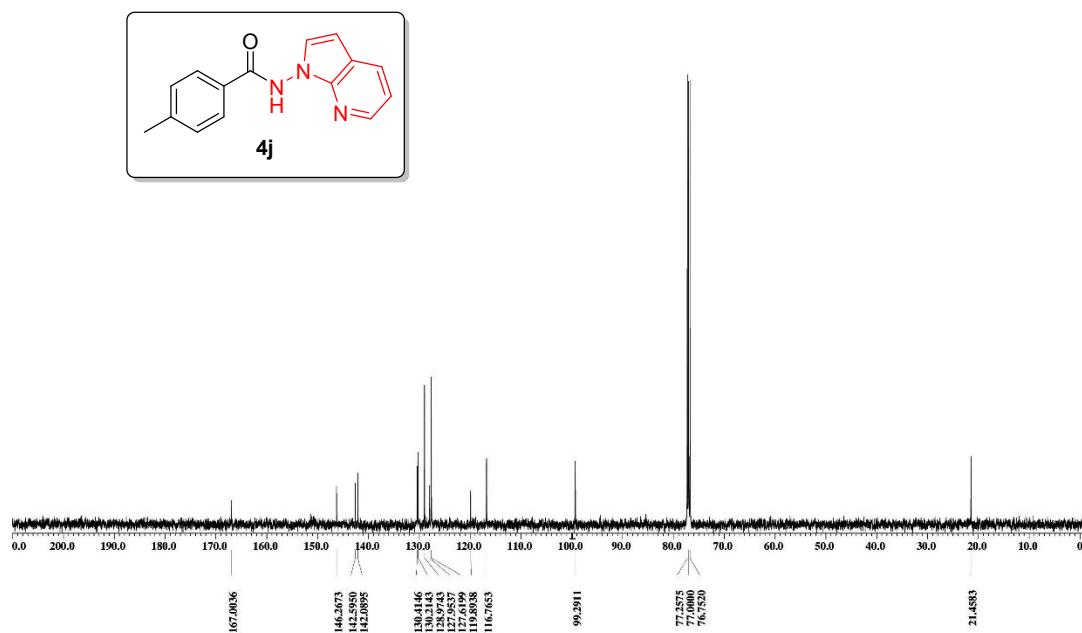


Fig. S22. ¹³C NMR spectrum of **4j** (125 MHz, CDCl₃)

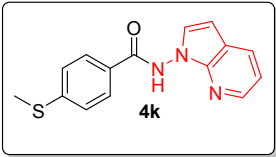
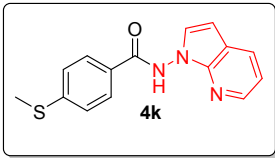


Fig. S24. ^{13}C NMR spectrum of **4k** (125 MHz, DMSO- d_6)



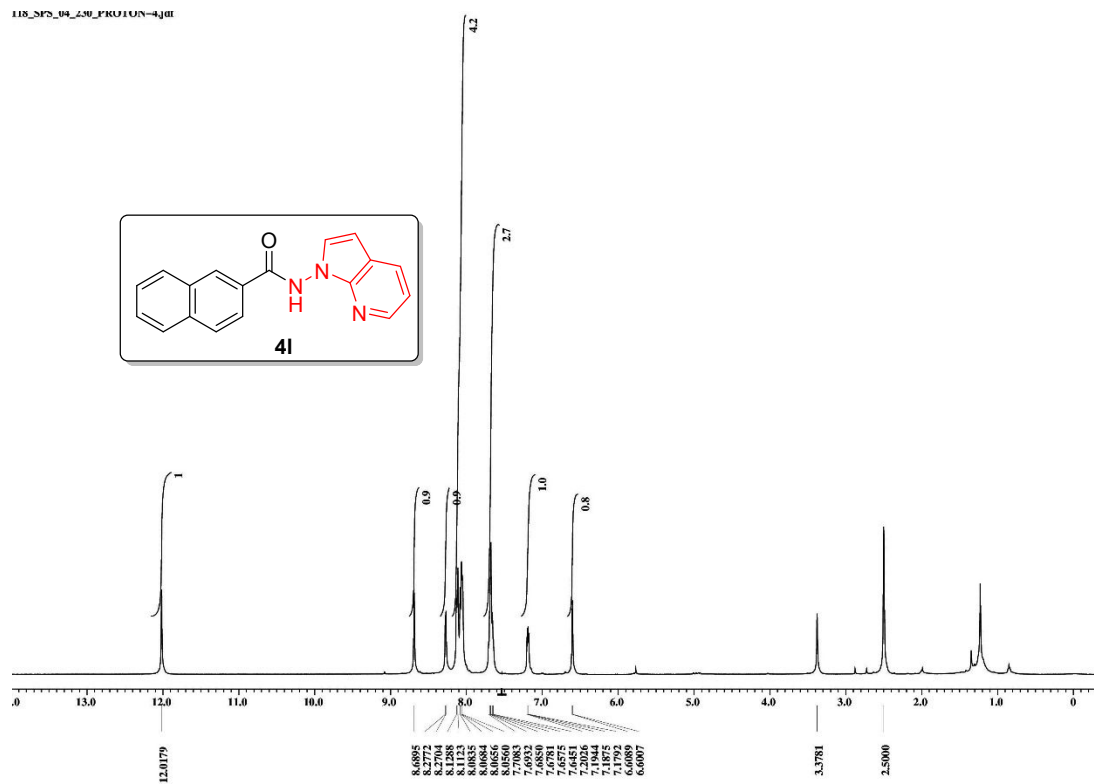


Fig. S25. ¹H NMR spectrum of **4I** (500 MHz, DMSO-d₆)

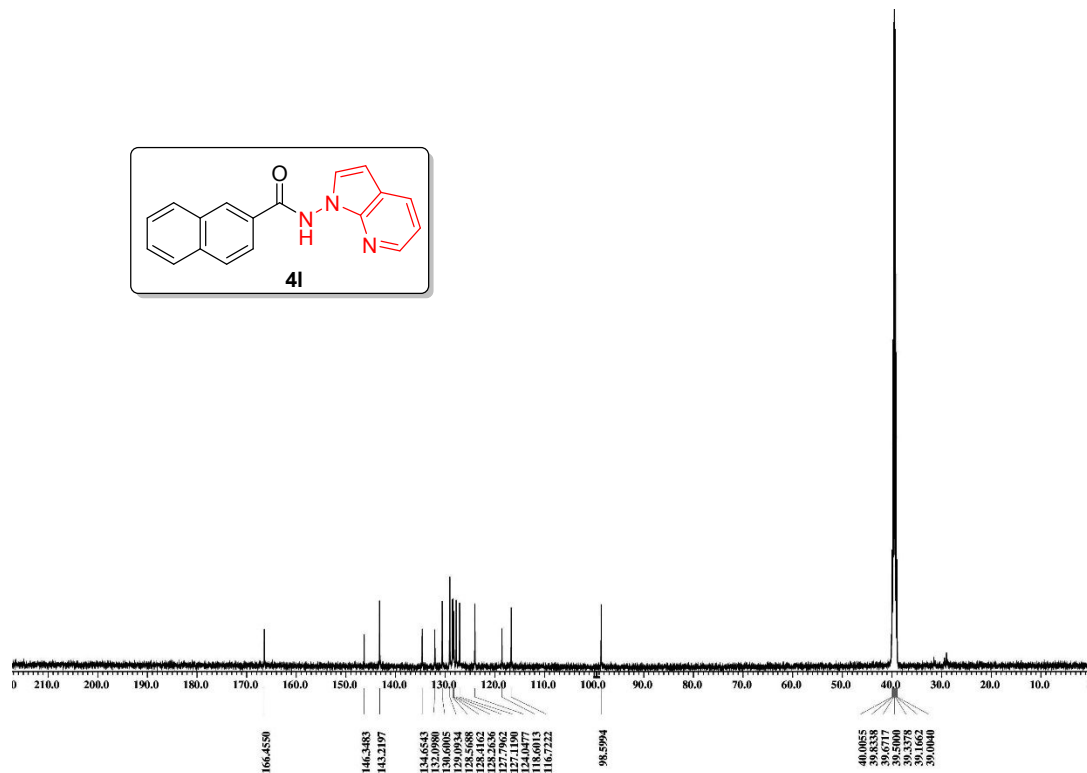


Fig. S26. ¹³C NMR spectrum of **4I** (125 MHz, DMSO-d₆)

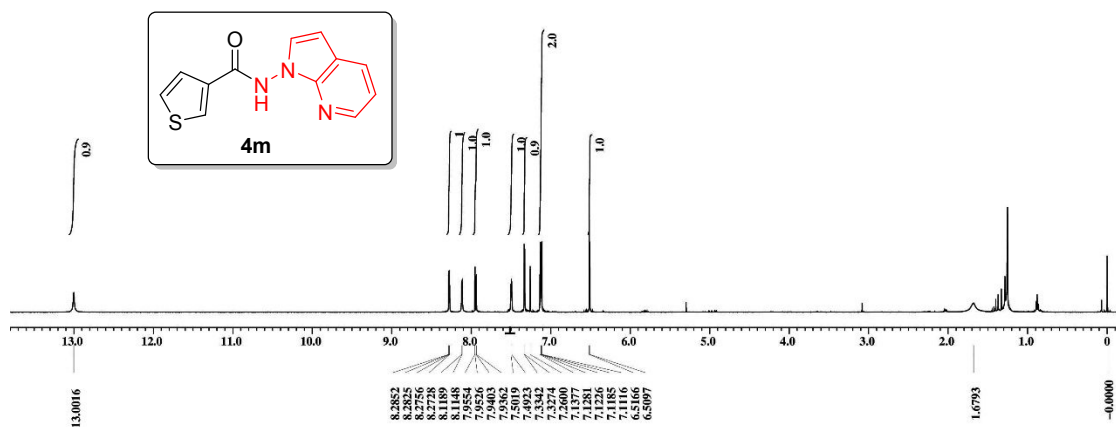


Fig. S27. ¹H NMR spectrum of **4m** (500 MHz, CDCl₃)

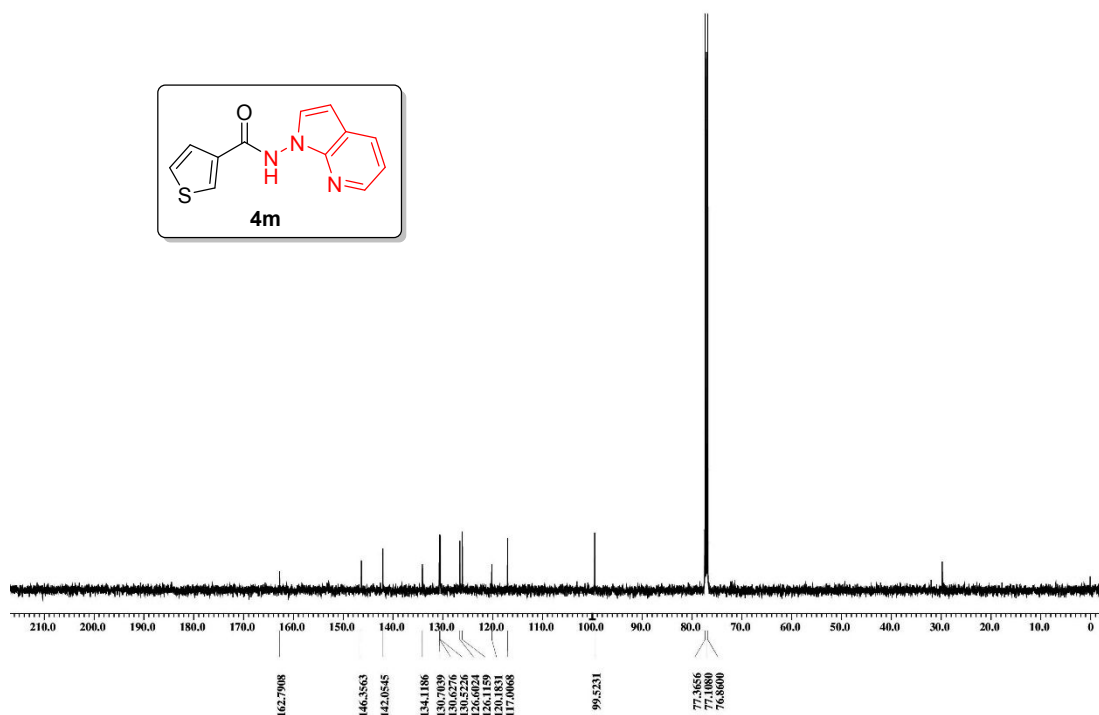


Fig. S28. ¹³C NMR spectrum of **4m** (125 MHz, CDCl₃)

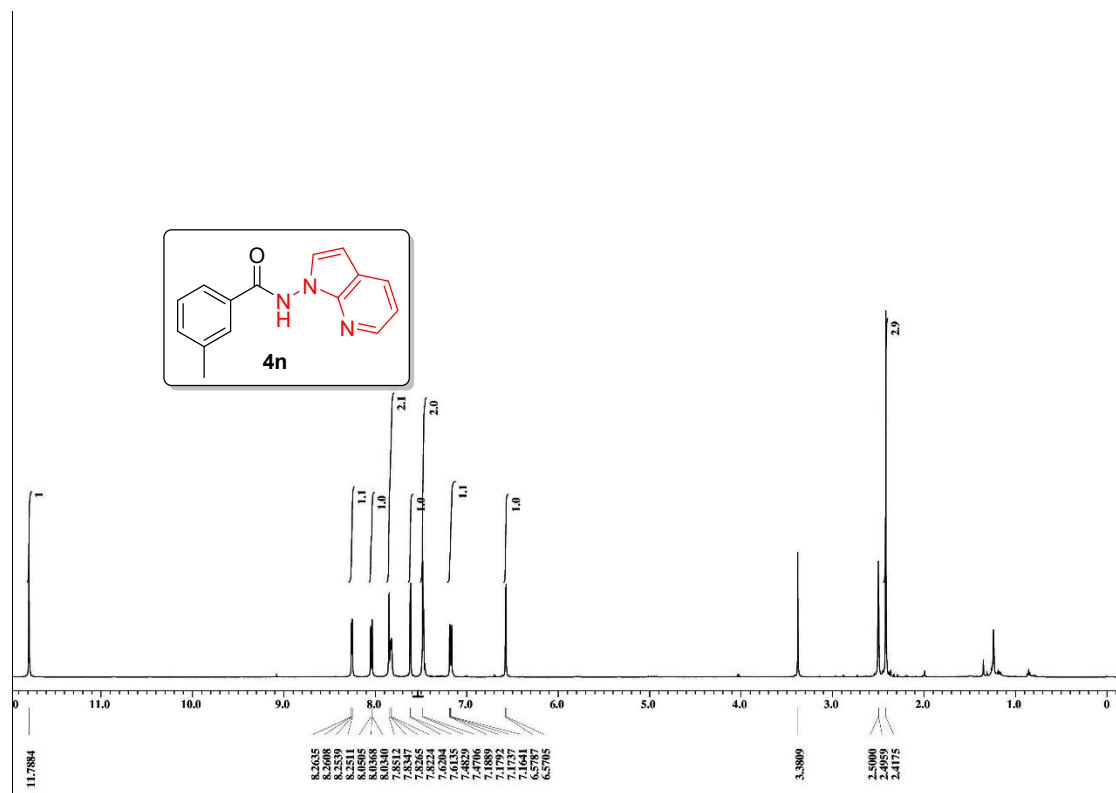


Fig. S29. ¹H NMR spectrum of **4n** (500 MHz, DMSO-d₆)

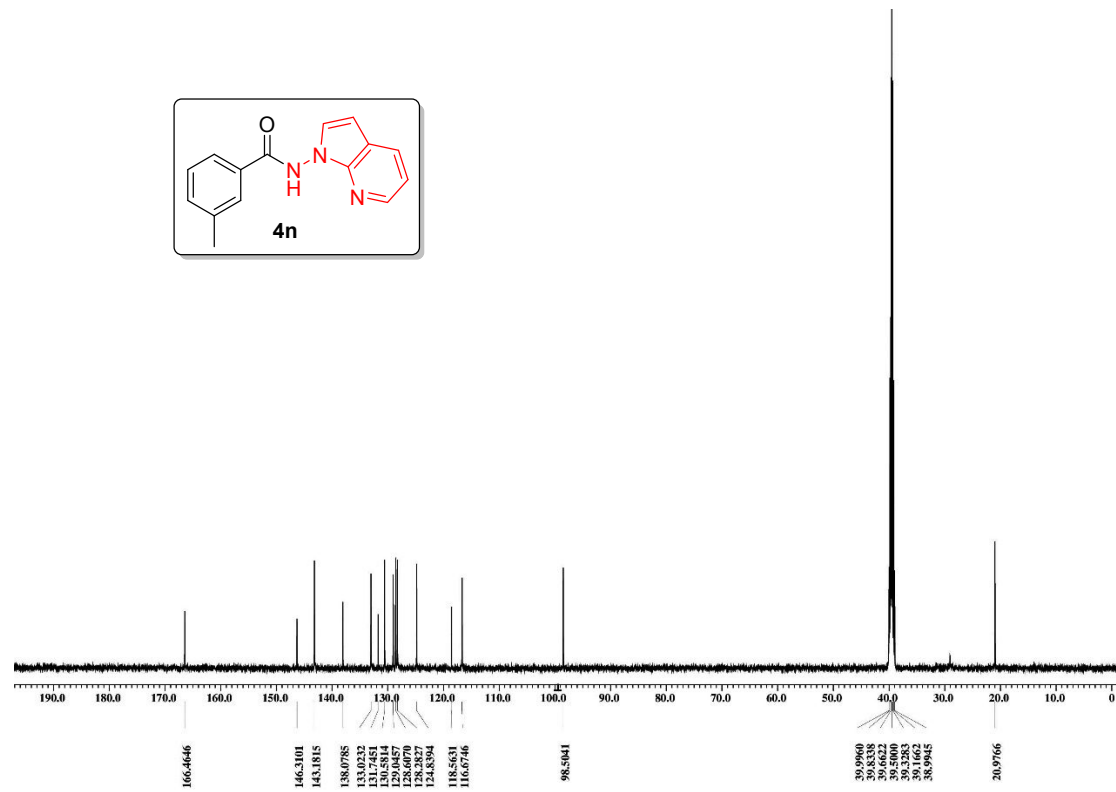


Fig. S30. ¹³C NMR spectrum of **4n** (125 MHz, DMSO-d₆)

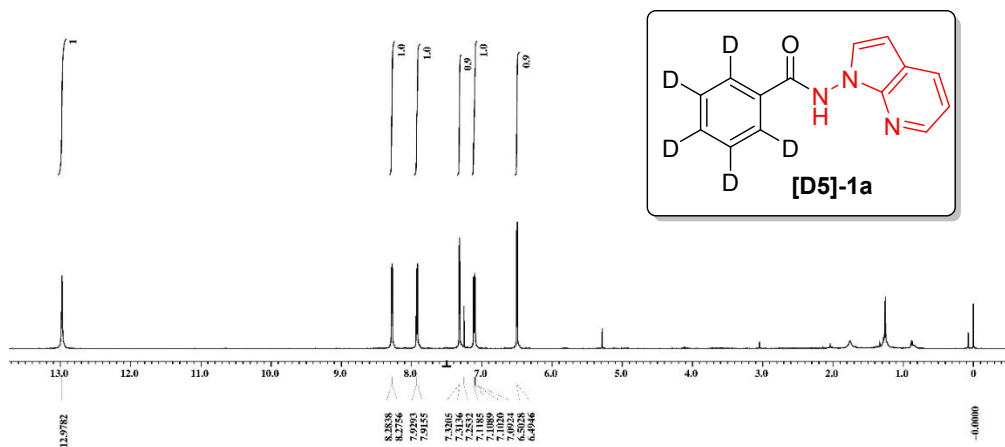


Fig. S31. ¹H NMR spectrum of **[D₅]-1a** (500 MHz, CDCl₃)

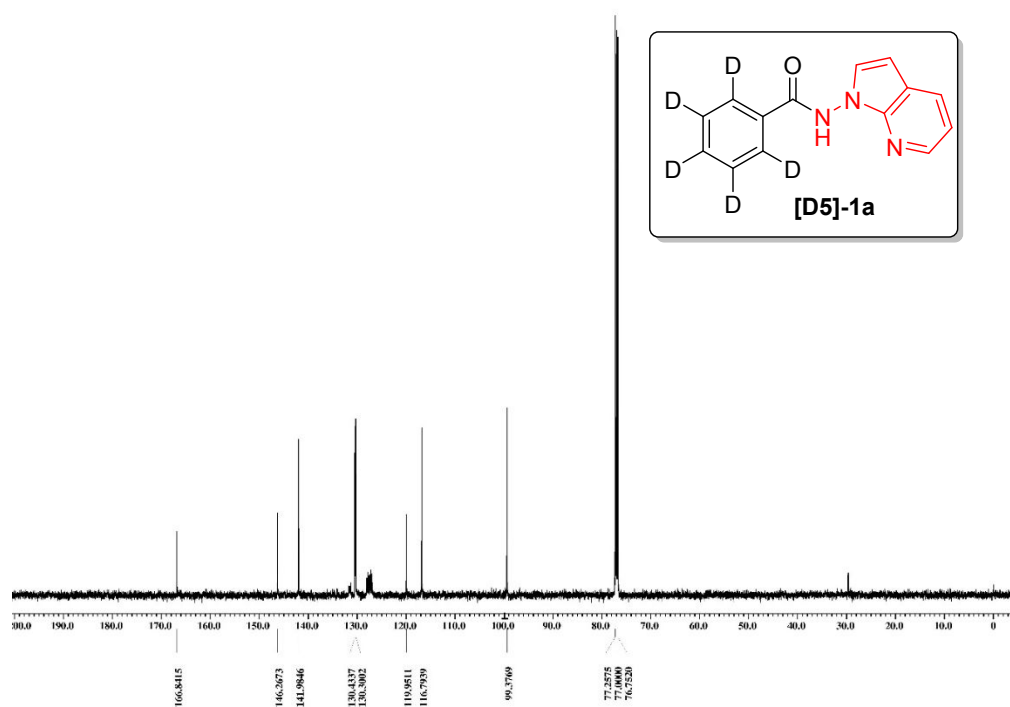


Fig. S32. ¹³C NMR spectrum of **[D₅]-1a** (125 MHz, CDCl₃)

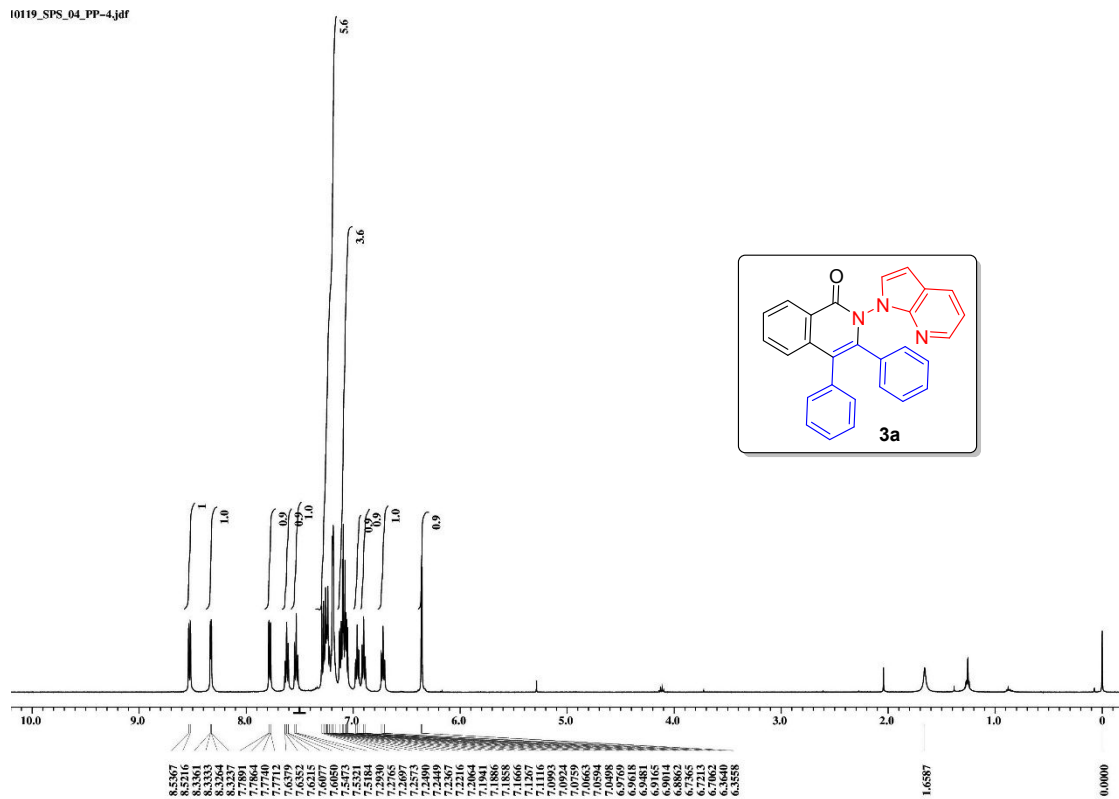


Fig. S33. ¹H NMR spectrum of **3a** (500 MHz, CDCl₃)

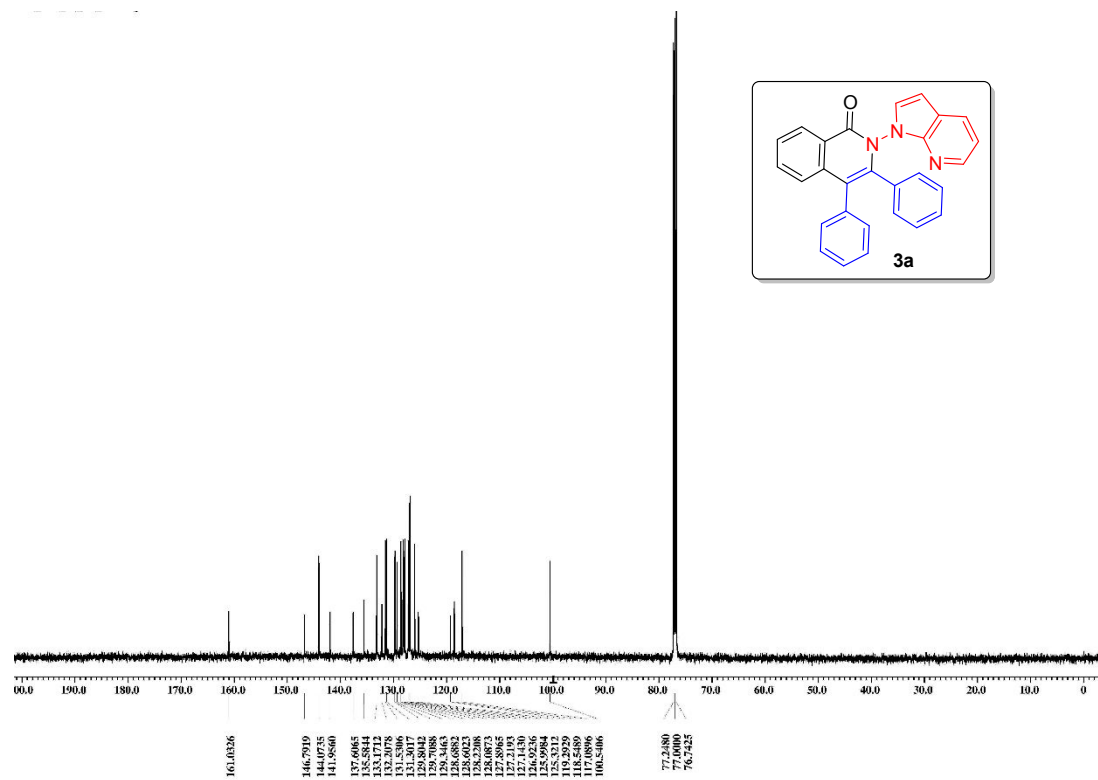


Fig. S34. ¹³C NMR spectrum of **3a** (125 MHz, CDCl₃)

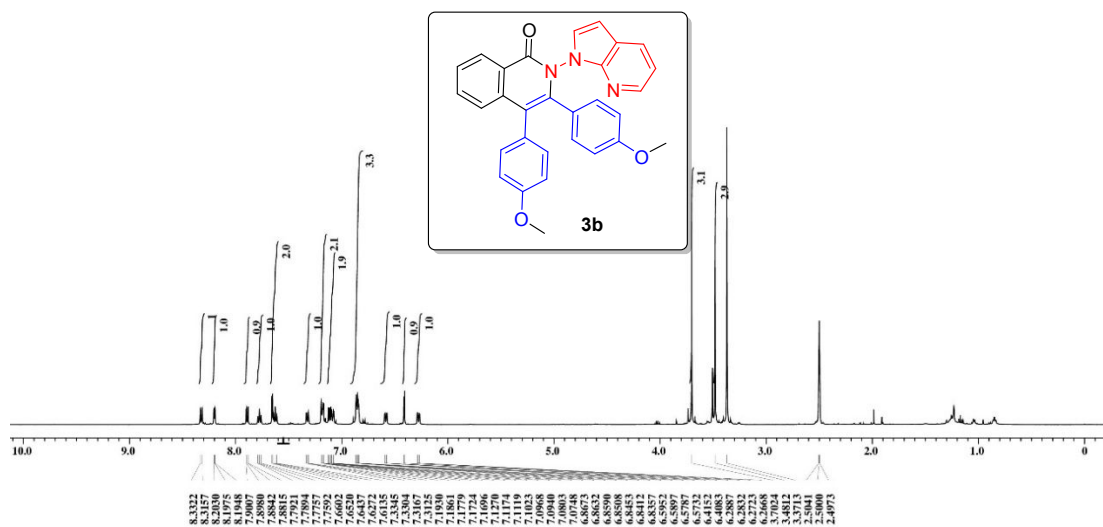


Fig. S35. ¹H NMR spectrum of **3b** (500 MHz, DMSO)

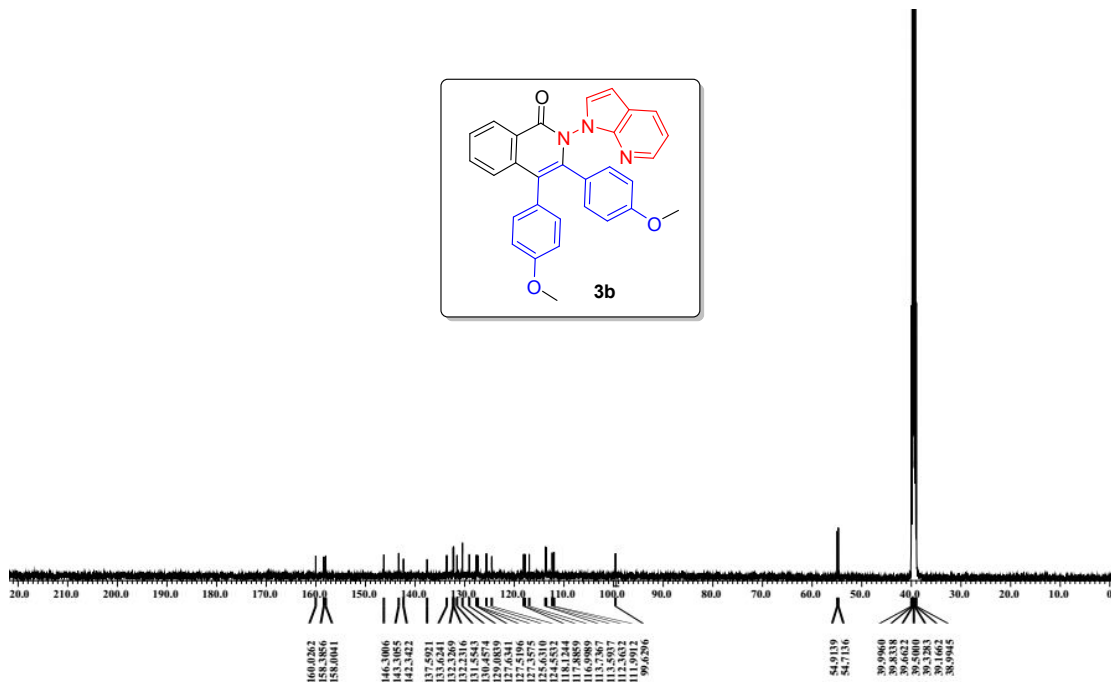


Fig. S36. ¹³C NMR spectrum of **3b** (125 MHz, DMSO)

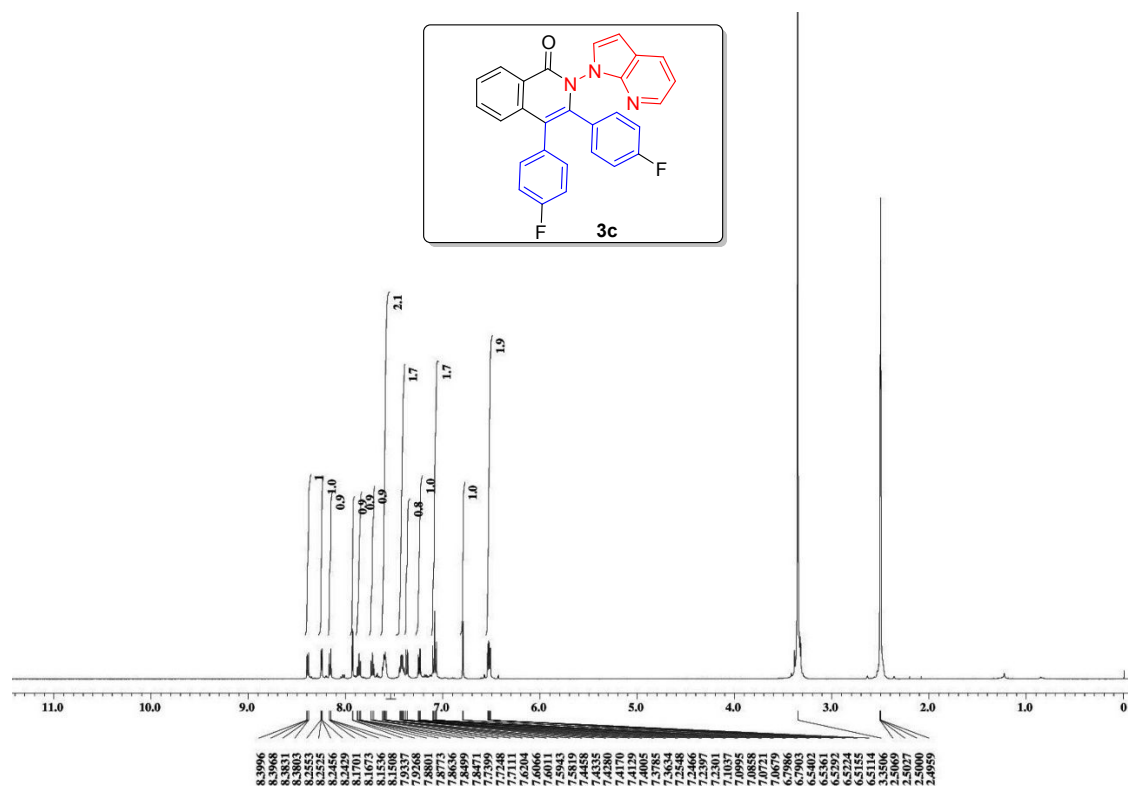


Fig. S37. ^1H NMR spectrum of **3c** (500 MHz, CDCl_3)

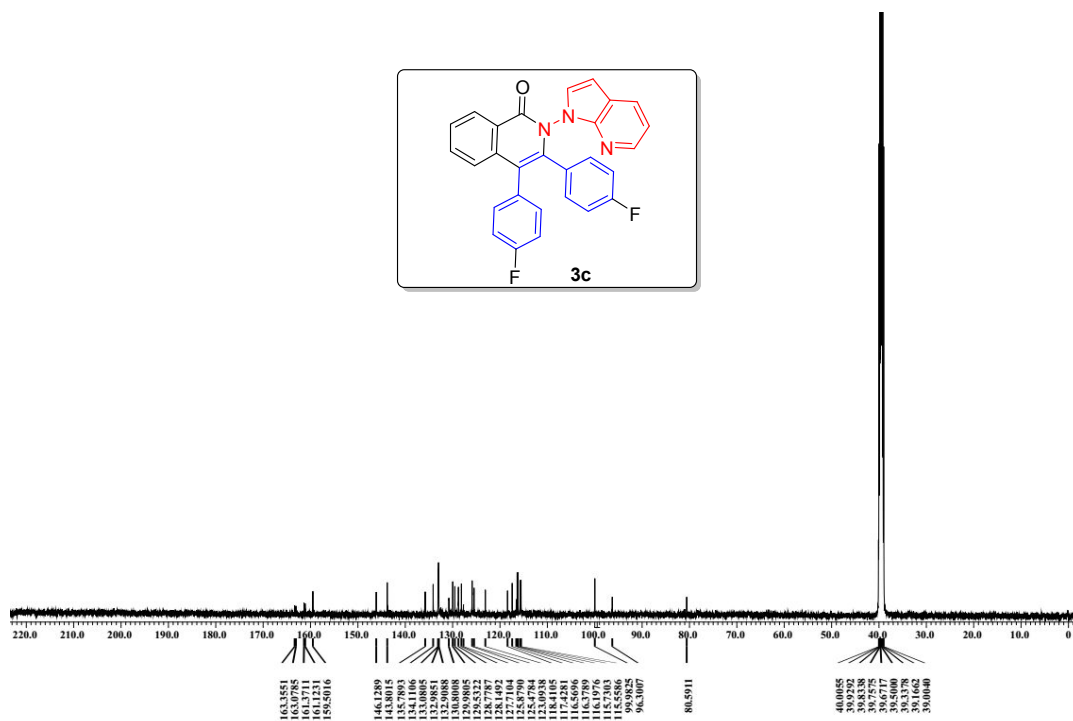


Fig. S38. ^{13}C NMR spectrum of **3c** (125 MHz, CDCl_3)

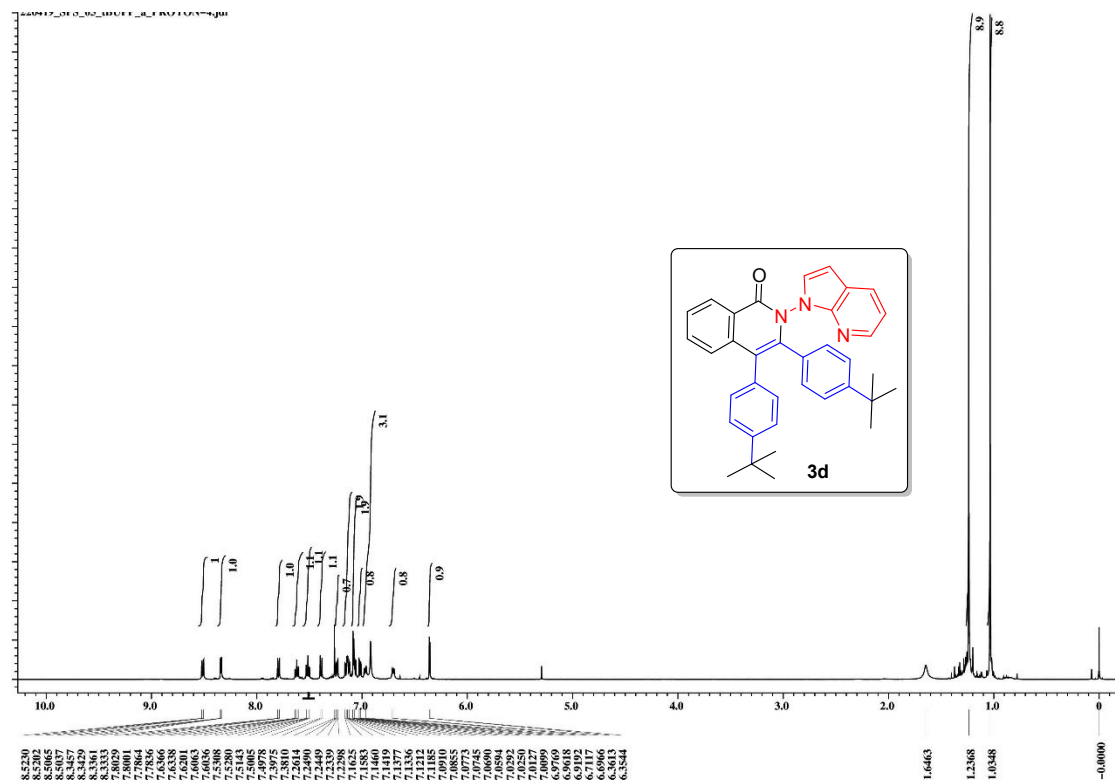


Fig. S39. ^1H NMR spectrum of **3d** (500 MHz, CDCl_3)

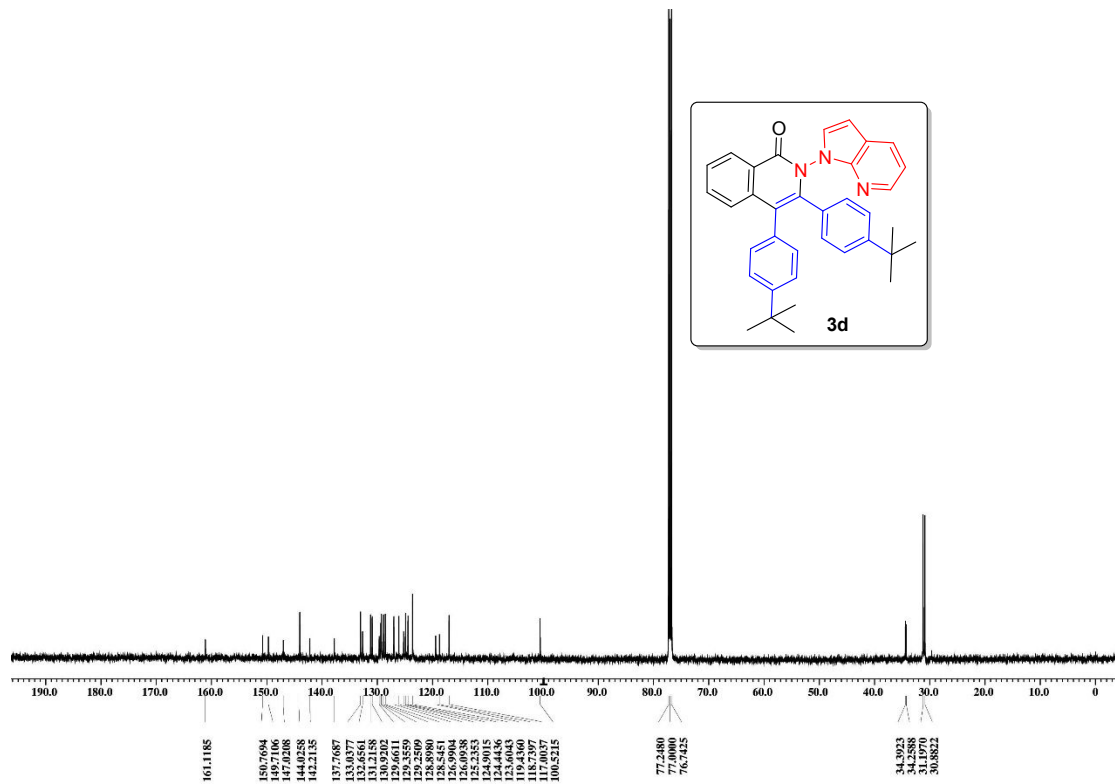


Fig. S40. ^{13}C NMR spectrum of **3d** (125 MHz, CDCl_3)

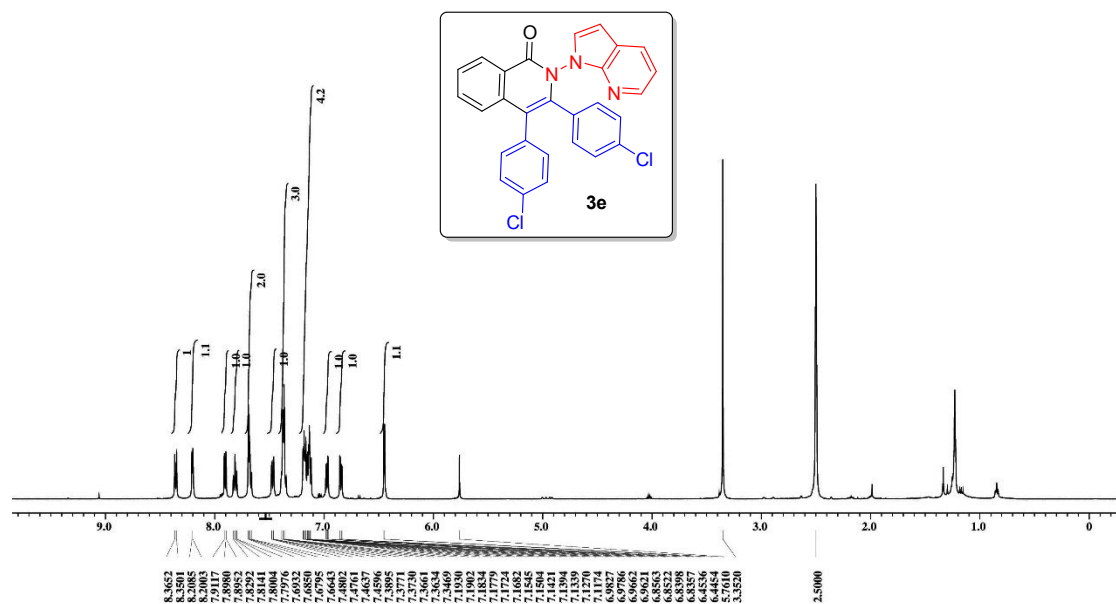


Fig. S41. ¹H NMR spectrum of **3e** (500 MHz, DMSO)

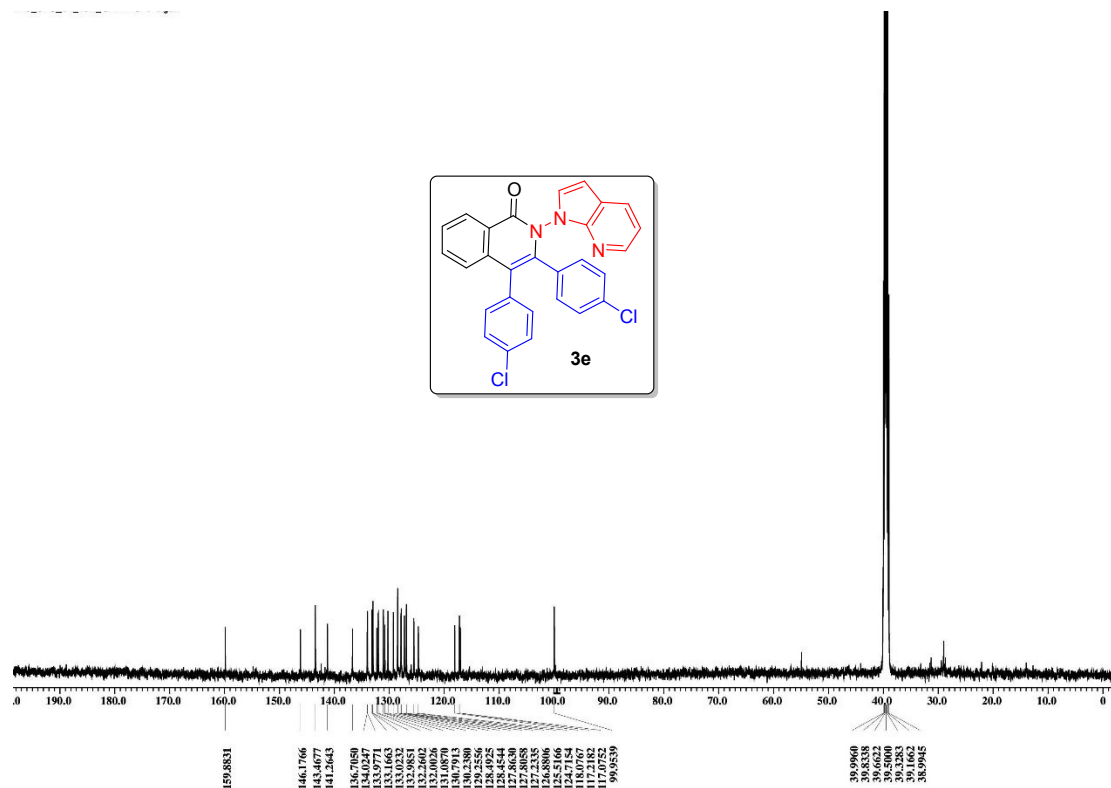


Fig. S42. ¹³C NMR spectrum of **3e** (125 MHz, DMSO)

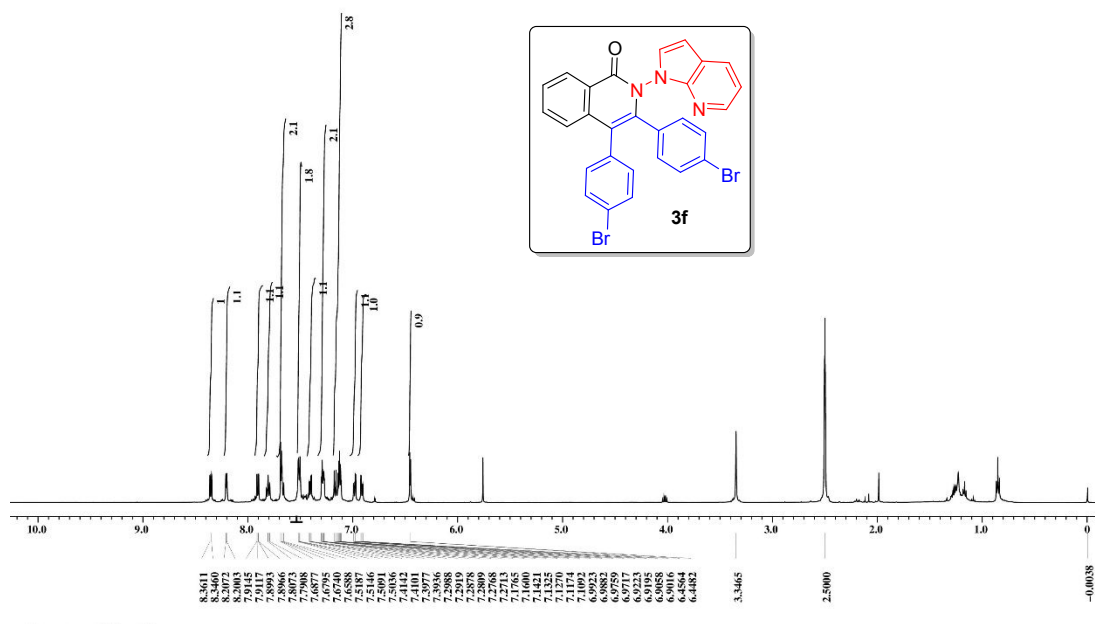


Fig. S43. ¹H NMR spectrum of **3f** (500 MHz, DMSO)

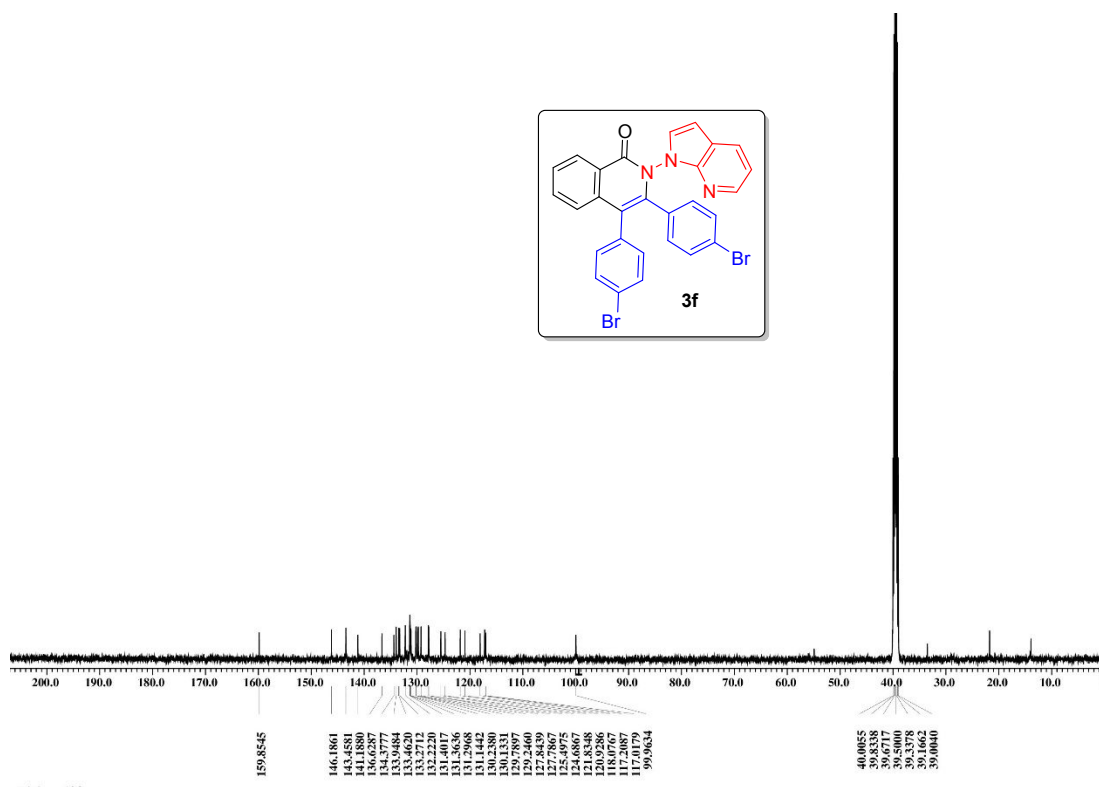


Fig. S44. ¹³C NMR spectrum of **3f** (125 MHz, DMSO)

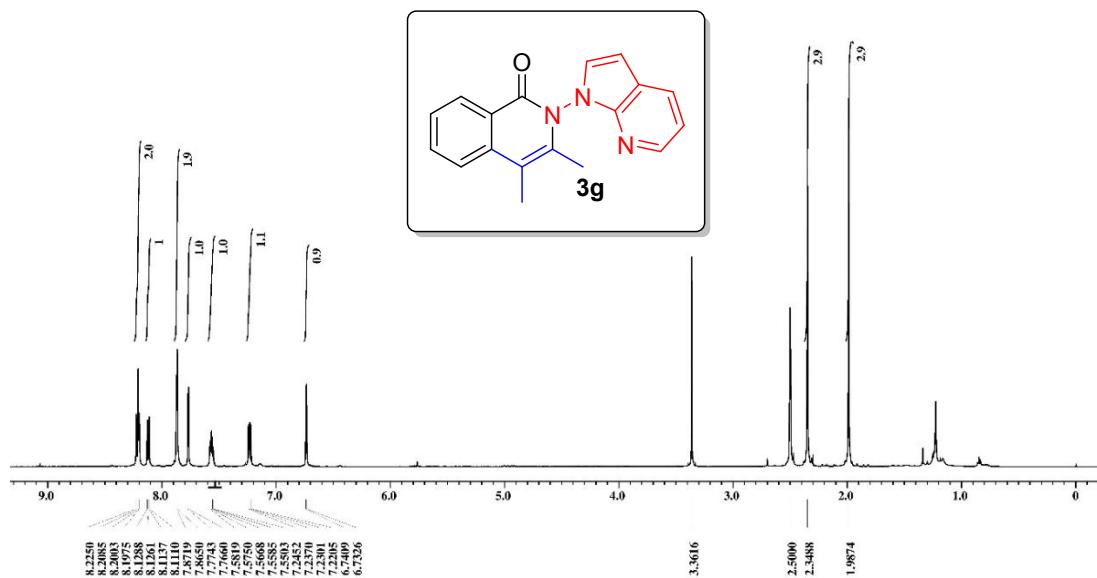


Fig. S45. ¹H NMR spectrum of **3g** (500 MHz, DMSO)

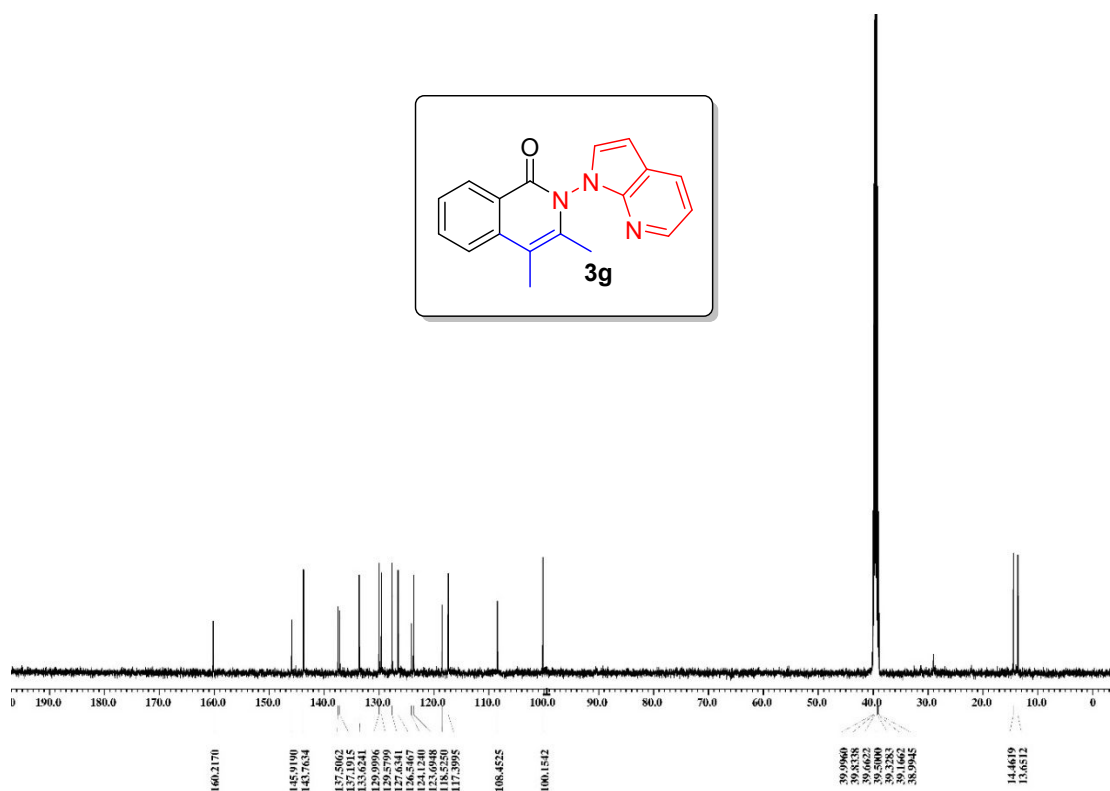


Fig. S46. ¹³C NMR spectrum of **3g** (125 MHz, DMSO)

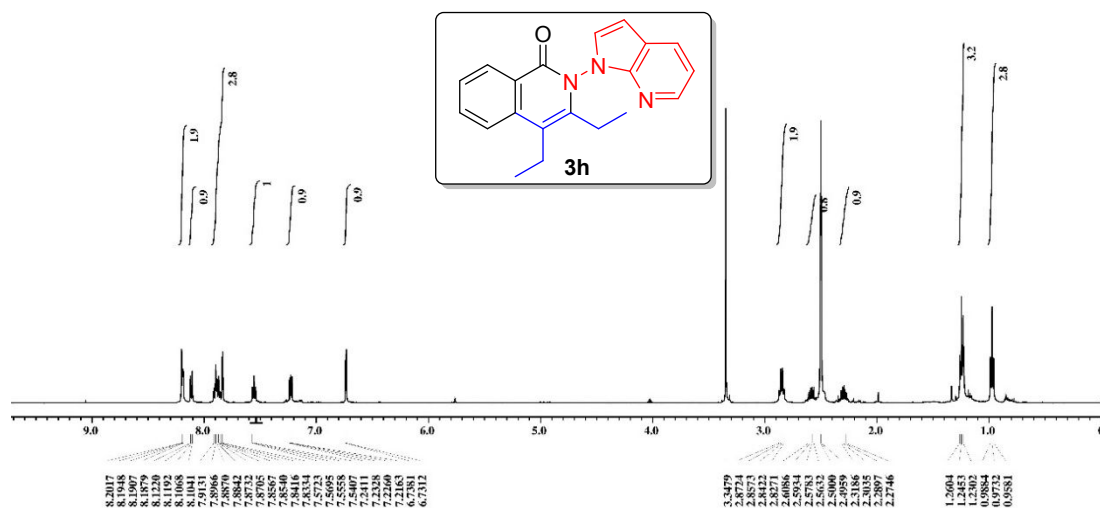


Fig. S47. ¹H NMR spectrum of **3h** (500 MHz, DMSO)

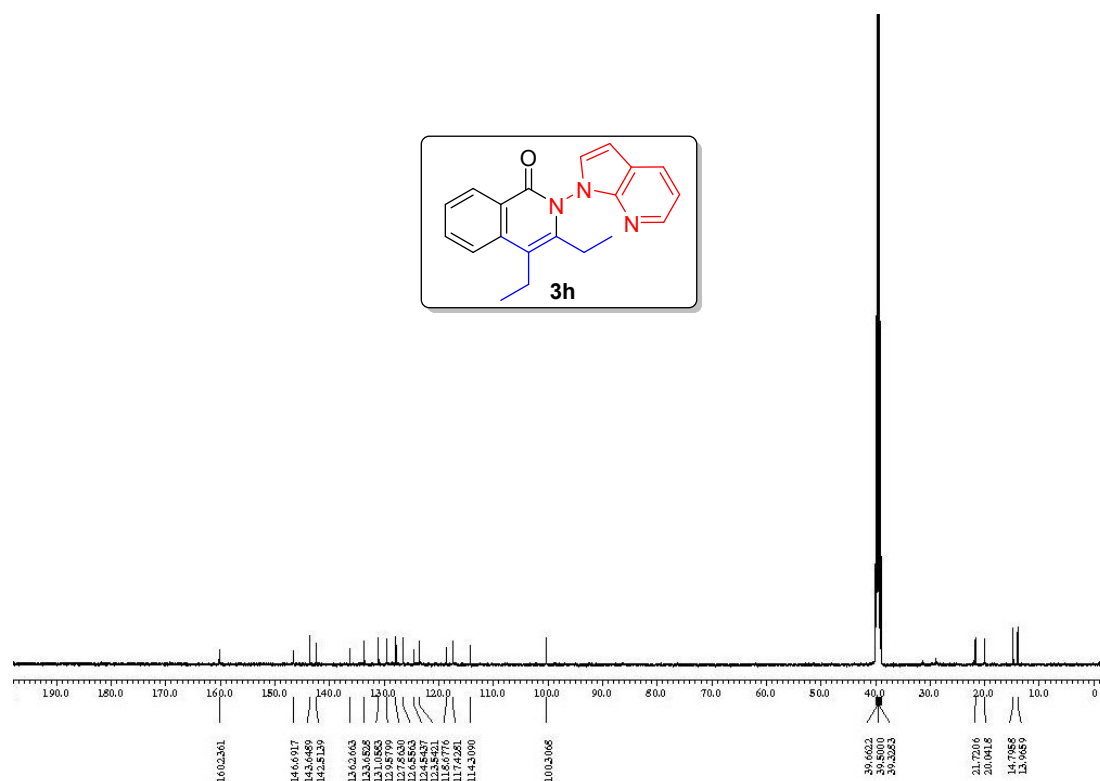


Fig. S48. ¹³C NMR spectrum of **3h** (125 MHz, DMSO)

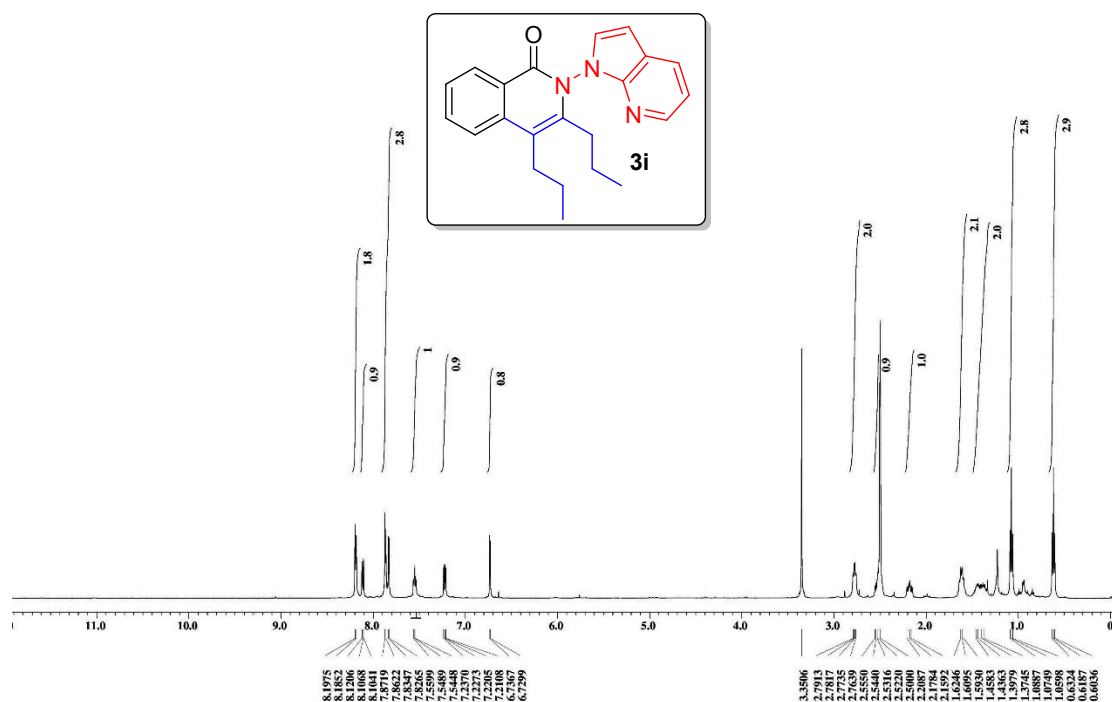


Fig. S49. ¹H NMR spectrum of **3i** (500 MHz, DMSO)

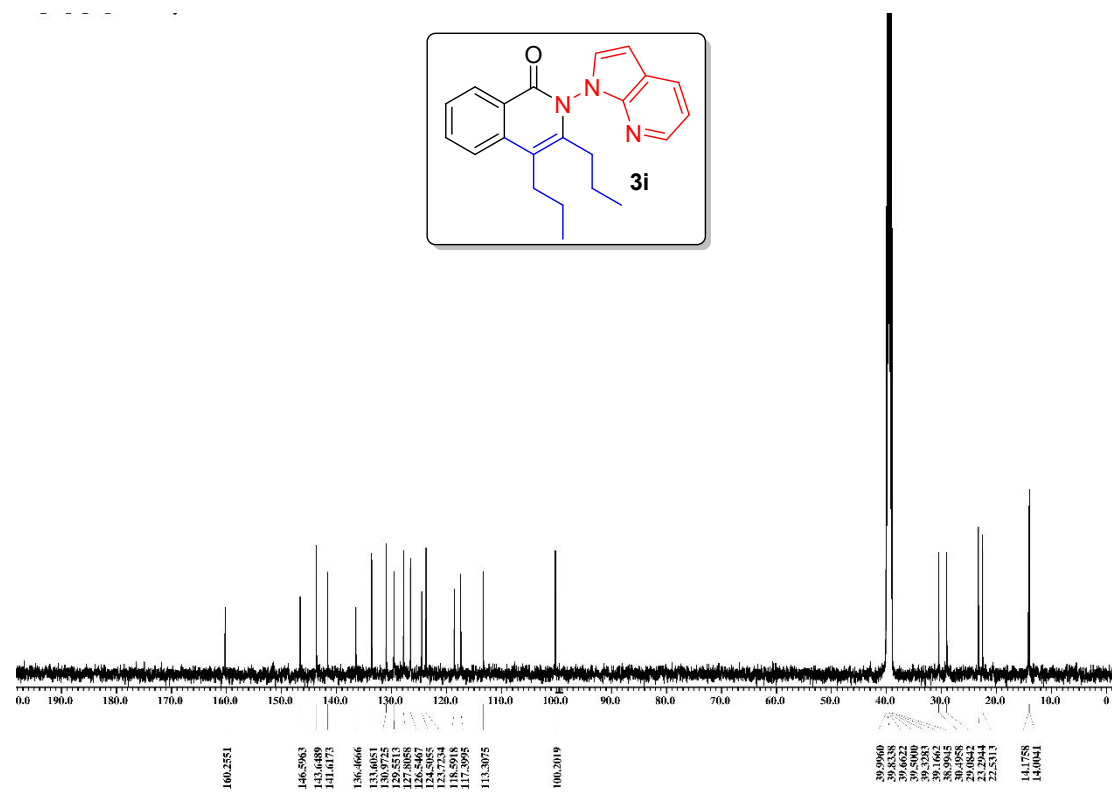


Fig. S50. ¹³C NMR spectrum of **3i** (125 MHz, DMSO)

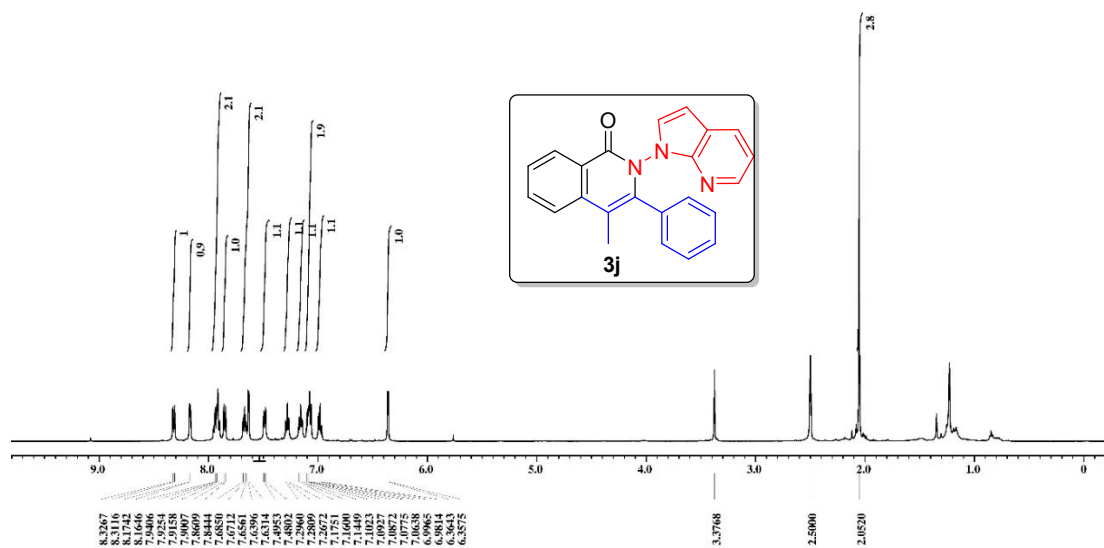


Fig. S51. ¹H NMR spectrum of **3j** (500 MHz, DMSO)

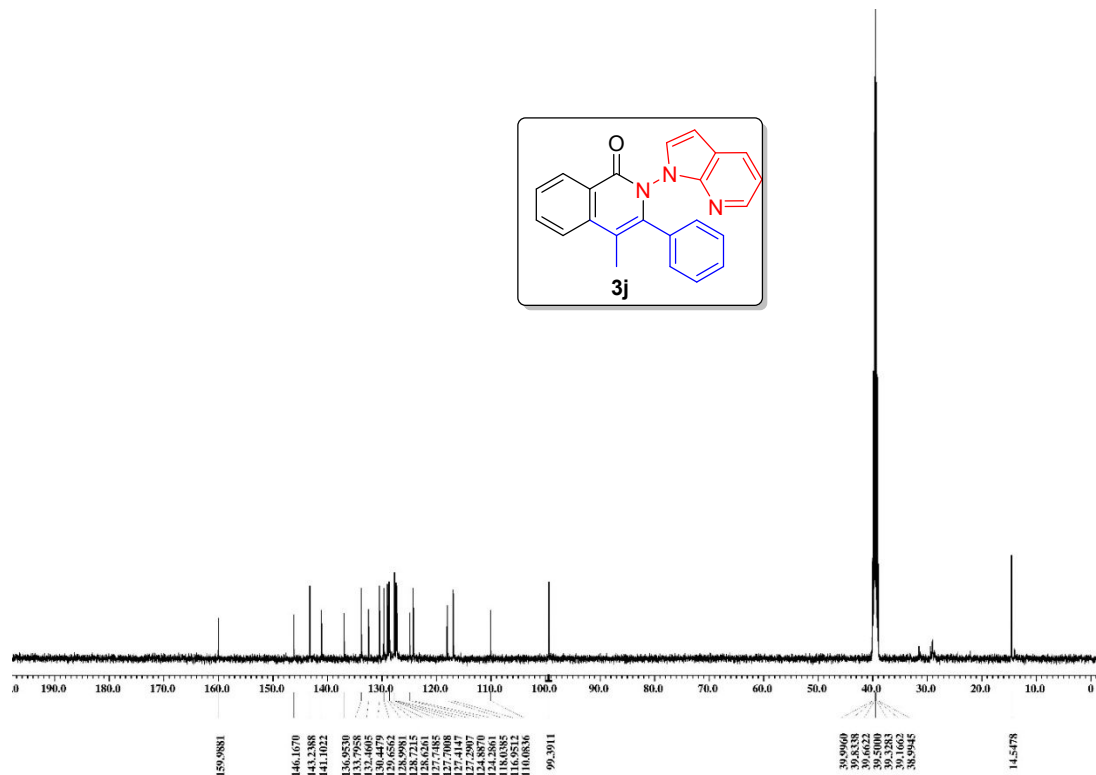


Fig. S52. ¹³C NMR spectrum of **3j** (125 MHz, DMSO)

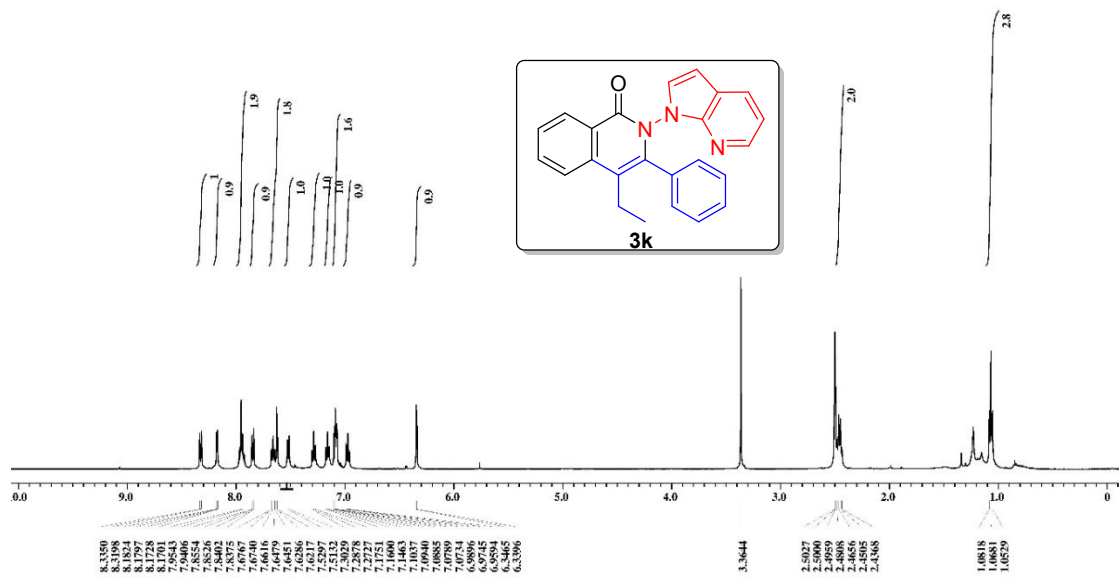


Fig. S53. ¹H NMR spectrum of **3k** (500 MHz, DMSO)

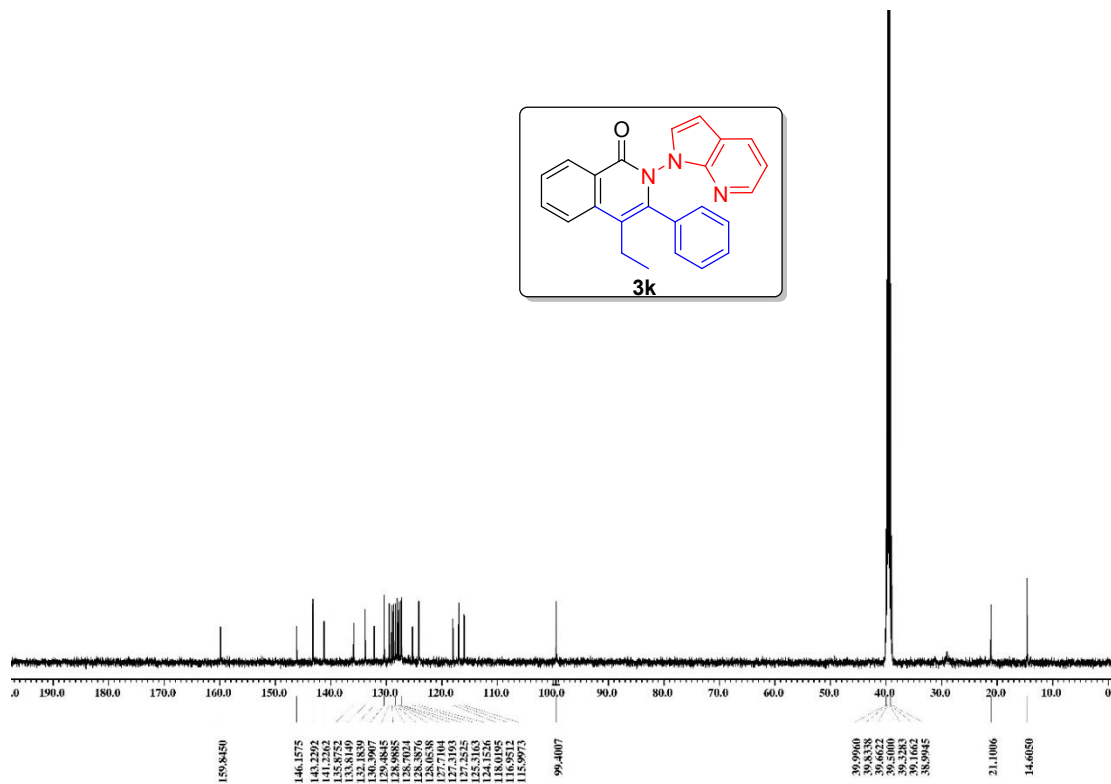


Fig. S54. ¹³C NMR spectrum of **3k** (125 MHz, DMSO)

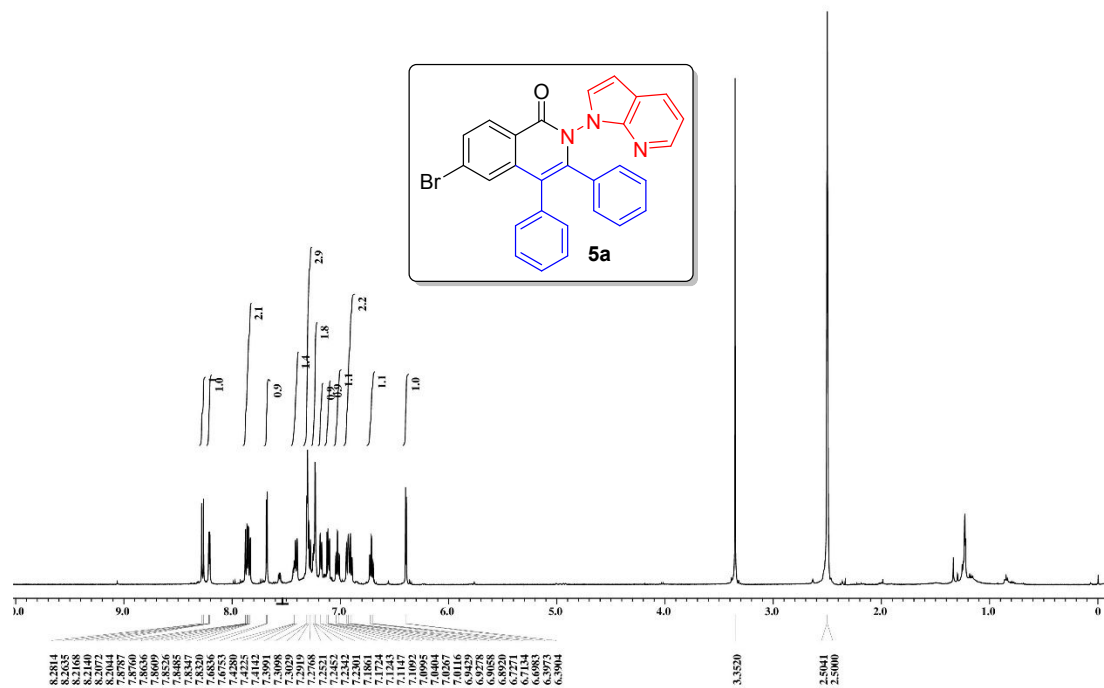


Fig. S55. ¹H NMR spectrum of **5a** (500 MHz, DMSO)

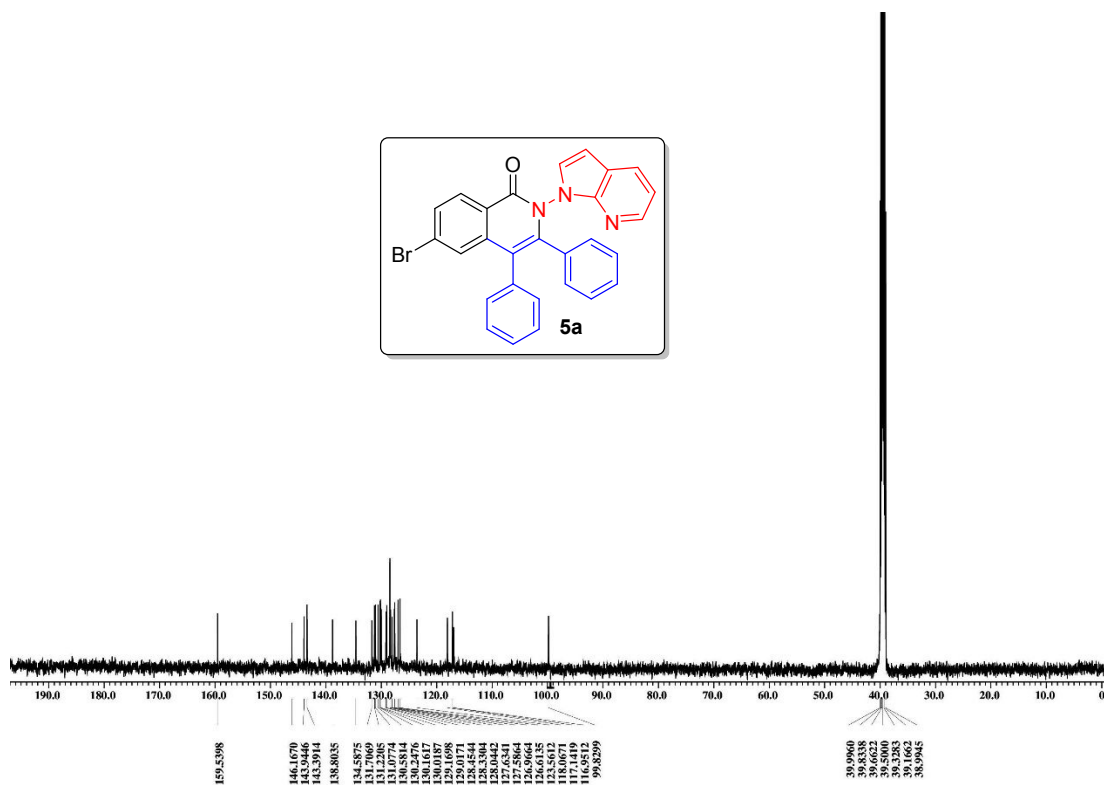


Fig. S56. ¹³C NMR spectrum of **5a** (125 MHz, DMSO)

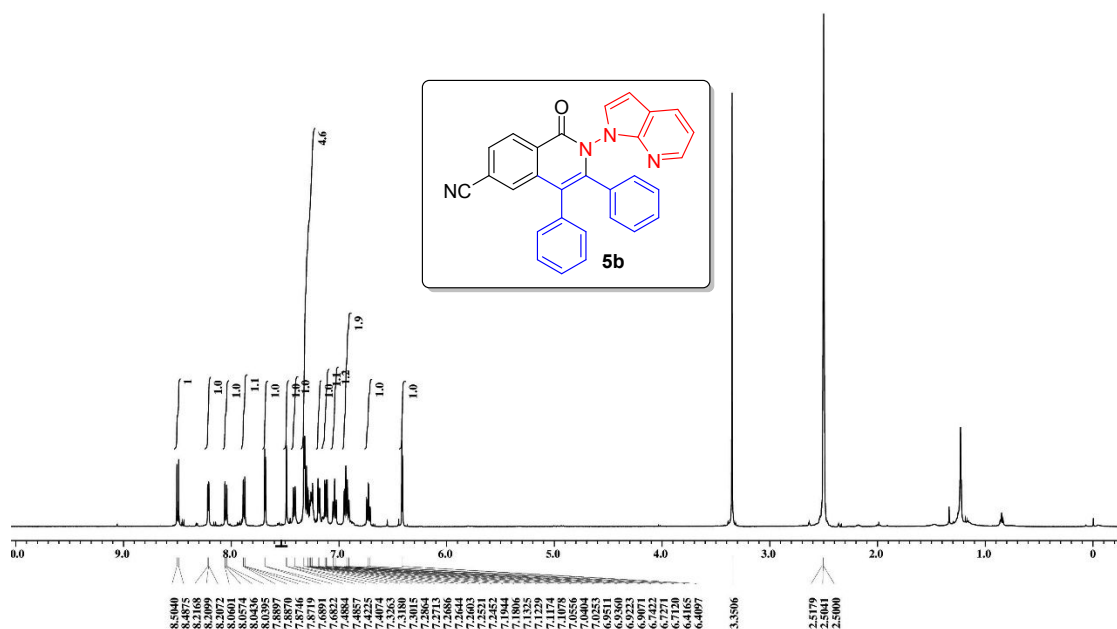


Fig. S57. ¹H NMR spectrum of **5b** (500 MHz, DMSO)

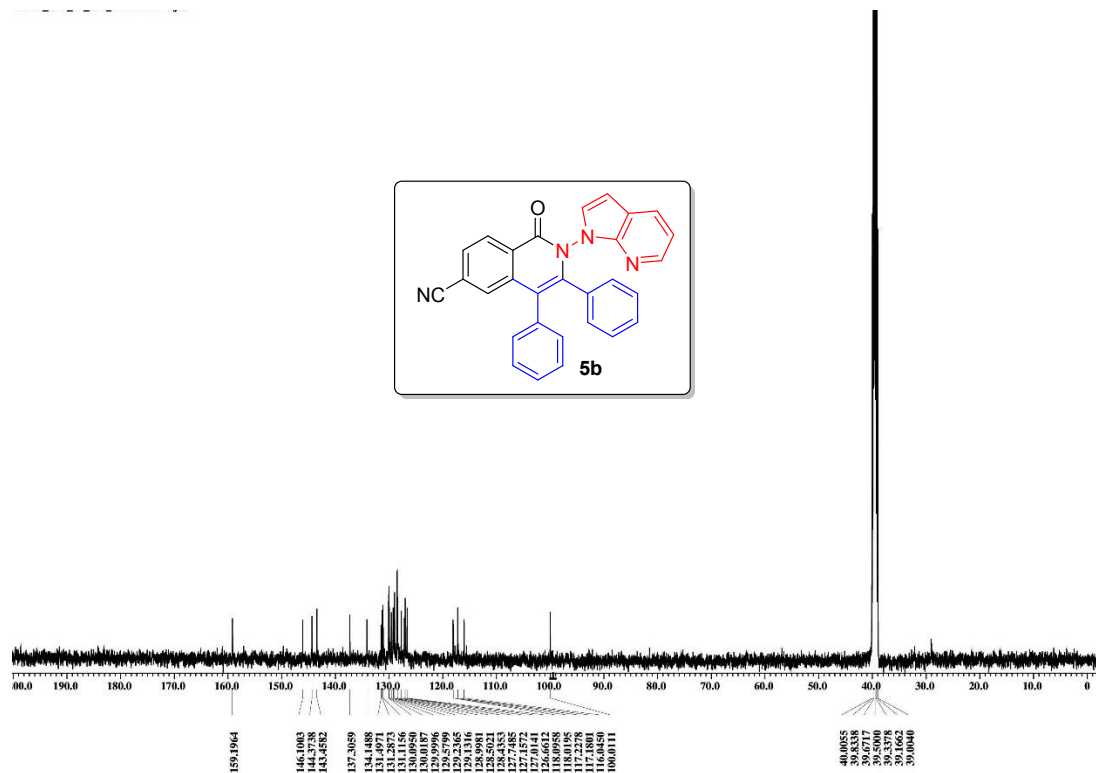


Fig. S58. ¹³C NMR spectrum of **5b** (125 MHz, DMSO)

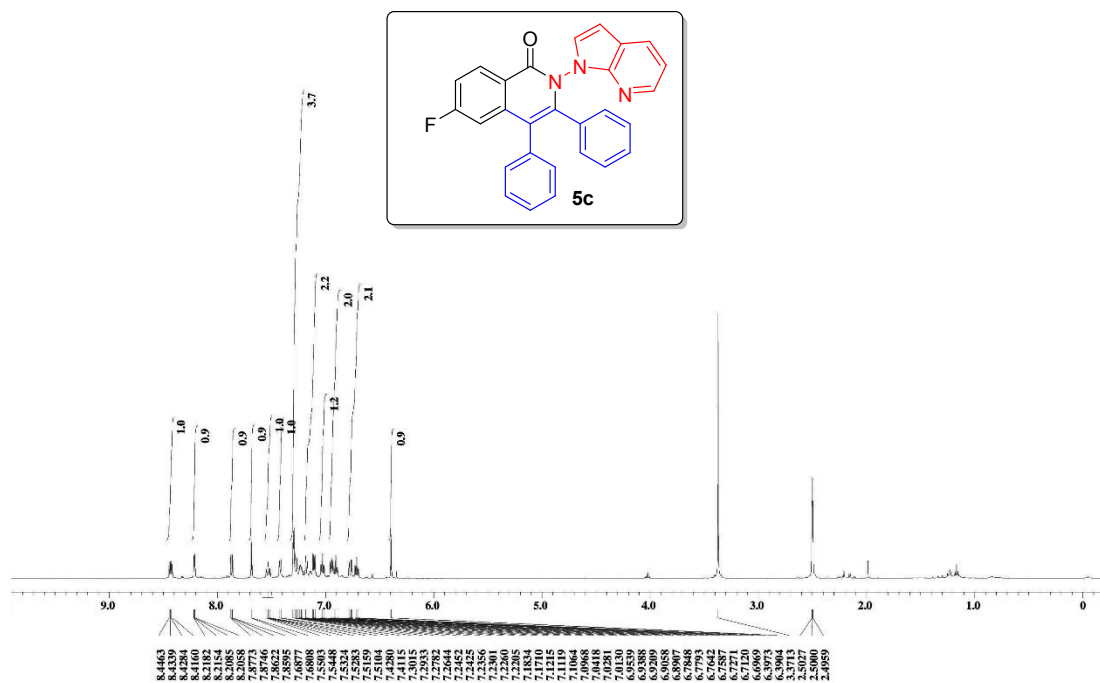


Fig. S59. ¹H NMR spectrum of **5c** (500 MHz, DMSO)

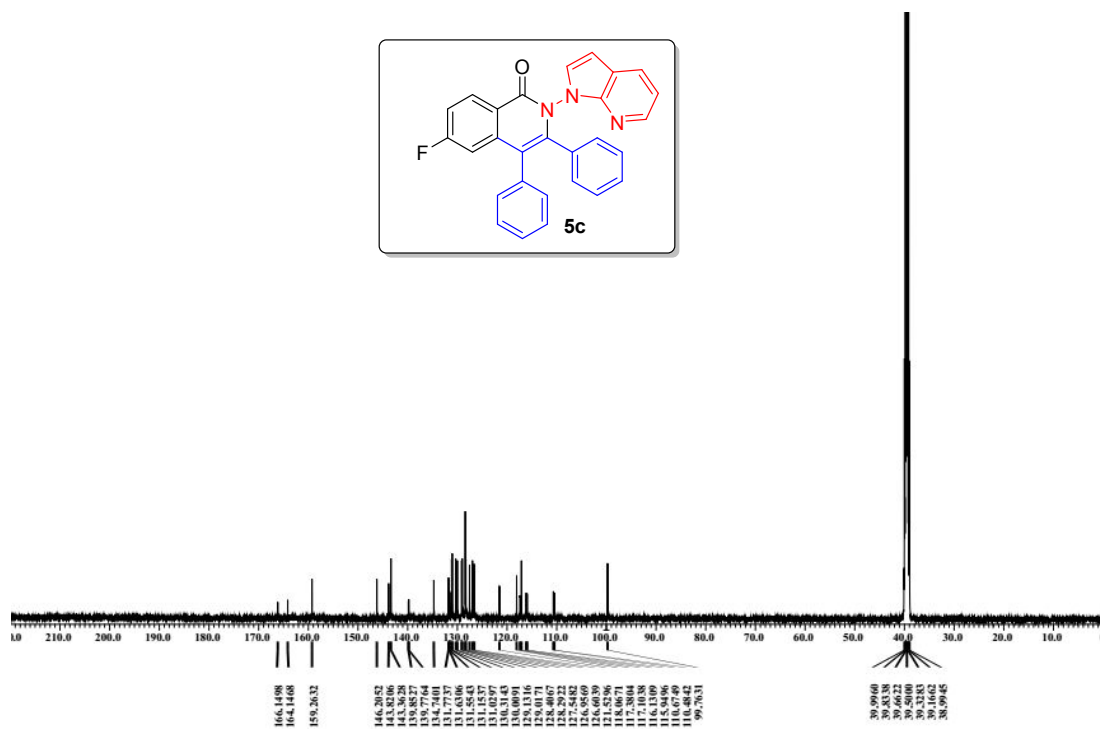


Fig. S60. ¹³C NMR spectrum of **5c** (125 MHz, DMSO)

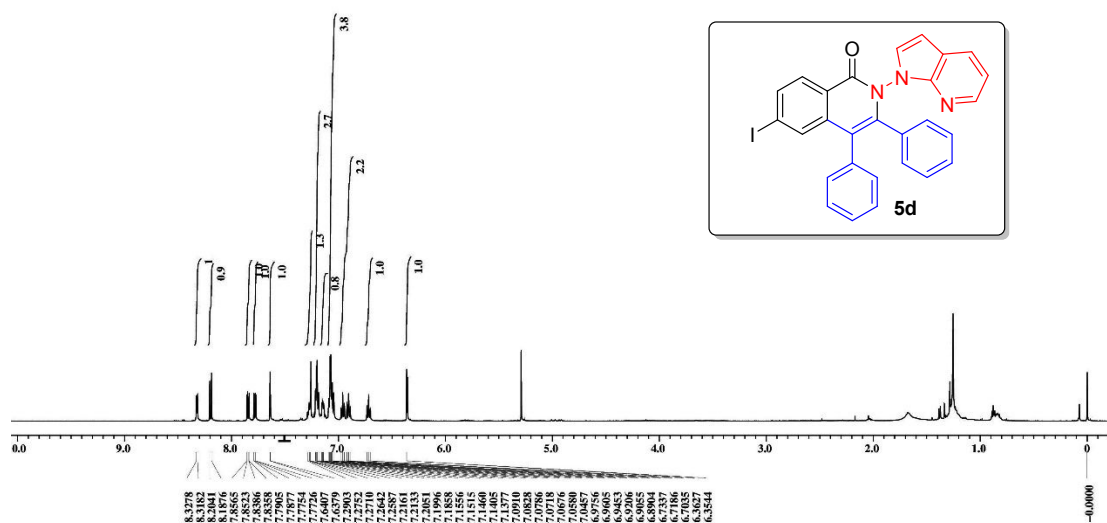


Fig. S61. ¹H NMR spectrum of **5d** (500 MHz, CDCl₃)

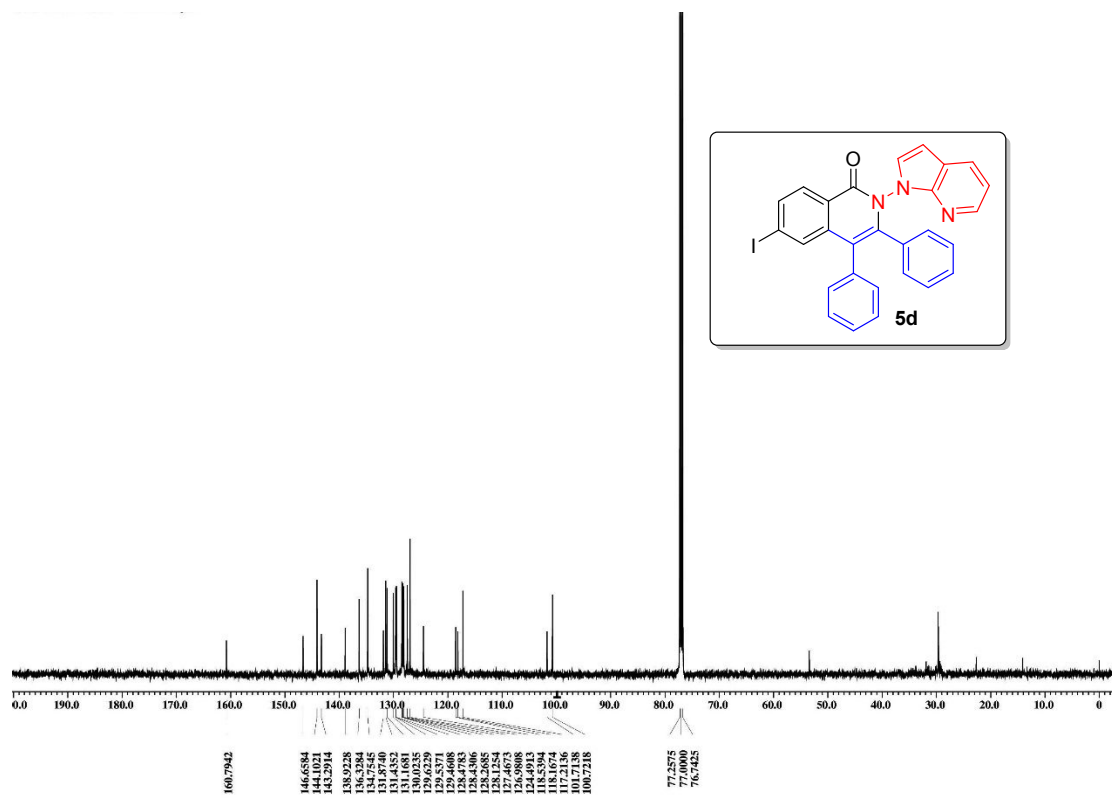


Fig. S62. ¹³C NMR spectrum of **5d** (125 MHz, CDCl₃)

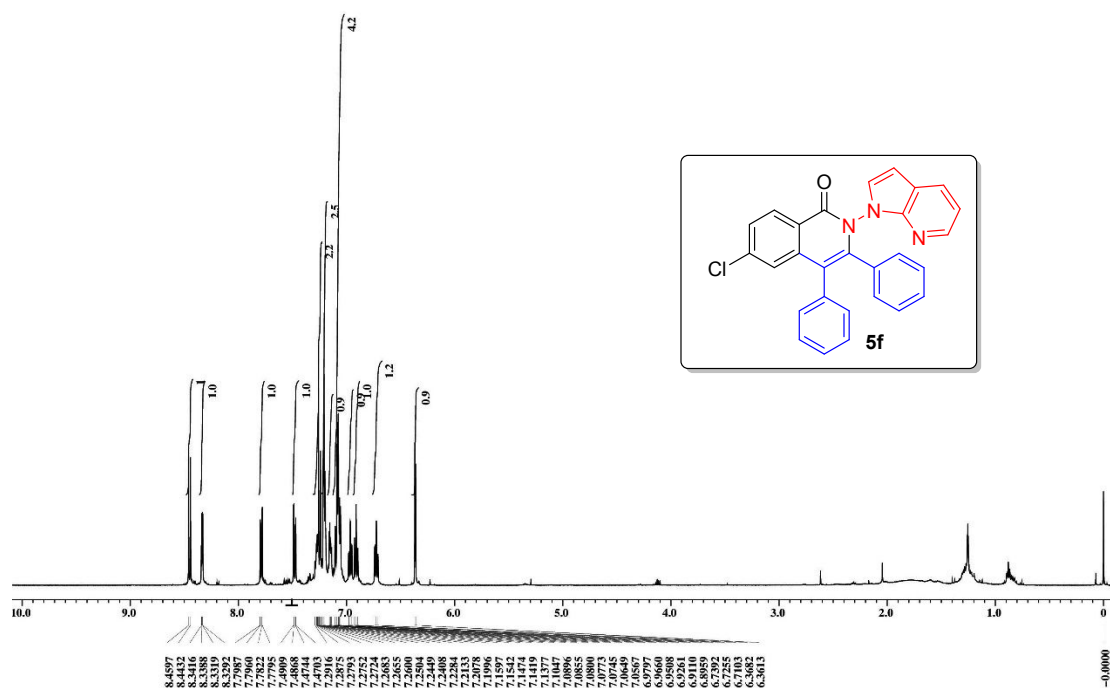


Fig. S65. ¹H NMR spectrum of **5f** (500 MHz, CDCl₃)

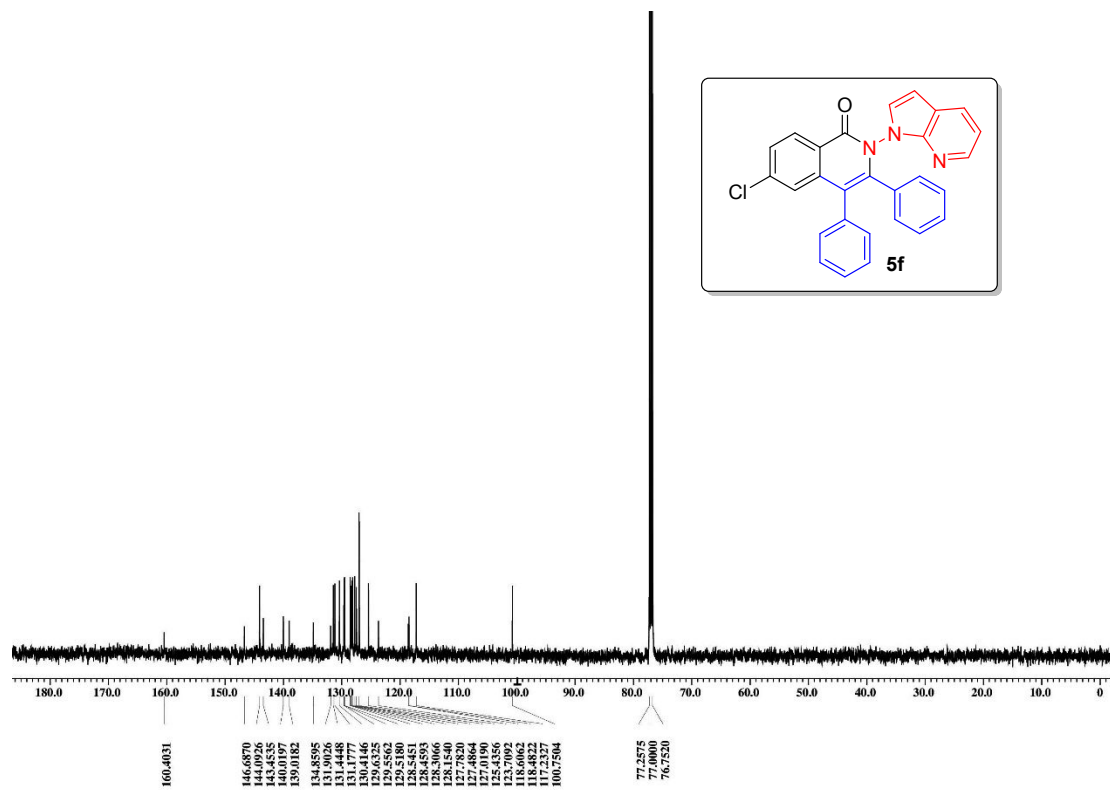


Fig. S66. ¹³C NMR spectrum of **5f** (125 MHz, CDCl₃)

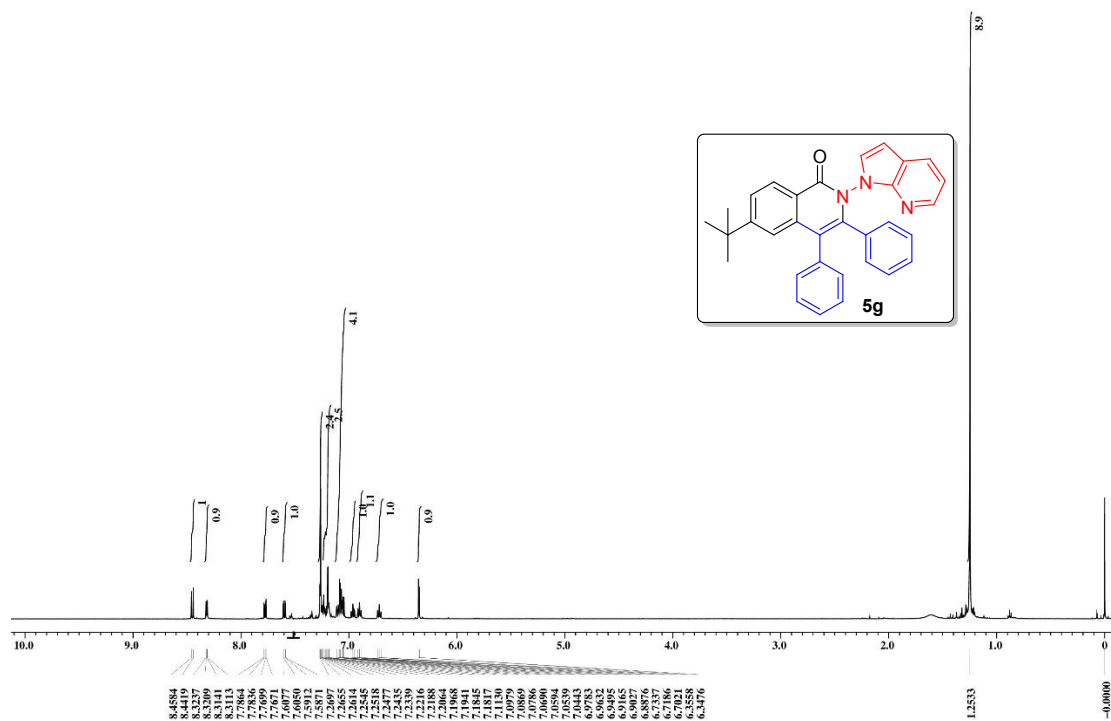


Fig. S67. ¹H NMR spectrum of **5g** (500 MHz, CDCl₃)

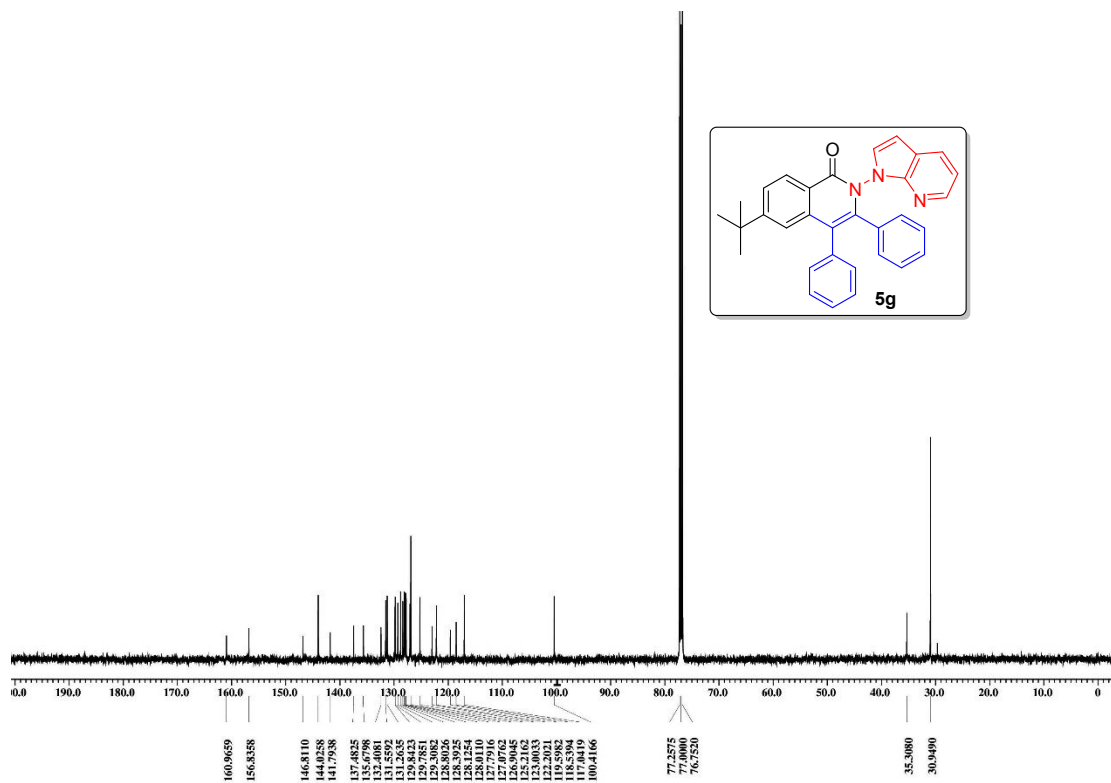


Fig. S68. ¹³C NMR spectrum of **5g** (125 MHz, CDCl₃)

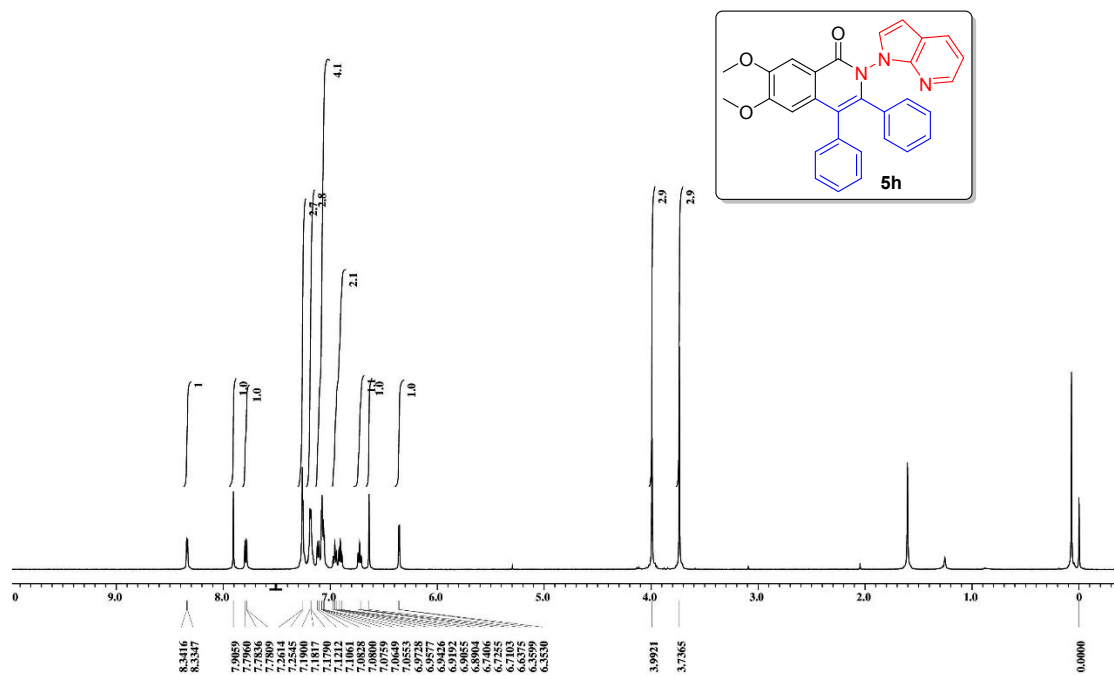


Fig. S69. ¹H NMR spectrum of **5h** (500 MHz, CDCl₃)

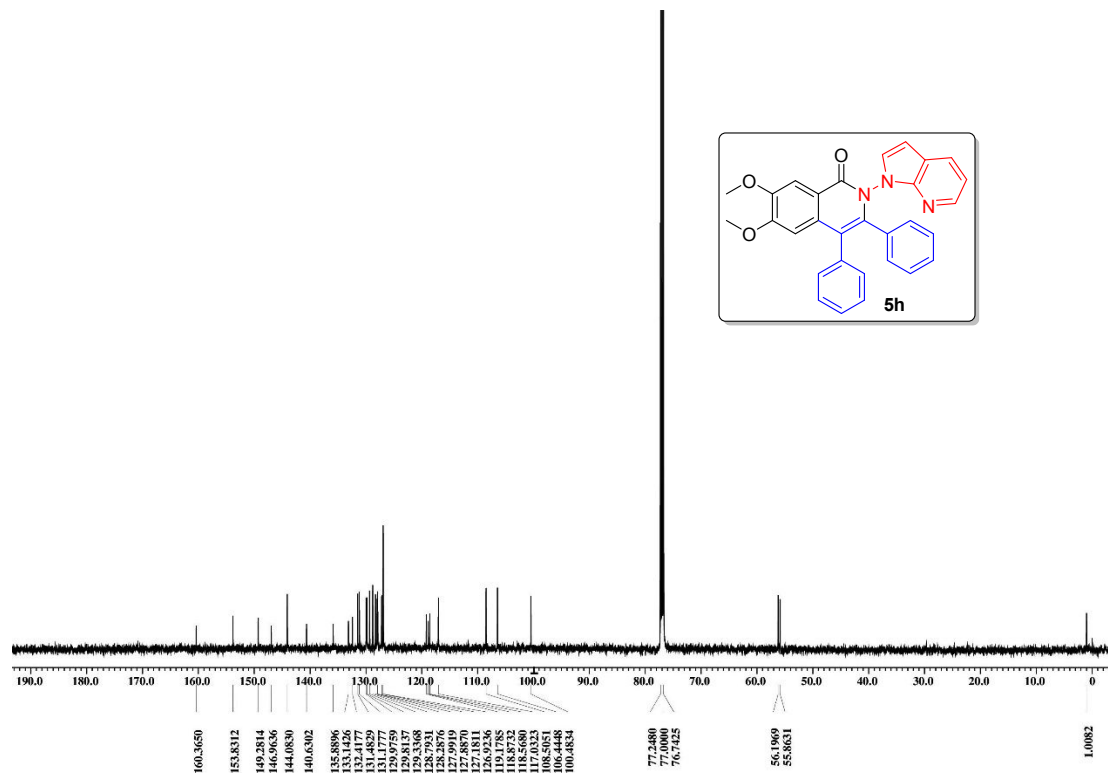
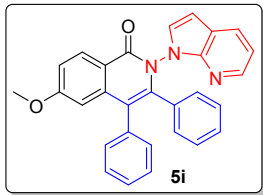


Fig. S70. ¹³C NMR spectrum of **5h** (125 MHz, CDCl₃)



Chemical structure of **5i** is shown in the inset. The structure is a benzimidazole derivative with a 4-methoxyphenyl group, a phenyl group, and a 1H-indol-1-yl group attached to the benzimidazole core.

¹H NMR spectrum (CDCl₃) of compound **5i**. The x-axis represents chemical shift in ppm, ranging from 0 to 10.0. The spectrum shows several peaks: a multiplet between 7.0 and 8.0 ppm, a sharp singlet at approximately 5.5 ppm, and a multiplet between 3.0 and 4.0 ppm. An inset shows the chemical structure of **5i**.

S43

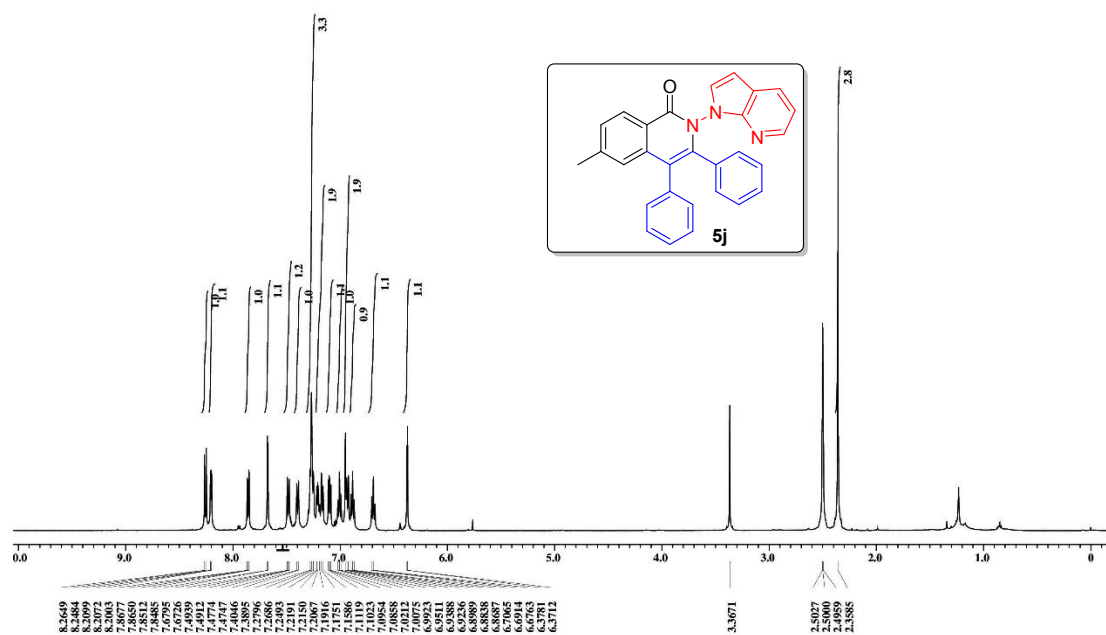


Fig. S73. ¹H NMR spectrum of **5j** (500 MHz, DMSO)

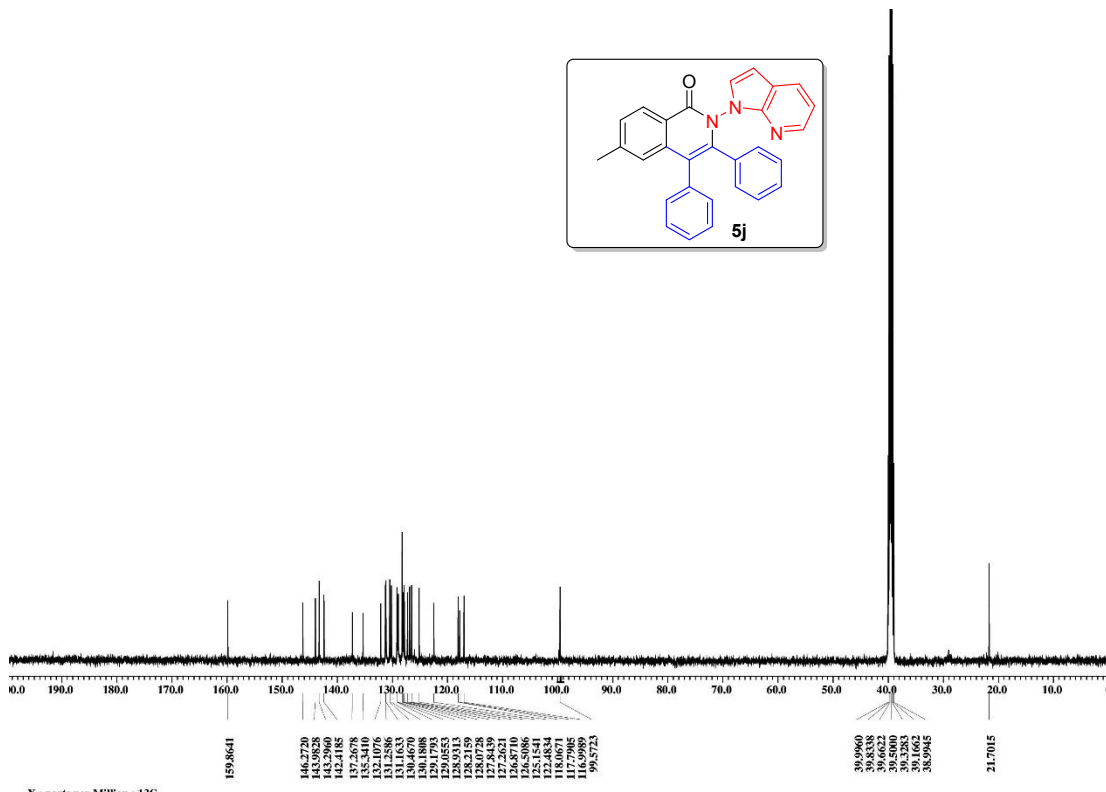


Fig. S74. ¹³C NMR spectrum of **5j** (125 MHz, DMSO)

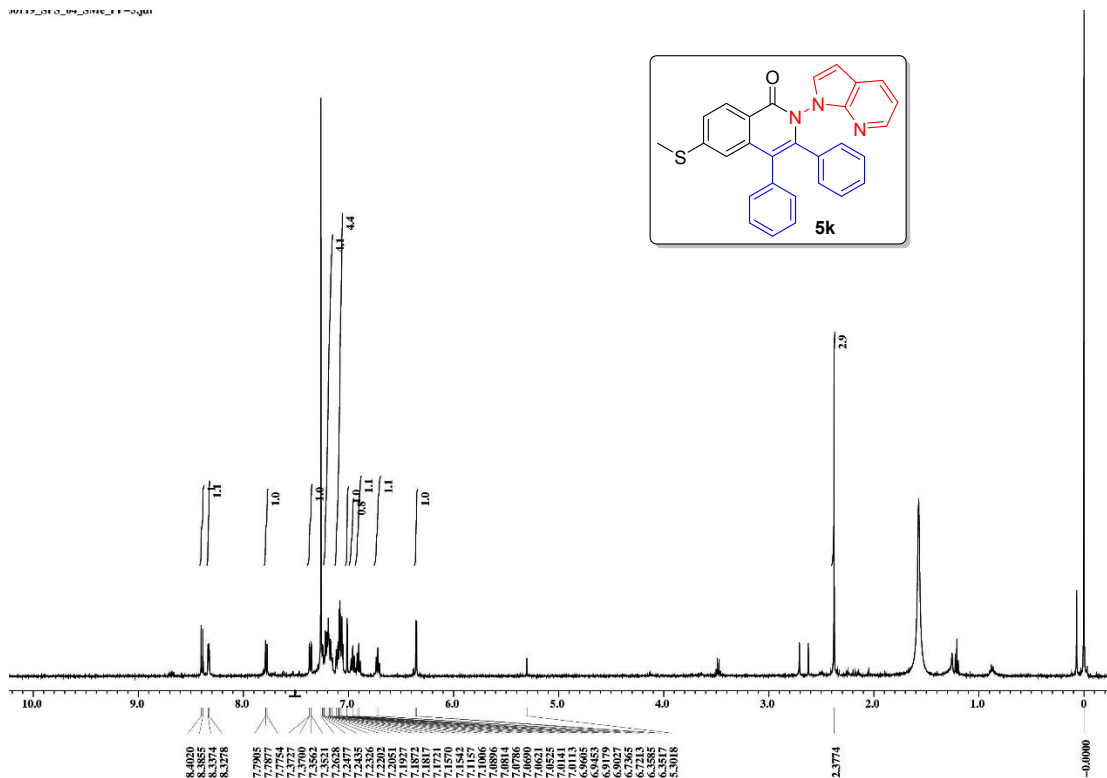


Fig. S75. ¹H NMR spectrum of **5k** (500 MHz, CDCl₃)

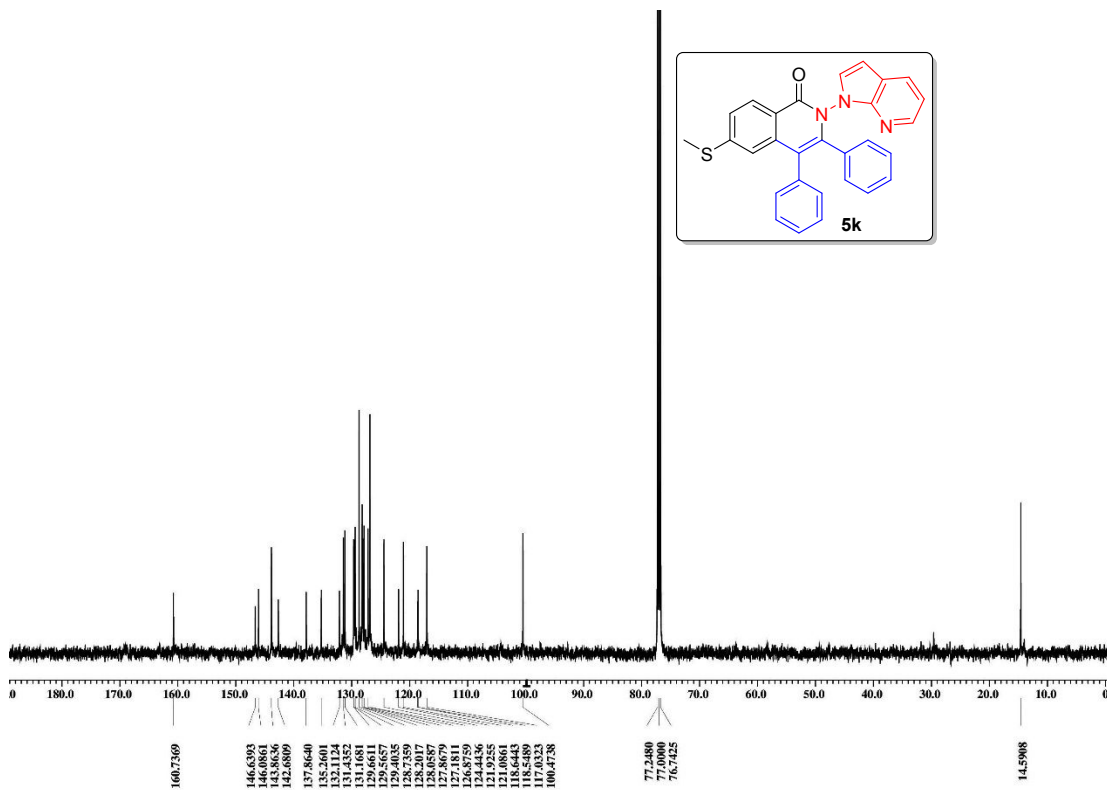


Fig. S76. ¹³C NMR spectrum of **5k** (125 MHz, CDCl₃)

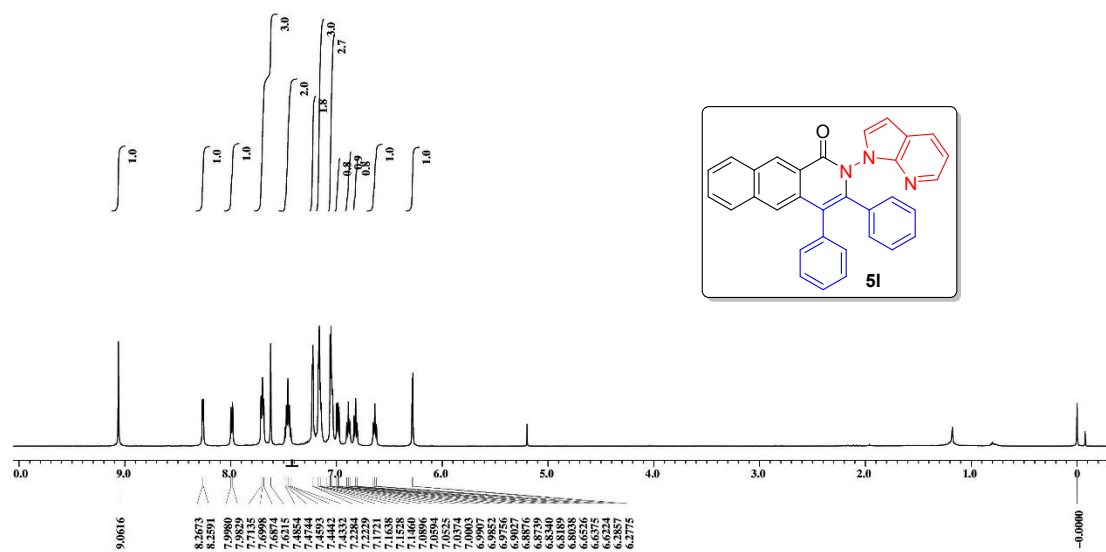


Fig. S77. ^1H NMR spectrum of **5I** (500 MHz, CDCl_3)

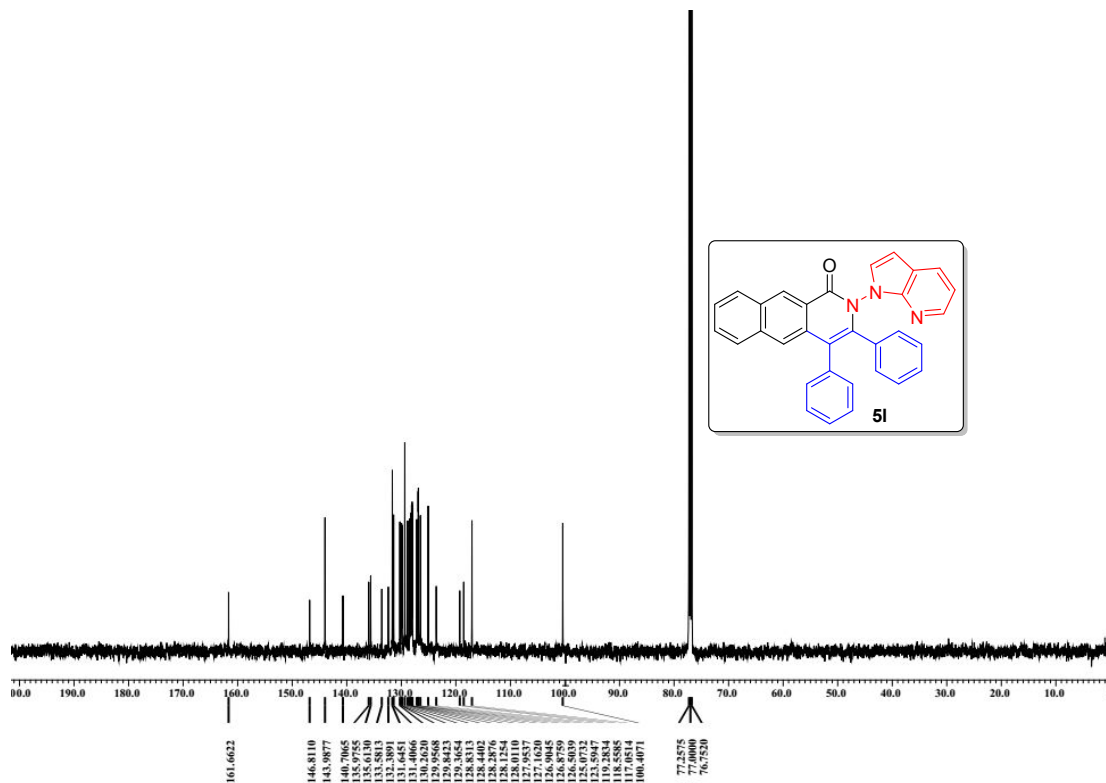


Fig. S78. ^{13}C NMR spectrum of **5I** (125 MHz, CDCl_3)

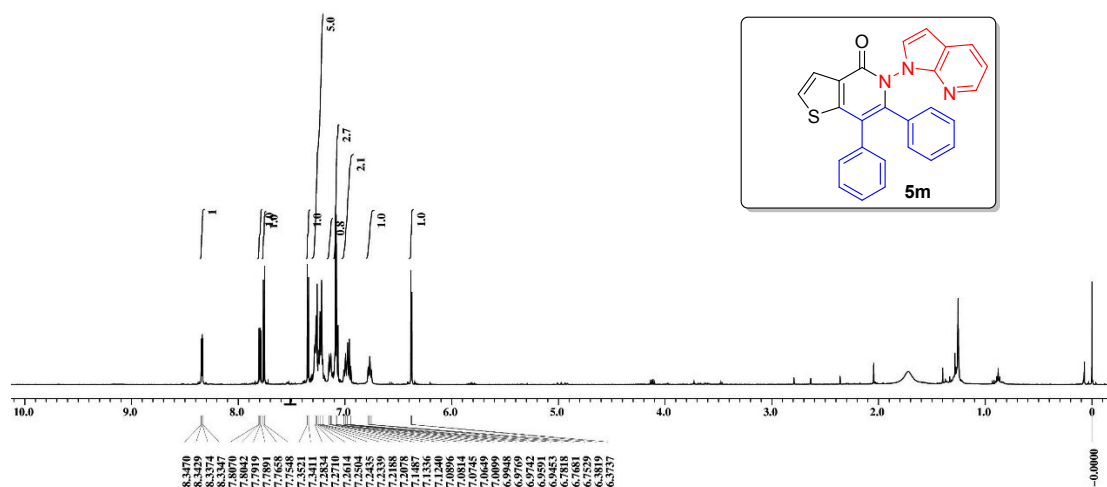


Fig. S79. ¹H NMR spectrum of **5m** (500 MHz, CDCl₃)

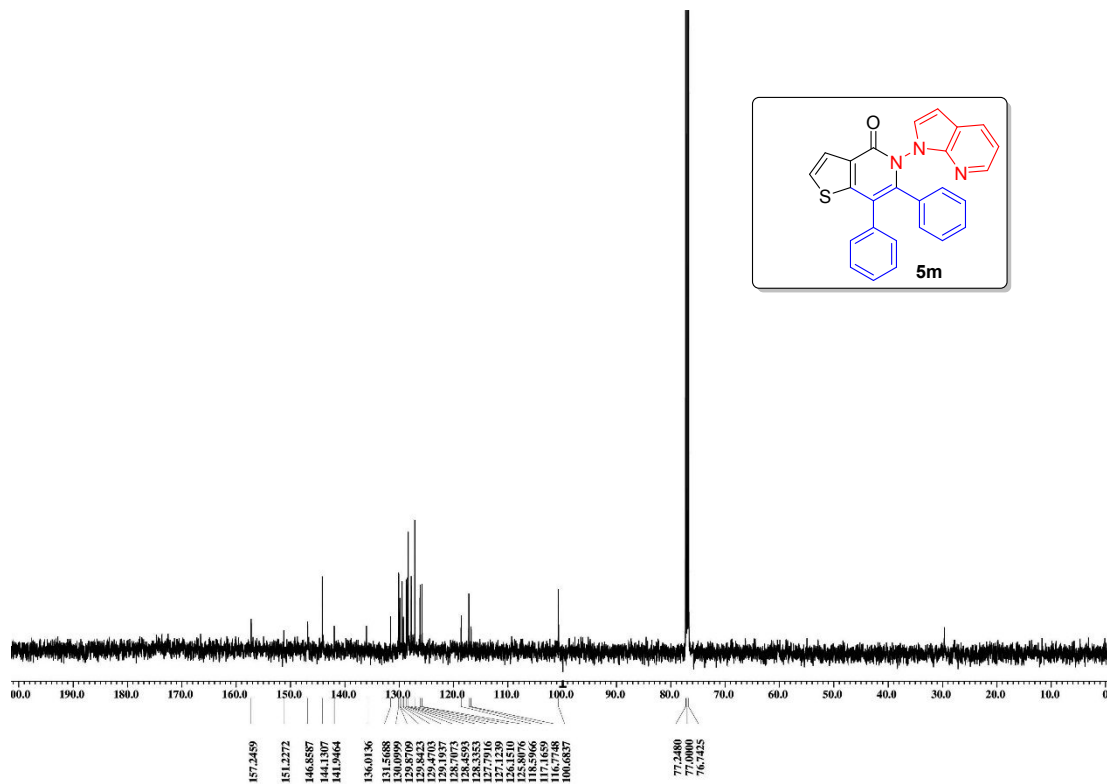


Fig. S80. ¹³C NMR spectrum of **5m** (125 MHz, CDCl₃)

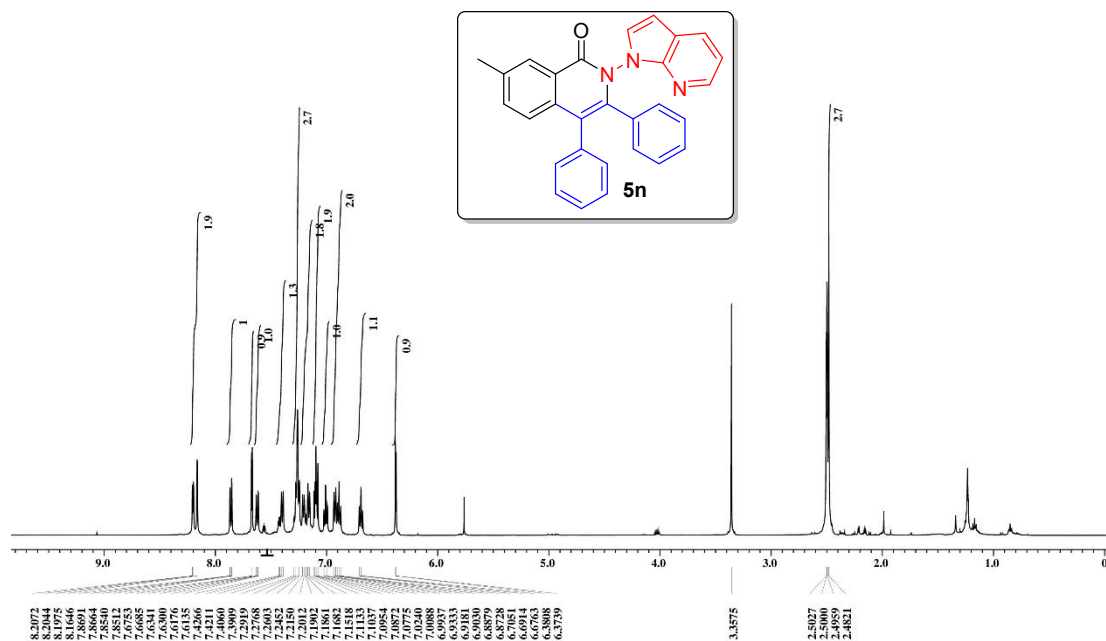


Fig. S81. ¹H NMR spectrum of **5n** (500 MHz, DMSO)

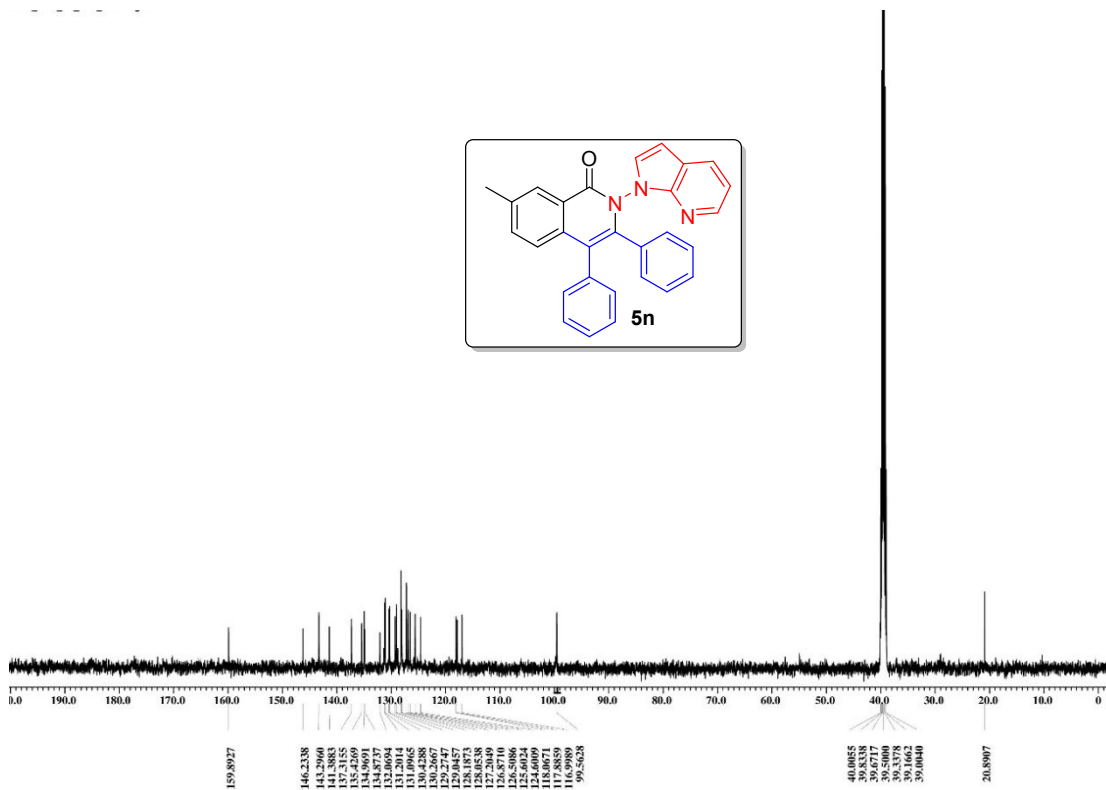


Fig. S82. ¹³C NMR spectrum of **5n** (125 MHz, DMSO)

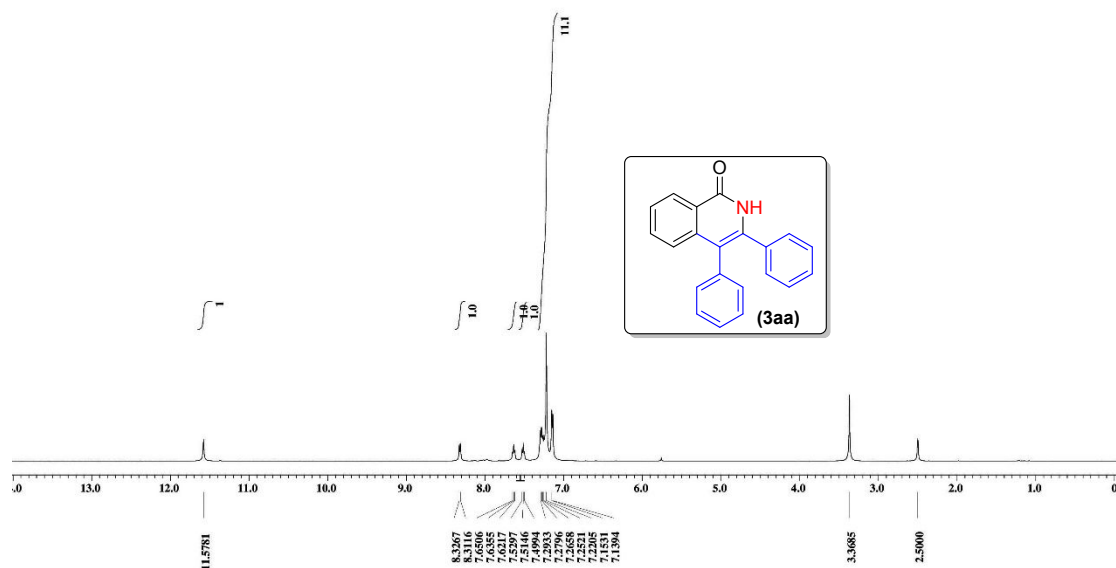


Fig. S83. ¹H NMR spectrum of **3aa** (500 MHz, DMSO)

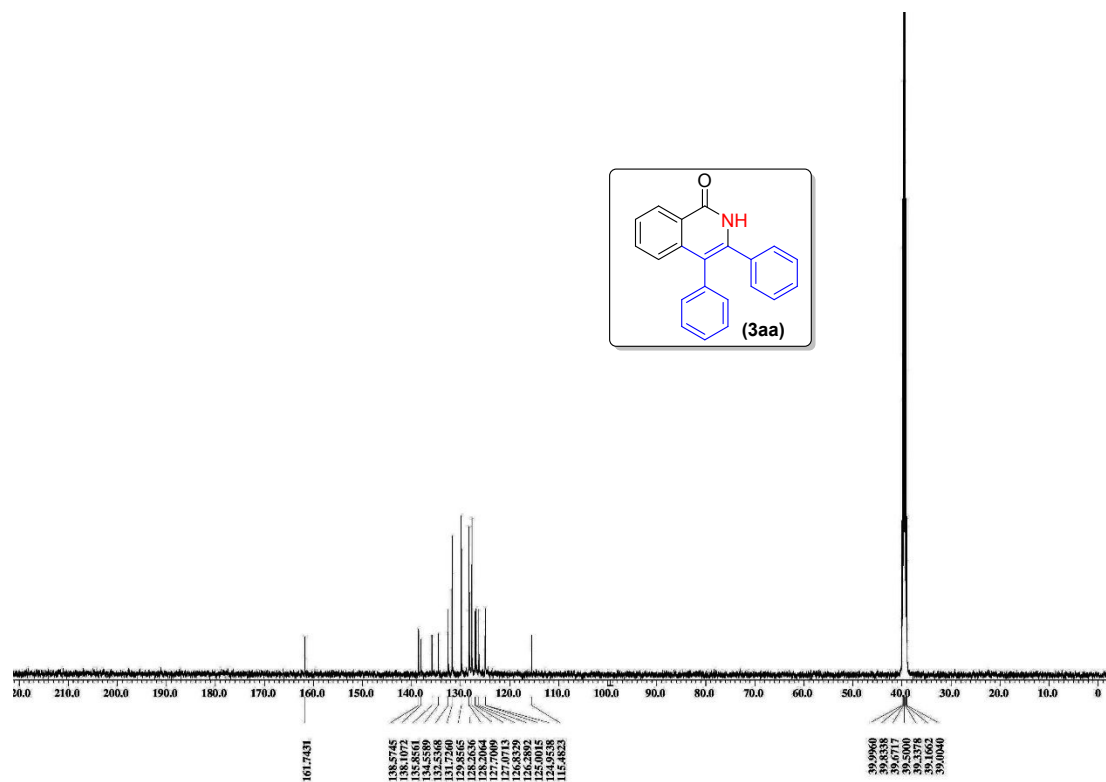


Fig. S84. ¹³C NMR spectrum of **3aa** (125 MHz, DMSO)

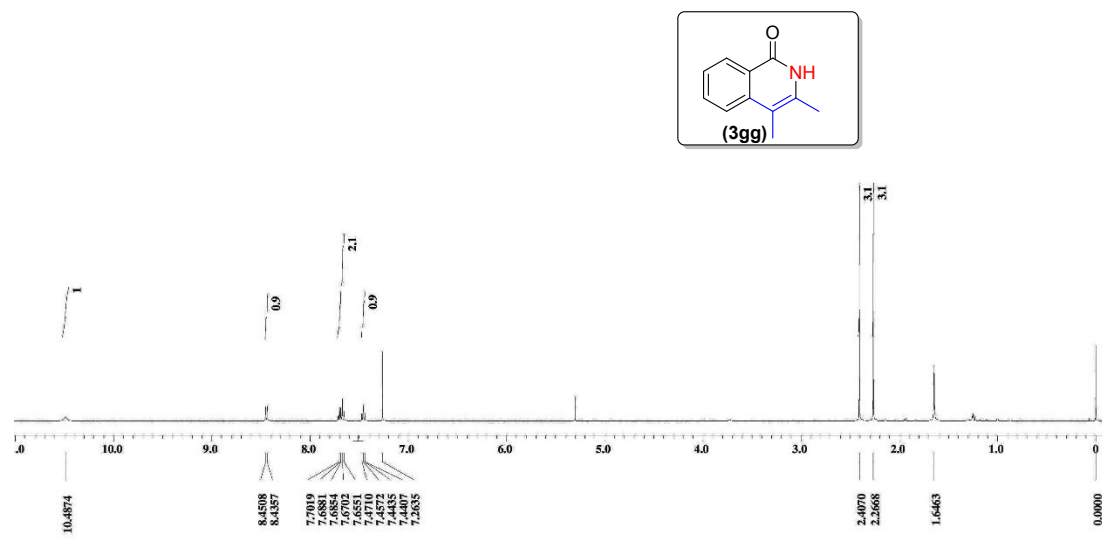


Fig. S85. ¹H NMR spectrum of **3gg** (500 MHz, CDCl₃)

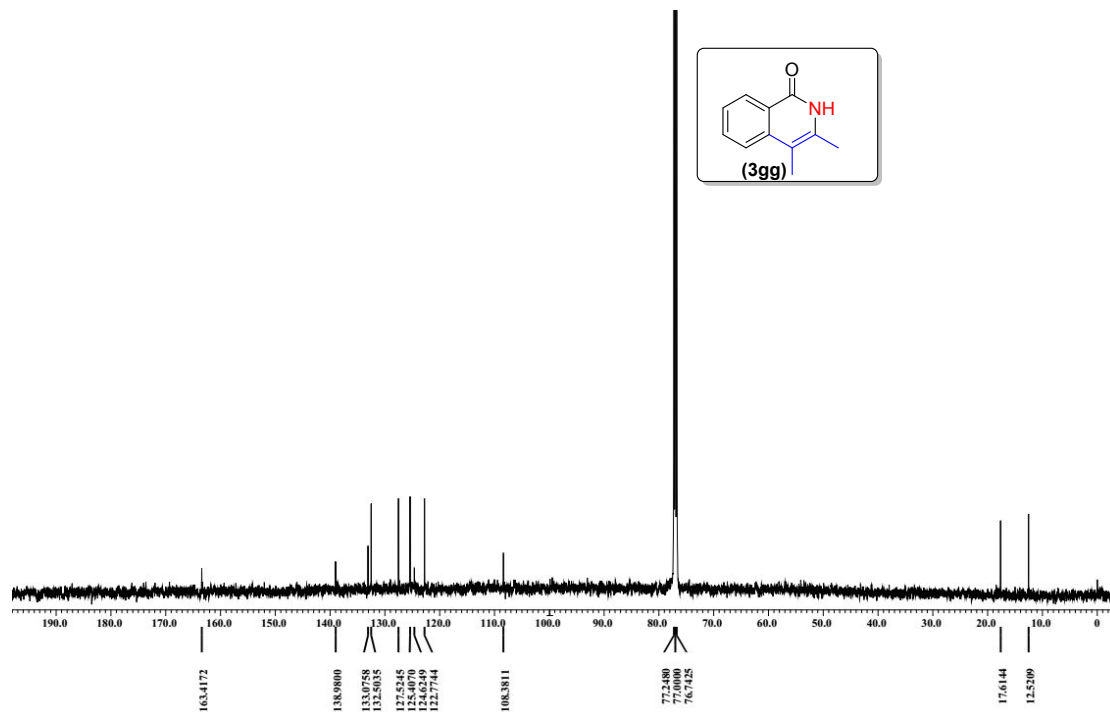


Fig. S86. ¹³C NMR spectrum of **3gg** (125 MHz, CDCl₃)

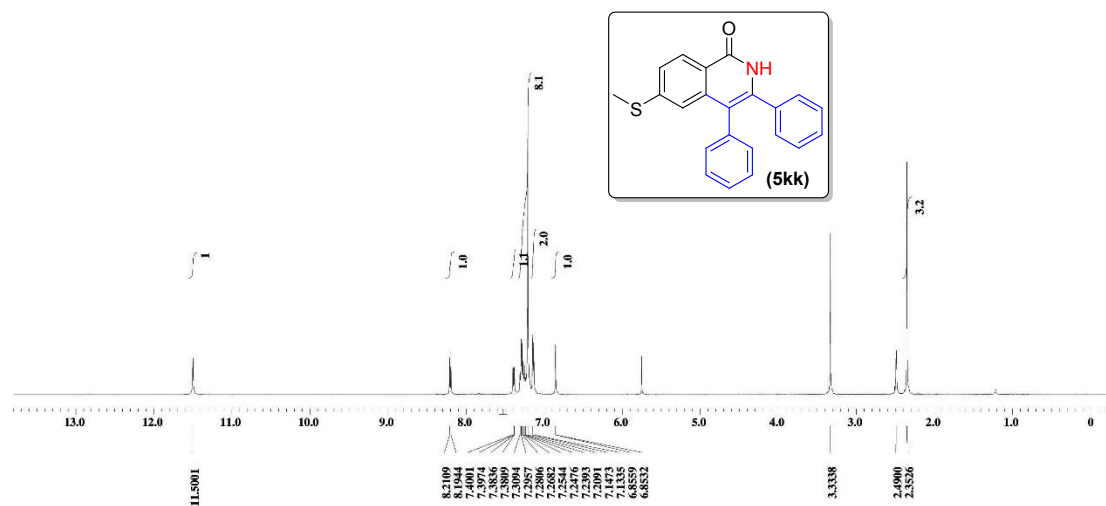


Fig. S87. ¹H NMR spectrum of **5kk** (500 MHz, DMSO)

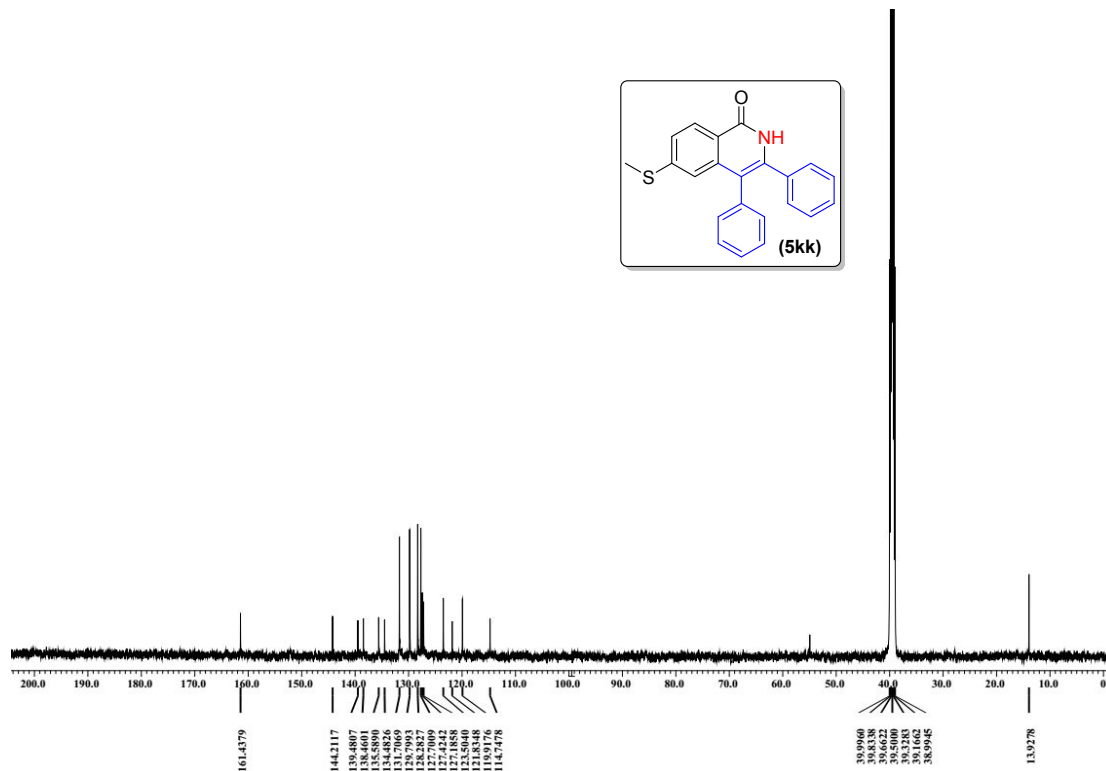


Fig. S88. ¹³C NMR spectrum of **5kk** (125 MHz, DMSO)

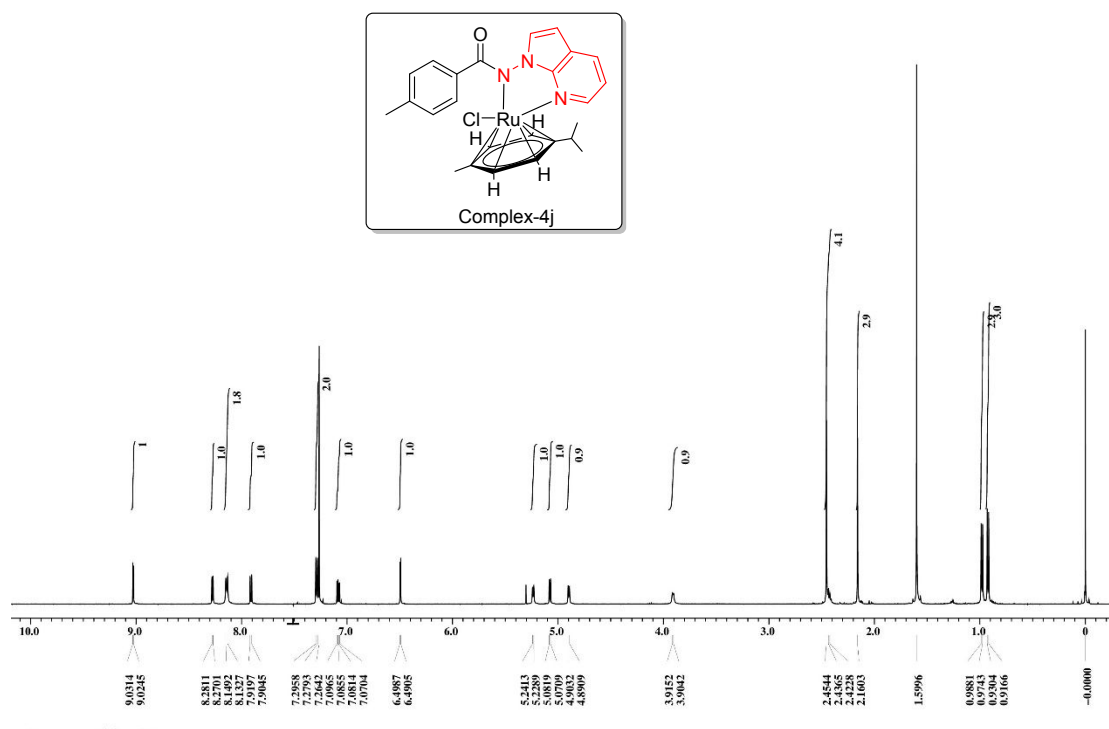


Fig. S89. ^1H NMR spectrum of **complex-4j** (500 MHz, CDCl_3)

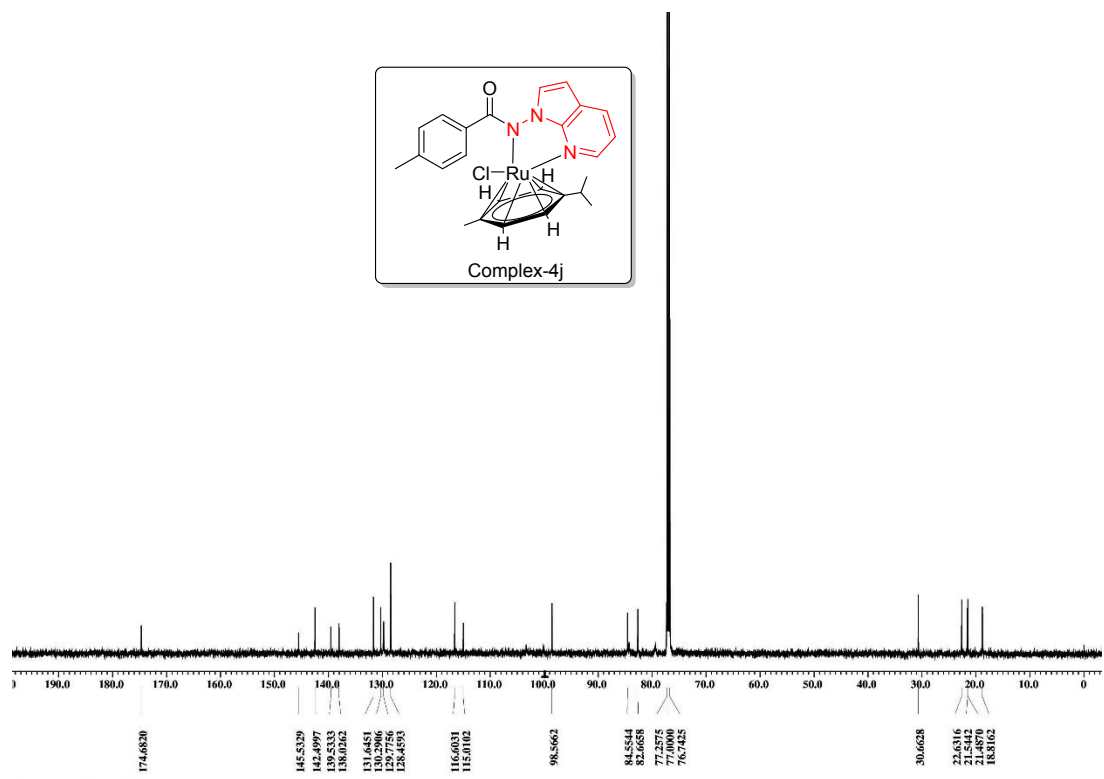
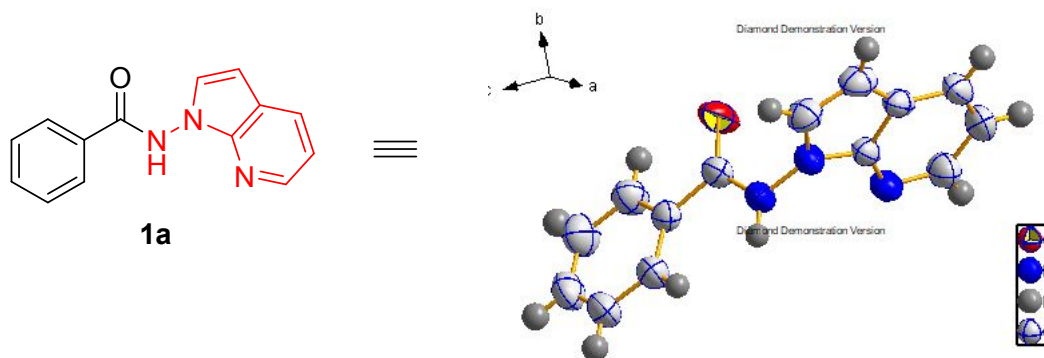


Fig. S90. ^{13}C NMR spectrum of **complex-4k** (125 MHz, CDCl_3)

4. X-ray Crystallographic Analysis of 1a, 3k & complex-4j’:

4a. crystal structure of 1a



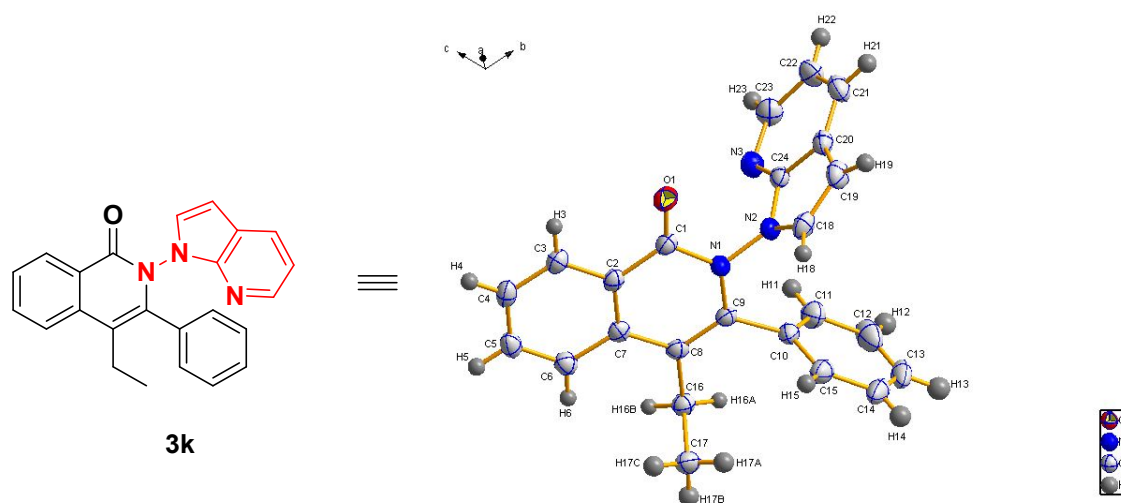
CCDC: 1868212

Table S2. Crystal data and structure refinement for

Identification code	
Empirical formula	C ₁₄ H ₁₁ N ₃ O
Formula weight	237.26
Temperature/K	293(2)
Crystal system	monoclinic
Space group	P2 ₁ /c
a/Å	11.8814(4)
b/Å	19.4570(4)
c/Å	11.4871(4)
α/°	90.00
β/°	115.705(4)
γ/°	90.00
Volume/Å ³	2392.74(11)
Z	8
ρ _{calc} /mg/mm ³	1.317
m/mm ⁻¹	0.698
F(000)	1136.0
Crystal size/mm ³	0.258 × 0.137 × 0.099
2θ range for data collection	8.26 to 133.66°
Index ranges	-13 ≤ h ≤ 14, -16 ≤ k ≤ 23, -13 ≤ l ≤ 10

Reflections collected	7757
Independent reflections	4196[R(int) = 0.0290]
Data/restraints/parameters	4196/0/325
Goodness-of-fit on F ²	1.035
Final R indexes [I>2σ (I)]	R ₁ = 0.0459, wR ₂ = 0.1248
Final R indexes [all data]	R ₁ = 0.0530, wR ₂ = 0.1335
Largest diff. peak/hole / e Å ⁻³	0.28/-0.28

4b. Crystal Structure of 3k



3k

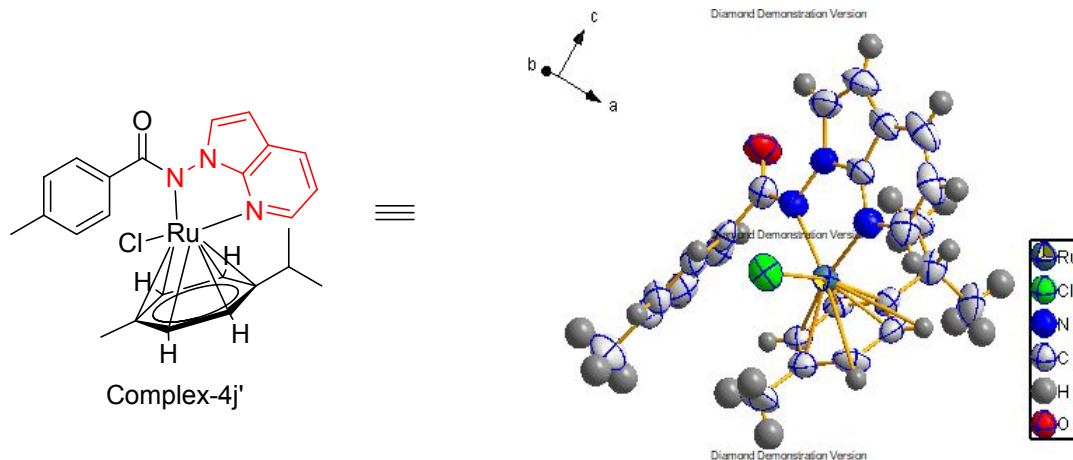
CCDC :1868597

Table S3. Crystal data and structure refinement for

Identification code	
Empirical formula	C ₂₄ H ₁₉ N ₃ O
Formula weight	365.42
Temperature/K	150.00(10)
Crystal system	triclinic
Space group	P-1
a/Å	9.2805(5)
b/Å	12.3962(7)
c/Å	16.8806(8)
α/°	107.261(5)
β/°	92.112(4)
γ/°	95.307(5)
Volume/Å ³	1842.38(17)

Z	4
ρ_{calc} mg/mm ³	1.314
m/mm ⁻¹	0.082
F(000)	878.0
Crystal size/mm ³	0.33 × 0.25 × 0.133
2 θ range for data collection	3.46 to 56.68°
Index ranges	-10 ≤ h ≤ 11, -16 ≤ k ≤ 15, -20 ≤ l ≤ 22
Reflections collected	11201
Independent reflections	7946[R(int) = 0.0186]
Data/restraints/parameters	7946/0/507
Goodness-of-fit on F ²	1.040
Final R indexes [I ≥ 2 σ (I)]	R ₁ = 0.0483, wR ₂ = 0.1073
Final R indexes [all data]	R ₁ = 0.0638, wR ₂ = 0.1187
Largest diff. peak/hole / e Å ⁻³	0.23/-0.21

4c. Crystal Structure of complex-4j'



CCDC: 1913752

Table S4. Crystal data and structure refinement for SPS_04_348.

Identification code	SPS_04_348
Empirical formula	C ₂₅ H ₂₆ ClN ₃ ORu
Formula weight	521.01

Temperature/K	293(2)
Crystal system	orthorhombic
Space group	Pca2 ₁
a/Å	9.1000(4)
b/Å	14.3544(4)
c/Å	17.5769(7)
$\alpha/^\circ$	90.00
$\beta/^\circ$	90.00
$\gamma/^\circ$	90.00
Volume/Å ³	2295.99(15)
Z	4
$\rho_{\text{calc}}/\text{g/cm}^3$	1.507
μ/mm^{-1}	6.764
F(000)	1064.0
Crystal size/mm ³	0.404 × 0.241 × 0.124
Radiation	CuK α (λ = 1.54184)
2 θ range for data collection/ $^\circ$	6.16 to 133.42
Index ranges	-10 ≤ h ≤ 10, -14 ≤ k ≤ 17, -20 ≤ l ≤ 20
Reflections collected	4575
Independent reflections	2616 [R_{int} = 0.0378, R_{sigma} = 0.0446]
Data/restraints/parameters	2616/1/284
Goodness-of-fit on F ²	1.064
Final R indexes [$I \geq 2\sigma(I)$]	R_1 = 0.0425, wR_2 = 0.1134
Final R indexes [all data]	R_1 = 0.0489, wR_2 = 0.1236
Largest diff. peak/hole / e Å ⁻³	0.40/-0.73
Flack parameter	-0.05(2)

Using Olex²¹, the structure was solved with the ShelXS² structure solution program using Direct Methods and refined with the ShelXL² refinement package using Least Squares minimisation.

1. Dolomanov, O. V; Bourhis, L. J.; Gildea, R. J.; Howard, J. A. K.; Puschmann, H. *{\it OLEX2}*: A Complete Structure Solution, Refinement and Analysis Program. *J. Appl. Crystallogr.* **2009**, 42 (2), 339–341.
2. Sheldrick, G. M. A Short History of *{\it SHELX}*. *Acta Crystallogr. Sect. A* **2008**, 64 (1), 112–122.