

# Analytical Chemistry

## Supporting Information

### **Photothermal Microfluidic Sensing Platform Using Near-Infrared Laser-Driven Multiplexed Dual-Mode Visual Quantitative Readout**

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## Experimental Section

**Materials and Instruments.** Acrylamide (AcAm) was the product of Aladdin Bio-Chem Technology Co., Ltd (Shanghai, China). N,N,N',N'-tetramethylethylenediamine (TEMED) was obtained from Aladdin Bio-Chem Technology Co., Ltd (Shanghai, China). Red food dye was obtained from AmeriColor Corp (CA, USA). Bisacrylamide and ammoniumpersulfate (APS) were acquired from Sigma-Aldrich (MO, USA). Dextran 70 was purchased from Aladdin Bio-Chem Technology Co., Ltd (Shanghai, China). Whatman chromatography papers (Grade No. 1) were the products of GE Healthcare Worldwide (Shanghai, China). Silver nitrate was the product of Shandong XiYa Chemical Industry Co., Ltd (Shandong, China). Ultrapure water (18.2 M $\Omega$  cm) was collected from a Milli-Q purification system (Bedford, MA). UV-vis absorption spectroscopic characterization was performed by using a Shimadzu UV-1800 spectrophotometer (Shimadzu, Kyoto, Japan). Photographs of  $\mu$ PADs were taken with a Canon EOS 600D camera (Canon Inc., Tokyo, Japan). Fourier transformed infrared (FT-IR) spectroscopic characterization of the hydrogels was carried out on a Nicolet 380 FT-IR spectrometer (Thermo Scientific, USA). Scanning electron microscopic (SEM) characterization of the hydrogels was performed by utilizing a FEI Quanta 250 scanning electron microscope (FEI Co., OR, USA). Thermal images were taken by using a FLIR C2 thermal camera (FLIR Systems Inc., CA, USA). Unless otherwise stated, all other chemicals were of analytical grade and were used as received.

**Synthesis of Blank and Photothermally Responsive Hydrogels.** The blank NIPAM-AcAm hydrogels were synthesized *via* the radical polymerization method according to the literature.<sup>1</sup> In brief, NIPAM (299 mg), AcAm (21 mg), bisarylamide (8 mg) and dextran 70 (40 mg) were dissolved in 2.0 ml of ultrapure water. After the addition of 100  $\mu$ L of APS solution (65 mM), the aqueous mixture was thoroughly purged with N<sub>2</sub> for 5.0 min. Afterwards, 24  $\mu$ L of TEMED was added into the above solution to initiate the polymerization reaction, followed by rapid filling of the aqueous mixture into plastic syringes (1.0 mL). The syringes were then stored at room temperature for 12 h to perform the polymerization reaction. To remove unreacted chemicals in the hydrogels, the as-obtained hydrogel rods were first incubated in water bath at 55 °C for 20 min to induce their complete collapse. The collapsed hydrogels were then stored in water at room temperature to achieve the swelling to their initial shape. The above washing procedure was repeated 3 times.

The photothermally responsive hydrogels (i.e. PB-NIPAM-AcAm) were prepared through an in-situ synthesis route. Briefly, the blank NIPAM-AcAm hydrogels were incubated in water bath at 55 °C for 20 min to induce their structural collapse, followed by successive storage of the hydrogels in aqueous solutions (25 mM citric acid) of FeCl<sub>3</sub> and potassium ferrocyanide for 3.0 hours at room temperature, respectively. Afterwards, the hydrogels were thoroughly washed with deionized water. The collapsed hydrogels were stored overnight in aqueous dye solutions at room temperature to load dye solutions in the hydrogel matrix. Unless otherwise stated, the

concentration of  $\text{FeCl}_3$  and potassium ferrocyanide used for synthesis of the composite hydrogel was 1.0 mM.

Before the loading of the hydrogels on  $\mu\text{PADs}$ , the as-obtained hydrogel rods of 6.0 mm in diameter were uniformly sliced into circular pieces of 2.0 mm in thickness, followed by drying of the hydrogel pieces on filter papers.

**Fabrication of Photothermal  $\mu\text{PAD}$ .** Patterns of  $\mu\text{PADs}$  were designed with the CorelDraw X4 software. The disk-like  $\mu\text{PAD}$  consisted of a central circular irradiation zone and multiple straight channels accessible to the irradiation zone. The central irradiation zone (2.0 cm in diameter) was composed of six circular inlets of 6.0 mm in diameter. Six straight channels of 2.5 mm in width and 5.0 cm in length were individually connected to corresponding inlets. The irradiation zone and channels were designed for multiplexed thermal image- and distance-based visual quantitative readout, respectively. Microfluidic hydrophobic barriers were printed on chromatography papers by using a wax printer (Xerox Phaser 8560, Fuji Xerox CO., LTD., Japan). The wax-printed chromatograph papers were heated in an oven for 3.0 min at 150 °C, followed by sealing of the back of the device with adhesive tapes.

**On-Chip Distance- and Thermal Image-Based Visual Quantitative Readout.**

Thermal images of the  $\mu\text{PADs}$  were taken before and after the laser irradiation, which were quantitatively analyzed with the FLIR Tools software for thermal image-based readout. After 10 minutes upon removal of the laser irradiation, photographs of the

$\mu$ PADs were taken to measure the travelling distance of dye solutions released in the channels.

**Determination of Silver Ions in Real Environmental Water Samples.** Aqueous solutions of  $\text{Ag}^+$  were prepared with Britton-Robinson (B-R) buffer solutions (pH=7.2). The hydrogel pieces were incubated in different concentrations of standard  $\text{Ag}^+$  solutions for 1.5 hours at room temperature, followed by loading of dye solutions into the hydrogels. The photothermal sensing performance of the hydrogels was then quantitatively analyzed using the dual-mode readout method on the  $\mu$ PAD. To evaluate the selectivity of the photothermal  $\mu$ PAD, the same concentration (i.e. 30  $\mu\text{M}$ ) of other metal ions, including  $\text{Mg}^{2+}$ ,  $\text{Cu}^{2+}$ ,  $\text{Fe}^{3+}$ ,  $\text{Zn}^{2+}$ ,  $\text{Ca}^{2+}$ ,  $\text{Ba}^{2+}$  and  $\text{Fe}^{2+}$ , were determined in parallel to  $\text{Ag}^+$ . Real water samples collected from a local lake were used to validate the practical analytical application of the  $\mu$ PAD. 1.5 mL of the water samples (5-fold diluted with B-R buffer solutions) were spiked with 10  $\mu\text{L}$  of different concentrations of standard  $\text{Ag}^+$  solutions to prepare water samples with final  $\text{Ag}^+$  concentrations of 5.0, 10 and 20  $\mu\text{M}$ , respectively. The concentrations of  $\text{Ag}^+$  were determined on the  $\mu$ PAD to calculate the analytical recoveries using the dual-mode readout method.

## Results and Discussion

**FTIR Characterization of Hydrogels.** The freeze-dried hydrogels were ground into powders and characterized by Fourier transform infrared spectroscopy (FTIR). As can

be seen in Figure S1, the composite hydrogel exhibited an obvious stretching band at  $2088\text{ cm}^{-1}$  that corresponded well with the CN stretching band in the  $[\text{Fe}^{\text{II}}\text{--CN--Fe}^{\text{III}}]$  structures in Prussian blue (PB),<sup>2, 3</sup> whereas no apparent stretching band was observed from the blank hydrogel in this region. The result confirmed the successful synthesis of PB in the composite hydrogel.

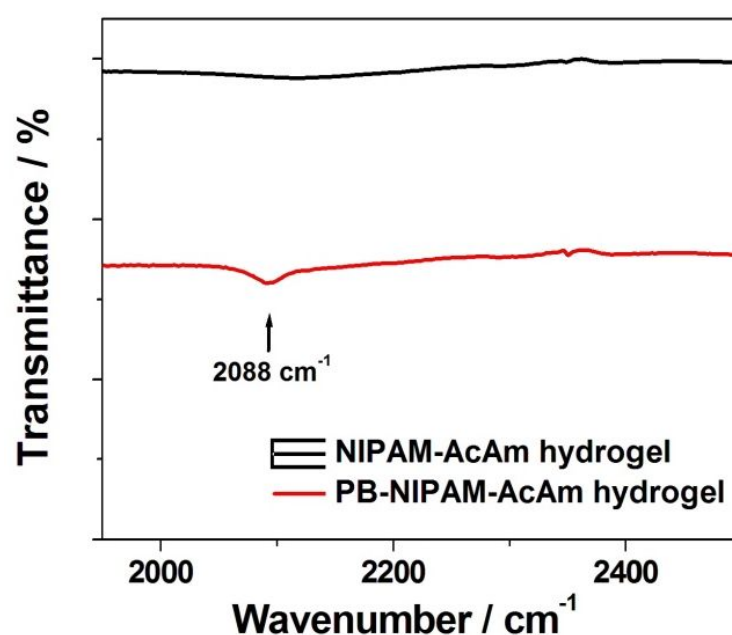


Figure S1. FTIR characterization of hydrogels. FTIR spectra of the blank (i.e., NIPAM-AcAm) and composite (i.e., PB-NIPAM-AcAm) hydrogels.

**Optical Absorption Properties of PB and Red Dye in NIR Region.** The optical absorption properties of PB nanoparticles (NPs) and red dye in the NIR region were studied by UV-vis absorption spectroscopy as shown in Figure S2. The aqueous dispersion of PB NPs showed strong optical absorption in the range of 500-1000 nm with the absorption peak at 710 nm, which can be ascribed to the charge transfer transition between Fe (II) and Fe (III) in PB NPs.<sup>3, 4</sup> In contrast, the red dye solution

showed no apparent optical absorption in the range of 600-1000 nm. Hence, under irradiation of an 808 nm laser, only PB served as the photothermally responsive element in the composite hydrogel without any photothermal interference from the red dye.

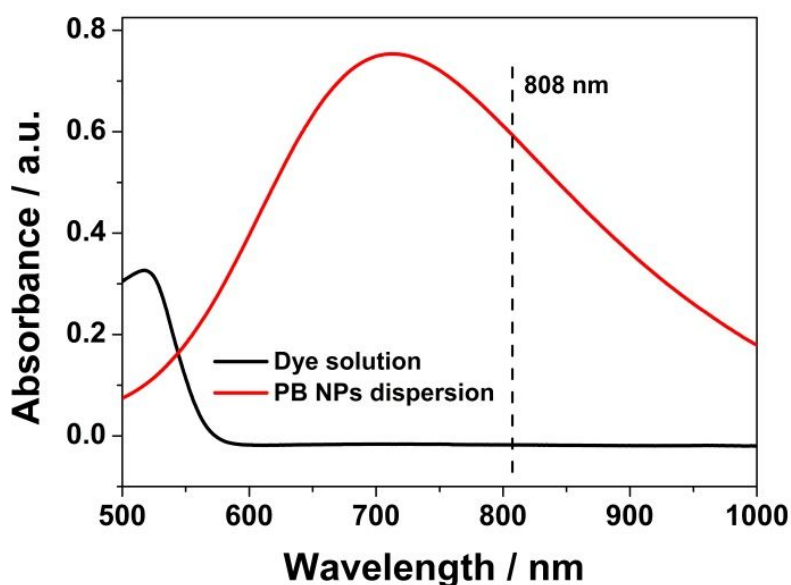


Figure S2. UV-vis absorption spectroscopic characterization of red dye and PB. UV-vis absorption spectra of red dye solution and aqueous dispersion of PB NPs (0.05 mg/mL).

### **Feasibility of Red Dye as On-Chip Quantitative Indicator of the Distance-Based**

**Readout Method.** On the  $\mu$ PAD, the capillary force can drive the flow of dye solutions in the channels, thus displaying visual red strips with quantitative lengths. Different volumes of the red dye solutions were pipetted to inlets of the disk-like  $\mu$ PAD to evaluate the feasibility of the read dye as the on-chip quantitative indicator of the distance-based readout method. As shown in Figure S3, the on-chip travelling distance of the dye solutions is proportional to the solution volume in the range from 2.0 to 20  $\mu$ L at the capillary transport temperature of both 4.0  $^{\circ}$ C and 29  $^{\circ}$ C, indicating



the feasibility of the read dye as the on-chip quantitative indicator of the distance-based readout method on the  $\mu$ PAD. In addition, the effect of temperature on on-chip capillary transport of the dye solution was assessed using 4.0 °C as the control (vs. room temperature of around 29 °C). It was found that the control  $\mu$ PAD (Figure S3B) showed longer dye-travelling distance than that obtained at the room temperature (Figure S3A) at various volumes. Accordingly, the control calibration plot (Figure S3C) exhibited a higher slope than that obtained at the room temperature, illustrating a negative effect of temperature on on-chip transport distance of the dye solution because of the influence on surface tension of the dye solution.

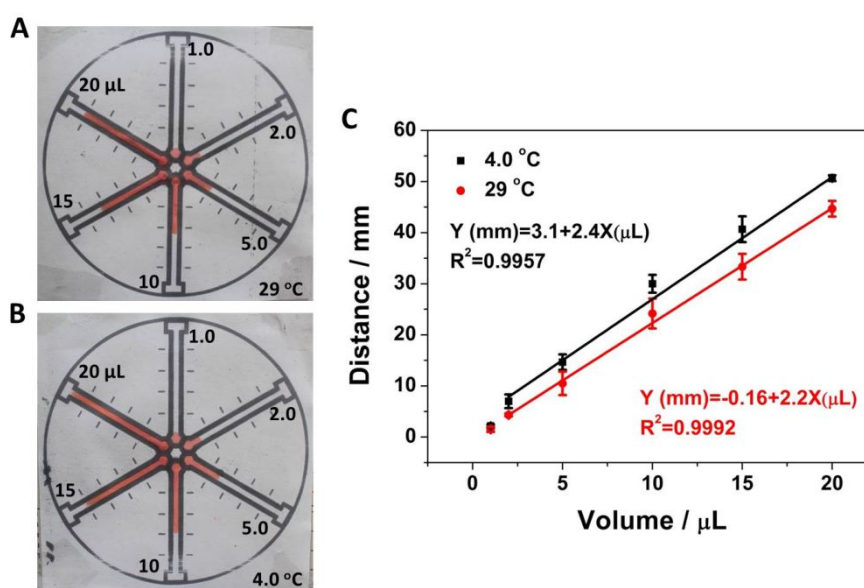


Figure S3. On-chip capillary force-driven travelling behavior of pure dye solution. (A) Photograph of the dye-loaded  $\mu$ PAD with the capillary transport at 29 °C; (B) Photograph of the control dye-loaded  $\mu$ PAD with the capillary transport at 4.0 °C; (C) Calibration plots of on-chip dye-travelling distance vs. volume of the dye solutions. Error bars represent standard deviations (n=3).

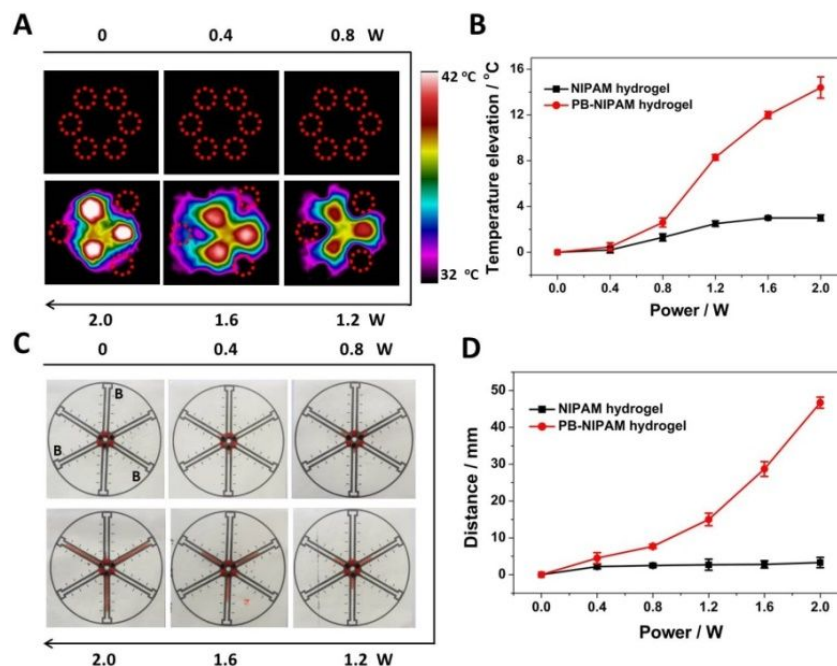


Figure S4. Effect of laser power on on-chip photothermal sensing performance of hydrogels. (A) Thermal images of the blank and composite hydrogels after the laser irradiation for 60 s at different laser powers; (B) Temperature elevations of the hydrogels after the irradiation; (C) Photographs of the hydrogel-loaded  $\mu$ PADs after the irradiation; (D) On-chip travelling distance of dye solutions as a function of laser power. Error bars represent standard deviations ( $n=3$ ).

### Effect of Laser Power on On-Chip Photothermal Sensing Performance of

### Hydrogels.

The effect of laser power on photothermal sensing performance of the composite hydrogel was studied using the dual-mode readout method as shown in Figure S4. With the increase of laser power in the range from 0 to 2.0 W, the composite hydrogels showed progressively increased temperature due to the increasing photothermal effect of PB in the hydrogels. Irradiation of the composite hydrogel at a laser power of 2.0 W resulted in a dramatic temperature elevation of 14.4 °C, while the blank hydrogel exhibited a minor temperature elevation of 3.7 °C.

In good agreement with the thermal image-based readout results, increasing

dye-travelling distance (i.e., 47 mm at 2.0 W) was observed from the composite hydrogels upon the increase of laser power, whereas the blank hydrogels exhibited minor dye-travelling distance of less than 3.3 mm at various laser powers. These results demonstrated a laser power-dependent character of the photothermal sensing performance of the composite hydrogel.

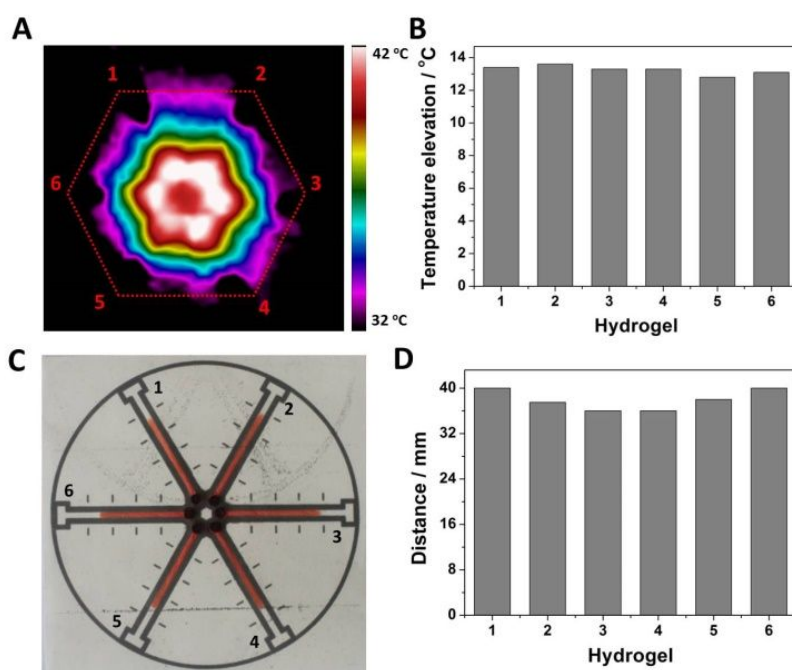


Figure S5. Reproducibility of on-chip photothermal sensing performance of composite hydrogel.

(A) Thermal image of six pieces of the composite hydrogels synthesized from six independent batches after the laser irradiation for 50 s at a power density of  $0.38 \text{ W} \cdot \text{cm}^{-2}$ ; (B) Temperature elevations of the composite hydrogels after the irradiation; (C) Photograph of the hydrogel-loaded  $\mu$ PAD after the irradiation; (D) On-chip travelling distance of dye solutions after the irradiation.

### Reproducibility of On-Chip Photothermal Sensing Performance of Hydrogels.

To study the reproducibility of on-chip photothermal sensing performance of the composite hydrogel, the composite hydrogels prepared in parallel from independent

batches were irradiated for the same time on the  $\mu$ PAD. As can be seen in Figure S5, after the laser irradiation for 50 s at a power density of  $0.38 \text{ W}\cdot\text{cm}^{-2}$ , uniform temperature elevations of six pieces of the composite hydrogels were monitored with a low relative standard deviation (RSD) of 2.0%. In addition, the composite hydrogels exhibited uniform on-chip dye-travelling distance with a relative standard deviation (RSD) of 4.8%. These results demonstrated good reproducibility of on-chip photothermal sensing performance of the composite hydrogel.

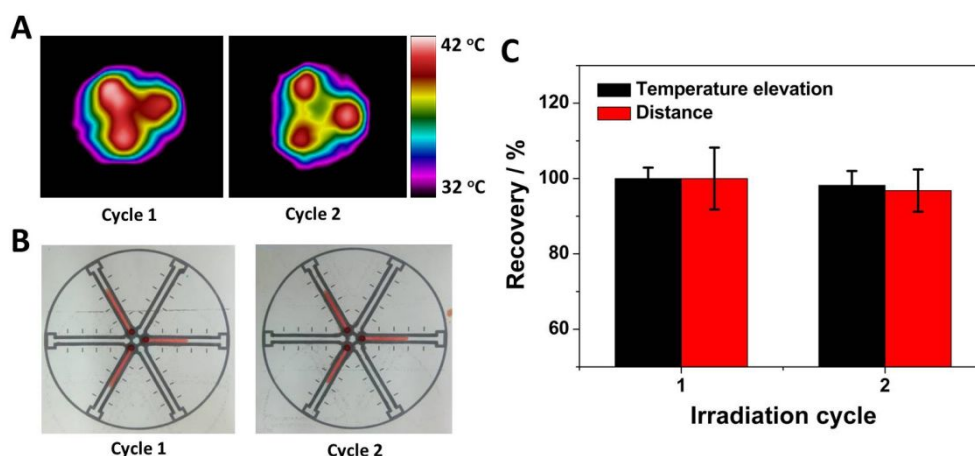


Figure S6. Recovery of on-chip photothermal sensing performance upon reuse of hydrogels. (A) Thermal images of the composite hydrogels after 2 cycles of laser irradiation for 40 s at a power density of  $0.38 \text{ W}\cdot\text{cm}^{-2}$ ; (B) Photographs of the hydrogel-loaded  $\mu$ PADs after the irradiation; (C) Temperature elevations of the hydrogels and on-chip travelling distance of dye solutions after the irradiation. Error bars represent standard deviations ( $n=3$ ).

**Recovery of On-Chip Photothermal Sensing Performance upon Reusage of Hydrogels.** It had been reported that the structural phase transition of NIPAM-derived thermoresponsive hydrogels was completely reversible in response to external temperature changes. Herein, the recovery of on-chip photothermal sensing

performance of the composite hydrogel was investigated upon multiple cycles of reuse of the composite hydrogel, as shown in Figure S6. The composite hydrogels on the  $\mu$ PAD were treated by two cycles of reuse. In each cycle, the hydrogels were first irradiated for 40 s, followed by off-chip re-storage of the hydrogels in red dye solutions at room temperature to allow their thorough swelling. The composite hydrogels showed no distinct changes in both the thermal image- and distance-based readout results after the above treatment, suggesting remarkable recovery of on-chip photothermal sensing performance of the composite hydrogel on the  $\mu$ PAD. The good recovery can be attributed to the reversible phase-transition behavior of the composite hydrogel upon thermal stimulus.<sup>5, 6</sup>

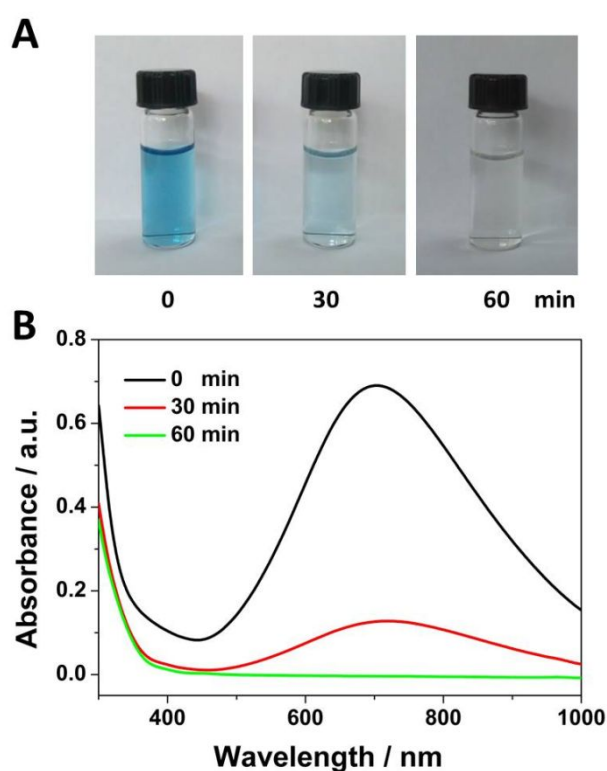


Figure S7. Confirmation of material conversion in the presence of  $\text{Ag}^+$ . (A) Photographs of aqueous dispersions of PB NPs (0.05 mg/mL) after incubation in  $\text{Ag}^+$  solutions (5.0  $\mu\text{M}$ ) for

different times; (B) UV-vis adsorption spectra of the PB NPs dispersions after the incubation in  $\text{Ag}^+$  solutions.

**Confirmation of Sensing Principle of Silver Ions.** The material conversion from photothermal PB to non-photothermal silver ferrocyanide in the presence of  $\text{Ag}^+$  was confirmed by UV-vis adsorption spectroscopy as shown in Figure S7. PB NPs were incubated in  $\text{Ag}^+$  solutions (5.0  $\mu\text{M}$ ) for different times to monitor their colorimetric and spectrophotometric changes. As can be seen, the color of the PB NPs dispersions degraded gradually with the increase of incubation time, which became colorless after the incubation for 1.0 hour. Meanwhile, the absorbance at 710 nm as well as in the NIR region (700-1000 nm) in UV-vis adsorption spectra of the PB NPs dispersions decreased dramatically with the increase of incubation time. These results confirmed the conversion of photothermal PB to colorless non-photothermal silver ferrocyanide in the presence of  $\text{Ag}^+$ . With a higher solubility product equilibrium constant ( $K_{\text{sp}}$ ) and worse stability, PB ( $\text{Fe}_4[\text{Fe}(\text{CN})_6]_3$ ) was transformed to silver ferrocyanide ( $\text{Ag}_4[\text{Fe}(\text{CN})_6]_3$ ) in the presence of  $\text{Ag}^+$  according to the dissolution-precipitation equilibrium mechanism.<sup>7</sup>

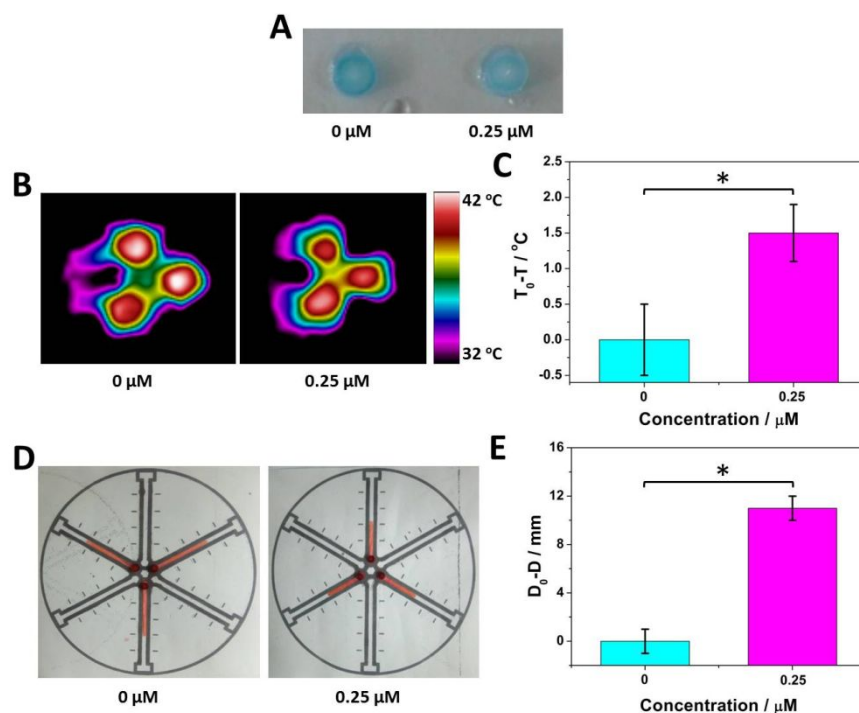


Figure S8. Determination of  $\text{Ag}^+$  upon prolonging of sample-pretreatment time. (A) Photograph of the composite hydrogels after incubation in different concentrations of  $\text{Ag}^+$  solutions for 10 hours; (B) Thermal images of the composite hydrogels after the laser irradiation for 60 s at a power density of  $0.38 \text{ W} \cdot \text{cm}^{-2}$ ; (C) Relative temperature changes ( $T_0 - T$ ) of the hydrogels after the irradiation; (D) Photographs of the hydrogel-loaded  $\mu\text{PADs}$  after the irradiation; (E) Relative changes in on-chip travelling distance ( $D_0 - D$ ) of dye solutions after the irradiation. Error bars represent standard deviations ( $n=3$ ). An asterisk indicates a statistically significant difference ( $t$ -test,  $*P < 0.05$ ).

**Determination of  $\text{Ag}^+$  upon Prolonging of Sample-Pretreatment Time.** To study the effect of sample-pretreatment time on detection limit of the photothermal  $\mu\text{PAD}$  for  $\text{Ag}^+$  detection, the composite hydrogels were incubated in  $\text{Ag}^+$  solutions (0.25  $\mu\text{M}$ ) for 10 hours. After the incubation, the composite hydrogels (0.25  $\mu\text{M}$ ) exhibited obviously degraded color in comparison with the control hydrogels (0  $\mu\text{M}$ ), as shown in Figure S8. After laser irradiation of the hydrogels on the  $\mu\text{PAD}$ , significant

difference between the control (0  $\mu\text{M}$ ) and the  $\text{Ag}^+$ -treated hydrogels (0.25  $\mu\text{M}$ ) was observed from both the thermal image- and distance-based readout results. Hence,  $\text{Ag}^+$  can be quantitatively determined with lower detection limits by properly prolonging the incubation time of the composite hydrogels in  $\text{Ag}^+$  solutions. It was worth noting that the control hydrogel displayed both lower temperature elevation and shorter on-chip dye-travelling distance in comparison with that of the control hydrogel with a shorter incubation time (i.e., 1.5 hours), which can be ascribed to the inevitable minor decomposition of PB at neutral conditions.

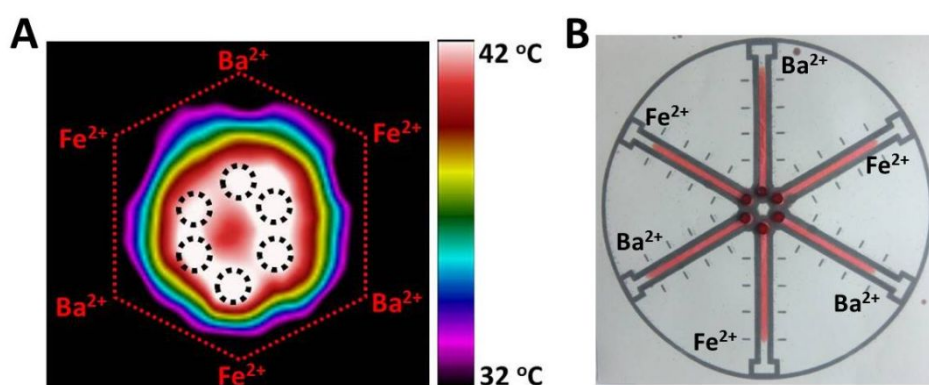


Figure S9. Selectivity of the photothermal  $\mu\text{PAD}$  towards  $\text{Ba}^{2+}$  and  $\text{Fe}^{2+}$ . (A) Thermal image of the  $\text{Ba}^{2+}$ - and  $\text{Fe}^{2+}$ -treated composite hydrogels after the laser irradiation for 60 s at a power density of  $0.38 \text{ W} \cdot \text{cm}^{-2}$ ; (B) Photograph of the hydrogel-loaded  $\mu\text{PAD}$  after the irradiation.



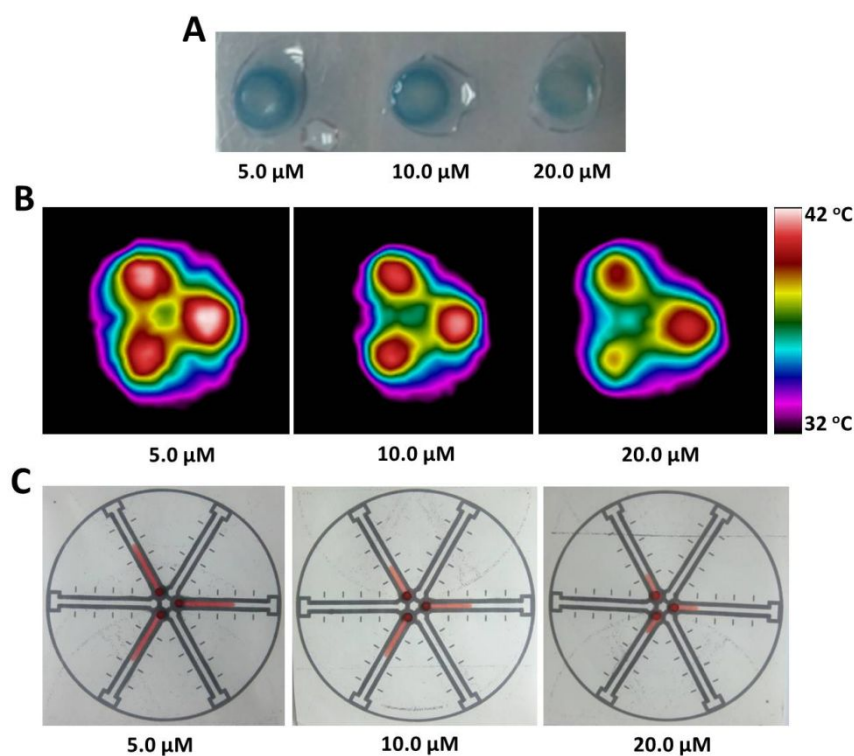


Figure S10. Determination of  $\text{Ag}^+$  in real water samples. (A) Photograph of the composite hydrogels after incubation with water samples spiked with different concentrations of  $\text{Ag}^+$  solutions; (B) Thermal images of the composite hydrogels after the laser irradiation for 60 s at a power density of  $0.38 \text{ W} \cdot \text{cm}^{-2}$ ; (C) Photographs of the hydrogel-loaded  $\mu\text{PADs}$  after the irradiation.

**Determination of  $\text{Ag}^+$  in Real Water Samples.** Real water samples were collected from a local lake (Jiulong Lake) and were diluted 5-fold with B-R buffer solutions ( $\text{pH}=7.2$ ). The samples were then spiked with different concentrations of standard  $\text{Ag}^+$  solutions to prepare water samples with final  $\text{Ag}^+$  concentrations of 5.0, 10, 20  $\mu\text{M}$ , respectively. After incubation of the composite hydrogels in the water samples,  $\text{Ag}^+$  concentrations were determined on the photothermal  $\mu\text{PAD}$  using the dual-mode readout method. As shown in Figure S10, with the increase of  $\text{Ag}^+$  concentration in the water samples, the

composite hydrogels exhibited increasingly degraded color. In addition, the quantitative detection results acquired from both the thermal image- and distance-based readout methods were in good agreement with the results obtained from Figure 5.

Table S1. Determination of  $\text{Ag}^+$  in spiked real water samples using the thermal image-based readout method on the photothermal  $\mu\text{PAD}$  (n=4).

Sample number	Spiked concentration ( $\mu\text{M}$ )	Detected concentration ( $\mu\text{M}$ )	Recovery (%)	RSD (%)
1	5.0	4.79 $\pm$ 0.10	95.8	2.09
2	10	10.2 $\pm$ 0.55	101.8	5.40
3	20	19.5 $\pm$ 0.90	97.5	4.62

Table S2. Determination of  $\text{Ag}^+$  in spiked real water samples using the distance-based readout method on the photothermal  $\mu\text{PAD}$  (n=4).

Sample number	Spiked concentration ( $\mu\text{M}$ )	Detected concentration ( $\mu\text{M}$ )	Recovery (%)	RSD (%)
1	5.0	4.79 $\pm$ 0.20	95.8	3.97
2	10	9.55 $\pm$ 0.65	95.5	6.81
3	20	19.3 $\pm$ 1.6	96.3	8.10

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