Supporting information

Nutrient-Based Chemical Library as a Source of Energy Metabolism Modulators

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Supplementary Chemistry Notes

General part

Unless otherwise noted all solvents and chemicals for chemical synthesis were purchased used without further purification. Some synthetic compounds were purified using a flash column system Shoko Scientific Purif $-\alpha 2$. Preparative layer chromatography was carried out with glass PLC plates, precoated with silica gel 60 F254 (Merck). Low -resolution mass spectra were obtained using a Shimadzu LCMS -2010EV in ESI mode. LC-MS conditions were as follows: Shimadzu Shim -pack XR -ODS column (2.2 µm, 3.0×50 mm); solvent gradient, A, 0.1% trifluoroacetic acid (TFA) in H2O; B, acetonitrile with gradient (0–3.5 min: B, 10–100%; 3.5-5 min: B, 100%; 5-5.5 min: B 10% at 40 °C); flow rate, 0.7 mL/min; detector, 220 nm. High resolution mass spectra (HRMS) were obtained using a JEOL MStation JMS -700 in FAB mode or Bruker SolariX in ESI mode. 1H -NMR spectra were collected by Bruker Avance III 800US Plus NMR or Bruker Avance III 600US Plus NMR. Chemical shifts were reported in δ (ppm) relative to tetramethylsilane.

Experimental procedures

Typical procedure for library compound synthesis

To a solution of an amine (0.14 mmol), a carboxylic acid (0.14 mmol) and an aldehyde (0.14 mmol) in EtOH (0.5 mL) and sat. NaHCO₃ aq. (0.1 mL) was added isocyanoethylacetate (0.14 mmol) and stirred for 2 h. The reaction mixture was charged on a PTLC plate and separated with CHCl₃-MeOH.

Modified synthesis of molecule 1



To a solution of D-(+)-glucosamine hydrochloride (150 mg), DHA (245 μ L), phenylacetoaldehyde (82 μ L) and LiOH • H₂O (30 mg) in water (5 mL) and EtOH (0.5 mL) was added ethyl isocyanoacetate (77 μ L). After stirring for 2 h, the reaction mixture was charged on a silica gel column and eluted with CHCl₃/MeOH (100:0 ~ 80:20) to give molecule **1** (25.8 mg, 5.1% yield, dr = 1:1). ¹H-NMR (600 MHz, CDCl₃) &: 7.39 - 7.26 (5H, m), 6.71 - 6.46 (1H, m), 5.63 - 5.29 (12H, m), 4.74 - 4.67 (1H, m), 4.23 - 4.13 (2H, m), 4.05 - 3.98 (1H, m), 3.97 - 3.96 (1H, m), 3.90 - 3.88 (2H, m), 3.80 - 3.72 (2H, m), 3.62 - 3.59 (1H, m), 3.47 - 3.39 (1H, m), 2.86 -2.80 (10H, m), 2.53 - 2.30 (4H, m), 2.07 (2H, quintet of doublet, *J* = 1.2, 0.6 Hz), 1.26 (3H, dt, *J* = 22.8, 7.2 Hz), 0.97 (3H, t, *J* = 7.2 Hz). ¹³C-NMR (150MHz, CDCl₃) &: 176.20, 174.38, 173.32, 172.71, 170.29, 170.27, 137.89, 137.44, 132.06, 129.48, 129.20, 129.18, 129.14, 129.02, 128.95, 128.59, 128.53, 128.31, 128.30, 128.27, 128.26, 128.24, 128.23, 128.21, 128.16, 128.11, 127.90, 127.89, 127.48, 127.39, 127.03, 93.97, 93.10, 75.37, 71.82, 71.68, 71.26, 71.12, 68.71, 65.18, 62.71, 62.49, 62.18, 62.03, 61.77, 61.66, 58.39, 42.06, 41.78, 35.56, 35.20, 34.14, 34.10, 25.68, 25.67, 25.66, 25.64, 25.63, 25.54, 22.68, 22.58, 20.57, 14.29, 14.11, 14.10. HRMS (FAB, m/z, [M+Na]⁺) calculated For C₄₁H₅₈N₂O₉Na: 745.4040 found 745.4043.

Synthesis of Molecule 3



To a solution of DHA (35 μ L), DL-Phenylalanylglycine ethyl ester **2** (30 mg) and HATU (42 mg) in DMF (1 mL) was added DIEA (80 μ L). After stirring for 1 h, the reaction mixture was quenched with water (5 mL) and extracted by EtOAc (2 mL×3). The organic layer was washed with sat. NaCl aq., dried over Na₂SO₃ and concentrated *in vacuo* to give a crude product. The crude product was purified by flash silica gel column chromatography with n-hexane/EtOAc (95:5 ~ 80:20) to give **3** (36.4 mg, 61% yield). ¹H-NMR (600 MHz, DMSO-d6) δ : 8.46 - 8.38 (1H, m), 8.09 (1H, d, *J* = 8.4 Hz), 7.18 - 7.17 (5H, m), 5.36 - 5.17 (12H, m), 4.58 - 4.48 (1H, m), 4.09 (2H, q, *J* = 7.2 Hz), 3.88 - 3.78 (2H, m), 3.03 (2H, dd, J = 13.8, 4.2), 2.85 - 2.68 (10H, m), 2.52 - 2.00 (6H, m), 1.19 (3H, t, *J* = 7.2 Hz), 0.91 (3H, t, *J* = 7.8 Hz). ¹³C-NMR (150MHz, DMSO-d6) δ : 171.84, 171.29, 169.56, 137.88, 131.44, 129.03, 128.99, 128.00, 127.96, 127.91, 127.87, 127.82, 127.79, 127.78, 127.76, 127.73, 127.62, 127.58, 126.03, 60.31, 53.43, 40.63, 37.49, 34.89, 25.10, 25.09, 25.08, 25.00, 25.00, 22.85, 19.92, 14.00, 13.94. HRMS (ESI, m/z, [M+Na]⁺) calculated For C₃₅H₄₈N₂O₄Na: 583.35063 found 583.35008.

Synthesis of Molecule 41



To a solution of DHA (98 µL), D-glucosamine hydrochloride (60 mg) and DIEA (53 µL) in EtOH (2 mL) was added DMT-MM (93 mg) and stirred for 3 h. The reaction mixture was charged on a PTLC plate and separated with CHCl₃ / MeOH (80:20) to give molecule **4** (9.8 mg, 7.2% yield). ¹H-NMR (600 MHz, DMSO-d6) δ : 7.57 (1H, d, *J* = 8.4 Hz), 6.43 - 6.38 (1H, m), 5.56 - 5.25 (12H, m), 4.82 - 4.77 (1H, m), 4.58 (1H, d, *J* = 5.4 Hz), 4.53 (1H, t, *J* = 5.4 Hz), 4.43 (1H, t, *J* = 4.8 Hz), 3.75 - 3.66 (1H, m), 3.64 - 3.55 (3H, m), 3.52 - 3.44 (2H, m), 2.85 - 2.77 (10H, m), 2.22 - 2.28 (2H, m), 2.12 - 2.19 (2H, m), 1.99 - 2.06 (2H, m), 0.92 (3H, t, *J* = 7.8 Hz). ¹³C-NMR (150MHz, DMSO-d6) δ : 161.78, 131.45, 129.04, 129.00, 128.06, 128.02, 127.81, 127.79, 127.77, 127.75, 127.67, 127.60, 126.84, 90.49, 71.04, 70.34, 69.67, 64.28, 61.02, 31.17, 25.11, 25.10, 25.05, 25.01, 24.37, 22.95, 21.97, 14.02. HRMS (ESI, m/z, [M+Na]⁺) calculated For C₂₈H₄₃NO₆Na: 512.29826 found 512.29839.

Synthesis of Molecule 5²



To a solution of D-glucosamine hydrochloride (107 mg), boric acid (31 mg), phenylacetoaldehyde (66μ L) and LiOH • H₂O (21 mg) in water (2 mL) was added ethyl isocyanoacetate (40μ L). After stirring for 2h, the reaction mixture was charged on PTLC plate and separated with CHCl₃/MeOH (80:20) to give molecule **5** (10.5 mg, 5.3% yield, dr = 1:1). ¹H-NMR (600 MHz, DMSO-d6) δ : 8.76 - 8.24, (1H, m), 7.33 - 7.16 (5H, m), 6.67 - 6.09 (1H, m), 4.97 - 4.91 (2H, m), 4.89 - 4.74 (1H, m), 4.51 - 4.30 (1H, m), 4.09 (2H, q, *J* = 7.2 Hz), 3.88 - 3.77 (2H, m), 3.75 - 3.48 (1H, m), 3.48 - 3.20 (6H, m, H₂O peak overwrapped) 3.09 - 2.98 (2H, m), 2.98 - 2.71 (1H, m), 1.21 - 1.18 (3H, m). ¹³C-NMR (200MHz, DMSO-d6) δ : 174.39, 174.04, 169.86, 169.58, 137.82, 137.67, 129.32, 129.07, 129.04, 128.91, 128.20, 127.93, 91.33, 89.64, 76.70, 75.96, 73.57, 72.11, 71.92, 71.72, 70.63, 69.67, 69.47, 63.08, 61.07, 60.99, 60.41, 60.31, 40.63, 40.57, 33.55, 33.07, 13.95, 13.84. HRMS (ESI, m/z, [M+Na]⁺) calculated For C₁₉H₂₈N₂O₈Na: 435.17379 found 435.17390.

Structural characterization

¹H (600 MHz) NMR spectrum of molecule 1 in CDCl₃



¹³C (150 MHz) NMR spectrum of molecule 1 in CDCl₃





¹H (600 MHz) NMR spectrum of molecule 3 in DMSO-d6



¹³C (150 MHz) NMR spectrum of molecule 3 in DMSO-d6



HRMS spectrum of molecule 3



¹H (600 MHz) NMR spectrum of molecule 4 in DMSO-d6



¹³C (150 MHz) NMR spectrum of molecule 4 in DMSO-d6





¹H (600 MHz) NMR spectrum of molecule 5 in DMSO-d6



¹³C (200 MHz) NMR spectrum of molecule 5 in DMSO-d6



HRMS spectrum of molecule 5



Supplementary Biology Notes

Antibodies

Anti SREBP-2 (IgG-7D4)³ were obtained from Division of Metabolic Medicine, Research Center for Advanced Science and Technology, The University of Tokyo. Anti β -actin (IgG-Ac-40) was obtained from Abcam. Anti AMPK- α (IgG-C20) was from Santa Cruz Biotechnology Inc. Anti phospho-AMPK α (Thr172) (IgG-40H9) was from Cell Signaling Technology.

Cell Culture

CHO-K1 cells were maintained in a medium A (1:1 mixture of Ham's F-12 medium and DMEM, supplemented with 100 units/mL penicillin, 100 mg/mL streptomycin sulfate, and 5% [v/v] fetal bovine serum) at 37°C in a humidified 5% CO2 incubator. Hepa 1-6 cells were maintained in a medium B (DMEM, supplemented with 100 units/mL penicillin, 100 mg/mL streptomycin sulfate, and 10% [v/v] fetal bovine serum) at 37°C in a humidified 5% CO2 incubator. Caco-2 cells were maintained in a medium C (DMEM, supplemented with 100 units/mL penicillin, 100 mg/mL streptomycin sulfate, 10% non-essential amino acids, and 10% [v/v] fetal bovine serum) at 37°C in a humidified 5% CO2 incubator.

Luciferase Reporter Assay

SREBP Reporter Assay

On day 0, CHO-K1 cells were added to 96-well plates at 1.0×10^4 cells per well in medium A. On day 1, the cells were co-transfected with an SRE-1-driven luciferase reporter plasmid (pSRE-Luc) and a β -gal reporter plasmid with a constitutively active actin promoter (pAc- β -gal) at a 20/1 ratio, using FuGENE HD Transfection Reagent (Promega), as instructed in the manufacturer's protocol. After 12-hr incubation, the cells were washed with PBS and then incubated in medium D (1:1 mixture of Ham's F-12 medium and DMEM, supplemented with 100 units/mL penicillin, 100 mg/mL streptomycin sulfate, 5% [v/v] lipid-depleted serum, 50 μ M compactin, and 50 μ M lithium mevalonate) containing the specific test compounds. Stock solutions of each compound in DMSO were prepared and added to medium A to give a 1000-fold (v/v) dilution (0.1% DMSO). After 24 h of incubation, the cells in each well were lysed by freeze-thaw with Reporter Lysis Buffer (Promega), and aliquots were used for meas-uring luciferase and β -gal activities. Luciferase activity was measured using the Steady-Glo Luciferase Assay System (Promega), and β -gal activity. LXR Reporter Assay

On day 0, CHO-K1 cells were added to 96-well plates at 1.0×10^4 cells per well in medium A. On day 1, the cells were co-transfected with an LXRdriven luciferase reporter plasmid (pLXR-Luc) and a β -gal reporter plasmid with a constitutively active actin promoter (pAc- β -gal) at a 20/1 ratio, using FuGENE HD Transfection Reagent (Promega), as instructed in the manufacturer's protocol. After 12-hr incubation, the cells were washed with PBS and then treated with medium A containing the specific test compounds. Stock solutions of each compound in DMSO were prepared and added to medium A to give a 1000-fold (v/v) dilution (0.1% [v/v] DMSO). After 24 h incubation, luciferase and β -gal activities were measured in a same manner as described above.

Western blot analysis

At the end of incubation, the cells were washed three times with cold PBS, and lysed with buffer A (50 mM Tris-HCl [pH 7.5], 150 mM NaCl, 1% [v/v] Nonidet P-40, 0.5% [w/v] sodium deoxycholate, 8 M urea, and protease inhibitor cocktail (Nacalai Tesque)). The cell lysates were passed 20 times through a 25G needle and centrifuged at 7,000×g at 4°C for 10 min. The supernatants were transferred to new tubes. The resulting lysate was mixed with 0.20 volume of 6×SDS sample buffer (Nacalai Tesque) and incubated at room temperature for 1 h. The samples were separated on a 4-15% Tris-HCl precast SDS PAGE gel (BIO-RAD) and blotted using specific antibodies. The specific bands were visualized using enhanced chemiluminescence (ECL Prime Western Blotting Detection Reagent, GE Healthcare) on an ImageQuant LAS 500 (GE Healthcare).

Glucose uptake assay

Glucose uptake was determined using Glucose Uptake Cell-Based Assay Kit (Cayman Chemical). On day 0, Caco-2 cells were added to 96-well plates at 1.0×10^4 cells per well in medium C. The medium was changed every 1-2 days, and the culture was carried out for 21 days. Then the cells were starved in medium E (glucose-free DMEM, supplemented with 100 units/mL penicillin, 100 mg/mL streptomycin sulfate) for 60 min. The cells were washed with PBS and then incubated for 3 h in medium E containing the specific test compounds (1000 fold dilution, 0.1% DMSO) and 2-NBDG (200 μ M). After three-time extensive wash with cold Cell-Based Assay Buffer (Cayman Chemical), fluorescent intensity was measured by a microplate reader (MTP-880) with the 490-nm excitation and 530-nm emission. Glucose uptake was quantified relative to DMSO control in %.

The selectivity of molecule **1** to GLUTs was evaluated in GLUT-overexpressing HEK293 cells. HEK293 cells were transiently transfected with an expression vector of each of GLUT1-4 (*SLC2A1-4*) and then treated with varied concentrations of molecule **1**. Uptake of fluorescently-labeled glucose analog 2-NBDG was measured as described above.

Mice glucose tolerant test

Prior to the test, male ICR mice were fasted for 24 h. Molecules 1 or 3 were orally administrated simultaneously or prior to oral administration of glucose. (i) Co-administration: D-glucose 12.5 mg with or without molecules 1 or 3 (6.25 μ mol) in distilled water (250 μ L) were orally administrated into the mice. (ii) Pretreatment: molecules 1 or 3 (6.25 μ mol) in distilled water (250 μ L) were orally administrated 10 min before administration of D-glucose (12.5mg) in distilled water (250 μ L). At 5, 15, 30, 60, 90 and 120 min after the D-glucose administration, blood was obtained from a tail (5 μ L) and assessed for glucose levels using a blood glucose self-monitoring kit (Eidia). Changes in continuous variables between the two examples were assessed using paired t-test.

Hexokinase inhibition assay

Effects of **1** (10 µM), molecules **3-5** (10 µM) and 3-bromopyruvic acid (positive control, 40 mM) on human hexokinase was measured using a Hexokinase Assay kit (BioVision) according to the manufacture's protocol. Briefly, glucose was converted to glucose-6-phosphate by hexokinase, and the glucose-6-phosphate is oxidized by glucose-6-phosphate dehydrogenase to form NADH, which reduces a colorless probe to a colored product with strong absorbance at 450 nm.

Supplementary Figures



Figure S1. Examples of nutrient conjugates. (A) Tocopherol nicotinate. (B) AA-2G. (C) NA-Gly.



Figure S2. Chemical structures of 5 screening hits and their IC_{5^0} values.



Figure S3. The ability of molecule 1 to block the glucose uptake in GLUT-overexpressing HEK293 cells. HEK293 cells were transiently transfected with an expression vector of each of GLUT1-4 (*SLC2A1-4*) and then treated with varied concentrations of molecule 1. Uptake of fluorescently-labeled glucose analog 2-NBDG was measured. After extensive wash, fluorescence of the cells was measured with a fluorescence plate reader (excitation: 490 nm; emission: 530 nm).



Figure S4. AMPK activation by molecule 1. Hepa 1-6 cells were treated with molecule 1 (10 μ M), phloretin (100 μ M), or molecules 3-5 (10 μ M), and the cell lysates were analyzed by western blots with an antibody specific for phosphor-Thr172 AMPK. It is evident that molecule 1 activates AMPK phosphorylation just as phloretin while molecules 3-5 have no detectable effects.



Figure S5. SREBP inhibitory activities of molecule 1, 6, 7 and molecules 10-12. Luciferase activities were measured at a concentration of 10 μ M after 24-h incubation.



Figure S6. SREBP inhibitory activities of molecule 1 and molecules 13-14. Luciferase activities were measured at a concentration of 10 μ M after 24-h incubation.



Figure S7. LXR reporter gene assay in CHO-K1 cells. CHO-K1 cells were co-transfected with an LXR-driven luciferase reporter plasmid (pLXR-Luc) and a β -gal reporter plasmid with a constitutively active actin promoter (pAc- β -gal). The cells were then treated with 5 μ M of molucle 1, molucles 6-9, 25-HC, or T0901317 (LXR agonist)⁴. After 24-h incubation, luciferase and β -gal activities were measured.



Figure S8. Effects on human hexokinase. Hexokinase activity was measured in the presence of of 1 (10 μ M), molecules 3-5 (10 μ) or a known inhibitor (3-bromopyruvic acid, 40 mM) using a Hexokinase Assay kit (BioVision). Glucose was converted to glucose-6-phosphate by hexokinase, and the glucose-6-phosphate is oxidized by glucose-6-phosphate dehydrogenase to form NADH, which reduces a colorless probe to a colored product with strong absorbance at 450 nm.

Supplementary Table

Table S1. Structures, chemical yield, LC-MS data, and SREBP inhibitory activity of library compounds.

	Structure		yield (%)		LC-MS	SREBP Inhibition	
No.		M.W.		RT (min)	m/z	Purity (area%)	@10 µM (%) (Primary Screening)
15		604.74	12.8	4.21	605 [M+H]⁺	96.8	19.6
16	HO, HOH HO, HOH	604.74	15.9	4.20	605 [M+H]⁺	81.5	6.3
17		602.77	5.3	3.57	603 [M+H]⁺	97.6	21.5
18		606.76	12.8	3.75	607 [M+H]+	95.5	-80.2

13	632.80	15.0	4.45	633 [M+H]+	91.9	4.7
19	608.77	23.6	4.51	609 [M+H]*	95.8	-124.0
20	582.74	16.4	4.17	583 [M+H]⁺	95.6	26.6
21	584.75	16.5	3.65	585 [M+H]⁺	97.4	9.2
22	548.67	10.9	3.08	549 [M+H]+	92.1	16.0
23	476.57	20.1	2.46	477 [M+H]*	87.0	-43.7
24	490.59	18.3	2.67	491 [M+H]*	94.2	9.1
25	504.62	14.6	2.91	505 [M+H]⁺	99.3	-6.4
26	560.73	3.1	3.98	561 [M+H]*	99.1	38.0
27	558.71	13.7	4.19	559 [M+H]⁺	94.6	45.6

28	HO, HO HO, HO HO	574.76	13.3	4.23	597 [M+Na] ⁺	93.3	40.6
29	HO, HO HO, HO HO	572.74	12.5	3.73	573 [M+H]+	87.5	51.7
30		588.78	12.7	4.54	589 [M+H]⁺	93.4	37-3
31		586.77	20.0	4.14	587 [M+H]⁺	96.7	41.5
32		586.77	12.2	4.01	587 [M+H]⁺	95.8	41.9
33		548.67	11.7	3.62	549 [M+H]+	94.0	21.7
34		523.54	6.5	1.23	524 [M+H]+	94.2	6.4
35	HO, COH	427.41	6.2	0.63	428 [M+H]+	99.3	-1.9
36		548.61	2.9	1.14	549 [M+H]+	88.9	1.6
37	HOV, HOH	694.87	2.8	4.62	695 [M+H]+	95.8	65.6

6	HO, HOH HO, HOH	694.87	3.3	4.59	695 [M+H]+	70.1	89.5
38		696.88	2.9	3.89	697 [M+H]+	89.9	79.9
1		722.92	5.1	4.04	745 [M+Na] ⁺	96.8	90.8
39		698.90	2.7	4.83	699 [M+H]+	97.2	77-3
7		672.86	3.3	4.59	673 [M+H]⁺	97.2	90.2
40		674.88	2.9	4.05	675 [M+H]+	84.6	81.6
41		638.80	4.6	3.57	639 [M+H]+	84.7	82.4
10		566.69	2.9	2.94	567 [M+H]*	72.0	62.4
11		580.72	2.7	3.16	581 [M+H]+	93.7	75.5
12		594.75	2.8	3.38	595 [M+H]+	91.7	36.6

42		517.54	5.0	1.33	518 [M+H]+	87.8	18.1
43	HOX, HOH	646.82	9.9	3.73	647 [M+H]+	85.8	33.8
44	HO, HOH HO HOH	646.82	11.9	3.71	647 [M+H]+	97.2	51.3
45		644.85	3.7	3.50	645 [M+H]⁺	77.6	44.6
46	H H H H H H H H H H H H H H H H H H H	648.84	2.9	3.70	649 [M+H]*	75.9	66.3
47		674.88	3.0	3.89	675 [M+H]⁺	66.9	67.3
48		650.85	16.5	3.97	651 [M+H]+	91.2	36.6
49	HO HO HO HO HO HO HO HO HO HO HO HO HO H	624.82	2.9	3.72	625 [M+H]+	93.6	43.7
50		626.83	3.1	4.09	649 [M+Na] ⁺	69.0	45.1
51		590.76	2.9	3.42	591 [M+H]+	92.7	30.5

52		518.65	3.3	2.85	519 [M+H]+	96.9	26.9
53		532.68	8.4	3.06	533 [M+H]⁺	96.3	24.5
54		546.70	7.7	3.28	547 [M+H]+	86.7	17.8
55		602.81	10.4	4.19	603 [M+H]⁺	99.1	26.0
56		600.79	4.8	3.84	601 [M+H]+	92.9	52.7
57	HO ₂ , OH HO ₂ , OH HO	616.84	2.8	4.43	617 [M+H]+	95.6	33.5
58		614.82	7.4	4.03	615 [M+H]+	96.0	50.3
59		630.86	5.4	4.75	653 [M+Na]⁺	96.2	27.8
60		628.85	3.5	4.25	651 [M+Na]+	83.0	37.1
61		628.85	6.5	4.27	629 [M+H]+	84.3	8.5

62		590.76	3.1	3.31	613 [M+Na]+	85.5	21.7
63		565.62	5.9	1.34	588 [M+Na] ⁺	99.6	14.6
64		469.49	13.7	1.18	470 [M+H]+	91.1	11.3
65		590.69	8.2	1.52	613 [M+Na]+	98.5	9.1
66		593.77	2.9	3.16	594 [M+H]+	96.3	38.2
67	HO NH HO	593.77	5.2	3.08	594 [M+H]+	98.4	49.5
68		591.79	3.3	2.75	592 [M+H]+	99.0	-3.1
69		595.78	4.8	3.07	596 [M+H]⁺	99.2	39.9
70		621.82	12.2	3.21	622 [M+H]+	99.0	-2.7
71		597.80	8.6	3.25	598 [M+H]+	99.5	30.0
72		571.76	11.2	3.09	572 [M+H]*	98.9	-16.0

73	573.78	11.7	3.26	574 [M+H]*	94.8	64.3
74	537.70	7.0	2.82	538 [M+H] ⁺	96.5	8.5
75	465.59	12.7	2.44	466 [M+H]+	99.0	34.8
76	479.62	12.1	2.60	480 [M+H]+	96.6	-1.1
77	493.65	10.9	2.78	494 [M+H]+	98.5	26.8
78	549.75	11.0	3.46	550 [M+H]+	96.9	-26.5
79	547.74	9.3	3.11	548 [M+H]+	99.1	51.1
80	563.78	3.3	4.73	564 [M+H]+	87.3	-5.0
81	561.76	10.3	3.26	562 [M+H]+	94.7	42.2
82	577.81	5.1	3.69	578 [M+H]⁺	87.5	12.3
83	575.79	7.4	3.44	576 [M+H]+	98.7	56.1

84		575.79	7.9	3.40	576 [M+H]⁺	87.4	65.3
85		537.70	4.9	2.73	538 [M+H]⁺	92.3	-1.8
86		512.56	8.4	1.03	513 [M+H]+	96.2	9.0
87		416.43	3.6	0.85	417 [M+H]+	96.4	5.1
88		537.63	6.4	1.24	538 [M+H]⁺	90.6	12.7
89		683.89	3.4	3.38	684 [M+H]*	98.0	70.9
90	HO N	683.89	6.1	3.25	684 [M+H]+	91.9	66.4
91		681.92	4.0	2.92	682 [M+H]+	87.9	77-4
92		685.91	6.1	3.22	686 [M+H]+	96.0	-58.1
93		711.94	5.2	3.36	712 [M+H] ⁺	99.0	-36.2
94		687.92	5.9	3.39	688 [M+H]+	98.5	-15.7

95		661.88	5.2	3.25	662 [M+H]+	99.1	64.4
96		663.90	6.6	3.44	664 [M+H]*	86.9	63.9
97		627.82	3.6	3.10	628 [M+H]+	93.9	50.9
98	or the second se	555.72	8.2	2.70	556 [M+H]⁺	92.3	40.7
99		569.74	5.5	2.84	570 [M+H]*	85.0	25.6
100		583.77	8.0	2.97	584 [M+H]⁺	88.2	50.8
101		639.88	6.5	3.56	640 [M+H]*	78.9	10.7
102		637.86	4.3	4.52	638 [M+H]+	92.4	67.4
103		653.91	3.6	4.91	654 [M+H]+	92.1	36.1
104		651.89	4.4	4.64	652 [M+H]*	91.4	84.4
105		667.93	2.9	5.10	668 [M+H]+	75.7	8.4

106	665.92	3.4	4.84	666 [M+H]*	99.6	81.8
107	665.92	3.6	4.79	666 [M+H]*	87.6	69.2
108	627.82	4.9	4.22	628 [M+H]+	76.6	52.9
109	602.69	2.8	1.43	638 [M+H]+	94.0	-34.1
110	506.56	11.0	1.35	507 [M+H]*	99.6	-3.6
111	627.76	3.3	1.59	628 [M+H]+	85.5	-14.5
112	635.85	7.2	3.28	636 [M+H]+	86.1	56.6
113	635.85	8 .o	3.10	636 [M+H]+	95.3	73.7
114	633.87	3.6	2.82	634 [M+H]+	97.9	75.2
115	637.86	7.5	3.13	638 [M+H]+	97.2	0.3
116	663.90	4.0	3.27	664 [M+H]+	97.4	13.6

117	639.88	3.7	3.30	640 [M+H]+	98.7	23.8
118	613.84	6.4	3.15	614 [M+H]+	99.7	42.2
119	615.86	8.0	3.29	616 [M+H]+	99.5	79.9
120	579.78	5.3	3.04	580 [M+H] ⁺	93.0	78.2
121	507.67	7.6	3.08	508 [M+H]⁺	98.0	33.3
122	521.70	8.1	3.59	522 [M+H]+	98.1	47.1
123	535.73	7.9	2.87	536 [M+H]+	99.9	60.7
124	591.83	4.0	3.46	592 [M+H]*	99.9	81.3
125	589.82	5.2	3.23	590 [M+H]⁺	97.9	79.1
126	605.86	5.1	3.60	606 [M+H]+	98.9	78.4
127	603.85	5.4	3.37	604 [M+H]+	99.2	77.0

128		619.89	7.0	3.75	620 [M+H]+	97.7	78.0
129		617.87	7.1	3.54	618 [M+H]*	99.0	82.9
130		617.87	6.1	3.51	618 [M+H]+	87.8	81.7
131		579.78	10.2	2.94	580 [M+H]⁺	96.4	83.4
132		554.64	13.4	1.30	555 [M+H]+	99.2	50.9
133		458.52	10.3	1.21	459 [M+H]+	98.0	28.7
134		579.71	3.0	1.47	580 [M+H]⁺	88.2	22.4
135	H H H H H H H H H H H H H H H H H H H	600.84	5.1	4.27	601 [M+H]⁺	94.4	20.7
136	Cet	556.74	83.0	4.11	557 [M+H]+	97.8	71.0
137		554.77	9.9	3.53	555 [M+H]*	94.6	52.4
138		558.76	12.0	3.96	559 [M+H]+	91.1	-57.4

139		584.80	5.1	4.11	585 [M+H]⁺	82.4	16.0
140		560.78	4.6	4.20	561 [M+H]+	95.8	-7.2
141		534.74	4.1	4.00	535 [M+H]+	84.0	72.7
142		536.75	10.2	4.20	537 [M+H]⁺	94.6	41.4
143		500.68	3.0	3.85	501 [M+H]+	96.8	15.0
144		428.57	9.5	3.24	429 [M+H]+	99.2	30.1
145		456.62	4.2	3.68	457 [M+H]⁺	99.1	-2.1
146		512.73	7.0	4.51	513 [M+H]†	96.8	26.3
147	OF HOLE	510.72	16.2	4.12	511 [M+H]+	99.8	24.2
148	OH O OF HOLET	500.68	7.3	3.70	501 [M+H]+	99.8	32.4
149		475-54	3.9	1.54	476 [M+H]+	94.8	24.5

150	379.41	17.5	1.31	378 [M+H]+	95.3	32.1
151	500.61	7.1	1.73	501 [M+H]+	98.2	13.0
152	646.87	4.1	4.56	647 [M+H]+	92.6	83.5
153	624.86	2.9	4.89	625 [M+H]+	78.7	60.9
154	626.88	3.0	4.48	627 [M+H]+	90.6	81.8
155	469.54	4.6	2.00	470 [M+H]*	98.3	43.3
156	590.74	2.7	2.35	591 [M+H]+	92.3	57.0
157	598.83	3.0	4.50	599 [M+H]⁺	99.4	74.0
158	596.85	3.2	3.99	597 [M+H]+	77.8	69.5
159	600.84	5.1	4.37	601 [M+H]+	93.5	48.4
160	626.88	5.6	4.55	627 [M+H]+	90.1	42.1

161	602.86	6.0	4.72	603 [M+H]+	77.5	48.0
162	576.82	3.6	4.44	577 [M+H]+	94.4	60.5
163	578.84	13.2	4.77	579 [M+H]*	94.3	41.4
164	542.76	13.4	4.25	543 [M+H]+	94.3	cell death
165	470.65	12.0	3.79	471 [M+H]+	98.8	cell death
8	498.71	15.2	4.14	499 [M+H]+	99.0	95.1
166	554.81	9.0	5.29	555 [M+H]+	99.0	cell death
167	552.80	16.2	4.64	553 [M+H]*	95.8	cell death
168	542.76	3.2	4.12	543 [M+H]+	87.9	cell death
169	517.62	2.8	2.00	518 [M+H] ⁺	83.4	57.9
170	421.49	17.8	1.84	422 [M+H]*	98.8	56.4

171		542.69	2.9	2.19	543 [M+H]+	86.2	44.3
172		602.83	5.0	4.35	603 [M+H] ⁺	87.8	41.9
173		604.85	3.0	4.09	605 [M+H]+	75.5	46.2
174		630.89	2.9	4.23	631 [M+H]+	79.0	21.3
175		606.86	3.6	4.47	607 [M+H]+	94.5	64.9
176		580.83	4.3	4.23	581 [M+H]*	84.7	4.1
177		582.84	2.7	4.55	582 [M+H]+	80.4	61.0
178	OH COEL	546.76	3.8	4.06	547 [M+H]+	91.2	14.8
179		502.71	3.0	4.00	503 [M+H]+	76.6	14.3
180		556.80	4.6	4.42	557 [M+H]*	98.5	45.5
181	HO, Y, H, H, OLOET OH H EIO S	521.63	6.6	1.89	522 [M+H]+	83.9	22.6
182		425.50	3.7	1.72	426 [M+H]+	94.4	3.0

183	546.70	6.5	2.07	547 [M+H]+	98.0	15.4
184	644.91	3.1	4.73	645 [M+H]+	78.9	57.3
185	646.93	3.3	4.57	647 [M+H]*	94.2	45.7
186	672.97	3.2	4.78	673 [M+H]+	84.2	32.9
187	648.94	4.1	4.94	649 [M+H]+	88.1	46.7
188	622.91	3.6	4.63	623 [M+H]+	86.6	51.4
189	624.92	4.8	4.91	625 [M+H]+	86.5	59.7
190	588.85	3.8	4.53	589 [M+H]+	88.3	46.6
191	516.74	3.7	3.97	517 [M+H]+	93.8	66.o
9	544.79	5.5	4.29	545 [M+H]+	99.0	74.3
192	598.88	3.1	4.91	599 [M+H]+	95.5	51.1
193	588.85	3.5	4.34	589 [M+H]+	89.6	67.2

194	HO, H,	563.71	8.9	2.34	564 [M+H]+	97.2	-17.6
195		467.58	4.7	2.24	468 [M+H]+	93.5	-9.8
196		588.78	7.9	2.48	589 [M+H] ⁺	96.9	-12.9
197	Contraction of the second seco	608.78	5.6	3.07	609 [M+H]+	81.9	18.5
198	HO, , , , , , , , , , , , , , , , , , ,	606.81	5.3	2.74	607 [M+H]*	87.7	23.2
199	C C C C C C C C C C C C C C C C C C C	610.80	8.3	3.02	611 [M+H]+	80.6	-7.5
20 0	C C C C C C C C C C C C C C C C C C C	636.83	7.2	3.18	637 [M+H]+	72.2	21.6
201	Contraction of the second seco	586.77	12.4	2.99	587 [M+H]+	87.5	16.3
202		588.79	9.9	3.16	589 [M+H]⁺	92.3	25.5
203	NH N=	480.61	13.1	2.40	481 [M+H]+	83.6	-11.9
20 4		508.66	10.3	2.68	509 [M+H]⁺	86.6	33.7

205		564.77	6.1	3.31	565 [M+H]⁺	89.1	25.8
20 6		562.75	2.7	2.56	563 [M+H]⁺	87.1	25.8
207		552.71	3.0	2.57	553 [M+H]⁺	84.3	0.4
20 8		431.45	9.1	1.12	432 [M+H]+	98.2	10.4
20 9		678.92	4.3	3.48	679 [M+H]+	73.7	24.0
210	A CONTRACT OF CONT	650.86	3.1	3.24	651 [M+H]⁺	96.8	45.4
211	HO,	648.89	4.5	2.90	649 [M+H]+	98.2	59.3
212		652.88	7.5	3.05	653 [M+H]⁺	98.0	4.8
213	NH NH NH	678.92	5.2	3.24	679 [M+H]+	96.3	4.2
214		628.86	5.0	3.09	629 [M+H]+	97.1	9.3

215	NH ^O CEI NH ^O CEI	630.87	6.9	3.42	631 [M+H]+	85.3	53.4
216	→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→	522.69	9.4	3.30	523 [M+H]+	99.8	6.8
217	o → → → → → → → → → → → → → → → → → → →	550.74	13.6	3.34	551 [M+H]+	92.4	58.1
218		606.85	3.2	3.42	607 [M+H]+	98.6	55-3
219	O C C C C C C C C C C C C C C C C C C C	604.83	5.8	3.30	605 [M+H]*	97.9	70.0
220	OH O OH	594.79	3.7	3.39	595 [M+H]+	94.5	9.1
221		569.66	9.2	1.50	570 [M+H]+	98.2	-13.5
222		618.82	3.2	4.50	619 [M+H]*	88.5	62.2
223		616.84	4.7	3.99	617 [M+H] ⁺	77.2	7.9
224		620.83	2.9	3.95	621 [M+H] ⁺	91.4	46.4
225	C C C C C C C C C C C C C C C C C C C	646.87	4.1	4.57	647 [M+H]*	86.8	22.0

226		622.85	3.4	4.69	623 [M+H]+	85.2	65.4
227		598.83	3.2	4.79	599 [M+H]⁺	96.0	27.9
228		490.64	2.8	3.84	491 [M+H]+	95.4	56.1
229	→→→→→↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓ ↓	518.70	2.9	4.17	519 [M+H]*	97.4	47.8
230	Contraction of the second seco	574.80	3.2	5.29	575 [M+H]⁺	99.8	63.3
231		572.79	2.7	4.64	573 [M+H]+	87.4	33.1
232		537.61	3.7	2.10	538 [M+H]⁺	87.5	0.0
233		441.48	5.8	2.00	442 [M+H]+	96.4	-6.6
234		562.68	3.2	2.26	563 [M+H]+	93.9	5.9
235		688.95	5.2	5.48	689 [M+H]+	87.1	22.5

236	531.61	4.0	2.70	532 [M+H]*	89.9	8.1
237	660.90	3.9	4.90	661 [M+H]*	91.5	70.9
238	662.91	3.3	4.71	663 [M+H]+	88.1	25.4
239	688.95	4.6	4.95	711 [M+Na]+	96.7	38.7
24 0	664.93	4.0	5.14	687 [M+Na] ⁺	94.8	33.6
241	638.89	4.5	4.77	661 [M+Na]+	89.1	15.3
242	640.91	4.8	5.18	641 [M+H]+	87.1	7.3
243	532.72	2.9	4.29	555 [M+Na]+	86.4	-25.9
244	560.78	3.3	4.72	583 [M+Na]+	98.5	67.5
245	616.88	2.9	3.57	617 [M+H] ⁺	94.2	-7.6

246		614.87	3.4	5.08	637 [M+Na]+	92.2	16.4
247		604.83	2.8	4.49	627 [M+Na]+	89.9	67.9
248		579.69	3.7	2.80	580 [M+H]+	87.2	3.0
249		483.57	2.7	2.44	484 [M+H]+	96.3	29.7
250		604.76	4.8	2.66	627 [M+Na] ⁺	91.8	18.4
251	Contraction of the second seco	657.85	15.7	4.23	658 [M+H]*	90.7	45.9
252		655.88	3.2	3.85	656 [M+H]⁺	84.0	54.7
253	G G G H H H H H H H H H H H H H H H H H	685.91	6.9	4.34	686 [M+H]*	98.5	21.8
254	Content of the second s	661.88	4.7	4.42	662 [M+H]*	97.1	32.5
255		635.85	19.0	4.21	636 [M+H]+	88.2	29.4

256		637.86	3.8	4.51	638 [M+H]+	91.9	43.2
257		557.73	4.1	4.00	558 [M+H]⁺	91.4	12.8
258		613.84	3.5	4.88	614 [M+H]*	85.1	24.6
259	o H O OEt	611.82	3.3	4.42	612 [M+H]*	96.0	53.0
26 0		601.72	4.9	2.24	602 [M+H]*	84.2	22.3
261		727.99	4.9	5.00	728 [M+H]+	79.9	17.8
262	Contraction of the second seco	699.93	5.2	4.41	700 [M+H]+	90.9	53.8
263		697.96	8.7	4.15	698 [M+H]⁺	81.4	2.9
264		701.95	3.1	4.25	702 [M+H]+	98.9	45.1
265	OCH NH	727.99	2.7	4.51	750 [M+Na] ⁺	99.6	21.7

2 66		703.97	2.9	4.62	704 [M+H]+	88.3	37.7
267		677.93	20.8	4.60	700 [M+Na] ⁺	96.8	17.7
26 8		679.94	3.2	4.73	702 [M+Na] ⁺	94.9	29.2
26 9		643.87	2.9	4.37	644 [M+H]+	92.4	33.4
270	→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→	571.76	6.9	4.01	572 [M+H]*	92.8	68.2
271	→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→→	599.81	5.7	4.34	600 [M+H]*	96.3	40.8
272	CHARACTER CONTRACTOR	655.92	9.8	5.10	678 [M+Na] ⁺	95.1	41.5
273	O H O OEt O H O OEt O DET O DET	653.91	4.9	4.61	676 [M+Na] ⁺	98.8	32.4
274		643.87	2.9	4.26	644 [M+H]*	89.6	60.1
275		618.73	2.8	2.38	641 [M+Na]+	83.6	6.3

276		522.60	4.1	2.88	523 [M+H]*	96.8	41.2
277		643.80	3.6	2.52	523 [M+H]+	93.3	2.9
278	$\begin{array}{c} \begin{array}{c} \begin{array}{c} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \\ \end{array} \\ \begin{array}{c} \\ \\ \\ \end{array} \\ \begin{array}{c} \\ \\ \\ \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \\ \end{array} $	765.01	4.9	4.17	788 [M+Na] ⁺	96.6	75.6
279		763.03	3.4	3.92	787 [M+Na]+	94.7	59.0
28 0		767.02	4.9	4.35	768 [M+H]+	99.2	52.4
281		793.06	8.1	4.56	816 [M+Na]+	97.4	18.2
282		769.04	5.4	4.69	770 [M+H]*	98.2	34.6
283		743.00	2.8	4.42	765 [M+Na] ⁺	99.2	cell death
284		745.02	5.9	4.75	746 [M+H]+	97.2	65.0

285		636.83	13.1	3.71	637 [M+H]+	97.7	26.4
28 6		664.89	8.3	4.11	665 [M+H]+	99.6	75.6
287		720.99	4.8	5.30	721 [M+H]*	97.8	cell death
288	HNN OF HNN OEL	679.86	7.8	2.90	680 [M+H]+	90.3	46.4
28 9		677.88	3.6	2.58	678 [M+H]+	97.1	24.4
29 0	HN O OCH NOCEL	681.88	4.5	2.91	682 [M+H]*	99.6	9.2
291	HN COEt HN COEt HN COEt HN COEt	707.91	2.9	3.06	708 [M+H]+	99.7	-0.2
292		683.89	3.0	3.06	684 [M+H]+	96.4	-4.6
293	Contraction of the contraction o	657.85	8.4	3.29	658 [M+H]+	98.8	-7.2

294	Correction North Correc	659.87	8.3	3.02	66o [M+H]+	95.5	16.6
295		623.79	5.6	2.72	624 [M+H]+	94.9	25.6
29 6		551.69	7.8	2.23	552 [M+H]+	99.1	18.7
297		579.74	10.3	2.53	580 [M+H]+	95.6	-1.3
29 8		635.85	3.7	3.13	636 [M+H]+	98.0	35.5
29 9		633.83	6.0	2.91	634 [M+H]+	96.8	7.5
30 0		623.79	2.9	2.61	624 [M+H]+	92.2	35.7
301	HN HN HN HN HN HN HN HN HN HN HN HN HN H	721.94	3.7	3.18	722 [M+H]*	96.1	9.4

302	HO HO HO HO HO HO HO HO HO HO HO HO HO H	719.97	5.6	2.79	720 [M+H]*	95.4	21.8
303	HN O OEt N HN	723.96	3.0	3.10	724 [M+H]+	98.9	3.9
304	TZ TZ TZ TZ TZ TZ TZ TZ TZ TZ TZ TZ TZ T	749-99	3.1	3.28	750 [M+H]*	81.6	7.6
305		725.97	2.7	3.29	726 [M+H]*	90.7	6.3
30 6		699.93	3.7	3.11	700 [M+H]*	98.5	16.6
307	HN COET	701.95	4.0	3.25	702 [M+H]*	96.0	42.2
30 8		665.87	4.0	3.02	666 [M+H]+	83.0	23.0
30 9		593.77	4.1	2.51	594 [M+H]+	98.5	-9.1

310	HN COEL HN COEL HN COEL HN COEL	621.82	3.3	2.83	622 [M+H]*	94.0	15.9
311	HN HN HN HN HN HN HN HN HN HN HN HN HN H	677.93	3.0	3.39	678 [M+H]+	99.3	32.4
312		675.91	3.1	3.18	676 [M+H]+	98.0	68.o
313		665.87	3.6	2.88	666 [M+H]+	88.1	21.8

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