Supporting Information

Palladium(0)-Lithium Iodide-Cocatalyzed Asymmetric Hydroalkylation of Conjugated Enynes with Pronucleophiles Leading to 1,3-Disubstituted Allenes

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Experimental Procedures

General Techniques. All commercially available reagents and anhydrous solvents including tetrahydrofuran (THF), dichloromethane (DCM), and xylene were purchased and used without further purification. Anhydrous N, N-dimethylformamide (DMF), methanol, and toluene were obtained by distillation from calcium hydride, magnesium, and sodium, respectively. All reactions were monitored by thin layer chromatography (TLC) performed on 0.25 mm silica gel glassplates (60 F254) using UV light and ethanolic p-anisaldehyde-sulfuric acid, ethanolic molybdatophosphoric acid, aqueous cerium sulfate-hexaammonium heptamolybdate-sulfuric acid, or aqueous potassium permanganate-potassium carbonate-sodium hydroxide solutions as visualizing agents. Flash column chromatography was carried out with silica gel (spherical, neutral, 63-210 µm grade). Preparative thin layer chromatography was performed on 0.75 mm Wakogel® B-5F PLC plates. Yields refer to chromatographically and spectroscopically homogenous materials. Melting points were measured on a melting point apparatus and were uncorrected. Only the strongest and/or structurally important absorptions of infrared (IR) spectra are reported in reciprocal centimeters (cm⁻¹). ¹H NMR spectra (400 MHz and 600 MHz) and ¹³C{¹H}NMR spectra (100 MHz and 151 MHz) were recorded in the indicated solvent. Chemical shifts (δ) are reported in delta (δ) units, parts per million (ppm). Chemical shifts for ¹H NMR spectra are given relative to signals for internal tetramethylsilane (0 ppm) or residual nondeuterated solvents, i.e., chloroform (7.26 ppm). Chemical shifts for ¹³C NMR spectra are given relative to the signal for chloroform-d (77.0 ppm). Chemical shifts for ³¹P NMR spectra are given relative to the signal for external 85% phosphoric acid (0 ppm). Multiplicities are reported by the following abbreviations: s (singlet), d (doublet), t (triplet), g (quartet), m (multiplet), br (broad). Coupling constants (J) are represented in hertz (Hz), ¹H and ¹³C NMR chemical shifts were assigned using a combination of COSY. NOESY. HMQC, and HMBC. Low and high-resolution mass spectra were measured on TOF-MS with EI, FAB, or ESI probe. Analytical HPLC was performed using Daicel Chiralpak Daicel Chiralpak AD-H, OD-H or AS-H. Optical rotations ([a]p) were measured on JASCO P-1010 polarimeter.

Enynes 7a-d were prepared according to the literature procedures.¹

General procedure for optimization of the hydroalkylation of 7a with 5A (Table S1)



Scheme S1. Optimization of the reaction conditions.

To a test tube containing enyne **7a** (1 equiv), (η^3 -allyl)(η^5 -Cp)Pd (10 mol%), and diphosphine ligand (15 mol%) were added anhydrous MeOH (0.25 M) and dimethyl malonate (**5A**, 3 equiv) under argon. The resulting mixture was sealed with a screw cap and stirred at 65 °C for the time described in Table S1. The reaction mixture was cooled to room temperature and concentrated *in vacuo*. NMR yield of **6aA** was determined by ¹H NMR analysis of the crude product using 1,1,2,2-tetrachloroethane as an internal standard. The residue was purified by preparative TLC eluting with 20% EtOAc/hexane to give **6aA**. The enantiomeric ratio (*er*) of **6aA** was determined by HPLC analysis with chiral stationary phase column (Daicel Chiralpak OD-H, hexane/*i*-propanol = 397/3, 0.5 mL/min, λ = 210 nm, $t_R(S)$ = 13.9 min, $t_R(R)$ = 14.7 min). Since it was reported that (*R*)-**6aA** exhibited levorotatory, the retention time of each enantiomer was assigned by that of a dextrorotatory 85:15 mixture of (*S*)- and (*R*)-**6aA** obtained in Table S2, Entry 12.

Table S1. Optimization of the reaction conditions.

Entry	Diphosphine ligand (mg)	(η³-allyl)(η⁵- Cp)Pd (mg) ^[b]	7a (mg)	Time (h)	6aA (%) ^a	Er (S:R)
1	Ph ₂ PCH ₂ PPh ₂ , 7.2	3.3	19.7	19	0	—
2	Ph ₂ P(CH ₂) ₂ PPh ₂ , 7.8	3.0	19.9	19	11	—
3	Ph ₂ P(CH ₂) ₃ PPh ₂ , 8.0	2.5	20.2	19	52	_
4	Ph ₂ P(CH ₂) ₄ PPh ₂ , 7.8	2.8	20.4	9.5	42	_
5	Ph ₂ P(CH ₂) ₆ PPh ₂ , 8.5	2.8	19.9	9.5	59	_
6	Cy ₂ P(CH ₂) ₄ PCy ₂ , 8.8	2.8	19.6	1.5	61	_
7	Xantphos ^b , 10.8	2.8	19.7	24	34	_
8	DPEphos ^c , 10.3	2.8	19.8	24	57	_
9	dppf ^d , 10.0	2.5	20.1	9.5	52	_
10 ^e	dppf ^d , 8.4	2.1	16.5	24	5	_
11	(S)-BINAP, 11.4	2.7	19.8	5	69	45:55
12	(<i>S</i>)-H ₈ -BINAP, 11.6	2.6	20.0	2.5	63	42:58
13	(S)-MeO-BIPHEP, 8.7	2.1	16.5	12	80 (75)	54:46
14	(S)-SEGPHOS, 11.6	2.7	19.8	18.5	74	56:44
15	(S)-Tol-BINAP, 12.8	2.7	19.9	1.5	71	49:51
16	(S)-DTBM-BINAP, 21.8	2.5	19.8	0.67	(90)	36:64
17	(S)-DTBM-MeO-BIPHEP, 21.0	2.6	20.0	0.67	(92)	32:68
18	(S)-DTBM-SEGPHOS, 22.4	2.6	20.0	0.83	(81)	29:71

^a NMR yields were determined by ¹H NMR analysis using 1,1,2,2-tetrachloroethane as an internal standard. Isolated yields are shown in parentheses. ^b 4,5-Bis(diphenylphophino)-9,9-dimethylxanthene. ^c Bis[2-(diphenylphosphino)phenyl] ether. ^d 1,1'-Bis(diphenylphosphino)ferrocene. ^e Reaction in THF instead of MeOH.

General procedure for optimization of the asymmetric hydroalkylation of 7a with 5A (Table 1 and S2)



Scheme S2. Optimization of the asymmetric hydroalkylation of 7a with 5A.

To a test tube containing enyne **7a** (1 equiv), Pd catalyst (X mol%), (S)-DTBM-SEGPHOS (1.5X mol%), and additive (Y mol%) were added anhydrous MeOH (0.25 M) and dimethyl malonate (**5A**, 3 equiv) under argon. The resulting mixture was sealed with a screw cap and stirred at 65 °C (entries 1–8), 50 °C (entries 9–11, 13) or rt (entry 12) for the time described in Table S2. The reaction mixture was cooled to room temperature and concentrated *in vacuo*. NMR yield of **6aA** was determined by ¹H NMR analysis of the crude product using 1,1,2,2-tetrachloroethane as an internal standard. The residue was purified by preparative TLC eluting with 20% EtOAc/hexane to give **6aA**. The enantiomeric ratio (*er*) of **6aA** was determined by HPLC analysis with chiral stationary phase column (Daicel Chiralpak OD-H, hexane/*i*-propanol = 397/3, 0.5 mL/min, λ = 210 nm, $t_R(S)$ = 13.9 min, $t_R(R)$ = 14.7 min).

Table S2. Optimization	of the asymr	netric hydroalky	ylation of 7a	with 5A
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Entry	Pd catalyst (mg, mol%)	Additive (mg, mol%)	7a (mg)	(S)-DTBM-SEGPHOS (mg, mol%)	Temp (°C)	Time (h)	6aA (%) ^a	Er (S:R)
1	(η ³ -allyl)(η ⁵ -Cp)Pd, 2.6, 10	none	20.0	22.4, 15	65	0.83	(81)	29:71
2	Pd ₂ dba ₃ • CHCl ₃ , 6.9, 5	none	19.9	21.4, 15	65	24	(83)	33:67
3	Pd(dba) ₂ , 7.0, 10	none	20.2	21.8, 15	65	0.83	(quant)	28:72
4	(η^3 -allyl)(η^5 -Cp)Pd, 2.1, 10	LiOAc, 2.7, 30	16.5	17.6, 15	65	0.25	94 (78)	28:72
5	(η^3 -allyl)(η^5 -Cp)Pd, 2.2, 10	KOAc, 4.5, 30	16.5	17.6, 15	65	0.25	100 (78)	25:75
6	(η^3 -allyl)(η^5 -Cp)Pd, 2.0, 10	LiCl, 1.7, 30	16.5	17.3, 15	65	0.25	95 (77)	68:32
7	(η ³ -allyl)(η ⁵ -Cp)Pd, 2.1, 10	LiBr, 3.2, 30	16.5	18.2, 15	65	0.25	93 (81)	76:24
8	(η^3 -allyl)(η^5 -Cp)Pd, 2.8, 10	Lil, 5.3, 30	20.2	21.8, 15	65	4.7	(90)	78:22
9	(η^3 -allyl)(η^5 -Cp)Pd, 2.2, 10	Lil, 5.5, 30	16.6	17.5, 15	50	0.67	94	83:17
10	(η³-allyl)(η⁵-Cp)Pd, 1.1, 5	Lil, 4.0, 30	16.5	8.9, 7.5	50	0.5	99	86:14
11	(η³-allyl)(η⁵-Cp)Pd, 1.1, 5	Lil, 2.0, 15	16.3	8.9, 7.5	50	0.5	quant	84:16
12	(η ³ -allyl)(η ⁵ -Cp)Pd, 1.0, 5	Lil, 2.1, 15	16.4	8.9, 7.5	rt	1	quant	86:14
13	(η³-allyl)(η⁵-Cp)Pd, 1.2, 5	Lil, 1.0, 5	16.6	8.9, 7.5	50	0.5	98 (84)	85:15

^a NMR yields were determined by ¹H NMR analysis using 1,1,2,2-tetrachloroethane as an internal standard. Isolated yields are shown in parentheses.

Dimethyl (S)-2-(5-oxododeca-2,3-dien-1-yl)malonate (6aA)^{2,3}



Yellow oil. Rf = 0.49 (17% EtOAc/hexane). $[\alpha]^{22}_{D}$ +43 (*c* 0.76 in hexane) (Table 2, Entry 11). IR (neat): 2954, 2926, 2855, 1964, 1756, 1739, 1436, 1262, 1231, 1152, 1042 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 5.20–5.04 (m, 2H), 3.74 (s, 6H), 3.51 (t, *J* = 7.6 Hz, 1H), 2.58 (ddd, *J* = 7.6, 6.1, 3.2 Hz, 2H), 1.95 (ddt, *J* = 3.0, 6.8, 6.8 Hz, 2H), 1.46–1.15 (m, 12H), 0.88 (d, *J* = 6.9 Hz, 3H). ¹³C-NMR (100 MHz, CDCl₃): δ 203.9, 169.4, 169.3, 93.0, 87.3, 52.5, 51.2, 31.8, 29.4, 29.2, 29.1, 29.0, 28.8, 28.0, 22.6, 14.0 (one carbon is missing due to overlap). LRMS m/z (relative intensity) 296 [M]⁺ (6), 233 (7), 198 (41), 166 (33), 138 (100). HRMS (EI) calcd for C₁₇H₂₈O₄ 296.1988, found 296.1982 [M]⁺.

HPLC chromatograms of 6aA



6aA (Table 2, Entry 11)



General procedure for scope of conjugated enynes 7b-d and pronucleophiles 5A-G (Figure 1)



Scheme S3. The scope of conjugated enynes 7b-d and pronucleophiles 5A-G investigated in the hydroalkylation reaction.

To a test tube containing enyne **7b–d** (1 equiv), (η^3 -allyl)(η^5 -Cp)Pd (5 mol%), (S)-DTBM-SEGPHOS (7.5 mol%) and Lil (5 mol%) were added anhydrous MeOH (0.25 M) and pronucleophile (**5A–G**, 3 equiv) under argon. The resulting mixture was sealed with a screw cap and stirred at the temperature and for the time described in Table S3. The reaction mixture was cooled to room temperature and concentrated *in vacuo*. The residue was purified by preparative TLC or silica gel column chromatography to give **6(b–d)(A–G)**. The authentic racemic mixture was obtained by the similar procedure using *rac*-MeO-BIPHEP instead of (S)-DTBM-SEGPHOS without Lil. The absolute configuration of **6(b–d)(A–G)** were determined by the sign of the optical rotation of literature known compounds or deduced by the Lowe-Brewster rule.^[4]

Table S3. The scope of	f conjugated enynes	7b-d and pronucleophiles 5A-	 G investigated in the 	hydroalkylation reaction.
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Entry	7 (mg)	5 (mg or µL)	(η³-allyl)(η⁵- Cp)Pd (mg)	(S)-DTBM-SEG- PHOS (mg)	Lil (mg)	Temp. (°C) Time (h)	Isolated yield of 6 (mg, %)	Er (S:R)
1	7b , 13.2	5Α , 34.5 μL	1.1	8.7	0.8	50 °C, 5 h	6bA: 26.5, quant	86:14
2	7c , 12.6	5Α , 34.5 μL	1.1	9.1	1.0	rt , 19 h	6cA: 20.9, 82	96:4
3	7d , 10.9 (66% purity) ^a	5Α , 34.5 μL	1.1	9.0	1.8	80 °C, 8 h	6dA : 15.2, 96	98:2
4	7d , 10.8 (81% purity) ^a	5B , 45.3 mg	1.1	9.3	0.8	80 °C, 6 h	6bB : 11.0, 54	94:6
5	7d , 10.8 (81% purity) ^a	5C , 64.2 mg	1.1	9.0	0.8	80 °C, 6 h	6dC: 16.7, 65	96:4
6	7d , 10.8 (81% purity) ^a	5D , 56.5 mg	1.0	9.3	1.0	80 °C, 6 h	6dD : 16.0, 65	95:5
7	7b , 13.2	5E , 90.0 mg	1.0	9.0	0.6	50 °C, 24 h	6bE : 39.7, 94	88:12
8	7d , 11.0 (81% purity) ^a	5E , 88.6 mg	1.3	9.2	1.2	80 °C, 3 h	6dE : 16.8, 51	94:6
9	7b , 13.3	5F , 30.8 μL	1.2	9.0	1.2	50 °C, 2 h; 65 °C, 11 h; 80 °C, 8 h	6bF : 11.3, 49	78:22
10	7b , 13.2	5G , 18.9 μL	1.1	9.0	1.4	50 °C, 24 h; 65 °C, 8 h	6bG : 5.1, 26	75:25

^a THF was contained.

Dimethyl (S)-2-(4-cyclohexylbuta-2,3-dien-1-yl)malonate (6bA)⁵

Isolated by preparative TLC eluting with 20% EtOAc/hexane. Yellow oil. Rf = 0.29 (17% EtOAc/hexane). $[\alpha]^{21}_{D}$ +50 (*c* 1.04 in CHCl₃). IR (neat): 2925, 2850, 1962, 1755, 1738, 1435, 1342, 1250, 1154, 1036 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 5.19–5.09

(m, 2H), 3.74 (s, 6H), 3.51 (t, J = 7.4 Hz, 1H), 2.62–2.54 (m, 2H), 1.98–1.85 (m, 1H), 1.80–1.50 (m, 4H), 1.45–0.96 (m, 6H).¹³C-NMR (100 MHz, CDCl₃): δ 202.8, 169.4, 169.3, 99.0, 88.3, 52.50, 52.48, 51.2, 37.2, 32.9, 32.8, 28.1, 26.1, 26.0. LRMS m/z (relative intensity) 266 [M]⁺ (15), 203 (26), 174 (15), 147 (27), 134 (100). HRMS (EI) calcd for C₁₅H₂₂O₄ 266.1518, found 266.1506 [M]⁺.

Chiral HPLC: (Daicel Chiralpak OD-H, hexane/*i*-propanol = 397/3, 0.5 mL/min, λ = 210 nm): $t_R(S)$ = 18.5 min (major enantiomer), $t_R(R)$ = 20.2 min (minor enantiomer).

HPLC chromatograms of 6bA





6bA

Dimethyl (S)-2-(4-phenylbuta-2,3-dien-1-yl)malonate (6cA)³



Isolated by preparative TLC eluting with 20% EtOAc/hexane. Yellow oil. Rf = 0.34 (17% EtOAc/hexane). $[\alpha]^{22}_{D}$ +118 (*c* 0.97 in CHCl₃). IR (neat): 2954, 2360, 2341, 1952, 1736, 1437, 1235, 1155, 1029, 695 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 7.34–7.15 (m, 5H), 6.20 (dt, *J* = 6.4, 3.2 Hz, 1H), 5.62 (dt, *J* = 6.4, 6.4 Hz, 1H), 3.73 (s, 3H), 3.59 (t, *J* = 7.4 Hz, 1H), 3.58 (s, 3H), 2.84–2.62 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃): δ 205.1, 169.3, 169.1, 134.0, 128.5, 127.1, 126.8, 96.5, 91.7, 52.6, 52.5, 50.9, 27.8. LRMS m/z (relative intensity) 260 [M]⁺ (42), 201 (14), 169 (23), 160 (48), 128 (100). HRMS (EI) calcd for C₁₅H₁₆O₄ 260.1049, found 260.1051 [M]⁺.

Chiral HPLC: (Daicel Chiralpak AD-H, hexane/*i*-propanol = 50/1, 1.0 mL/min, λ = 254 nm): $t_R(S)$ = 30.9 min (major enantiomer), $t_R(R)$ = 42.8 min (minor enantiomer).

HPLC chromatograms of 6cA





Dimethyl (S)-2-(5,5-dimethylhexa-2,3-dien-1-yl)malonate (6dA)^{6,7}



Isolated by silica gel column chromatography eluting with 3% EtOAc/hexane. Colorless oil. Rf = 0.40 (17% EtOAc/hexane). $[\alpha]^{26}_{D}$ +68.6 (*c* 0.51 in CHCl₃). IR (neat): 2959, 1964, 1737, 1457, 1246, 1201, 1111, 877 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 5.21–5.11 (m, 2H), 3.74 (s, 3H), 3.74 (s, 3H), 3.49 (t, *J* = 7.4 Hz, 1H), 2.60 (ddd, *J* = 7.4, 5.9, 3.2 Hz, 2H), 1.01 (s, 9H). ¹³C-NMR (100 MHz, CDCl₃): δ 201.2, 169.41, 169.37, 104.9, 89.2, 52.6, 52.5, 51.3, 31.7, 30.0, 28.3. LRMS m/z (relative intensity) 240 [M]⁺ (52), 225 (7), 177 (32), 165 (18), 132 (100). HRMS (EI) calcd for C₁₃H₂₀O₄ 240.1362, found 240.1372 [M]⁺.

Chiral HPLC: (Daicel Chiralpak OD-H, hexane/*i*-propanol = 397/3, 0.5 mL/min, λ = 210 nm): $t_R(S)$ = 15.3 min (major enantiomer), $t_R(R)$ = 16.8 min (minor enantiomer).

HPLC chromatograms of 6dA





Dimethyl (S)-2-(5,5-dimethylhexa-2,3-dien-1-yl)-2-methylmalonate (6dB)⁶⁻⁸



Isolated by silica gel column chromatography eluting with 3% EtOAc/hexane. Yellow oil. Rf = 0.55 (17% EtOAc/hexane). $[\alpha]^{18}$ D +30.3 (*c* 0.23 in CHCl₃). IR (neat): 2959, 1964, 1737, 1457, 1246, 1201, 1111, 877 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 5.10 (dt, *J* = 6.3, 2.4 Hz, 1H), 5.00 (dt, *J* = 6.3, 7.6 Hz, 1H), 3.72 (s, 6H), 2.61–2.56 (m, 2H), 1.45 (s, 3H), 1.02 (s, 9H). ¹³C-NMR (100 MHz, CDCl₃): δ 203.2, 172.4, 172.3, 102.9, 86.9, 53.8, 52.52, 52.47, 36.3, 31.7, 30.1, 19.9. HRMS (ESI) calcd for C₁₄H₂₃O₄ 255.1596, found 255.1587 [M+H]⁺.

Chiral HPLC: (Daicel Chiralpak AD-H, hexane/*i*-propanol = 397/3, 0.5 mL/min, λ = 210 nm): $t_R(R)$ = 11.4 min (minor enantiomer), $t_R(S)$ = 12.1 min (major enantiomer).

HPLC chromatograms of 6dB





Dimethyl (S)-2-(5,5-dimethylhexa-2,3-dien-1-yl)-2-phenylmalonate (6dC)⁶



Isolated by preparative TLC eluting with 20% EtOAc/hexane. Yellow oil. Rf = 0.47 (17% EtOAc/hexane). $[\alpha]^{19}{}_{D}$ +40.8 (*c* 0.79 in CHCl₃). IR (neat): 2956, 2864, 1961, 1737, 1499, 1435, 1362, 1233, 1175, 1074, 878, 730, 696 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 7.46–7.37 (m, 2H), 7.37–7.23 (m, 3H), 5.12–4.99 (m, 2H), 3.75 (s, 6H), 3.07–3.03 (m, 2H), 0.96 (s, 9H). ¹³C-NMR (100 MHz, CDCl₃): δ 202.9, 170.8, 170.7, 136.5, 128.2, 128.1, 127.6, 102.9, 87.4, 62.9, 52.7, 36.4, 31.6, 30.1 (one carbon is missing due to overlap). HRMS (ESI) calcd for C₁₉H₂₅O₄ 317.1753, found 317.1741 [M+H]⁺.

Chiral HPLC: (Daicel Chiralpak AD-H, hexane/*i*-propanol = 397/3, 0.5 mL/min, λ = 210 nm): $t_{R}(R)$ = 23.2 min (minor enantiomer), $t_{R}(S)$ = 27.1 min (major enantiomer).

HPLC chromatograms of 6dC





Dimethyl (S)-2-acetamido-2-(5,5-dimethylhexa-2,3-dien-1-yl)malonate (6dD)



Isolated by silica gel column chromatography eluting with 33% EtOAc/hexane. Colorless oil. Rf = 0.41 (50% EtOAc/hexane). $[\alpha]^{20}_{D}$ +24.1 (*c* 0.66 in CHCl₃). IR (neat): 3297, 2959, 1961, 1748, 1683, 1506, 1437, 1305, 1214, 1065, 979, 886 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 6.78 (s, 1H), 5.52 (dt, *J* = 7.6, 2.2 Hz, 1H), 4.89 (dt, *J* = 7.6, 7.6 Hz, 1H), 3.78 (s, 6H), 3.03 (dd, *J* = 7.6, 2.2 Hz, 2H), 2.03 (s, 3H), 1.00 (s, 9H). ¹³C-NMR (100 MHz, CDCl₃): δ 203.2, 169.0, 168.2, 168.1, 103.2, 85.5, 66.3, 53.5, 53.4, 33.2, 31.5, 30.0, 23.0. HRMS (ESI) calcd for C₁₅H₂₄NO₅ 298.1654, found 298.1642 [M+H]⁺.

Chiral HPLC: (Daicel Chiralpak AD-H, hexane/*i*-propanol = 19/1, 0.5 mL/min, λ = 210 nm): $t_{R}(R)$ = 29.9 min (minor enantiomer), $t_{R}(S)$ = 32.8 min (major enantiomer).

HPLC chromatograms of 6dD





(S)-(5-Cyclohexylpenta-3,4-diene-1,1-diyldisulfonyl)dibenzene (6bE)



Isolated by preparative TLC eluting with 40% EtOAc/hexane. Yellow oil. Rf = 0.13 (17% EtOAc/hexane). [α]²⁵_D +71.8 (*c* 1.80 in CHCl₃). IR (neat): 3065, 2924, 2850, 2358, 2343, 1961, 1583, 1447, 1331, 1312, 1156, 1079, 1023, 999, 892, 738, 724, 687 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 8.01–7.89 (m, 4H), 7.75–7.65 (m, 2H), 7.65–7.52 (m, 4H), 5.19 (ddt, *J* = 6.2, 3.0, 6.2 Hz, 1H), 5.12 (ddt, *J* = 6.2, 6.2, 3.0 Hz, 1H), 4.57 (t, *J* = 6.2 Hz, 1H), 2.88 (dddd, *J* = 6.2, 6.2, 3.0, 1.0 Hz, 2H), 1.96–1.83 (m, 1H), 1.74–1.58 (m, 4H), 1.36–1.15 (m, 4H), 0.99–0.96 (m, 2H). ¹³C-NMR (100 MHz, CDCl₃): δ 202.7, 138.2, 138.0, 134.6, 134.5, 129.6, 129.5, 129.1, 100.5, 87.9, 83.0, 37.0, 32.92, 32.89, 25.95, 25.87, 25.86, 25.3. HRMS (ESI) calcd for C₂₃H₂₇O₄S₂ 431.1351, found 431.1334 [M+H]⁺.

Chiral HPLC: (Daicel Chiralpak AD-H, hexane/*i*-propanol = 9/1, 0.5 mL/min, λ = 254 nm): $t_R(R)$ = 47.0 min (minor enantiomer), $t_R(S)$ = 77.2 min (major enantiomer).

HPLC chromatograms of 6bE





(S)-(6,6-Dimethylhepta-3,4-diene-1,1-diyldisulfonyl)dibenzene (6dE)

Isolated by preparative TLC eluting with 50% EtOAc/hexane. Yellow oil. Rf = 0.17 (17% EtOAc/hexane). $[\alpha]^{24}_{D}$ +74.8 (c 0.75 in CHCl₃). IR (neat): 3066, 2959, 2866, 1962, 1584, 1447, 1332, 1156, 1079, 743, 724, 687 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 8.02–7.88 (m, 4H), 7.75–7.64 (m, 2H), 7.64–7.55 (m, 4H), 5.23 (dt, *J* = 6.1, 6.1 Hz, 1H), 5.14 (dt, *J* = 6.1, 3.0 Hz, 1H), 4.55 (t, *J* = 6.1 Hz, 1H), 2.88 (ddd, J = 6.1, 6.1, 3.0 Hz, 2H), 0.97 (s, 9H). ¹³C-NMR (100 MHz, CDCl₃): δ 201.2, 139.0, 138.3, 137.9, 134.6, 134.5, 129.7, 129.5, 129.1, 106.4, 88.9, 83.1, 31.9, 30.0, 25.5. HRMS (ESI) calcd for C₂₁H₂₅O₄S₂ 405.1194, found 405.1180 [M+H]⁺.

Chiral HPLC: (Daicel Chiralpak OD-H, hexane/*i*-propanol = 9/1, 0.5 mL/min, λ = 254 nm): $t_R(S)$ = 31.6 min (major enantiomer), $t_R(R)$ = 35.2 min (minor enantiomer).

HPLC chromatograms of 6dE





(S)-3-(4-Cyclohexylbuta-2,3-dien-1-yl)pentane-2,4-dione (6bF)



Isolated by silica gel column chromatography eluting with 3% EtOAc/hexane. Yellow oil. Rf = 0.50 (17% EtOAc/hexane). $[\alpha]^{24}$ D +81.4 (c 0.57 in CHCl₃). IR (neat): 2925, 2851, 1961, 1755, 1738, 1437, 1341, 1258, 1231, 1152, 1034, 891 cm⁻¹. ¹H-NMR (400

MHz, CDCl₃): δ 5.24–5.02 (m, 2H), 2.92 (t, *J* = 4.5 Hz, 1H), 2.58–2.47 (m, 1H), 2.33–2.02 (m, 7H), 2.02–1.83 (m, 1H), 1.83–1.51 (m, 4H), 1.36–0.91 (m, 6H). ¹³C-NMR (100 MHz, CDCl₃): δ 203.8, 203.7, 202.9, 202.8, 191.3, 99.33, 99.25, 90.5, 88.5, 67.9, 37.4, 37.2, 33.1, 33.03, 33.00, 32.8, 29.2, 29.1, 27.3, 27.0, 26.14, 26.08, 26.07, 26.01, 25.99, 25.96, 25.94, 22.8 (as a tautomeric mixture of the keto and enol forms). LRMS m/z (relative intensity) 234 [M]⁺ (9), 191 (81), 134 (47), 121 (100). HRMS (EI) calcd for C₁₅H₂₂O₂ 234.1620, found 234.1615 [M]⁺.

Chiral HPLC: (Daicel Chiralpak AD-H, hexane/*i*-propanol =397/3, 0.5 mL/min, λ = 190 nm): $t_{R}(R)$ = 11.2 min (minor enantiomer), $t_{R}(S)$ = 12.0 min (minor enantiomer).

HPLC chromatograms of 6bF





S17

(S)-2-(4-Cyclohexylbuta-2,3-dien-1-yl)malononitrile (6bG)9



Isolated by silica gel column chromatography eluting with 3% EtOAc/hexane. Yellow solid. Mp = 48.3–49.9 °C. Rf = 0.50 (17% EtOAc/hexane). [α]²⁵_D +45.3 (*c* 0.28 in CHCl₃). IR (neat): 2925, 2851, 2256, 1963, 1962, 1725, 1448, 1260, 891, 868, 722 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 5.40 (ddt, *J* = 6.4, 6.4, 3.1 Hz, 1H), 5.23 (ddt, *J* = 3.0, 6.4, 6.4 Hz, 1H), 3.77 (t, *J* = 7.0 Hz, 1H), 2.73–2.64 (m, 2H), 2.13–1.99 (m, 1H), 1.85–1.60 (m, 4H), 1.38–1.02 (m, 6H). ¹³C-NMR (100 MHz, CDCl₃): δ 203.8, 112.38, 112.35, 101.5, 85.0, 37.1, 32.9, 32.8, 30.5, 26.0, 25.9, 22.4. LRMS m/z (relative intensity) 200 [M]⁺ (10), 135 (100), 93 (85), 81 (86). HRMS (EI) calcd for C₁₃H₁₆N₂ 200.1313, found 200.1311 [M]⁺.

Chiral HPLC: (Daicel Chiralpak OD-H, hexane/*i*-propanol = 50/1, 1.0 mL/min, λ = 214 nm): $t_R(S)$ = 18.3 min (major enantiomer), $t_R(R)$ = 20.4 min (minor enantiomer).

HPLC chromatograms of 6bG





Preparation of 7b-d

Preparation of (E)-(4-cyclohexylbut-1-en-3-yn-1-yl)trimethylsilane



Scheme S4. Preparation of (E)-(4-cyclohexylbut-1-en-3-yn-1-yl)trimethylsilane.

To a mixture of ethynylcyclohexane (4.13 g, 38.2 mmol), Pd(PPh₃)₄ (1.02 g, 0.882 mmol), and Cul (336 mg, 1.76 mmol) in anhydrous THF (29 mL) were added piperidine (14.5 mL, 147 mmol) and a solution of alkenyl iodide¹⁰ (6.64 g, 29.4 mmol) in anhydrous THF (30 mL) at 0 °C under argon. The mixture was allowed to warm up to room temperature, stirred at 50 °C for 12 h, and then treated with water. The aqueous layer was extracted with Et₂O three times. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated *in vacuo*. The residue was diluted with hexane, filtered through a pad of SiO₂ and concentrated *in vacuo* to give (*E*)-(4-cyclohexylbut-1-en-3-yn-1-yl)trimethylsilane (5.62 g, 93%) as a red-black oil. The oil was pure enough for analysis.

Rf = 0.51 (hexane). IR (neat): 2931, 2854, 2359, 2342, 2208, 1574, 1448, 1249, 975, 865, 839 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 6.32 (d, *J* = 19.3 Hz, 1H), 5.96 (dd, *J* = 19.3, 1.9 Hz, 1H), 2.53–2.43 (m, 1H), 1.87–1.76 (m, 2H), 1.76–1.64 (m, 2H), 1.64–1.21 (m, 6H), 0.07 (s, 9H). ¹³C-NMR (100 MHz, CDCl₃): δ 143.4, 124.1, 95.2, 80.9, 32.7, 29.7, 25.9, 24.9, –1.6. LRMS m/z (relative intensity) 206 [M]⁺ (24), 191 (100), 125 (44), 109 (28). HRMS (EI) calcd for C₁₃H₂₂Si 206.1491, found 206.1517 [M]⁺.

Preparation of (E)-(but-3-en-1-yn-1-yl-4-d)cyclohexane (7b-d)



Scheme S5. Preparation of (E)-(but-3-en-1-yn-1-yl-4-d)cyclohexane (7b-d).

To a solution of (*E*)-(4-cyclohexylbut-1-en-3-yn-1-yl)trimethylsilane (5.62 g, 27.2 mmol) in anhydrous DCM (109 mL) were added D_2O (2.45 mL, 136 mmol) and TFAA (18.9 mL, 136 mmol) at 0 °C under argon.¹¹ The mixture was allowed to warm up to room temperature, stirred for 7 h, and then treated with saturated aqueous NaHCO₃ slowly at 0 °C. The aqueous layer was extracted with DCM three times. The combined organic layers were dried over MgSO₄ and concentrated *in vacuo*. The residue was diluted with hexane and filtered through a pad of SiO₂ (thoroughly washed with hexane). The filtrate was concentrated *in vacuo* and then purified by bulb-to-bulb distillation (120 °C/40 mmHg) to give **7b**-*d* (2.38 g, 65%) as a pale yellow oil.

Rf = 0.56 (hexane).¹H-NMR (400 MHz, CDCl₃): δ 5.79 (d, *J* = 17.4 Hz, 1H), 5.52 (d, *J* = 17.4 Hz, 1H), 5.36 (dd, *J* = 10.9, 2.0 Hz, 0.08 H), 2.57–2.34 (m, 1H), 1.92–1.61 (m, 4H), 1.61–1.06 (m, 6H). ¹³C-NMR (100 MHz, CDCl₃): δ 124.9 (t, *J* = 24.5 Hz), 117.6, 95.1, 79.2, 32.6, 29.6, 25.9, 24.9 (additional small peak at 125.2 ppm due to undeuterated olefinic carbon). LRMS m/z (relative intensity) 135 [M]⁺ (34), 120 (16), 106 (35), 92 (100). HRMS (EI) calcd for C₁₀H₁₃D 135.1158, found 135.1159 [M]⁺.

Preparation of 8

Preparation of tetraethyl [1,1'-biphenyl]-2,2'-diylbis(phosphonate)



Scheme S6. Preparation of tetraethyl [1,1'-biphenyl]-2,2'-diylbis(phosphonate).

To a 200 mL two-necked flask containing Pd(OAc)₂ (397 mg, 1.77 mmol), dppf (1.07 g, 1.93 mmol), and KOAc (172 mg, 1.75 mmol) was added anhydrous DMF (9 mL) under argon. The resulting mixture was stirred at 60 °C for 30 min and cooled to room temperature. Then the mixture was treated with DIPEA (7.80 mL, 45.6 mmol) and HPO(OEt)₂ (5.45 mL, 42.2 mmol) and stirred at room temperature for 15 min.¹² A solution of [1,1'-biphenyl]-2,2'-diyl bis(trifluoromethanesulfonate)¹³ (7.92 g, 17.6 mmol) in anhydrous DMF (30 mL + 6 mL) was added to the mixture. The mixture was stirred at 100 °C for 15 h and cooled to room temperature. Then, the resulting mixture was diluted with EtOAc (180 mL) and filtered through a pad of Celite (thoroughly washed with EtOAc). The filtrate was washed with water (90 mL x 1, 45 mL x 1) and brine (45 mL), dried over MgSO₄, and concentrated *in vacuo*. The residue was purified by silica gel column chromatography eluting with 10% EtOH/EtOAc to give tetraethyl [1,1'-biphenyl]-2,2'-diylbis(phosphonate) (5.82 g, 78%) as a black solid. The analytical sample was obtained as a pale yellow oil after purification using silica gel column chromatography eluting with 3% MeOH/CHCl₃.



Rf = 0.64 (10% CHCl₃/MeOH). IR (neat): 3670–3120 (br), 2983, 1660, 1589, 1430, 1392, 1231, 1141, 1055, 1025, 971, 765 cm⁻¹. ¹H NMR (400 MHz, CDCl₃): δ 8.10–7.97 (m, 2H), 7.52 (t, J = 7.3 Hz, 2H), 7.50–7.39 (m, 4H), 4.08–3.87 (m, 6H), 3.85–3.72 (m, 1H), 1.15 (t, J = 7.2 Hz, 6H), 1.10 (t, J = 7.2 Hz, 6H). ¹³C NMR (151 MHz, CDCl₃): δ 144.0 (dd, J = 9.2, 4.2 Hz), 133.0 (d, J = 9.8 Hz), 131.7 (d, J = 13.7 Hz), 130.8 (d, J = 2.9 Hz), 127.7 (d, J = 190 Hz), 127.2 (d, J = 14.8 Hz), 61.8 (d, J = 6.1 Hz), 61.6 (dd, J = 6.1 Hz), 16.2 (d, J = 6.5 Hz), 16.1 (dd, J = 6.5 Hz). ³¹P-NMR (243 MHz, CDCl₃): δ 18.0. HRMS (ESI) calcd for C₂₀H₂₈NaO₆P₂ 449.1253, found 449.1247 (M+Na)⁺.

Preparation of [1,1'-biphenyl]-2,2'-diylbis(bis(3,5-di-tert-butyl-4-methoxyphenyl)phosphine oxide)



Scheme S7. Preparation of [1,1'-biphenyl]-2,2'-diylbis(bis(3,5-di-tert-butyl-4-methoxyphenyl)phosphine oxide).

To a 50 mL recovery flask containing tetraethyl [1,1'-biphenyl]-2,2'-diylbis(phosphonate) (2.03 g, 4.76 mmol) were added SOCl₂ (5.2 mL, 72 mmol) and anhydrous DMF (0.55 mL, 7.1 mmol) under argon. The resulting mixture was stirred at 100 °C for 24 h and cooled to room temperature. Then the mixture was concentrated *in vacuo* and evaporated with toluene (10 mL x 3) azeotropically to give a crude [1,1'-biphenyl]-2,2'-diylbis(phosphonic dichloride), which was used for the next reaction without further purification.

0.63 M Grignard solution in THF prepared from Mg (1.15 g, 47.3 matom), bromoarene¹⁴ (11.4 g, 38.1 mmol), anhydrous THF 60 mL, and 1,2-dibromoethene (0.33 mL, 3.8 mmol) was treated with a solution of the crude [1,1'-biphenyl]-2,2'-diylbis(phosphonic dichloride) in THF (20 mL + 2 mL) at -78 °C. The resulting mixture was allowed to warm up to room temperature over 12 h. The mixture was cooled to -78 °C and treated with sat. aq. NH₄Cl (80 mL). The aqueous layer was extracted with EtOAc two times. The combined organic layers were washed with brine (40 mL), dried over MgSO4, and concentrated in vacuo. The residue was purified silica column chromatography eluting 16% EtOAc/hexane bv ael with to give [1,1'-biphenyl]-2,2'-diylbis(bis(3,5-di-tert-butyl-4-methoxyphenyl)phosphine oxide) (2.22 g, 41%) as a white solid.



Mp = 107–108 °C. Rf = 0.40 (33% EtOAc/hexane). IR (neat): 3398 (br), 2964, 2871, 1585, 1448, 1411, 1395, 1362, 1265, 1200, 1148, 1118, 1011, 887, 811, 793, 761, 714, 617 cm⁻¹. ¹H-NMR (600 MHz, CDCl₃): δ 7.70 (d, J = 12.4 Hz, 4H), 7.42 (d, J = 12.0 Hz, 4H), 7.28–7.22 (m, 2H), 7.20 (dd, J = 7.2, 7.6 Hz, 2H), 7.12 (dd, J = 7.6, 7.6 Hz, 4H), 6.80–6.74 (m, 2H), 3.69 (s, 6H), 3.68 (s, 6H), 1.35 (s, 36H), 1.33 (s, 36H). ¹³C-NMR (151 MHz, CDCl₃): δ 161.92 (d, J = 3.1 Hz), 161.89 (d, J = 3.3 Hz), 145.7 (dd, J = 6.6, 3.7 Hz), 143.2 (d, J = 11.9 Hz), 143.1 (d, J = 12.3 Hz), 132.6 (d, J = 102.2 Hz), 132.4 (d, J = 12.3 Hz), 131.9 (d, J = 11.6 Hz), 131.0 (d, J = 9.8 Hz), 130.6 (d, J = 10.8 Hz), 129.19 (d, J = 104.9 Hz), 129.18 (d, J = 2.3 Hz), 128.1 (d, J = 108.2 Hz), 126.1 (d, J = 12.5 Hz), 64.4, 64.2, 35.93, 35.87, 31.99, 31.97. ³¹P-NMR (243 MHz, CDCl₃): δ 27.4. HRMS (ESI) calcd for C₇₂H₁₀₁O₆P₂ 1123.7068, found 1123.7035 [M+H]⁺.

Preparation of 2,2'-bis(bis(3,5-di-tert-butyl-4-methoxyphenyl)phosphaneyl)-1,1'-biphenyl (8)



Scheme S8. Preparation of 2,2'-bis(bis(3,5-di-tert-butyl-4-methoxyphenyl)phosphaneyl)-1,1'-biphenyl (8).

To a 50 mL recovery flask containing [1,1'-biphenyl]-2,2'-diylbis(bis(3,5-di-*tert*-butyl-4-methoxyphenyl)phosphine oxide) (1.12 g, 0.997 mmol) were added anhydrous xylene (10 mL) and tributylamine (2.4 mL, 10 mmol) under argon. The resulting mixture was stirred at 0 °C and treated with HSiCl₃ (1.0 mL, 9.9 mmol).¹⁵ The mixture was stirred at 0 °C for 15 min, refluxed for 8 h, cooled to

0 °C, and treated with 7.5 M aqueous NaOH (15 mL). The immiscible mixture was stirred at 60 °C for 2 h, cooled to room temperature, and extracted with toluene. The organic phase was washed with water (10 mL), 1 M aqueous HCI (15 mL), water (10 mL), and brine (10 mL), dried over MgSO₄, and concentrated *in vacuo*. The residue was purified by silica gel column chromatography eluting with 4% EtOAc/hexane to give **8** (894 mg, 82%) as a white solid. The analytical sample was obtained by recrystallization from MeOH.

PAr₂ Ar =
$$C_6H_2$$
-3,5-
PAr₂ di-^tBu-4-OMe

Mp = 116–117 °C. Rf = 0.50 (10% EtOAc/hexane). IR (neat): 2962, 2870, 1578, 1447, 1409, 1361, 1263, 1223, 1140, 1117, 885, 754 cm⁻¹. ¹H-NMR (600 MHz, CDCl₃): δ 7.31 (dd, *J* = 3.6, 3.6 Hz, 4H), 7.19 (ddd, *J* = 7.6, 7.6, 1.4 Hz, 2H), 7.06 (dd, *J* = 7.6, 1.0 Hz, 2H), 7.03 (ddd, *J* = 7.2, 7.6, 1.0 Hz, 2H), 7.00 (dd, *J* = 3.6, 3.6 Hz, 4H), 6.66 (d, *J* = 7.2 Hz, 2H), 3.68 (s, 6H), 3.63 (s, 6H), 1.32 (s, 36H), 1.29 (s, 36H). ¹³C-NMR (151 MHz, CDCl₃): δ 159.9, 159.4, 147.1 (t, *J* = 17.5 Hz), 142.8 (t, *J* = 3.1 Hz), 142.6 (t, *J* = 3.8 Hz), 139.0 (dd, *J* = 6.1, 5.8 Hz), 133.0, 132.8 (t, *J* = 11.5 Hz), 132.3 (t, *J* = 11.5 Hz), 132.0 (dd, *J* = 6.8, 6.5 Hz), 131.4 (dd, *J* = 6.3, 5.0 Hz), 130.3 (t, *J* = 3.7 Hz), 64.2, 64.1, 35.8, 35.7, 32.2, 32.0. ³¹P-NMR (243 MHz, CDCl₃): δ -13.9. HRMS (ESI) calcd for C₇₂H₁₀₁O₄P₂ 1092.7203, found 1092.7174 [M+H]⁺.

Hydroalkylation of 7b-d with 5A under the catalysis of Pd-achiral diphosphine 8 (Scheme 2)



Scheme S9. Hydroalkylation of 7b-d with 5A under the catalysis of Pd-achiral diphosphine 8.

To a test tube containing enyne **7b**-*d* (84.9 mg, 0.628 mmol), (η^3 -allyl)(η^5 -Cp)Pd (6.7 mg, 0.031 mmol), and diphosphine **8** (51.4 mg, 0.0471 mmol) were added anhydrous MeOH (2.5 mL) and dimethyl malonate (216 µL, 1.88 mmol) under argon. The resulting mixture was sealed with a screw cap and stirred at 50 °C for 3 h. The reaction mixture was cooled to room temperature and concentrated *in vacuo*. The residue was purified by silica gel column chromatography eluting with 3% EtOAc/hexane and preparative TLC eluting with 20% EtOAc/hexane to give **6bA**-*d* (128 mg, 76%) as a pale yellow oil. The diastereomeric ratio and stereochemistry of **6bA**-*d* were determined using NMR analysis of γ -lactone **10**-*d* prepared according to the following procedure.

Dimethyl 2-(4-cyclohexylbuta-2,3-dien-1-yl-1-d)malonate (6bA-d) as a 73:27 diastereomeric mixture

Rf = 0.29 (17% EtOAc/hexane). IR (neat): 2925, 2850, 1962, 1737, 1435, 1250, 1153, 1030, 891 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 5.15–5.12 (m, 2H), 3.740 (s, 3H), 3.737 (s, 3H), 3.50 (d, *J* = 7.3 Hz, 1H), 2.62–2.51 (m, 1H), 1.99–1.86 (m, 1H), 1.79–1.58 (m, 4H), 1.34–0.96 (m, 6H). ¹³C-NMR (100 MHz, CDCl₃): δ 202.8, 169.4, 169.3, 98.9, 88.2, 52.5, 52.4, 51.2, 37.1, 32.9, 32.8, 27.8 (t, *J* = 20.2 Hz), 26.1, 26.0 (additional small peak at 28.1 ppm due to undeuterated allylic carbon). HRMS (ESI) calcd for C₁₅H₂₂DO₄ 268.1659, found 268.1649 [M+H]⁺.

Conversion of 6bA-d into stereodefined iodolactone 10-d and 11-d (Scheme 2)



Scheme S10. Conversion of 6bA-d into stereodefined iodolactone 10-d and 10-d.

To a solution of **6bA**-*d* (62.9 mg, 0.235 mmol) in anhydrous MeOH (0.9 mL) were added KOH (56.7 mg, 0.940 mmol) and H₂O (24 μ L).² The resulting mixture was stirred at 70 °C for 30 min, and then treated with aqueous 3 M HCl. The aqueous layer was extracted with EtOAc twice. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated *in vacuo* to give a crude dicarboxylic acid, which was used for the next reaction without further purification.

A solution of the crude dicarboxylic acid in anhydrous DMF (0.5 mL) was sealed with a screw cap and stirred at 80 °C for 30 h.¹⁶ The reaction mixture was cooled to room temperature and concentrated *in vacuo* to give a crude monocarboxylic acid **9**-*d*, which was used for the next reaction without further purification.

To a solution of the crude acid **9**-*d* in anhydrous DCM (2.8 mL) was added Cs₂CO₃ (115 mg, 0.323 mmol) under argon. The resulting mixture was stirred at room temperature for 1.5 h, cooled to -60 °C, and then treated with NIS (159 mg, 0.705 mmol).¹⁷ The reaction mixture was stirred at the same temperature for 15 h, and then treated with water and saturated aqueous Na₂S₂O₃. The aqueous layer was extracted with Et₂O three times. The combined organic layers were washed with brine, dried over Na₂SO₄, and concentrated *in vacuo*. The residue was purified by silica gel column chromatography eluting with 7–10% EtOAc/hexane and then HPLC (eluent: hexane/*i*-propanol = 95/5, flow rate: 2 mL/min, *t*_R = 14.5 min (minor **11**-*d*) and *t*_R = 16.0 min (major **10**-*d*)) to give **11**-*d* (8.5 mg, 11%) and **10**-*d* (30.7 mg, 41%) as a white solid and a white solid, respectively.

The stereochemistries of **10**-*d* and **11**-*d* were established by their NOESY spectra. While the methine proton at γ -position in **10**-*d* had NOE with vinyl proton and no NOE with major residual proton as X at β -position, the γ -proton in **11**-*d* had NOE with major residual proton as X and no NOE with vinyl proton. The chemical shifts of geminal protons at β -position in **10**-*d* and **11**-*d* also agreed with those reported for γ -vinyl- γ -butyrolactone, where the vinyl group shielded and deshielded *cis*- and *trans*-protons, respectively.¹⁸ The shielded X and deshielded Y were also observed after reductive deiodination of **10**-*d* (vide infra). The corresponding undeuterated iodolactone **10** was also prepared from **6bA** by the same procedure described above and its NOESY spectrum showed a strong NOE between the methine δ -proton and β -proton located trans to the adjacent alkenyl group at 2.40 ppm.



(4R*,5R*)-5-((Z)-2-Cyclohexyl-1-iodovinyl)dihydrofuran-2(3H)-one-4-d (10-d)

White solid. Mp = 101 °C. Rf = 0.24 (17% EtOAc/hexane). IR (neat): 2924, 2849, 1781, 1635, 1447, 1317, 1258, 1165, 1006, 965, 896, 661 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 5.85 (d, *J* = 8.5 Hz, 1H), 4.76 (d, *J* = 6.8 Hz, 1H), 2.66 (dd, *J* = 18.0, 10.0, 1H), 2.53 (dd, *J* = 10.0, 8.8 Hz, 1H), 2.45–2.36 (m, 0.27H), 2.36–2.24 (m, 1H), 2.22–2.09 (m, 0.73H), 1.82–1.60 (m, 4H), 1.41–1.04 (m, 6H).

¹³C-NMR (100 MHz, CDCl₃): δ 176.4, 143.0, 104.3, 84.2, 44.6, 31.31, 31.29, 28.4 (t, J = 13.9 Hz), 28.0, 25.8, 25.5 (additional small peak at 28.7 ppm due to undeuterated homoallylic carbon). LRMS m/z (relative intensity) 321 [M]⁺ (46), 240 (49), 194 (100), 152 (65). HRMS (EI) calcd for C₁₂H₁₆DIO₂ 321.0336 found 321.0333 [M]⁺.

 $(4R^*, 5S^*)$ -5-((E)-2-Cyclohexyl-1-iodovinyl)dihydrofuran-2(3H)-one-4-d (**11**-d)

White solid. Rf = 0.24 (17% EtOAc/hexane). IR (neat): 2924, 2850, 1783, 1541, 1448, 1418, 1363, 1318, 1263, 1197, 1156, 1010, 961, 899, 802, 740, 697, 657 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 6.33 (d, *J* = 9.6 Hz, 1H), 5.90 (d, *J* = 7.2 Hz, 1H), 2.79–2.71 (m, 1H), 2.59 (dd, *J* = 18.6, 10.2, 1H), 2.49–2.39 (m, 1H), 2.39–2.30 (m, 0.72H), 2.20–2.10 (m, 0.28H), 1.80–1.55 (m, 4H), 1.35–1.12 (m, 6H). ¹³C-NMR (100 MHz, CDCl₃): δ 176.2, 151.8, 101.5, 77.2, 41.1, 33.1, 32.7, 28.5 (t, *J* = 20.9 Hz), 28.3, 25.53, 25.50, 25.38 (additional small peak at 28.8 ppm due to undeuterated homoallylic carbon). HRMS (ESI) calcd for C₁₂H₁₇DIO₂ 322.0414 found 322.0409 [M+H]⁺.

Reductive Deiodination of 10-d



Scheme S11. Conversion of 6bA-d into stereodefined iodolactone 10-d and 10-d.

To a solution of **10**-*d* (22.0 mg, 0.0687 mmol) in anhydrous PhMe (1.3 mL) were added Bu₃SnH (66.7 µL, 0.412 mmol) and AIBN (4.3 mg, 0.026 mmol) under argon. The mixture was stirred at 80 °C for 8.5 h. The reaction mixture was cooled to room temperature and concentrated in *vacuo*. The residue was purified by silica gel column chromatography eluting with 7–10% EtOAc/hexane and then HPLC (eluent: 5% i-PrOH/hexane, flow rate: 2 mL/min) to give **12** (6.1 mg, 45%) as a colorless oil.

(4R*,5R*)-5-((E)-2-Cyclohexylvinyl)dihydrofuran-2(3H)-one-4-d

Rf = 0.44 (25% EtOAc/PhMe). ¹H-NMR (600 MHz, CDCl₃): δ 5.75 (dd, *J* = 15.6, 6.5 Hz, 1H), 5.44 (dd, *J* = 15.6, 7.2 Hz, 1H), 4.88 (dd, *J* = 7.2, 7.2 Hz, 1H), 2.57–2.46 (m, 2H), 2.37–2.31 (m, 0.3H), 2.02–1.91 (1.7H), 1.78–1.59 (m, 4H), 1.37–1.00 (m, 6H). ¹³C-NMR (151 MHz, CDCl₃): δ 177.1, 141.3, 125.0, 81.3, 40.3, 32.6, 32.6, 29.8, 29.4, 29.0, 28.8, 28.7, 28.7, 28.5, 26.1, 26.0. LRMS m/z (relative intensity) 195 [M]⁺ (27), 135 (48), 121 (39), 112 (100). HRMS (EI) calcd for C₁₂H₁₇DO₂ 195.1370 found 195.1375 [M]⁺.

Preparation of (Sa, S)-2b-d

(1*R*, 2*R*)-4-Cyclohexylbut-3-yne-1-*d*-1,2-diol was prepared by asymmetric dihydroxylation of **7b**-*d* according to the literature procedure.¹⁹

White solid. Mp = 42.6 °C. Rf = 0.53 (67% EtOAc/hexane). $[\alpha]^{24}_{D}$ -13.6 (*c* 1.00 in CHCl₃). IR (neat): 3353 (br), 2929, 2853, 2656, 2361, 2332, 2227, 1450, 1444, 1087, 1048, 1030, 889, 865 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 4.45 (d, *J* = 6.6 Hz, 1H), 3.61 (d, *J* = 6.6 Hz, 1H), 2.60–2.11 (br, 3H), 1.86–1.59 (m, 4H), 1.59–1.04 (m, 6H). ¹³C-NMR (100 MHz, CDCl₃): δ 91.3, 77.5, 66.5 (t, *J* = 20.0 Hz), 63.4, 32.5, 29.0, 25.7, 24.8 (additional small peak at 66.9 ppm due to undeuterated homopropargylic carbon). LRMS m/z (relative intensity) 169 [M]⁺ (0.4), 137 (100), 119 (11), 91 (33). HRMS (EI) calcd for C₁₀H₁₅DO₂ 169.1213, found 169.1213 [M]⁺.

(1R, 2R)-1-((tert-Butyldimethylsilyl)oxy)-4-cyclohexylbut-3-yn-1-d-2-ol



Scheme S12. Monosilylation of diol.

To a solution of diol (846 mg, 5.00 mmol) in anhydrous DCM (30 mL) were added Et_3N (1.4 mL, 10.0 mmol), Me_2SnCl_2 (165 mg, 0.750 mmol), and TBSCI (1.09 g, 10.0 mmol) at 0 °C under argon.²⁰ The mixture was allowed to warm to room temperature and stirred for 2.5 h, and then treated with water. The aqueous layer was extracted with DCM three times. The combined organic layers were dried over MgSO₄ and concentrated *in vacuo*. The residue was purified by silica gel column chromatography eluting with 2–3% EtOAc/hexane to give TBS ether (1.43 g, quant) as a colorless oil.

Rf = 0.56 (17% EtOAc/hexane). $[a]^{24}_{D}$ –12.3 (c 0.98 in CHCl₃). IR (neat): 2930, 2856, 2361, 2342, 2231, 2151, 1254, 1125, 1050, 837, 777, 668 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 4.39 (ddd, *J* = 7.3, 4.4, 1.6 Hz, 1H), 3.58 (d, *J* = 7.3 Hz, 1H), 2.53 (d, *J* = 4.4 Hz, 1H), 2.43–2.31 (m, 1H), 1.85–1.61 (m, 4H), 1.61–1.20 (m, 6H), 0.91 (s, 9H), 0.10 (s, 3H), 0.90 (s, 3H). ¹³C-NMR (100 MHz, CDCl₃): δ 90.2, 77.7, 67.0 (t, *J* = 20.0 Hz), 63.2, 32.5, 29.0, 25.82, 25.80, 24.9, 18.3, -5.35, -5.39 (additional small peak at 67.4 ppm due to undeuterated homopropargylic carbon). LRMS m/z (relative intensity) 283 [M]⁺ (2), 251 (10), 226 (100), 134 (40). HRMS (EI) calcd for C₁₆H₂₉DO₂Si 283.2078, found 283.2084 [M]⁺.

(1R, 2R)-1-((tert-Butyldimethylsilyl)oxy)-4-cyclohexylbut-3-yn-2-yl-1-d methyl carbonate



Scheme S13. Introduction of leaving group.

To a solution of the alcohol (140 mg, 0.494 mmol) in anhydrous DCM (0.8 mL) were added pyridine (63 μ L, 0.78 mmol) and methyl chloroformate (61 μ L, 0.78 mmol) at 0 °C under argon. The mixture was stirred at room temperature for 4.5 h, and then treated with water. The aqueous layer was extracted with Et₂O twice. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated *in vacuo*. The residue was purified by preparative TLC eluting with 6% EtOAc/hexane to give carbonate (164 mg, 97%) as a colorless oil.

$$c_y = \bigvee_{H}^{OCO_2Me} OTBS$$

Rf = 0.68 (17% EtOAc/hexane). $[\alpha]^{18}_{D}$ -41.7 (*c* 1.21 in CHCl₃). IR (neat): 2931, 2856, 2239, 2158, 1757, 1577, 1472, 1463, 1443, 1409, 1389, 1362, 1340, 1268, 1140, 1106, 1084, 1007, 986, 939, 890, 873, 838, 815, 778, 740, 667 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 5.33 (dd, *J* = 8.0, 1.7 Hz, 1H), 3.98–3.75 (m, 1H), 3.79 (s, 3H), 2.47–2.33 (m, 1H), 1.86–1.25 (m, 10H), 0.88 (s, 9H), 0.08 (s, 6H). ¹³C-NMR (100 MHz, CDCl₃): δ 155.2, 92.1, 74.3, 69.4, 65.0 (t, *J* = 22.0 Hz), 54.8, 32.3, 29.0, 25.8, 25.7, 24.7, 18.2, -5.4 (additional small peak at 65.3 ppm due to undeuterated homopropargylic carbon). LRMS m/z (relative intensity) 341 [M]⁺ (0.04), 310 (0.3), 284 (3), 240 (10), 133 (100). HRMS (EI) calcd for C₁₈H₃₁DO₄Si 341.2133 found 341.2138 [M]⁺.

tert-Butyl(((1R, 3S)-4-cyclohexylbuta-2,3-dien-1-yl-1-d)oxy)dimethylsilane



Scheme S14. SN2' anti reduction of propargyl carbonate.

To a test tube containing CuCl (7.0 mg, 0.0707 mmol), IBiox12·HOTf (42.5 mg, 0.0700 mmol), and NaO'Bu (20.2 mg, 0.210 mmol) was added anhydrous PhMe (2.8 mL) under argon.²¹ The mixture was stirred at 40 °C for 1 h, treated with PMHS (23.7 μ L, 1.40 mmol), and stirred at room temperature for 5 min. To the resulting mixture was added carbonate (239 mg, 0.700 mmol). The mixture was stirred at 80 °C for 24 h and cooled to room temperature. The mixture was poured into saturated aqueous Na₂CO₃ and Et₂O, and was stirred for 1 h. The aqueous layer was extracted with Et₂O twice. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated *in vacuo*. The residue was purified by preparative TLC eluting with 2% EtOAc/hexane to give allene (54.7 mg, 29%) as a colorless oil and the recovered substrate (140 mg, 59%).

Rf = 0.16 (hexane). $[α]^{23}_{D}$ +48.6 (*c* 1.08 in CHCl₃). IR (neat): 2927, 2854, 2363, 2141, 1962, 1471, 1448, 1361, 1254, 1095, 1006, 951, 836, 775, 704 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 5.23 (ddd, *J* = 6.1, 6.1, 2.7 Hz, 1H), 5.16 (ddd, *J* = 6.1, 6.1, 2.7 Hz, 1H), 4.19–4.14 (m, 1H), 2.05–1.92 (m, 1H), 1.80–1.53 (m, 4H), 1.34–1.01 (m, 6H), 0.90 (s, 9H), 0.08 (s, 6H). ¹³C-NMR (100 MHz, CDCl₃): δ 202.4, 98.3, 92.5, 61.9 (t, *J* = 22.5 Hz), 37.1, 33.12, 33.10, 26.2, 26.0, 18.4, -5.06, -5.08 (additional small peak at 62.2 ppm due to undeuterated allylic carbon). LRMS m/z (relative intensity) 267 [M]⁺ (0.3), 210 (100), 209 (33), 128 (62). HRMS (EI) calcd for C₁₆H₂₉DOSi 267.2129 found 267.2123 [M]⁺.

(1R, 3S)-4-Cyclohexylbuta-2,3-dien-1-d-1-ol



Scheme S15. Removal of TBS group.

To a test tube containing substrate (54.7 mg, 0.204 mmol) were added anhydrous THF (0.8 mL) and 1 M Bu₄NF solution in THF (408 μ L, 0.41 mmol) under argon. The resulting mixture was sealed with a screw cap, stirred at room temperature for 1.5 h, and then treated with saturated aqueous NH₄Cl. The aqueous layer was extracted with Et₂O twice. The combined organic layers were washed with brine, dried over MgSO₄ and concentrated *in vacuo*. The residue was purified by preparative TLC eluting with 20% EtOAc/hexane to give allenic alcohol (30.1 mg, 96%) as a colorless oil. The enantiomeric ratio (*er*) of **6aA** was (1*R*, 3*S*):(1*S*, 3*R*) = 84:16, which was determined by HPLC analysis with chiral stationary phase column (Daicel Chiralpak AS-H, hexane/*i*-propanol = 50/1, 0.6 mL/min, λ = 214 nm, *t*_R(1*S*, 3*R* - minor enantiomer) = 14.9 min, *t*_R(1*R*, 3*S* - major enantiomer) = 16.1 min).²²



Rf = 0.30 (17% EtOAc/hexane). [α]²⁷_D +43.8 (*c* 0.69 in CHCl₃). IR (neat): 3341 (br), 2924, 2850, 2158, 1961, 1448, 1290, 1066, 1046, 1017, 890 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 5.45–5.12 (m, 2H), 4.22–4.00 (m, 1H), 2.09–1.84 (m, 1H), 1.84–1.47 (m, 4H), 1.38–0.98 (m, 6H). ¹³C-NMR (100 MHz, CDCl₃): δ 201.9, 100.0, 92.6, 60.5 (t, *J* = 22.5 Hz), 37.0, 33.04, 32.98, 26.0, 25.92, 25.91 (additional small peak at 60.8 ppm due to undeuterated allylic carbon). LRMS m/z (relative intensity) 153 [M]⁺ (4), 137 (27), 121 (36), 108 (100). HRMS (EI) calcd for C₁₀H₁₅DO 153.1264 found 153.1246 [M]⁺.

(1R, 3S)-4-Cyclohexylbuta-2,3-dien-1-yl-1-d methyl carbonate ((Sa, R)-2b-d)



Scheme S16. Formation of methyl carbonate (Sa, R)-2b-d.

To a solution of the allenic alcohol (30.1 mg, 0.196 mmol) in anhydrous DCM (0.8 mL) were added pyridine (63 μ L, 0.78 mmol) and methyl chloroformate (61 μ L, 0.78 mmol) under argon. The mixture was stirred at room temperature for 18 h and then treated with water. The aqueous layer was extracted with Et₂O twice. The combined organic layers were washed with brine, dried over MgSO₄, and concentrated *in vacuo*. The residue was purified by preparative TLC eluting with 10% EtOAc/hexane to give allenic carbonate (34.9 mg, 84%) as a colorless oil.

Rf = 0.68 (17% EtOAc/hexane). [α]²³_D +31.0 (*c* 1.06 in CHCl₃). IR (neat): 2925, 2851, 1964, 1750, 1444, 1340, 1265, 946, 791 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 5.36–5.19 (m, 2H), 4.63–4.50 (m, 1H), 3.79 (s, 3H), 2.05–1.93 (m, 1H), 1.79–1.51 (m, 6H), 1.40–0.96 (m, 4H). ¹³C-NMR (100 MHz, CDCl₃): δ 204.7, 155.6, 99.0, 87.2, 66.1 (t, *J* = 23.0 Hz), 54.7, 36.8, 32.8, 26.0, 25.90, 25.88 (additional small peak at 66.4 ppm due to undeuterated allylic carbon). LRMS m/z (relative intensity) 136 [M–OCO₂Me]⁺ (3), 119 (15), 107 (43), 92 (43), 83 (100). HRMS (EI) calcd for C₁₀H₁₄D 136.1237 found 136.1235 [M–OCO₂Me]⁺.

Substitution reaction of (Sa, R)-2b-d with 5A under the catalysis of Pd-diphosphine 8 in the absence or presence of Lil

In the absence of Lil



Scheme S17. Substitution reaction of (Sa, R)-2b-d with 5A under the catalysis of Pd-diphosphine 8 in the absence of Lil.

To a test tube containing (Sa, *R*)-**2b**-*d* (25.0 mg, 0.118 mmol), (η^3 -allyl)(η^5 -Cp)Pd (0.8 mg, 0.004 mmol), and **8** (5.8 mg, 0.0053 mmol) were added anhydrous MeOH (1.2 mL) and dimethyl malonate (41 µL, 0.354 mmol) under argon. The resulting mixture was sealed with a screw cap and stirred at 65 °C for 18 h. The reaction mixture was cooled to room temperature and concentrated *in vacuo*. The residue was purified by preparative TLC eluting with 10% EtOAc/hexane to give **6bA**-*d* (18.5 mg, 59%) as a colorless oil. The enantiomeric ratio (*er*) of **6bA**-*d* was (1*R*, 3*S*):(1*S*, 3*R*) = 64:36, which was determined by HPLC analysis with chiral stationary phase column (Daicel Chiralpak OD-H, hexane/*i*-propanol = 397/3, 0.5 mL/min, λ = 210 nm, $t_R(3S - 10^{-1})$

major enantiomer) = 17.5 min, $t_R(3R - \text{minor enantiomer}) = 18.7 \text{ min})$. The diastereomeric ratio and stereochemistry of **6bA**-*d* were determined using NMR analysis of γ -lactone **10**-*d* prepared according to the following procedure.

Dimethyl 2-((1R,3S)-4-cyclohexylbuta-2,3-dien-1-yl-1-d)malonate (6bA-d)

$$Cy$$
 CO_2Me
 X CO_2Me
 $X: 85 atom % D$
 $Y: 15 atom % D$

. .

Rf = 0.29 (17% EtOAc/hexane). [α]²³_D +33.4 (*c* 0.50 in CHCl₃). IR (neat): 2925, 2851, 1961, 1755, 1738, 1436, 1262, 1154, 1038, 892 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 5.14 (m, 2H), 3.74 (s, 6H), 3.50 (d, *J* = 7.6 Hz, 1H), 2.66–2.52 (m, 1H), 2.02–1.81 (m, 1H), 1.81–1.58 (m, 4H), 1.34–0.95 (m, 6H). ¹³C-NMR (100 MHz, CDCl₃): δ 202.9, 169.42, 169.37, 99.0, 88.2, 52.49, 52.47, 51.2, 37.2, 32.94, 32.88, 27.9 (t, *J* = 20.5 Hz), 26.1, 26.0 (additional small peak at 28.1 ppm due to undeuterated allylic carbon). HRMS (ESI) calcd for C₁₅H₂₂DO₄ 268.1659, found 268.1649 [M+H]⁺.



6bA-*d* obtained from (Sa, R)-**2b**-*d* was also converted into iodolactone **10**-*d* (6% over 3 steps) and overall retention of the stereoselectivity in the substitution reaction of (Sa, R)-**2b**-*d* with **5A** was confirmed.

Conversion of 6bA-d into stereodefined iodolactone 10-d



Scheme S18. Conversion of 6bA-d into stereodefined iodolactone 10-d.

(4S, 5R)-5-((Z)-2-Cyclohexyl-1-iodovinyl)dihydrofuran-2(3H)-one-4-d (10-d)



White solid. Rf = 0.24 (17% EtOAc/hexane). IR (neat): 2923, 2849, 1780, 1450, 1317, 1165, 1009 cm⁻¹. ¹H-NMR (400 MHz, CDCl₃): δ 5.85 (d, *J* = 8.8 Hz, 1H), 4.76 (d, *J* = 7.6 Hz, 1H), 2.82–2.60 (m, 1H), 2.60–2.47 (m, 1H), 2.47–2.37 (m, 0.85H), 2.37–2.23 (m, 1H), 2.23–2.10 (m, 0.15H) 1.60 (m, 4H), 1.20 (m, 6H). ¹³C-NMR (100 MHz, CDCl₃): δ 176.5, 143.1, 104.3, 84.2, 44.6, 31.3, 31.2, 28.4 (t, *J* = 21.0 Hz), 28.0, 25.7, 25.4 (additional small peak at 28.7 ppm due to undeuterated homoallylic carbon). LRMS m/z (relative intensity) 321 [M]⁺ (37), 240 (35), 194 (100), 152 (50). HRMS (EI) calcd for C₁₂H₁₆DIO₂ 321.0336 found 321.0329 [M]⁺.

In the presence of Lil



Scheme S19. Substitution reaction of (Sa, R)-2b-d with 5A under the catalysis of Pd-diphosphine 8 in the presence of Lil.

To a test tube containing (Sa, *R*)-**2b**-*d* (0.9 mg, 4 µmol), (η^3 -allyl)(η^5 -Cp)Pd (0.2 mg, 0.9 µmol), **8** (0.3 mg, 0.3 µmol), and Lil (0.6 mg, 4 µmol) were added anhydrous MeOH (34 µL) and dimethyl malonate (1.5 µL, 13 µmol) under argon. The resulting mixture was sealed with a screw cap and stirred at 65 °C for 3 h. The reaction mixture was cooled to room temperature and concentrated *in vacuo*. The residue was purified by silica gel column chromatography eluting with 3% EtOAc/hexane to give **6bA**-*d* (0.3 mg, 26%) as a colorless oil. The enantiomeric ratio (*er*) of **6bA**-*d* was (1*R*, 3*S*):(1*S*, 3*R*) = 51:49, which was determined by HPLC analysis with chiral stationary phase column (Daicel Chiralpak OD-H, hexane/*i*-propanol = 397/3, 0.5 mL/min, λ = 210 nm, *t*_R(3*S* - major enantiomer) = 17.1 min, *t*_R(3*R* - minor enantiomer) = 18.6 min).



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¹H NMR spectrum of (*E*)-(4-cyclohexylbut-1-en-3-yn-1-yl)trimethylsilane



¹³C NMR spectrum of (*E*)-(4-cyclohexylbut-1-en-3-yn-1-yl)trimethylsilane





¹³C NMR spectrum of **7b**-d





¹H NMR spectrum of tetraethyl [1,1'-biphenyl]-2,2'-diylbis(phosphonate)



¹³C NMR spectrum of tetraethyl [1,1'-biphenyl]-2,2'-diylbis(phosphonate



³¹P NMR spectrum of tetraethyl [1,1'-biphenyl]-2,2'-diylbis(phosphonate



¹H NMR spectrum of [1,1'-biphenyl]-2,2'-diylbis(bis(3,5-di-*tert*-butyl-4-methoxyphenyl)phosphine oxide)



¹³C NMR spectrum of [1,1'-biphenyl]-2,2'-diylbis(bis(3,5-di-*tert*-butyl-4-methoxyphenyl)phosphine oxide)



³¹P NMR spectrum of [1,1'-biphenyl]-2,2'-diylbis(bis(3,5-di-*tert*-butyl-4-methoxyphenyl)phosphine oxide)









¹H NMR spectrum of **6bA**-*d* prepared from **7b**-*d*



¹³C NMR spectrum of **6bA**-*d* prepared from **7b**-*d*



¹H NMR spectrum of **10**-*d* prepared from **7b**-*d*



¹³C NMR spectrum of **10**-*d* prepared from **7b**-*d*



COSY spectrum of **10**-*d* prepared from **7b**-*d*



NOESY spectrum of **10**-*d* prepared from **7b**-*d*



COSY spectrum of **10**




¹H NMR spectrum of **11**-*d* prepared from **7b**-*d*



¹³C NMR spectrum of **11**-*d* prepared from **7b**-*d*



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COSY spectrum of **11**-*d* prepared from **7b**-*d*



NOESY spectrum of **11**-*d* prepared from **7b**-*d*



¹H NMR spectrum of (4*R**,5*R**)-5-((*E*)-2-cyclohexylvinyl)dihydrofuran-2(3*H*)-one-4-*d*



¹³C NMR spectrum of (4*R**,5*R**)-5-((*E*)-2-cyclohexylvinyl)dihydrofuran-2(3*H*)-one-4-*d*









NOESY spectrum of (4R*,5R*)-5-((E)-2-cyclohexylvinyl)dihydrofuran-2(3H)-one-4-d

¹H NMR spectrum of (1*R*, 2*R*)-4-cyclohexylbut-3-yne-1-*d*-1,2-diol





¹H NMR spectrum of (1*R*, 2*R*)-1-((*tert*-butyldimethylsilyl)oxy)-4-cyclohexylbut-3-yn-1-*d*-2-ol



¹³C NMR spectrum of (1*R*, 2*R*)-1-((*tert*-butyldimethylsilyl)oxy)-4-cyclohexylbut-3-yn-1-*d*-2-ol



¹H NMR spectrum of (1*R*, 2*R*)-1-((*tert*-butyldimethylsilyl)oxy)-4-cyclohexylbut-3-yn-2-yl-1-*d* methyl carbonate



¹³C NMR spectrum of (1*R*, 2*R*)-1-((*tert*-butyldimethylsilyl)oxy)-4-cyclohexylbut-3-yn-2-yl-1-*d* methyl carbonate



¹H NMR spectrum of *tert*-butyl(((1*R*, 3*S*)-4-cyclohexylbuta-2,3-dien-1-yl-1-*d*)oxy)dimethylsilane



¹³C NMR spectrum of *tert*-butyl(((1*R*, 3*S*)-4-cyclohexylbuta-2,3-dien-1-yl-1-*d*)oxy)dimethylsilane



¹H NMR spectrum of (1*R*, 3*S*)-4-cyclohexylbuta-2,3-dien-1-*d*-1-ol



¹³C NMR spectrum of (1*R*, 3*S*)-4-cyclohexylbuta-2,3-dien-1-*d*-1-ol







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¹H NMR spectrum of **6bA**-*d* prepared from (*S*a, *R*)-**2b**-*d*



¹³C NMR spectrum of **6bA**-*d* prepared from (*S*a, *R*)-**2b**-*d*



¹H NMR spectrum of **10**-*d* prepared from (*S*a, *R*)-**2b**-*d*



¹³C NMR spectrum of **10**-*d* prepared from (*S*a, *R*)-**2b**-*d*

