

DNA Structural Alteration Leading to Antibacterial Properties of 6-Nitroquinoxaline Derivatives

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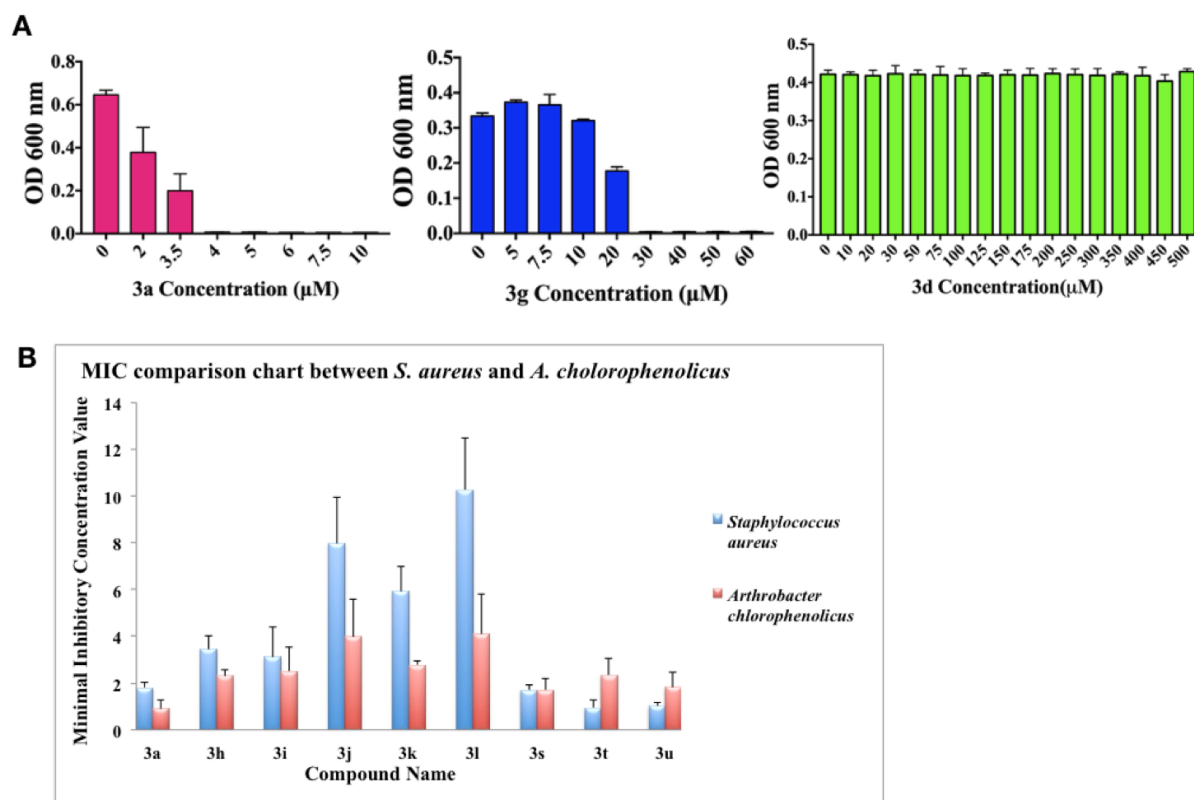
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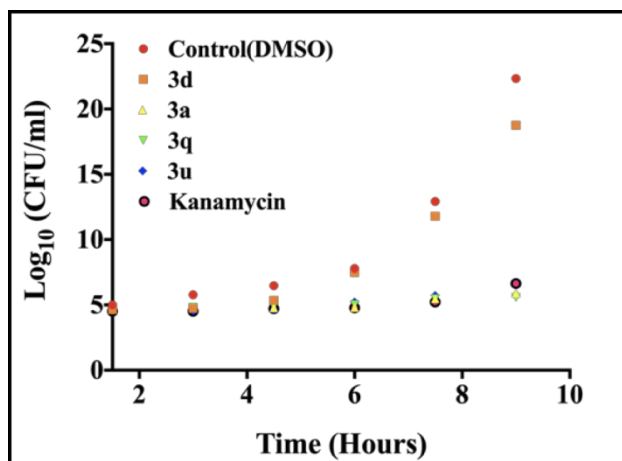
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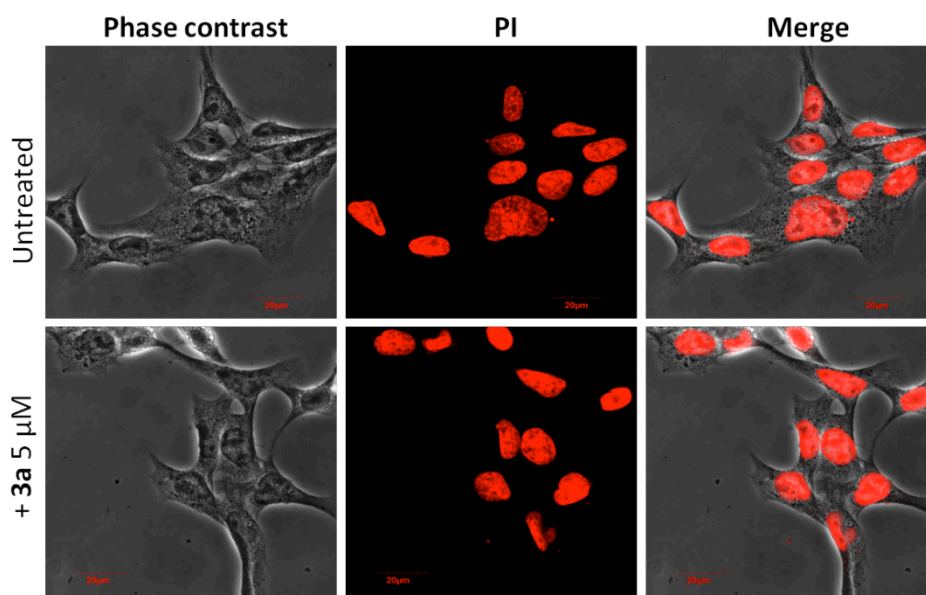
Supporting Figure S1:(A) MIC values of *S. aureus* cells treated with indicated compounds (**3a**, **3g**, **3d**) and incubated for 6 h at 37 °C.^{1, 2} (B) Comparison of MIC values of para substituted derivatives (**3a**, **3h**, **3i**, **3j**, **3k**, **3l**) and disubstituted derivatives (**3s**, **3t**, **3u**) between *S. aureus* and *A. chlorophenolicus*.

Time-kill assay:

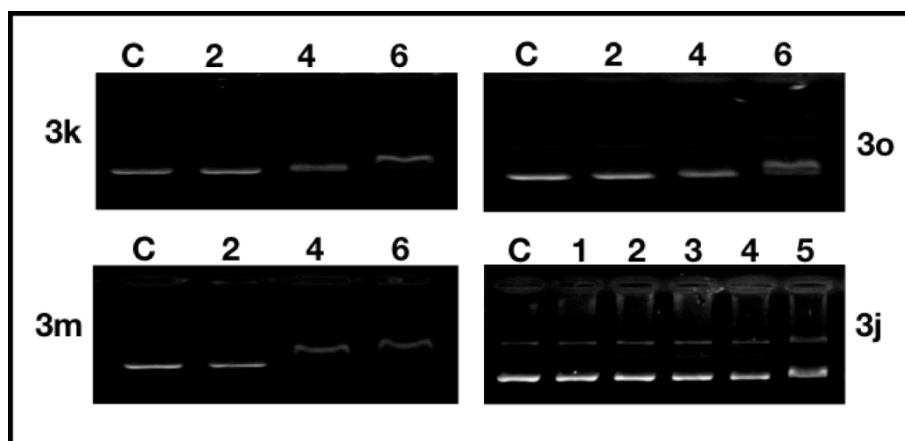
Monitoring the rate of bacteriocidal or bacteriostatic activity in the presence of varying concentrations of the antimicrobial compounds is very crucial in determining the actual effect of the drug against a growing population of the bacterial cells.³ In this study, the growth kinetics of *S. aureus* cells in the presence of designed compounds were evaluated by an *in-vitro* time-kill assay. Log phase *S. aureus* cells (3×10^5 cells/mL) were treated with **3a**, **3d**, **3u**, **3q** and kanamycin (as a positive control), incubated at 37 °C and OD was measured (at 600 nm) at 1.5 hours interval. Results indicate that the compounds **3a**, **3q** and **3u** were able to inhibit the growth of the bacterial cell at double-MIC concentrations while **3d** was unable to inhibit the growth of the cells (Figure S2).



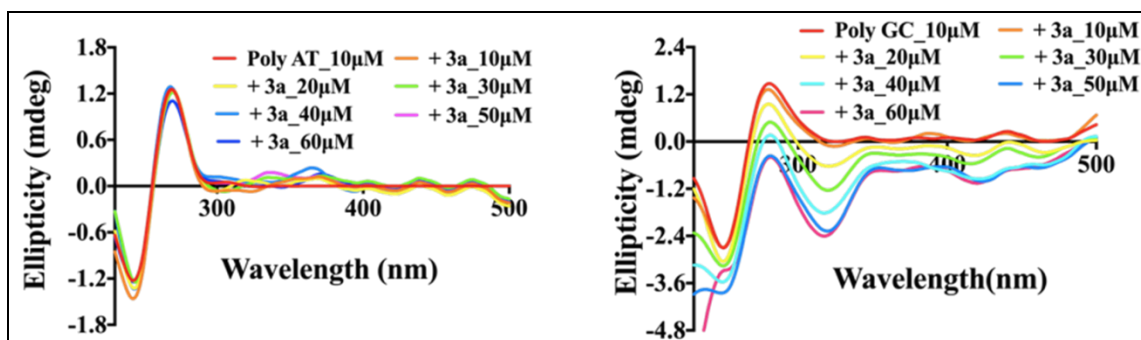
Supporting Figure S2: Time-kill kinetic study of the compounds (**3a**, **3d**, **3q**, **3u** and kanamycin) against *S. aureus*. 3×10^5 cells were inoculated, treated with DMSO (0.5 %) or indicated compounds. Log CFU/mL was plotted against different time (hour).



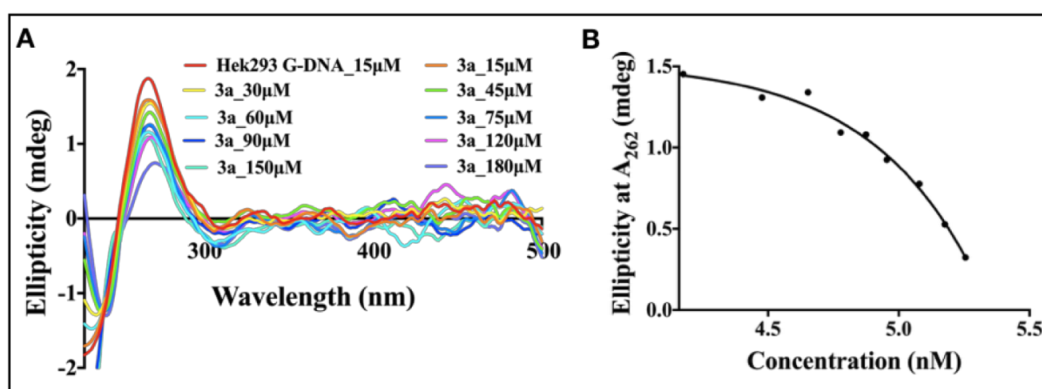
Supporting Figure S3: Cellular morphology of HEK 293 cells, untreated or treated with **3a**, stained with propidium iodide (PI, nuclear stain) cells were visualized in 60x magnification. Scale bar- 20 μm .³



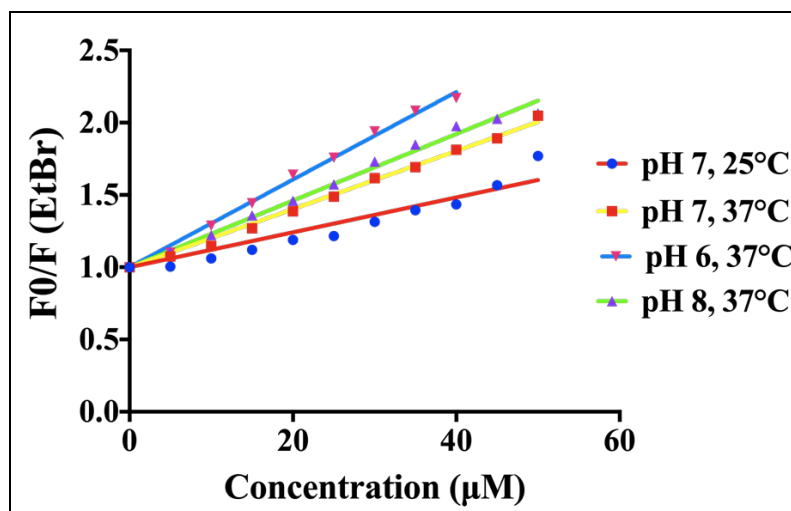
Supporting Figure S4: Agarose gel shift assay with **3k**, **3o**, **3m** and **3j**. Lane number indicates [compound]/[DNA base pair] ratios.⁴



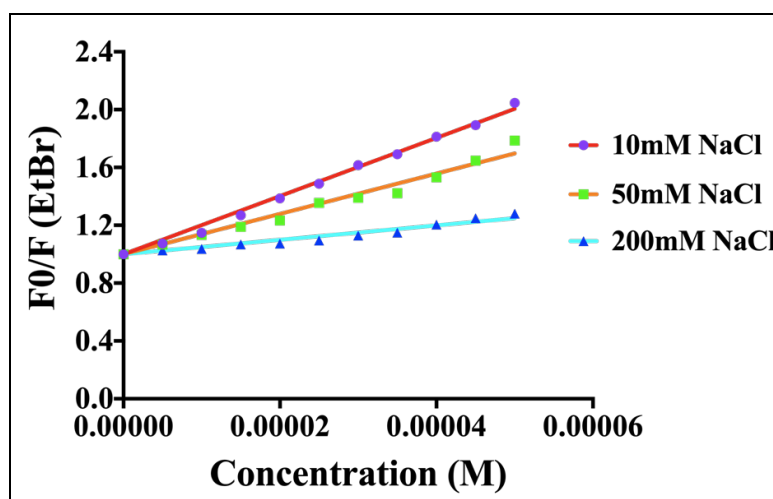
Supporting Figure S5: Circular dichroism (CD) spectra of 10 μ M poly-AT DNA (left) and poly-GC DNA (right) titrated with **3a** in an increasing concentrations from 10 to 60 μ M.⁴



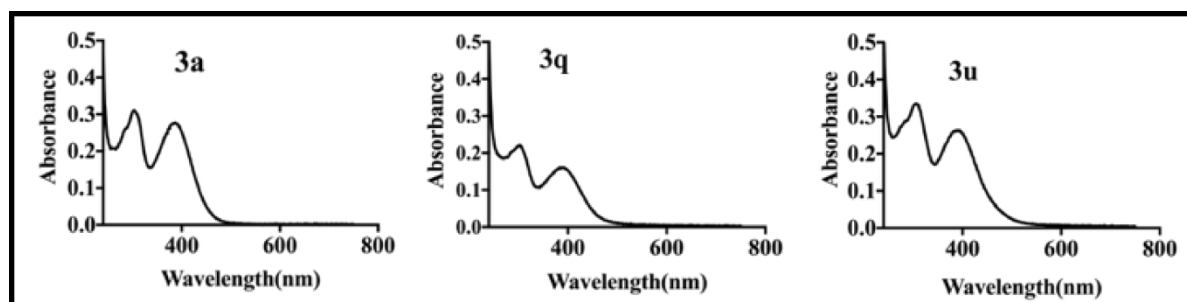
Supporting Figure S6: (A) Circular dichroism (CD) spectra of 15 μ M mammalian genomic DNA (isolated from HEK 293 cell line) titrated with **3a** in an increasing concentrations of 15, 30, 45, 60, 75, 90, 120, 150, 180 μ M. (B) Ellipticity plot at 262 nm bands of CD spectra.



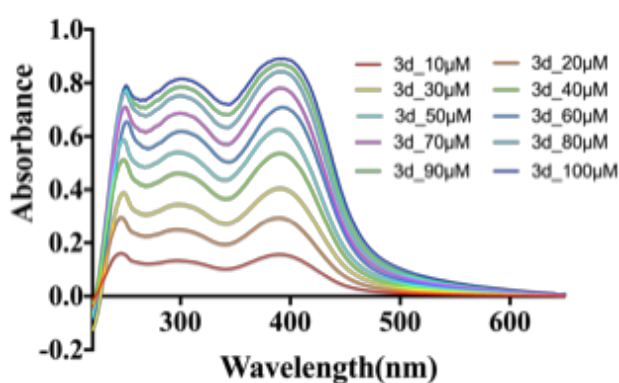
Supporting Figure S7: Quenching plot of fluorescence intercalator displacement assay (FID) with 10 μM *S. aureus* genomic DNA and 5 μM EtBr ($\lambda_{\text{ex}} = 480 \text{ nm}$) titrated with **3a** (2.5-50 μM) at indicated pH and temperature. K_{SV} values at (pH 8, 37 °C is $2.3 \times 10^4 \text{ M}^{-1}$, pH 7, 37 °C is $2.0 \times 10^4 \text{ M}^{-1}$, pH 6, 37 °C is $3.0 \times 10^4 \text{ M}^{-1}$).



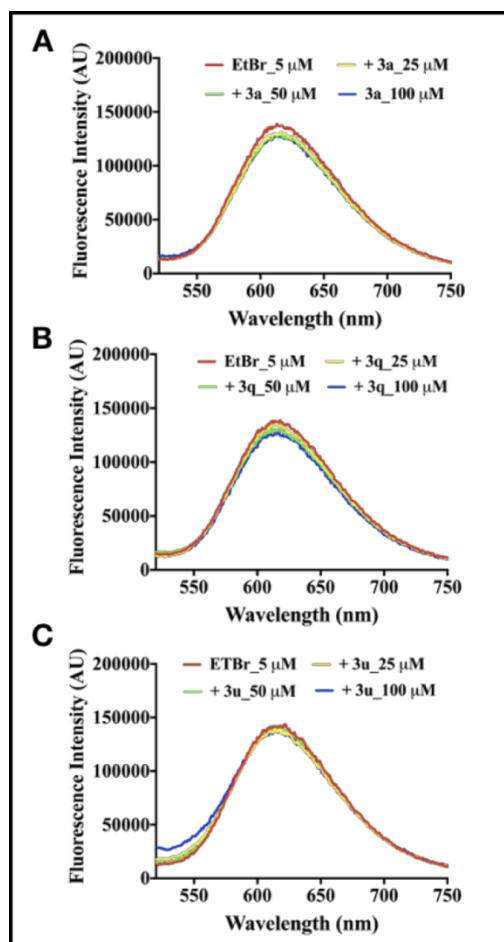
Supporting Figure S8: Quenching plot of fluorescence intercalator displacement assay (FID) with 10 μM *S. aureus* genomic DNA and 5 μM EtBr ($\lambda_{\text{ex}} = 480 \text{ nm}$) with indicated salt concentration.



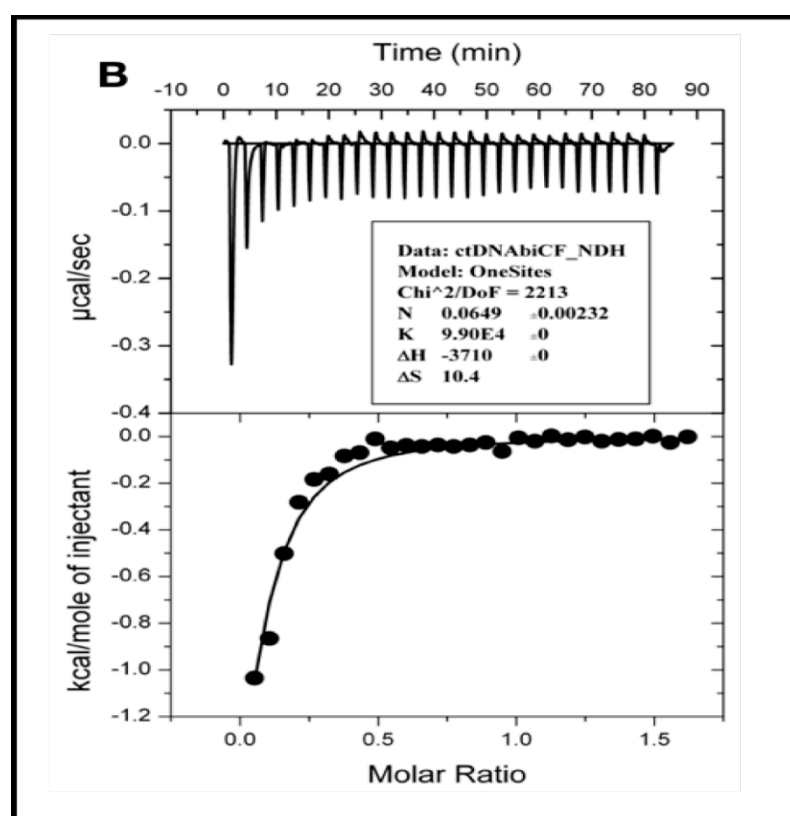
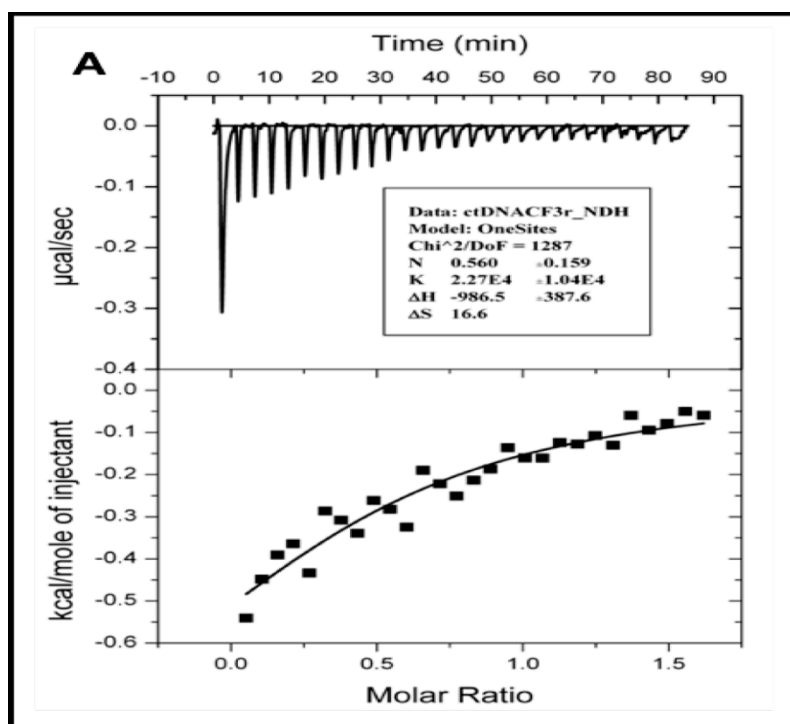
Supporting Figure S9: UV absorption spectra of compounds **3a**, **3q**, **3u** in 10 mM phosphate buffer at pH 7.0 with 10 mM NaCl and 1 % DMSO at 25⁰ C.

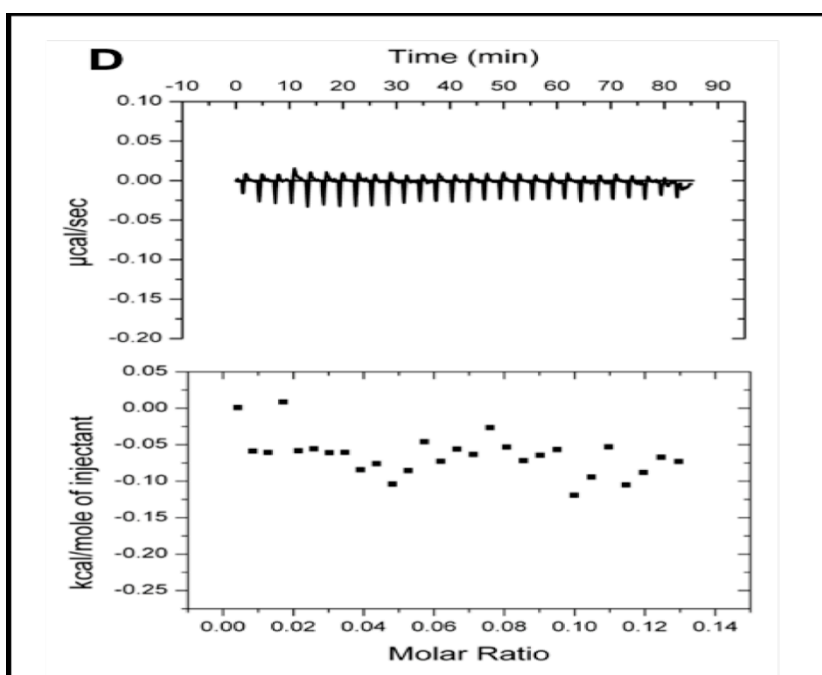
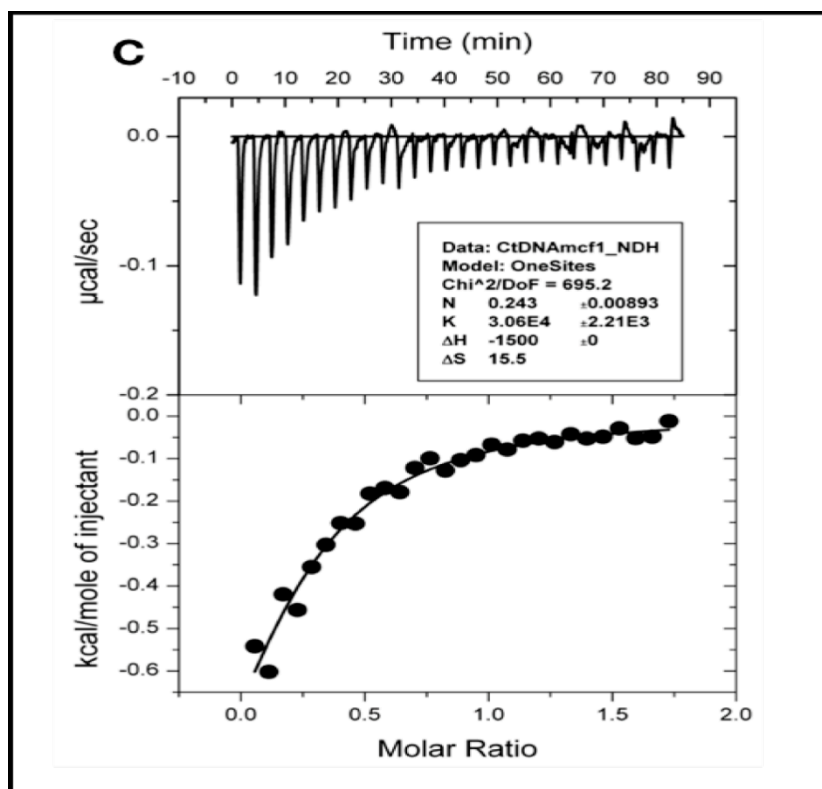


Supporting Figure S10: UV absorption spectra of compound **3d** (in 10 mM phosphate buffer at pH 7.0 with 10 mM NaCl and 1 % DMSO at 25⁰ C) with an increasing concentration (10 μM to 100 μM).

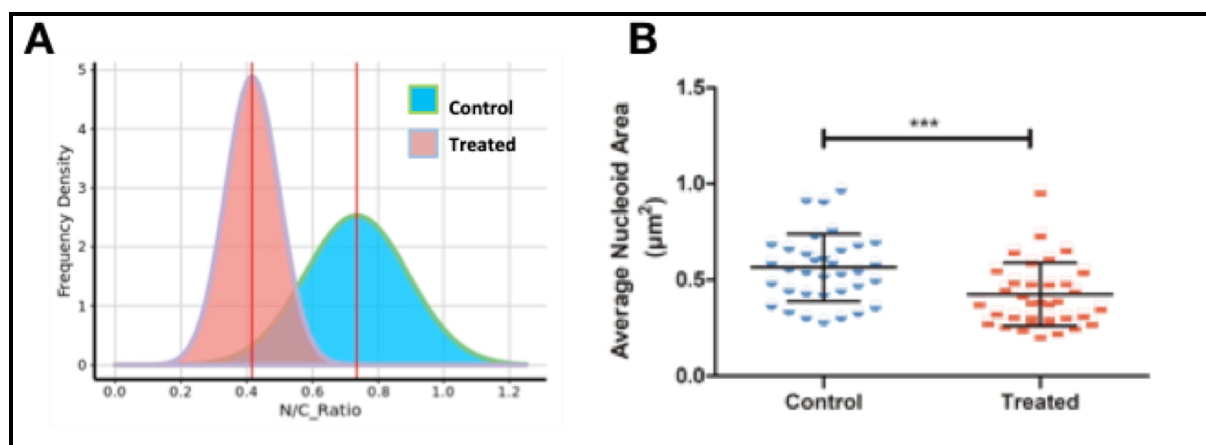


Supporting Figure S11: EtBr fluorescence quenching studies for compounds (A) **3a**, (B) **3q** and (C) **3u** in the presence of EtBr only in buffer (10 mM Na-P pH 7, 10 mM NaCl and 1 % DMSO)

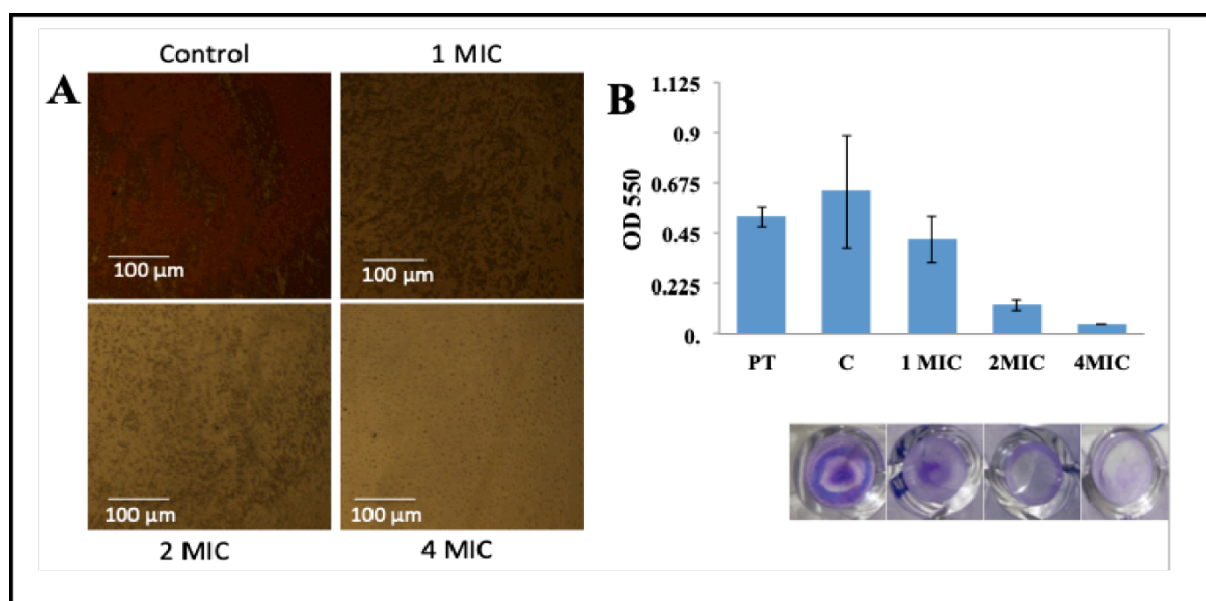




Supporting Figure S12: Isothermal titration calorimetric analysis of CT-DNA with (A) **3a**, (B) **3u**, (C) **3q** and (D) Buffer correction. Plots were drawn using Origin 7 software.⁵



Supporting Figure S13: (A) Frequency density plot shows significantly different distribution for untreated and treated *S. aureus* cells with **3a** (control mean N/C = 0.73, treated mean N/C = 0.41). (B) Average nucleoid area also shows significant reduction in treated (0.423) *S. aureus* cells compared to control (0.56). (n=35).



Supporting Figure S14: Disruption of preformed biofilm of *S. epidermidis* by **3a**. (A) Visualization of compound **3a** treated (1, 2, 4 MIC) or untreated *S. epidermidis* biofilm under light microscope at 20 \times . (B) Quantification and visualization of biofilm formation in bacteria using crystal violet stain.⁶

Supporting Table 1: Comparison Table of our synthetic derivatives with already available analogues.

Sl. No.	Synthetic compounds/natural products	MIC (µg/mL)			
		Gram positive bacteria		Gram negative bacteria	
		<i>S. aureus</i>	<i>S. epidermidis</i>	<i>E. coli</i>	<i>P. aeruginosa</i>
1.	3a	1.79	2.24	3.36	6.73
2.	3b	3.78	3.78	3.78	>10.07
3.	3c	3.47	3.47	2.31	>9.24
4.	3f	3.03	4.32	4.32	10.80
5.	3h	3.45	3.45	4.59	>11.48
6.	3m	3.36	2.24	>22.42	>22.42
7.	3q	0.89	0.89	>8.96	>6.72
8.	3s	1.69	0.96	7.24	>7.24
9.	3t	0.93	1.63	>9.33	>6.99
10.	3u	1.03	0.52	>7.75	>7.75
11.	3-(5-Hexyl-3-phenylimidazo[1,5-a]quinoxalin-4-on-1-yl)-1-nonylpyridinium iodide. ⁷	0.78	—	>500	>500
12.	2,3-Bis(bromomethyl)-6-(trifluoromethyl)quinoxaline. ⁸	12.5	—	>100	>100
13.	N-(2,3-di(furan-2-yl)quinoxalin-6-yl)-4-Nitrobenzenesulfonamide. ⁹	15	—	25	30
14.	Choles-5-en-3[thiazolo [4,5-b]quinoxaline-2-ylhydrazone. ¹	0.78	—	0.39	—
15.	4-tert-Butyl-3-hydroxy-1,4-dihydrobenzo[g]quinoxaline-5,10-dione. ¹¹	12.5	—	12.5	12.5
16.	Echinomycin (quinomycin A). ¹	0.03125	—	25	>100
17.	Triostin A. ¹	0.625	—	>100	>100

REFERENCE:

1. Cornish, A.; Fox, K. R.; Waring, M. J. Preparation and DNA-binding properties of substituted triostin antibiotics. *Antimicrob. Agents Chemother.* **1983**, *23*, 221-231.
2. Zhao, Y.; Tian, Y.; Cui, Y.; Liu, W.; Ma, W.; Jiang, X. Small molecule-capped gold nanoparticles as potent antibacterial agents that target gram-negative bacteria. *J. Am. Chem. Soc.* **2010**, *132*, 12349–12356.
3. Niu, Y.; Wang, M.; Cao, Y.; Nimmagadda, A.; Hu, J.; Wu, Y.; Cai, J.; Ye, X. Rational design of dimeric lysine N-alkylamides as potent and broad-spectrum antibacterial agents. *J. Med. Chem.* **2018**, *61*, 2865–2874.
4. Mahata, T.; Kanungo, A.; Ganguly, S.; Modugula, E. K.; Choudhury, S.; Pal, S. K.; Basu, G.; Dutta, S. The benzyl moiety in a quinoxaline-based scaffold acts as a DNA intercalation switch. *Angew. Chem. Int. Ed.* **2016**, *128*, 7864-7867.
5. Kwasny, S. M.; Opperman, T. J. Static biofilm cultures of gram-positive pathogens grown in a microtiter format used for anti-biofilm drug discovery. *Curr. prot. pharmacol.* **2010**, *50*, 13A.8.1-13A.8.23.
6. Buurma, N. J.; Haq, I. Advances in the analysis of isothermal titration calorimetry data for ligand–DNA interactions. *Methods.* **2007**, *42*, 162-172.
7. Kalinin, A. A.; Voloshina, A. D.; Kulik, N. V.; Zobov, V. V.; Mamedov, V. A. Antimicrobial activity of imidazo[1,5-a]quinoxaline derivatives with pyridinium moiety. *Eur. J. Med. Chem.* **2013**, *66*, 345-354.
8. Ishikawa, H.; Sugiyama, T.; Kurita, K.; Yokoyama, A. Synthesis and antimicrobial activity of 2,3-bis(bromomethyl)quinoxaline derivatives. *Bioorg. Chem.* **2012**, *41*, 1-5.
9. Kamal, A.; Babu, K. S.; Faazil, S.; Hussaini, S. A.; Shaik, A. B. L-Proline mediated synthesis of quinoxalines; evaluation of cytotoxic and antimicrobial activity. *RSC Adv.* **2014**, *4*, 46369-46377.

10. Khan, S. A. Synthesis, characterization and *in vitro* antibacterial activity of new steroidal 5-en-3-oxazolo and thiazoloquinoxaline. *Eur. J. Med. Chem.* **2008**, *43*, 2040- 2044.
11. Tandon, V. K.; Yadav, D. B.; Maurya, H. K.; Chaturvedi, A. K.; Shukla, P. K. Design, synthesis, and biological evaluation of 1,2,3-trisubstituted-1,4-dihydrobenzo[g]quinoxaline-5,10-diones and related compounds as antifungal and antibacterial agents. *Bioorg. Med. Chem.* **2006**, *14*, 6120- 6126.
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