

**Supplementary Information for:**

**A Case Study Comparing Heterogeneous Lysine- and Site-Specific Cysteine-Conjugated  
Maytansinoid Antibody-Drug Conjugates (ADCs) Illustrates Benefits of Lysine  
Conjugation**

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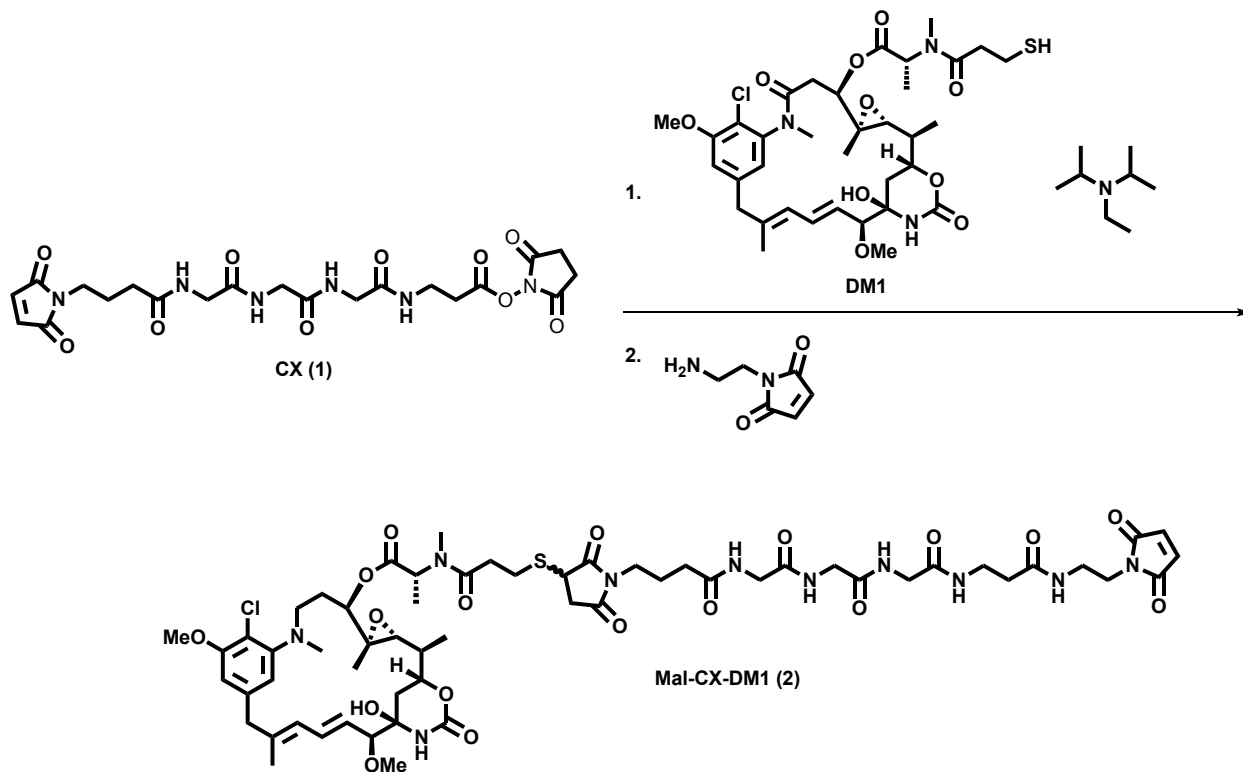
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## General Synthetic Methods.

DM1 was manufactured at Sicor Inc. Synthesis of the core heterobifunctional CX linker (Supplementary Figure 1) has been previously reported [Singh MCT 2016]. All other reagents were obtained from Sigma-Aldrich. Nuclear magnetic resonance (NMR) spectra were acquired on a Bruker Ascend 400 spectrometer operating at 400 MHz for  $^1\text{H}$  and 100 MHz for  $^{13}\text{C}$ . NMR chemical shifts are reported in  $\delta$  values relative to the utilized NMR solvent. High resolution mass spectra were acquired on a Thermo Fisher Q-Exactive mass spectrometer. HPLC purifications were performed using a Varian ProStar preparative HPLC system.

## Synthesis of Mal-CX-DM1 (2).



**Supplementary Figure 1.** Scheme for chemical synthesis of compound 2 (Mal-CX-DM1).

A 10 mL round bottom flask equipped with a stir bar was charged with DM1 (145.4 mg, 0.197 mmol) and DMF (1 mL). CX linker (103 mg, 0.197 mmol) and *N,N*-diisopropylethylamine (DIPEA) (0.069 mL, 0.394 mmol) were sequentially added. The reaction was placed under an argon atmosphere and stirred for ~45 minutes at room temperature. 1-(2-aminoethylmaleimide) hydrochloride (34.6 mg, 0.197 mmol) was added directly to this crude reaction. Stirring was continued for ~1h at room temperature. Upon completion, no work-up was performed and the desired product was isolated by semi-preparative C18 HPLC using a Targa C18 10  $\mu$ m, 20 x 250 mM column. The column was eluted with a gradient of acetonitrile in water containing 0.1% formic acid (see Supplementary Table S1).

Time (min)	% Acetonitrile
0	15
3.75	32
6.25	48
0:00	58
15	100
18	100
18.5	15
20	15

**Supplementary Table S1.** HPLC gradient used for purification of **2**.

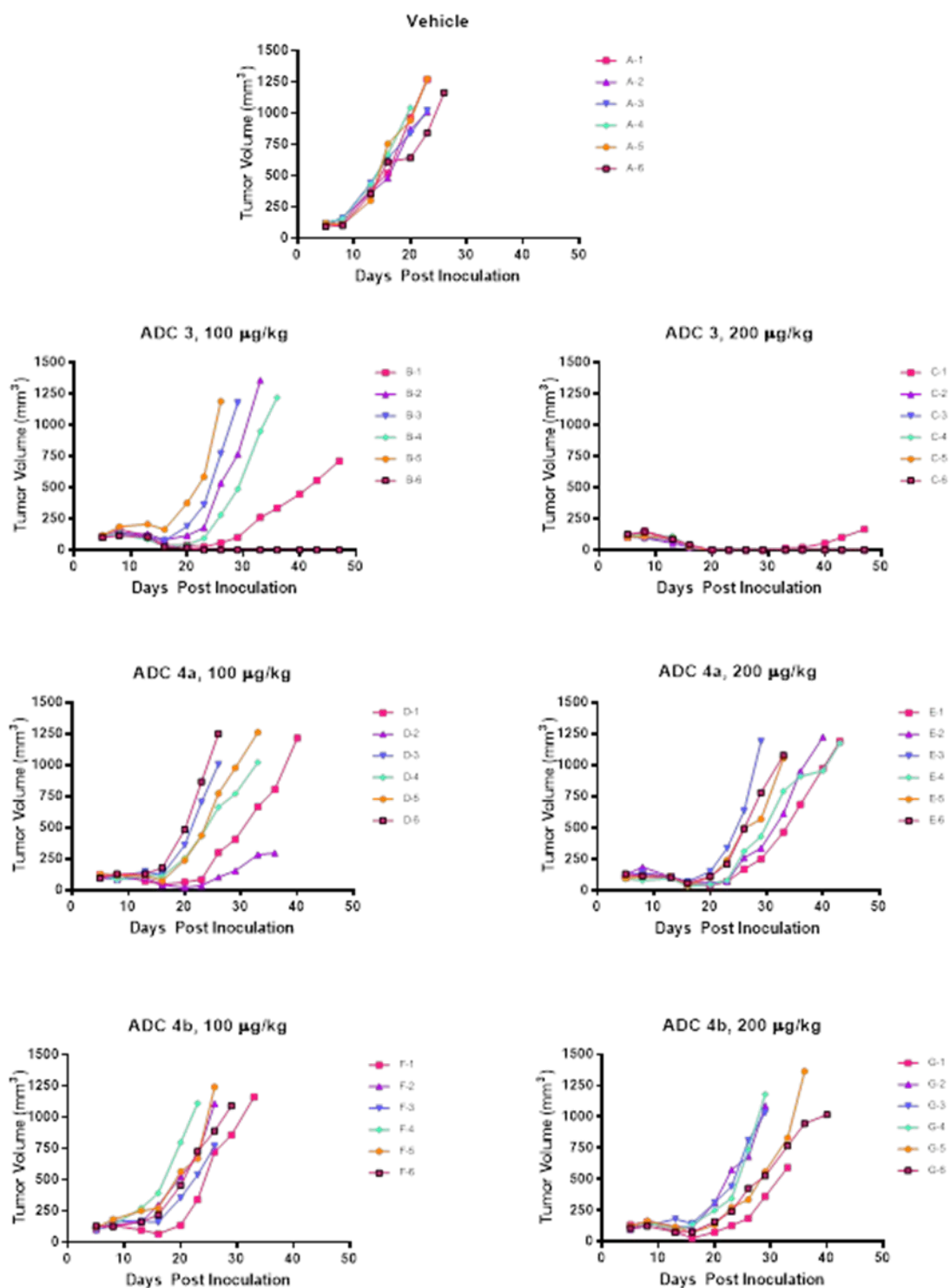
Product containing fractions were combined and concentrated *in vacuo* to give 130.2 mg of the desired Mal-CX-DM1. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  0.78 (d, *J* = 2.0 Hz, 3H), 1.12 (d, *J* = 6.4 Hz, 3H), 1.18 (d, *J* = 6.8 Hz, 3H), 1.22– 1.30 (m, 2H), 1.45 (tt, *J* = 10.2, 5.0 Hz, 2H), 1.59 (s, 3H), 1.61 – 1.72 (m, 3H), 2.04 (dt, *J* = 14.3, 2.7 Hz, 1H), 2.08 – 2.18 (m, 4H), 2.30 – 2.41 (m, 1H), 2.64 – 2.70 (m, 1H), 2.72 (s, 2H), 2.76 – 3.06 (m, 3H), 3.10 (d, *J* = 1.6 Hz, 2H), 3.14 – 3.23 (m, 4H), 3.32 – 3.55 (m, 10H), 3.64 (dd, *J* = 5.9, 3.2 Hz, 2H), 3.71 (t, *J* = 5.3 Hz, 4H), 3.86 (dd, *J* = 9.0, 3.9 Hz, 1H), 3.90 – 3.96 (m, 3H), 4.03 – 4.12 (m, 1H), 4.53 (dd, *J* = 12.0, 2.9 Hz, 1H),

5.31 (dp,  $J = 5.6, 3.9, 2.7$  Hz, 1H), 5.51 – 5.60 (m, 1H), 5.75 (s, 2H), 6.50 – 6.62 (m, 3H), 6.85 (s, 1H), 7.00 (s, 2H), 7.16 (td,  $J = 7.0, 6.4, 1.8$  Hz, 1H), 7.72 (t,  $J = 5.7$  Hz, 1H), 7.95 (t,  $J = 6.1$  Hz, 1H), 8.01 – 8.14 (m, 2H).  $^{13}\text{C}$  NMR (101 MHz, DMSO- $d_6$ )  $\delta$  11.86, 13.56, 14.88, 15.55, 23.59, 32.82, 35.68, 35.75, 37.30, 37.60, 38.19, 39.51, 40.57, 42.46, 42.58, 55.37, 56.60, 57.05, 60.50, 67.22, 73.65, 80.48, 88.69, 114.45, 117.62, 128.92, 133.07, 134.97, 141.71, 151.68, 155.77, 169.00, 169.50, 169.98, 170.91, 170.98, 171.52, 175.37. HRMS (ESI)  $m/z$ :  $[\text{M}+\text{H}]^+$  Calcd for  $\text{C}_{58}\text{H}_{77}\text{ClN}_{10}\text{O}_{19}\text{S}$  1285.4848; found 1285.4827

### Biochemical characterization of ADCs

ADC	UV DAR	SEC DAR	Monomer (%)	Free Drug (%)	H2L2 (%)	Yield (%)
<b>3</b> (Lys-conjugated M9436A-CX-DM1)	3.3	3.5	99.5	< 1	ND	91
<b>4a</b> (M9436A-C205-Mal- CX-DM1)	2.0	1.9	99.7	1.8	94	85
<b>4b</b> (M9436A-C400-Mal- CX-DM1)	1.7	2.1	99.0	< 1	93	96
<b>4c</b> (M9346A-C118-Mal- CX-DM1)	2.1	2.1	99.3	1.2	94	85
<b>4d</b> (M9346A-C239-Mal- CX-DM1)	2.0	2.1	99.6	3.5	92	87

**Supplementary Table S2.** Biochemical characterization of ADCs.



**Supplementary Figure S2.** Tumor growth curves for individual mice from the KB xenograft efficacy study depicted in Figure 7.