Synthesis of Cryptophane-223 Type Derivatives with dual functionalization

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- S4: ¹H NMR (400 MHz) spectrum of compound 4 recorded in CDCl₃ at 298 K.
- S5: ¹³C NMR (100.6 MHz) spectrum of compound 4 recorded in CDCl₃ at 298 K.
- S6: ¹H NMR (400 MHz) spectrum of CTB **5** recorded in CDCl₃ at 298 K.
- S7: ¹³C NMR (100.6 MHz) spectrum of CTB **5** recorded in CDCl₃ at 298 K.
- S8: ¹H NMR (400 MHz) spectrum of CTB **7** recorded in CDCl₃ at 298 K.
- S9: ¹³C NMR (100.6 MHz) spectrum of CTB 7 recorded in CDCl₃ at 298 K.
- S10: ¹H NMR (400 MHz) spectrum of compound **8** recorded in CDCl₃ at 298 K.
- S11: ¹³C NMR (100.6 MHz) spectrum of compound 8 recorded in CDCl₃ at 298 K.
- S12: ¹H NMR (400 MHz) spectrum of compound **9** recorded in CDCl₃ at 298 K.
- S13: ¹³C NMR (100.6 MHz) spectrum of compound **9** recorded in CDCl₃ at 298 K.
- S14: ¹H NMR (400 MHz) spectrum of compound **10** recorded in CDCl₃ at 298 K.
- S15: ¹³C NMR (100.6 MHz) spectrum of compound **10** recorded in CDCl₃ at 298 K.
- S16: ¹H NMR (400 MHz) spectrum of compound **11** recorded in DMSO-*d*₆ at 298 K.
- S17: 13 C NMR (100.6 MHz) spectrum of compound 11 recorded in DMSO- d_6 at 298K.

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- S18: ¹H NMR (400 MHz) spectrum of compound **14** recorded in CDCl₃ at 298 K.
- S19: ¹³C NMR (100.6 MHz) spectrum of compound **14** recorded in CDCl₃ at 298 K.
- S20: HSQC (400 MHz) spectrum of compound 14 recorded in CDCl₃ at 298 K.
- S21:¹H NMR (400 MHz) spectrum of compound 15 recorded in CDCl₃ at 298 K.
- S22: ¹³C NMR (100.6 MHz) spectrum of compound **15** recorded in CDCl₃ at 298 K.
- S23: HSQC (400 MHz) spectrum of compound 15 recorded in CDCl₃ at 298 K.
- S24: ¹H NMR (400 MHz) spectrum of compound **18** recorded in CDCl₃ at 298 K.
- S25: ¹³C NMR (100.6 MHz) spectrum of compound **18** recorded in CDCl₃ at 298 K.
- S26: HSQC (400 MHz) spectrum of compound 18 recorded in CDCl₃ at 298 K.
- S27: ¹H NMR (400 MHz) spectrum of compound **19** recorded in CDCl₃ at 298 K.
- S28: ¹³C NMR (100.6 MHz) spectrum of compound **19** recorded in CDCl₃ at 298 K.
- S29: HSQC (400 MHz) spectrum of compound 19 recorded in CDCl₃ at 298 K.
- S30: ¹H NMR (400 MHz) spectrum of compound **21** recorded in CDCl₃ at 298 K.
- S31: ¹³C NMR (100.6 MHz) spectrum of compound **21** recorded in CDCl₃ at 298 K.
- S32: HSQC (400 MHz) spectrum of compound 21 recorded in CDCl₃ at 298 K.
- S33: ¹H NMR (400 MHz) spectrum of compound **2** recorded in DMSO-*d*₆ at 298 K.
- S34: ¹³C NMR (100.6 MHz) spectrum of compound **2** recorded in DMSO-*d*₆ at 298 K.
- S35: HSQC (400 MHz) spectrum of compound 2 recorded in DMSO-d₆ at 298 K.
- S36: COSY (400 MHz) spectrum of compound 2 recorded in DMSO-d₆ at 298 K.
- S37: ¹H NMR (400 MHz) spectrum of compound **23** recorded in CDCl₃ at 298 K.
- S38: ¹³C NMR (100.6 MHz) spectrum of compound **23** recorded in CDCl₃ at 298 K.
- S39: HSQC (400 MHz) spectrum of compound 23 recorded in CDCl₃ at 298 K.
- S40: ORTEP representation of compound **9** (hydrogen atoms and solvent molecule were omitted for clarity. The displacement ellipsoids were plotted at 30% probability level).
- S41: Calorimetric titration of compound 2 in $H_2O/TRIS$ (20 mM; pH = 7.6). The solution host (c = 0.08 mM) was placed into the calorimeter cell (1.4 mL) and 28 successive aliquots (10 μ L) of Ni^{2+} solution (c = 1.0 mM) were added at 3 min intervals.

S42: Calorimetric titration of Cryptophane-222 hexacarboxylate in $H_2O/TRIS$ (20 mM; pH = 7.0). The solution host (c = 0.08 mM) was placed into the calorimeter cell (1.4 mL) and 28 successive aliquots (10 μ L) of Ni^{2+} solution (c = 1.0 mM) were added at 3 min intervals.

S43: Hyperpolarized 129-xenon spectra of compound 2 in TRIS buffer (20 mM, pH = 7.5). a) in absence of Zn^{2+} (green spectrum). b) in presence of 1 equiv. of Zn^{2+} (blue spectrum). c) in presence of 5 equiv. of Zn^{2+} (red spectrum). Spectra recorded at 25°C.

S44: hyperpolarized ¹²⁹Xe NMR spectrum of compound **10** recorded at 298 K in C₂D₂Cl₄.

Table S1: Crystallographic data of cryptophane 9.

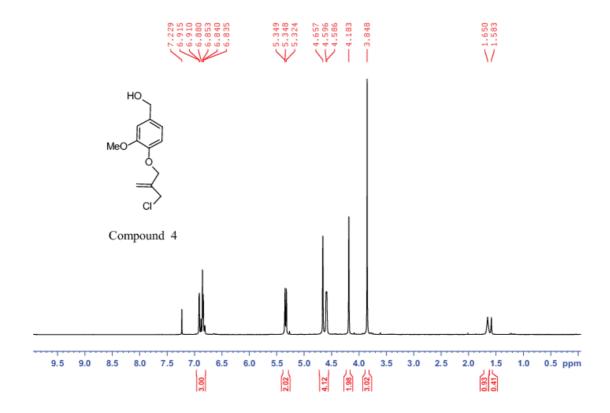


Figure S1: ¹H NMR (400 MHz) spectrum of compound **4** recorded in CDCl₃ at 298 K.

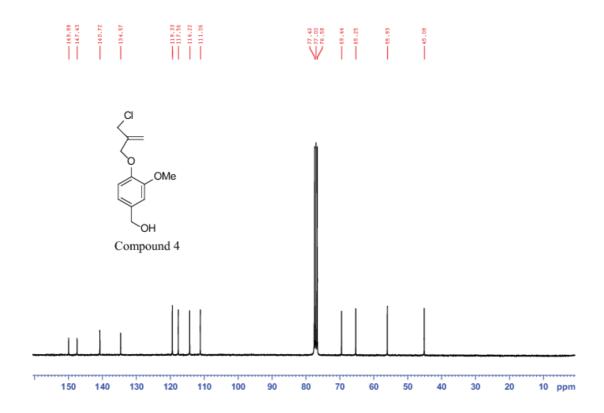


Figure S2: ¹³C NMR (100.6 MHz) spectrum of compound **4** recorded in CDCl₃ at 298 K.

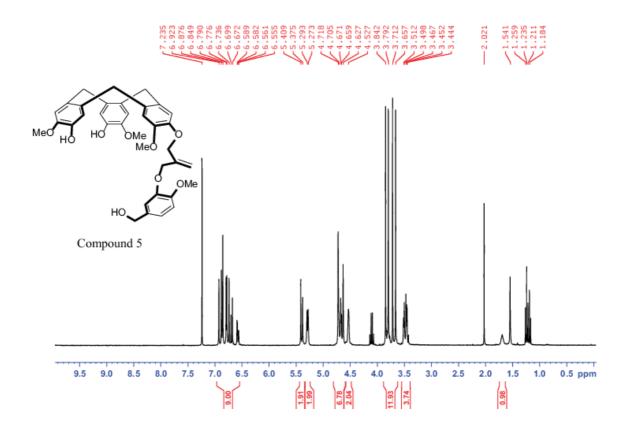


Figure S3: ¹H NMR (400 MHz) spectrum of CTB **5** recorded in CDCl₃ at 298 K.

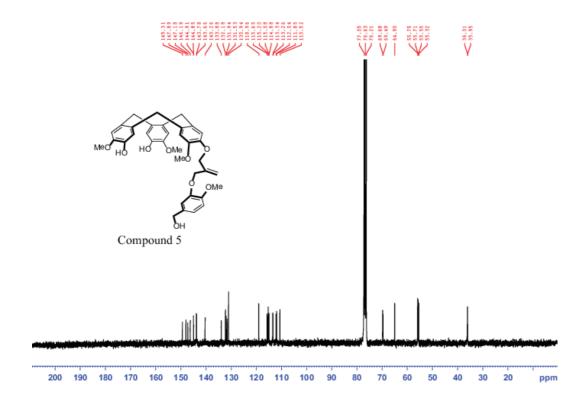


Figure S4: ¹³C NMR (100.6 MHz) spectrum of CTB **5** recorded in CDCl₃ at 298 K.

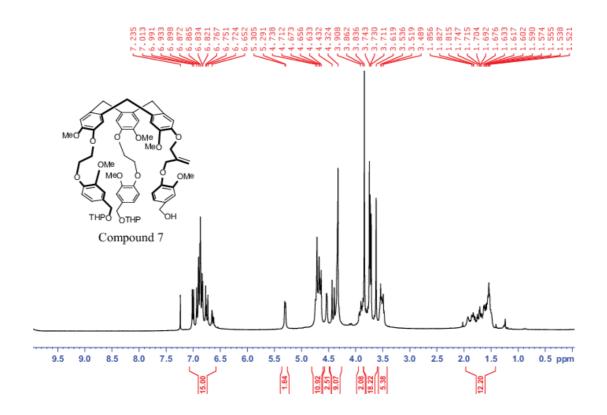


Figure S5: ¹H NMR (400 MHz) spectrum of CTB **7** recorded in CDCl₃ at 298 K.

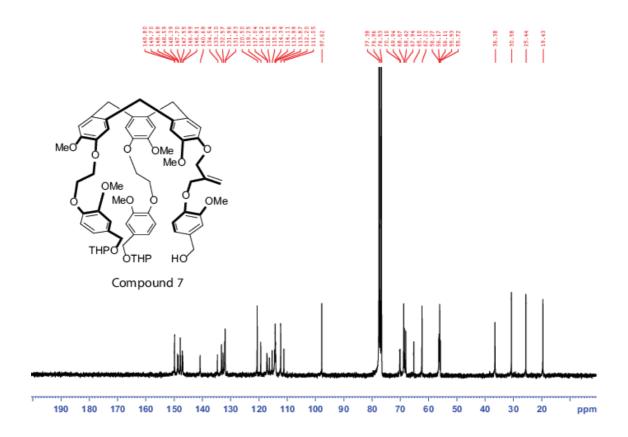


Figure S6: ¹³C NMR (100.6 MHz) spectrum of CTB **7** recorded in CDCl₃ at 298 K.

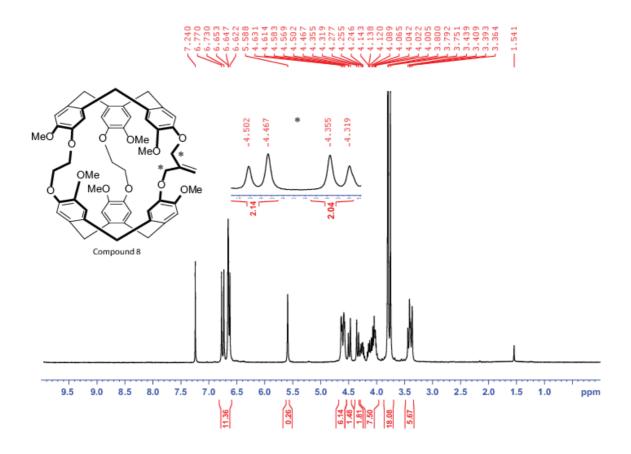


Figure S7: ¹H NMR (400 MHz) spectrum of compound 8 recorded in CDCl₃ at 298 K.

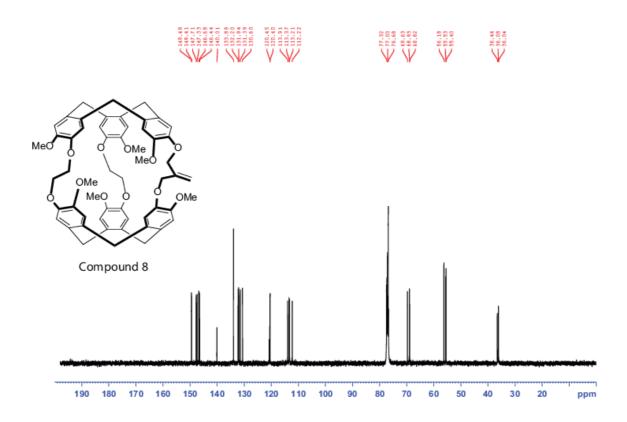


Figure S8: ¹³C NMR (100.6 MHz) spectrum of compound **8** recorded in CDCl₃ at 298 K.

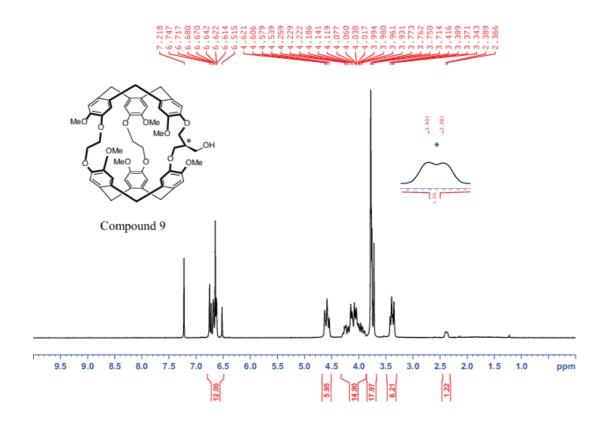


Figure S9: ¹H NMR (400 MHz) spectrum of compound **9** recorded in CDCl₃ at 298 K.

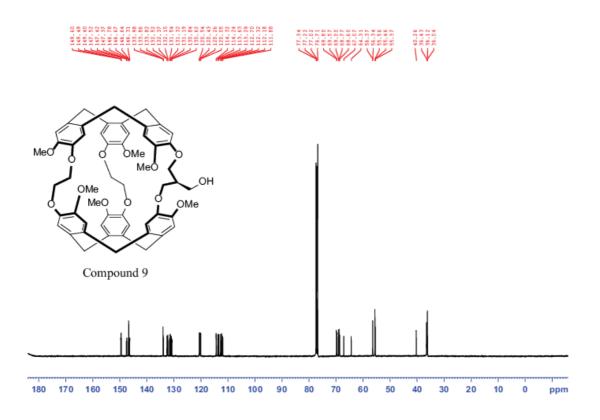


Figure S10: ¹³C NMR (100.6 MHz) spectrum of compound **9** recorded in CDCl₃ at 298 K.

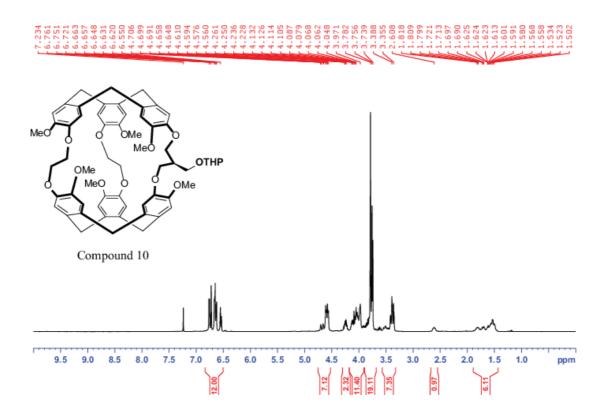


Figure S11: ¹H NMR (400 MHz) spectrum of compound **10** recorded in CDCl₃ at 298 K.

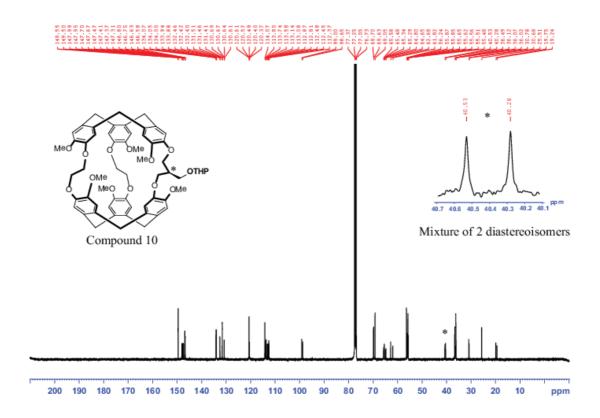


Figure S12: ¹³C NMR (100.6 MHz) spectrum of compound **10** recorded in CDCl₃ at 298 K.

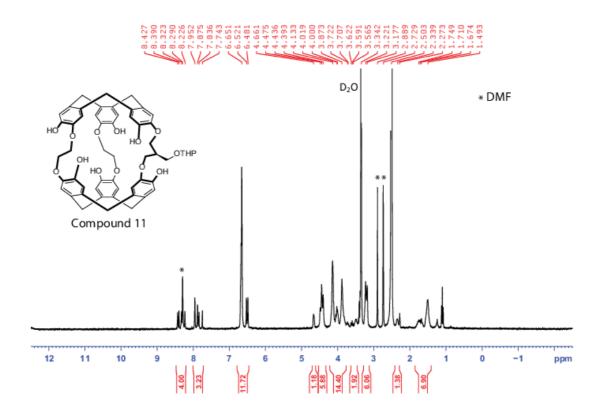


Figure S13: 1 H NMR (400 MHz) spectrum of compound **11** recorded in DMSO- d_{6} at 298 K.

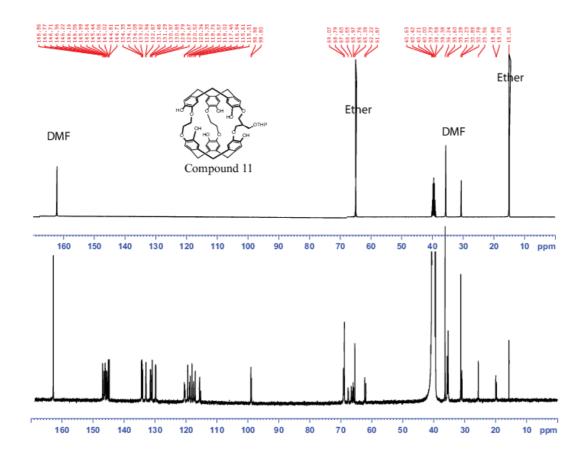


Figure S14: 13 C NMR (100.6 MHz) spectrum of compound **11** recorded in DMSO- d_6 at 298K.

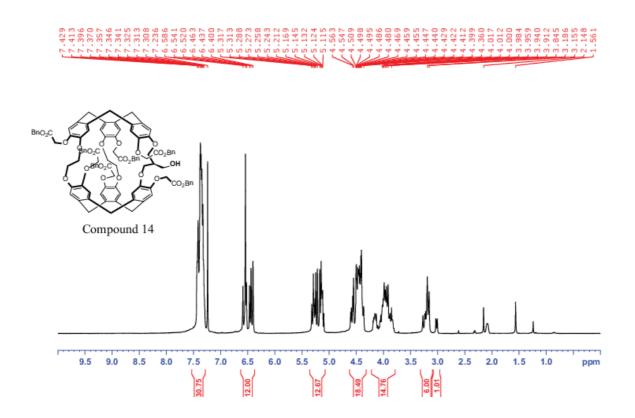


Figure S15: ¹H NMR (400 MHz) spectrum of compound **14** recorded in CDCl₃ at 298 K.

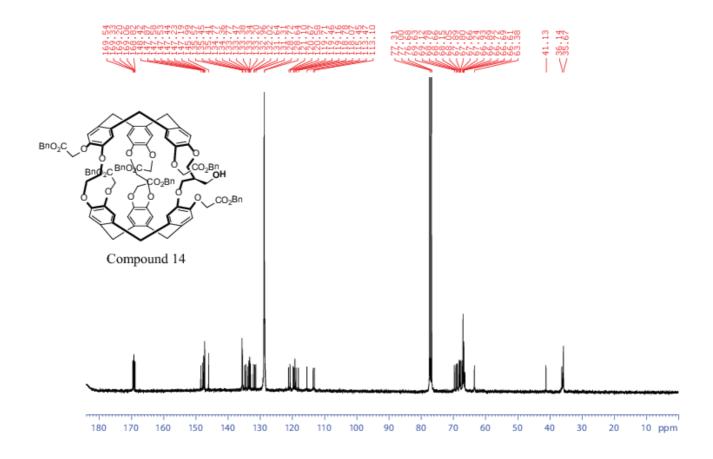


Figure S16: ¹³C NMR (100.6 MHz) spectrum of compound **14** recorded in CDCl₃ at 298 K.

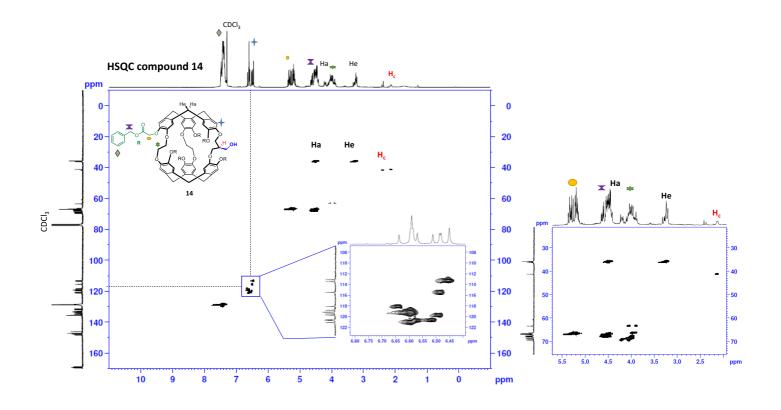


Figure S17: HSQC (400 MHz) spectrum of compound 14 recorded in CDCl₃ at 298 K.

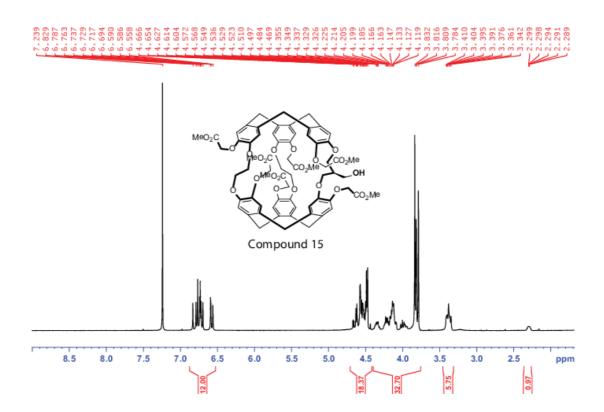


Figure S18: ¹H NMR (400 MHz) spectrum of compound **15** recorded in CDCl₃ at 298 K.

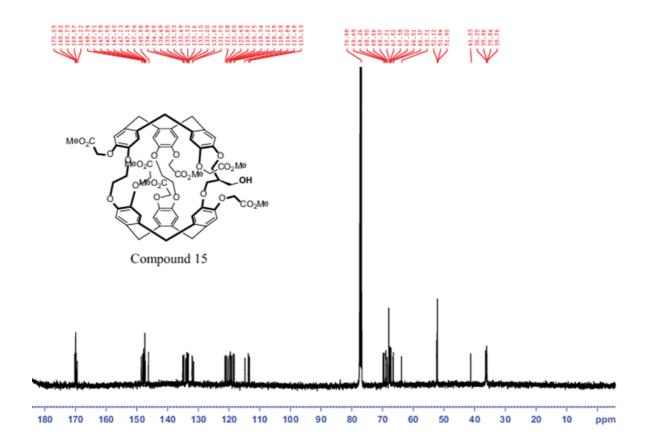


Figure S19: ¹³C NMR (100.6 MHz) spectrum of compound **15** recorded in CDCl₃ at 298 K.

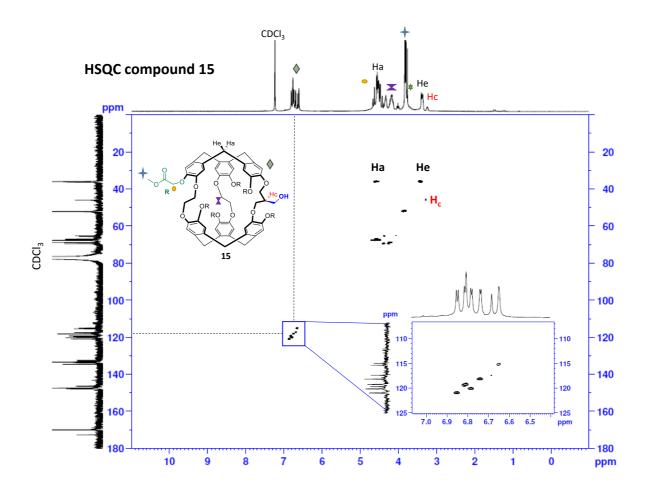


Figure S20: HSQC (400 MHz) spectrum of compound 15 recorded in CDCl₃ at 298 K.

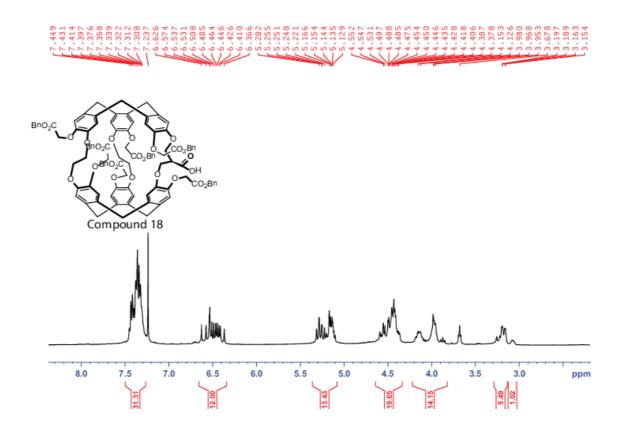


Figure S21: ¹H NMR (400 MHz) spectrum of compound **18** recorded in CDCl₃ at 298 K.

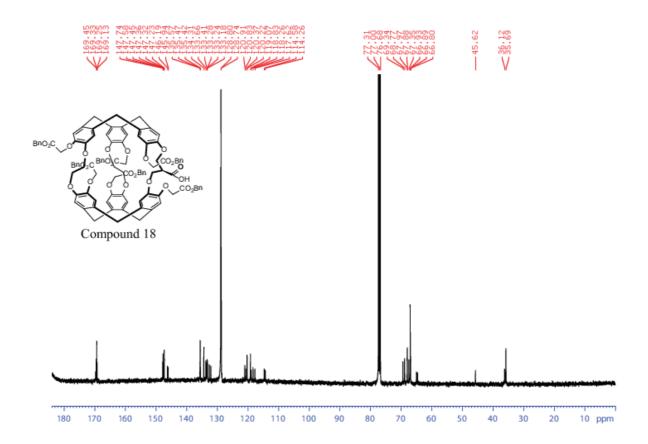


Figure S22: ¹³C NMR (100.6 MHz) spectrum of compound **18** recorded in CDCl₃ at 298 K.

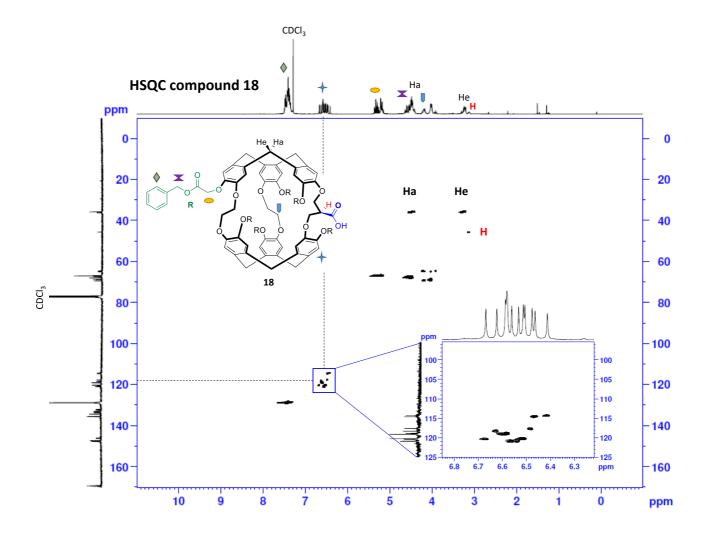


Figure S23: HSQC (400 MHz) spectrum of compound 18 recorded in CDCl₃ at 298 K.

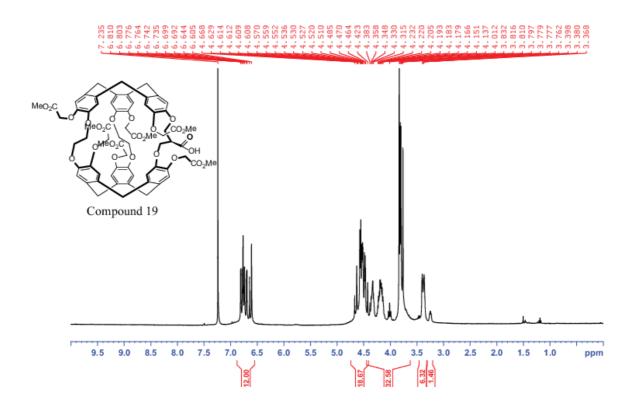


Figure S24: ¹H NMR (400 MHz) spectrum of compound **19** recorded in CDCl₃ at 298 K.

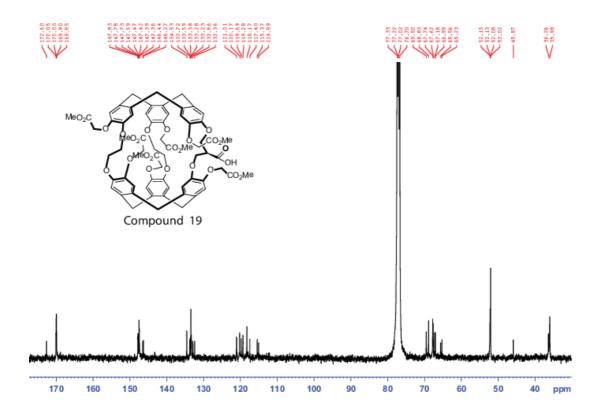


Figure S25: ¹³C NMR (100.6 MHz) spectrum of compound **19** recorded in CDCl₃ at 298 K.

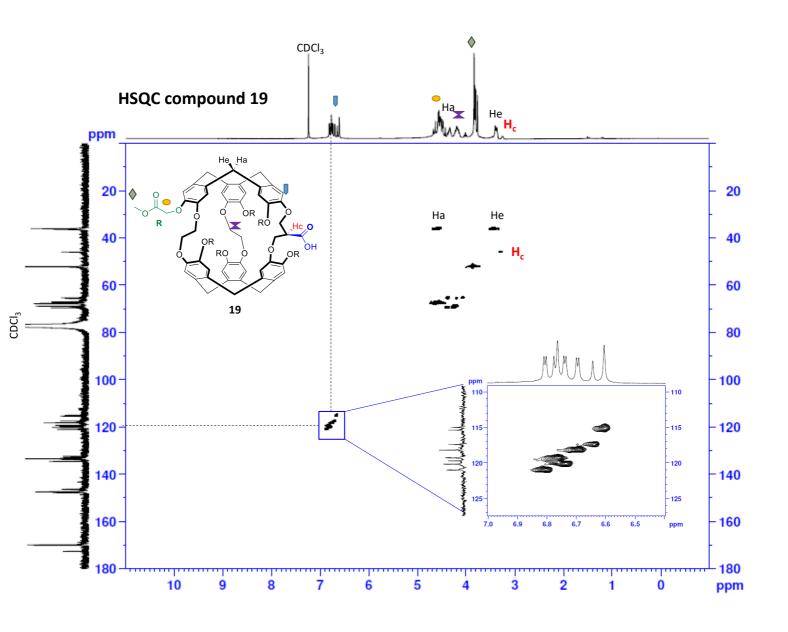


Figure S26: HSQC (400 MHz) spectrum of compound 19 recorded in CDCl₃ at 298 K.

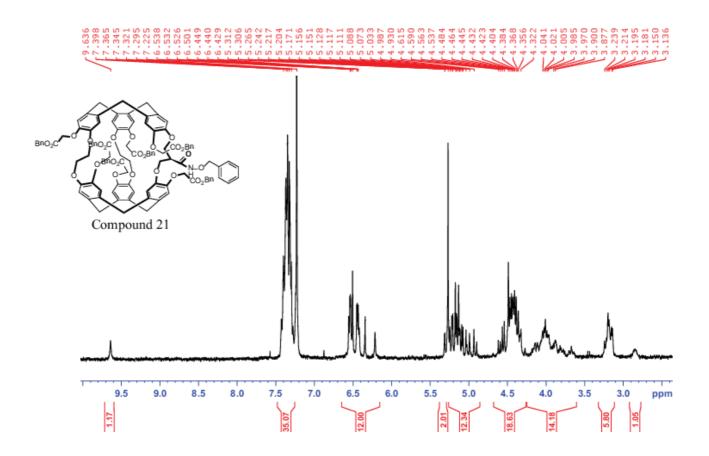


Figure S27: ¹H NMR (400 MHz) spectrum of compound **21** recorded in CDCl₃ at 298 K.

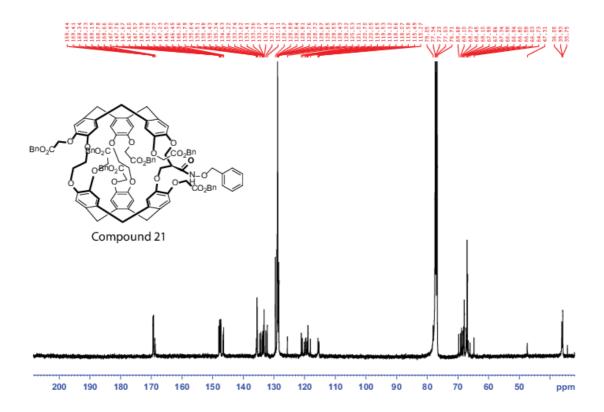


Figure S28: ¹³C NMR (400 MHz) spectrum of compound **21** recorded in CDCl₃ at 298 K.

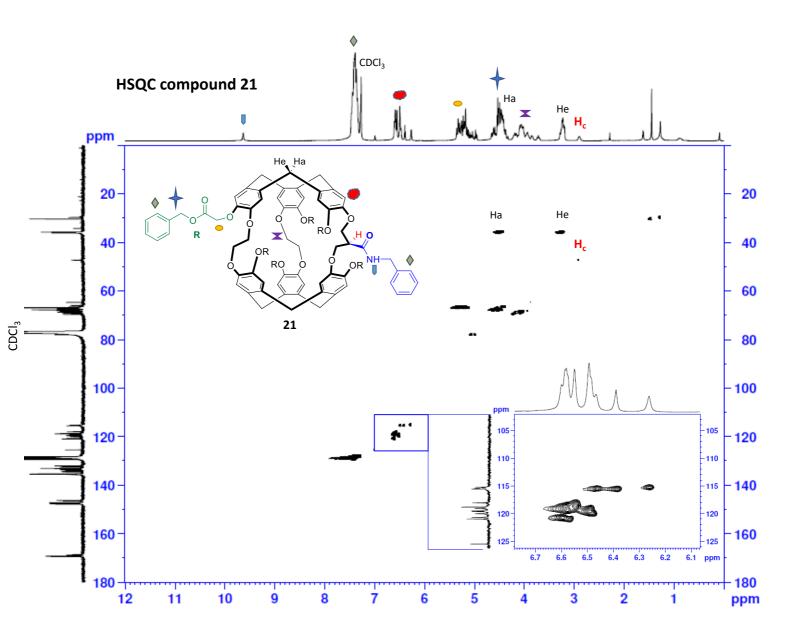


Figure S29: HSQC (400 MHz) spectrum of compound 21 recorded in CDCl₃ at 298 K.

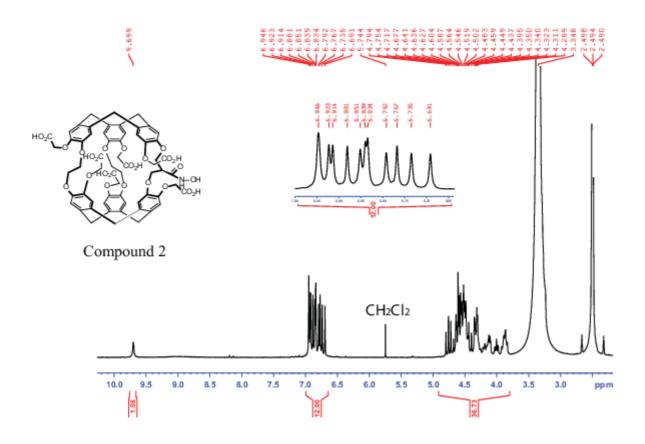


Figure S30: 1 H NMR (400 MHz) spectrum of compound **2** recorded in DMSO- d_{6} at 298 K.

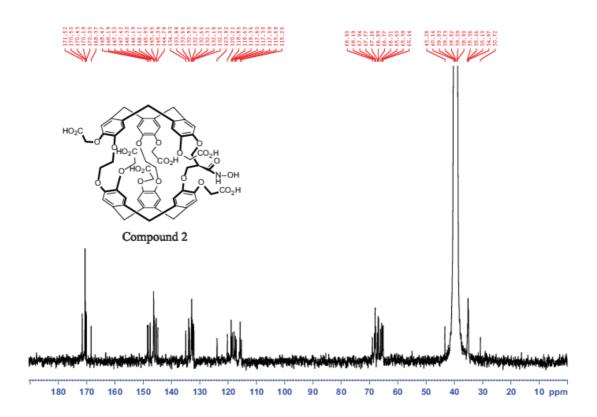


Figure S31: 13 C NMR (100.6 MHz) spectrum of compound **2** recorded in DMSO- d_6 at 298 K.

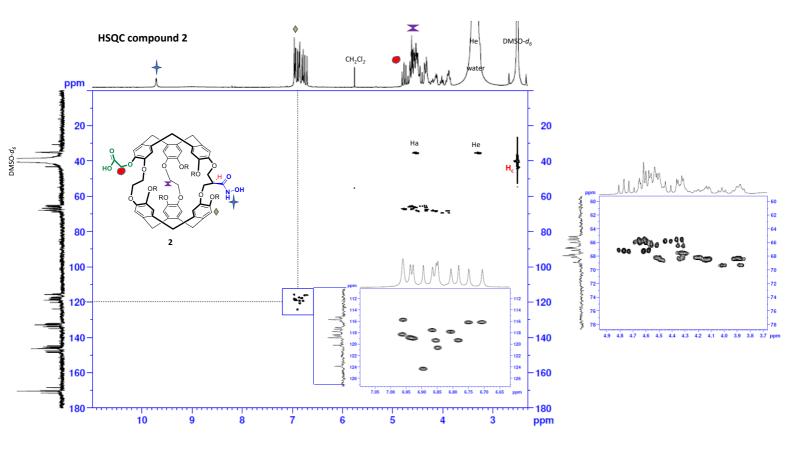


Figure S32: HSQC (400 MHz) spectrum of compound $\bf 2$ recorded in DMSO- d_6 at 298 K.

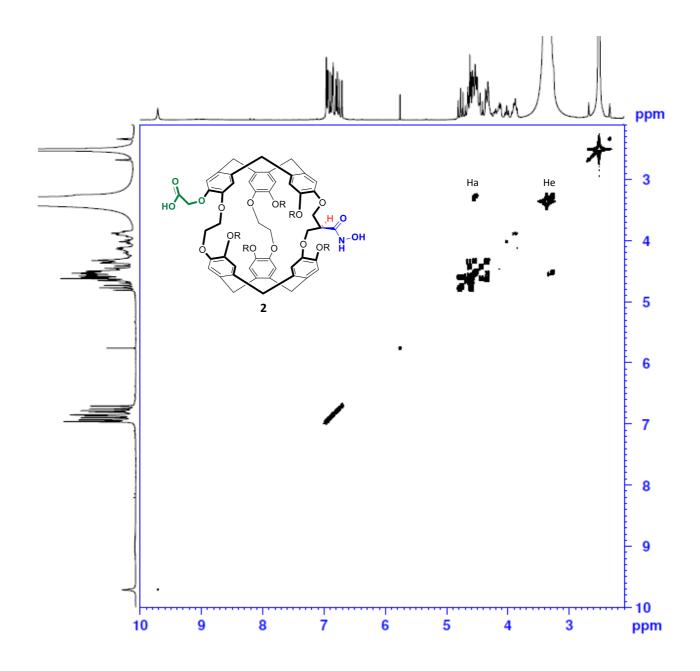


Figure S33: COSY (400 MHz) spectrum of compound 2 recorded in DMSO-d₆ at 298 K.

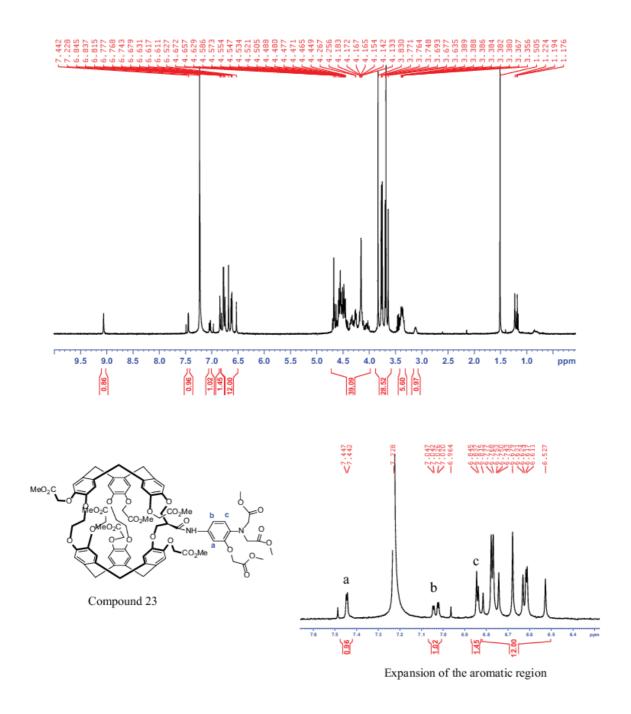


Figure S34: ¹H NMR (400 MHz) spectrum of compound **23** recorded in CDCl₃ at 298 K.

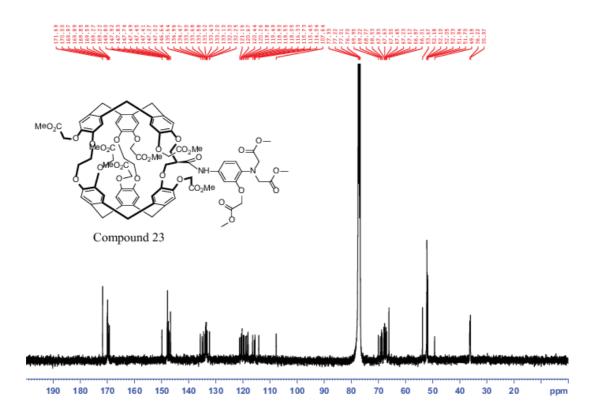


Figure S35: ¹³C NMR (100.6 MHz) spectrum of compound **23** recorded in CDCl₃ at 298 K.

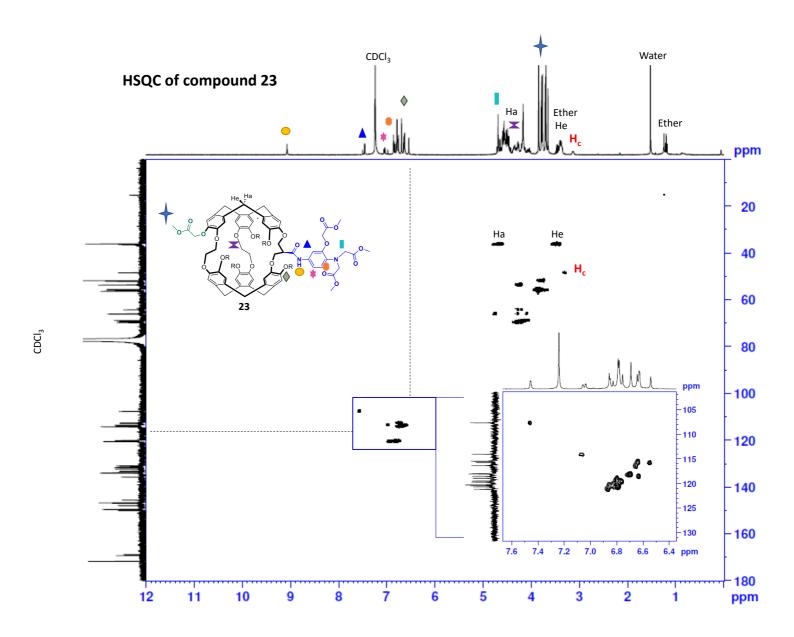


Figure S36: HSQC (400 MHz) spectrum of compound 23 recorded in CDCl₃ at 298 K.

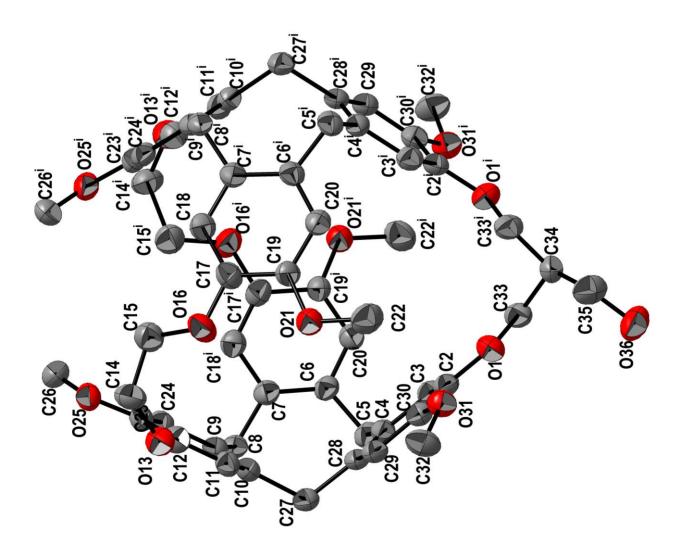


Figure S37: ORTEP representation of compound **9** (hydrogen atoms and solvent molecule were omitted for clarity. The displacement ellipsoids were plotted at 30% probability level).

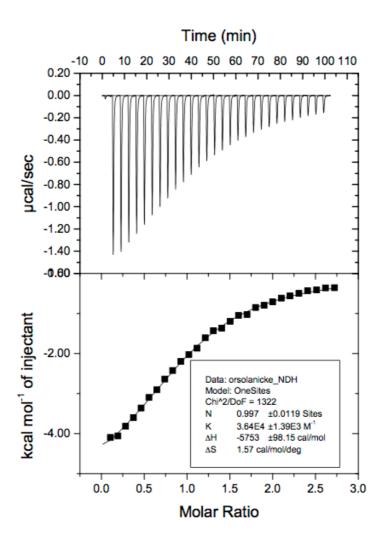


Figure S38: Calorimetric titration of compound 2 in $H_2O/TRIS$ (20 mM; pH = 7.6). The solution host (c = 0.08 mM) was placed into the calorimeter cell (1.4 mL) and 28 successive aliquots (10 μ L) of Ni^{2+} solution (c = 1.0 mM) were added at 3 min intervals.

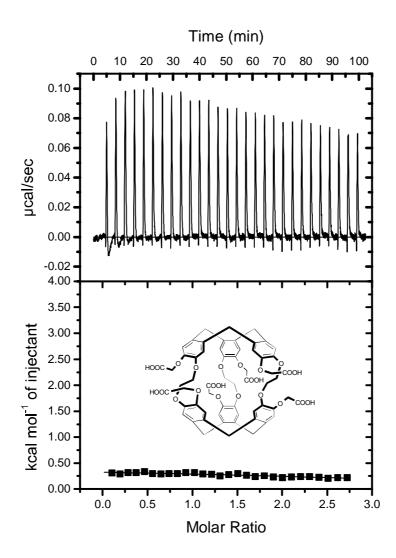


Figure S39: Calorimetric titration of Cryptophane-222 hexacarboxylate in $H_2O/TRIS$ (20 mM; pH = 7.0). The solution host (c = 0.08 mM) was placed into the calorimeter cell (1.4 mL) and 28 successive aliquots (10 μ L) of Zn^{2+} solution (c = 1.0 mM) were added at 3 min intervals.

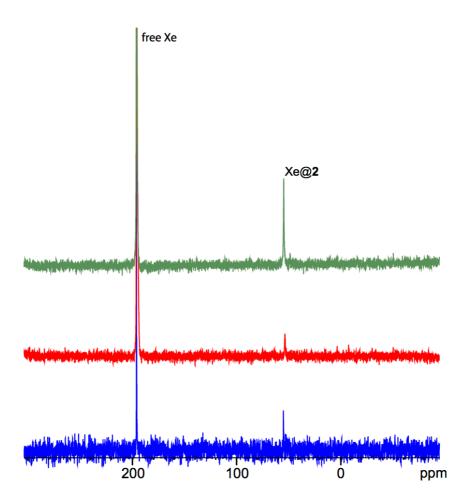


Figure S40: Hyperpolarized 129-xenon spectra (1 scann) of compound **2** in TRIS buffer (20 mM, pH = 7.5). a) in absence of Zn^{2+} (green spectrum). b) in presence of 1.0 equiv. of Zn^{2+} (blue spectrum). c) in presence of 5.0 equiv. of Zn^{2+} (red spectrum). Spectra recorded at 25°C. 0.39 mg of **1** in 1000 μ L of TRIS buffer (20 mM)/10% D_2O pH 7.5 (c = 0.32 mM).

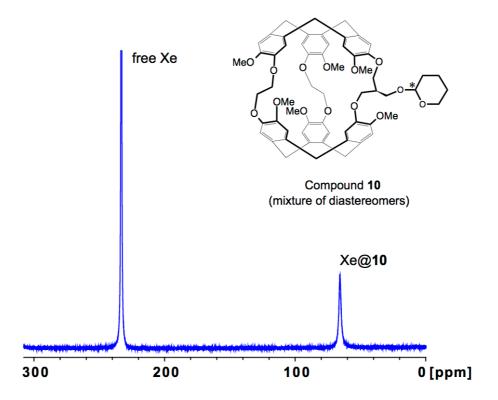


Figure S41: hyperpolarized 129 Xe NMR spectrum of compound **10** recorded at 298 K in $C_2D_2Cl_4$. 1.8 mg of cryptophane **10** in 600 μ L of 1,1,2,2-tetrachloroethane- d_2 .

Crystal data

| $C_{56}H_{58}O_{13}$ | $D_{\rm x} = 0.987 \; {\rm Mg \; m^{-3}}$ |
|----------------------------------|---|
| $M_r = 939.07$ | Cu Ka radiation, l = 1.54184 Å |
| Cubic, <i>I</i> 2 ₁ 3 | Cell parameters from 3819 reflections |
| Hall symbol: I 2b 2c 3 | q = 3.2–66.9° |
| a = 33.5971 (16) Å | $m = 0.57 \text{ mm}^{-1}$ |
| $V = 37923 (5) \text{ Å}^3$ | T = 150 K |
| Z = 24 | Plate, colorless |
| F(000) = 11952 | 0.40 × 0.32 × 0.19 mm |

Data collection

| Xcalibur, Atlas, Gemini ultra diffractometer | 10442 independent reflections |
|--|---|
| Radiation source: fine-focus sealed X-ray tube, Enhance Ultra (Cu) X-ray Source | 6339 reflections with $I > 2.0s(I)$ |
| Mirror monochromator | $R_{\rm int} = 0.069$ |
| Detector resolution: 10.4685 pixels mm ⁻¹ | $q_{\text{max}} = 67.0^{\circ}, q_{\text{min}} = 3.2^{\circ}$ |
| w scans | h = -38®28 |
| Absorption correction: analytical CrysAlis PRO 1.171.38.46 (Rigaku Oxford Diffraction, 2015) Analytical numeric absorption correction using a multifaceted crystal model based on expressions derived by R.C. Clark & J.S. Reid. (Clark, R. C. & Reid, J. S. (1995). Acta Cryst. A51, 887-897) Empirical absorption correction using spherical harmonics, implemented in SCALE3 ABSPACK scaling algorithm. | $k = -40 \circledast 10$ |
| $T_{\min} = 0.863, T_{\max} = 0.912$ | l = -29®26 |
| 25070 measured reflections | |

Refinement

| Refinement on F^2 | Hydrogen site location: difference Fourier map |
|----------------------------|--|
| Least-squares matrix: full | H-atom parameters constrained |
| $R[F^2 > 2s(F^2)] = 0.078$ | Method, part 1, Chebychev polynomial, (Watkin, 1994, P rince, 1982) [weight] = $1.0/[A_0*T_0(x) + A_1*T_1(x) \cdots + A_{n-1}]*T_{n-1}(x)]$ where A_i are the Chebychev coefficients listed below and $x = F$ //Fmax Method = Robust Weighting (P rince, 1982) W = [weight] * [1-(delta F /6*sigma F) 2] 2 A_i are: 347. 529. 246. 53.0 |
| $wR(F^2) = 0.189$ | $(D/s)_{max} = 0.003$ |
| S = 1.00 | $D\rho_{max} = 0.53 \text{ e Å}^{-3}$ |
| 645 parameters | Absolute structure: Parsons, Flack & Wagner (2013), 3517 Friedel Pairs |
| 60 restraints | Absolute structure parameter: 0.473 (13) |

Table S1: Crystallographic data of cryptophane 9.

Primary atom site location: other