

Combining Bidentate Lewis Acid Catalysis and Photochemistry: Formal Insertion of *o*-Xylene Into an Enamine Double Bond

Sebastian Ahles,^{†,‡} Julia Ruhl,^{†,‡} Marcel A. Strauss,^{†,‡} Hermann A. Wegner^{*,†,‡}

[†]Institute of Organic Chemistry, Justus Liebig University Giessen, Heinrich-Buff-Ring 17, 35392 Giessen, Germany

[‡]Center for Materials Research (LaMa), Justus Liebig University Giessen, Heinrich-Buff-Ring 16, 35392 Giessen, Germany

hermann.a.wegner@org.chemie.uni-giessen.de

General Information:	2
Synthesis:	4
Optimization of Irradiation Wavelength for the Domino IEDDA/PIRO Reaction:	23
Isomerization Monitoring of IEDDA/PIRO Product:	24
Degradation of BDLA-Phthalazine Complex by Irradiation:	25
NMR Spectra:	26
Computations:	56
References:	66

General Information:

Domino-IEDDA-reactions were set up in a nitrogen filled MBRAUN UNIIlab glove box. Other air and/or water sensitive reactions were carried out in a fume hood under Schlenk conditions.

NMR:

NMR spectra were measured on a Bruker Avance II 200 MHz, Avance II 400 MHz, Avance III 400 MHz HD or Avance III 600 MHz spectrometer at 25 °C if not otherwise noted. The ¹H (7.26 ppm) or ¹³C (77.16 ppm) chemical shift of internal residual CHCl₃ from CDCl₃ was used as reference. Boron trifluoride diethyl etherate was used for ¹⁹F (-153 ppm) as external reference.

UV/Vis:

Spectra were measured with a SPECORD® 200 PLUS UV/Vis spectrophotometer equipped with two automatic eightfold cell changers and a Peltier thermostat system for temperature control manufactured by Analytik Jena. The spectrophotometer system was operated by the software ASpec UV from Analytik Jena. The samples were measured in QS High Precision Cells made of Quartz Suprasil® by Hellma Analytics with a light path of 10 mm.

MS:

ESI-MS spectra were measured on a Bruker Micro TOF.

Chemicals:

The chemicals were purchased from Sigma-Aldrich, Acros Organics, Alfa Aesar and TCI Europe.

Anhydrous solvents were purchased from Acros Organics.

Deuterated solvents were purchased from Euriso - Top GmbH.

Solids were dried over Sicapent® and under high vacuum if necessary.

Technical grade solvents, used during work-up and purification, were distilled prior to use.

Aldehydes **6** were purified by distillation, degassed by freeze-pump-thaw-cycles, and stored in a nitrogen filled glove box.

Amines **7** were degassed by freeze-pump-thaw-cycles, dried either by distillation over CaH₂ or storage over molecular sieve 3 Å, and were stored in a nitrogen filled glove box.

Bidentate Lewis acid **BDLA** was synthesized as described in literature, and was stored in a nitrogen filled glove box.^{1,2}

Substituted phthalazines **1b-f**,^{3,4} and aldehyde **6d**⁵ were synthesized as described in literature.

Column Chromatography:

Flash column chromatography was carried out with Silica 60 M (0.04 – 0.063 mm) from Macherey-Nagel GmbH & Co. KG.

Thin layer chromatography was carried out on Polygram® SIL G/UV₂₅₄ from Macherey-Nagel GmbH & Co. KG.

In cases when NEt₃ was added to the eluent, the TLC plates were washed with cyclohexane containing 1% NEt₃ and dried prior to use. For a good separation the correct amount of NEt₃ is essential. It depends strongly on the acidity of used Silica.

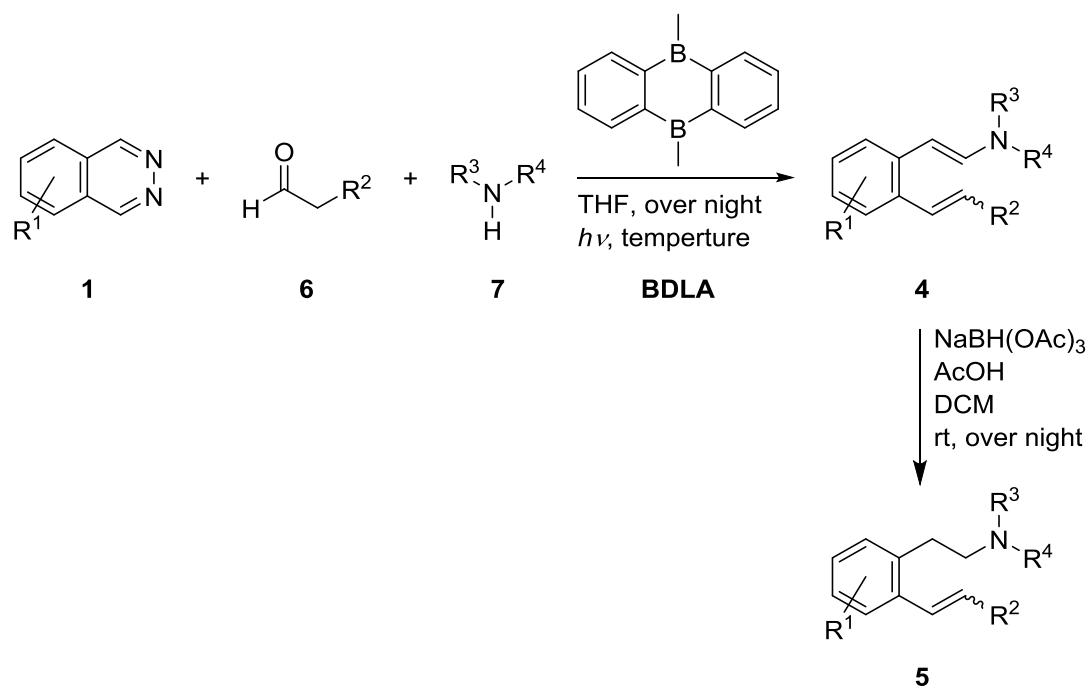
LEDs:

Following LEDs were used for irradiation:

$\lambda_{\text{max}} / \text{nm}$	$\Delta\lambda_{\text{FWHM}} / \text{nm}$	Typ
385	10	NCSU276AT-U385
405	12	NCSU276AT-U405
425 – 430	14	LHUV-0425-0650
448	20	LXML-PR01-0500
470	20	LXML-PB01-0030
500	30	NCSE119AT

Synthesis:

General Procedure for Domino IEDDA/PIRO Reaction Catalyzed by BDLA:



The reaction was set up in a nitrogen filled glovebox.

Phthalazine **1** and **BDLA** were suspended in THF (dry, degassed) (2.00 mL). Then, amine **7** was added, the reaction vessel was sealed and the remaining part of the reaction was carried out in a fume hood. The reaction mixture was heated/cooled to the reported temperature, and irradiated with a LED. Afterwards aldehyde **6** was added. Gas evolution was observed after a few minutes. For electron deficient phthalazines **1**, the reaction mixture was initially cooled, and the temperature was slowly increased until gas evolution started. The reaction was kept irradiated at this temperature overnight. Then, the yellow solution was concentrated under reduced pressure (10^{-1} mbar, volatile compounds were condensed into a cooling trap cooled by liquid nitrogen). The remaining oil (in some cases suspension) was transferred with DCM (dry, degassed) (3 x 2.00 mL) into a Schlenk tube containing $\text{NaBH}(\text{OAc})_3$. Then, AcOH was added. The yellow white suspension was stirred at rt overnight. Afterwards it was quenched by the addition of 1 M NaOH solution (15.0 mL). After gas evolution ceased the layers were separated. The aqueous one was extracted with DCM (3 x 10.0 mL). The combined organic fractions were washed with brine (20.0 mL), dried over Na_2SO_4 , filtered and concentrated under reduced pressure. The crude product was purified by flash column chromatography.

Phenethylamine **5a**



Starting materials:

Phthalazine (**1a**) (98.6 mg, 750 μmol , 1.00 equiv); **BDLA** (3.80 mg, 18.6 μmol , 2.49 mol %); pyrrolidine (**7a**) (74.7 μL , 900 μmol , 1.20 equiv); butyraldehyde (**6a**) (100 μL , 1.09 mmol, 1.45 equiv); $\text{NaBH}(\text{OAc})_3$ (328 mg, 1.50 mmol, 2.00 equiv); AcOH (90.4 μL , 1.50 mmol, 2.00 equiv).

LED:

425 – 430 nm

Temperature:

30 °C

Purification:

Flash column chromatography (2 times) (SiO_2 : 20 g, cyclohexane/ EtOAc + 1% NEt_3 , 2:1).

Yield: mixture of *E/Z*-isomers **E-5a/Z-5a** (1.00 : 1.78)

139 mg (604 μmol , 81%); pale yellow oil.

***E-5a*:**

^1H NMR (400 MHz, CDCl_3 with 0.03% v/v TMS): δ 7.46 – 7.39 (m, 1H, H4), 7.24 – 7.12 (m, 3H, H1, H2, H3), 6.65 (d, J = 15.6 Hz, 1H, H13), 6.14 (dt, J = 15.7, 6.6 Hz, 1H, H14), 2.95 – 2.86 (m, 2H, H7), 2.66 – 2.53 (m, 6H, H8 – H10), 2.31 – 2.20 (m, 4H, H15), 1.87 – 1.76 (m, 4H, H11, H12), 1.10 (t, J = 7.5 Hz, 3H, H16).

$^{13}\text{C}\{\text{H}\}$ NMR (101 MHz, CDCl_3 with 0.03% v/v TMS): δ 137.3 (C6), 137.0 (C5), 134.6 (C14), 129.9 (C1), 127.1 (C3), 126.6 (C4), 126.4 (C13), 126.0 (C2), 57.7 (C8), 54.4 (C9, C10), 33.4 (C7), 26.5 (C15), 23.6 (C11, C12), 14.0 (C16).

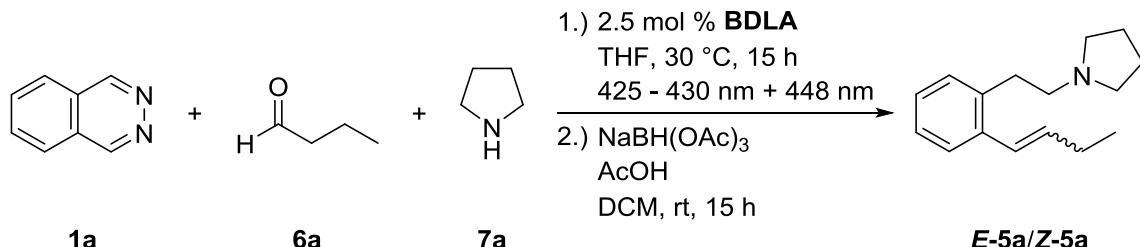
***Z-5a*:**

^1H NMR (400 MHz, CDCl_3 with 0.03% v/v TMS): δ 7.24 – 7.12 (m, 4H, H1' – H4'), 6.50 (d, J = 11.4 Hz, 1H, H13'), 5.71 (dt, J = 11.5, 7.4 Hz, 1H, H14'), 2.86 – 2.80 (m, 2H, H7'), 2.66 – 2.53 (m, 6H, H8' – H10'), 2.14 (tdd, J = 7.5, 7.4, 1.6 Hz, 2H, H15'), 0.99 (t, J = 7.5 Hz, 8H, H16').

$^{13}\text{C}\{\text{H}\}$ NMR (101 MHz, CDCl_3 with 0.03% v/v TMS): δ 138.7 (C6'), 136.8 (C5'), 135.2 (C14'), 129.7 (C4'), 129.4 (C1'), 127.1 (C13'), 127.1 (C3'), 125.9 (C2'), 57.4 (C8'), 54.3 (C9', C10'), 33.5 (C7'), 23.6 (C11', C12'), 21.9 (C15'), 14.5 (C16').

HRMS (ESI) m/z [M+H] $^+$ calc. for $\text{C}_{16}\text{H}_{24}\text{N}^+$: 230.1903; found: 230.1904.

Scaled up Synthesis of Phenethylamine **5a**



In case the reaction is scaled up it has to be ensured that the employed light source is strong enough or the reaction vessel is designed for photoreactions, i.e. thin diameter/large surface area. Insufficient irradiation led to the formation of IEDDA/amine group transfer product⁶, among others (see 3.00 mmol-Scale below).

1.50 mmol-Scale:

The reaction was set up in a nitrogen filled glovebox.

Phthalazine **1a** (394 mg, 3.00 mmol, 1.00 equiv) and **BDLA** (15.4 mg, 75.5 µmol, 2.52 mol %) were suspended in THF (dry, degassed) (4.00 mL). Then, pyrrolidine (**7a**) (300 µL, 3.62 µmol, 1.21 equiv) was added, the reaction vessel was sealed and the remaining part of the reaction was carried out in a fume hood. The colorless solution was heated to 30 °C, and irradiated with two LEDs (425 – 430 nm + 448 nm). Afterwards butyraldehyde (**6a**) (400 µL, 4.35 mmol, 1.45 equiv) was added. Gas evolution was observed almost immediately. The reaction was kept irradiated at 30 °C overnight. Then, the pale yellow solution was concentrated under reduced pressure (10⁻¹ mbar, volatile compounds were condensed into a cooling trap cooled by liquid nitrogen). The remaining yellow suspension was transferred with DCM (dry, degassed) (3 x 4.00 mL) into a Schlenk tube containing NaBH(OAc)₃ (670 mg, 1.50 mmol, 2.00 equiv). Then, AcOH (181 µL, 3.00 mmol, 2.00 equiv) was added, and the yellow white suspension was stirred at rt overnight. Afterwards it was quenched by the addition of 1 M NaOH solution (30.0 mL). After gas evolution ceased the layers were separated. The aqueous one was extracted with DCM (3 x 20.0 mL). The combined organic fractions were washed with brine (40.0 mL), dried over Na₂SO₄, filtered and concentrated under reduced pressure. The crude product was purified by flash column chromatography (SiO₂: 50 g, cyclohexane/EtOAc + 1% NEt₃, 2:1).

Yield:

mixture of *E/Z*-isomers **E-5a/Z-5a** (1.00 : 1.23)

269 mg (1.17 mmol, 78%); pale yellow oil.

3.00 mmol-Scale:

Starting materials (solvents were scaled up accordingly):

Phthalazine (**1a**) (394 mg, 3.00 mmol, 1.00 equiv); **BDLA** (15.4 mg, 75.5 μ mol, 2.52 mol %); pyrrolidine (**7a**) (300 μ L, 3.62 μ mol, 1.21 equiv); butyraldehyde (**6a**) (400 μ L, 4.35 mmol, 1.45 equiv); NaBH(OAc)₃ (1.34 g, 6.00 mmol, 2.00 equiv); AcOH (361 μ L, 6.00 mmol, 2.00 equiv).

LED (3x):

425 – 430 nm + 2 x 448 nm

Temperature:

30 °C

Purification:

Flash column chromatography (SiO₂: 100 g, cyclohexane/EtOAc + 1% NEt₃, 2:1).

Yield:

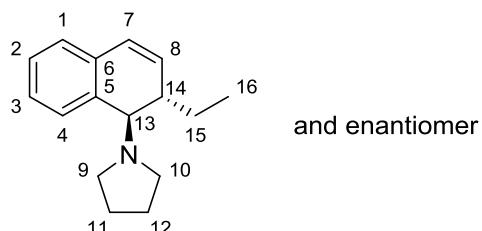
IEDDA/PIRO product

mixture of *E/Z*-isomers **E-5a/Z-5a** (1.10 : 1.00)

447 mg (1.95 mmol, 65%); pale yellow oil.

IEDDA/amine group transfer product⁶

38.3 mg (0.168 mmol, 6%); pale yellow oil.



¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS) : δ 7.22 (td, *J* = 7.3, 1.6 Hz, 1H, H2), 7.16 (td, *J* = 7.3, 1.5 Hz, 1H, H3), 7.10 (dd, *J* = 7.4, 1.5 Hz, 1H, H4), 7.07 (dd, *J* = 7.4, 1.4 Hz, 1H, H1), 6.44 (d, *J* = 9.6 Hz, 1H, H7), 6.07 (ddd, *J* = 9.6, 6.0, 1.1 Hz, 1H, H8), 3.47 (s, 1H, H13), 2.53 – 2.39 (m, 5H, H9, H10, H14), 1.72 – 1.59 (m, 4H, H11, H12), 1.39 – 1.23 (m, 2H, H15), 0.92 (t, *J* = 7.4 Hz, 3H, H16).

Phenethylamine **5b**



Starting materials:

Phthalazine (**1a**) (98.6 mg, 750 μmol , 1.00 equiv); **BDLA** (3.80 mg, 18.6 μmol , 2.49 mol %); pyrrolidine (**7a**) (74.7 μL , 900 μmol , 1.20 equiv); isovaleraldehyde (**6b**) (120 μL , 1.12 mmol, 1.49 equiv); NaBH(OAc)₃ (328 mg, 1.50 mmol, 2.00 equiv); AcOH (90.4 μL , 1.50 mmol, 2.00 equiv).

LED:

470 nm

Temperature:

30 °C

Purification:

Flash column chromatography (SiO₂: 20 g, cyclohexane/EtOAc + 1% NEt₃, 5:1).

Yield: mixture of *E*/*Z*-isomers **E-5b/Z-5b** (1.16 : 1.00)

132 mg (543 μmol , 72%); pale yellow oil.

***E-5b*:**

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 7.45 – 7.39 (m, H4), 7.23 – 7.12 (m, 3H, H1, H2, H3), 6.62 (dd, *J* = 15.7, 0.9 Hz, 1H, H13), 6.06 (dd, *J* = 15.7, 7.0 Hz, 1H, H14), 2.95 – 2.88 (m, 2H, H7), 2.67 – 2.55 (m, 6H, H8 – H10), 2.55 – 2.44 (m, 1H, H15), 1.88 – 1.77 (m, 7H, H11, H12), 1.10 (d, *J* = 6.7 Hz, 4H, H16, H17).

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 140.1 (14), 138.51 (C_{Ar} or C_{Ar'}'), 137.27 (C_{Ar} or C_{Ar'}'), 137.11 (C_{Ar} or C_{Ar'}'), 137.01 (C_{Ar} or C_{Ar'}'), 129.87 (C_{Ar} or C_{Ar'}'), 129.60 (C_{Ar} or C_{Ar'}'), 129.41 (C_{Ar} or C_{Ar'}'), 127.08 (C_{Ar} or C_{Ar'}'), 127.06 (C_{Ar} or C_{Ar'}'), 126.60 (C_{Ar} or C_{Ar'}'), 126.04 (C4), 125.95 (C_{Ar} or C_{Ar'}'), 124.4 (C13), 57.6 (C8), 54.4 (C9, C10), 33.3 (C7), 31.9 (C15), 23.6 (C11, C12), 22.7 (C16, C17).

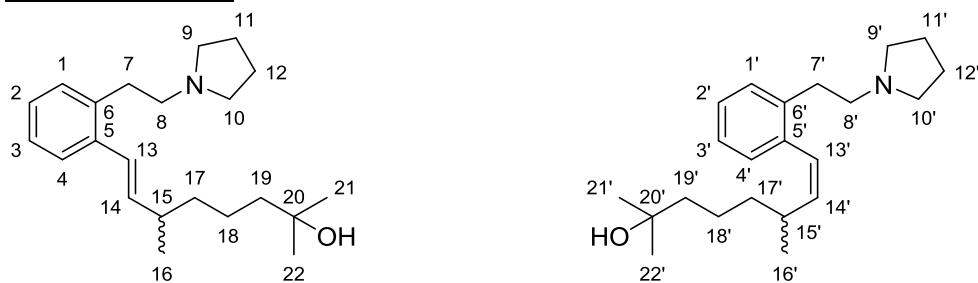
***Z-5b*:**

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 7.23 – 7.12 (m, 4H, H1' – H4'), 6.41 (d, *J* = 11.4 Hz, 1H, H13'), 5.53 (dd, *J* = 11.3, 10.3 Hz, 1H, H14'), 2.87 – 2.80 (m, 2H, H7'), 2.67 – 2.55 (m, 7H, H8' – H10', H15'), 1.88 – 1.77 (m, 7H, H11', H12'), 0.98 (d, *J* = 6.6 Hz, 4H, H16', H17').

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 140.9 (C14'), 138.51 (C_{Ar} or C_{Ar'}'), 137.27 (C_{Ar} or C_{Ar'}'), 137.11 (C_{Ar} or C_{Ar'}'), 137.01 (C_{Ar} or C_{Ar'}'), 129.87 (C_{Ar} or C_{Ar'}'), 129.60 (C_{Ar} or C_{Ar'}'), 129.41 (C_{Ar} or C_{Ar'}'), 127.08 (C_{Ar} or C_{Ar'}'), 127.06 (C_{Ar} or C_{Ar'}'), 126.60 (C_{Ar} or C_{Ar'}'), 126.04 (C_{Ar} or C_{Ar'}'), 125.95 (C4), 125.4 (C13'), 57.4 (C8'), 54.3 (C9', C10'), 33.4 (C7'), 27.3 (C15'), 23.6 (C11', C12'), 23.25 (C16', C17').

HRMS (ESI) m/z [M+H]⁺ calc. for C₁₇H₂₆N⁺: 244.2060; found: 244.2057.

Phenethylamine **5c**



Starting materials:

Phthalazine (**1a**) (98.6 mg, 750 μmol , 1.00 equiv); **BDLA** (3.80 mg, 18.6 μmol , 2.49 mol %); pyrrolidine (**7a**) (74.7 μL , 900 μmol , 1.20 equiv); 3,7-dimethyl-7-hydroxyoctanal (**6c**) (200 μL , 1.06 mmol, 1.41 equiv); $\text{NaBH}(\text{OAc})_3$ (328 mg, 1.50 mmol, 2.00 equiv); AcOH (90.4 μL , 1.50 mmol, 2.00 equiv).

LED:

470 nm

Temperature:

50 °C

Purification:

Flash column chromatography (SiO_2 : 20 g, cyclohexane/EtOAc + 1% NEt_3 , 2:1 to 1:1).

Yield: mixture of *E/Z*-isomers **E-5c/Z-5c** (1.46 : 1.00)

175 mg (531 μmol , 71%); pale yellow oil.

***E-5c*:**

^1H NMR (400 MHz, CDCl_3 with 0.03% v/v TMS): δ 7.44 – 7.36 (m, 1H, H4), 7.21 – 7.11 (m, 3H, 1, 2, 3), 6.62 (d, J = 15.6 Hz, 1H, H13), 5.91 (dd, J = 15.6, 8.0 Hz, 1H, H14), 2.94 – 2.86 (m, 2H, H7), 2.67 – 2.54 (m, 6H, H8, H9, H10), 2.34 (dt, J = 13.2, 6.7 Hz, 1H, H15), 1.99 (s, br, -OH), 1.88 – 1.76 (m, 4H, 11, 12), 1.57 – 1.21 (m, 6H, 17, 18, 19), 1.14 (s, 4H, H21, H22), 1.09 (d, J = 6.7 Hz, 3H, H16).

$^{13}\text{C}\{\text{H}\}$ NMR (101 MHz, CDCl_3 with 0.03% v/v TMS): δ 138.8 (C14), 137.3 (C6), 137.2 (C5), 129.8 (C1 or C2 or C3), 127.1 (C1 or C2 or C3), 126.6 (C1 or C2 or C3), 126.3 (C4), 126.1 (C13), 125.89, 70.8 (C20), 57.8 (C8), 54.3 (C9, C10), 44.1 (C19), 37.6 (C17), 37.5 (C15), 33.3 (C7), 29.4 (C21, C22), 23.5 (C11, C12), 22.1 (C18), 21.1 (C16).

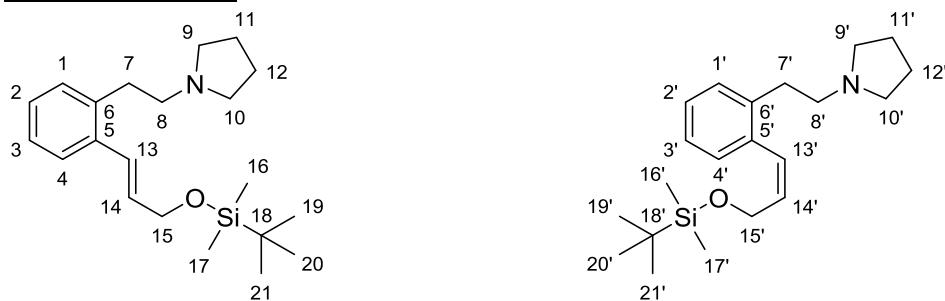
***Z-5c*:**

^1H NMR (400 MHz, CDCl_3 with 0.03% v/v TMS): δ 7.21 – 7.11 (m, 4H, 1', 2', 3', 4'), 6.47 (d, J = 11.5 Hz, 1H, H13'), 5.49 (dd, J = 11.4, 10.5 Hz, 1H, H14'), 2.87 – 2.80 (m, 2H, H7'), 2.67 – 2.54 (m, 6H, 8', 9', 10'), 2.48 (dq, J = 12.8, 6.4 Hz, 1H, H15'), 1.99 (s, br, -OH), 1.88 – 1.76 (m, 4H, 11', 12'), 1.57 – 1.21 (m, 6H, 17', 18', 19'), 1.20 (s, 6H, 21', 22'), 1.00 (d, J = 6.6 Hz, 2H, 16').

$^{13}\text{C}\{\text{H}\}$ NMR (101 MHz, CDCl_3 with 0.03% v/v TMS): δ 139.7 (C14'), 138.5 (C6'), 137.0 (C5'), 129.5 (C1' or C2' or C3' or C4'), 129.2 (C1' or C2' or C3' or C4'), 127.1 (C1' or C2' or C3' or C4'), 126.5 (C13'), 125.9 (C1' or C2' or C3' or C4'), 70.9 (C20'), 57.2 (C8'), 54.3 (C9', C10'), 44.2 (C19'), 38.2 (C17'), 33.2 (C7'), 32.3 (C15'), 29.5 (C21', C22'), 23.5 (C11', C12'), 22.3 (C18'), 21.3 (C16').

HRMS (ESI) m/z [M+H] $^+$ calc. for $\text{C}_{22}\text{H}_{36}\text{NO}^+$: 330.2791; found: 330.2790.

Phenethylamine **5d**



Starting materials:

Phthalazine (1a) (98.6 mg, 750 μmol , 1.00 equiv); **BDLA** (3.80 mg, 18.6 μmol , 2.49 mol %); pyrrolidine (**7a**) (74.7 μL , 900 μmol , 1.20 equiv); aldehyde **6d** (207 mg, 1.09 mmol, 1.45 equiv); $\text{NaBH}(\text{OAc})_3$ (328 mg, 1.50 mmol, 2.00 equiv); AcOH (90.4 μL , 1.50 mmol, 2.00 equiv).

LED:

425 – 430 nm

Temperature:

30 °C

Purification:

Flash column chromatography (2 times) (SiO_2 : 20 g, cyclohexane/ EtOAc + 3% NEt_3 , 10:1).

Yield: mixture of *E*/*Z*-isomers **E-5d/Z-5d** (1.00 : 1.53)

129 mg (374 μmol , 50%); pale yellow oil.

***E-5d*:**

^1H NMR (400 MHz, CDCl_3 with 0.03% v/v TMS): δ 7.49 – 7.41 (m, 1H, H4), 7.23 – 7.13 (m, 3H, H1, H2, H3), 6.88 (dt, J = 15.7, 1.9 Hz, 1H, H13), 6.19 (dt, J = 15.6, 5.1 Hz, 1H, H14), 4.37 (dd, J = 5.0, 1.8 Hz, 2H, H15), 2.95 – 2.87 (m, 2H, H7), 2.67 – 2.53 (m, 6H, H8 – H10), 1.86 – 1.75 (m, 4H, H11, H12), 0.94 (s, 9H, H19 – H21), 0.12 (s, 6H, H16, H17).

$^{13}\text{C}\{\text{H}\}$ NMR (101 MHz, CDCl_3 with 0.03% v/v TMS): δ 137.9 (C6), 136.1 (C5), 131.1 (C14), 130.0 (C1), 127.6 (C2 or C3), 126.9 (C13), 126.6 (C2 or C3), 126.2 (C4), 64.2 (15) 57.8 (C8), 54.3 (H9, H10), 33.3 (C7), 26.1 (C19 – C21), 23.6 (C11, C12), 18.6 (C18), -4.95 (C16, C17).

***Z-5d*:**

^1H NMR (400 MHz, CDCl_3 with 0.03% v/v TMS): δ 7.23 – 7.13 (m, 3H, H1', H2', H3'), 7.07 (d, J = 7.1 Hz, 1H, H4'), 6.64 (dt, J = 11.5, 1.7 Hz, 1H, H13'), 5.87 (dt, J = 11.6, 6.3 Hz, 1H, H14'), 4.29 (dd, J = 6.3, 1.6 Hz, 2H, H15'), 2.87 – 2.81 (m, 2H, H7'), 2.67 – 2.53 (m, 6H, H8' – H10'), 1.86 – 1.75 (m, 4H, H11', H12'), 0.87 (s, 9H, H19' – H21'), 0.01 (s, 6H, H16', H17').

$^{13}\text{C}\{\text{H}\}$ NMR (101 MHz, CDCl_3 with 0.03% v/v TMS): δ 138.7 (C6'), 135.7 (C5'), 132.8 (C14'), 129.6 (C1'), 129.6 (C4'), 128.5 (C13'), 127.6 (C2'), 125.9 (C3'), 60.4 (C15'), 57.4 (C8'), 54.3 (C9', C10'), 33.5 (C7'), 26.1 (C19' – C21'), 23.6 (C11', C12'), 18.4 (C18'), -4.98 (C16', C17').

HRMS (ESI) m/z [M+H] $^+$ calc. for $\text{C}_{21}\text{H}_{36}\text{NOSi}^+$: 346.2561; found: 346.2565.

Phenethylamine **5e**



Starting materials:

Phthalazine (1a) (98.6 mg, 750 μ mol, 1.00 equiv); **BDLA** (3.80 mg, 18.6 μ mol, 2.49 mol %); pyrrolidine (**7a**) (74.7 μ L, 900 μ mol, 1.20 equiv); pent-4-enal (**6e**) (100 μ L, 982 μ mol, 1.31 equiv); $\text{NaBH}(\text{OAc})_3$ (328 mg, 1.50 mmol, 2.00 equiv); AcOH (90.4 μ L, 1.50 mmol, 2.00 equiv).

LED:

448 nm

Temperature:

30 °C

Purification:

Flash column chromatography (2 times) (SiO_2 : 20 g, cyclohexane/ EtOAc + 1% NEt_3 , 10:1 to 5:1).

Yield: mixture of *E/Z*-isomers **E-5e/Z-5e** (1.24 : 1.00)

138 mg (573 μ mol, 76%).

E-5e:

^1H NMR (400 MHz, CDCl_3 with 0.03% v/v TMS): δ 7.46 – 7.40 (m, 1H, H4), 7.24 – 7.13 (m, 3H, H1, H2, H3), 6.69 (dt, J = 15.6, 1.7 Hz, 1H, H13), 6.11 (dt, J = 15.6, 6.7 Hz, 1H, H14), 5.97 – 5.80 (m, 1H, H16), 5.15 – 4.99 (m, 2H, H17), 3.01 – 2.95 (m, 2H, H15), 2.94 – 2.79 (m, 2H, H7), 2.67 – 2.52 (m, 6H, H8, H9, H10), 1.87 – 1.76 (m, 4H, H11, H12).

$^{13}\text{C}\{\text{H}\}$ NMR (101 MHz, CDCl_3 with 0.03% v/v TMS): δ 137.4 (C6), 136.7 (C16), 136.3 (C5), 130.21 (C_{Ar} or C_{Ar'}'), 129.9 (C14), 129.52 (C_{Ar} or C_{Ar'}'), 129.46 (C_{Ar} or C_{Ar'}'), 128.6 (C13), 127.33 (C_{Ar} or C_{Ar'}'), 127.32 (C_{Ar} or C_{Ar'}'), 126.6 (C_{Ar} or C_{Ar'}'), 126.1 (C4), 126.0 (C_{Ar} or C_{Ar'}'), 115.8 (C17), 57.7 (C8), 54.4 (C9, C10), 37.4 (C15), 33.3 (C7), 23.6 (C11, C12).

Z-5e:

^1H NMR (400 MHz, CDCl_3 with 0.03% v/v TMS): δ 7.24 – 7.13 (m, 4H, H_{Ar'}'), 6.64 (dt, J = 11.4, 1.6 Hz, 1H, H13'), 5.97 – 5.80 (m, 1H, H16'), 5.77 (dt, J = 11.4, 7.6 Hz, 1H, H14'), 5.15 – 4.99 (m, 2H, H17'), 2.94 – 2.79 (m, 4H, H7', H15'), 2.67 – 2.52 (m, 6H, H8', H9', H10'), 1.87 – 1.76 (m, 4H, H11', H12').

$^{13}\text{C}\{\text{H}\}$ NMR (101 MHz, CDCl_3 with 0.03% v/v TMS): δ 138.7 (C6'), 136.9 (C16'), 130.2 (C_{Ar} or C_{Ar'}'), 130.1 (C14'), 129.52 (C_{Ar} or C_{Ar'}'), 129.46 (C_{Ar} or C_{Ar'}'), 129.0 (C13'), 127.33 (C_{Ar} or C_{Ar'}'), 127.32 (C_{Ar} or C_{Ar'}'), 126.6 (C_{Ar} or C_{Ar'}'), 126.1 (C_{Ar} or C_{Ar'}'), 126.0 (C_{Ar} or C_{Ar'}'), 115.2 (C17'), 57.4 (C8'), 54.3 (C9', C10'), 33.5 (C7'), 32.7 (C15'), 23.6 (C11', C12').

HRMS (ESI) m/z [M+H]⁺ calc. for $\text{C}_{17}\text{H}_{24}\text{N}^+$: 242.1903; found: 242.1902.

Phenethylamine **5f**



Starting materials:

Phthalazine (**1a**) (98.6 mg, 750 μ mol, 1.00 equiv); **BDLA** (3.80 mg, 18.6 μ mol, 2.49 mol %); morpholine (**7b**) (79.6 μ L, 900 μ mol, 1.20 equiv); butyraldehyde (**6a**) (100 μ L, 1.09 mmol, 1.45 equiv); NaBH(OAc)₃ (328 mg, 1.50 mmol, 2.00 equiv); AcOH (90.4 μ L, 1.50 mmol, 2.00 equiv).

LED:

470 nm

Temperature:

45 °C

Purification:

Flash column chromatography (2 times) (SiO₂: 20 g, cyclohexane/EtOAc + 1% NEt₃, 2:1).

Yield: mixture of *E/Z*-isomers **E-5f/Z-5f** (1.00 : 1.12)

149 mg, (606 mmol, 81%); pale yellow oil.

***E-5f*:**

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 7.44 – 7.40 (m, 1H, H4), 7.22 – 7.12 (m, 3H, H1, H2, H3), 6.62 (dt, J = 15.5, 1.7 Hz, 1H, H13), 6.15 (dt, J = 15.6, 6.6 Hz, 1H, H14), 3.79 – 3.72 (m, 4H, H11, H12), 2.90 – 2.84 (m, 2H, H7), 2.57 – 2.47 (m, 6H, H8, H9, H10), 2.32 – 2.21 (m, 2H, H15), 1.10 (t, J = 7.5 Hz, 3H, H16).

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 136.9 (C5), 136.8 (C6), 134.8 (C14), 129.9 (C1), 129.7 (C_{Ar} or C_{Ar'}'), 127.13 (C_{Ar} or C_{Ar'}'), 127.10 (C_{Ar} or C_{Ar'}'), 126.7 (C_{Ar} or C_{Ar'}'), 126.3 (C13), 126.1 (C4), 126.0 (C_{Ar} or C_{Ar'}'), 67.1 (C11, C12), 60.2 (C8), 53.8 (C9, C10), 30.7 (C7), 26.5 (C15), 13.9 (C16).

***Z-5f*:**

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 7.22 – 7.12 (m, 4H, H_{Ar'}'), 6.48 (dt, J = 11.4, 1.7 Hz, 1H, H13'), 5.73 (dt, J = 11.4, 7.4 Hz, 1H, H14'), 3.79 – 3.72 (m, 4H, H11', H12'), 2.82 – 2.77 (m, 2H, H7'), 2.57 – 2.47 (m, 6H, H8', H9', H10'), 2.19 – 2.09 (m, 3H, H15'), 1.00 (t, J = 7.5 Hz, 4H, H16').

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 138.3 (C6'), 137.0 (C5'), 135.3 (C14'), 129.7 (C_{Ar} or C_{Ar'}'), 129.5 (C1'), 127.13 (C_{Ar} or C_{Ar'}'), 127.10 (C_{Ar} or C_{Ar'}'), 127.0 (C13'), 126.7 (C_{Ar} or C_{Ar'}'), 126.0 (C_{Ar} or C_{Ar'}'), 67.1 (C11', C12'), 59.9 (C8'), 53.8 (C9', C10'), 30.9 (C7'), 21.9 (C15'), 14.5 (C16').

HRMS (ESI) m/z [M+H]⁺ calc. for C₁₆H₂₄NO⁺: 246.1852; found: 246.1849.

Phenethylamine **5g**



Starting materials:

Phthalazine (**1a**) (98.6 mg, 750 μmol , 1.00 equiv); **BDLA** (3.80 mg, 18.6 μmol , 2.49 mol %); dipropylamine (**7c**) (125 μL , 900 μmol , 1.20 equiv); butyraldehyde (**6a**) (100 μL , 1.09 mmol, 1.45 equiv); $\text{NaBH}(\text{OAc})_3$ (328 mg, 1.50 mmol, 2.00 equiv); AcOH (90.4 μL , 1.50 mmol, 2.00 equiv).

LED:

470 nm

Temperature:

50 °C

Purification:

Flash column chromatography (2 times) (SiO_2 : 20 g, cyclohexane/ EtOAc + 1% NEt_3 , 30:1).

Yield: mixture of *E/Z*-isomers **E-5g/Z-5g** (5.83 : 1.00)

161 mg (621 μmol , 83%); pale yellow oil.

***E*-5g:**

^1H NMR (400 MHz, CDCl_3 with 0.03% v/v TMS): δ 7.44 – 7.39 (m, 1H, H4), 7.19 – 7.09 (m, 3H, H1, H2, H3), 6.64 (d, J = 15.6 Hz, 1H, H15), 6.14 (dt, J = 15.5, 6.5 Hz, 1H, H16), 2.84 – 2.78 (m, 2H, H7), 2.65 – 2.57 (m, 2H, H8), 2.52 – 2.42 (m, 4H, H9, H10), 2.25 (p, J = 7.3 Hz, 2H, H17), 1.57 – 1.43 (m, 4H, H11, H12), 1.11 (td, J = 7.4, 1.0 Hz, 3H, H18), 0.95 – 0.86 (m, 6H, H13, H14).

$^{13}\text{C}\{\text{H}\}$ NMR (101 MHz, CDCl_3 with 0.03% v/v TMS): δ 137.8 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 137.0 (C5), 135.1 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 134.6 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 134.5 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 130.0 (C1), 129.7 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 129.6 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 127.2 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 127.1 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 127.0 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 126.46 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 126.45 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 126.1 (C4), 125.7 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 56.4 (C9, C10), 55.2 (C8), 31.0 (C7), 26.5 (C17), 20.6 (C11, C12), 13.8 (C18), 12.1 (C13, C14).

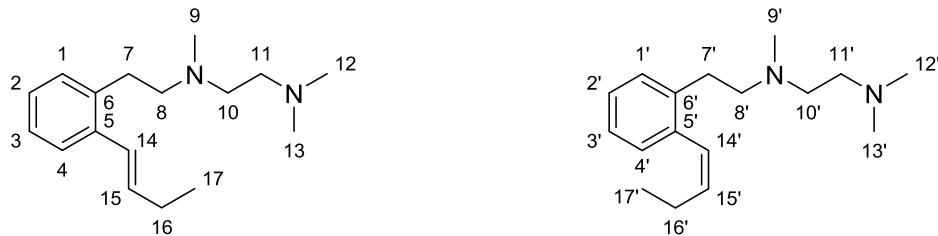
***Z*-5g:**

^1H NMR (400 MHz, CDCl_3 with 0.03% v/v TMS): δ 7.19 – 7.09 (m, 4H, H1', H2', H3', H4'), 6.50 (d, J = 11.4 Hz, 1H, H15'), 5.72 (dt, J = 11.4, 7.4 Hz, 1H, H16'), 2.78 – 2.70 (m, 2H, H7'), 2.65 – 2.57 (m, 2H, H8'), 2.52 – 2.42 (m, 4H, H9', H10'), 2.15 (p, J = 7.4 Hz, 2H, H17'), 1.57 – 1.43 (m, 4H, H11', H12'), 1.04 – 0.96 (td, J = 7.5, 0.9 Hz, 3H, H18'), 0.95 – 0.86 (m, 6H, H13', H14').

$^{13}\text{C}\{\text{H}\}$ NMR (101 MHz, CDCl_3 with 0.03% v/v TMS): δ 137.8 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 135.1 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 134.6 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 134.5 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 129.7 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 129.6 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 127.2 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 127.1 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 127.0 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 127.0 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 126.5 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 125.7 (C_{Ar} or $\text{C}_{\text{Ar}'}$), 56.3 (C9', C10'), 54.9 (C8'), 31.0 (C7'), 21.9 (C17'), 20.6 (C11', C12'), 14.4 (C18'), 12.1 (C13', C14').

HRMS (ESI) m/z [M+H]⁺ calc. for $\text{C}_{18}\text{H}_{30}\text{N}^+$: 260.2373; found: 260.2374.

Phenethylamine **5h**



Starting materials:

Phthalazine (**1a**) (98.6 mg, 750 μmol , 1.00 equiv); **BDLA** (3.80 mg, 18.6 μmol , 2.49 mol %); N,N,N'-trimethylethylenediamine (**7d**) (121 μL , 900 μmol , 1.20 equiv); butyraldehyde (**6a**) (100 μL , 1.09 mmol, 1.45 equiv); NaBH(OAc)₃ (328 mg, 1.50 mmol, 2.00 equiv); AcOH (90.4 μL , 1.50 mmol, 2.00 equiv).

LED:

470 nm

Temperature:

30 °C

Purification:

Flash column chromatography (2 times) (SiO₂: 20 g, cyclohexane/EtOAc/NEt₃, 10:10:1 to 5:5:1).

Yield: mixture of *E/Z*-isomers **E-5h/Z-5h** (2.09 : 1.00)

134 mg (515 μmol , 69%); pale yellow oil.

***E-5h*:**

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 7.44 – 7.39 (m, 1H, H4), 7.20 – 7.10 (m, 3H, H1, H2, H3), 6.62 (d, *J* = 15.6 Hz, 1H, H14) 6.14 (dt, *J* = 15.6, 6.5 Hz, 1H, H15), 2.88 – 2.83 (m, 2H, H7), 2.59 – 2.51 (m, 4H, H8, H10), 2.45 – 2.39 (m, 2H, H11), 2.36 (s, 3H, H9), 2.25 (s, 6H, H12, H13), 2.31 – 2.19 (m, 2H, H16), 1.10 (t, *J* = 7.5 Hz, 3H, H17).

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 137.3 (C6), 137.0 (C5), 134.6 (C15), 129.9 (C1), 127.1 (C2), 126.6 (C3), 126.4 (C14), 126.1 (C4), 59.3 (C8 or C10), 57.60 (C11), 55.6 (C8 or C10), 46.0 (C12, C13), 42.7 (C9), 31.3 (C7), 26.5 (C16), 13.9 (C17).

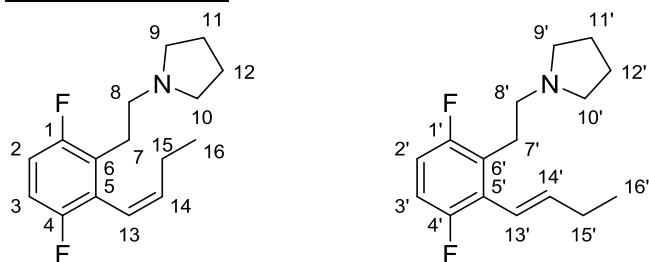
***Z-5h*:**

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 7.20 – 7.10 (m, 4H, H1', H2', H3', H4'), 6.48 (dt, *J* = 11.4, 1.5 Hz, 1H, H14'), 5.72 (dt, *J* = 11.4, 7.4 Hz, 1H, H15'), 2.80 – 2.75 (m, 2H, H7'), 2.59 – 2.51 (m, 4H, H8', H10'), 2.45 – 2.39 (m, 2H, H11'), 2.33 (s, 3H, H9'), 2.25 (s, 6H, H12', H13'), 2.15 (qd, *J* = 7.5, 1.6 Hz, 2H, H16'), 0.99 (t, *J* = 7.5 Hz, 3H, H17').

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 138.7 (C6'), 136.8 (C5'), 135.2 (C15'), 129.7 (C1' or C2' or C3' or C4'), 129.5 (C1' or C2' or C3' or C4'), 127.12 (C1' or C2' or C3' or C4' or C14'), 127.09 (C1' or C2' or C3' or C4' or C14'), 125.8 (C1' or C2' or C3' or C4'), 59.0 (C8' or C10'), 57.6 (C11'), 55.4 (C8' or C10'), 46.0 (C12', C13'), 42.6 (C9'), 31.2 (C7'), 21.9 (C16'), 14.5 (C17').

HRMS (ESI) m/z [M+H]⁺ calc. for C₁₇H₂₉N₂⁺: 261.2325; found: 261.2322.

Phenethylamine 5i



Starting materials:

5,8-Difluorophthalazine (**1b**) (62.3 mg, 375 μ mol, 1.00 equiv); **BDLA** (1.90 mg, 9.30 μ mol, 2.49 mol %); pyrrolidine (**7a**) (37.3 μ L, 450 μ mol, 1.20 equiv); butyraldehyde (50 μ L, 544 μ mol, 1.45 equiv); NaBH(OAc)₃ (164 mg, 735 μ mol, 2.00 equiv); AcOH (45.2 μ L, 750 μ mol, 2.00 equiv).

LED:

448 nm

Temperature:

-15 °C to 10 °C

Purification:

Flash column chromatography (2 times) (SiO₂: 20 g, cyclohexane/EtOAc + 1% NEt₃, 8:1).

Yield: mixture of E/Z-isomers E-5h/Z-5h (1.00 : 5.71)

58.0 mg (219 mmol, 58%); pale yellow oil.

E-5i:

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 6.93 – 6.78 (m, 2H, H₂, H₃), 6.30 – 6.25 (m, 2H, H₁₃, H₁₄), 2.97 – 2.88 (m, 2H, H₇), 2.64 – 2.47 (m, 6H, H₈, H₉, H₁₀), 2.32 – 2.22 (m, 2H, H₁₅), 1.86 – 1.73 (m, 4H, H₁₁, H₁₂), 1.10 (t, J = 7.5 Hz, 3H, H₁₆).

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 157.3 (dd, J = 240.3, 2.5 Hz, C₁, detected by hmbc experiment), 155.6 (dd, J = 240.2, 2.2 Hz, C₄, detected by hmbc experiment), 140.8 (d, J = 8.8 Hz, C₁₄), 126.7 (C₅), 119.3 (d, J = 2.7 Hz, C₁₃), 114.1 (dd, J = 27.1, 9.9 Hz, C₂), 113.6 (dd, J = 26.0, 9.4 Hz, C₃), 55.8 (C₈), 54.3 (C₉, C₁₀) 27.1 (C₁₅), 25.8 (C₇), 23.6 (C₁₁, C₁₂), 13.6 (C₁₆).

Z-5i:

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 6.93 – 6.78 (m, 2H, H_{2'}, H_{3'}), 6.13 (d, J = 11.2 Hz, 1H, H_{13'}), 5.89 (dt, J = 11.2, 7.3 Hz, 1H, H_{14'}), 2.86 – 2.79 (m, 2H, H_{7'}), 2.64 – 2.47 (m, 6H, H_{8'}, H_{9'}, H_{10'}), 1.94 (p, J = 7.4 Hz, 2H, H_{15'}), 1.86 – 1.73 (m, 4H, H_{11'}, H_{12'}), 0.96 (t, J = 7.5 Hz, 1H, H_{16'}).

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 157.3 (dd, J = 240.3, 2.5 Hz, C_{1'}), 155.6 (dd, J = 240.2, 2.2 Hz, C_{4'}), 139.14 (d, J = 1.4 Hz, C_{14'}), 127.64 (dd, J = 17.7, 3.3 Hz, C_{6'}), 126.2 (dd, J = 19.1, 4.9 Hz, C_{5'}) 118.8 (d, J = 2.1 Hz, C_{13'}), 114.5 (dd, J = 25.7, 9.0 Hz, C_{2'}), 113.8 (dd, J = 26.3, 9.2 Hz, C_{3'}), 55.5 (C_{8'}), 54.2(C_{9'}, C_{10'}), 26.3 (C_{7'}) 23.6 (C_{11'}, C_{12'}), 22.8 (d, J = 2.4 Hz, C_{15'}), 13.5 (C_{16'}).

HRMS (ESI) m/z [M+H]⁺ calc. for C₁₆H₂₂F₂N⁺: 266.1715; found: 266.1713.

Phenethylamine 5j



Starting materials:

6-(Trifluoromethyl)phthalazine (**1c**) (149 mg, 750 μmol , 1.00 equiv); **BDLA** (3.80 mg, 18.6 μmol , 2.49 mol %); pyrrolidine (**7a**) (74.7 μL , 900 μmol , 1.20 equiv); butyraldehyde (100 μL , 1.09 mmol, 1.45 equiv); $\text{NaBH}(\text{OAc})_3$ (328 mg, 1.50 mmol, 2.00 equiv); AcOH (90.4 μL , 1.50 mmol, 2.00 equiv).

LED:

448 nm

Temperature:

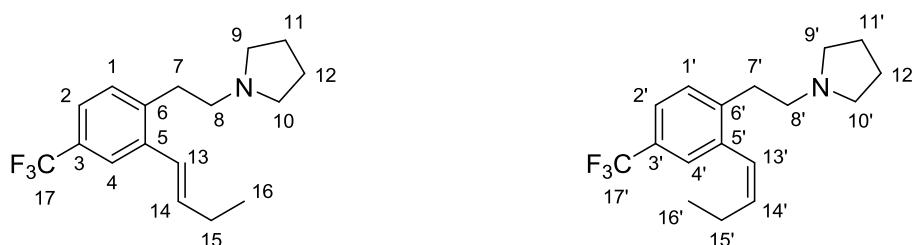
-10 °C to 0 °C

Purification:

Flash column chromatography (2 times) (SiO_2 : 20 g, cyclohexane/EtOAc + 1% NEt_3 , 8:1).

Yield: mixture of *E/Z*-isomers *E*-5j/*Z*-5j (9.00 : 1.00)

67.0 mg (225 μmol , 30%); pale yellow oil.



***E*-5j:**

^1H NMR (400 MHz, CDCl_3 with 0.03% v/v TMS): δ 7.64 (d, $J = 1.9$ Hz, 1H, H4), 7.37 (dd, $J = 8.1, 1.9$ Hz, 1H, H2), 7.25 (d, $J = 7.8$ Hz, 1H, H1), 6.64 (dt, $J = 15.6, 1.7$ Hz, 1H, H13), 6.21 (dt, $J = 15.6, 6.5$ Hz, 1H, H14), 2.97 – 2.90 (m, 2H, H7), 2.66 – 2.54 (m, 6H, H8, H9, H10), 2.31 – 2.22 (m, 2H, H15), 1.87 – 1.78 (m, 4H, H11, H12), 1.11 (t, $J = 7.5$ Hz, 3H, H16).

$^{13}\text{C}\{\text{H}\}$ NMR (101 MHz, CDCl_3 with 0.03% v/v TMS): δ 141.1 (q, $J = 1.2$ Hz, C6, only visible as doublet due to low intensity), 137.7 (C5), 136.5 (C14), 130.2 (C1), 128.9 (q, $J = 32.0$ Hz, C3), 125.4 (C13), 124.5 (q, $J = 272.0$ Hz, C17), 123.5 (q, $J = 3.8$ Hz, C2), 122.9 (q, $J = 3.8$ Hz, C4), 57.1 (C8), 54.4 (C9, C10), 33.2 (C7), 26.5 (C15), 23.6 (C11, C12), 13.8 (C16).

***Z*-5j:**

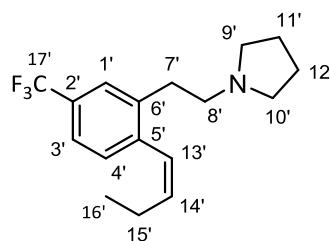
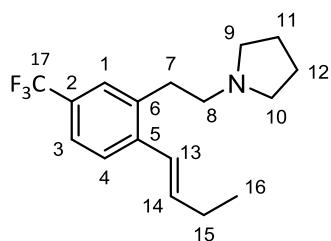
^1H NMR (400 MHz, CDCl_3 with 0.03% v/v TMS): δ 7.42 (dd, $J = 8.1, 2.0$ Hz, 1H, H2'), 7.38 (s, 1H, H4', detected by HMQC experiment), 7.30 (d, $J = 8.0$ Hz, 1H, H1'), 6.47 (d, $J = 11.5$ Hz, 1H, H13'), 5.79 (dt, $J = 11.4, 7.4$ Hz, 1H, H14'), 2.89 – 2.82 (m, 2H, H7'), 2.66 – 2.54 (m, 6H, H8', H9', H10'), 2.11 (pd, $J = 7.5, 1.7$ Hz, 2H, H15'), 1.87 – 1.78 (m, 4H, H11', H12'), 1.00 (t, $J = 7.5$ Hz, 3H, H16').

$^{13}\text{C}\{\text{H}\}$ NMR (101 MHz, CDCl_3 with 0.03% v/v TMS): δ 142.8 (C6'), 137.4 (C5'), 136.6 (C14'), 129.8 (C1'), 126.4 (q, $J = 3.6$ Hz, C4', only visible as doublet due to low intensity), 126.0 (C13') 123.8 (q, $J = 3.7$ Hz, C2', only visible as doublet due to low intensity), 56.9 (C8'), 54.3 (C9', C10'), 33.3 (C7'), 23.6 (C11', C12'), 21.9 (C15'), 14.3 (C16').

HRMS (ESI) m/z [M+H]⁺ calc. for C₁₇H₂₃F₃N⁺: 298.1777; found: 298.1780.

Yield: mixture of *E/Z*-isomers *E*-5j'/*Z*-5j' (3.12 : 1.00)

54.0 mg (182 µmol, 24%); pale yellow oil.



***E*-5j':**

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 7.50 (d, *J* = 8.5 Hz, 1H, H4), 7.45 – 7.36 (m, 2H, H1, H3), 6.65 (d, *J* = 15.7 Hz, 1H, H13), 6.22 (dt, *J* = 15.6, 6.6 Hz, 1H, H14), 2.98 – 2.91 (m, 2H, H7), 2.68 – 2.55 (m, 6H, H8, H9, H10), 2.32 – 2.21 (m, 2H, H15), 1.88 – 1.78 (m, 4H, H11, H12), 1.11 (t, *J* = 7.4 Hz, 3H, H16).

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 140.6 (q, *J* = 1.5 Hz, C5, only visible as doublet due to low intensity), 137.7 (C6), 137.1 (C14), 128.9 (q, *J* = 32.2 Hz, C2), 126.5 (q, *J* = 3.8 Hz, C1), 126.4 (C4), 125.4 (C13), 124.5 (q, *J* = 271.8 Hz, C17), 123.3 (q, *J* = 3.8 Hz, C3), 57.2 (C8), 54.3 (C9, C10), 33.1 (C7), 26.5 (C15), 23.60 (C11, C12), 13.7 (C16).

***Z*-5j':**

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 7.45 – 7.36 (m, 2H, H1', H3'), 7.24 (d, *J* = 8.0 Hz, 1H, H4'), 6.48 (d, *J* = 11.4 Hz, 1H, H13'), 5.80 (dt, *J* = 11.5, 7.5 Hz, 1H, H14'), 2.90 – 2.83 (m, 1H, H7'), 2.68 – 2.55 (m, 6H, H8', H9', H10'), 2.11 (pd, *J* = 7.5, 1.7 Hz, 2H, H15'), 1.88 – 1.78 (m, 4H, H11', H12'), 0.99 (t, *J* = 7.5 Hz, 1H, H16').

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 140.5 (q, *J* = 1.2 Hz, C5', only visible as doublet due to low intensity), 139.4 (C6'), 136.7 (C14'), 123.0 (C4'), 129.1 (q, *J* = 32.1 Hz, C2', only visible as doublet due to low intensity), 126.1 (q, *J* = 3.9 Hz, C1', only visible as doublet due to low intensity), 126.0 (C13'), 122.7 (q, *J* = 3.9 Hz, C3', only visible as doublet due to low intensity), 56.9 (C8'), 54.3 (C9', C10'), 33.2 (C7'), 23.6 (C11', C12'), 21.9 (C15'), 14.3 (C16').

HRMS (ESI) m/z [M+H]⁺ calc. for C₁₇H₂₃F₃N⁺: 298.1777; found: 298.1780.

Phenethylamine **5k**



Starting materials:

6-Chlorophthalazine (**1d**) (123 mg, 750 µmol, 1.00 equiv); **BDLA** (3.80 mg, 18.6 µmol, 2.49 mol %); pyrrolidine (**7a**) (74.7 µL, 900 µmol, 1.20 equiv); butyraldehyde (**6a**) (100 µL, 1.09 mmol, 1.45 equiv); NaBH(OAc)₃ (328 mg, 1.50 mmol, 2.00 equiv); AcOH (90.4 µL, 1.50 mmol, 2.00 equiv).

LED:

448 nm

Temperature:

-15 °C to 10 °C

Purification:

Flash column chromatography (2 times) (SiO₂: 20 g, cyclohexane/EtOAc + 1% NEt₃, 3:1).

Yield: mixture of *E/Z*- **E-5k/Z-5k** (5.73 : 1.00)

and the corresponding constitutional isomers (**E-5k'/Z-5k'**).

142 mg (537 µmol, 72%); pale yellow oil.

NMR spectra could not be assigned unambiguously

E-5k and E-5k':

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 7.38 (d, *J* = 2.1 Hz, 1H, H4), 7.33 (d, *J* = 8.8 Hz, 1H, H4), 7.15 – 7.03 (m, 6H, H1, H2, H3), 6.56 (dt, *J* = 15.6, 1.7 Hz, 2H, H13), 6.13 (ddt, *J* = 15.6, 13.5, 6.6 Hz, 2H, H14), 2.90 – 2.80 (m, 4H, H7), 2.65 – 2.52 (m, 12H, H8, H9, H10), 2.30 – 2.17 (m, 4H, H15), 1.86 – 1.77 (m, 8H, H11, H12), 1.09 (td, *J* = 7.4, 1.9 Hz, 6H, H16).

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 139.0 (C_{Ar}), 138.7 (C_{Ar}), 135.91 (C14), 135.7 (C_{Ar}), 135.5 (C_{Ar}), 135.2 (C14), 132.4 (C_{Ar}), 132.2 (C_{Ar}), 131.2 (C_{Ar}), 129.6 (C_{Ar}), 127.35 (C4), 126.9 (C_{Ar}), 126.6 (C_{Ar}), 125.90 (C4), 125.4 (C13), 57.5 (C8), 57.3 (C8), 54.4 (C9, C10), 33.1 (C7), 32.7 (C7), 26.45 (C15), 26.42 (C15), 23.6 (C11, C12), 13.83 (C16), 13.79 (C16).

Z-5k and Z-5k':

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 7.18 (d, *J* = 2.2 Hz, 1H, H_{Ar}'), 7.15 – 7.03 (m, 5H, H_{Ar}'), 6.41 (dd, *J* = 11.4, 1.7 Hz, 2H, H13'), 5.73 (dtd, *J* = 12.2, 7.4, 4.9 Hz, 2H, H14'), 2.82 – 2.73 (m, 4H, H7'), 2.65 – 2.52 (m, 12H, H8', H9', H10'), 2.19 – 2.03 (m, 4H, H15'), 1.86 – 1.77 (m, 8H, H11', H12'), 0.99 (td, *J* = 7.5, 4.9 Hz, 6H, H16').

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 140.6 (C_{Ar}'), 138.5 (C_{Ar}'), 137.2 (C_{Ar}'), 136.2 (C14'), 135.87 (C14'), 132.48 (C_{Ar}'), 131.4 (C_{Ar}'), 130.9 (C_{Ar}'), 130.7 (C_{Ar}'), 129.33 (C_{Ar}'), 129.29 (C_{Ar}'), 127.0 (C_z'), 126.03 (C13' or CAr'), 126.00 (C13' or CAr'), 125.93 (C13' or CAr'), 57.2 (C8'), 57.0 (C8'), 54.31 (C9', C10'), 33.25 (C7'), 32.84 (C7'), 23.59 (C11', C12'), 21.9 (C15'), 14.34 (C16'), 14.30 (C16')

HRMS (ESI) m/z [M+H]⁺ calc. for C₁₆H₂₃ClN⁺: 264.1514; found: 264.1514.

Phenethylamine **5I**



Starting materials:

6-Methylphthalazine (**1e**) (149 mg, 750 μ mol, 1.00 equiv); **BDLA** (3.80 mg, 18.6 μ mol, 2.49 mol %); pyrrolidine (**7a**) (74.7 μ L, 900 μ mol, 1.20 equiv); butyraldehyde (**6a**) (100 μ L, 1.09 mmol, 1.45 equiv); NaBH(OAc)₃ (328 mg, 1.50 mmol, 2.00 equiv); AcOH (90.4 μ L, 1.50 mmol, 2.00 equiv).

LED:

448 nm

Temperature:

40 °C

Purification:

Flash column chromatography (2 times) (SiO₂: 20 g, cyclohexane/EtOAc + 5% NEt₃, 20:1).

Yield: mixture of *E/Z*-isomers **E-5I/Z-5I** (1.71 : 1.00)
and the corresponding constitutional isomers (**E-5I'/Z-5I'**).
142 mg (584 μ mol, 78%); pale yellow oil.

NMR spectra could not be assigned unambiguously

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 7.33 (d, *J* = 7.7 Hz, 1H, H_{Ar} or H_{Ar'}), 7.25 (s, 1H, H_{Ar} or H_{Ar'}), 7.12 – 6.93 (m, 10H, H_{Ar} or H_{Ar'}), 6.62 (dd, *J* = 15.7, 5.3 Hz, 2H, H13), 6.47 (d, *J* = 11.4 Hz, 2H, H13'), 6.13 (dt, *J* = 15.5, 6.6 Hz, 2H, H14), 5.69 (dtd, *J* = 11.7, 7.3, 4.7 Hz, 2H, H14'), 2.91 – 2.83 (m, 4H, H7), 2.84 – 2.75 (m, 4H, H7'), 2.66 – 2.53 (m, 24H, H8, H9, H10, H8', H9', H10'), 2.31 (overlapping singlets, look like a t, *J* = 4.1 Hz, 12H, H17, H17'), 2.30 – 2.17 (m, 4H, H15), 2.19 – 2.10 (m, 4H, H15'), 1.88 – 1.75 (m, 16H, H11, H12, H11', H12'), 1.09 (td, *J* = 7.5, 3.0 Hz, 6H, H16), 0.99 (td, *J* = 7.5, 1.8 Hz, 6H, H16').

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 138.5, 137.1, 136.7, 136.6, 135.9, 135.6, 135.3, 135.0, 134.9, 134.3, 134.3, 134.1, 133.8, 133.7, 130.6, 130.3, 130.2, 129.8, 129.6, 129.4, 127.9, 127.8, 127.4, 127.2, 126.9, 126.64, 126.59, 126.4, 126.2, 125.9, 57.9, 57.8, 57.6, 57.5, 54.4, 54.3, 33.4, 33.2, 33.0, 32.9, 26.5, 26.5, 23.6, 21.9, 21.24, 21.20, 21.17, 14.49, 14.47, 14.00, 13.99.

HRMS (ESI) m/z [M+H]⁺ calc. for C₁₇H₂₆N⁺: 244.2060; found 244.2058.

Phenethylamine **5m**



Starting materials:

6-Methoxyphthalazine (**1f**) (120 mg, 750 μ mol, 1.00 equiv); **BDLA** (3.80 mg, 18.6 μ mol, 2.49 mol %); pyrrolidine (**7a**) (74.7 μ L, 900 μ mol, 1.20 equiv); butyraldehyde (**6a**) (100 μ L, 1.09 mmol, 1.45 equiv); NaBH(OAc)₃ (328 mg, 1.50 mmol, 2.00 equiv); AcOH (90.4 μ L, 1.50 mmol, 2.00 equiv).

LED:

448 nm

Temperature:

40 °C

Purification:

Flash column chromatography (2 times) (SiO₂: 20 g, cyclohexane/EtOAc + 5% NEt₃, 10:1).

Yield: mixture of *E/Z*-isomers **E-5m/Z-5m** (2.95 : 1.00)
and the corresponding constitutional isomers (**E-5m'/Z-5m'**).
142.4 mg (549 μ mol, 73%); pale yellow oil.

NMR spectra could not be assigned unambiguously

***E-5m* and *E-5m'*:**

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 7.35 (d, *J* = 8.2 Hz, 1H, H1 or H2 or H3 or H4), 7.07 (dd, *J* = 10.4, 8.8 Hz, 1H, H1 or H2 or H3 or H4), 6.98 – 6.95 (m, 2H, H1 or H2 or H3 or H4), 6.79 – 6.68 (m, 2H, H1 or H2 or H3 or H4), 6.59 (dd, *J* = 19.4, 15.6 Hz, 2H, H13), 6.14 (dt, *J* = 15.5, 6.6 Hz, 1H, H14), 6.03 (dt, *J* = 15.5, 6.6 Hz, 1H, H14), 3.81 – 3.77 (m, 6H, H17), 2.91 – 2.73 (m, 4H, H7), 2.66 – 2.54 (m, 12H, H8, H9, H10), 2.30 – 2.18 (m, 4H, H15), 1.85 – 1.77 (m, 8H, H11, H12), 1.09 (td, *J* = 7.4, 5.6 Hz, 6H, H16).

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 158.6 (C_{Ar}-OMe), 158.3 (C_{Ar}-OMe), 140.14, 138.67, 137.94, 137.83, 134.7 (14), 132.8 (14), 130.87, 130.63, 130.33, 129.75, 129.72, 129.27, 127.13 (C4), 126.4 (C13), 125.8 (C13), 115.22, 115.13, 115.05, 112.82, 112.30, 112.06, 111.08, 110.99, 58.0 (C8), 57.6 (C8), 55.4 (C17), 55.3 (C17), 54.4 (C9, C10), 33.5 (C7), 32.5 (C7), 26.5 (C15), 26.4 (C15), 23.6 (C11, C12), 14.1 (C16), 13.9 (C16).

Z-5m and Z-5m':

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 7.35 (d, J = 8.2 Hz, 1H, H1' or H2' or H3' or H4'), 7.07 (dd, J = 10.4, 8.8 Hz, 1H, H1' or H2' or H3' or H4'), 6.98 – 6.95 (m, 2H, H1' or H2' or H3' or H4'), 6.79 – 6.68 (m, 2H, H1' or H2' or H3' or H4'), 6.44 (dd, J = 17.2, 11.4 Hz, 2H, H13'), 5.74 – 5.60 (m, 2H, H14'), 3.81 – 3.77 (m, 6H, H17'), 2.91 – 2.73 (m, 4H, H7'), 2.66 – 2.54 (m, 12H, H8', H9', H10'), 2.14 (pd, J = 7.4, 1.5 Hz, 4H, H15'), 1.85 – 1.77 (m, 8H, H11', H12'), 0.99 (td, J = 7.5, 4.4 Hz, 6H, H16').

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 158.7 (C'_{Ar}-OMe), 140.14, 138.67, 137.94, 137.83, 135.3 (C14'), 134.6 (C14'), 130.87, 130.63, 130.33, 129.75, 129.72, 129.27, 127.1 (C13'), 126.6 (C13'), 115.22, 115.13, 115.05, 112.82, 112.30, 112.06, 111.08, 110.99, 57.7 (C8'), 57.3 (C8'), 55.33 (C17'), 55.30 (C17'), 54.3 (C9', C10'), 33.7 (C7'), 32.6 (C7'), 23.6 (C11', C12'), 21.9 (C15'), 21.9 (C15'), 14.5 (C16'), 14.4 (C16').

HRMS (ESI) m/z [M+H]⁺ calc. for C₁₇H₂₆NO⁺: 260.2009; found: 260.2012.

Phenethylenamine **4a**



The reaction was set up in a nitrogen filled glovebox.

Phthalazine (**1a**) (105 mg, 795 µmol, 1.00 equiv) and **BDLA** (4.00 mg, 19.6 µmol, 2.47 mol %) were suspended in THF (dry, degassed) (2.00 mL). Then pyrrolidine (**7a**) (79.6 µL, 960 µmol, 1.21 equiv) was added (yellow suspension turned colorless). The reaction vessel was sealed and the reaming steps of the reaction were carried out in a fume hood. The reaction mixture was irradiated by a 425 – 430 nm LED and butyraldehyde (**6a**) (110 µL, 1.20 mmol, 1.50 equiv) was added dropwise. Gas evolution started almost immediately. The yellow solution was stirred at rt for 15 h. Afterwards all volatile compounds were removed under vacuum (rt, <1 mbar, reconddensed into a liquid N₂ cooling trap). The remaining orange solution was purified by distillation (Kugelrohr-Apperatus). A pale yellow oil was obtained (150 °C at 4x10⁻² mbar).

Yield: mixture of *E*/*Z*-isomers **E-4a/Z-4a** (1.00 : 2.60)

95.2 mg (418 µmol, 54%); pale yellow oil.

***E*-4a:**

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 7.35 – 7.29 (m, 1H, H2), 7.25 (d, J = 6.4 Hz, 1H, H1), 7.14 – 7.06 (m, 1H, H4), 7.00 – 6.91 (m, 1H, H3), 6.92 (d, J = 13.7 Hz, 1H, H8), 6.67 (d, J = 15.7 Hz, 1H, H13), 6.09 (dt, J = 15.6, 6.5 Hz, 1H, H14), 5.24 (d, J = 13.7 Hz, 1H, H7), 3.29 – 3.19 (m, 4H, H9, H10), 2.31 – 2.22 (m, 2H, H15), 1.96 – 1.90 (m, 4H, H11, H12), 1.12 (t, J = 7.5 Hz, 3H, H16).

¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 137.5 (C6), 137.1 (C8), 133.9 (C5), 133.3 (C14), 127.7 (C13), 127.0 (C4), 126.6 (C2), 123.5 (C1), 123.5 (C3), 95.0 (C7), 49.1 (C9, C10), 26.5 (C15), 25.4 (C11, C12), 14.1 (C16).

***Z*-4a:**

¹H NMR (400 MHz, CDCl₃ with 0.03% v/v TMS): δ 7.35 – 7.29 (m, 1H, H1'), 7.14 – 7.06 (m, 2H, H2', H4'), 7.00 (d, J = 13.8 Hz, 1H, H8'), 7.00 – 6.91 (m, 1H, H3'), 6.43 (d, J = 11.4 Hz, 1H, H13'), 5.67 (dt, J = 11.4, 7.4 Hz, 1H, H14'), 5.14 (d, J = 13.8 Hz, 1H, H7'), 3.29 – 3.19 (m, 4H, H9', H10'), 2.19 (qd, J = 7.4, 1.7 Hz, 2H, H15'), 1.96 – 1.90 (m, 4H, H11', H12'), 1.02 (t, J = 7.5 Hz, 3H, H16').

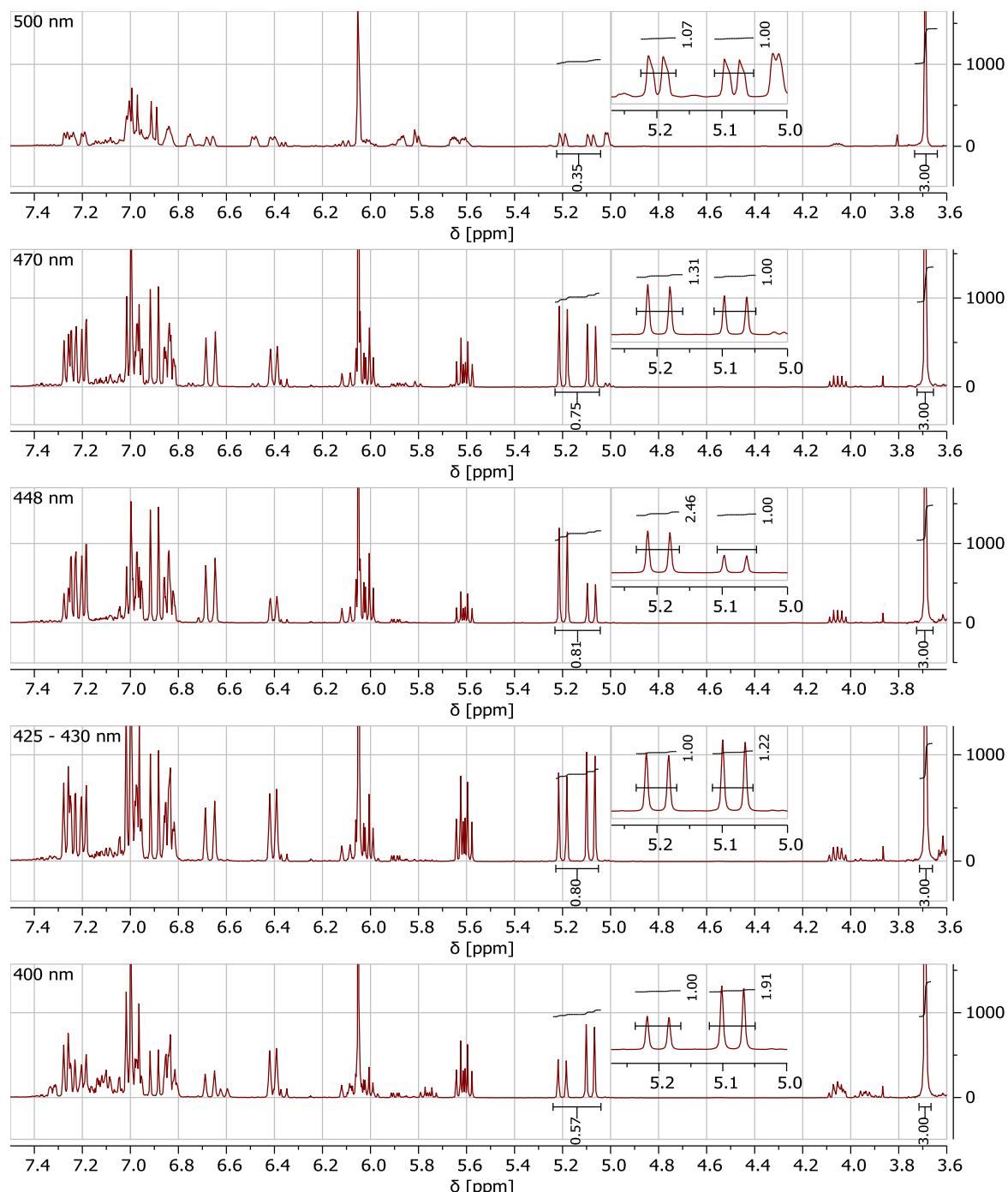
¹³C{¹H} NMR (101 MHz, CDCl₃ with 0.03% v/v TMS): δ 138.3 (C6'), 136.6 (C8'), 133.9 (C14'), 133.1 (C5'), 129.7 (C4'), 128.2 (C13'), 127.0 (C2'), 122.5 (C3'), 122.3 (C1'), 95.4 (C7'), 49.1 (C9', C10'), 25.4 (C11', C12'), 22.0 (C15'), 14.6 (C16').

HRMS (ESI) m/z [M+H]⁺ calc. for C₁₆H₂₂N⁺: 228.1747; found: 228.1751.

Optimization of Irradiation Wavelength for the Domino IEDDA/PIRO Reaction:

The reaction was set up in a nitrogen filled glovebox.

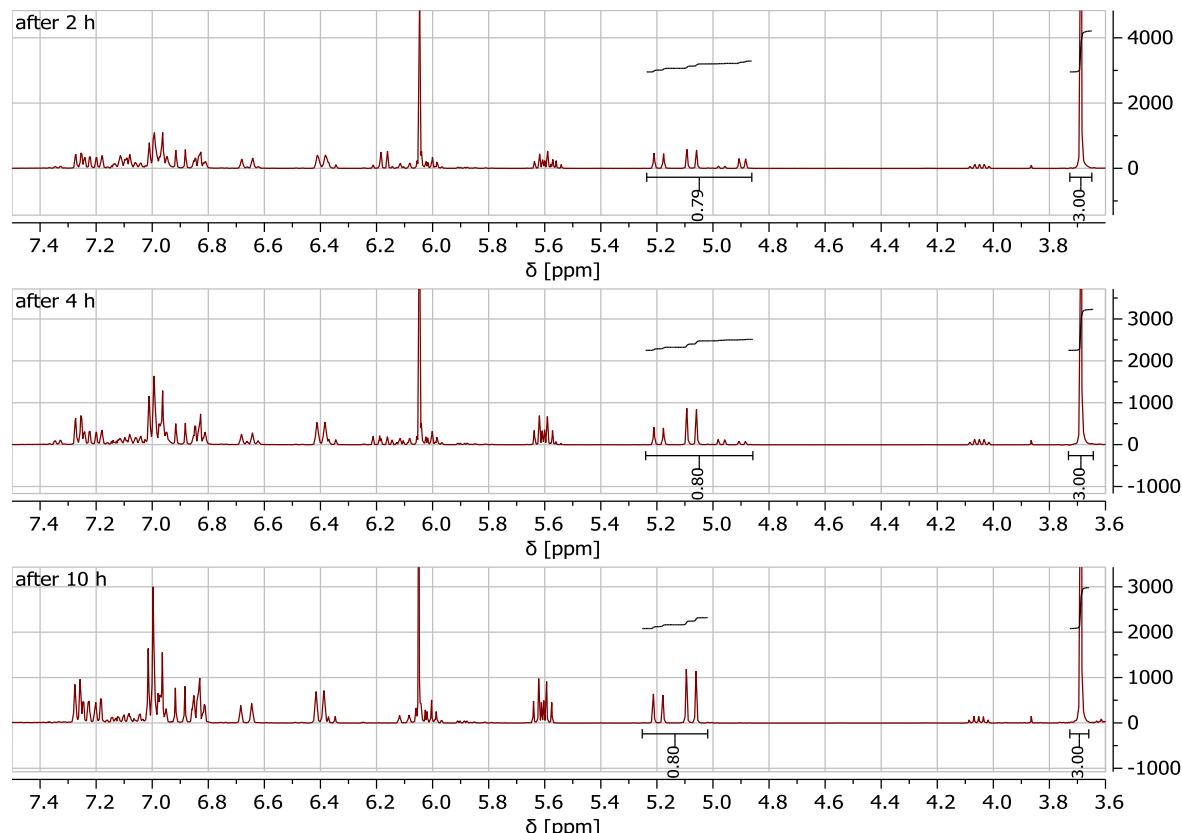
Phthalazine (**1a**) (49.3 mg, 375 μmol , 1.00 equiv), 1,3,5-trimethoxybenzene (TMB) (21.0 mg, 124 μmol , 0.333 equiv), and **BDLA** (1.90 mg, 9.32 μmol , 2.49 equiv) were suspended in THF-d₈ (dry, degassed) (1.00 mL). Then, pyrrolidine (**7a**) (37.3 μL , 450 μmol , 1.20 equiv) was added, the reaction vessel was sealed and the reaction was continued in a fume hood. The reaction mixture was heated to 30 °C and irradiated with a LED. Then butyraldehyde (**6a**) (50.0 μL , 544 μmol , 1.45 equiv) was added. Gas evolution was observed after a few minutes. The reaction was kept irradiated overnight. Afterwards the vessel was sealed, put back into the glovebox and a NMR sample was prepared.



Isomerization Monitoring of IEDDA/PIRO Product:

The reaction was set up in a nitrogen filled glovebox.

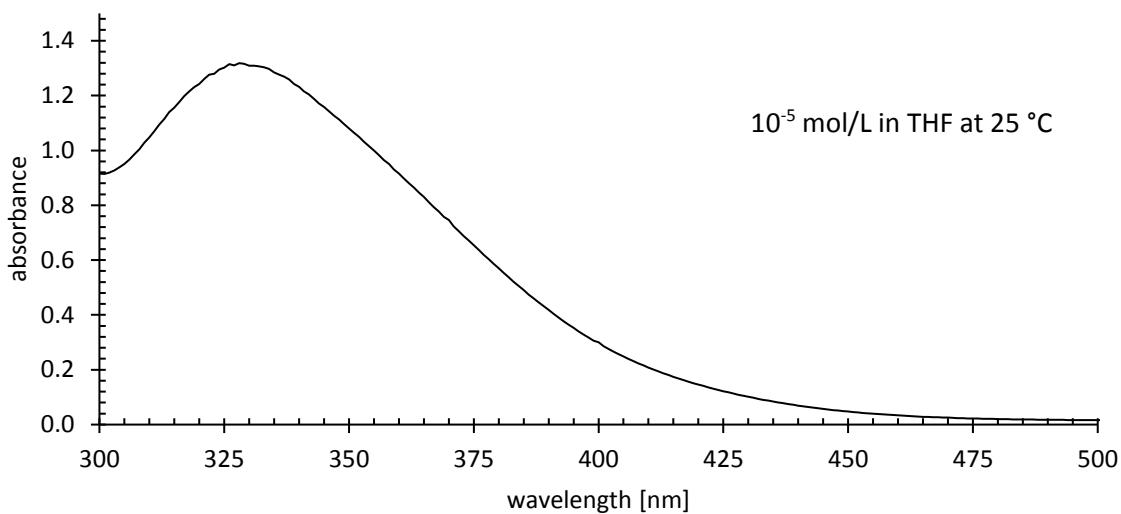
Phthalazine (**1a**) (148 mg, 1.13 mmol, 1.00 equiv), 1,3,5-trimethoxybenzene (TMB) (63.0 mg, 371 μ mol, 0.333 equiv), and **BDLA** (5.70 mg, 28.0 μ mol, 2.49 mol %) were suspended in THF-d₈ (dry, degassed) (3.00 mL). Then, pyrrolidine (**7a**) (112 μ L, 1.35 mmol, 1.20 equiv) was added, the reaction vessel was sealed and the reaction was continued in a fume hood. The reaction mixture was heated to 30 °C and irradiated with a 425 – 430 nm LED. Then butyraldehyde (**6a**) (150 μ L, 1.63 mmol, 1.45 equiv) was added. Gas evolution was observed after a few minutes. Aliquots (0.70 mL) were taken in intervals (2 h, 4 h, 10 h) under Schlenk conditions.



Degradation of BDLA-Phthalazine Complex by Irradiation:

UV/Vis Spectrum of BDLA-Phthalazine Complex

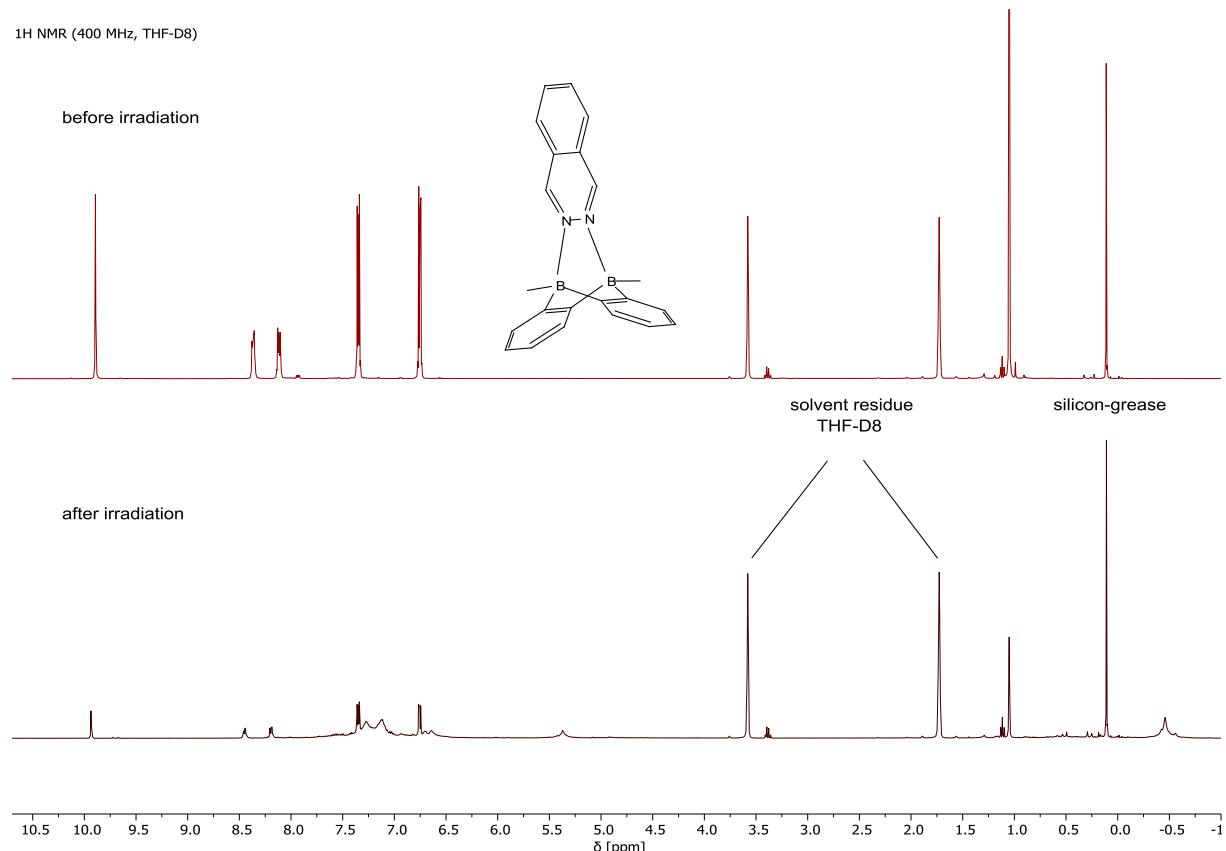
Sample was prepared in a nitrogen filled glovebox.



NMR analysis of BDLA-Phthalazine Complex under irradiation.

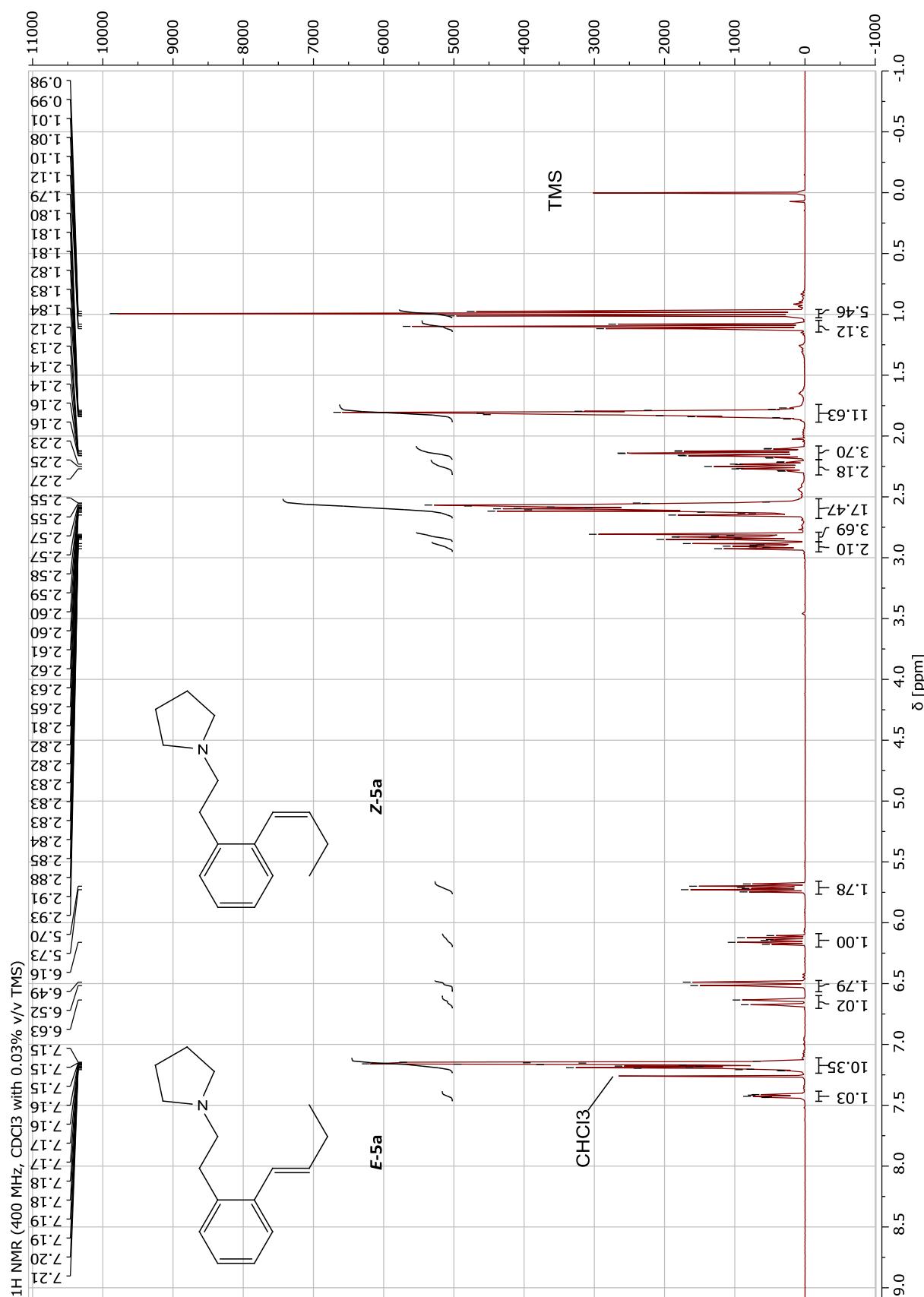
Reaction was set up in a nitrogen filled glovebox.

Phthalazine (**1a**) (3.30 mg, 25.1 μ mol, 1.00 equiv) and **BDLA** (5.10 mg, 25.0 μ mol, 1.00 equiv) were suspended in THF-d₈ (dry, degassed) (1.00 mL) in a screw cap NMR tube, resulting in a saturated solution. A ¹H NMR was measured before it was irradiated by a 425 – 430 nm LED for 15 h at 30 °C. Afterwards a second ¹H NMR was measured.

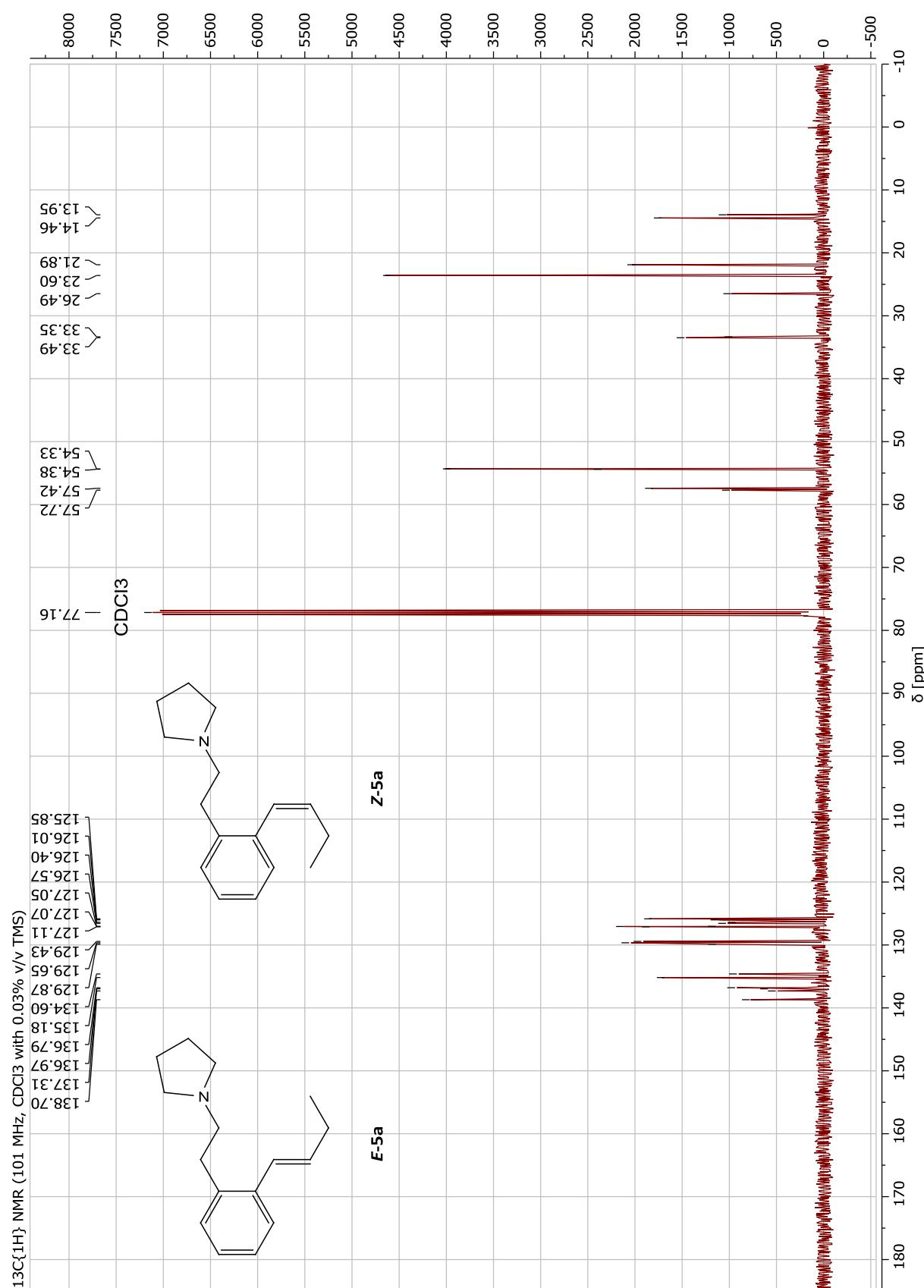


NMR Spectra:

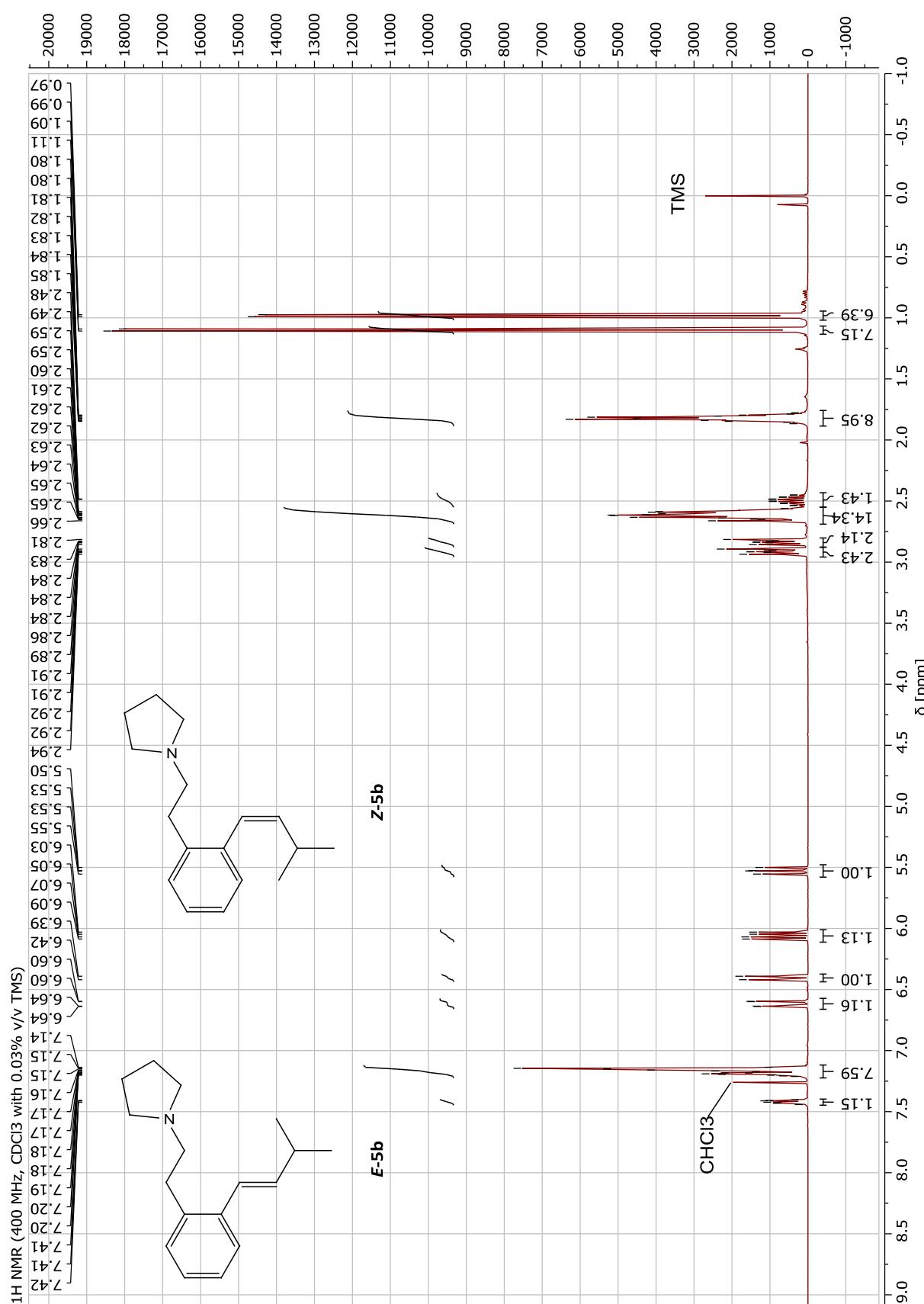
¹H NMR of 5a



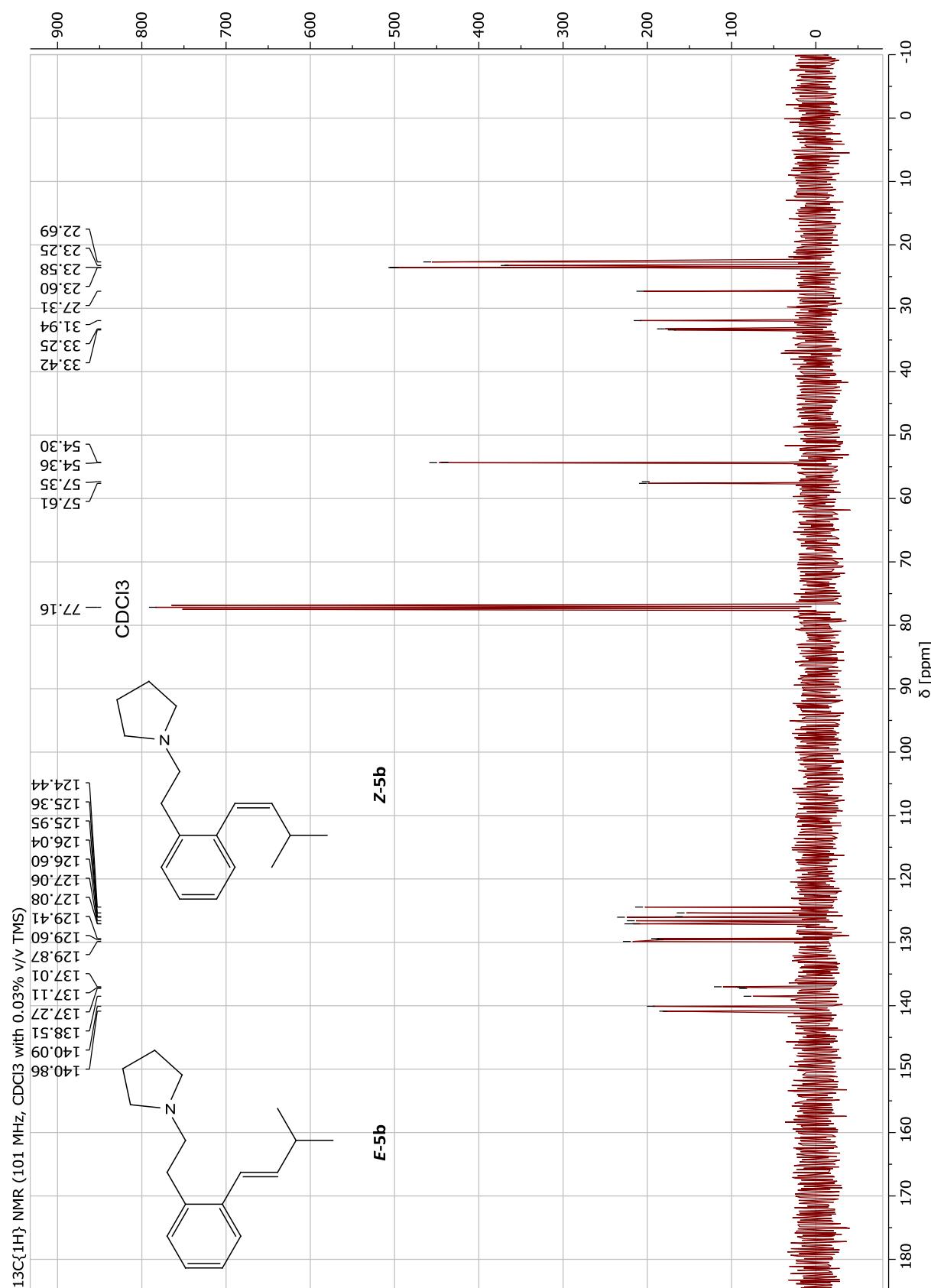
$^{13}\text{C}\{^1\text{H}\}$ NMR of 5a



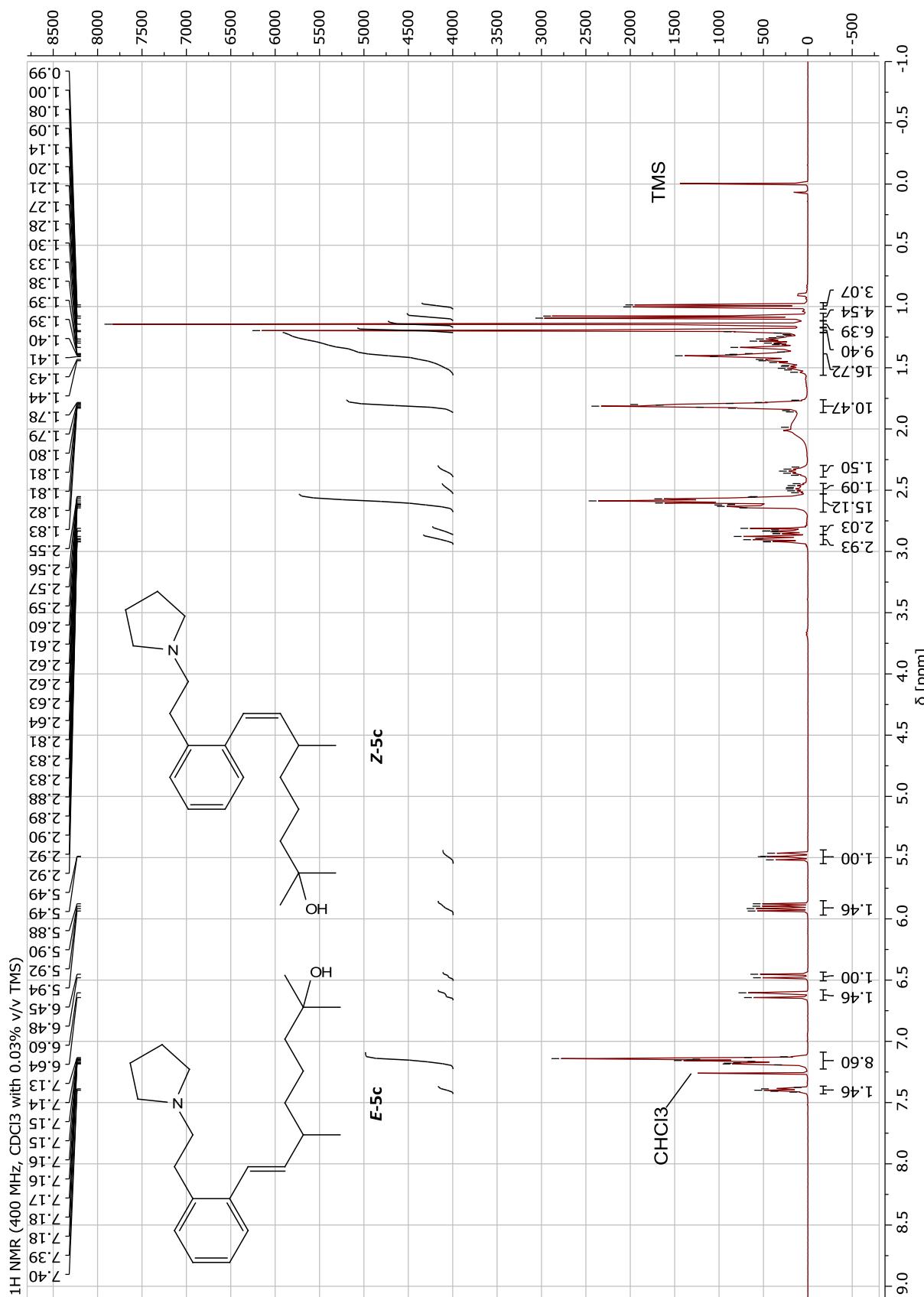
¹H NMR of 5b

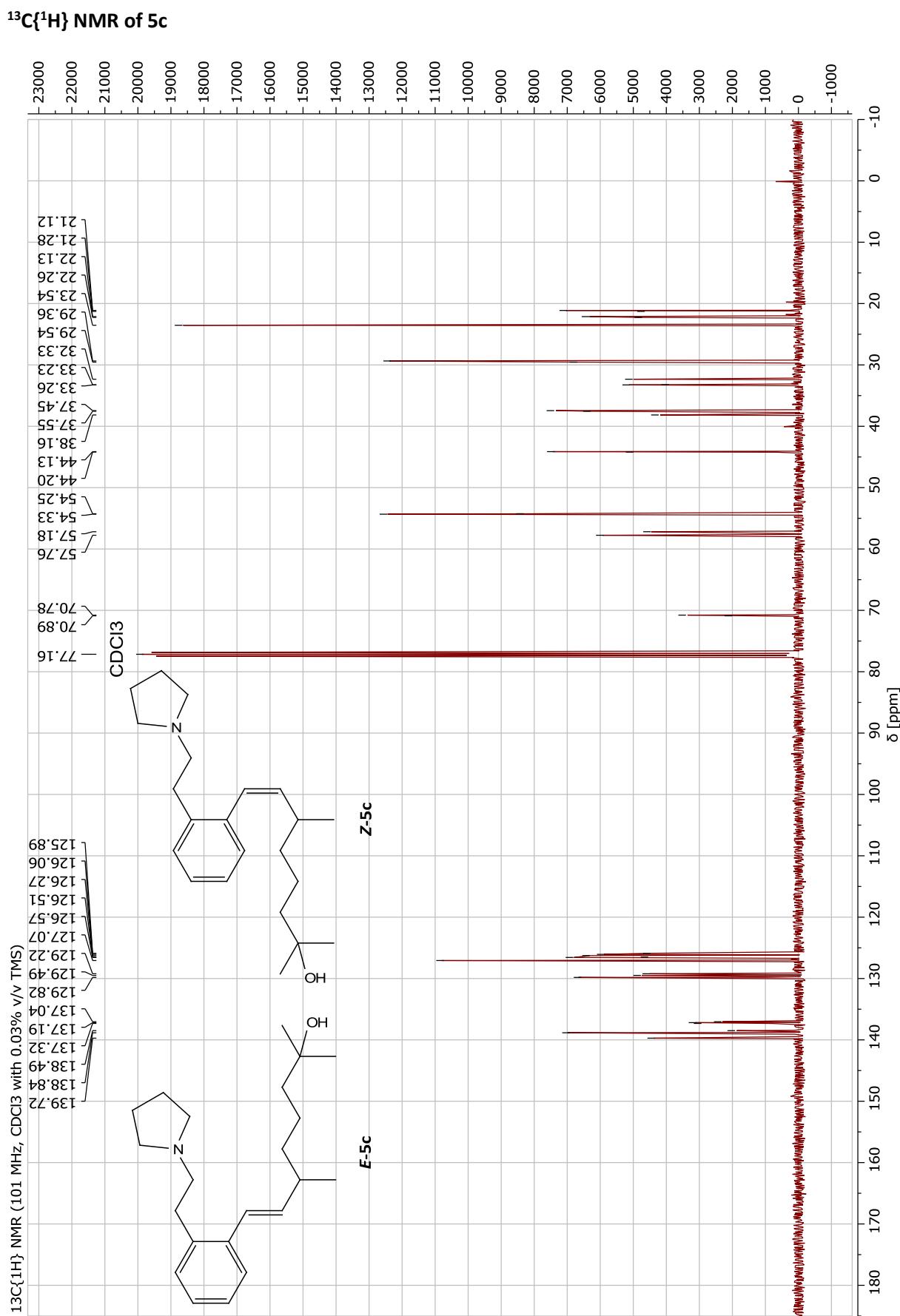


$^{13}\text{C}\{^1\text{H}\}$ NMR of 5b

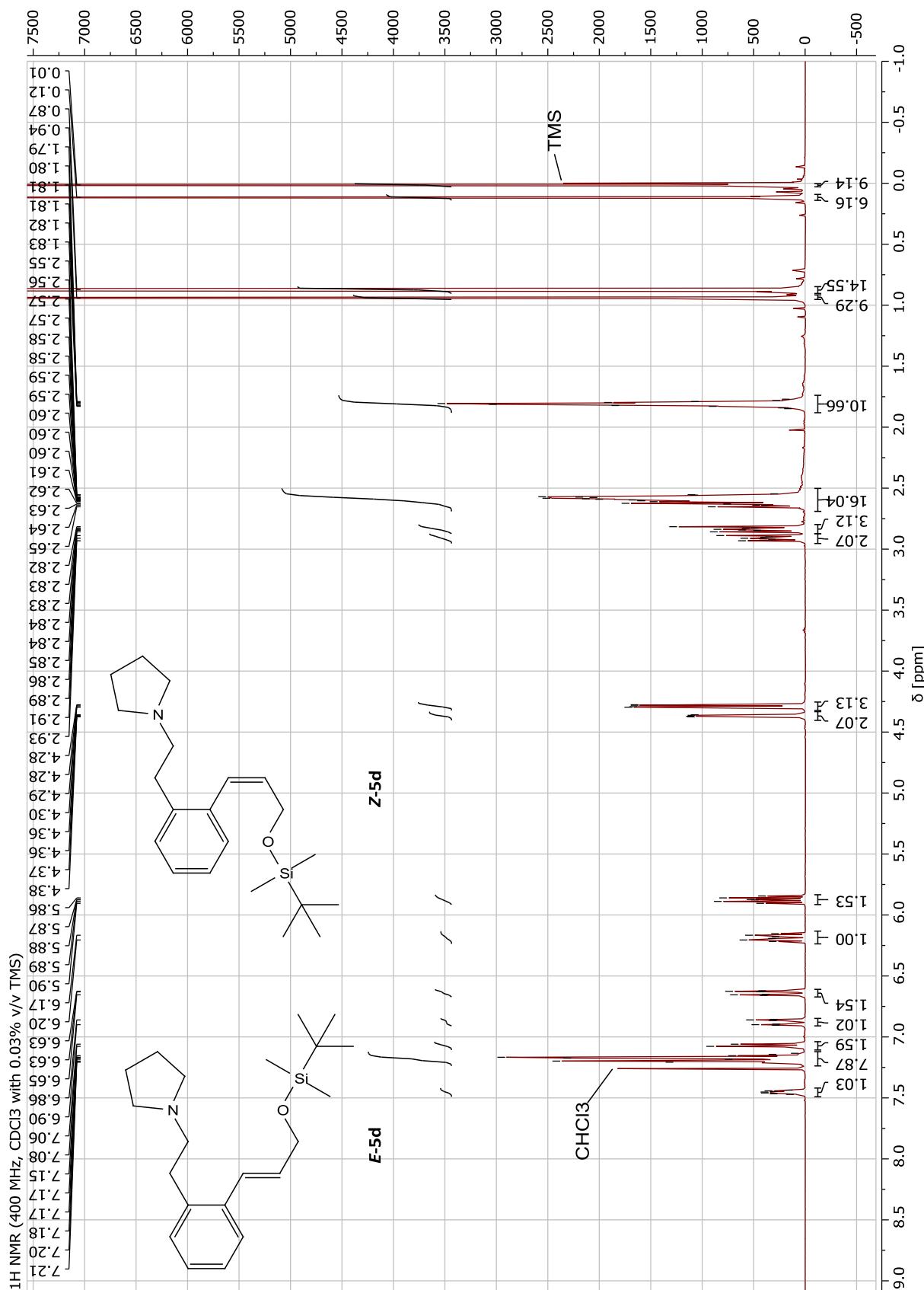


¹H NMR of 5c

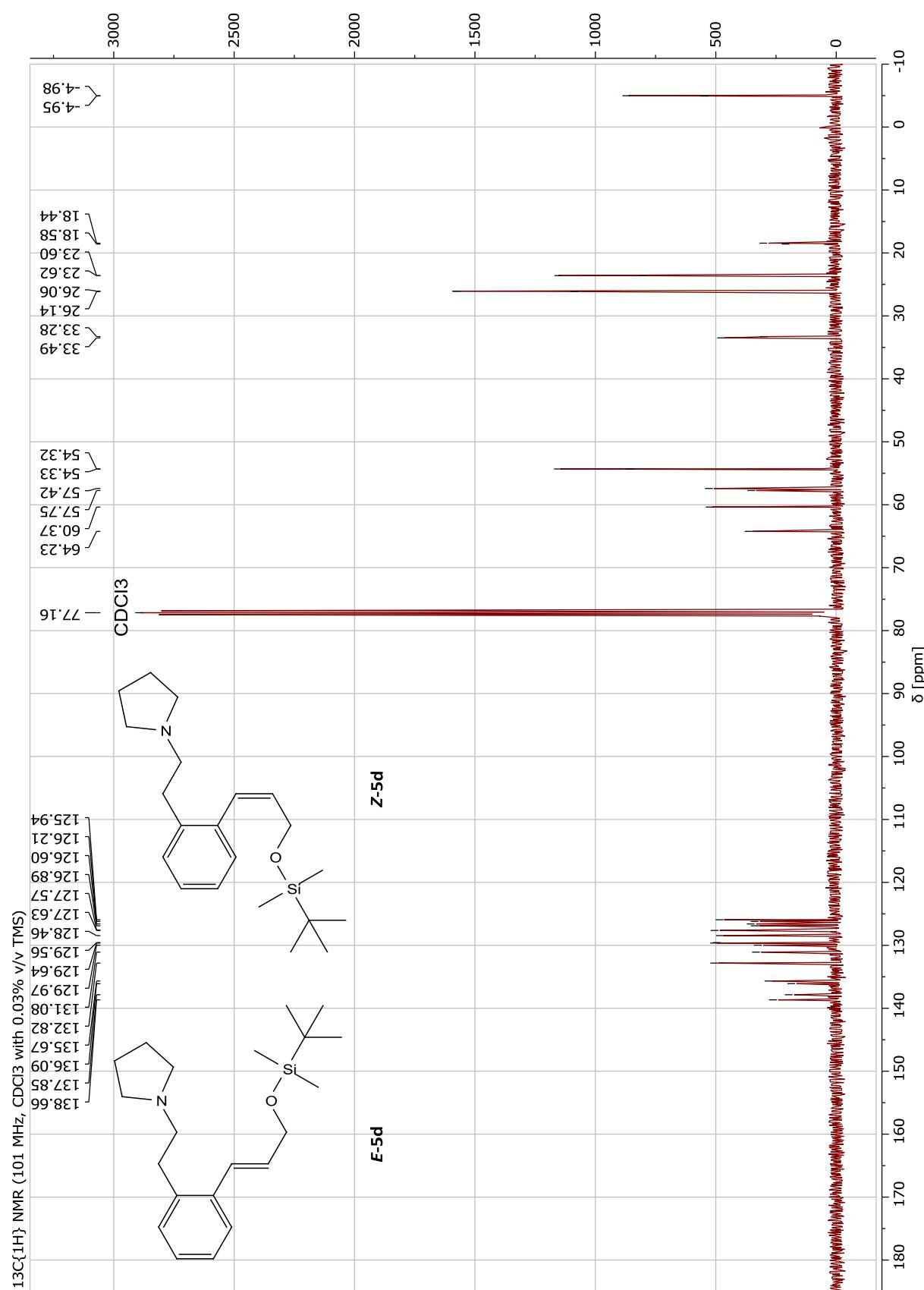




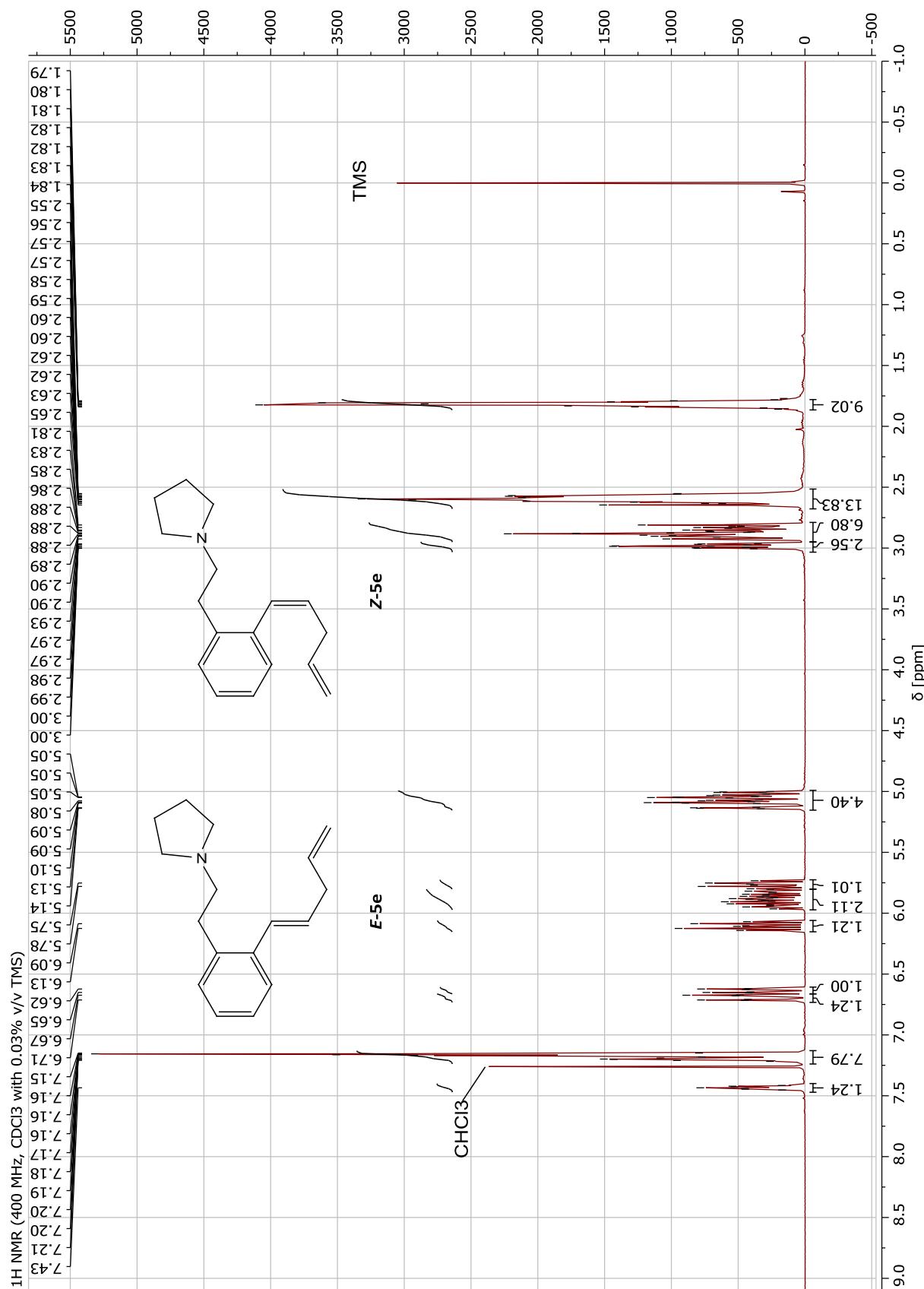
¹H NMR of 5d



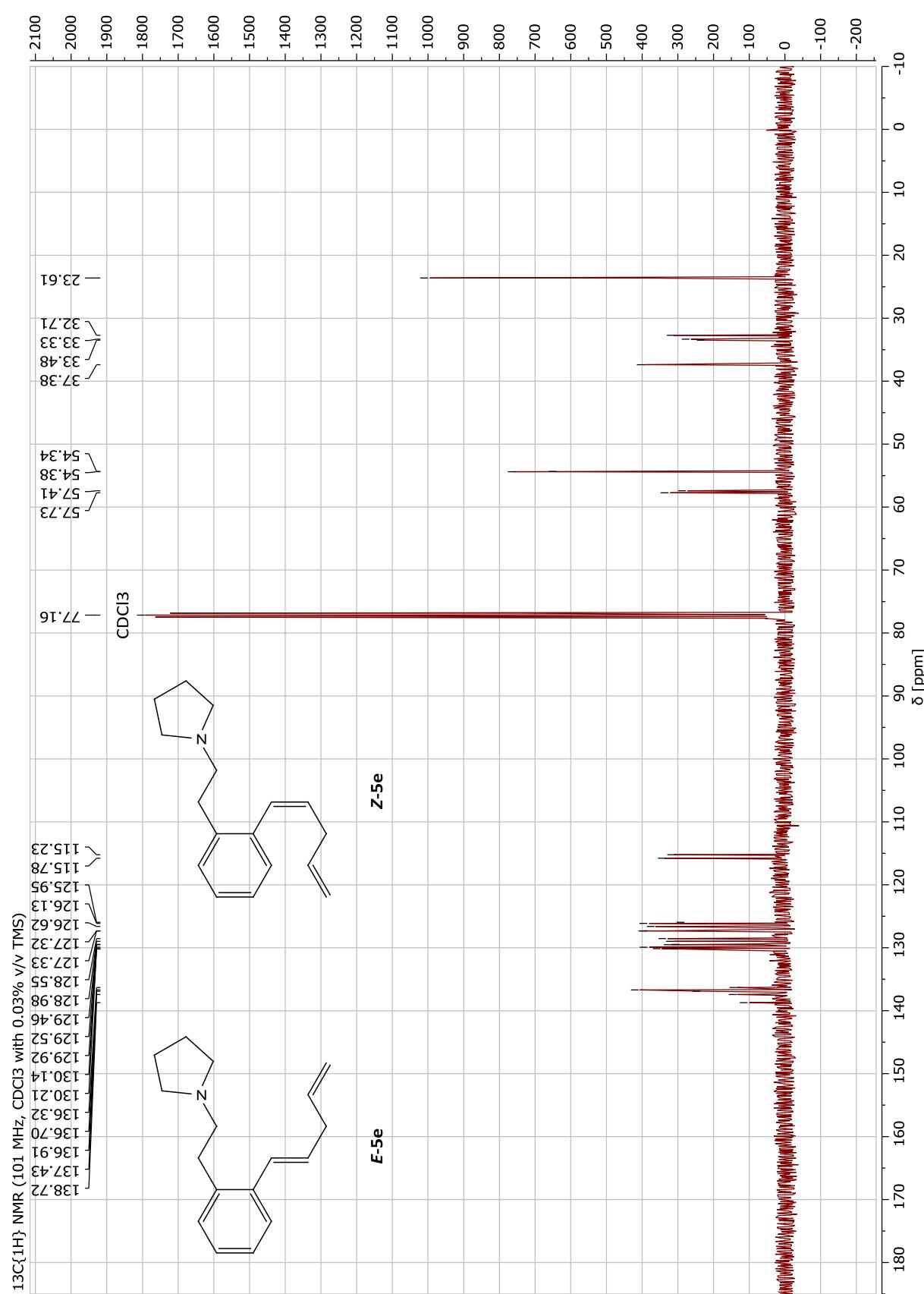
$^{13}\text{C}\{^1\text{H}\}$ NMR of 5d



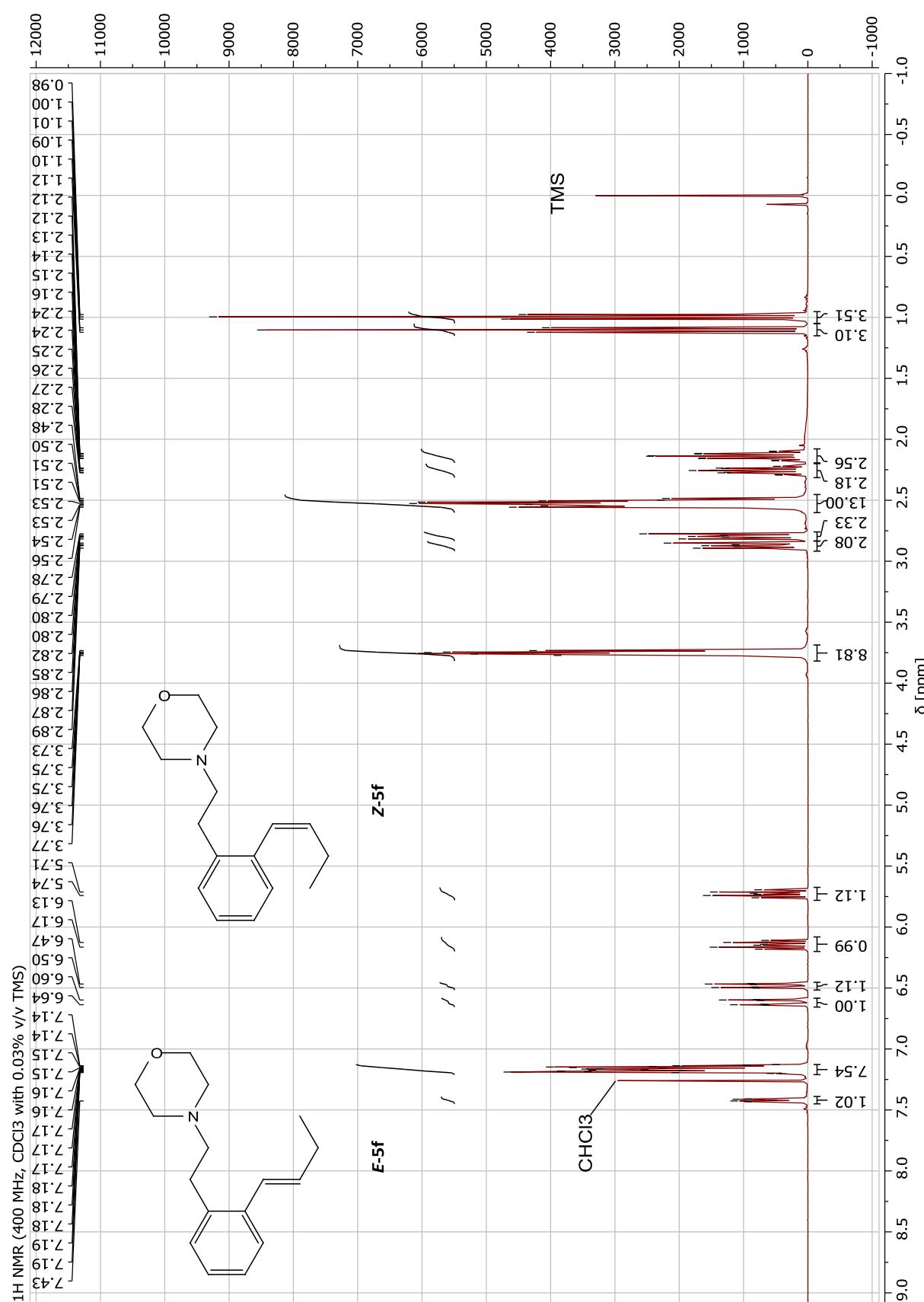
¹H NMR of 5e



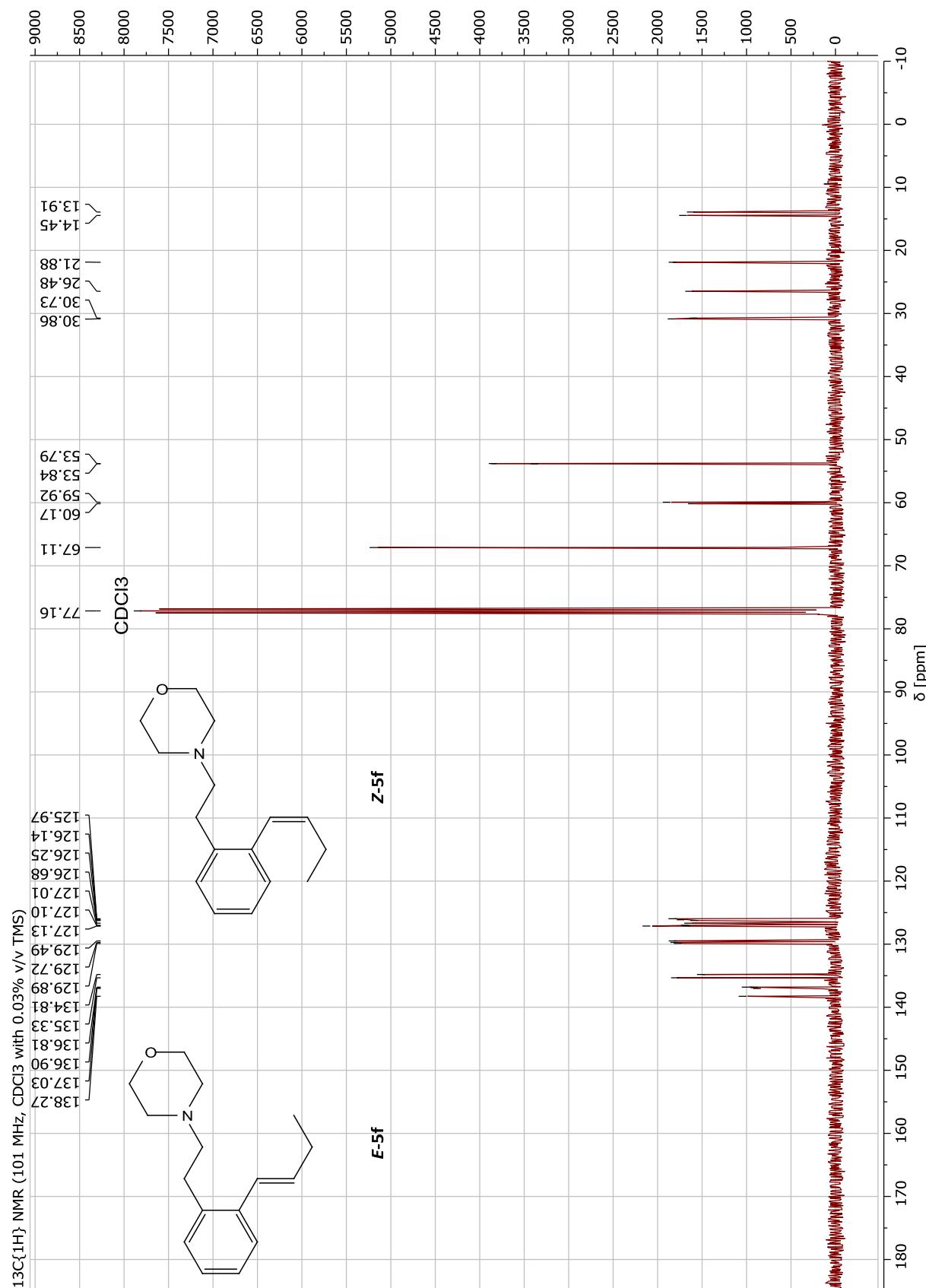
$^{13}\text{C}\{^1\text{H}\}$ NMR of 5e



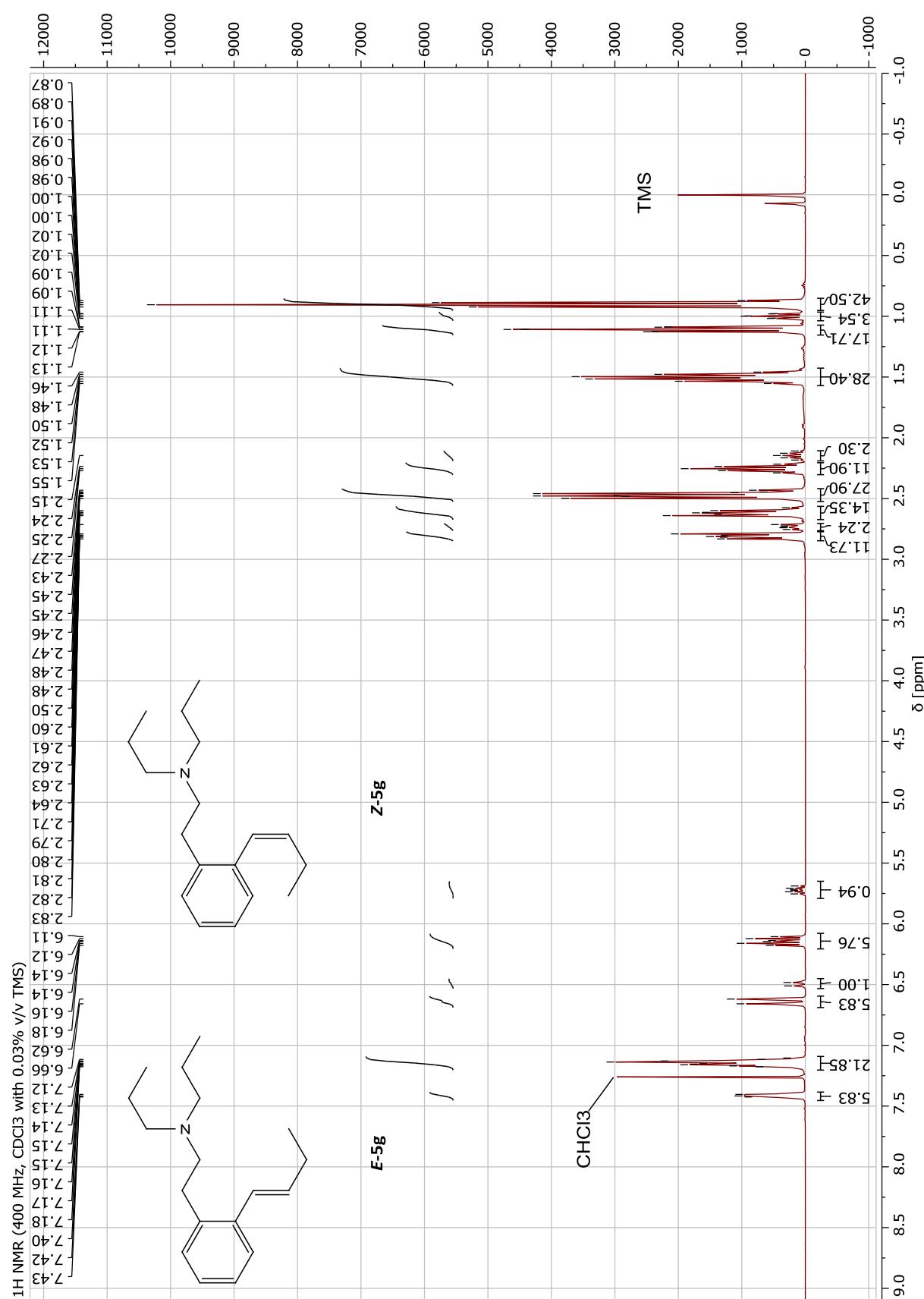
¹H NMR of 5f



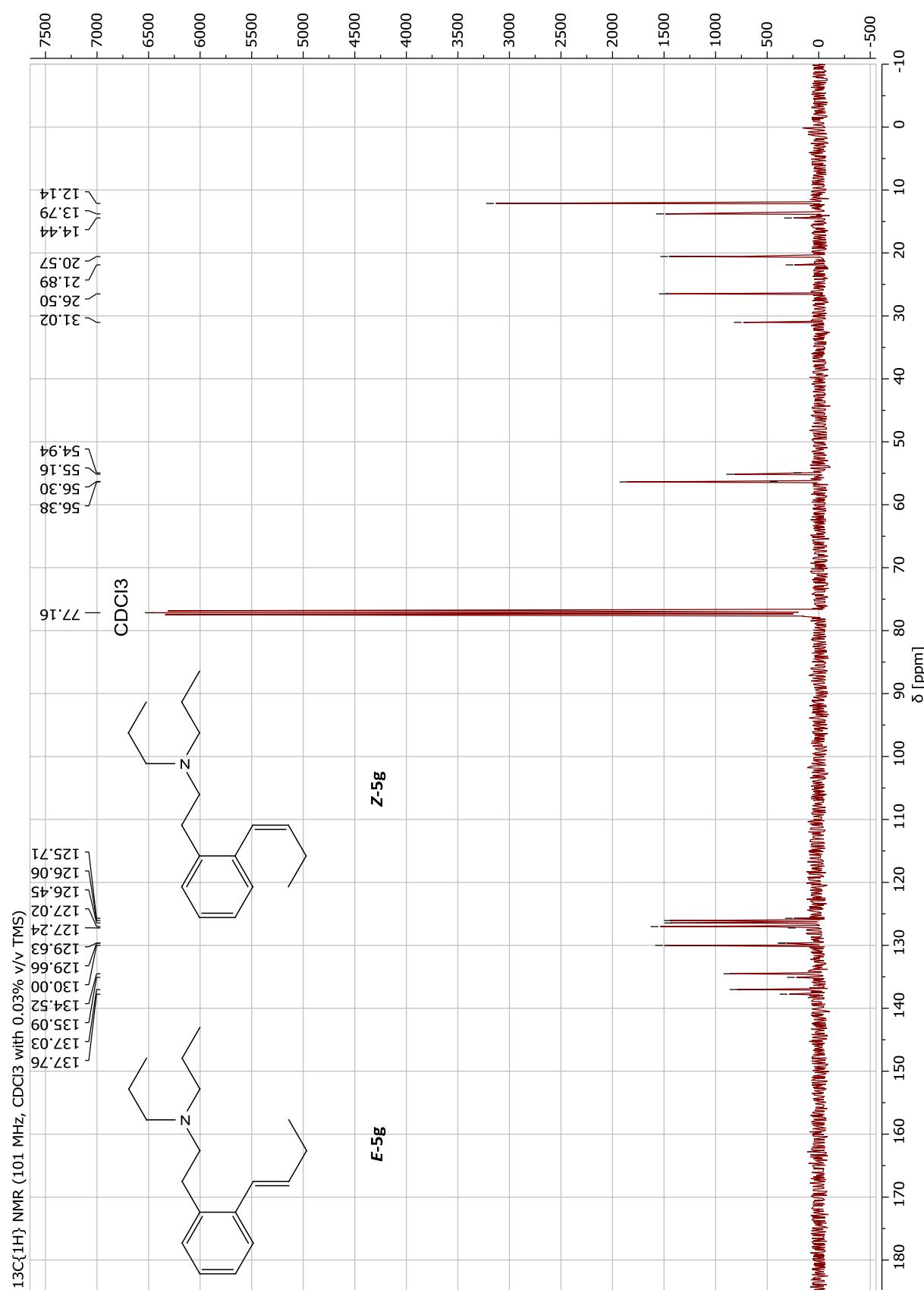
$^{13}\text{C}\{^1\text{H}\}$ NMR of 5f



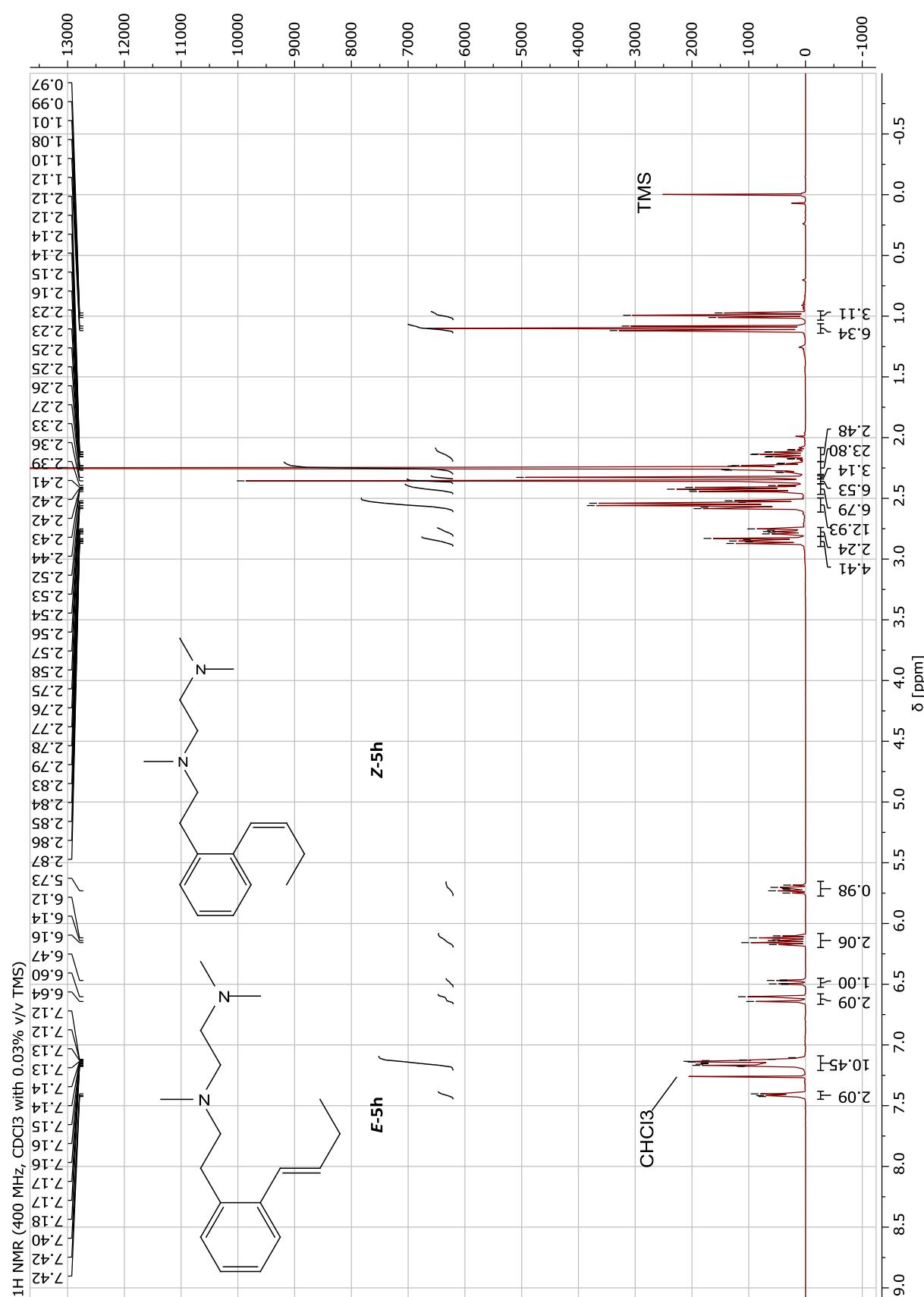
¹H NMR of 5g



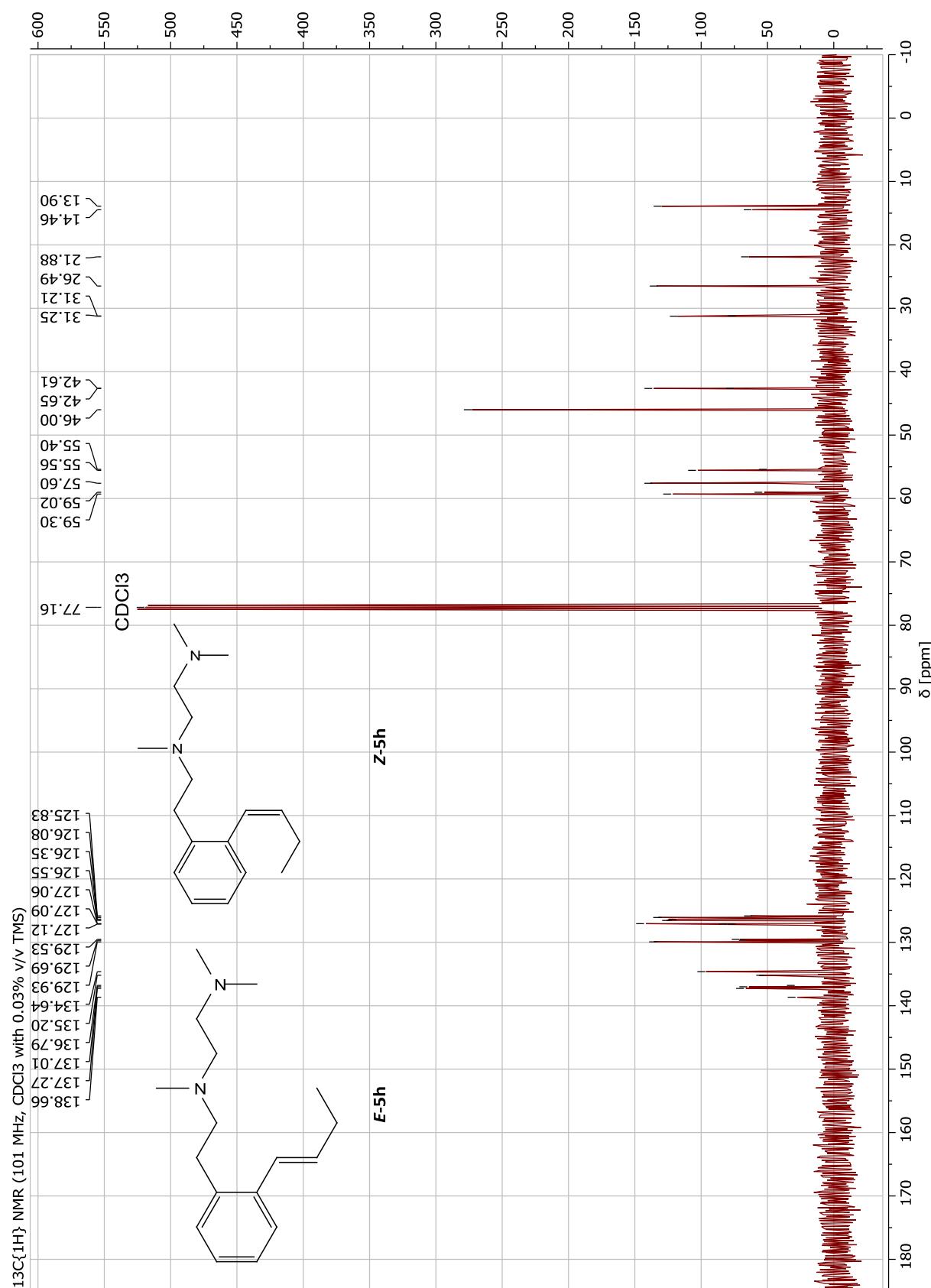
$^{13}\text{C}\{^1\text{H}\}$ NMR of 5g



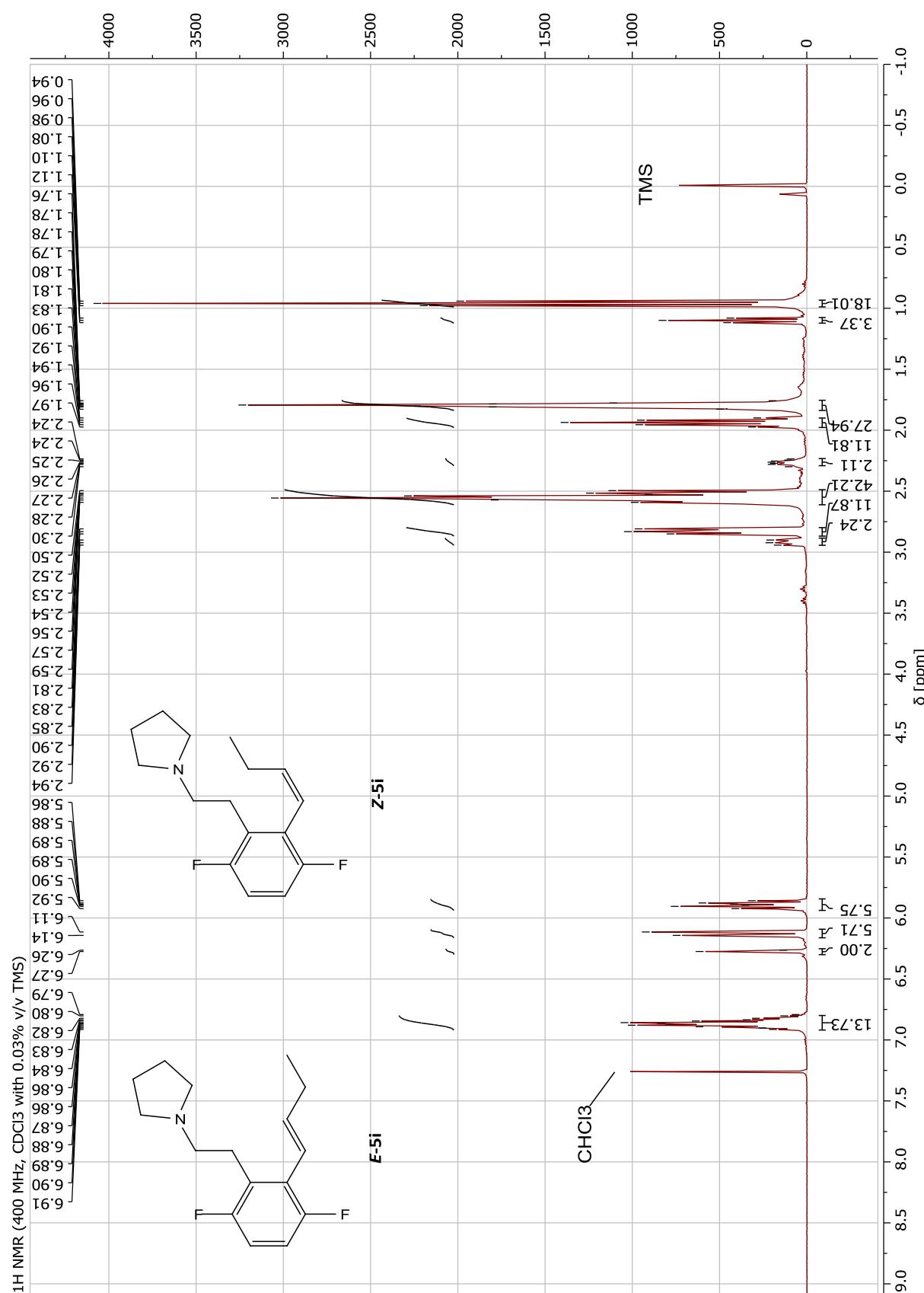
¹H NMR of 5h



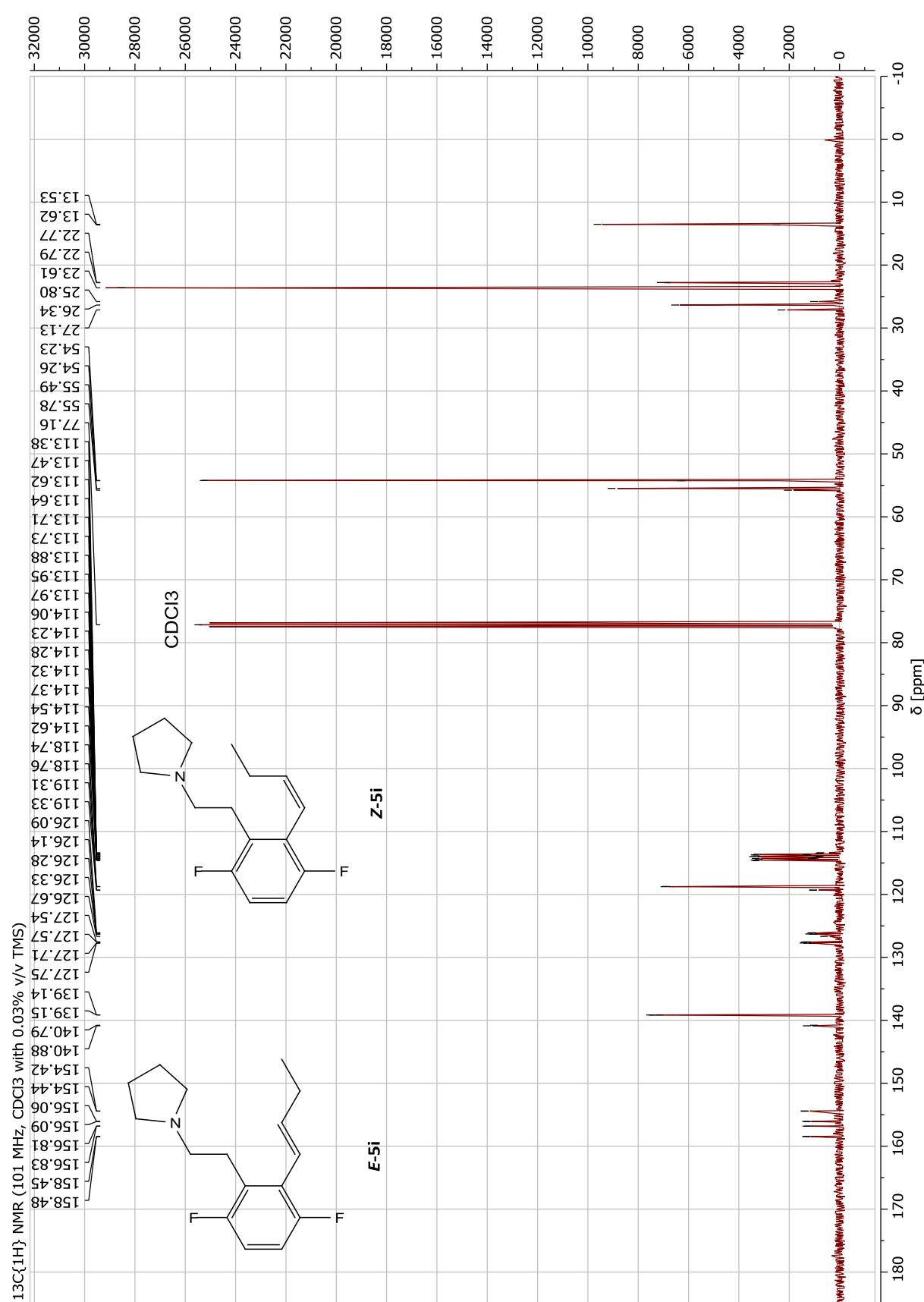
$^{13}\text{C}\{^1\text{H}\}$ NMR of 5h



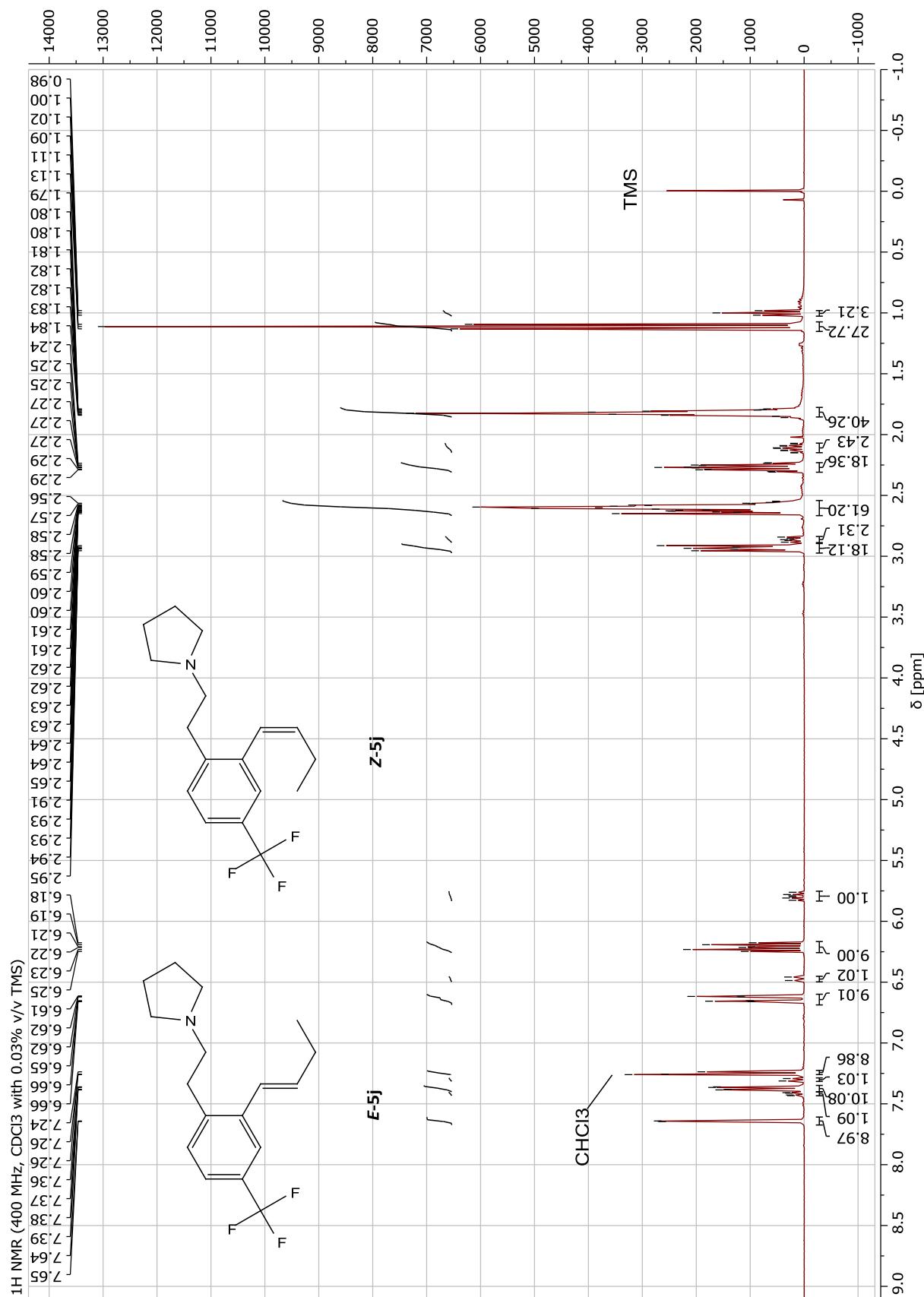
¹H NMR of 5i



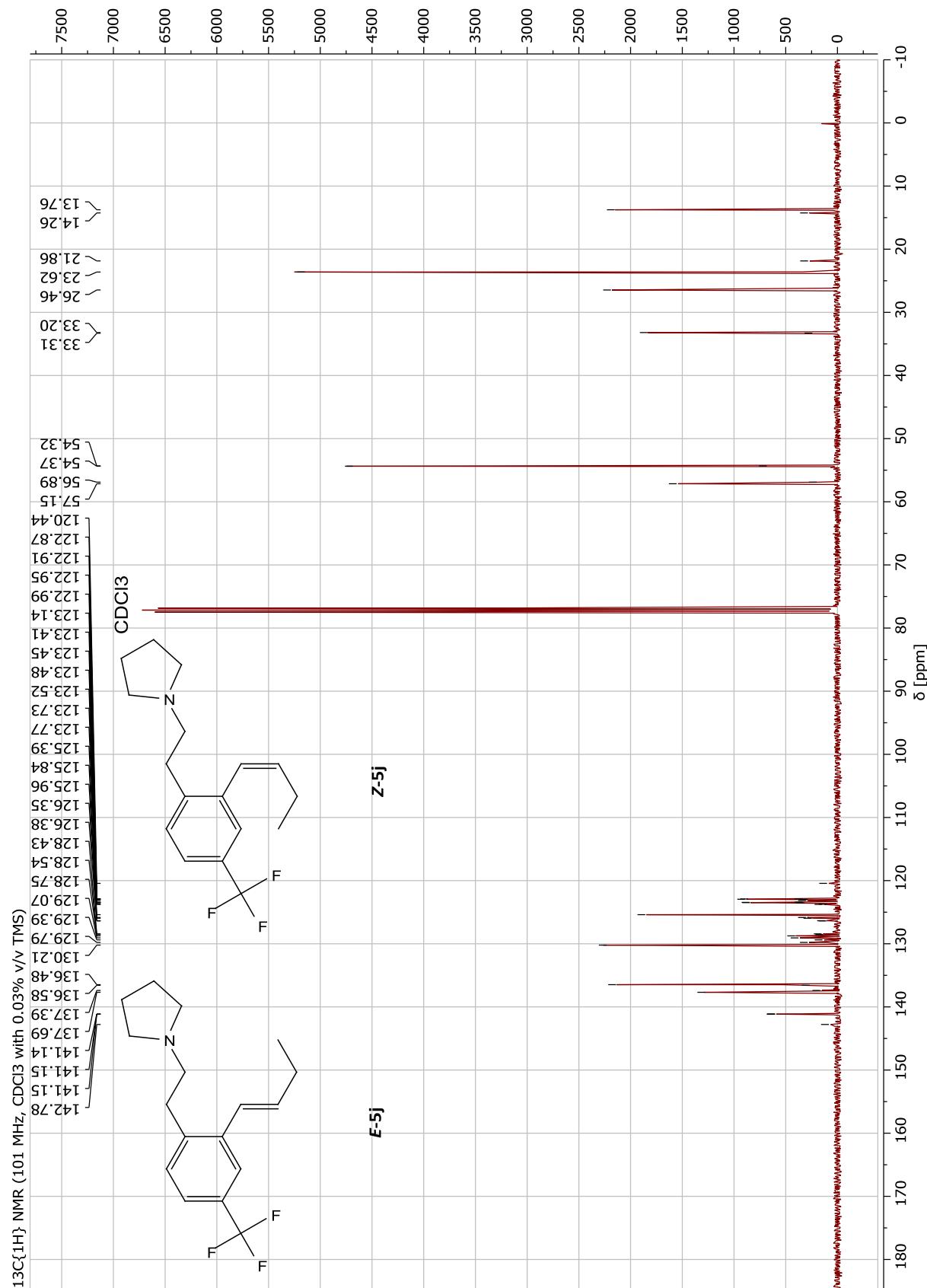
$^{13}\text{C}\{^1\text{H}\}$ NMR of 5i



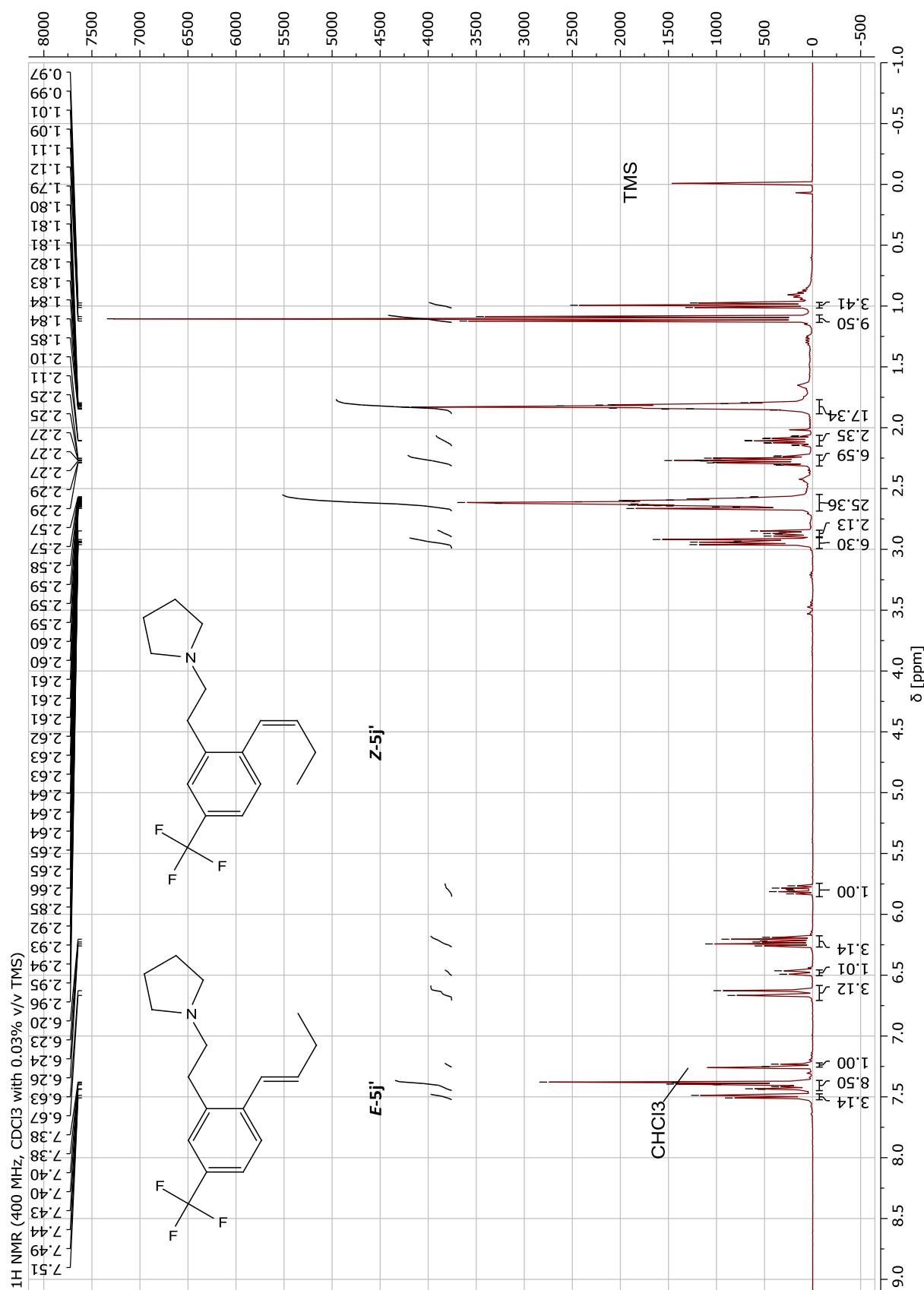
¹H NMR of 5j



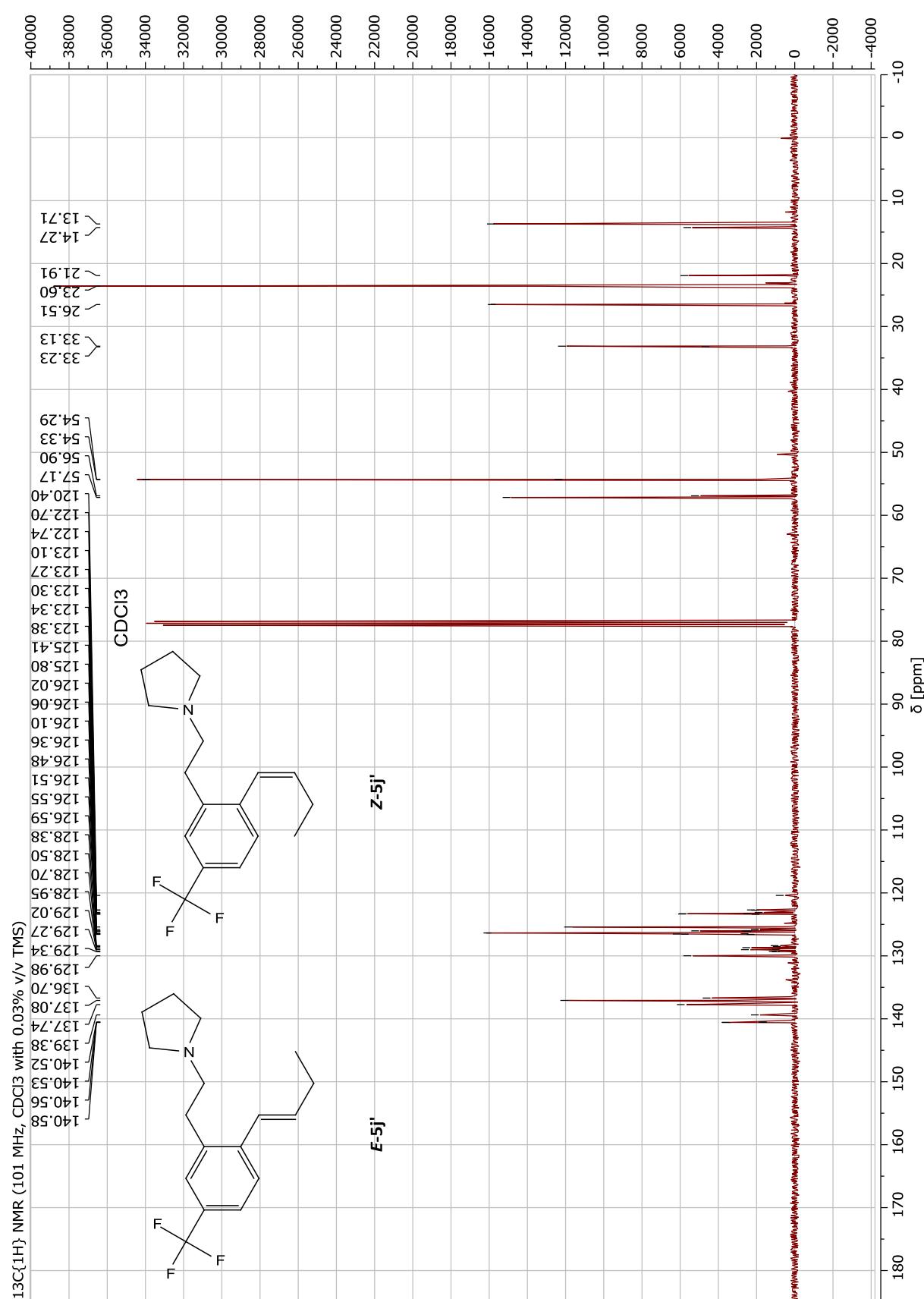
$^{13}\text{C}\{\text{H}\}$ NMR of 5j



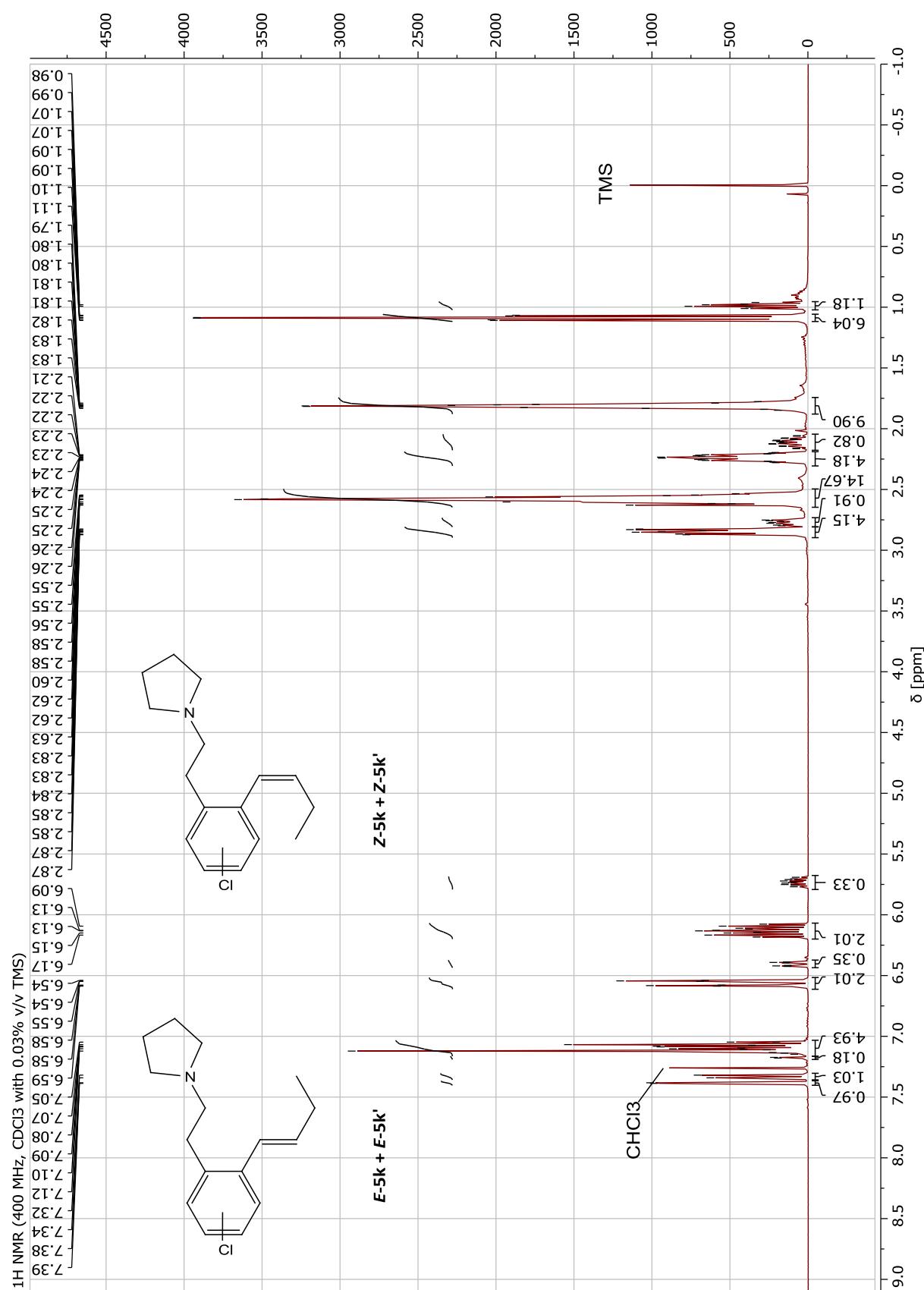
¹H NMR of 5j'

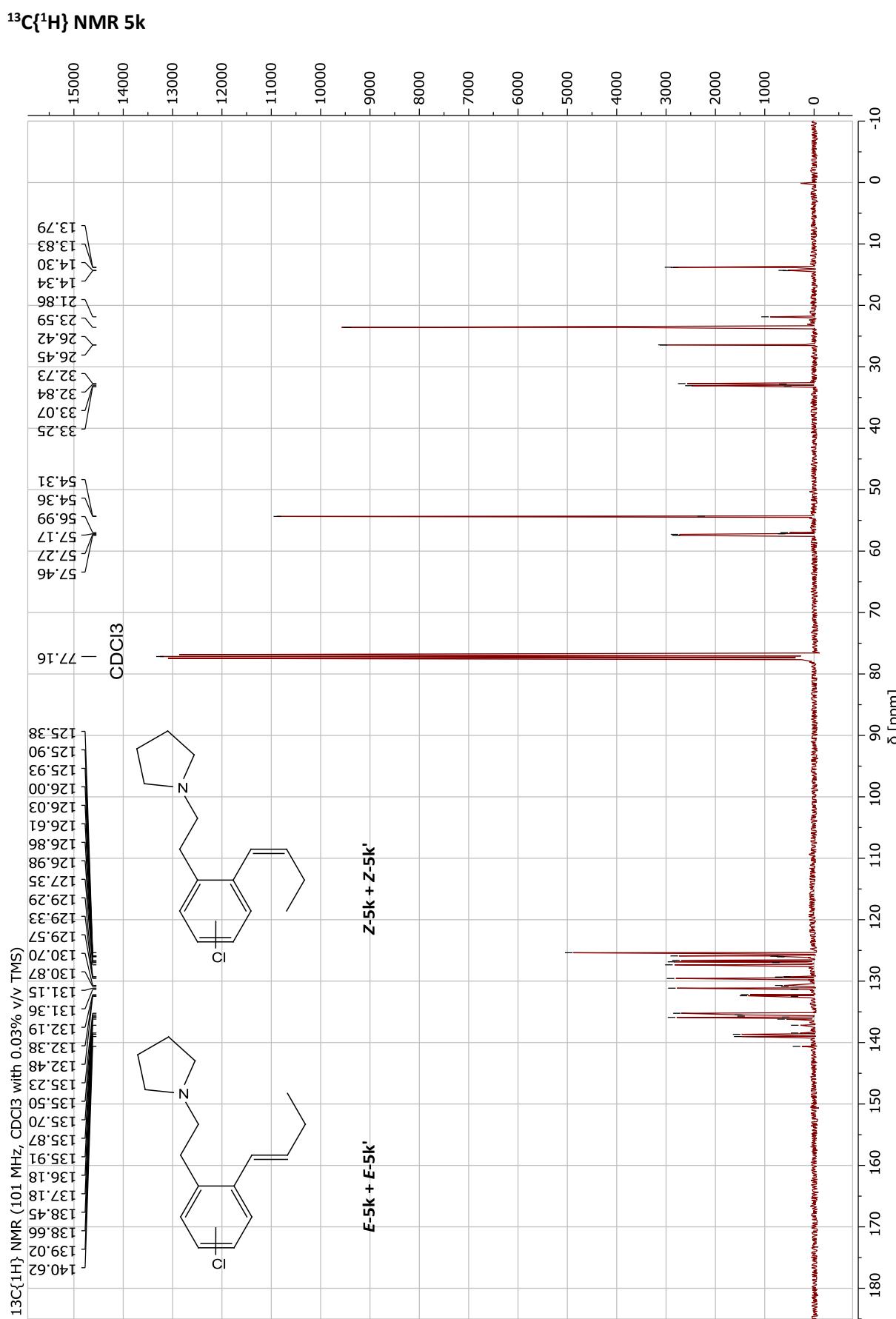


$^{13}\text{C}\{\text{H}\}$ NMR of 5j'

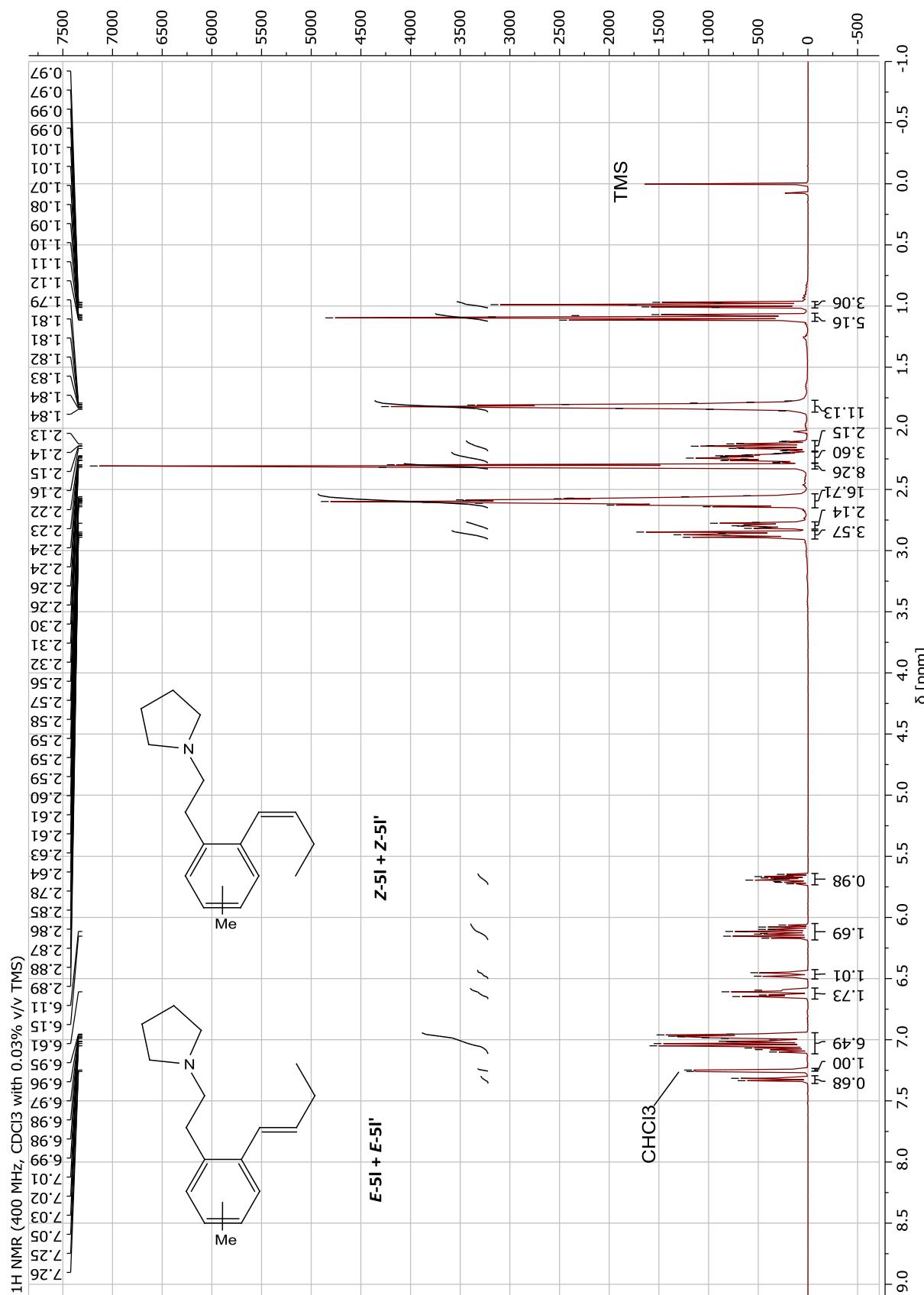


¹H NMR of 5k

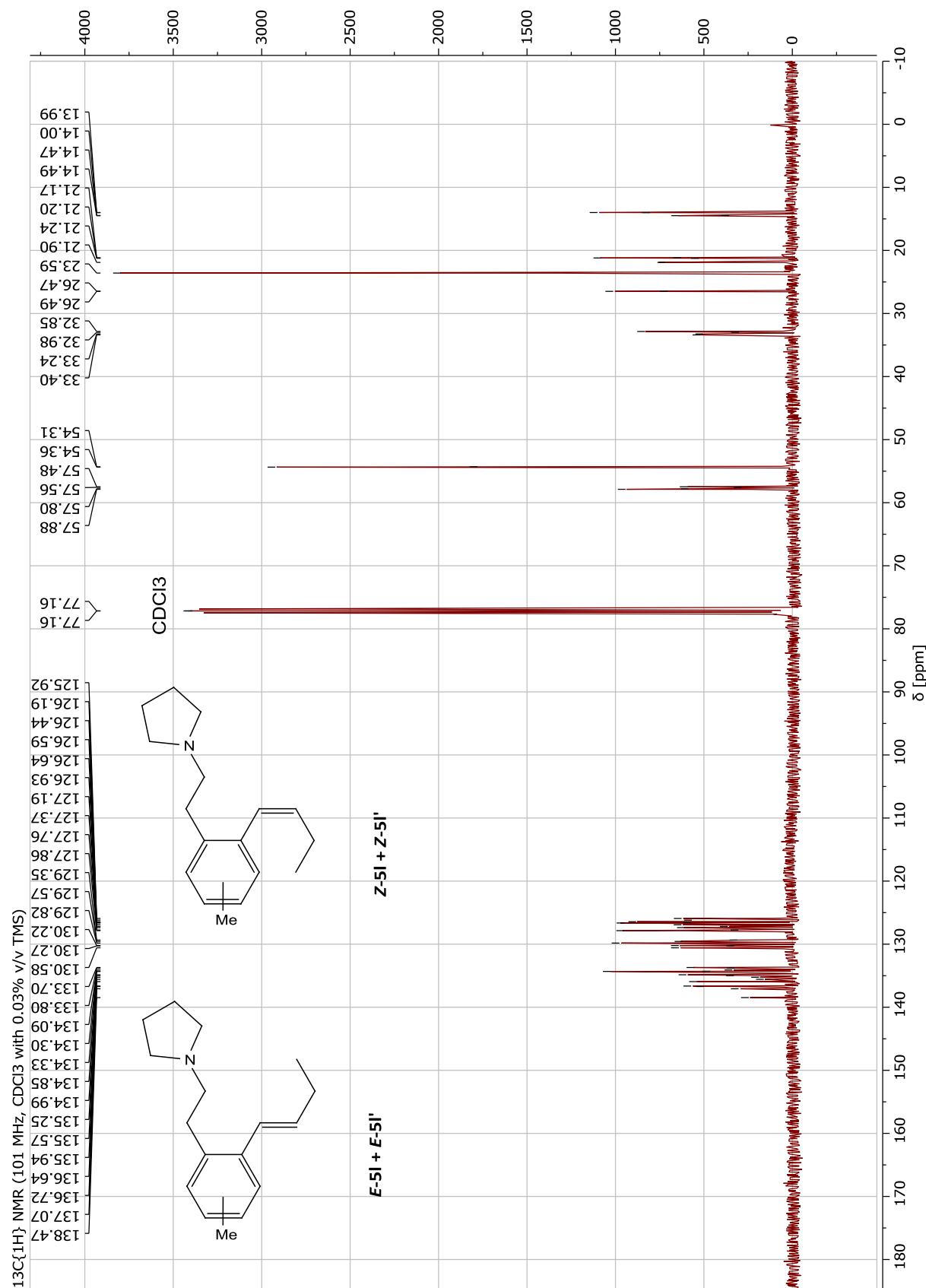




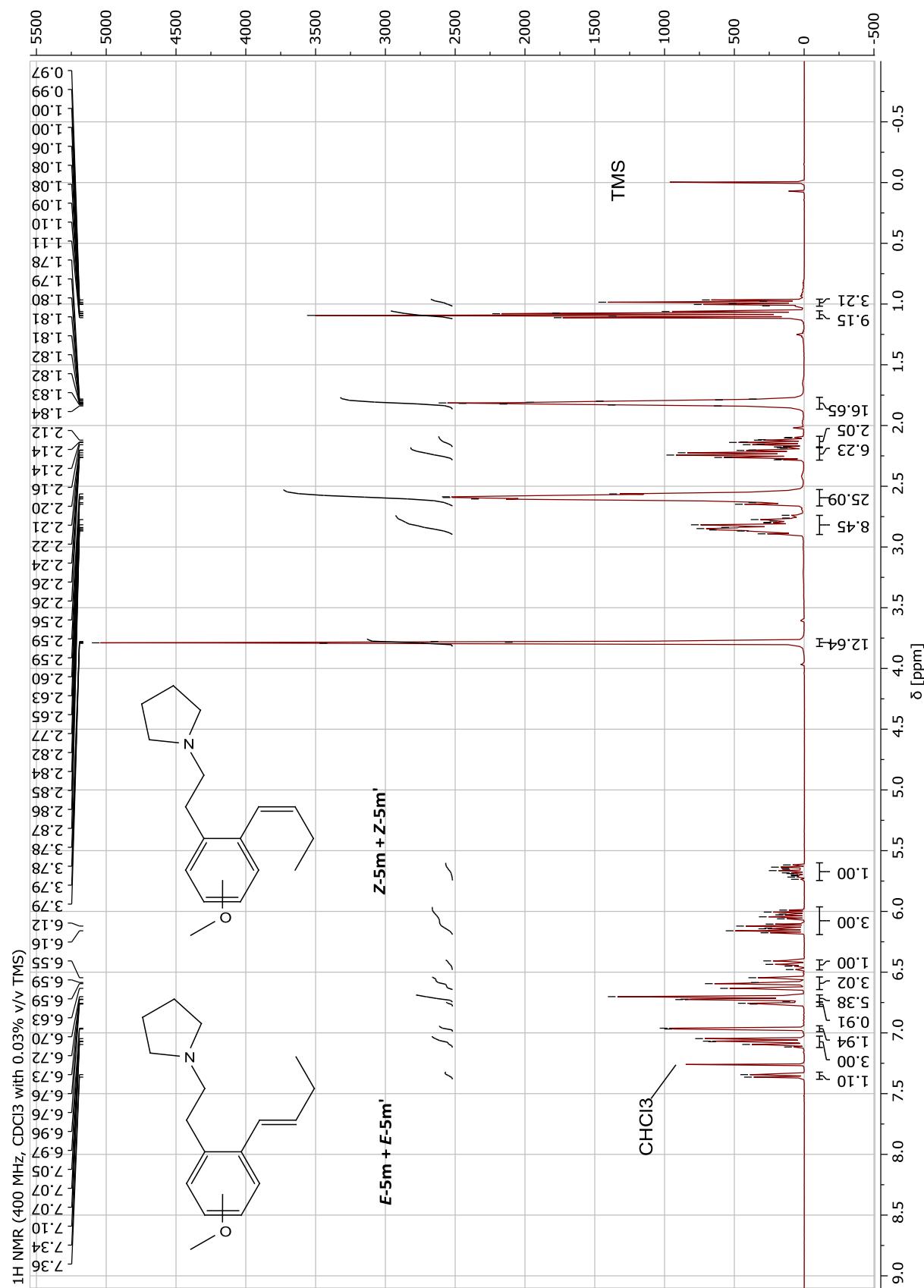
¹H NMR of 5I



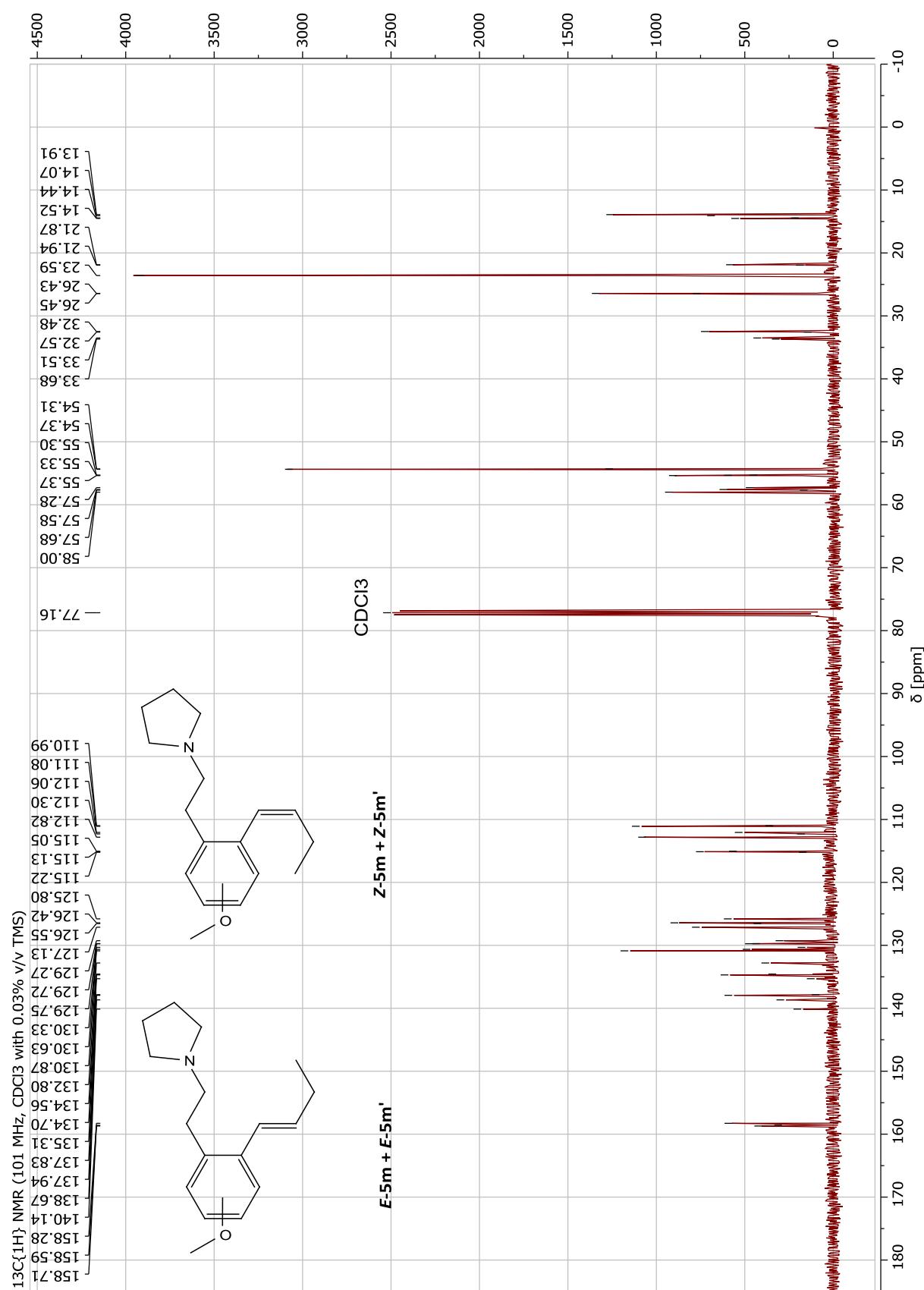
$^{13}\text{C}\{^1\text{H}\}$ NMR of 5



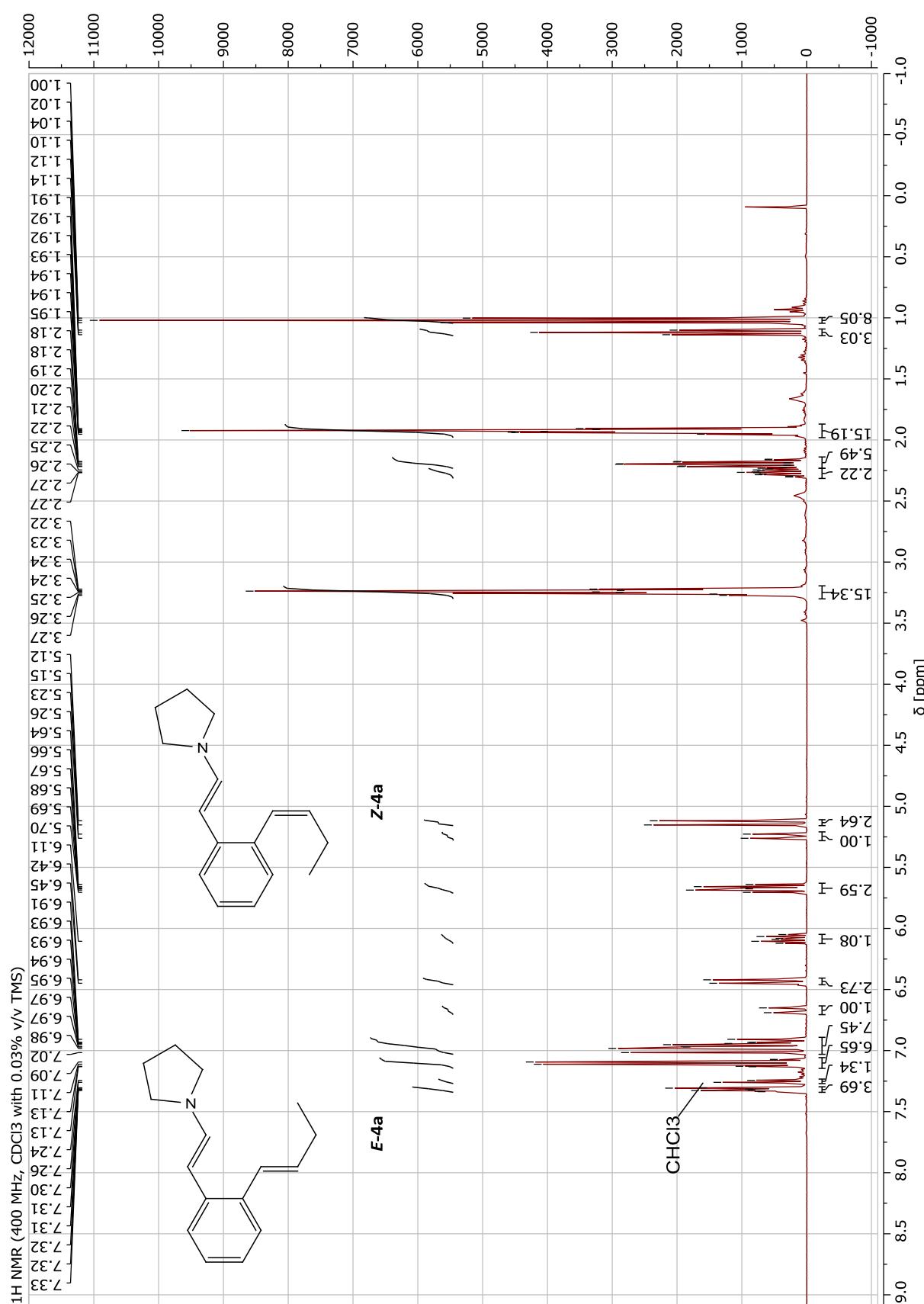
¹H NMR of 5m



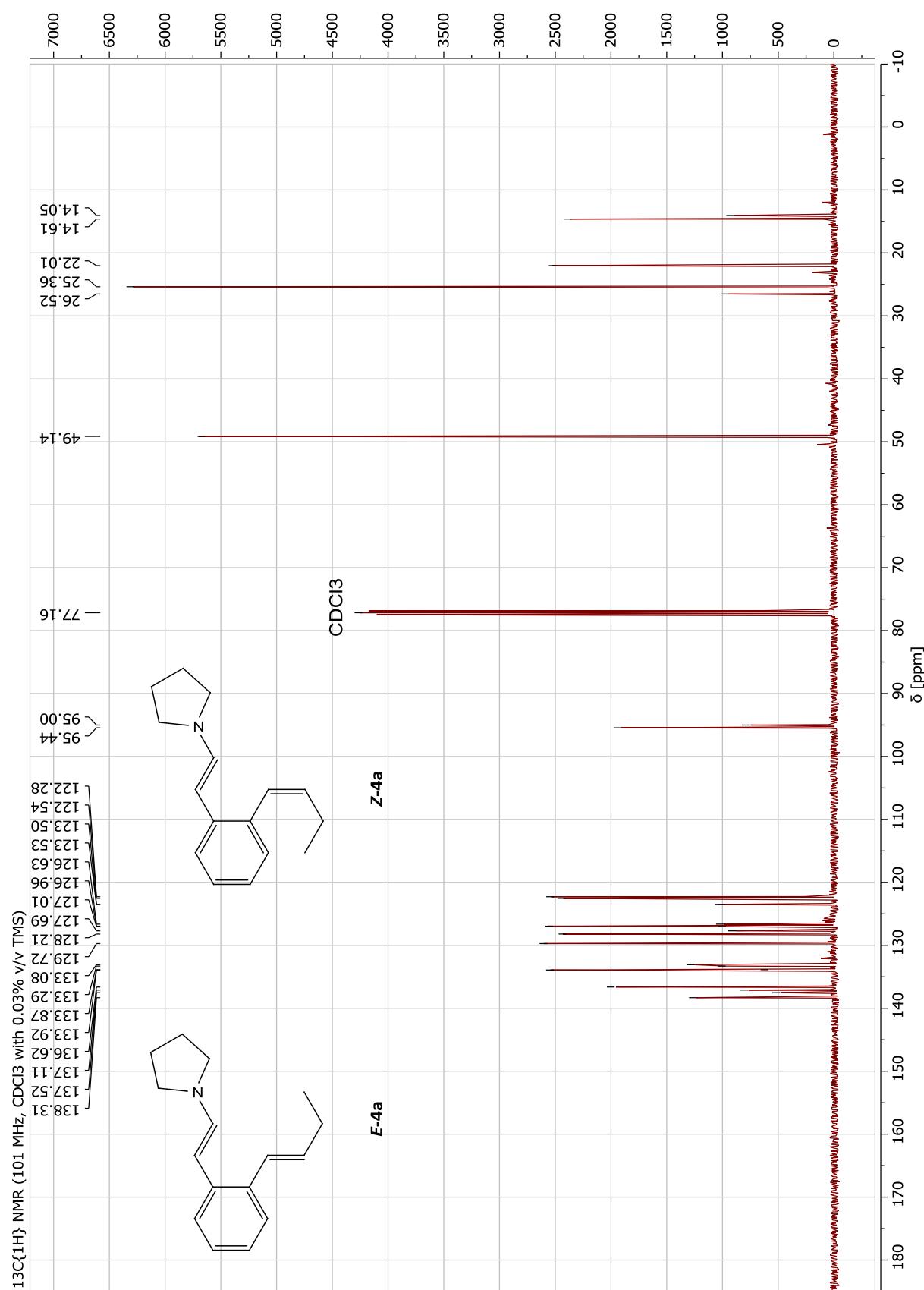
$^{13}\text{C}\{^1\text{H}\}$ NMR of 5m



¹H NMR of 4a



$^{13}\text{C}\{^1\text{H}\}$ NMR of 4a



Computations:

All computations were carried out with the Gaussian 16 software package.⁷ Geometry optimizations were performed on a B3LYP-D3 level using the def2-TZVP basis set with the IEFPCM solvent model.⁸⁻¹¹ Additionally, the opt=tight and int=ultrafine keywords were applied. The absence of imaginary frequencies in ground state was checked for every optimized geometry. All possible orientations of the substituents were taken into consideration. For each structure the substituent arrangement with the lowest energy was used. Molecular structures were visualized using the CYLview software.¹² For the time-dependent DFT computations also the previously optimized B3LYP structures were used.¹³ Therefore, the Cam-B3LYP method with the cc-pVTZ basis set and the IEFPCM solvent model was applied.¹⁴⁻¹⁶ Using these parameters, the experimentally observed absorption for *o*-quinodimethane in acetonitrile solution by Trahanovsky *et al.* could be reproduced with high accuracy.¹⁷

Table S1: Computed vertical excitation energies of the low-lying excited state.

The values are given in nm.

Compound	<i>o</i> -quinodimethane		3a	3i	3m-c2	3m-c3
	computed (ACN)	experimental (ACN)	computed (THF)			
HOMO-LUMO transition	378	367	417	419	411	412

Table S2: Coordinates and energy parameters of *o*-quinodimethane (B3LYP-D3/def2-TZVP, IEFPCM=ACN).

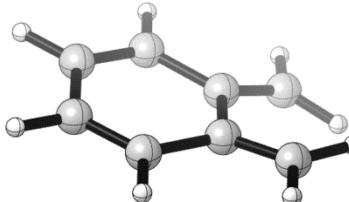
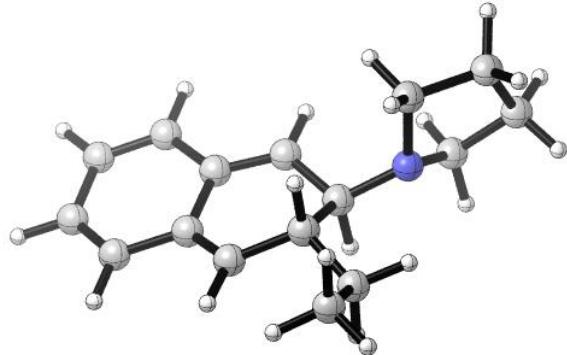
B3LYP-D3/def2-TZVP (SCRF=IEFPCM=ACN)			
XYZ coordinates			
C	1.829131	0.716942	-0.104094
C	0.672225	1.403511	-0.162687
C	-0.620029	0.746388	0.002351
C	-0.620029	-0.746388	-0.002351
C	0.672225	-1.403511	0.162687
C	1.829131	-0.716942	0.104094
H	2.776781	1.233332	-0.191780
H	0.675277	2.480530	-0.281547
H	0.675277	-2.480530	0.281546
H	2.776781	-1.233332	0.191780
C	-1.727914	1.485517	0.207508
H	-2.689366	1.041824	0.426515
H	-1.683172	2.566631	0.174642
C	-1.727914	-1.485517	-0.207508
H	-1.683172	-2.566631	-0.174642
H	-2.689366	-1.041824	-0.426515
			
E (Hartree/Particle)	-309.736208297		
Zero-point correction	0.131937		
Thermal correction to Energy	0.138660		
Thermal correction to Enthalpy	0.139605		
Thermal correction to Gibbs Free Energy	0.101131		
Sum of electronic and zero-point Energies	-309.604271		
Sum of electronic and thermal Energies	-309.597548		
Sum of electronic and thermal Enthalpies	-309.596604		
Sum of electronic and thermal Free Energies	-309.635077		

Table S3: Coordinates and energy parameters of **3a** (B3LYP-D3/def2-TZVP, IEFPCM=THF).

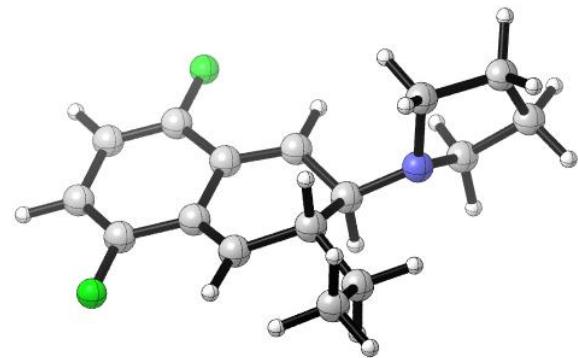
B3LYP-D3/def2-TZVP (SCRF=IEFPCM=THF)			
XYZ coordinates			
C	1.658649	-0.931785	0.039550
C	2.191582	0.443343	-0.098287
C	0.333633	-1.141883	-0.105875
C	1.337362	1.489717	-0.094451
H	-0.060693	-2.149048	-0.041269
H	1.735275	2.497105	-0.137053
C	4.461284	-0.443832	0.013994
H	5.534628	-0.302735	-0.011243
C	3.937926	-1.775127	0.249375
H	4.635200	-2.589827	0.400380
C	3.632157	0.606719	-0.155457
C	2.608734	-2.004722	0.267278
H	2.220088	-3.004781	0.421561
H	4.025835	1.605002	-0.308627
C	-0.963197	2.500154	-0.437346
H	-0.710867	2.653317	-1.491950
H	-2.016935	2.219762	-0.407310
C	-0.600859	-0.023099	-0.512064
C	-0.145342	1.315469	0.096906
H	-0.312923	1.262361	1.186887
N	-2.015064	-0.283696	-0.264142
C	-2.628811	-1.328465	-1.076884
H	-2.114321	-2.299445	-0.975826
H	-2.605332	-1.052929	-2.133106
C	-4.045295	-1.426878	-0.503106
H	-4.465018	-2.425635	-0.622953
H	-4.704533	-0.727048	-1.017447
C	-3.881808	-1.020258	0.984962
H	-4.095033	-1.846807	1.662829
H	-4.562199	-0.207927	1.240014
C	-2.415203	-0.568449	1.112564
H	-2.298835	0.314026	1.742910
H	-1.803853	-1.370356	1.558308
C	-0.757468	3.796776	0.343394
H	-1.028883	3.664370	1.394005
H	0.278389	4.140899	0.311241
H	-1.379155	4.597112	-0.062767
H	-0.505453	0.082374	-1.606900



E (Hartree/Particle)	-677.331485824
Zero-point correction	0.339940
Thermal correction to Energy	0.349593
Thermal correction to Enthalpy	0.350537
Thermal correction to Gibbs Free Energy	0.290135
Sum of electronic and zero-point Energies	-676.997492
Sum of electronic and thermal Energies	-676.981893
Sum of electronic and thermal Enthalpies	-676.980949
Sum of electronic and thermal Free Energies	-677.041351

Table S4: Coordinates and energy parameters of **3i** (B3LYP-D3/def2-TZVP, IEFPCM=THF).

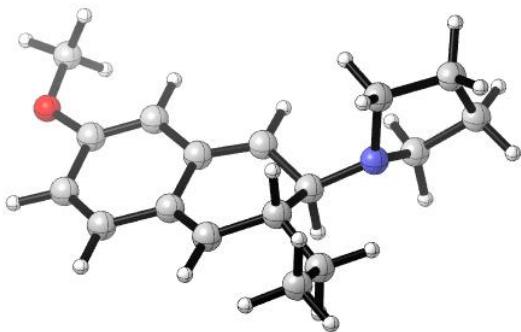
B3LYP-D3/def2-TZVP (SCRF=IEFPCM=THF)			
XYZ coordinates			
C	1.278764	-0.873349	-0.006287
C	1.836254	0.496499	-0.072046
C	-0.045154	-1.060521	-0.166201
C	1.008478	1.559522	-0.025392
H	-0.451345	-2.062411	-0.149242
H	1.429004	2.555902	-0.014181
C	4.105600	-0.441639	0.023227
H	5.176077	-0.292784	0.021649
C	3.557831	-1.773684	0.175306
H	4.218657	-2.622306	0.281927
C	3.278065	0.607140	-0.099427
C	2.228073	-1.950463	0.167866
C	-1.266470	2.620021	-0.364858
H	-0.999823	2.802163	-1.411086
H	-2.324841	2.357406	-0.354140
C	-0.956926	0.094253	-0.521071
C	-0.478113	1.401785	0.138630
H	-0.656403	1.317450	1.224384
N	-2.372933	-0.150243	-0.279722
C	-3.010320	-1.145337	-1.136354
H	-2.515947	-2.130204	-1.080486
H	-2.984512	-0.822966	-2.179082
C	-4.425706	-1.238577	-0.561079
H	-4.872842	-2.217380	-0.733714
H	-5.067118	-0.493231	-1.032186
C	-4.244853	-0.919507	0.945862
H	-4.456240	-1.783453	1.575808
H	-4.917725	-0.120110	1.255441
C	-2.773227	-0.486470	1.086124
H	-2.646928	0.369023	1.750835
H	-2.168255	-1.311075	1.497383
C	-1.044973	3.885580	0.460767
H	-1.329557	3.723770	1.503630
H	-0.003206	4.212625	0.449430
H	-1.648398	4.709271	0.074724
H	-0.858825	0.237434	-1.610816
F	1.707165	-3.195324	0.287110
F	3.782259	1.858775	-0.218513



E (Hartree/Particle)	-875.896715562
Zero-point correction	0.317910
Thermal correction to Energy	0.335271
Thermal correction to Enthalpy	0.336215
Thermal correction to Gibbs Free Energy	0.271258
Sum of electronic and zero-point Energies	-875.578805
Sum of electronic and thermal Energies	-875.561445
Sum of electronic and thermal Enthalpies	-875.560501
Sum of electronic and thermal Free Energies	-875.625457

Table S5: Coordinates and energy parameters of **3m-c2** (B3LYP-D3/def2-TZVP, IEFPCM=THF).

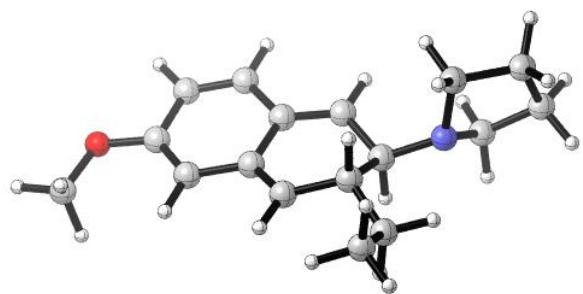
B3LYP-D3/def2-TZVP (SCRF=IEFPCM=THF)			
XYZ coordinates			
C	1.229209	-0.164485	-0.091448
C	1.306504	1.313241	-0.151207
C	0.023958	-0.757254	-0.225467
C	0.177873	2.048870	-0.071141
H	-0.044589	-1.838556	-0.225591
H	0.252500	3.130286	-0.061275
C	3.738711	1.172720	-0.124615
H	4.724833	1.617838	-0.152346
C	3.658881	-0.268984	0.036892
C	2.623277	1.917533	-0.218478
C	2.466012	-0.908867	0.059087
H	2.399510	-1.983526	0.151318
H	2.688551	2.994719	-0.316959
C	-2.327396	2.331427	-0.330567
H	-2.163452	2.602420	-1.378901
H	-3.246448	1.744590	-0.300732
C	-1.223965	0.040076	-0.540017
C	-1.176015	1.425466	0.129654
H	-1.287358	1.275263	1.217496
N	-2.479156	-0.653084	-0.259331
C	-2.787892	-1.793306	-1.115445
H	-1.998263	-2.563927	-1.093554
H	-2.903515	-1.471393	-2.152347
C	-4.076198	-2.349938	-0.502100
H	-4.177063	-3.422299	-0.669573
H	-4.942804	-1.862402	-0.949653
C	-3.968758	-1.985844	1.002062
H	-3.889693	-2.870043	1.634703
H	-4.848490	-1.431284	1.328453
C	-2.701593	-1.117395	1.108599
H	-2.823516	-0.275813	1.791625
H	-1.854621	-1.717782	1.479129
C	-2.500463	3.592246	0.514283
H	-2.688417	3.335836	1.560182
H	-1.618406	4.235441	0.485890
H	-3.346382	4.184338	0.159455
H	-1.225368	0.218945	-1.629166
O	4.881715	-0.853371	0.144606
C	4.936242	-2.266476	0.308091
H	4.494892	-2.779440	-0.551059
H	5.990338	-2.521707	0.382692
H	4.417818	-2.577128	1.219572



E (Hartree/Particle)	-791.910158560
Zero-point correction	0.366465
Thermal correction to Energy	0.384656
Thermal correction to Enthalpy	0.385600
Thermal correction to Gibbs Free Energy	0.319186
Sum of electronic and zero-point Energies	-791.543693
Sum of electronic and thermal Energies	-791.525502
Sum of electronic and thermal Enthalpies	-791.524558
Sum of electronic and thermal Free Energies	-791.590973

Table S6: Coordinates and energy parameters of **3m-c3** (B3LYP-D3/def2-TZVP, IEFPCM=THF).

B3LYP-D3/def2-TZVP (SCRF=IEFPCM=THF)			
XYZ coordinates			
C	0.914047	-0.991869	0.059828
C	1.498044	0.363823	-0.057713
C	-0.413695	-1.161457	-0.102755
C	0.673176	1.433941	-0.055031
H	-0.837849	-2.157127	-0.049252
H	1.099982	2.429963	-0.079446
C	3.728904	-0.606056	0.058682
C	3.154728	-1.922120	0.280142
H	3.840031	-2.747028	0.426152
C	2.943605	0.483981	-0.107333
C	1.822026	-2.100244	0.286264
H	1.399835	-3.087818	0.430195
H	3.363237	1.470412	-0.243884
C	-1.589473	2.517064	-0.426372
H	-1.316344	2.663655	-1.476842
H	-2.652437	2.271966	-0.411954
C	-1.310758	-0.012617	-0.505748
C	-0.818371	1.305711	0.117424
H	-1.010232	1.254457	1.203261
N	-2.734294	-0.235170	-0.271290
C	-3.371662	-1.250709	-1.102617
H	-2.886010	-2.237531	-1.011216
H	-3.332183	-0.961901	-2.154818
C	-4.794883	-1.316336	-0.540704
H	-5.241375	-2.301415	-0.676134
H	-5.430310	-0.592095	-1.051201
C	-4.631742	-0.932677	0.953517
H	-4.876181	-1.760117	1.619684
H	-5.288902	-0.102681	1.212646
C	-3.153170	-0.527651	1.098272
H	-3.014418	0.341348	1.742437
H	-2.570304	-1.354669	1.536388
C	-1.353569	3.805691	0.359221
H	-1.641248	3.679868	1.406375
H	-0.307276	4.117517	0.339906
H	-1.945674	4.625606	-0.052312
H	-1.202341	0.098543	-1.598622
O	5.088748	-0.620232	0.055381
C	5.769926	0.612009	-0.154315
H	6.831446	0.378756	-0.129440
H	5.511597	1.043858	-1.125360
H	5.534233	1.331571	0.634855



E (Hartree/Particle)	-791.909988218
Zero-point correction	0.366506
Thermal correction to Energy	0.384668
Thermal correction to Enthalpy	0.385612
Thermal correction to Gibbs Free Energy	0.319447
Sum of electronic and zero-point Energies	-791.543483
Sum of electronic and thermal Energies	-791.525320
Sum of electronic and thermal Enthalpies	-791.524376
Sum of electronic and thermal Free Energies	-791.590541

References:

- (1) Ahles S.; Wegner H. A. "Recipe for the Preparation of a Bidentate Lewis Acid Catalyst", **2015**. www.beilstein.tv/video/recipe-for-the-preparation-of-a-bidentate-lewis-acid-catalyst/ (accessed March 16th, 2019).
- (2) Kessler, S. N.; Neuburger, M.; Wegner, H. A. *Eur. J. Org. Chem.* **2011**, *2011*, 3238–3245.
- (3) Kessler, S. N.; Wegner, H. A. *Org. Lett.* **2012**, *14*, 3268–3271.
- (4) Kessler, S. N.; Neuburger, M.; Wegner, H. A. *J. Am. Chem. Soc.* **2012**, *134*, 17885–17888.
- (5) Allegretti, P. A.; Ferreira, E. M. *Chem. Sci.* **2013**, *4*, 1053–1058.
- (6) Ahles, S.; Götz, S.; Schweighauser, L.; Brodsky, M.; Kessler, S. N.; Heindl, A. H.; Wegner, H. A. *Org. Lett.* **2018**, *20*, 7034–7038.
- (7) Frisch, M. J.; Trucks, G. W.; Schlegel, H. B.; Scuseria, G. E.; Robb, M. A.; Cheeseman, J. R.; Scalmani, G.; Barone, V.; Petersson, G. A.; Nakatsuji, H.; Li, X.; Caricato, M.; Marenich, A. V.; Bloino, J.; Janesko, B. G.; Gomperts, R.; Mennucci, B.; Hratchian, H. P.; Ortiz, J. V.; Izmaylov, A. F.; Sonnenberg, J. L.; Williams; Ding, F.; Lipparini, F.; Egidi, F.; Goings, J.; Peng, B.; Petrone, A.; Henderson, T.; Ranasinghe, D.; Zakrzewski, V. G.; Gao, J.; Rega, N.; Zheng, G.; Liang, W.; Hada, M.; Ehara, M.; Toyota, K.; Fukuda, R.; Hasegawa, J.; Ishida, M.; Nakajima, T.; Honda, Y.; Kitao, O.; Nakai, H.; Vreven, T.; Throssell, K.; Montgomery Jr., J. A.; Peralta, J. E.; Ogliaro, F.; Bearpark, M. J.; Heyd, J. J.; Brothers, E. N.; Kudin, K. N.; Staroverov, V. N.; Keith, T. A.; Kobayashi, R.; Normand, J.; Raghavachari, K.; Rendell, A. P.; Burant, J. C.; Iyengar, S. S.; Tomasi, J.; Cossi, M.; Millam, J. M.; Klene, M.; Adamo, C.; Cammi, R.; Ochterski, J. W.; Martin, R. L.; Morokuma, K.; Farkas, O.; Foresman, J. B.; Fox,D. J. *Gaussian 16 Rev. B.01*; Wallingford, CT, **2016**.
- (8) Becke, A. D. *J. Chem. Phys.* **1993**, *98*, 5648–5652.
- (9) Lee, C.; Yang, W.; Parr, R. G. *Phys. Rev. B* **1988**, *37*, 785–789.
- (10) Grimme, S. *J. Comput. Chem.* **2004**, *25*, 1463–1473.
- (11) Caricato, M. *J. Chem. Theory Comput.* **2012**, *8*, 4494–4502.
- (12) Legault, C. Y. *CYLview*; Université de Sherbrooke: Quebec, Canada, **2009**. www.Cylview.org.
- (13) Runge, E.; Gross, E. K. U. *Phys. Rev. Lett.* **1984**, *52*, 997–1000.
- (14) Davidson, E. R. *Chem. Phys. Lett.* **1996**, *260*, 514–518.
- (15) Dunning, T. H. *J. Chem. Phys.* **1989**, *90*, 1007–1023.
- (16) Yanai, T.; Tew, D. P.; Handy, N. C. *Chem. Phys. Lett.* **2004**, *393*, 51–57.
- (17) Trahanovsky, W. S.; Macias, J. R. *J. Am. Chem. Soc.* **1986**, *108*, 6820–6821.