SUPPLEMENTARY METHODS

1. Analytical Instrumentation.

NMR spectra were recorded on a Bruker Avance III 400 spectrometer at 400 MHz for ¹H NMR and 100 MHz for ¹³C NMR using residual CHCl₃ as an internal reference. Low-resolution MS was recorded on a ThermoScientific LXQ mass spectrometer.

2. Extraction and Isolation.

Dried and powdered aerial parts (1004.3 g) of aeroponically grown¹ *Physalis acutifolia* (family: Solanaceae) was extracted with CH₃OH (4.6 L × 2) for 2 h at room temperature in an ultrasonic bath. Evaporation of CH₃OH under reduced pressure yielded the crude extract (122.36 g) which was subjected to solvent-solvent partitioning with hexanes (500 mL) and 80% aqueous CH₃OH (4:1, v/v) (500 mL). The 80% aqueous CH₃OH layer was diluted with H₂O to make it to 50% aqueous CH₃OH, which was then extracted with CHCl₃ (500 mL × 2). The CHCl₃ layer was evaporated to yield the CHCl₃ fraction (4.1 g) which was found to contain physalins. This fraction was subjected to chromatography over HP-20SS resin (500 g) and successively eluted with CH₂Cl₂/hexanes (4:1, 700 mL), CH₂Cl₂/acetone (4:1, 600 mL), CH₂Cl₂/acetone (2:3, 600 mL), and CH₃OH (600 mL) to yield seven fractions [A (134 mg), B (139 mg), C (751 mg), D (699 mg), E (343 mg), F (547 mg), and G (1452 mg)] combined based on their TLC profiles. Fraction C was separated by silica gel chromatography (60 g) and elution with mixtures of hexane/EtOAc (6:1, 4:1, and 2:1, each 400 mL) to provide ten combined fractions, C1–C10 of which C7 was found to contain physalins. Further fractionation of C7 (428 mg) by silica gel (40 g) column chromatography and elution with CH₂Cl₂/EtOAc (10:1, 700 mL) afforded a white amorphous solid (331 mg) which was identified as physalin F by comparison of its ¹H NMR, ¹³C NMR and MS data with those reported.^{2,3}

References

- Xu, Y., Bunting, D. P., Liu, M. X., Bandaranayake, H. A., and Gunatilaka, A. A. L. (2016) 17β-Hydroxy-18-acetoxywithanolides from aeroponically grown *Physalis crassifolia* and their potent and selective cytotoxicity for prostate cancer cells, *J. Nat. Prod.* 79, 821–30.
- (2) Ozawa, M., Morita, M., Hirai, G., Tamura, S., Kawai, M., Tsuchiya, A., Oonuma, K., Maruoka, K., and Sodeoka, M. (2013) Contribution of cage-shaped structure of physalins to their mode of action in inhibition of NF-κB activation. ACS Med. Chem. Lett. 4, 730–35.
- (3) Jacobo-Herrera, N. J., Bremner, P., Márquez, N., Gupta, M. P., Gibbons, S., Muñoz, E., and Heinrich, M.
 (2006) Physalins from *Witheringia solanacea* as modulators of the NF-κB cascade. *J. Nat. Prod.* 69, 328–31.