

*Supplementary material related to the article*

**GALLOYL CARBOHYDRATES WITH ANTIANGIOGENIC ACTIVITY MEDIATED BY  
CAPILLARY MORPHOGENESIS GENE 2 (CMG2) PROTEIN BINDING**

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Includes:

1. Structural assignments
2. Inhibitory effects of **4**, **7**, **8**, **13**, **19** and **29** with respect to PA-CMG2 inhibition. Dose-response curves determined by a FRET interaction assay
3. Selected <sup>1</sup>H and <sup>13</sup>C NMR spectra

## 1. Structural assignments

The structures of the novel carbohydrate derivatives were assigned by  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectroscopic analysis. In the case of the gallic acid monosaccharide series, compounds **3-10**, the spectroscopic data were identical to those reported previously.<sup>1</sup>

For the novel 2,3,4-trihydroxybenzoyl monosaccharide and disaccharide compounds well-known NMR rules,<sup>2</sup> together with bidimensional NOESY and ROESY experiments and  $^{13}\text{C}$ - $^1\text{H}$  techniques (gHMBC<sup>3</sup> and gHSQC<sup>4</sup>) have been applied for the unequivocal assignment of the structures.

Table 1 showed all the chemical shifts and coupling constants observed for 2,3,4-trihydroxybenzoyl monosaccharides series.

**Table 1.** NMR data (chemical shifts and coupling constants) observed for 2,3,4-trihydroxybenzoyl monosaccharide series.

	<b>13</b> $\alpha/\beta$ -D-Glcp*	<b>15</b> $\alpha$ -D-Galf**	<b>18</b> $\alpha$ -D-Manp	<b>19</b> $\beta$ -D-Manp	<b>22</b> $\alpha$ -D-Ribp	<b>23</b> $\beta$ -D-Ribp
$\delta$ (ppm)						
C-1	91.76/93.97	95.85	92.65	92.13	90.71	93.84
C-2	71.95/72.45	77.38	70.27	70.72	68.22	68.88 or 69.07
C-3	72.12/74.28	75.47	71.35	72.47	68.99	67.73
C-4	70.17/ 70.75	80.68	66.60	67.25	67.41	68.88 or 69.07
C-5	72.23/74.20	71.60	72.44	73.90		64.76
C-6	63.89/64.09	63.95	62.69	62.98	-	-
H-1	6.85/6.39	6.80	6.57	6.53	6.56	6.57
H-2	5.70/5.75	5.94	5.93	6.08	5.76	5.62-5.70
H-3	6.20/6.08	6.26	5.98	5.92	6.15	6.01
H-4	5.82/5.75	4.79-4.85	6.16	6.05	5.60	5.62-5.70
H-5	4.50-4.70	5.85	4.52-4.68	4.49	4.13 ( $5_A$ ), 4.45 ( $5_B$ )	4.21 ( $5_A$ ), 4.46 ( $5_B$ )
H-6 <sub>A</sub>	4.50-4.70	4.65	4.52-4.68	4.62		
H-6 <sub>B</sub>	4.50-4.70	4.79-4.85	4.52-4.68	4.71		
$^3J_{\text{HH}}$ (Hz)						
$^3J_{1,2}$	3.7/7.9	4.8	1.9	1.1	3.9	2.7
$^3J_{2,3}$	9.9/9.6	6.4	3.2	3.2	3.5	3.9
$^3J_{3,4}$	9.9/9.6	6.4	10.2	9.9	4.0	3.9
$^3J_{4,5}$	9.9/9.3	3.6	10.1	9.9	4.2 ( $^3J_{4,5A}$ ), 9.1 ( $^3J_{4,5B}$ )	3.5
$^3J_{5,6A}$	-	6.0	-	3.0	11.5 ( $^3J_{5A,5B}$ )	13.3 ( $^3J_{5A,5B}$ )
$^3J_{5,6B}$	-	5.6	-	2.5		
$^2J_{6A,6B}$	-	12.4	-	12.5		

\*p: Pyranose form

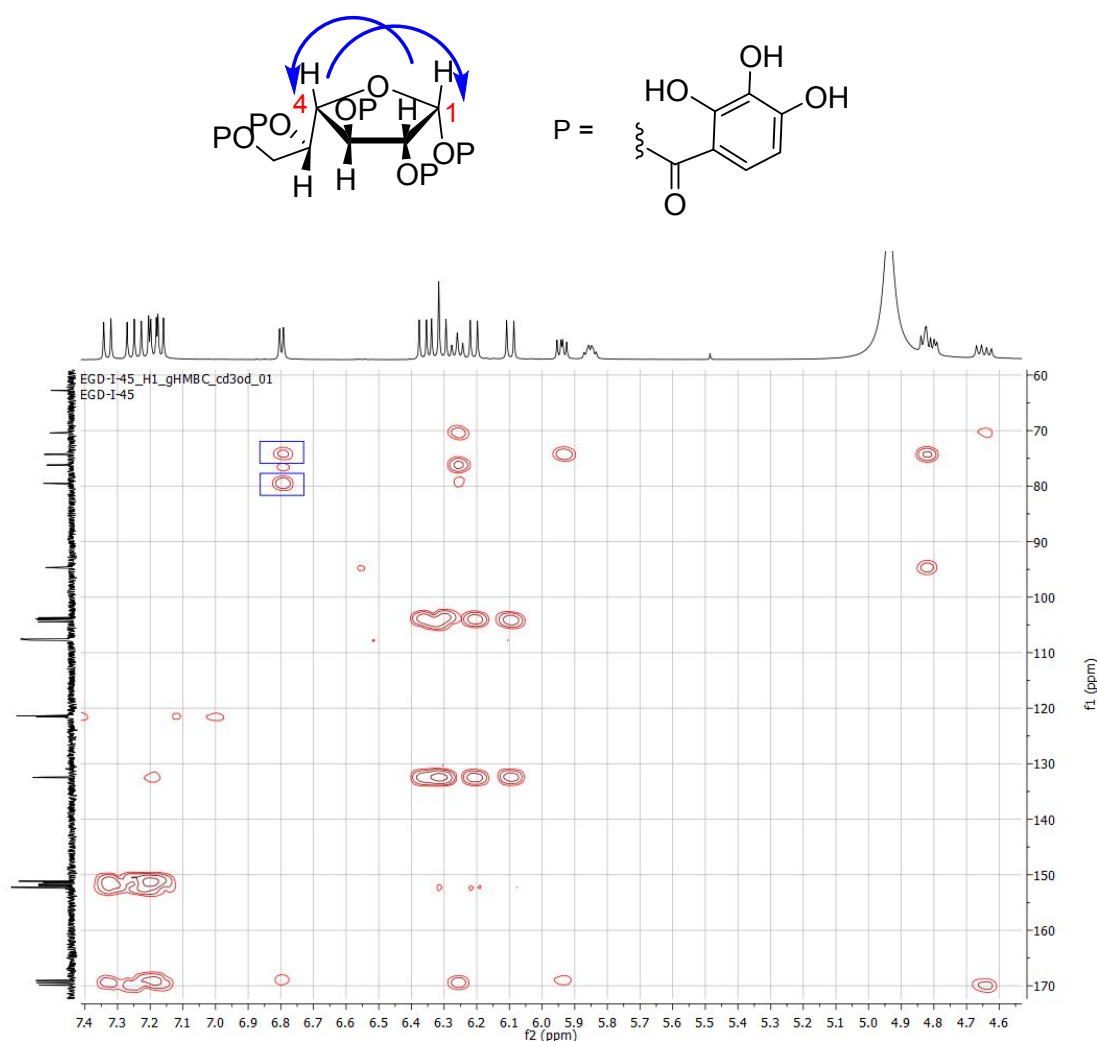
\*\*f: Furanose form

According to the NMR carbohydrate rules,<sup>2</sup> for pyranoses in a  $^4\text{C}_1$  conformation, a large coupling constant ( $J_{1,2} = 7\text{-}10$  Hz) between the anomeric H1 and the H2 indicates that both protons are in an axial configuration, whereas if they are equatorial-axial the coupling constant is smaller ( $J_{1,2} \sim 4$  Hz) and for axial-equatorial or equatorial-equatorial oriented protons even smaller coupling constants are observed ( $J_{1,2} < 2$  Hz). Following these rules the glucopyranose derivative **13** were confirmed as an anomeric  $\alpha/\beta$  mixture 1:2.

In the case of the galactose derivative **15**, the  $^{13}\text{C}$  NMR spectrum shows that the chemical shifts are in agreement with those reported for a furanose with  $\alpha$  configuration ( $\alpha$ -Galf) instead of a pyranose form (Table

1).<sup>5-7</sup> The multiplicity and  $J$  value of the H-3,  $\delta = 6.26$  (t,  $J = 6.4$  Hz, 1H, H-3) is also consistent with a furanose form. Moreover, the  $J_{1,2}$  value obtained for this compound (4.8 Hz) is characteristic of a furanose ( $\alpha$ -Gal $f$ ) configuration. Furanose conformation was corroborated using a bidimensional HMBC experiment. Thus, the 2D-HMBC spectrum (Figure 1) shows two intense correlation signals, between C-1/H-4 and C-4/H-1, only possible if the sugar is in the furanose form (C1 and C4 separated by three bonds).

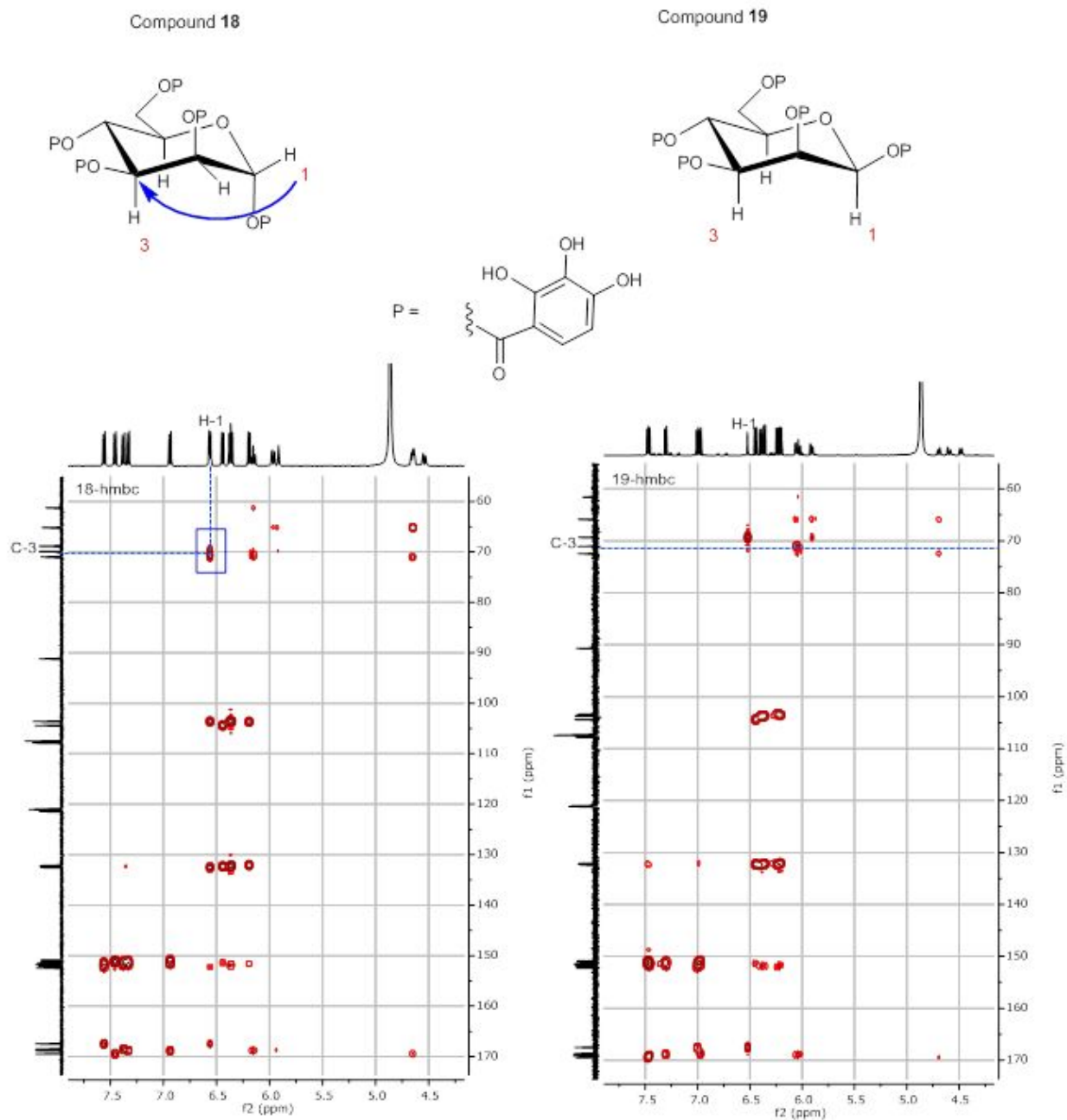
#### Compound 15



**Figure 1.** Structural assignments and HMBC correlations observed for galactose derivative **15**

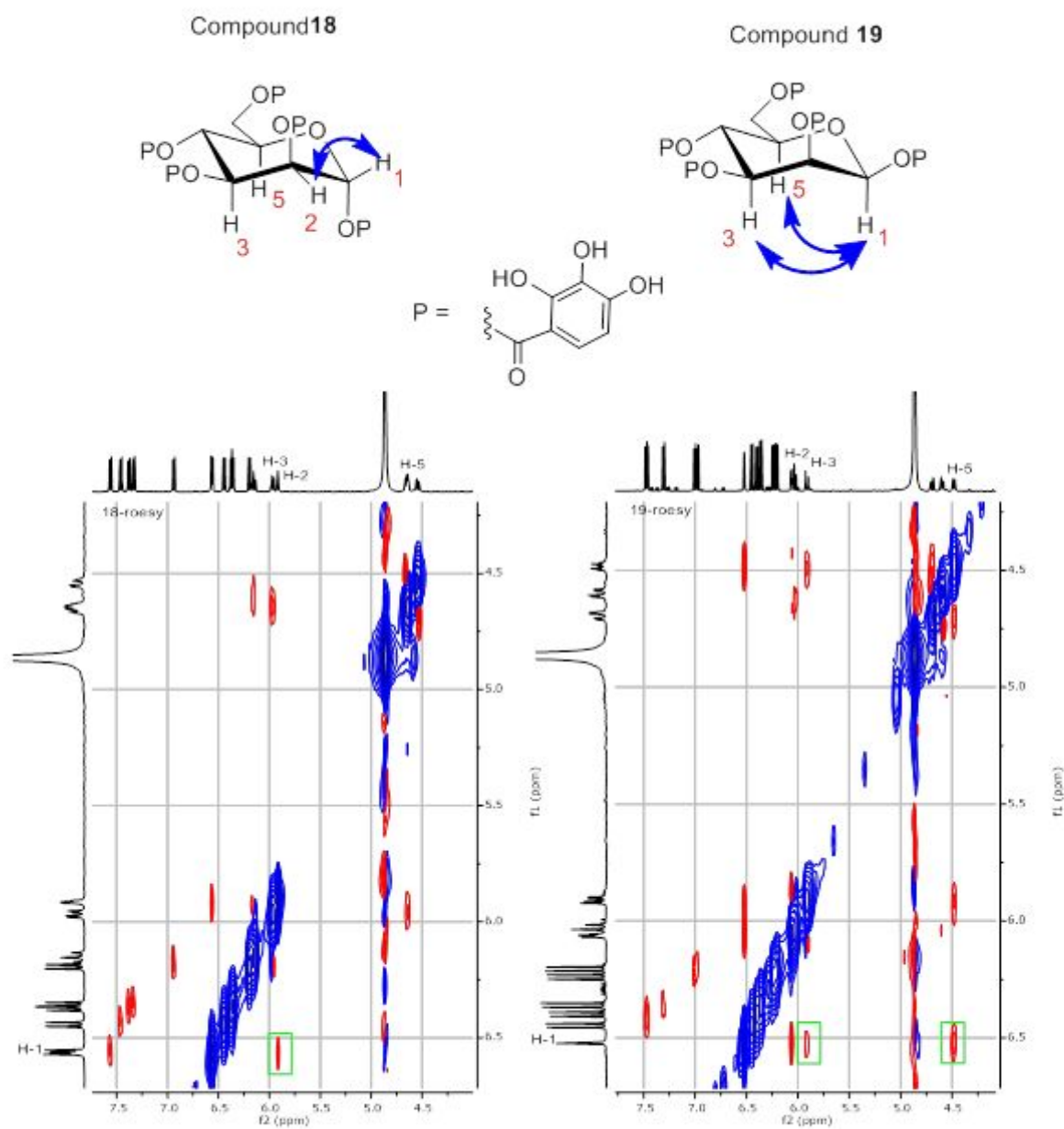
For mannose derivatives **18** and **19**, the  $J_{1,2}$  values obtained were very similar (1.9 Hz and 1.1 Hz, respectively) and  $\alpha$  and  $\beta$  assignment was not easy (Table 1).

In the HMBC spectrum for compound **18** a correlation between H-1 and C-3 was observed indicating that H-1 is situated in an equatorial position ( $\alpha$ -configuration), giving the appropriate dihedral angle value (C1, C2 and C3 in W disposition) (Figure 2 left). However HMBC for **19** ( $\beta$ -configuration) did not show such correlation (Figure 2, right).



**Figure 2.** Structural assignments and HMBC correlations observed for mannose derivatives **18** (left) and **19** (right)

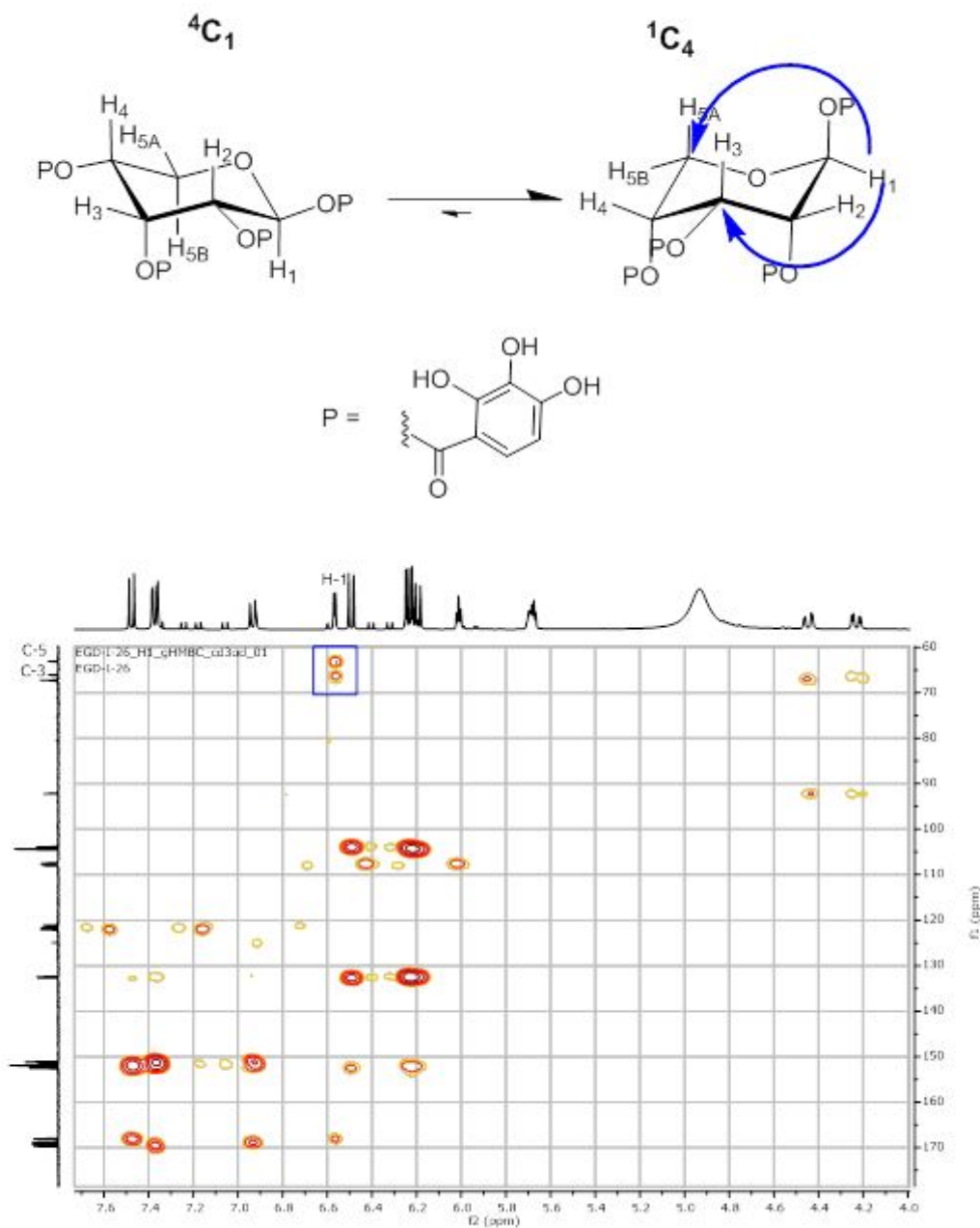
In the 2D-ROESY spectrum of **19** correlations H-1/H-3 and H-1/H-5 were observed. However, in compound **18**, H-1 only correlates with H-2 (Figure 3). These data also support the proposed structures.



**Figure 3.** Structural assignments and ROESY correlations observed for mannose derivatives **18** and **19**

The  ${}^4C_1$  conformation and  $\alpha$  configuration of the ribopyranose derivative **22** was supported by a small coupling constant ( $J_{1,2}=3.9$  Hz) (Table 1). However the smaller coupling constant observed for **23** ( $J_{1,2}=2.7$  Hz) does not match with a  ${}^4C_1$   $\beta$  configuration. For this compound gHMBC experiments (Figure 4) revealed two long-range correlations between the H1 proton ( $\delta$  6.57 ppm) and the C3 ( $\delta$  67.73 ppm) and C5 carbons ( $\delta$  64.76 ppm). These data allowed to propose for **23** a different conformational state ( ${}^1C_4$ ) and a  $\beta$  configuration.

Compound 23

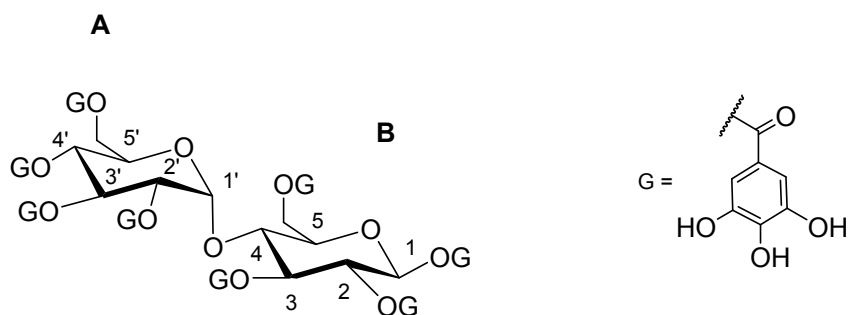


**Figure 4.** Structural assignments and HMBC correlation, observed for ribose derivative **23**

For the disaccharide compounds **25**, **27**, **29** and **31**, homonuclear COSY spectra were used in the identification of individual monosaccharide residues.

In the galloyl maltose derivative **25**, a coupling constant  $J_{1,2} = 3.9$  Hz allowed the unambiguous assignment of a  $\beta$  configuration for residue B (numbered 1-6), while a coupling constant  $J_{1,2} = 8.3$  Hz corroborates the  $\alpha$  configuration of residue A (numbered 1'-6'). The same was observed for the 2,3,4-trihydroxybenzoyl maltose derivative **29**.

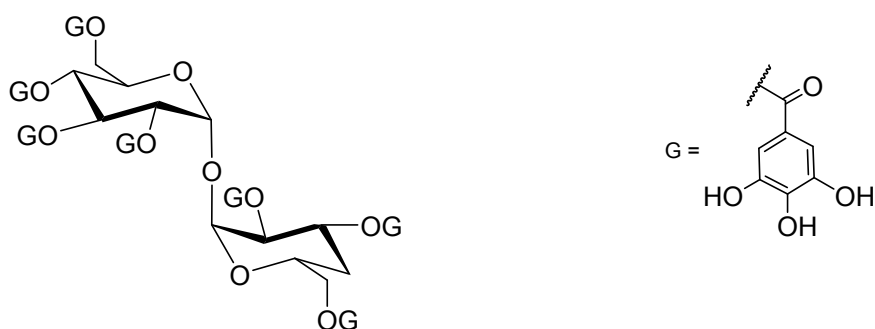
Compound **25**



**Figure 5.** Structure of compound **25** as a representative example of a maltose derivative

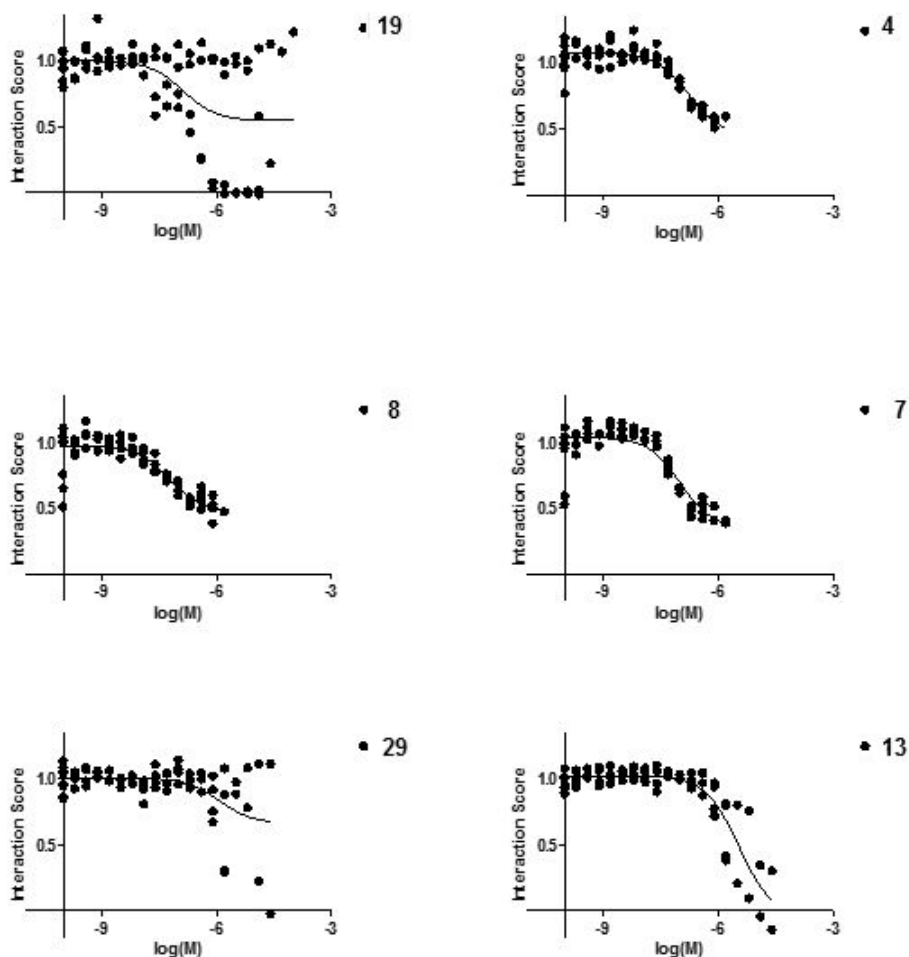
In trehalose derivatives, **27** and **31**, two coupling constants  $J_{1,2} = 3.7$  Hz allowed the unambiguous assignment of a  $\alpha,\alpha$  configuration for the two sugar residues.

Compound **27**



**Figure 6.** Structure of compound **27** as a representative example of a trehalose derivative

**2. Inhibitory effects of 4, 7, 8, 13, 19 and 29 with respect to PA-CMG2 inhibition. Dose-response curves determined by a FRET interaction assay**



**Figure 7.** Dose-response curves for synthesized compounds 4, 7, 8, 13, 19 and 29

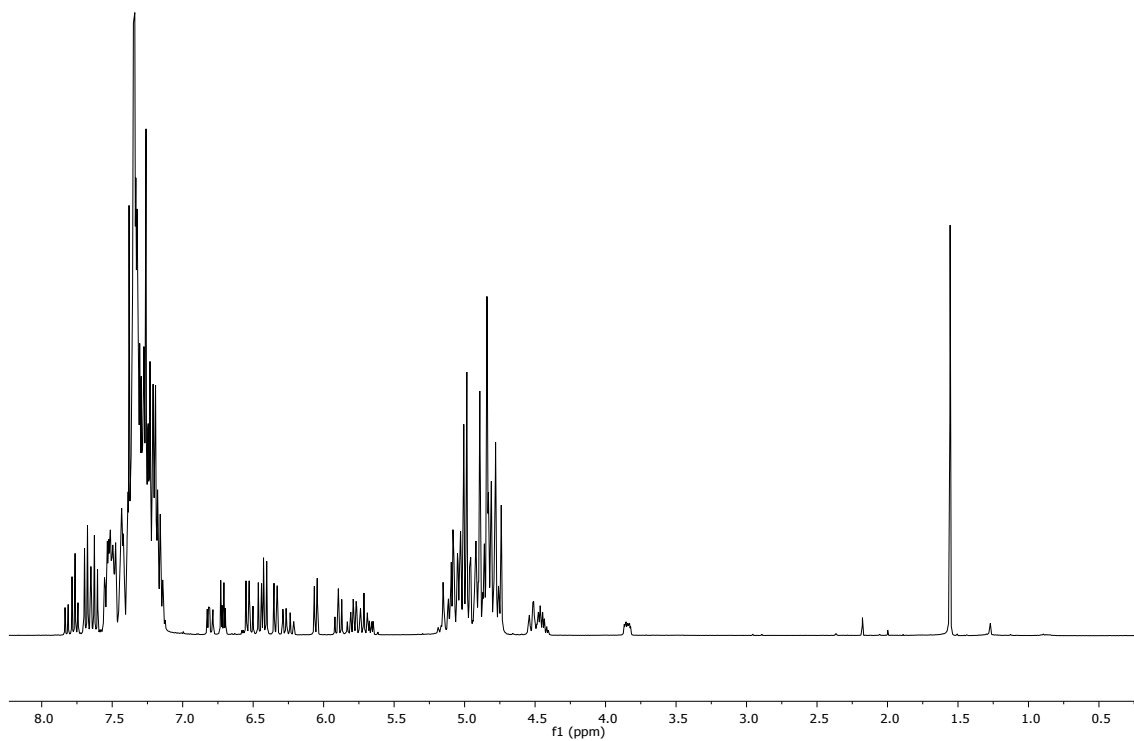
Interaction score represents the fraction of control interaction exhibited in a given well. Curves are best-fit binding isotherms with Hill coefficient of 1.



### 3. Selected $^1\text{H}$ and $^{13}\text{C}$ NMR spectra

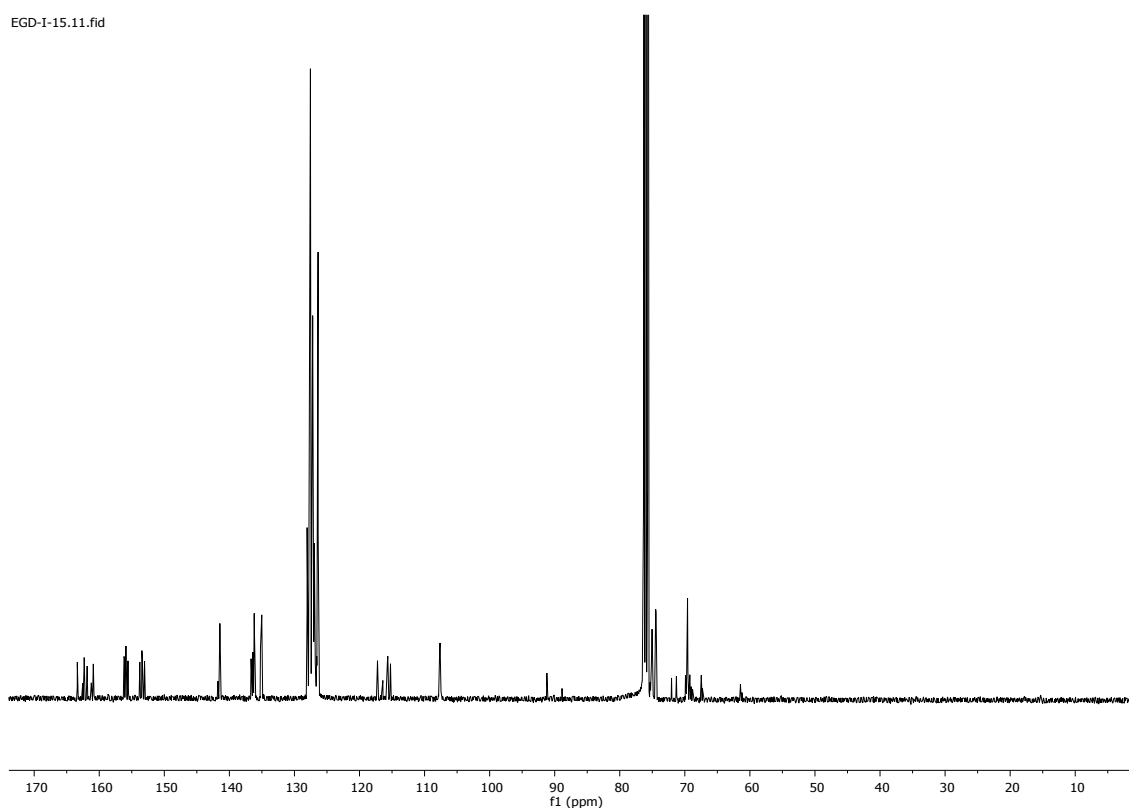
#### Compound 12 ( $^1\text{H}$ -NMR)

EGD-I-15.10.fid



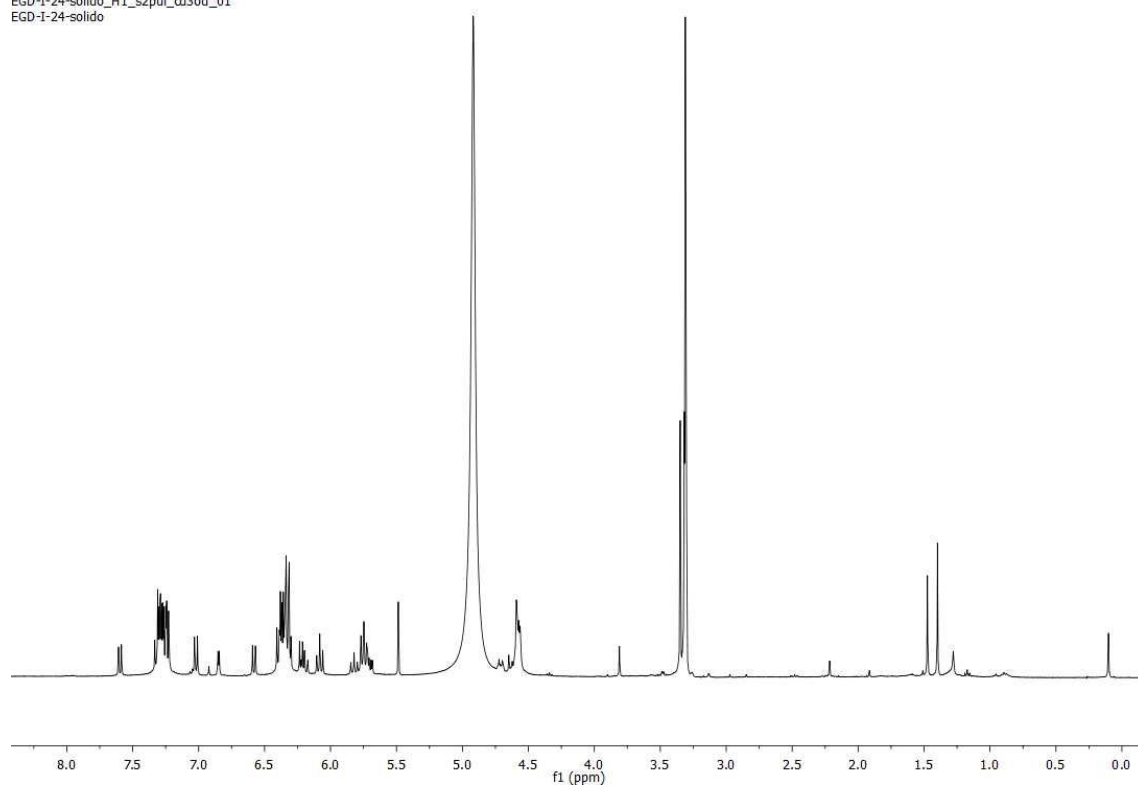
#### Compound 12 ( $^{13}\text{C}$ -NMR)

EGD-I-15.11.fid



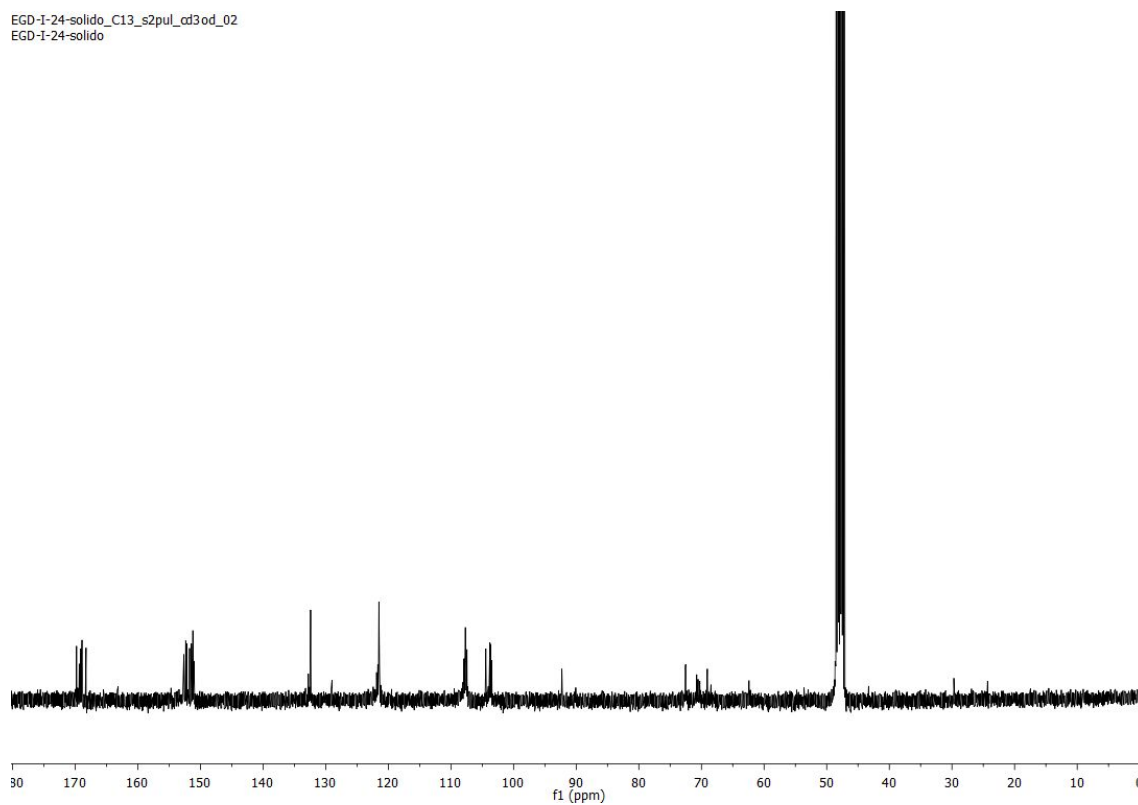
### Compound 13 ( $^1\text{H}$ -NMR)

EGD-I-24-solido\_H1\_s2pul\_cd3od\_01  
EGD-I-24-solido



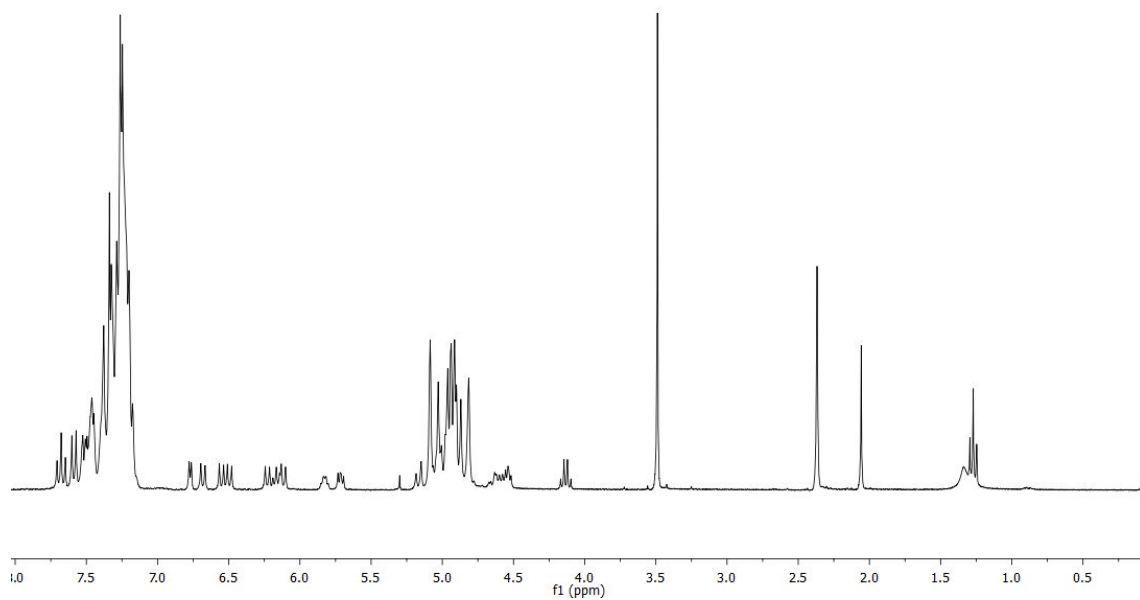
### Compound 13 ( $^{13}\text{C}$ -NMR)

EGD-I-24-solido\_C13\_s2pul\_cd3od\_02  
EGD-I-24-solido



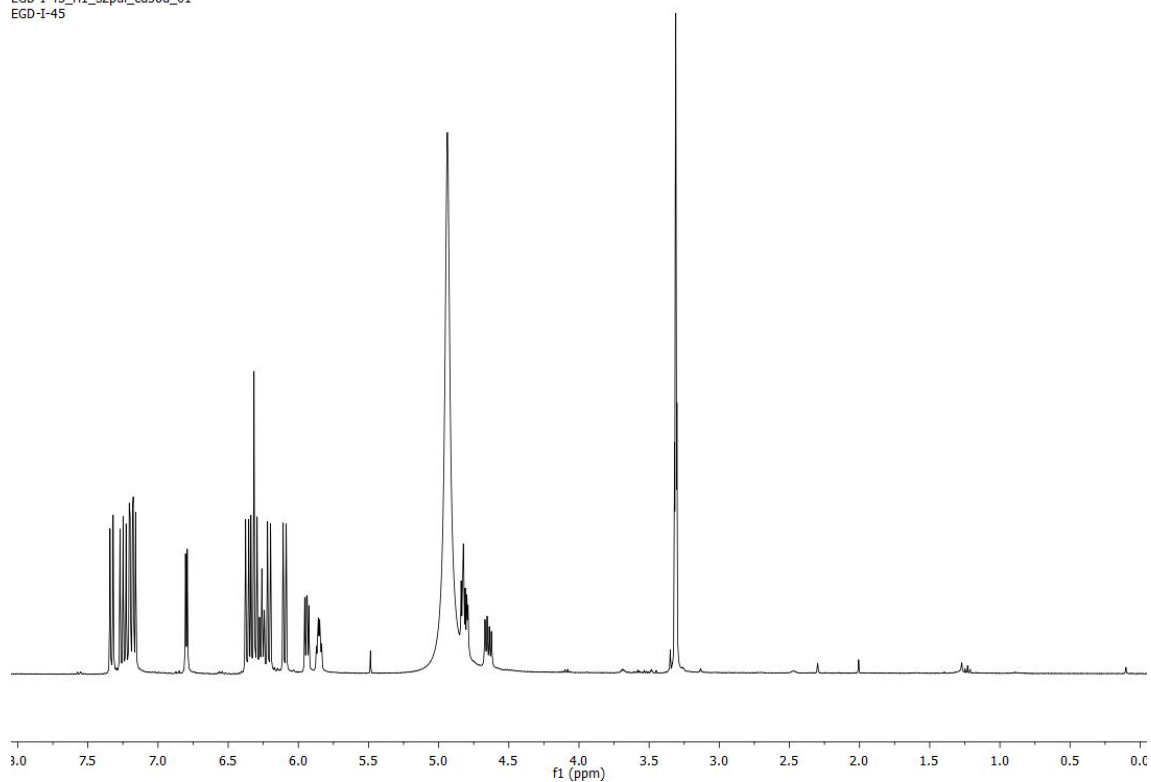
### Compound 14 ( $^1\text{H}$ - NMR)

EGD-I-18-F5-cdcd3  
EGD-I-18-F5-cdcd3



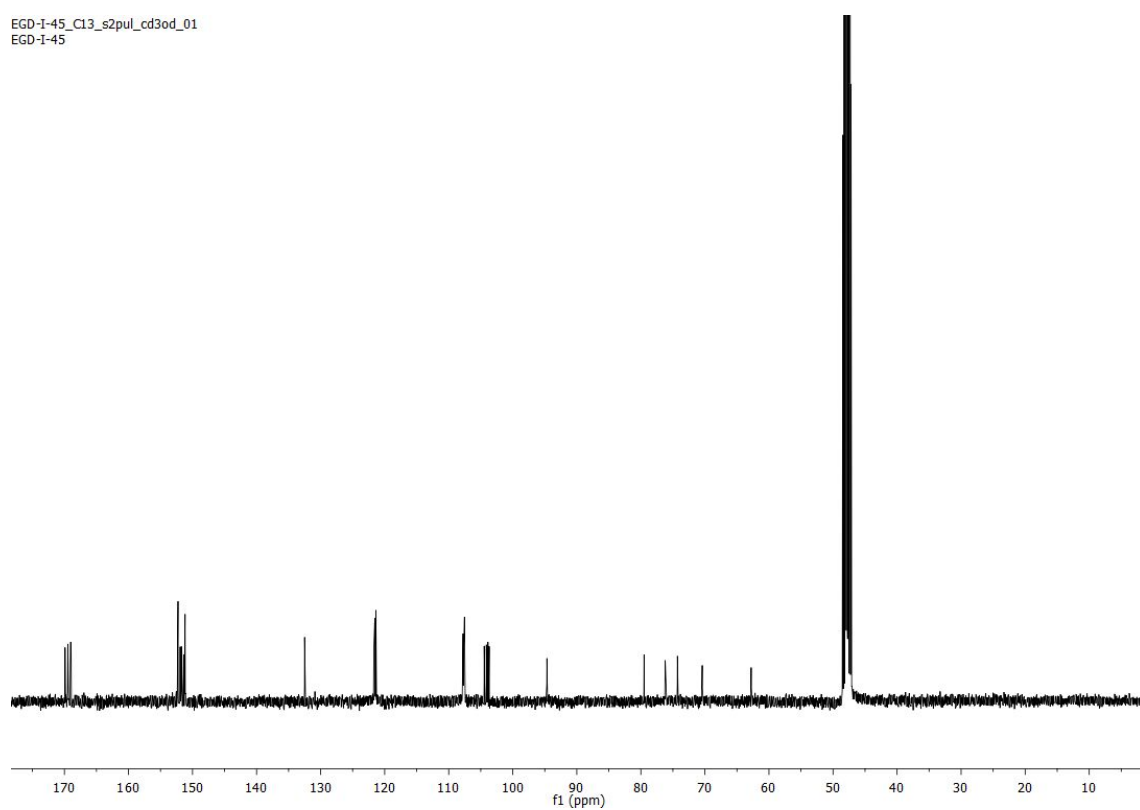
### Compound 15 ( $^1\text{H}$ - NMR)

EGD-I-45\_H1\_s2pul\_cd3od\_01  
EGD-I-45



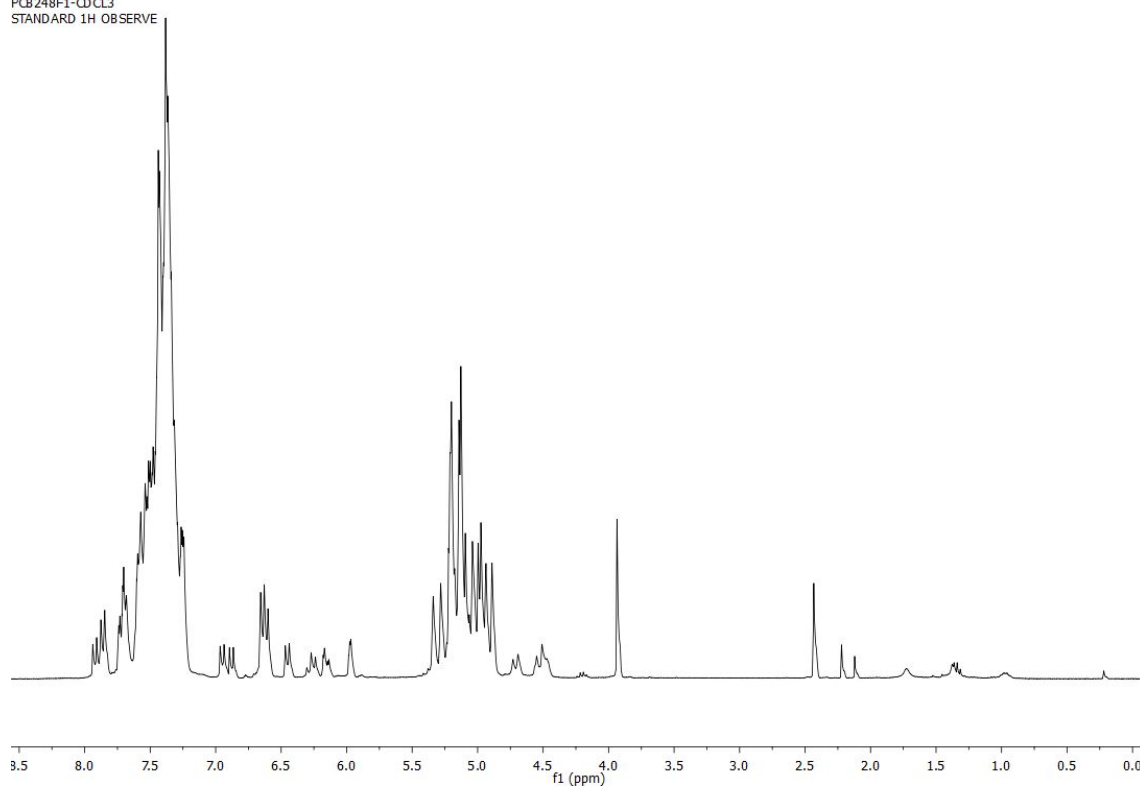
### Compound 15 ( $^{13}\text{C}$ - NMR)

EGD-I-45\_C13\_s2pul\_cd3od\_01  
EGD-I-45



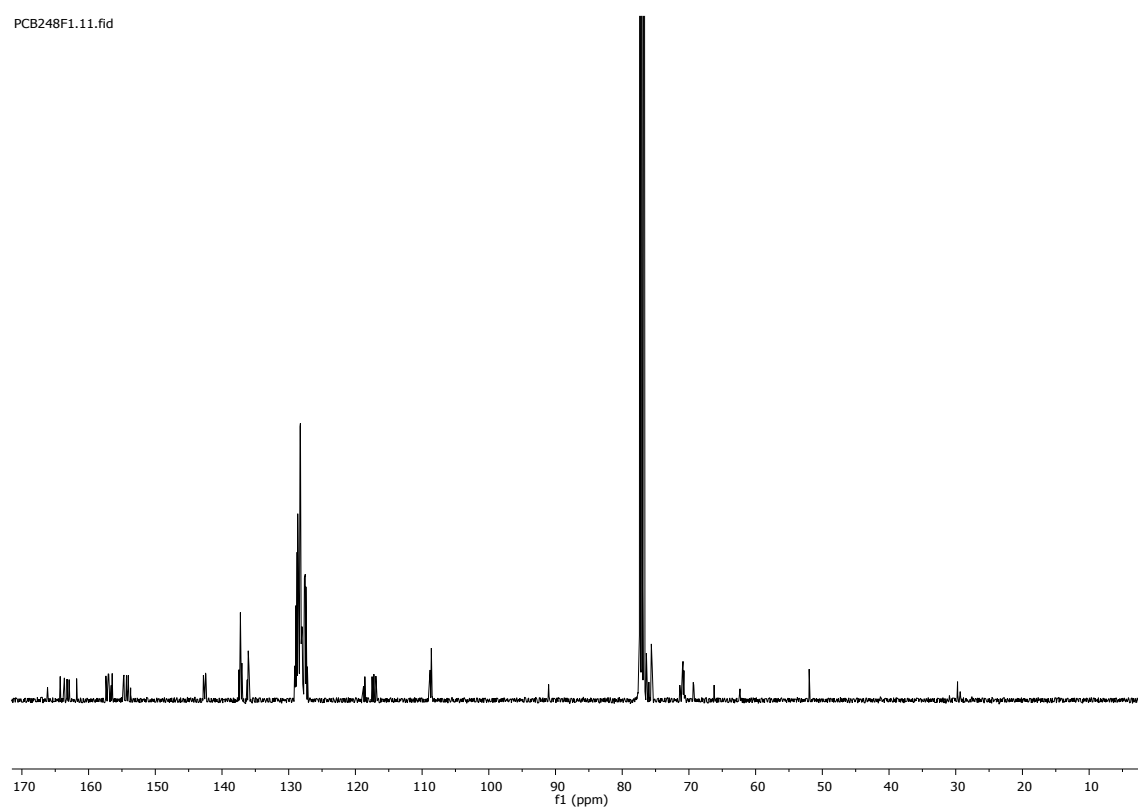
### Compound 16 ( $^1\text{H}$ - NMR)

PCB248F1-CDCL3  
STANDARD 1H OBSERVE



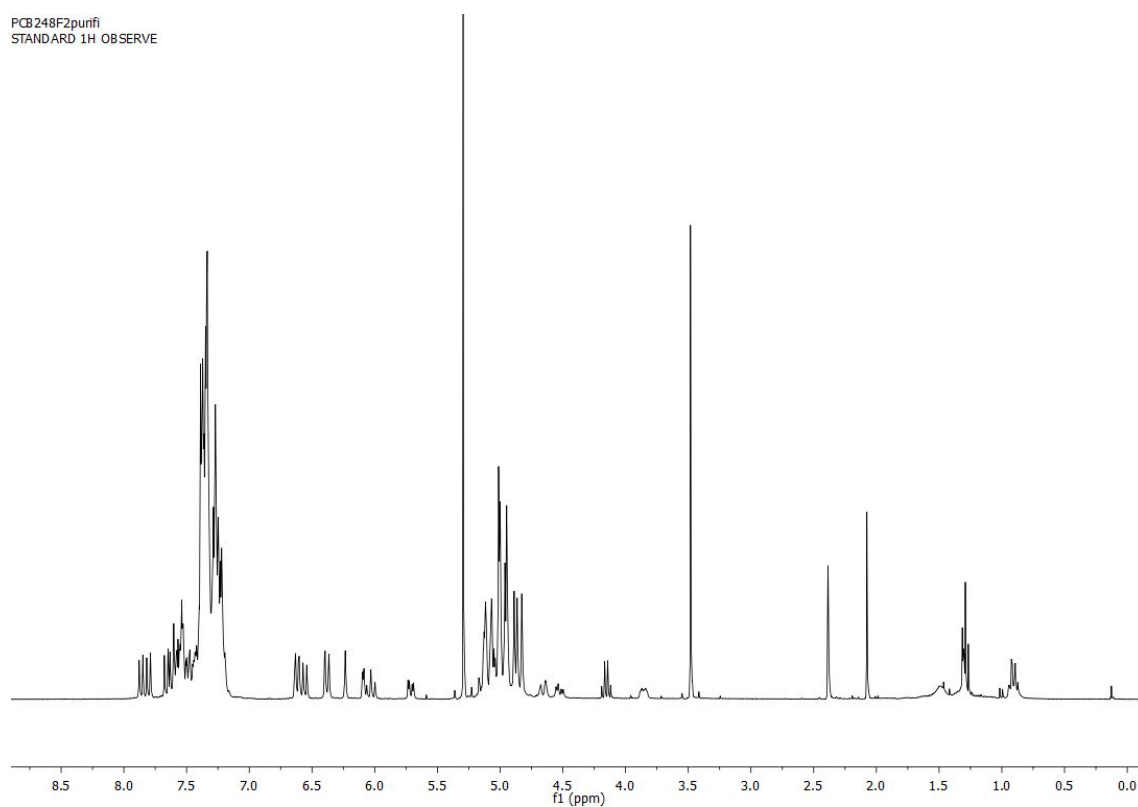
### Compound 16 ( $^{13}\text{C}$ - NMR)

PCB248F1.111.fid



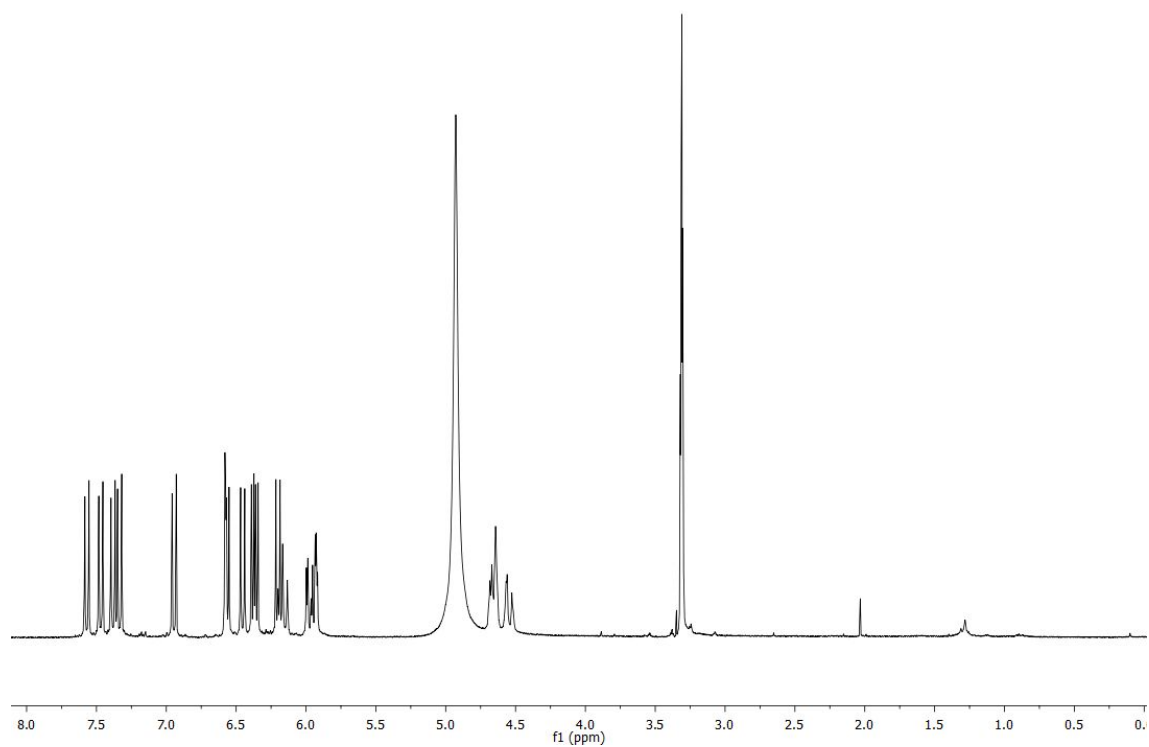
### Compound 17 ( $^1\text{H}$ - NMR)

PCB248F2purifi  
STANDARD 1H OBSERVE



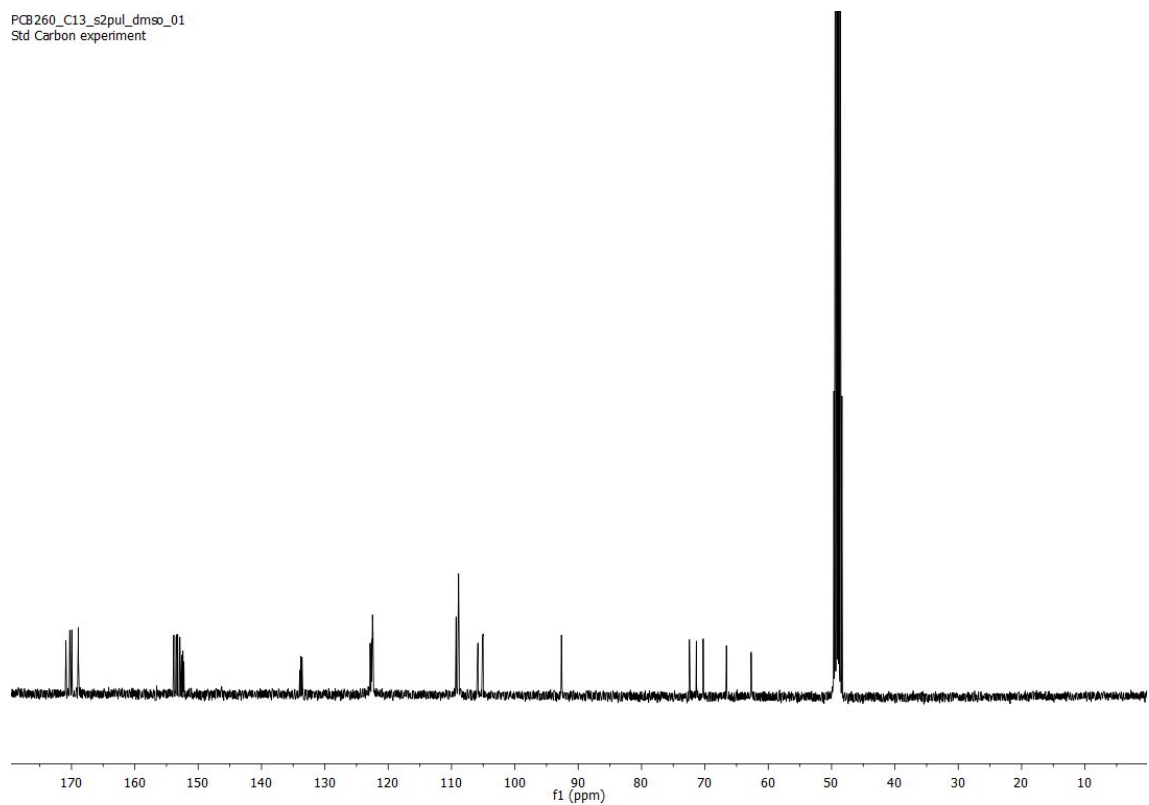
### Compound 18 ( $^1\text{H}$ - NMR)

PCB260t16-23  
STANDARD 1H OBSERVE



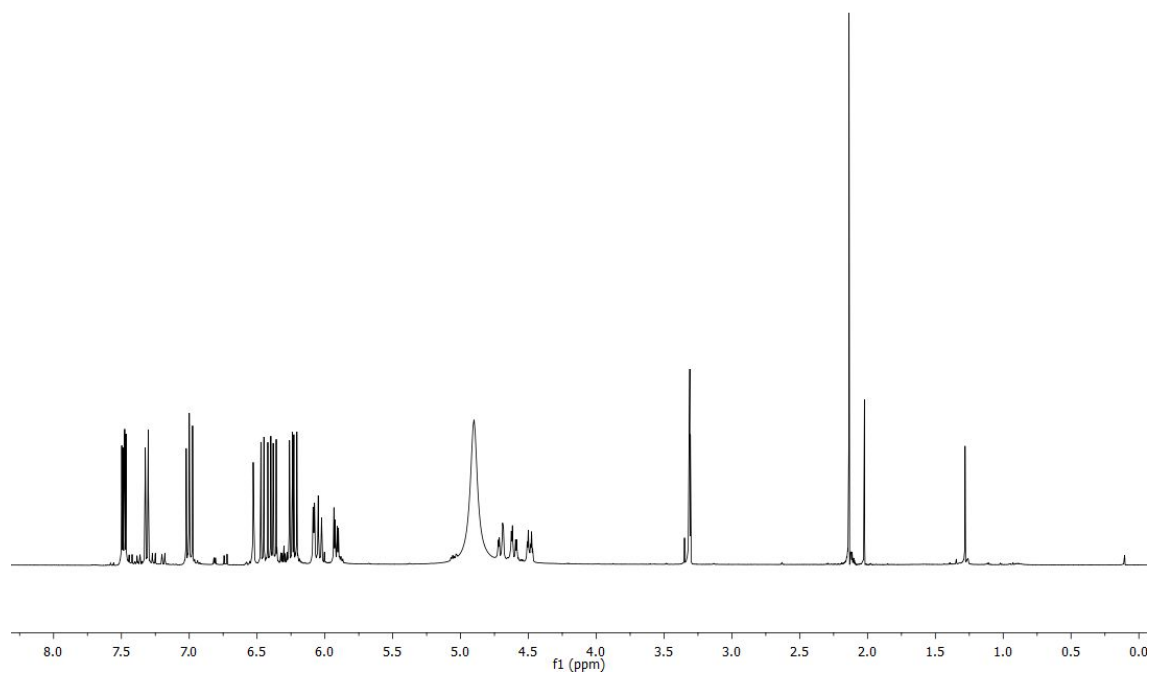
### Compound 18 ( $^{13}\text{C}$ - NMR)

PCB260\_C13\_s2pul\_dmsd\_01  
Std Carbon experiment



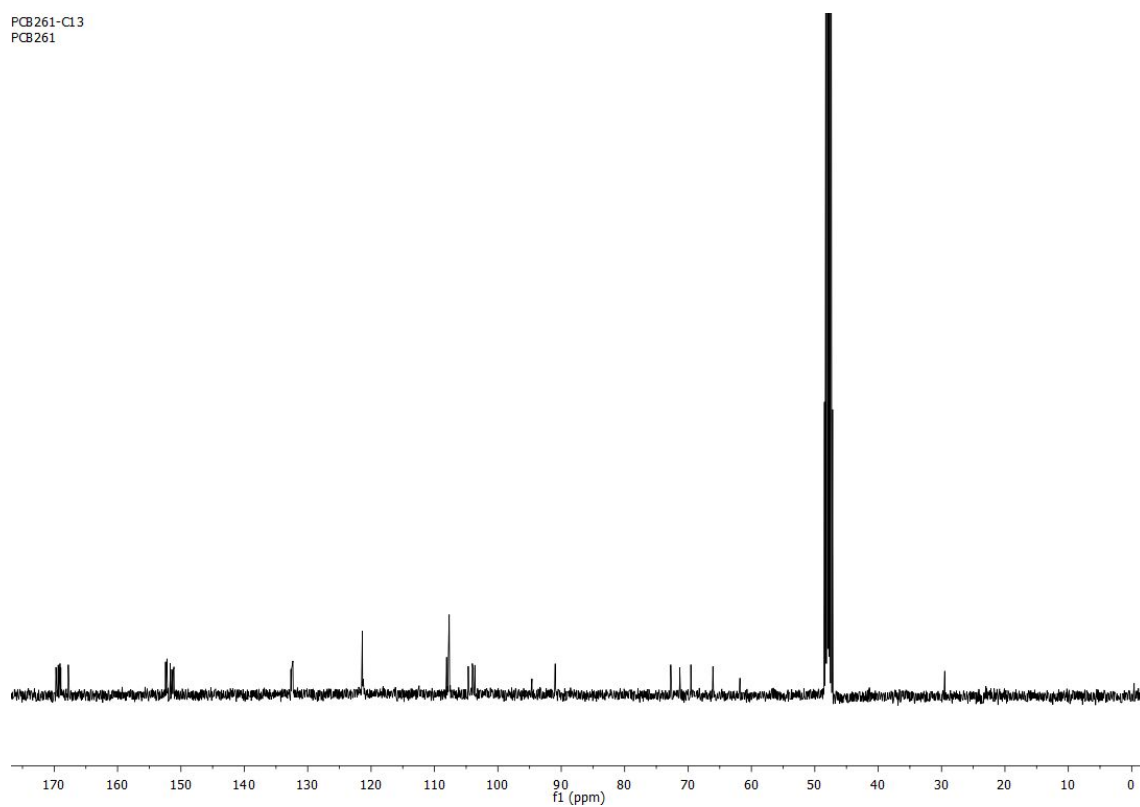
### Compound 19 ( $^1\text{H}$ - NMR)

PCB261-1H  
PCB261



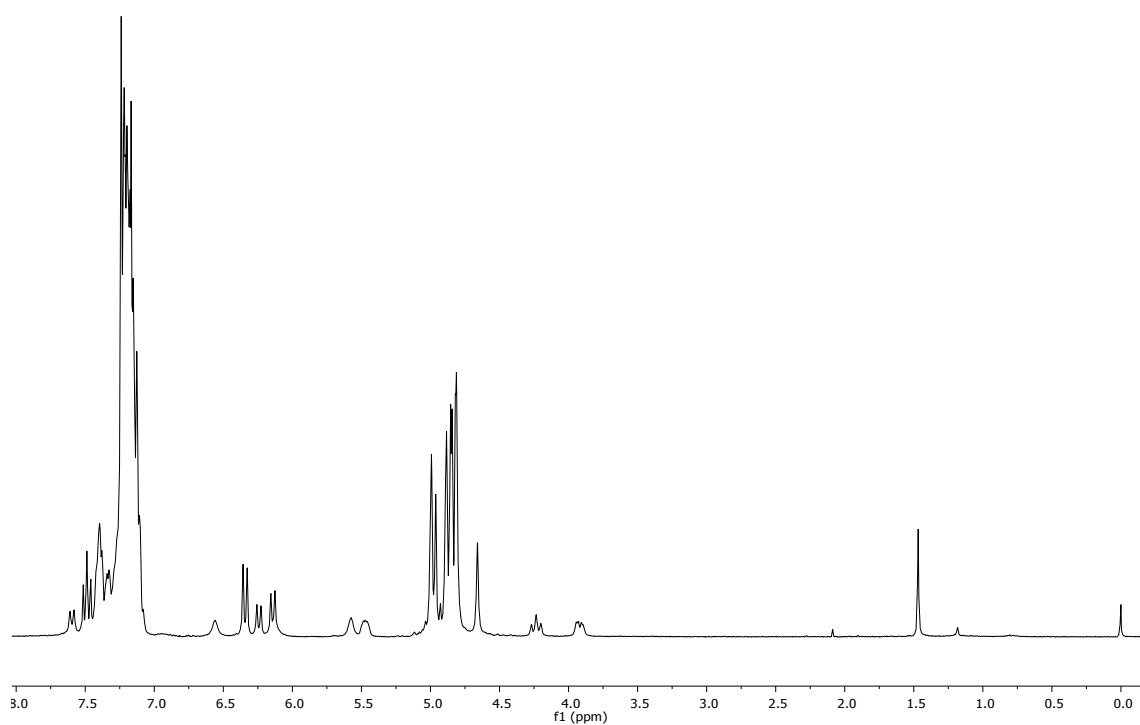
### Compound 19 ( $^{13}\text{C}$ - NMR)

PCB261-Cl3  
PCB261



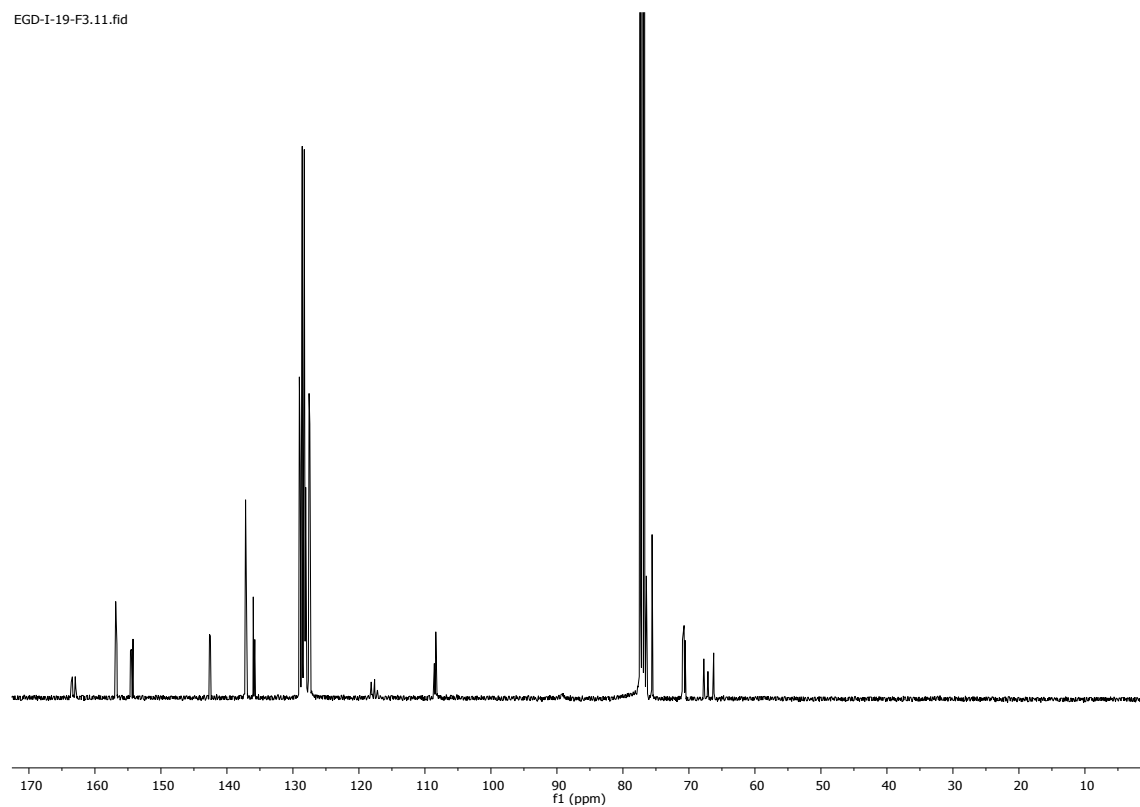
### Compound 20 ( $^1\text{H}$ - NMR)

EGD-I-19-F3  
EGD-I-19-F3-ribosa



### Compound 20 ( $^{13}\text{C}$ - NMR)

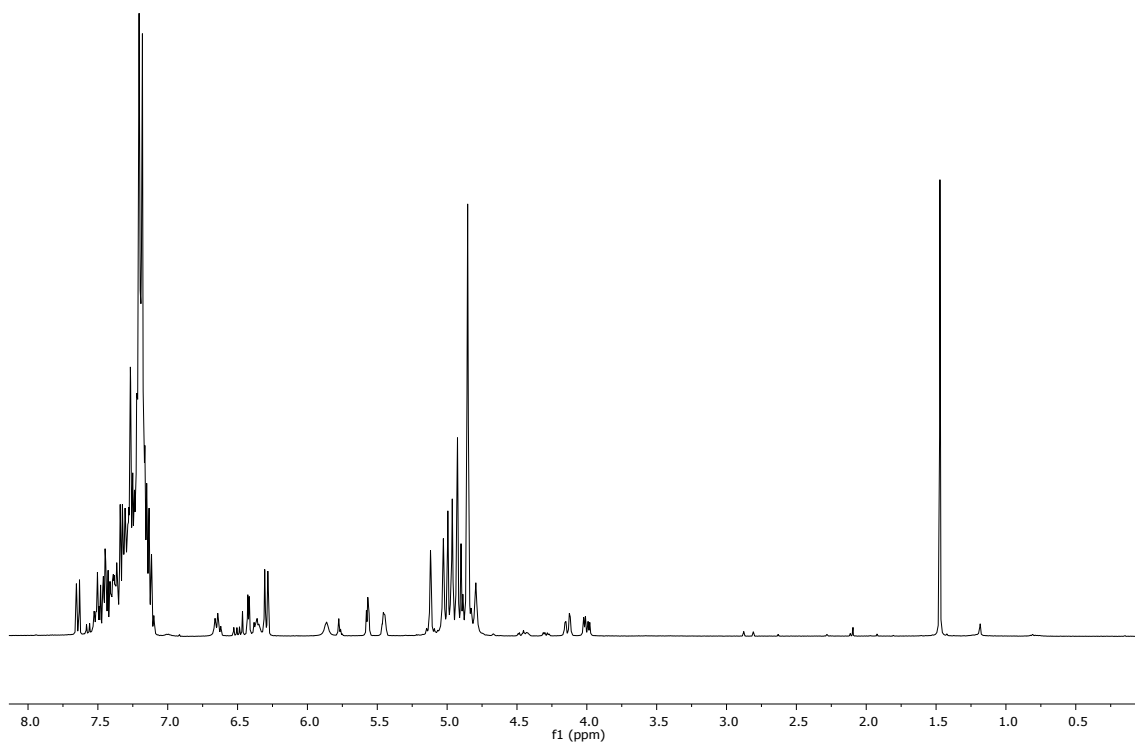
EGD-I-19-F3.11.fid





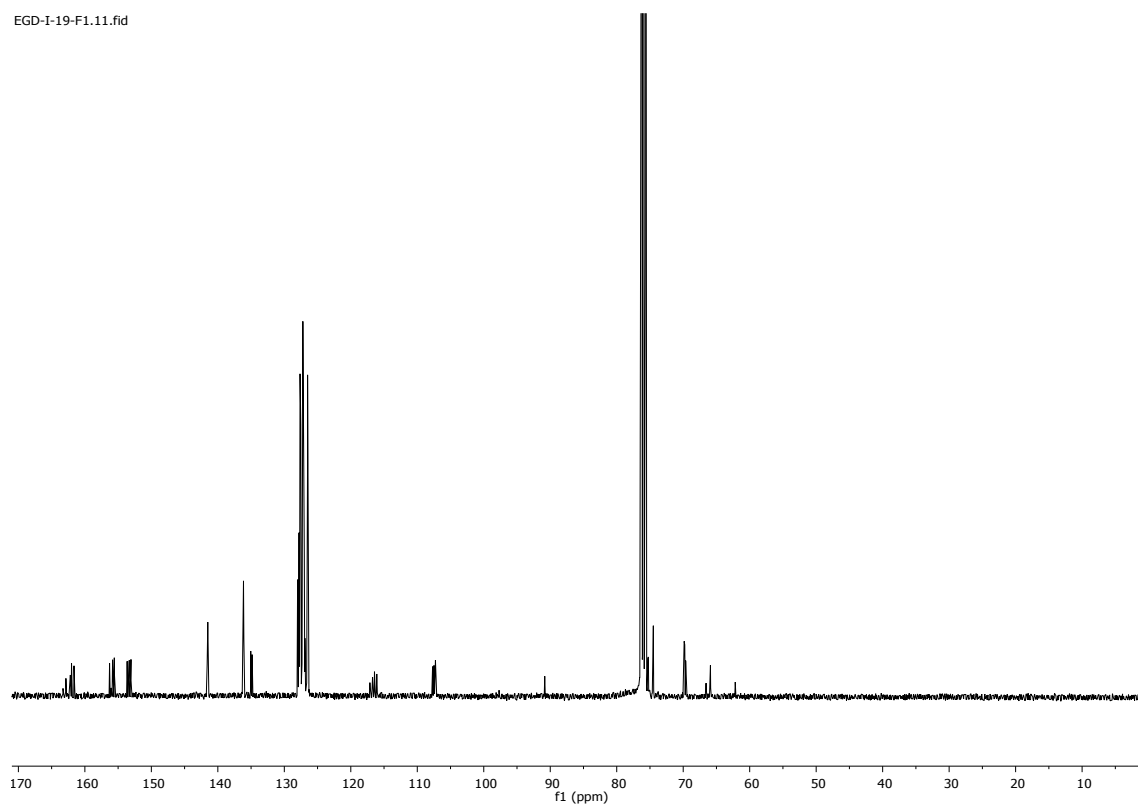
### Compound 21 ( $^1\text{H}$ -NMR)

EGD-I-19-F1.10.fid



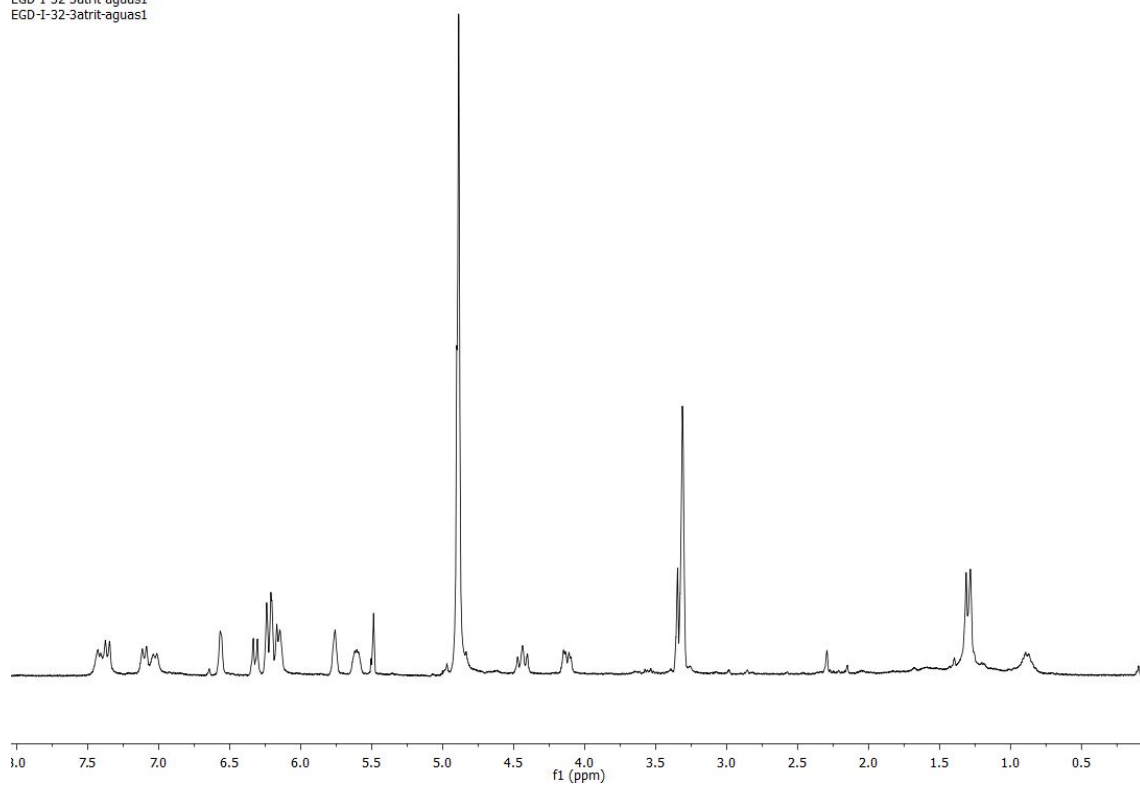
### Compound 21 ( $^{13}\text{C}$ -NMR)

EGD-I-19-F1.11.fid



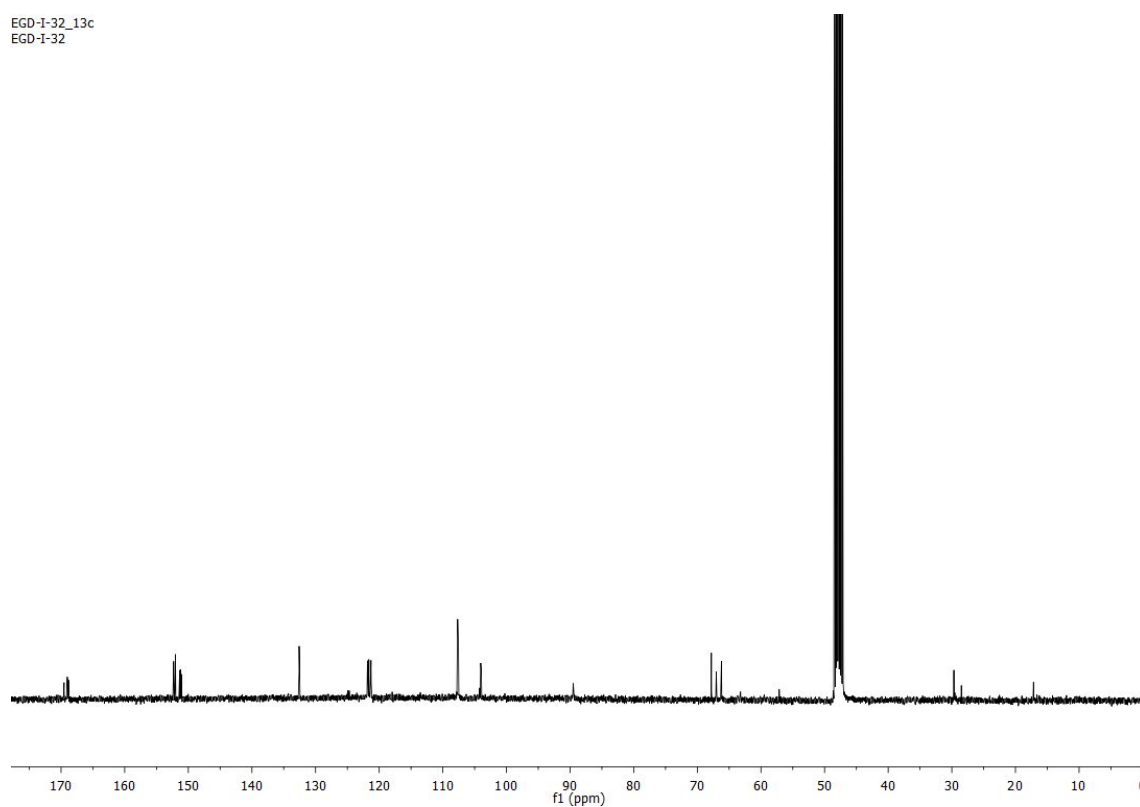
### Compound 22 ( $^1\text{H}$ - NMR)

EGD-I-32-3atrit-aguas1  
EGD-I-32-3atrit-aguas1



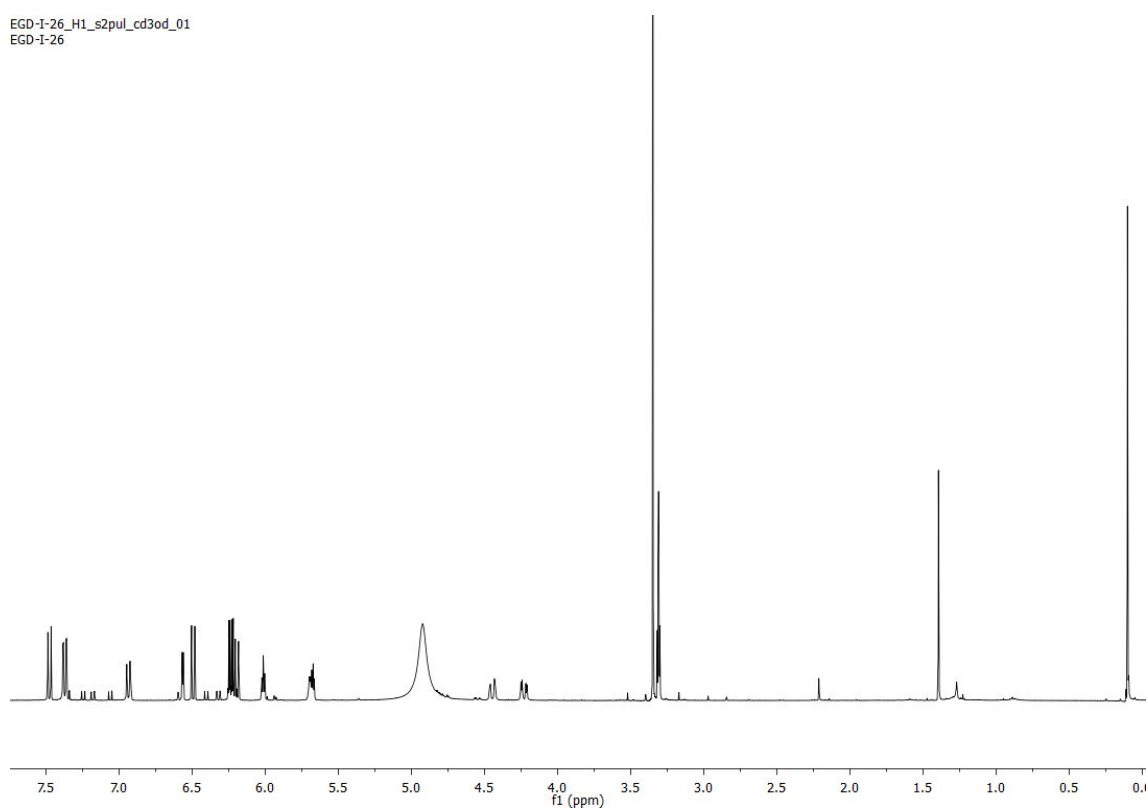
### Compound 22 ( $^{13}\text{C}$ - NMR)

EGD-I-32\_13c  
EGD-I-32



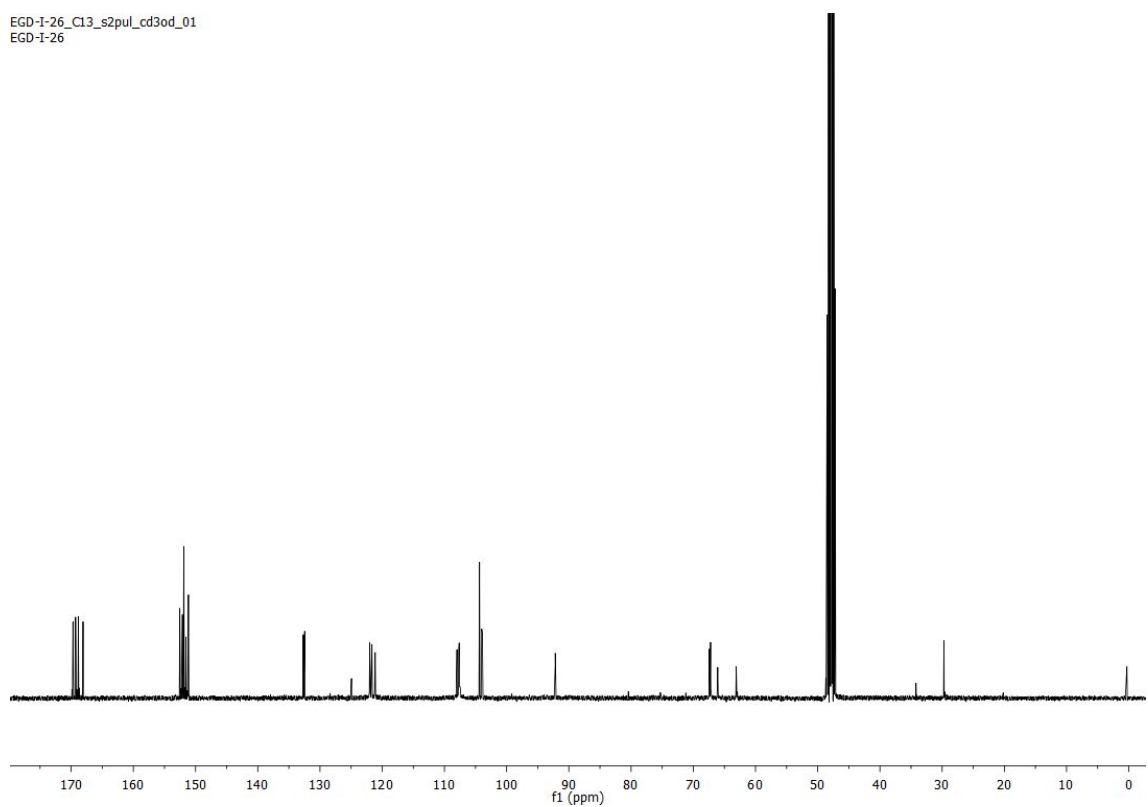
### Compound 23 ( $^1\text{H}$ - NMR)

EGD-I-26\_H1\_s2pul\_cd3od\_01  
EGD-I-26

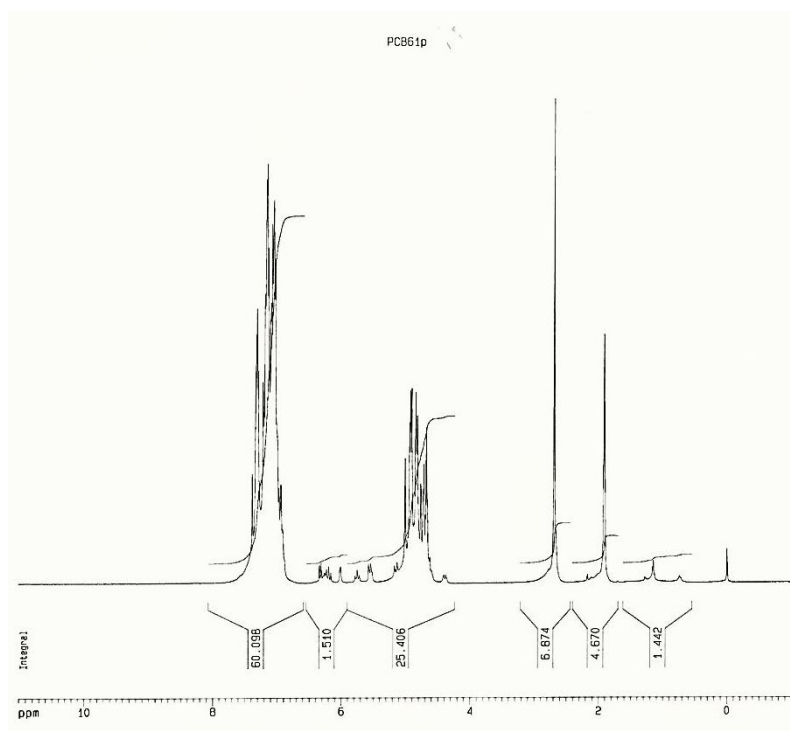


### Compound 23 ( $^{13}\text{C}$ - NMR)

EGD-I-26\_C13\_s2pul\_cd3od\_01  
EGD-I-26

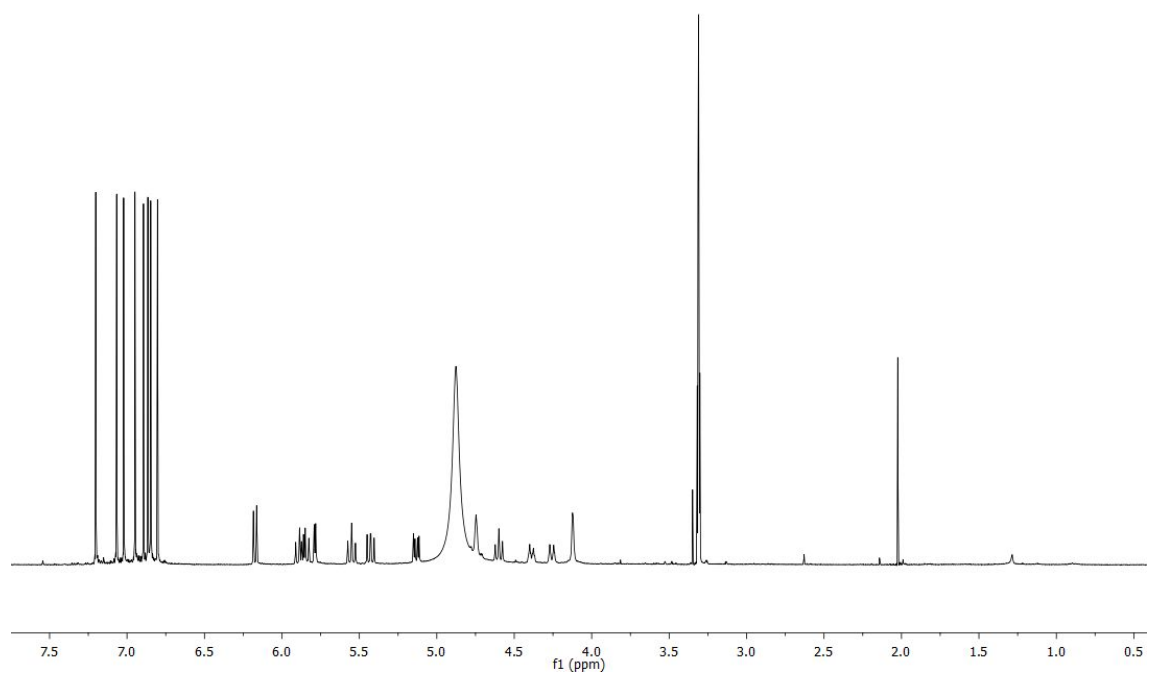


### Compound 24 ( $^1\text{H}$ - NMR)



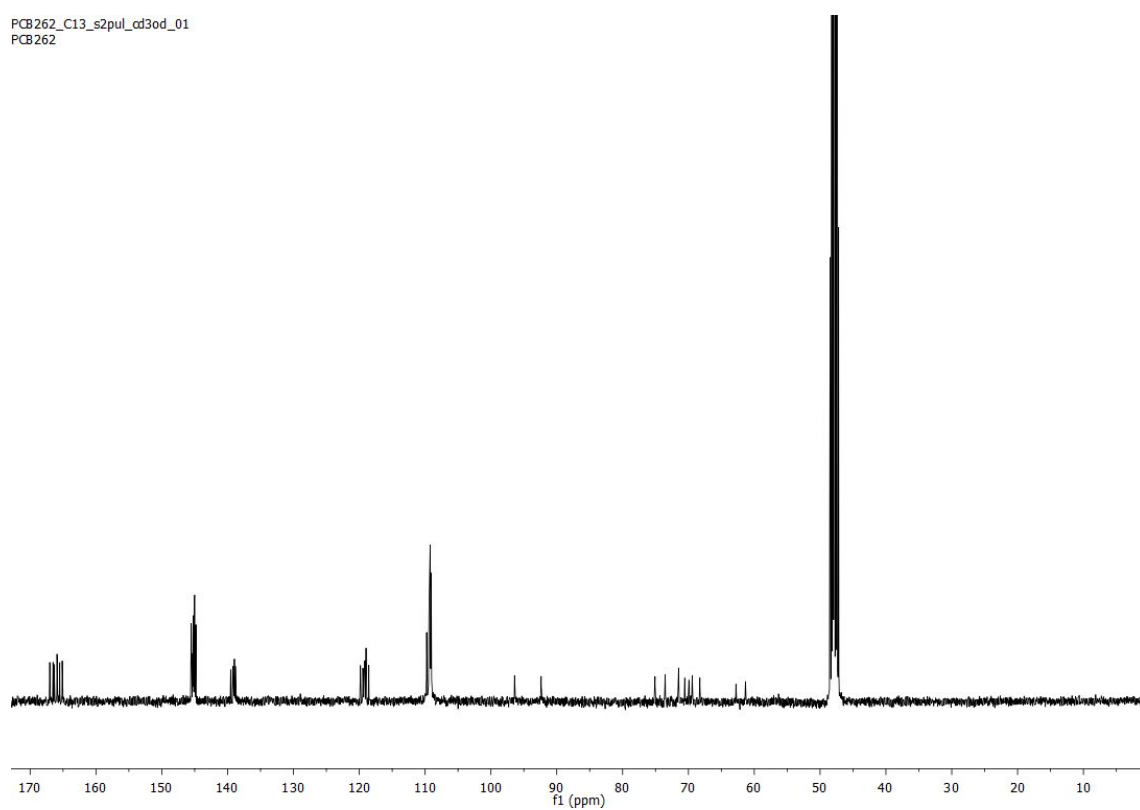
### Compound 25 ( $^1\text{H}$ - NMR)

PCB262\_H1\_s2pul\_cd3od\_01  
PCB262



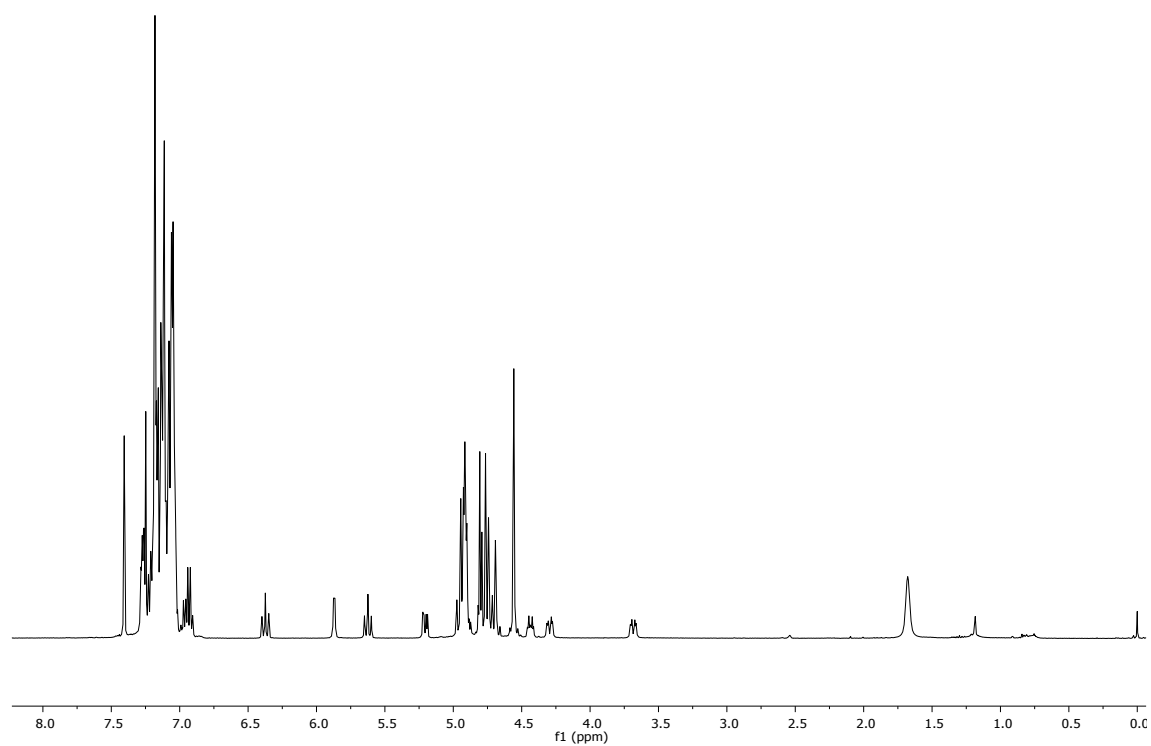
### Compound 25 ( $^{13}\text{C}$ - NMR)

PCB262\_C13\_s2pul\_cd3od\_01  
PCB262



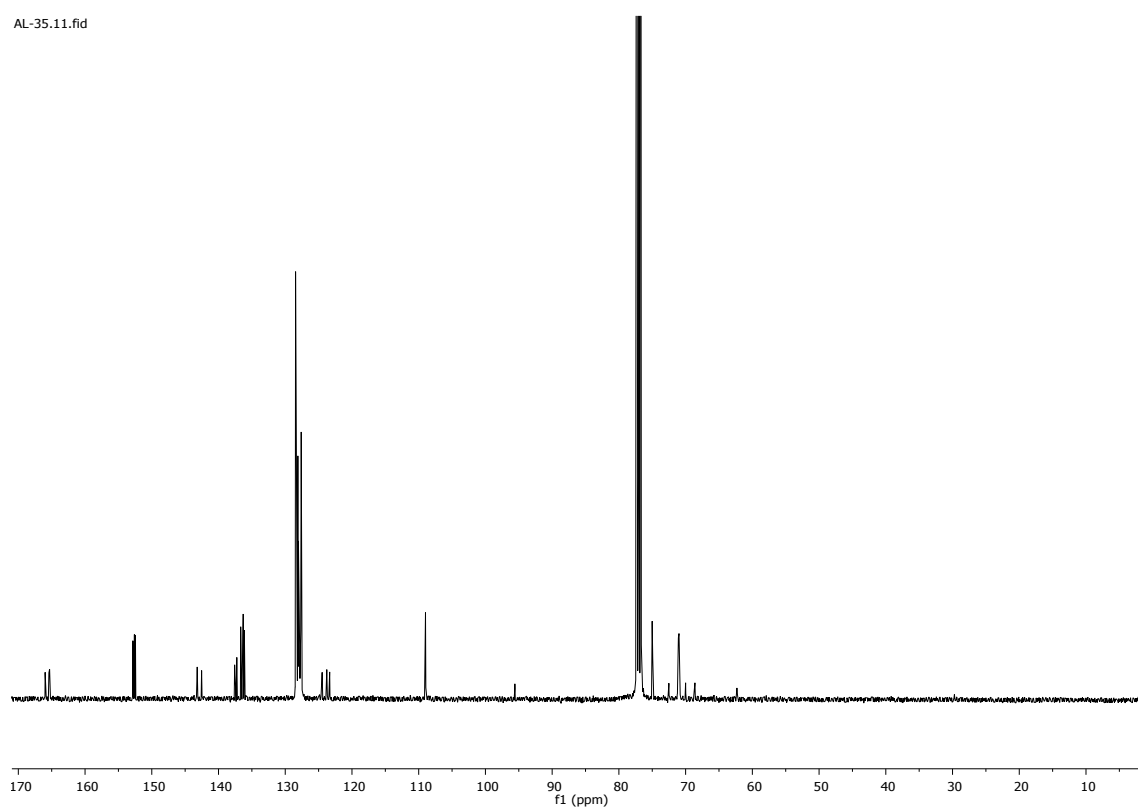
### Compound 26 ( $^1\text{H}$ - NMR)

AL-35.10.fid



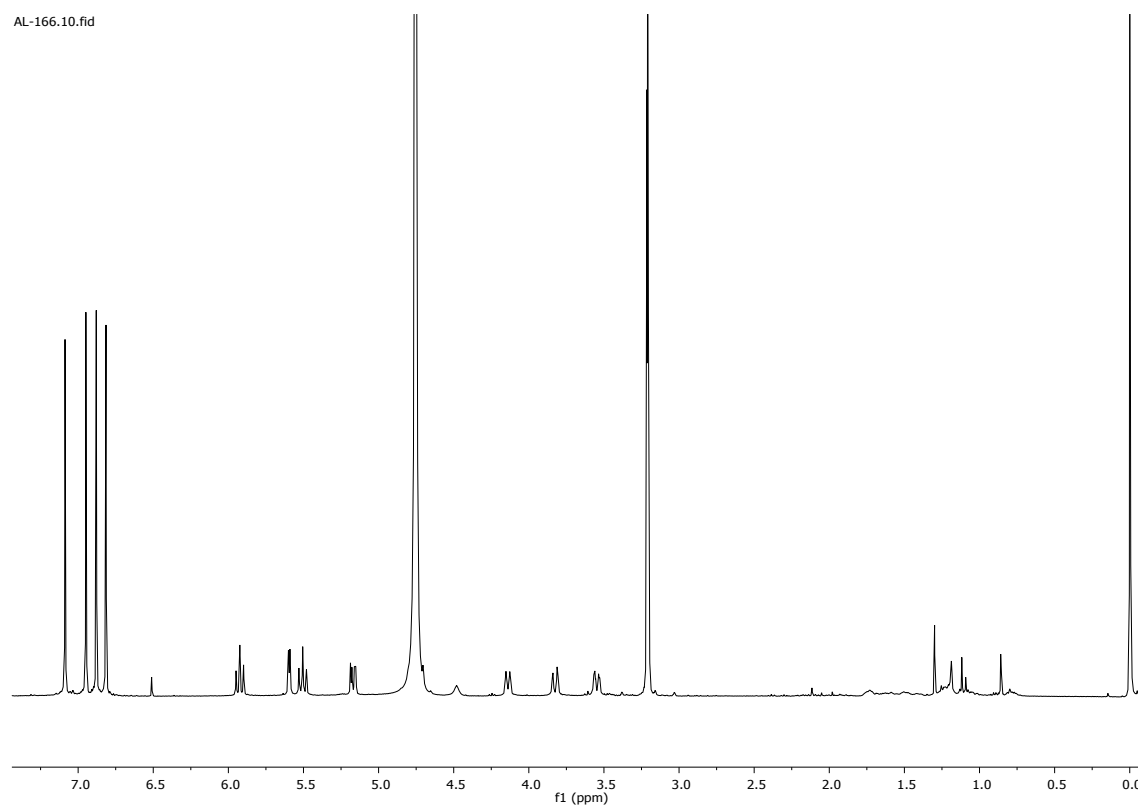
### Compound 26 ( $^{13}\text{C}$ - NMR)

AL-35.11.fid



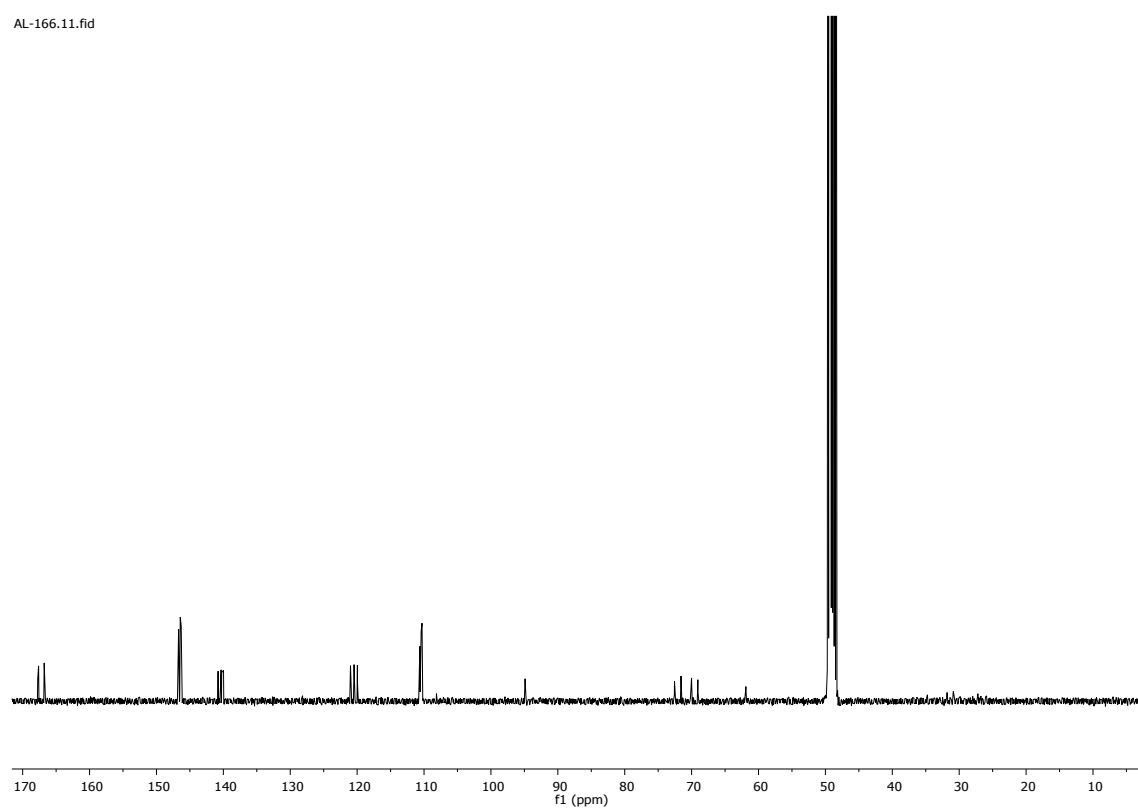
### Compound 27 ( $^1\text{H}$ - NMR)

AL-166.10.fid



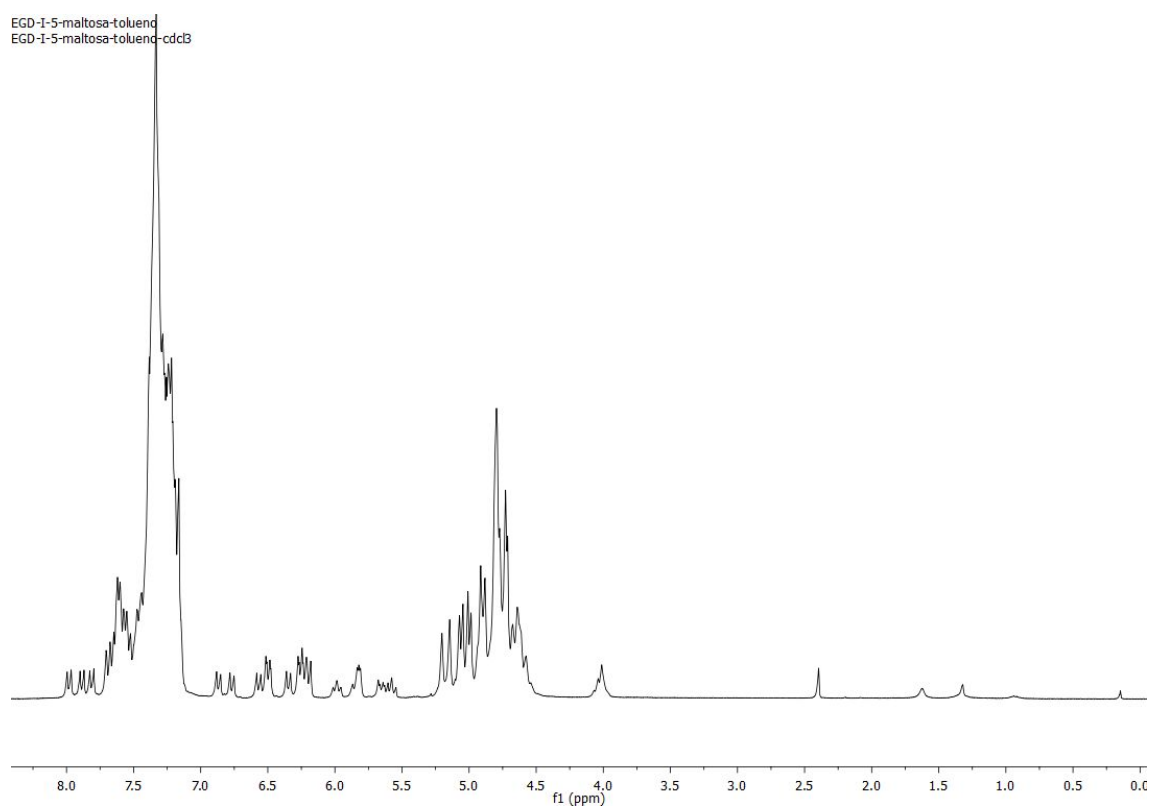
### Compound 27 ( $^{13}\text{C}$ - NMR)

AL-166.11.fid



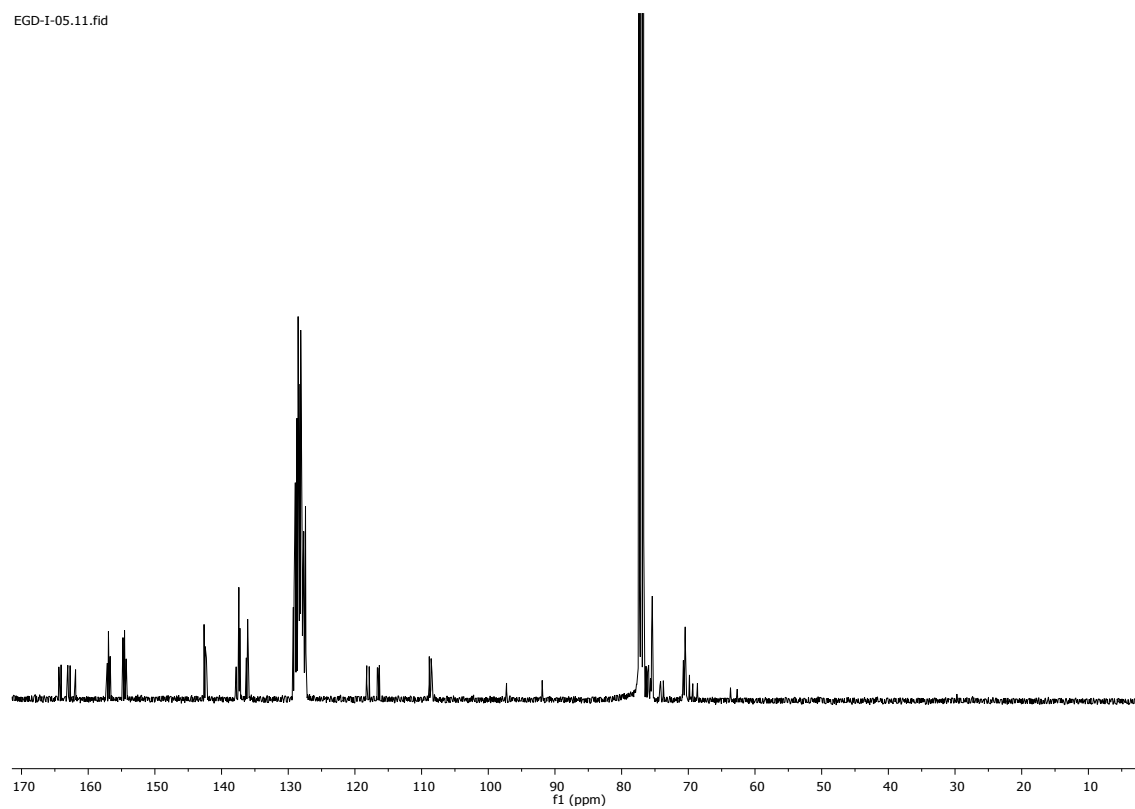
### Compound 28 ( $^1\text{H}$ - NMR)

EGD-I-5-maltosa-toluene  
EGD-I-5-maltosa-toluene-cdcl3



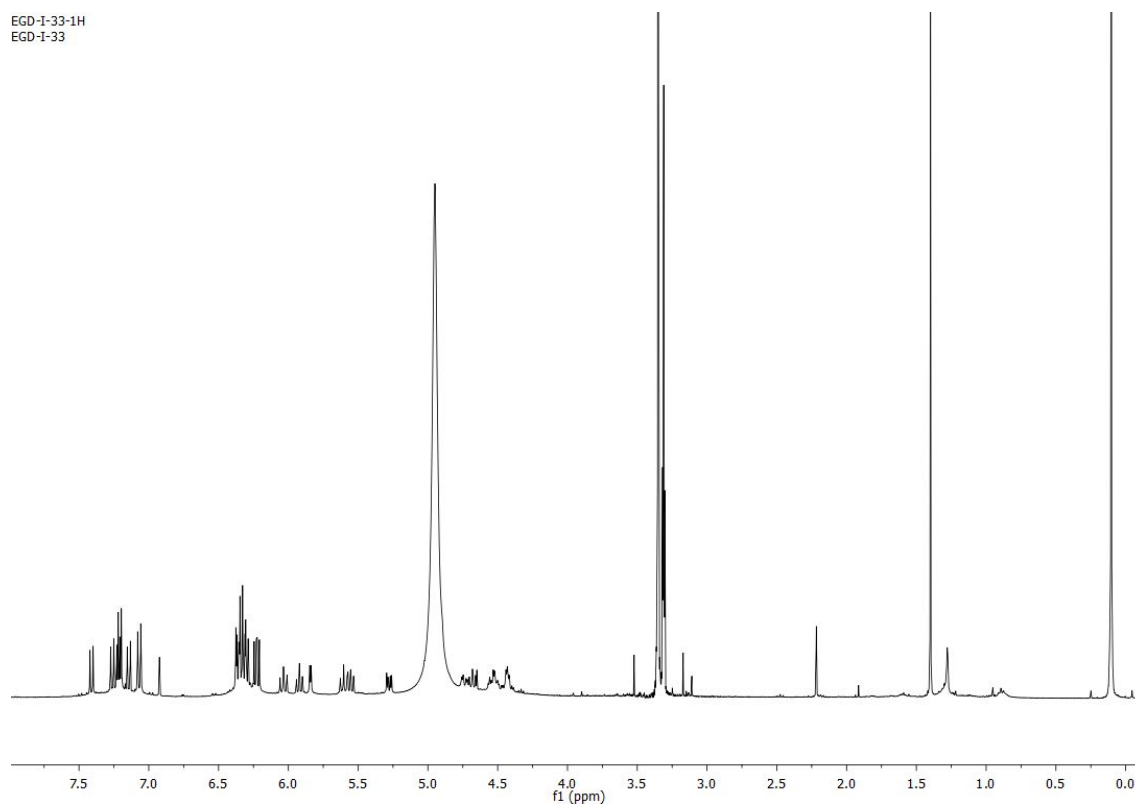
### Compound 28 ( $^{13}\text{C}$ -NMR)

EGD-I-05.11.fid



### Compound 29 ( $^1\text{H}$ -NMR)

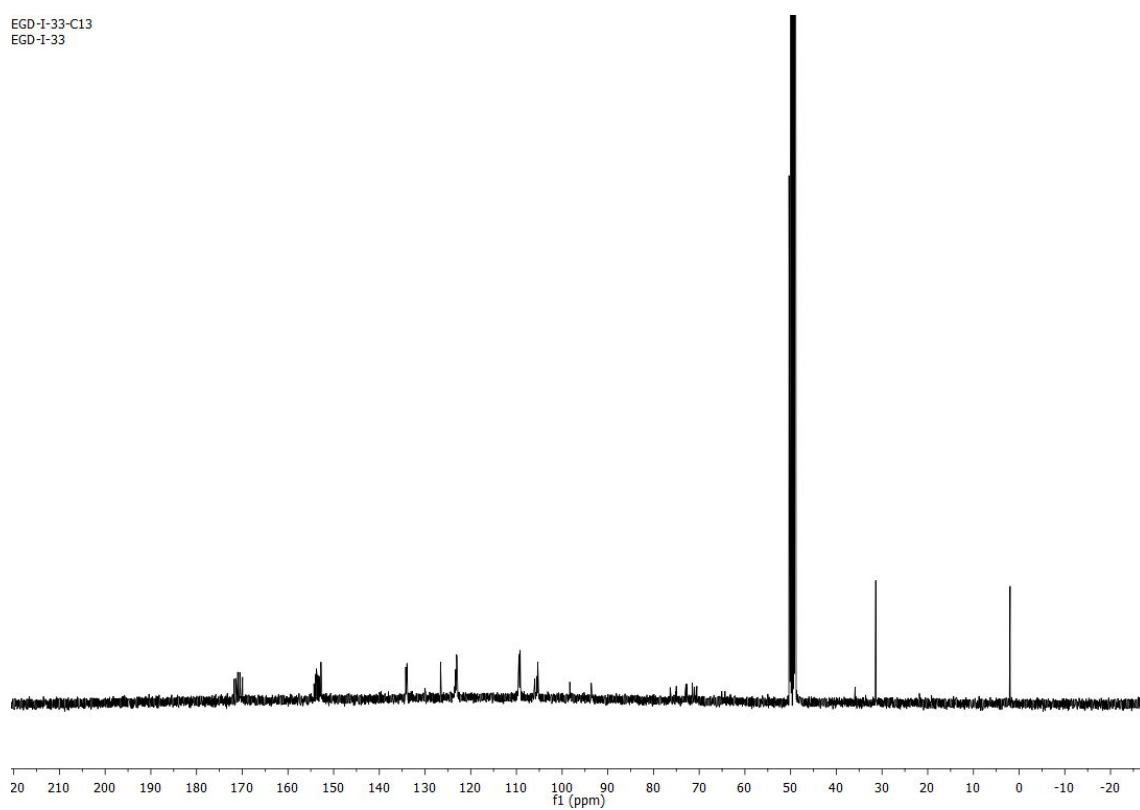
EGD-I-33-1H  
EGD-I-33





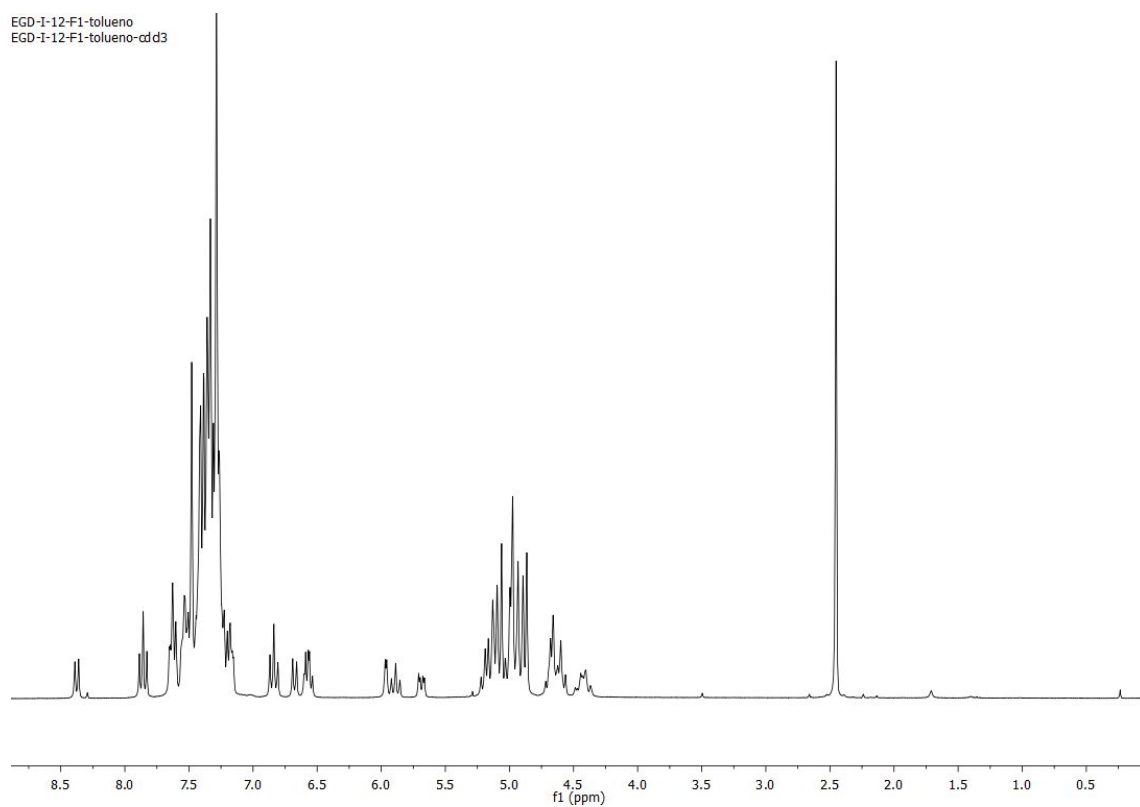
### Compound 29 ( $^{13}\text{C}$ -NMR)

EGD-I-33-C13  
EGD-I-33



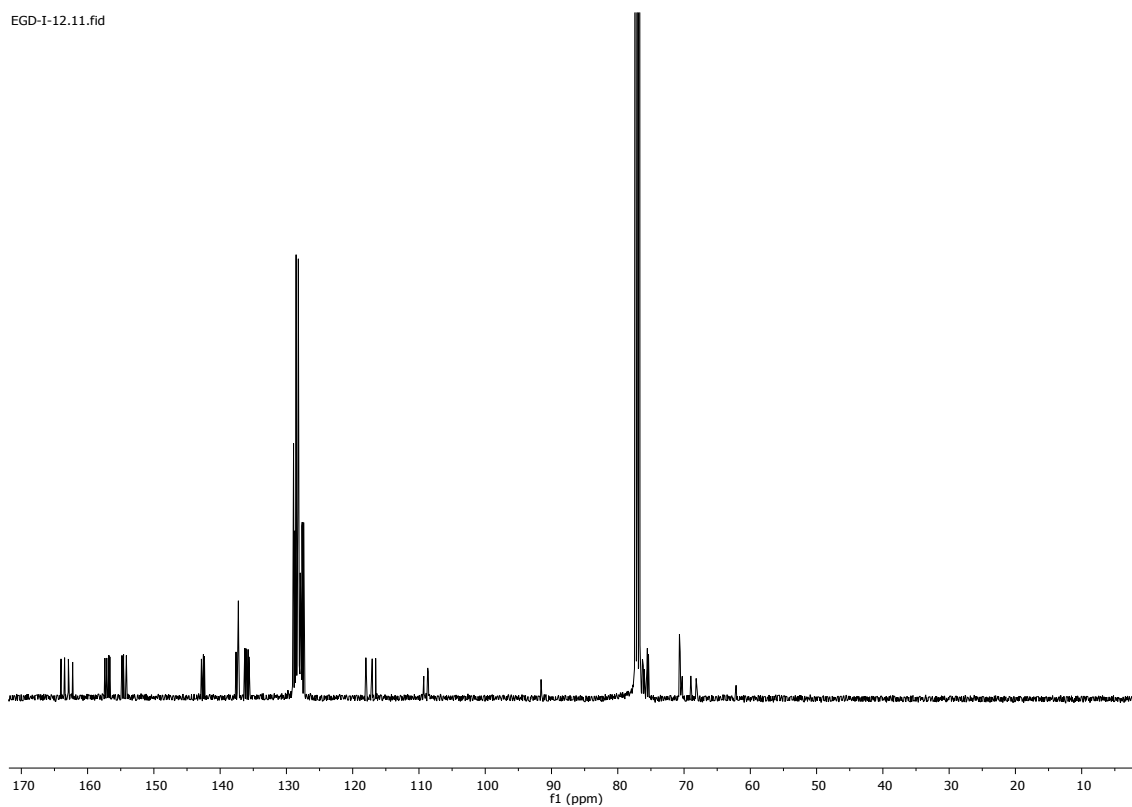
### Compound 30 ( $^1\text{H}$ -NMR)

EGD-I-12-F1-toluene  
EGD-I-12-F1-toluene- $\alpha$ -d<sub>3</sub>



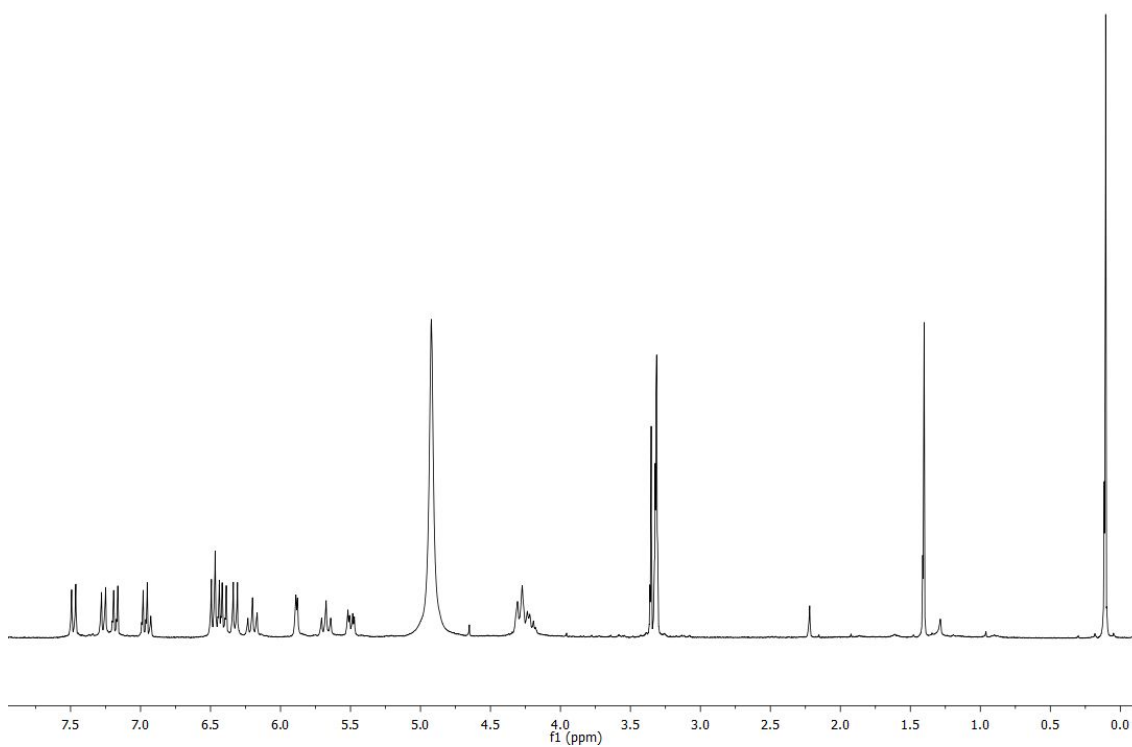
### Compound 30 ( $^{13}\text{C}$ -NMR)

EGD-I-12.11.fid



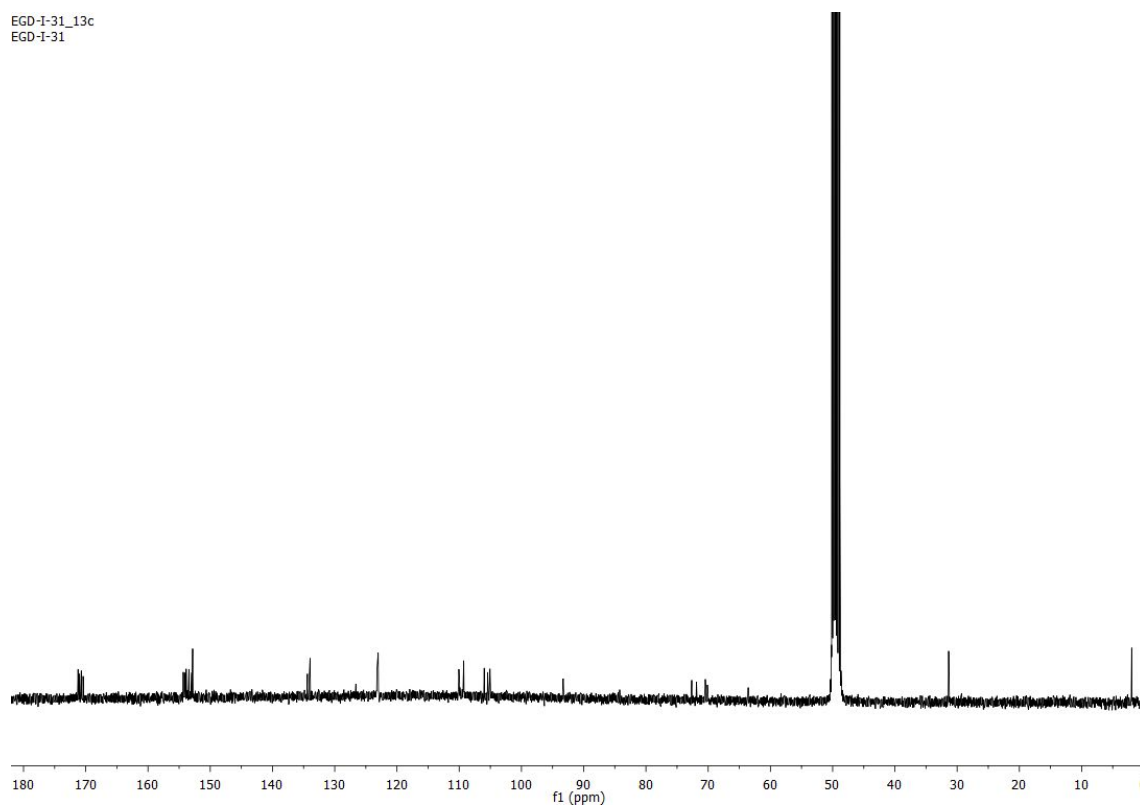
### Compound 31 ( $^1\text{H}$ -NMR)

EGD-I-31-trehalosa  
EGD-I-31-trehalosa-cd3od



## Compound 31 (<sup>13</sup>C-NMR)

EGD-I-31\_13c  
EGD-I-31



## References

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