Supplementary material

Metabolic activation of myristicin and its role in cellular toxicity

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1 Chemical syntheses of 1'-hydroxymyristicin

The synthesis of 1'-hydroxymyristicin was completed by Professor Hong-bo Qin's 2 laboratory. 1'-Hydroxymyristicin was synthesized using nucleophilic addition of 3 5-Methoxypiperonal. To a solution of 5-methoxypiperonal (101 mg, 0.56 mmol) in 4 anhydrous tetrahydrofuran (2 mL) under N2 was supplemented Vinylmagnesium 5 6 bromide (0.56 mL, 1 mol/L, 0.56 mmol) dropwise at 0 °C. After being stirred for 1h at 25 °C, the mixture was terminated by saturated aqueous NH₄Cl and extracted with 7 ethyl acetate (EtOAc) three times. Combined organic phase was washed sequentially 8 9 with saturated aqueous sodium carbonate solution, water and brine, and dried over 10 Na₂SO₄. Filtration and concentration afforded the crude product. Purification by column chromatography using EtOAc/Petroleum ether (1:10) as mobile phase gave 11 alcohol product (96 mg, 82%) as a colorless oil. The purity of 1'-hydroxymyristicin 12 was > 98% measured by high-performance liquid chromatography. NMR spectra 13 data were operated on 600 MHz for ¹H-NMR spectrum and 150 MHz for ¹³C-NMR 14 15 spectral. Deuterochloroform (CDCl₃) was used as solvents for NMR detection. The structural identification of 1'-hydroxymyristicin was characterized by ¹H- and 16 ¹³C-NMR spectral (Supplementary Fig. 1A and 1B). ¹H-NMR (CDCl₃, 600 MHz): δ 17 18 3.88 (3H, s, OCH₃), 5.93 (2H, s, -OCH₂O-), 6.55 (1H, s, 2-H), 6.53 (1H, s, 4-H), 5.32 (1H, d, H1'), 5.97 (1H, d, H2'), 5.16/5.05 (2H, d, CH₂=) (Supplementary 19 Fig.1A). ¹³C-NMR (CDCl₃, 150MHz): δ 143.53 (C-1), 134.53 (C-2), 148.83 (C-3), 20 101.42 (C-4), 137.52 (C-5), 100.59 (C-6), 75.08 (C-1'), 140.13 (C-2'), 114.52 (C-3'), 21 56.50 (-OCH₃), 105.70 (-OCH₂O-) (Supplementary Fig.1B). HR-ESI-MS: m/z 22

23 231.0633 $[M+Na]^+$ (calculated for $C_{11}H_{12}O_4$).

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25 Sample preparation method of urine, feces and blood

For urine samples preparation, 180 µL of 50% aqueous acetonitrile containing 5 µM 26 of chlorpropamide (used as internal standard) was added into 20 µL of urine sample, 27 28 and vortexed for 1 min. Then the samples were centrifuged at 18000 g for 20 min at 4 °C to remove proteins and particulates. For plasma samples preparation, 190 µL of 29 50% aqueous acetonitrile containing 5 µM of chlorpropamide (used as internal 30 standard) was added into 10 µL of plasma sample, and vortexed for 1 min. Then the 31 samples were centrifuged at 18 000 g for 20 min at 4 °C to remove proteins and 32 particulates. For feces samples preparation, 20 mg of each feces sample was 33 homogenized with 10-fold of 50% aqueous acetonitrile (containing 5 uM 34 35 chlorpropamide), shaken for 20 min at room temperature. Then the samples were centrifuged at 18000 g for 20 min at 4 °C. After that, 100 µL of each supernatant was 36 transferred to new tube. Then 200 µL of 50% aqueous acetonitrile was added into 37 each sample. The diluted samples were again centrifuged at 18000 g for 20 min at 38 4 °C. The supernatants were transferred to new tubes and 5 µL aliquot of each 39 sample was injected for UPLC-ESI-QTOFMS analysis. 40

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42 Structural identification of 1'-hydroxymyristicin and its metabolites

43 The chromatic pattern and fragment pathway of H0 were consistent with

44	1'-hydroxymyristicin (Supplementary Fig. 5A). Metabolite H1 was detected at m/z
45	209.0808 in full scan mode of MLMs sample at t_{R} of 8.42 min, which matched
46	structural formula of $C_{11}H_{12}O_4$. It was similar to H0, demonstrating that H1 may be
47	isomerization product of 1'-hydroxymyristicin. Different from the fragment pathway
48	of 1'-hydroxymyristicin, H1 gave various daughter ions included m/z 193.0555 (loss
49	of O atom), 179.0677 (loss of CH ₂ O moiety), 165.0522 (loss of CH ₂ O+CH ₂ moiety)
50	and 137.0928, which implied H1 was hydroxyl isomerization product at site of ally
51	(Supplementary Fig. 5B). The retention time and fragments of metabolite H2 were
51 52	(Supplementary Fig. 5B). The retention time and fragments of metabolite H2 were consistent with M6, suggesting that H2 and M6 were the same metabolites from
52	consistent with M6, suggesting that H2 and M6 were the same metabolites from
52 53	consistent with M6, suggesting that H2 and M6 were the same metabolites from myristicin and 1'-hydroxymyristicin metabolism, respectively. The MS/MS spectrum
52 53 54	consistent with M6, suggesting that H2 and M6 were the same metabolites from myristicin and 1'-hydroxymyristicin metabolism, respectively. The MS/MS spectrum of metabolite H3 revealed a molecular ion $[M+H]^+$ of m/z 179.0703, and was 30 Da

Metabolite H5 yielded $[M+H]^+$ at m/z 225.0757, and fragments of m/z 207 58 (dehydration), 184 [(M+H)-CO-CH₂]⁺ and 165 (6-vinylbenzo[d][1,3]dioxol-4-ol 59 residue) were observed in the MS/MS spectrum. The characteristic ion of m/z 184 60 (7-methoxybenzo[d][1,3]dioxol-5-yl)methanol) was formed via neutral losses of 61 acetaldehyde in site of chain, suggesting H5 62 the was 3-hydroxy-3-(7-methoxybenzo[d][1,3]dioxol-5-yl)propanal. Metabolite H4 ($t_R = 6.94$ min, 63 observed $[M+H]^+$ at m/z 223.0601) gave a match for the molecular formula $C_{11}H_{10}O_5$, 64 and was 2 Da (H₂) lower than H5, suggesting H4 was dehydrogenated product of H5. 65

According to the major two related fragment ions of H4 at m/z 205 [M+H-H₂O]⁺, 66 (6-vinylbenzo[d][1,3]dioxol-4-ol residue), it was 165^{+} identified H4 67 as 2',3'-dicarbonylated myristicin. Metabolite H6 exhibited an abundant parent ion 68 $[M+H]^+$ at m/z 207.0652, which accurately matched $C_{11}H_{10}O_4$. It was 2 Da (H₂) lower 69 than parent, implying that H6 was dehydrogenated product via hydroxyl oxidation. 70 Two daughter ions of H6 at m/z 177 [M+H-CH₂O]⁺ and 149 [M+H-(CH₂O+CO)]⁺ 71 were observed on the MS^2 spectra. Metabolite H7 exhibited quasi-molecular $[M+H]^+$ 72 ion at m/z 195.0662, which gave a match for the molecular formula of C₁₀H₁₀O₄ with 73 74 the error of -5.15 ppm. It was lower 14 Da than parent, suggesting that H7 was demethylated 1'-hydroxymyristicin. H7 yielded the major daughter ions at m/z 153 75 of CH₂=CH-OH), and 123 (cleavage of CH₂=CH-CHOH+O) 76 (cleavage 77 (Supplementary Fig. 5C).

The structural formula of H8 was speculated as C₉H₈O₄ from protonated molecular 78 $[M+H]^+$ ion at m/z 181.0495. It was lower 28 Da (CH₂=CH₂) than 79 1'-hydroxymyristicin, implying that H8 was devinylated 1'-hydroxymyristicin. It 80 produced prominent fragment ions at m/z 153 (cleavage of CH₂=CH-OH), and 123 81 82 (cleavage of CH₂=CH-CHOH+O). The chromatic pattern and fragment pathways of H9 were consistent with M3. The daughter ion spectra of H10 showed the diagnostic 83 neutral loss of 101 Da (m/z 328 \rightarrow 207), indicating the presence of a cysteine moiety 84 via the linker of sulfur atom. Furthermore, protonated cysteine ion m/z at 122 was 85 discovered in MS/MS spectrum of H10. Thereby, H10 was characterized as 86 1'-hydroxymyristicin plus cysteine conjugate (Supplementary Fig. 5D). 87

88	Figure Captions
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89	Supplementary Fig. 1 NMR spectral of 1'-hydroxymyristicin. (A) ¹ H-NMR
90	spectrum of 1'-hydroxymyristicin. (B) ¹³ C-NMR spectrum of 1'-hydroxymyristicin.
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92	Supplementary Fig. 2 The relative abundance of myristicin and its metabolites in
93	urine (A), in plasma (B) and in microsomes (C).
94	
95	Supplementary Fig. 3 The relative abundance of 1'-hydroxymyristicin and its
96	metabolites in urine (A), in plasma (B) and in microsomes (C).
97	
98	Supplementary Fig. 4 MS ² spectrums and fragmentation pathways of M0 (A), M1
99	(B), M2 (C) and M6 (D).
100	
101	Supplementary Fig. 5 MS ² spectrums and fragmentation pathways of H0 (A), H1
102	(B), H7 (C) and H10 (D).
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Metabolits (ID)	Rt (min)	Observed [M+H] ⁺	Molecular formula	Mass error (ppm)	Major fragment ions	Reaction	Identity	Source
M0	9.95	193.0859	$C_{11}H_{12}O_3$	0.43	163,152,135	-	Myristicin	M,H,U,P
M1*	6.12	195.0973	$C_{11}H_{14}O_3$	-5.15	181,165	1	2',3'-Reduced-myristicin	M,H,U
M2*	5.97	179.0683	$C_{10}H_{10}O_3$	7.39	161,151,147	2	3-Hydroxysafrole	M,H,U
M3	6.02	227.0903	$C_{11}H_{14}O_5$	7.59	167,109,81	3	2',3'-Dihydroxy myristicin	M,H,U
M4	5.99	209.0812	$C_{11}H_{12}O_4$	-8.02	179,168,94	4	4-or 6-Hydroxy myristicin	M,H,U
M5	6.61	181.0862	$C_{10}H_{12}O_3$	2.76	154,135,121	5	5-Allyl-1-methoxy-2,3-dihydroxybenzene	M,H,U
M6 *	5.54	191.0686	$C_{11}H_{10}O_3$	7.85	161,133	6	2',3'-Dehydro-myristicin	M,H,U
M7 *	5.62	165.0552	$C_9H_8O_3$	-2.27	131,115	7	4-Allyl-2-methoxyphenol	M,H
M8	6.87	231.0633Na ⁺	$C_{11}H_{12}O_4$	-4.33	213,191,165	4	1'-Hydroxymyristicin	M,H
M9	5.97	385.1092	$C_{17}H_{20}O_{10}$	5.52	209,165,147	4+Gluc	Myristicin-Gluc adduct	U
M10*	6.39	370.0947	C16H19NS7	-1.62	207,164,122	4+ NAC	Myristicin-NAC adduct	U

Supplementary Table1. Summary of metabolites of myristicin produced in vivo and in vitro metabolism

1, Reduction; 2, Demethylation; 3, Alkenes to dihydrodiol; 4, Hydroxylation; 5, open-ring; 6, Desaturation; 7, Demethylation; Gluc, glucuronide; NAC, *N*-acetylcysteine; U, urine; P, plasma; M, mouse liver microsomes; H, human liver microsomes. *Indicate novel metabolites found in this study.

Metabolites	Rt	Observed	Molecular	Mass	Major		Identity	
(ID)	(min)	[M+H] ⁺	formula	error (ppm)	fragment ions	Reaction		Source
HO	6.87	231.0633Na ⁺	$C_{11}H_{12}O_4$	0.67	213,191,165,123	-	1'-Hydroxymyristicin	М, Н, Р
H1	8.42	209.0808	$C_{11}H_{12}O_4$	1.76	193,179,165,137	-	3'-Hydroxymyristicin	M,H,U
H2*	5.55	191.0682	$C_{11}H_{10}O_3$	1.17	161,133,97,69	1	2',3'-Dehydro-myristicin	M,H,P,U
H3*	6.28	179.0703	$C_{10}H_{10}O_3$	1.30	161,137	2	Demethoxy -1'-hydroxymyristicin	М, Н
H4*	6.94	223.0601	$C_{11}H_{10}O_5$	7.63	205,165	3	1'3'-Dioxo-myristicin	M,H,P
H5*	6.73	225.0757	$C_{11}H_{12}O_5$	3.34	207,184,165	4	1'-Hydro-3'-oxo-mxymyristicin	Р
H6*	5.74	207.0652	$C_{11}H_{10}O_4$	0.56	177,149,81	5	1'-Oxo-mxymyristicin	Р
H7*	7.27	195.0662	$C_{10}H_{10}O_4$	-3.97	153,123	6	Dimethyl-myristicin	U
H8*	6.99	181.0495	$C_9H_8O_4$	-5.15	153.123,93	7	Myristic aldehyde	U
H9	6.02	227.0942	$C_{11}H_{14}O_5$	-2.97	167,109,81	8	1'3'-Dihydroxymyristicin	U
H10*	5.02	328.0862	$C_{14}H_{17}NS_6$	-0.23	207,122,85,55	H0 + <i>S</i> -Cys	Hydroxymyristicin-Cys adduct	U
H11*	6.40	370.0952	$C_{16}H_{19}NS_7$	-0.74	207,164,122,55	H0+ NACys	Myristicin-NAC adduct	U

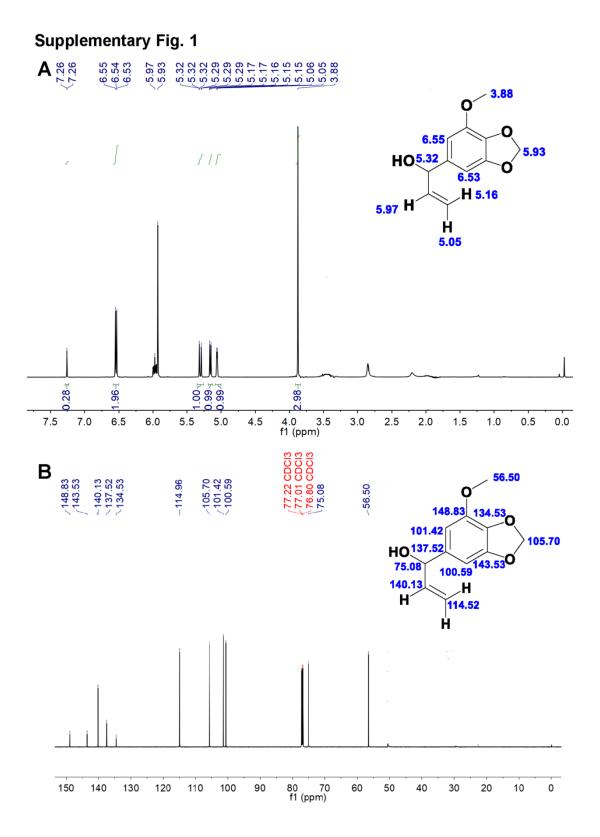
Supplementary Table 2. Summary of metabolites of 1'-hydroxymyristicin produced in vivo and in vitro metabolism

1, Alcohols dehydration; 2, hydroxymethylene loss; 3, Hydroxylation+desaturation; 4, Hydroxylation; 5, desaturation; 6, Demethylation; 7, Deethylation; 8, hydration; S-Cys, cysteine; NAC, acetylcysteine; U, urine; P, plasma; M, mouse liver microsomes; H, human liver microsomes. *Indicate novel metabolites found in this study.

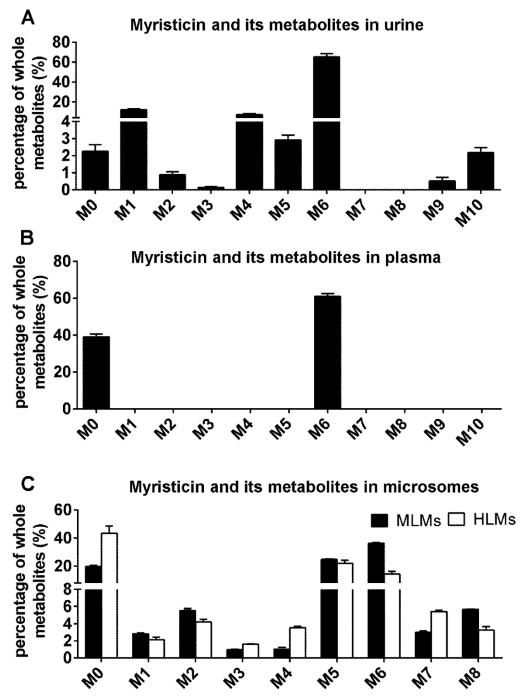
No.	Enzyme	M1	M2	M3	M4	M5	M6	M7	M8
0	Control	-	-	-	-	-	-	-	-
1	CYP1A1	-	36.49	17.25	30.64	8.99	42.73	38.78	47.48
2	CYP1A2	29.27	-	-	-	47.69	10.99	-	-
3	CYP1B1	20.15	63.51	-	18.89	-	16.52	61.21	13.65
4	CYP2A6	-	-	17.75	-	-	9.28	-	-
5	CYP2B6	-	-	-	-	-	-	-	-
6	CYP2C19	-	-	-	-	-	-	-	-
7	CYP2C8	-	-	-	-	-	-	-	-
8	CYP2C9	-	-	27.04	-	-	-	-	21.82
9	CYP2D6	10.19	-	-	-	-	-	-	-
10	CYP2E1	-	-	-	-	-	-	-	-
11	CYP3A4	16.75	-	17.02	17.60	43.31	5.09	-	-
12	CYP3A5	23.62	-	20.93	32.86	-	15.38	-	17.05

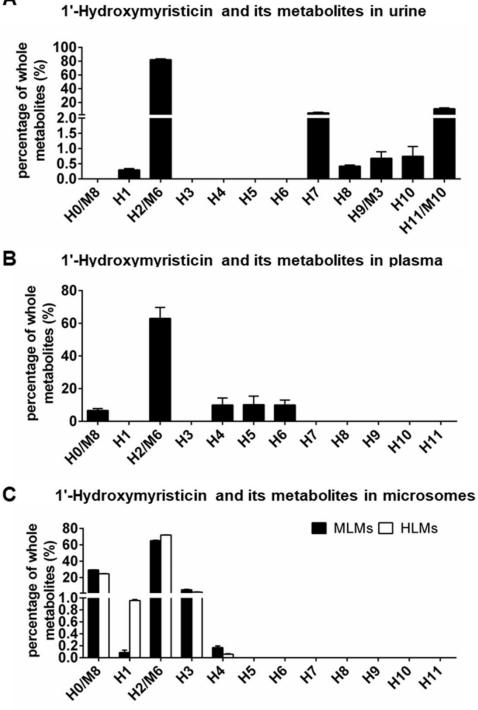
Supplementary Table 3. Role of CYP450s in the formation of myristicin metabolites

cDNA-expressed CYPs (control, CYP1A1, 1A2, 1B1, 2A6, 2B6, 2C19, 2C8, 2C9, 2D6, 2E1, 3A4, 3A5, and 4A11) were used to examine the roles of in individual CYPs in myristicin metabolism. All samples were analyzed by UPLC-ESI-QTOF-MS. The total peak areas of each metabolites of myristicin form all the CYPs were set as 100%. All data are expressed as mean (n=3).



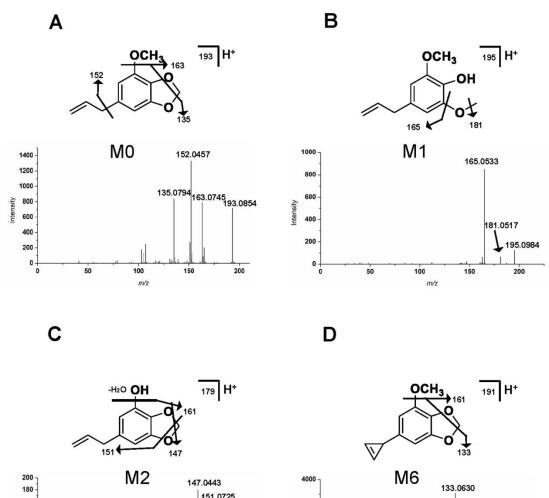
Supplementary Fig. 2

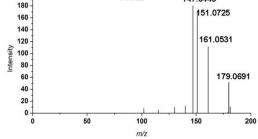


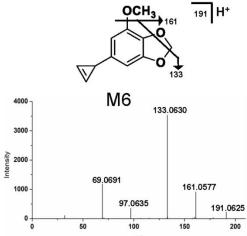


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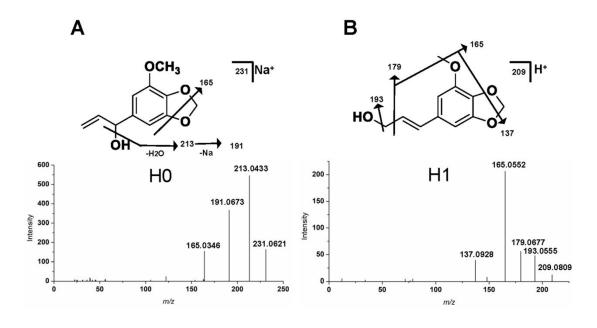
Supplementary Fig. 4







m/z



С

D

