### Supporting Information for

# Corannurylene Pentapetalae

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#### **Materials and Measurements**

Perylene-3,4,9,10-tetracarboxylic diimides,1-bromo-perylene-3,4,9,10-tetracarboxylic diimides were synthesized using known procedures.<sup>1</sup> All chemicals and solvents were purchased from commercial suppliers and used without further purification unless otherwise specified.

<sup>1</sup>H NMR (500 MHz) and <sup>13</sup>C NMR (125 MHz) spectra were recorded in deuterated solvents on a Bruker ADVANCE 500 NMR Spectrometer. *J* values are expressed in Hz and quoted chemical shifts are in ppm downfield from tetramethylsilane (TMS) reference using the residual protonated solvent as an internal standard. The signals have been designated as follows: s (singlet), d (doublet), t (triplet), q (quartet), sd (singlet doublet), dd (doublet doublet) and m (multiplets). High resolution mass spectra (HRMS) were determined on IonSpec 4.7 Tesla Fourier Transform Mass Spectrometer.

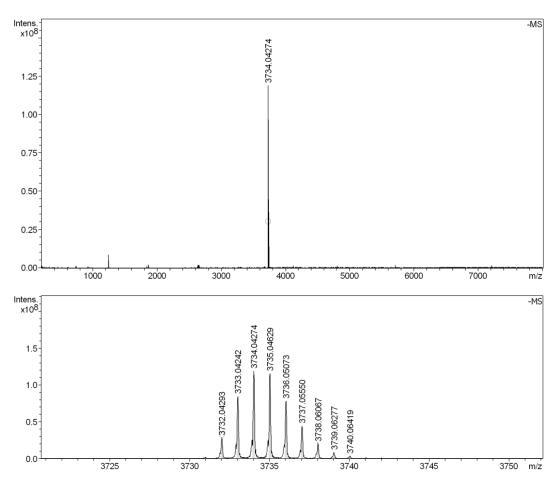
UV-vis spectra were measured with Hitachi (Model U-3010) UV-vis spectrophotometer in a 1-cm quartz cell. Cyclic voltammograms (CVs) were recorded on a Zahner IM6e electrochemical workstation using glassy carbon discs as the working electrode, Pt wire as the counter electrode, Ag/AgCl electrode as the of 100 reference electrode at scanning rate mV/s. 0.1 tetrabutylammoniumhexafluorophosphate (Bu<sub>4</sub>NPF<sub>6</sub>) dissolved in CH<sub>2</sub>Cl<sub>2</sub> employed supporting electrolyte, which was calibrated by as the ferrocene/ferroncenium (Fc/Fc<sup>+</sup>) as the redox couple. CH<sub>2</sub>Cl<sub>2</sub> was freshly distilled prior to use. The microscope images of all the micro-/nano-crystals were acquired by an optical microscope (Vision Engineering Co., UK), which was coupled to a CCD camera.

### **Synthetic Details**

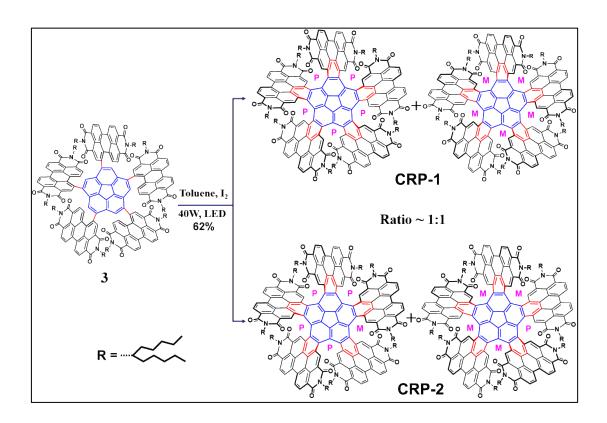
Compound 3: A Schlenk flask charged with was 1,3,5,7,9-pentakis(pinacolatoboryl)corannulene (2: 100 mg, 0.11 mmol ,1.0 equiv), 1-bromo-perylene-3,4,9,10-tetracarboxylic diimides (1: 486 mg, 0.62 mmol, 5.5 equiv), THF (6 ml) and Na<sub>2</sub>CO<sub>3</sub> aqueous solution (2M, 3 ml). The mixture was degassed with argon for 15min. PdCl<sub>2</sub>(dppf) (30 mg, 0.03 mmol, 0.3 equiv) was added under an argon atmosphere. The mixture was refluxed for 72 h and then cooled down to room temperature. The cooled mixture was poured into 1M HCl aqueous solution (500 ml) and stirred for 2 h. The precipitate was collected by vacuum filtration, washed with water and CH<sub>3</sub>OH, dried, and purified by column chromatography on silica gel, eluted with petroleum ether/CH<sub>2</sub>Cl<sub>2</sub>(1:1 to 1:5 v/v) to afford compound 3 (254 mg, 60% yield) as dark red solids.

<sup>1</sup>H NMR (500 MHz,  $C_2D_2Cl_4$ , 298.5 K)  $\delta = 8.81-8.39$  (m, 26 H), 8.07-7.24 (m, 14 H), 5.23-4.71 (m, 10 H), 2.36-1.82 (m, 35 H), 1.37-1.18 (m, 105 H), 0.96-0.28 (m, 72 H); <sup>13</sup>C NMR (125 MHz,  $C_2D_2Cl_4$ , 298.5 K):  $\delta = 163.88$ , 163.38, 162.78, 162.14, 161.30, 144.71, 137.39, 136.47, 135.71, 134.55, 134.13, 133.47, 132.93, 131.70, 131.09, 130.56, 130.28, 130.01, 129.06, 128.87, 128.24, 126.84, 126.24, 124.44, 123.56, 122.62, 121.93, 54.79, 53.53, 52.42, 51.73, 33.71, 33.07, 32.58, 32.01, 31.52,

30.11, 26.47, 25.23, 24.65, 24.39, 23.03, 22.76, 22.50, 21.91, 21.50, 14.54, 14.33, 14.15, 13.84, 13.13, 12.83, 12.24; HRMS (MALDI(N), 100%): calcd (%) for  $C_{250}H_{270}N_{10}O_{20}$ : 3732.04234; found, 3732.04293.

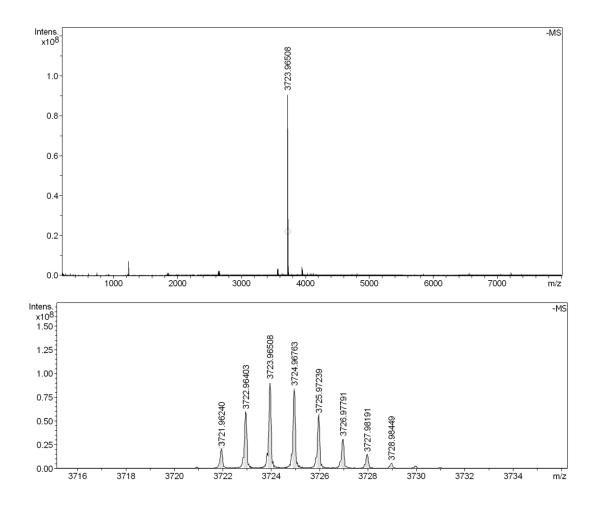


HR-MALDI-TOF mass spectrum of Compound 3.



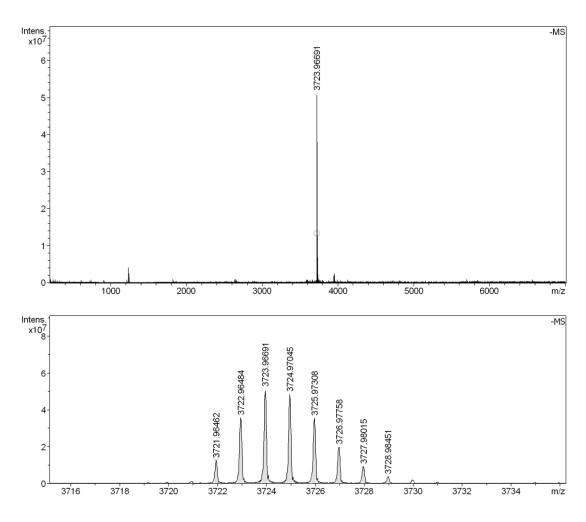
**Compound CRP-1 and CRP-2:** To a standard photocyclization glassware were added compound **3** (500 mg,  $10 \times 50$  mg), toluene (500 ml,  $10 \times 50$  ml) and  $I_2$  (50 mg,  $10 \times 5$  mg). The mixture was illuminated by 40 W mercury lamp for 72 h at 110 °C. The mixture was extracted with saturated sodium sulfite solution ( $2 \times 500$  ml) and concentrated under reduced pressure. The crude product was purified by column chromatography on silica gel, eluted with petroleum ether/CH<sub>2</sub>Cl<sub>2</sub> (3:2 to 1:1 v/v) to afford **compound CRP-1** (first part) and **CRP-2** (second part) as red solids (Ratio ~ 1:1,309 mg, 62% total yields).

Compound CRP-1: <sup>1</sup>H NMR (500 MHz,  $C_2D_2Cl_4$ , 373.2 K)  $\delta = 9.59$  (s, 10 H), 9.46-9.44 (d, J = 10 Hz, 10 H), 9.15-9.13 (d, J = 10 Hz, 10 H), 4.63 (s, 10 H), 1.64-1.57 (m, 25 H), 1.42 (s, 10 H), 1.25-1.20 (m, 85 H), 0.99-0.92 (m, 110 H); <sup>13</sup>C NMR (125 MHz,  $C_2D_2Cl_4$ , 373.2 K):  $\delta = 162.70$ , 162.25, 133.79, 133.11, 132.11, 132.03, 129.04, 128.32, 126.04, 124.77, 123.57, 122.48, 122.27, 121.64, 120.90, 53.26, 30.94, 30.88, 30.58, 30.40, 25.34, 25.08, 21.36, 21.17, 12.88; HRMS (MALDI(N), 100%): calcd (%) for  $C_{250}H_{260}N_{10}O_{20}$ : 3721.96409; found, 3721.96240.



HR-MALDI-TOF mass spectrum of **CRP-1**.

Compound CRP-2: <sup>1</sup>H NMR (500 MHz,  $C_2D_2Cl_4$ , 373.2 K) δ = 10.59 (s, 2 H), 10.42 (s, 2 H), 9.74 (s, 2 H), 9.55-9.51 (q,  $J_I$  = 5 Hz,  $J_2$  = 10 Hz,  $J_3$  = 5 Hz, 4 H), 9.48 (s, 2 H), 9.45-9.44 (d, J = 5 Hz, 3 H), 9.42-9.40 (m, 5 H), 9.26-9.25 (d, J = 5 Hz, 2 H), 9.17-9.11 (q,  $J_I$  = 10 Hz,  $J_2$  = 10 Hz,  $J_3$  = 10 Hz, 6 H), 9.05-9.04 (d, J = 5 Hz, 2 H), 5.08 (s, 2 H), 4.83-4.66 (m, 8 H), 2.14-1.87 (m, 9 H), 1.65 (m, 22 H), 1.35-0.78 (m, 199 H); <sup>13</sup>C NMR (125 MHz,  $C_2D_2Cl_4$ , 373.2 K): δ = 163.79, 162.97, 136.01, 135.62, 135.17, 134.51, 134.22, 134.15, 133.99, 133.71, 133.10, 132.52, 131.05, 130.04, 129.10, 127.91, 127.55, 127.20, 127.07, 126.70, 126.02, 125.91, 125.71, 125.48, 125.31, 124.80, 124.53, 123.46, 122.68, 121.89, 54.50, 54.35, 32.25, 32.11, 31.98, 31.66, 31.60, 31.51, 31.42, 31.27, 29.61, 26.51, 26.43, 26.22, 22.37, 22.26, 22.13, 13.85; HRMS (MALDI(N), 100%): calcd (%) for  $C_{250}H_{260}N_{10}O_{20}$  : 3721.96409; found, 3721.96462.



HR-MALDI-TOF mass spectrum of CRP-2.

## **Computational Methods Details:**

**I.** The structural and energetic analyses of the molecular systems described in this study were investigated with two types of density functional methods, B3LYP-D3<sup>2,3</sup> and wB97-D<sup>4</sup>, and several basis set types, specifically shown here are 6-311G(2d,p)<sup>5-7</sup> and Def2-TZVP<sup>8</sup> basis sets. All provided consistent results; B3LYP-D3/6-311G(2d,p) and B3LYP-D3/Def2-TZVP are shown here, and the former used in the discussion in the paper. All calculations employed an ultrafine grid. Full geometry optimizations were performed and uniquely characterized via second derivatives (Hessian) analysis to establish stationary points and effects of zero point energy. Visualization and analysis of structural and property results were obtained using Avogadro.<sup>9</sup>

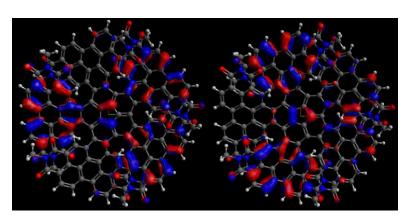
**Table S1.** The energetic analyses of four isomers.

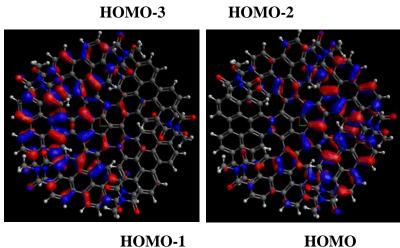
Molecule	B3LYP-D3/6	-311G(2d,p)	B3LYP-D3/	Def2-TZVP
	(kcal/mol)		(kcal/	/mol)
	$\Delta \mathbf{H}$	$\Delta \mathbf{G}$	$\Delta \mathbf{H}$	$\Delta \mathbf{G}$
CRP-1	0.0	0.0	0.0	0.0
CRP-2	5.00	5.53	5.7	6.2
CRP-3	5.56	6.03	6.3	6.8
CRP-4	9.95	11.4	11.2	12.6

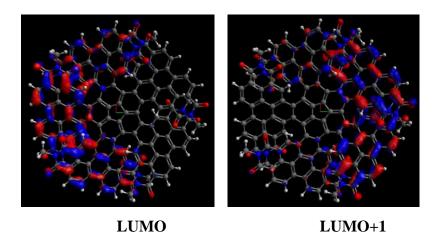
## Orbital structure for optimized CRP-1, $D_5$ symmetry

HOMO-3 HOMO-2 HOMO-1 HOMO
592 593 594 595
(E1)--0 (E1)--0 (E2)--0 (E2)--0
Eigenvalues -- -0.23555 -0.23555 -0.22300 -0.22300

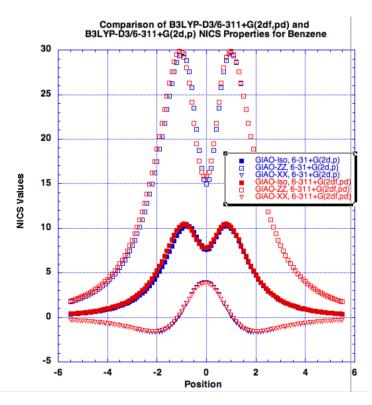
LUMO LUMO+1
596 597
(E2)--V (E2)--V
Eigenvalues -- -0.13721 -0.13721





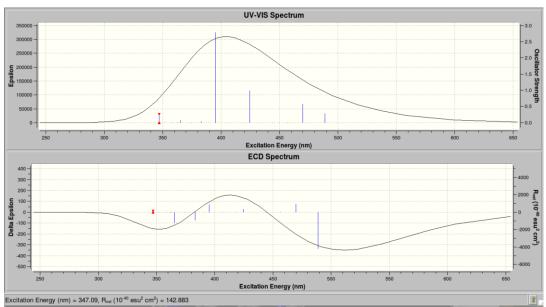


II. Nucleus-independent chemical shifts (NICS) properties, 10 including NICS(Iso), 11,12 NICS(In-plane), and NICS(ZZ) 13 values, were investigated using the method<sup>14,15</sup> B3LYP-D3/6-311+G(2d,p), 5-7 **GIAO** compared and at  $B3LYP-D3/6-311+G(2df,pd), ^{5\text{-}7} \ and \ B3LYP-D3/Def2-TZVP \ levels \ of \ theory, \ all \ of \ and \ baseline and \ bas$ which provided consistent results. The isotropic chemical shift values are separated into their in-plane (NICS(Iso)) and out-of-plane (NICS(ZZ)) components, by consideration of the eigenvalues of the chemical shift tensors. The NICS(ZZ) values, more closely related to the current density, reflect the magnetic response of a molecule toward a magnetic field applied perpendicular to the plane (i.e., z direction), and are therefore considered primarily associated with the  $\pi$  contribution. Graphical visualization of the results were carried out with the KaleidaGraph software.



**Figure S1.** B3LYP/6-311+G(2d,p) (blue) and B3LYP/6-311+G(2df,pd) (red) calculated NICS values along the symmetry axis for benzene  $D_{6h}$  symmetry.

**III.** Absorption energies were computed at the TD-B97xD<sup>16</sup>/6-311G(2d,p), TD-B97xD<sup>16</sup>/Def2-TZVP and TD-camB3LYP<sup>17</sup>/6-311G(2d,p), TD-camB3LYP<sup>17</sup>/Def2-TZVP levels of theory in gas phase and in chloroform, with consistent results. Representative spectra shown here carried out in chloroform.



**Figure S2.** TD-B97xD/6-311G(2d,p) (chloroform) UV-Vis and ECD spectra for CRP-1.

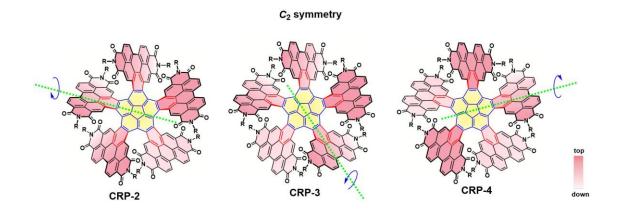
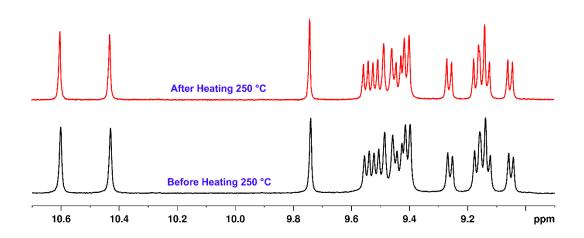


Figure S3.  $C_2$  axis position of CRP-2, CRP-3 and CRP-4.



**Figure S4.** The <sup>1</sup>H NMR spectra of **CRP-2** before and after heating in the diphenyl ether at 250 °C for 6 hours. (500 MHz, C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub>, 373.2 K)

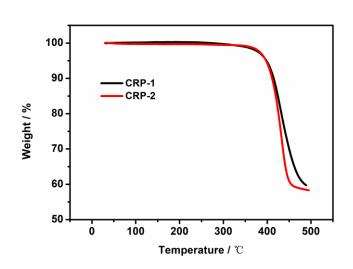
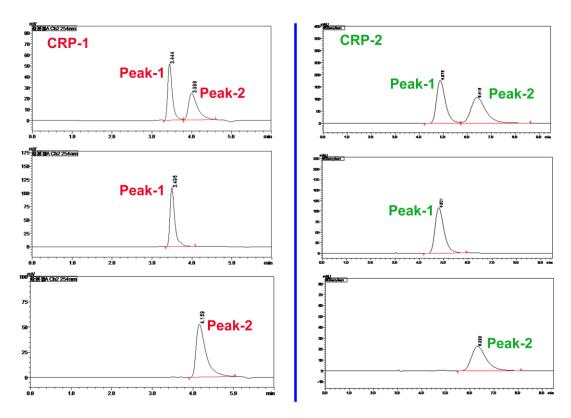
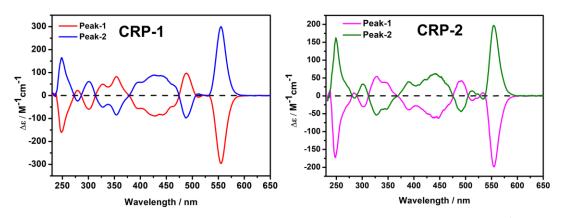


Figure S5. TGA curves of CRP-1 and CRP-2 under nitrogen flow.



**Figure S6.** Chiral HPLC chromatogram of **CRP-1** (red) and **CRP-2** (green). The separation conditions: for **CRP-1**, CHIRALPAK AS-H (ASH0CE-PG009) column with n-hexane/Isopropyl alcohol/ Diethanolamine (99.8/0.2/0.05(V/V/V)) as eluent; for **CRP-2**, CHIRALPAK IG column with Hexane/EtOH (90/10(V/V)) as eluent.



**Figure S7.** The CD spectra of **CRP-1** and **CRP-2** in CHCl<sub>3</sub> solution  $(1.0 \times 10^{-5} \text{ M})$ .

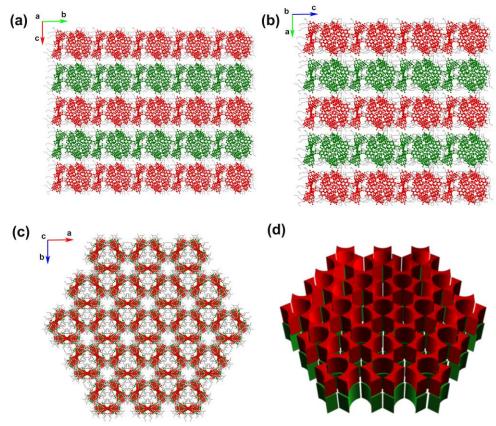
## X-ray Crystal Structure

The single crystals were obtained by slow vapor diffusion method under the THF (good solvent) and CH<sub>3</sub>OH (poor solvent). Single crystal data collections were performed at about 100 K and 170 K for CRP-2 and CRP-1 on a MD2 diffractometer and MM007HF Saturn724+ diffractometer, using adjustable graphite-monochromated radiation ( $\lambda$ =0.88557 Å) and Mo K $\alpha$  radiation (0.71073 Å), respectively. Using Olex2, these structures were solved with the ShelXS and refined with the ShelXL-2014 refinement package using Least Squares minimization. Refinement was performed on  $F^2$  anisotropically for all the non-hydrogen atoms by the full-matrix least-squares method. The hydrogen atoms were placed at the calculated positions and were included in the structure calculation without further refinement of the parameters. Because of the very large thermal motion and disorder of side chains in the lattice, the diffuse residual electron density is difficult to be accurately modeled and thus a treatment by SQUEEZE (from PLATON) was used to dissolve the disorder in the side chains. Despite many attempts, the diffraction intensity of CRP-1 and CRP-2 is very weak, which lead to their high R values and alerts of level A in checkCIF reports. Fortunately, the diffraction from the conjugated backbone could be easily confirmed, and we did not find any errors from the conjugated backbone.

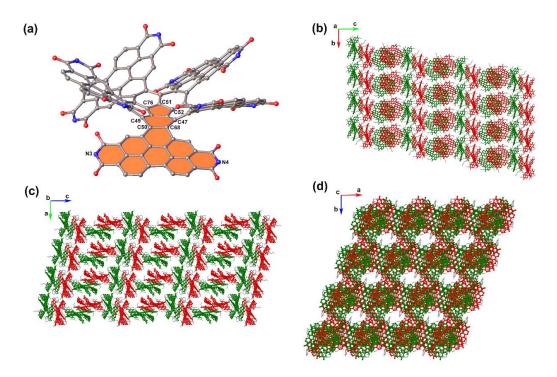
Table S2. Crystallographic data and structure refinement details for CRP-1 and CRP-2

CRP-1	CRP-2
173 (2)	100
0.71073	0.88557
Trigonal	Triclinic
P3c1	P-1
31.791(5)	21.504(4)
31.791(5)	22.883(5)
45.798(9)	44.815(9)
90.00	96.640(4)
90.00	100.370(6)
	173 (2) 0.71073 Trigonal  P3c1 31.791(5) 31.791(5) 45.798(9) 90.00

γ, (deg)	120.00	109.800(9)
$V$ , ( $\mathring{\mathrm{A}}^3$ )	40085(11)	20035(7)
Z	6	2
$D_{\rm calc}$ , (g / cm <sup>3</sup> )	0.731	0.960
$\mu \left(mm^{\text{-}1}\right)$	0.047	0.063
F(000)	9252	6072
Theta range, (deg)	0.74-25.01	0.94-20.99
reflns collected	345743	256413
indep reflns/ $R_{\rm int}$	47156 / 0.1201	39076 / 0.1962
params	2035	4002
GOF on $F^2$	1.519	1.973
$R_1$ , w $R_2$ [I>2 $\sigma$ (I)]	0.2756 / 0.5701	0.2807 / 0.5710
$R_1$ , w $R_2$ (all data)	0.3206 / 0.6104	0.3473 / 0.6103



**Figure S8.** The crystal 3D-stacking model of **CRP-1** with side chains (a) (b) (c) and schematic diagram (d) (along the c axis).



**Figure S9.** (a) The relatively planar component of **CRP-2** consisting of down PDI petal and part of the corannulene core (orange part) and (b) (c) (d) the crystal 3D-stacking model of **CRP-2** with part of side chains.

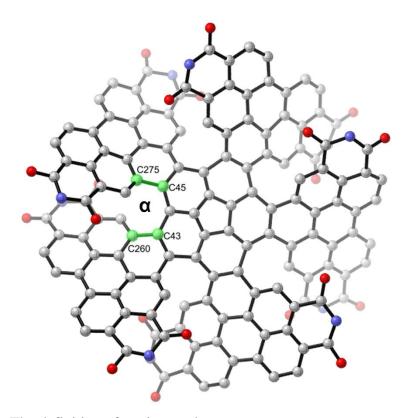


Figure S10. The definition of torsion angle  $\alpha$ .

**Transistor Device Fabrication and Characterization:** The substrates used here were successively cleaned with pure water, piranha solution ( $H_2SO_4/H_2O_2 = 7:3$ ), pure water, pure isopropyl alcohol, and finally blown dry with high-purity nitrogen gas. Treatment of the  $Si/SiO_2$  wafers with OTS was carried out by the vapor-deposition method. The clean wafers were dried under vacuum at 90 °C for 0.5 h in order to eliminate the influence of moisture. When the temperature decreased to 70 °C, a small drop of OTS was placed around the wafers. Subsequently, this system was heated to 120 °C and maintained for 2 h under vacuum.

Micro-/nano-crystals were fabricated by the use of a dilute CHCl<sub>3</sub> solution (1 mg mL<sup>-1</sup>). Bottom-gate top-contact OFETs based on the micro-/nano-crystals were constructed on an OTS modified Si/SiO<sub>2</sub> substrate (n-type Si wafer containing 300 nm-thick SiO<sub>2</sub>) using an "organic ribbon mask" technique. Prior to the self-assembly of micro-/nano-crystals, the OTS modified Si/SiO<sub>2</sub> substrate was cleaned with pure n-hexane, pure chloroform, and pure isopropyl alcohol. Then, micro-/nano-crystals were produced on Si/SiO<sub>2</sub> substrates through drop casting. Subsequently, 40 nm thick source and drain electrodes were deposited on the micro-/nano-crystals by thermal evaporation. Electrical characteristics of the devices were tested with a Keithley 4200-SCS semiconductor parameter analyzer and a Micromanipulator 6150 probe station in a glove box at room temperature. The mobilities were calculated from the saturation region with the following equation:  $I_{DS} = (W/2L) C_i \mu (V_G - V_T)^2$ , where  $I_{DS}$ is the drain-source current, W is the channel width, L is the channel length,  $\mu$  is the field-effect mobility,  $C_i$  is the capacitance per unit area of the gate dielectric layer, and  $V_{\rm G}$  and  $V_{\rm T}$  are the gate voltage and threshold voltage, respectively. This equation defines the important characteristics of electron mobility ( $\mu$ ), on/off ratio ( $I_{\text{on/off}}$ ), and threshold voltage  $(V_T)$ , which could be deduced by the equation from the plot of current-voltage.

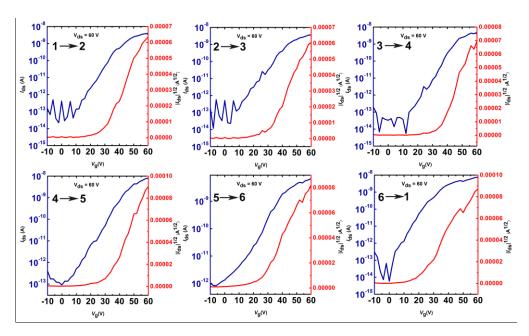


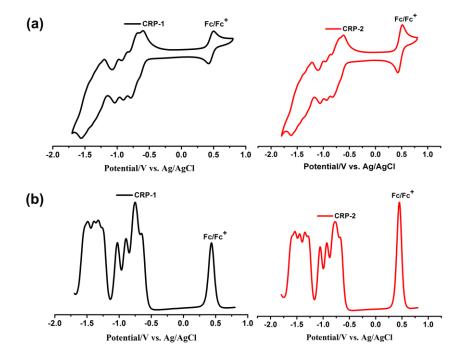
Figure S11 Transfer curves of six OFET devices with different crystal planes.

**Table S3.** Field-effect electron (e) mobility of six OFET devices with different crystal planes.

Channels	1 ← 2	2 ←→ 3	3 ← → 4	4 ← → 5	5 ← 6	6←→1
$\mu \left[ \text{cm}^2/(\text{V} \cdot \text{s}) \right]$	5.92×10 <sup>-4</sup>	5.69×10 <sup>-4</sup>	7.32×10 <sup>-4</sup>	6.69×10 <sup>-4</sup>	9.64×10 <sup>-4</sup>	6.92×10 <sup>-4</sup>
$I_{on}/I_{off}$	$1.82 \times 10^5$	$3.08 \times 10^5$	$8.84 \times 10^5$	$8.84 \times 10^4$	$8.17 \times 10^3$	$1.30 \times 10^6$

**Table S4.** The mobilities in thin-film devices made of CRP-1 and CRP-2 by spin-coating method (Solvent: CHCl<sub>3</sub>; 3000 rpm; Concentration: 10 mg/ml).

Devices	$W/L$ ( $\mu m$ )	Mobility <sup>ave</sup> (cm <sup>2</sup> V <sup>-1</sup> s <sup>-1</sup> )	Mobility <sup>max</sup> $(cm^2 V^{-1} s^{-1})$	$V_{\mathrm{T}}(\mathrm{V})$	On/off Ratio
CRP-1	1400/10	9.22×10 <sup>-4</sup>	1.32×10 <sup>-3</sup>	-5.1	$3.2 \times 10^6$
CRP-2		$7.03 \times 10^{-4}$	$8.19 \times 10^{-4}$	-5.4	$4.9 \times 10^5$



**Figure S12**. Cyclic voltammetries (a) and DPV profiles (b) of **CRP-1** and **CRP-2** in  $CH_2Cl_2$ .

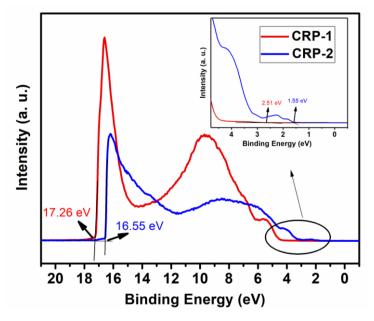
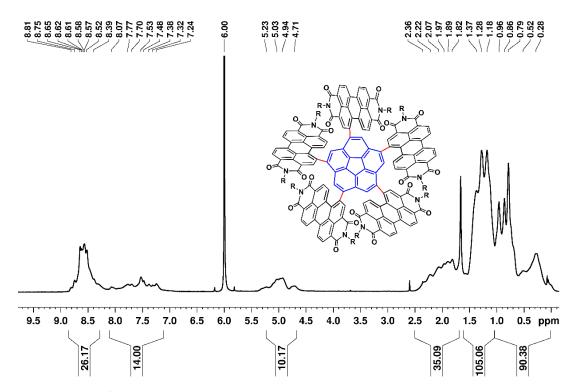


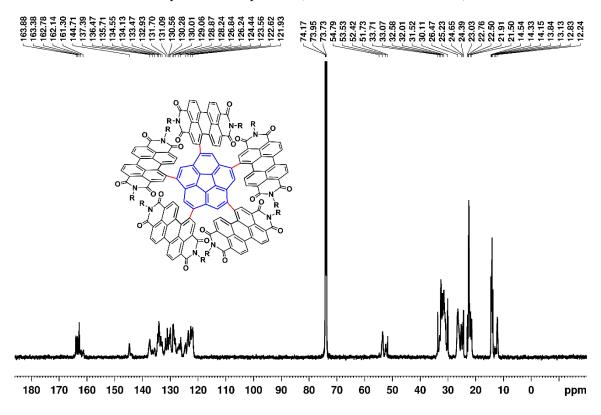
Figure S13. UPS characterization of CRP-1 and CRP-2 solids

The work function (WF) and ionization potential (IP) of CRP-1 and CRP-2 samples were measured by ultraviolet photoemission spectroscopy (UPS). As shown in Figure S13, the binding energy cutoff ( $E_{cutoff}$ ) values of CRP-1 and CRP-2 samples are 17.26 and 16.55 eV, respectively. Accordingly, the calculated WF and IP of CRP-1 samples are 3.96 eV and 6.47 eV, CRP-2 samples are 4.67 eV and 6.22 eV, respectively.

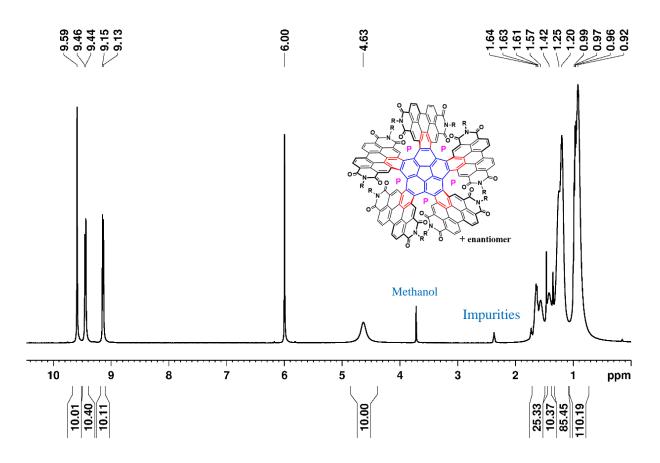
## <sup>1</sup>H NMR and <sup>13</sup>C NMR Spectra of Products



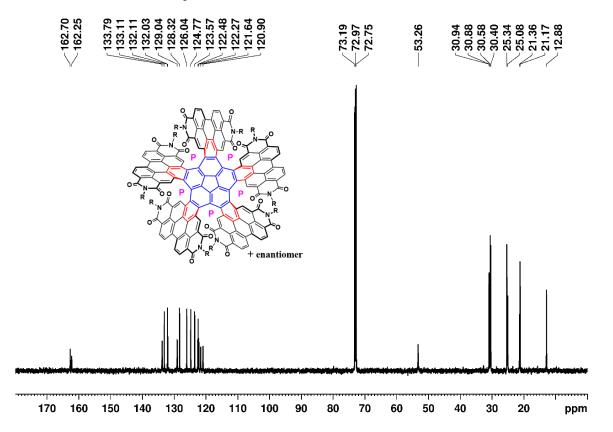
 $^{1}\text{H}$  NMR Spectra of Compound 3 (500 MHz,  $C_{2}D_{2}Cl_{4}$ , 298.5 K)



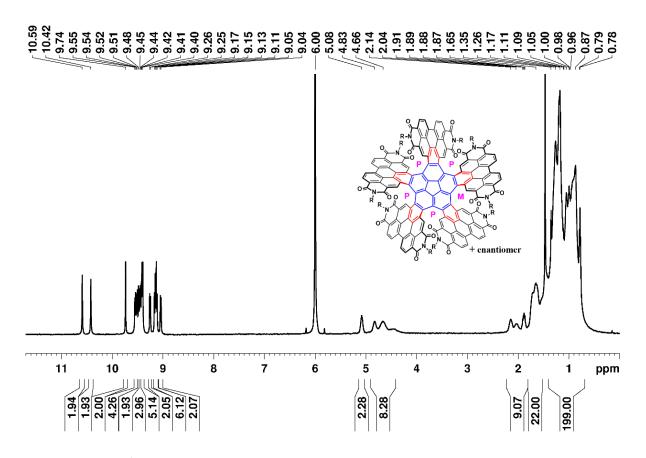
 $^{13}\text{C}$  NMR Spectra of Compound 3 (500 MHz,  $C_2D_2Cl_4,\,298.5$  K)



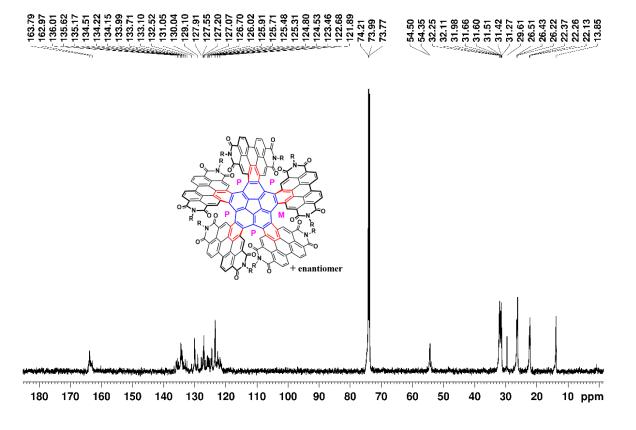
<sup>1</sup>H NMR Spectra of **CRP-1** (500 MHz, C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub>, 373.2 K)



 $^{13}C$  NMR Spectra of CRP-1 (500 MHz,  $C_2D_2Cl_4,\,373.2~K)$ 



<sup>1</sup>H NMR Spectra of **CRP-2** (500 MHz, C<sub>2</sub>D<sub>2</sub>Cl<sub>4</sub>, 373.2 K)



 $^{13}\text{C}$  NMR Spectra of CRP-2 (500 MHz,  $C_2D_2Cl_4,\,373.2~\text{K})$ 

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