

Novel aryloxyethyl derivatives of
1-(1-benzoylpiperidin-4-yl)methanamine as the
Extracellular Regulated Kinases 1/2 (ERK1/2)
phosphorylation-preferring serotonin 5-HT_{1A}
receptor biased agonists with robust antidepressant-
like activity

Joanna Sniecikowska^a, Monika Gluch-Lutwin^a, Adam Bucki^a, Anna Więckowska^a, Agata Siwek^a, Magdalena Jastrzebska-Wiesek^a, Anna Partyka^a, Daria Wilczyńska^a, Karolina Pytka^a, Krzysztof Pociecha^a, Agnieszka Cios^a, Elżbieta Wyska^a, Anna Wesółowska^a, Maciej Pawłowski^a, Mark Varney^b, Adrian Newman-Tancredi^b, Marcin Kolaczowski^{a}*

^a Faculty of Pharmacy, Jagiellonian University Medical College, 9 Medyczna St., 30 688 Kraków, Poland;

^b Neurolix Inc. 34145 Pacific Coast Highway #504, Dana Point, CA 92629 USA

*Corresponding Author Information:

Phone: (48)126205460 Fax: (48)126205458.

E-mail: marcin.kolaczowski@uj.edu.pl

Contents

Page number

Functional activity results at 5-HT _{1A} receptors with the SEM values.	S3
<i>E</i> _{max} and EC ₅₀ values (transformed from pEC ₅₀ values) used to calculate Bias Factors for compounds 10–17, 19, 20, 22, 27 and references	S4
Radioligand binding assay results and methodology for 5-HT _{2A} , M ₁ and H ₁ receptors	S5
Effect of compound 17 in the forced swimming test in rats (15 and 60 minutes after oral administration).	S9
Dose-response curves for functional activity.	S10

Functional activity results at 5-HT_{1A} receptors with the SEM values.

Table S1. Functional activity of compounds **10–17, 19, 20, 22, 27** and references at 5-HT_{1A} receptors.

Compd	5-HT _{1A} functional activity							
	ERK		cAMP		β-arrestin		Ca ²⁺	
	E _{MAX}	pEC ₅₀	E _{MAX}	pEC ₅₀	E _{MAX}	pEC ₅₀	E _{MAX}	pEC ₅₀
10	77% ± 7.9%	7.88 ± 0.07	91% ± 5.3%	7.10 ± 0.11	99% ± 0.5%	6.35 ± 0.02	74% ± 2.4%	7.07 ± 0.02
11	95% ± 3.5%	7.90 ± 0.08	74% ± 5.1%	7.33 ± 0.06	101% ± 0.5%	6.94 ± 0.11	87% ± 7.5%	7.84 ± 0.06
12	81% ± 6.3%	7.59 ± 0.09	79% ± 2.7%	6.73 ± 0.01	104% ± 2.1%	5.70 ± 0.07	65% ± 3.0%	6.98 ± 0.06
13	83% ± 4.6%	7.76 ± 0.14	86% ± 4.8%	7.26 ± 0.10	99% ± 0.8%	6.58 ± 0.09	52% ± 0.2%	7.13 ± 0.01
14	85% ± 1.3%	6.77 ± 0.02	71% ± 4.7%	5.61 ± 0.11	70% ± 2.9%	5.44 ± 0.03	NT	NT
15	84% ± 2.3%	7.89 ± 0.09	76% ± 7.4%	7.18 ± 0.19	102% ± 2.4%	6.54 ± 0.15	73% ± 9.8%	6.95 ± 0.01
16	92% ± 2.7%	9.10 ± 0.14	90% ± 2.3%	8.09 ± 0.12	96% ± 1.0%	7.98 ± 0.02	52% ± 2.0%	7.39 ± 0.17
17	81% ± 7.2%	9.37 ± 0.19	89% ± 3.3%	8.05 ± 0.09	98% ± 1.0%	7.68 ± 0.11	57% ± 5.8%	7.76 ± 0.05
19	85% ± 0.6%	9.06 ± 0.17	71% ± 7.3%	8.04 ± 0.06	99% ± 0.9%	6.76 ± 0.13	58% ± 1.5%	7.41 ± 0.09
20	99% ± 2.5%	7.62 ± 0.16	100% ± 4.0%	7.32 ± 0.20	101% ± 1.1%	6.87 ± 0.09	98% ± 1.4%	7.34 ± 0.06
22	100% ± 5.2%	8.31 ± 0.17	90% ± 6.4%	7.68 ± 0.08	97% ± 2.8%	7.60 ± 0.17	89% ± 5.3%	7.54 ± 0.20
27	72% ± 8.2%	7.28 ± 0.02	41% ± 1.4%	6.03 ± 0.03	75% ± 3.7%	5.56 ± 0.06	NT	NT
1	100% ± 4.3%	8.33 ± 0.14	92% ± 6.9%	7.22 ± 0.08	98% ± 1.0%	6.71 ± 0.07	60% ± 1.7%	6.61 ± 0.10
8-OH-DPAT	93% ± 3.6%	8.09 ± 0.11	63% ± 2.3%	7.50 ± 0.05	101% ± 1.5%	7.84 ± 0.18	NT	NT
Buspirone	44% ± 5.5%	7.82 ± 0.13	49% ± 5.7%	7.14 ± 0.12	100% ± 1.1%	6.73 ± 0.08	NT	NT
Serotonin	100% ± 0.0	7.48 ± 0.08	100% ± 0.0%	7.51 ± 0.05	100% ± 0.0%	6.89 ± 0.04	100% ± 0.0%	7.30 ± 0.08

E_{\max} and EC_{50} values (transformed from pEC_{50} values) used to calculate Bias Factors for compounds 10–17, 19, 20, 22, 27 and references

Table S2. E_{\max} and EC_{50} values (transformed from pEC_{50} values) used to calculate Bias Factors for compounds 10–17, 19, 20, 22, 27 and references .

Compd	5-HT _{1A} functional activity							
	ERK		cAMP		β -arrestin		Ca ²⁺	
	E_{\max}	EC_{50}	E_{\max}	EC_{50}	E_{\max}	EC_{50}	E_{\max}	pEC_{50}
10	0,77	1.31E-08	0.91	7.96E-08	0.99	4.46E-07	0.74	8.58E-08
11	0.95	1.25E-08	0.74	4.71E-08	1.01	1.15E-07	0.87	1.43E-08
12	0.81	2.55E-08	0.79	1.86E-07	1.04	1.99E-06	0.65	1.04E-07
13	0.83	1.75E-08	0.86	5.44E-08	0.99	2.60E-07	0.52	7.47E-08
14	0.85	1.69E-07	0.71	2.44E-06	0.70	3.61E-06	NT	NT
15	0.84	1.29E-08	0.76	6.62E-08	1.02	2.88E-07	0.73	1.11E-07
16	0.92	7.93E-10	0.90	8.09E-09	0.96	1.05E-08	0.52	4.11E-08
17	0.81	4.29E-10	0.89	8.85E-09	0.98	2.09E-08	0.57	1.72E-08
19	0.85	8.72E-10	0.71	9.10E-09	0.99	1.73E-07	0.58	3.85E-08
20	0.99	2.39E-08	1.00	4.79E-08	1.01	1.36E-07	0.98	4.54E-08
22	1.00	4.94E-09	0.90	2.10E-08	0.97	2.51E-08	0.89	2.86E-08
27	0.72	5.22E-08	0.41	9.25E-07	0.75	2.74E-06	NT	NT
1	1.00	4.71E-09	0.92	6.01E-08	0.98	1.93E-07	0.60	2.47E-07
8-OH-DPAT	0.93	8.17E-09	0.63	3.19E-08	1.01	1.44E-08	NT	NT
Buspirone	0.44	1.51E-08	0.49	7.20E-08	1.00	1.86E-07	NT	NT
Serotonin	1.00	3.28E-08	1.00	3.07E-08	1.00	1.30E-07	1.00	4.99E-08

Radioligand binding assay results and methodology for 5-HT_{2A}, M₁ and H₁ receptors

Table S3. Radioligand binding assay results for 5-HT_{2A}, M₁ and H₁ receptors using [³H]-ketanserin, [³H]-scopolamine and [³H]-pyrilamine, respectively.

Compound	percentage of control specific binding at 1 μ M concentration		
	5-HT _{2A} R	M ₁ R	H ₁ R
10	23%	10%	0%
11	0%	6%	0%
12	3%	10%	0%
13	12%	0%	0%
14	3%	4%	0%
15	0%	0%	0%
16	2%	0%	0%
17	4%	-1%	0%
18	0%	0%	0%
19	4%	0%	17%
20	44%	0%	40%
21	0%	0%	45%
22	4%	8%	1%
23	0%	6%	0%
24	0%	1%	0%
25	0%	2%	0%
26	0%	0%	0%
27	0%	0%	1%
1	0%	0%	0%
Buspirone	17%	0%	50%
Methiothepin	100%	23%	99%
Mepyramine	–	–	99%
Scopolamine	–	100%	–

Preparation of solutions of test and reference compounds

10 mM stock solutions of tested compounds were prepared in DMSO. Serial dilutions of compounds were prepared in 96-well microplate in assay buffers using automated pipetting system epMotion 5070 (Eppendorf). Each compound was tested in a screening assay at final concentration of 1 μ M.

5-HT_{2A} Receptor Binding Assay

Radioligand binding was performed using membranes from CHO-K1 cells stably transfected with the human 5-HT_{2A} receptor (PerkinElmer). All assays were carried out in duplicates. 50 μ L working solution of the tested compounds, 50 μ L [³H]-ketanserin (final concentration 1 nM) and 150 μ L diluted membranes (7 μ g protein per well) prepared in assay buffer (50 mM Tris, pH 7.4, 4 mM CaCl₂, 0.1% ascorbic acid) were transferred to polypropylene 96-well microplate using 96-wells pipetting station Rainin Liquidator (MettlerToledo). Mianserin (10 μ M) was used to define nonspecific binding. Microplate was covered with a sealing tape, mixed and incubated for 60 minutes at 27 °C. The reaction was terminated by rapid filtration through GF/B filter mate presoaked with 0.5% polyethyleneimine for 30 minutes. Ten rapid washes with 200 μ L 50 mM Tris buffer (4 °C, pH 7.4) were performed using automated harvester system Harvester-96 MACH III FM (Tomtec). The filter mates were dried at 37 °C in forced air fan incubator and then solid scintillator MeltiLex was melted on filter mates at 90 °C for 5 minutes. Radioactivity was counted in MicroBeta2 scintillation counter (PerkinElmer). Data were fitted to a one-site curve-fitting equation with Prism 6 (GraphPad Software).

M₁ Receptor Binding Assay

Radioligand binding was performed using membranes from CHO-K1 cells stably transfected with the human M₁ receptor. All assays were carried out in duplicates. 50 µL working solution of the tested compounds, 50 µL [³H]-scopolamine (spec. act. 84.1 Ci/mmol, final concentration 0.2 nM) and 150 µL diluted membranes (35 µg protein per well) prepared in assay buffer (PBS, pH 7.4) were transferred to polypropylene 96-well microplate using 96-wells pipetting station Rainin Liquidator (MettlerToledo). Atropine (10 µM) was used to define nonspecific binding. Microplate was covered with a sealing tape, mixed and incubated for 120 minutes at 27 °C. The reaction was terminated by rapid filtration through GF/A filter mate presoaked with 0.5% polyethyleneimine for 30 minutes. Five rapid washes with 300 µL 50 mM Tris buffer (4 °C, pH 7.4) were performed using 96-well FilterMate harvester (PerkinElmer, USA). The filter mates were dried at 37 °C in forced air fan incubator and then solid scintillator MeltiLex was melted on filter mates at 90 °C for 4 minutes. Radioactivity was counted in MicroBeta2 scintillation counter (PerkinElmer). Data were fitted to a one-site curve-fitting equation with Prism 5 (GraphPad Software).

H₁ Receptor Binding Assay

Radioligand binding was performed using membranes from CHO-K1 cells stably transfected with the human histaminergic H₁ receptor. All assays were carried out in duplicates. 50 µL working solution of the tested compounds, 50 µL [³H]-Pyrilamine (spec. act. 20.0 Ci/mmol, final concentration 1.5 nM) and 150 µL diluted membranes (5 µg protein per well) prepared in assay buffer (50 mM Tris-HCl, pH 7.4; 5 mM MgCl₂) were transferred to polypropylene 96-well microplate using 96-wells pipetting station Rainin Liquidator (MettlerToledo). Mepyramine (10 µM) was used to define nonspecific binding. Microplate was covered with a sealing tape, mixed and incubated for 60 minutes at 27 °C. The reaction was terminated by rapid filtration through

GF/B filter mate presoaked with 0.5% polyethyleneimine for 30 minutes. Five rapid washes with 300 μ L 50 mM Tris buffer (4 °C, pH 7.4) were performed using 96-well FilterMate harvester (PerkinElmer, USA). The filter mates were dried at 37 °C in forced air fan incubator and then solid scintillator MeltiLex was melted on filter mates at 90 °C for 4 minutes. Radioactivity was counted in MicroBeta2 scintillation counter (PerkinElmer). Data were fitted to a one-site curve-fitting equation with Prism 5 (GraphPad Software).

Effect of compound 17 in the forced swimming test in rats (15 and 60 minutes after oral administration).

Table S4. Effect of compound 17 in the forced swimming test in rats (15 and 60 minutes after oral administration).

Treatment	Dose (mg/kg)	15 minutes after p.o. administration	60 minutes after p.o. administration
		Immobility time (s) (mean \pm SEM)	
Vehicle (water)	0	252.50 \pm 12.19	212.63 \pm 17.1
Compd 17	0.04	212.33 \pm 23.53; ns	174.75 \pm 10.2; ns
	0.16	189.67 \pm 10.47; p<0.05	145.43 \pm 13.9; p<0.01
	0.63	76.17 \pm 17.11; p<0.00001	67.75 \pm 12.5; p<0.00001
	2.5	2.38 \pm 0.46; p<0.00001	6.86 \pm 2.3; p<0.00001
		F ^a (4,29)=63.220; p<0.0000001	F ^a (4,33)=43.736; p<0.000001
Vehicle (water) + vehicle	0 + 0	253.20 \pm 11.70	207.14 \pm 16.2
Compd 17 + vehicle	2.5 + 0 (15 minutes)/ 0.16 + 0 (60 minutes)	2.38 \pm 0.46; p<0.001 vs veh p<0.001 vs WAY p<0.001 vs WAY+ Compd 17	114.00 \pm 9.4 p<0.001 vs veh p<0.001 vs WAY + Compd 17
WAY-100635 + vehicle	0.63 + 0	260.43 \pm 8.65; ns vs veh p<0.001 vs NLX204 p<0.001 vs WAY+NLX204	260.43 \pm 8.7 p<0.05 vs veh p<0.001 vs WAY + Compd 17
WAY-100635 + Compd 17	0.63 + 2.5 (15 minutes)/ 0.63 + 0.16 (60 minutes)	142.38 \pm 17.57; p<0.001 vs veh p<0.001 vs WAY p<0.001 vs Compd 17	210.00 \pm 12.7 p<0.01 vs WAY p<0.001 vs Compd 17
		F ^b (1,27)=31.616; p<0.0001	F ^b (1,24)=3.1208; NS

Compound 17 was given p.o. 15 min or 60 min, while WAY-100635 was injected s.c. 30 min or 75 min, respectively before the test. Values represent the mean \pm SEM of the immobility time during 5-min test session compared to: the respective vehicle group (^aone-way ANOVA followed by Bonferroni's post hoc test – when only one drug was given); or the respective groups (^b two-way ANOVA followed by Newman-Keuls post hoc test, when two drugs were injected); N=6-8

Dose-response curves for functional activity.



