

Supporting Information

Acoustic Droplet Ejection Enabled Automated Reaction Scouting

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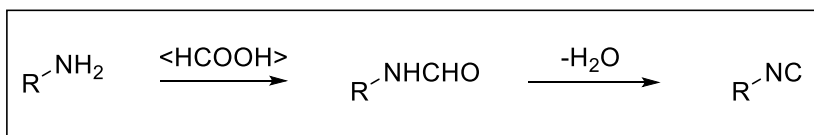
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1. General information

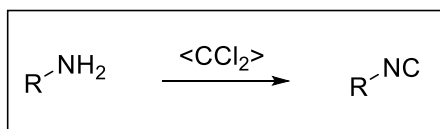
1.1. Materials and methods

All reagents and solvents were purchased from commercial suppliers and used without any purification unless otherwise noted. All isocyanides were prepared in house by either performing the Ugi,¹⁻⁴ Hoffman^{5,6} or our recently described Leukart-Wallach reductive amination procedure⁷ (Scheme S1).

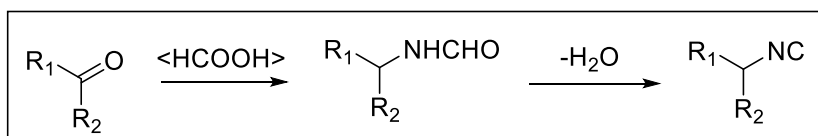
Ugi Method



Hoffmann Method



Leukart Wallach Reductive Amination Method



Scheme S1. Isocyanide Syntheses.

Other reagents were purchased from Sigma Aldrich, ABCR, Acros and AK Scientific and were used without further purification. Nuclear magnetic resonance spectra (NMR) were recorded on a Bruker Avance 500 spectrometer (¹H NMR (500 MHz), ¹³C NMR (126 MHz)). Chemical shifts for ¹H NMR were reported as δ values and coupling constants were in hertz (Hz). The following abbreviations were used for spin multiplicity: s = singlet, bs = broad singlet, d = doublet, t = triplet, q = quartet, m = multiplet. Chemical shifts for ¹³C NMR reported in ppm relative to the solvent peak. Flash chromatography was performed using RediSep R_f Normal-phase Silica Flash Columns (Silica Gel 60 Å, 230-400 mesh). Electrospray ionization mass spectra (ESI-MS) were recorded on a Waters Investigator Semi-prep 15 SFC-MS instrument. High resolution mass spectra were recorded using an LTQ-Orbitrap-XL (Thermo Fisher Scientific; ESI⁺ mode) at a resolution of 60000@m/z400.

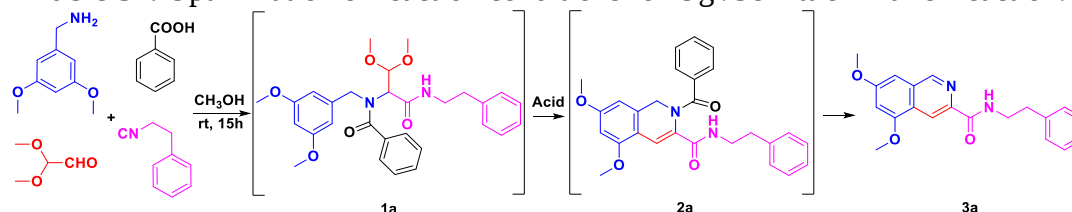
Analytical chemicals and reagents: MeOH (technical grade), CH₃CN (HPLC gradient grade), dioxane (99+% extra pure, stabilized), ethylene glycol (99+% extra pure) and fuming HCl (37%, for analysis). Formic acid (≥98.0%), acetic acid (≥99.8%), trifluoroacetic acid (≥99.0%), methanesulfonic acid (≥99.0%), benzoic acid (≥99.5%), 2,2-dimethoxyacetaldehyde (60 wt. % in H₂O), 3,4,5-trimethoxybenzylamine (98%), 3,4-dimethoxybenzylamine (97%), piperonylamine (97%), 2,3-dimethoxybenzylamine (99%), 2,5-dimethoxybenzylamine (97%).

2. Optimization of the novel isoquinoline reaction

In a model reaction, (3,5-dimethoxyphenyl)methane amine (1 mmol), 2,2-dimethoxy acetaldehyde (1 mmol), benzoic acid (1 mmol) and phenylethyl isocyanide (1 mmol) were stirred at room temperature in MeOH (1 M) for 15 h. Upon completion (TLC control), solvent was removed under vacuum. The Ugi adduct **1a** was directly used without any purification in the acid-catalyzed cyclisation/oxidation reaction.

In order to optimize the cyclisation/oxidation reaction, various acidic conditions (1 mL) were screened (Table S1). The desired product **3a** was formed in 68% yield in the presence of 37% HCl_(aq) solution in dioxane (1 mL, 1:1, v/v) as solvent at room temperature (Table S1, entry 10).

Table S1. Optimization of reaction conditions for Ugi/Schlittler-Müller reaction.



Entry	Solvent	Isolated yield (%)
1	HCOOH	0
2	CH ₃ COOH	0
3	CF ₃ COOH	0
4	CH ₃ SO ₃ H (10 eq)/ CH ₃ CN	0
5	CH ₃ SO ₃ H	0
6	CH ₃ COOH/conc. H ₂ SO ₄ (2:1, v/v)	33
7	CH ₃ COOH/ conc. H ₂ SO ₄ (1:1, v/v)	26
8	HCl _(aq) / dioxane (1:4, v/v)	34
9	HCl _(aq) / dioxane (1:2, v/v)	43
10	HCl_(aq)/ dioxane (1:1, v/v)	68
11	HCl _(aq)	52

3. General workflow for nanoscale synthesis

The general method typically follows the steps summarized in Table S2, with the implicit details for each step provided in the following text.

Table S2. General workflow.

No.	Step	Short Description
1	Stock solution preparation	Stock solutions were prepared at 0.5 M in ethylene glycol or 2-methoxy ethanol and sealed and kept at -20 °C.
2	Source plate preparation	The stock solutions were pipetted into the 384 well source plates.
3	Nanoscale synthesis	Sequence tables and methods were loaded into Echo 555 software and automatic transfer of the reagents started.
4	QC by SFC	384 Well synthesis plate was diluted with methanol and SFC-MS analytic performed with autosampler.

5	QC by TLC	384 Well synthesis plate was diluted with methanol and spotted onto silica TLC plates and eluted. The UV active spots were eluted and transferred into the Advion desktop MS.
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4. Nanomole-scale chemical reactions

4.1. General materials

Stock solutions were prepared in glass flat bottom vials (Screening devices, Catalog#: 9920-812FBT, 2.0 mL (Topas) Plate) and they were kept at -20 °C.

Nanomole-scale chemistry was performed using Echo qualified 384-well polypropylene microplate (Labcyte, Catalog#: PP-0200, clear, flat bottom) according to the producers' manual.

384-Well source and destination plates were sealed by a sealing tape (Thermo Scientific, Catalog#: 232701, polyolefin acrylate) and were kept at -20 °C.

4.2. Instrumentation

The Echo 555 liquid handler (Labcyte) was used in order to transfer nL droplets of starting materials from the 384-well source plate to the 384-well destination plate.

4.3. Nanomole-scale automated chemistry

Stock solutions of aldehyde, acid, amines (A1-A7) and isocyanides (I1-I3, I6-I8, I10-16, I18, I19, I21, I28, I29, I31, I34, I35, I42-I44, I48, I50-I62) were prepared as 0.5 M ethylene glycole. Due to the insolubility of some isocyanides (I4, I5, I9, I17, I20, I22-I27, I30, I32, I33, I36-I41, I45-I47, I49) in ethylene glycole, their stock solutions were instead prepared as 0.5 M in 2-methoxyethanol.

The stock solutions were dispensed to a 384-well source plate using Eppendorf multi-channel pipettes. The Echo 555 was used to transfer 750 nL (375 nmol) of each starting material into the corresponding well in the destination plate. Labcyte Echo plate reformat software using custom mapping mode with the run protocol as defined by a pick list was used (Fig. S1B).

In order to generate a random library of products (N=384), a modified version of our previously reported program RandReactor was used.⁸ The smiles files of the starting materials with the corresponding location in the source plate and mrv file of reaction were the input of the RandReactor program. The smiles file of the randomly generated products with their corresponding locations in the source and destination plate were the output of the RandReactor program. The smiles file was converted to a csv file which was the required format for Labcyte Echo plate reformat software (Fig. S1A).

Once the starting materials transfer was completed (~150 min), the destination plate was covered with the sealing film and was then placed for 15 h at 23 °C on an orbital shaker. Then, 10 µL of 37% HCl_(aq)/dioxane solution (1:1, v/v) was added to each well using a multichannel

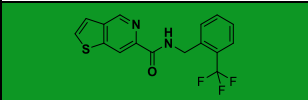
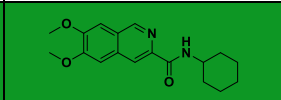
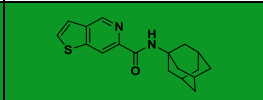
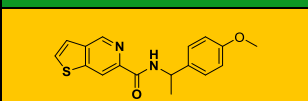
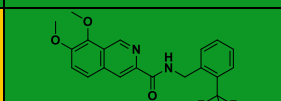
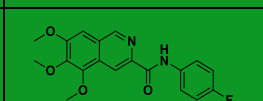
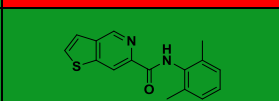
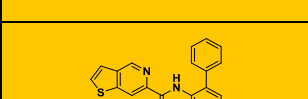
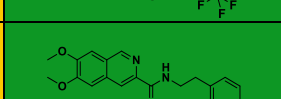
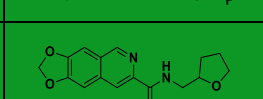
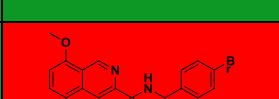
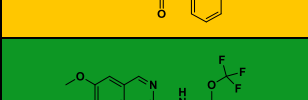
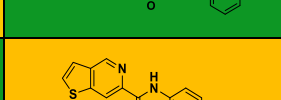
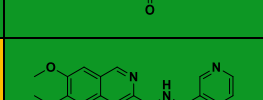
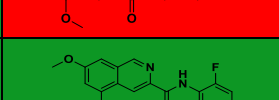
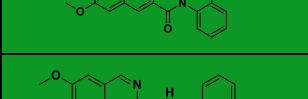
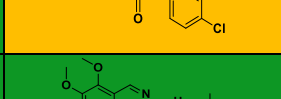
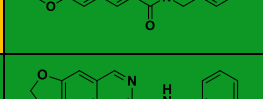
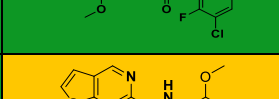
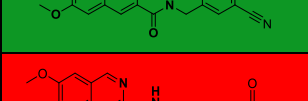
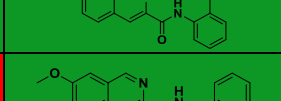
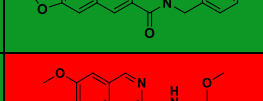
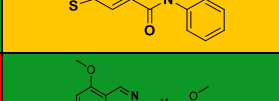
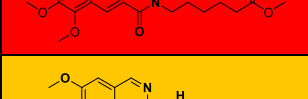
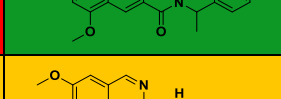
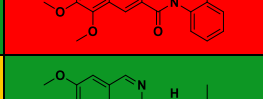
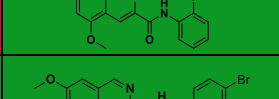
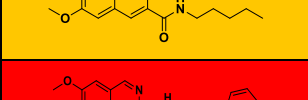
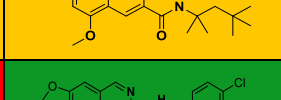
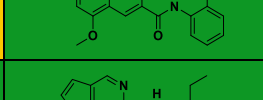
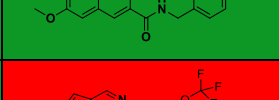
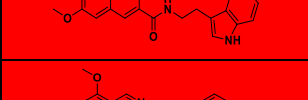
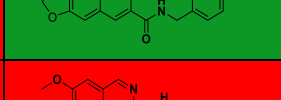
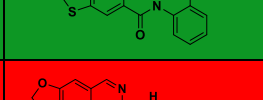
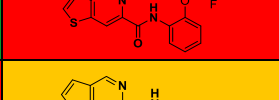
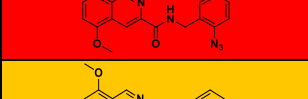
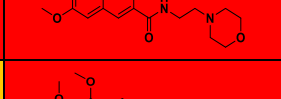
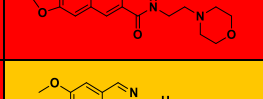
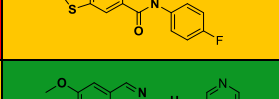
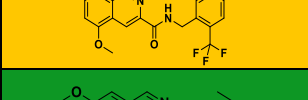
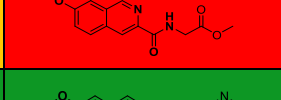
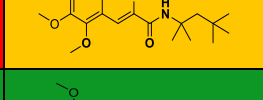
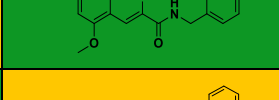
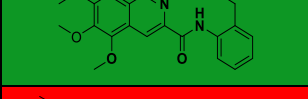
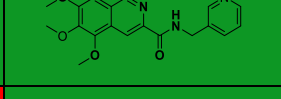
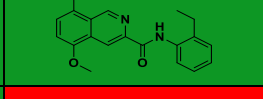
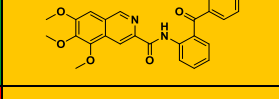
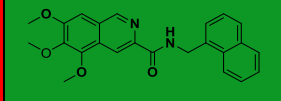
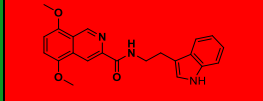
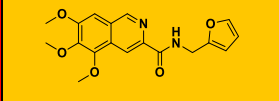
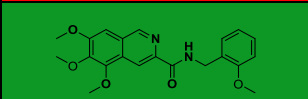
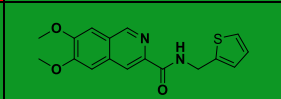
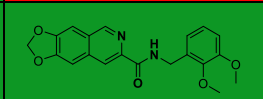
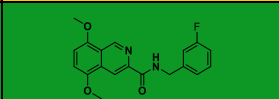
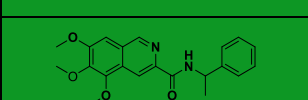
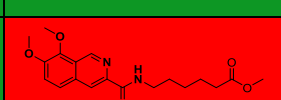
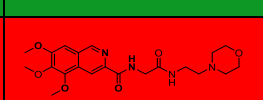
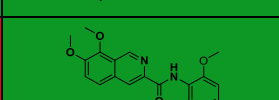
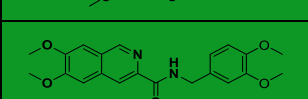
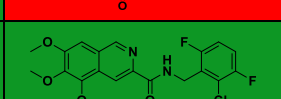
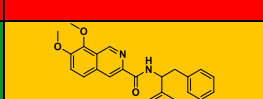
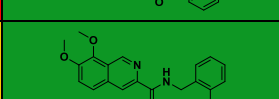
pipettor and the plate was sealed and kept at 23 °C for another 12 h. Then the plate was dried from the solvent by applying a mild stream of nitrogen. The plates were sealed and stored at -20 °C for further processing. The structures of the products are shown in Fig. S2.

A

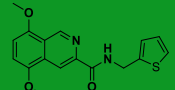
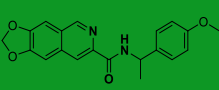
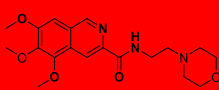
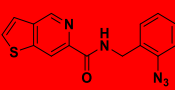
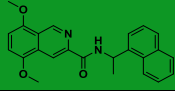
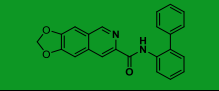
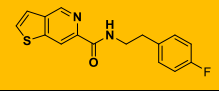
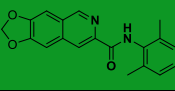
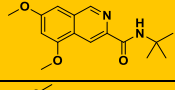
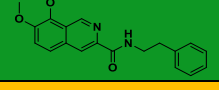
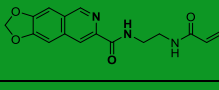
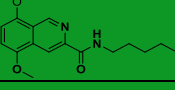
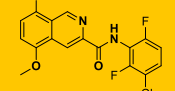
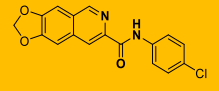
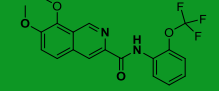
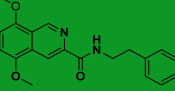
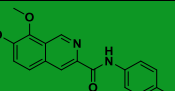
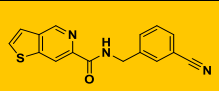
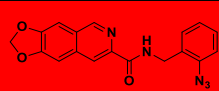
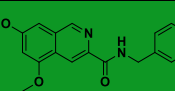
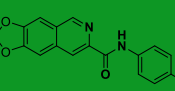
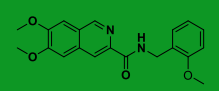
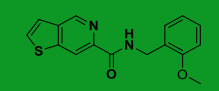
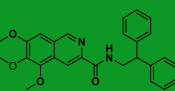
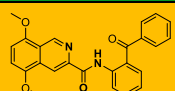
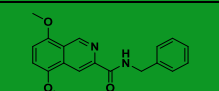
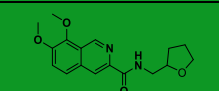
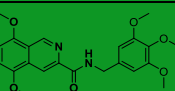
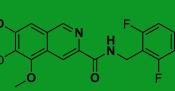
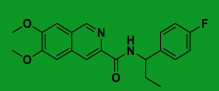
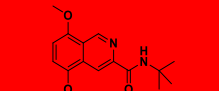
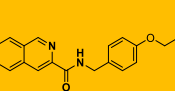
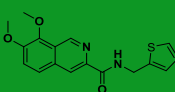
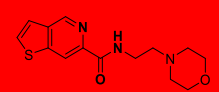
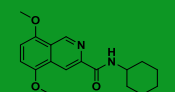
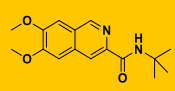
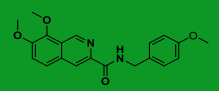
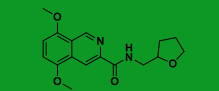
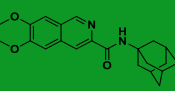
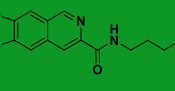
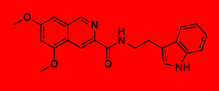
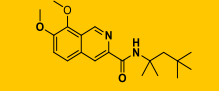
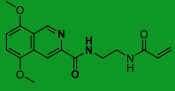
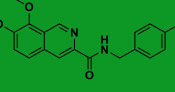
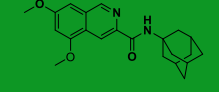
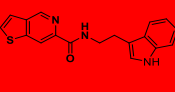
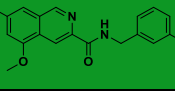
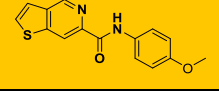
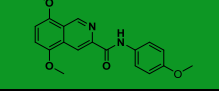
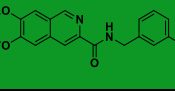
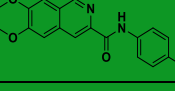
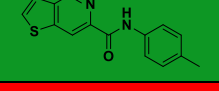
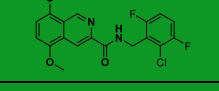
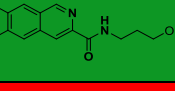
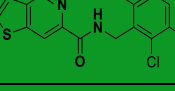
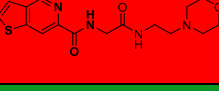
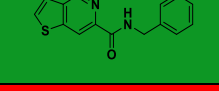
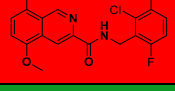
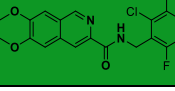
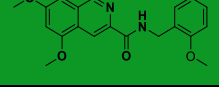
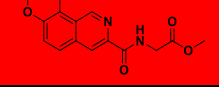
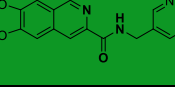
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3	Sample Plate 1	A1	Destination 1	A1	750						
4	Sample Plate 1	A2	Destination 1	A1	750						
5	Sample Plate 1	B14	Destination 1	A1	750						
6	Sample Plate 1	F20	Destination 1	A2	750						
7	Sample Plate 1	A1	Destination 1	A2	750						
8	Sample Plate 1	A2	Destination 1	A2	750						
9	Sample Plate 1	A17	Destination 1	A2	750						
10	Sample Plate 1	H20	Destination 1	A3	750						
11	Sample Plate 1	A1	Destination 1	A3	750						
12	Sample Plate 1	A2	Destination 1	A3	750						
13	Sample Plate 1	A17	Destination 1	A3	750						
14	Sample Plate 1	G19	Destination 1	A4	750						
15	Sample Plate 1	A1	Destination 1	A4	750						
16	Sample Plate 1	A2	Destination 1	A4	750						
17	Sample Plate 1	D16	Destination 1	A4	750						
18	Sample Plate 1	F20	Destination 1	A5	750						
19	Sample Plate 1	A1	Destination 1	A5	750						
20	Sample Plate 1	A2	Destination 1	A5	750						
21	Sample Plate 1	B14	Destination 1	A5	750						
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23	Sample Plate 1	A1	Destination 1	A6	750						

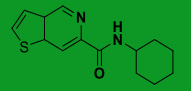
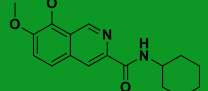
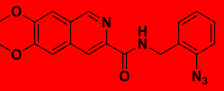
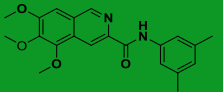
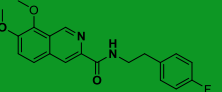
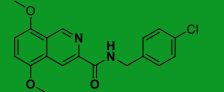
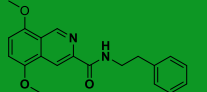
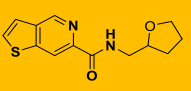
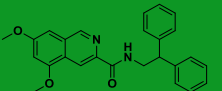
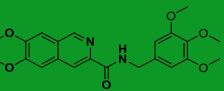
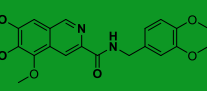
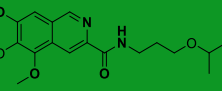
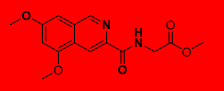
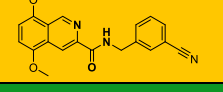
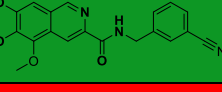
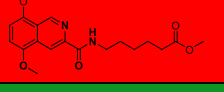
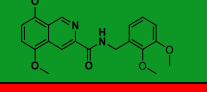
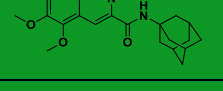
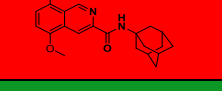
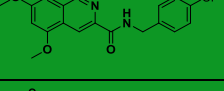
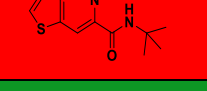
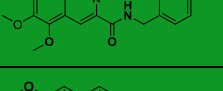
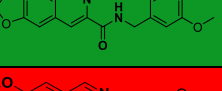
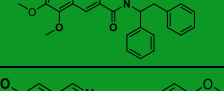
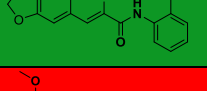
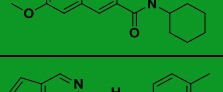
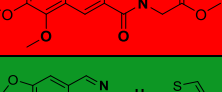
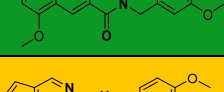
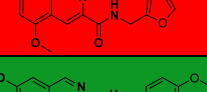
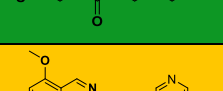
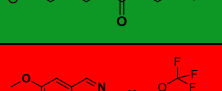
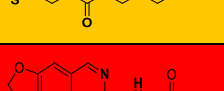
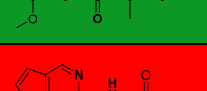
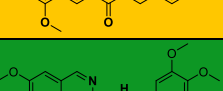
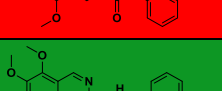
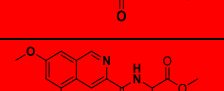
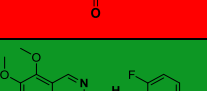
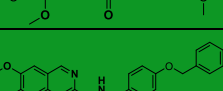
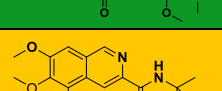
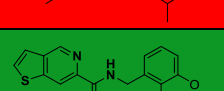
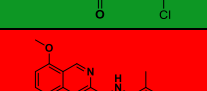
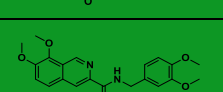
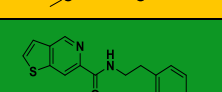
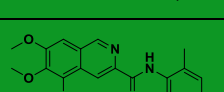
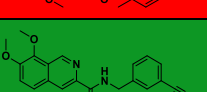
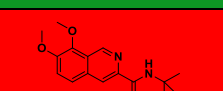
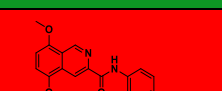
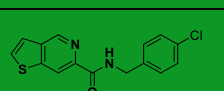
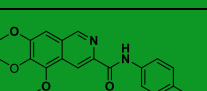
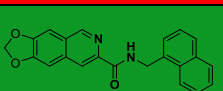
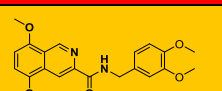
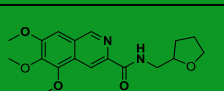
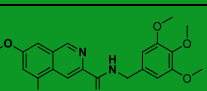
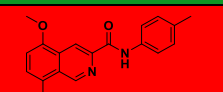
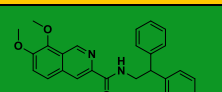
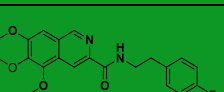
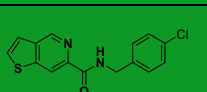
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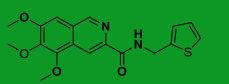
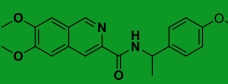
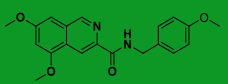
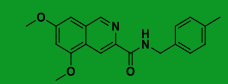
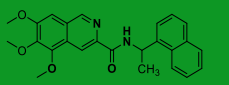
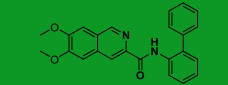
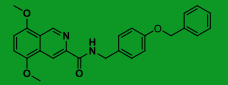
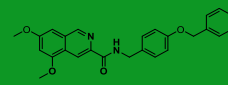
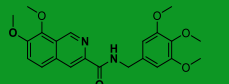
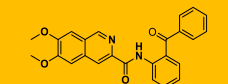
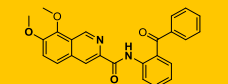
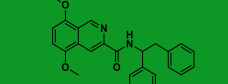
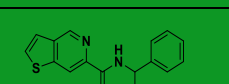
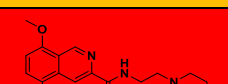
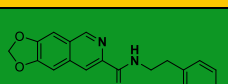
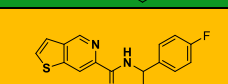
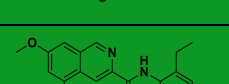
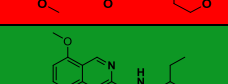
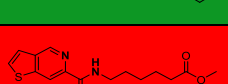
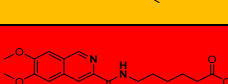
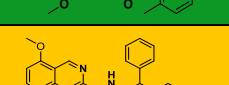
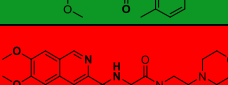
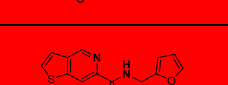
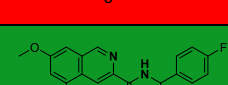
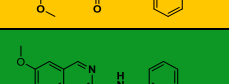
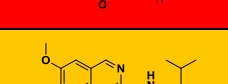
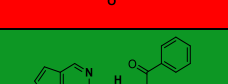
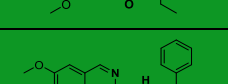
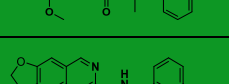
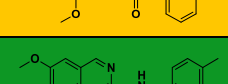
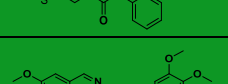
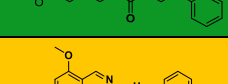
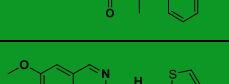
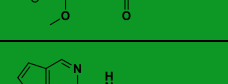
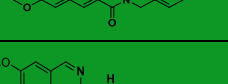
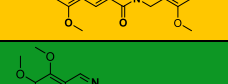
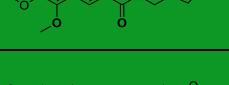
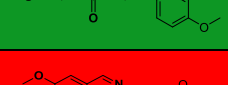
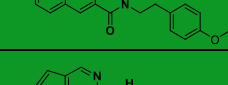
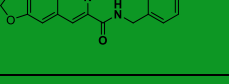
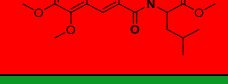
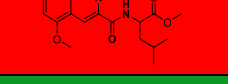
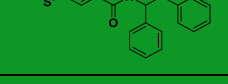
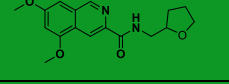
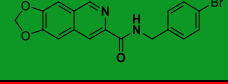
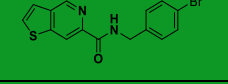
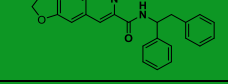
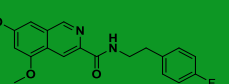
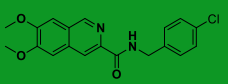
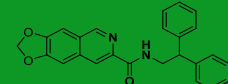
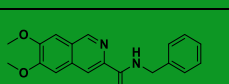
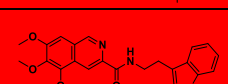
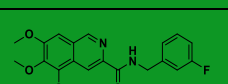
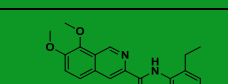
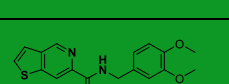
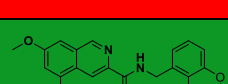
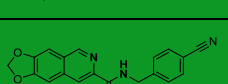
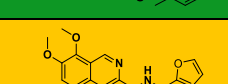
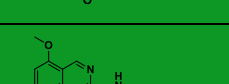
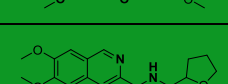
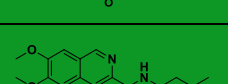
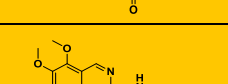
Fig. S1. A: Pick list in csv format required for Labcyte Echo plate reformat software; **B:** Labcyte Echo plate reformat software, showing on top the source plate and below the destination plate.

	1	2	3	4
A				
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	5	6	7	8
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	9	10	11	12
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K				
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M				
N				
O				
P				

	13	14	15	16
A				
B				
C				
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M				
N				
O				
P				

	17	18	19	20
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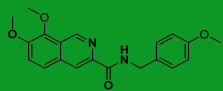
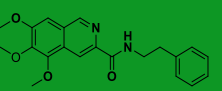
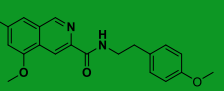
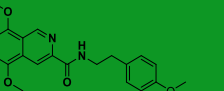
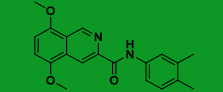
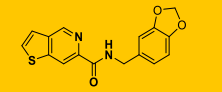
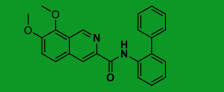
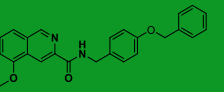
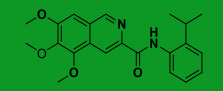
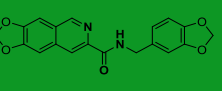
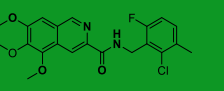
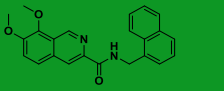
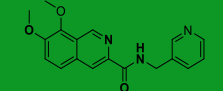
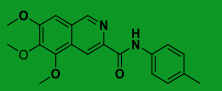
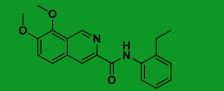
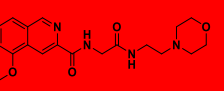
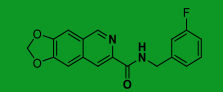
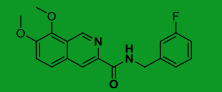
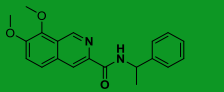
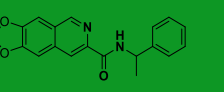
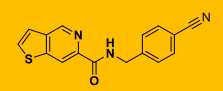
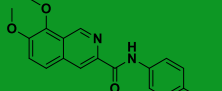
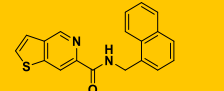
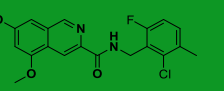
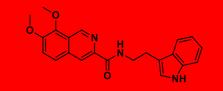
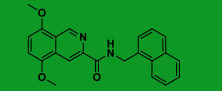
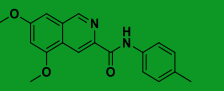
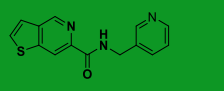
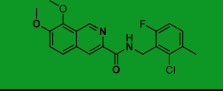
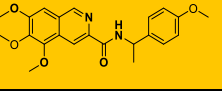
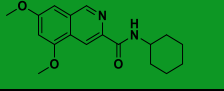
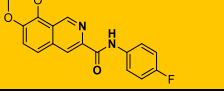
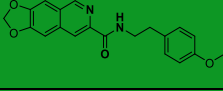
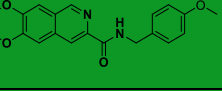
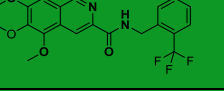
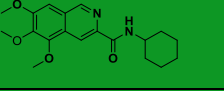
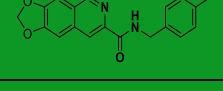
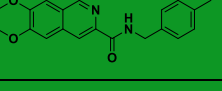
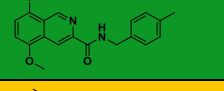
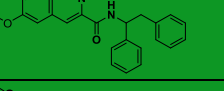
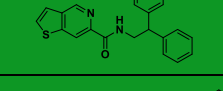
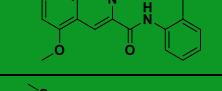
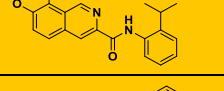
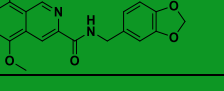
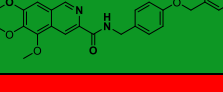
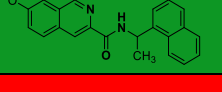
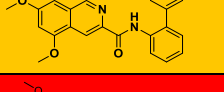
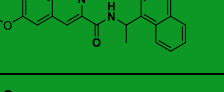
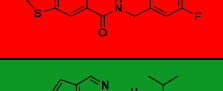
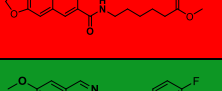
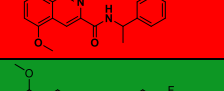
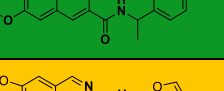
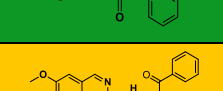
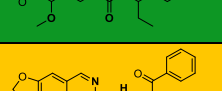
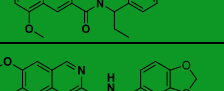
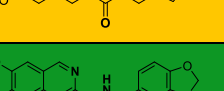
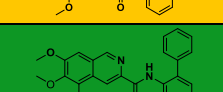
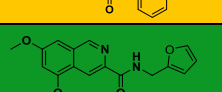
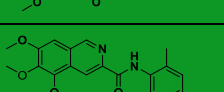
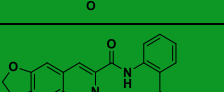
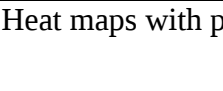
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Fig. S2. Heat maps with product structures.

4.4. Quality control (QC)

The analytics of all wells was performed by two complementary methods, SFC-UV-MS and TLC-UV-MS.

4.4.1. SFC-UV-MS analysis

Mass spectra were measured on a Waters Investigator Supercritical Fluid Chromatograph with a 3100 MS Detector (ESI⁺) on a Kromasil SFC-2.5-2EP (3.0 × 50 mm) column and MassLynx software.

Conditions; mobile phase: CO₂ with 35% MeOH (isocratic), run time: 1 min, flow rate: 4 mL/min, temperature: 40 °C, pressure: 120 bar.

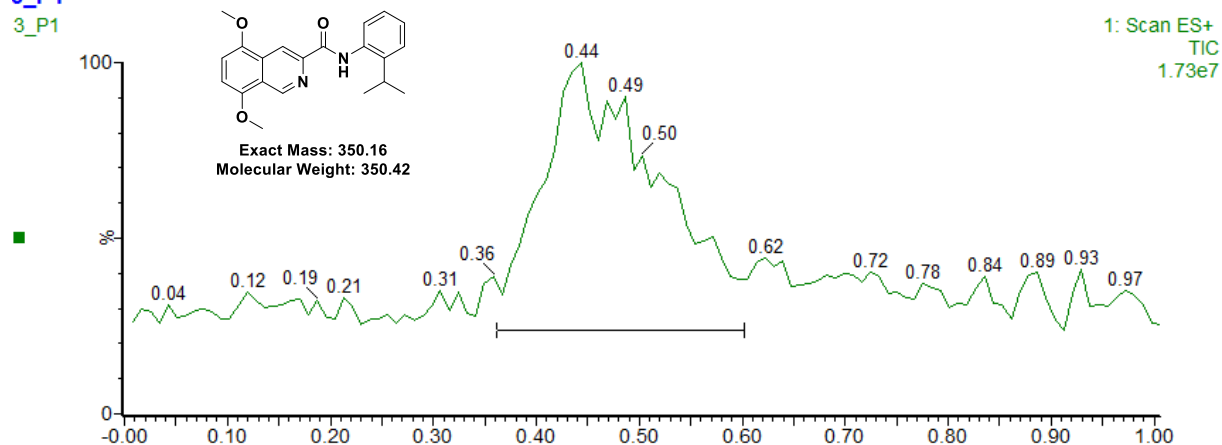
Each well of the destination plate was diluted with 100 µL MeOH and then the chromatographic analysis was done by SFC-UV-MS using an autosampler. A right-click and drag operation of the total ion current (TIC) spectrum generated a mass chromatogram for the selected range. If the peak corresponding to M+1 was the major peak, the well got a green designation and otherwise yellow. If the peak of M+1 was absent, the well got a red designation.

The SFC analytic of one well took ~1 min, resulting in an overall measuring time for the 384 well plate of less than one night.

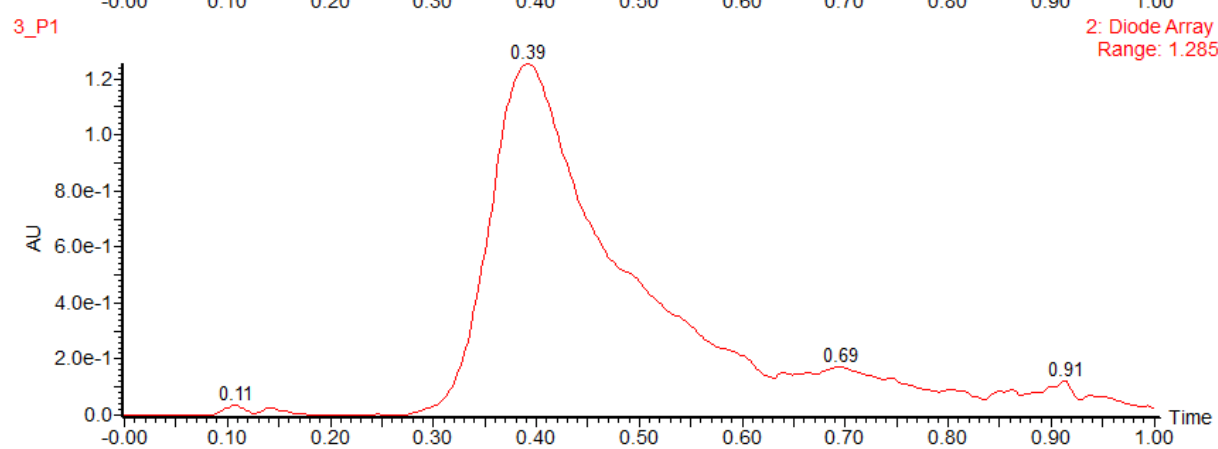
Examples of SFC-UV-MS analytics of the row P1-P24

3_P1

3_P1

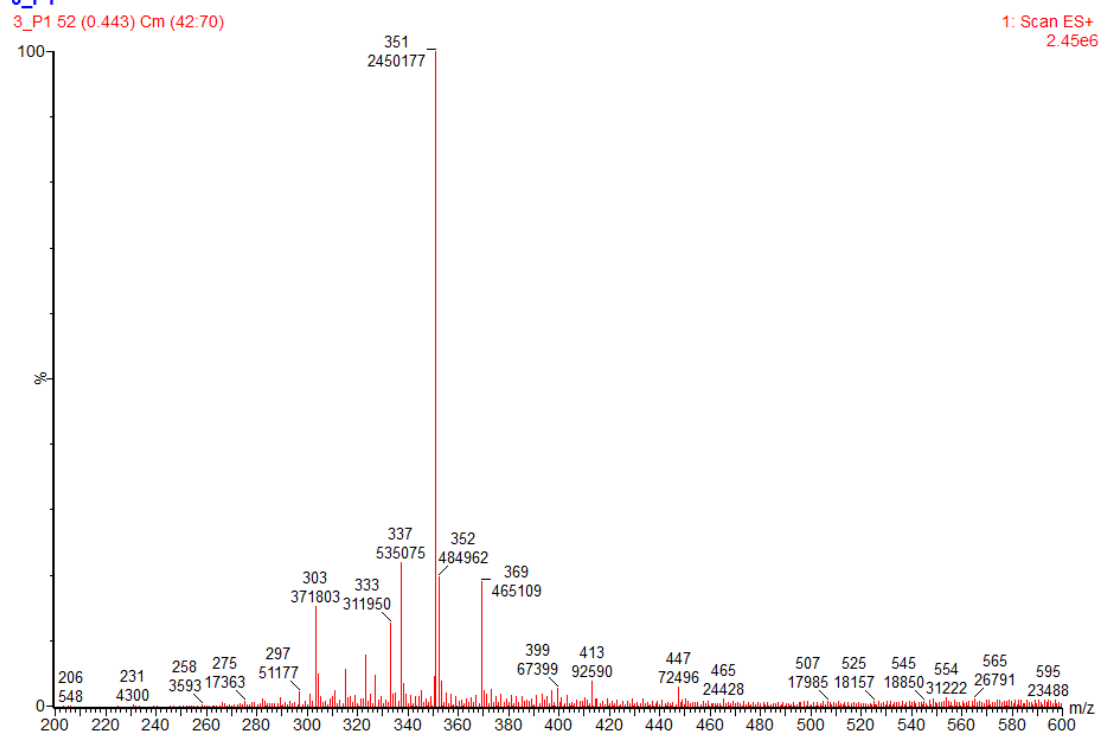


3_P1



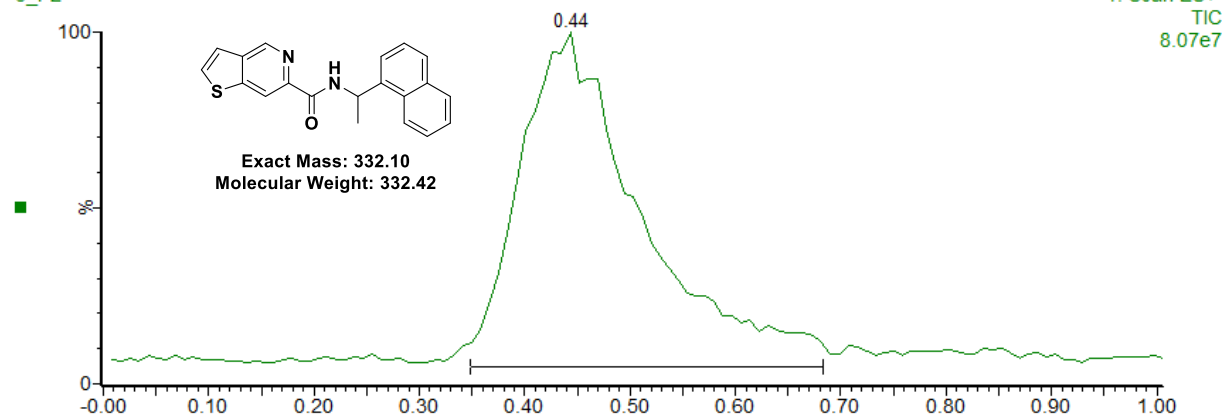
3_P1

3_P1 52 (0.443) Cm (42:70)

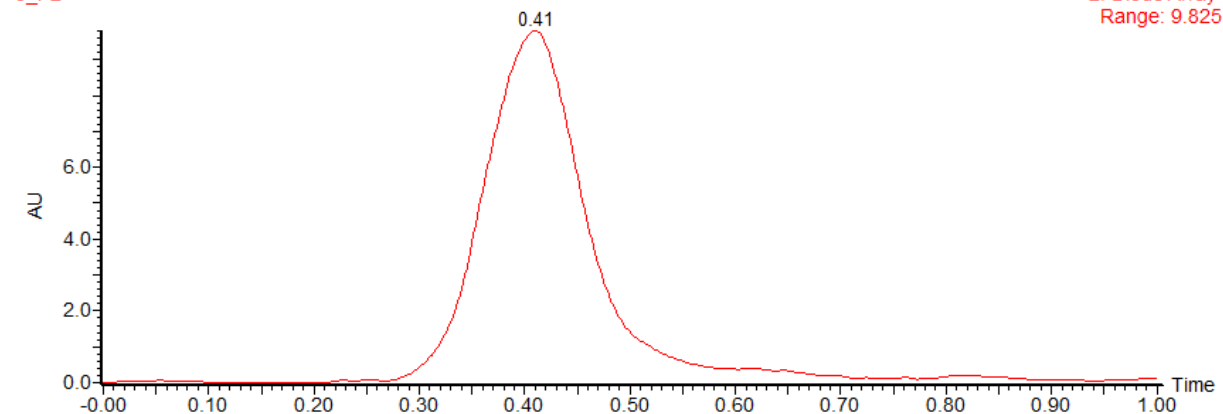


3_P2

3_P2

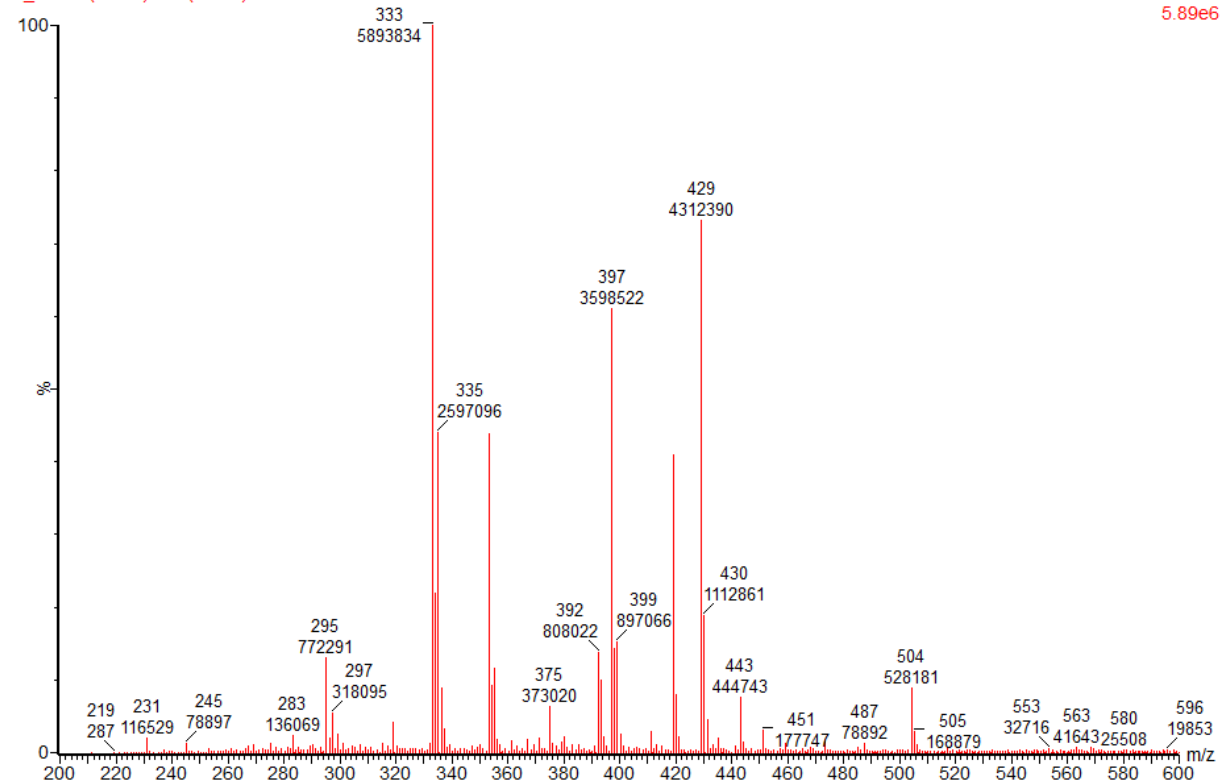


3_P2



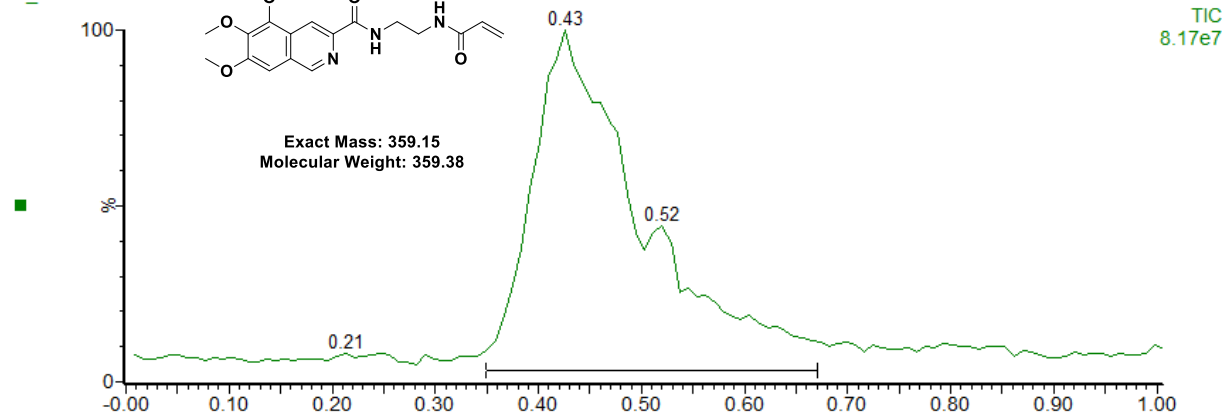
3_P2

3_P2 52 (0.443) Cm (41:69)

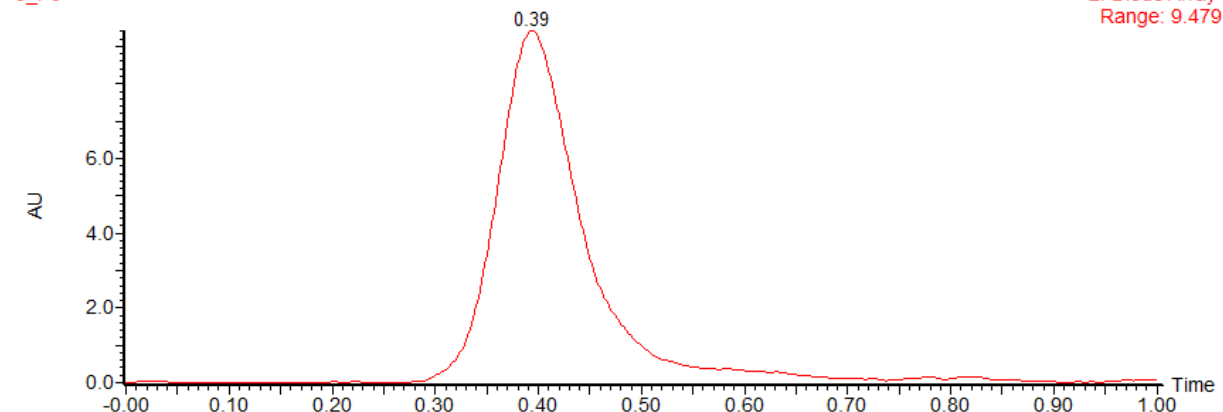


3_P3

3_P3

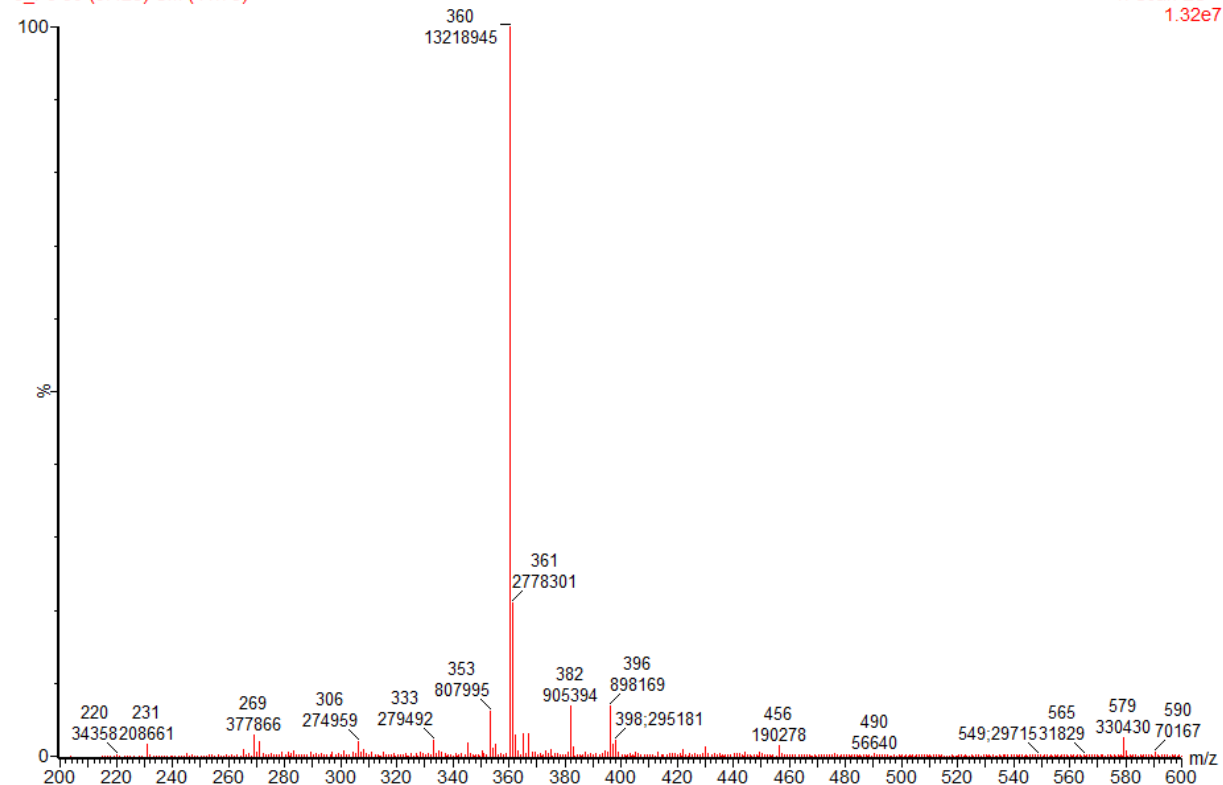


3_P3



3_P3

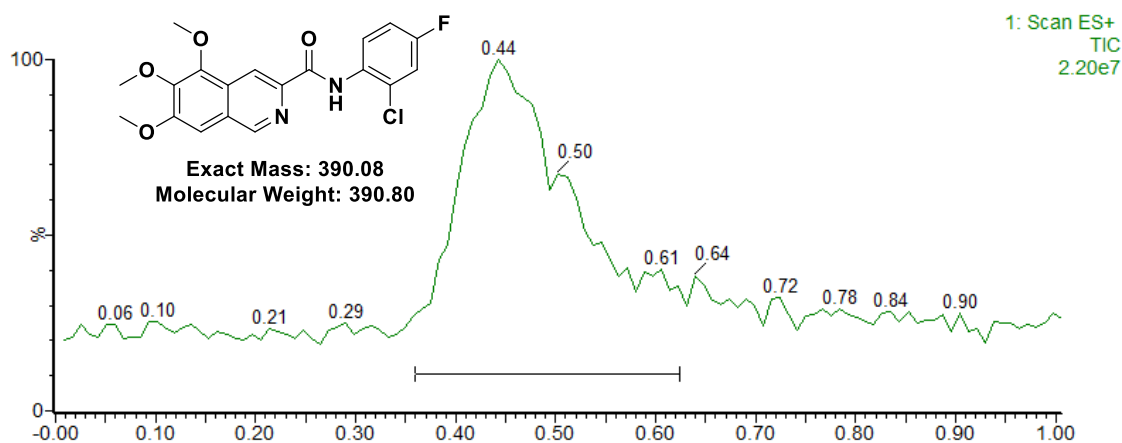
3_P3 50 (0.426) Cm (41:78)



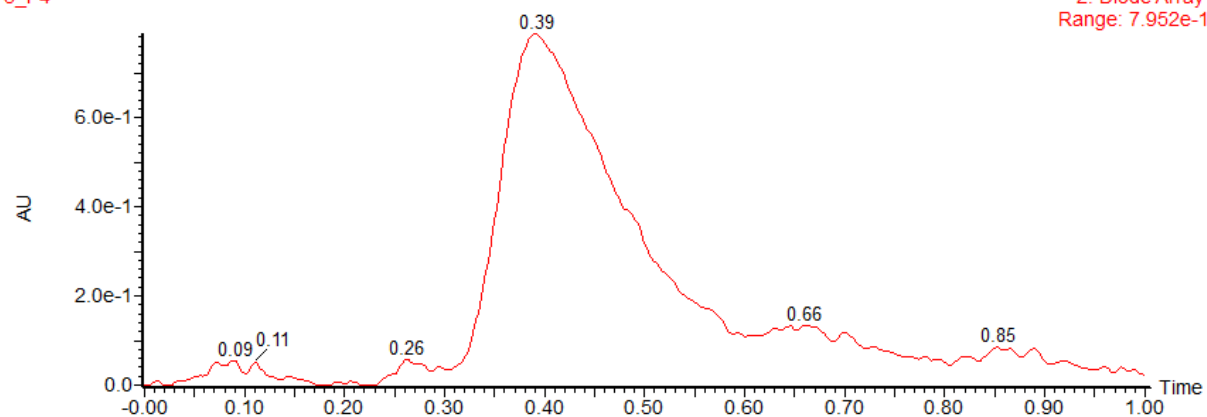
3_P4

3_P4

■

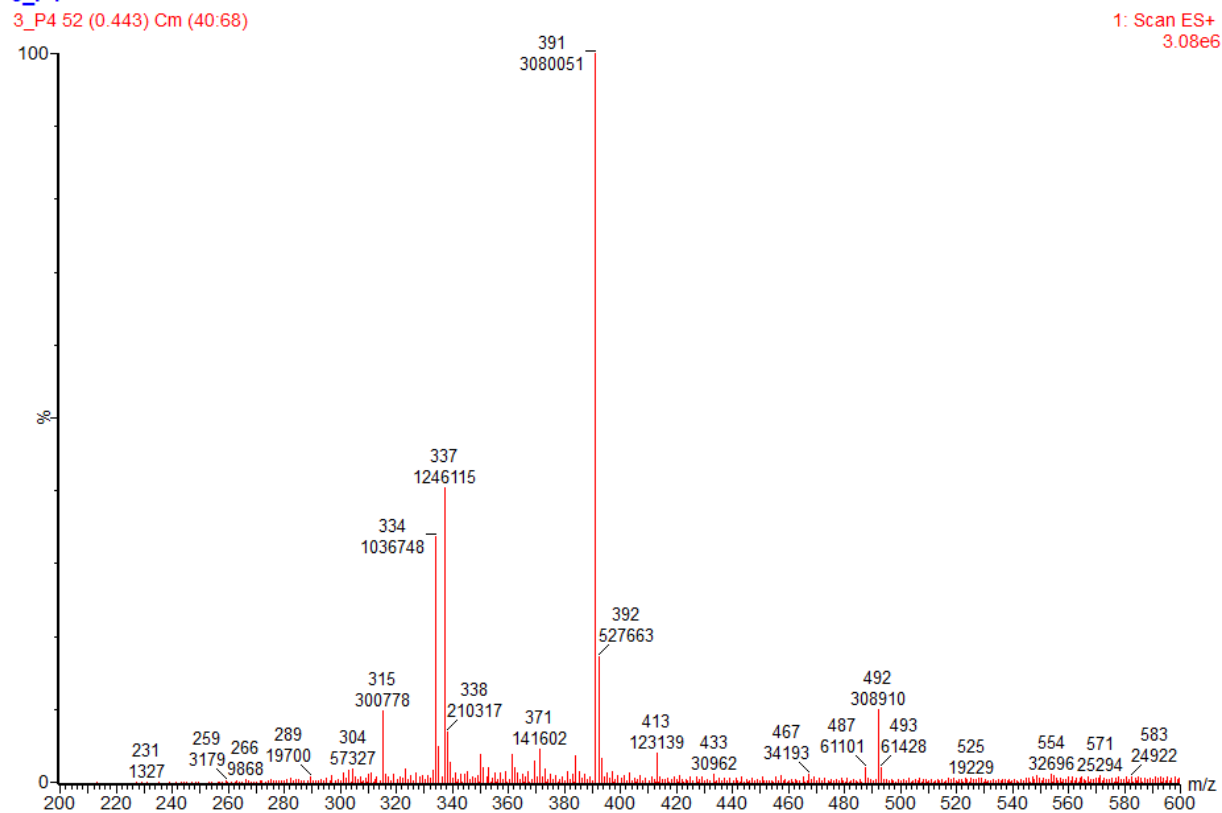


3_P4



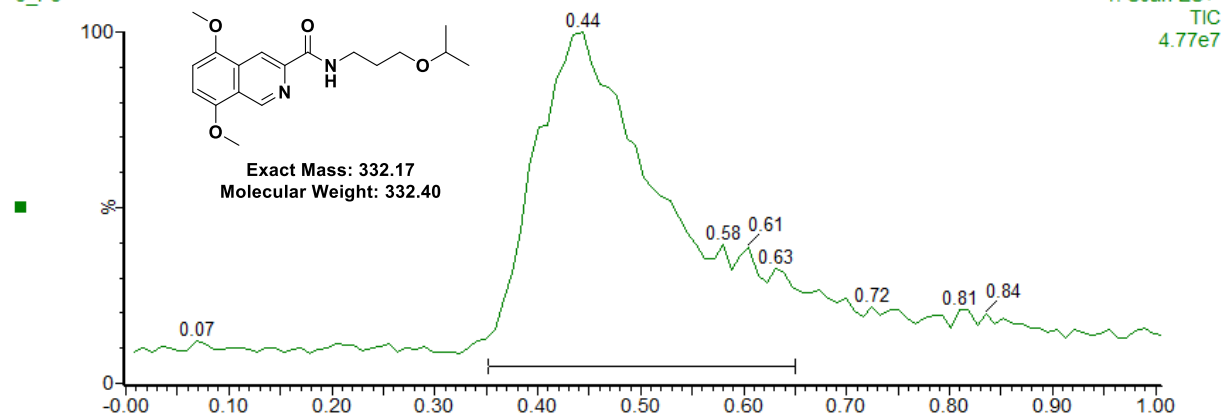
3_P4

3_P4 52 (0.443) Cm (40:68)

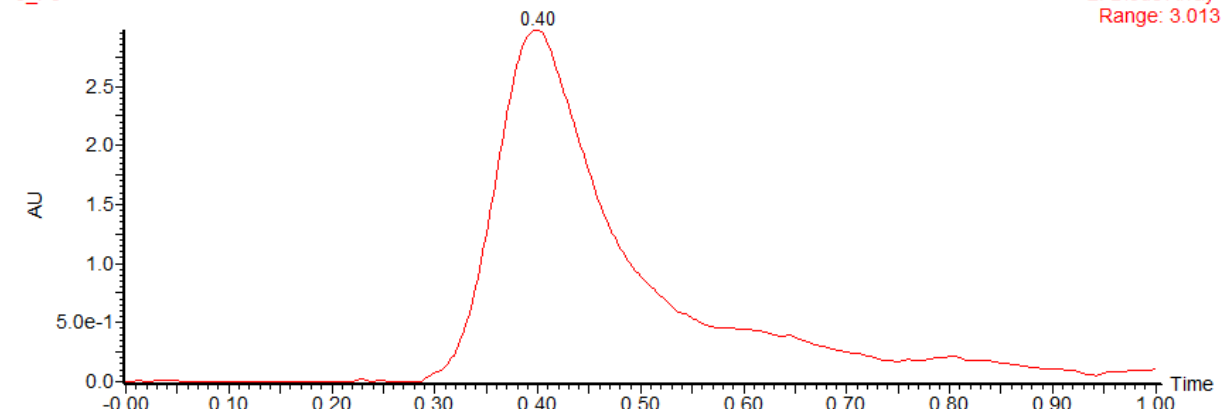


3_P5

3_P5

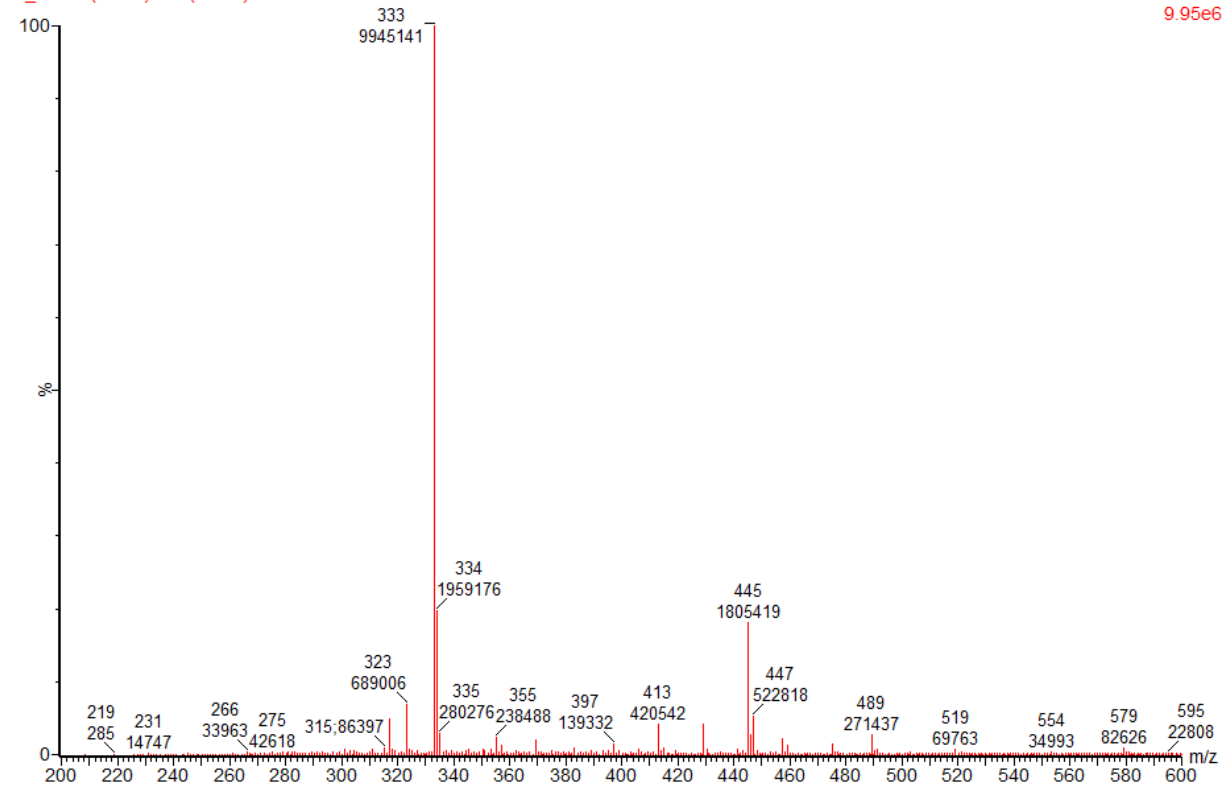


3_P5



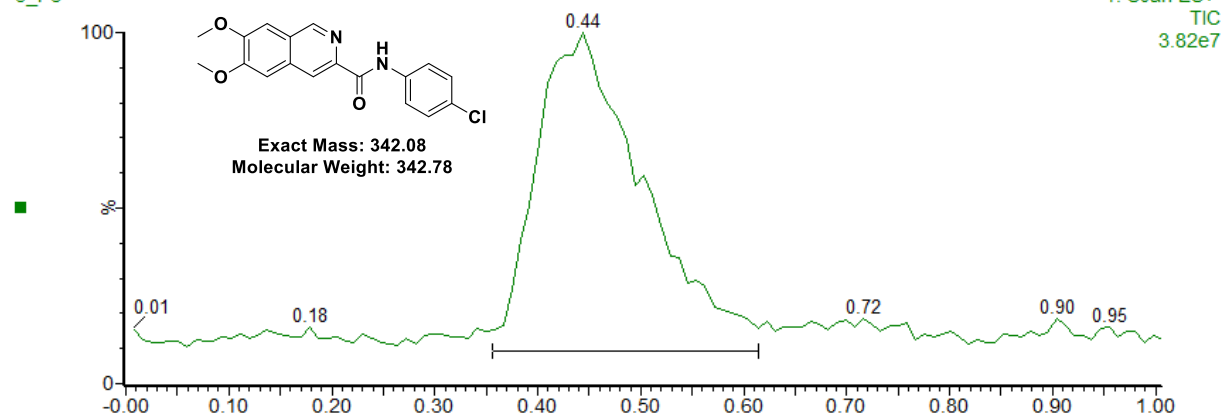
3_P5

3_P5 52 (0.443) Cm (42.76)

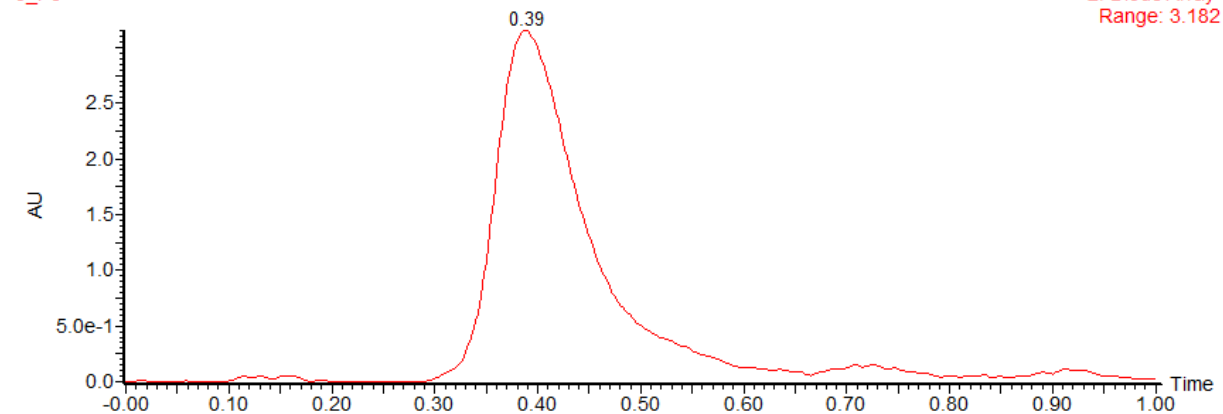


3_P6

3_P6

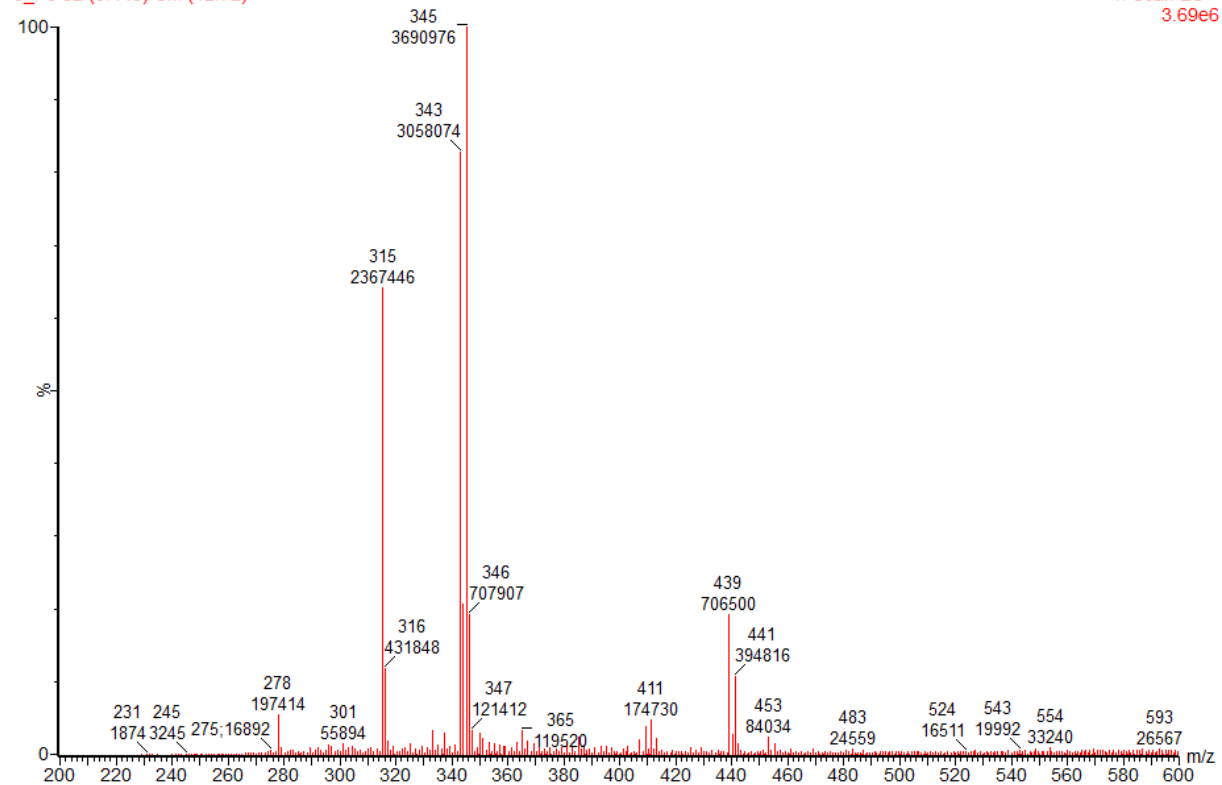


3_P6



3_P6

3_P6 52 (0.443) Cm (42:72)

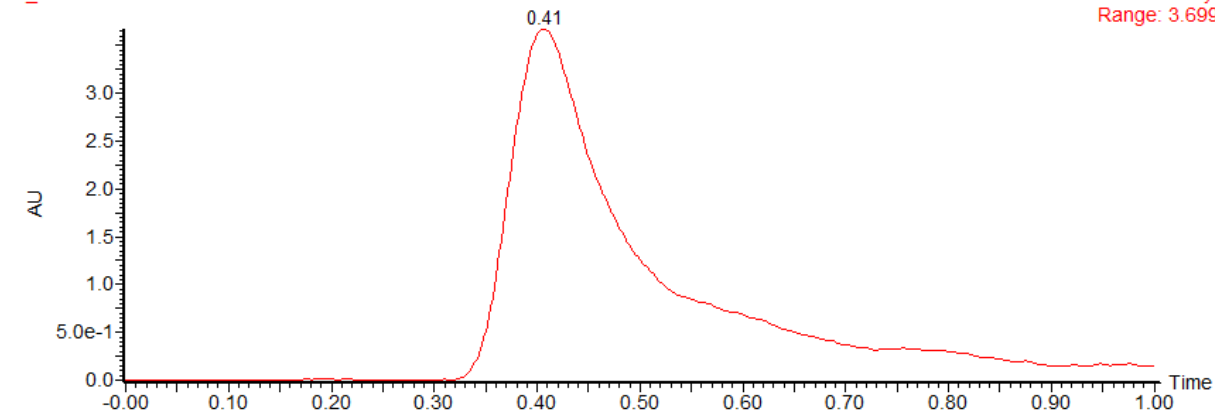


3_P7



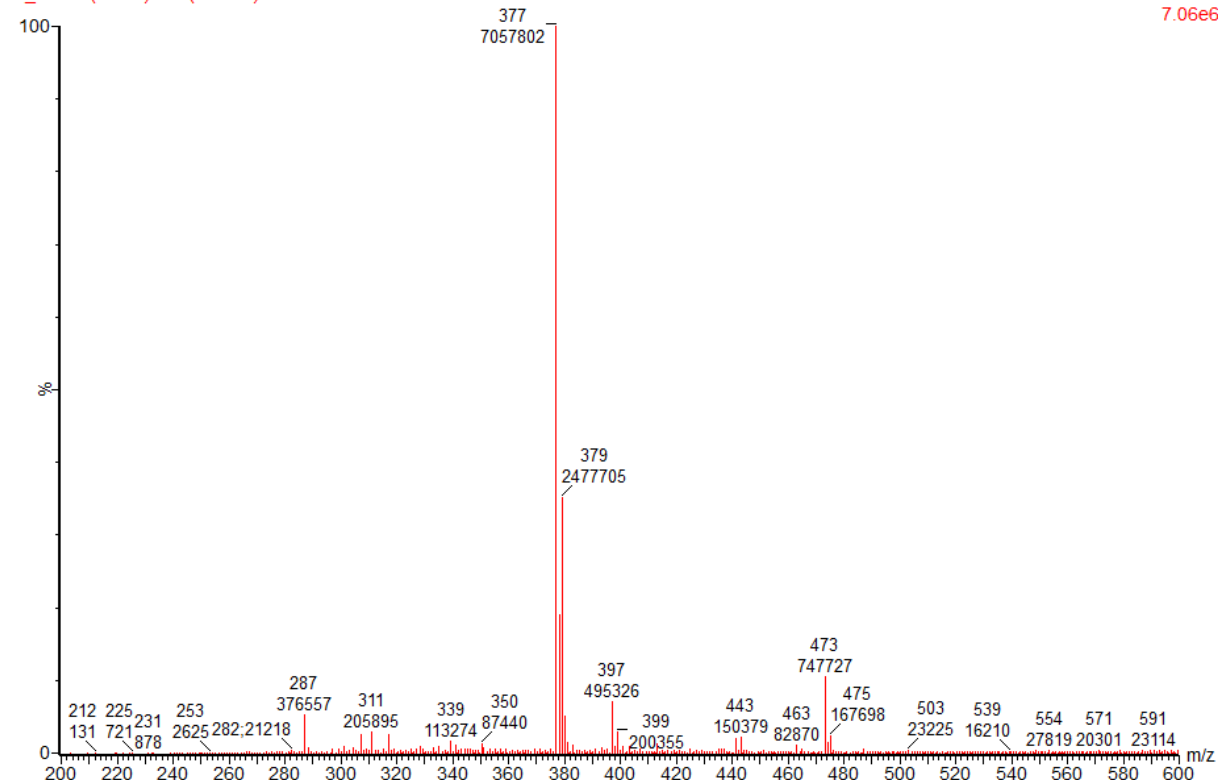
Exact Mass: 376,07
Molecular Weight: 376,29
TIC
4.39e7

2: Diode Array
Range: 3.699



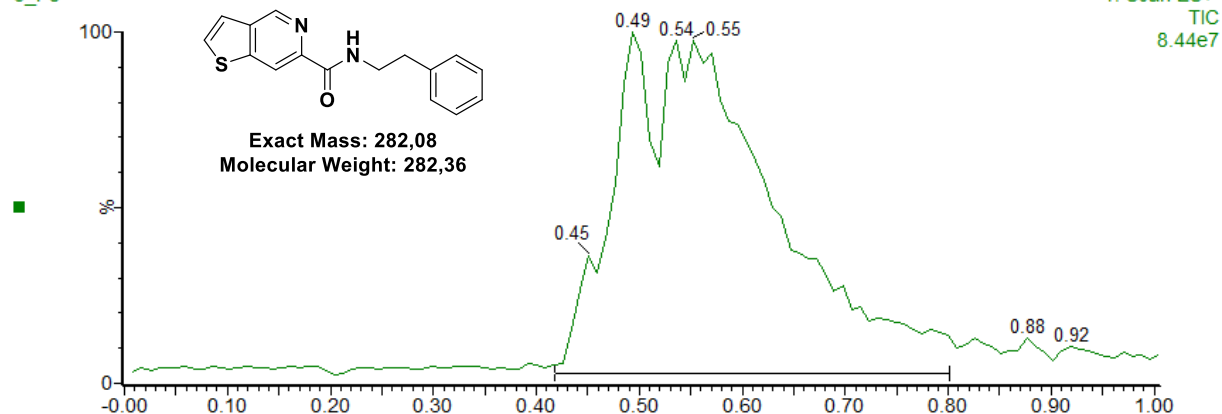
3_P7 52 (0.442) Cm (43:103)

1: Scan ES+
7.06e6

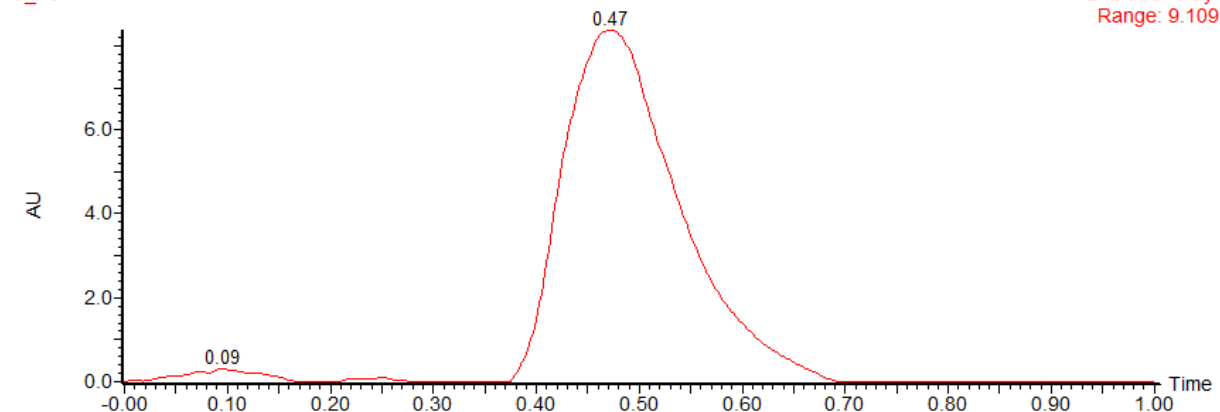


3_P8

3_P8

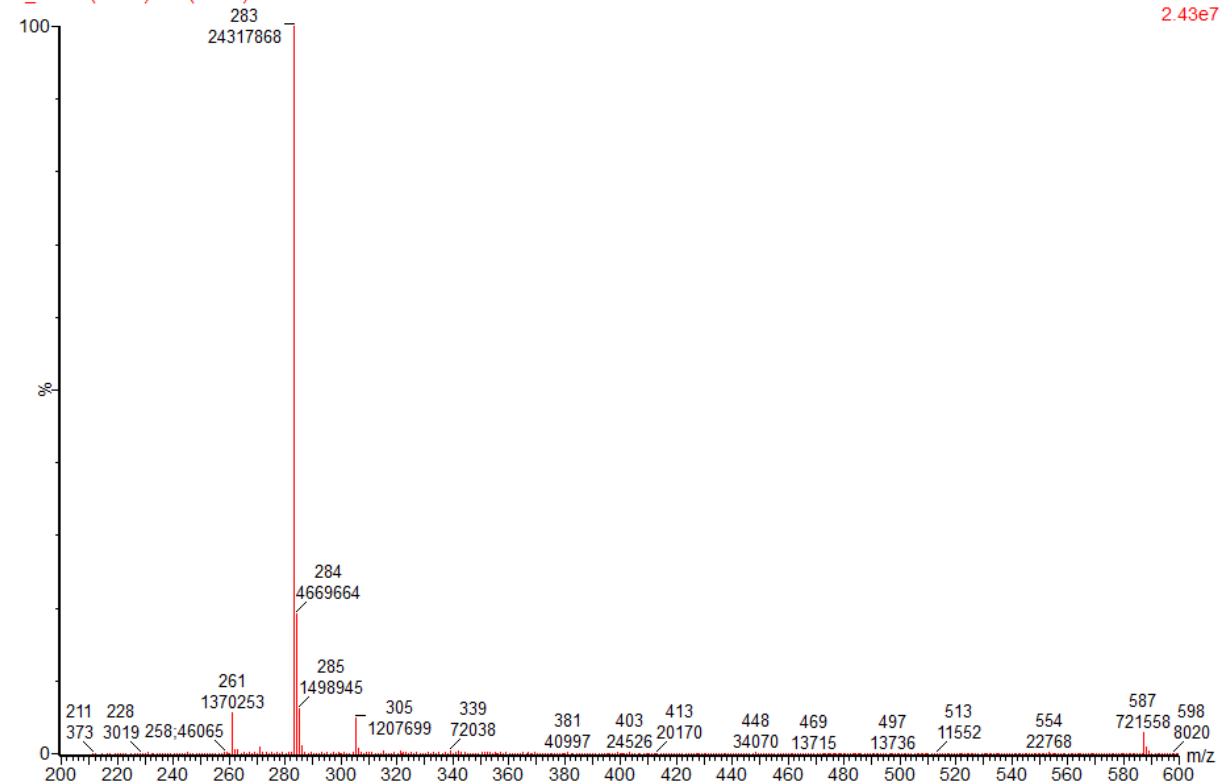


3_P8



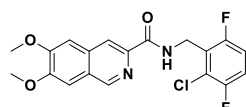
3_P8

3_P8 58 (0.493) Cm (49.93)



3_P9

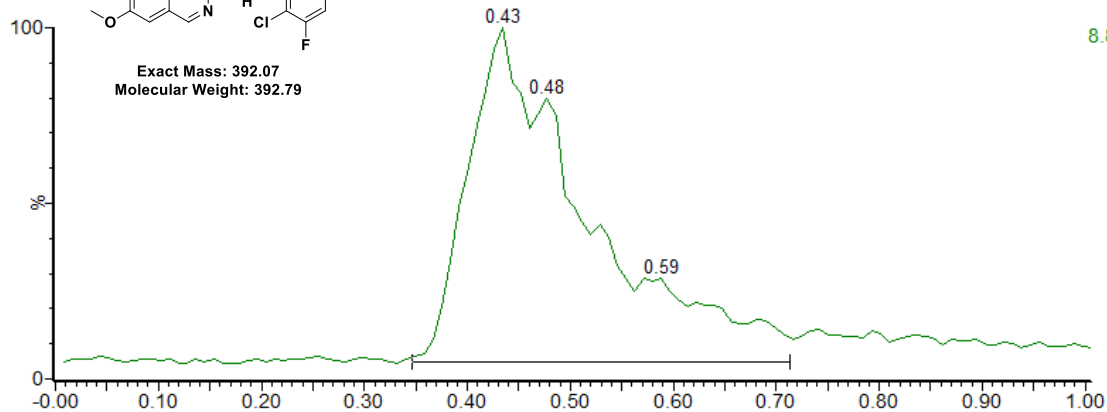
3_P9



Exact Mass: 392.07
Molecular Weight: 392.79

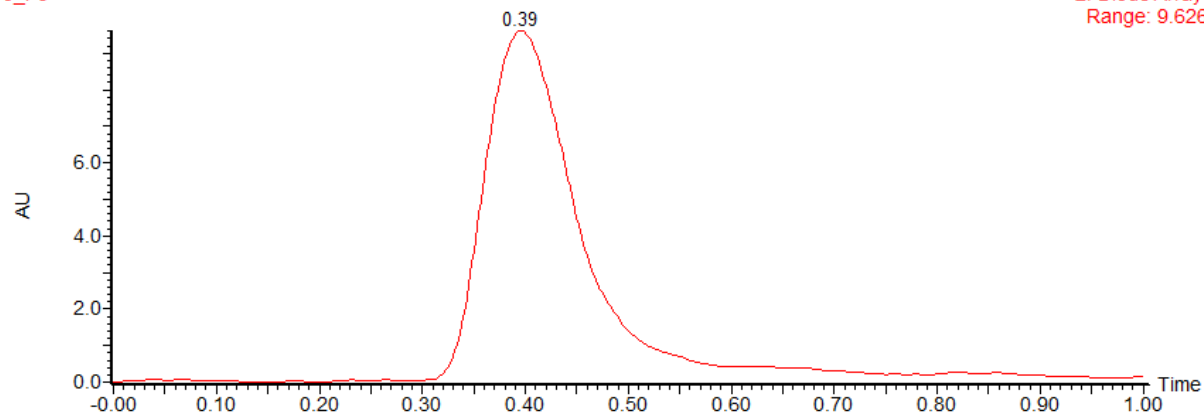
1: Scan ES+
TIC
8.81e7

■



3_P9

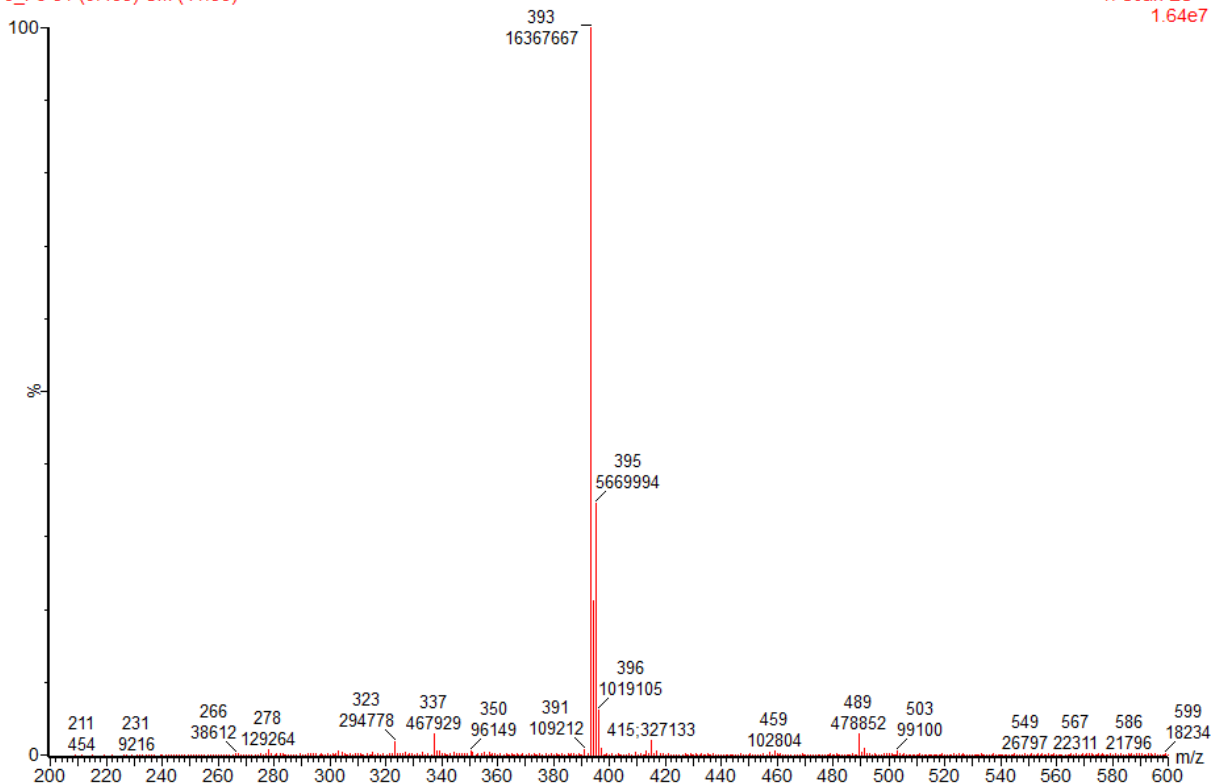
2: Diode Array
Range: 9.626



3_P9

3_P9 51 (0.435) Cm (41:83)

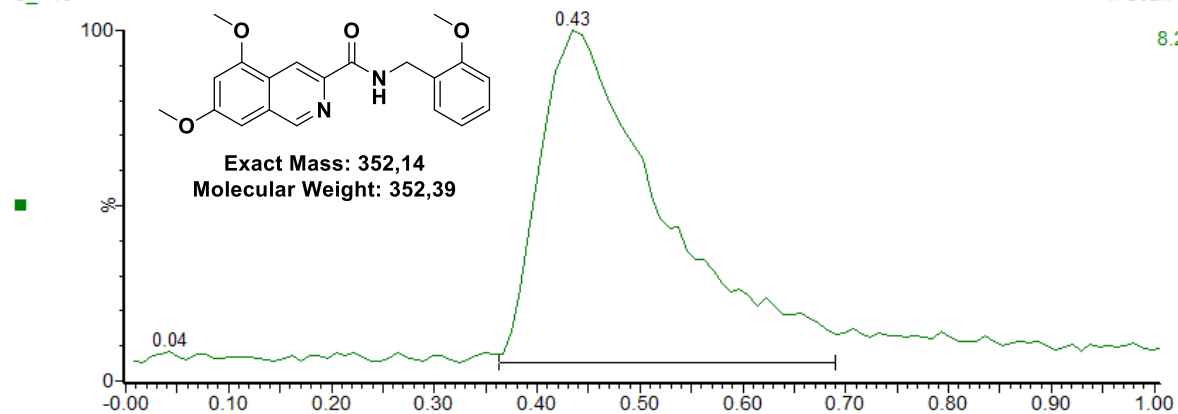
1: Scan ES+
1.64e7



3_P10

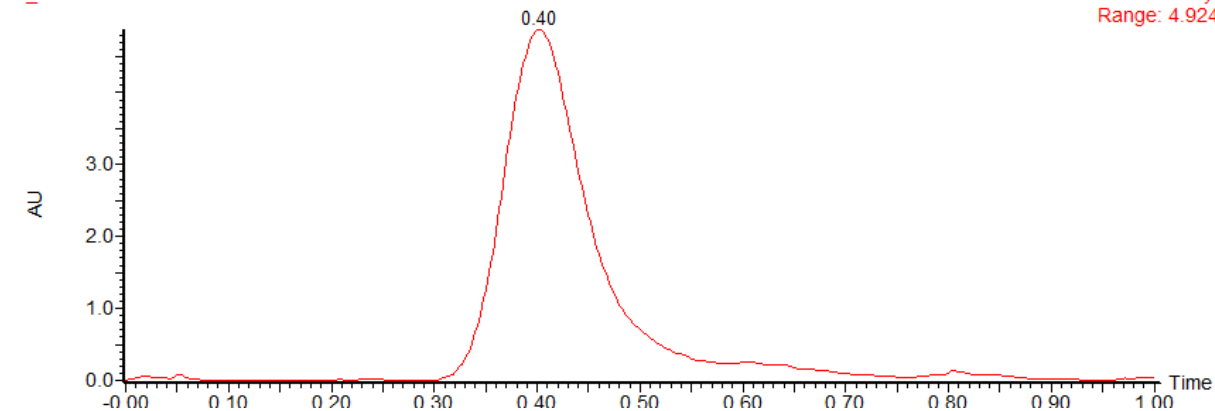
3_P10

1: Scan ES+
TIC
8.21e7



3_P10

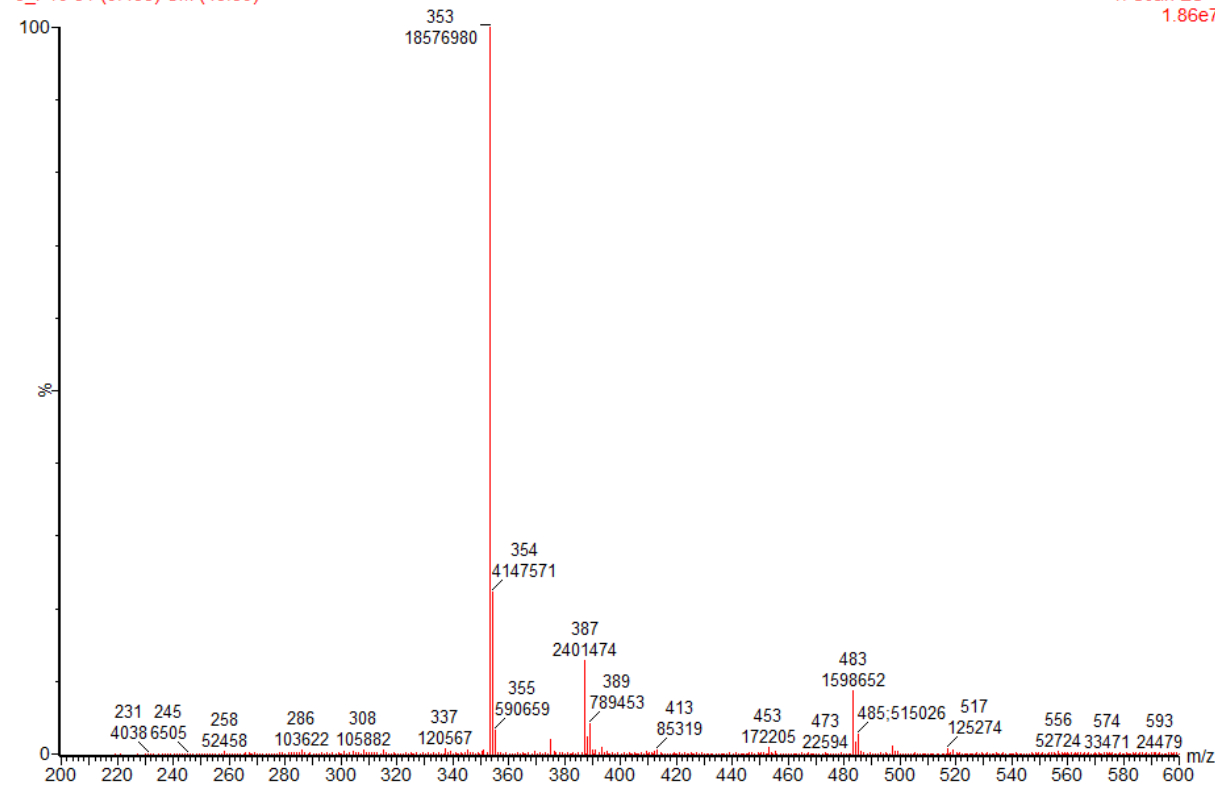
2: Diode Array
Range: 4.924



3_P10

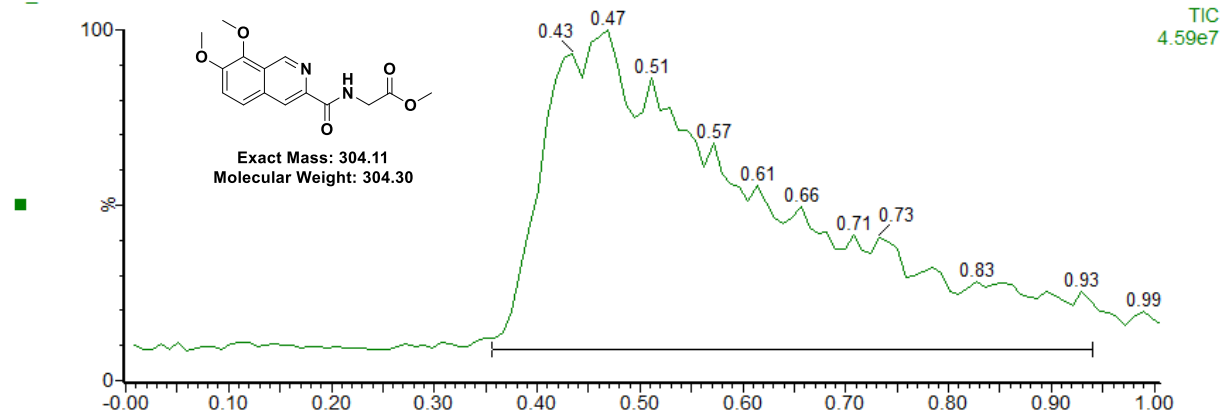
3_P10 51 (0.435) Cm (43:80)

1: Scan ES+
1.86e7

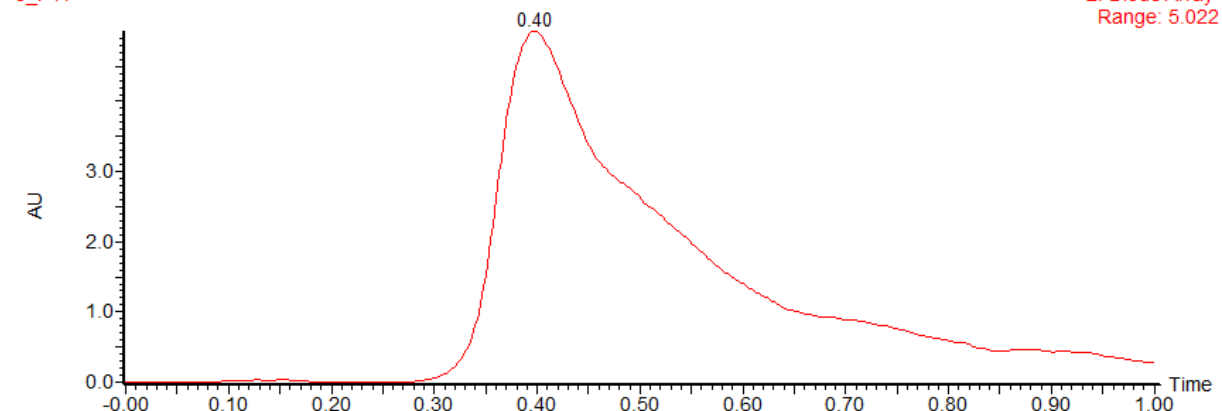


3_P11

3_P11

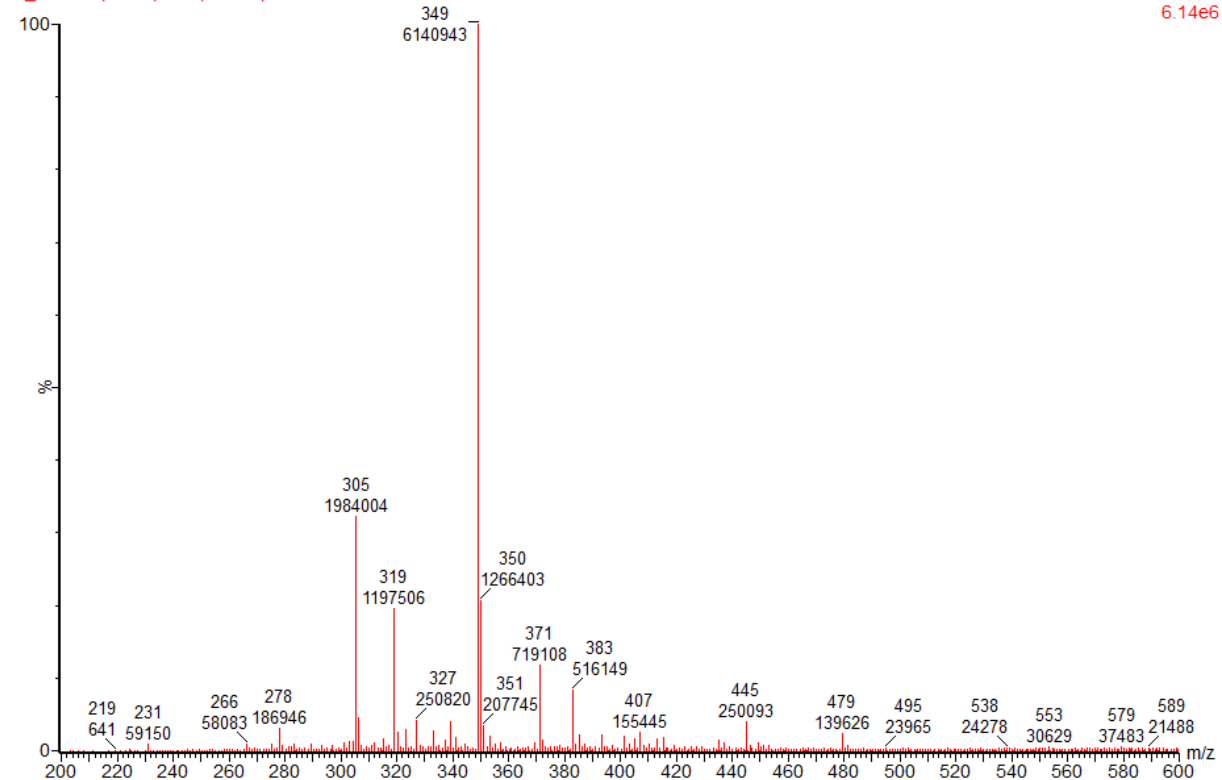


3_P11



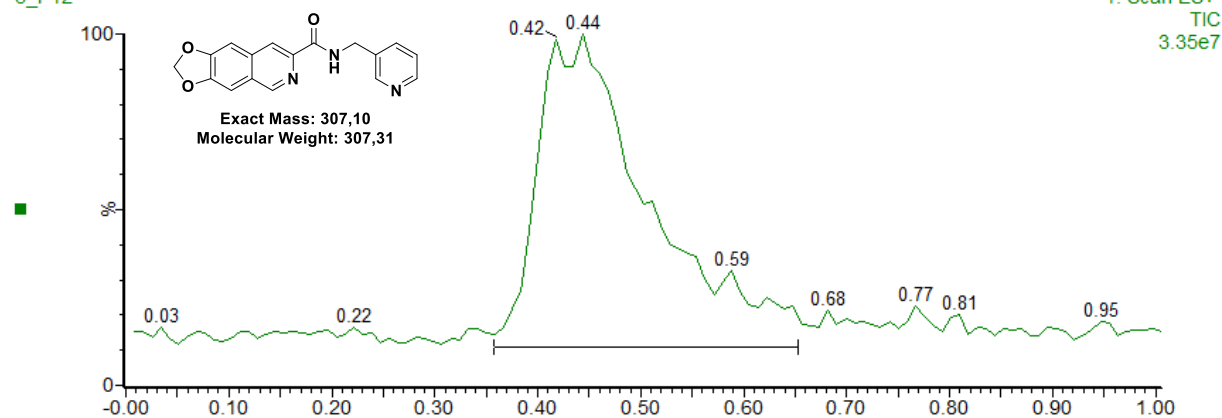
3_P11

3_P11 55 (0.469) Cm (42:110)

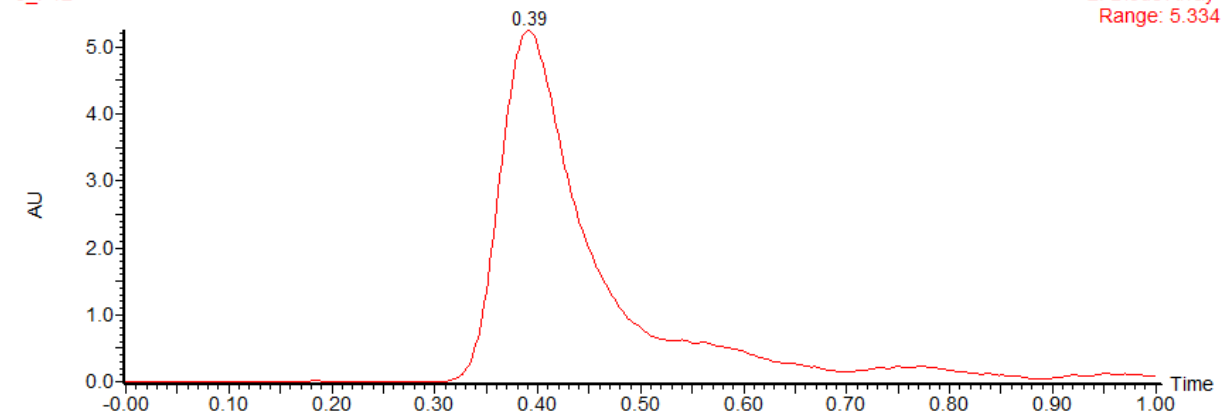


3_P12

3_P12

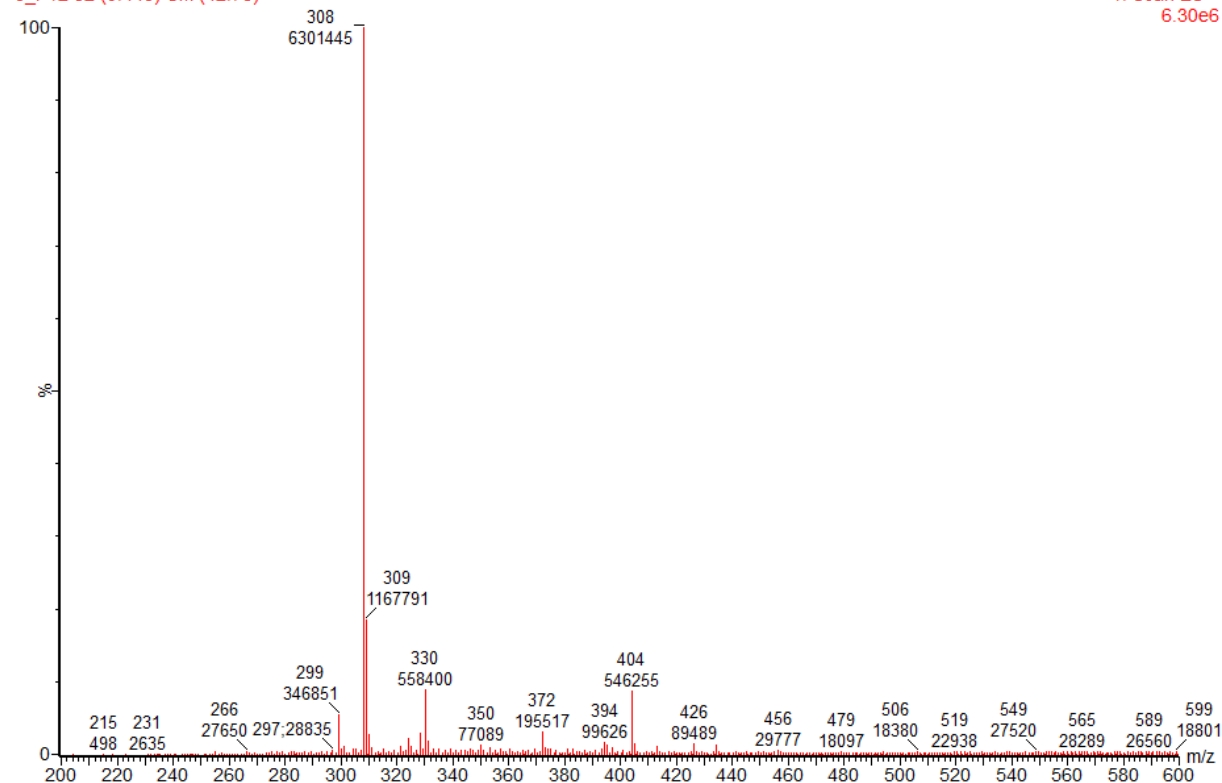


3_P12



3_P12

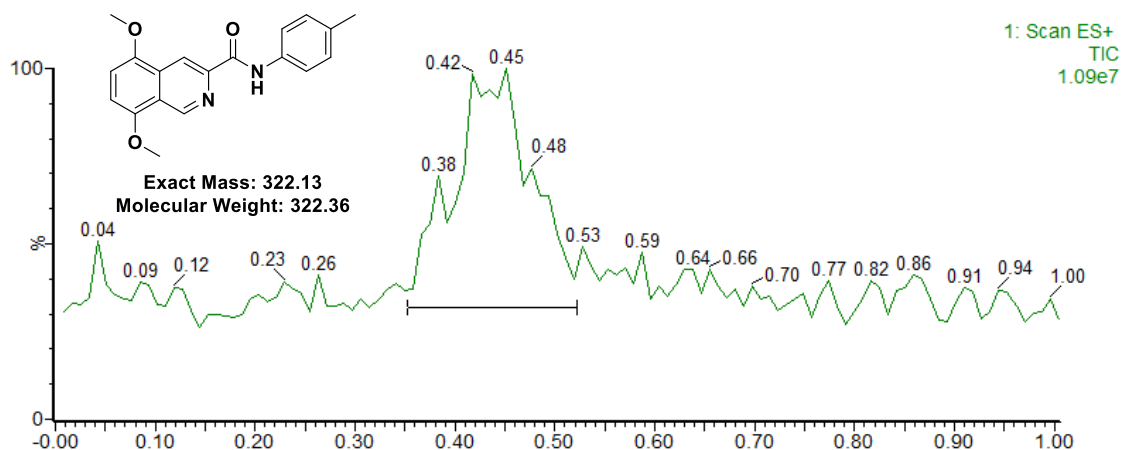
3_P12 52 (0.443) Cm (42:76)



3_P13

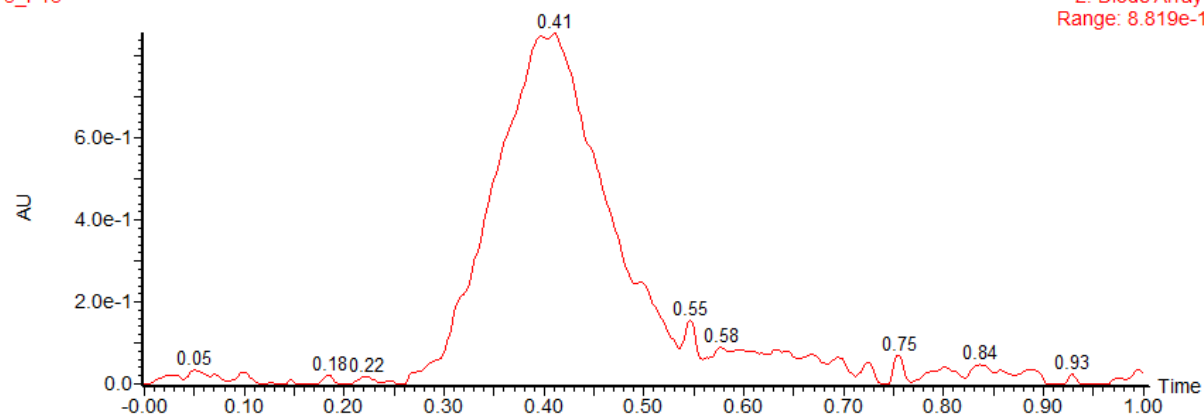
3_P13

■



3_P13

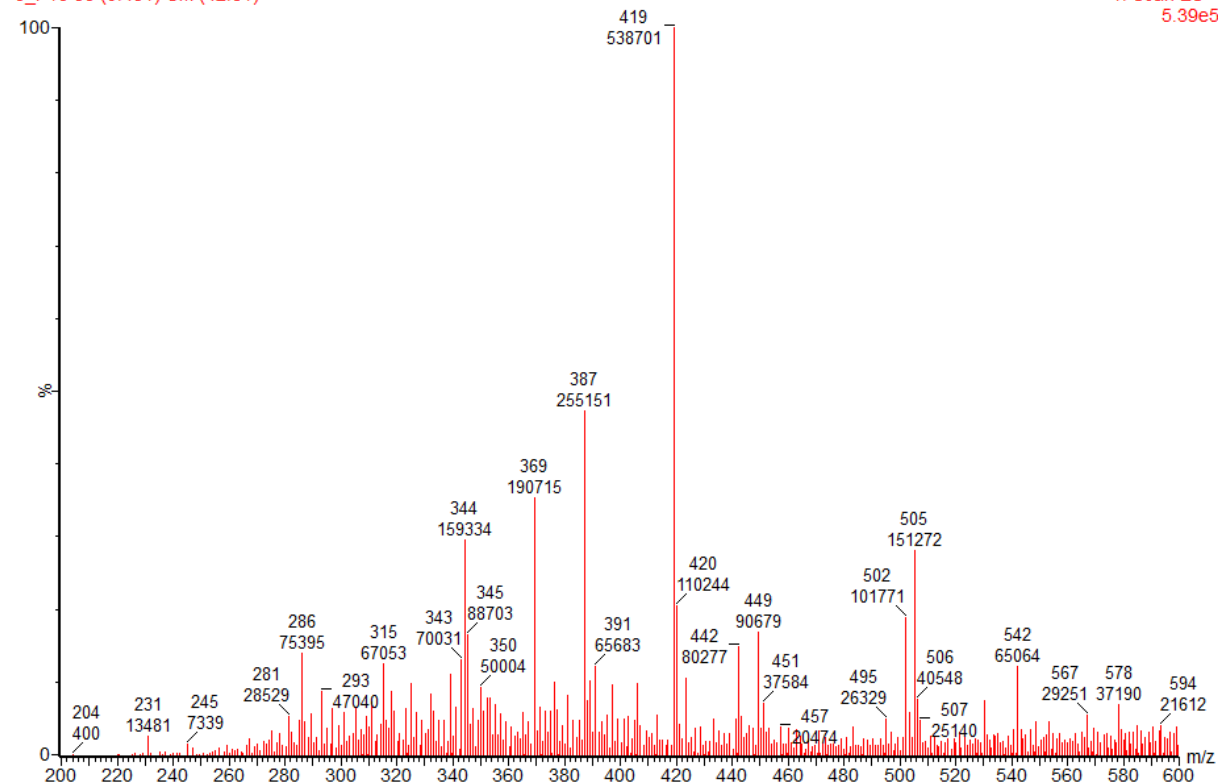
2: Diode Array
Range: 8.819e-1



3_P13

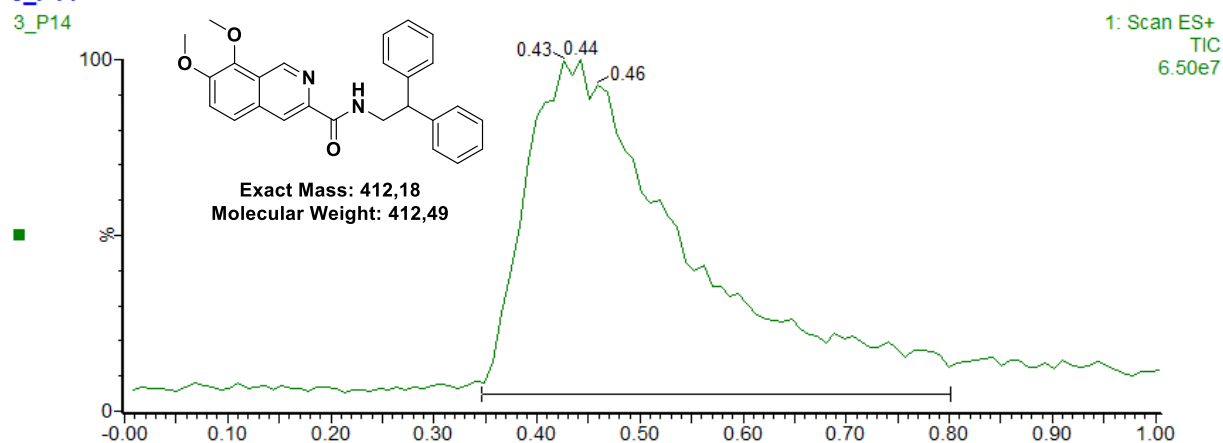
3_P13 53 (0.451) Cm (42.61)

1: Scan ES+
5.39e5

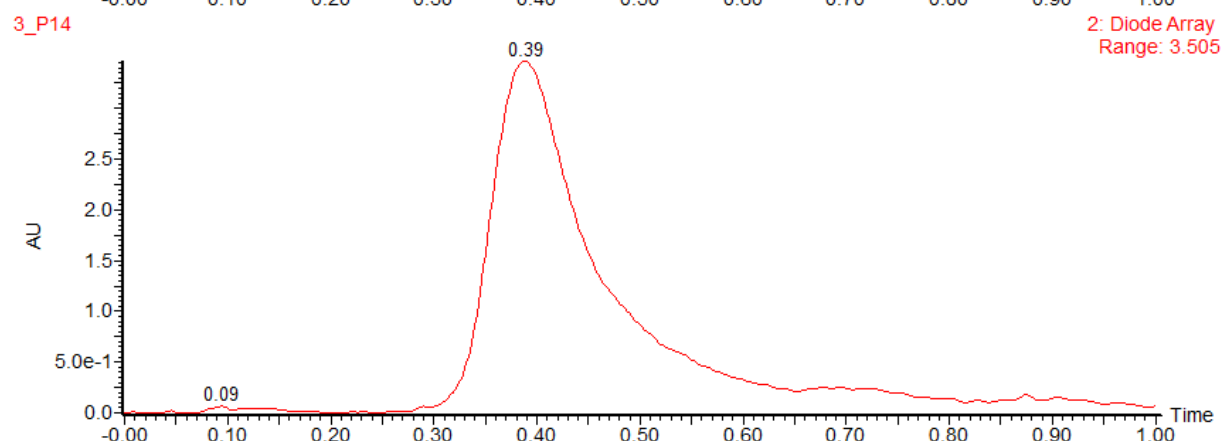


3_P14

3_P14

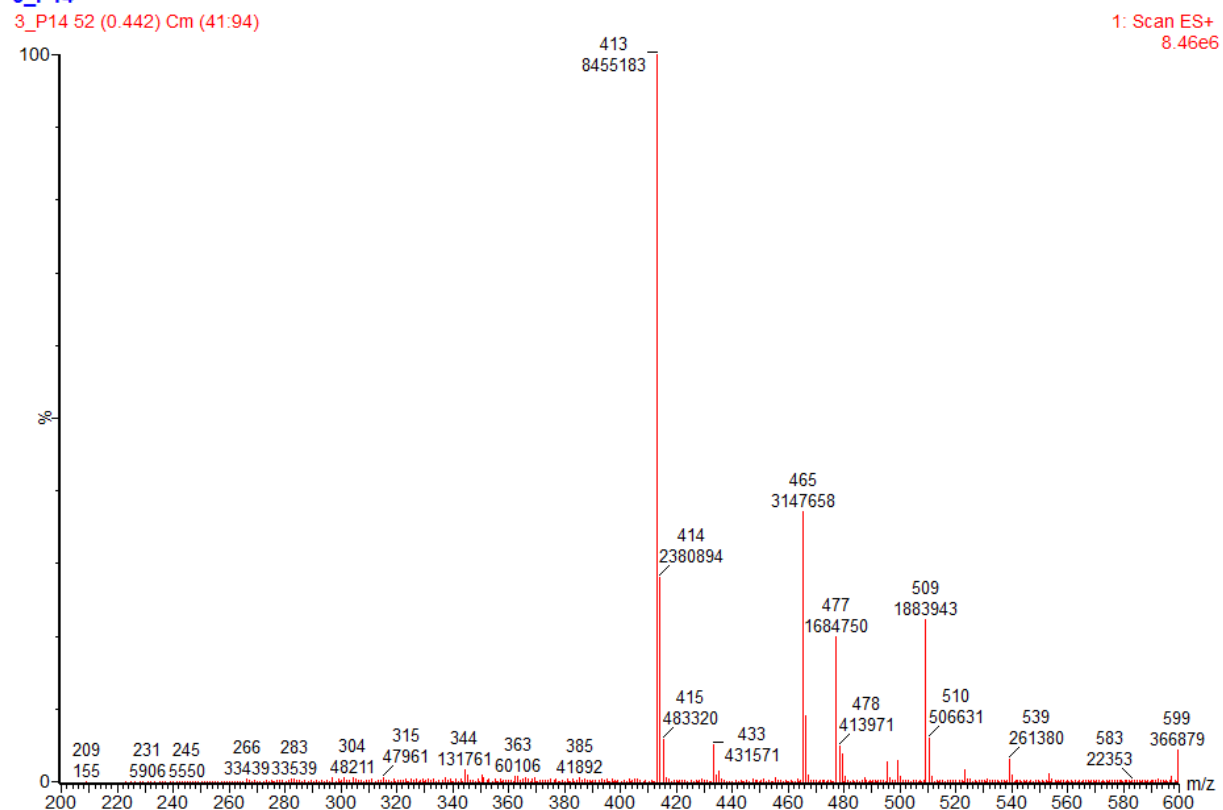


3_P14



3_P14

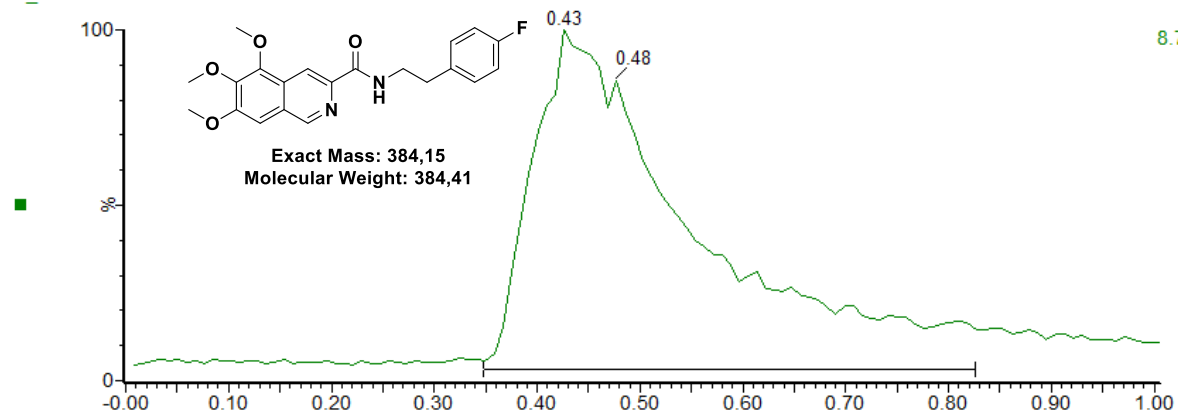
3_P14 52 (0.442) Cm (41:94)



3_P15

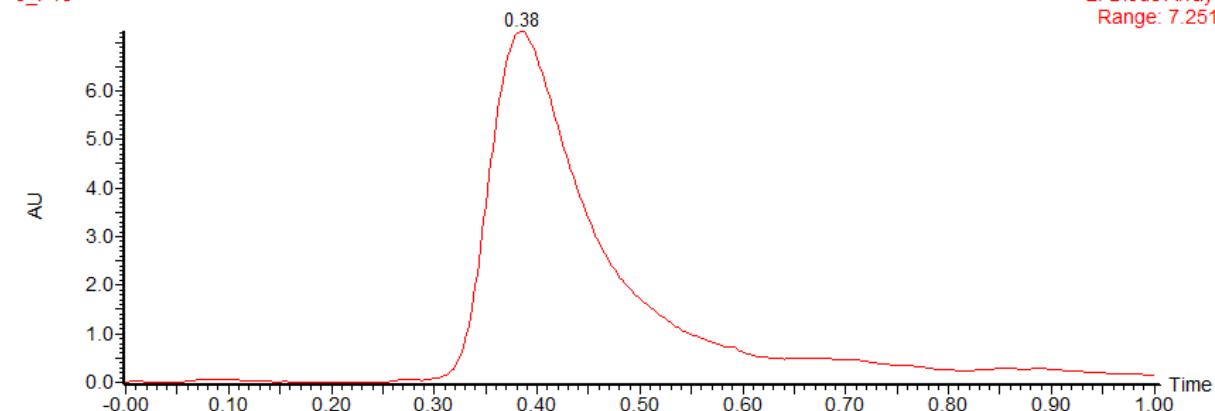
3_P15

1: Scan ES+
TIC
8.78e7



3_P15

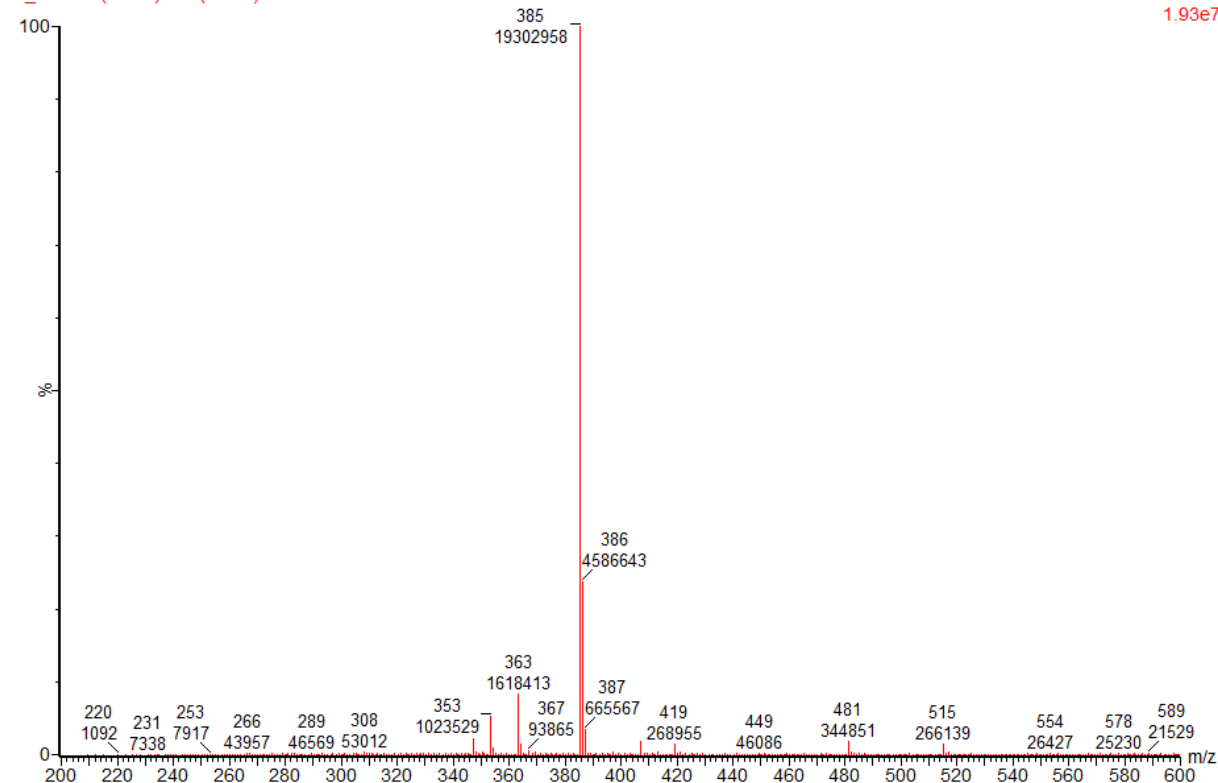
2: Diode Array
Range: 7.251



3_P15

3_P15 50 (0.426) Cm (41:96)

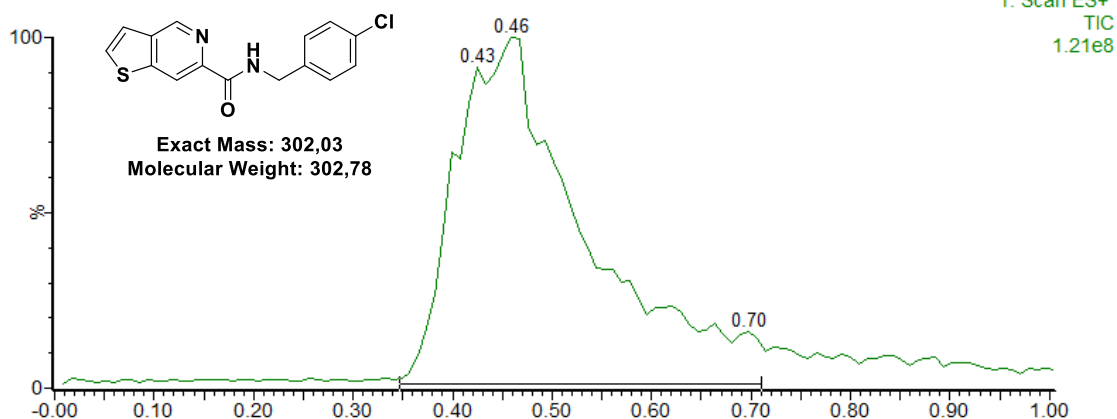
1: Scan ES+
1.93e7



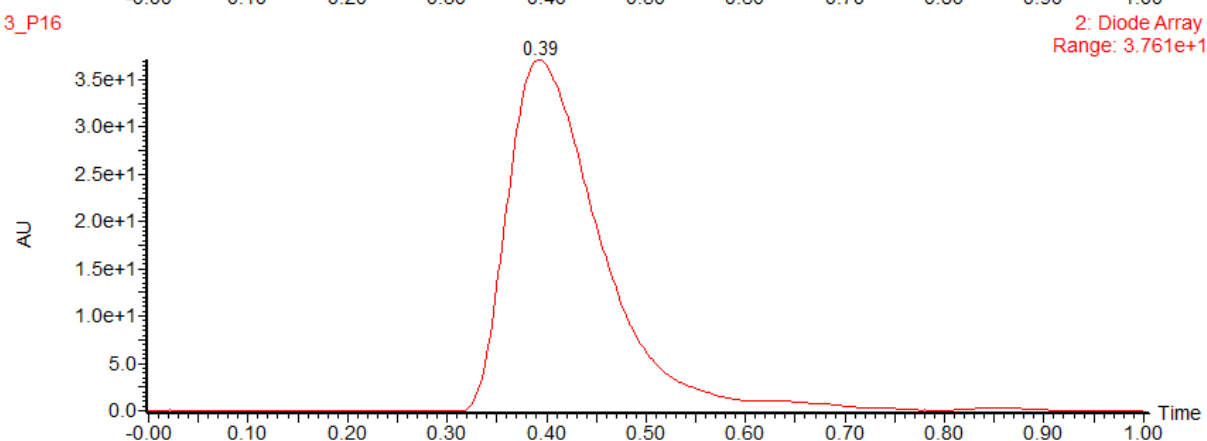
3_P16

3_P16

■

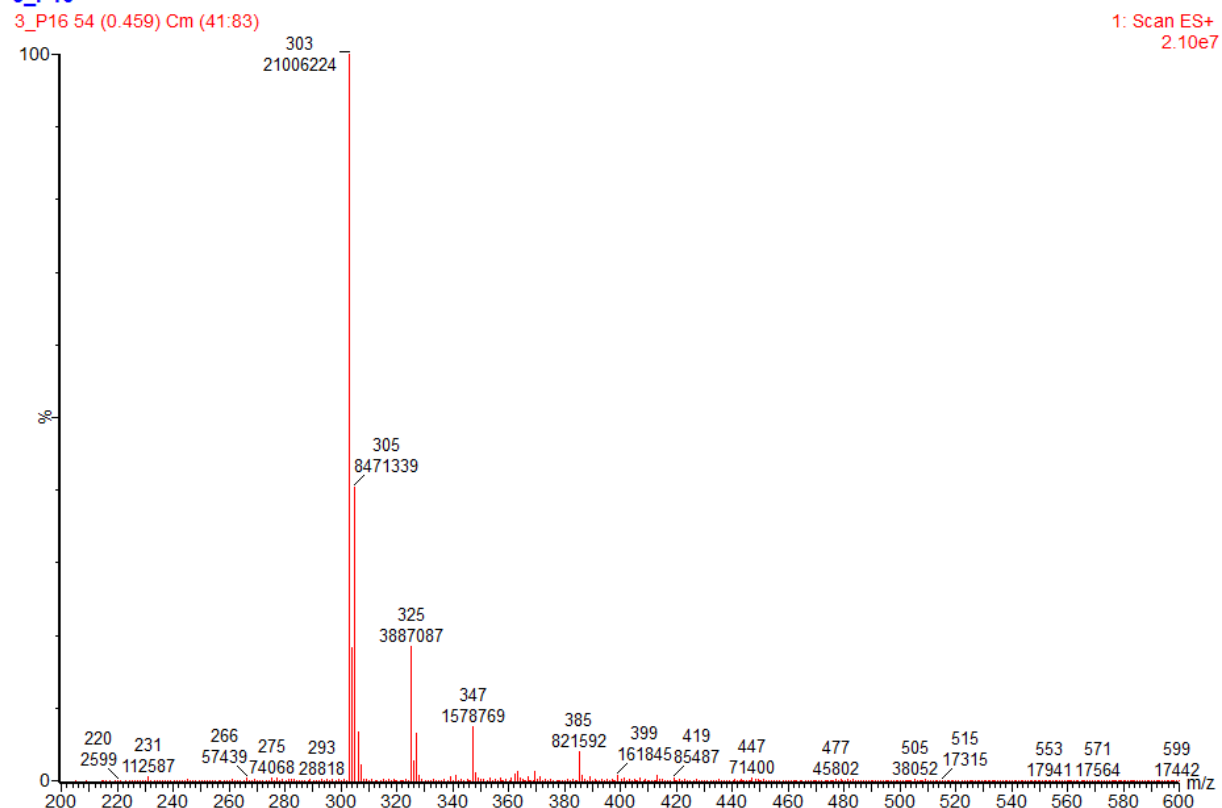


3_P16



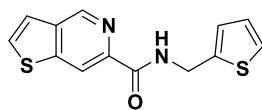
3_P16

3_P16 54 (0.459) Cm (41:83)



3_P17

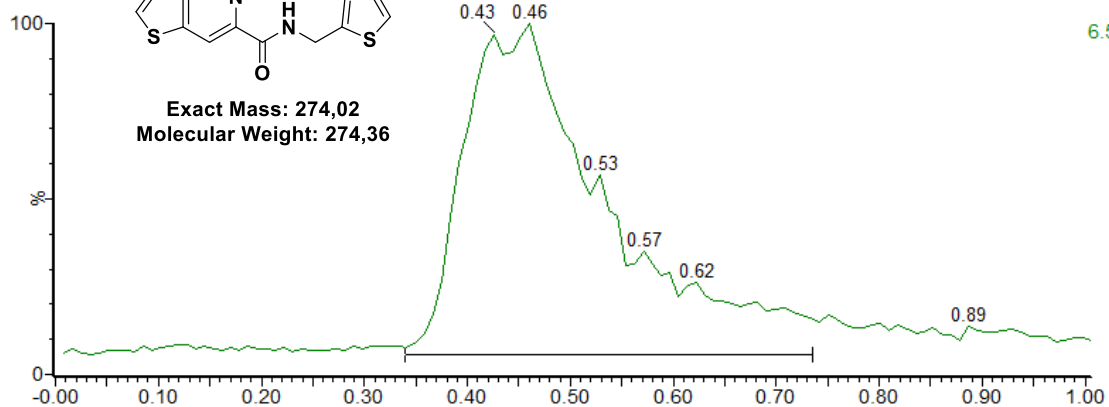
3_P17



Exact Mass: 274,02
Molecular Weight: 274,36

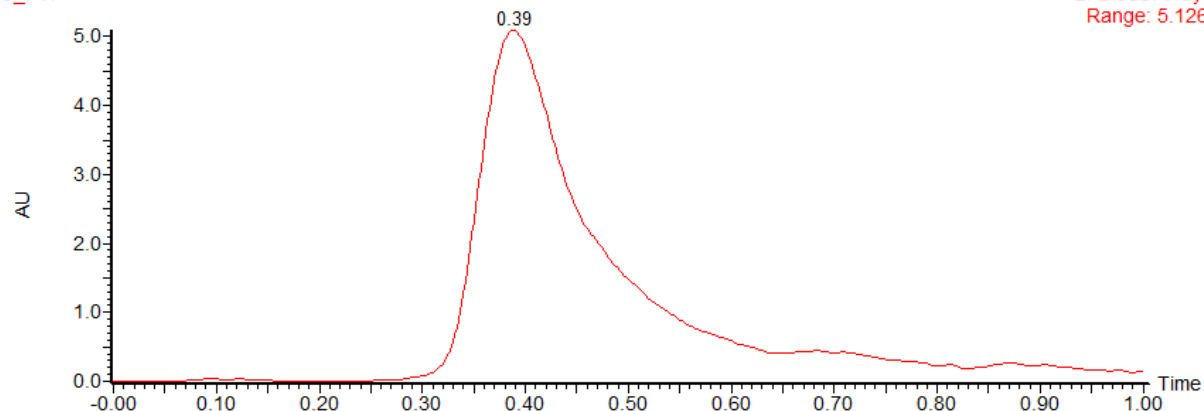
1: Scan ES+
TIC
6.54e7

■



3_P17

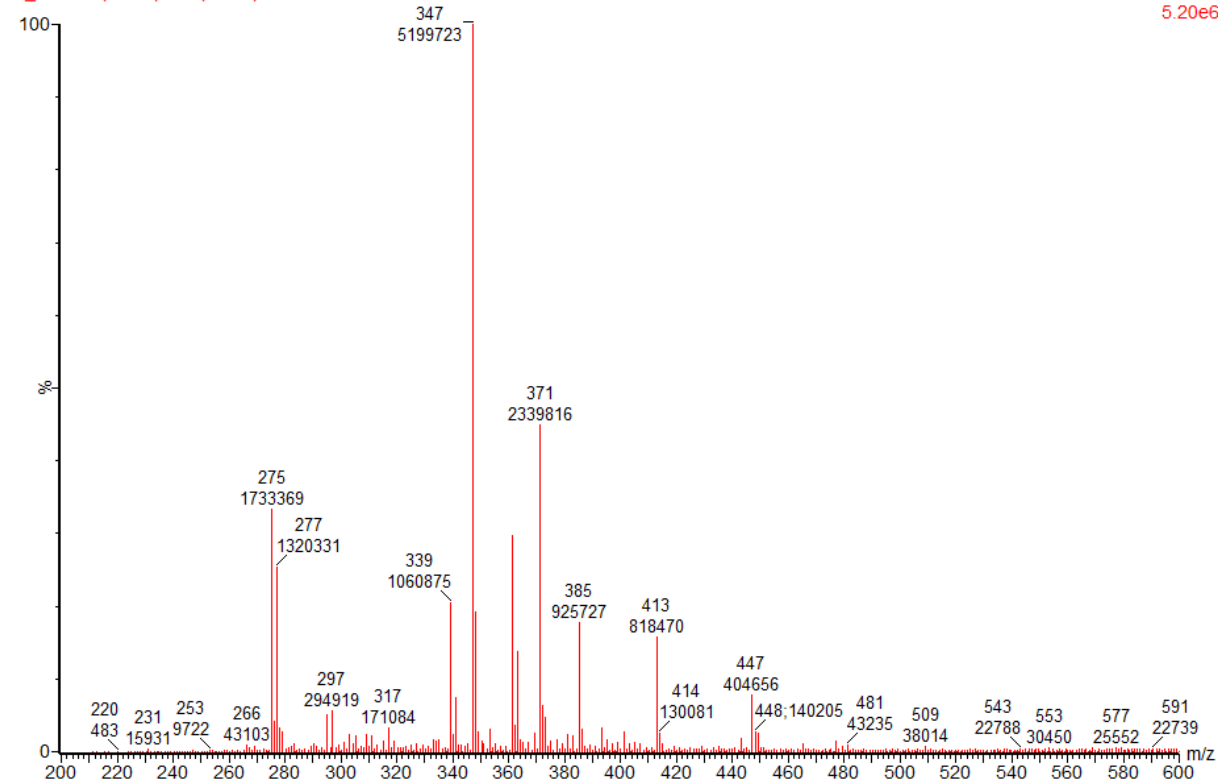
2: Diode Array
Range: 5.126



3_P17

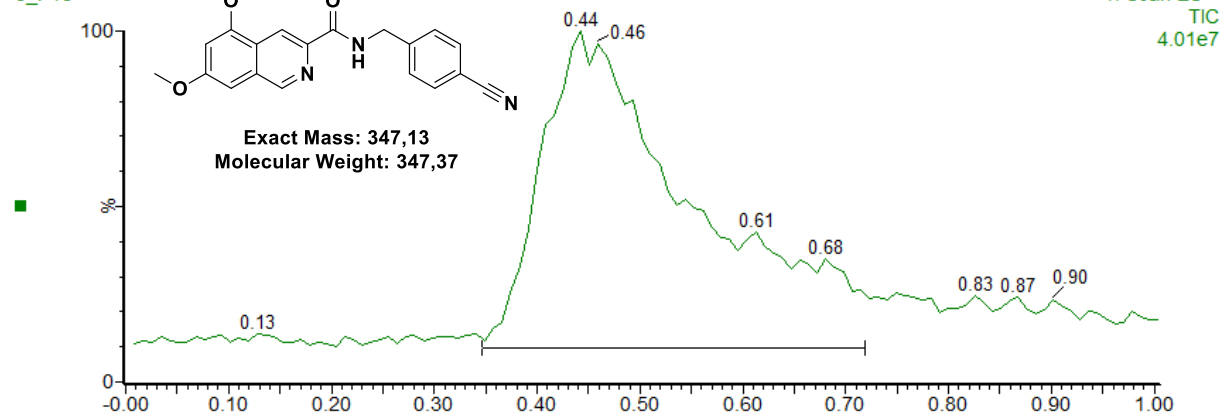
3_P17 54 (0.460) Cm (40:86)

1: Scan ES+
5.20e6

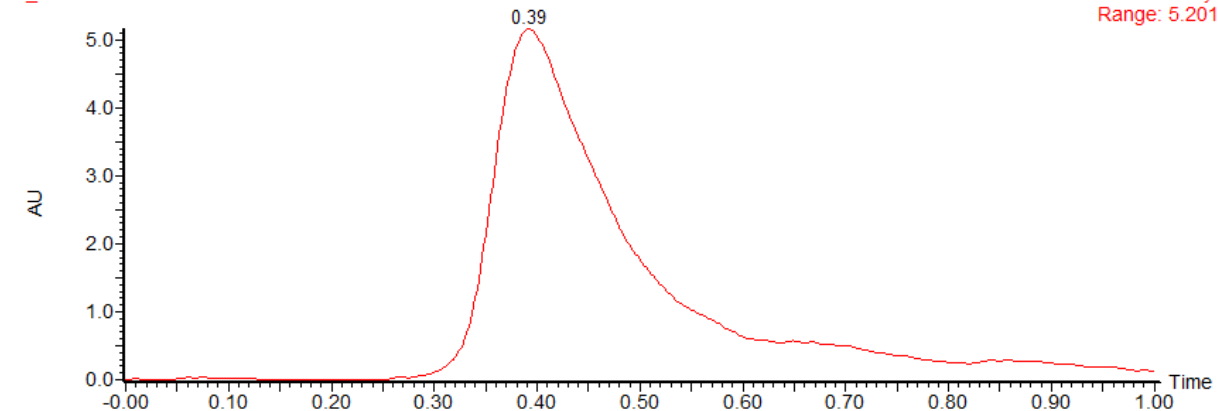


3_P18

3_P18

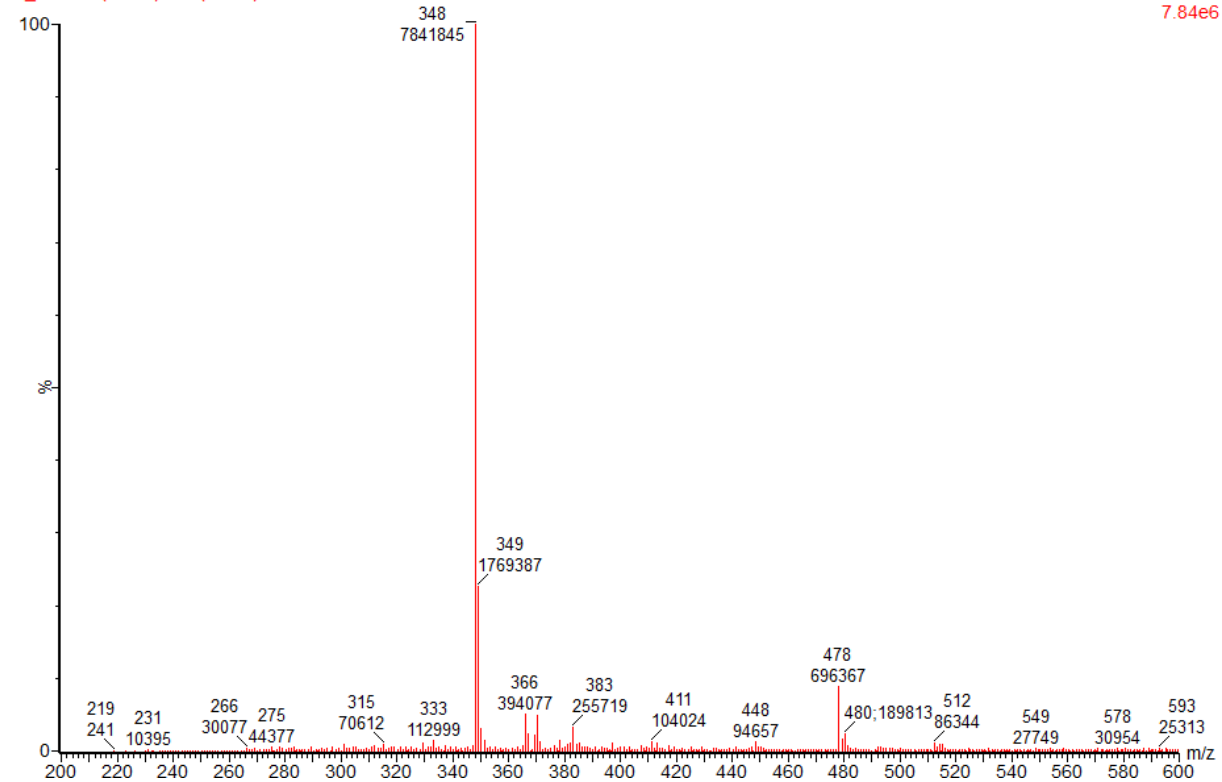


3_P18



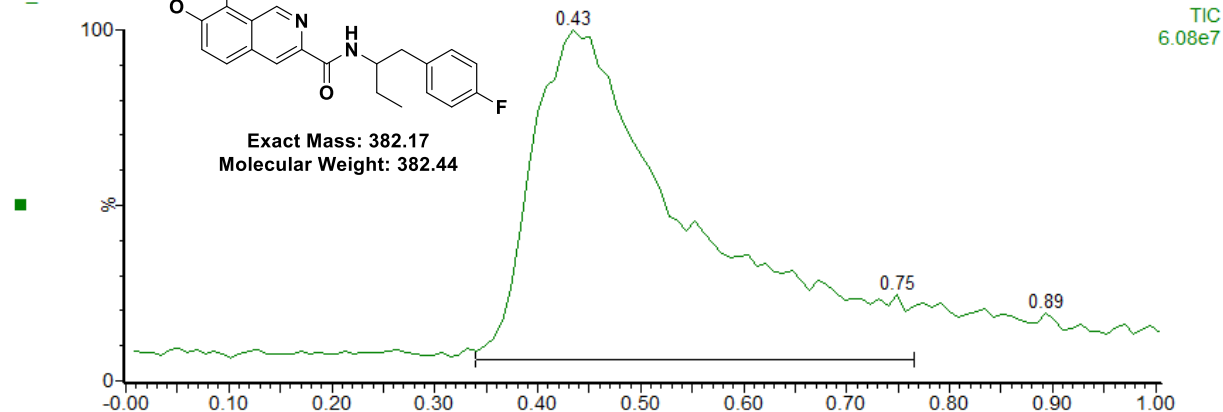
3_P18

3_P18 52 (0.442) Cm (41:84)

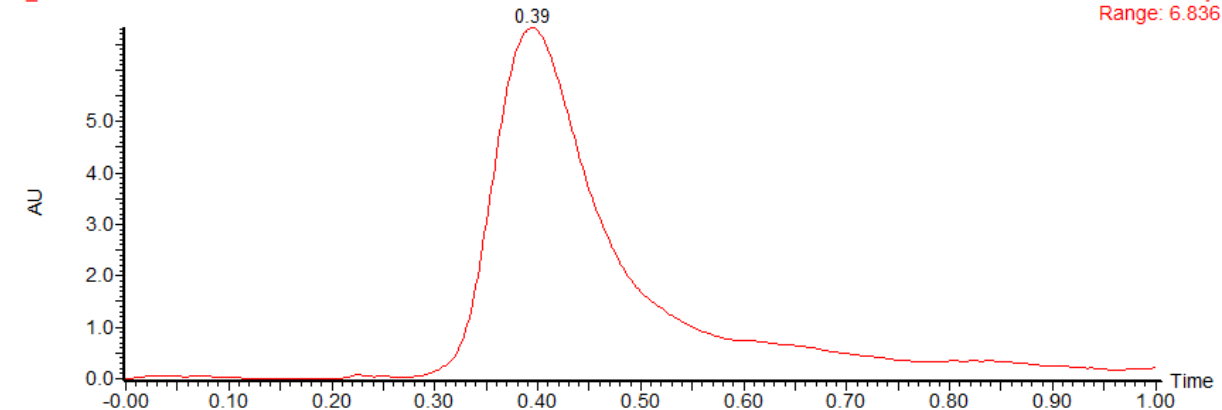


3_P19

3_P19

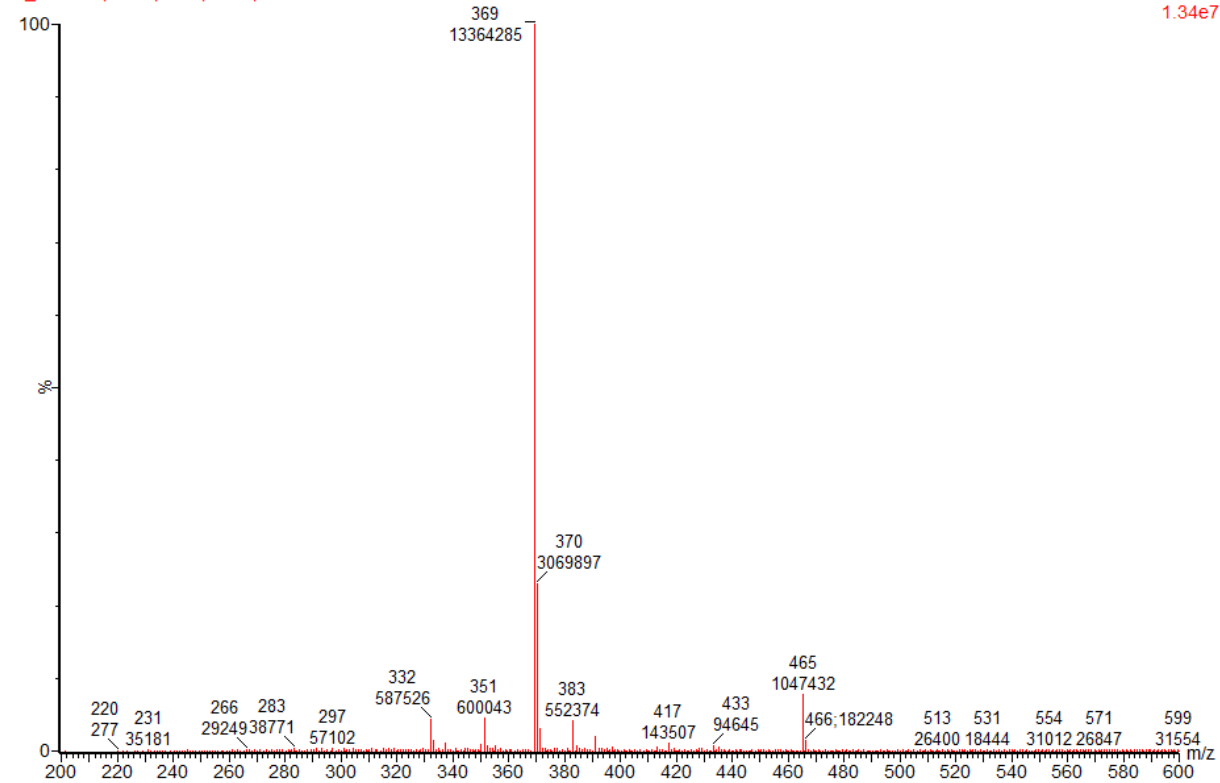


3_P19



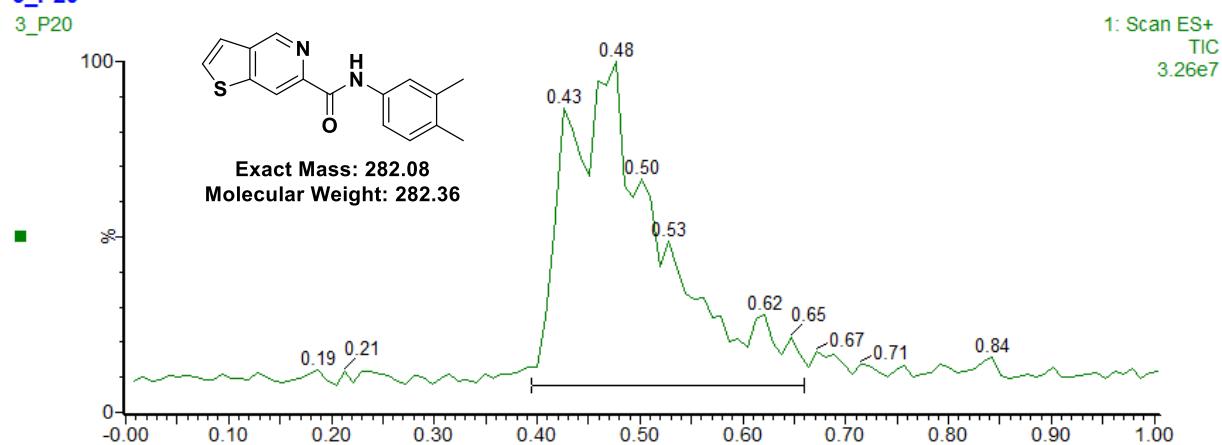
3_P19

3_P19 51 (0.434) Cm (40:89)

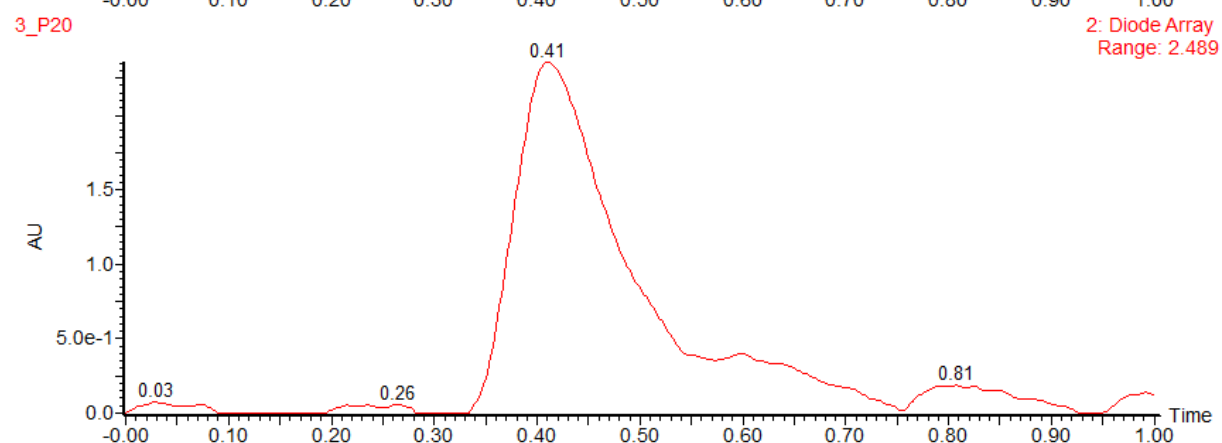


3_P20

3_P20

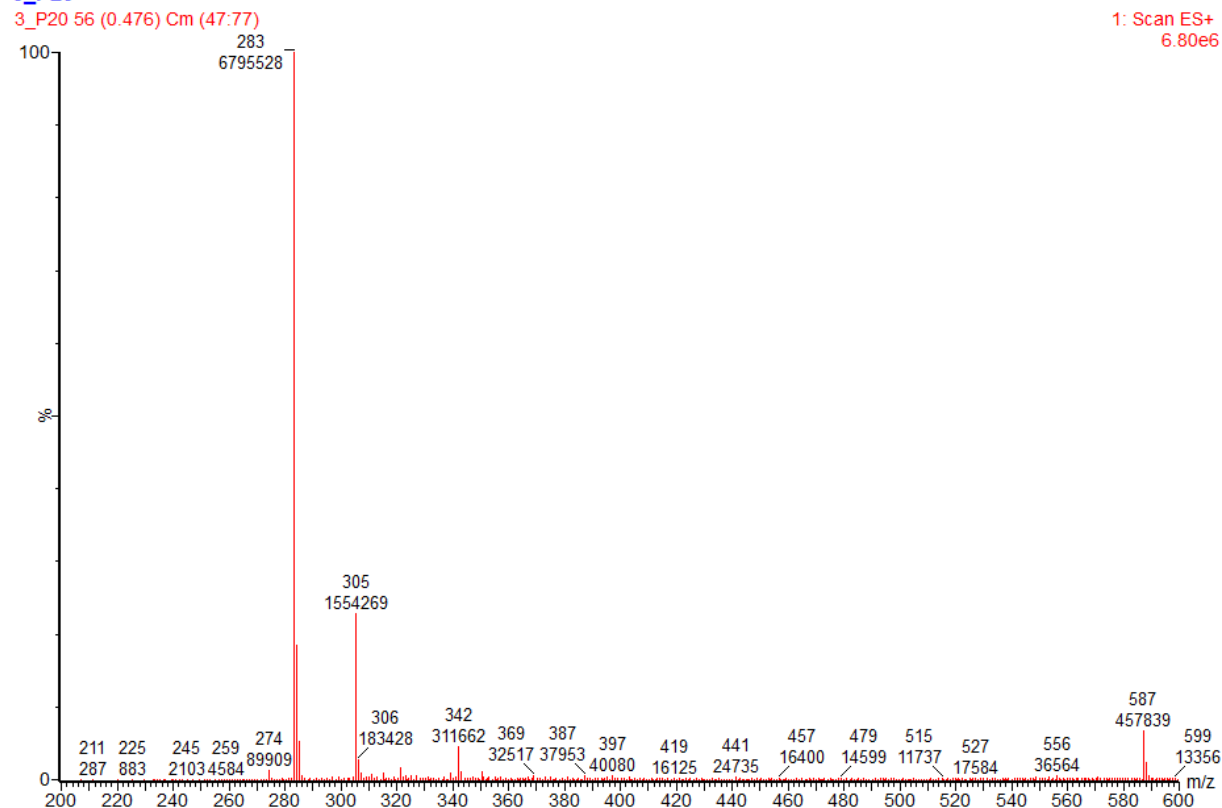


3_P20



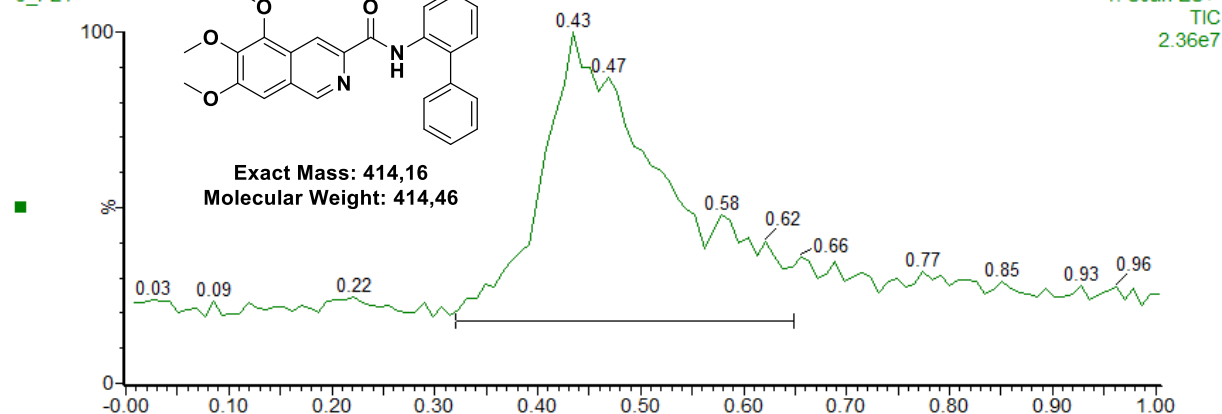
3_P20

3_P20 56 (0.476) Cm (47.77)

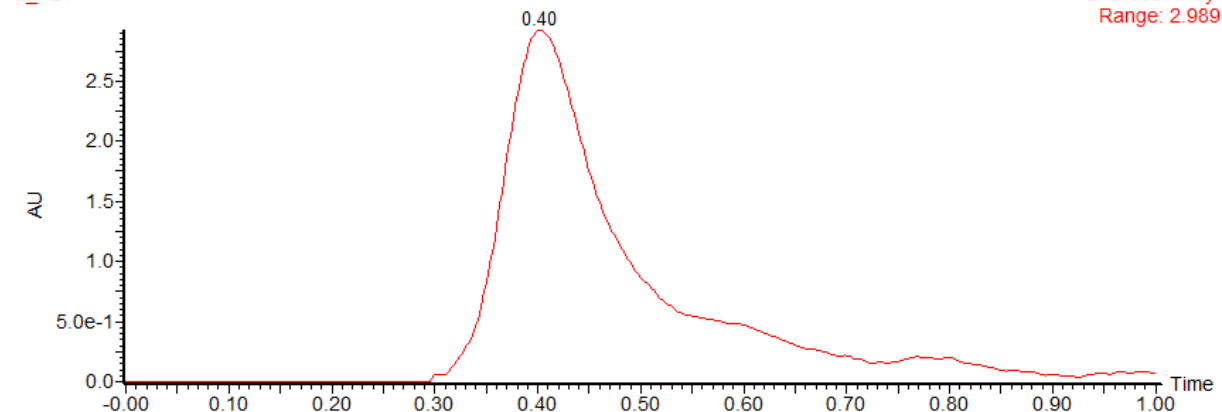


3_P21

3_P21

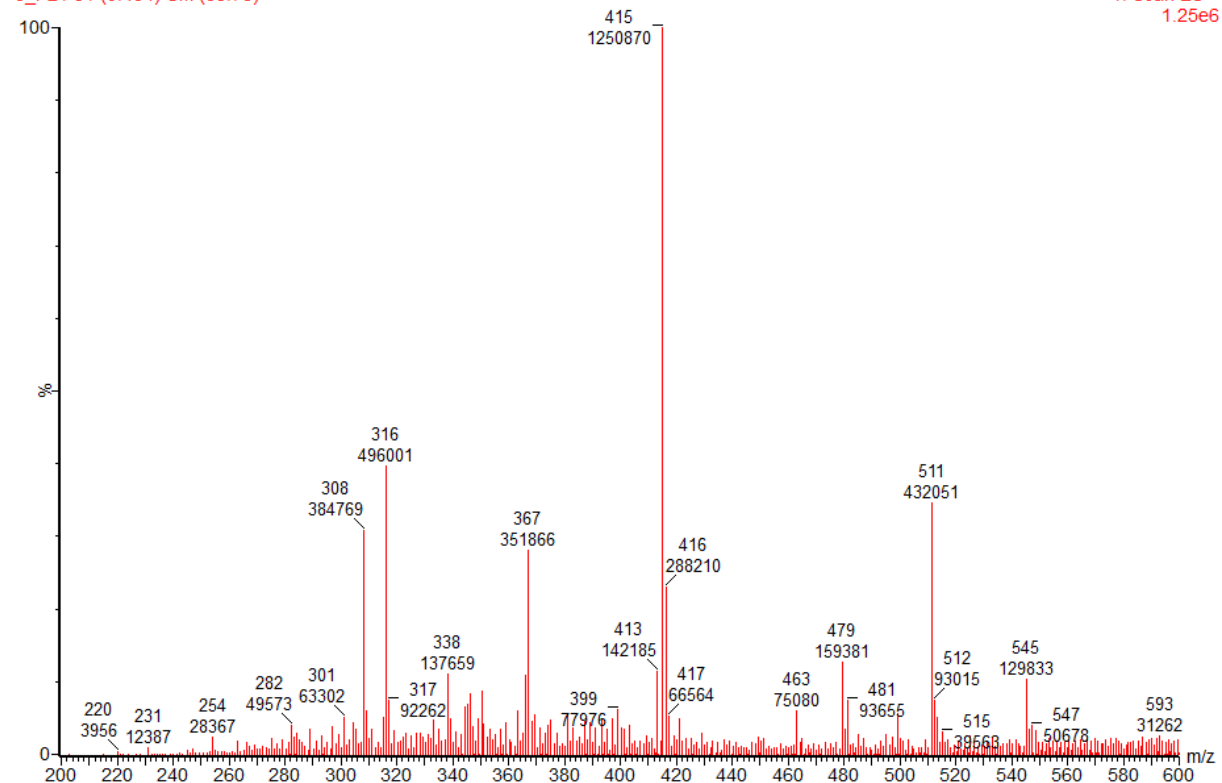


3_P21



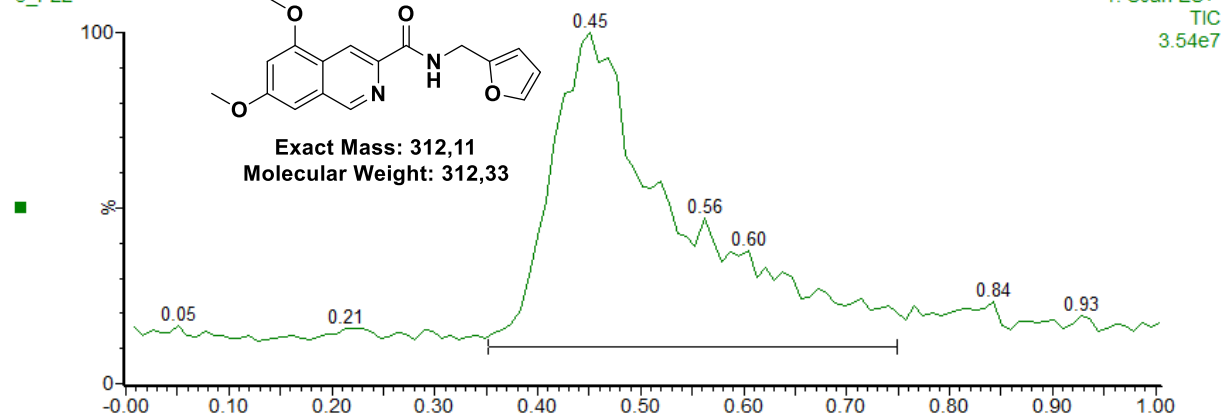
3_P21

3_P21 51 (0.434) Cm (38.76)

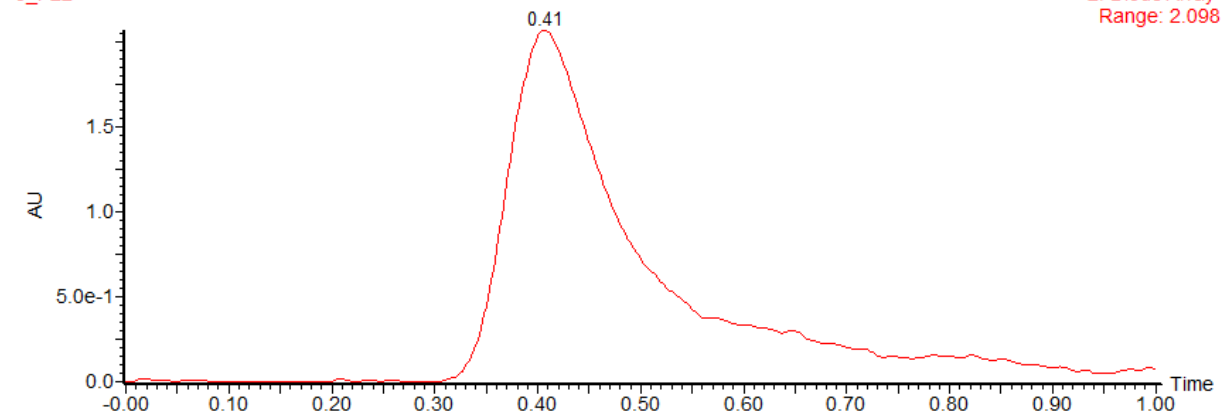


3_P22

3_P22

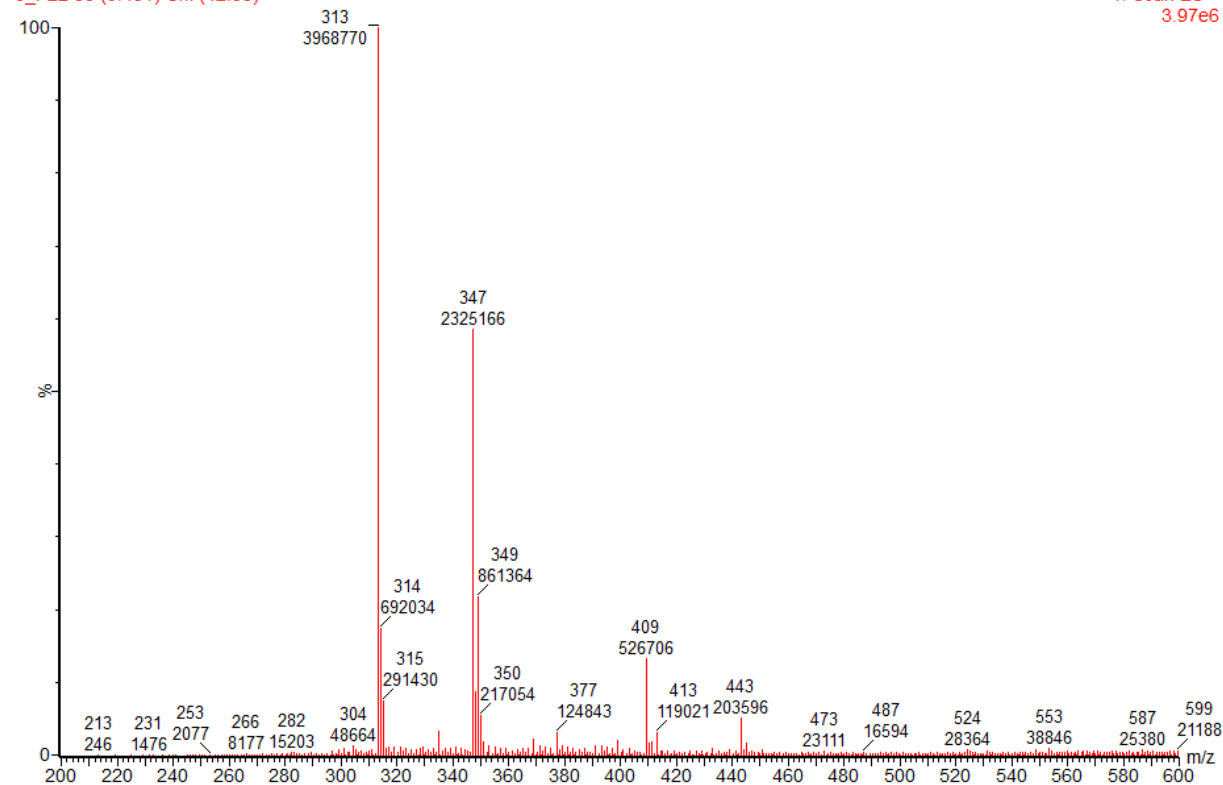


3_P22



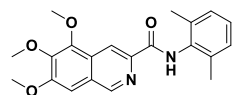
3_P22

3_P22 53 (0.451) Cm (42.88)



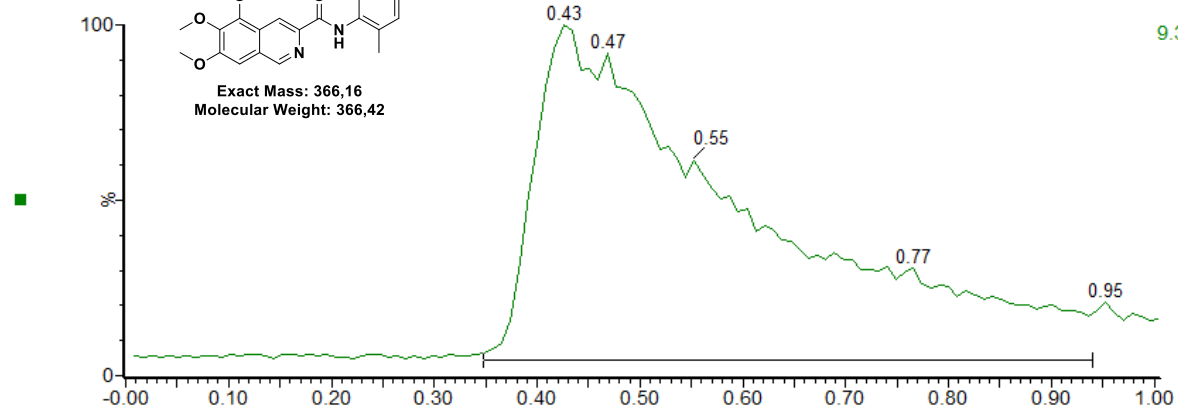
3_P23

3_P23



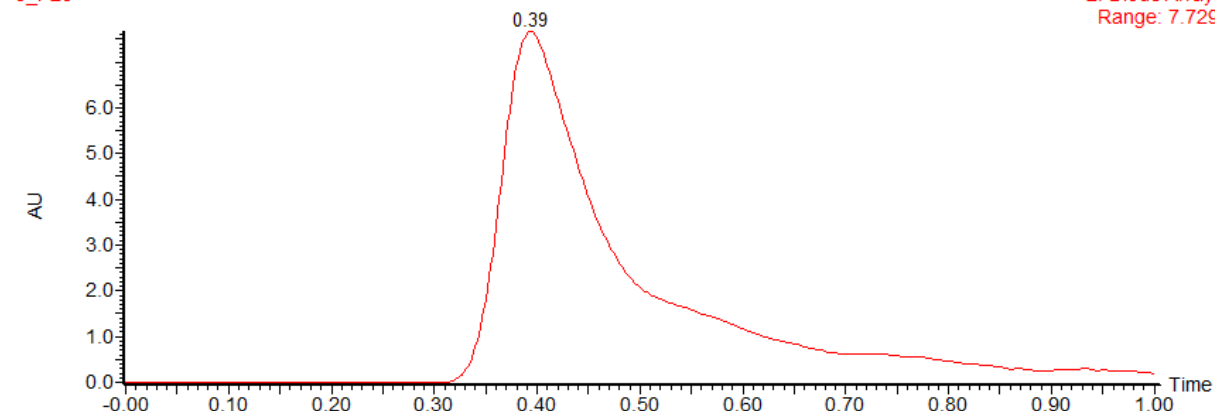
Exact Mass: 366,16
Molecular Weight: 366,42

1: Scan ES+
TIC
9.35e7



3_P23

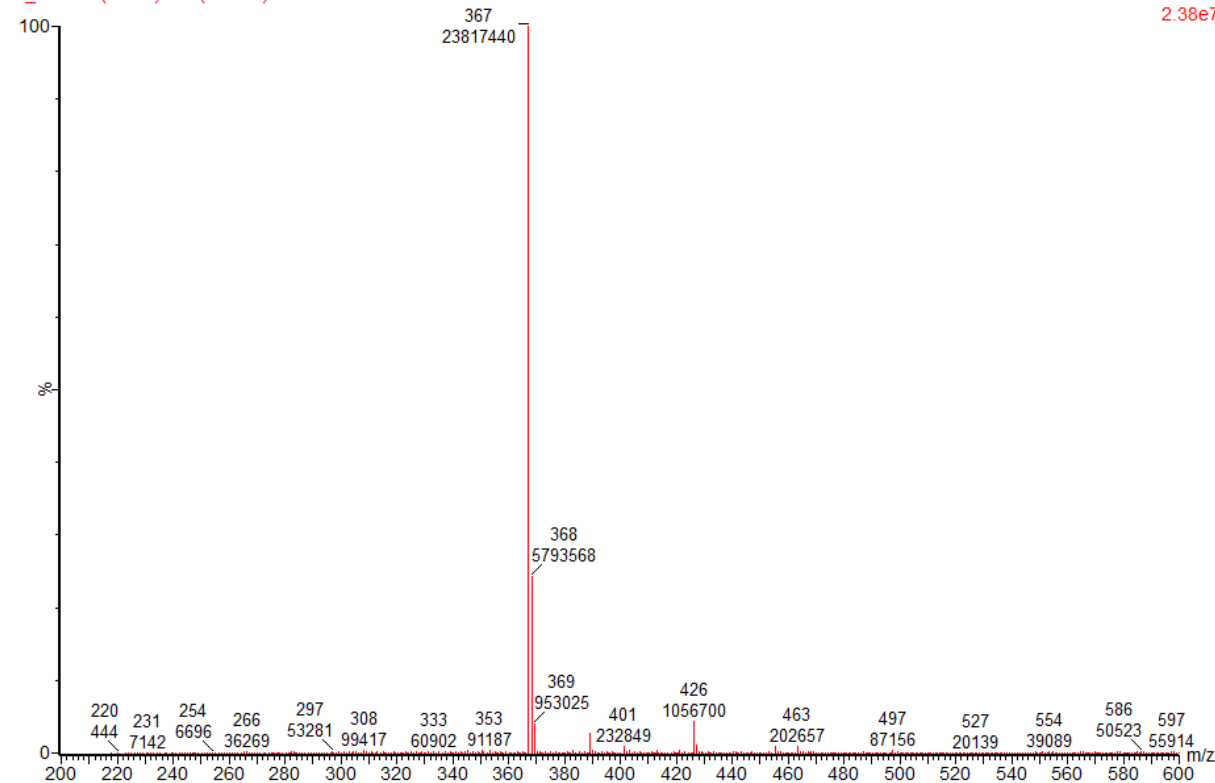
2: Diode Array
Range: 7.729



3_P23

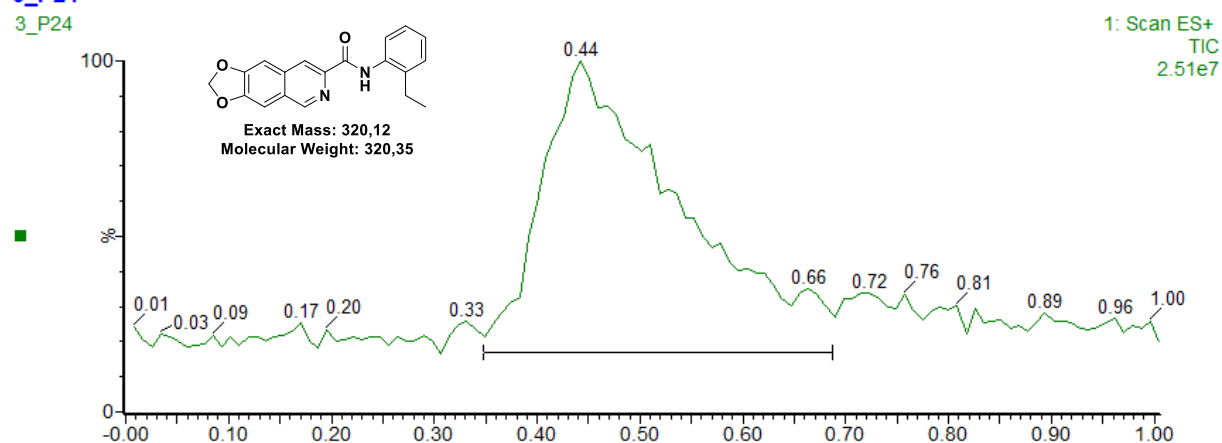
3_P23 50 (0.425) Cm (41:110)

1: Scan ES+
2.38e7

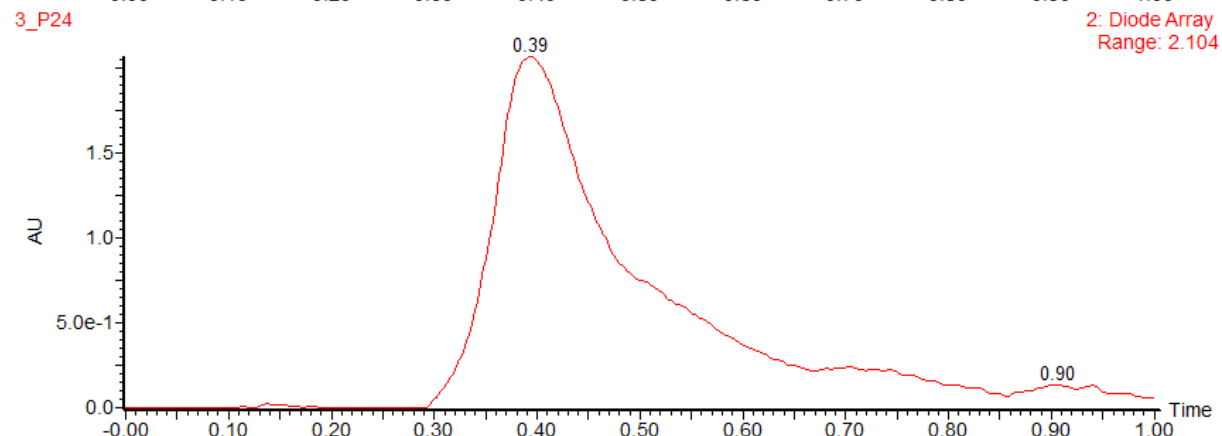


3_P24

3_P24

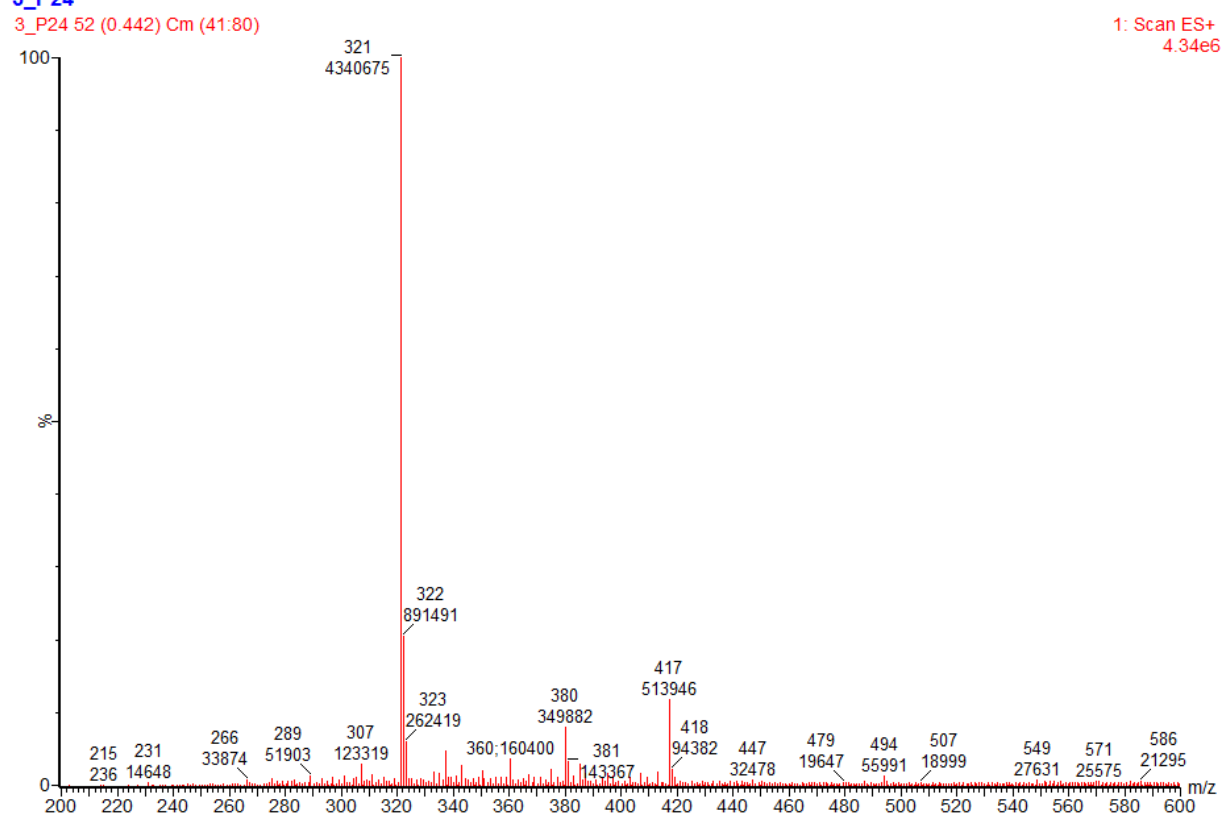


3_P24



3_P24

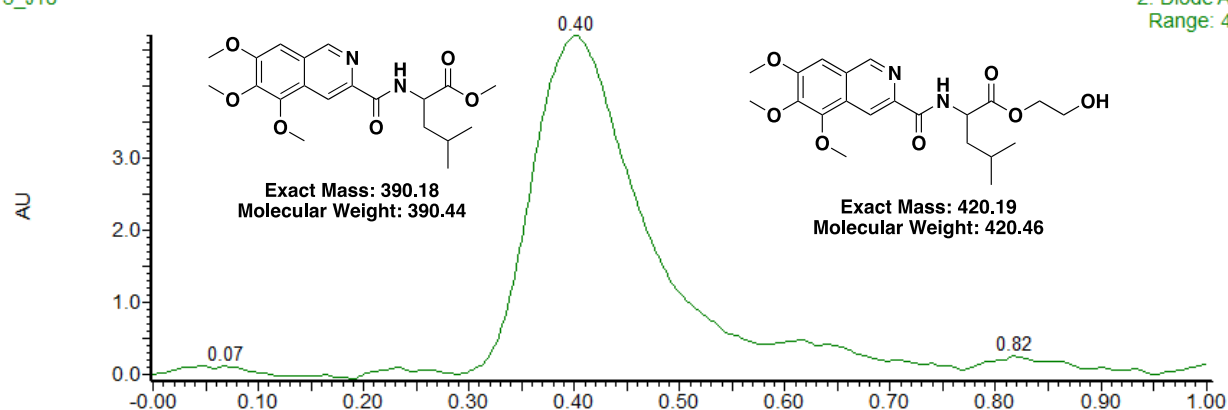
3_P24 52 (0.442) Cm (41:80)



3_J18

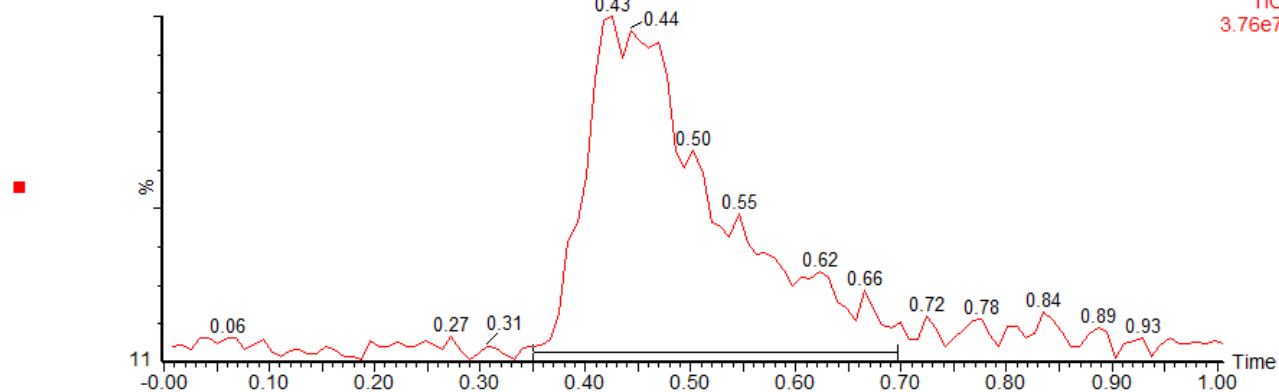
3_J18

2: Diode Array
Range: 4.764



3_J18

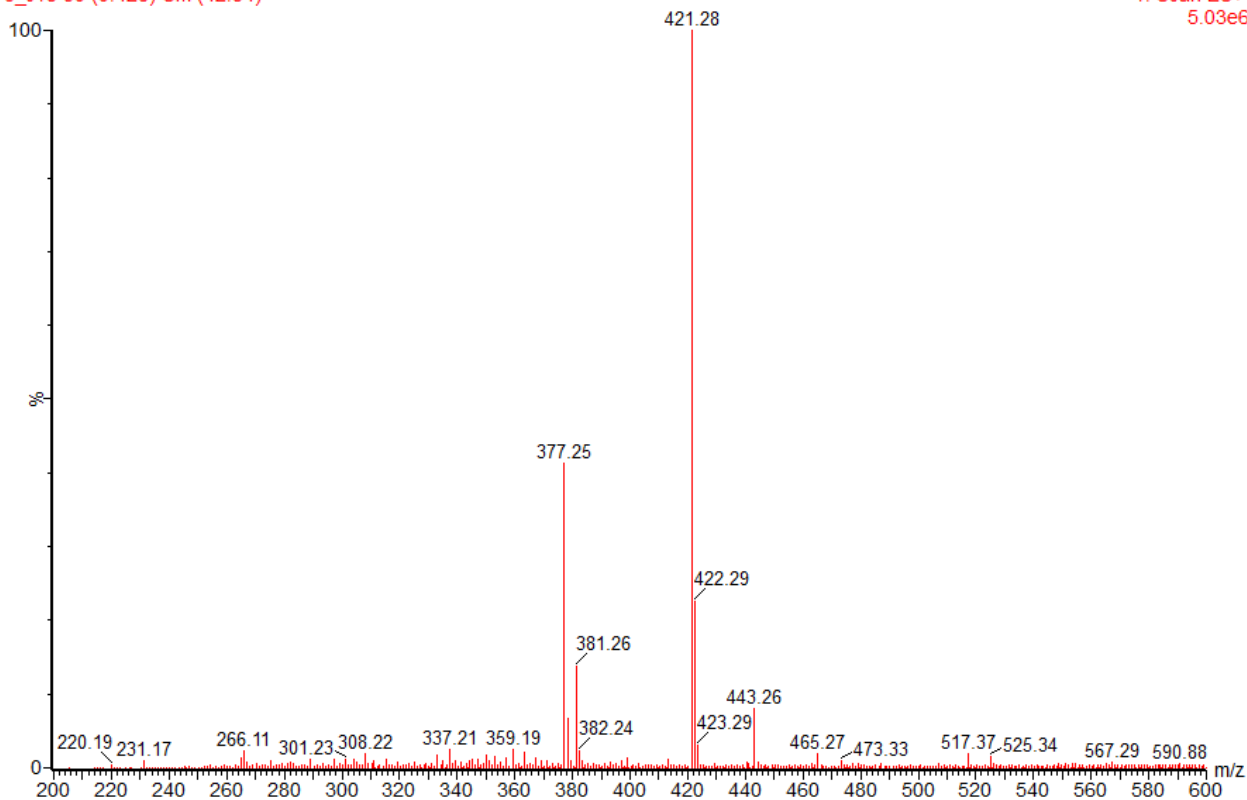
1: Scan ES+
TIC
3.76e7



3_J18

3_J18 50 (0.426) Cm (42:84)

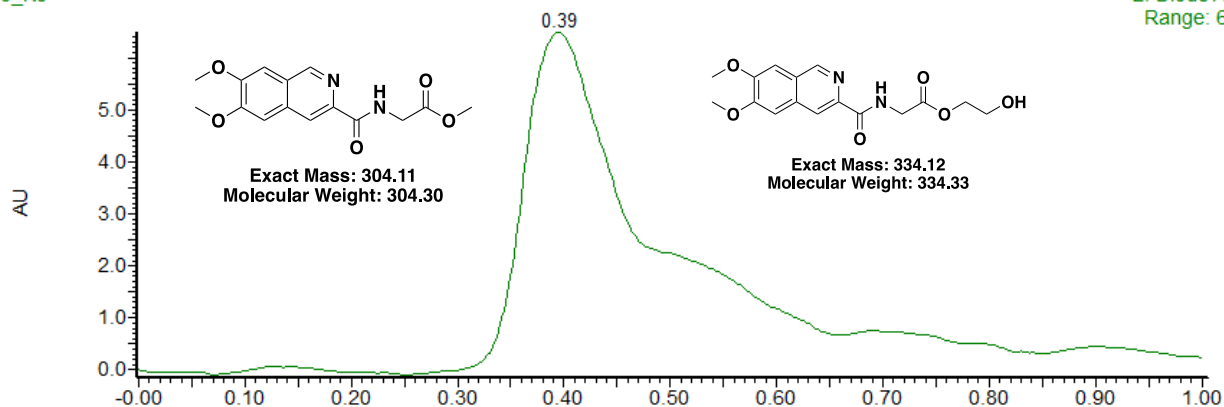
1: Scan ES+
5.03e6



3_K5

3_K5

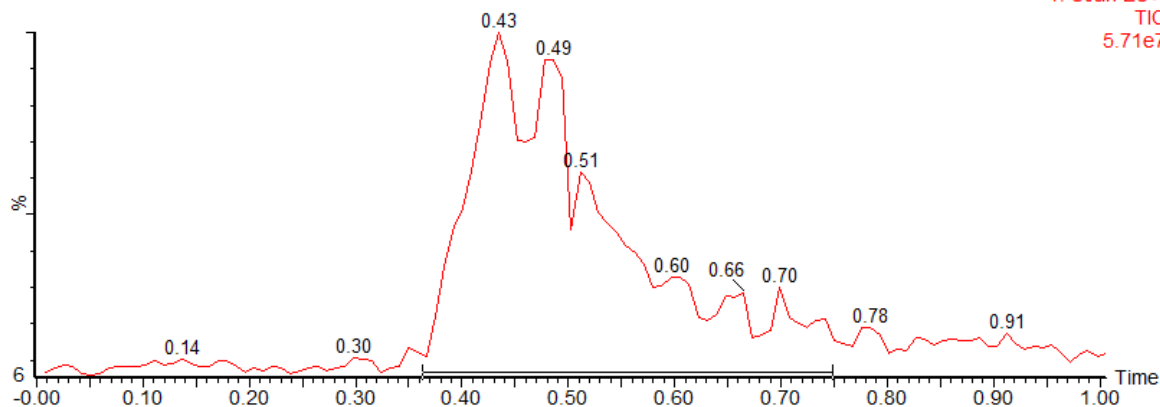
2: Diode Array
Range: 6.585



3_K5

1: Scan ES+
TIC
5.71e7

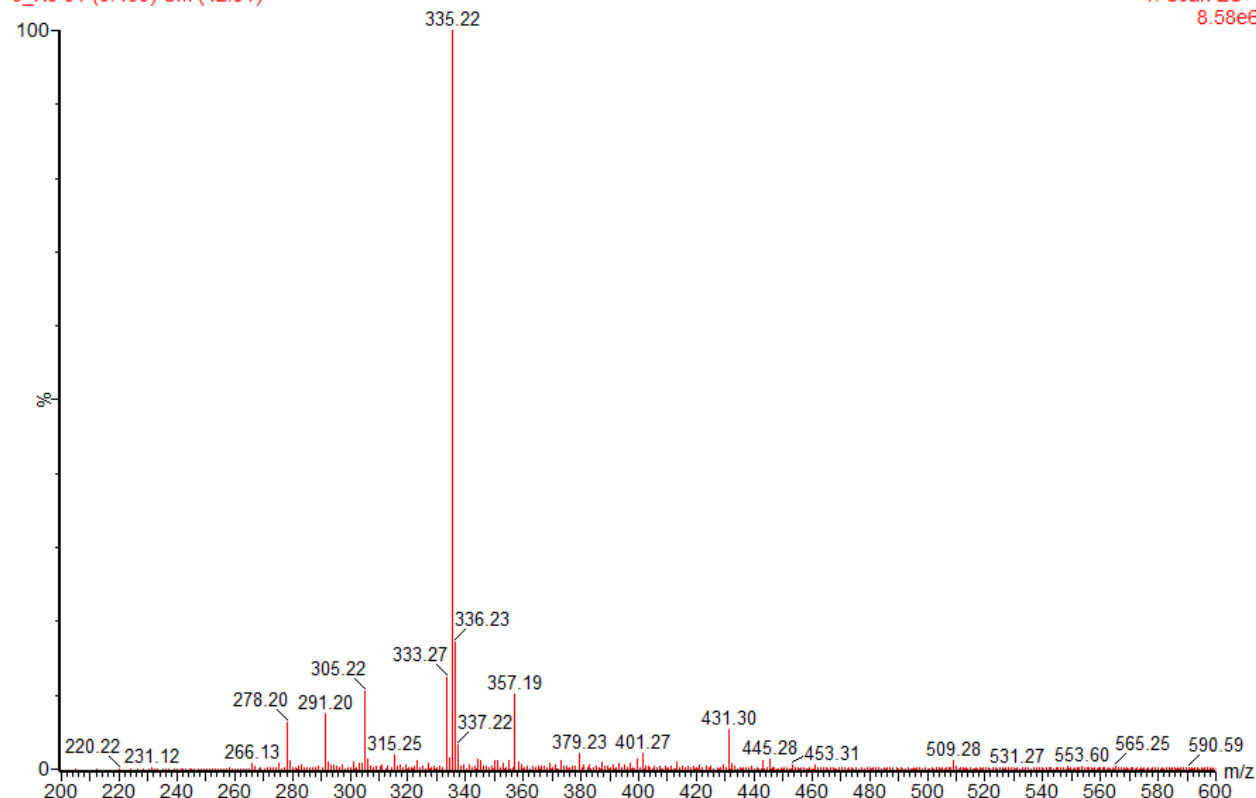
■



3_K5

3_K5 51 (0.435) Cm (42:91)

1: Scan ES+
8.58e6



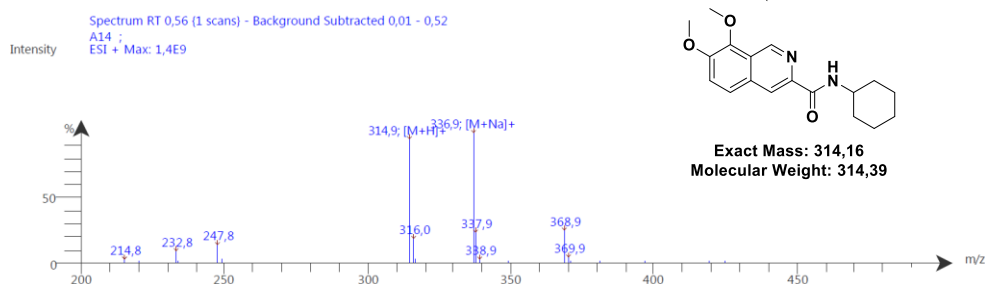
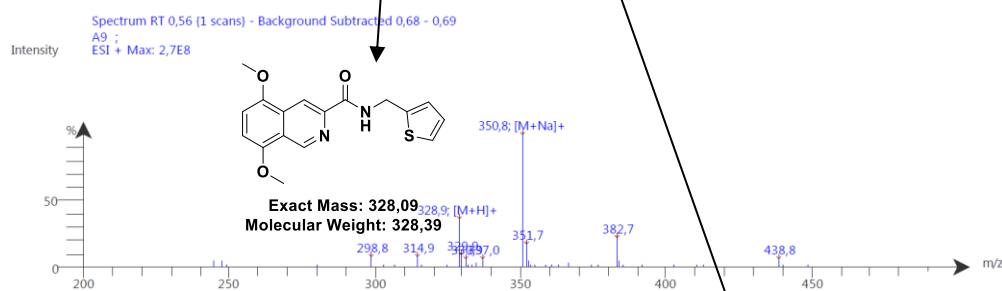
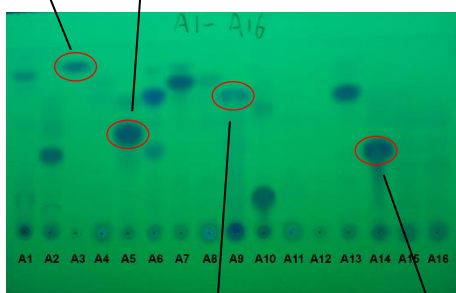
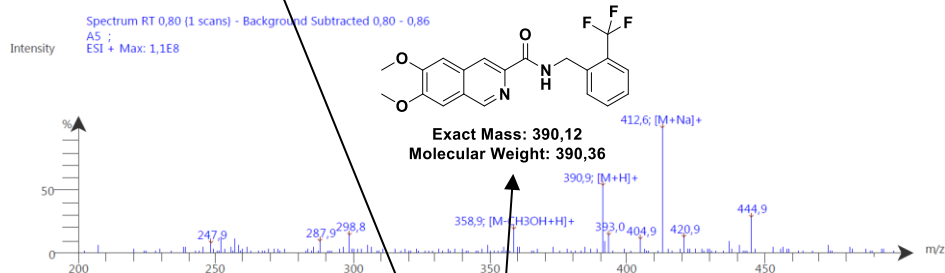
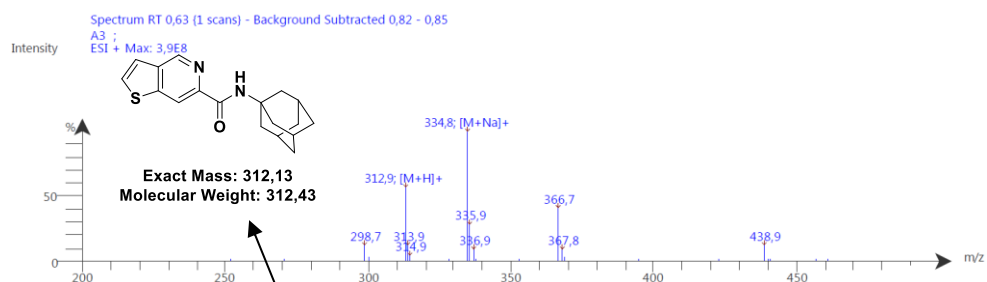
4.4.2. TLC-UV-MS analysis

Thin layer chromatography was performed on silica gel plates (0.20 mm thick, particle size 25 μm , Merck). Mass analysis of TLC plates was performed on an Advion Plate Express connected to an Advion compact mass spectrometer (CMS) fitted with an electrospray ionization (ESI^+) and using Mass Express software.

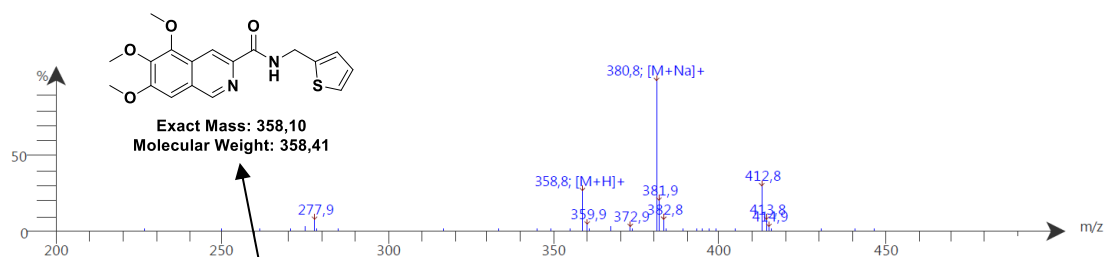
The isocratic elution was performed with MeOH, 5% H_2O , 0.1% formic acid, at a flow rate of 0.2 mL/min. Full scan mass spectra were recorded in the positive ionization mode (ESI^+) using a capillary temperature of 250 $^\circ\text{C}$, voltage of 150 V, source voltage offset of 20 V, source voltage span of 30, source gas temperature of 200 $^\circ\text{C}$, and an ESI voltage of 3.5 kV.

Each well of the 384 well destination plate was diluted with 20 μL MeOH. In order to analyze the 384 well destination plate, 24 TLC plates with each 16 spots (one row of the 384 well plate) were developed with petroleum ether - ethyl acetate (1:1, v/v). The developed TLC plates were placed into the plate express. Major spots of interest detected by UV were marked with a soft pencil on the plate and then directly transferred to the CMS. Advion Mass Express and Data Express software was used for mass measurements and data processing.

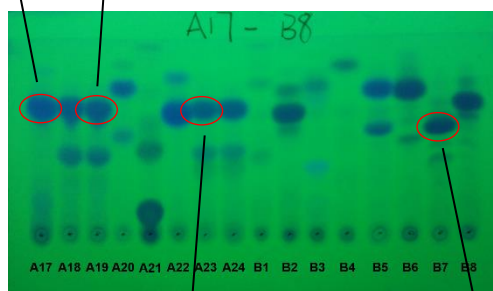
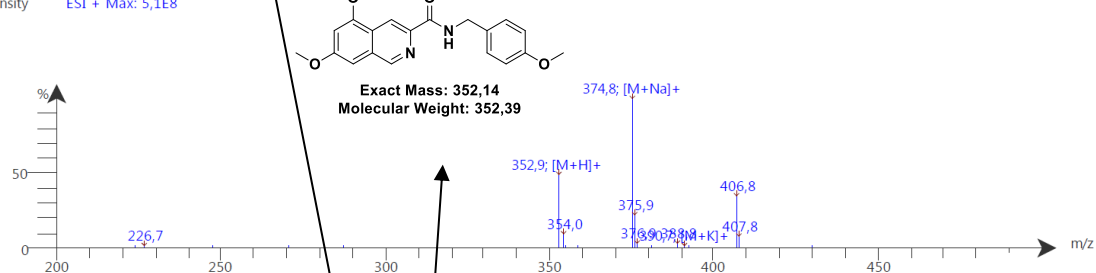
Examples of TLC-UV-MS analytics



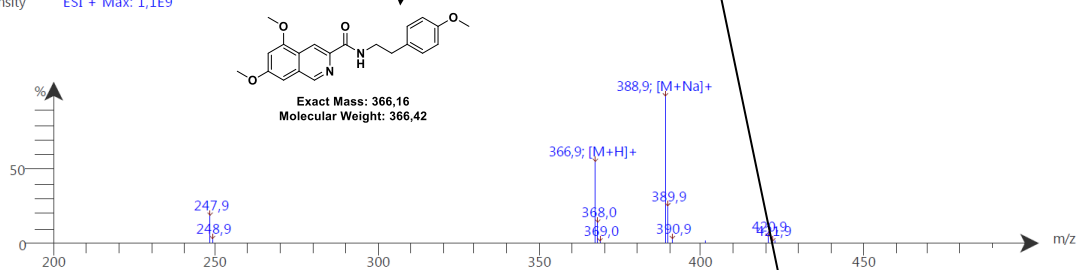
Spectrum RT 0,87 (1 scans) - Background Subtracted 0,02 - 0,68
A17 ;
ESI + Max: 6,5E8



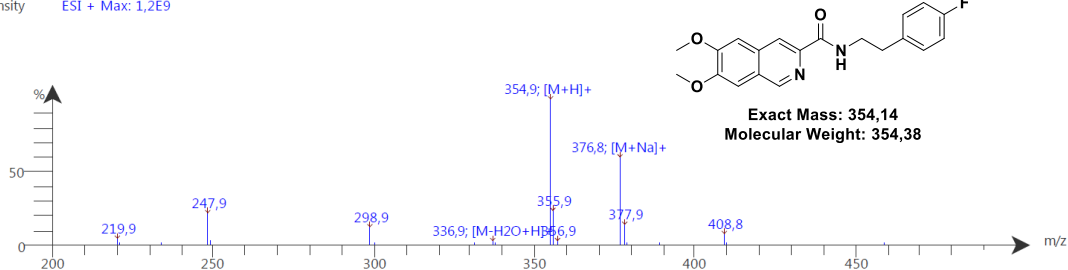
Spectrum RT 0,87 (1 scans) - Background Subtracted 0,03 - 0,76
A19 ;
ESI + Max: 5,1E8



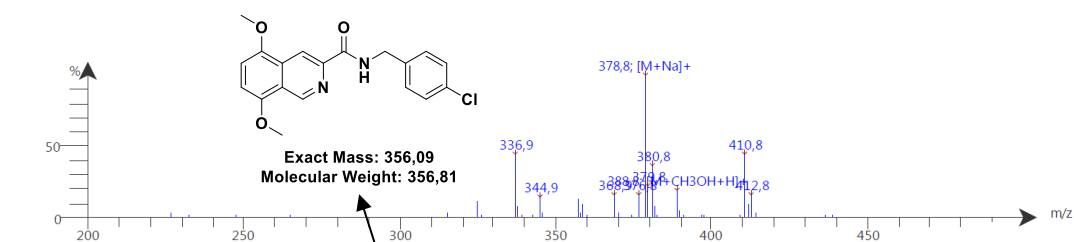
Spectrum RT 0,79 (1 scans) - Background Subtracted 0,02 - 0,65
A23 ;
ESI + Max: 1,1E9



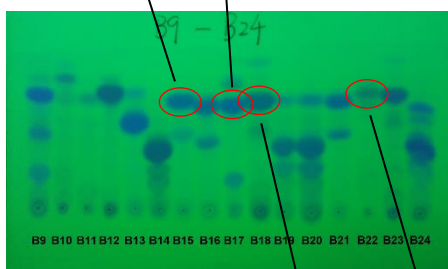
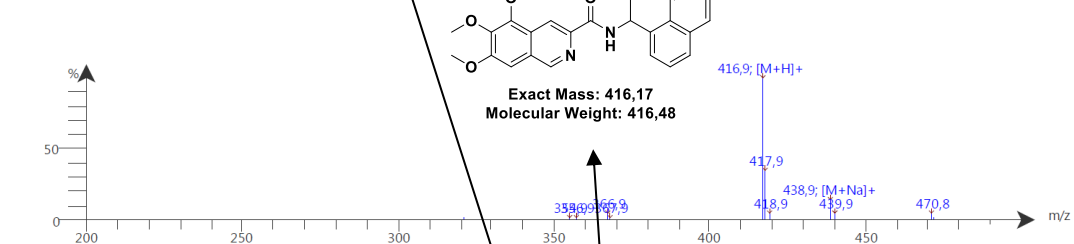
Spectrum RT 0,87 (1 scans) - Background Subtracted 0,02 - 0,69
B7 ;
ESI + Max: 1,2E9



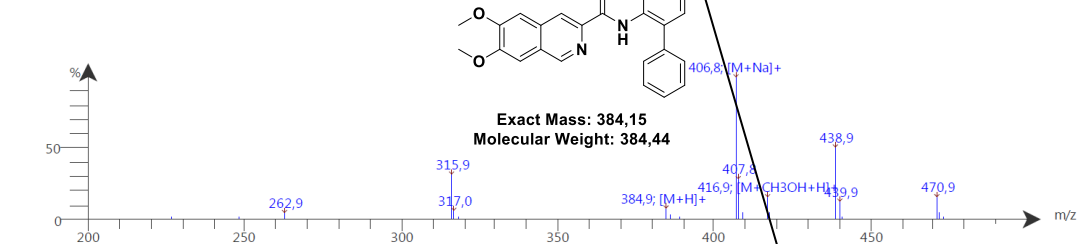
Spectrum RT 0,70 (1 scans) - Background Subtracted 0,02 - 0,64
B15 ;
ESI + Max: 8,3E8



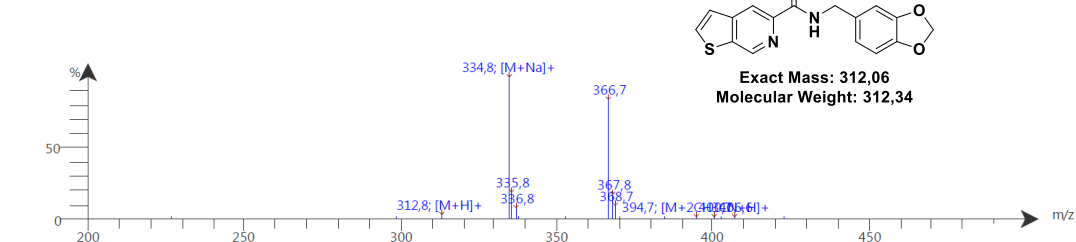
Spectrum RT 0,87 (1 scans) - Background Subtracted 0,02 - 0,79
B17 ;
ESI + Max: 2,1E9



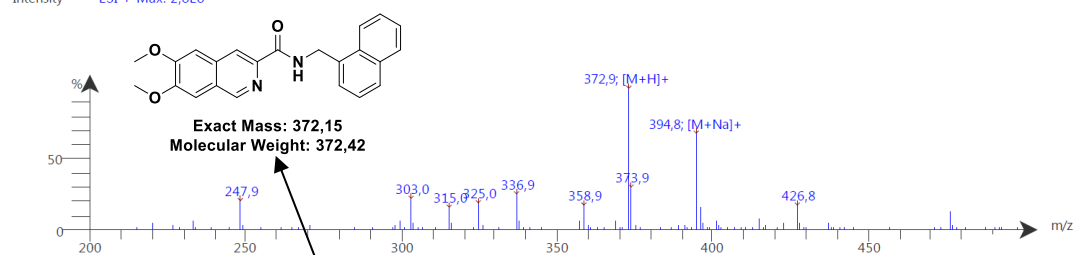
Spectrum RT 0,92 (1 scans) - Background Subtracted 0,03 - 0,80
B18 ;
ESI + Max: 1,1E9



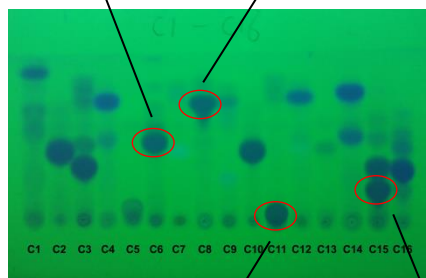
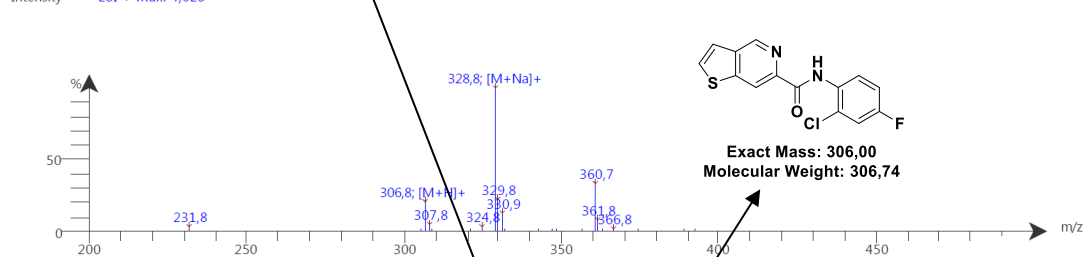
Spectrum RT 0,63 (1 scans) - Background Subtracted 0,01 - 0,56
B22 ;
ESI + Max: 7E8



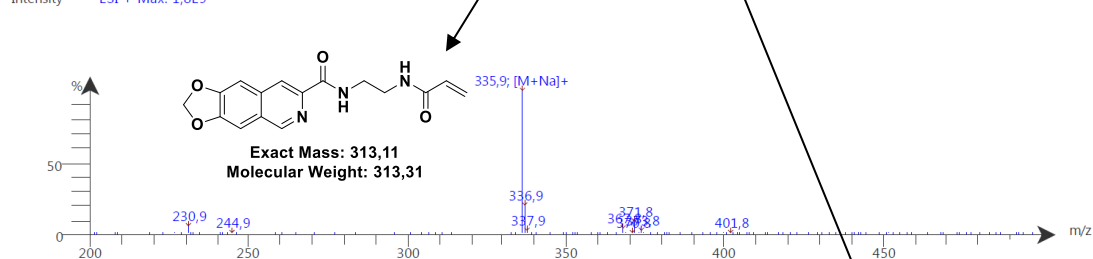
Spectrum RT 0,96 (1 scans) - Background Subtracted 0,02 - 0,86
C6 ;
ESI + Max: 2,8E8



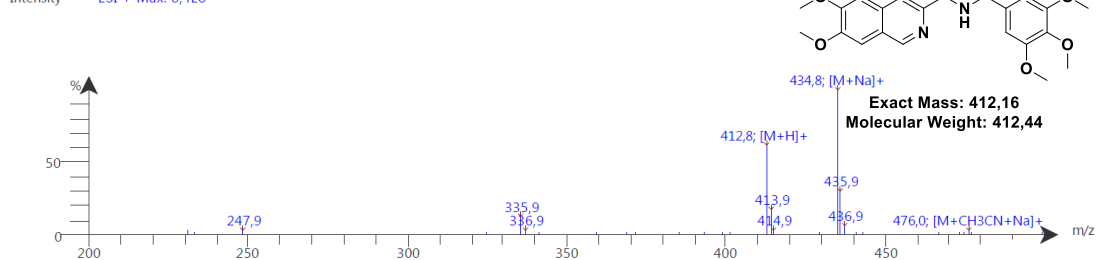
Spectrum RT 0,58 (1 scans) - Background Subtracted 0,73 - 0,76
C8 ;
ESI + Max: 4,6E8

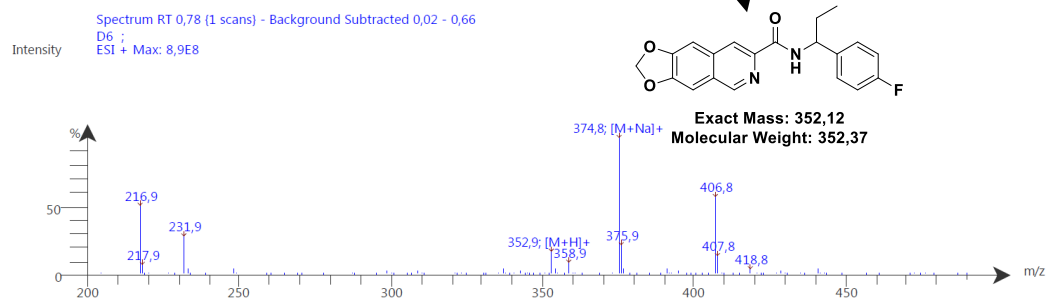
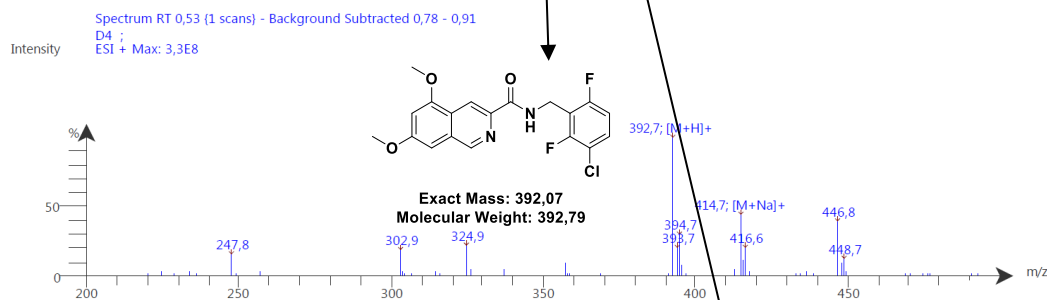
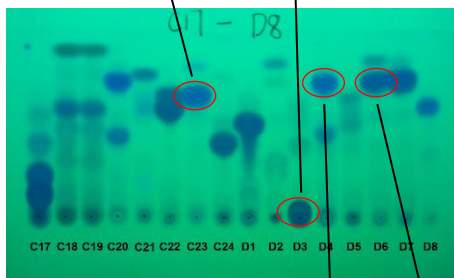
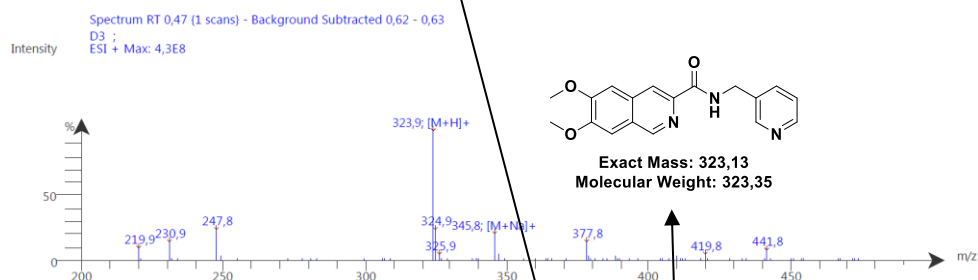
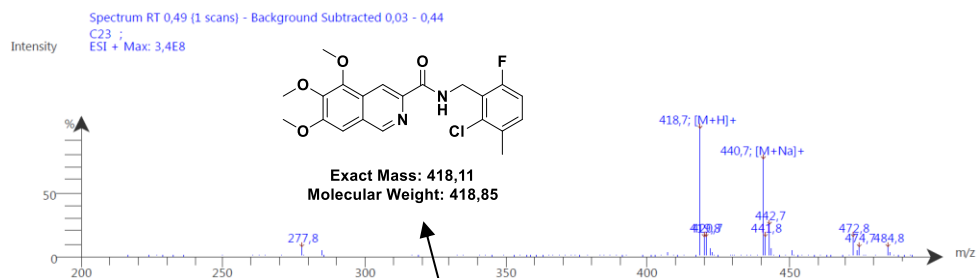


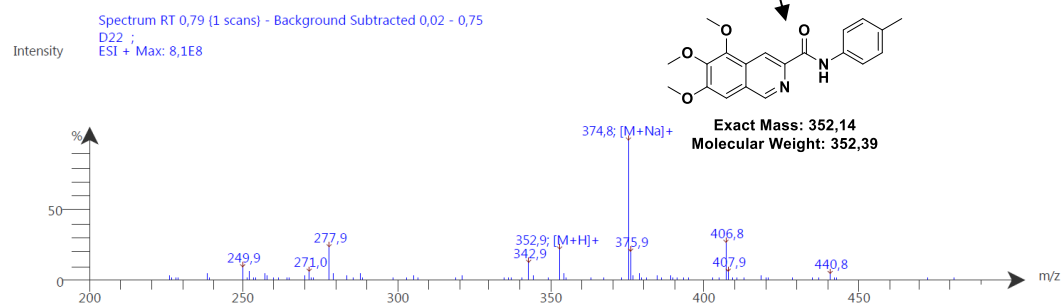
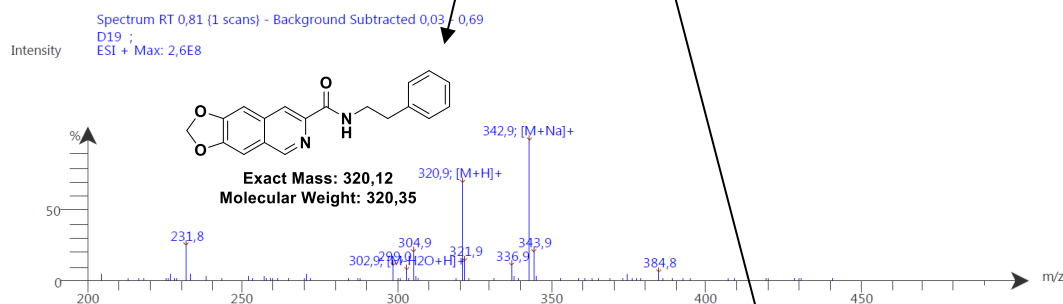
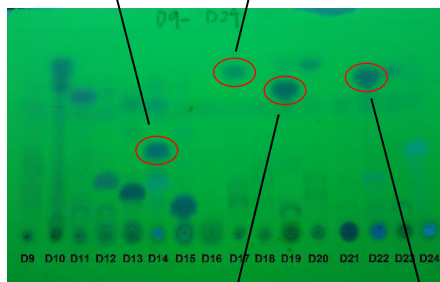
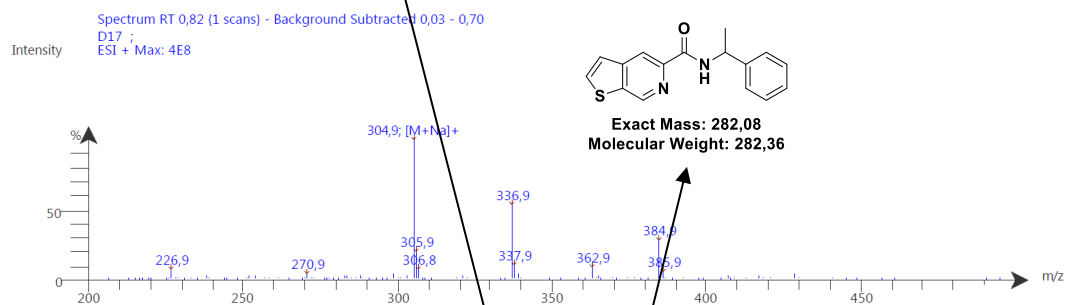
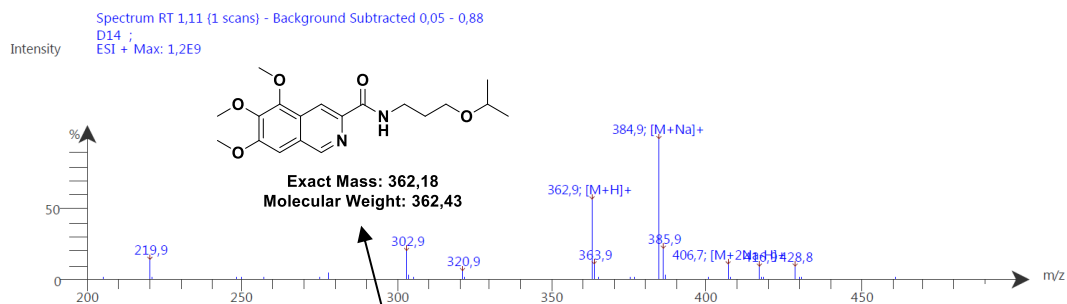
Spectrum RT 0,69 (1 scans) - Background Subtracted 0,03 - 0,59
C11 ;
ESI + Max: 1,8E9



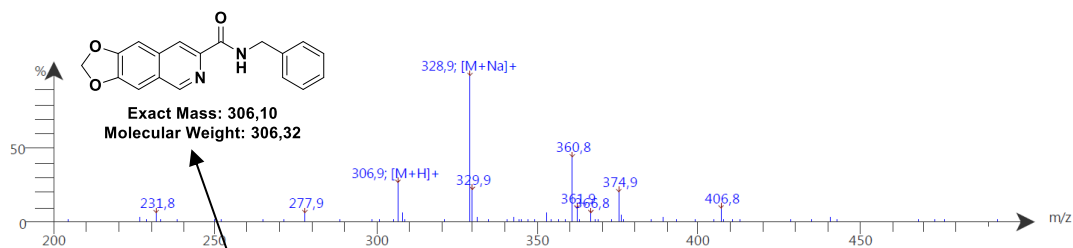
Spectrum RT 0,75 (1 scans) - Background Subtracted 0,03 - 0,65
C15 ;
ESI + Max: 8,4E8



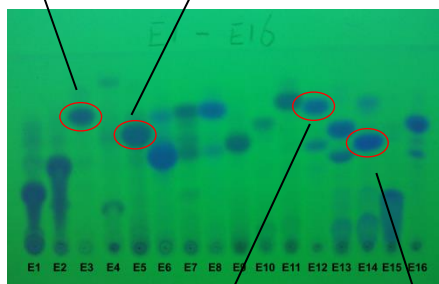
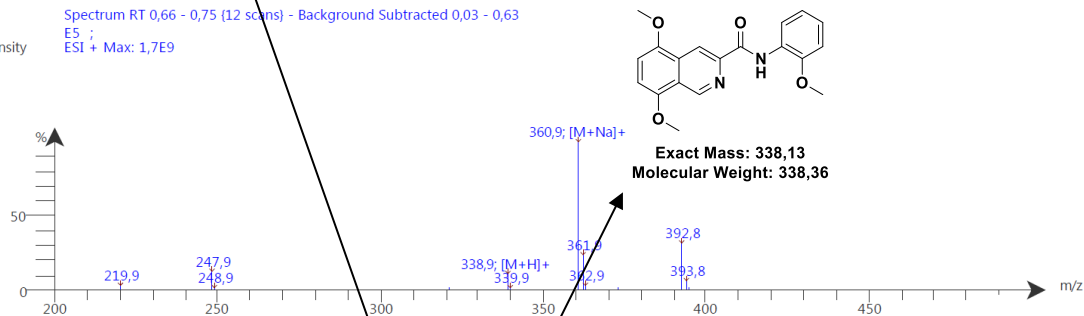




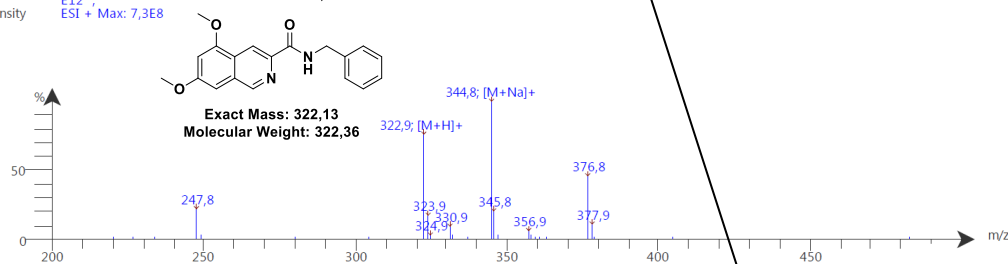
Spectrum RT 0,77 (1 scans) - Background Subtracted 0,03 - 0,66
E3 ;
ESI + Max: 5,8E8



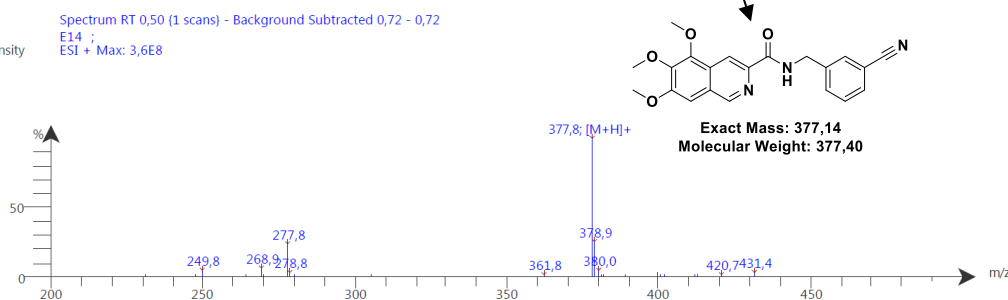
Spectrum RT 0,66 - 0,75 (12 scans) - Background Subtracted 0,03 - 0,63
E5 ;
ESI + Max: 1,7E9

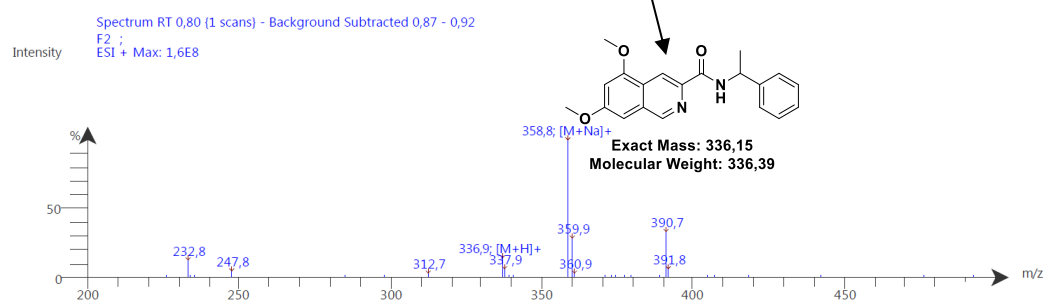
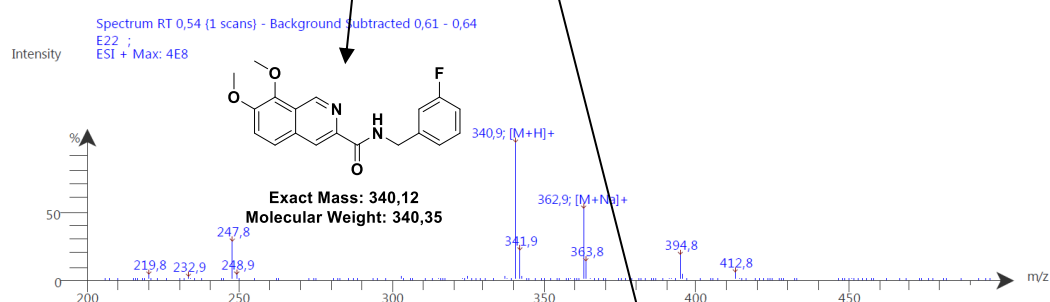
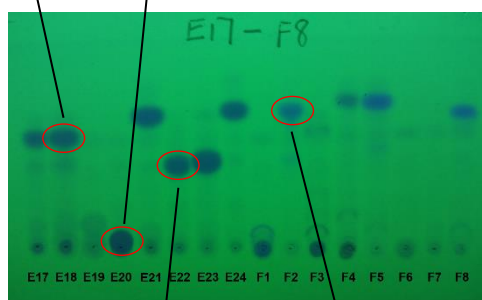
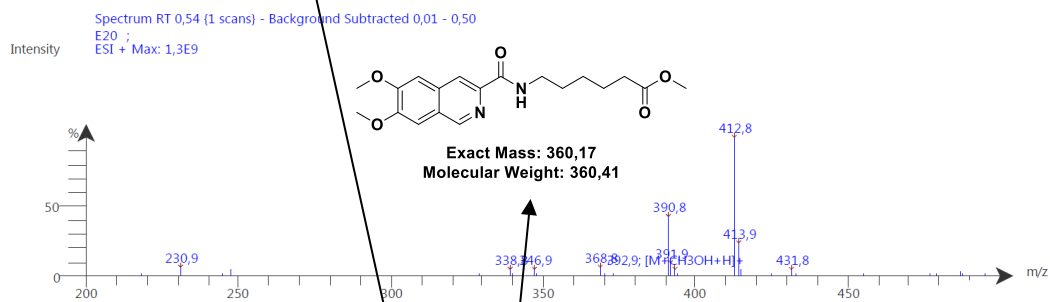
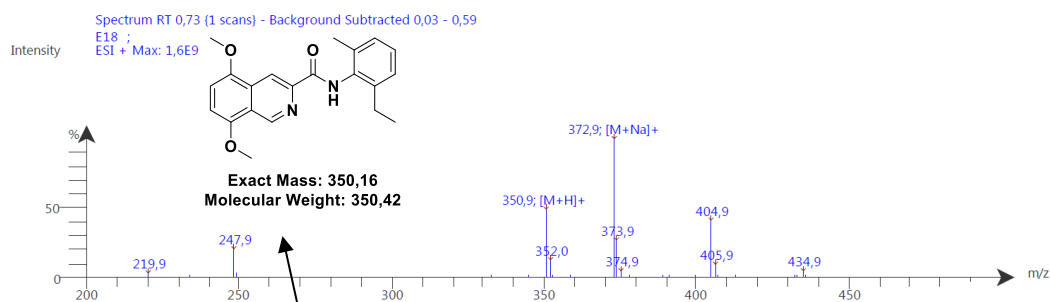


Spectrum RT 0,52 (1 scans) - Background Subtracted 0,01 - 0,49
E12 ;
ESI + Max: 7,3E8

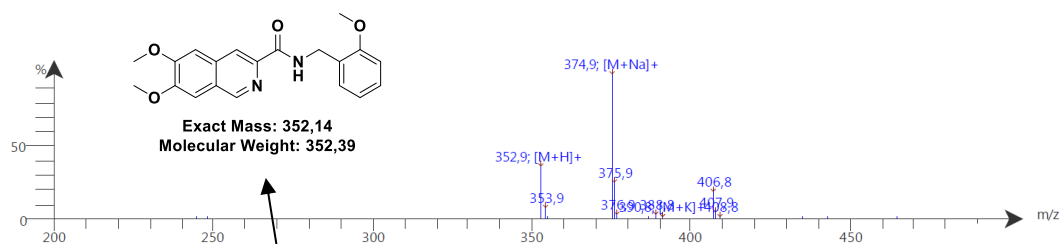


Spectrum RT 0,50 (1 scans) - Background Subtracted 0,72 - 0,72
E14 ;
ESI + Max: 3,6E8

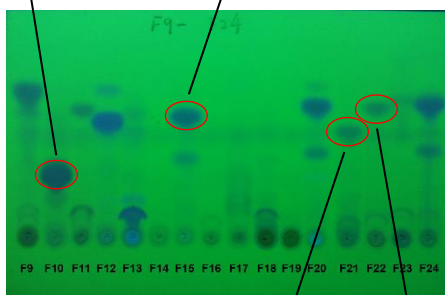
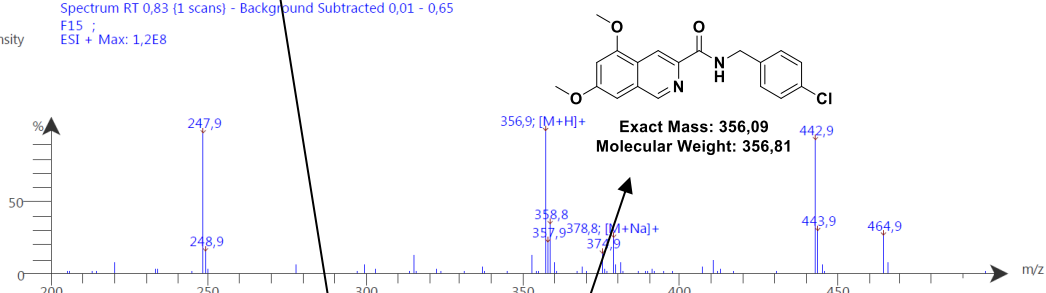




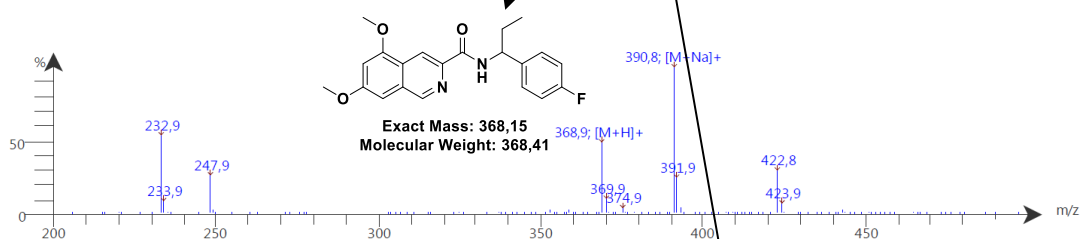
Spectrum RT 0,77 (1 scans) - Background Subtracted 0,03 - 0,69
F10 ;
ESI + Max: 1,9E9



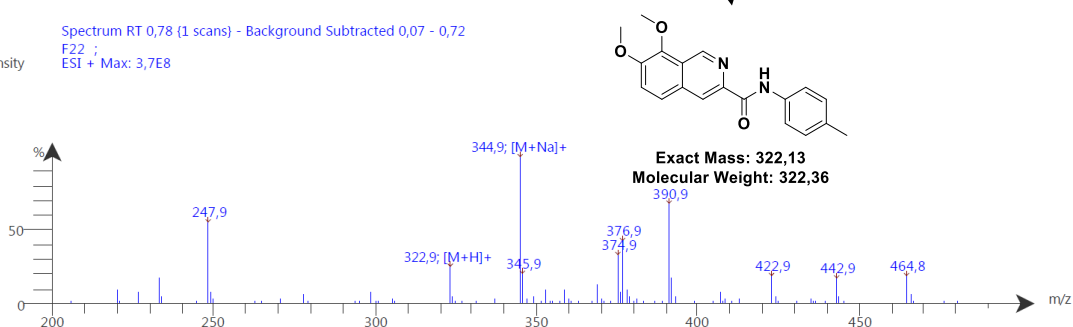
Spectrum RT 0,83 (1 scans) - Background Subtracted 0,01 - 0,65
F15 ;
ESI + Max: 1,2E8



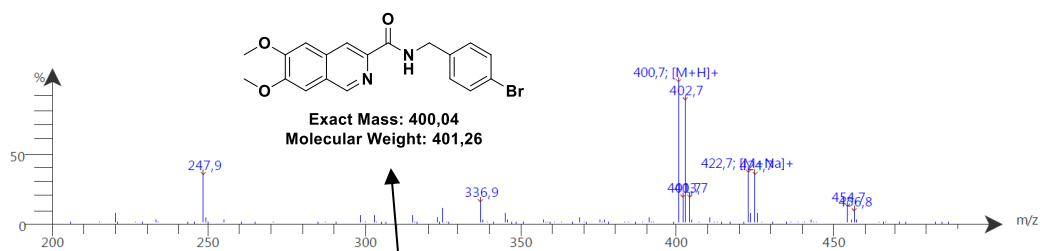
Spectrum RT 0,77 (1 scans) - Background Subtracted 0,01 - 0,75
F20 ;
ESI + Max: 1,7E9



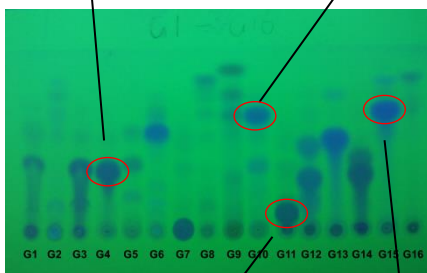
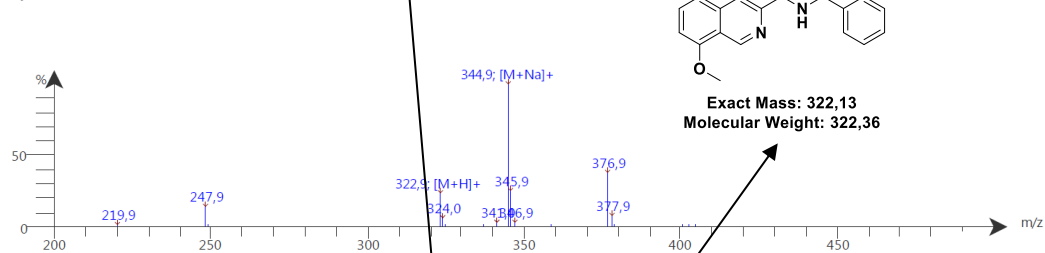
Spectrum RT 0,78 (1 scans) - Background Subtracted 0,07 - 0,72
F22 ;
ESI + Max: 3,7E8



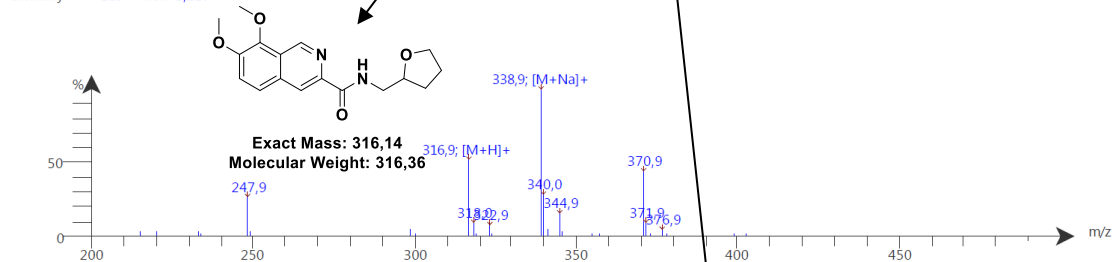
Spectrum RT 0,83 (1 scans) - Background Subtracted 0,03 - 0,72
G4 ;
ESI + Max: 4,2E8



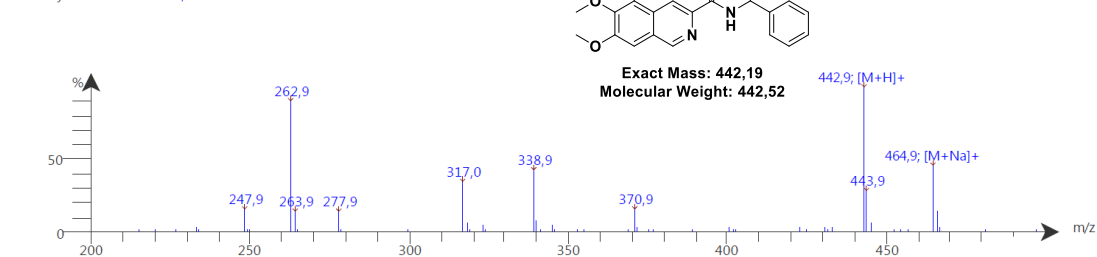
Spectrum RT 0,73 (1 scans) - Background Subtracted 0,01 - 0,68
G10 ;
ESI + Max: 2,2E9

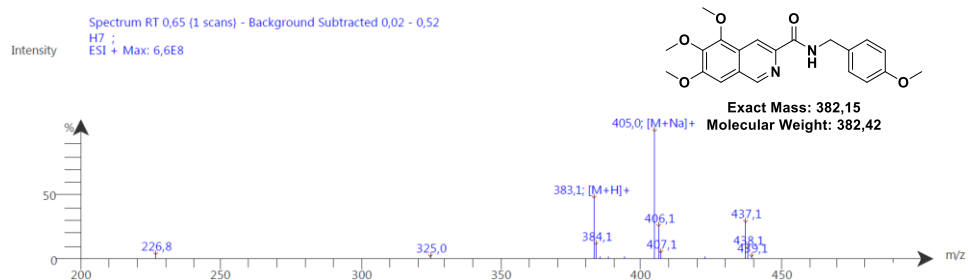
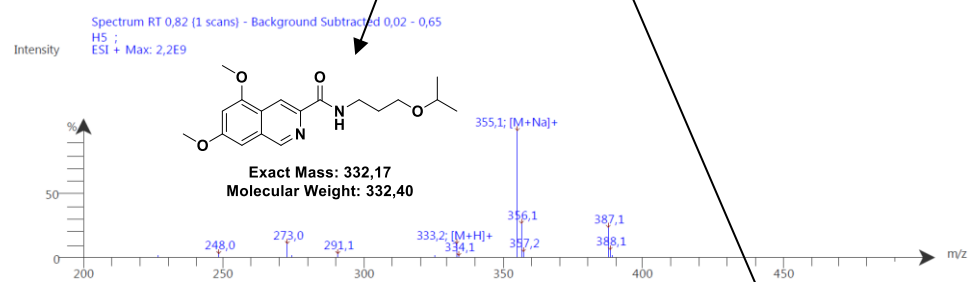
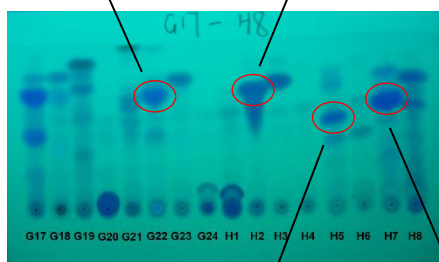
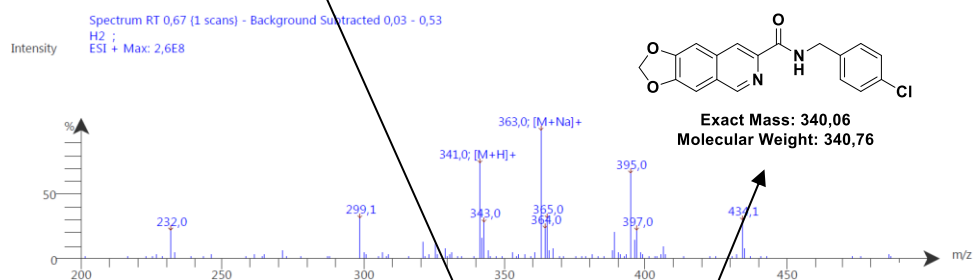
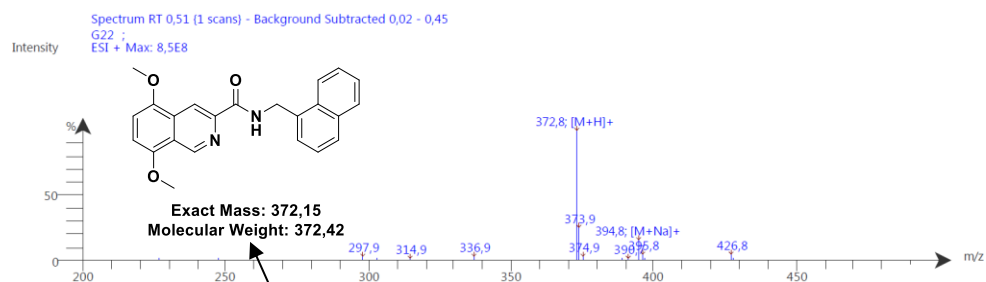


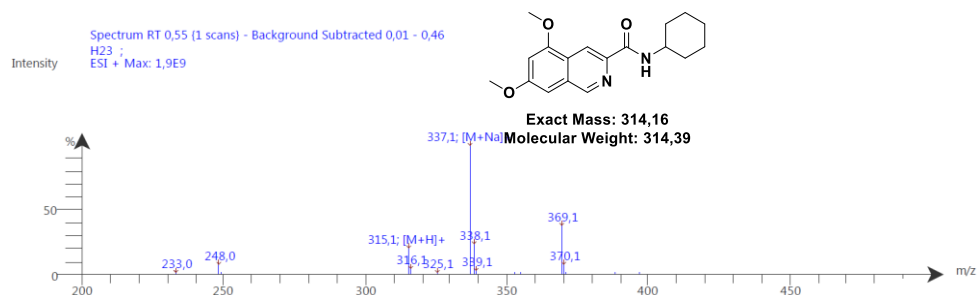
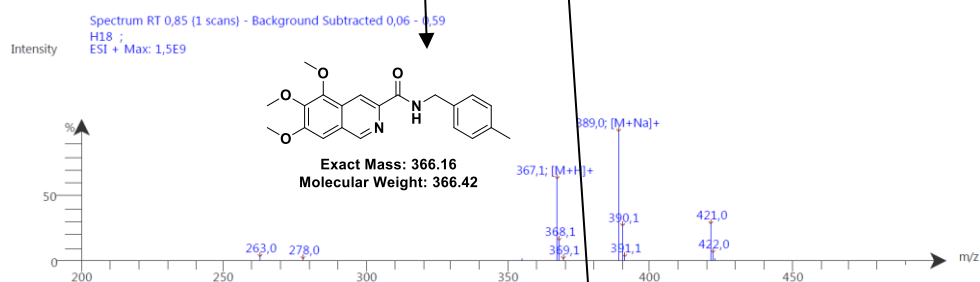
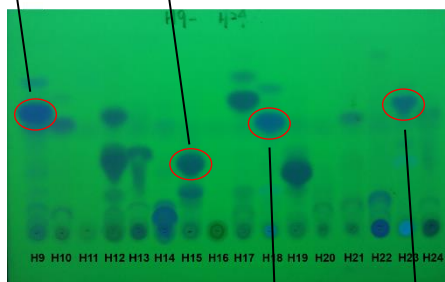
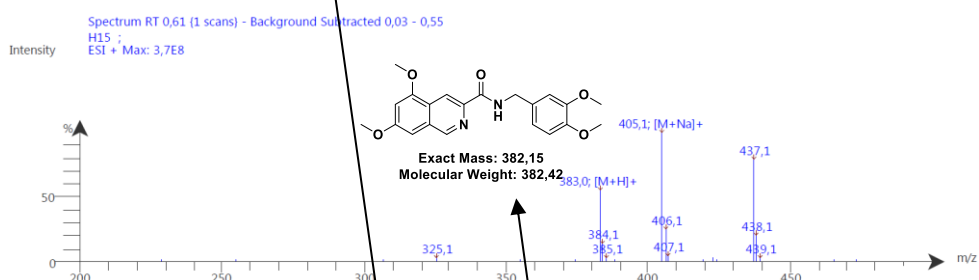
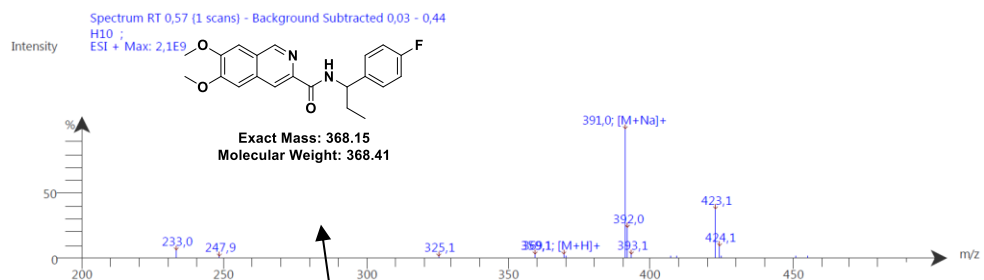
Spectrum RT 0,80 (1 scans) - Background Subtracted 0,03 - 0,71
G11 ;
ESI + Max: 2,2E9

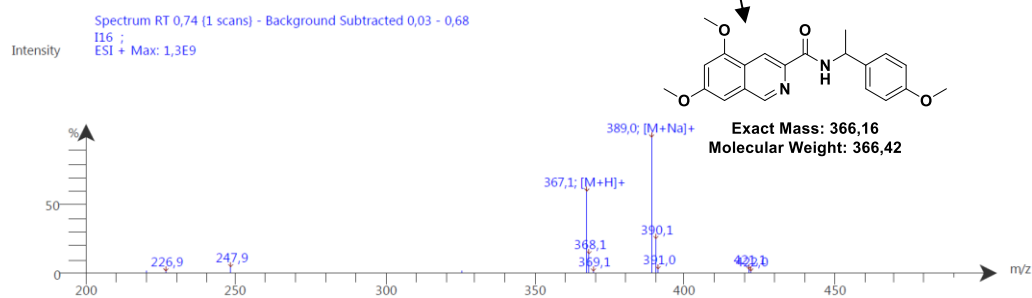
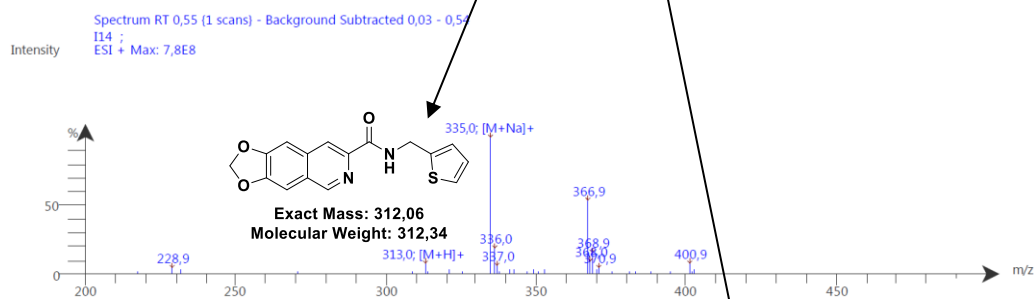
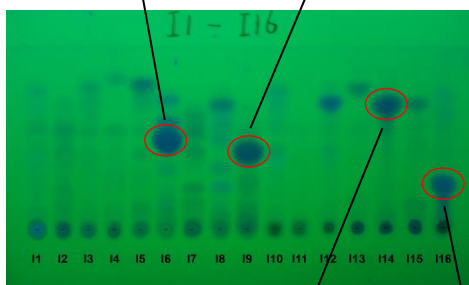
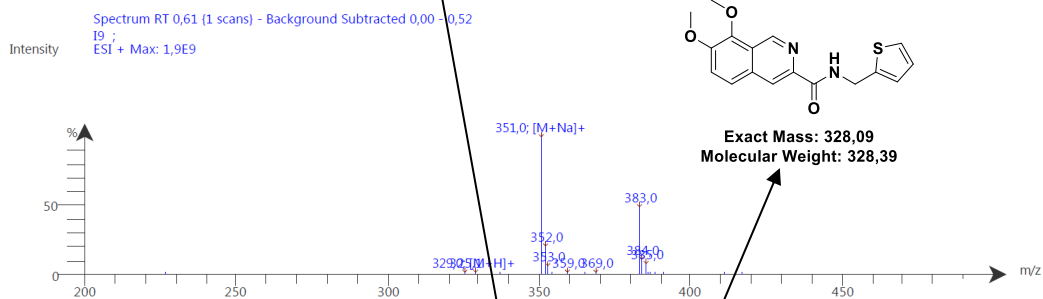
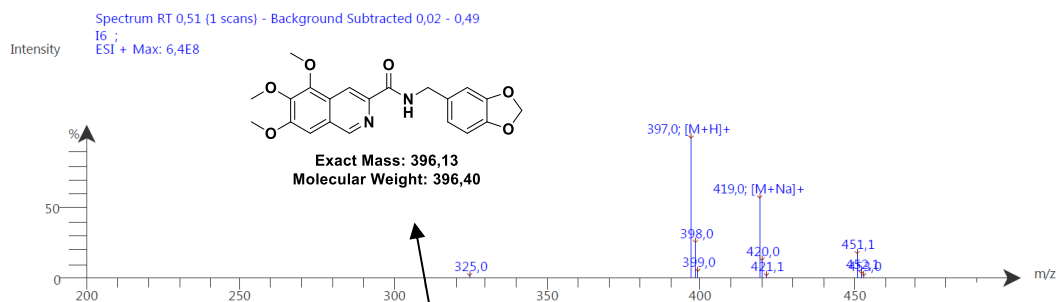


Spectrum RT 1,09 (1 scans) - Background Subtracted 0,00 - 1,05
G15 ;
ESI + Max: 5,8E8

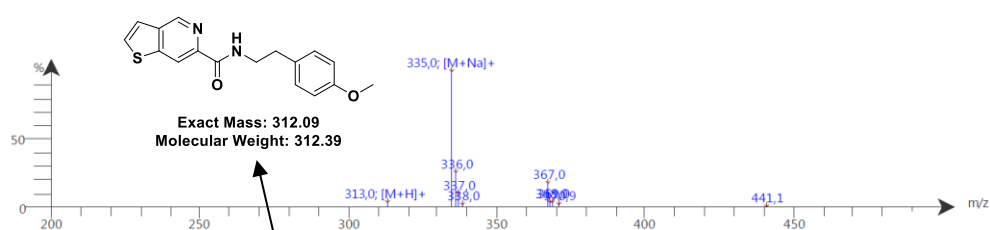




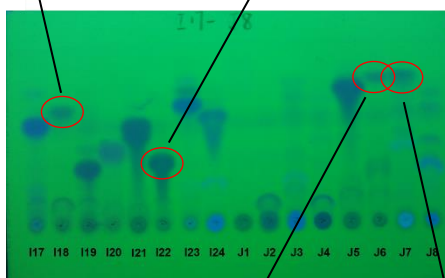
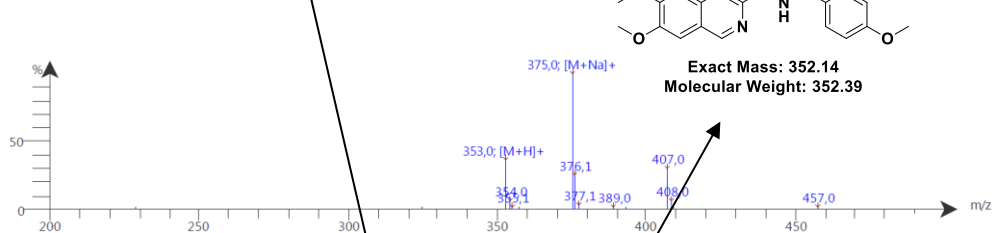




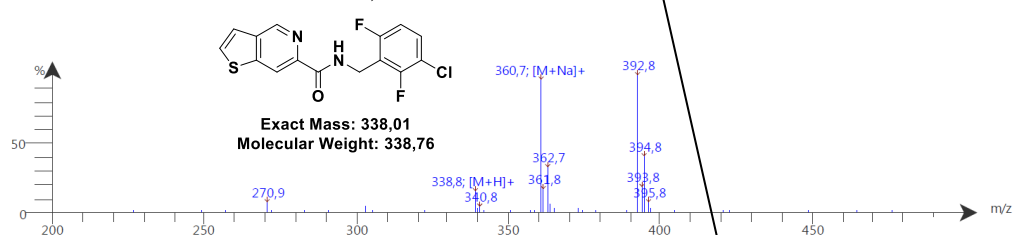
Spectrum RT 0,58 (1 scans) - Background Subtracted 0,02 - 0,51
I18 ;
ESI + Max: 2,2E9



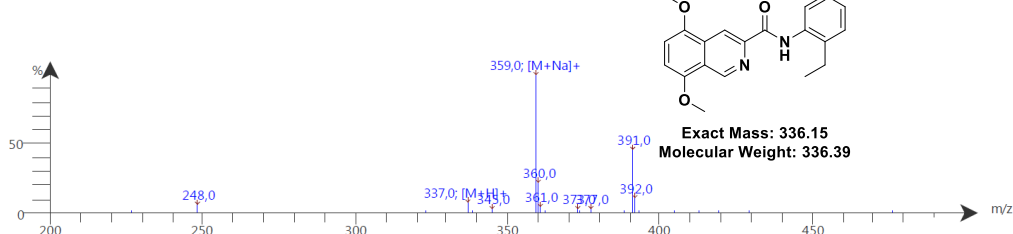
Spectrum RT 0,53 (1 scans) - Background Subtracted 0,03 - 0,48
I22 ;
ESI + Max: 8,4E8

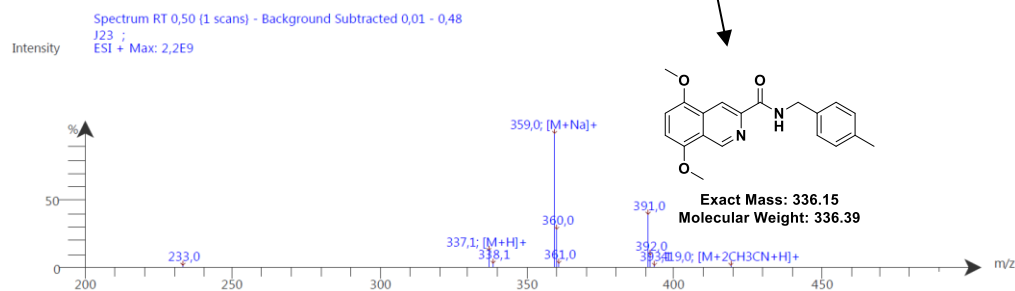
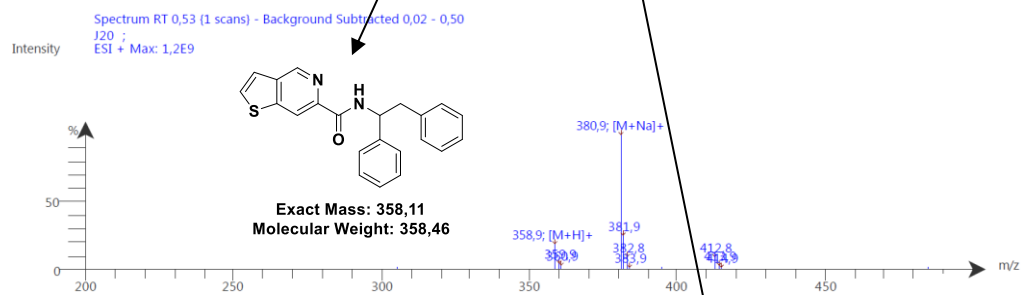
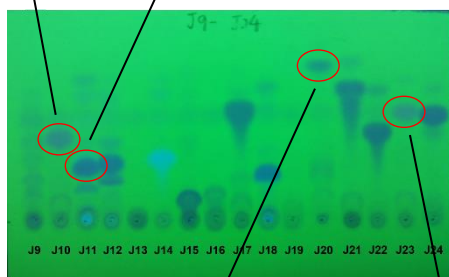
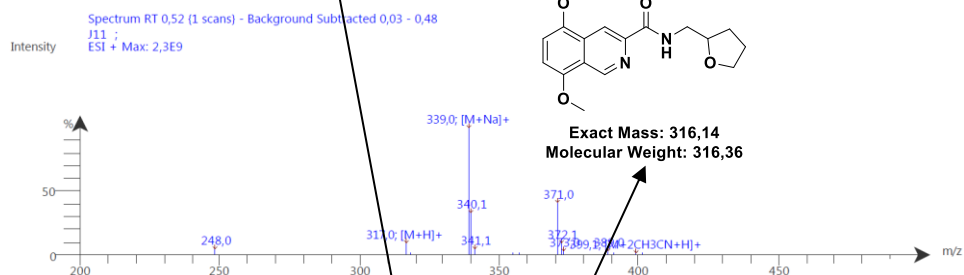
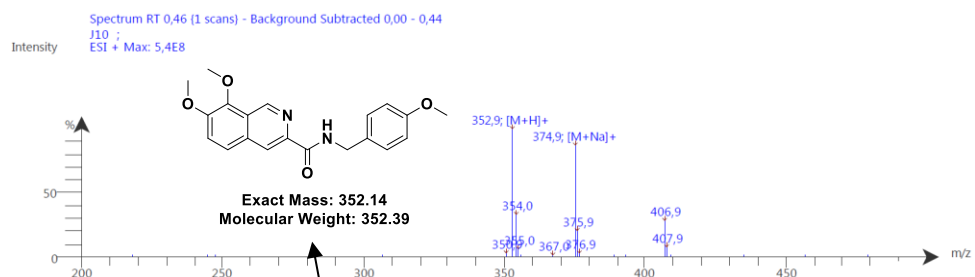


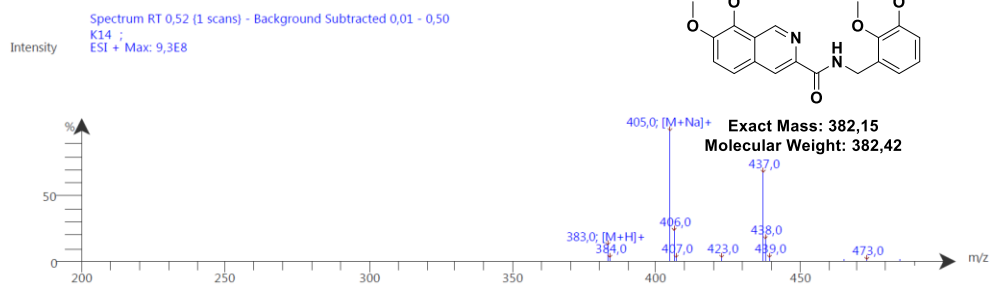
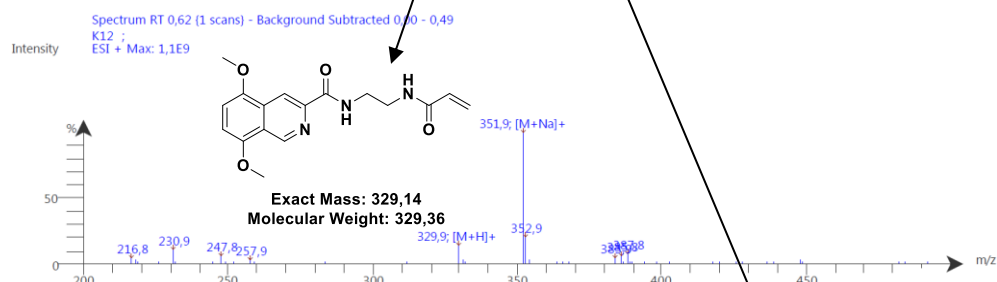
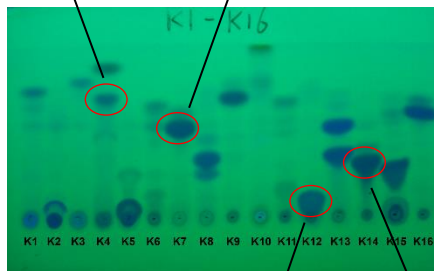
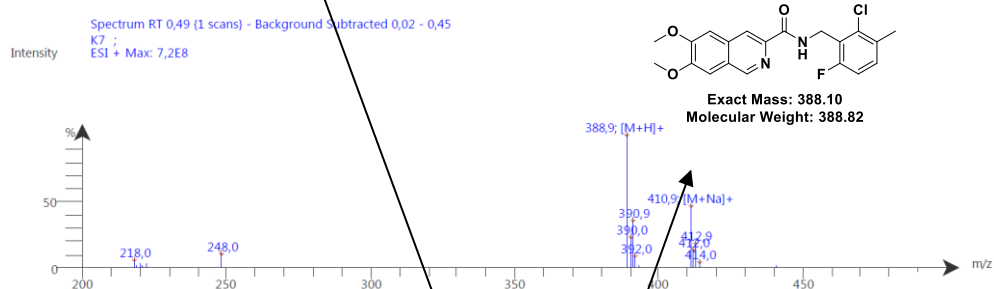
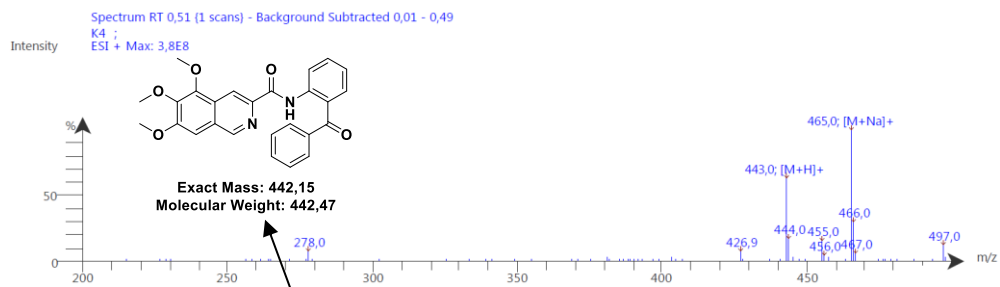
Spectrum RT 0,52 (1 scans) - Background Subtracted 0,03 - 0,48
J6 ;
ESI + Max: 7,9E8



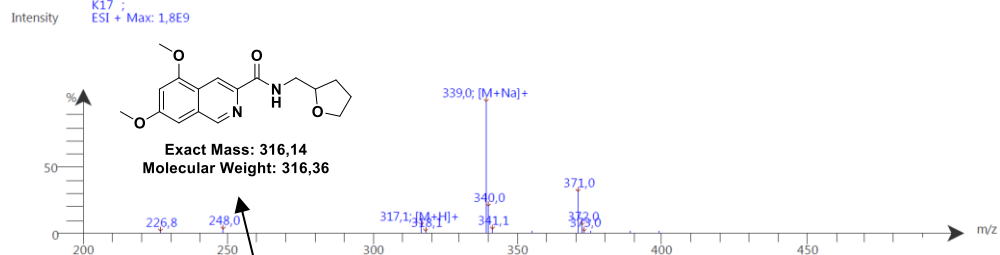
Spectrum RT 1,29 (1 scans) - Background Subtracted 0,13 - 1,23
J7 ;
ESI + Max: 1E9



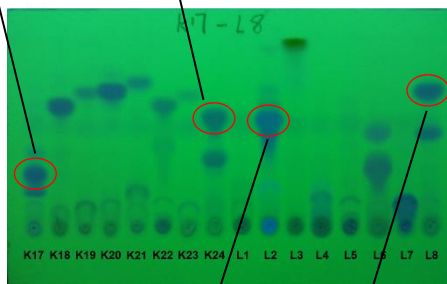
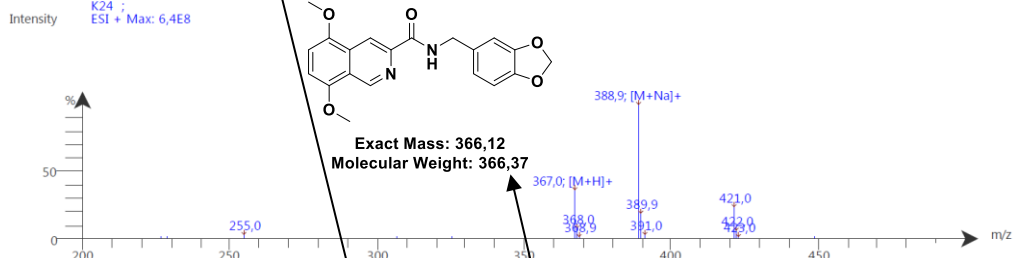




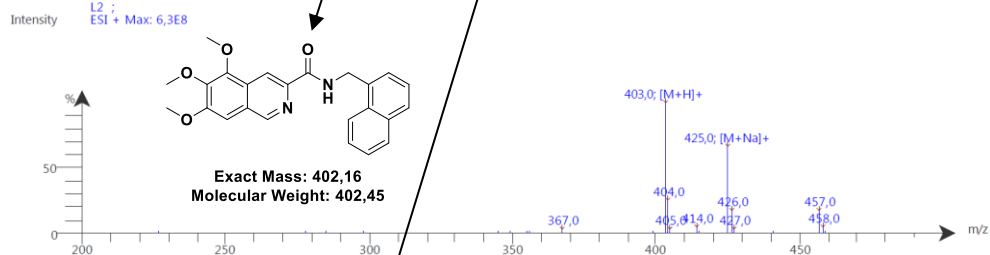
Spectrum RT 0,70 (1 scans) - Background Subtracted 0,02 - 0,46
K17 ;
ESI + Max: 1,8E9



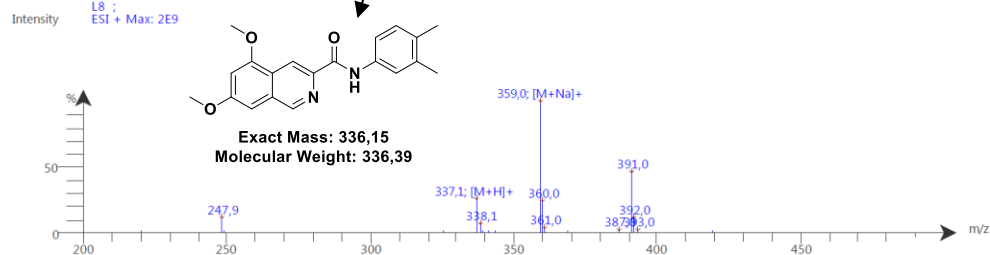
Spectrum RT 0,51 (1 scans) - Background Subtracted 0,08 - 0,49
K24 ;
ESI + Max: 6,4E8



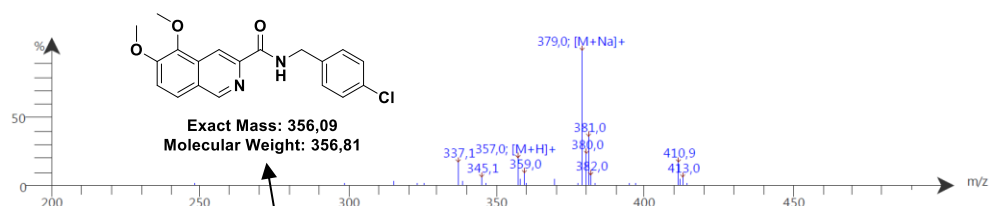
Spectrum RT 0,52 (1 scans) - Background Subtracted 0,03 - 0,48
L2 ;
ESI + Max: 6,3E8



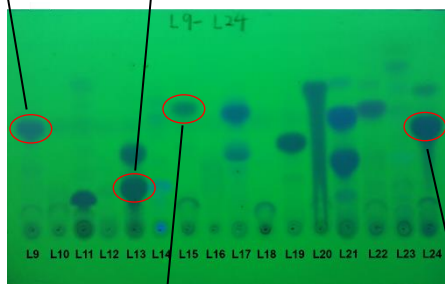
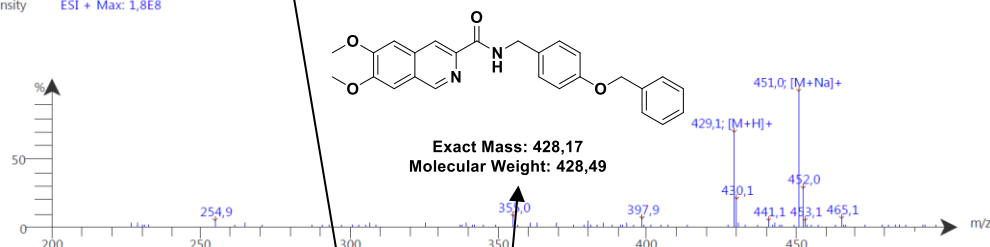
Spectrum RT 0,50 (1 scans) - Background Subtracted 0,04 - 0,45
L8 ;
ESI + Max: 2E9



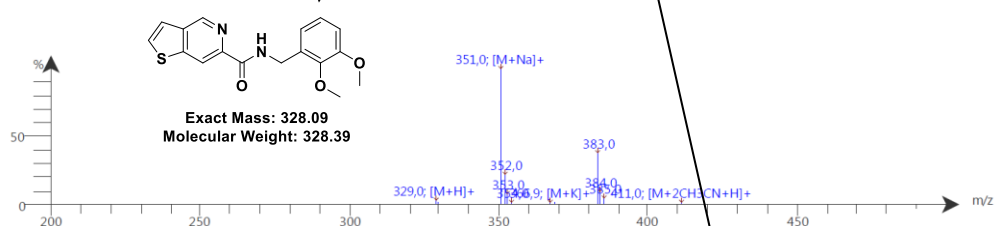
Spectrum RT 0,55 (1 scans) - Background Subtracted 0,02 - 0,52
L9 ;
ESI + Max: 1,8E9



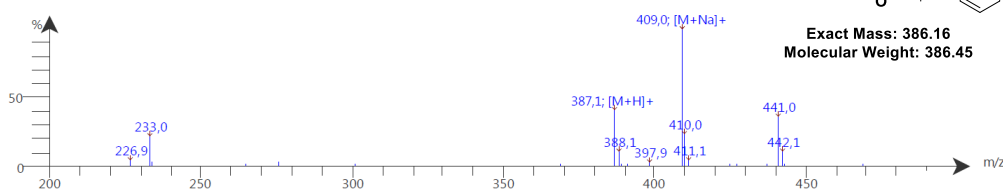
Spectrum RT 0,55 (1 scans) - Background Subtracted 0,01 - 0,50
L13 ;
ESI + Max: 1,8E8



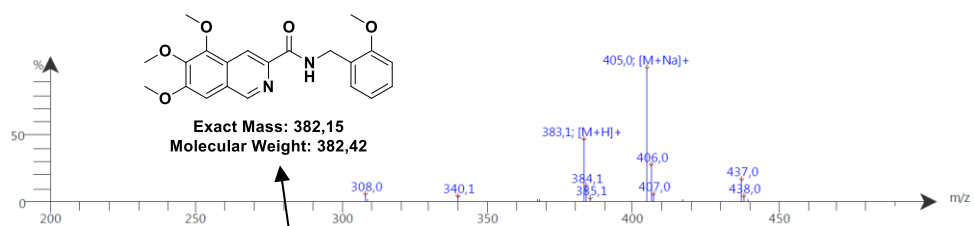
Spectrum RT 0,55 (1 scans) - Background Subtracted 0,01 - 0,53
L15 ;
ESI + Max: 2,1E9



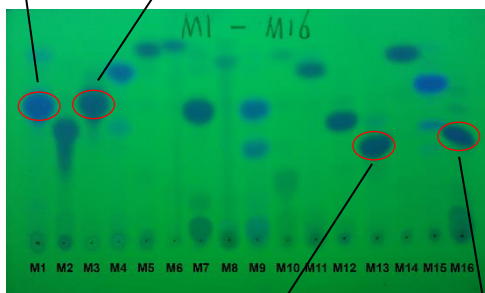
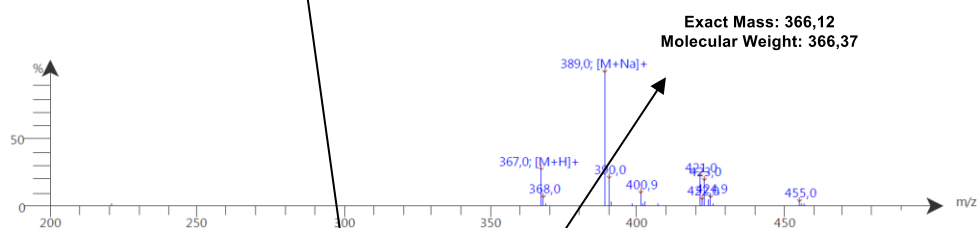
Spectrum RT 0,61 (1 scans) - Background Subtracted 0,03 - 0,48
L22 ;
ESI + Max: 5,6E8



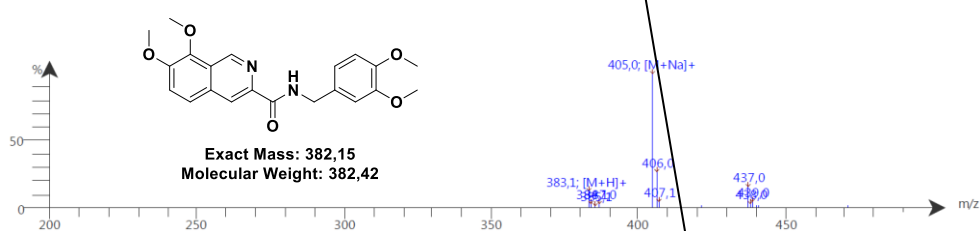
Spectrum RT 0,52 (1 scans) - Background Subtracted 0,01 - 0,47
M1 ;
ESI + Max: 1,2E9



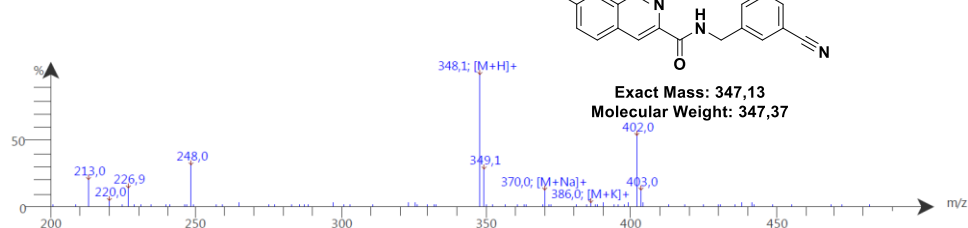
Spectrum RT 0,51 (1 scans) - Background Subtracted 0,03 - 0,45
M3 ;
ESI + Max: 8,6E8

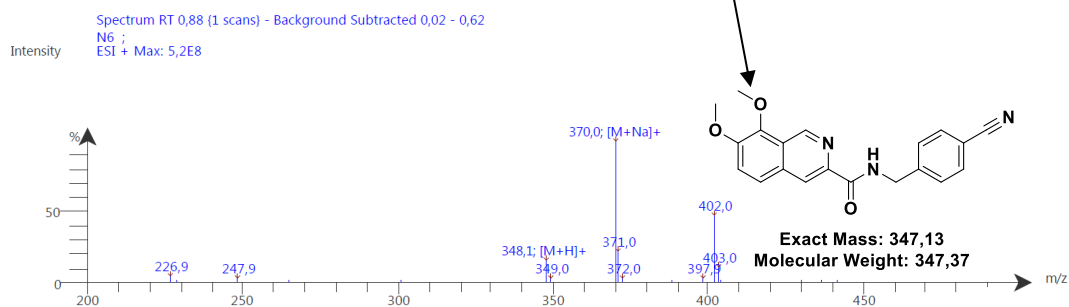
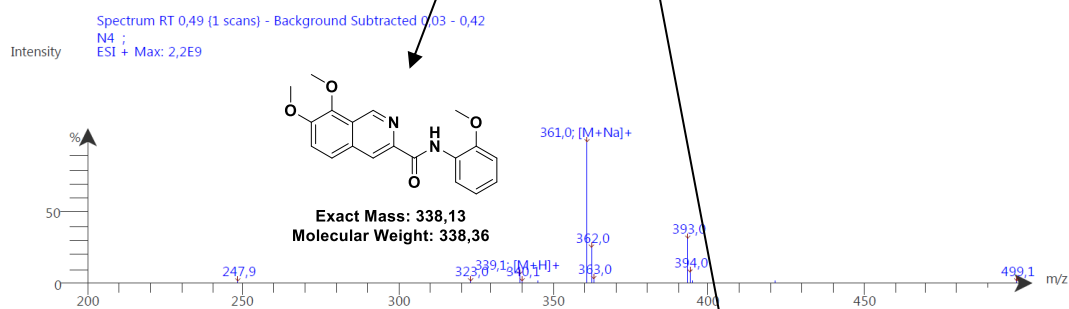
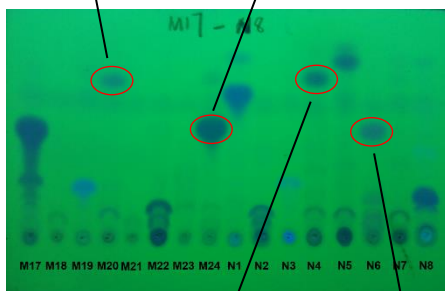
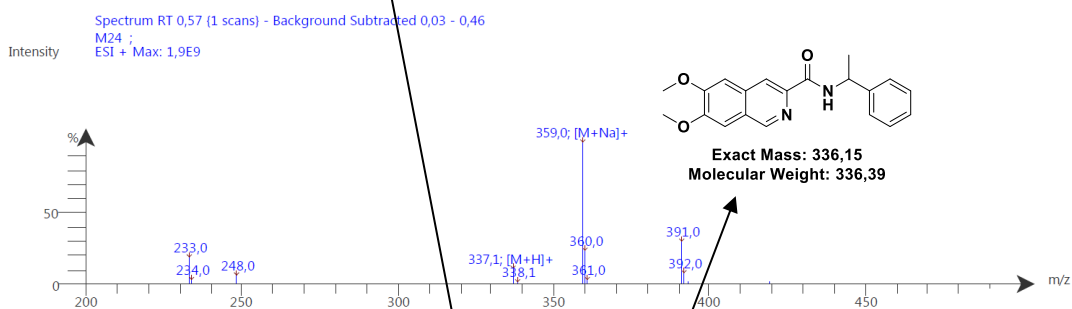
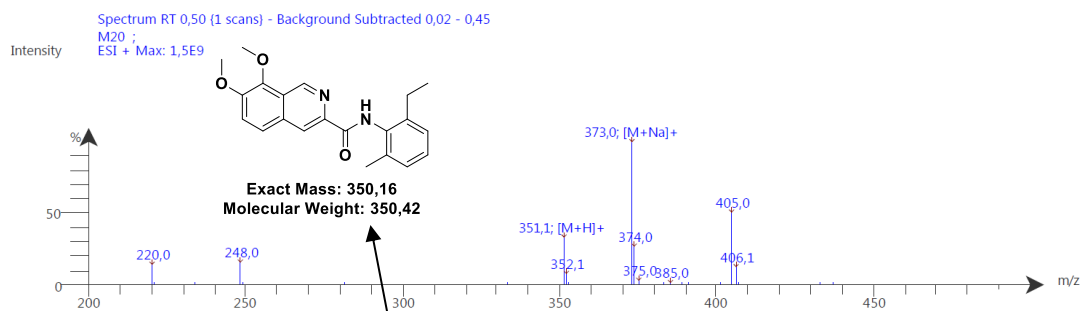


Spectrum RT 0,55 (1 scans) - Background Subtracted 0,02 - 0,49
M13 ;
ESI + Max: 1,8E9

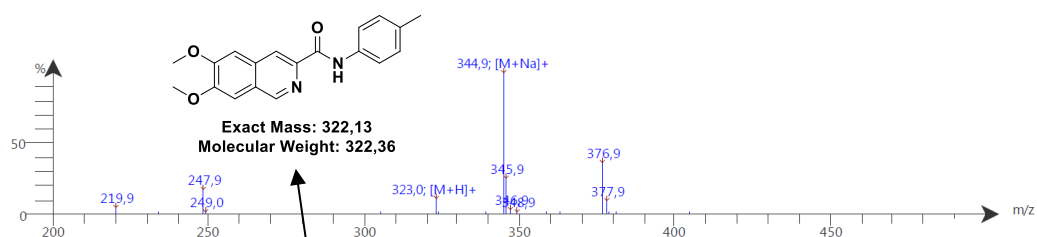


Spectrum RT 0,85 (1 scans) - Background Subtracted 0,03 - 0,47
M16 ;
ESI + Max: 6,6E7

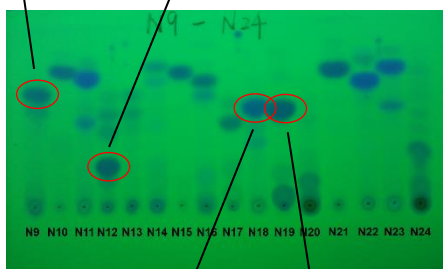
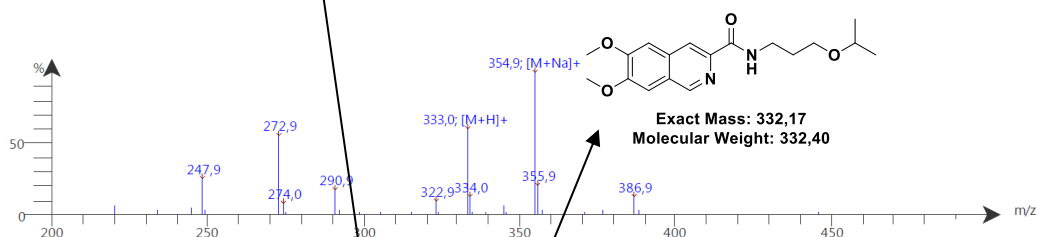




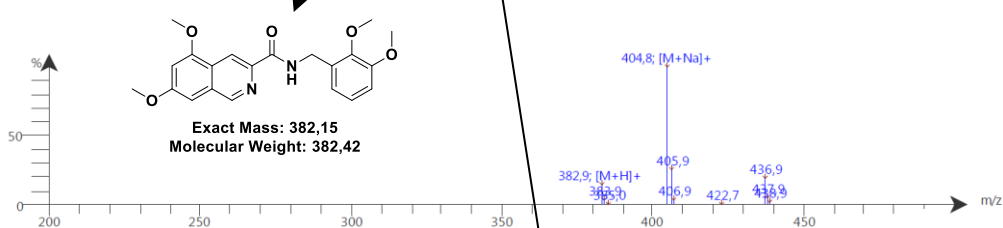
Spectrum RT 0,77 (1 scans) - Background Subtracted 0,05 - 0,60
N9 ;
ESI + Max: 2,1E9



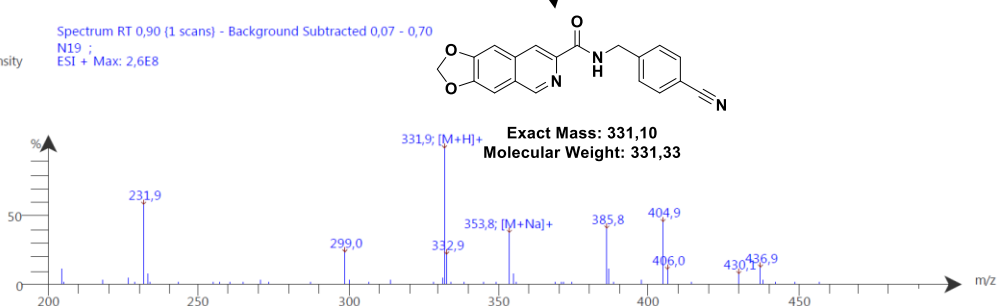
Spectrum RT 0,84 - 0,86 (3 scans) - Background Subtracted 0,03 - 0,78
N12 ;
ESI + Max: 1,3E9

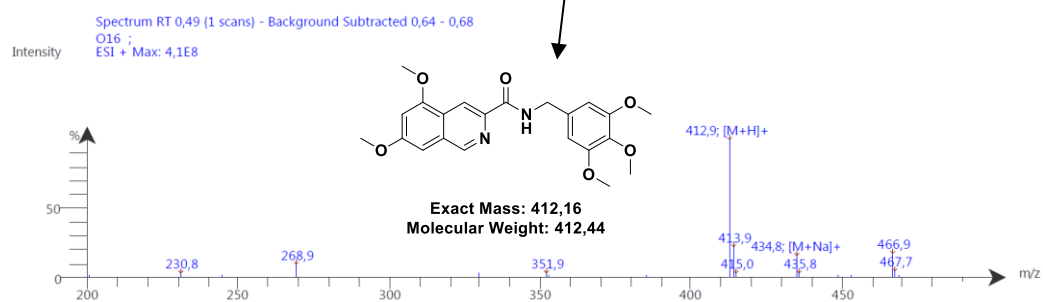
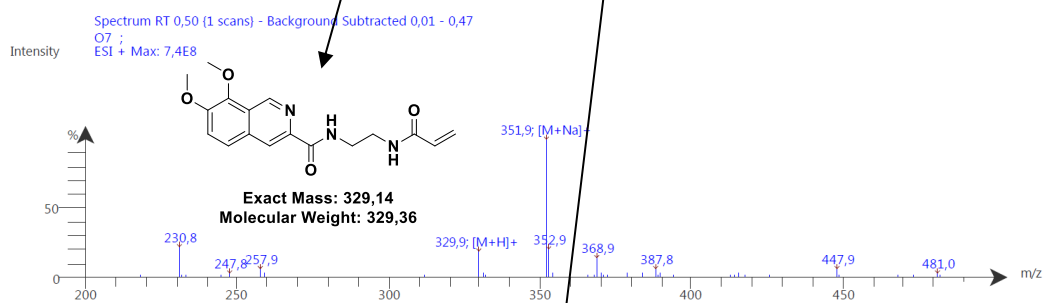
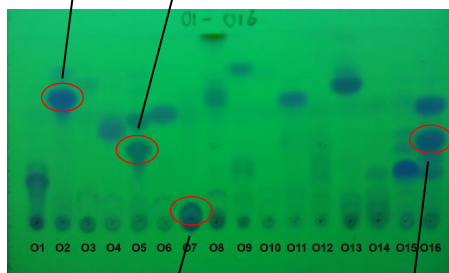
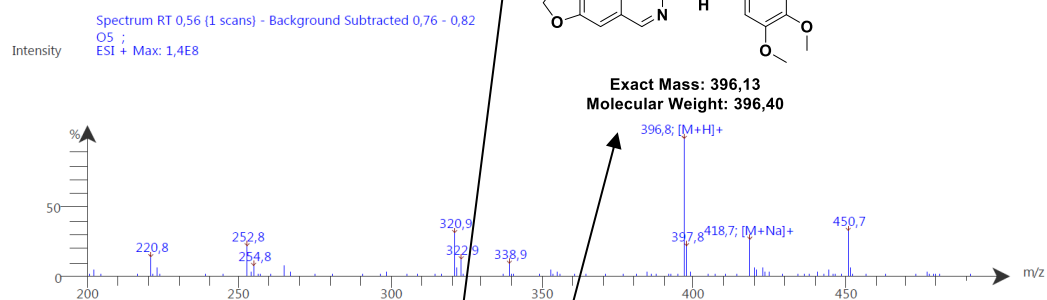
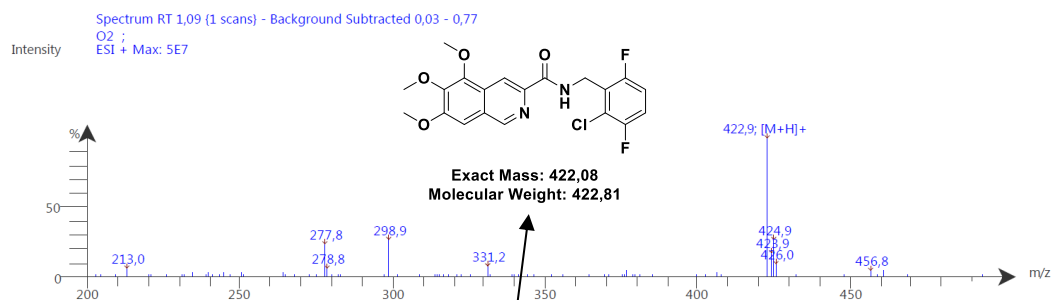


Spectrum RT 0,83 (1 scans) - Background Subtracted 0,03 - 0,71
N18 ;
ESI + Max: 2E9

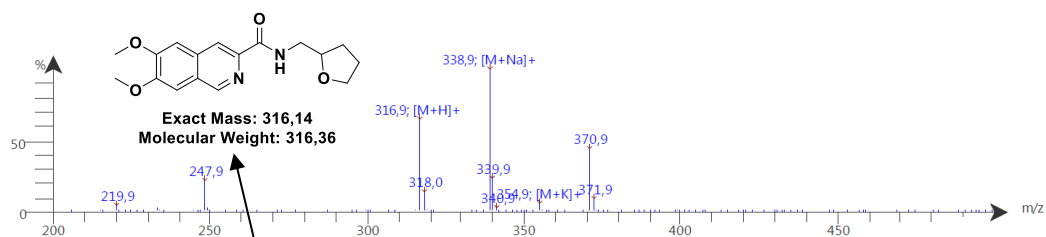


Spectrum RT 0,90 (1 scans) - Background Subtracted 0,07 - 0,70
N19 ;
ESI + Max: 2,6E8

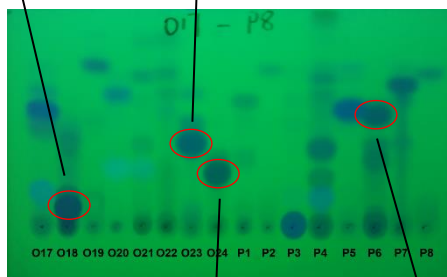
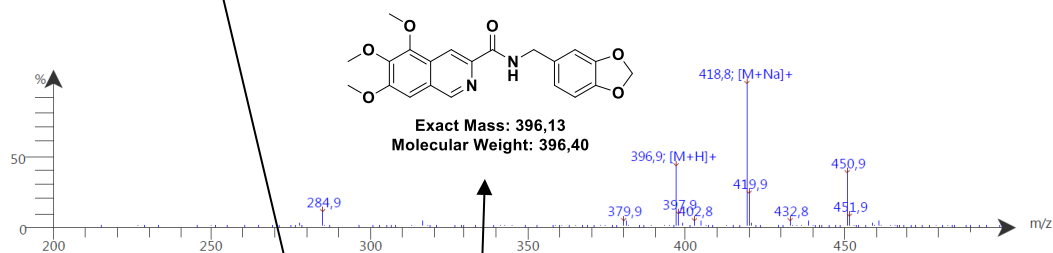




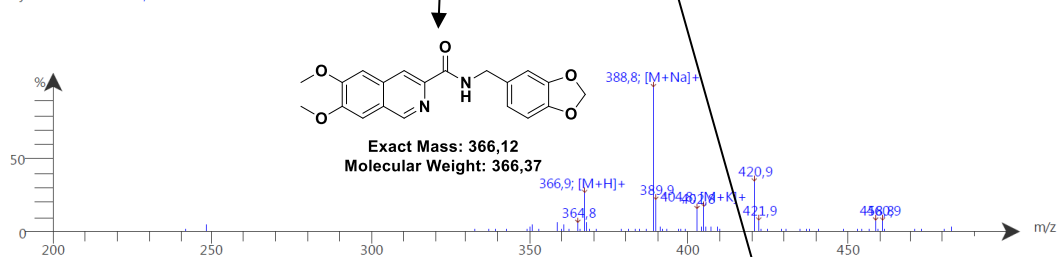
Spectrum RT 0,92 (1 scans) - Background Subtracted 0,05 - 0,72
O18 ;
ESI + Max: 2,1E9



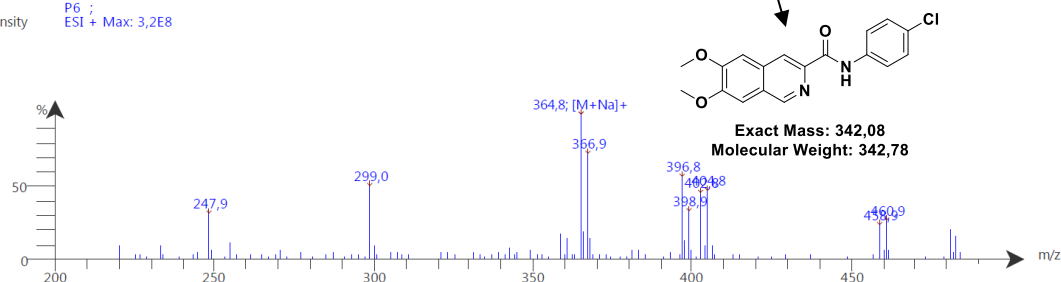
Spectrum RT 0,81 (1 scans) - Background Subtracted 0,03 - 0,75
O23 ;
ESI + Max: 7,3E8

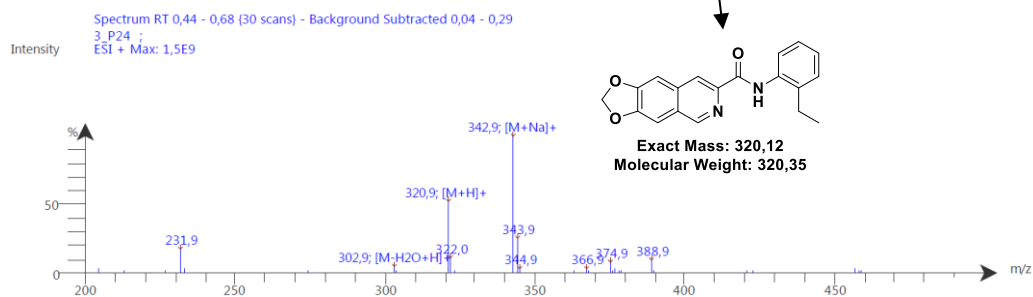
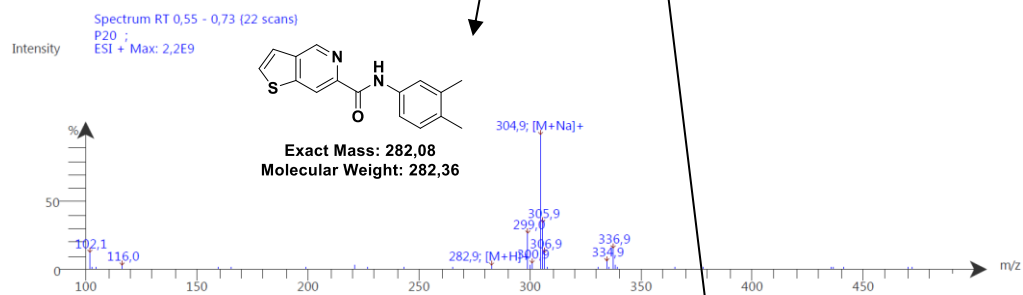
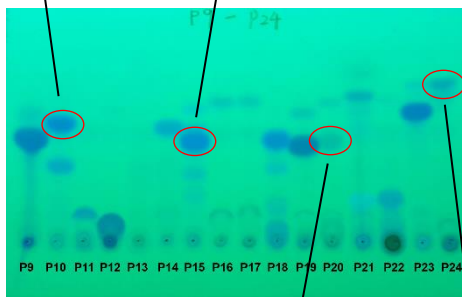
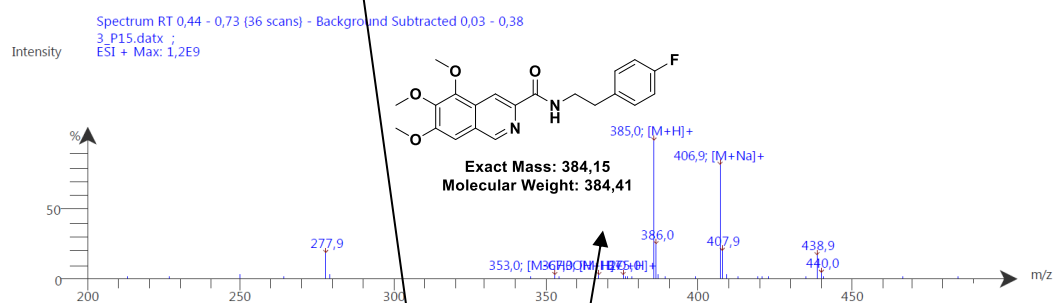
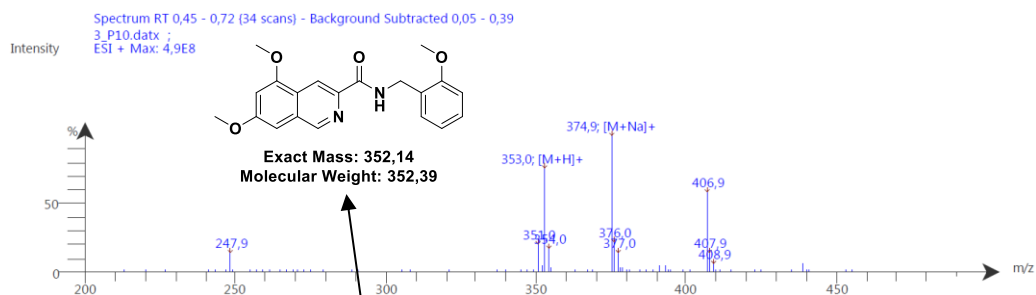


Spectrum RT 0,72 (1 scans) - Background Subtracted 0,03 - 0,66
O24 ;
ESI + Max: 6,1E8

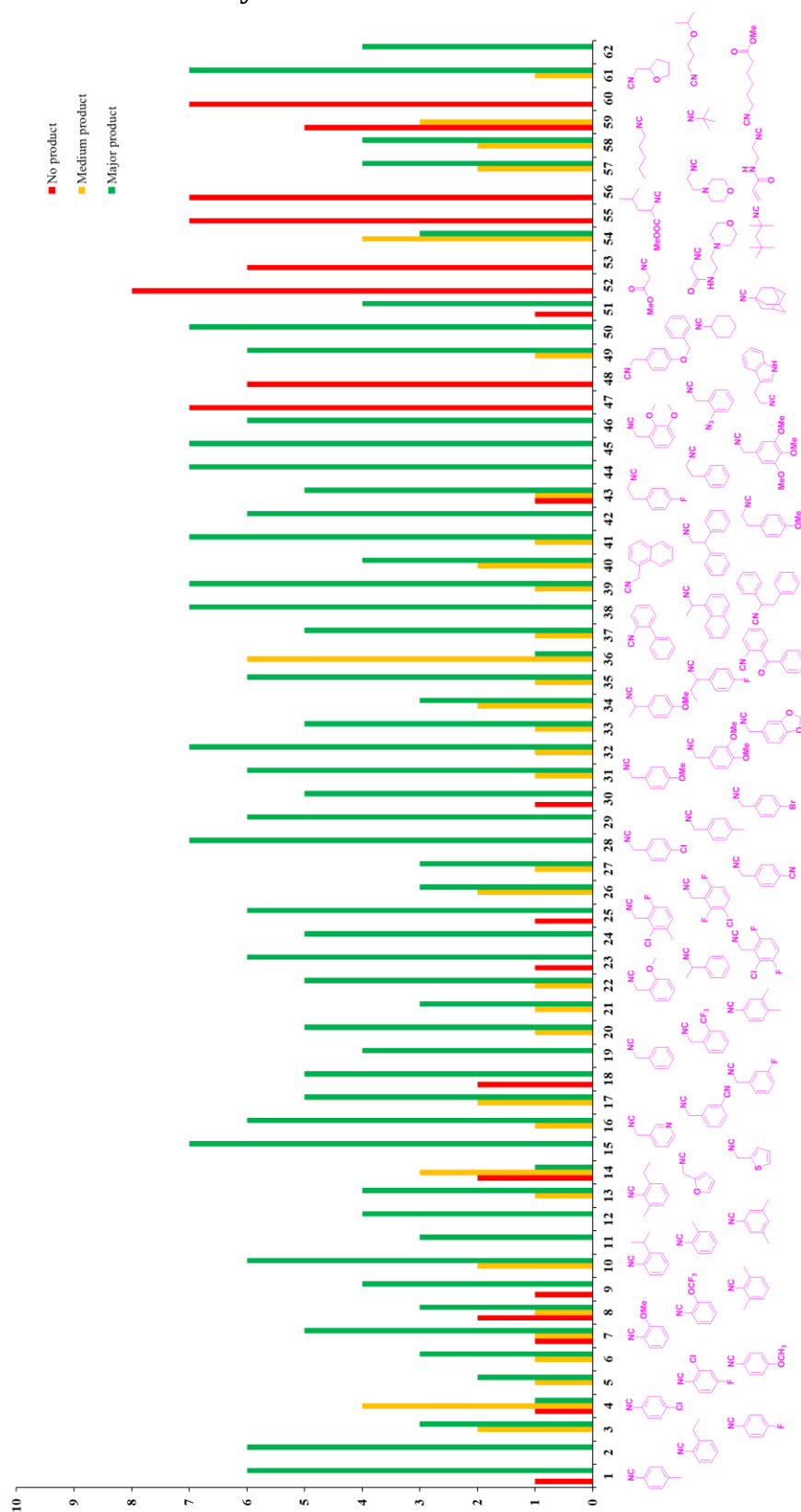


Spectrum RT 0,82 (1 scans) - Background Subtracted 0,07 - 0,68
P6 ;
ESI + Max: 3,2E8

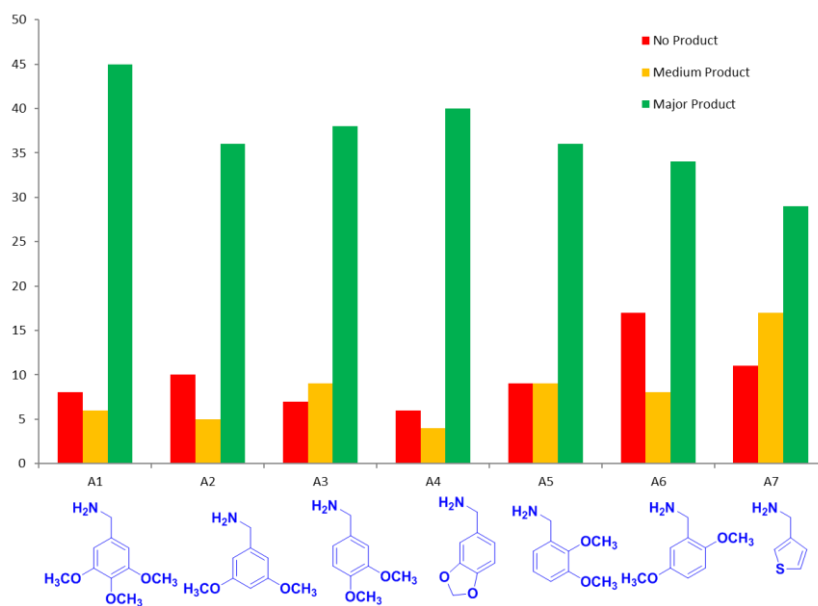




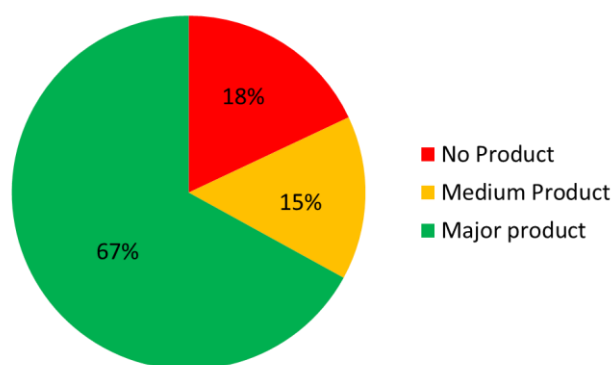
5. Statistical reaction analysis



Scheme S2. Frequency of the isocyanides in the plate along with the QC results.



Scheme S3. Frequency of the benzyl amines in the plate along with the QC results.



Scheme S4. QC results for the 384-destination plate.

6. Mg scale reactions

6.1. General procedure

To a stirred solution of 2,2-dimethoxyacetaldehyde (1 mmol) in MeOH (1M) at room temperature, amine (1 mmol), benzoic acid (1 mmol) and isocyanide (1 mmol) were added. The resulting mixture was stirred at room temperature for 15 h. Upon completion, the solvent was evaporated under vacuum. Then, the crude Ugi-adduct was dissolved in 37% HCl_(aq) solution in dioxane (1 mL, 1:1, v/v) and was stirred at room temperature for 12 h. The reaction was diluted with dichloromethane (20 mL) and washed with saturated sodium bicarbonate solution (3 x 10 mL). Finally, the solvent was evaporated under vacuum and the crude product was purified by flash column chromatography using petroleum ether/ethyl acetate (Fig. S3).

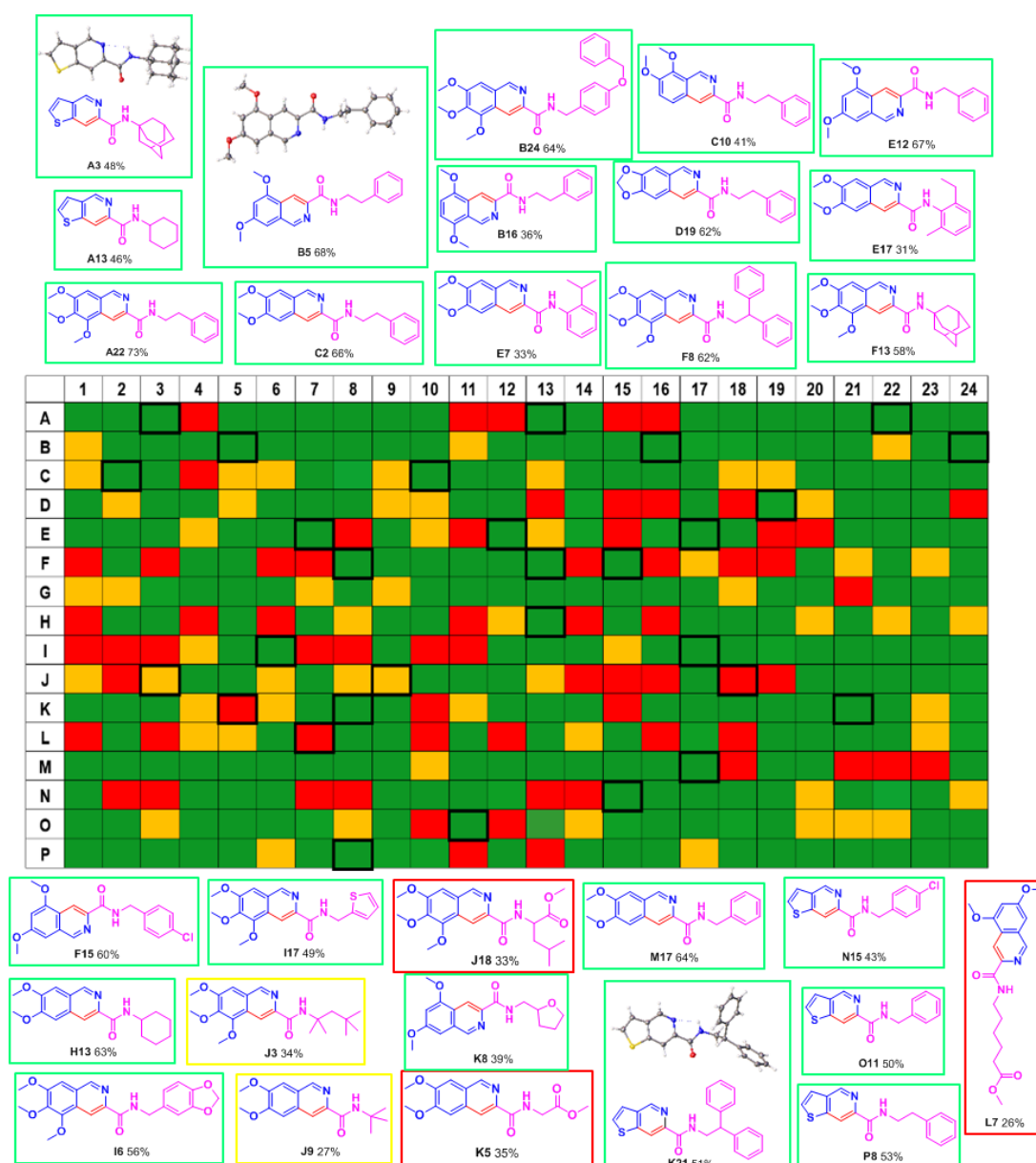
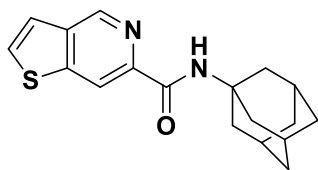


Fig. S3. Structures of the randomly resynthesized isoquinolines on mg scale and selected X-ray structures.

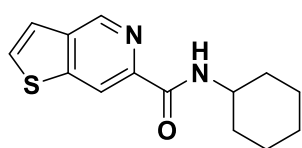
6.2. Characterization of the products

N-((1*S*,3*S*)-adamantan-1-yl)thieno[3,2-*c*]pyridine-6-carboxamide (A3)



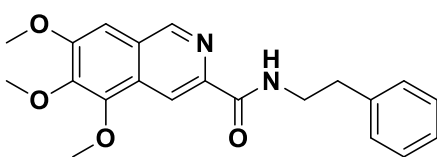
White solid (149 mg, 48% yield), M.P.= 221 – 223 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.97 (d, *J* = 0.9 Hz, 1H), 8.68 (d, *J* = 1.0 Hz, 1H), 8.03 (s, 1H), 7.62 (d, *J* = 5.5 Hz, 1H), 7.50 – 7.47 (m, 1H), 2.19 (d, *J* = 2.8 Hz, 6H), 2.16 – 2.11 (m, 3H), 1.79 – 1.67 (m, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 163.4, 148.2, 144.5, 143.5, 137.2, 130.1, 122.4, 116.1, 51.7, 41.6, 36.4, 29.5; HRMS (ESI) *m/z* calculated for C₁₈H₂₁N₂OS [M+H]⁺: 313.1369; found [M+H]⁺: 313.1367.

N-cyclohexylthieno[3,2-*c*]pyridine-6-carboxamide (A13)



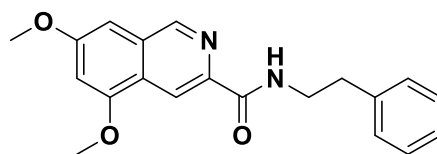
White solid (119 mg, 46% yield), M.P.= 109 – 111 °C; ¹H NMR (500 MHz, CDCl₃) δ 9.0 (d, *J* = 0.9 Hz, 1H), 8.7 (s, 1H), 8.1 (d, *J* = 8.6 Hz, 1H), 7.6 (d, *J* = 5.5 Hz, 1H), 7.5 – 7.5 (m, 1H), 4.1 – 3.9 (m, 1H), 2.1 – 2.0 (m, 2H), 1.9 – 1.7 (m, 2H), 1.7 (m, 1H), 1.5 – 1.4 (m, 2H), 1.4 – 1.3 (m, 2H), 1.3 – 1.2 (m, 1H); ¹³C NMR (126 MHz, CDCl₃) δ 163.7, 148.2, 143.9, 137.5, 130.4, 130.2, 122.4, 116.7, 48.4, 33.3, 25.7, 25.7; HRMS (ESI) *m/z* calculated for C₁₄H₁₇N₂OS [M+H]⁺: 261.1056; found [M+H]⁺: 261.1058.

5,6,7-Trimethoxy-*N*-phenethylisoquinoline-3-carboxamide (A22)



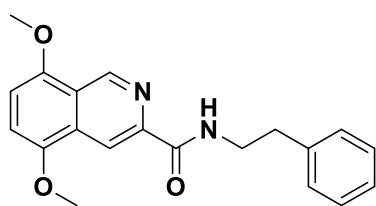
White solid (267 mg, 73% yield), M.P.= 121 – 123 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.90 (s, 1H), 8.76 (s, 1H), 8.28 (t, *J* = 6.2 Hz, 1H), 7.33 – 7.20 (m, 5H), 7.05 (s, 1H), 4.06 (s, 3H), 4.01 (s, 3H), 4.00 (s, 3H), 3.81 – 3.74 (m, 2H), 2.97 (t, *J* = 7.3 Hz, 2H); ¹³C NMR (126 MHz, CDCl₃) δ 165.2, 155.3, 148.8, 148.1, 144.6, 142.2, 139.2, 128.9, 128.6, 127.9, 127.1, 126.5, 114.7, 101.6, 61.8, 61.3, 56.2, 40.9, 36.1; HRMS (ESI) *m/z* calculated for C₂₁H₂₃N₂O₄ [M+H]⁺: 367.1652; found [M+H]⁺: 367.1652.

5,7-Dimethoxy-*N*-phenethylisoquinoline-3-carboxamide (B5)



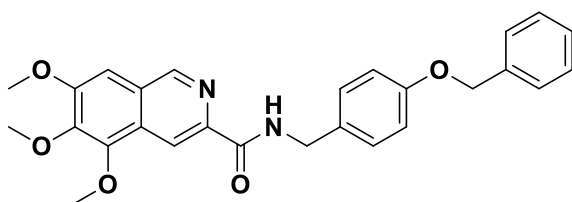
Grey solid (229 mg, 68% yield), M.P.= 159 – 161 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.90 (s, 1H), 8.83 (s, 1H), 8.24 (t, *J* = 6.2 Hz, 1H), 7.35 – 7.20 (m, 5H), 6.79 (d, *J* = 2.2 Hz, 1H), 6.65 (d, *J* = 2.1 Hz, 1H), 3.97 (s, 3H), 3.93 (s, 3H), 3.81 – 3.74 (m, 2H), 2.98 (t, *J* = 7.3 Hz, 2H); ¹³C NMR (126 MHz, CDCl₃) δ 165.2, 160.7, 157.0, 148.9, 141.8, 139.3, 131.6, 128.9, 128.7, 126.5, 125.0, 115.3, 101.9, 96.8, 55.9, 55.7, 40.9, 36.2; HRMS (ESI) *m/z* calculated for C₂₀H₂₁N₂O₃ [M+H]⁺: 337.1547; found [M+H]⁺: 337.1545.

5,8-Dimethoxy-*N*-phenethylisoquinoline-3-carboxamide (B16)



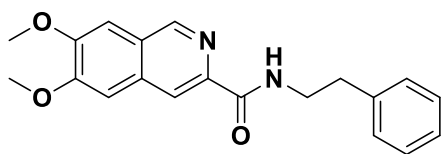
Yellow solid (121 mg, 36% yield), M.P.= 120 – 122 °C; ^1H NMR (500 MHz, CDCl_3) δ 9.41 (s, 1H), 8.89 (s, 1H), 8.36 (t, J = 6.2 Hz, 1H), 7.34 – 7.20 (m, 5H), 6.91 (d, J = 8.5 Hz, 1H), 6.83 (d, J = 8.5 Hz, 1H), 3.97 (s, 3H), 3.95 (s, 3H), 3.83 – 3.75 (m, 2H), 2.99 (t, J = 7.3 Hz, 2H); ^{13}C NMR (126 MHz, CDCl_3) δ 165.0, 150.1, 149.5, 146.0, 144.0, 139.3, 129.5, 129.0, 128.7, 126.5, 122.0, 114.8, 108.5, 106.5, 56.0, 55.9, 41.0, 36.2; HRMS (ESI) m/z calculated for $\text{C}_{20}\text{H}_{21}\text{N}_2\text{O}_3$ $[\text{M}+\text{H}]^+$: 337.1547; found $[\text{M}+\text{H}]^+$: 337.1546.

***N*-(4-(benzyloxy)benzyl)-5,6,7-trimethoxyisoquinoline-3-carboxamide (B24)**



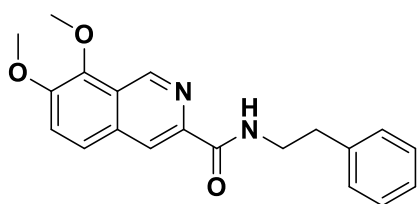
Grey solid (293 mg, 64% yield), M.P.= 110 – 112 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.91 (s, 1H), 8.80 (s, 1H), 8.45 (t, J = 5.9 Hz, 1H), 7.45 – 7.29 (m, 7H), 7.06 (s, 1H), 6.95 (d, J = 8.2 Hz, 2H), 5.05 (s, 2H), 4.65 (d, J = 5.9 Hz, 2H), 4.07 (s, 3H), 4.02 (s, 3H), 4.01 (s, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 165.0, 158.2, 155.4, 148.8, 148.1, 144.7, 142.2, 137.1, 131.1, 129.4, 128.7, 128.0, 128.0, 127.5, 127.2, 115.1, 115.0, 101.6, 70.1, 61.9, 61.4, 56.3, 43.1; HRMS (ESI) m/z calculated for $\text{C}_{27}\text{H}_{27}\text{N}_2\text{O}_5$ $[\text{M}+\text{H}]^+$: 459.1915; found $[\text{M}+\text{H}]^+$: 459.1914.

6,7-Dimethoxy-*N*-phenethylisoquinoline-3-carboxamide (C2)



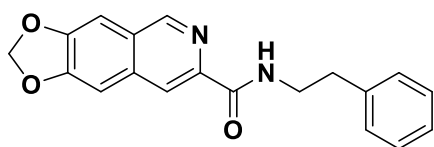
White solid (220 mg, 66% yield), M.P.= 158 – 160 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.9 (s, 1H), 8.8 (s, 1H), 8.2 (t, J = 6.2 Hz, 1H), 7.3 – 7.2 (m, 5H), 6.8 (s, 1H), 6.6 (s, 1H), 4.0 (s, 3H), 3.9 (s, 3H), 3.8 – 3.7 (m, 2H), 3.0 (t, J = 7.3 Hz, 2H); ^{13}C NMR (126 MHz, CDCl_3) δ 165.3, 153.5, 151.7, 148.5, 142.9, 139.3, 132.9, 129.0, 128.7, 126.5, 126.1, 119.1, 106.1, 105.4, 56.4, 56.3, 40.9, 36.2; HRMS (ESI) m/z calculated for $\text{C}_{20}\text{H}_{21}\text{N}_2\text{O}_3$ $[\text{M}+\text{H}]^+$: 337.1547; found $[\text{M}+\text{H}]^+$: 337.1545.

7,8-Dimethoxy-*N*-phenethylisoquinoline-3-carboxamide (C10)



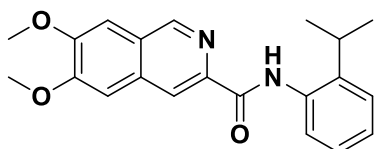
Color-less semi-solid (138 mg, 41% yield); ^1H NMR (500 MHz, CDCl_3) δ 9.40 (s, 1H), 8.52 (s, 1H), 8.33 (t, J = 6.3 Hz, 1H), 7.73 (d, J = 8.9 Hz, 1H), 7.54 (d, J = 8.9 Hz, 1H), 7.36 – 7.22 (m, 5H), 4.07 (s, 3H), 4.03 (s, 3H), 3.79 (q, J = 7.0 Hz, 2H), 2.98 (t, J = 7.3 Hz, 2H); ^{13}C NMR (126 MHz, CDCl_3) δ 165.2, 150.4, 146.1, 144.0, 142.2, 139.2, 131.5, 129.0, 128.7, 126.6, 125.2, 124.6, 120.1, 119.9, 61.8, 56.9, 40.9, 36.2; HRMS (ESI) m/z calculated for $\text{C}_{20}\text{H}_{21}\text{N}_2\text{O}_3$ $[\text{M}+\text{H}]^+$: 337.1547; found $[\text{M}+\text{H}]^+$: 337.1546.

***N*-phenethyl-[1,3]dioxolo[4,5-*f*]isoquinoline-8-carboxamide (D19)**



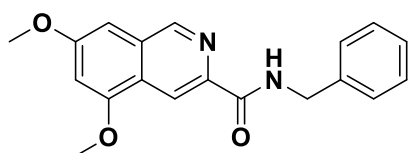
White solid (198 mg, 62% yield), M.P.= 138 – 140 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.80 (s, 1H), 8.40 (s, 1H), 8.27 (t, J = 6.3 Hz, 1H), 7.33 – 7.21 (m, 5H), 7.17 (d, J = 8.3 Hz, 2H), 6.10 (s, 2H), 3.82 – 3.70 (m, 2H), 2.97 (t, J = 7.3 Hz, 2H); ^{13}C NMR (126 MHz, CDCl_3) δ 165.1, 151.5, 149.7, 148.7, 143.1, 139.2, 134.5, 128.9, 128.7, 127.4, 126.5, 119.6, 103.9, 103.3, 102.1, 40.9, 36.1; HRMS (ESI) m/z calculated for $\text{C}_{19}\text{H}_{17}\text{N}_2\text{O}_3$ $[\text{M}+\text{H}]^+$: 321.1234; found $[\text{M}+\text{H}]^+$: 321.1234.

***N*-(2-isopropylphenyl)-6,7-dimethoxyisoquinoline-3-carboxamide (E7)**



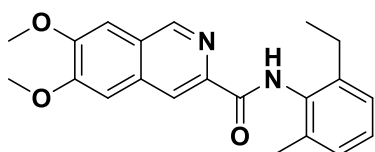
White solid (115 mg, 33% yield), M.P.= 180 – 182 °C; ^1H NMR (500 MHz, CDCl_3) δ 10.36 (s, 1H), 8.99 (s, 1H), 8.56 (s, 1H), 8.27 – 8.22 (m, 1H), 7.35 – 7.31 (m, 1H), 7.30 – 7.26 (m, 2H), 7.22 (s, 1H), 7.20 – 7.15 (m, 1H), 4.05 (s, 3H), 4.05 (s, 3H), 3.32 – 3.24 (m, 1H), 1.36 (s, 3H), 1.34 (s, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 163.2, 153.7, 151.9, 148.6, 143.1, 139.1, 134.9, 133.0, 126.7, 126.3, 125.7, 125.2, 122.8, 119.5, 106.2, 105.6, 56.5, 56.4, 28.4, 23.1; HRMS (ESI) m/z calculated for $\text{C}_{21}\text{H}_{23}\text{N}_2\text{O}_3$ $[\text{M}+\text{H}]^+$: 351.1703; found $[\text{M}+\text{H}]^+$: 351.1701.

***N*-(benzyl)-5,7-dimethoxyisoquinoline-3-carboxamide (E12)**



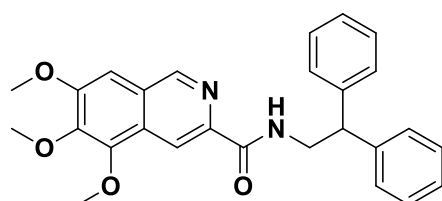
White solid (215 mg, 67% yield), M.P.= 146 – 148 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.90 (s, 1H), 8.87 (s, 1H), 7.45 – 7.23 (m, 5H), 6.79 (s, 1H), 6.69 – 6.56 (m, 1H), 4.72 (d, J = 6.0 Hz, 2H), 3.97 (s, 3H), 3.93 (s, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 165.1, 160.8, 157.0, 148.9, 141.6, 138.7, 131.6, 128.7, 128.0, 127.4, 124.9, 115.5, 101.9, 96.8, 55.9, 55.7, 43.6; HRMS (ESI) m/z calculated for $\text{C}_{19}\text{H}_{19}\text{N}_2\text{O}_3$ $[\text{M}+\text{H}]^+$: 323.1390; found $[\text{M}+\text{H}]^+$: 323.1390.

***N*-(2-ethyl-6-methylphenyl)-6,7-dimethoxyisoquinoline-3-carboxamide (E17)**



White solid (109 mg, 31% yield), M.P.= 225 – 227 °C; ^1H NMR (500 MHz, CDCl_3) δ 9.64 (s, 1H), 9.01 (s, 1H), 8.57 (s, 1H), 7.29 (s, 1H), 7.22 (s, 1H), 7.21 – 7.17 (m, 1H), 7.17 – 7.13 (m, 2H), 4.07 (s, 3H), 4.04 (s, 3H), 2.69 (q, J = 7.6 Hz, 2H), 2.32 (s, 3H), 1.21 (t, J = 7.6 Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 163.8, 153.6, 151.8, 148.6, 142.7, 141.3, 136.1, 133.7, 132.9, 128.3, 127.5, 126.4, 126.3, 119.5, 106.1, 105.5, 56.4, 56.3, 25.2, 18.8, 14.6; HRMS (ESI) m/z calculated for $\text{C}_{21}\text{H}_{23}\text{N}_2\text{O}_3$ $[\text{M}+\text{H}]^+$: 351.1703; found $[\text{M}+\text{H}]^+$: 351.1702.

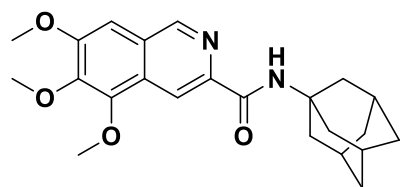
***N*-(2,2-diphenylethyl)-5,6,7-trimethoxyisoquinoline-3-carboxamide (F8)**



White solid (274 mg, 62% yield), M.P.= 171 – 172 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.84 (s, 1H), 8.75 (s, 1H), 8.21 (t, J = 6.1 Hz, 1H), 7.35 – 7.30 (m, 8H), 7.25 – 7.20 (m, 2H), 7.03 (s, 1H), 4.38 (t, J = 7.8 Hz, 1H), 4.19 – 4.14 (m, 2H), 4.06 (s, 3H), 4.01 (s, 3H), 4.01 (s, 3H); ^{13}C NMR

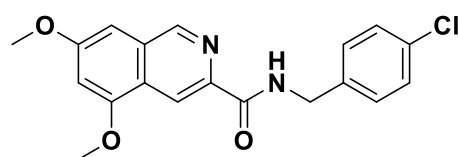
(126 MHz, CDCl₃) 165.3, 155.4, 148.8, 148.1, 144.7, 142.4, 142.2, 128.8, 128.3, 128.0, 127.2, 126.8, 114.8, 101.6, 61.9, 61.4, 56.3, 51.0, 44.1; HRMS (ESI) *m/z* calculated for C₂₇H₂₇N₂O₄ [M+H]⁺: 443.1965; found [M+H]⁺: 443.1957.

***N*-((1*s*,3*s*)-adamantan-1-yl)-5,6,7-trimethoxyisoquinoline-3-carboxamide (F13)**



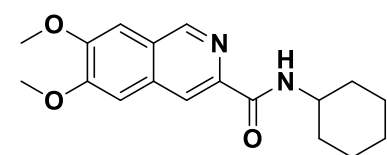
White solid (229 mg, 58% yield), M.P.= 167 – 169 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.92 (s, 1H), 8.73 (s, 1H), 8.01 (s, 1H), 7.07 (s, 1H), 4.04 (s, 3H), 4.02 (s, 6H), 2.21 (d, *J* = 2.8 Hz, 6H), 2.17 – 2.12 (m, 3H), 1.84 – 1.66 (m, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 164.1, 155.2, 148.5, 148.1, 144.8, 143.3, 128.2, 127.0, 114.3, 101.6, 61.9, 61.4, 56.3, 51.7, 41.8, 36.6, 29.7; HRMS (ESI) *m/z* calculated for C₂₃H₂₉N₂O₄ [M+H]⁺: 397.2122; found [M+H]⁺: 397.2117.

***N*-(4-chlorobenzyl)-5,7-dimethoxyisoquinoline-3-carboxamide (F15)**



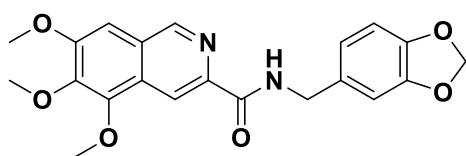
Color-less semi-solid (213 mg, 60% yield); ¹H NMR (500 MHz, CDCl₃) δ 8.91 (s, 1H), 8.86 (s, 1H), 8.49 (t, *J* = 6.2 Hz, 1H), 7.34 – 7.27 (m, 4H), 6.81 (d, *J* = 2.1 Hz, 1H), 6.66 (d, *J* = 2.1 Hz, 1H), 4.67 (d, *J* = 6.2 Hz, 2H), 3.97 (s, 3H), 3.93 (s, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 165.2, 160.9, 157.1, 149.0, 141.5, 137.3, 133.2, 131.7, 129.3, 128.8, 125.0, 115.7, 102.1, 102.0, 96.9, 96.8, 55.9, 55.8, 42.9; HRMS (ESI) *m/z* calculated for C₁₉H₁₈ClN₂O₃ [M+H]⁺: 357.1001; found [M+H]⁺: 357.1001.

***N*-cyclohexyl-6,7-dimethoxyisoquinoline-3-carboxamide (H13)**



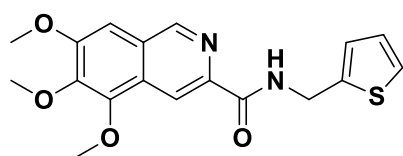
White solid (198 mg, 63% yield), M.P.= 179 – 180 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.89 (s, 1H), 8.43 (s, 1H), 8.08 (d, *J* = 8.6 Hz, 1H), 7.21 (s, 1H), 7.16 (s, 1H), 4.01 (s, 3H), 4.01 (s, 3H), 4.00 – 3.96 (m, 1H), 2.07 – 1.97 (m, 2H), 1.80 – 1.73 (m, 2H), 1.67 – 1.60 (m, 1H), 1.47 – 1.37 (m, 2H), 1.37 – 1.28 (m, 2H), 1.26 – 1.18 (m, 1H); ¹³C NMR (126 MHz, CDCl₃) δ 164.2, 153.4, 151.5, 148.3, 143.1, 132.8, 126.0, 119.0, 106.0, 105.4, 57.3, 56.2, 48.2, 33.3, 25.7, 25.0; HRMS (ESI) *m/z* calculated for C₁₈H₂₃N₂O₃ [M+H]⁺: 315.1703; found [M+H]⁺: 315.1702.

***N*-(benzo[*d*][1,3]dioxol-5-ylmethyl)-5,6,7-trimethoxyisoquinoline-3-carboxamide (I6)**



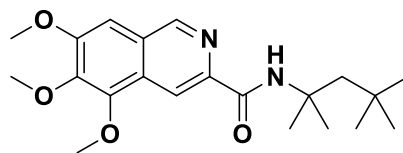
Light yellow solid (221 mg, 56% yield), M.P.= 123 – 125 °C; ¹H NMR (500 MHz, CDCl₃) δ 8.92 (s, 1H), 8.79 (s, 1H), 8.44 (t, *J* = 6.2 Hz, 1H), 7.07 (s, 1H), 6.90 (d, *J* = 1.7 Hz, 1H), 6.87 – 6.84 (m, 1H), 6.77 (d, *J* = 7.9 Hz, 1H), 5.93 (s, 2H), 4.62 (d, *J* = 6.0 Hz, 2H), 4.07 (s, 3H), 4.02 (d, *J* = 2.1 Hz, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 165.1, 155.5, 148.8, 148.1, 148.0, 147.0, 144.8, 142.1, 132.6, 128.0, 127.2, 121.3, 115.1, 108.7, 108.4, 101.6, 101.1, 61.9, 61.4, 56.3, 43.5; HRMS (ESI) *m/z* calculated for C₂₁H₂₁N₂O₆ [M+H]⁺: 397.1394; found [M+H]⁺: 397.1390.

5,6,7-Trimethoxy-*N*-(thiophen-2-ylmethyl)isoquinoline-3-carboxamide (I17)



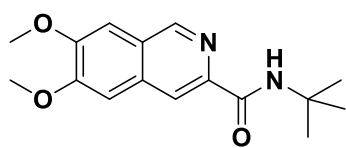
White solid (175 mg, 49% yield), M.P.= 115 – 117 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.92 (d, J = 0.9 Hz, 1H), 8.80 (s, 1H), 8.51 (t, J = 6.1 Hz, 1H), 7.24 – 7.22 (m, 1H), 7.08 – 7.05 (m, 2H), 6.98 – 6.95 (m, 1H), 4.90 – 4.86 (m, 2H), 4.07 (s, 3H), 4.02 (d, J = 1.7 Hz, 6H); ^{13}C NMR (126 MHz, CDCl_3) δ 165.0, 155.5, 148.9, 148.1, 144.7, 142.0, 141.3, 128.0, 127.3, 127.0, 126.2, 125.2, 115.2, 101.6, 61.9, 61.4, 56.3, 38.4; HRMS (ESI) m/z calculated for $\text{C}_{18}\text{H}_{19}\text{N}_2\text{O}_4\text{S}$ $[\text{M}+\text{H}]^+$: 359.1060; found $[\text{M}+\text{H}]^+$: 359.1057.

5,6,7-Trimethoxy-*N*-(2,4,4-trimethylpentan-2-yl)isoquinoline-3-carboxamide (J3)



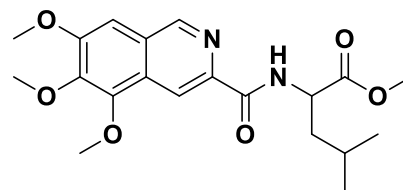
Color-less semi-solid (127 mg, 34% yield); ^1H NMR (500 MHz, CDCl_3) δ 8.91 (s, 1H), 8.74 (s, 1H), 8.22 (s, 1H), 7.05 (s, 1H), 4.03 (s, 3H), 4.01 (d, J = 1.2 Hz, 6H), 1.91 (s, 2H), 1.58 (s, 6H), 1.03 (s, 9H); ^{13}C NMR (126 MHz, CDCl_3) δ 164.08, 155.20, 148.58, 148.10, 144.70, 143.33, 128.18, 126.97, 114.16, 101.57, 77.36, 61.85, 61.36, 56.27, 54.79, 51.98, 31.85, 31.62, 29.42; HRMS (ESI) m/z calculated for $\text{C}_{21}\text{H}_{31}\text{N}_2\text{O}_4$ $[\text{M}+\text{H}]^+$: 375.2278; found $[\text{M}+\text{H}]^+$: 375.2279.

N-(tert-butyl)-6,7-dimethoxyisoquinoline-3-carboxamide (J9)



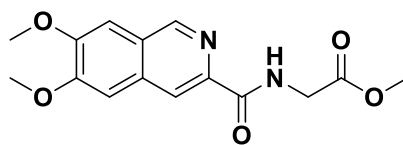
White solid (78 mg, 27% yield), M.P.= 188 – 190 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.89 (s, 1H), 8.41 (s, 1H), 8.14 (s, 1H), 7.22 (s, 1H), 7.16 (s, 1H), 4.03 (s, 6H), 1.52 (s, 9H); ^{13}C NMR (126 MHz, CDCl_3) δ 164.4, 153.4, 151.5, 148.1, 143.7, 132.9, 125.9, 118.4, 106.0, 105.4, 56.3, 56.2, 50.9, 29.0; HRMS (ESI) m/z calculated for $\text{C}_{16}\text{H}_{21}\text{N}_2\text{O}_3$ $[\text{M}+\text{H}]^+$: 289.1547; found $[\text{M}+\text{H}]^+$: 289.1547.

Methyl (5,6,7-trimethoxyisoquinoline-3-carbonyl)leucinate (J18)



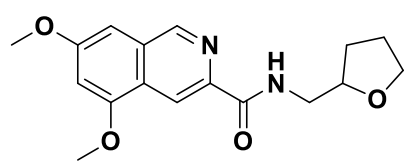
Color-less semi-solid (129 mg, 33% yield); ^1H NMR (500 MHz, CDCl_3) δ 8.97 (s, 1H), 8.75 (s, 1H), 8.48 (d, J = 8.7 Hz, 1H), 7.08 (s, 1H), 4.96 – 4.84 (m, 1H), 4.04 (s, 3H), 4.02 (s, 3H), 4.01 (s, 3H), 3.76 (s, 3H), 1.87 – 1.69 (m, 3H), 1.00 (d, J = 6.0 Hz, 3H), 0.98 (d, J = 6.0 Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 173.5, 164.9, 155.4, 148.8, 148.0, 144.6, 141.6, 127.8, 127.2, 115.1, 101.6, 61.8, 61.3, 56.2, 52.3, 50.8, 41.8, 25.0, 23.0, 21.9; HRMS (ESI) m/z calculated for $\text{C}_{20}\text{H}_{27}\text{N}_2\text{O}_6$ $[\text{M}+\text{H}]^+$: 391.1864; found $[\text{M}+\text{H}]^+$: 391.1867.

Methyl (6,7-dimethoxyisoquinoline-3-carbonyl) glycinate (K5)



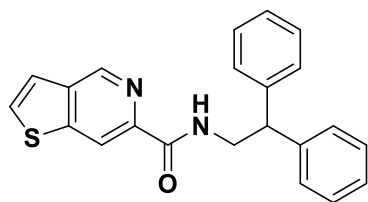
Yellow solid (106 mg, 35% yield), M.P.= 197 – 198 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.95 (s, 1H), 8.61 (t, J = 5.7 Hz, 1H), 8.44 (s, 1H), 7.25 (s, 1H), 7.19 (s, 1H), 4.32 (d, J = 5.7 Hz, 2H), 4.05 (s, 3H), 4.04 (s, 3H), 3.80 (s, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 170.5, 165.6, 153.6, 151.8, 148.7, 142.2, 132.7, 126.3, 119.3, 106.1, 105.5, 56.4, 56.3, 52.5, 41.5; HRMS (ESI) m/z calculated for $\text{C}_{15}\text{H}_{17}\text{N}_2\text{O}_5$ $[\text{M}+\text{H}]^+$: 305.1132; found $[\text{M}+\text{H}]^+$: 305.1132.

5,7-Dimethoxy-*N*-((tetrahydrofuran-2-yl)methyl)isoquinoline-3-carboxamide (K8)



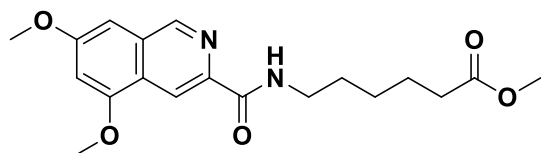
White solid (123 mg, 39% yield), M.P.= 119 – 121 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.94 (s, 1H), 8.81 (s, 1H), 8.44 (t, J = 6.1 Hz, 1H), 6.81 (d, J = 2.4 Hz, 1H), 6.64 (d, J = 2.1 Hz, 1H), 4.17 – 4.05 (m, 1H), 3.96 (s, 3H), 3.93 (s, 3H), 3.84 – 3.71 (m, 2H), 3.51 – 3.40 (m, 1H), 2.11 – 1.98 (m, 1H), 1.98 – 1.85 (m, 2H), 1.75 – 1.58 (m, 1H); ^{13}C NMR (126 MHz, CDCl_3) δ 165.4, 160.7, 157.0, 149.0, 141.8, 131.6, 125.0, 115.4, 102.0, 96.9, 78.0, 68.3, 55.9, 55.7, 43.3, 28.9, 26.0; HRMS (ESI) m/z calculated for $\text{C}_{17}\text{H}_{21}\text{N}_2\text{O}_4$ $[\text{M}+\text{H}]^+$: 317.1496; found $[\text{M}+\text{H}]^+$: 317.1497.

N-(2,2-diphenylethyl)thieno[3,2-*c*]pyridine-6-carboxamide (K21)



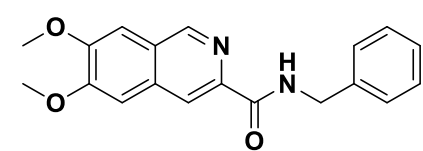
Yellow solid (182 mg, 51% yield), M.P.= 175 – 177 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.89 (s, 1H), 8.72 (s, 1H), 8.23 (t, J = 6.2 Hz, 1H), 7.64 – 7.58 (m, 1H), 7.45 (d, J = 5.4 Hz, 1H), 7.35 – 7.29 (m, 8H), 7.27 – 7.19 (m, 2H), 4.38 (t, J = 7.8 Hz, 1H), 4.21 – 4.13 (m, 2H); ^{13}C NMR (126 MHz, CDCl_3) δ 164.7, 148.0, 144.0, 143.4, 142.2, 137.5, 130.5, 128.8, 128.2, 126.8, 122.5, 122.4, 116.7, 50.9, 44.1; HRMS (ESI) m/z calculated for $\text{C}_{22}\text{H}_{19}\text{N}_2\text{OS}$ $[\text{M}+\text{H}]^+$: 359.1213; found $[\text{M}+\text{H}]^+$: 359.1212.

Methyl 6-(5,7-dimethoxyisoquinoline-3-carboxamido)hexanoate (L7)



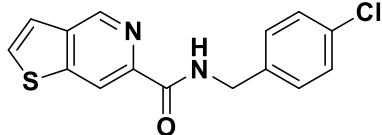
Color-less semi-solid (94 mg, 26% yield); ^1H NMR (500 MHz, CDCl_3) ^1H NMR (500 MHz, Chloroform- d) δ 8.94 (s, 1H), 8.83 (s, 1H), 8.15 (s, 1H), 6.83 (s, 1H), 6.67 (s, 1H), 3.98 (s, 3H), 3.95 (s, 3H), 3.66 (s, 3H), 3.55 – 3.49 (m, 2H), 2.33 (t, J = 7.5 Hz, 2H), 1.74 – 1.64 (m, 4H), 1.49 – 1.41 (m, 2H); ^{13}C NMR (126 MHz, CDCl_3) δ 174.2, 165.2, 160.8, 157.1, 148.9, 141.9, 131.6, 125.1, 115.4, 102.1, 96.9, 55.9, 51.7, 39.3, 34.1, 29.7, 26.7, 24.8; HRMS (ESI) m/z calculated for $\text{C}_{19}\text{H}_{25}\text{N}_2\text{O}_5$ $[\text{M}+\text{H}]^+$: 361.1758; found $[\text{M}+\text{H}]^+$: 361.1756.

N-benzyl-6,7-dimethoxyisoquinoline-3-carboxamide (M17)



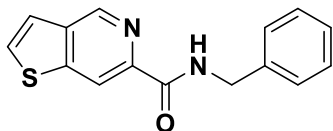
Grey solid (206 mg, 64% yield), M.P.= 160 – 162 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.90 (s, 1H), 8.58 – 8.46 (m, 2H), 7.43 – 7.27 (m, 5H), 7.23 (s, 1H), 7.20 (s, 1H), 4.72 (d, J = 6.0 Hz, 2H), 4.04 (s, 3H), 4.04 (s, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 165.2, 153.5, 151.7, 148.5, 142.7, 138.6, 132.8, 128.8, 128.0, 127.5, 126.2, 119.3, 106.1, 105.5, 56.4, 56.3, 43.7; HRMS (ESI) m/z calculated for $\text{C}_{19}\text{H}_{19}\text{N}_2\text{O}_3$ $[\text{M}+\text{H}]^+$: 323.1390; found $[\text{M}+\text{H}]^+$: 323.1390.

N-(4-chlorobenzyl)thieno[3,2-*c*]pyridine-6-carboxamide (N15)



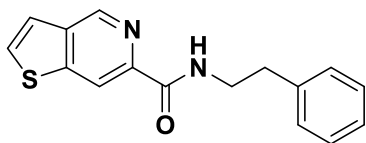
White solid (129 mg, 43% yield), M.P.= 129 – 131 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.99 (d, J = 1.0 Hz, 1H), 8.77 (s, 1H), 8.53 (s, 1H), 7.66 (d, J = 5.4 Hz, 1H), 7.56 – 7.46 (m, 1H), 7.32 – 7.30 (m, 4H), 4.67 (d, J = 6.2 Hz, 2H); ^{13}C NMR (126 MHz, CDCl_3) δ 164.7, 148.2, 144.0, 143.2, 137.7, 137.1, 133.3, 130.7, 130.5, 129.3, 128.9, 122.6, 122.5, 117.0, 43.0; HRMS (ESI) m/z calculated for $\text{C}_{15}\text{H}_{12}\text{ClN}_2\text{OS}$ $[\text{M}+\text{H}]^+$: 303.0353; found $[\text{M}+\text{H}]^+$: 303.0354.

***N*-benzylthieno[3,2-*c*]pyridine-6-carboxamide (O11)**



White solid (134 mg, 50% yield), M.P.= 100 – 102 °C; ^1H NMR (500 MHz, CDCl_3) δ 9.0 (d, J = 0.9 Hz, 1H), 8.8 (s, 1H), 8.5 (s, 1H), 7.7 (d, J = 5.4 Hz, 1H), 7.5 – 7.5 (m, 1H), 7.4 – 7.4 (m, 2H), 7.4 – 7.3 (m, 2H), 7.3 – 7.3 (m, 1H), 4.7 (d, J = 6.1 Hz, 2H); ^{13}C NMR (126 MHz, CDCl_3) δ 164.7, 148.2, 144.0, 143.5, 138.5, 137.6, 130.6, 130.4, 128.9, 128.1, 127.6, 122.6, 122.4, 117.0, 43.8; HRMS (ESI) m/z calculated for $\text{C}_{15}\text{H}_{13}\text{N}_2\text{OS}$ $[\text{M}+\text{H}]^+$: 269.0743; found $[\text{M}+\text{H}]^+$: 269.0743.

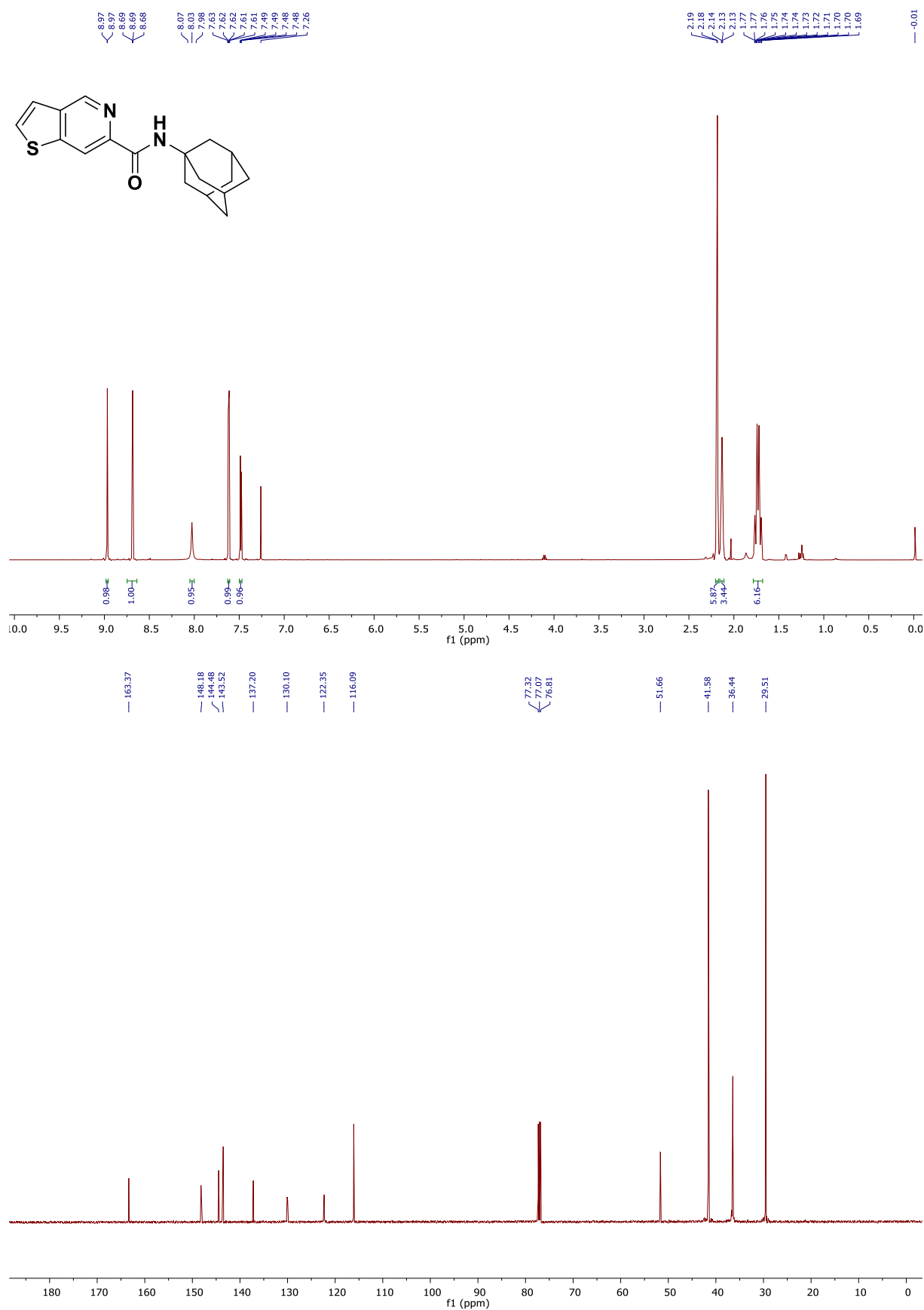
***N*-phenethylthieno[3,2-*c*]pyridine-6-carboxamide (P8)**



White solid (149 mg, 53% yield), M.P.= 108 – 109 °C; ^1H NMR (500 MHz, CDCl_3) δ 8.98 (d, J = 0.9 Hz, 1H), 8.75 (d, J = 1.0 Hz, 1H), 8.28 (s, 1H), 7.65 (d, J = 5.5 Hz, 1H), 7.51 – 7.49 (m, 1H), 7.35 – 7.22 (m, 5H), 3.82 – 3.73 (m, 2H), 2.98 (t, J = 7.3 Hz, 2H); ^{13}C NMR (126 MHz, CDCl_3) δ 164.7, 148.1, 144.0, 143.6, 139.2, 137.6, 130.5, 128.9, 128.7, 126.6, 122.5, 116.7, 41.0, 36.1; HRMS (ESI) m/z calculated for $\text{C}_{16}\text{H}_{15}\text{N}_2\text{OS}$ $[\text{M}+\text{H}]^+$: 283.0900; found $[\text{M}+\text{H}]^+$: 283.0899.

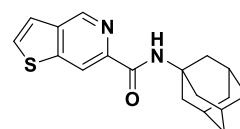
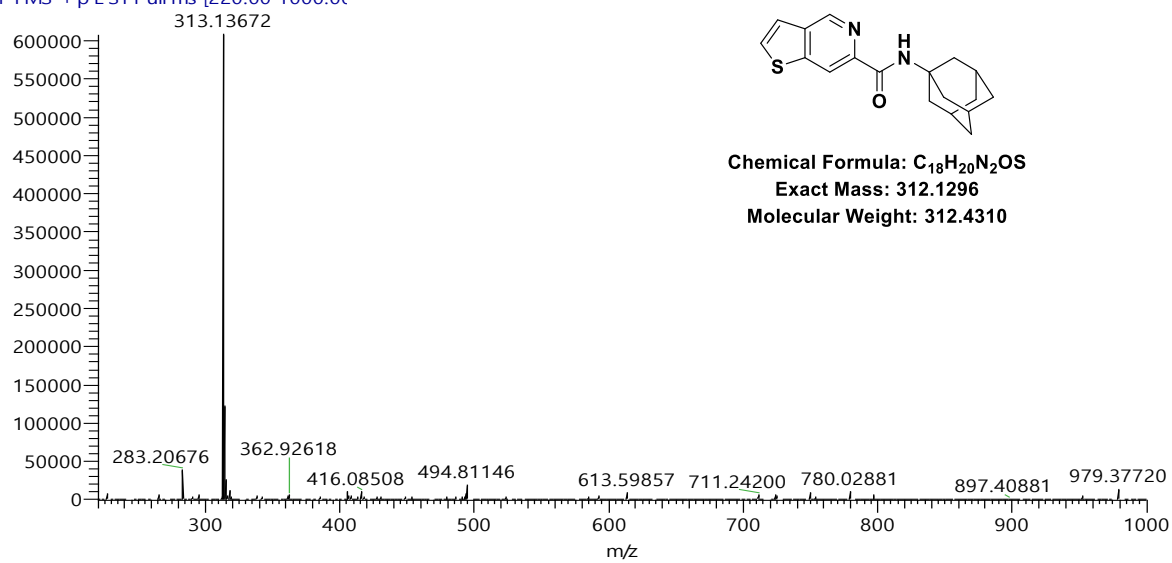
6.3. ^1H and ^{13}C NMR spectra, HRMS

N-((1*S*,3*S*)-adamantan-1-yl)thieno[3,2-*c*]pyridine-6-carboxamide (A3)



18mdv071-YZ479C #12 RT: 0.21658 /
T: FTMS +p ESI Full ms [220.00-1000.00]

:5

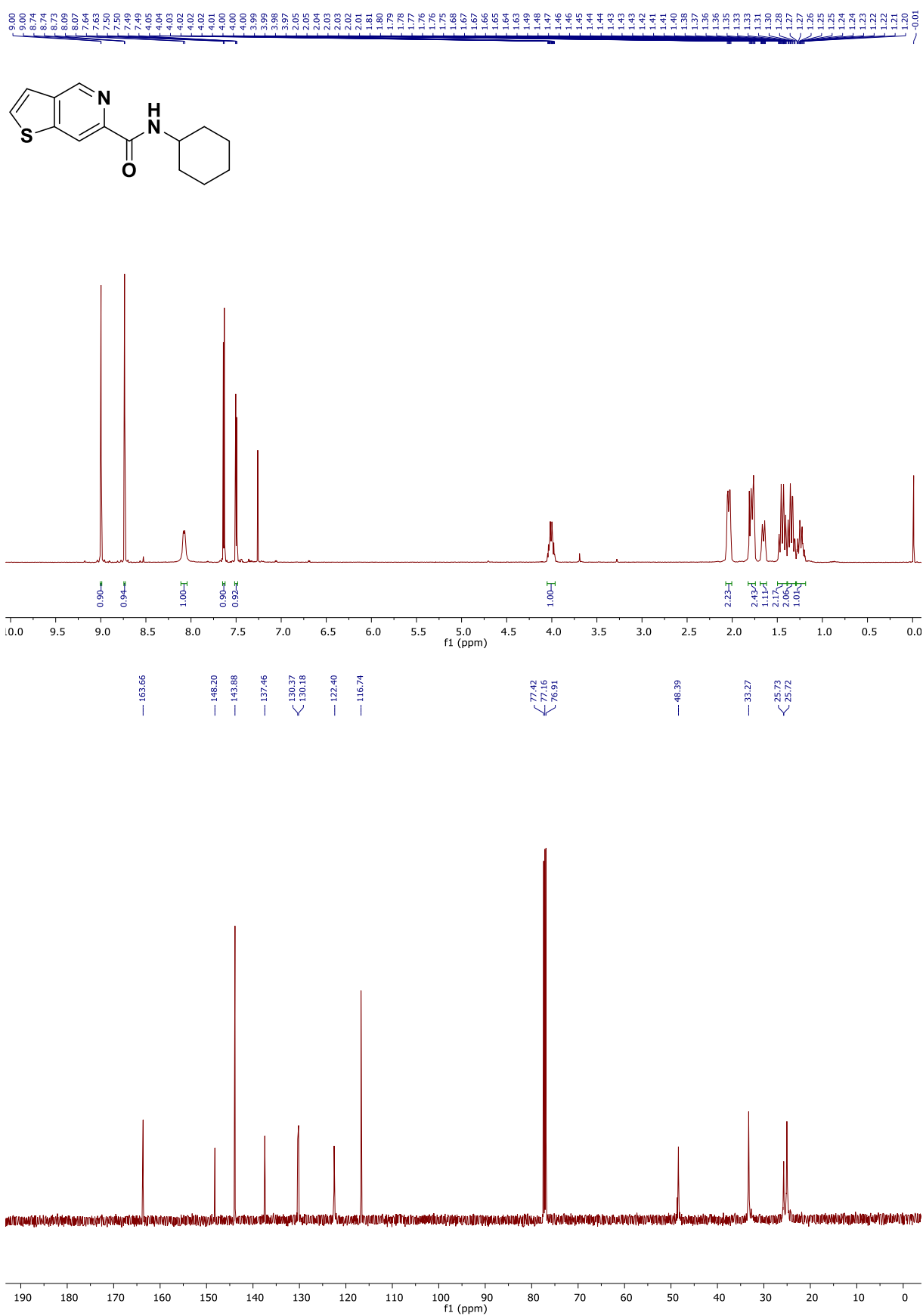


Chemical Formula: $C_{18}H_{20}N_2OS$

Exact Mass: 312.1296

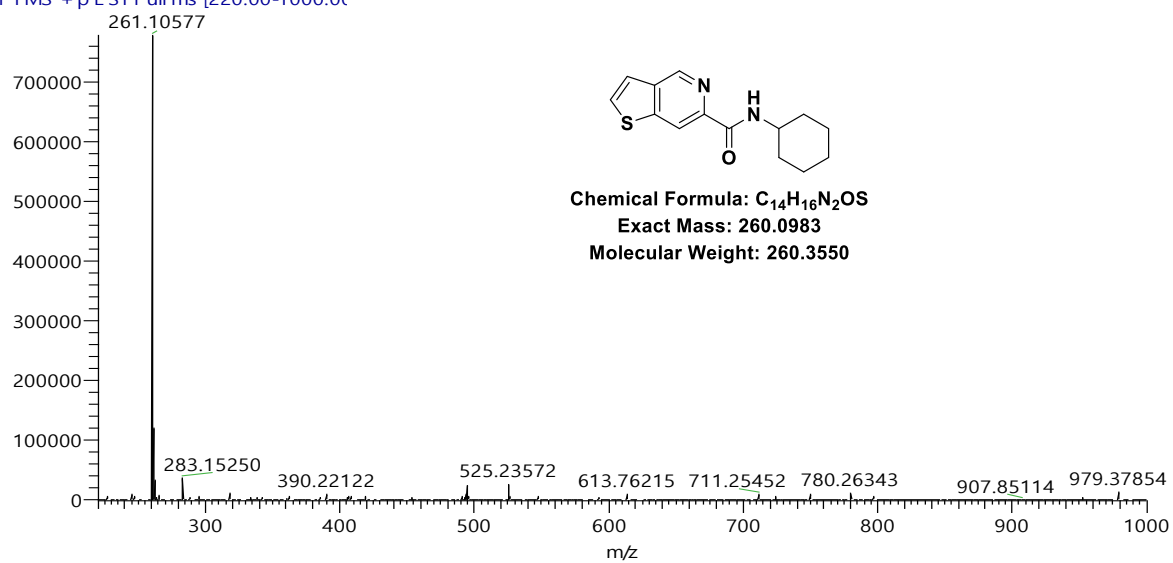
Molecular Weight: 312.4310

***N*-cyclohexylthieno[3,2-*c*]pyridine-6-carboxamide (A13)**

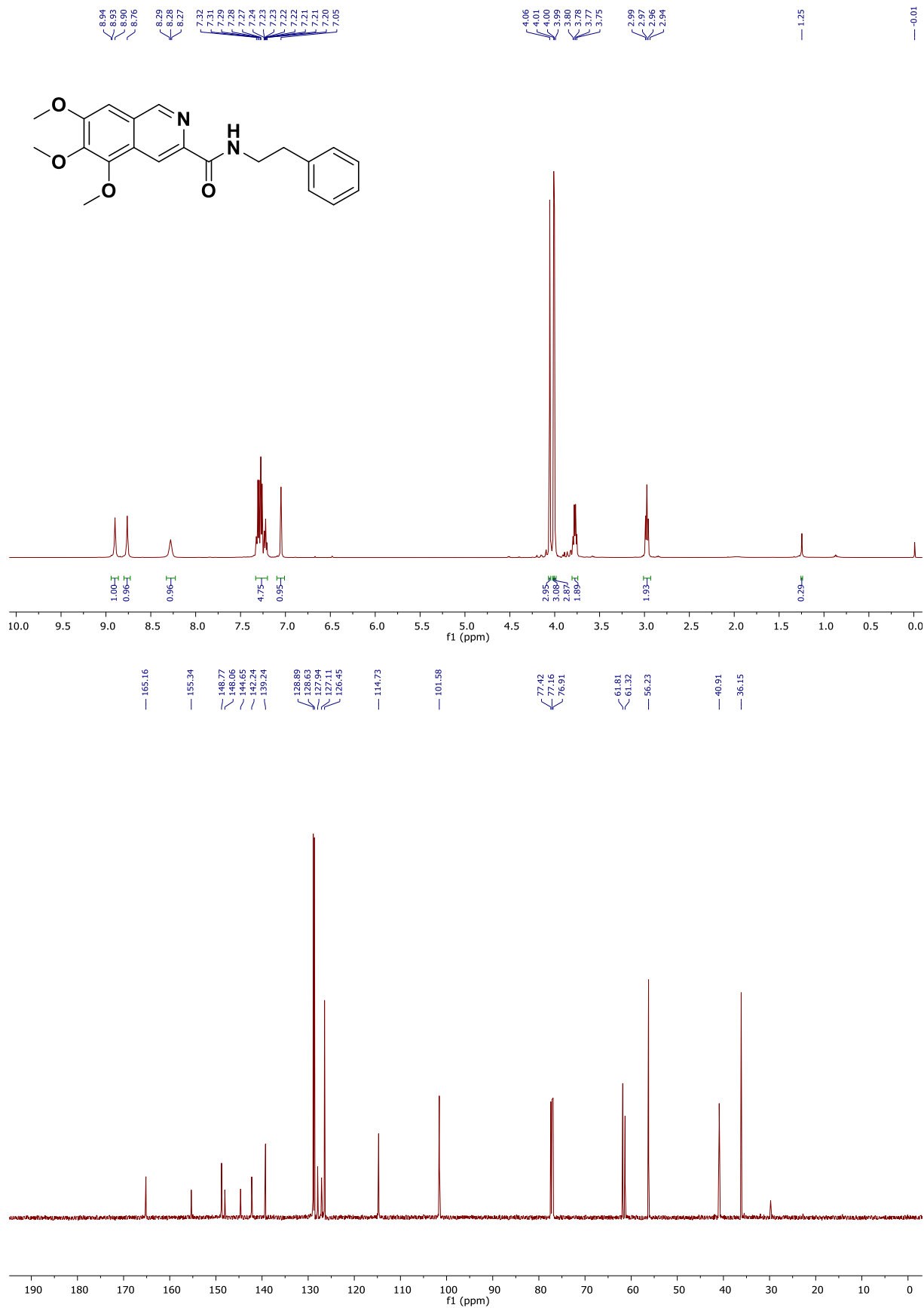


18mdv071-yz478C #12 RT: 0.20842 A
T: FTMS +p ESI Full ms [220.00-1000.00]

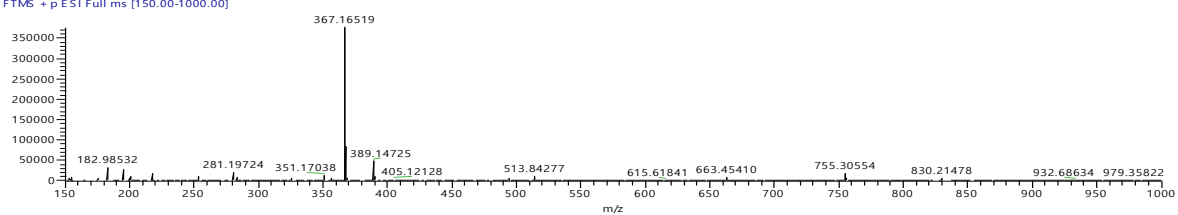
5



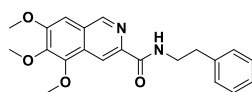
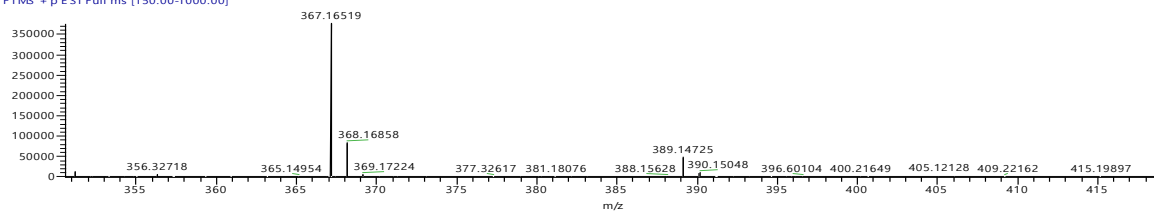
5,6,7-Trimethoxy-*N*-phenethylisoquinoline-3-carboxamide (A22)



17mdv196-YZ399C #32 RT: 0.58726 AV: 1 NL: 3.76E5
T: FTMS + p ESI Full ms [150.00-1000.00]



17mdv196-YZ399C #32 RT: 0.58726 AV: 1 NL: 3.76E5
T: FTMS + p ESI Full ms [150.00-1000.00]

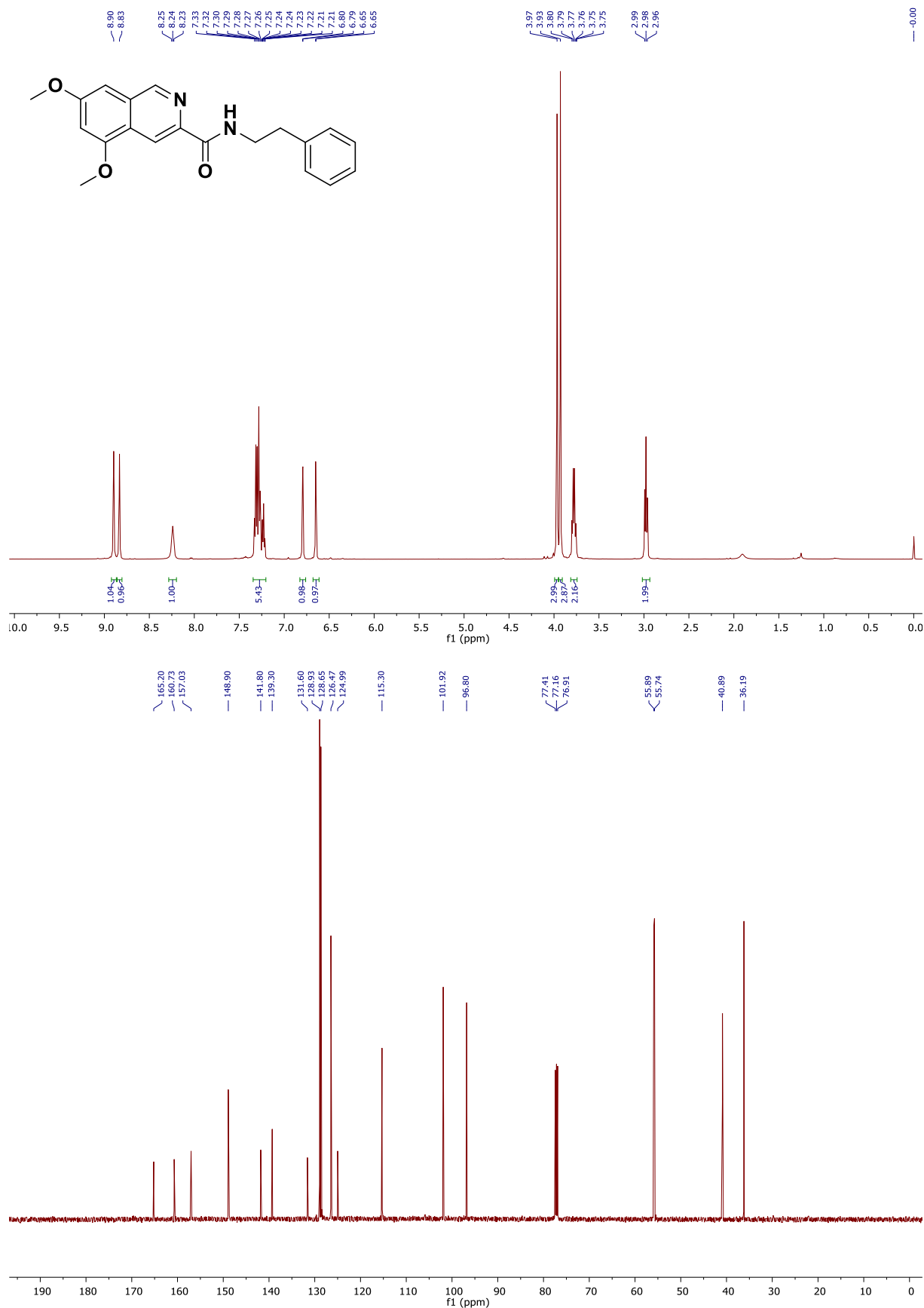


Chemical Formula: $C_{21}H_{22}N_2O_4$

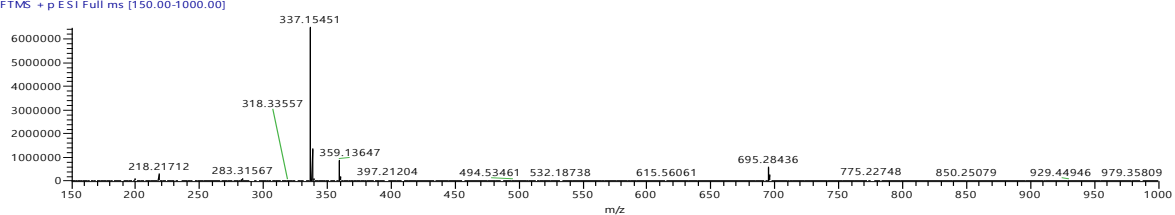
Exact Mass: 366.1580

Molecular Weight: 366.4170

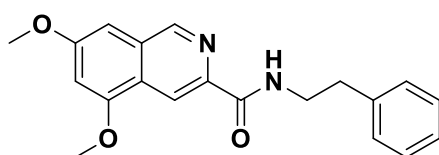
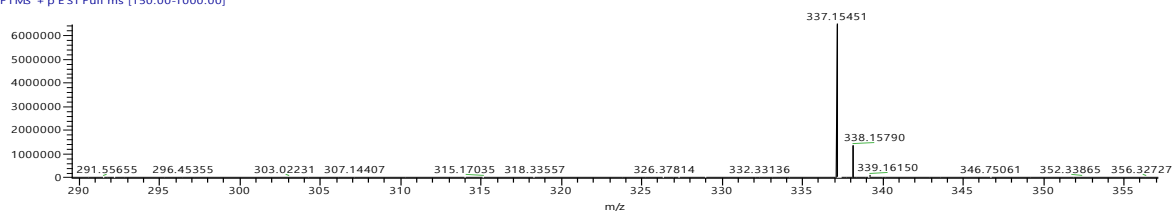
5,7-Dimethoxy-N-phenethylisoquinoline-3-carboxamide (B5)



17mdv196-YZ397C #25 RT: 0.41685 AV: 1 NL: 6.48E6
T: FTMS + p ESI Full ms [150.00-1000.00]



17mdv196-YZ397C #25 RT: 0.41685 AV: 1 NL: 6.48E6
T: FTMS + p ESI Full ms [150.00-1000.00]

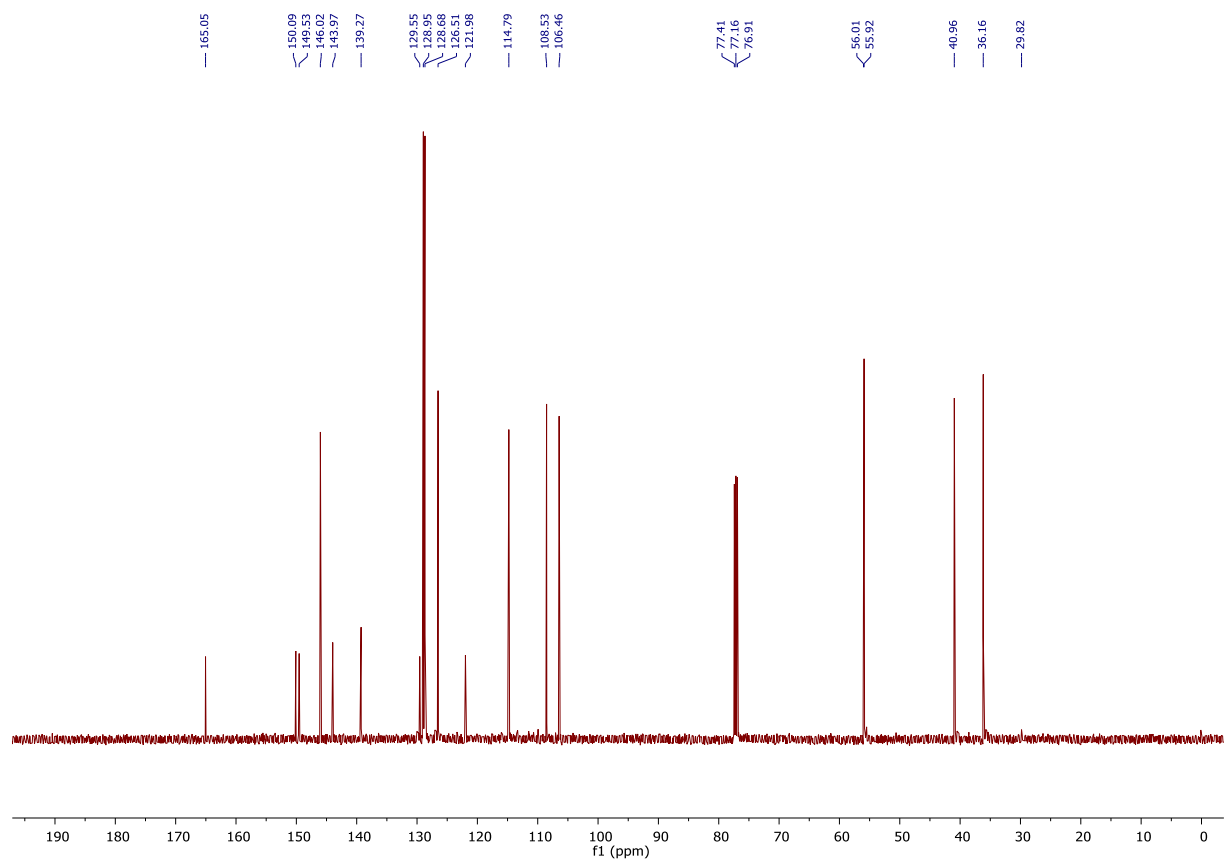
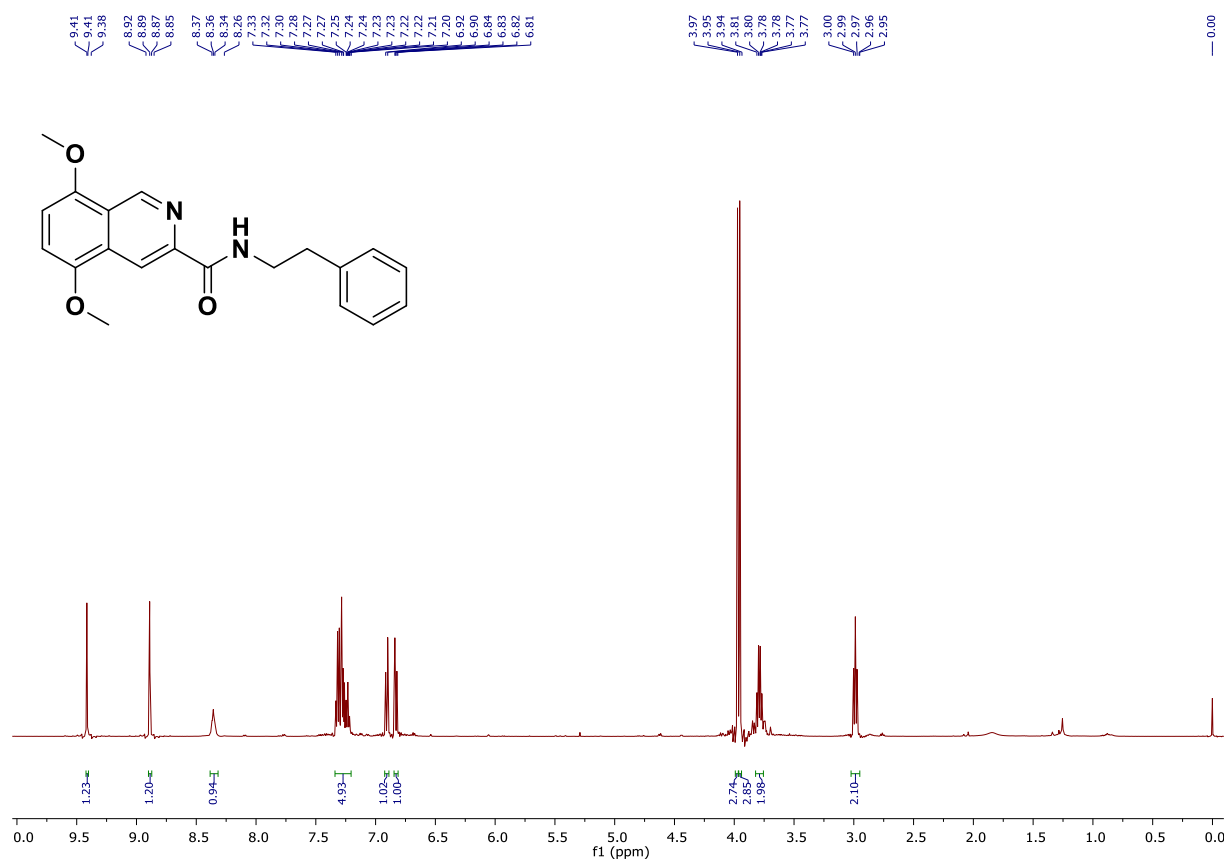


Chemical Formula: $C_{20}H_{20}N_2O_3$

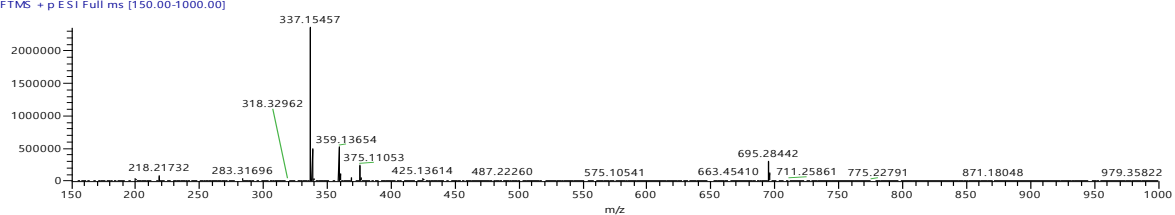
Exact Mass: 336,15

Molecular Weight: 336,39

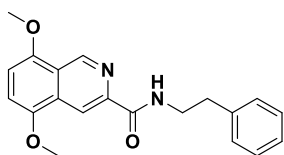
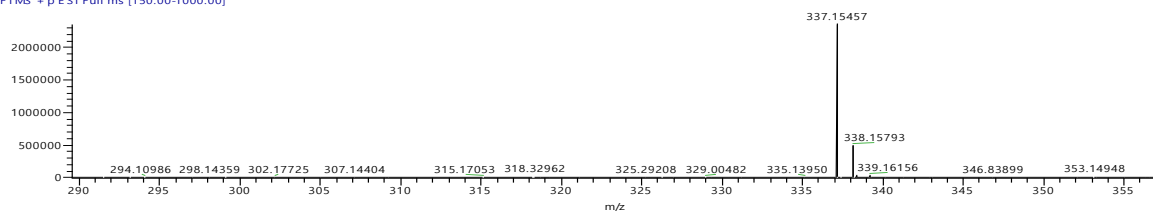
5,8-Dimethoxy-*N*-phenethylisoquinoline-3-carboxamide (B16)



17mdv196-YZ410C #24 RT: 0.41727 AV: 1 NL: 2.36E6
T: FTMS + p ESI Full ms [150.00-1000.00]



17mdv196-YZ410C #24 RT: 0.41727 AV: 1 NL: 2.36E6
T: FTMS + p ESI Full ms [150.00-1000.00]

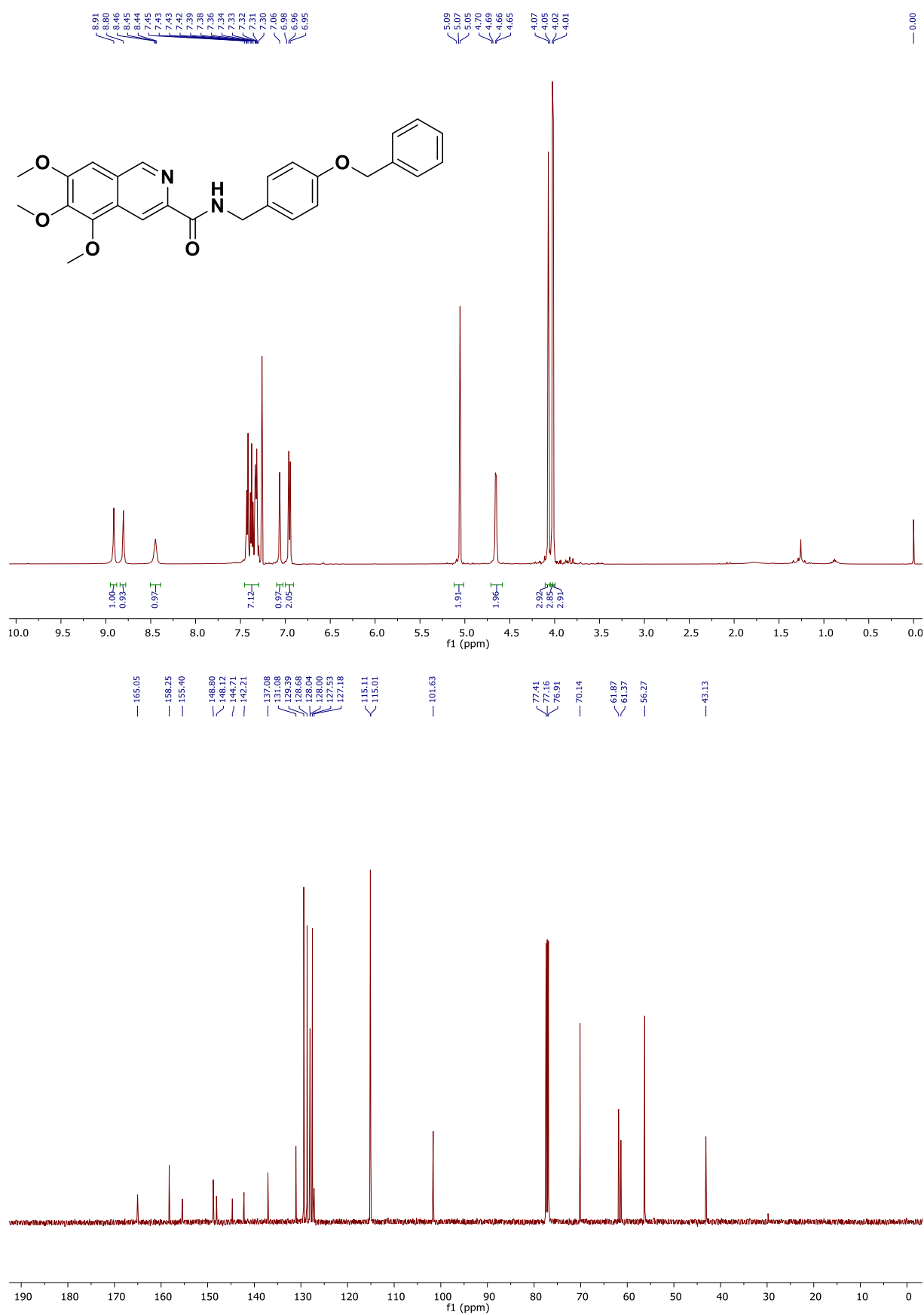


Chemical Formula: $C_{20}H_{20}N_2O_3$

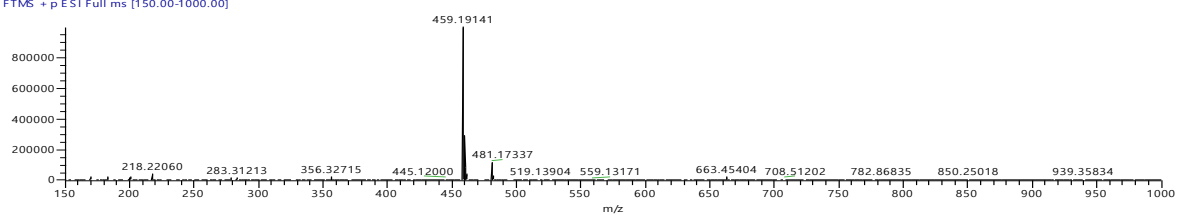
Exact Mass: 336.1474

Molecular Weight: 336.3910

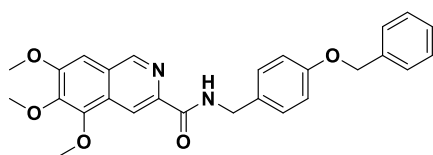
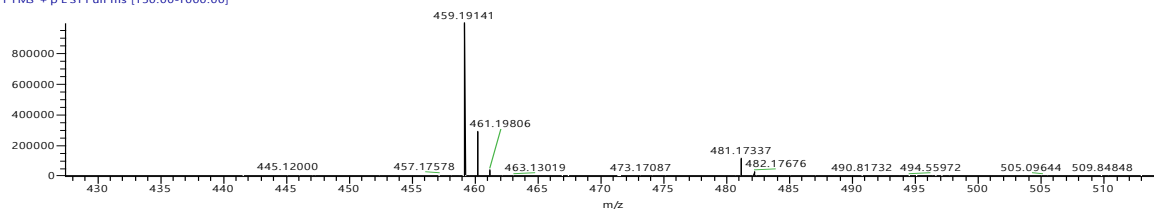
***N*-(4-(benzyloxy)benzyl)-5,6,7-trimethoxyisoquinoline-3-carboxamide (B24)**



17mdv196-YZ440C #34 RT: 0.58348 AV: 1 NL: 9.98E5
T: FTMS + p ESI Full ms [150.00-1000.00]



17mdv196-YZ440C #34 RT: 0.58348 AV: 1 NL: 9.98E5
T: FTMS + p ESI Full ms [150.00-1000.00]

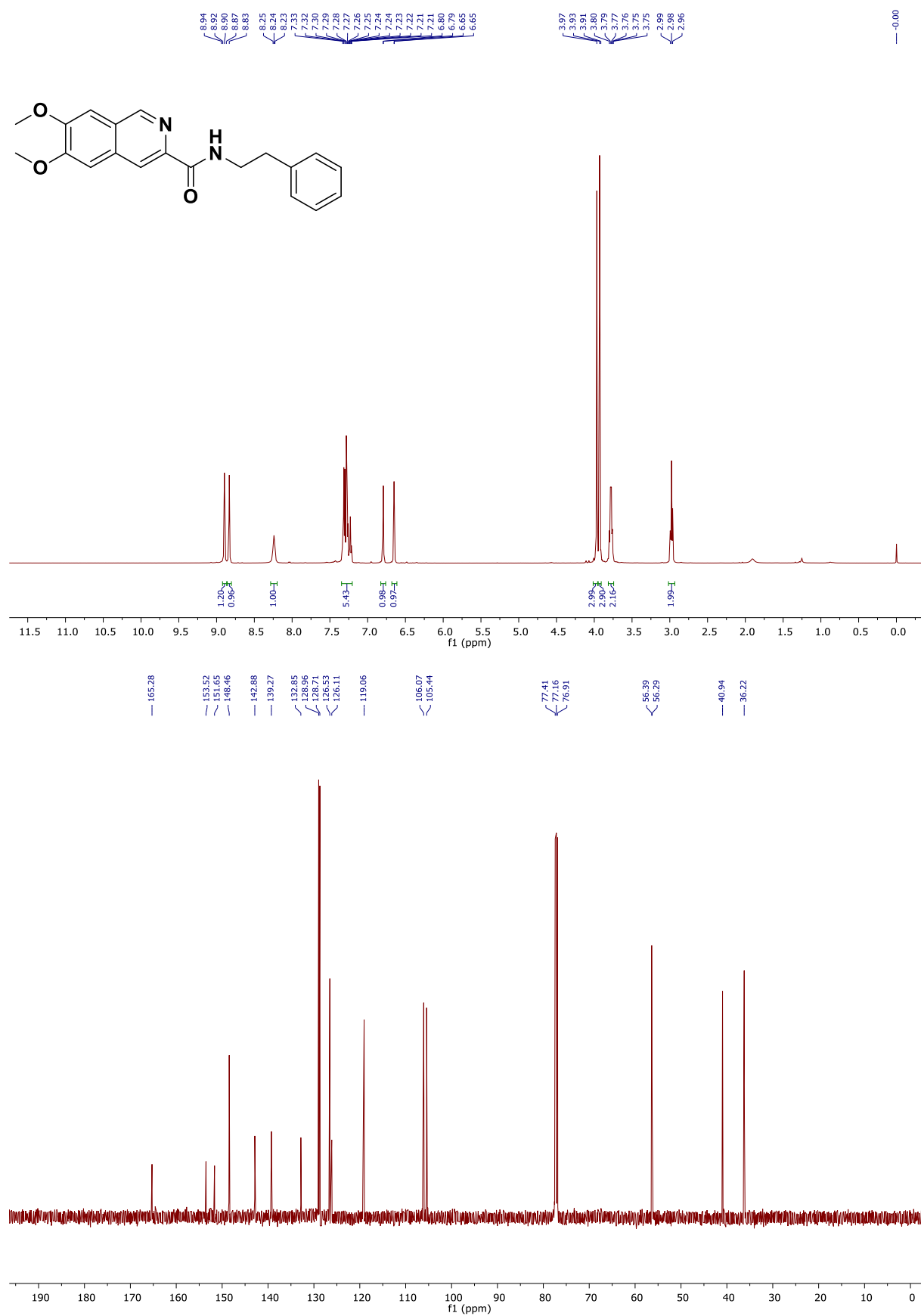


Chemical Formula: $C_{27}H_{26}N_2O_5$

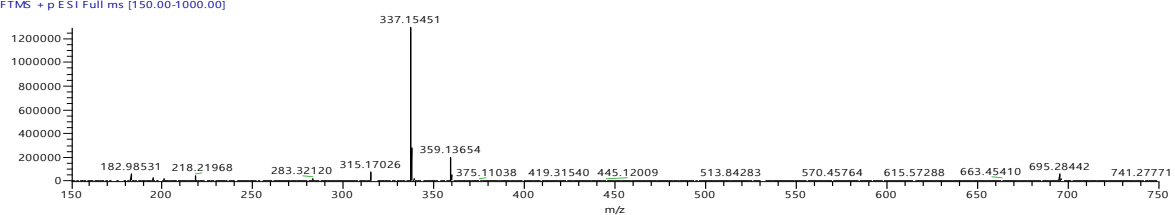
Exact Mass: 458.1842

Molecular Weight: 458.5140

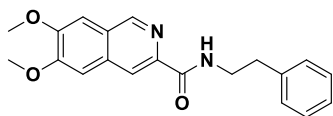
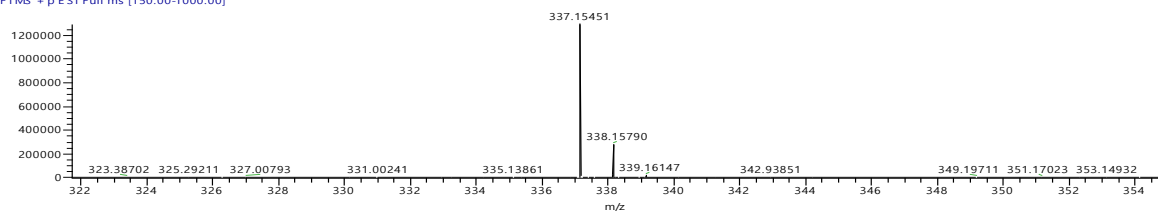
6,7-Dimethoxy-*N*-phenethylisoquinoline-3-carboxamide (C2)



17mdv196-YZ386C #29 RT: 0.50314 AV: 1 NL: 1.29E6
T: FTMS + p ESI Full ms [150.00-1000.00]



17mdv196-YZ386C #29 RT: 0.50314 AV: 1 NL: 1.29E6
T: FTMS + p ESI Full ms [150.00-1000.00]

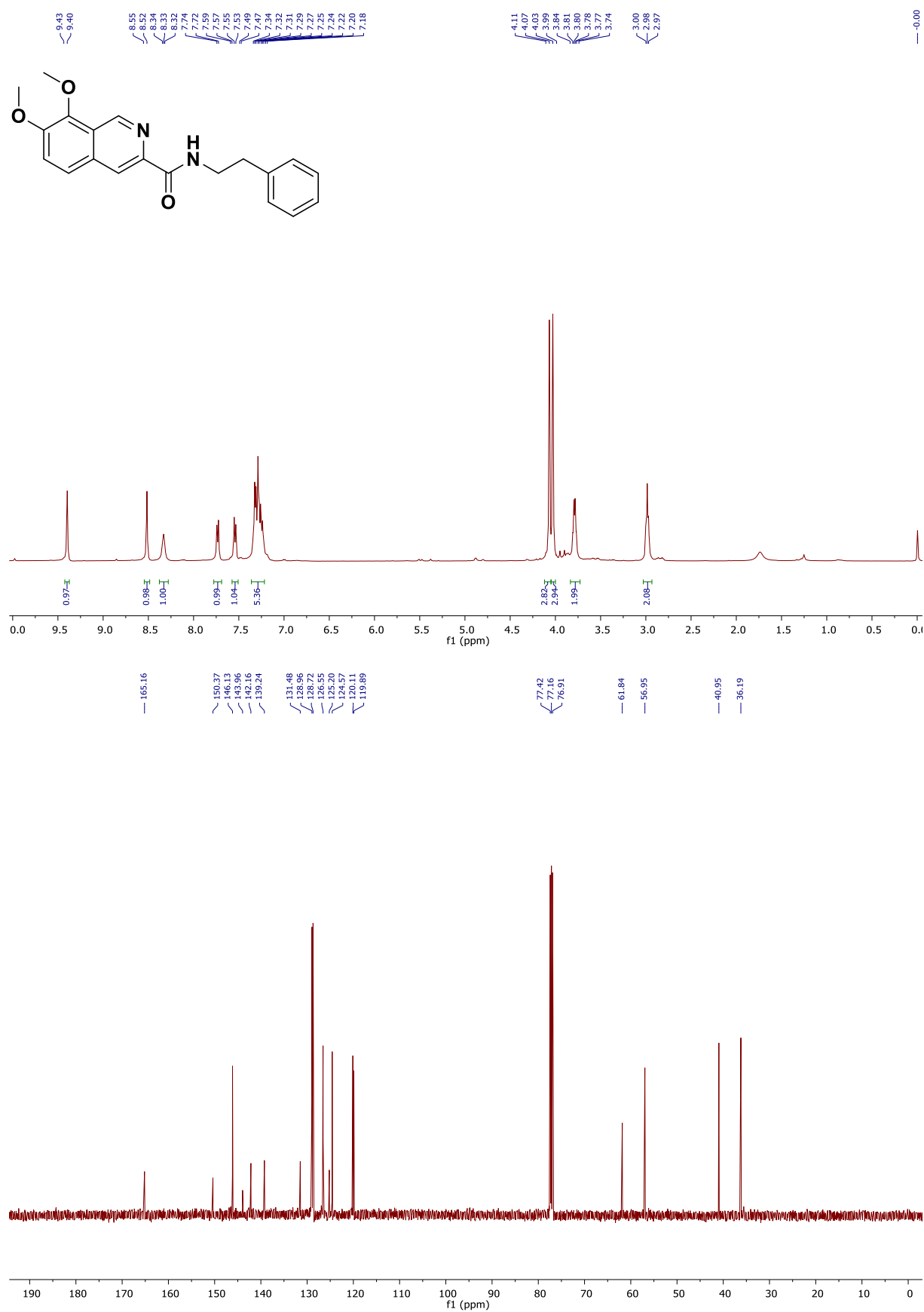


Chemical Formula: $C_{20}H_{20}N_2O_3$

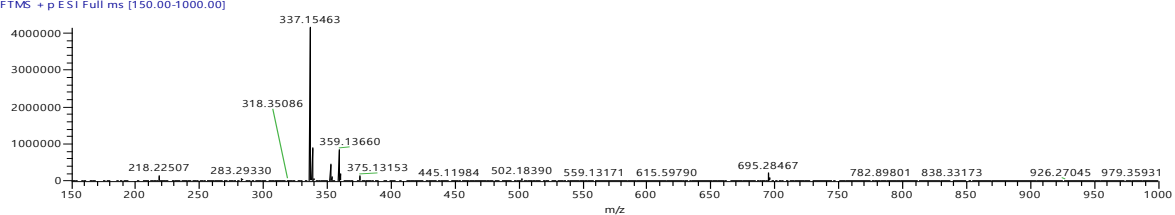
Exact Mass: 336.1474

Molecular Weight: 336.3910

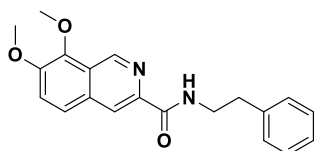
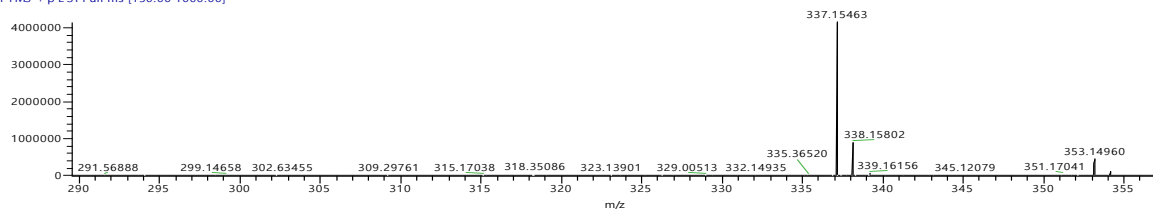
7,8-Dimethoxy-*N*-phenethylisoquinoline-3-carboxamide (C10)



17mdv196-YZ408C #24 RT: 0.40957 AV: 1 NL: 4.15E6
T: FTMS + p ESI Full ms [150.00-1000.00]



17mdv196-YZ408C #24 RT: 0.40957 AV: 1 NL: 4.15E6
T: FTMS + p ESI Full ms [150.00-1000.00]

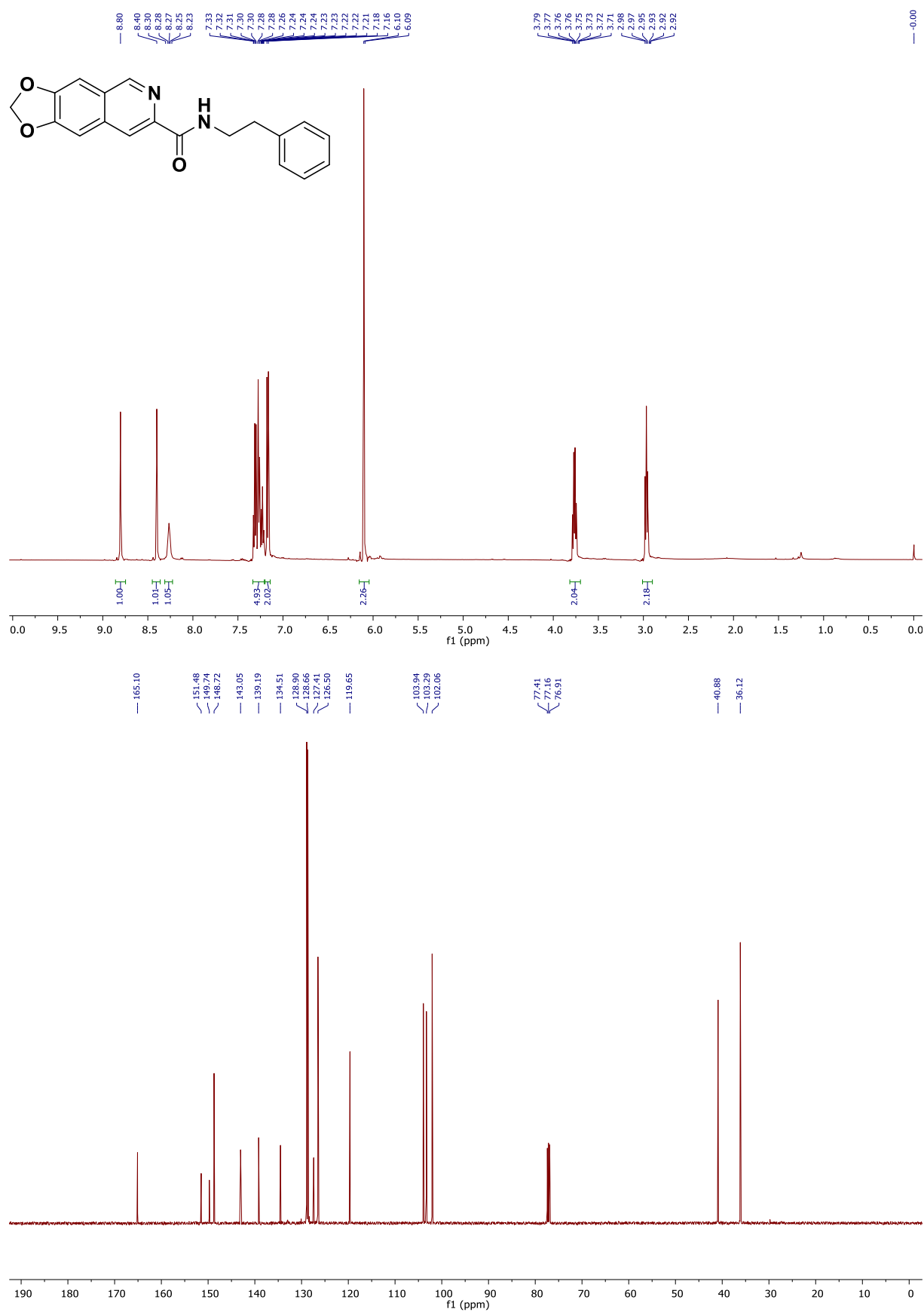


Chemical Formula: $C_{20}H_{20}N_2O_3$

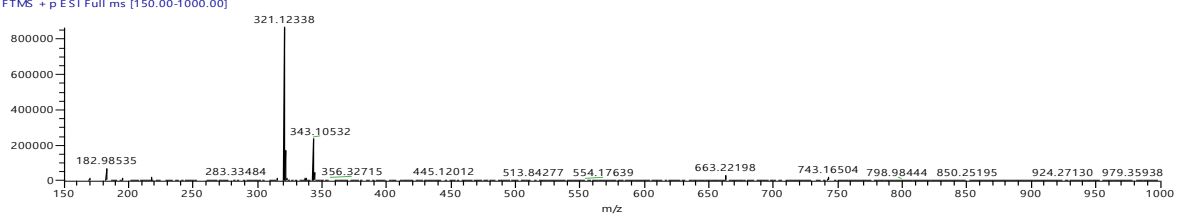
Exact Mass: 336.1474

Molecular Weight: 336.3910

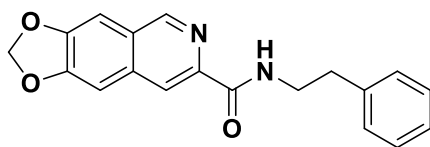
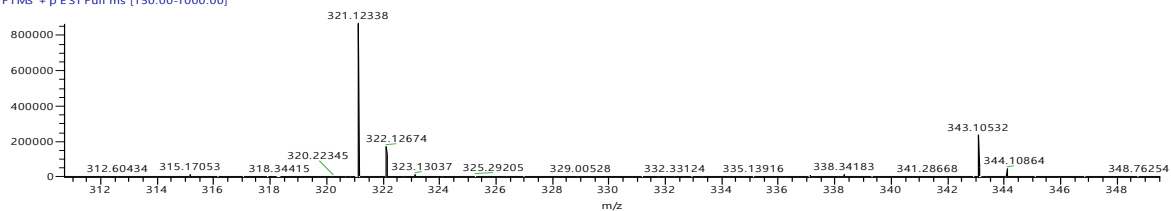
***N*-phenethyl-[1,3]dioxolo[4,5-*f*]isoquinoline-8-carboxamide (D19)**



17mdv196-YZ421C #24 RT: 0.41906 AV: 1 NL: 8.65E5
T: FTMS + p ESI Full ms [150.00-1000.00]



17mdv196-YZ421C #24 RT: 0.41906 AV: 1 NL: 8.65E5
T: FTMS + p ESI Full ms [150.00-1000.00]

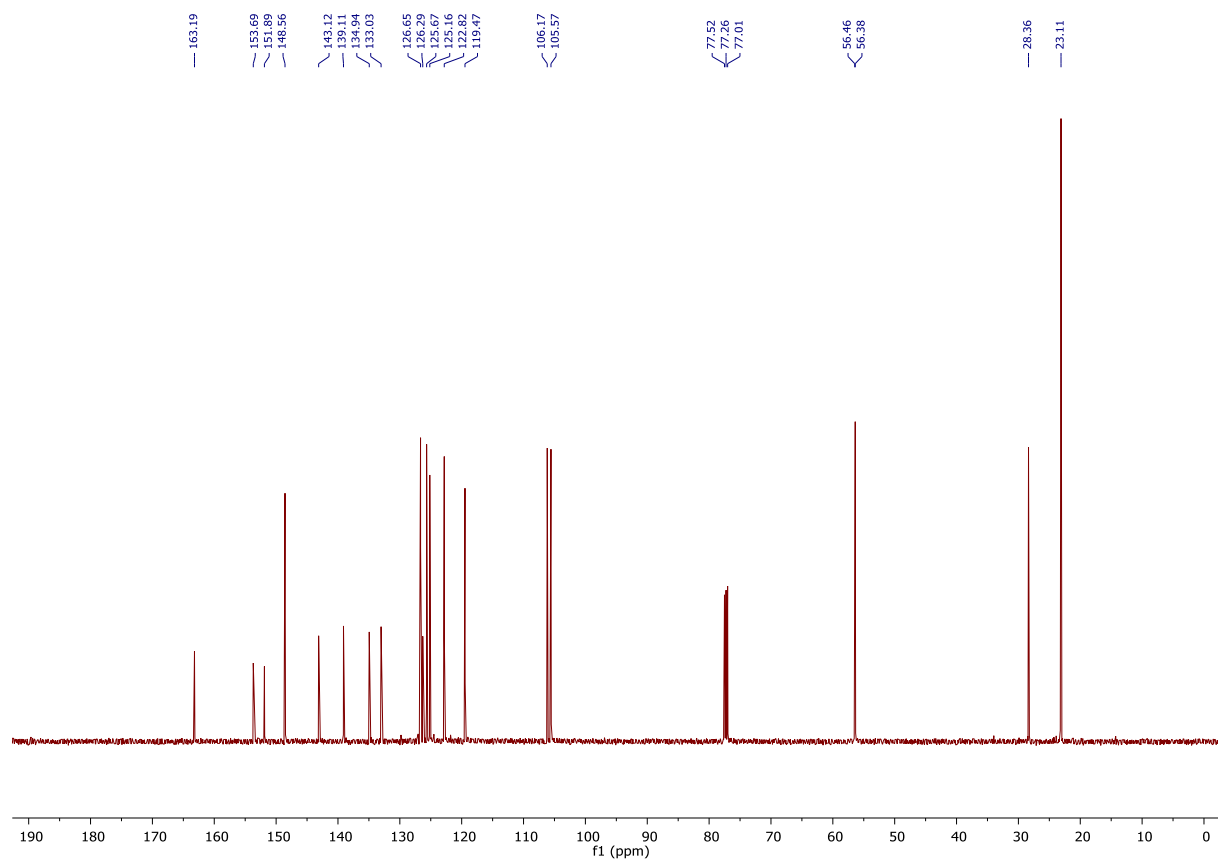
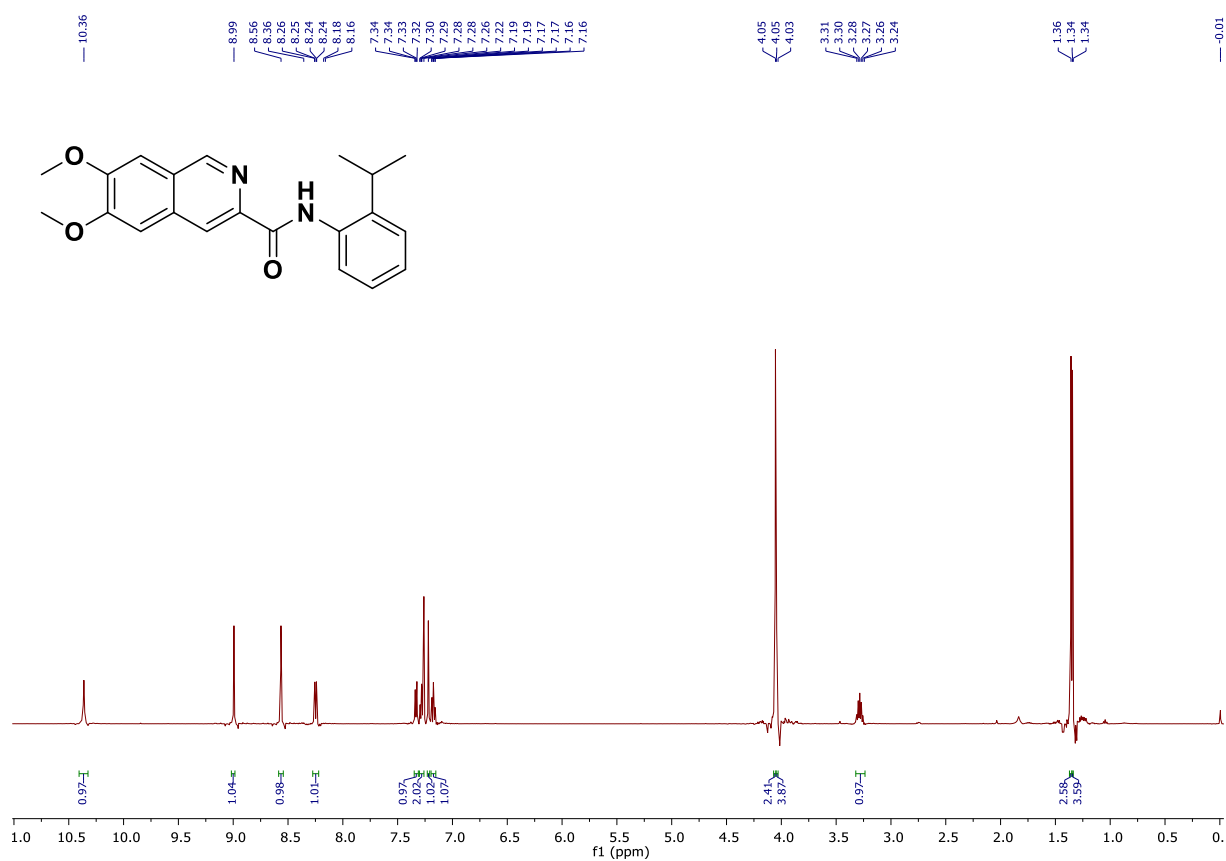


Chemical Formula: $C_{19}H_{16}N_2O_3$

Exact Mass: 320,1161

Molecular Weight: 320,3480

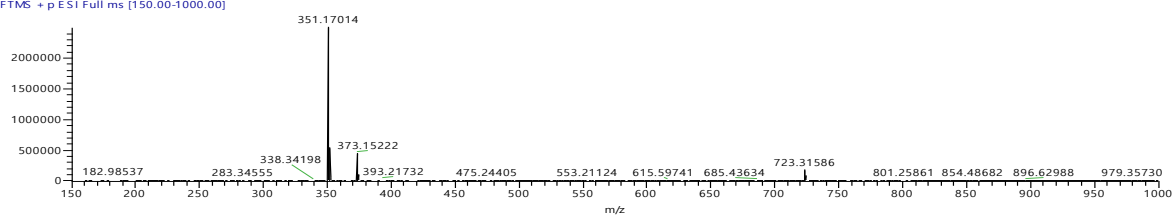
***N*-(2-isopropylphenyl)-6,7-dimethoxyisoquinoline-3-carboxamide (E7)**



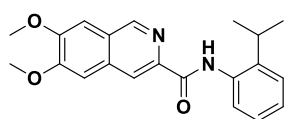
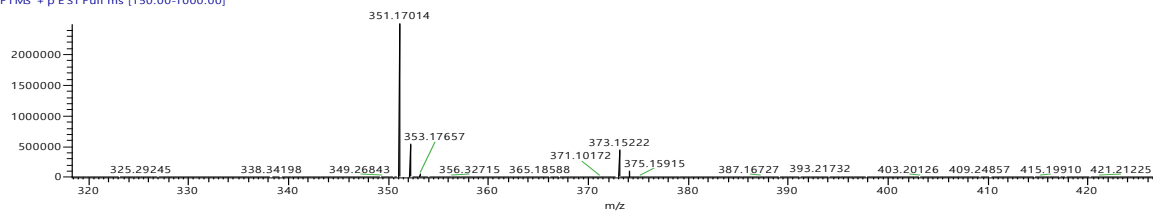
17mdv196-YZ423C

10/06/17 12:40:27

17mdv196-YZ423C #24 RT: 0.40346 AV: 1 NL: 2.50E6
T: FTMS + p ESI Full ms [150.00-1000.00]



17mdv196-YZ423C #24 RT: 0.40346 AV: 1 NL: 2.50E6
T: FTMS + p ESI Full ms [150.00-1000.00]

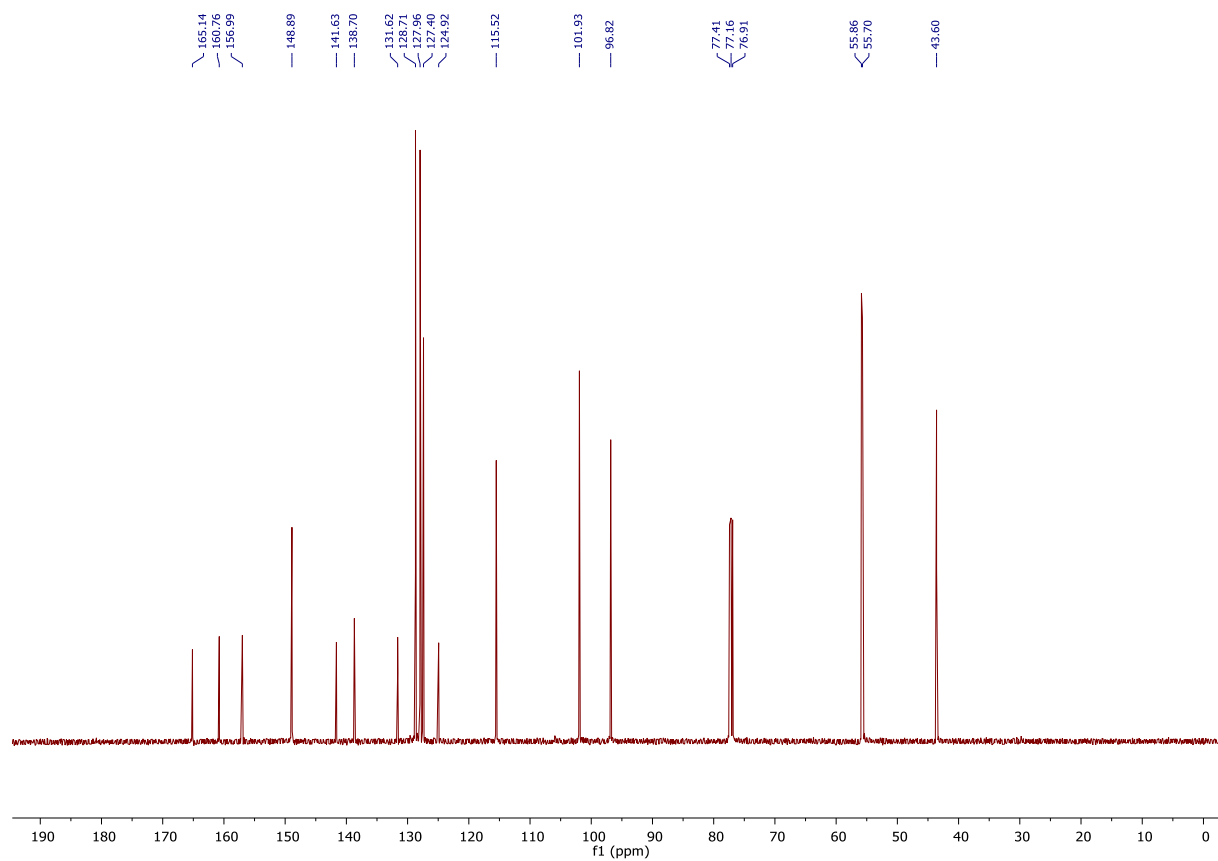
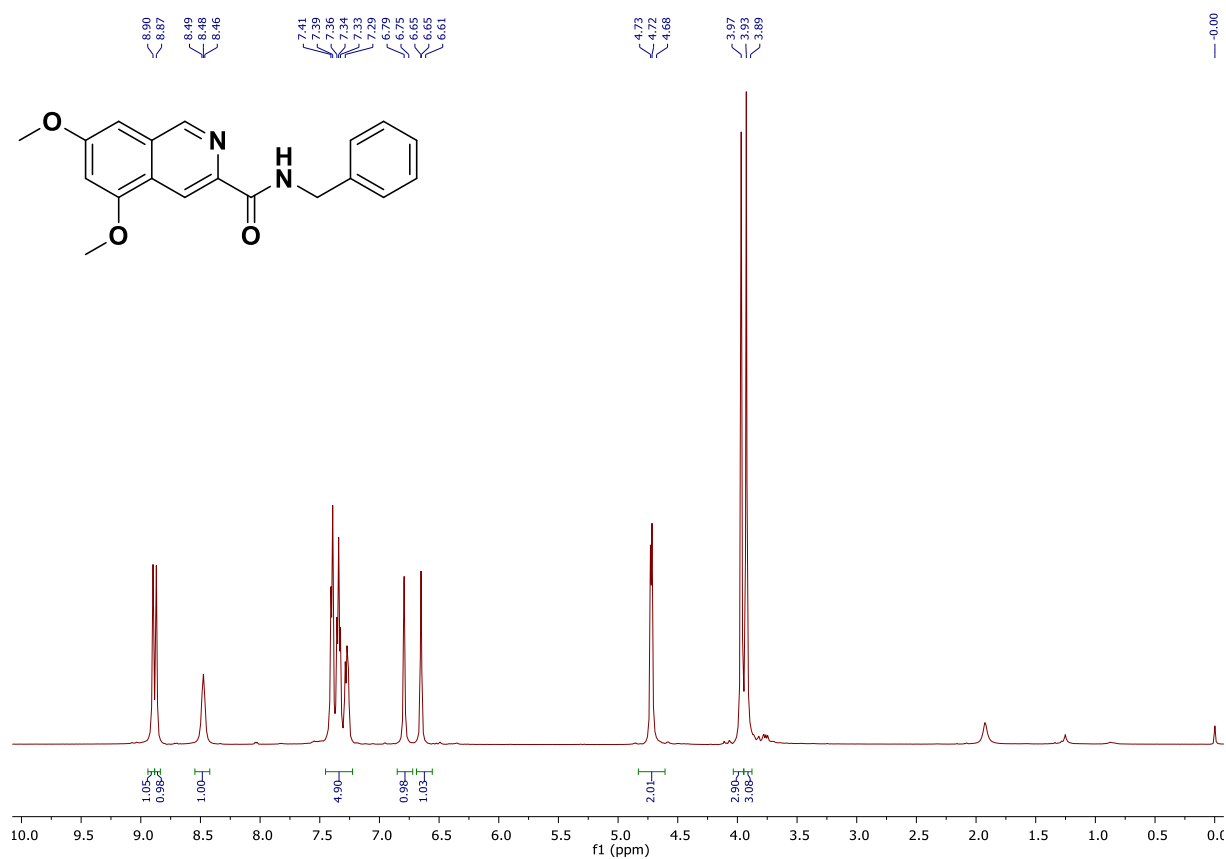


Chemical Formula: $C_{21}H_{22}N_2O_3$

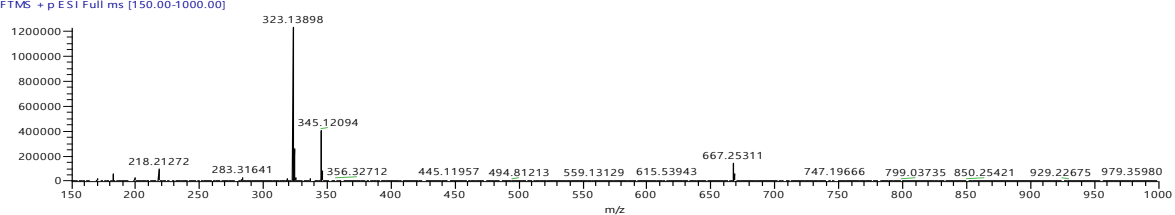
Exact Mass: 350.1630

Molecular Weight: 350.4180

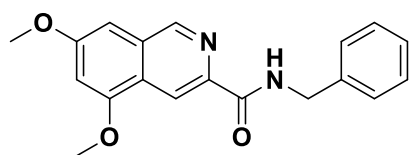
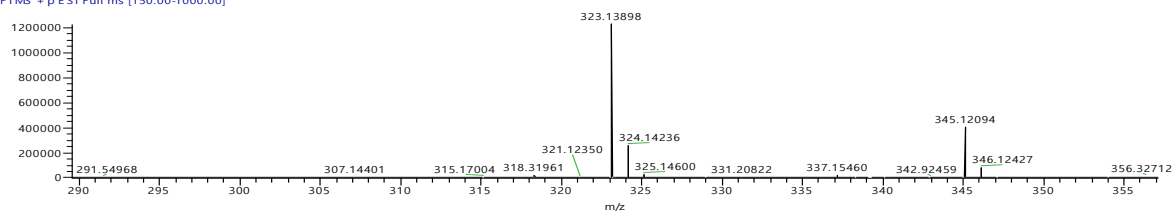
***N*-benzyl-5,7-dimethoxyisoquinoline-3-carboxamide (E12)**



17mdv196-YZ398C #24 RT: 0.41302 AV: 1 NL: 1.22E6
T: FTMS + p ESI Full ms [150.00-1000.00]



17mdv196-YZ398C #24 RT: 0.41302 AV: 1 NL: 1.22E6
T: FTMS + p ESI Full ms [150.00-1000.00]

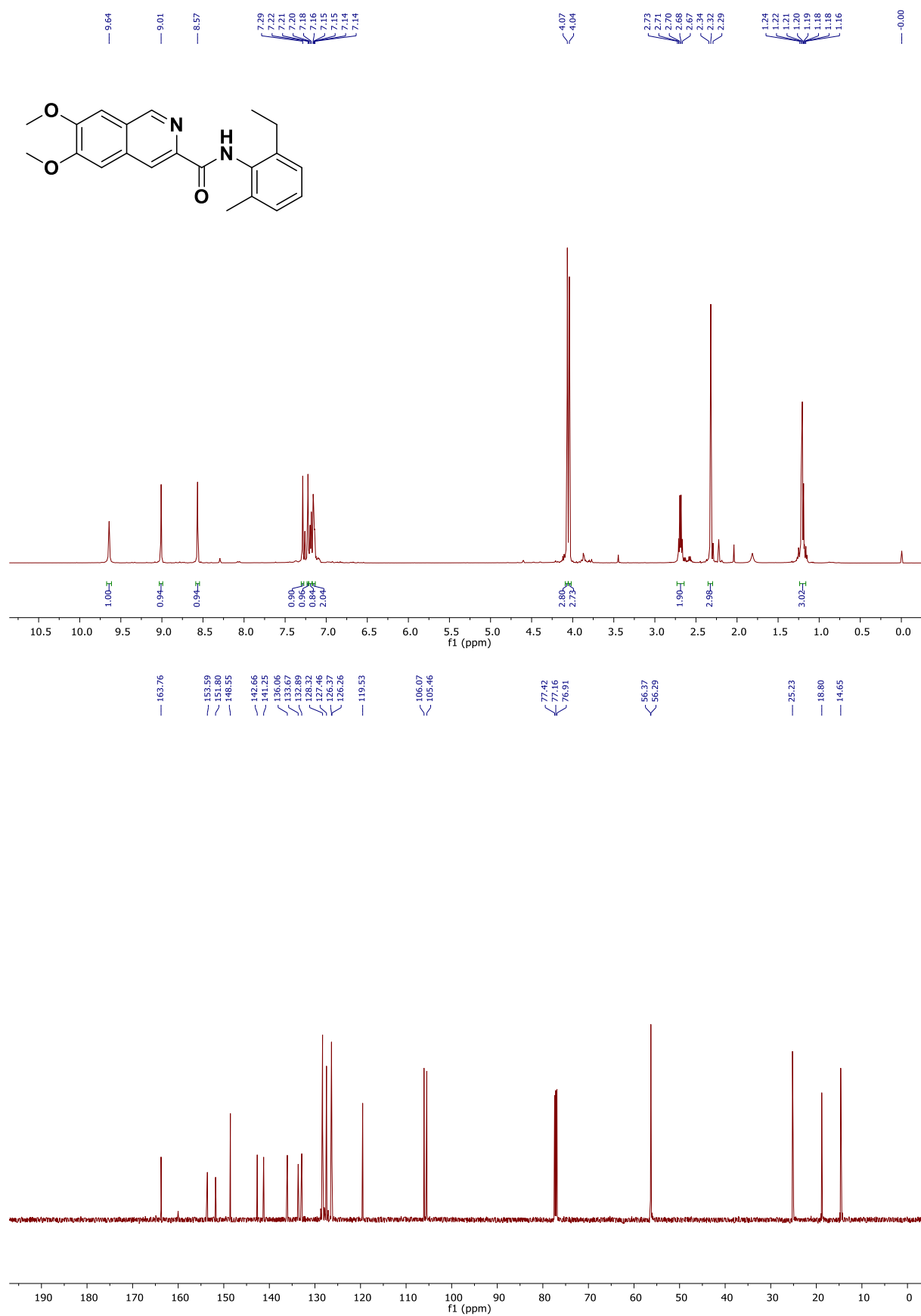


Chemical Formula: C₁₉H₁₈N₂O₃

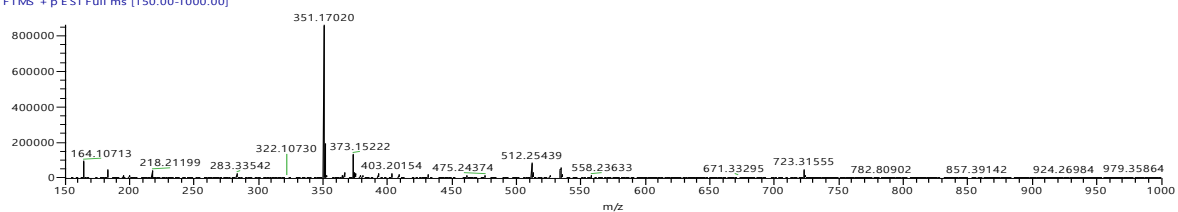
Exact Mass: 322,13

Molecular Weight: 322,36

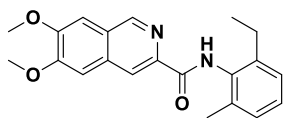
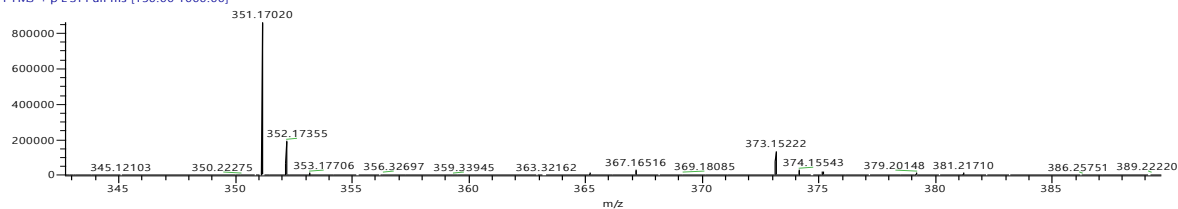
***N*-(2-ethyl-6-methylphenyl)-6,7-dimethoxyisoquinoline-3-carboxamide (E17)**



17mdv196-YZ422C #24 RT: 0.40985 AV: 1 NL: 8.62E5
T: FTMS + p ESI Full ms [150.00-1000.00]



17mdv196-YZ422C #24 RT: 0.40985 AV: 1 NL: 8.62E5
T: FTMS + p ESI Full ms [150.00-1000.00]

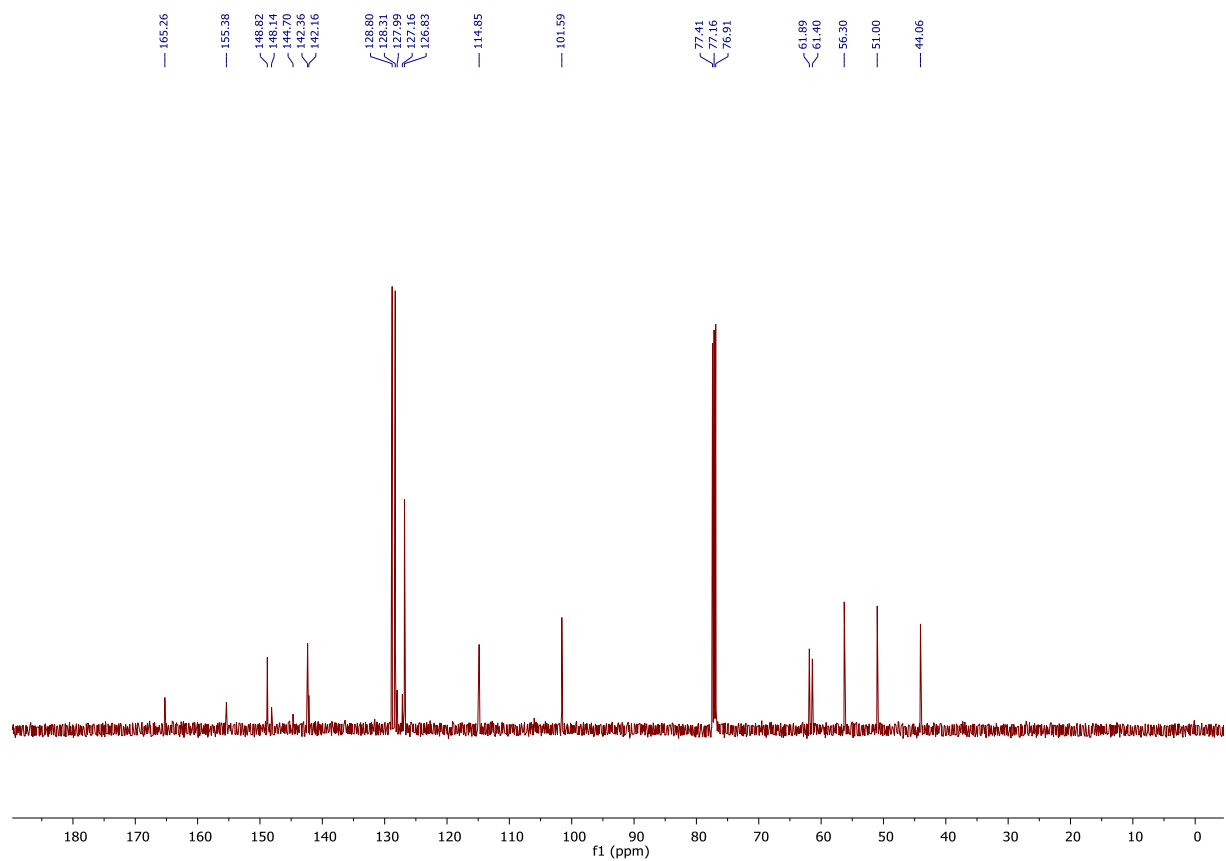
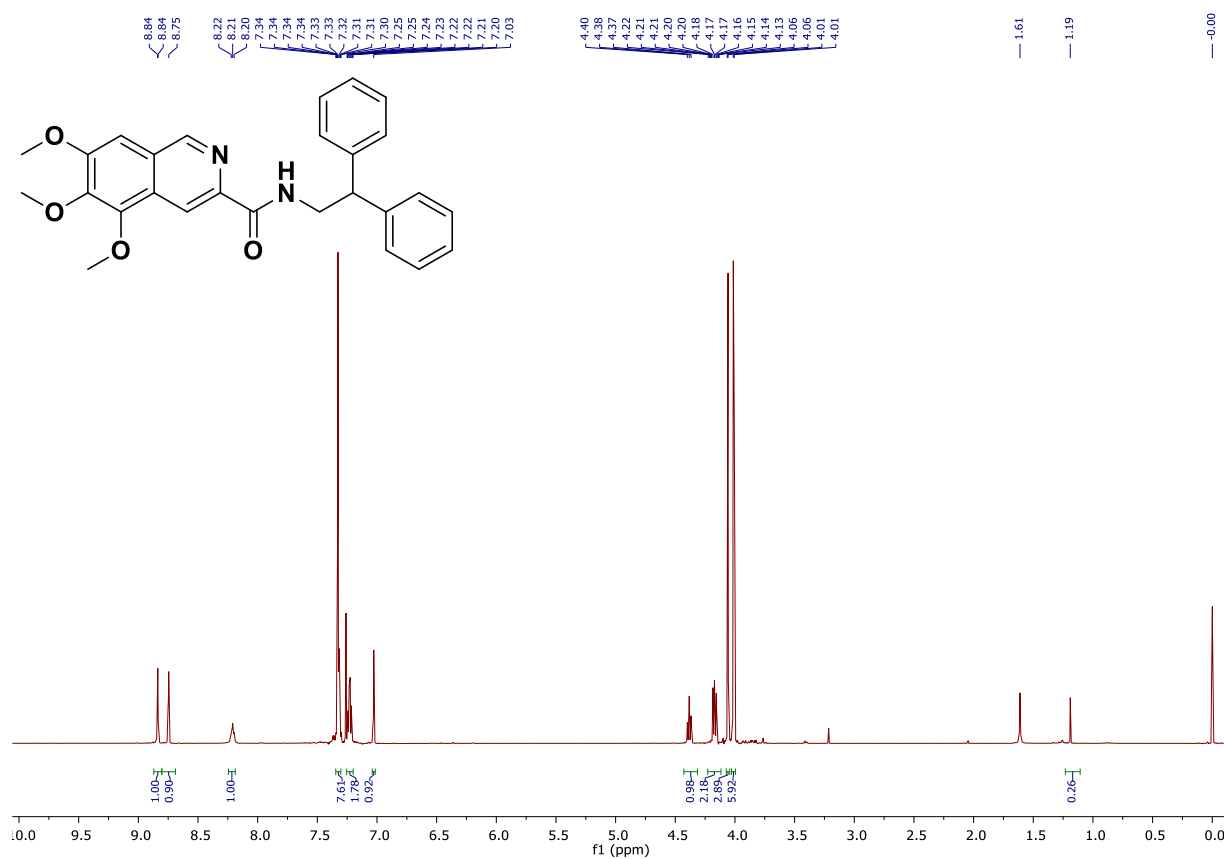


Chemical Formula: $C_{21}H_{22}N_2O_3$

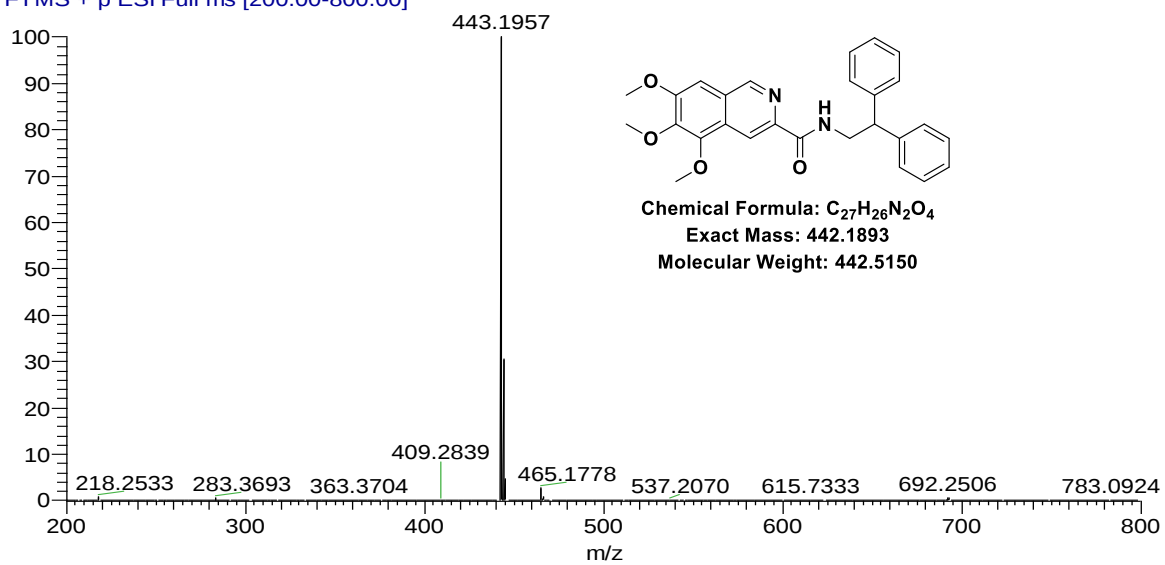
Exact Mass: 350.1630

Molecular Weight: 350.4180

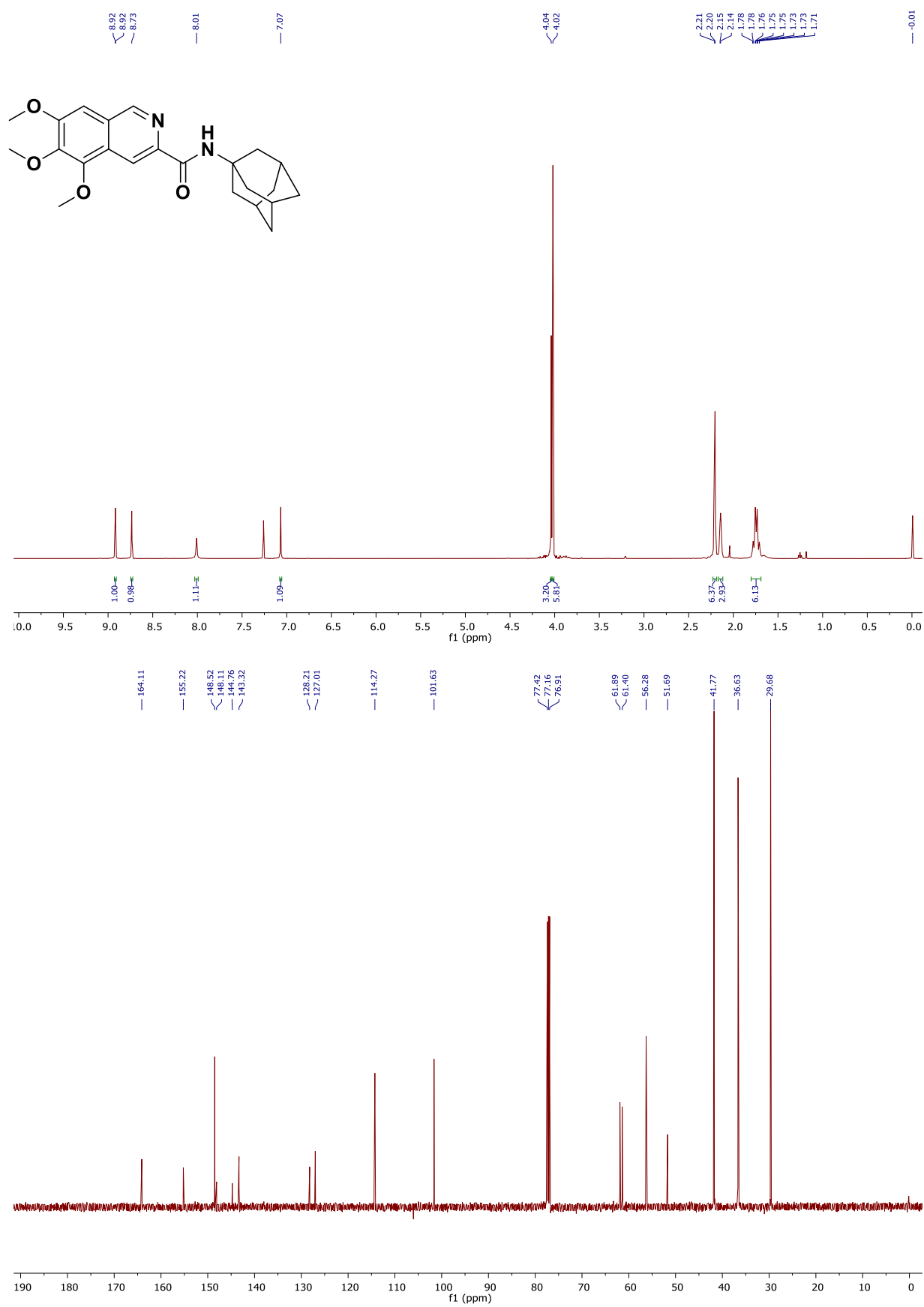
***N*-(2,2-diphenylethyl)-5,6,7-trimethoxyisoquinoline-3-carboxamide (F8)**



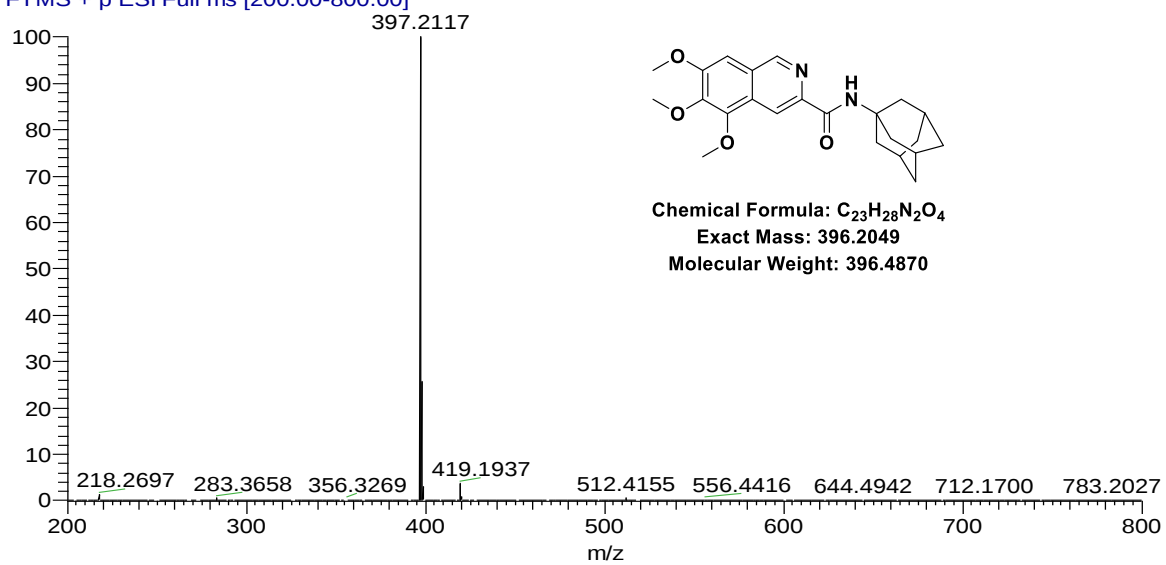
17mdv207-yz444c #280 RT: 5.25 AV: 1 NL: 2.69E8
T: FTMS + p ESI Full ms [200.00-800.00]



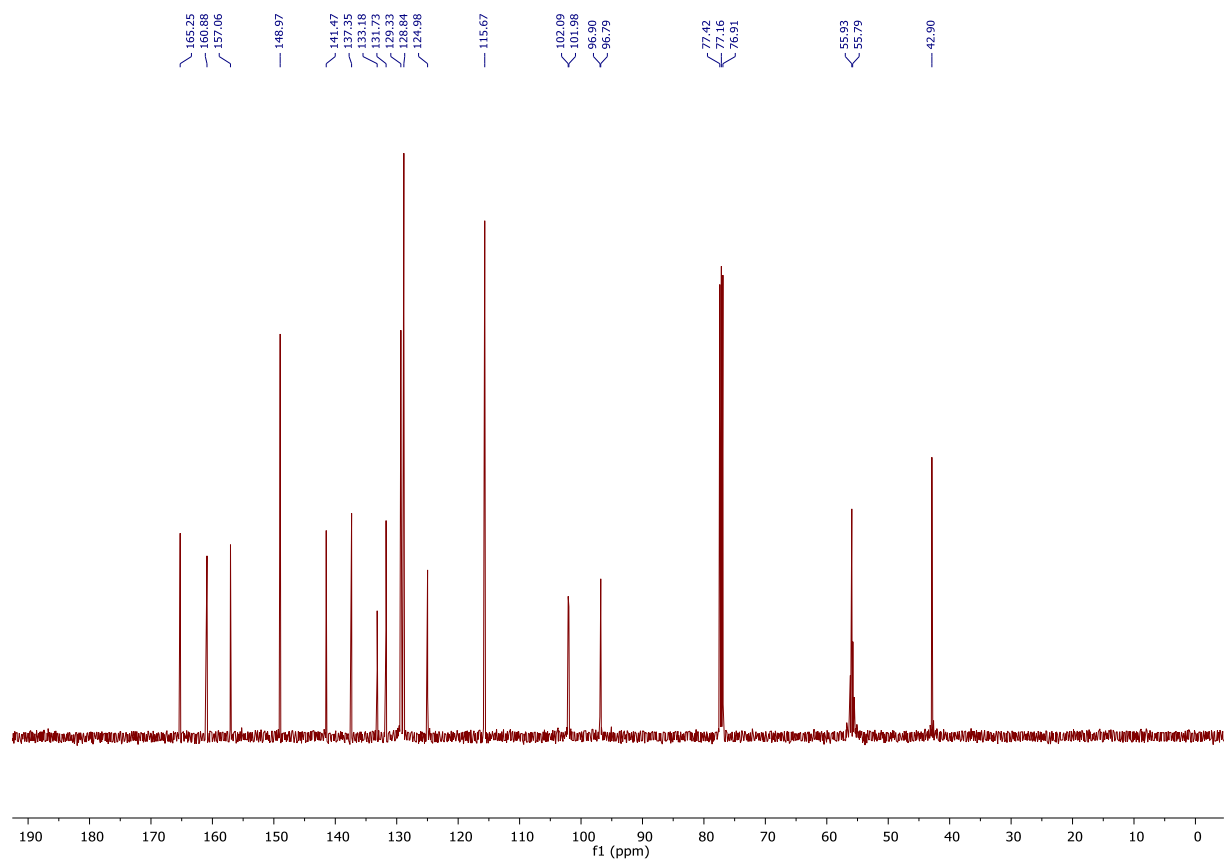
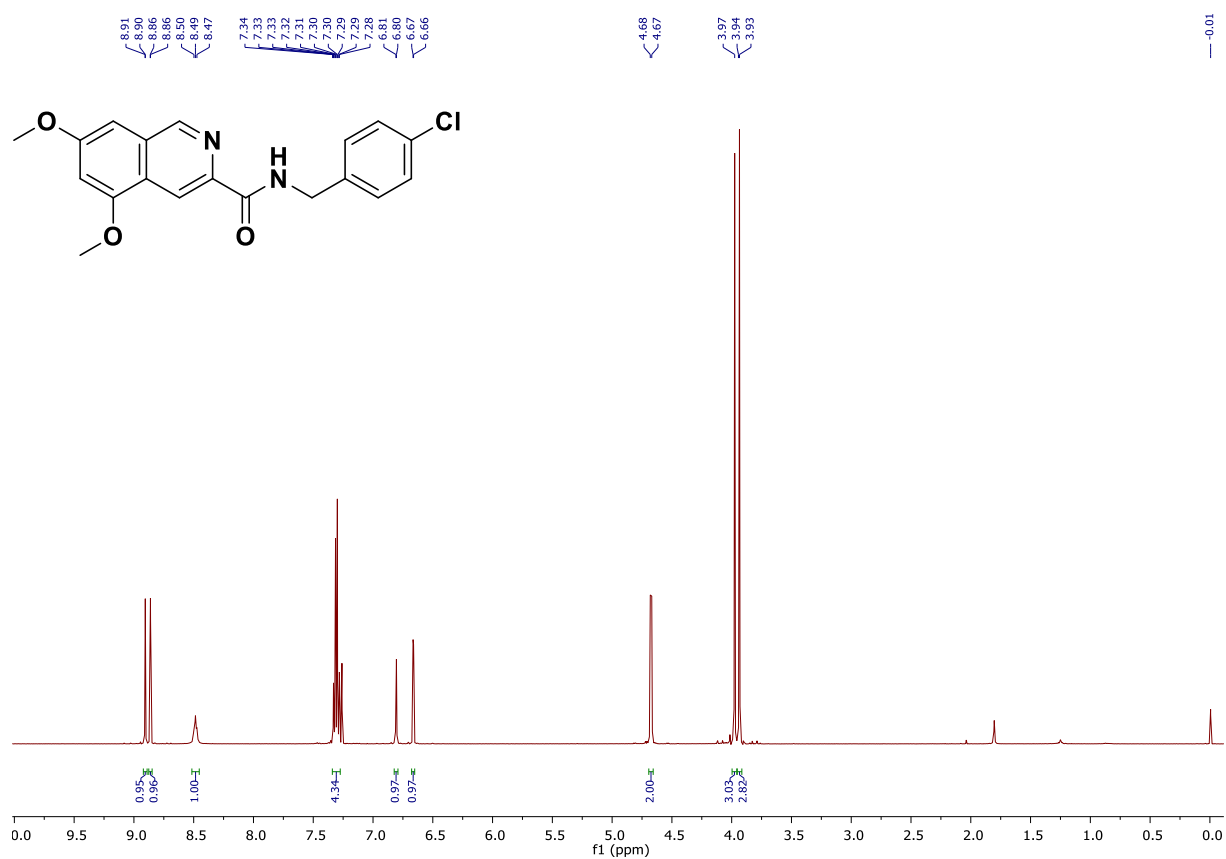
***N*-((1*s*,3*s*)-adamantan-1-yl)-5,6,7-trimethoxyisoquinoline-3-carboxamide (F13)**



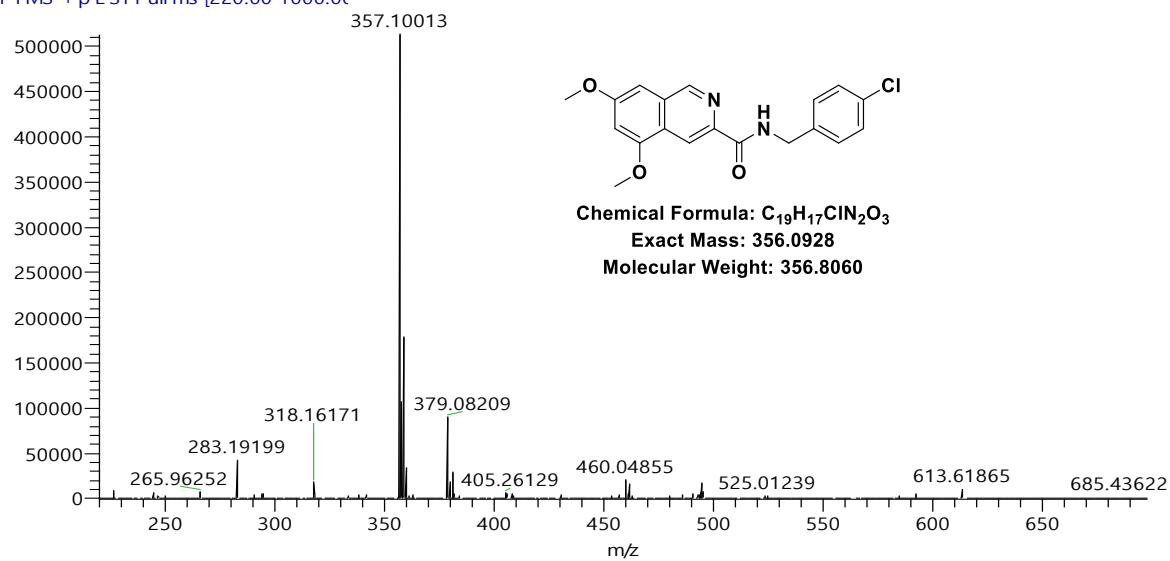
17mdv207-yz450c #283 RT: 5.69 AV: 1 NL: 1.25E7
T: FTMS + p ESI Full ms [200.00-800.00]



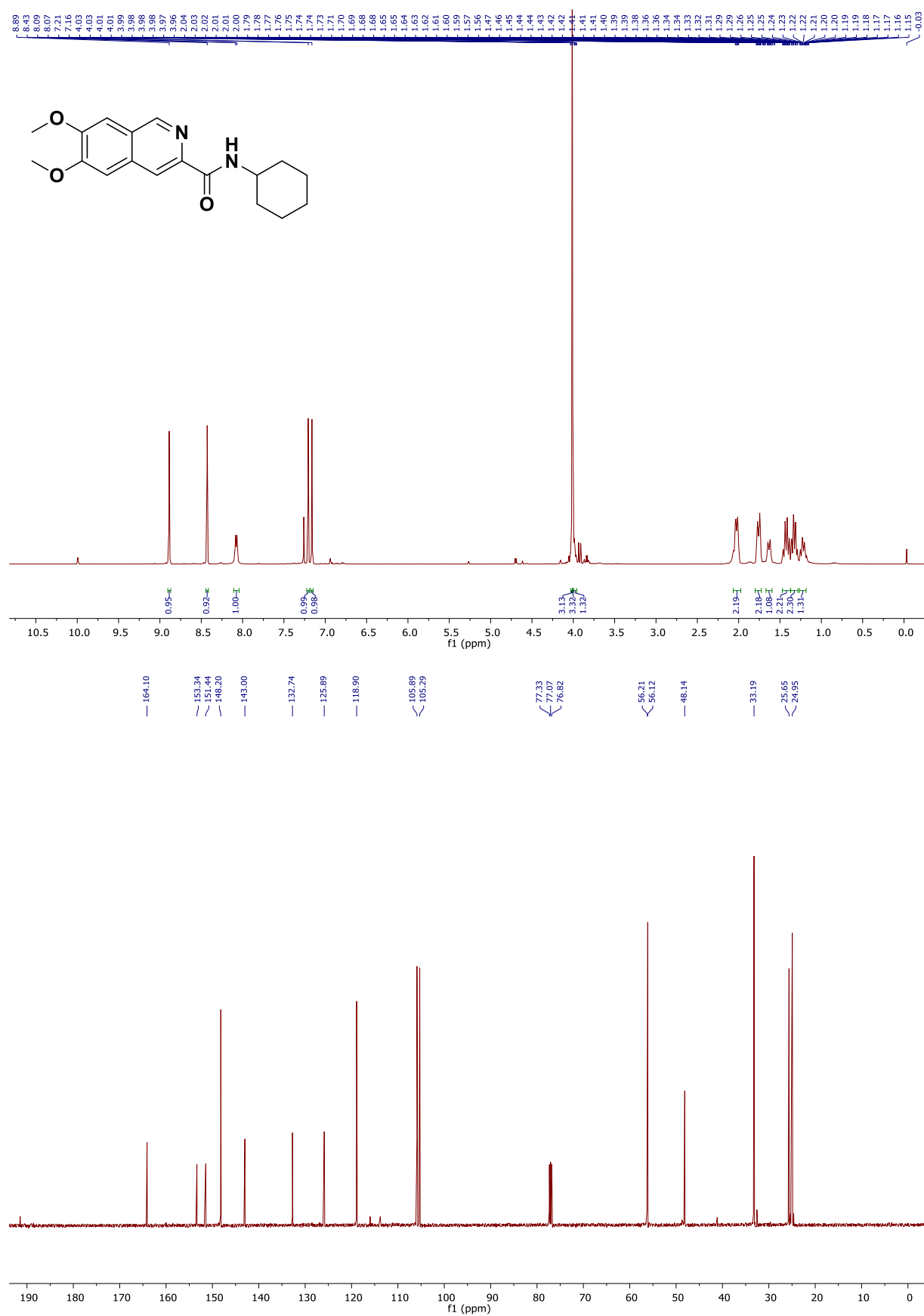
***N*-(4-chlorobenzyl)-5,7-dimethoxyisoquinoline-3-carboxamide (F15)**



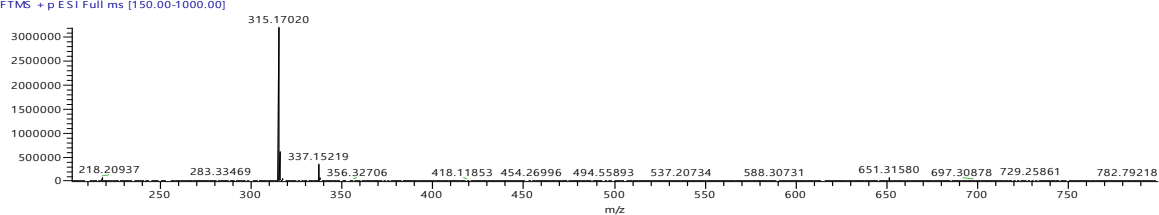
18mdv071-YZ482C #11 RT: 0.19645 / :5
T: FTMS +p ESI Full ms [220.00-1000.00]



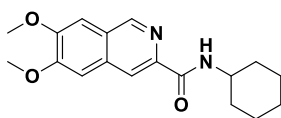
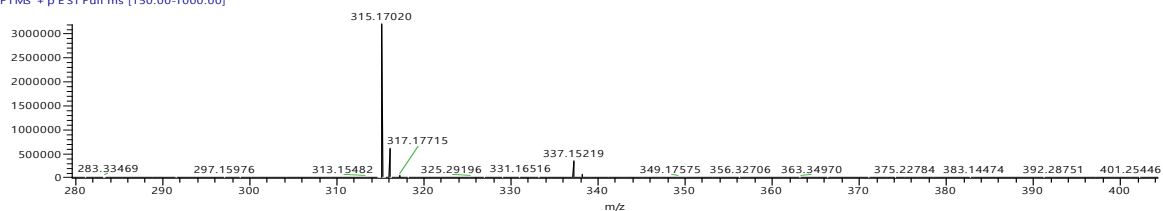
***N*-cyclohexyl-6,7-dimethoxyisoquinoline-3-carboxamide (H13)**



17mdv196-YZ395C #29 RT: 0.50034 AV: 1 NL: 3.20E6
T: FTMS + p ESI Full ms [150.00-1000.00]



17mdv196-YZ395C #29 RT: 0.50034 AV: 1 NL: 3.20E6
T: FTMS + p ESI Full ms [150.00-1000.00]

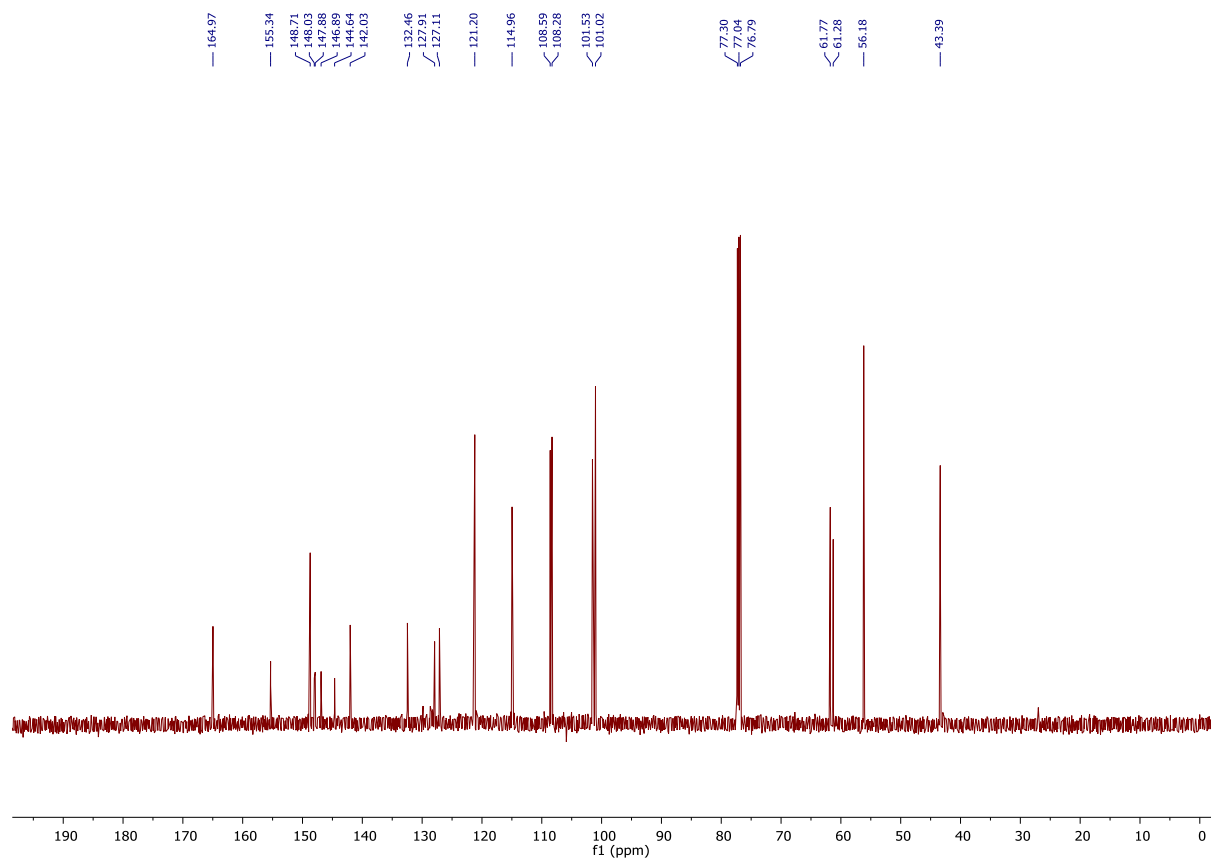
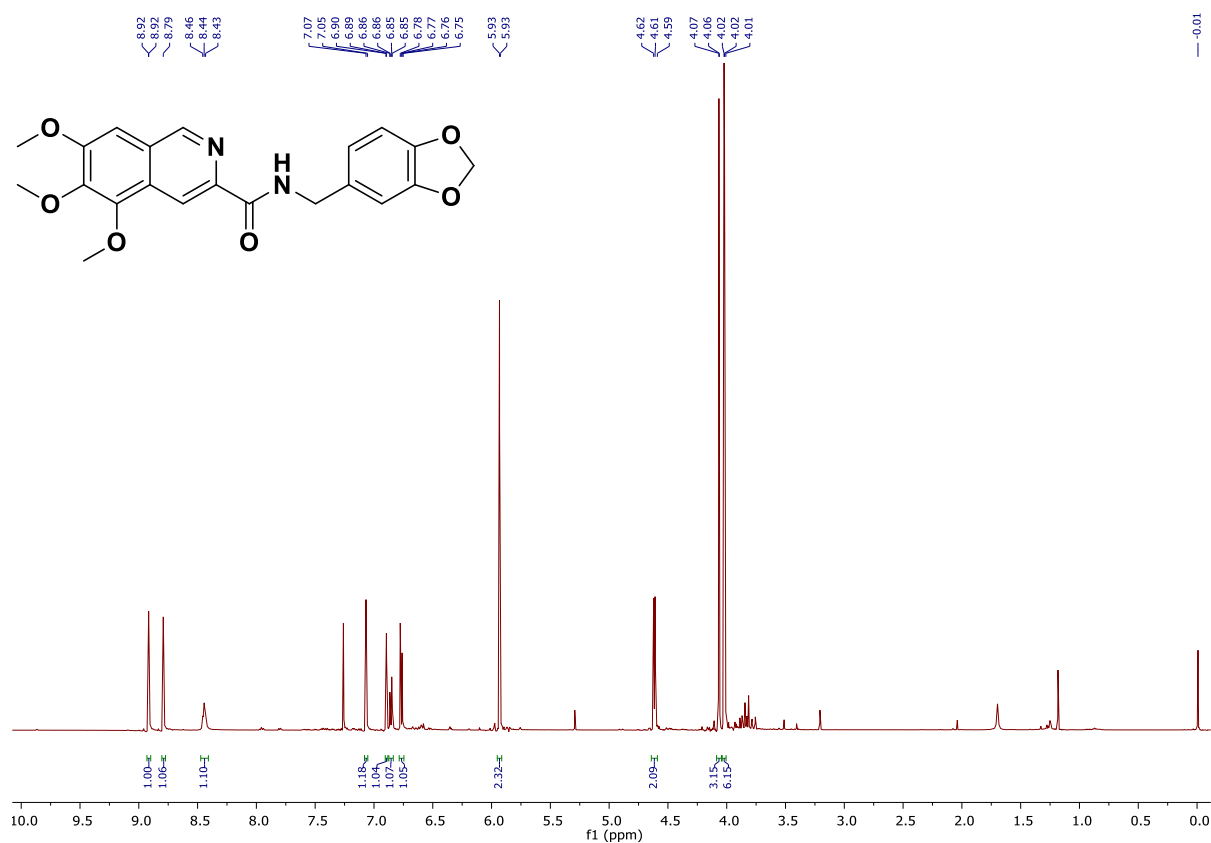


Chemical Formula: $C_{18}H_{22}N_2O_3$

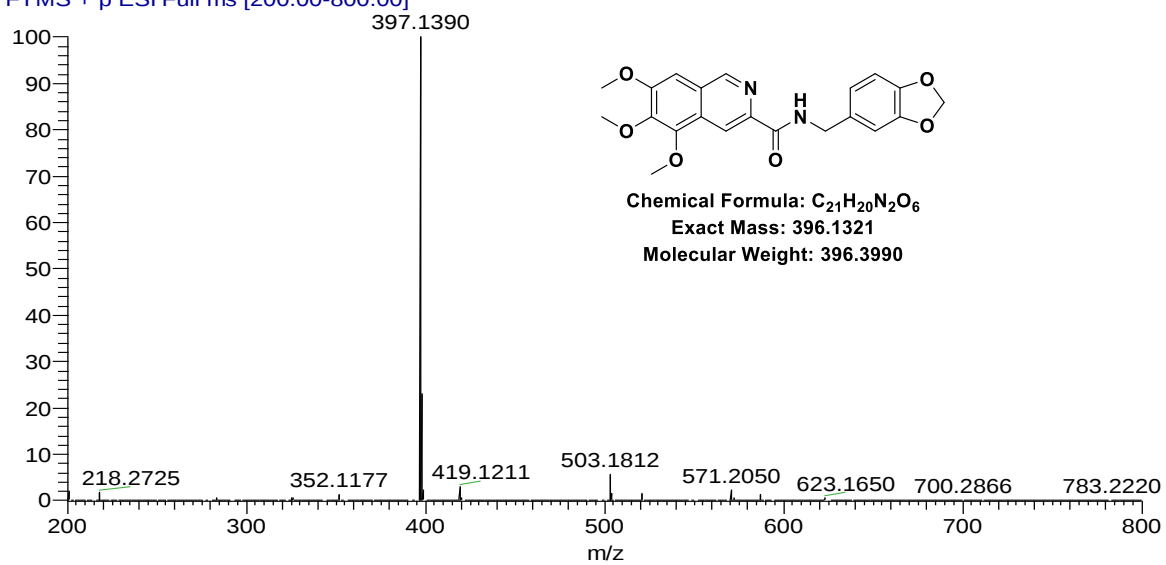
Exact Mass: 314.1630

Molecular Weight: 314.3850

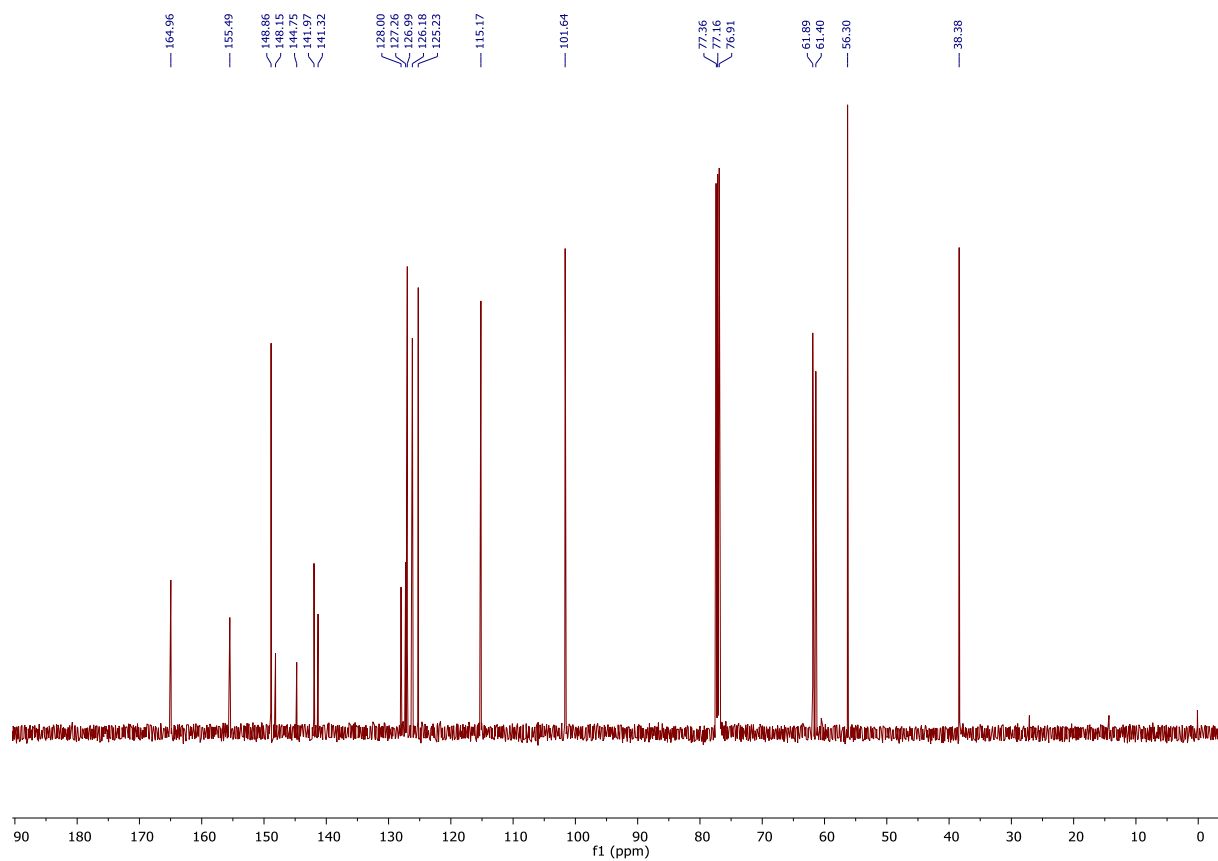
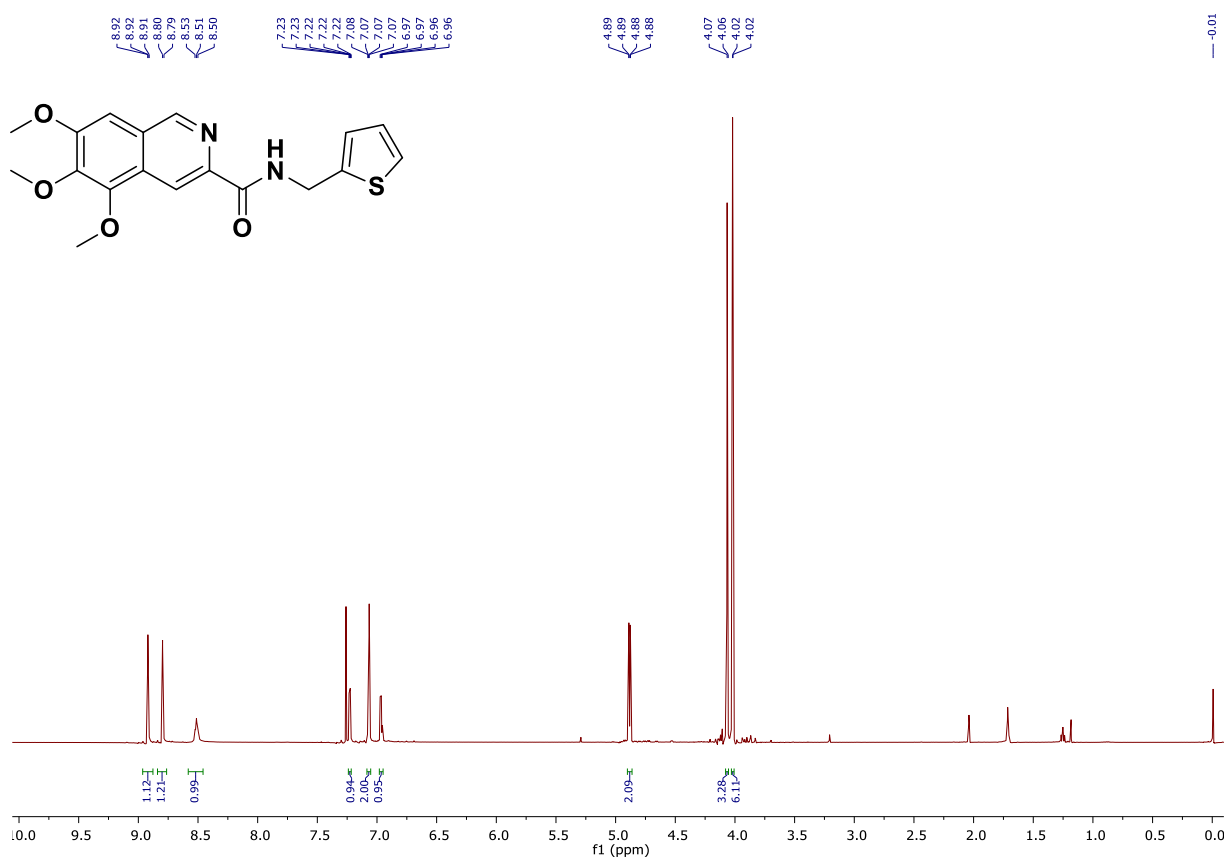
***N*-(benzo[d][1,3]dioxol-5-ylmethyl)-5,6,7-trimethoxyisoquinoline-3-carboxamide (I6)**



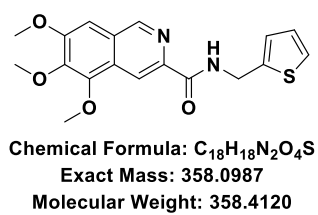
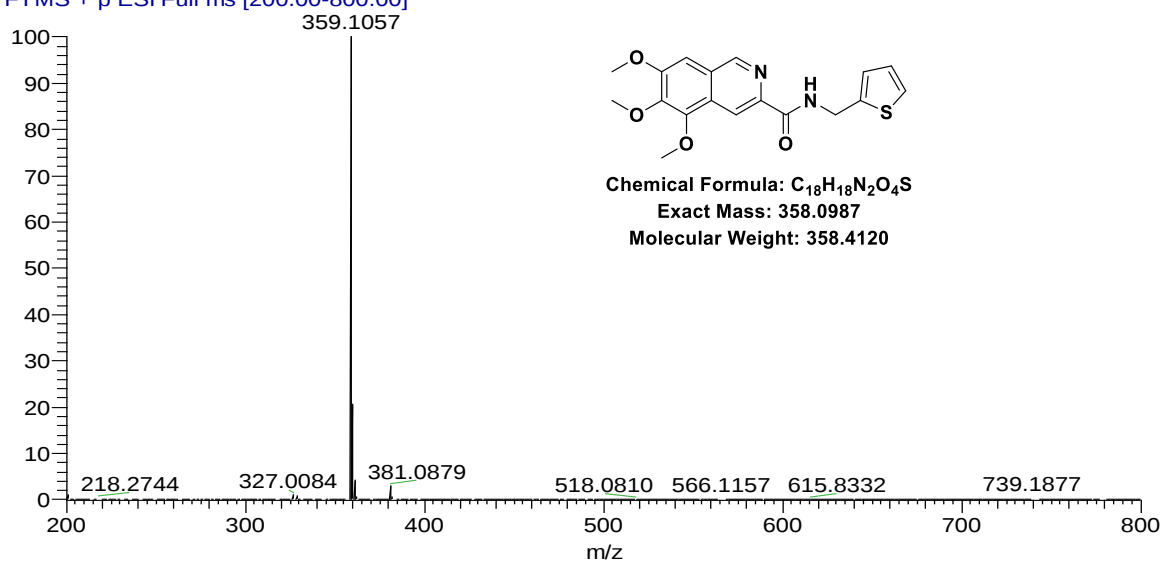
17mdv207-yz447 #226 RT: 4.62 AV: 1 NL: 6.02E6
T: FTMS + p ESI Full ms [200.00-800.00]



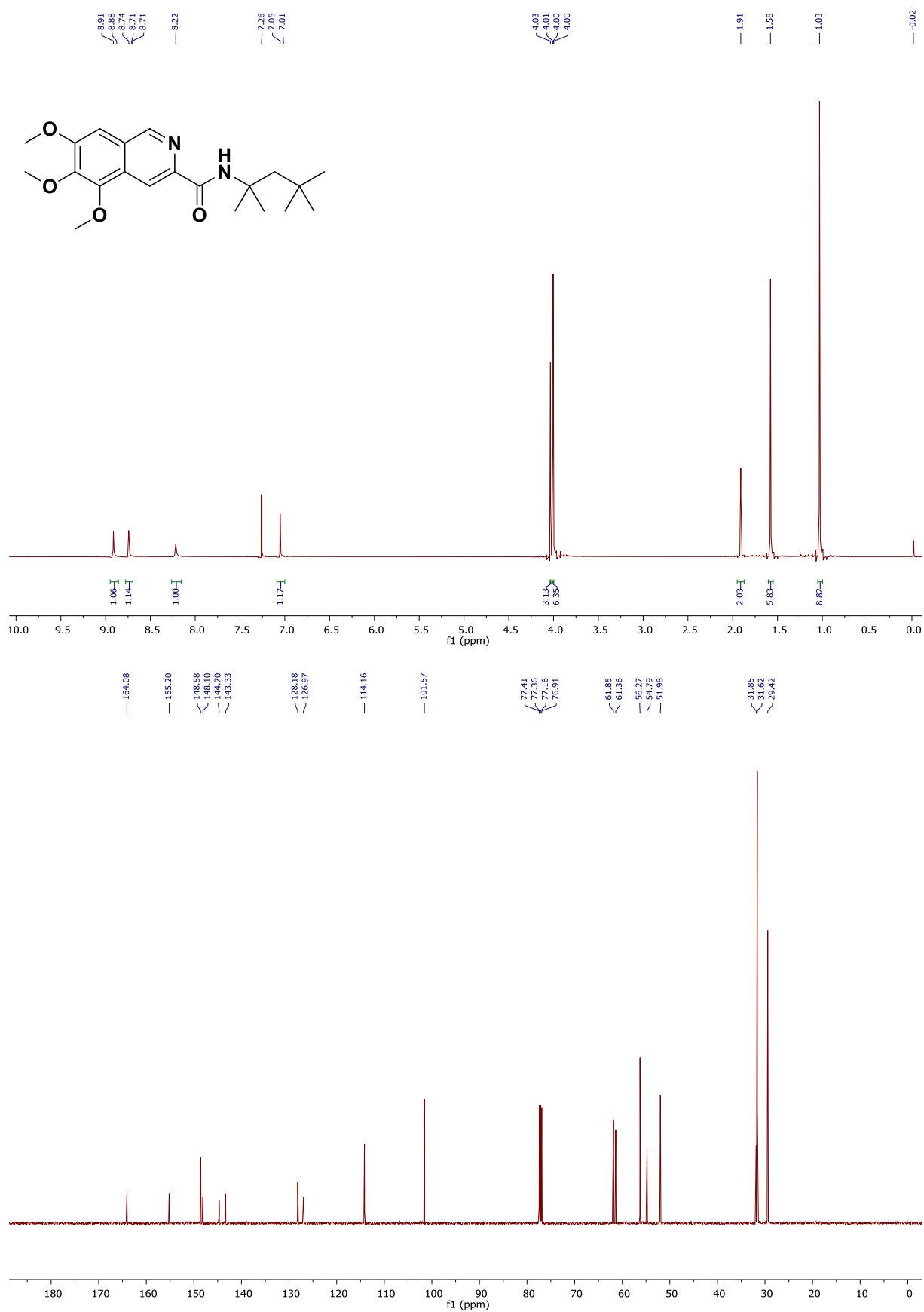
5,6,7-Trimethoxy-*N*-(thiophen-2-ylmethyl)isoquinoline-3-carboxamide (I17)



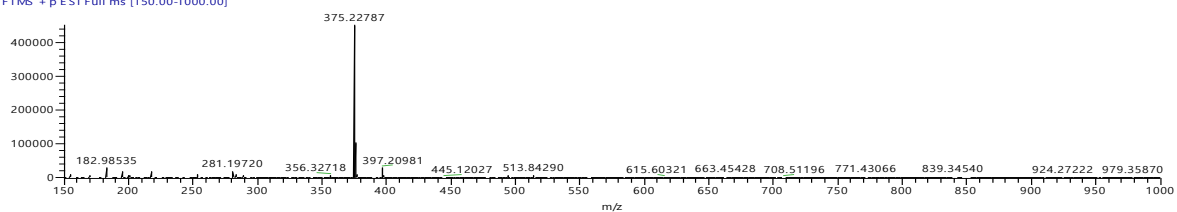
17mdv207-yz443c #217 RT: 4.55 AV: 1 NL: 8.98E6
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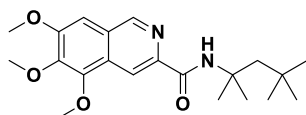
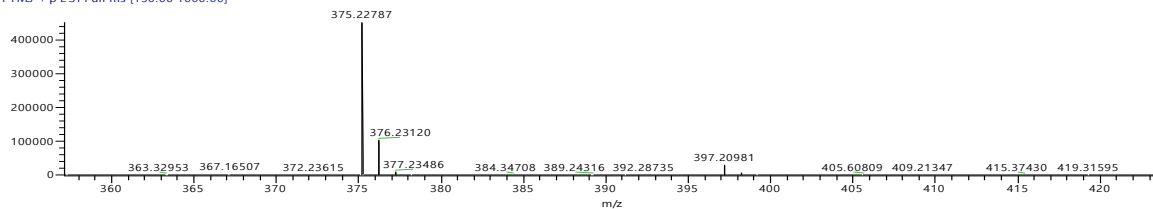
5,6,7-Trimethoxy-*N*-(2,4,4-trimethylpentan-2-yl)isoquinoline-3-carboxamide (J3)



17mdv196-YZ441C #32 RT: 0.59382 AV: 1 NL: 4.53E5
T: FTMS + p ESI Full ms [150.00-1000.00]



17mdv196-YZ441C #32 RT: 0.59382 AV: 1 NL: 4.53E5
T: FTMS + p ESI Full ms [150.00-1000.00]

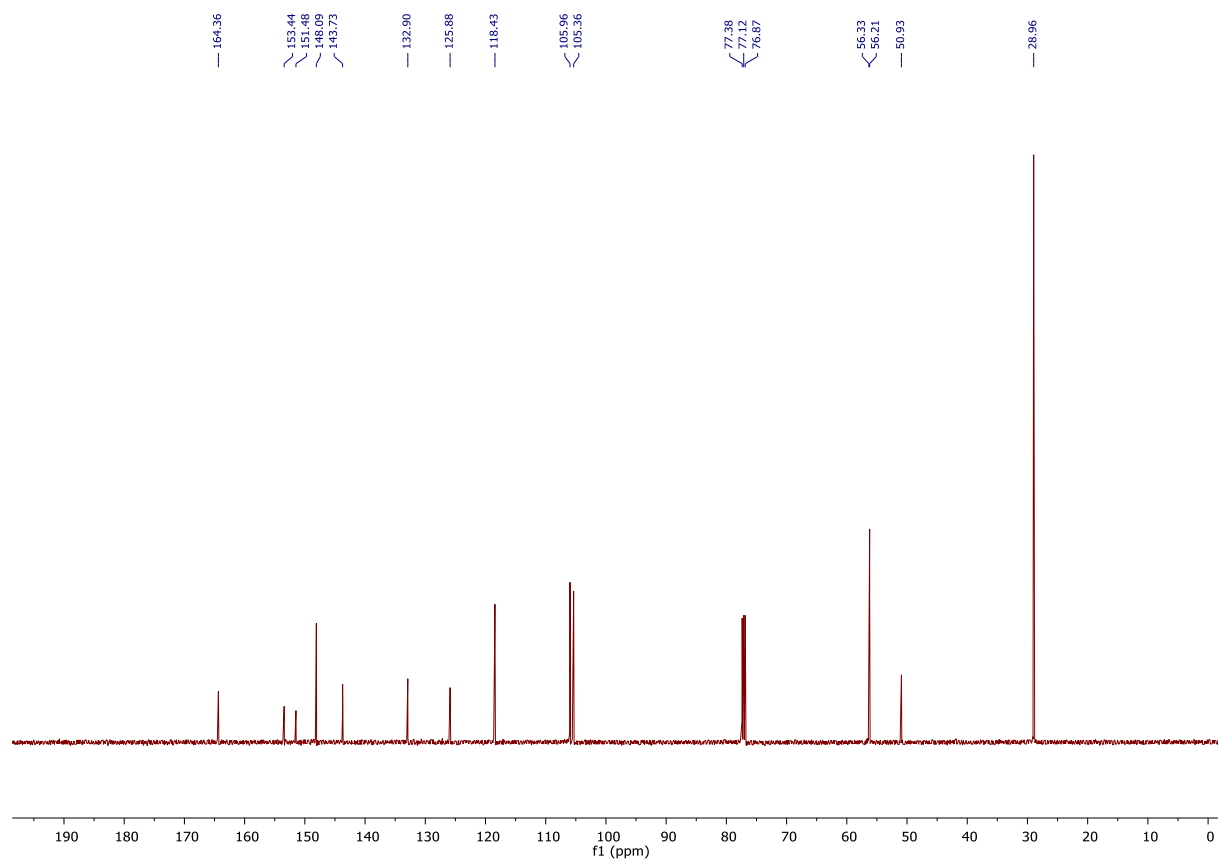
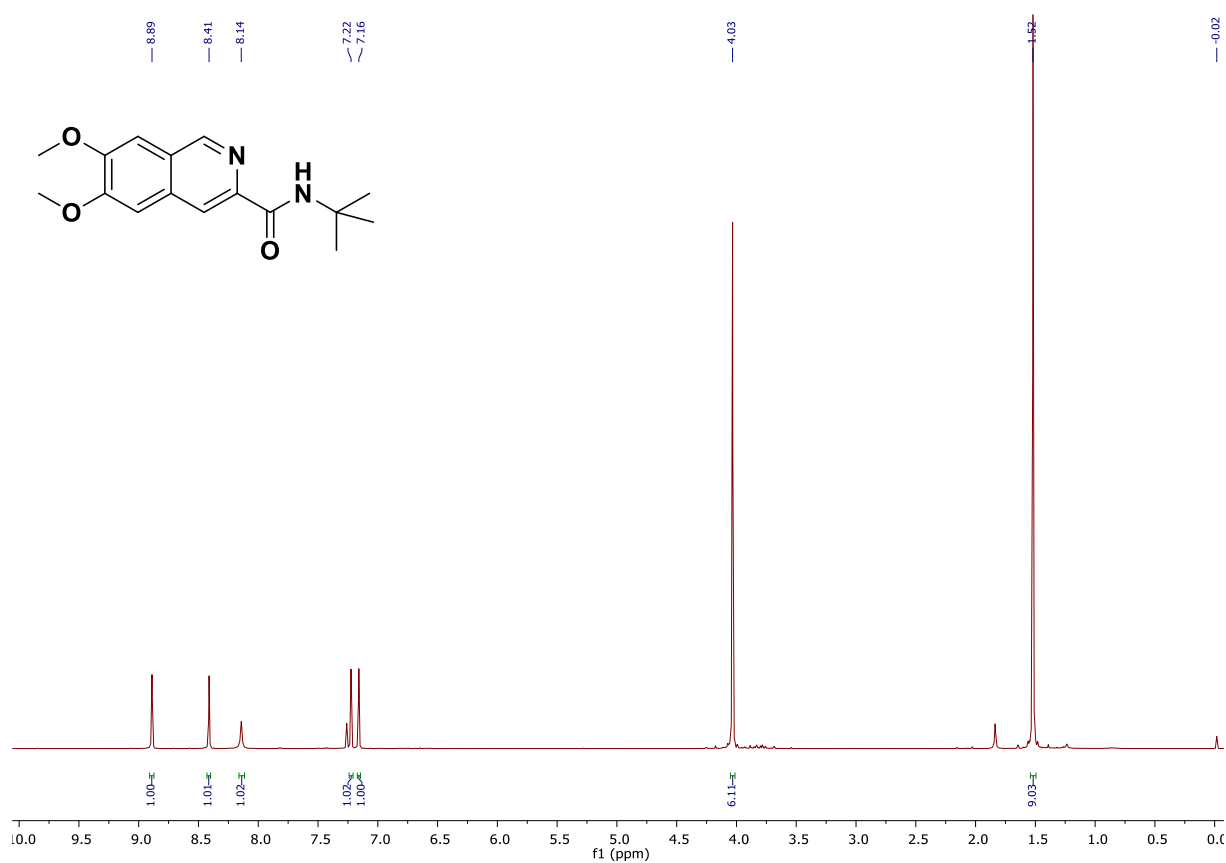


Chemical Formula: $C_{21}H_{30}N_2O_4$

Exact Mass: 374.2206

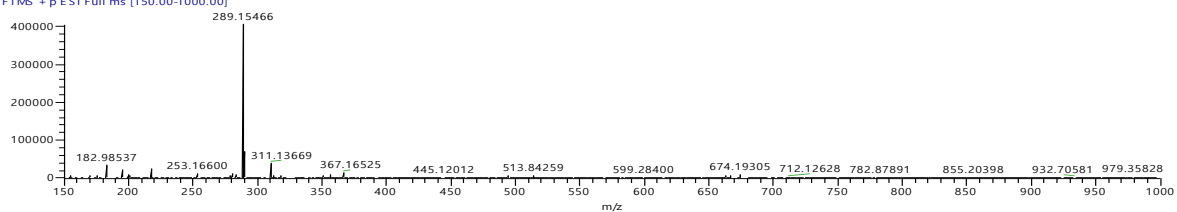
Molecular Weight: 374.4810

***N*-(*tert*-butyl)-6,7-dimethoxyisoquinoline-3-carboxamide (J9)**



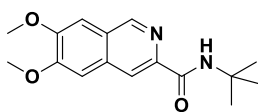
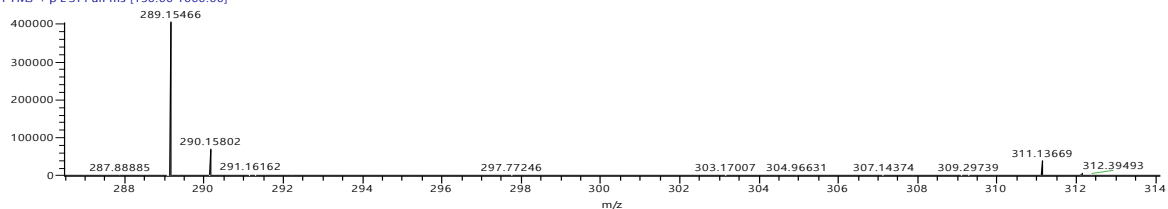
17mdv196-YZ420C #30 RT: 0.53962 AV: 1 NL: 4.04E5

T: FTMS + p ESI Full ms [150.00-1000.00]

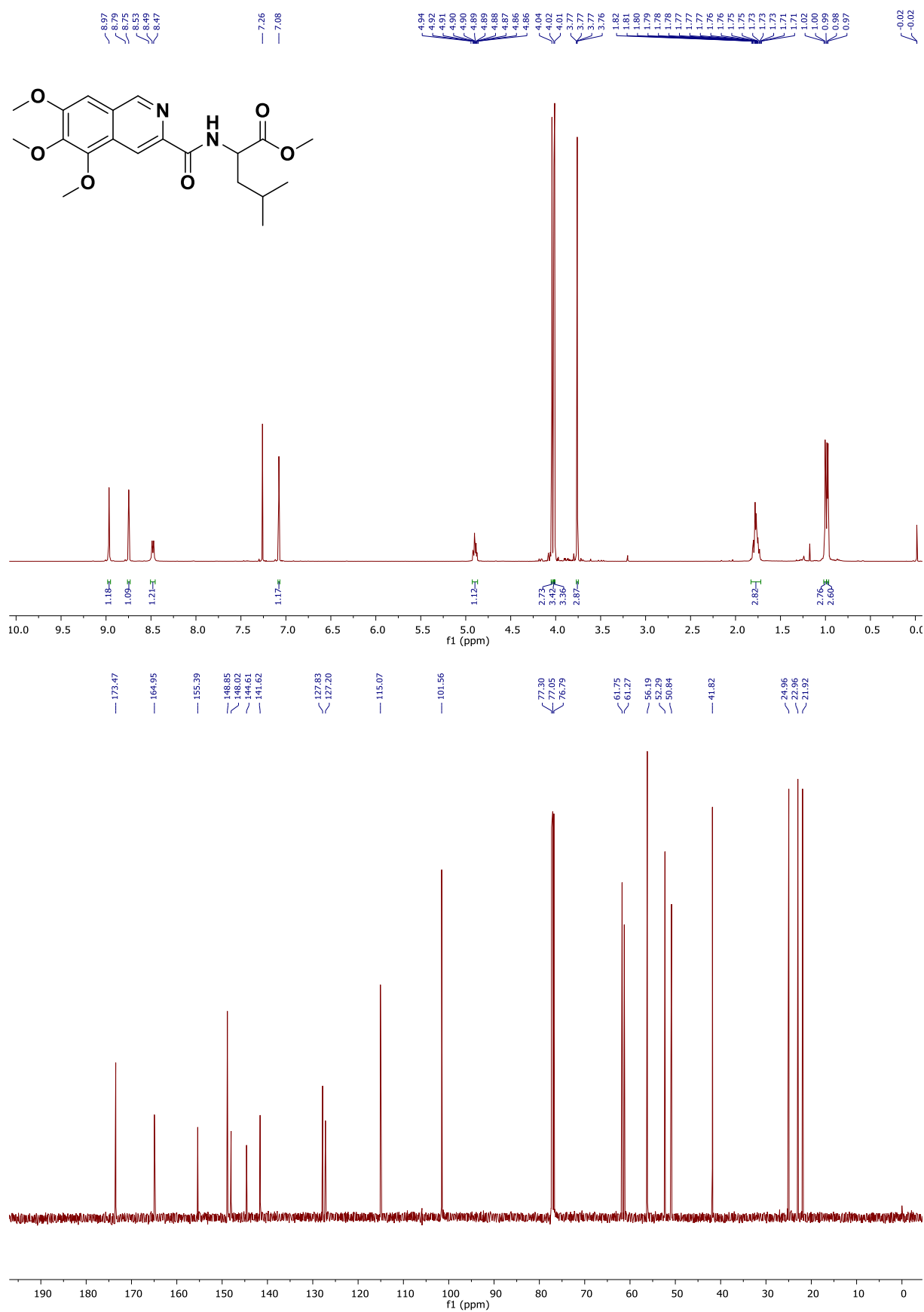


17mdv196-YZ420C #30 RT: 0.53962 AV: 1 NL: 4.04E5

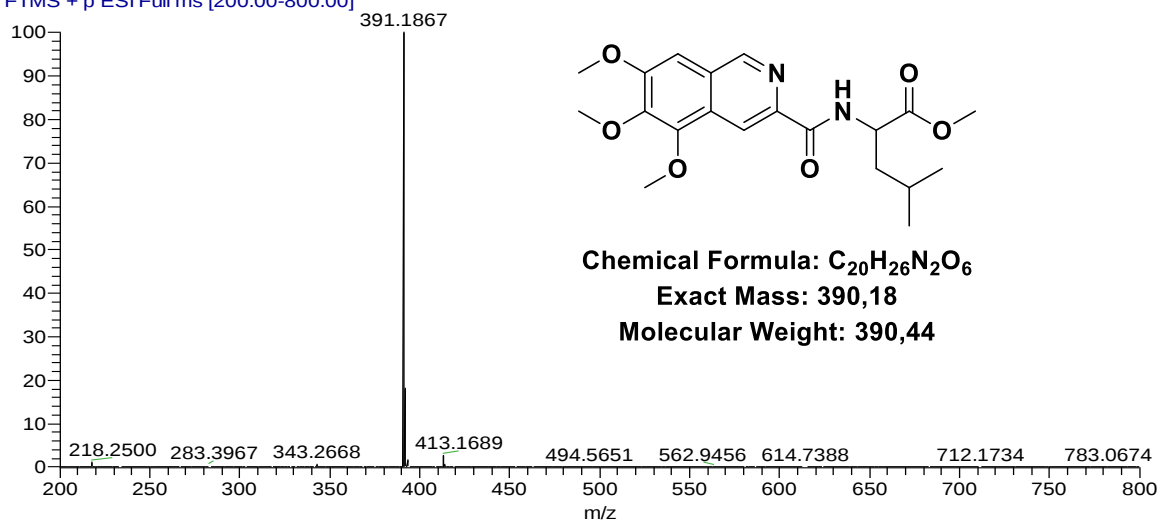
T: FTMS + p ESI Full ms [150.00-1000.00]

**Chemical Formula:** C₁₆H₂₀N₂O₃**Exact Mass:** 288.1474**Molecular Weight:** 288.3470

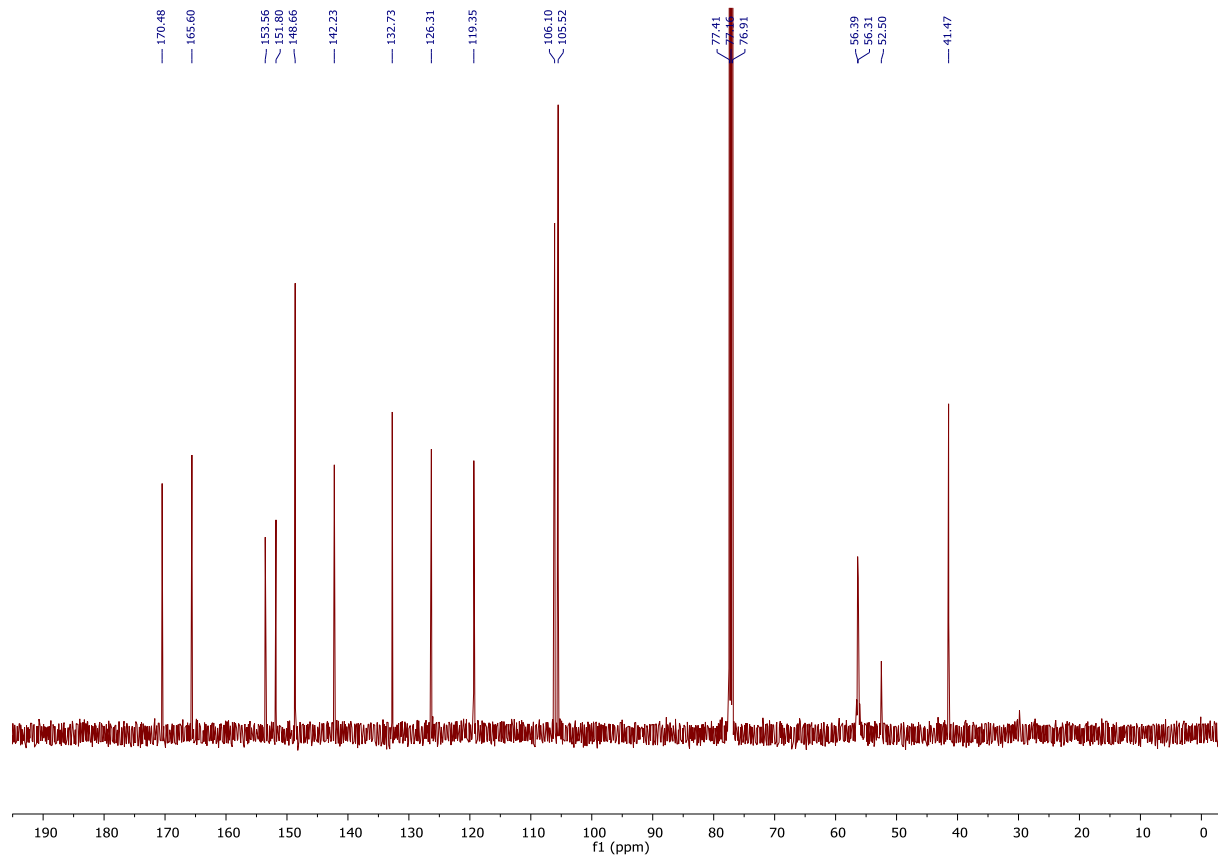
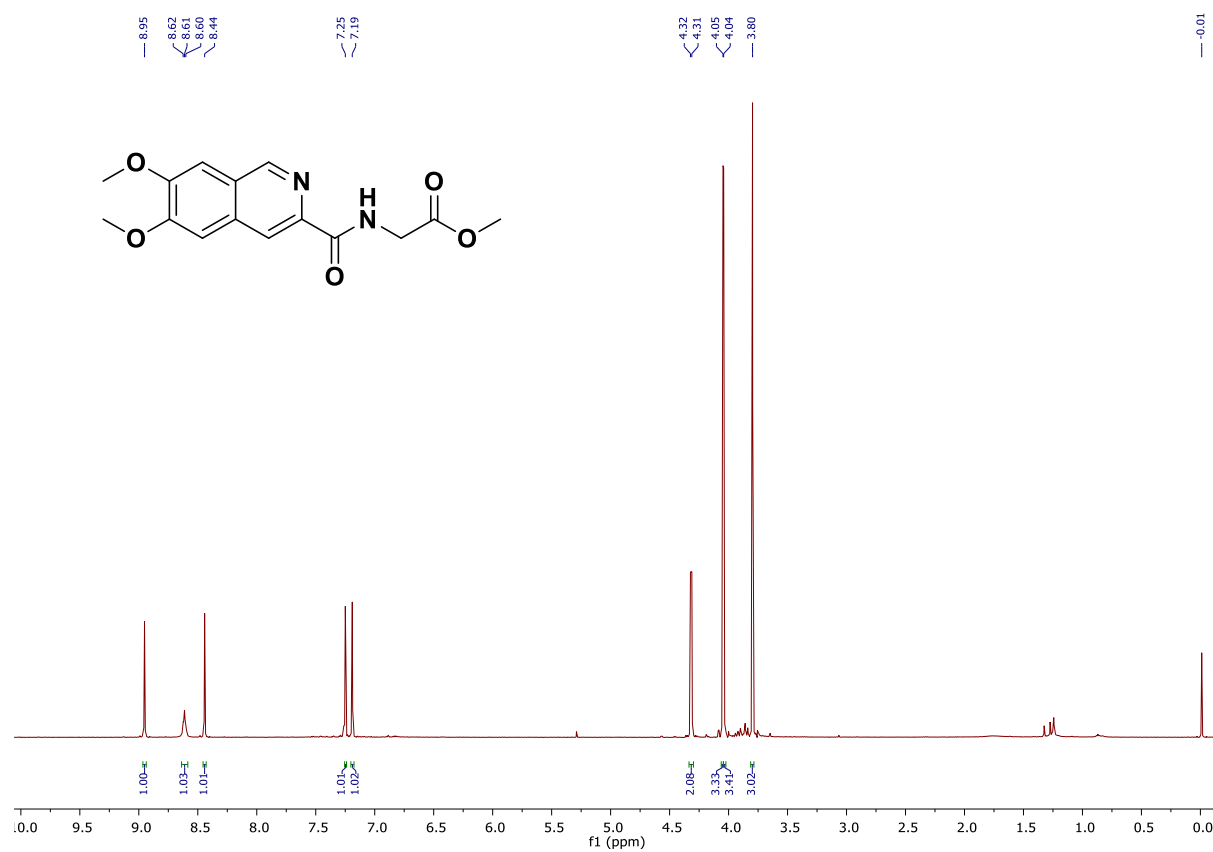
Methyl (5,6,7-trimethoxyisoquinoline-3-carbonyl)leucinate (J18)



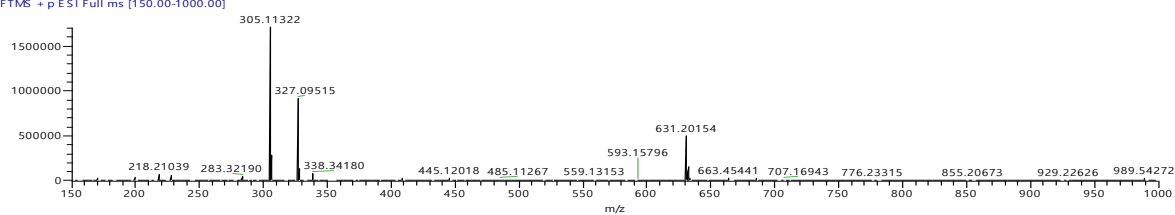
17mdv207-yz449c #275 RT: 4.89 AV: 1 NL: 3
T: FTMS + p ESI Full ms [200.00-800.00]



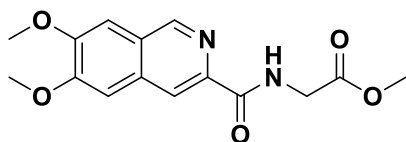
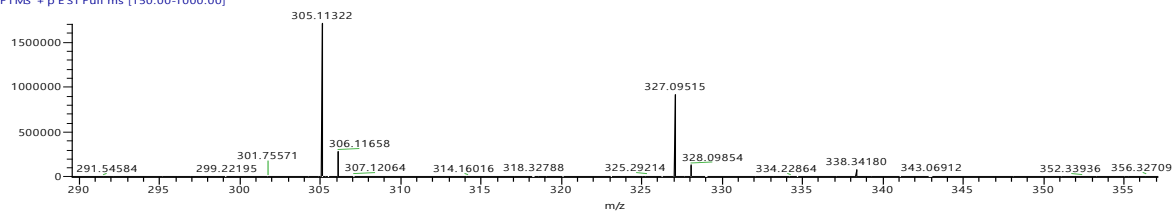
Methyl (6,7-dimethoxyisoquinoline-3-carbonyl) glycinate (K5)



17mdv196-YZ424C #24 RT: 0.40534 AV: 1 NL: 1.70E6
T: FTMS + p ESI Full ms [150.00-1000.00]



17mdv196-YZ424C #24 RT: 0.40534 AV: 1 NL: 1.70E6
T: FTMS + p ESI Full ms [150.00-1000.00]

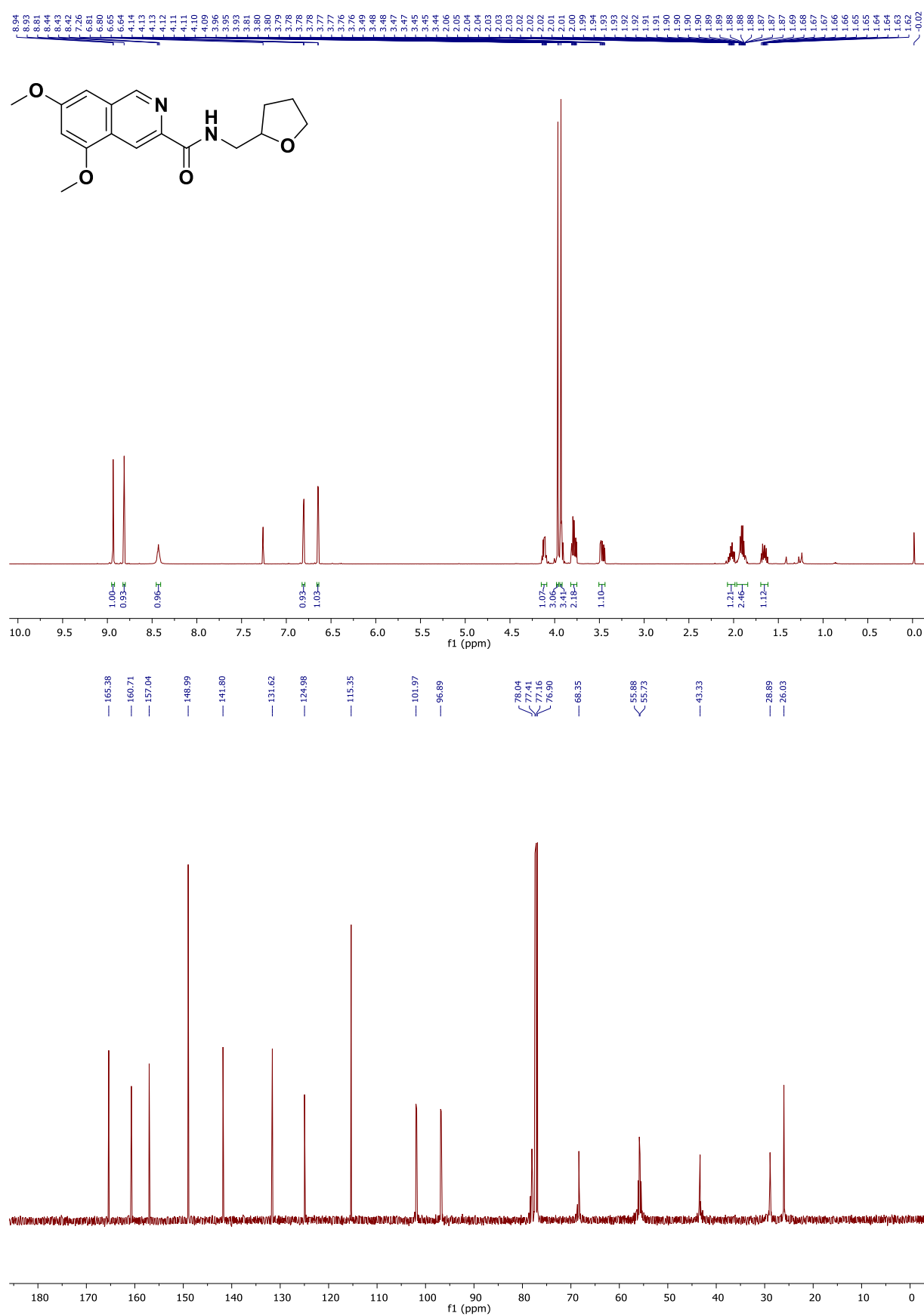


Chemical Formula: C₁₅H₁₆N₂O₅

Exact Mass: 304,11

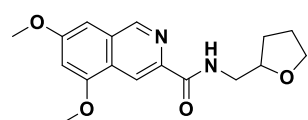
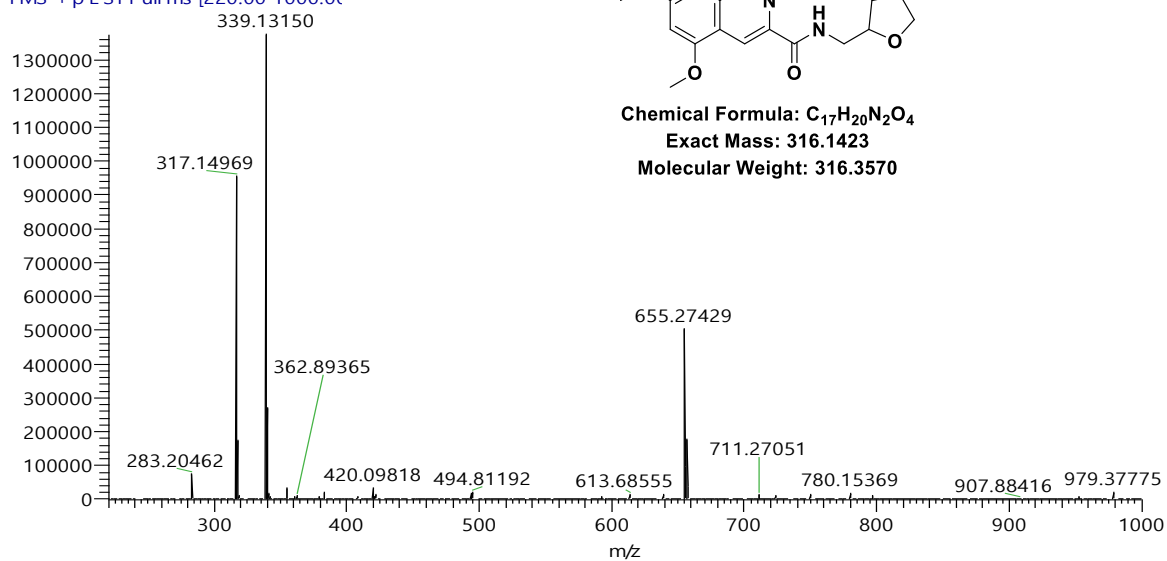
Molecular Weight: 304,30

5,7-Dimethoxy-*N*-((tetrahydrofuran-2-yl)methyl)isoquinoline-3-carboxamide (K8)



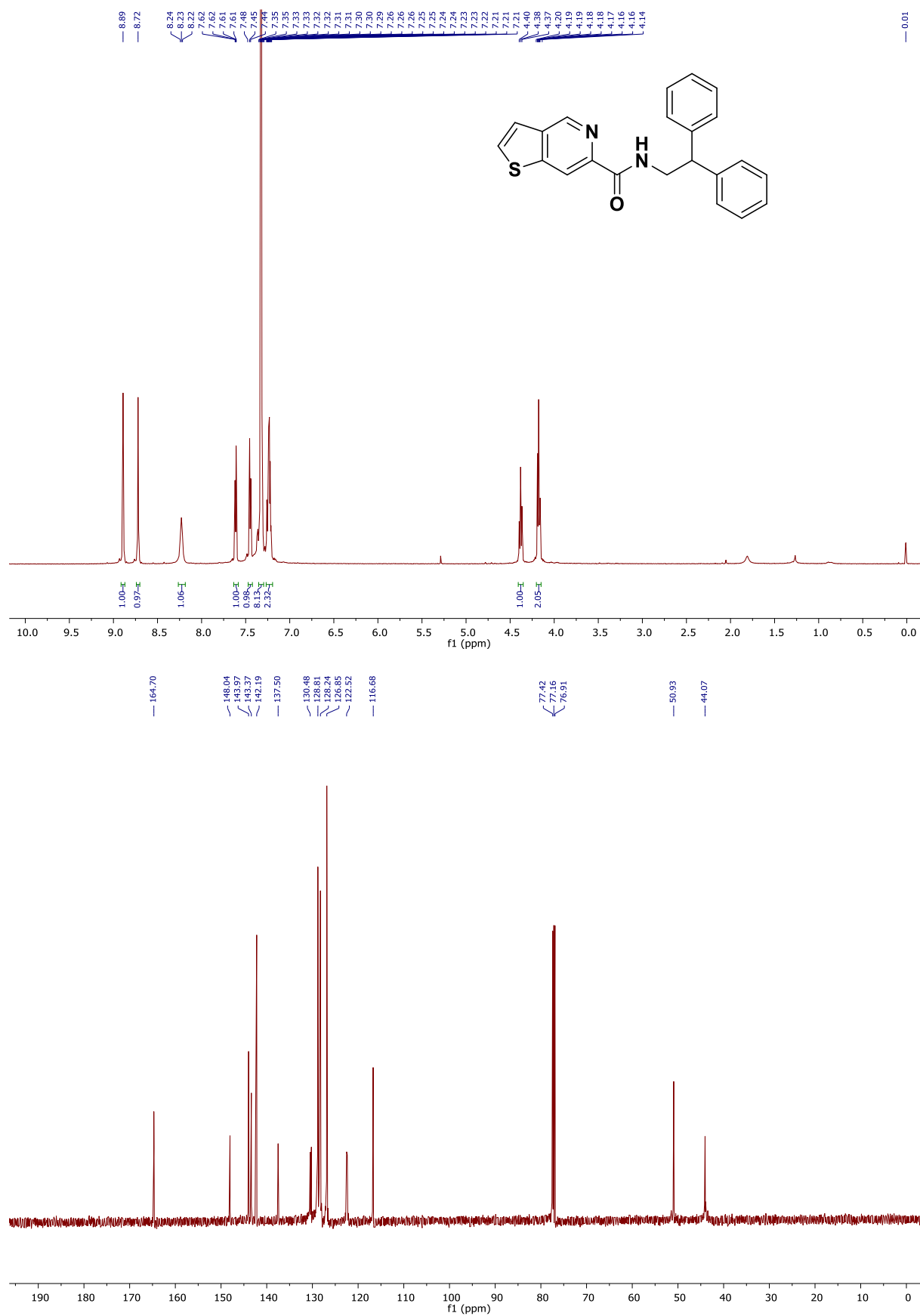
18mdv071-YZ484C #12 RT: 0.20567 /
T: FTMS +p ESI Full ms [220.00-1000.00]

:6



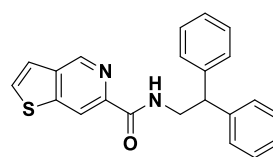
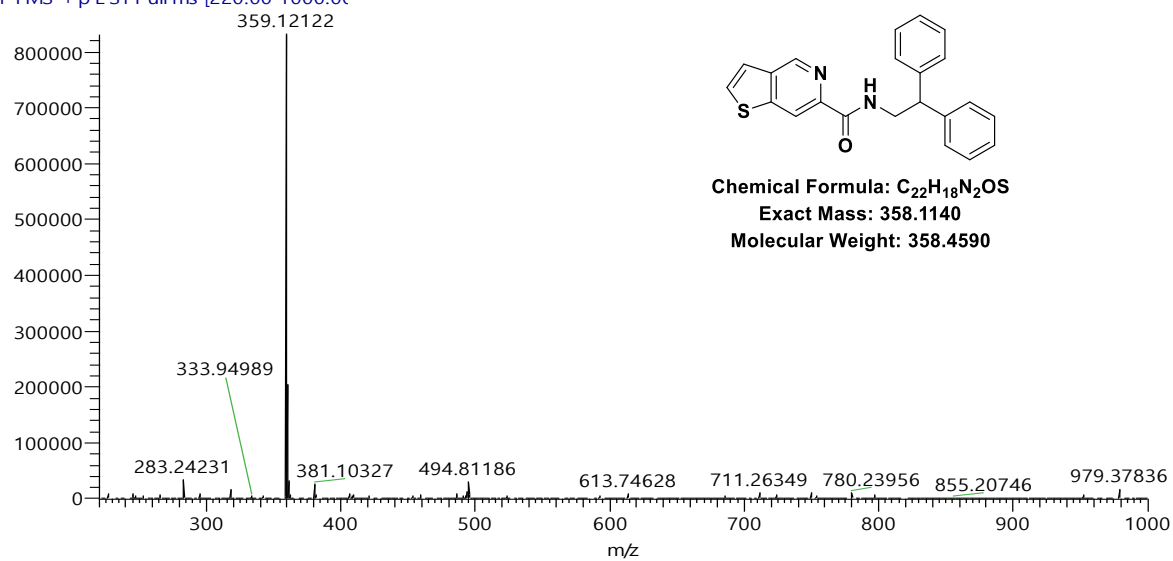
Chemical Formula: C₁₇H₂₀N₂O₄
Exact Mass: 316.1423
Molecular Weight: 316.3570

***N*-(2,2-diphenylethyl)thieno[3,2-*c*]pyridine-6-carboxamide (K21)**



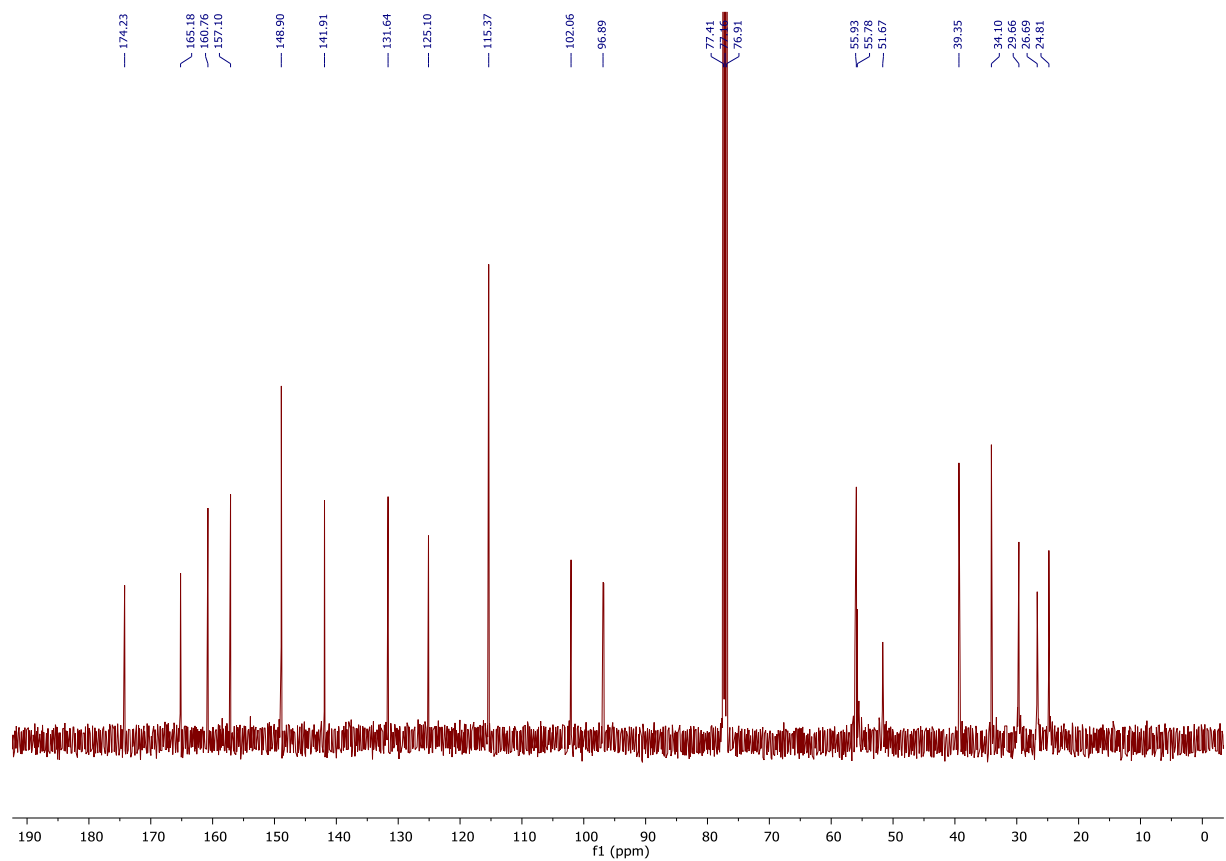
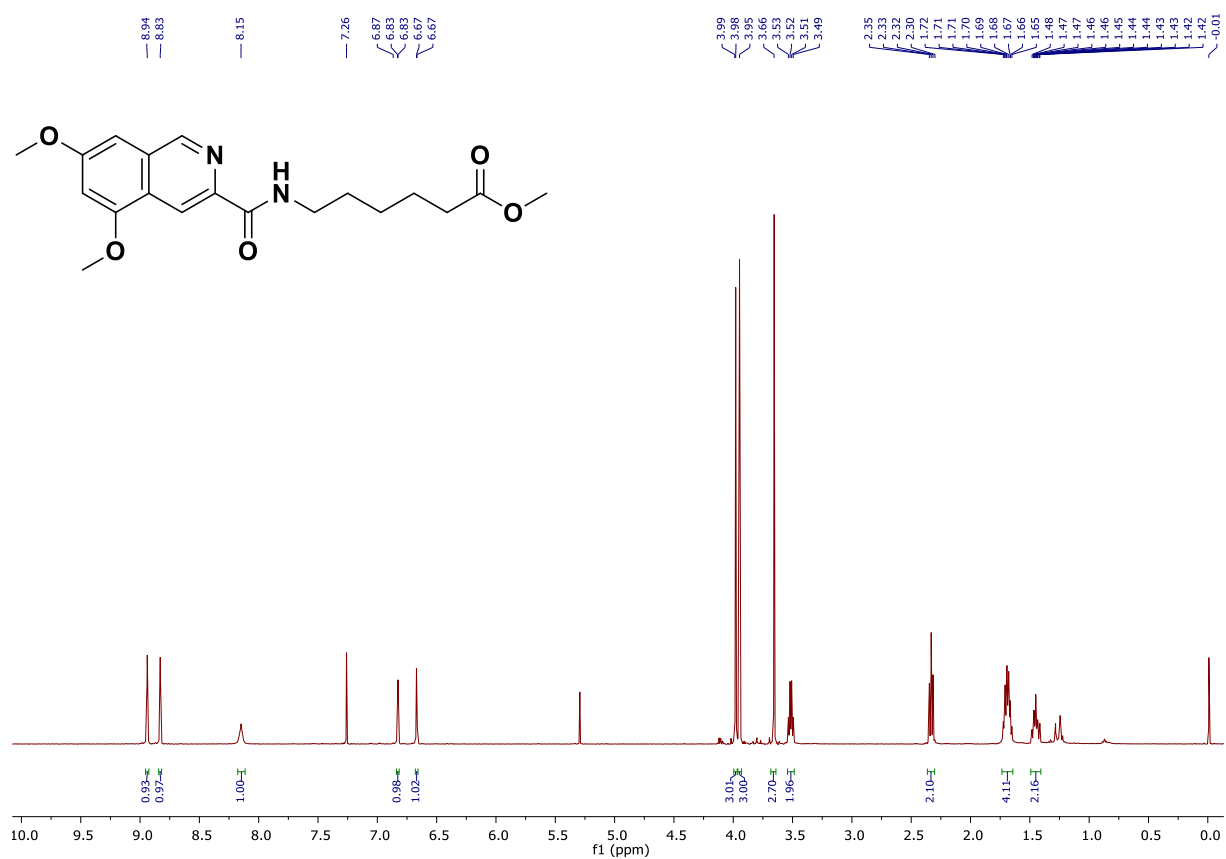
18mdv071-yz480C #10 RT: 0.18045 A
T: FTMS +p ESI Full ms [220.00-1000.00]

5

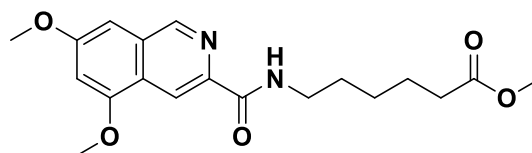
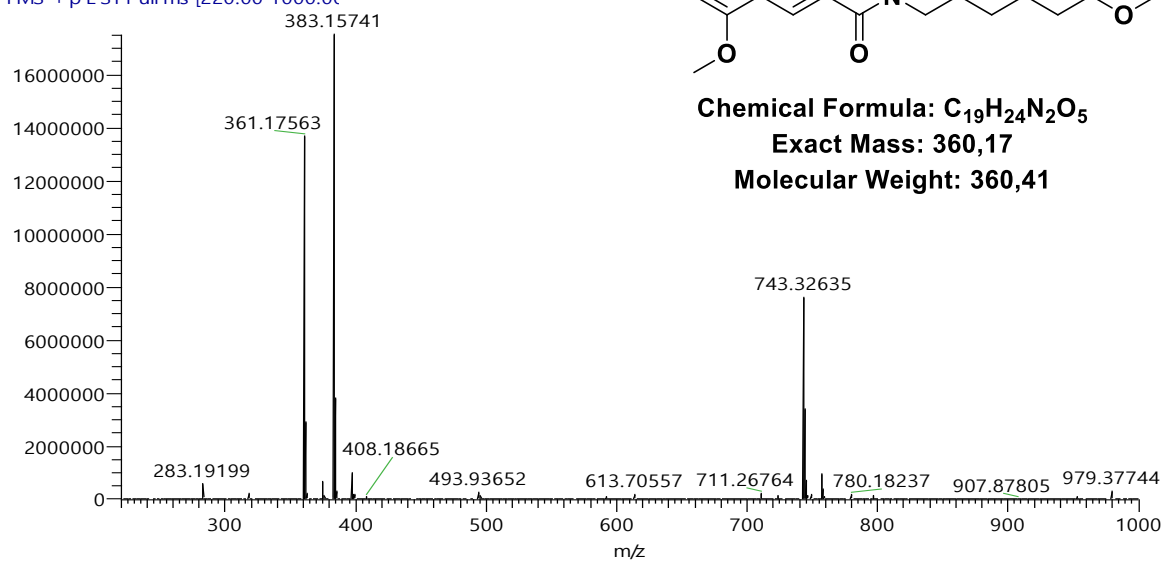


Chemical Formula: $C_{22}H_{18}N_2OS$
Exact Mass: 358.1140
Molecular Weight: 358.4590

Methyl 6-(5,7-dimethoxyisoquinoline-3-carboxamido)hexanoate (L7)



18mdv071-YZ485c #6 RT: 0.10171 AV
T: FTMS +p ESI Full ms [220.00-1000.00]

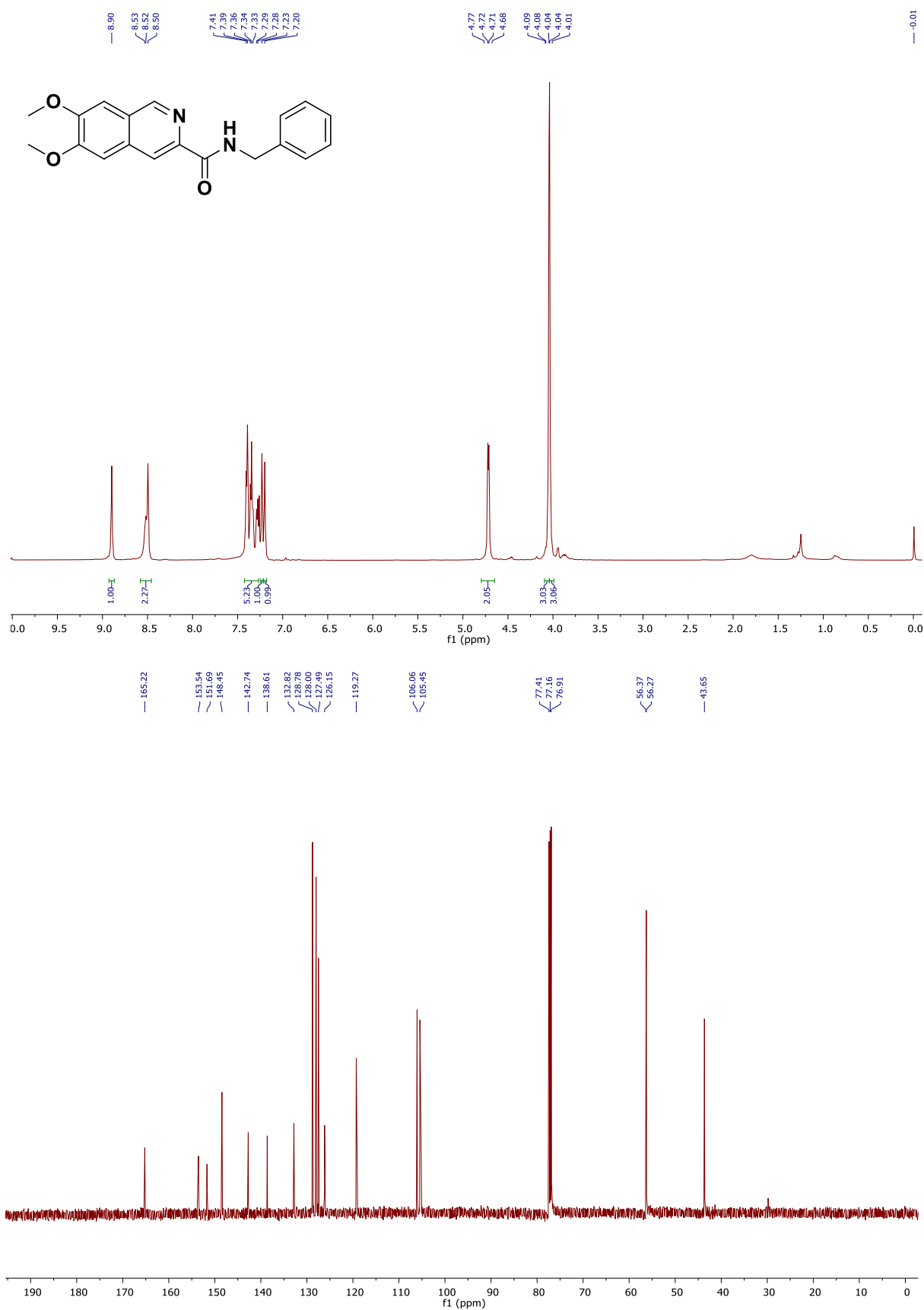


Chemical Formula: $C_{19}H_{24}N_2O_5$

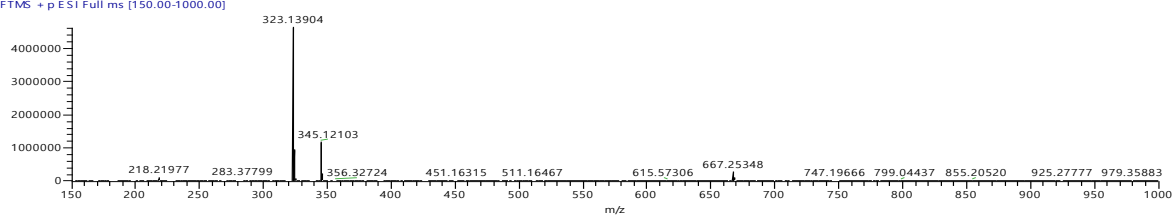
Exact Mass: 360,17

Molecular Weight: 360,41

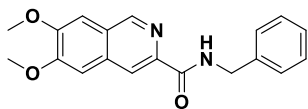
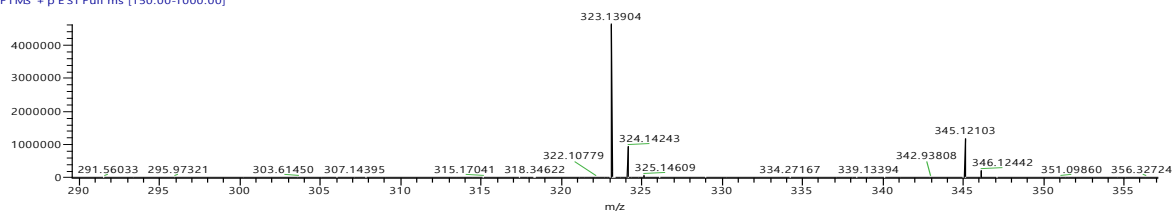
***N*-benzyl-6,7-dimethoxyisoquinoline-3-carboxamide (M17)**



17mdv196-YZ394C #24 RT: 0.40483 AV: 1 NL: 4.63E6
T: FTMS + p ESI Full ms [150.00-1000.00]



17mdv196-YZ394C #24 RT: 0.40483 AV: 1 NL: 4.63E6
T: FTMS + p ESI Full ms [150.00-1000.00]

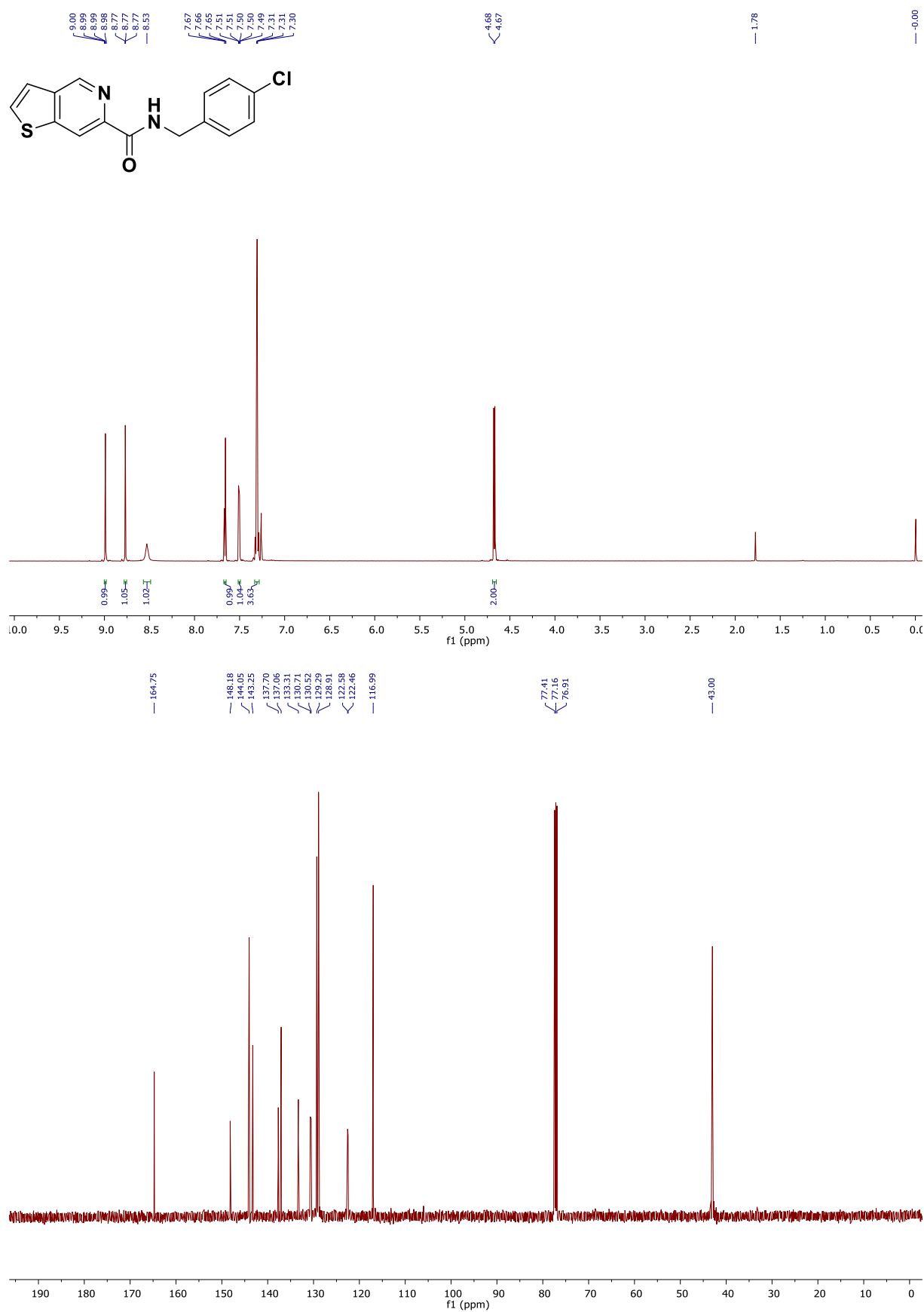


Chemical Formula: $C_{19}H_{18}N_2O_3$

Exact Mass: 322.1317

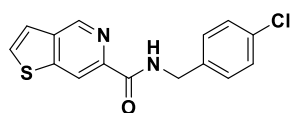
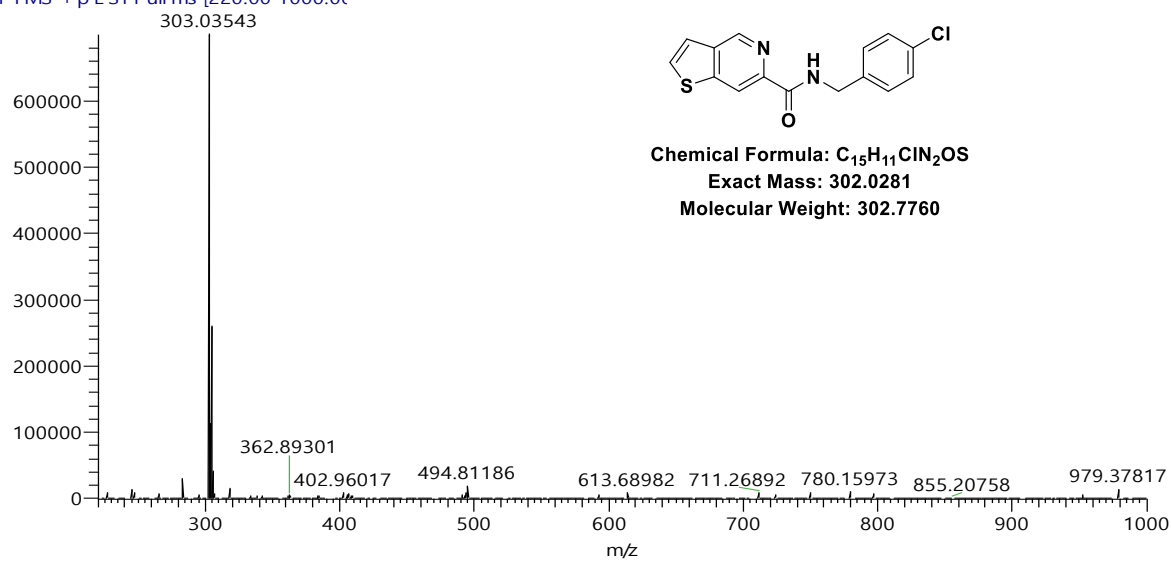
Molecular Weight: 322.3640

***N*-(4-chlorobenzyl)thieno[3,2-*c*]pyridine-6-carboxamide (N15)**



18mdv071-YZ481C #11 RT: 0.19473 /
T: FTMS +p ESI Full ms [220.00-1000.00]

:5

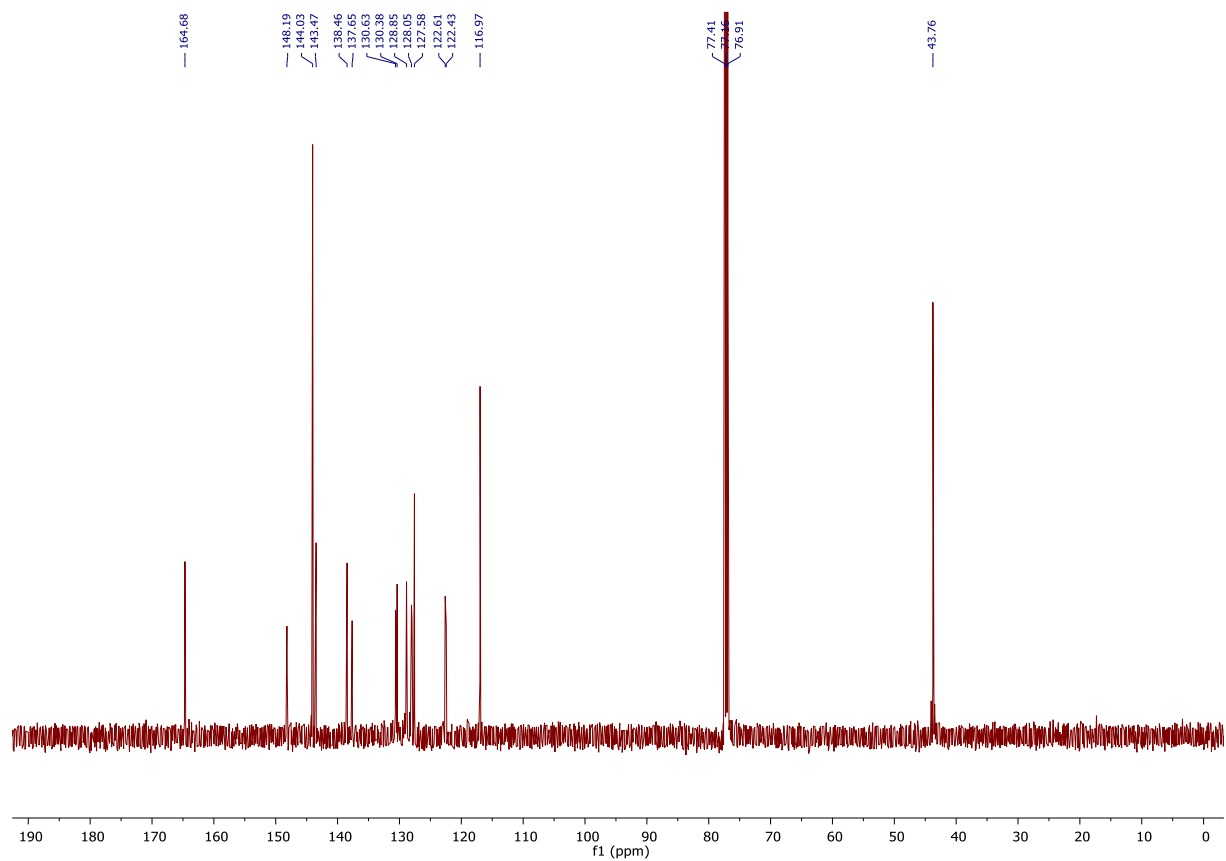
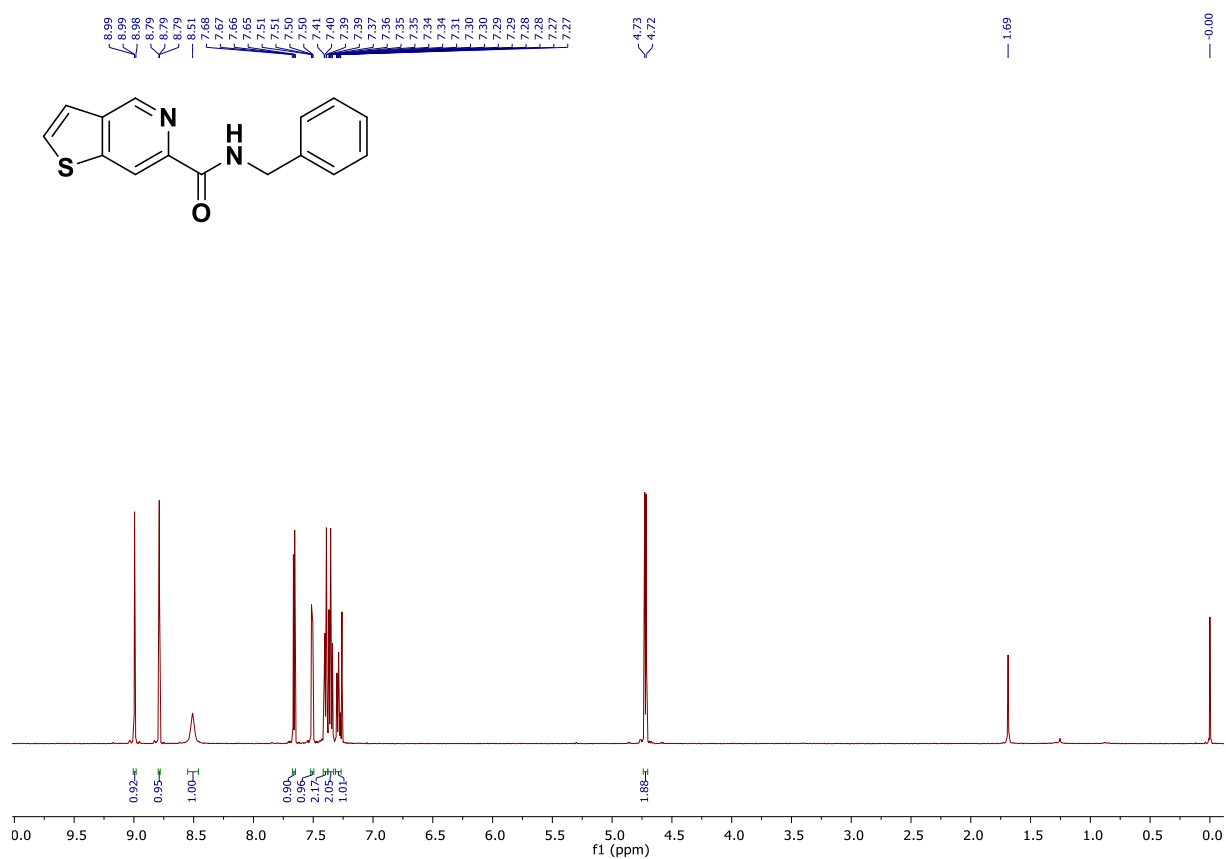


Chemical Formula: C₁₅H₁₁ClN₂OS

Exact Mass: 302.0281

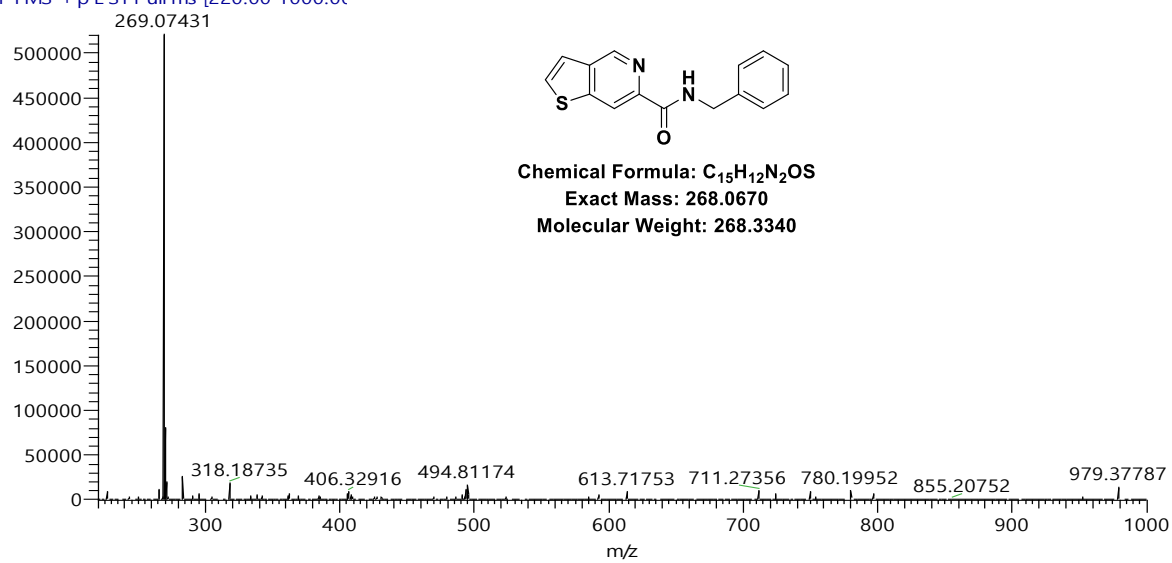
Molecular Weight: 302.7760

***N*-benzylthieno[3,2-*c*]pyridine-6-carboxamide (O11)**

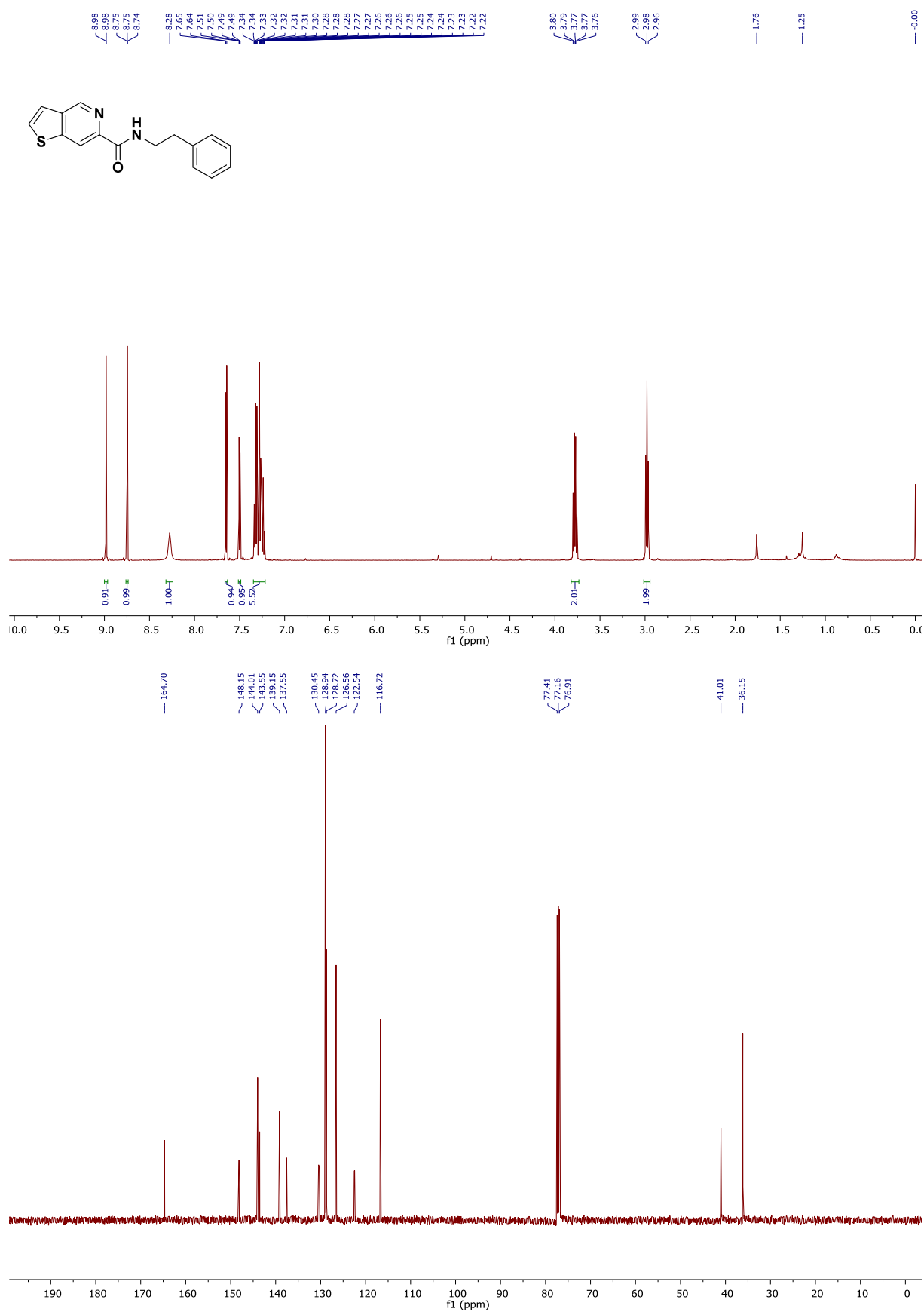


18mdv071-YZ477C #12 RT: 0.21681 /
T: FTMS +p ESI Full ms [220.00-1000.00]

:5

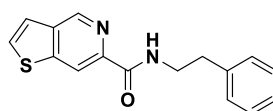
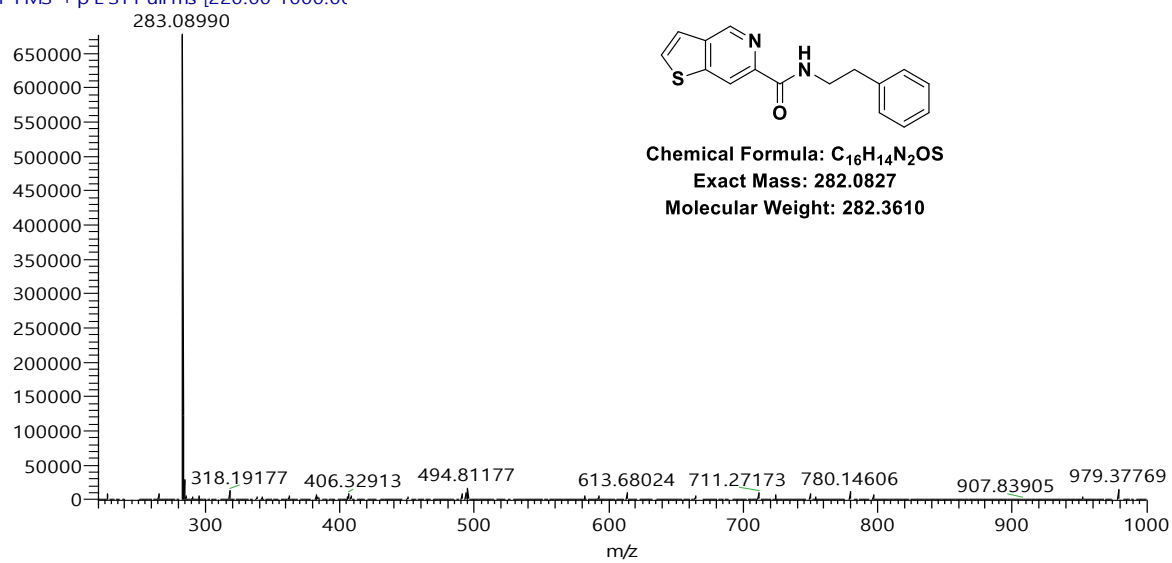


***N*-phenethylthieno[3,2-*c*]pyridine-6-carboxamide (P8)**



18mdv071-YZ473C #13 RT: 0.22783 /
T: FTMS +p ESI Full ms [220.00-1000.00]

:5



Chemical Formula: $C_{16}H_{14}N_2OS$

Exact Mass: 282.0827

Molecular Weight: 282.3610

7. Gram scale reaction procedure for the synthesis of 6,7-dimethoxy-*N*-phenethylisoquinoline-3-carboxamide

A round-bottomed flask (100 mL) equipped with a Teflon-coated magnetic stir bar was charged with 2,2-dimethoxyacetaldehyde (4.52 mL, 30 mmol) and 3,4-dimethoxybenzylamine (5.01 g, 30 mmol) in MeOH (20 mL). The reaction mixture was stirred at room temperature for 20 min, then, benzoic acid (3.66 g, 30 mmol) and phenylethylisocyanide (3.93g, 30 mmol) were added. The reaction mixture was stirred at room temperature for 15 h. Upon completion, the solvent was removed under vacuum. Then, the reaction mixture was dissolved in dioxane (15 mL) and 37% HCl_(aq) (15 mL) was added dropwise via a pressure equalizing dropping funnel (60 mL). The reaction mixture was stirred at room temperature for 12 h. The precipitated product was dried under vacuum. Then, it was dissolved in dichloromethane (100 mL), transferred to a separatory funnel (250 mL), washed with saturated NaHCO₃ solution (3 x 50 mL), water (1 x 50 mL), brine (1 x 50 mL) and dried over MgSO₄. The solvent was removed under vacuum and the crude product was washed with cold tert-butyl methyl ether (7.26 g, yield: 72%, Fig. S4). The purity of the obtained product was determined by qHNMR using benzyl benzoate as internal standard.

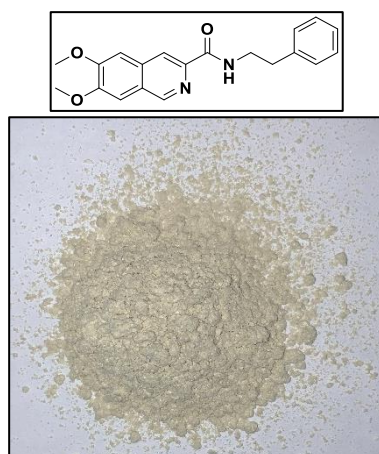


Fig. S4. Multi-gram synthesis.

7.1. qHNMR

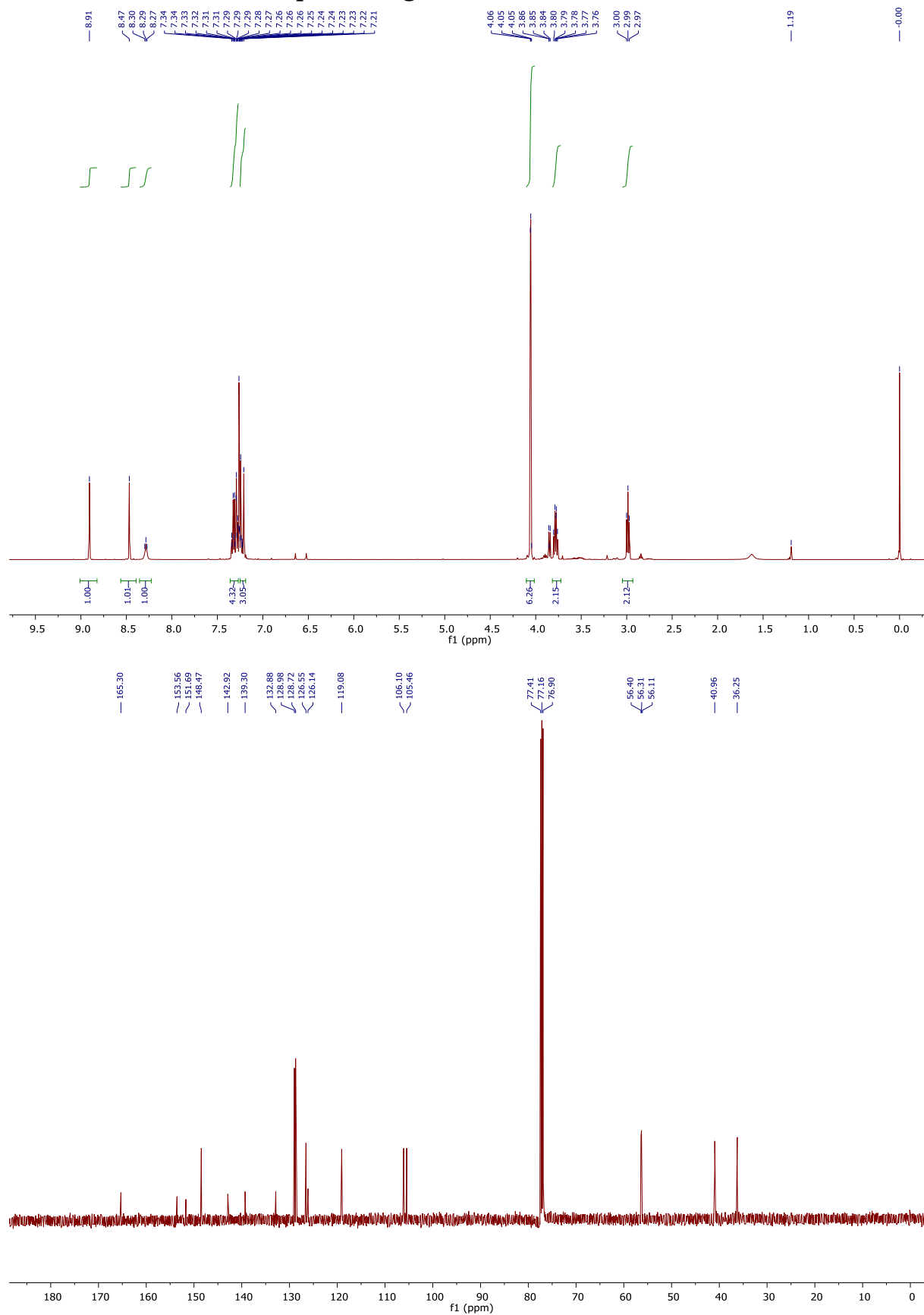
For the ¹H NMR experiment, 42.10 mg (~1/8 mmol) of the gram scale reaction sample and 28.67 mg (~1/8 mmol) of the internal standard were dissolved in CDCl₃.

The following equation was used for quantification:⁹

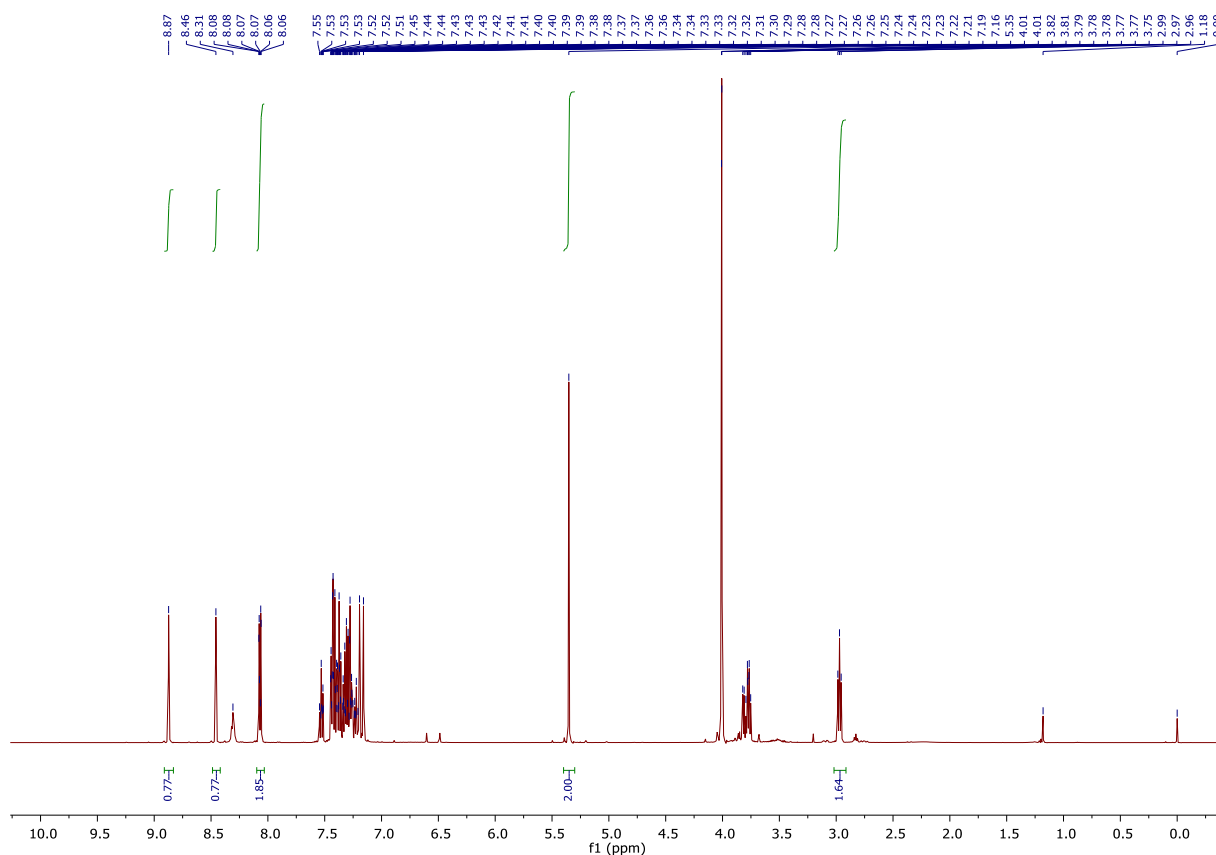
$$P_x = \frac{I_x}{I_{std}} \frac{N_{std}}{N_x} \frac{M_x}{M_{std}} \frac{m_{std}}{m_x} P_{std} = \frac{3.18}{3.85} \frac{4}{4} \frac{336.39}{212.25} \frac{28.67}{42.10} 100 = 89\%$$

where I_x and I_{std} correspond to the integrated signal area of the NMR line of the sample and standard, respectively. N , M , m and P are number of spins (protons), molecular mass, weighed mass and the purity, respectively.

7.2. ^1H and ^{13}C NMR spectra of gram scale reaction



7.3. qHNMR spectrum of gram scale reaction



8. Crystal structure determination

X-ray diffraction data for single crystals of compounds **A3**, **B5** and **K21** were collected using SuperNova (Rigaku - Oxford Diffraction) four circle diffractometer with a mirror monochromator and a microfocus MoK α radiation source ($\lambda = 0.71073$ Å) which was used for monocrystals of **A3** and **K21**. Additionally, the diffractometer was equipped with a CryoJet HT cryostat system (Oxford Instruments) allowing low temperature experiments, performed at 130(2) K. The obtained data sets were processed with CrysAlisPro software.¹⁰ The phase problem was solved with direct methods using SIR2004¹¹ or SUPERFLIP.¹² Parameters of obtained models were refined by full-matrix least-squares on F^2 using SHELXL-2014/6.¹³ Calculations were performed using WinGX integrated system (ver. 2014.1).¹⁴ Figure was prepared with Mercury 3.7 software.¹⁵

All non-hydrogen atoms were refined anisotropically. All hydrogen atoms attached to carbon atoms were positioned with the idealised geometry and refined using the riding model with the isotropic displacement parameter $U_{\text{iso}}[\text{H}] = 1.2 U_{\text{eq}}[\text{C}]$. The difference Fourier map was inspected in order to find position of hydrogens linked to nitrogen atoms. These hydrogen atoms were refined with no restraints on the isotropic displacement parameters. Crystal data and structure refinement results for presented crystal structures are shown in Table S3. The molecular geometry (asymmetric units) observed in presented crystal structures are shown in Fig. S5.

Crystals of compound **A3** exhibited the twinning phenomena. Obtained data show the two-component twin with approximately 52% and 48% of component 1 and component 2, respectively. Data was processed with twin option of the CrysAlisPro software⁹. The obtained model was refined against HKLF4.

Crystallographic data have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication nos.: CCDC1828773 (**B5**), CCDC 1827864 (**K21**) and CCDC 1827863 (**A3**). Copies of the data can be obtained, free of charge, on application to CCDC, 12 Union Road, Cambridge CB2 1EZ, UK, (fax: +44-(0)1223-336033 or e-mail: deposit@ccdc.cam.ac.uk).

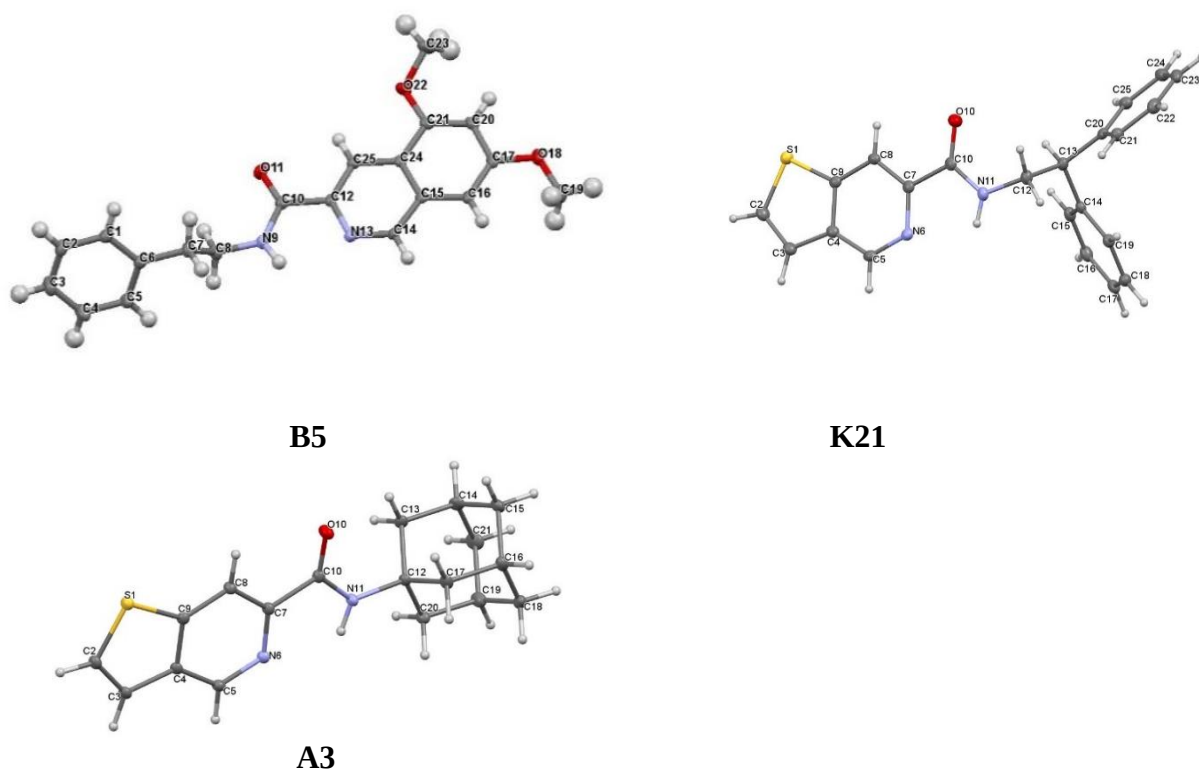


Fig. S5. Molecular geometry observed in crystal structures of compounds **B5**, **K21** and **A3**, showing the atom labelling scheme (here asymmetric units are presented except for **B5**, for which three independent molecules are observed in the asymmetric unit). Displacement ellipsoids of non-hydrogen atoms are drawn at the 30% probability level. H atoms are presented as small spheres with an arbitrary radius.

Table S3. Crystal data and structure refinement results for compounds **B5**, **K21** and **A3**.

	B5	K21	A3
Empirical moiety formula	3x (C ₂₀ H ₂₀ N ₂ O ₃)	C ₂₂ H ₁₈ N ₂ O S	C ₁₈ H ₂₀ N ₂ O S
Formula weight [g/mol]	336.38	358.44	312.42
Crystal system	Monoclinic	Monoclinic	Monoclinic
Space group	P2 ₁ /a	P2 ₁ /n	P2 ₁ /c
Unit cell dimensions	a = 9.3950(4) Å b = 43.7250(14) Å c = 12.6567(4) Å β = 96.256(3)°	a = 8.9525(2) Å b = 19.7350(4) Å c = 10.1928(3) Å β = 100.181(2)°	a = 11.8199(10) Å b = 11.5116(6) Å c = 11.5293(9) Å β = 101.081(9)°
Volume [Å ³]	5168.4(3)	1772.48(8)	1539.5(2)
Z	12	4	4
D _{calc} [Mg/m ³]	1.297	1.343	1.348
μ [mm ⁻¹]	0.088	0.196	0.214
F(000)	2136	752	664
Crystal size [mm ³]	0.4 x 0.3 x 0.1	0.5 x 0.3 x 0.1	0.5 x 0.5 x 0.1
Θ range	2.83° to 28.57°	2.90° to 28.60°	3.27° to 28.67°
Index ranges	-12 ≤ h ≤ 7, -57 ≤ k ≤ 54, -16 ≤ l ≤ 15	-11 ≤ h ≤ 12, -25 ≤ k ≤ 25, -11 ≤ l ≤ 13	-13 ≤ h ≤ 14, -15 ≤ k ≤ 9, -14 ≤ l ≤ 14
Refl. collected	33394	14761	8756
Independent reflections	12007 [R(int) = 0.0298]	4168 [R(int) = 0.0370]	2885 [R(int) = 0.1021]
Completeness [%] to Θ	99.7 (Θ 26.3°)	99.9 (Θ 25.2°)	98.7 (Θ 25.0°)
Absorption correction	Multi-scan	Multi-scan	Multi-scan
Tmin. and Tmax.	0.872 and 1.000	0.757 and 1.000	0.784 and 1.000
Data/ restraints/parameters	12007 / 0 / 683	4168 / 6 / 239	2885 / 0 / 203
GooF on F ²	1.062	1.040	1.049
Final R indices [I > 2σ(I)]	R1 = 0.0534, wR2 = 0.1157	R1 = 0.0405, wR2 = 0.0860	R1 = 0.0548, wR2 = 0.1388
R indices (all data)	R1 = 0.0777, wR2 = 0.1296	R1 = 0.0667, wR2 = 0.0995	R1 = 0.0629, wR2 = 0.1464
Δρ _{max} , Δρ _{min} [e·Å ⁻³]	0.25 and -0.20	0.29 and -0.30	0.38 and -0.45

References

- (1) Obrecht, R.; Herrmann, R.; Ugi, I. Isocyanide synthesis with phosphoryl chloride and diisopropylamine. *Synthesis* **1985**, 1985 (04), 400-402.
- (2) Skorna, G.; Ugi, I. Isocyanide Synthesis with Diphosgene. *Angew. Chem. Int. Ed. Engl* **1977**, 16, 259-260.
- (3) Ugi, I.; Fetzer, U.; Eholzer, U.; Knupfer, H.; Offermann, K. Isonitrile syntheses. *Angew. Chem. Int. Ed. Engl.* **1965**, 4 (6), 472-484.
- (4) Ugi, I.; Meyr, R. o-Tolyl Isocyanide. *Org. Synth.* **1961**, 101-101.
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