

Supporting Information

A highly efficient and enzyme-recoverable method for enzymatic concentrating omega-3 fatty acids generated by hydrolysis of fish oil in a substrate-constituted three-liquid phase system

Zhigang Li,^{a, c} Hua Chen,^a Jinfen Su,^a Weifei Wang,^b Huayong Chen,^{a, c} Bo Yang,^{*}
^{a, c} and Yonghua Wang^{* d}

^a School of Biology and Biological engineering, South China University of Technology, Guangzhou 510006, China.e-mail: yangbo@scut.edu.cn

^b Sericultural & Agri-food Research Institute, Guangdong Academy of Agricultural Sciences, Key Laboratory of Functional Food, Ministry of Agriculture, Guangdong Key Laboratory of Agricultural Products Processing Guangzhou 510610, China.

^c Guangdong Provincial Key Laboratory of Fermentation and Enzyme Engineering,South China University of Technology, Guangzhou 510006, China.

^d School of Light Industry and Food Sciences, South China University of Technology, Guangzhou 510641, China. e-mail: yonghw@scut.edu.cn.

*** Corresponding author.** Yonghua Wang

Table S1 The main fatty acid composition of tuna oil.

fatty acid	Content (w, %)
C14:0	4.26
C15:0	0.85
C16:0	18.54
C18:0	4.85
C20:0	0.52
C16:1	6.42
C18:1	11.00
C20:1	1.06
C22	0.34
C18:2	1.99
C20:2	1.56
C22:2	0.47
C18:3n3	0.81
C18:3n6	0.20
C20:4n6	1.72
EPA	8.25
DPA	1.30
DHA	23.17

Table S2 Number of phases in different systems

	PEG-400	PEG-600	[BMIM]BF ₄	[EMIM]EtSO ₄	methanol	ethanol	isopropanol	isoamyl alcohol
(NH ₄) ₂ SO ₄	III	III	III	III	II	III	III	II
Na ₂ SO ₄	III	III	III	III	II	III	III	II

Table S3 Partition coefficients and recoveries of lipases as well as the yields of the FFAs in different SC-TLPSs contained ionic liquid.

	[DMIM]Cl/ [OMIM]Cl/ [EMIM]EtSO ₄ / [OMIM]Cl/ [EMIM]EtSO ₄ / [BMIM]Br / [EMIM]EtSO ₄ /	Na ₂ SO ₄ ^a	Na ₂ SO ₄ ^a	Na ₂ SO ₄ ^a	Na ₂ SO ₄ ^b	(NH ₄) ₂ SO ₄ ^b	(NH ₄) ₂ SO ₄ ^c	(NH ₄) ₂ SO ₄ ^c
K _{lipase}	573.3	37.9	0	49.3	0	3.6	0	
Y _{lipase (%)}	99.9	99.3	0	96.6	0	76.4	0	
Y _{FFA (%)}	0	0	0.33	0	0.26	0.63	1.01	

^a: lipase AYS; ^b: lipase PL20000; ^c: lipase TL-100L. Y_{FFA}: the yields of the FFAs

Table S4 Changes in fatty acid content of the glycerides fractions and FFAs as well as the recoveries of n3 PUFA obtained by multi-repeated hydrolysis in the SC-TLPS.

	W _{C14:0}	W _{C16:0}	W _{C18:1}	W _{C20:4n6}	W _{EPA}	W _{DPA}	W _{DHA}	W _{PUFA}	R _{PUFA}	
I	GB-FAs (%)	3.00	10.96	9.43	2.74	10.93	2.02	35.91	48.86	87.95
	FFA (%)	5.13	24.90	21.45	0.97	5.65	0.78	10.36	16.79	12.05
II	GB-FAs (%)	2.21	7.53	4.57	3.17	12.02	2.38	42.49	56.89	77.03
	FFA (%)	4.39	19.08	15.61	1.62	7.93	1.37	21.99	31.29	10.92
III	GB-FAs (%)	1.629	5.14	3.21	3.43	12.56	2.57	47.83	62.96	70.27
	FFA (%)	4.24	16.39	8.46	2.07	10.31	1.64	23.76	35.71	6.76
IV	GB-FAs (%)	1.17	3.51	3.05	3.66	12.74	2.705	52.52	67.97	62.58
	FFA (%)	3.72	14.41	6.94	2.37	12.22	1.84	25.48	39.54	7.68
V	GB-FAs (%)	1.06	3.28	2.64	3.68	12.24	2.72	54.59	69.55	58.54
	FFA (%)	3.46	12.26	6.10	2.70	14.26	2.04	26.92	43.23	4.04

GB-FAs: FAs on the glycerol backbone; W_{FA}: mass percent content of the FA; R_{PUFA}: recovery of FA.

Table S5 Diameters and specific surfaces of oil in water, middle phase and bottom phase of SC-TLPS.

	water	middle phase	bottom phase
D-specific surface (μm)	57.030	20.585	31.588
D-volume (μm)	65.991	28.236	49.531
specific surface ($\text{m}^2 \text{ g}^{-1}$)	0.105	0.291	0.19

Table S6 Comparison of achieved hydrolytic ratios of oils in different enzymatic reaction systems.

Reaction system	lipase origin	catalyst form	Substrate	T (°C)	Time (h)	C _{n3-PUFA} (%)	catalyst reuse and stability	ref.
SC-TLPS	<i>Candida rugosa</i>	free lipase (3.25%)	tuna oil	37	8	67.97	no significant losses of activity after 8 uses	this paper
reversed micelles	<i>Candida rugosa</i>	surfactant-coated lipase (3.3%)	tuna oil	40	24	49.8	-	1
O/W system	<i>Candida rugosa</i>	free lipase (6%)	salmon oil	45	8	50.58	-	2
O/W system	<i>Cryptococcus sp.</i>	free lipase (1000U/ml)	sardine oil	25	72	42	-	3
reversed micelles	<i>Candida rugosa</i>	immobilized lipase (30%)	tuna oil	40	10	16.5	80% activity after 5 uses (washed with tBA)	4
cyclohexane /water system	<i>Rhizomucor miehei</i>	immobilized lipases (25%)	fish oil	25	24	-	85% activity after 5 uses (washed with cyclohexane)	5
Reverse micelles	<i>Yarrowia lipolytica</i>	immobilized lipase (500U/g)	fish oil	25	24	36.8 (DHA)	80% activity after 20 uses (washed with PBS)	6
O/W system	<i>Yarrowia lipolytica</i>	immobilized lipase (64%)	<i>Chlorella protothecoides</i> oil	30	16	31.5 (DHA)	90% activity after 10uses (washed with PBS)	7

T: temperature; Y: the hydrolysis rate of oil; (DHA): C_{DHA}

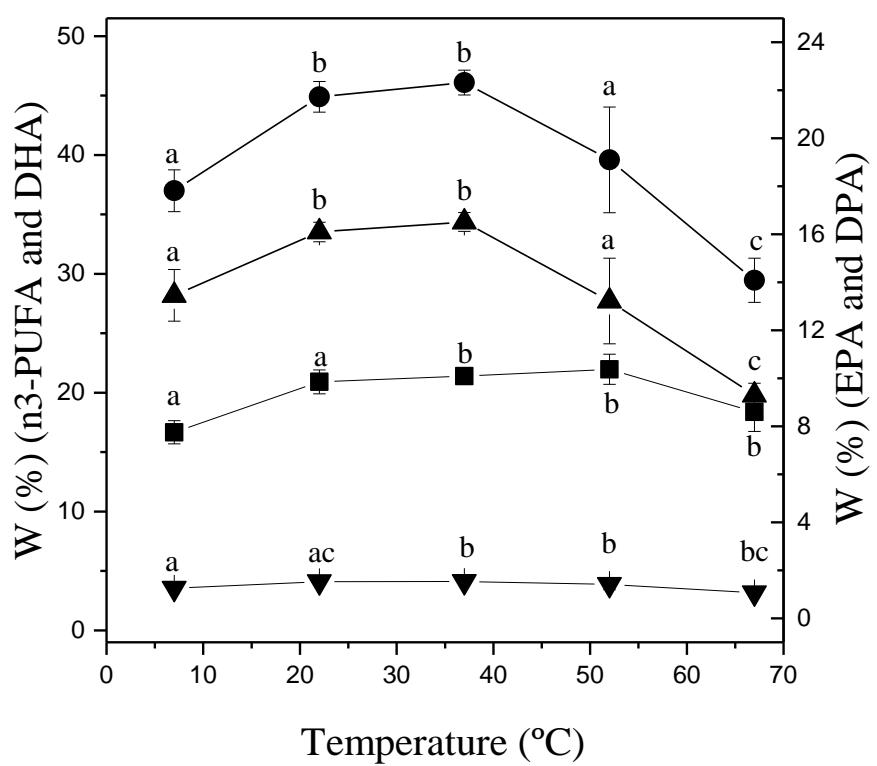


Figure S1. Effects of temperature of FFA in the PEG-400/Na₂SO₄ SC-TLPS contained lipase AYS. (●: n-3 PUFA; ▲: DHA; ■: EPA; ▼: DPA).

References

- 1.Ko W C, Wang H J, Hwang J S, et al. Efficient hydrolysis of tuna oil by a surfactant-coated lipase in a two-phase system[J]. Journal of agricultural and food chemistry, 2006, 54(5): 1849-1853.
- 2.Kahveci D, Xu X. Repeated hydrolysis process is effective for enrichment of omega 3 polyunsaturated fatty acids in salmon oil by Candida rugosa lipase[J]. Food Chemistry, 2011, 129(4): 1552-1558.
- 3.Aarthy M, Saravanan P, Ayyadurai N, et al. A two step process for production of omega 3-polyunsaturated fatty acid concentrates from sardine oil using Cryptococcus sp. MTCC 5455 lipase[J]. Journal of Molecular Catalysis B: Enzymatic, 2016, 125: 25-33.
- 4.Yan J, Liu S, Hu J, et al. Enzymatic enrichment of polyunsaturated fatty acids using novel lipase preparations modified by combination of immobilization and fish oil treatment[J]. Bioresource technology, 2011, 102(14): 7154-7158.
- 5.Mohammadi M, Habibi Z, Dezhvarei S, et al. Selective enrichment of polyunsaturated fatty acids by hydrolysis of fish oil using immobilized and stabilized Rhizomucor miehei lipase preparations[J]. Food and Bioproducts Processing, 2015, 94: 414-421.
6. Liu T, Zhao Y, Wang X, et al. A novel oriented immobilized lipase on magnetic nanoparticles in reverse micelles system and its application in the enrichment of polyunsaturated fatty acids[J]. Bioresource technology, 2013, 132: 99-102.
- 7.Yan Y, Zhang X, Chen D. Enhanced catalysis of Yarrowia lipolytica lipase LIP2 immobilized on macroporous resin and its application in enrichment of polyunsaturated fatty

acids[J]. Bioresource technology, 2013, 131: 179-1.