

Supporting Information for

Absolute binding free energy calculation and design of a subnanomolar inhibitor of phosphodiesterase-10

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Section S1. PDE4, PDE5, and PDE9 inhibitors used in this study

PDE4 inhibitors: There are 20 PDE4 inhibitors examined in this study. The structures of the first 8 molecules share certain similarity, whereas the rest structures are quite different from each other. Structures of all the PDE4 inhibitors are given in Fig. S1.

PDE5 inhibitors: 11 PDE5 inhibitors were used in this study. Their structures are given in Fig. S2.

PDE9 inhibitors: Seven PDE9 inhibitors were used in this study. Their structures are given in Fig. S3.

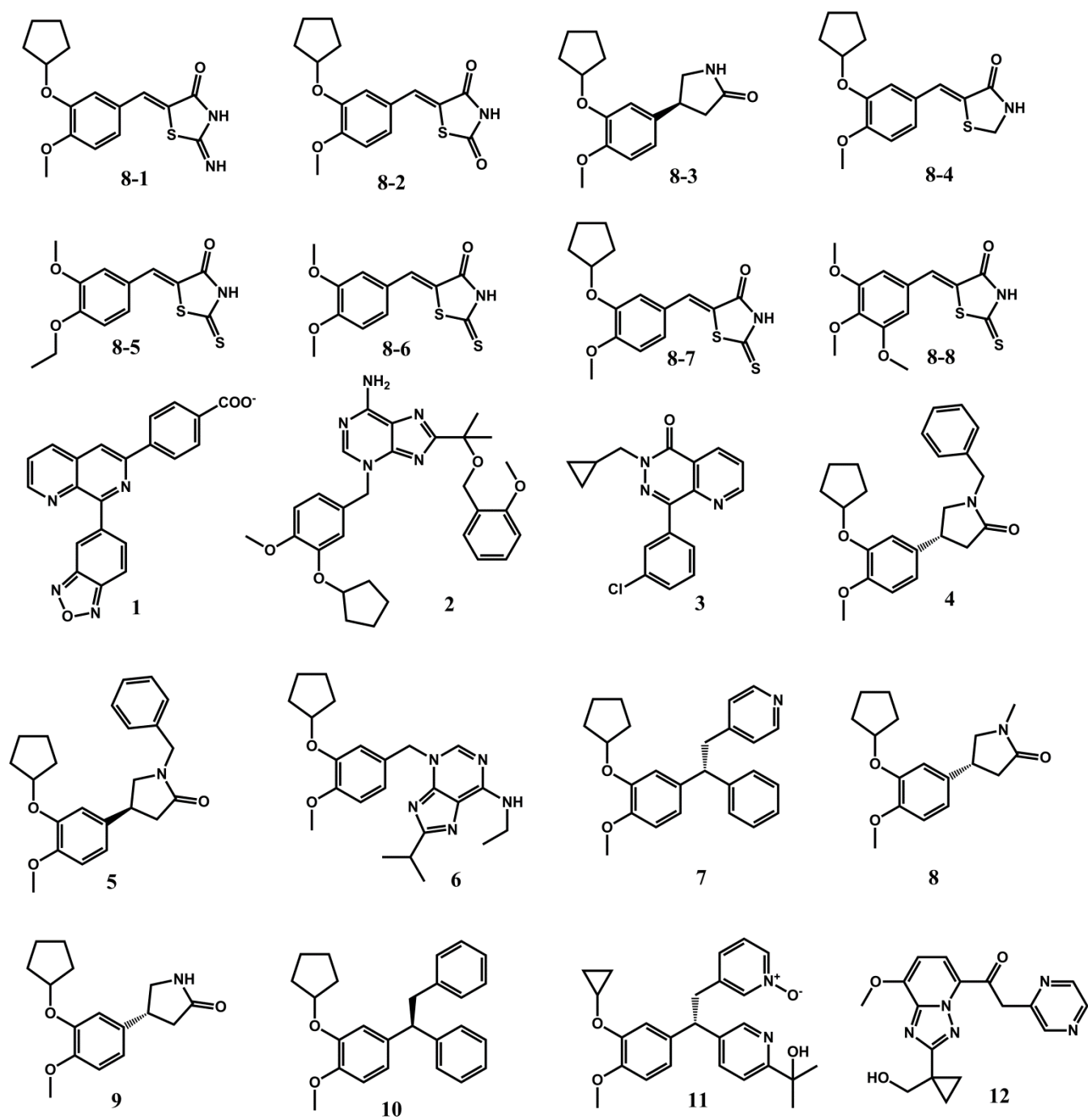


Fig. S1. Structures of PDE4 inhibitors used in this study.

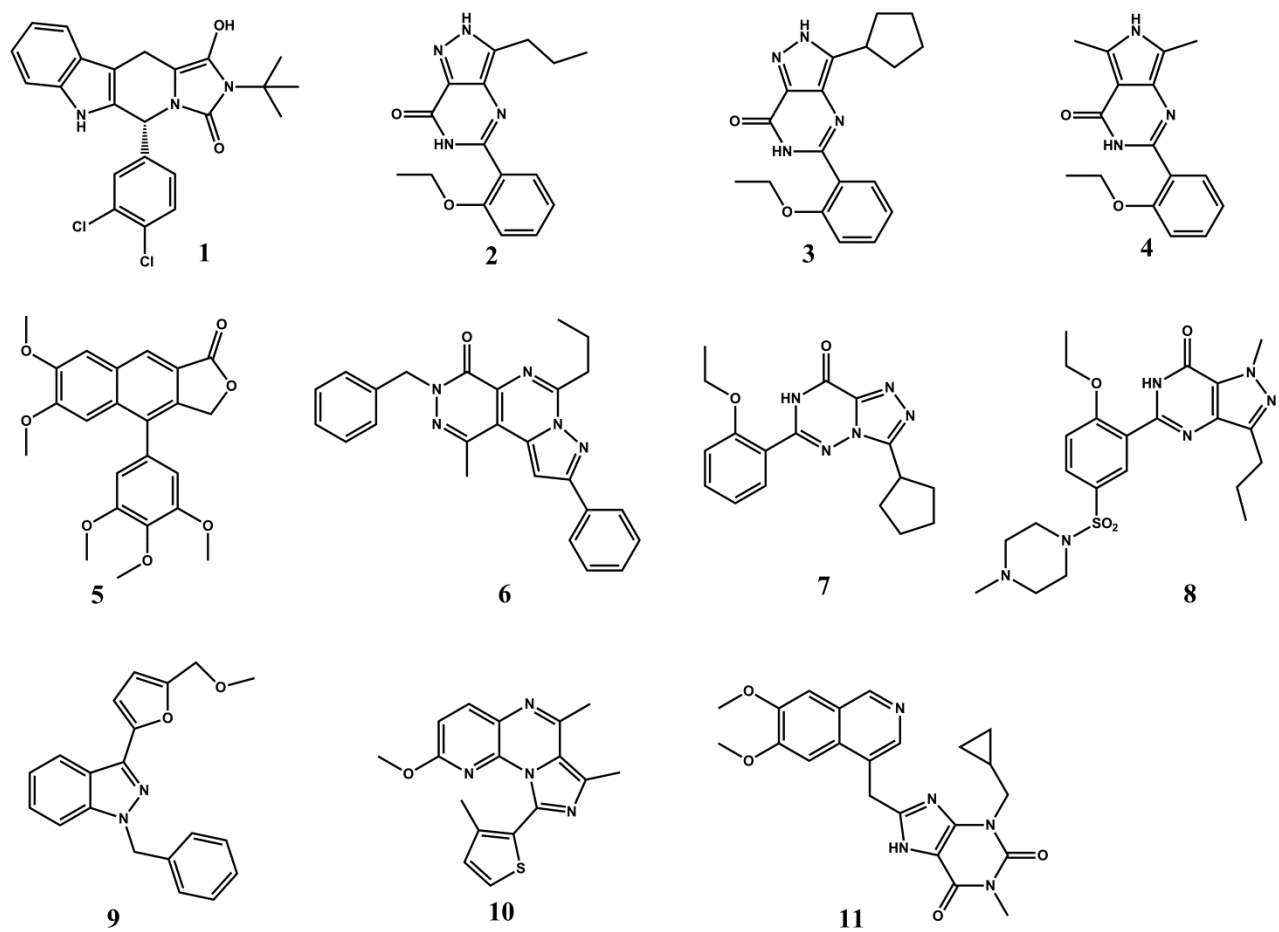
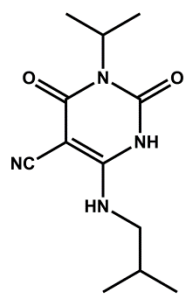
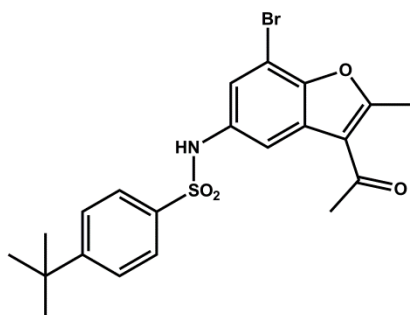


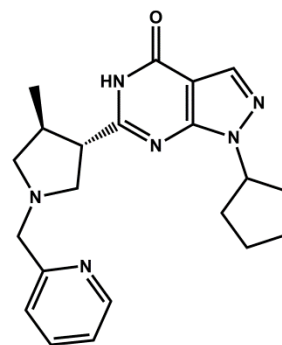
Fig. S2. Structures of PDE5 inhibitors used in this study.



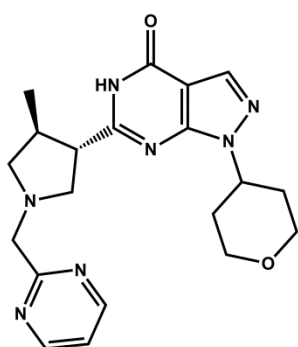
1



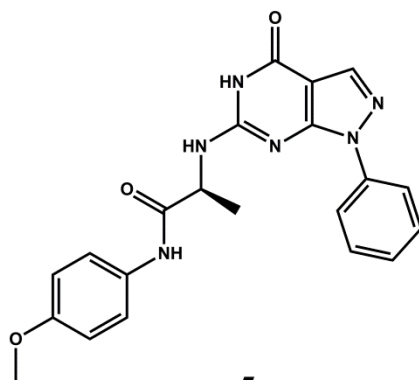
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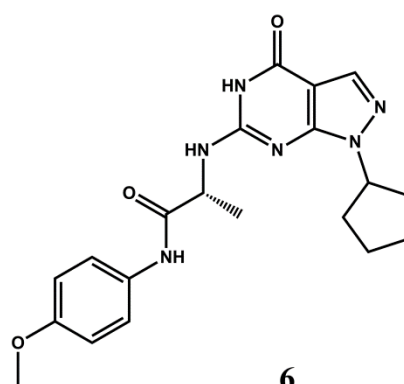
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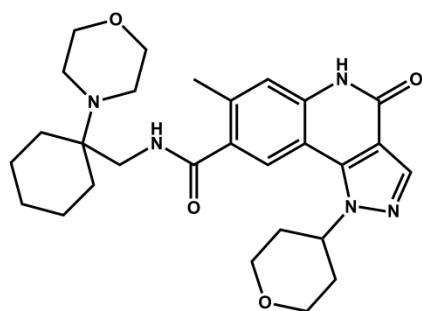
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5



6



7

Fig. S3. Structures of PDE9 inhibitors used in this study.

Section S2. An example to show how the GA-FEP calculations work

By using lig_16 of CDK2 as an example, the fitted probability distribution $P(\Delta U)$ is shown as Fig. S4. Picture (a) represents the complex system (Rec-Lig), and picture (b) represents the Lig system. The probability distributions of both forward ΔU and backward ΔU are included, and shown in the same color for the same alchemical state. The 10 probability distributions depicted by black, red, green, blue and cyan in Fig. S4 correspond to the first 5 states that annihilated the electrostatic interactions. The rest probability distributions in the center of the figure correspond to the following 5 states that deal with annihilating vdW interactions. To improve calculation accuracy, for each state, the forward and backward perturbation energies (calculated by equation (2)), and BAR(2, 3) energies that combines the forward and backward results (calculated by equation (6) and (7)) were calculated based on the fitted probability distribution. For each state, if the energies of the forward and backward perturbations differ too much, the final result would be unreliable, and more simulations should be considered. Table S1 listed the calculation result of lig_16 of CDK2. In this table, c_fwd, c_bwd, l_fwd, l_bwd represent the forward and backward energy results calculated by basic FEP equation (2), c_dif and l_dif represents the energy difference between forward energy and backward energy, c_bar and l_bar represents the energy results calculated by BAR, and ene represents the final energy calculation result. As seen from the data in the table, for all the states, the forward and backward energies were similar to each other. The energy difference between the final state and the state prior to it was calculated by using the basic FEP equation instead of BAR due to the backward energy calculation from the final target state will face with 'end-point catastrophes'(4). The c_tot and l_tot are the summations of the energetic results using BAR method (except for the final state which was calculated by using the basic FEP equation), and the final binding free energy was the difference between l_tot and c_tot. The standard deviation of the electrostatic energy was evaluated based on the forward, backward and BAR energies. The standard deviation of the total interaction energies was evaluated based on the forward and BAR energies, because the backward energy may be calculated for the final state.

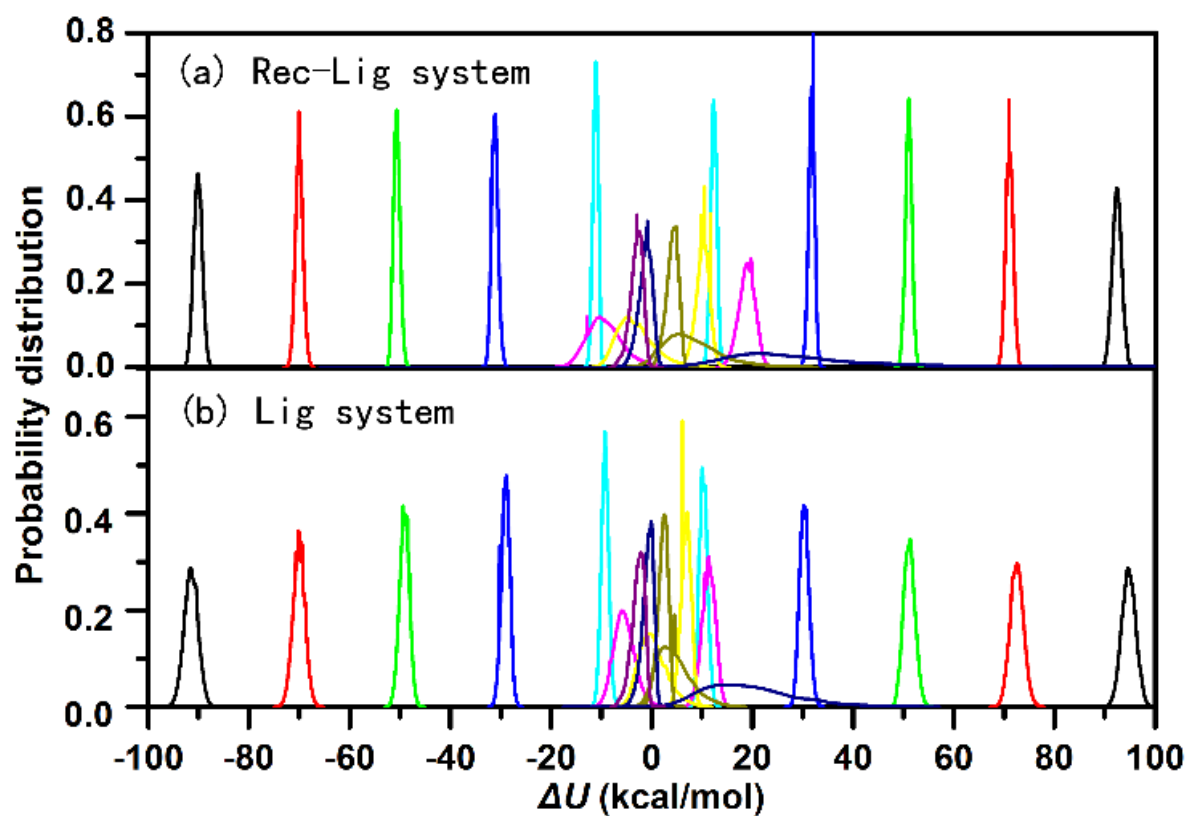


Fig. S4. The fitted $P(\Delta U)$ of all the states of lig_16 of CDK2. The $P(\Delta U)$ for both the forward and backward calculations are included in this figure, and they are almost symmetric to each other with respect to the origin.

Table S1. Calculation result of lig_16 of CDK2. In this table, c_fwd, c_bwd, l_fwd, l_bwd represent the forward and backward energy results calculated by basic FEP equation, c_dif and l_dif represent the energy difference between the forward and backward energies, c_bar and l_bar refer to the energy results calculated by BAR, and ene represents the final energy calculation result. c_tot and l_tot refer to the summations of the energy results using BAR method (except for the final state which was calculated by using basic FEP equation), and the final binding free energy was the difference between l_tot and c_tot.

| state | c_fwd ¹ | c_bwd | c_bar | c_dif | l_fwd ² | l_bwd | l_bar | l_dif | c_tot | l_tot | ene |
|----------------------|--------------------|---------|--------|--------|--------------------|---------|--------|--------|---------|---------|---------|
| 00_reference | 91.608 | -90.593 | 91.14 | 1.015 | 93.106 | -93.139 | 93.122 | -0.033 | 91.14 | 93.122 | 1.982 |
| 01_charge_0.2 | 70.56 | -70.351 | 70.454 | 0.209 | 71.138 | -71.308 | 71.262 | -0.17 | 161.594 | 164.384 | 2.79 |
| 02_charge_0.4 | 50.662 | -50.883 | 50.787 | -0.221 | 50.023 | -49.95 | 50.084 | 0.073 | 212.381 | 214.468 | 2.087 |
| 03_charge_0.6 | 31.443 | -31.434 | 31.429 | 0.009 | 29.6 | -29.632 | 29.642 | -0.032 | 243.81 | 244.11 | 0.3 |
| 04_charge_0.8 | 11.992 | -11.352 | 11.688 | 0.64 | 9.772 | -9.77 | 9.768 | 0.002 | 255.498 | 253.878 | -1.62 |
| 05_charge_1.0 | 16.762 | -16.75 | 15.359 | 0.012 | 9.688 | -8.182 | 8.789 | 1.506 | 270.857 | 262.667 | -8.19 |
| 06_vdw_0.2 | 8.361 | -8.719 | 7.769 | -0.358 | 5.845 | -3.807 | 4.386 | 2.038 | 278.626 | 267.053 | -11.573 |
| 07_vdw_0.4 | 2.624 | -1.597 | 0.936 | 1.027 | 1.024 | -0.782 | 0.122 | 0.242 | 279.562 | 267.175 | -12.387 |
| 08_vdw_0.6 | -3.193 | 0.751 | -5.566 | -2.442 | -2.373 | 2.019 | -4.361 | -0.354 | 273.996 | 262.814 | -11.182 |
| 09_vdw_0.8 | -4.721 | --- | --- | -4.721 | -4.493 | --- | --- | --- | 269.275 | 258.321 | -10.954 |

¹: c in c_fwd, c_bwd, c_bar and c_tot represents the Rec-Lig complex

²: l in l_fwd, l_bwd, l_bar and l_tot represents the Lig system

Section S3. Using 10 lambda windows could reach to similar accuracy with that of 20 or 39 lambda windows.

The overall alchemical transformation of either Rec-Lig system or Lig system contains 10 alchemical states (lambda windows), in which the first 5 states were used to decouple the electrostatic interactions and the rest were used to decouple the vdW interactions. To show the effect of increasing the number of intermediate states to the energy calculation results, we calculated ABFE for 3 CDK2 inhibitors, including **30**, **28** and **1oiy**, with doubled and quadrupled number of lambda windows. Table S2 shows the calculation details of ligand **30**, and calculation details of all other ligands could be found in Supporting Dataset S1. As could be seen from this table, the convergence of the FEP-ABFE calculations were pretty good. When lambda of charge interaction reached to 1.0 and lambda of vdW interaction reached to 0.999936, the energies for (Rec-Lig system, Lig system) calculated with 39, 20 and 10 lambda values were (320.002, 305.880) kcal/mol, (319.376, 305.848) kcal/mol and (320.444, 306.577) kcal/mol, respectively, indicating the reliability of 10-lambda-windows FEP calculation. The energy for the last step of 10-lambda-windows calculation (lambda of vdW interaction from 0.999936 to 1.0) was calculated based on forward perturbation instead of BAR, and the calculated FEP-ABFE result (-14.119 kcal/mol) was similar to that of 20-lambda-windows calculation (-13.988 kcal/mol) and 39-lambda-windows calculation (-14.234 kcal/mol). Calculation results for **28** and **1oiy** (see Supporting Dataset S1 for details) also showed the good convergence and reliability of the 10-lambda-windows FEP calculation.

We further increased the simulation time from 4 ns to 20 ns for each lambda window for these three ligands, and the sampled probability distributions of the last 18 ns were used for energy calculation, and the ABFE results wasn't affected too much (Supporting Dataset S1).

Table S2. Calculation details of CDK2 inhibitor 30 using different number of λ values. com_ene represents the energy of Rec-Lig system; lig_ene represents the energy of Lig system; FEP_ene represents the FEP energy calculated by equation (1). In the first several steps, charge interactions were decoupled with λ_{chg} gradually increased to 1.00, while the vdW interactions were gradually decoupled in the following steps. Part of the table was omitted, and the full table could be found in Supporting Dataset S1.

| λ values | | 39 λ | | | 20 λ | | | 10 λ | | |
|------------------------|------------------------|--------------|---------|---------|--------------|---------|---------|--------------|---------|---------|
| λ_{chg} | λ_{vdw} | com_ene | lig_ene | FEP_ene | com_ene | lig_ene | FEP_ene | com_ene | lig_ene | FEP_ene |
| 0.05 | 0.000000 | 28.791 | 29.525 | 0.734 | | | | | | |
| 0.10 | 0.000000 | 56.044 | 57.329 | 1.285 | 56.001 | 57.294 | 1.293 | | | |
| 0.15 | 0.000000 | 81.770 | 83.523 | 1.753 | | | | | | |
| 0.20 | 0.000000 | 105.936 | 108.121 | 2.185 | 105.936 | 108.093 | 2.157 | 105.992 | 109.034 | 3.042 |
| ... | ... | ... | ... | ... | ... | ... | ... | ... | ... | ... |
| 1.00 | 0.000000 | 294.846 | 293.617 | -1.229 | 294.988 | 293.590 | -1.398 | 295.142 | 294.826 | -0.316 |
| 1.00 | 0.264908 | 301.030 | 296.720 | -4.310 | | | | | | |
| 1.00 | 0.468559 | 306.490 | 299.561 | -6.929 | 306.217 | 299.525 | -6.692 | | | |
| 1.00 | 0.622850 | 311.184 | 302.071 | -9.113 | | | | | | |
| 1.00 | 0.737856 | 315.151 | 304.124 | -11.027 | 314.820 | 303.962 | -10.858 | 315.194 | 305.726 | -9.468 |
| 1.00 | 0.822021 | 318.501 | 305.902 | -12.599 | | | | | | |
| 1.00 | 0.882351 | 321.411 | 307.568 | -13.843 | 320.790 | 307.188 | -13.602 | | | |
| 1.00 | 0.924581 | 323.666 | 308.980 | -14.686 | | | | | | |
| 1.00 | 0.953344 | 325.305 | 309.982 | -15.323 | 324.558 | 309.309 | -15.249 | 325.283 | 311.295 | -13.988 |
| 1.00 | 0.972319 | 326.316 | 310.592 | -15.724 | | | | | | |
| 1.00 | 0.984375 | 326.908 | 310.863 | -16.045 | 326.328 | 310.396 | -15.932 | | | |
| 1.00 | 0.991696 | 327.036 | 310.759 | -16.277 | | | | | | |
| 1.00 | 0.995904 | 326.775 | 310.452 | -16.323 | 325.972 | 310.460 | -15.512 | 326.552 | 311.584 | -14.968 |
| 1.00 | 0.998162 | 326.100 | 309.856 | -16.244 | | | | | | |
| 1.00 | 0.999271 | 324.825 | 308.937 | -15.888 | 323.653 | 308.961 | -14.692 | | | |
| 1.00 | 0.999756 | 322.830 | 307.680 | -15.150 | | | | | | |
| 1.00 | 0.999936 | 320.002 | 305.880 | -14.122 | 319.376 | 305.848 | -13.528 | 320.444 | 306.577 | -13.867 |
| 1.00 | 0.999989 | 316.871 | 303.339 | -13.532 | | | | | | |
| 1.00 | 0.999999 | 313.305 | 299.746 | -13.559 | 313.125 | 299.812 | -13.313 | | | |
| 1.00 | 1.000000 | 309.043 | 294.809 | -14.234 | 308.863 | 294.875 | -13.988 | 315.126 | 301.007 | -14.119 |

Section S4. Comparison between ABFE and RBFE for the same data set

To show the accuracy of the ABFE calculation method and compare it with the reported RBFE method, the calculation result of each target is given here separately, as shown in Fig. S5. On the left side are the results calculated by our ABFE method, and on the right side are the results calculated by RBFE method reported by Abel and Wang (1). The pairwise comparison between the two methods shows that they have comparable accuracy. For some targets, such as CDK2, the ABFE method performed even better than the RBFE method.

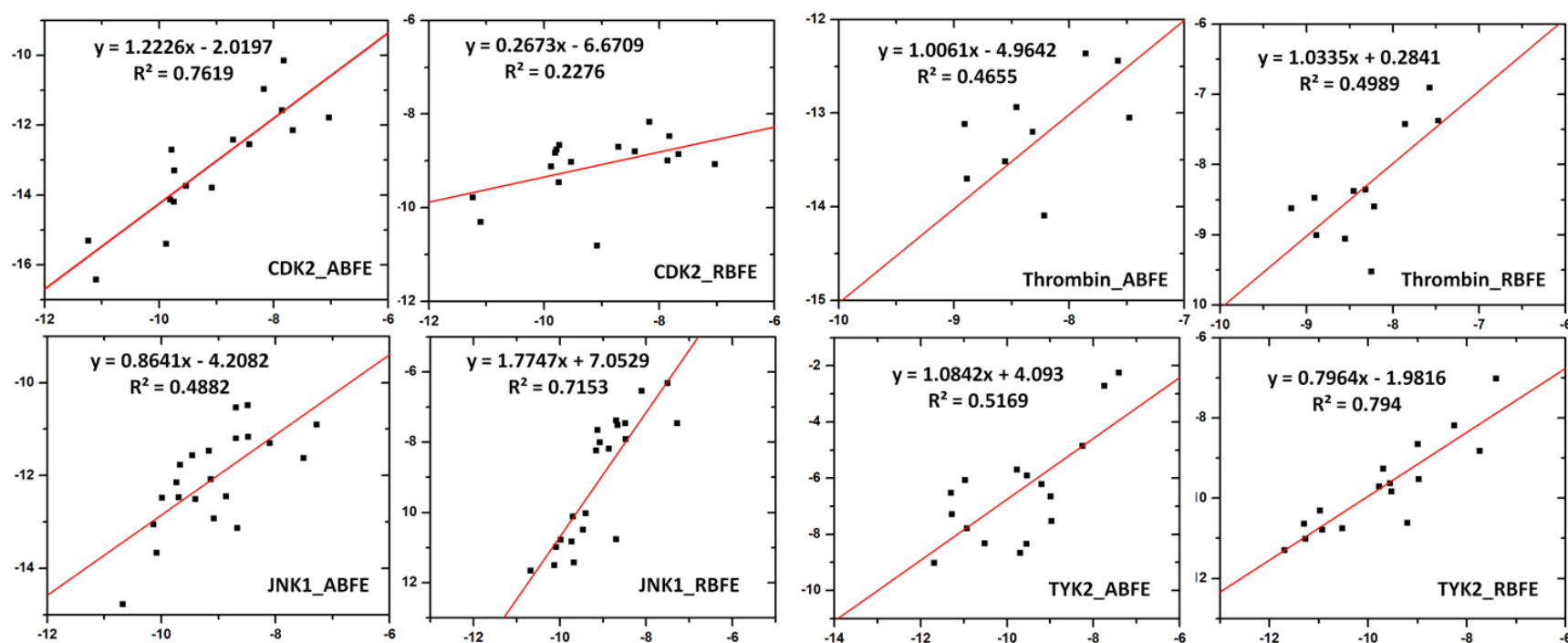


Fig. S5. Comparison between the ABFE and RBFE data for the same data set.

Section S5. Residual distribution of the GA-FEP/ABFE method.

to find out the distribution property of the calculated results of our GA-FEP method, we tried to put all the residual distributions of the 100+ results in Figure 5 together, and the distribution is just like the histogram in Fig. S6 below. We have also added a Gaussian distribution with the same deviation (SD = 1.03) and AUC on this figure, and we found that the distribution of the calculated result is just very much like a Gaussian distribution.

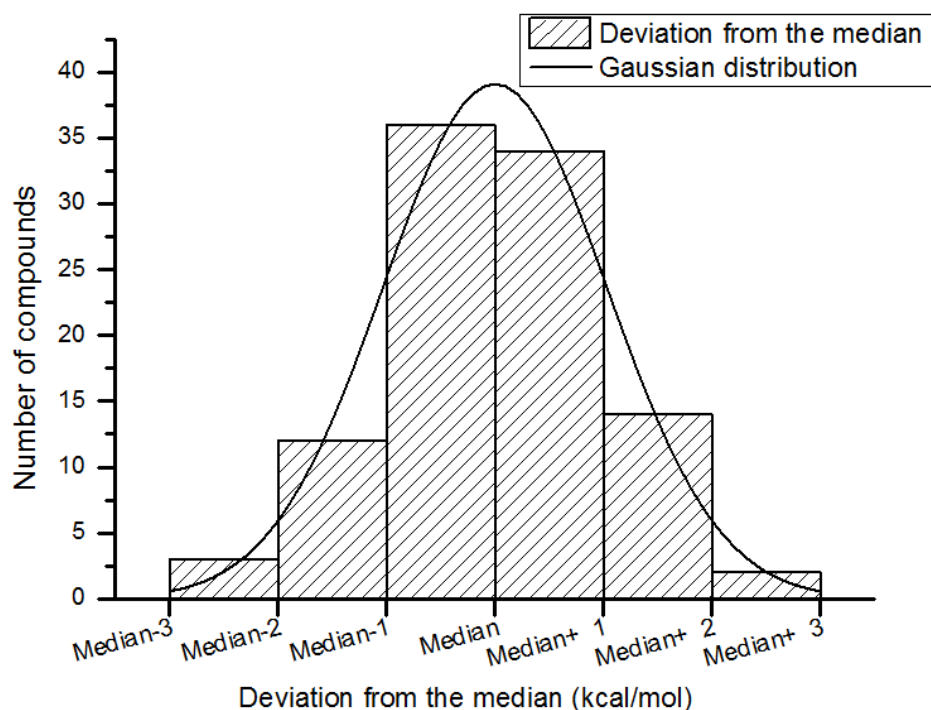


Fig. S6. Residual distributions of all the > 100 ABFE results.

Section S6. Designed PDE10 inhibitors. Molecular structures of designed PDE10 inhibitors and their inhibitory activity (IC_{50}) are given in Table S3. The FEP calculated ABFE value are given in Table S4.

Table S3. Molecular structures of designed PDE10 inhibitors and their inhibitory activity (IC_{50})

| Ligand | Structure | IC_{50} (nM) | Ligand | Structure | IC_{50} (nM) |
|--------|-----------|-------------------|--------|-----------|-------------------|
| LHB-1 | | 1800 | LHB-6 | | 1.7 |
| LHB-2 | | 890 | LHB-7 | | 1.1 |
| LHB-3 | | 403 | LHB-8 | | 2.0 |
| LHB-4 | | 5.9 | LHB-9 | | 21 |
| LHB-5 | | 73 | LHB-10 | | 0.87 |

Table S4. Calculation and experimental results of designed PDE10 inhibitors. exp stands for Experimental results, and FEP-cal stands for the energy calculated by FEP. Correlation constant R between calculation and experimental results is 0.86.

| ligand_ID | IC50(nM) | exp (kcal/mol) | FEP-cal (kcal/mol) | ene_corrected_by_linear_regression (kcal/mol)¹ |
|------------------|-----------------|---------------------------|-------------------------------|--|
| LHB-1 | 1800 | -7.855 | -9.208 | -6.37 |
| LHB-2 | 890 | -8.273 | -12.121 | -8.39 |
| LHB-3 | 403 | -8.744 | -12.603 | -8.73 |
| LHB-4 | 5.9 | -11.252 | -17.07 | -11.83 |
| LHB-5 | 73 | -9.758 | -16.209 | -11.23 |
| LHB-6 | 1.7 | -11.991 | -16.779 | -11.63 |
| LHB-7 | 1.1 | -12.250 | -17.648 | -12.24 |
| LHB-8 | 2 | -11.895 | -18.039 | -12.51 |
| LHB-9 | 21 | -10.498 | -16.394 | -11.36 |
| LHB-10 | 0.87 | -12.389 | -15.328 | -10.62 |

¹: The linear regression function between exp(x) and FEP-cal(y) is $y = 1.4379x - 0.0545$.

ene_corrected_by_linear_regression is the corresponding predicted x value for each ligand based on the linear regression function.

Section S7. Crystal structure of PDE10 with LHB-6 was determined to verify the predicted binding mode. The coordinate and structure factors have been deposited in the Protein Data Bank with PDB ID **5ZNL**. The diffraction data is given in Table S5.

Table S5. Diffraction data and structure refinement statistic for PDE10A-LHB-6 structure

| <i>Data collection</i> | PDE10A-LHB-6 |
|---|---|
| Wavelength (Å) | 1.5418 |
| Temperature (K) | 100 |
| Resolution (Å) | 24.11-2.80 |
| Space group | P2 ₁ 2 ₁ 2 ₁ |
| Unit Cell | |
| <i>a</i> , <i>b</i> , <i>c</i> (Å) | 49.151,81.328,158.207 |
| α , β , γ (°) | 90.00, 90.00, 90.00 |
| No. reflections | 73182(15928) |
| Completeness (%) | 97.86(97.69) |
| <i>R</i> _{merge} | 0.107(0.274) |
| $\langle I/\sigma(I) \rangle$ | 11.7(3.7) |
| Redundancy | 4.6(3.34) |
| <i>Structure refinement</i> | |
| R-factor/R-free | 0.23/0.30 |
| RMS deviations | |
| Bond lengths | 0.0081 Å |
| Bond angles | 1.1754 |
| Average B-factor (Å ²)(atoms) | |
| Protein | 34.8(5003) |
| Inhibitor | 30.6(62) |
| Zn | 33.6(2) |
| Mg | 17.6(2) |
| Waters | 16.8(80) |
| Ramachandran plot | |
| Preferred | 95.70% |
| Allowed | 4.14% |

Section S8. Details about fitting probability distribution. In the post-simulation processing steps, for each FEP state, the last 2 ns were saved into production MD trajectory files with an interval of 100 fs, which resulted in 20,000 snapshots. As shown in Fig. S7, all the 20,000 data points of ΔU was transformed to its distribution $P(\Delta U)_{data}$ with 150 bins. As a result, we get 150 data points of $P(\Delta U)_{data}$. Since there are 15 parameters to fit, the number of data points is 10 times of that of the parameters, and thus it's unlikely to be overfitted.

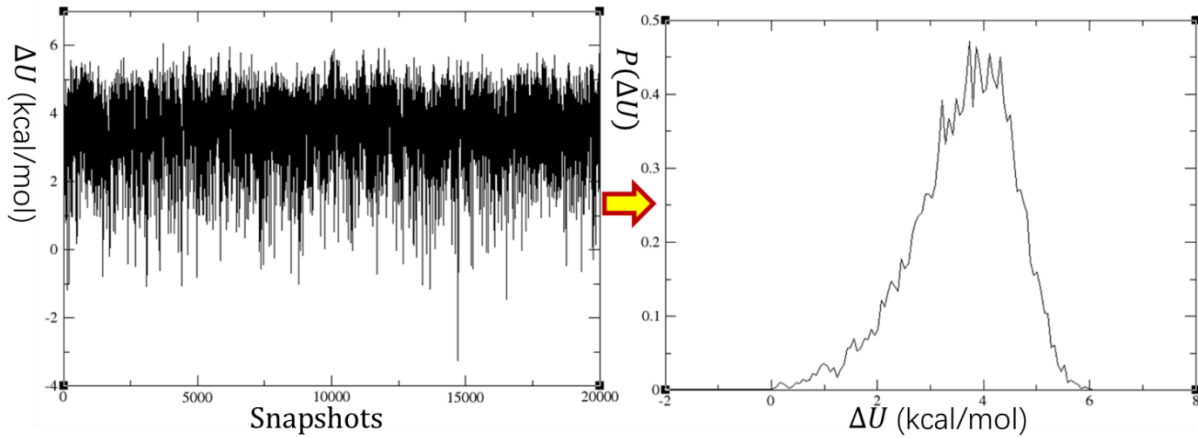


Fig. S7. ΔU data points transformed to the distribution $P(\Delta U)_{data}$

To fit these parameters, we used the following least square procedures:

1. Since the function to be fitted is in the following form:

$$P(\Delta U)_{func} = \sum_{i=1}^5 c_i \exp\left(-\frac{(\Delta U - \mu_i)^2}{2\sigma_i^2}\right)$$

We designed a cost function in the form of

$$\begin{aligned} Cost(c_i, \mu_i, \sigma_i, \Delta U, P(\Delta U)_{data}) &= \sum_{\text{over all data points}} (P(\Delta U)_{data} - P(\Delta U)_{func})^2 \\ &= \sum_{\text{over all data points}} \left(P(\Delta U)_{data} - \sum_{i=1}^5 c_i \exp\left(-\frac{(\Delta U - \mu_i)^2}{2\sigma_i^2}\right) \right)^2, \end{aligned}$$

where $P(\Delta U)_{data}$ is the distribution data of the original FEP simulation.

2. We minimized the value of the cost function by using Stochastic Steepest Descent (SGD) method and get the optimized parameters for the fitted function. Of course, other

minimization method can also be applied, which include Mini-batch Steepest Descent, Stochastic Steepest Descent with momentum, Nesterov Accelerated Gradient, Adagrad, Adadelta, RMSprop, etc. However, SGD method just worked well in this situation.

SGD method optimize the 15 parameters c_i, μ_i, σ_i where $i = 1, 2, 3, 4, 5$ by the following procedure:

$$(c_i, \mu_i, \sigma_i)_t = (c_i, \mu_i, \sigma_i)_{t-1} - \lambda \nabla_{c_i, \mu_i, \sigma_i} \text{Cost}(c_i, \mu_i, \sigma_i, \Delta U, P(\Delta U)_{data})$$

Loop until the parameters in the t th cycle $(c_i, \mu_i, \sigma_i)_t$ are very close to $(c_i, \mu_i, \sigma_i)_{t-1}$, and the resulted $(c_i, \mu_i, \sigma_i)_t$ are the optimized parameters for the fitted function.

The fitted probability distribution will be like Fig. S8:

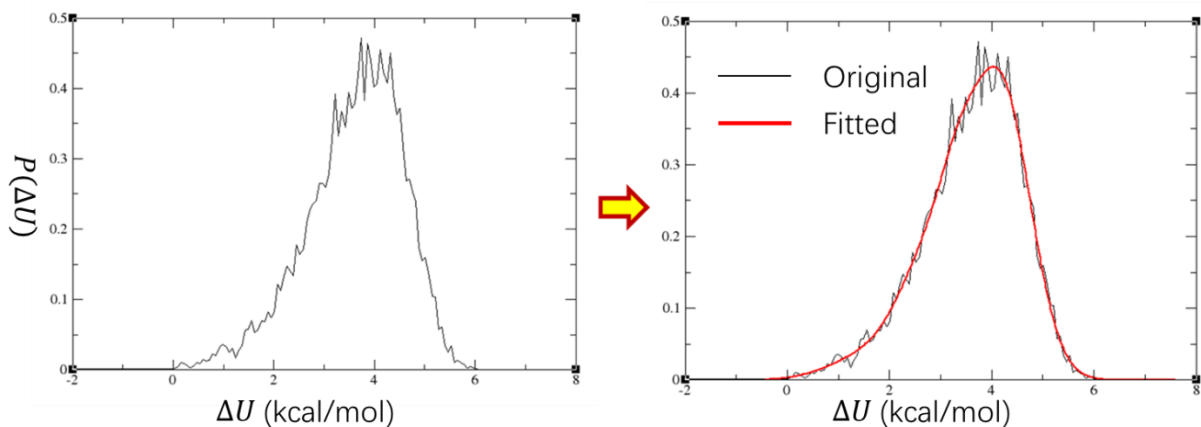


Fig. S8. Fitting the probability distribution.

Note: The probability distribution used as the example is a little bit “skewed” because this is a perturbation of ligand system from ($\lambda_{\text{chg}} = 1.00, \lambda_{\text{vdw}} = 0.953344$) to ($\lambda_{\text{chg}} = 1.00, \lambda_{\text{vdw}} = 0.995904$). The ligand is nearly totally annihilated, and the surrounding water molecules can get closer to the atoms of the ligand. The unphysical state will skew the resulted probability distribution. We are using this example here just want to show that 5 Gaussians can work well even for skewed distributions near the end of the perturbation.

Here are some other examples of fitted probability distributions in Fig. S9:

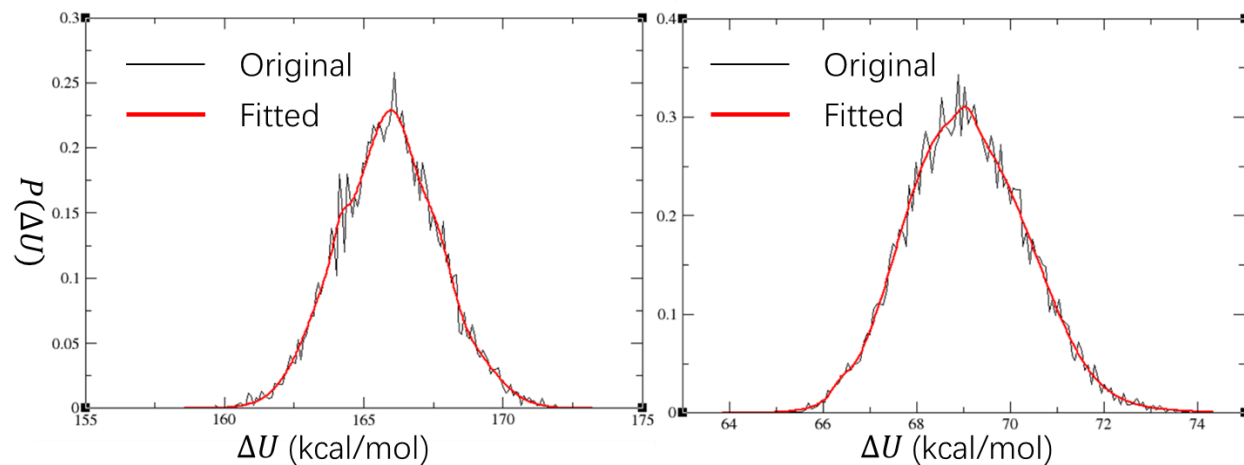


Fig. S9. Examples of some fitted probability distributions

3. Based on the fitted function, we generate 500,000 to 1,000,000 new data points. The newly generated data points are used for further energy calculations based on either BAR method (Eq. 6 and 7) or traditional FEP method (Eq. 2).

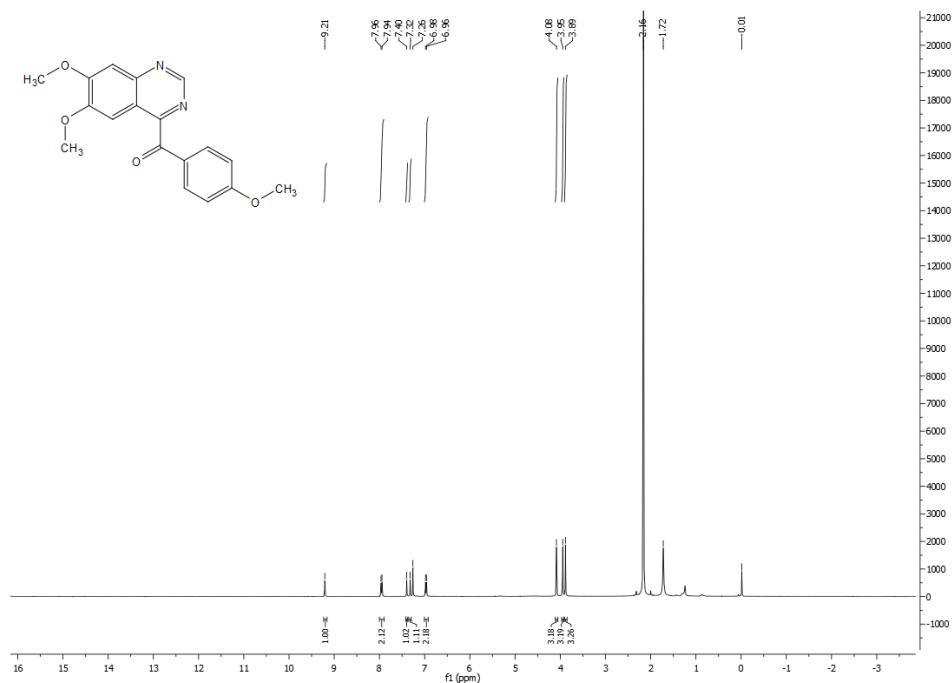
Section S9. ¹H NMR and ¹³C NMR spectrums of the designed PDE10 inhibitors

Fig. S10. ^1H -NMR spectrum of compound **LHB-2**

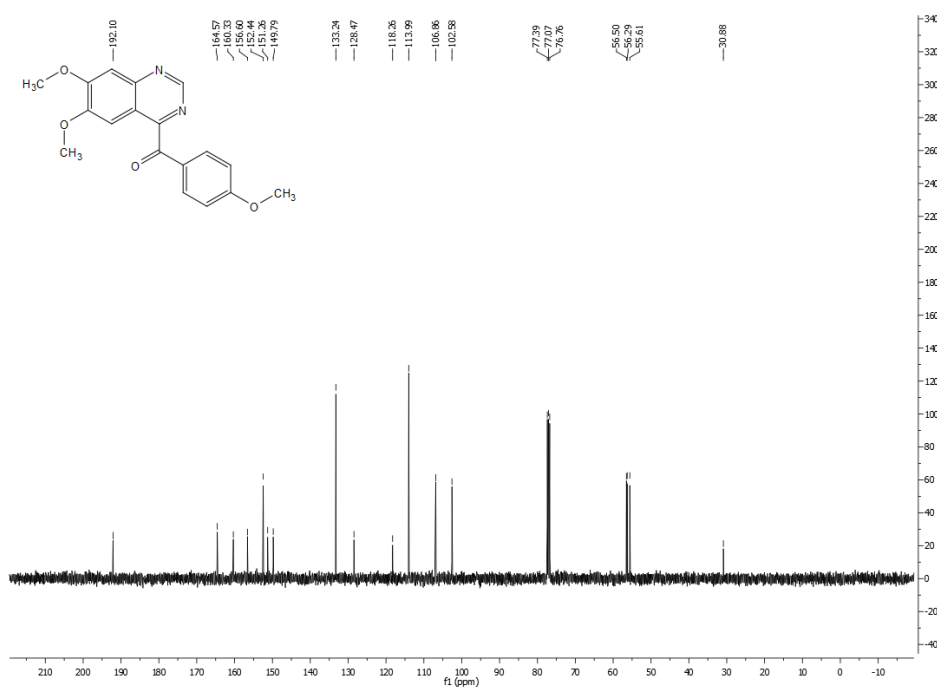


Fig. S11. ^{13}C -NMR spectrum of compound **LHB-2**

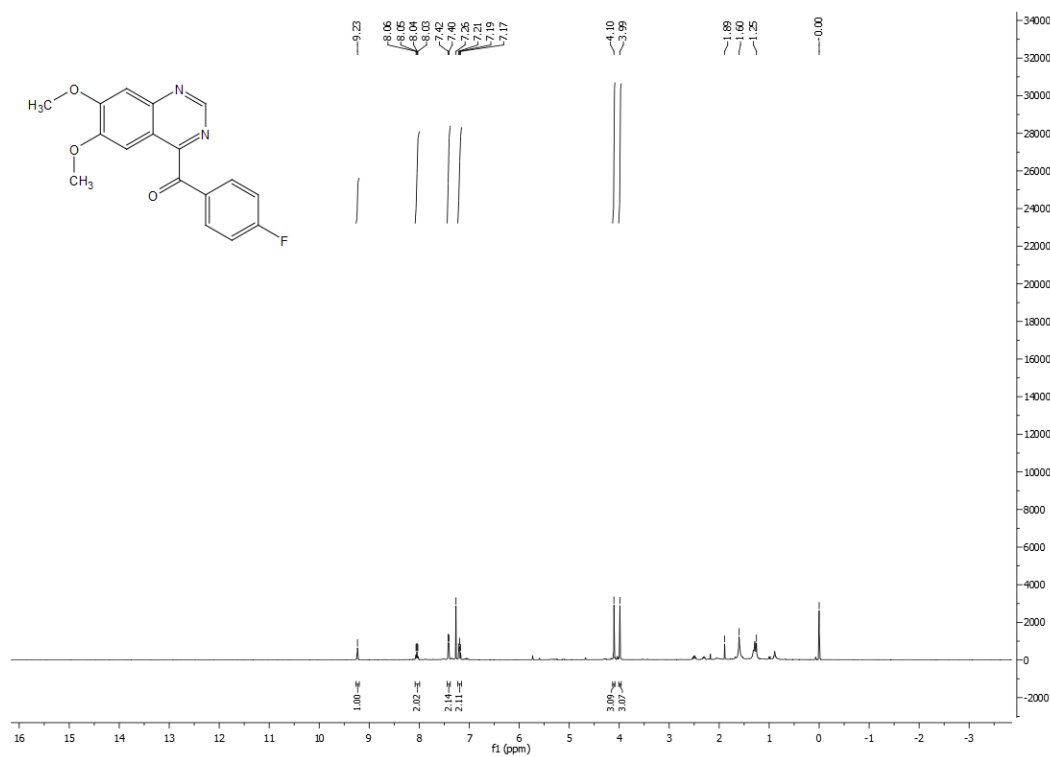


Fig. S12. ¹H-NMR spectrum of compound **LHB-3**

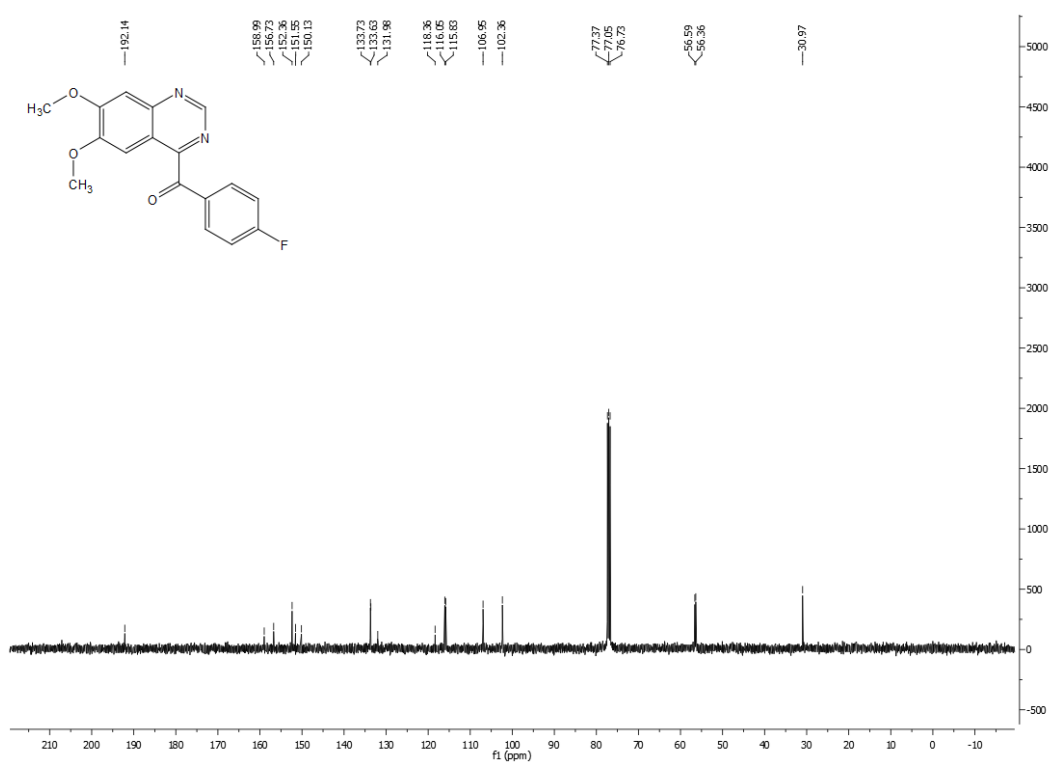


Fig. S13. ¹³C-NMR spectrum of compound **LHB-3**

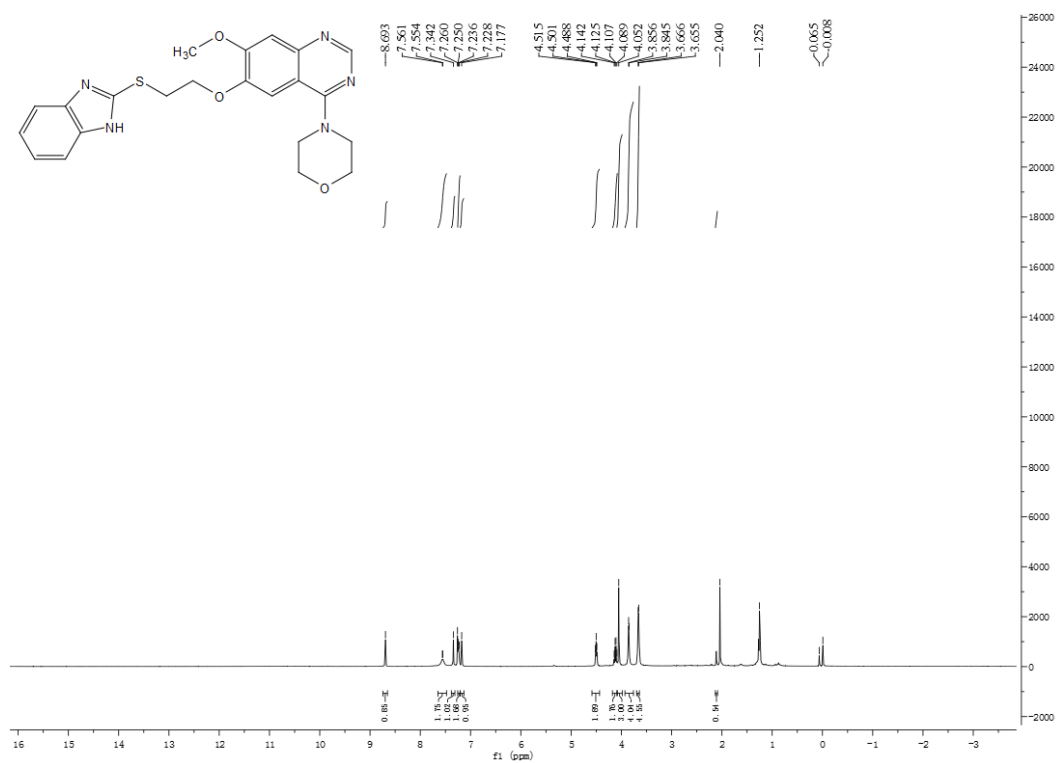


Fig. S14. ¹H-NMR spectrum of compound **LHB-4**

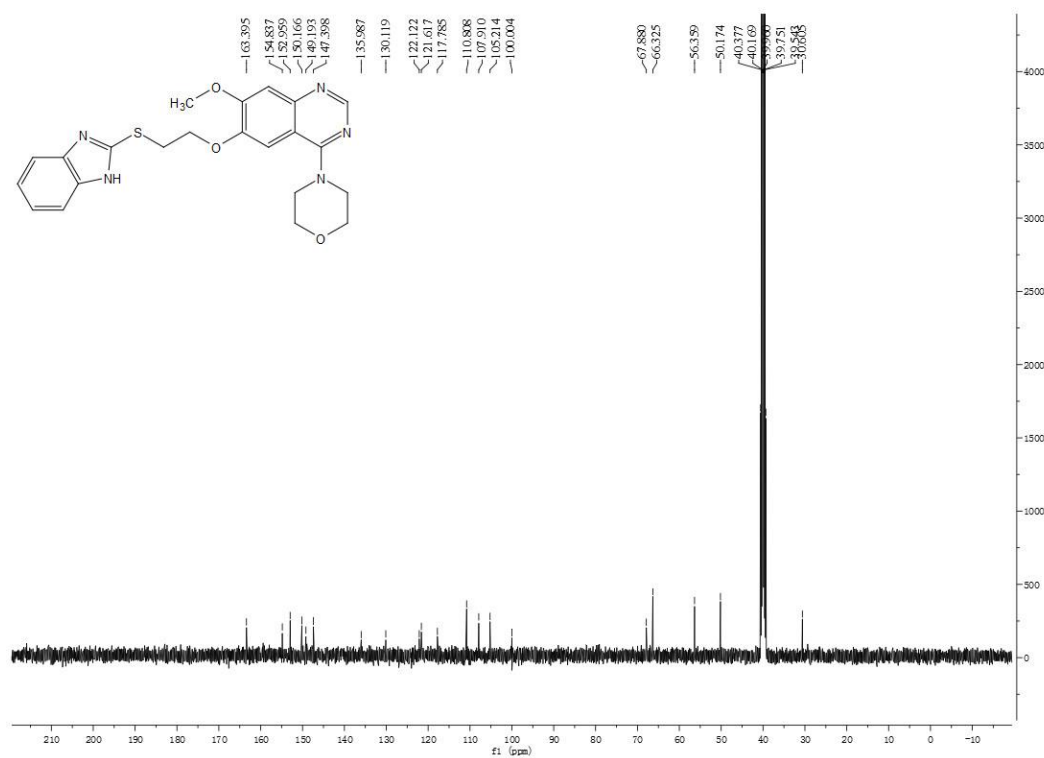
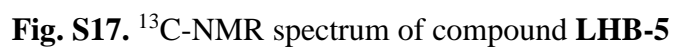
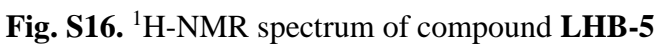


Fig. S15. ¹³C-NMR spectrum of compound **LHB-4**



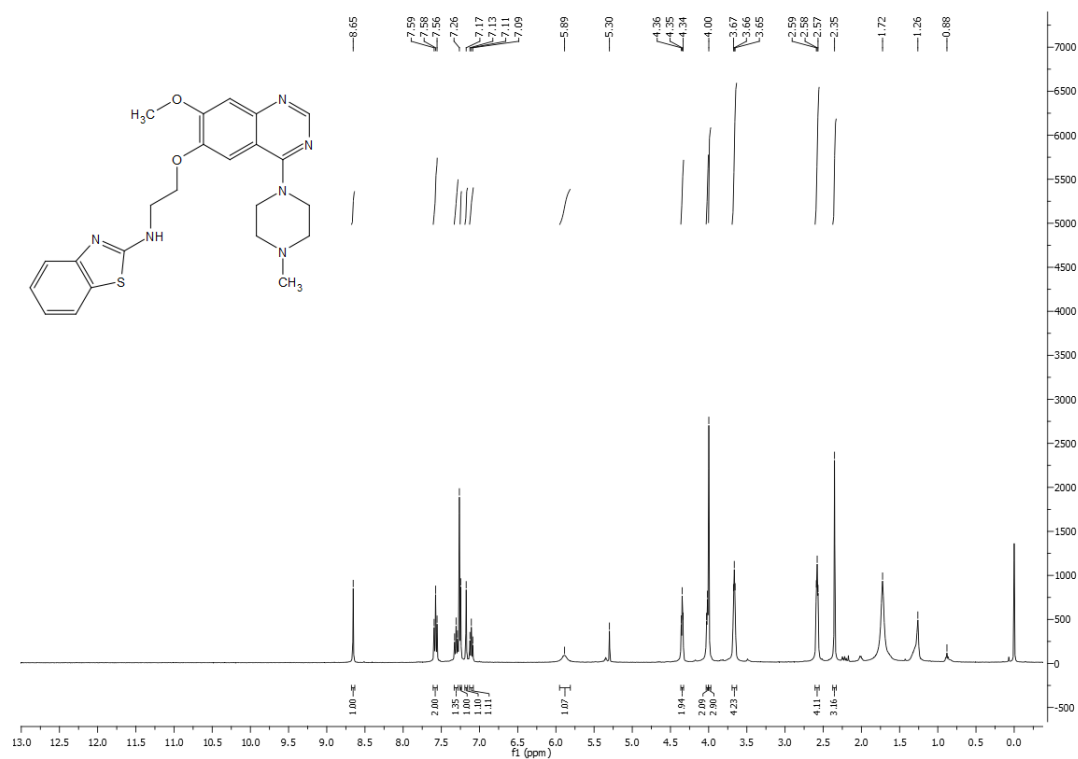


Fig. S20. ¹³C-NMR spectrum of compound **LHB-7**

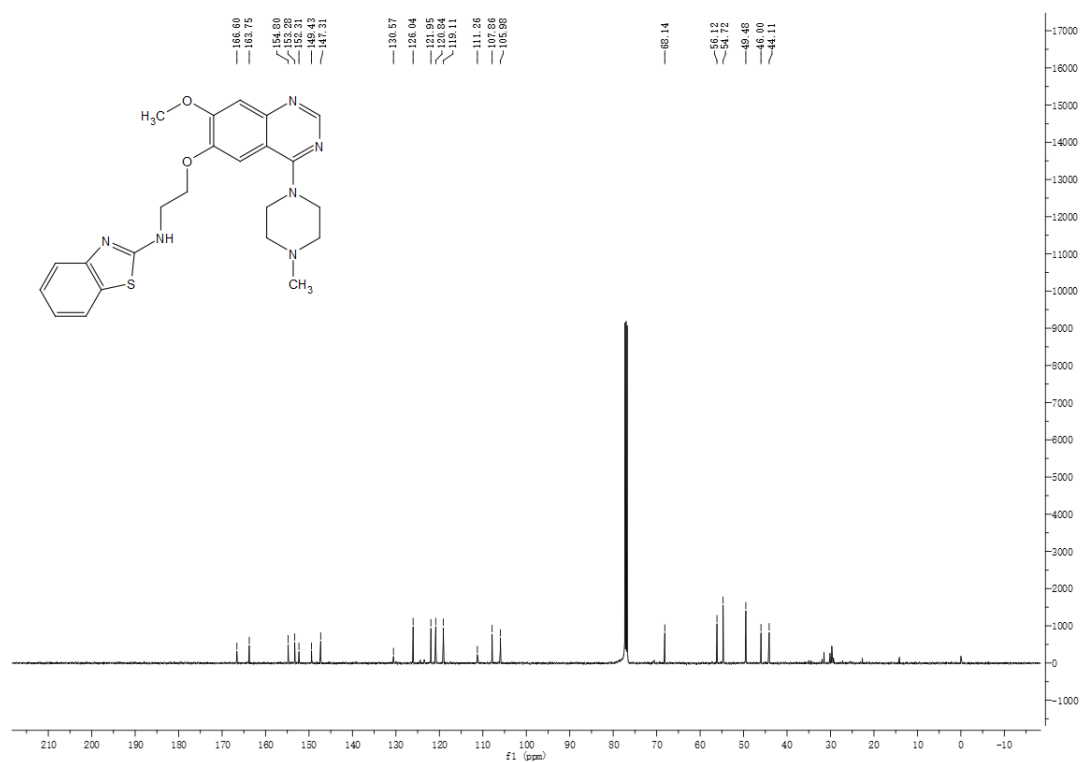
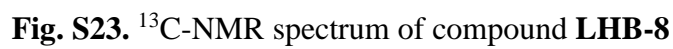
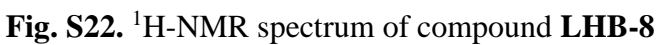
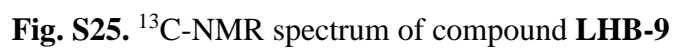
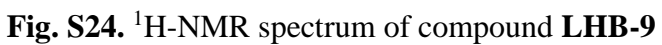


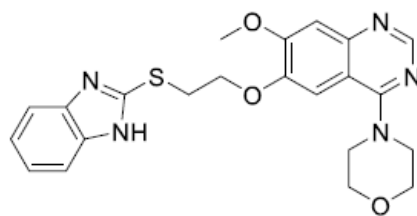
Fig. S21. ¹³C-NMR spectrum of compound **LHB-7**



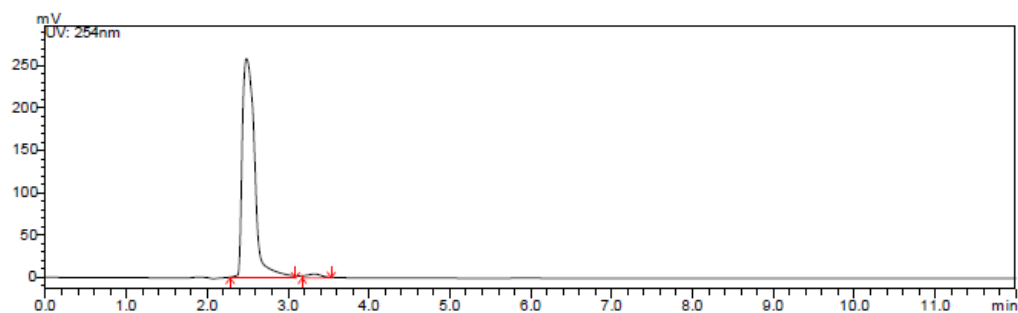


Section S10. HPLC spectrum for the purity of representative target compounds

SHIMADZU LC-20AT (column, Hypersil BDS C18, 5.0 μ m, 4.6 mm \times 150 mm (Elite); detector, SPD-20A UV/vis detector, UV detection at 254 nm; elution, MeOH in water (50%,v/v); T=25°C; flow rate = 1.0 mL/min).



LHB-4

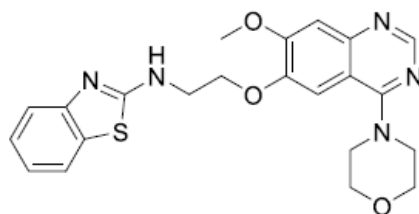


| Peak# | Retention Time | Peak area | Peak height | Peak area% |
|-------|----------------|-----------|-------------|------------|
| 1 | 2.481 | 2780685 | 259226 | 97.8756 |
| 2 | 3.320 | 60355 | 4508 | 2.1244 |

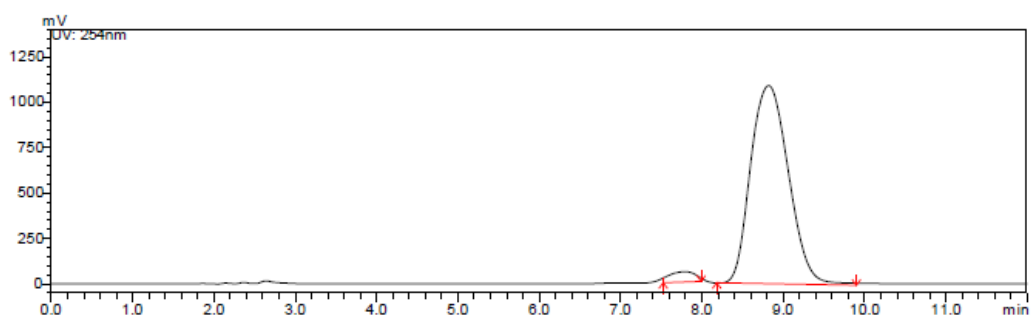
Purity: 97.9%

Fig. S28. HPLC spectrum for the purity of **LHB-4**

SHIMADZU LC-20AT (column, Hypersil BDS C18, 5.0 μm , 4.6 mm \times 150 mm (Elite); detector, SPD-20A UV/vis detector, UV detection at 254 nm; elution, MeOH in water (50%,v/v); T=25°C; flow rate = 1.0 mL/min).



LHB-6

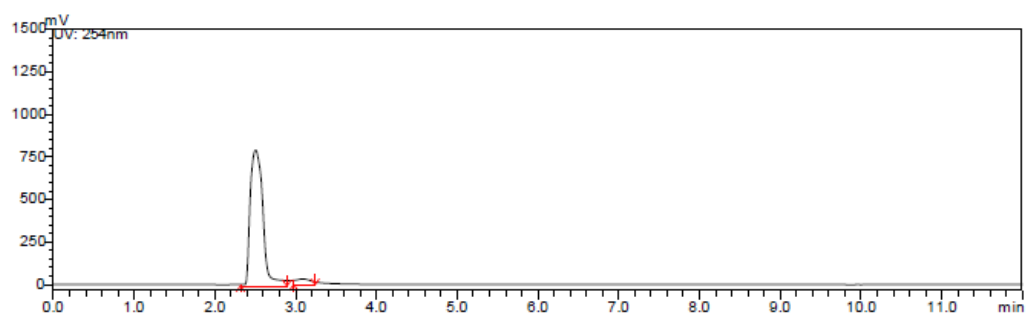
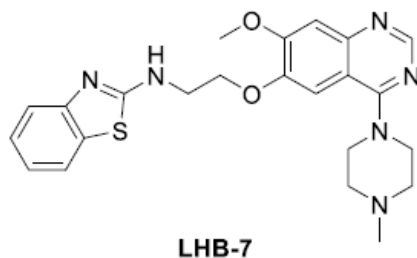


| Peak# | Retention Time | Peak area | Peak height | Peak area% |
|-------|----------------|-----------|-------------|------------|
| 1 | 7.788 | 1255231 | 56914 | 3.4254 |
| 2 | 8.817 | 35389720 | 1091410 | 96.5746 |

Purity: 96.6%

Fig. S29. HPLC spectrum for the purity of **LHB-6**

SHIMADZU LC-20AT (column, Hypersil BDS C18, 5.0 μm , 4.6 mm \times 150 mm (Elite); detector, SPD-20A UV/vis detector, UV detection at 254 nm; elution, MeOH in water (50%,v/v); T=25°C; flow rate = 1.0 mL/min).

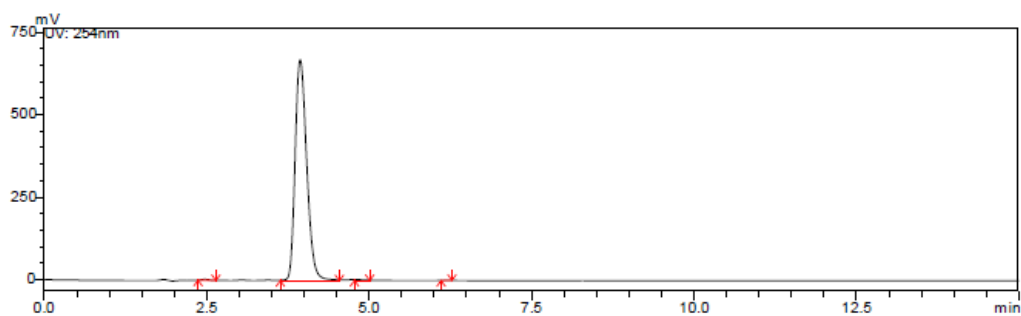
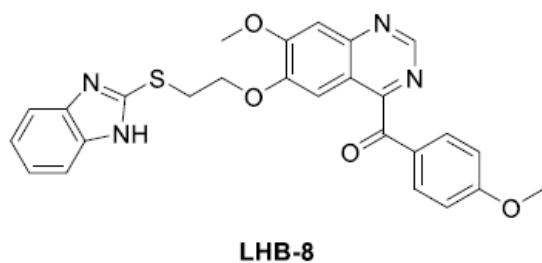


| Peak# | Retention Time | Peak area | Peak height | Peak area% |
|-------|----------------|-----------|-------------|------------|
| 1 | 2.500 | 8769921 | 797416 | 95.3530 |
| 2 | 3.085 | 427404 | 33065 | 4.6470 |

Purity: 95.4%

Fig. S30. HPLC spectrum for the purity of **LHB-7**

SHIMADZU LC-20AT (column, Hypersil BDS C18, 5.0 μ m, 4.6 mm \times 150 mm (Elite); detector, SPD-20A UV/vis detector, UV detection at 254 nm; elution, MeOH in water (60%,v/v); T=25°C; flow rate = 1.0 mL/min).

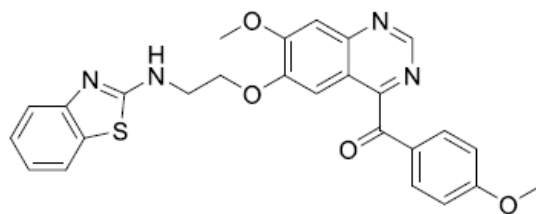


| Peak# | Retention Time | Peak area | Peak height | Peak area% |
|-------|----------------|-----------|-------------|------------|
| 1 | 2.468 | 24300 | 2893 | 0.2897 |
| 2 | 3.935 | 8334977 | 669950 | 99.3730 |
| 3 | 4.782 | 26419 | 2668 | 0.3150 |
| 4 | 6.181 | 1874 | 319 | 0.0223 |

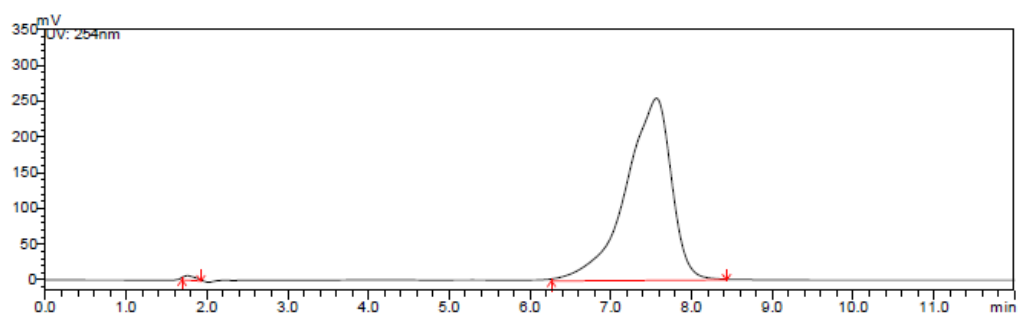
Purity: 99.4%

Fig. S31. HPLC spectrum for the purity of **LHB-8**

SHIMADZU LC-20AT (column, Hypersil BDS C18, 5.0 μ m, 4.6 mm \times 150 mm (Elite); detector, SPD-20A UV/vis detector, UV detection at 254 nm; elution, MeOH in water (55%,v/v); T=25°C; flow rate = 1.0 mL/min).



LHB-10



| Peak# | Retention Time | Peak area | Peak height | Peak area% |
|-------|----------------|-----------|-------------|------------|
| 1 | 1.756 | 65065 | 6282 | 0.6450 |
| 2 | 7.562 | 10022295 | 254729 | 99.3550 |

Purity: 99.4%

Fig. S32. HPLC spectrum for the purity of **LHB-10**

Section S11. High-resolution mass spectra (HRMS) spectrums of representative compounds

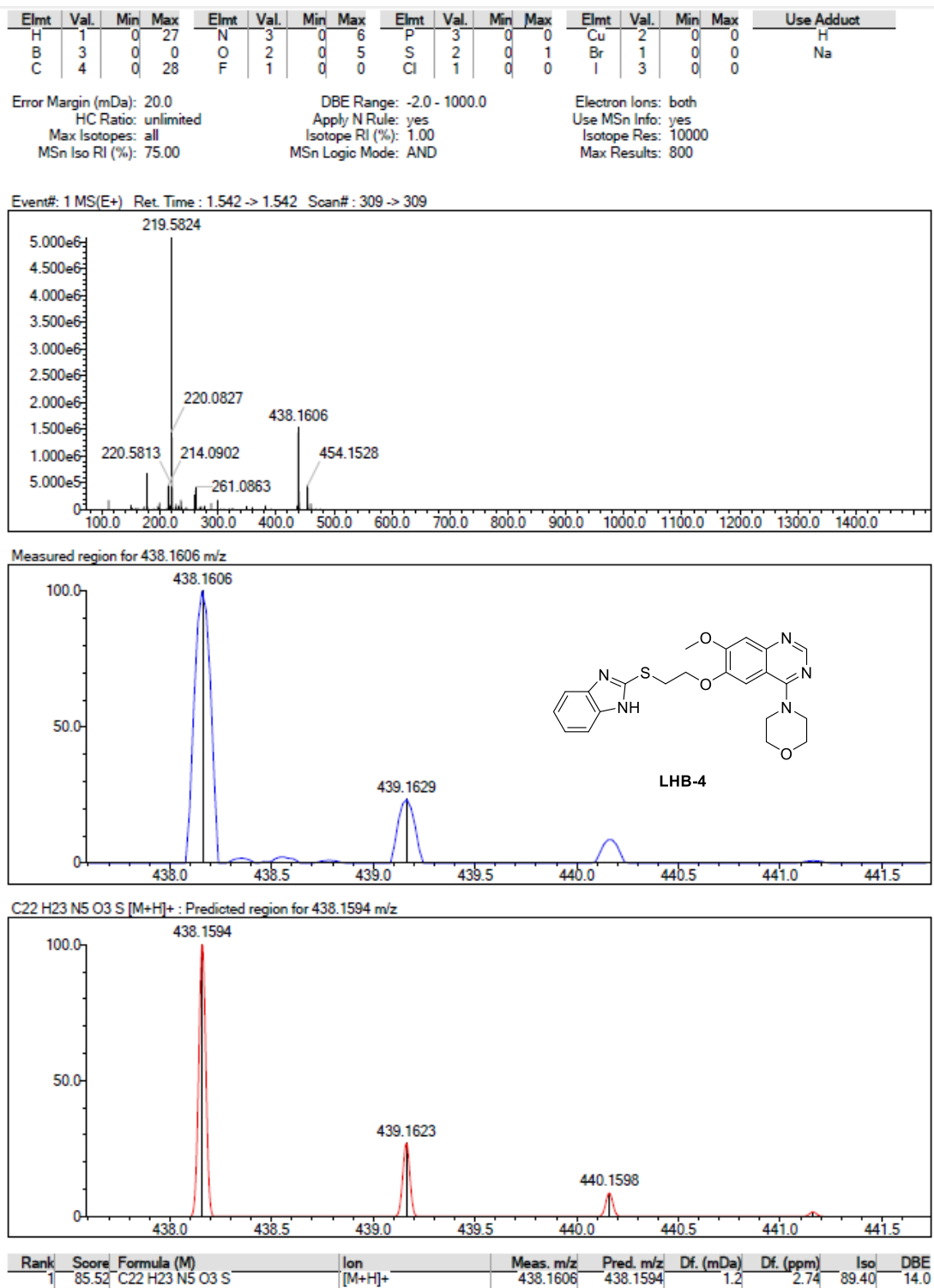


Fig. S33. HRMS spectrum of **LHB-4**

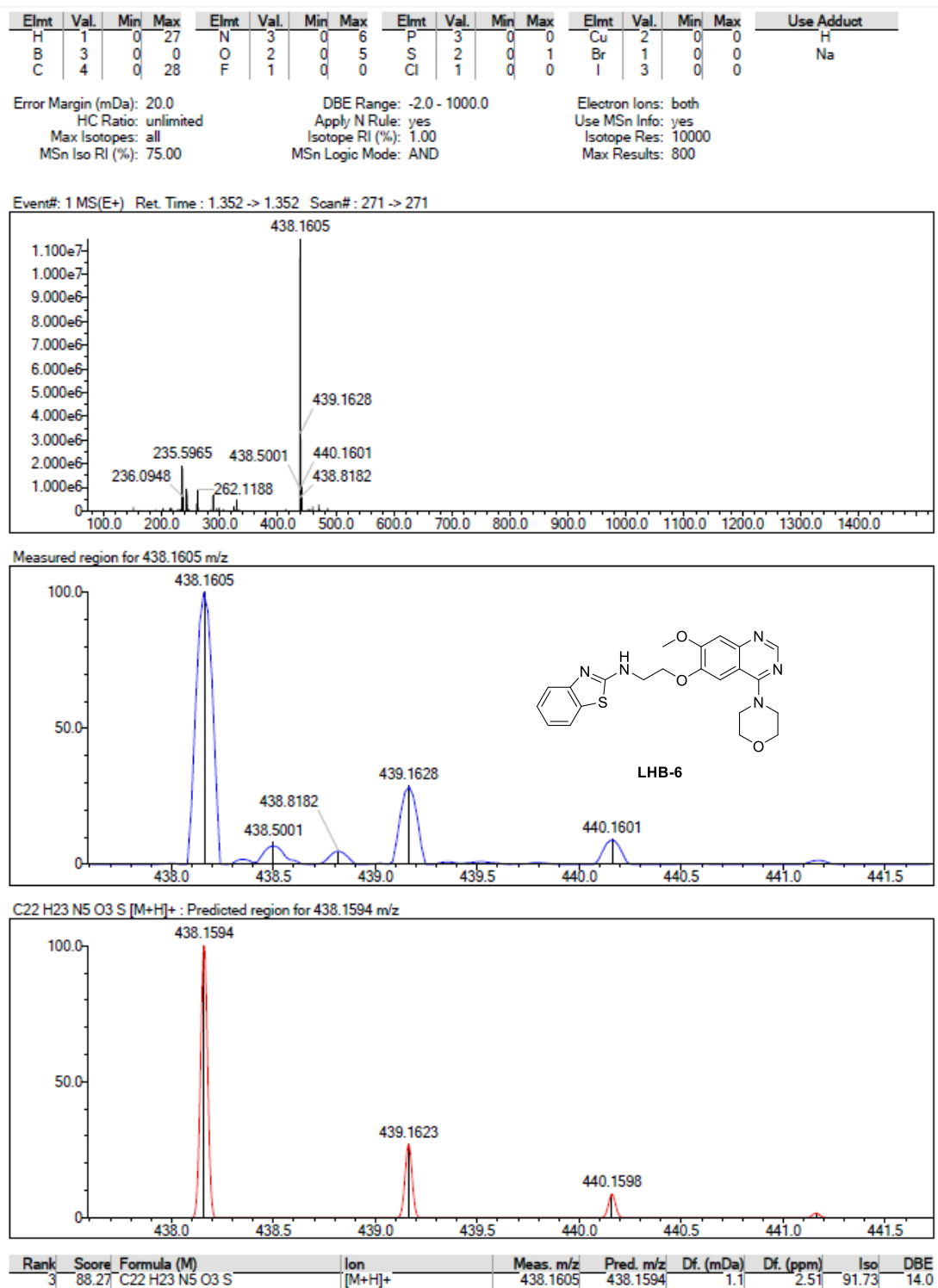


Fig. S34. HRMS spectrum of **LHB-6**

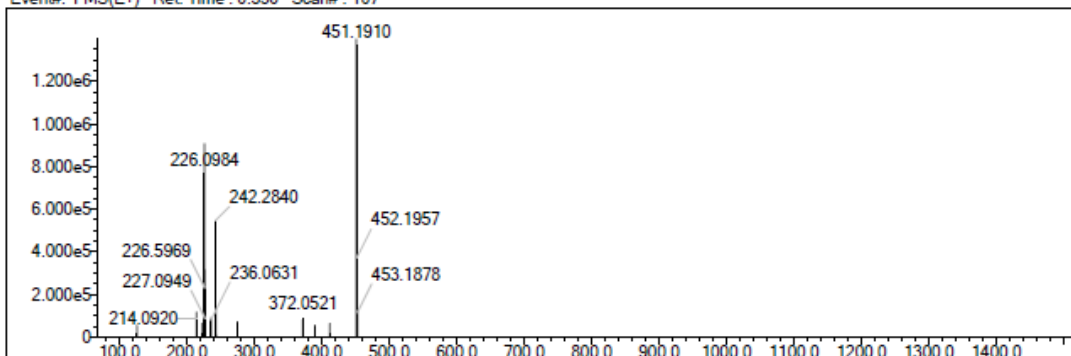
| Elmt | Val. | Min | Max | Elmt | Val. | Min | Max | Elmt | Val. | Min | Max | Elmt | Val. | Min | Max | Use Adduct |
|------|------|-----|-----|------|------|-----|-----|------|------|-----|-----|------|------|-----|-----|------------|
| H | 1 | 15 | 48 | O | 2 | 0 | 3 | K | 1 | 0 | 0 | Pd | 2 | 0 | 0 | H |
| 2H | 1 | 0 | 0 | F | 1 | 0 | 0 | Mn | 3 | 0 | 0 | I | 3 | 0 | 0 | Na |
| 3H | 1 | 0 | 0 | Si | 4 | 0 | 0 | Fe | 2 | 0 | 0 | Pt | 2 | 0 | 0 | |
| B | 3 | 0 | 0 | P | 3 | 0 | 0 | Cu | 2 | 0 | 0 | Au | 1 | 0 | 0 | |
| C | 4 | 13 | 30 | S | 2 | 1 | 2 | Se | 2 | 0 | 0 | | | | | |
| N | 3 | 0 | 8 | Cl | 1 | 0 | 0 | Br | 1 | 0 | 0 | | | | | |

Error Margin (ppm): 100
 HC Ratio: 0.0 - 20.0
 Max Isotopes: all
 MSn Iso RI (%): 75.00

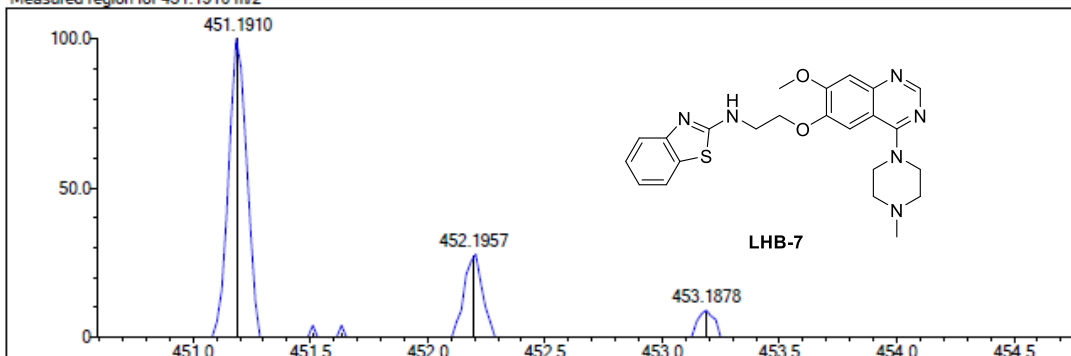
DBE Range: -100.0 - 200.0
 Apply N Rule: no
 Isotope RI (%): 1.00
 MSn Logic Mode: AND

Electron Ions: both
 Use MSn Info: no
 Isotope Res: 10000
 Max Results: 100

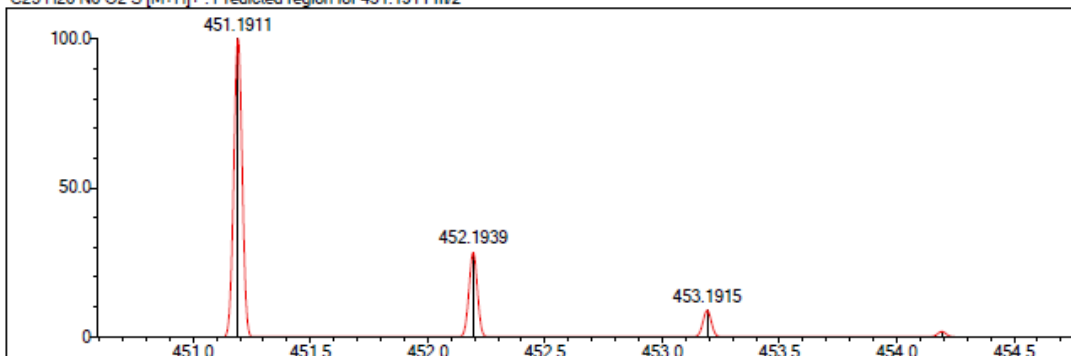
Event#: 1 MS(E+) Ret. Time : 0.530 Scan#: 107



Measured region for 451.1910 m/z



C23 H26 N6 O2 S [M+H]⁺ : Predicted region for 451.1911 m/z



| Rank | Score | Formula (M) | Ion | Meas. m/z | Pred. m/z | Df. (mDa) | Df. (ppm) | Iso | DBE |
|------|-------|-----------------|--------------------|-----------|-----------|-----------|-----------|-------|------|
| 3 | 84.14 | C23 H26 N6 O2 S | [M+H] ⁺ | 451.1910 | 451.1911 | -0.1 | -0.22 | 84.14 | 14.0 |

Fig. S35. HRMS spectrum of **LHB-7**

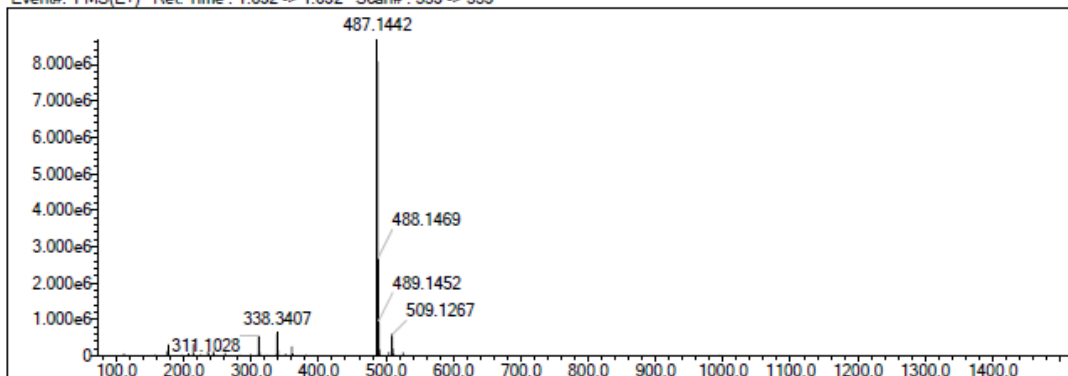
| Elmt | Val. | Min | Max | Elmt | Val. | Min | Max | Elmt | Val. | Min | Max | Elmt | Val. | Min | Max | Use Adduct |
|------|------|-----|-----|------|------|-----|-----|------|------|-----|-----|------|------|-----|-----|------------|
| H | 1 | 0 | 27 | N | 3 | 0 | 6 | P | 3 | 0 | 0 | Cu | 2 | 0 | 0 | H |
| B | 3 | 0 | 0 | O | 2 | 0 | 5 | S | 2 | 0 | 2 | Br | 1 | 0 | 0 | Na |
| C | 4 | 0 | 28 | F | 1 | 0 | 1 | Cl | 1 | 0 | 0 | I | 3 | 0 | 0 | |

Error Margin (mDa): 20.0
 HC Ratio: unlimited
 Max Isotopes: all
 MSn Iso RI (%): 75.00

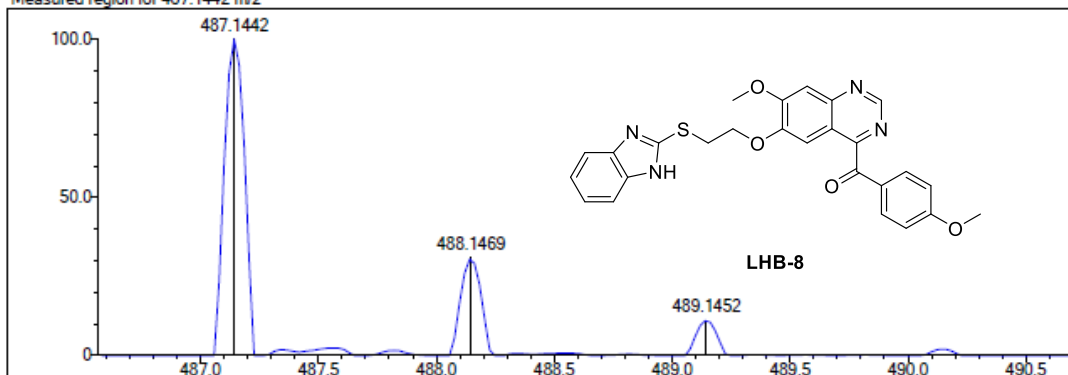
DBE Range: -2.0 - 1000.0
 Apply N Rule: yes
 Isotope RI (%): 1.00
 MSn Logic Mode: AND

Electron Ions: both
 Use MSn Info: yes
 Isotope Res: 10000
 Max Results: 800

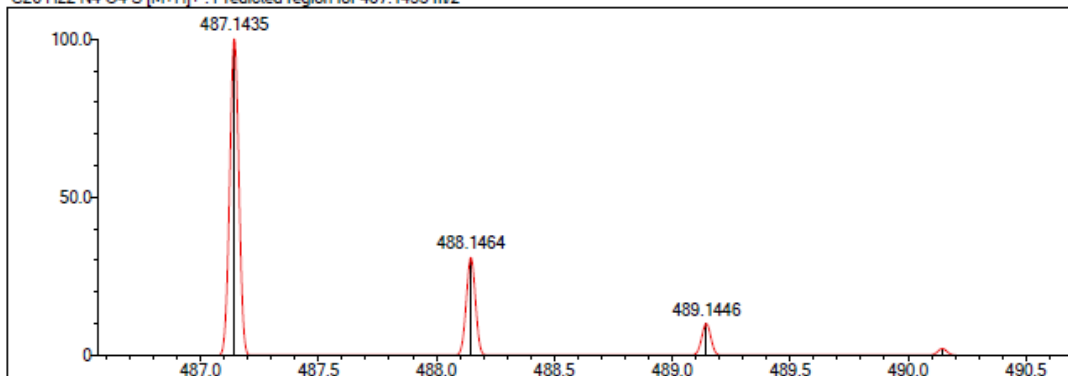
Event#: 1 MS(E+) Ret. Time: 1.692 -> 1.692 Scan#: 339 -> 339



Measured region for 487.1442 m/z



C26 H22 N4 O4 S [M+H]⁺: Predicted region for 487.1435 m/z



| Rank | Score | Formula (M) | Ion | Meas. m/z | Pred. m/z | Df. (mDa) | Df. (ppm) | Iso | DBE |
|------|-------|-----------------|--------------------|-----------|-----------|-----------|-----------|--------|------|
| 2 | 98.90 | C26 H22 N4 O4 S | [M+H] ⁺ | 487.1442 | 487.1435 | 0.7 | 1.44 | 100.00 | 18.0 |

Fig. S36. HRMS spectrum of **LHB-8**

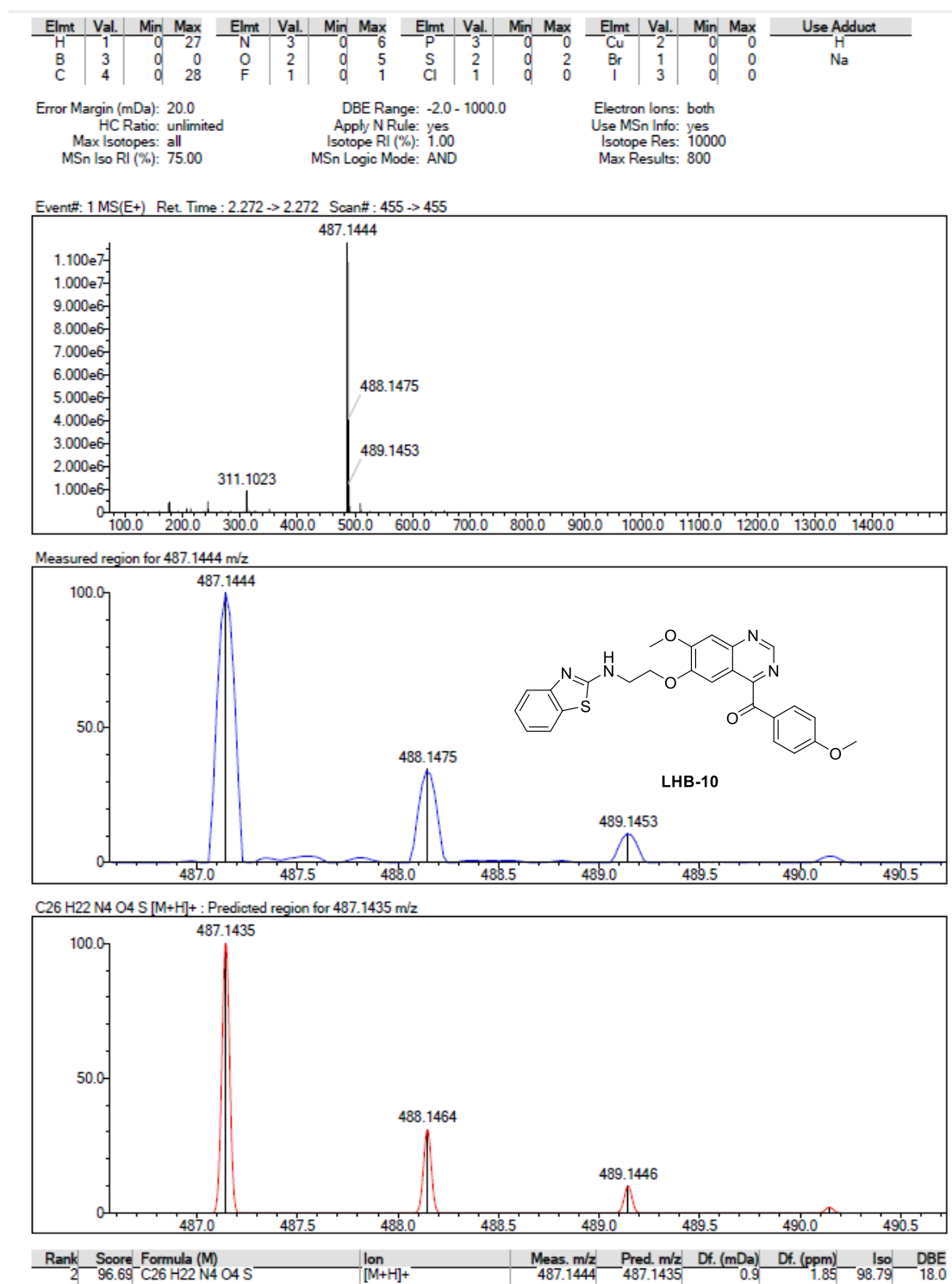


Fig. S37. HRMS spectrum of **LHB-10**

Supporting References

1. Neuhaus J-M, Sitcher L, Meins F, Jr, Boller T (1991) A short C-terminal sequence is necessary and sufficient for the targeting of chitinases to the plant vacuole. *Proc Natl Acad Sci USA* 88:10362–10366.
2. Bennett CH (1976) Efficient estimation of free energy differences from Monte Carlo data. *J Comput Phys* 22(2):245-268.
3. Shirts MR, Chodera JD (2008) Statistically optimal analysis of samples from multiple equilibrium states. *J Chem Phys* 129(12):124105.
4. Pitera JW, van Gunsteren WF (2002) A comparison of non-bonded scaling approaches for free energy calculations. *Mol Simul* 28(1-2):45-65.