

**Vasodilator Hydralazine Promotes Nanoparticle Penetration in Advanced Desmoplastic Tumor**

Yanzuo Chen,<sup>†, ‡</sup> Wantong Song,<sup>†, §, \*</sup> Limei Shen,<sup>†</sup> Nasha Qiu,<sup>†, ||</sup> Mengying Hu,<sup>†</sup> Yun Liu,<sup>†</sup> Qi Liu<sup>†</sup> and Leaf Huang<sup>†, \*</sup>

<sup>†</sup> Division of Pharmacoengineering and Molecular Pharmaceutics and Center for Nanotechnology in Drug Delivery,

Eshelman School of Pharmacy, University of North Carolina at Chapel Hill, Chapel Hill, North Carolina 27599, United States

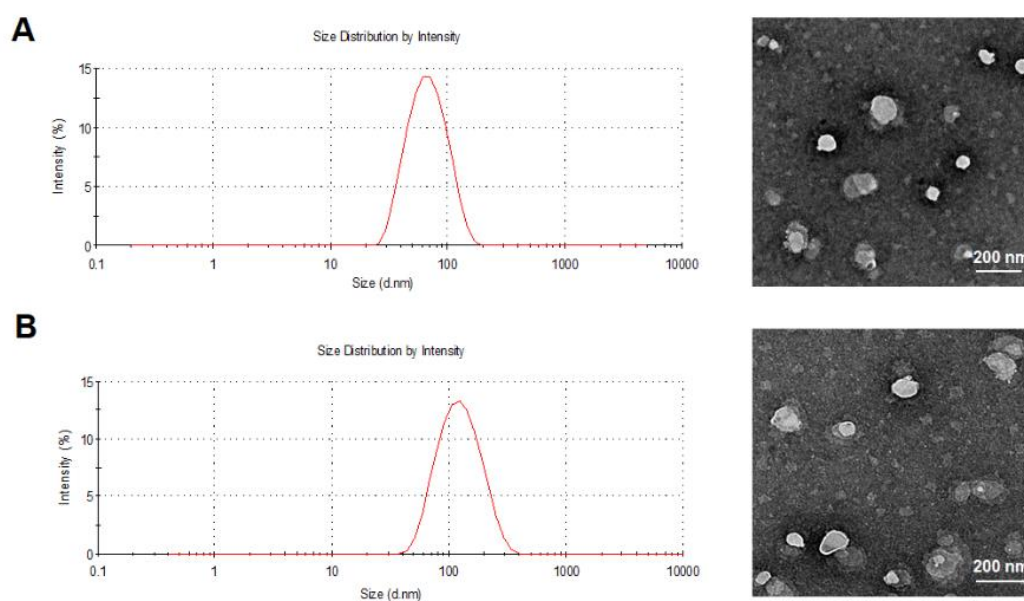
<sup>‡</sup> Department of Pharmaceutics, School of Pharmacy, East China University of Science and Technology, Shanghai 200237, China

<sup>§</sup> Key Laboratory of Polymer Ecomaterials, Changchun Institute of Applied Chemistry, Chinese Academy of Sciences, Changchun 130022, China

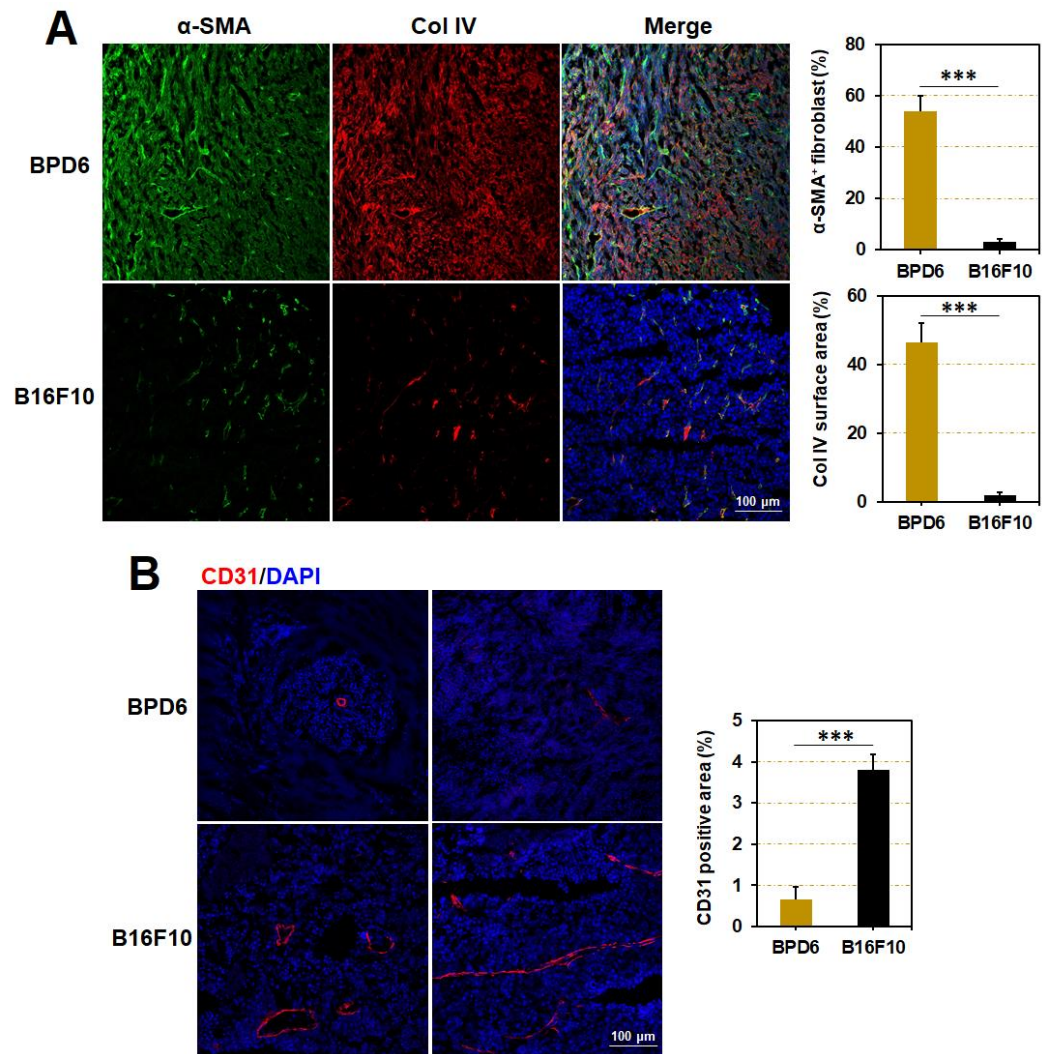
<sup>||</sup> Institute of Pharmaceutics, College of Pharmaceutical Sciences, Zhejiang University, Hangzhou 310058, China

\*Corresponding authors: wtsong@ciac.ac.cn, leafh@email.unc.edu

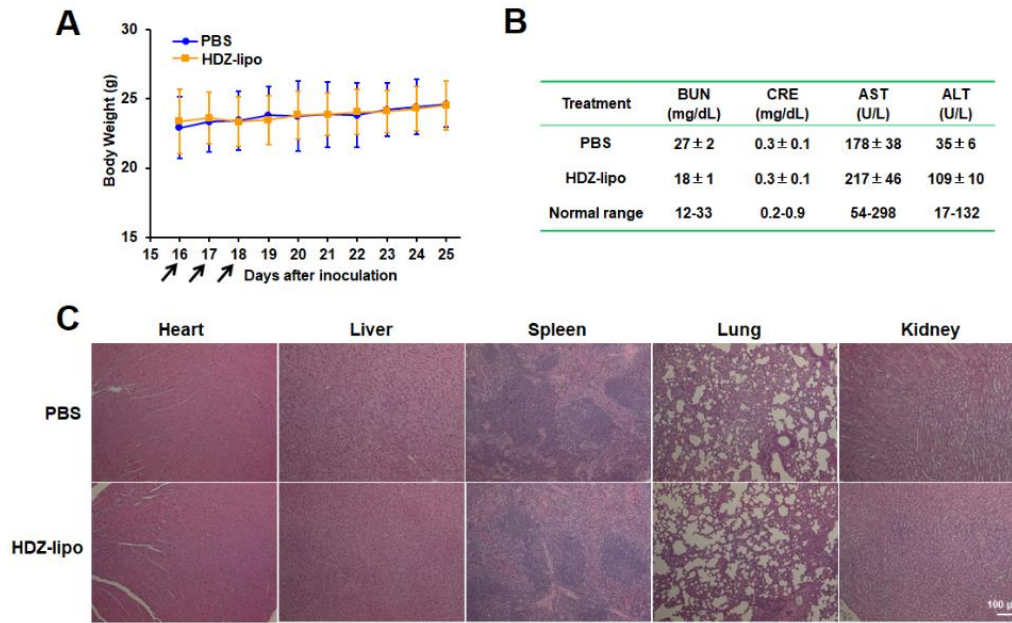
## Supporting Information



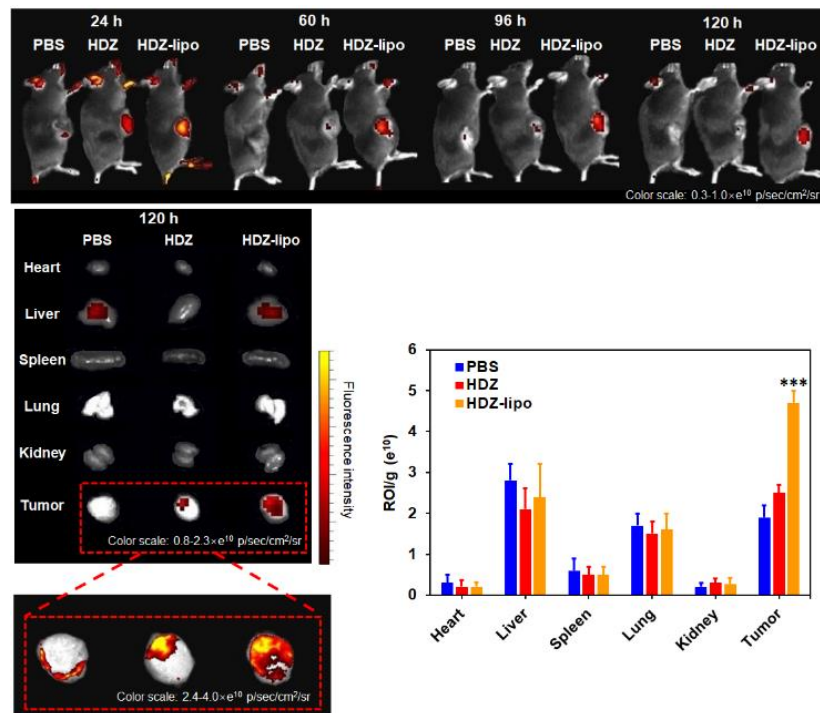
**Figure S1. DLS and TEM results of the HDZ-liposomes.** DLS and TEM depicting size and morphology of (A) empty-liposomes after extrusion ( $\sim 75$  nm), and (B) HDZ-liposomes after 30 days' storage ( $\sim 101$  nm), respectively.



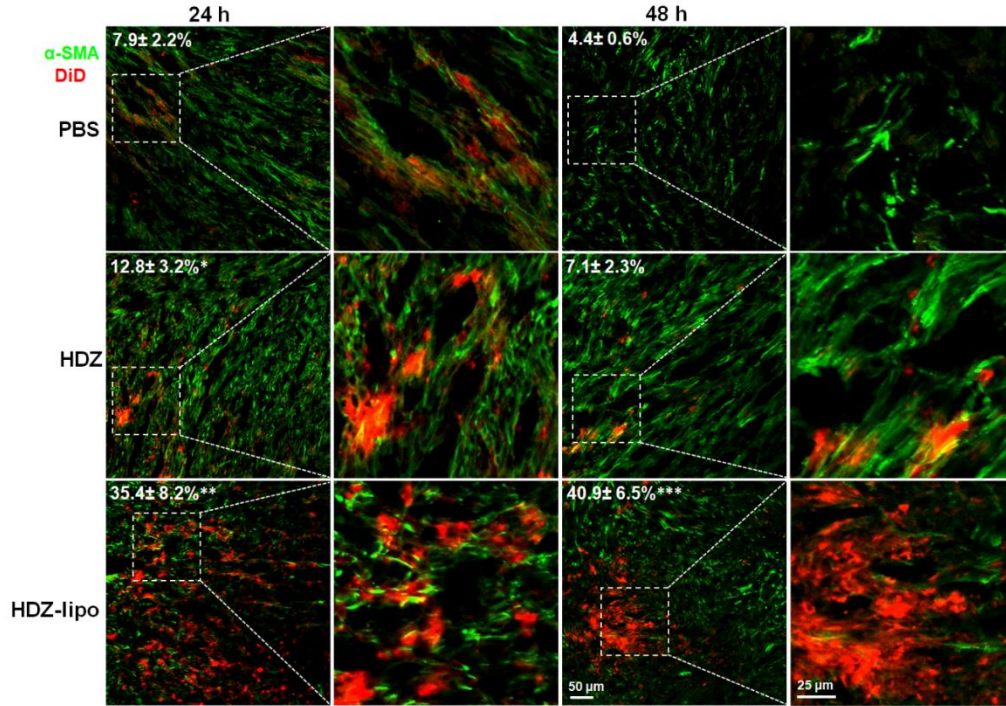
**Figure S2. Histological analysis of desmoplastic (BPD6) and wide-type (B16F10) melanoma tumors.** (A) Immunostaining of  $\alpha$ -SMA (green), collagen IV (red), and DAPI (blue). (B) Immunostaining of CD31 positive (red) vessels. n = 5; \*p < 0.05, \*\* p < 0.01, \*\*\* p < 0.001.



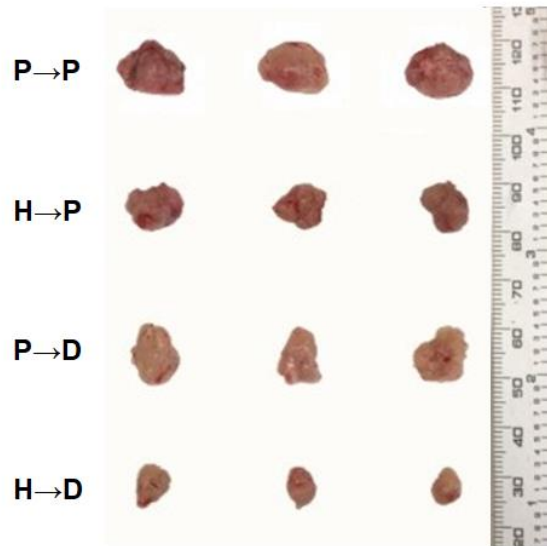
**Figure S3. Toxicity evaluation of HDZ-liposomes.** (A) BPD6 tumor-bearing mice body weight changes after three i.v. injections of HDZ-liposomes (black arrows). (B) Serum BUN, CRE, AST and ALT levels. (C) Pathological analyses of heart, liver, spleen, lung and kidney of the BPD6 tumor-bearing mice in the PBS and HDZ-liposome treated groups 7 days post injection.



**Figure S4. *In vivo* and *ex vivo* images of DiD-loaded liposomes in BPD6 tumor-bearing mice at 24 to 120 h post injection.** Mice were pre-treated with PBS, HDZ and HDZ-liposomes, respectively. n = 3 mice per group; \*p < 0.05, \*\*p < 0.01, \*\*\*p < 0.001, compared to PBS group.

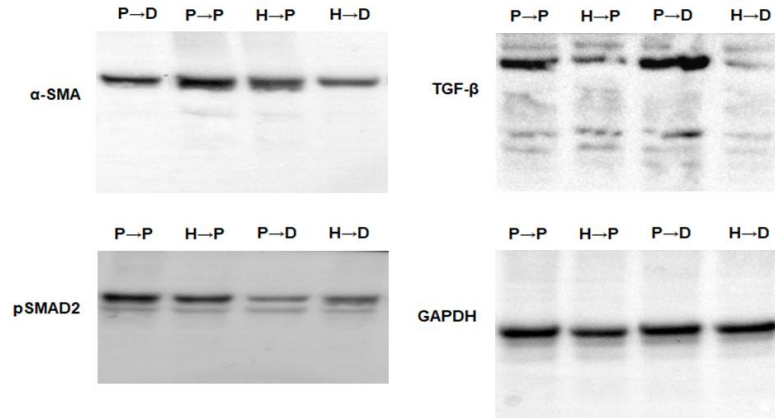


**Figure S5.** DiD-loaded liposome distribution in TAFs ( $\alpha$ -SMA staining) at 24 and 48 h post injection. Mice were pre-treated with PBS, HDZ, and HDZ-liposomes, respectively.  $n = 5$ . \* $p < 0.05$ , \*\* $p < 0.01$ , \*\*\* $p < 0.001$ , compared to PBS group.

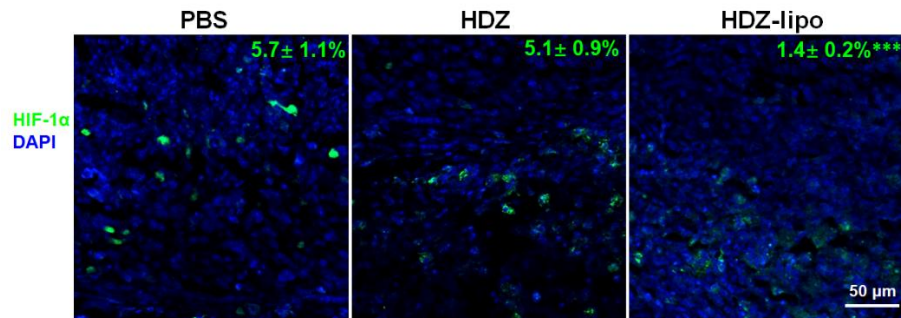


**Figure S6.** Representative *ex vivo* images of BPD6 tumors after various treatments. PBS followed by PBS (P→P), HDZ-liposomes followed by PBS (H→P), PBS followed by DOX-liposomes (P→D) and HDZ-liposomes followed by DOX-liposomes (H→D).

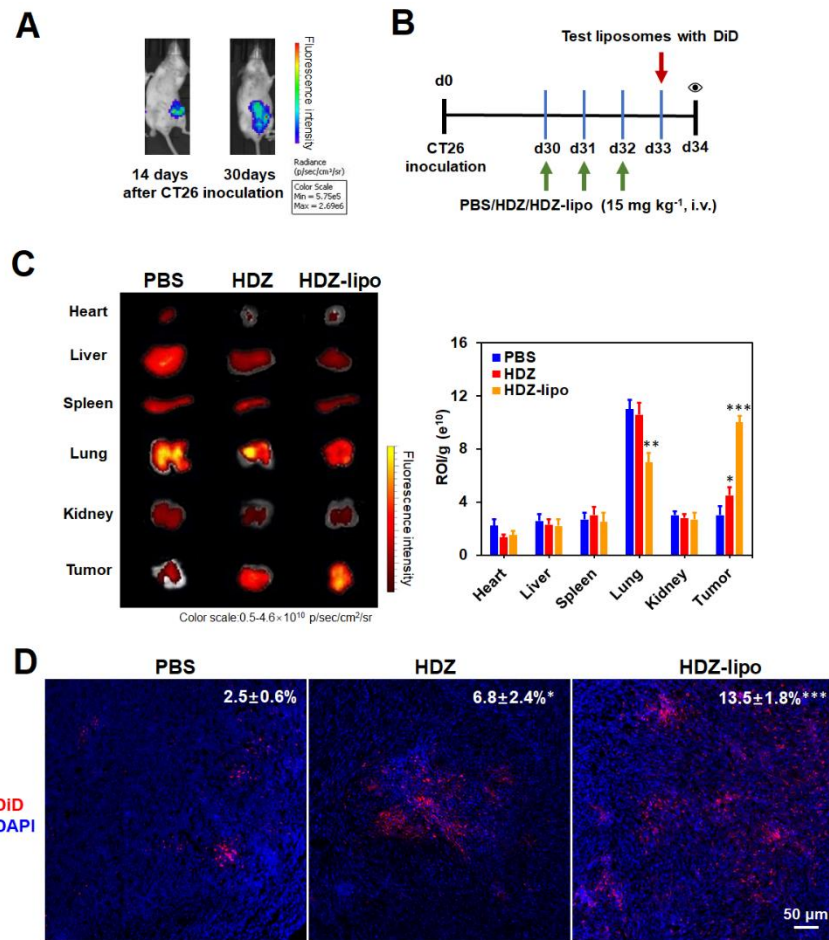




**Figure S7.** Original photographs of western blotting data in Figure 6E.



**Figure S8.** Immunofluorescence staining of HIF-1 $\alpha$  in 4T1 tumor after various treatments. n = 5. \*p < 0.05, \*\*p < 0.01, \*\*\*p < 0.001, compared to PBS group.



**Figure S9. HDZ-liposomes promote the nanoparticle penetration in orthotopic CT26-FL3 tumor model.** (A) Advanced orthotopic colorectal tumors establishment on day 30 after CT26-FL3 cells inoculated into the mouse cecum wall. The tumor burden was monitored by bioluminescent analysis. (B) Treatment schedule. (C) *Ex vivo* IVIS image and quantitative analysis of DiD-loaded liposomes in tumor and major organs at 24 h post injection; n = 3 mice per group. (D) Distribution of HDZ-liposomes inside tumor in various treatment groups; n = 5. \*p < 0.05, \*\*p < 0.01, \*\*\*p < 0.001, compared to PBS group.

**Table S1.** Antibodies Used in the Study

<b>Antibodies</b>	<b>Company</b>	<b>Catalog</b>	<b>Application</b>
Alexa Fluor®647 Anti-CD3	BioLegend	100209	IF
Alexa Fluor®488 Anti- $\alpha$ -SMA	Invitrogen	4332501	IF
Alexa Fluor®594 Anti-CD31	BioLegend	102432	IF
Anti-collagen IV	Abcam	Ab6586	IF
Anti-fibronectin	Abcam	Ab2413	IF
Anti-HIF-1 $\alpha$	Santa Cruz Biotechnology	SC-10790	IF
Goat Anti-Rabbit IgG, Alexa Fluor®488	Abcam	Ab150077	IF
Goat Anti-Rabbit IgG, Alexa Fluor®594	Abcam	Ab150080	IF
Anti-pSMAD2	Cell signaling	3108S	WB
Anti- $\alpha$ -SMA	Abcam	Ab124964	WB
Anti-TGF $\beta$	Santa Cruz Biotechnology	SC-146	WB
GAPDH	Santa Cruz Biotechnology	SC-25778	WB
Goat anti-rabbit HRP	Abcam	Ab205718	WB
FITC anti-CD8	BioLegend	100705	flow cyt
BV421 anti-CD4	BioLegend	100543	flow cyt
PE/Cy7 anti-CD3	BioLegend	100219	flow cyt
PE/Cy7 anti- NK1.1	BioLegend	552878	flow cyt
PE/Cy7 anti-CD11c	BioLegend	117317	flow cyt
PerCp anti-MHCII	BioLegend	107623	flow cyt
APC/Cy7 anti-CD11b	BioLegend	101225	flow cyt
APC anti-Gr-1	BioLegend	108411	flow cyt
PerCp/Cy5.5 anti-CD206	BioLegend	141715	flow cyt
PE anti-F4/80	BioLegend	123110	flow cyt



**Table S2.** Primers for real-time PCR used in this study

<b>Primer</b>	<b>Applied Biosystems</b>
Mouse GAPDH	Mm99999915_g1
Mouse CCL2	Mm00441242_m1
Mouse CCL5	Mm01302427_m1
Mouse IL-4	Mm00445259_m1
Mouse IL-10	Mm01288386_m1
Mouse IL-12a	Mm00434169_m1
Mouse IL-1 $\beta$	Mm00434228_m1
Mouse CXCL9	Mm00434946_m1
Mouse CXCL10	Mm00445235_m1
Mouse CXCL12	Mm00445553_m1
Mouse CXCL13	Mm01208154_g1
Mouse TNF- $\alpha$	Mm00443260_g1
Mouse IFN- $\gamma$	Mm01168134_m1
Mouse TGF- $\beta$	Mm01178820_m1