

SUPPORTING INFORMATION

A tandem mass spectrometry sequence database search method for identification of O-fucosylated proteins by mass spectrometry.

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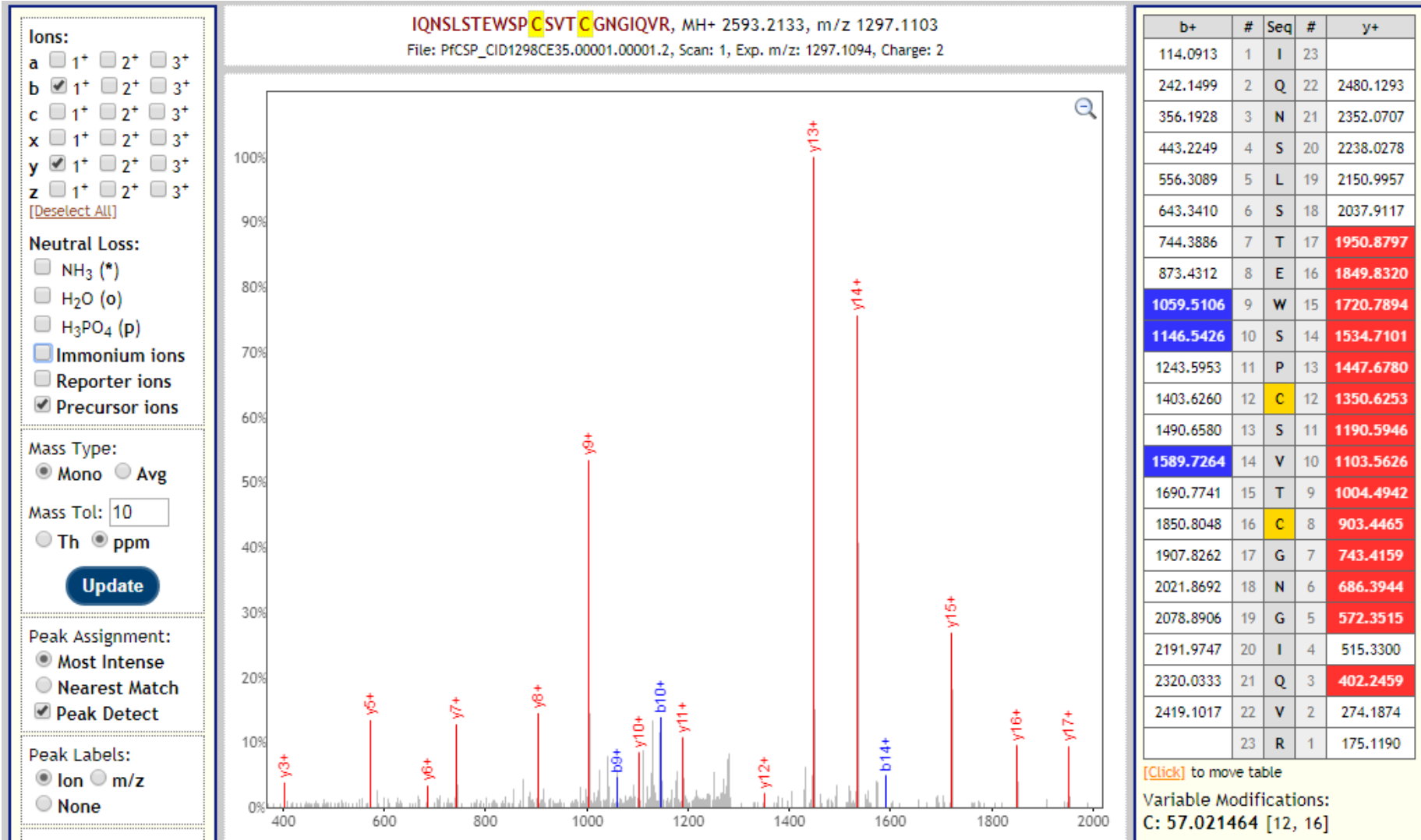
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Fig S15. Example false positive peptides showing neutral loss of O-linked glycans.

Figure S1. MSⁿ confirming the identity of fragment spectra arising from neutral loss of O-linked glycans. A tryptic digest of recombinant *P. falciparum* circumsporozoite protein (CSP) TSR domain was infused into an LTQ-Orbitrap Elite by nanospray. Precursor ions for the unmodified or O-glucosylfucosylated peptide were isolated and fragmented. All spectra are collision-induced dissociation (CID) collected at high resolution in the Orbitrap. CE = normalized collision energy. Annotated fragment spectra were visualized using a development version of Lorikeet as implemented in the Trans-Proteomic Pipeline.



S1a. CID of the unmodified peptide, [M+2H]⁺⁺ = 1297 *m/z*, CE = 35 %. Note that the dominant peaks are y₁₃ and y₁₄, the y-ion peaks containing the Pro residue. The y₁₃ and y₁₄ peaks are the only species visible in the spectra in Figure 1 still bearing the O-linked disaccharide.

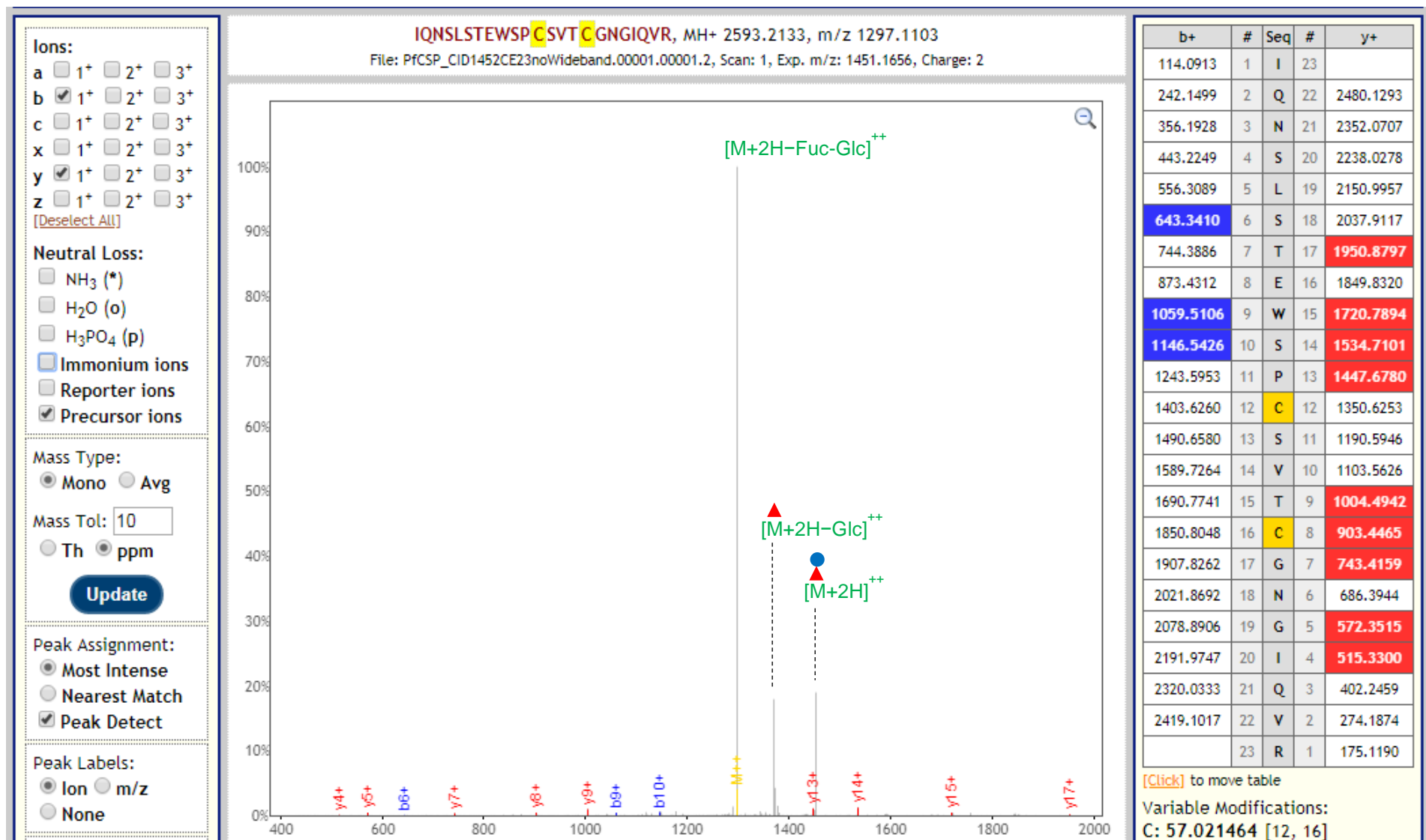


Figure S1b. CID of the peptide modified with O-Fuc-Glc, $[M+2H]^{++} = 1451.17$ m/z , CE = 23 %. At this CE, some of the precursor is still intact ($[M+2H]^{++}$, red triangle = O-Fuc, blue circle = β 1,3-Glc). Neutral loss of Glc can be seen as a peak at 1370.14 m/z ($[M+2H-Glc]^{++}$, red triangle = O-Fuc). The dominant peak at 1297.11 m/z is the precursor with neutral loss of the O-Fuc-Glc disaccharide ($[M+2H-Fuc-Glc]^{++}$). Peptide fragments are present at low abundance, identifying the sequence of the unmodified peptide.

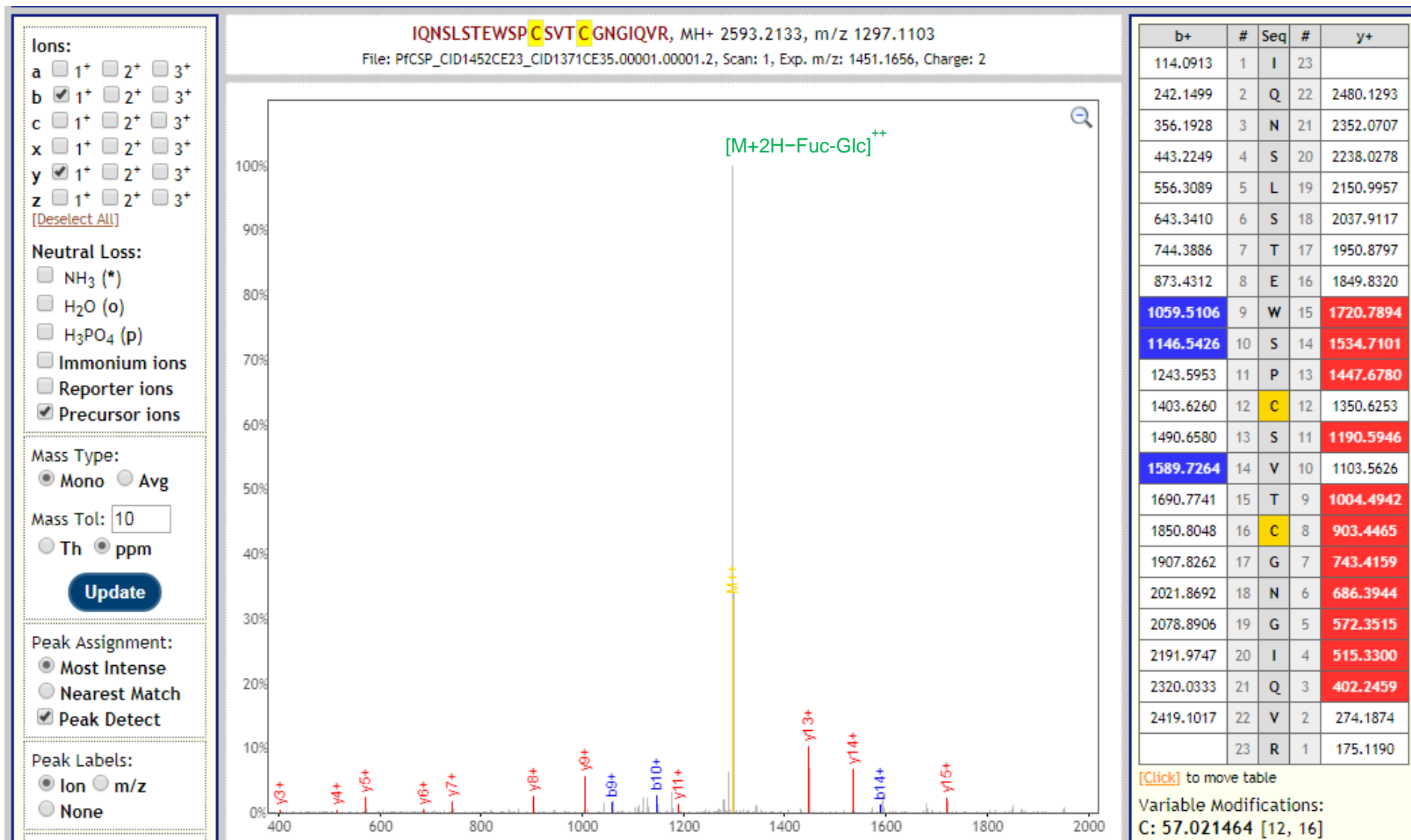


Figure S1c. MS³ (CE = 35 %) of the 1370.14 *m/z* fragment seen in Figure S1b produces a dominant peak at 1297.11 *m/z* matching the mass of the unmodified peptide. Many unmodified peptide fragment peaks confirm the sequence of the peptide. This spectrum confirms that the 1370.14 *m/z* peak seen in Figure S1b arises from neutral loss of Glc from the O-Fuc-Glc dissaccharide.

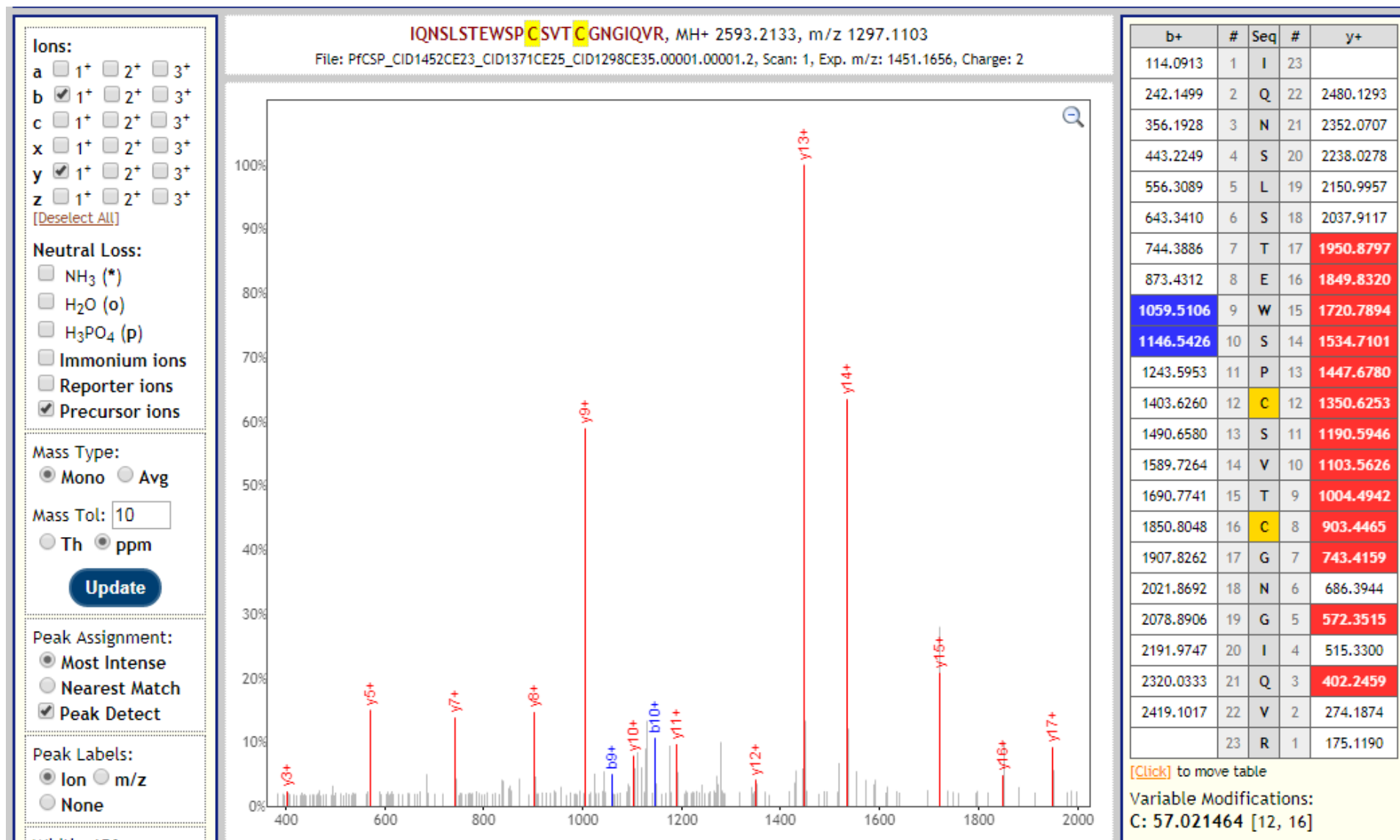


Figure S1d. MS⁴ of the 1298.11 *m/z* peak seen in Figure S1c (MS² CE = 23 %, MS³ CE = 25 %, MS⁴ CE = 35 %). This peak arises from neutral loss of O-Fuc after neutral loss of Glc from the O-Fuc-Glc disaccharide. Fragment ions confirm the sequence of peptide. This spectrum further confirms that the 1370.14 *m/z* peak seen in Figure S1b arises from neutral loss of Glc from the O-Fuc-Glc disaccharide.

Figure S2. Neutral loss of O-linked and C-linked glycans at increasing collision energies.

A tryptic digest of recombinant *P. falciparum* thrombospondin related anonymous protein (TRAP) was analyzed by LC-MS/MS with an LTQ-Orbitrap Elite. Precursor ions for the TSR peptide modified with C-Man and O-Fuc-Glc were isolated and fragmented at sequentially increasing normalized collision energies (CE). All spectra are collision-induced dissociation (CID) collected at high resolution in the Orbitrap. Annotated fragment spectra were visualized using a development version of Lorikeet as implemented in the Trans-Proteomic Pipeline.

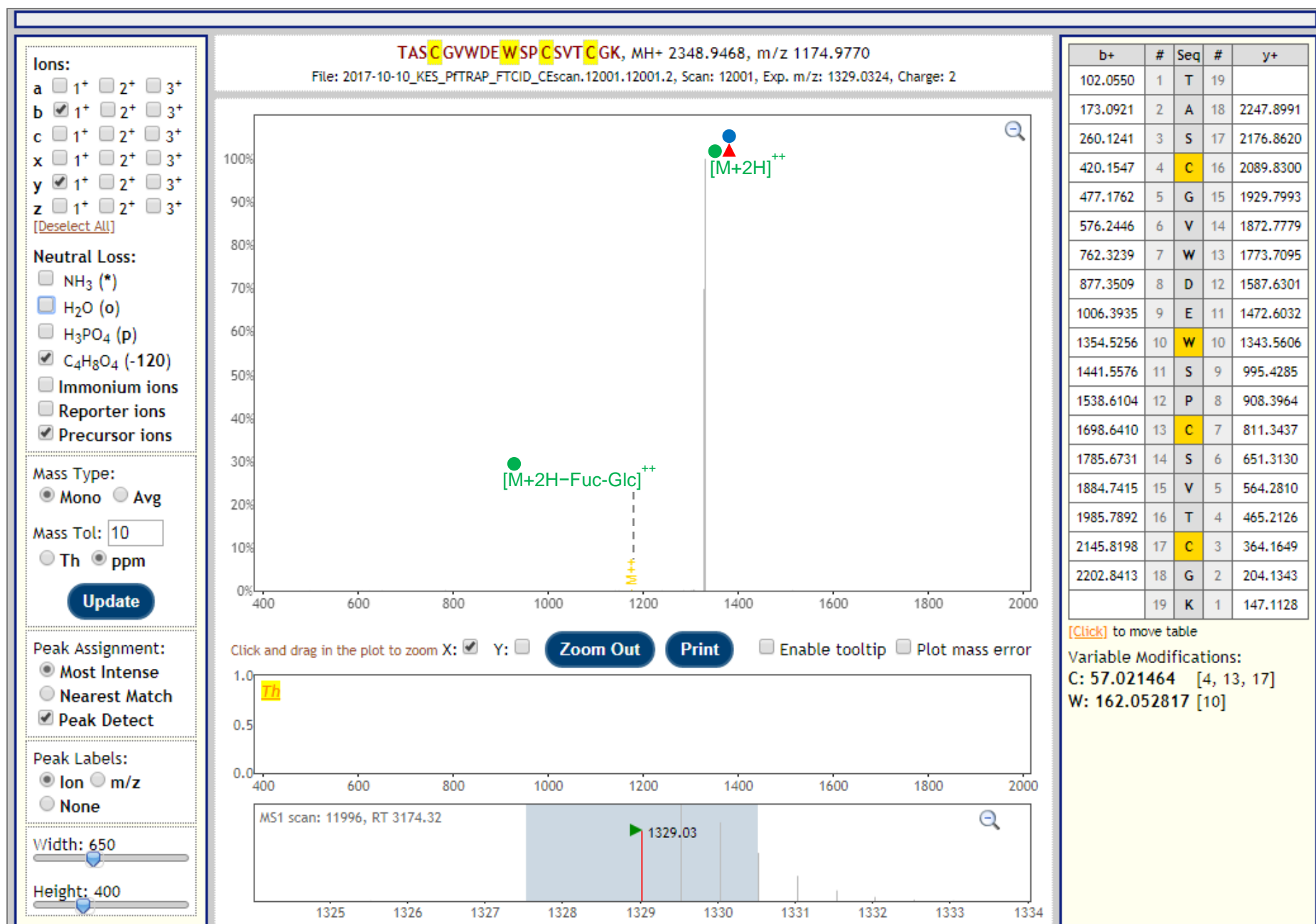


Figure S2a. CID at 1329.03 *m/z*, CE = 20 %. The precursor peak is largely unreacted ([M+2H]⁺⁺, green circle = C-Man, red triangle = O-Fuc, blue circle = β1,3-Glc), though a small peak (~0.3% relative abundance) matches the mass of the peptide after neutral loss of the O-linked disaccharide (1174.98 *m/z*, yellow, automatically annotated by Lorikeet as the precursor peak M⁺⁺).

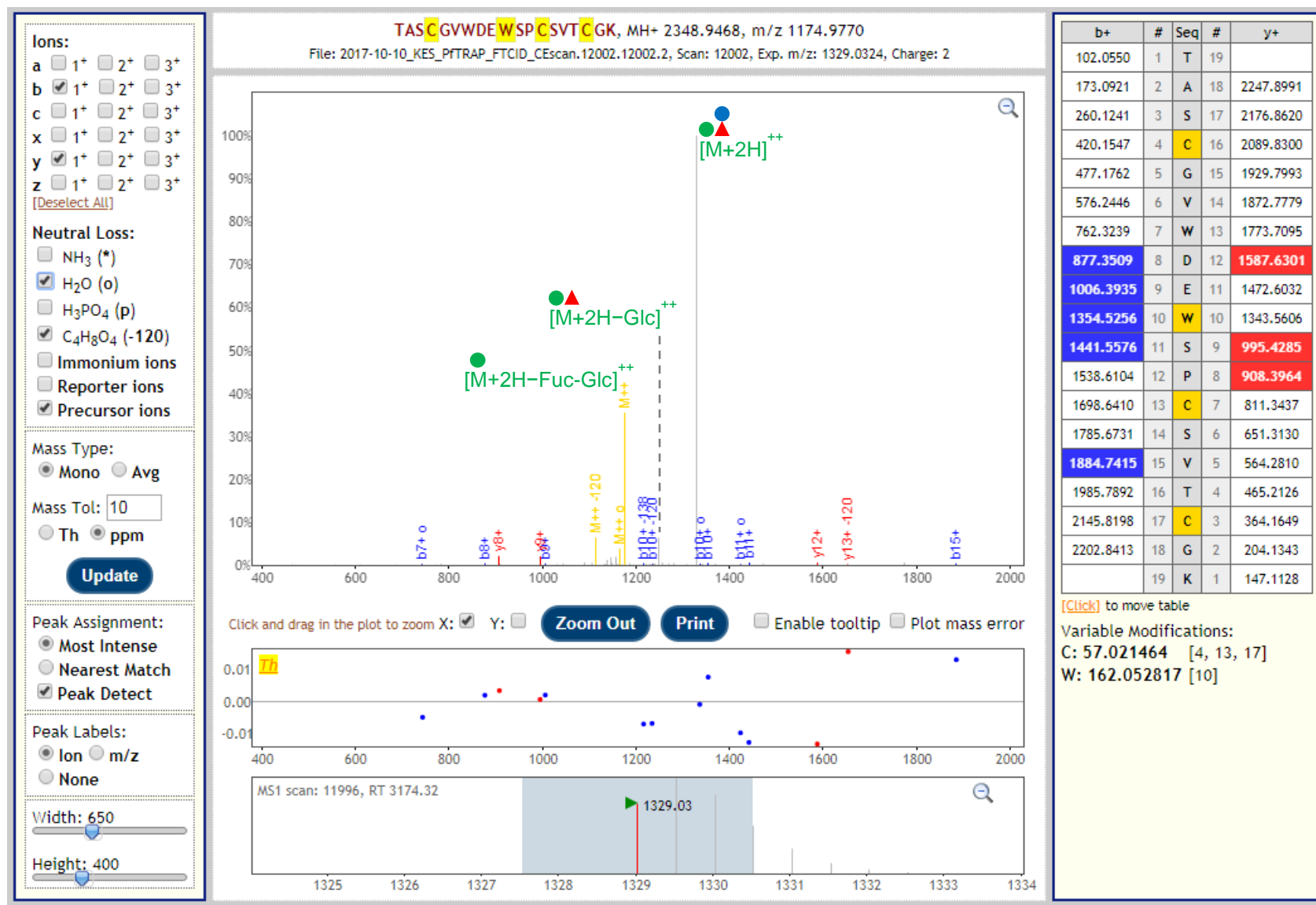


Figure S2b. CID at 1329.03 *m/z*, CE = 25 %. The dominant peak is still unreacted precursor ([M+2H]⁺⁺, green circle = C-Man, red triangle = O-Fuc, blue circle = β1,3-Glc). A smaller peak at 1248.01 *m/z* matches the mass of the precursor after neutral loss of Glc from the O-Fuc-Glc disaccharide ([M+2H-Glc]⁺⁺). A third peak at 1174.98 *m/z* (yellow, automatically annotated by Lorikeet as the precursor peak M++) matches the mass of the peptide precursor ion after neutral loss of the O-Fuc-Glc disaccharide ([M+2H-Fuc-Glc]⁺⁺). Note that peptide fragment ions appearing at this CE identify the sequence of the peptide with C-Man intact, positively localizing the glycan to the C-terminal Trp residue. Neutral loss of 120.04 Da from cross-ring cleavage is seen on the precursor and fragment peaks.

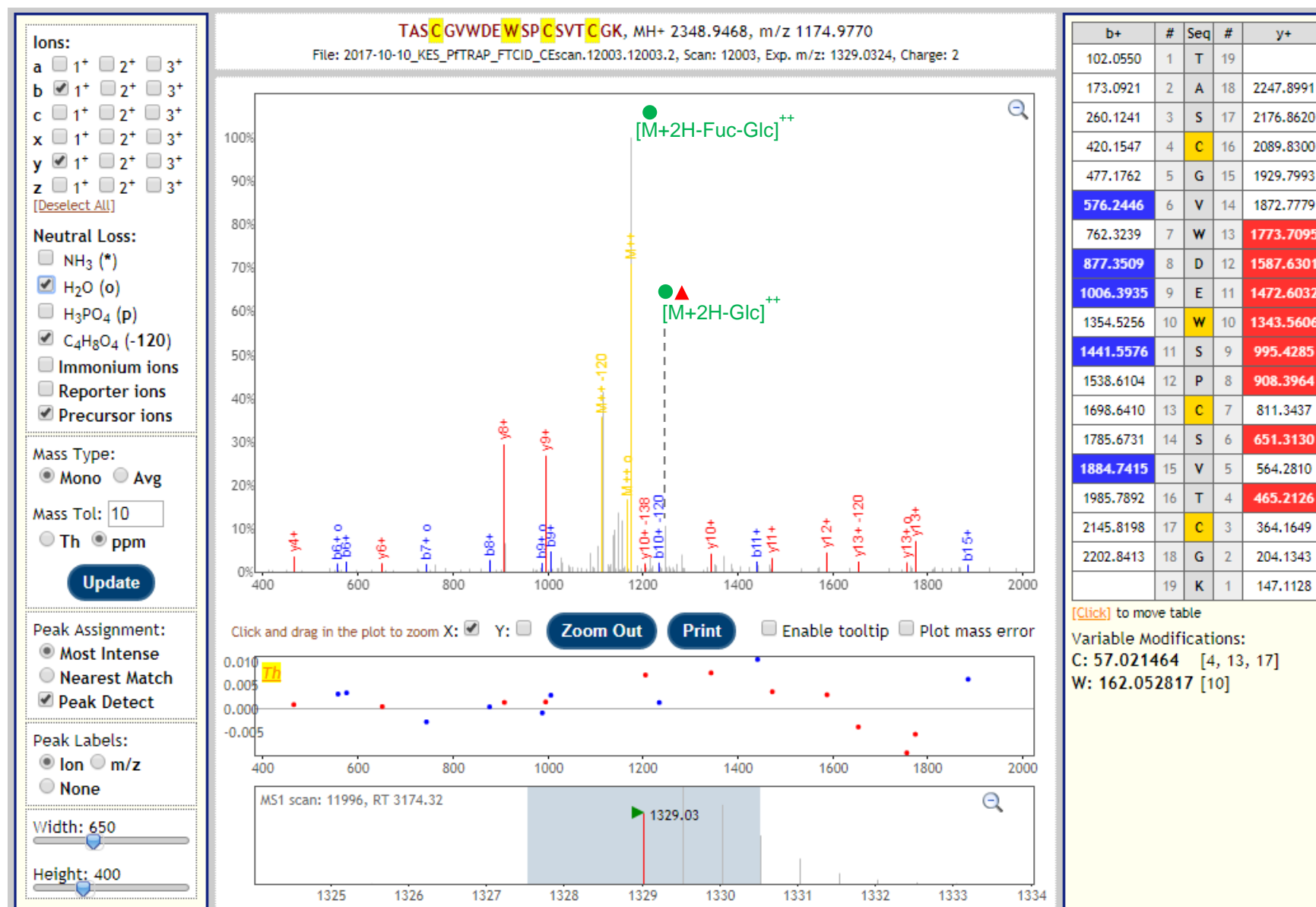


Figure S2c. CID at 1329.03 *m/z*, CE = 30 %. At this CE all parent precursor ion has been fragmented. The small peak at 1248.01 *m/z* remains, the mass of the precursor after neutral loss of Glc from the O-Fuc-Glc disaccharide ([M+2H-Glc]⁺⁺). The dominant peak is now 1174.98 *m/z* (yellow M⁺⁺), the peptide precursor ion after neutral loss of the O-Fuc-Glc disaccharide ([M+2H-Fuc-Glc]⁺⁺). Further peptide fragment ions confidently identify the sequence of the peptide with C-Man intact.

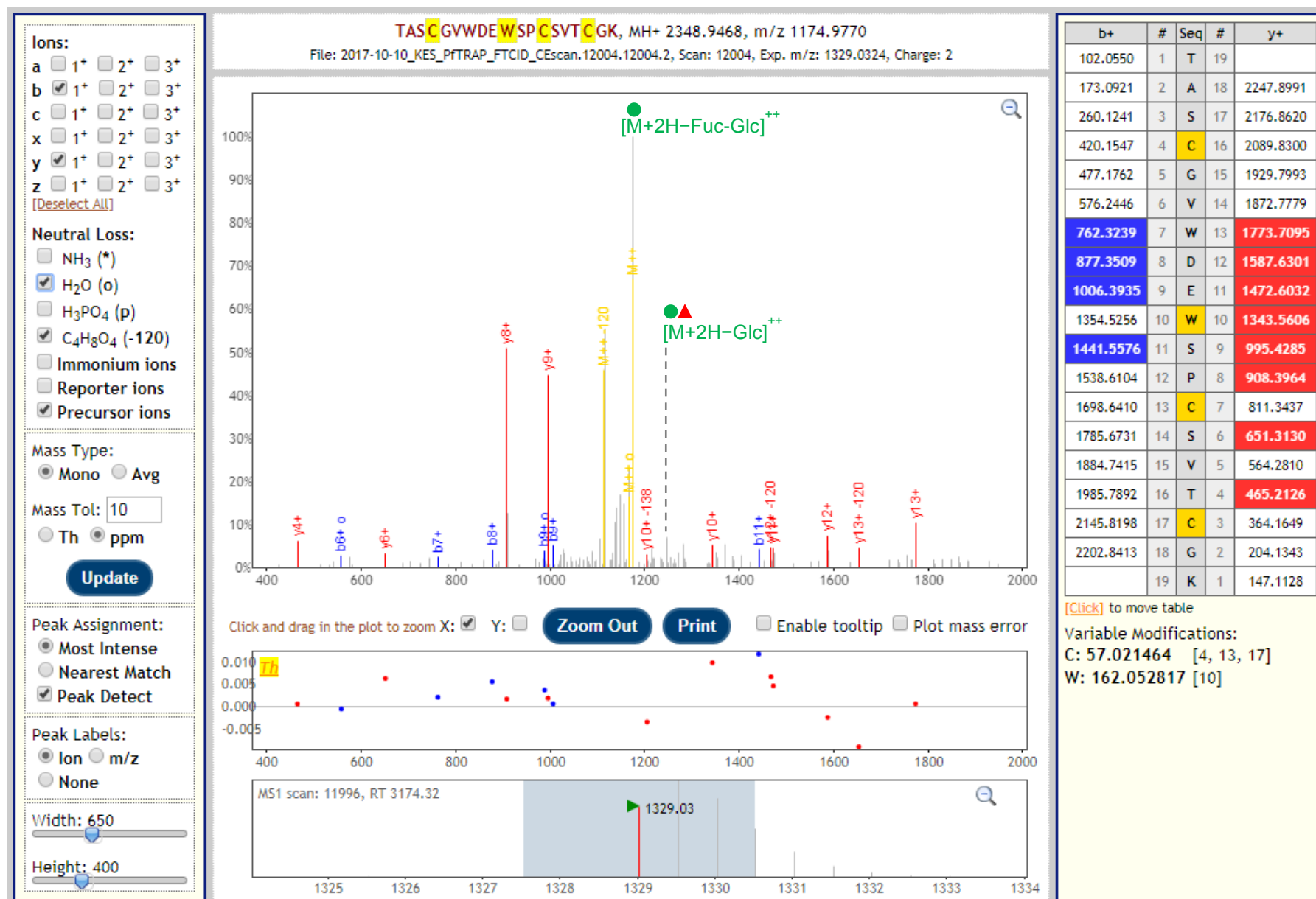


Figure S2d. CID at 1329.03 *m/z*, CE = 35 %. This is the CE typically used in shotgun proteomics methods. The C-mannosylated peptide precursor lacking the O-linked glycan remains the dominant peak, but peptide fragment ions are abundant enough to confidently sequence the peptide. Unlike the recombinant *PfCSP* spectra in Figure 1 and supplemental Figure S1b, no peptide fragment ions with the O-linked glycan intact are visible at any CE. This is likely a signal-to-noise issue; the spectra in Figure 1 and Figure S1 were acquired by direct infusion of the peptide, allowing signal and ion fill time to be maximized, whereas these spectra were acquired on-the-fly by LC-MS/MS.

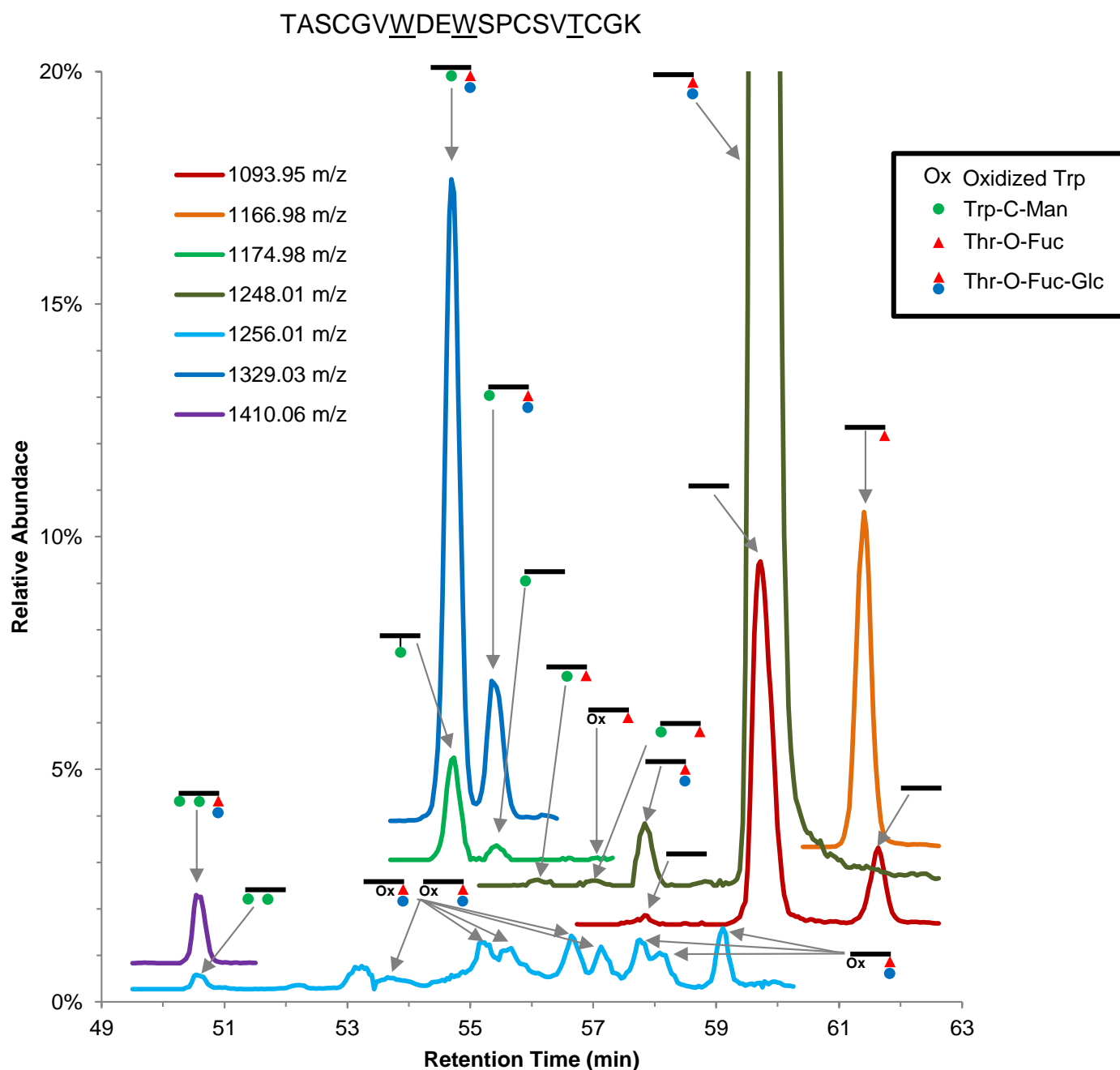


Figure S3. Glycoforms identified by automated sequence database search of data from LC-MS/MS analysis of recombinant *P. falciparum* TRAP. The variably C-mannosylated or oxidized Trp residues and the variably O-fucosylated Thr are underlined. Traces are offset for clarity. Peaks indicated with arrows were positively identified by PSMs. Both positional isoforms of the oxidized peptide were identified at several co-eluting retention times. See table S2.

Figure S4. Indistinguishable isobaric glycopeptides from recombinant *P. falciparum* TRAP.



Figure S4a,b. Representative MS² of the two 1248.01 m/z peaks eluting at a) 57.8 min and b) 58.9 min. The MS² from both species appear to identify the same species, i.e. the peptide modified with a gas-phase-labile moiety having a mass matching that of O-Fuc-Glc. The dominant species of the MS² spectra, 1093.95 m/z, matches the mass of the unmodified peptide. Additional PSMs are listed in Table S2.

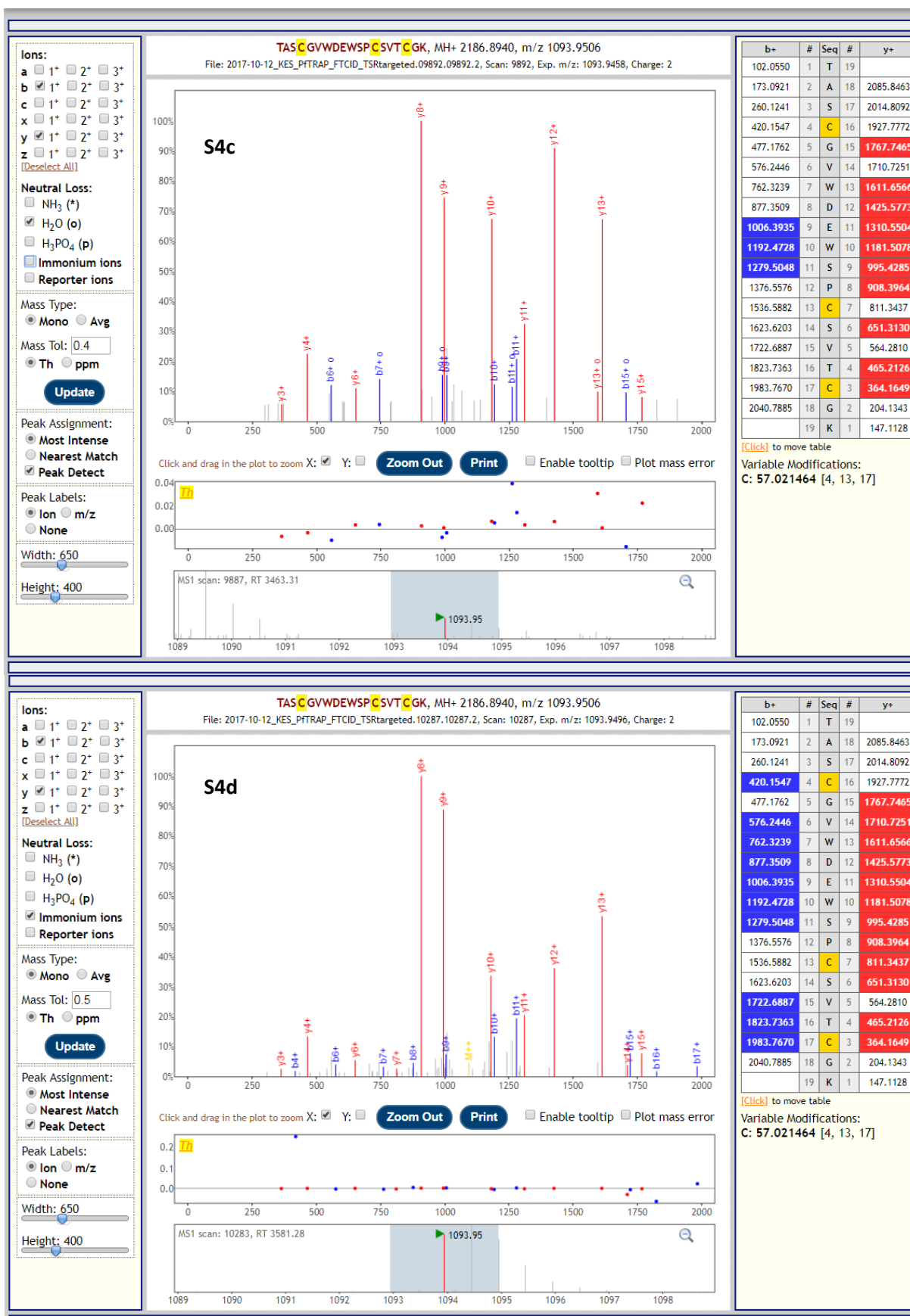


Figure S4c,d. Representative MS² of the two 1093.5 *m/z* peaks eluting at **c)** 57.8 min and **d)** 58.9 min. The MS² from both species appear to identify the unmodified peptide. Both species co-elute with the 1248.01 *m/z* species identified in Fig. S4a and b, consistent with loss of the gas-phase-labile O-Fuc-Glc from in-source fragmentation. Additional PSMs are listed in Table S2.

S5. Misidentification of a glycoform in LC-MS/MS of recombinant *P. falciparum* TRAP

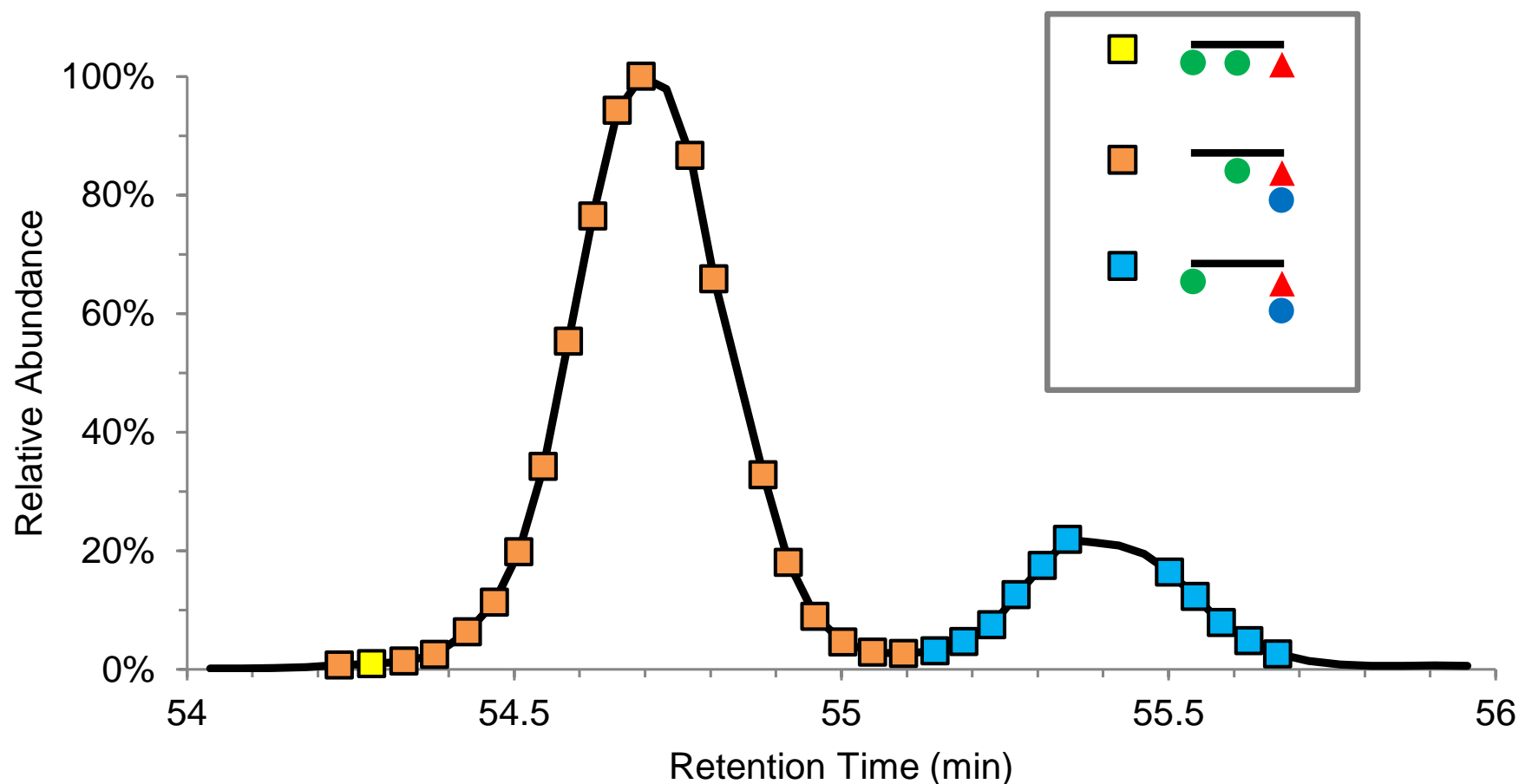


Figure S5a. Extracted Ion Chromatogram (XIC) of the precursor ion matching multiple isobaric glycoforms. The XIC of 1329.03 m/z (black line) is overlaid with squares showing points where MS² resulted in high-quality peptide spectrum matches (PSMs) identifying glycoforms of the recombinant *Pf*TRAP TSR peptide TASC₁GVDEWSPCSVT₂CGK modified with combinations of C-linked mannose at tryptophan (green circle), O-linked fucose at the C-terminal threonine (red triangle) and β 1,3-linked glucose (blue circle) added to the fucose. See Table S2.

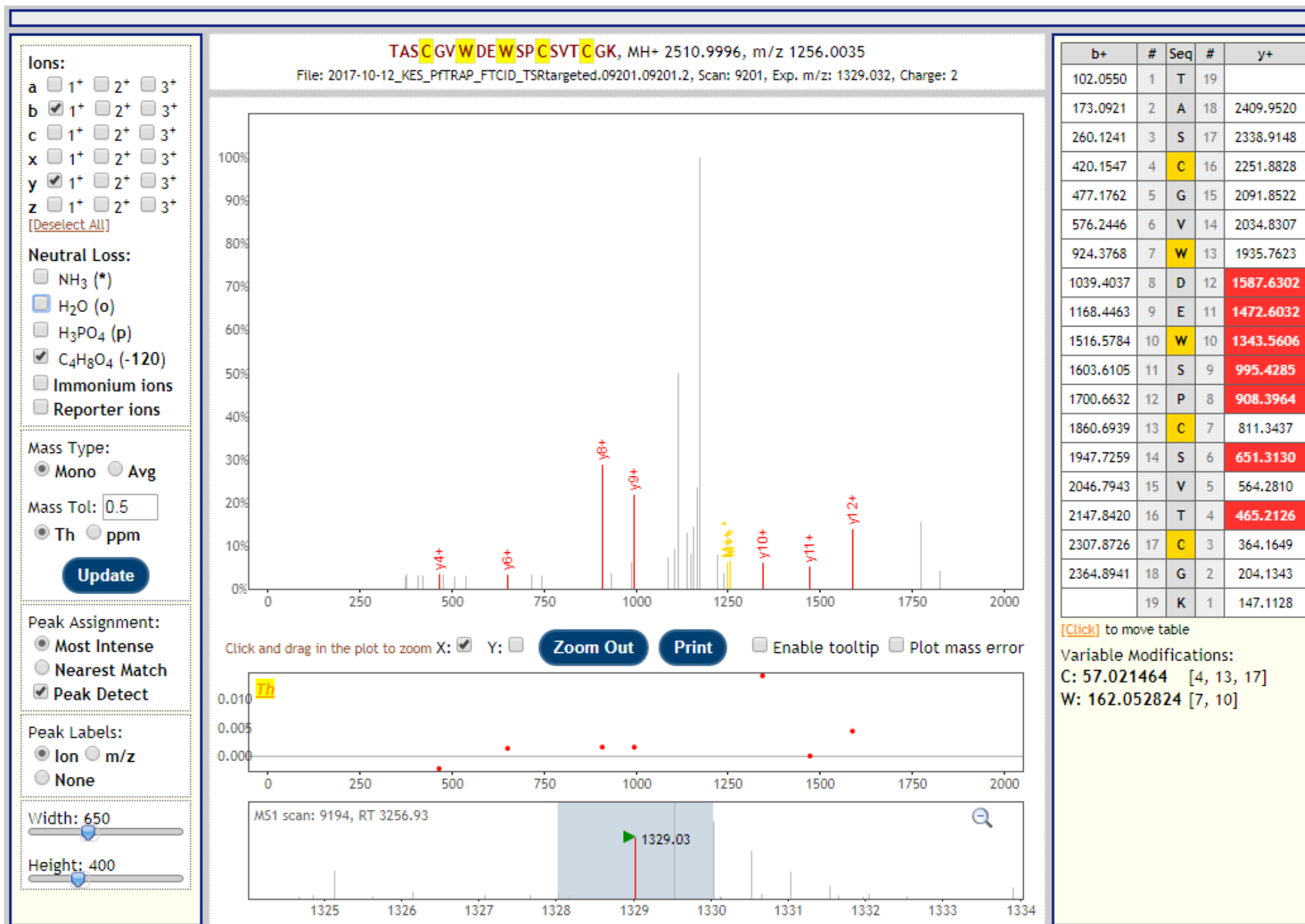


Figure S5b A single PSM (the yellow square in Figure S5a) was incorrectly identified as the glycoform of the peptide featuring double mannosylation and an O-Fuc, with the MS² annotated as shown here.

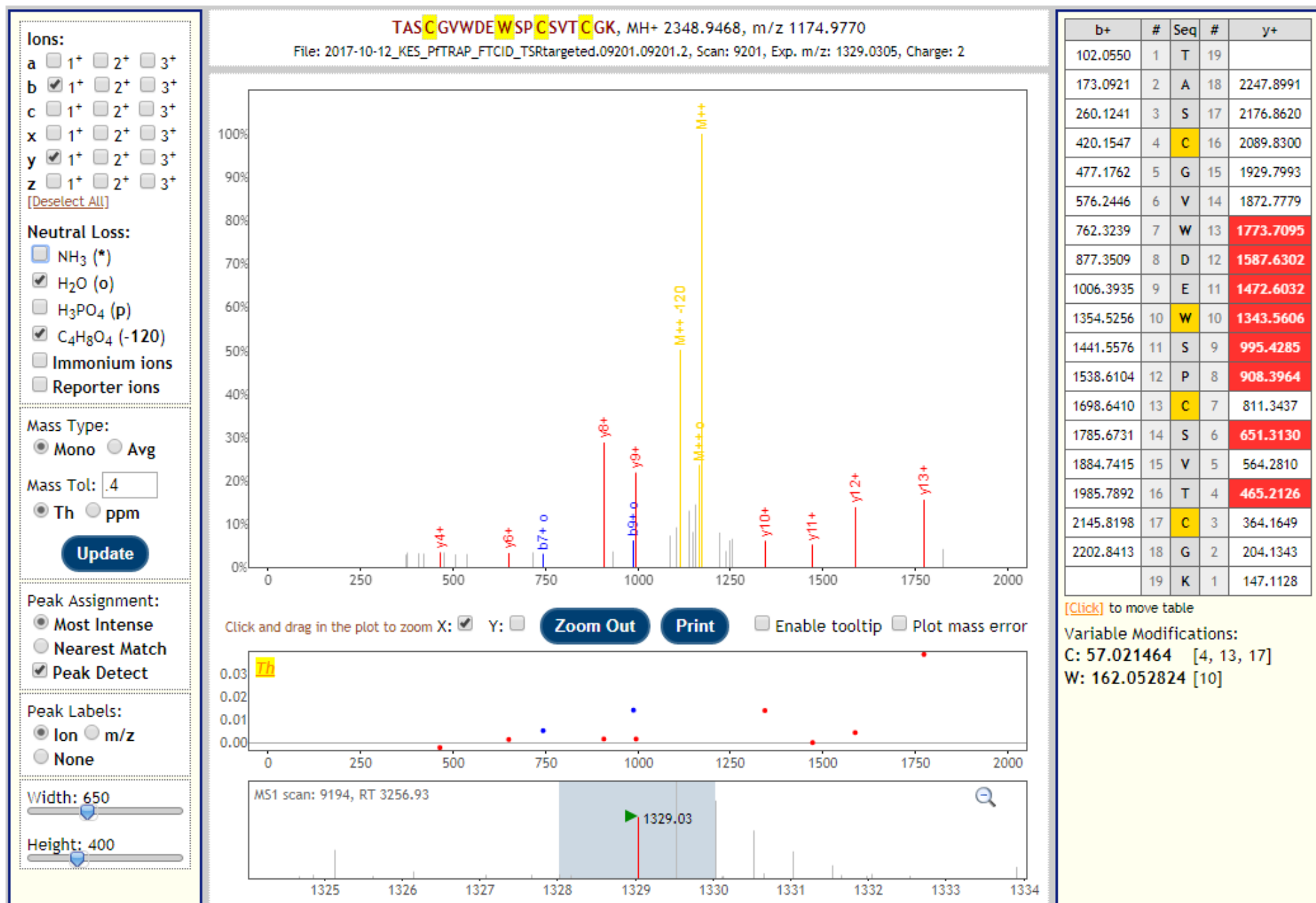


Figure S5c The same MS² from Figure S5b, re-annotated with the correct glycoform. While most of the fragment ions are the same as for the incorrect annotation (except y₁₃), the precursor fragment exhibiting neutral loss of O-Fuc-Glc (the yellow M⁺⁺ peak) as well as cross-ring cleavage of C-man (yellow M⁺⁺-120 peak) confirm that this is the correct annotation.

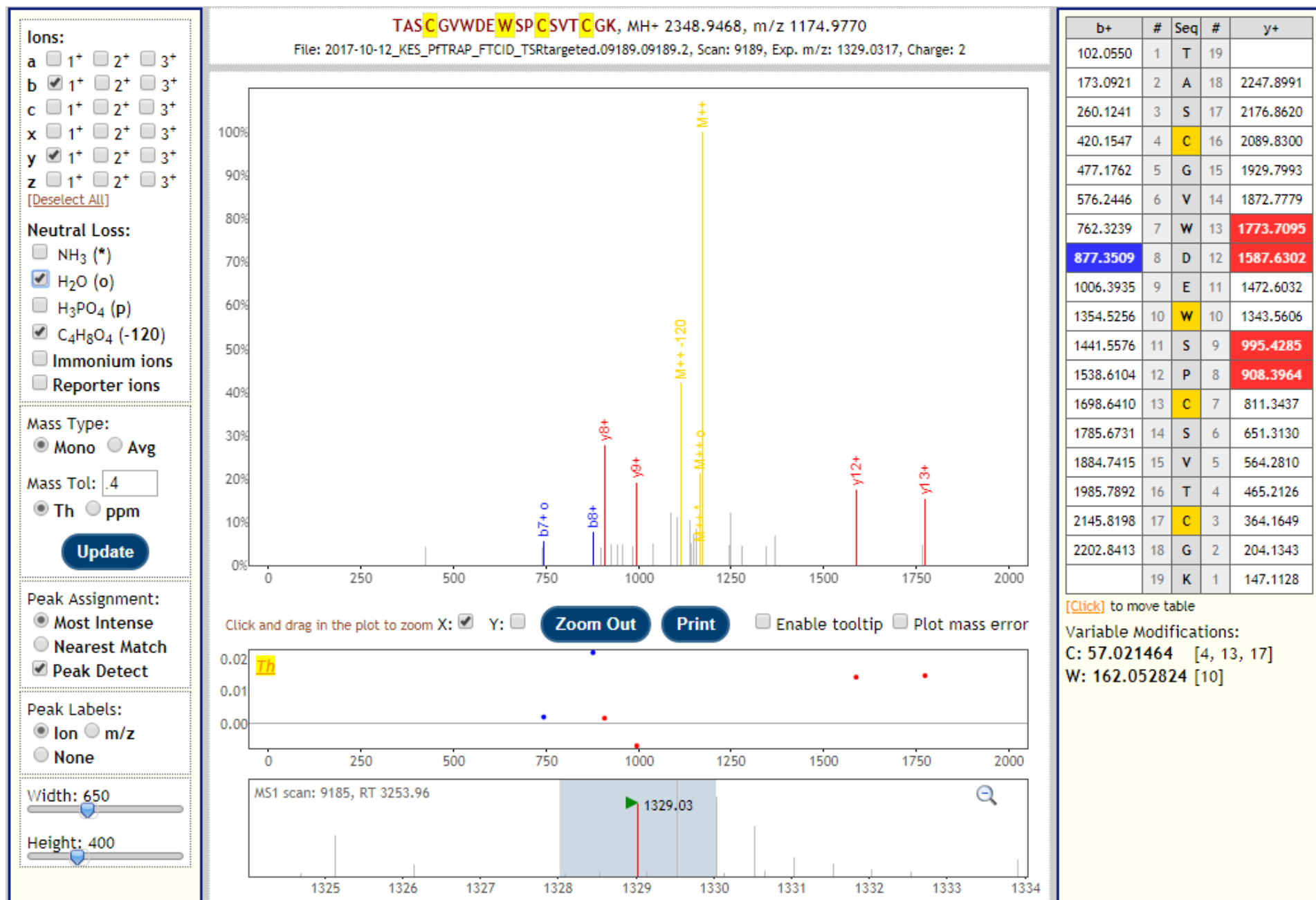


Figure S5d. The MS² for the same parent ion that immediately precedes the incorrect PSM, though sparse, identifies the correct glycoform.

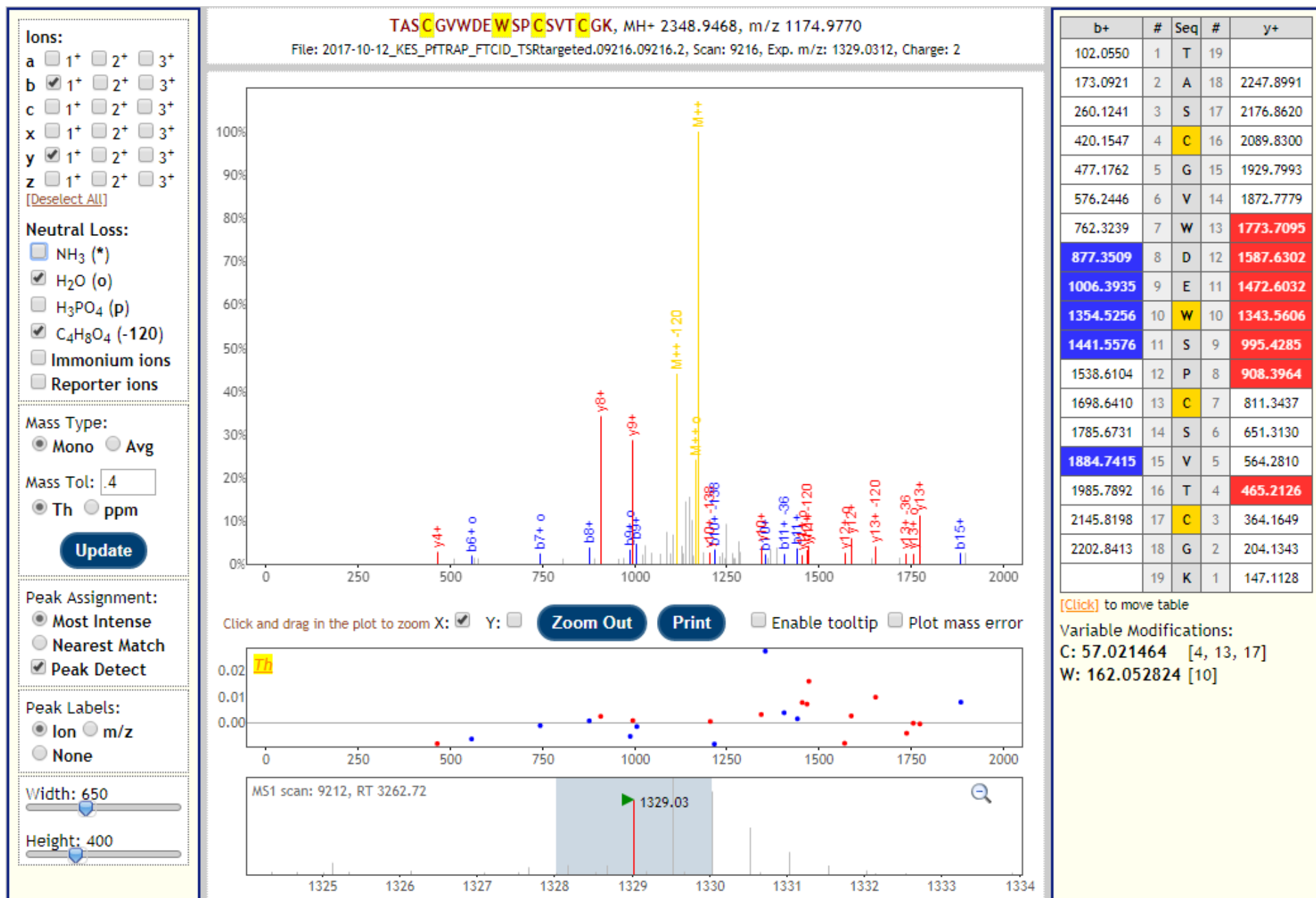


Figure S5e. The MS² for the same parent ion immediately following the incorrect PSM identifies the correct glycoform, as do the remaining PSMs for the parent ion in the same chromatographic peak.

IQNSLSTEWSPCSVICGNGIQVR

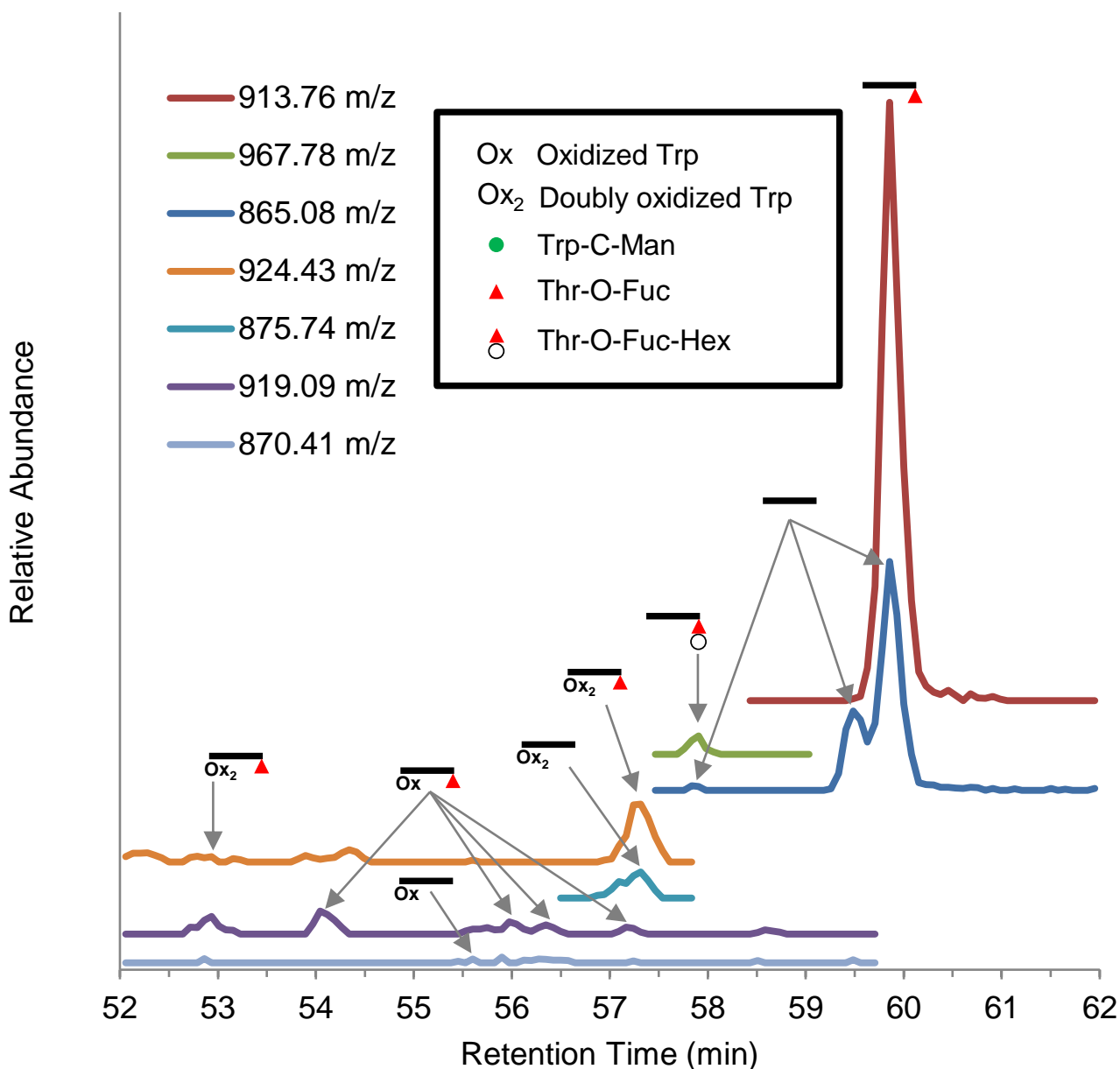
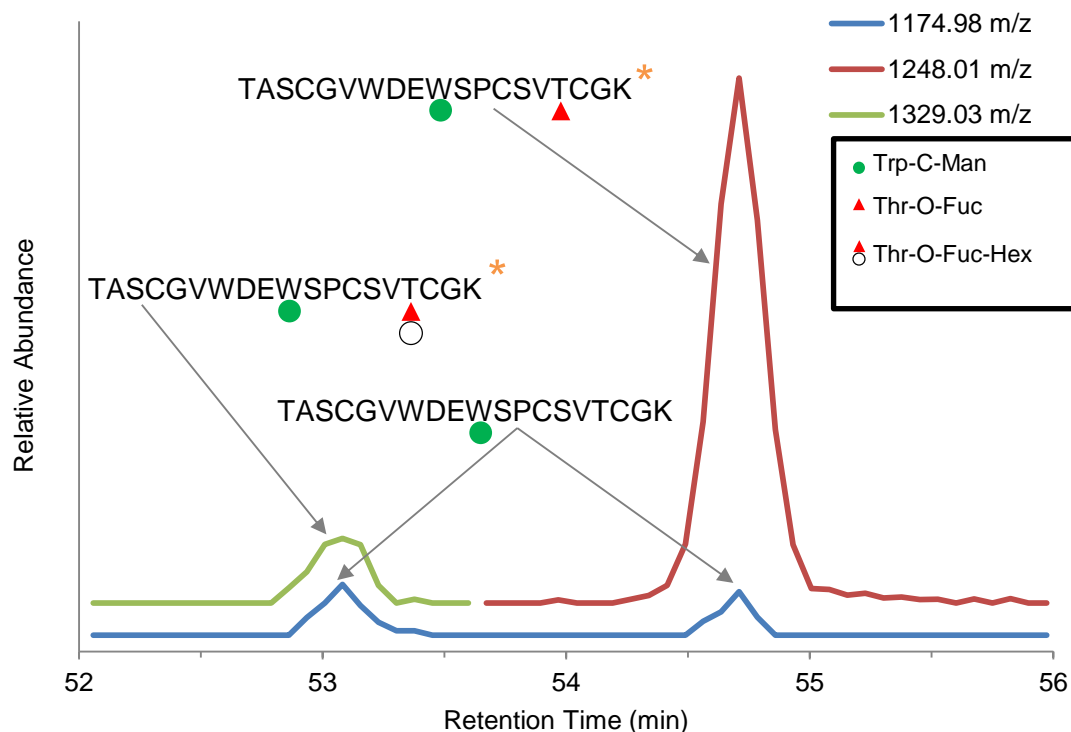


Figure S6. Representative XIC of glycosylated CSP from *P. falciparum* salivary gland sporozoites. The variably oxidized Trp and the variably O-fucosylated Thr are underlined. Traces are offset for clarity. Peaks indicated with arrows were positively identified by PSMs. The Hex of the O-Fuc-Hex disaccharide is indicated as a generic hexose because the identity of the glycan has not been confirmed.

***P. falciparum* salivary gland sporozoites**
Whole proteome



***P. falciparum* salivary gland sporozoites**
Surface-enriched

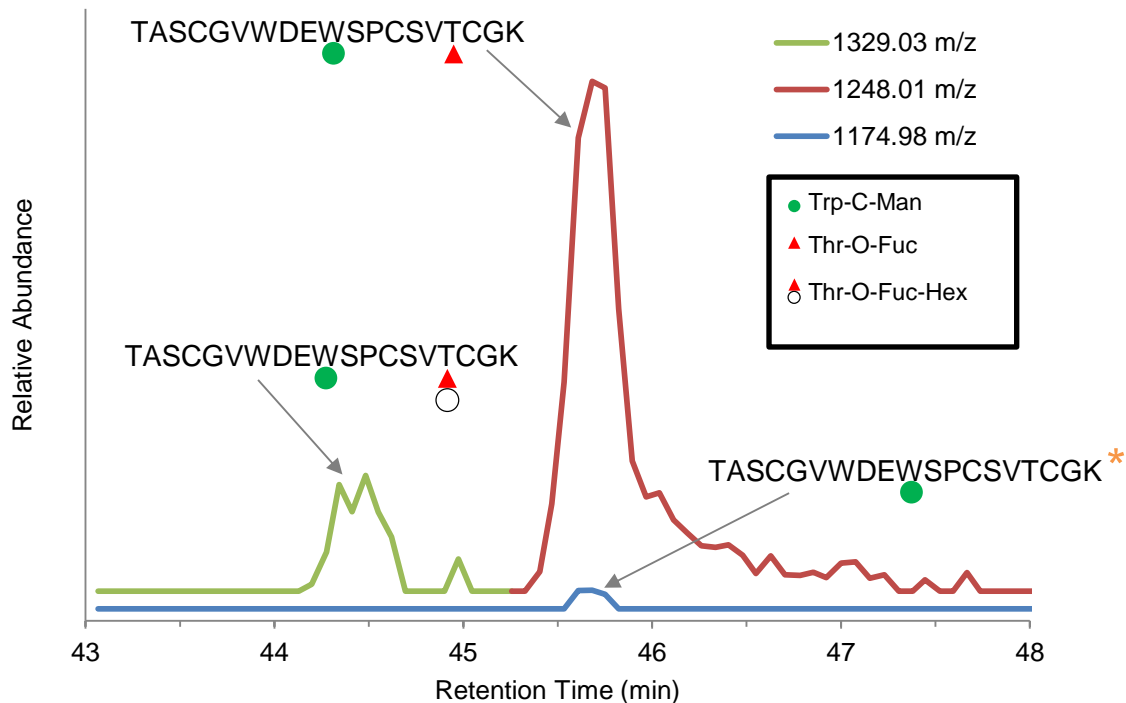


Figure S7. Representative XIC of glycosylated TRAP from *P. falciparum* salivary gland sporozoites. Traces are offset for clarity. Peaks marked with an orange asterisk (*) were not identified by PSMs because the precursor was not selected for MS², but their retention time and mass correspond with the indicated glycoform. The Hex of the O-Fuc-Hex disaccharide is indicated as a generic hexose because the identity of the glycan has not been confirmed.

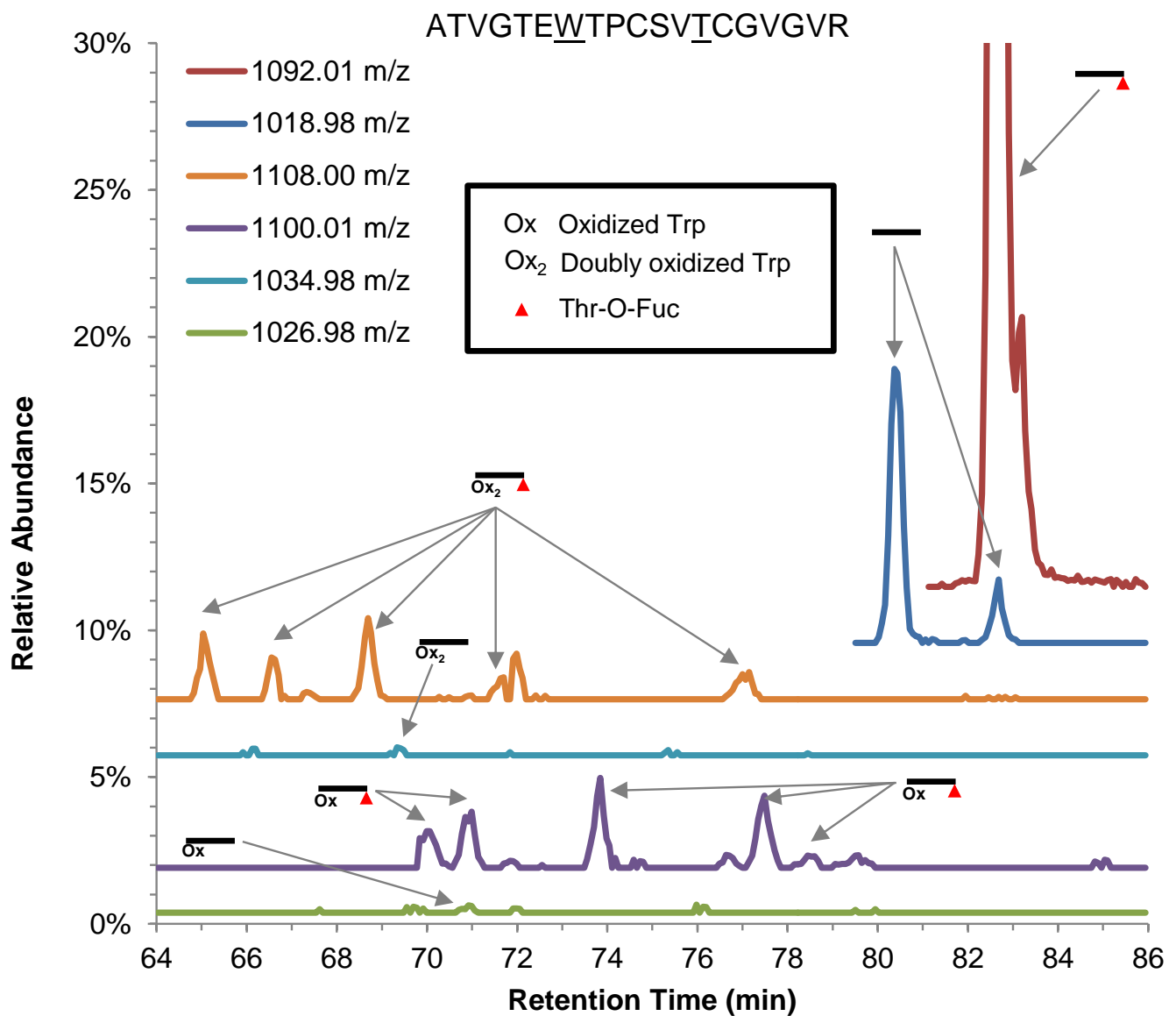


Figure S8. Representative XIC of glycosylated CSP from *P. vivax* VK210 salivary gland sporozoites. The variably oxidized Trp and the variably O-fucosylated Thr are underlined. Traces are offset for clarity. Peaks indicated with arrows were positively identified by PSMs.

VANCGPWDPWTACSVTCGR

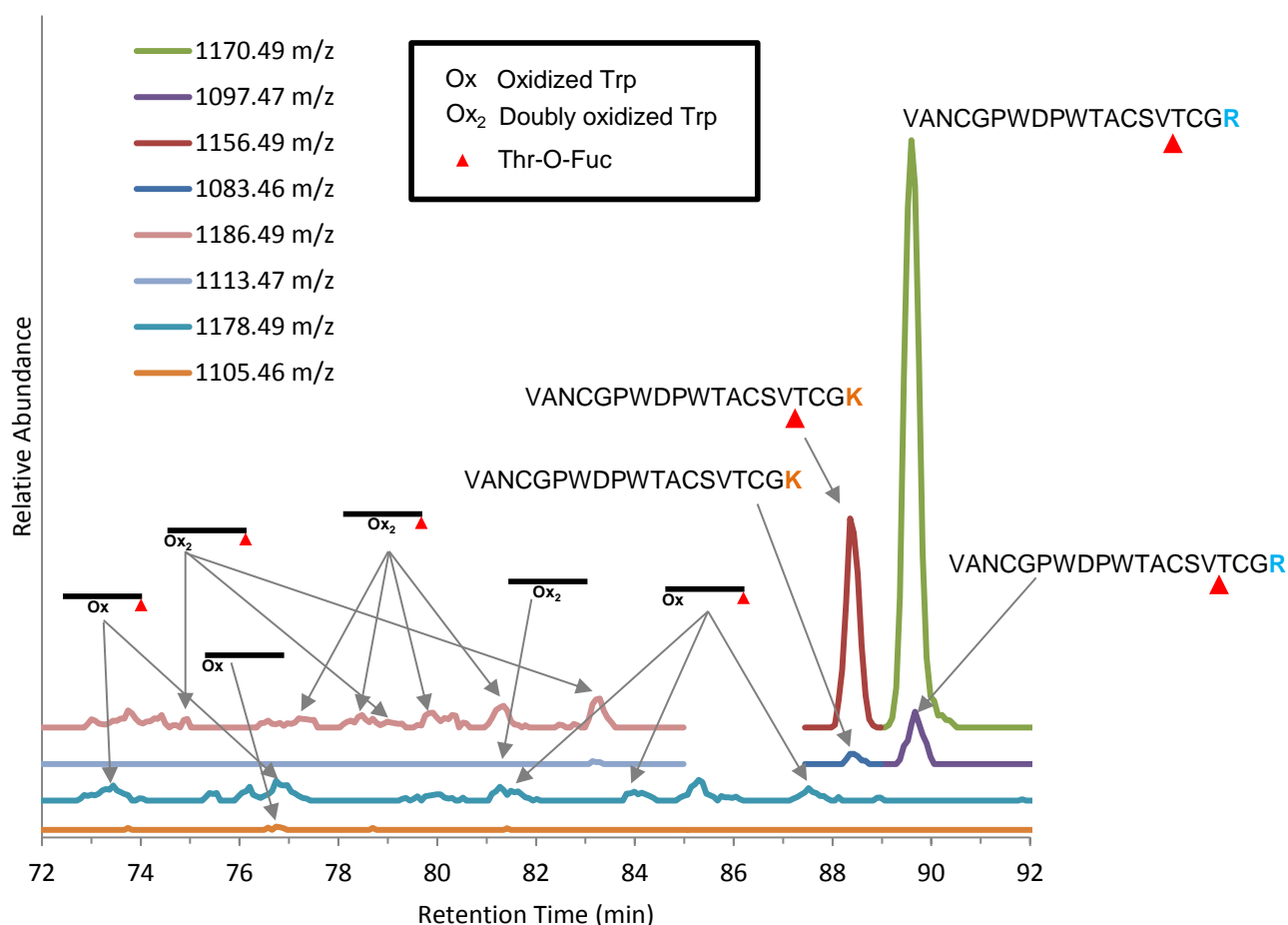


Figure S9. Representative XIC of glycosylated TRAP from *P. vivax* VK210 salivary gland sporozoites. The variably oxidized Trp residues and the variably O-fucosylated Thr are underlined. Traces are offset for clarity. Peaks indicated with arrows were positively identified by PSMs. Note that this sample contained a mixture of two field isolates, each of which carried a different version of the TRAP gene with either a Lys or Arg at the C-terminus of the glycopeptide.

ATVGTEWTPCSVICGVGVR

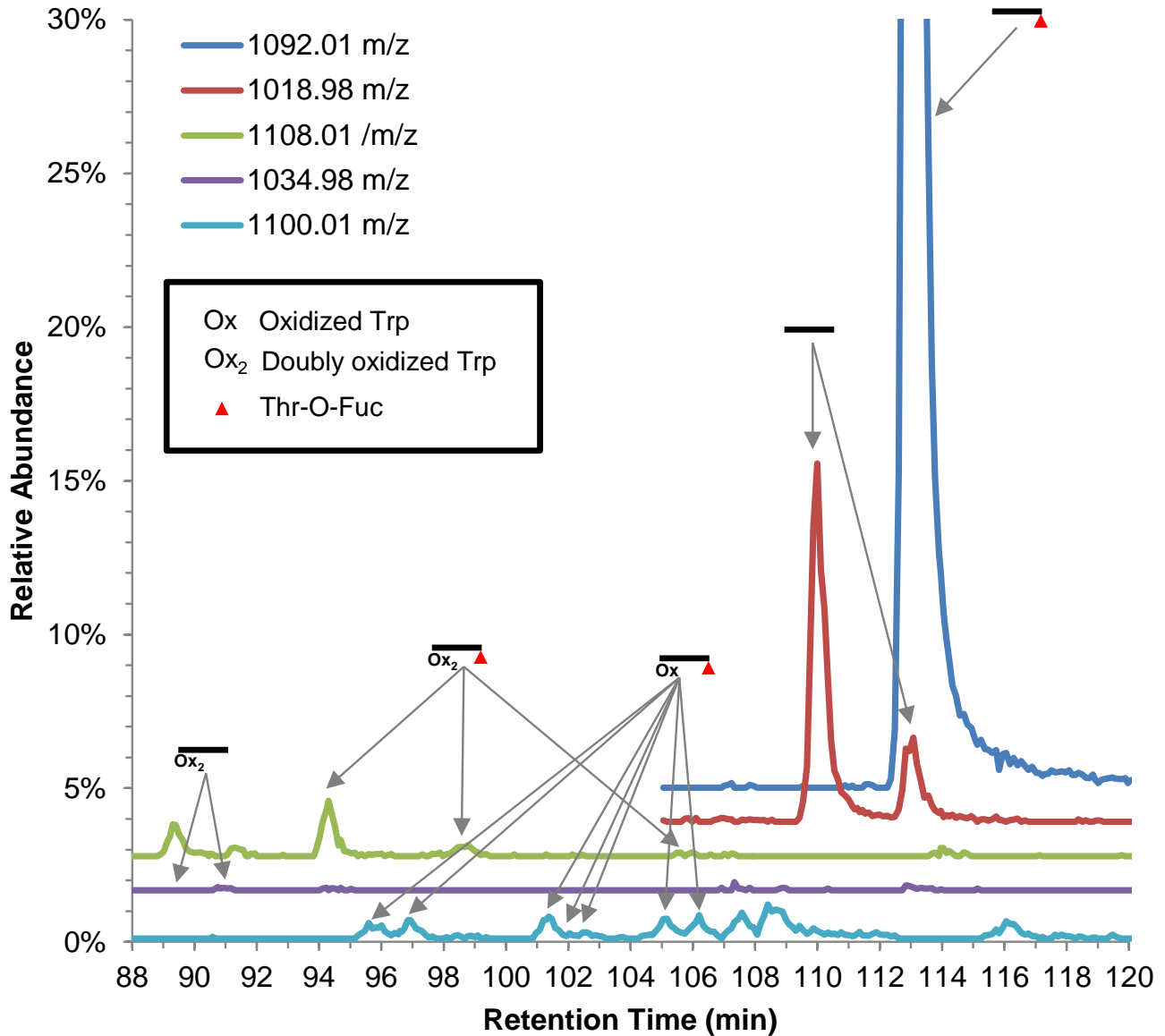


Figure S10. Representative XIC of glycosylated CSP from *P. vivax* VK247 salivary gland sporozoites. The variably oxidized Trp and the variably O-fucosylated Thr are underlined. Traces are offset for clarity. Peaks indicated with arrows were positively identified by PSMs.

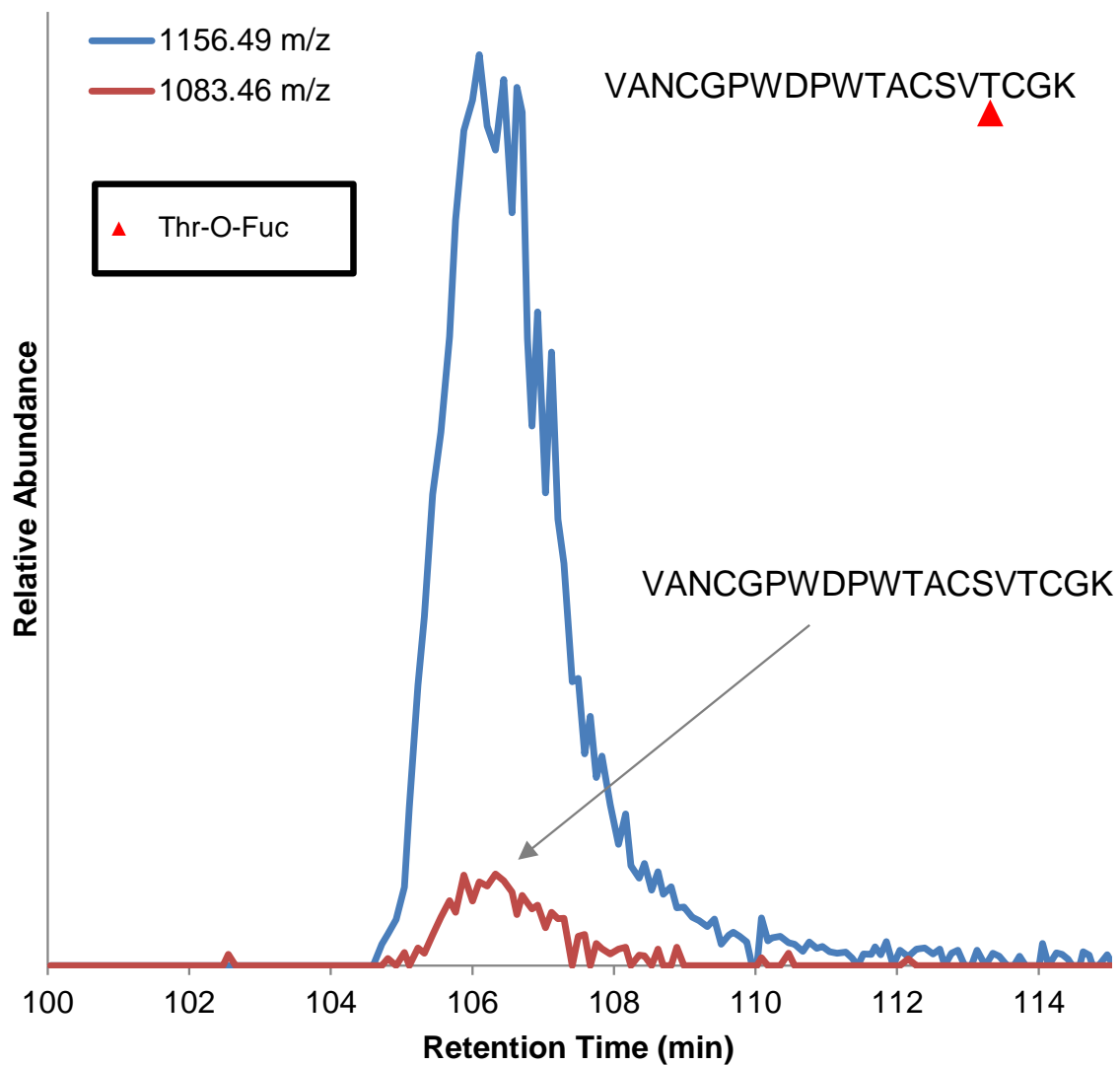


Figure S11. Representative XIC of glycosylated TRAP from *P. vivax* VK247 salivary gland sporozoites.

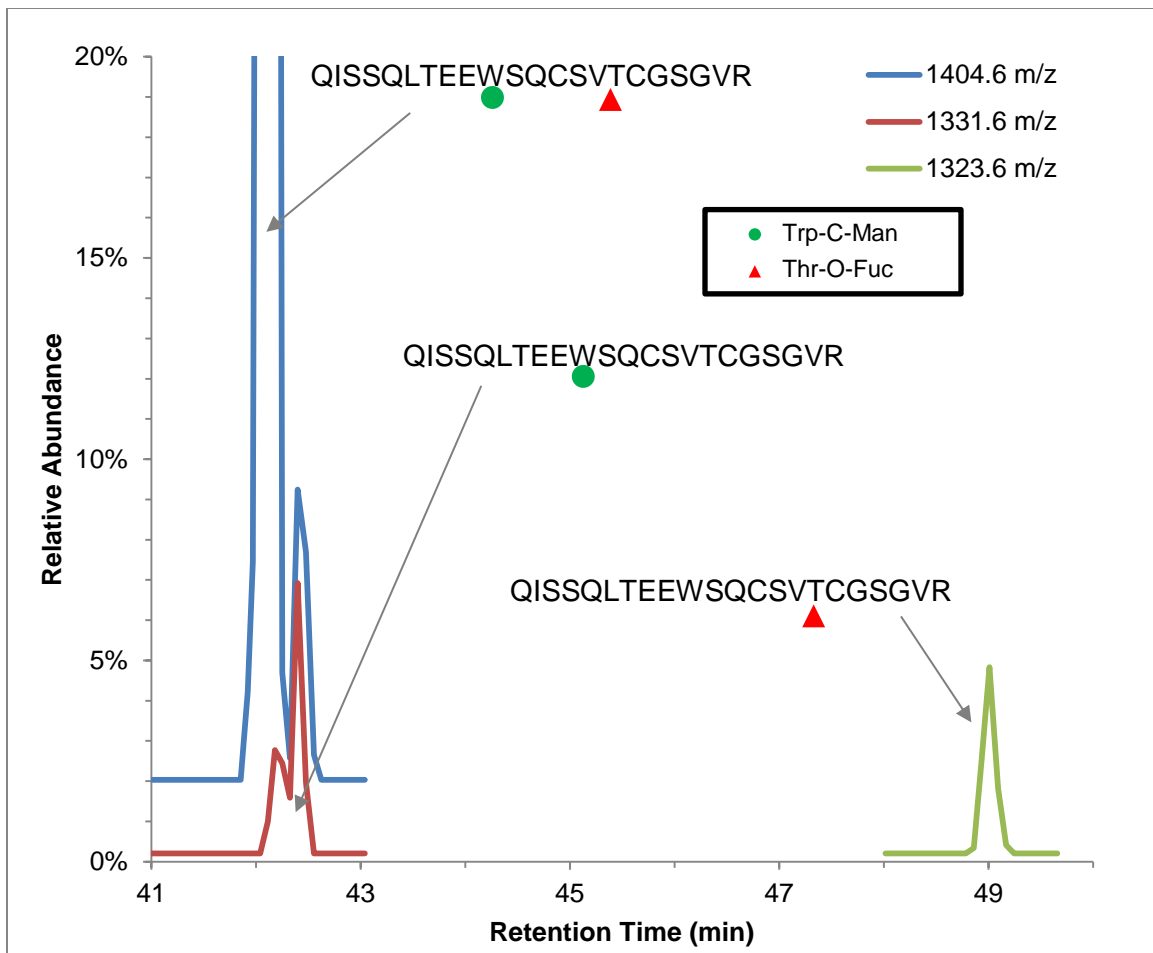


Figure S12. Representative XIC of glycosylated CSP from *P. yoelii* salivary gland sporozoites. Traces are offset for clarity. The reason for the doublet shape of the 1404.6 *m/z* and co-eluting 1331.6 *m/z* peaks is unknown, and may reflect spray instability or closely eluting isobaric species. Only the indicated glycopeptides were identified from MS² of these masses collected during the elution window of the peaks.

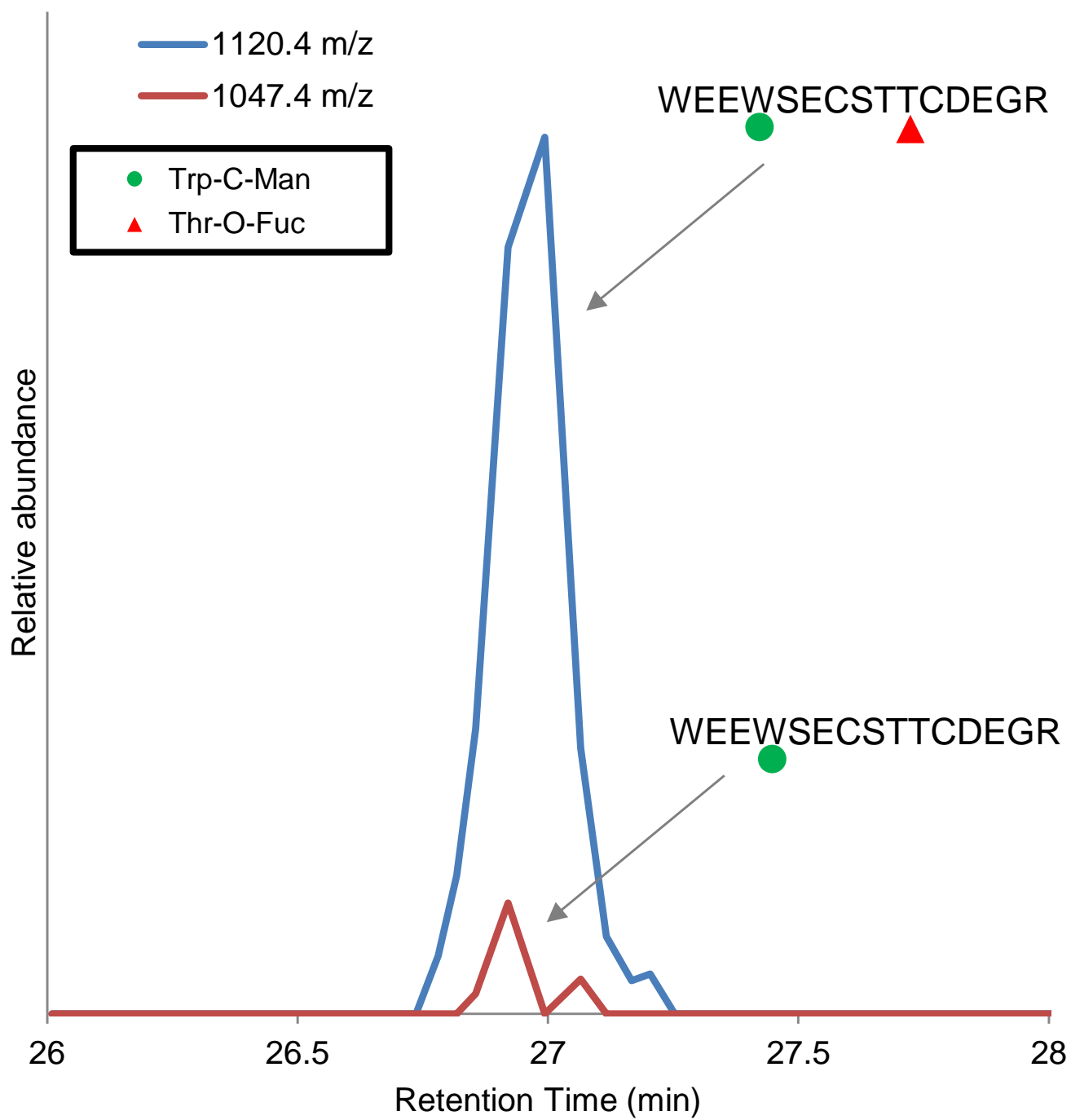


Figure S13. Representative XIC of glycosylated TRAP from *P. yoelii* salivary gland sporozoites.

Ions:

a ☐ 1+ ☐ 2+ ☐ 3+
b ☒ 1+ ☐ 2+ ☐ 3+
c ☐ 1+ ☐ 2+ ☐ 3+
x ☐ 1+ ☐ 2+ ☐ 3+
y ☒ 1+ ☐ 2+ ☐ 3+
z ☐ 1+ ☐ 2+ ☐ 3+
[\[Deselect All\]](#)

Neutral Loss:

☐ NH₃ (*)
☒ H₂O (o)
☐ H₃PO₄ (p)
☒ C₄H₈O₄ (-120)
☐ Immonium ions
☐ Reporter ions
☒ Precursor ions

Mass Type:

☒ Mono ☐ Avg

Mass Tol: 0.5

☒ Th ☐ ppm

Update

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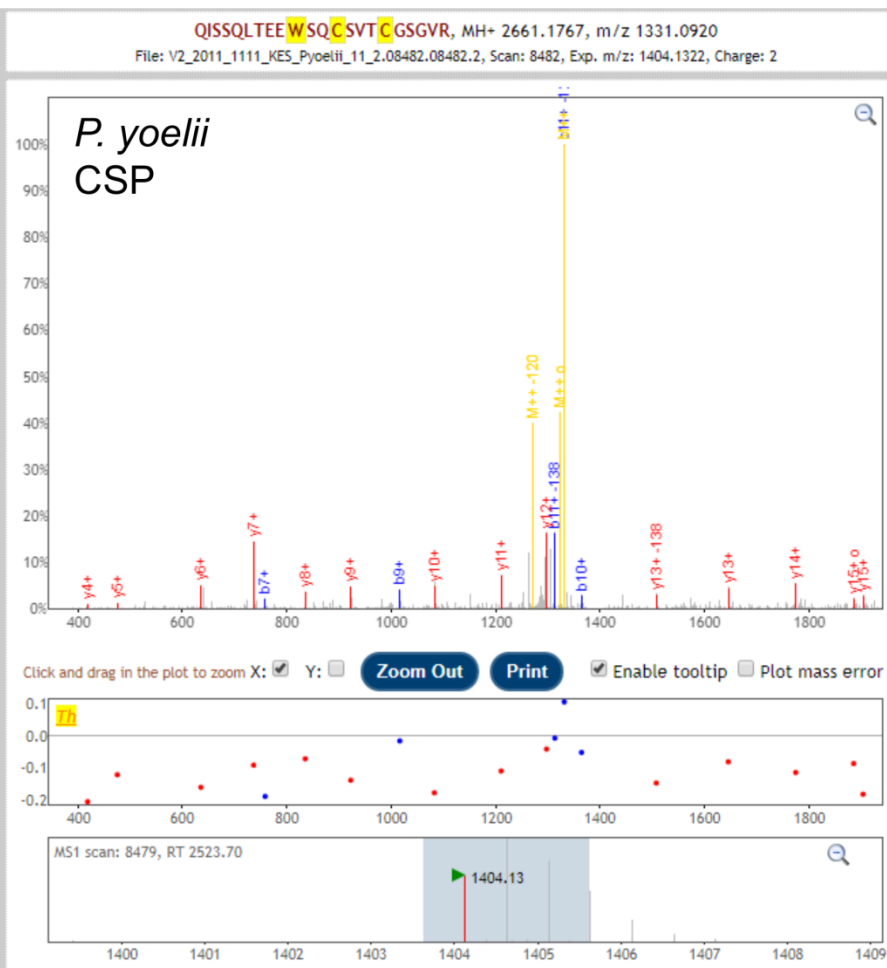
☒ Most Intense
☐ Nearest Match
☒ Peak Detect

Peak Labels:

☒ Ion ☐ m/z
☐ None

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Height: 400



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416.2140	4	S	19	2333.0020
544.2726	5	Q	18	2245.9700
657.3566	6	L	17	2117.9114
758.4043	7	T	16	2004.8273
887.4469	8	E	15	1903.7797
1016.4895	9	E	14	1774.7371
1364.6216	10	W	13	1645.6945
1451.6536	11	S	12	1297.5623
1579.7122	12	Q	11	1210.5303
1739.7429	13	C	10	1082.4717
1826.7749	14	S	9	922.4411
1925.8433	15	V	8	835.4091
2026.8910	16	T	7	736.3406
2186.9216	17	C	6	635.2930
2243.9431	18	G	5	475.2623
2330.9751	19	S	4	418.2409
2387.9966	20	G	3	331.2088
2487.0650	21	V	2	274.1874
	22	R	1	175.1190

[\[Click\]](#) to move table

Variable Modifications:
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W: 162.052824 [10]

Ions:

a ☐ 1+ ☐ 2+ ☐ 3+
b ☒ 1+ ☐ 2+ ☐ 3+
c ☐ 1+ ☐ 2+ ☐ 3+
x ☐ 1+ ☐ 2+ ☐ 3+
y ☒ 1+ ☐ 2+ ☐ 3+
z ☐ 1+ ☐ 2+ ☐ 3+
[\[Deselect All\]](#)

Neutral Loss:

☐ NH₃ (*)
☒ H₂O (o)
☐ H₃PO₄ (p)
☒ C₄H₈O₄ (-120)
☐ Immonium ions
☐ Reporter ions
☒ Precursor ions

Mass Type:

☒ Mono ☐ Avg

Mass Tol: .4

☒ Th ☐ ppm

Update

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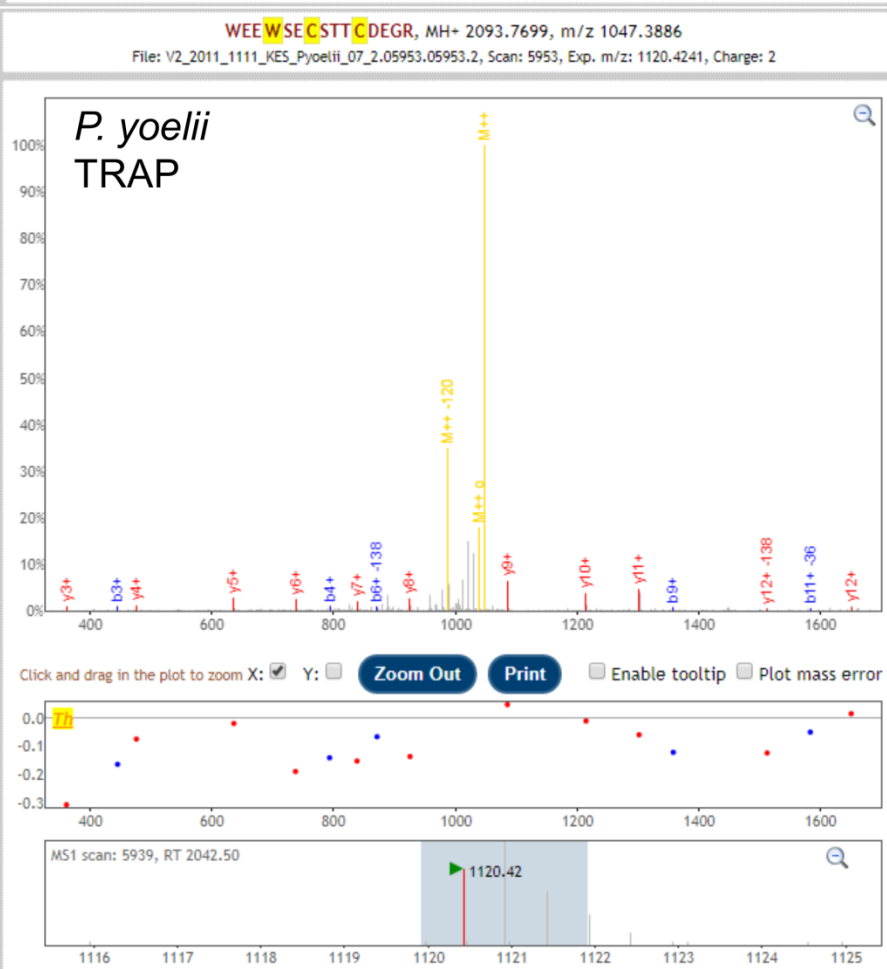
☒ Most Intense
☐ Nearest Match
☒ Peak Detect

Peak Labels:

☒ Ion ☐ m/z
☐ None

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Height: 400



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445.1718	3	E	13	1778.6480
793.3039	4	W	12	1649.6054
880.3359	5	S	11	1301.4733
1009.3785	6	E	10	1214.4412
1169.4092	7	C	9	1085.3986
1256.4412	8	S	8	925.3680
1357.4889	9	T	7	838.3360
1458.5366	10	T	6	737.2883
1618.5672	11	C	5	636.2406
1733.5942	12	D	4	476.2100
1862.6368	13	E	3	361.1830
1919.6582	14	G	2	232.1404
	15	R	1	175.1190

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Variable Modifications:
C: 57.021464 [7, 11]
W: 162.052824 [4]

Figure S14. Automated identification of TSR glycosylation in *P. yoelii* salivary gland sporozoites. A previously published proteomic analysis of salivary gland sporozoites of the rodent-infective *Plasmodium* species *P. yoelii* (Lindner and Swearingen *et al.* 2013 PMID:23325771) was re-analyzed with the mass offset approach described here, providing the first reported evidence of O-fucosylation and C-mannosylation of TSR domains in that species. Sample MS² spectra for the CSP peptide (top) and the TRAP peptide (bottom) are shown as automatically annotated by the Lorikeet spectrum viewer incorporated into the TPP. Both examples show a mass difference of -146.06 Da between the observed precursor *m/z* and that of the matched peptide, corresponding to neutral loss of O-Fuc. The C-mannosylated peptide precursor after neutral loss of O-Fuc is the dominant peak in both spectra (yellow peak annotated M++). In the CSP spectrum, this peak is isobaric with the b11-120 Da peak). Neutral loss of 120.04 Da from the M++ peak due to cross-ring cleavage of C-Man is also indicated.

Figure S15. Example false positive peptides showing neutral loss of O-linked glycans.

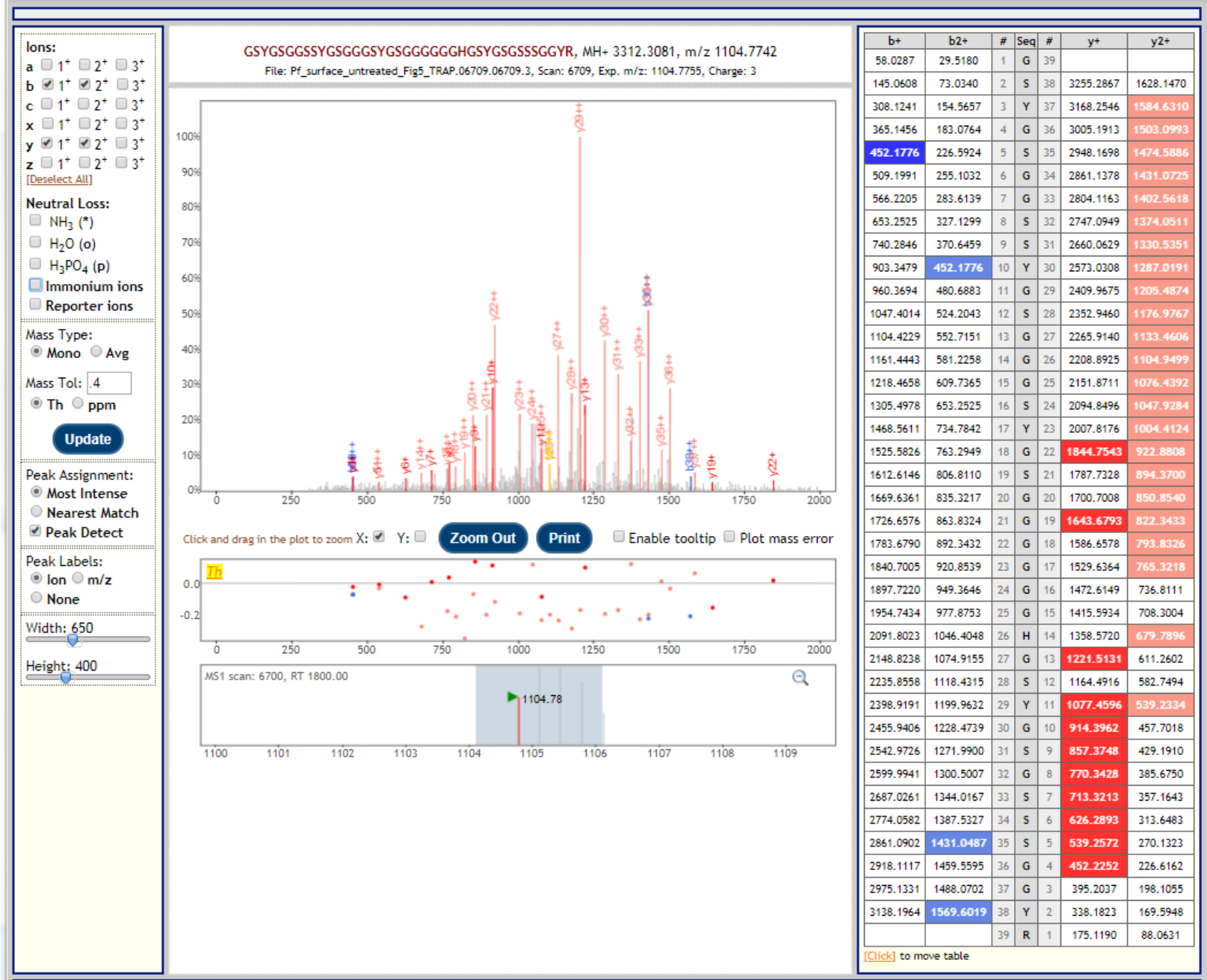


Figure S15a. A sample high-quality PSM identified the fully tryptic human keratin peptide GSYGSGGSSYSGGGSYSGGGGGGHGSYSGSSSGGYR.

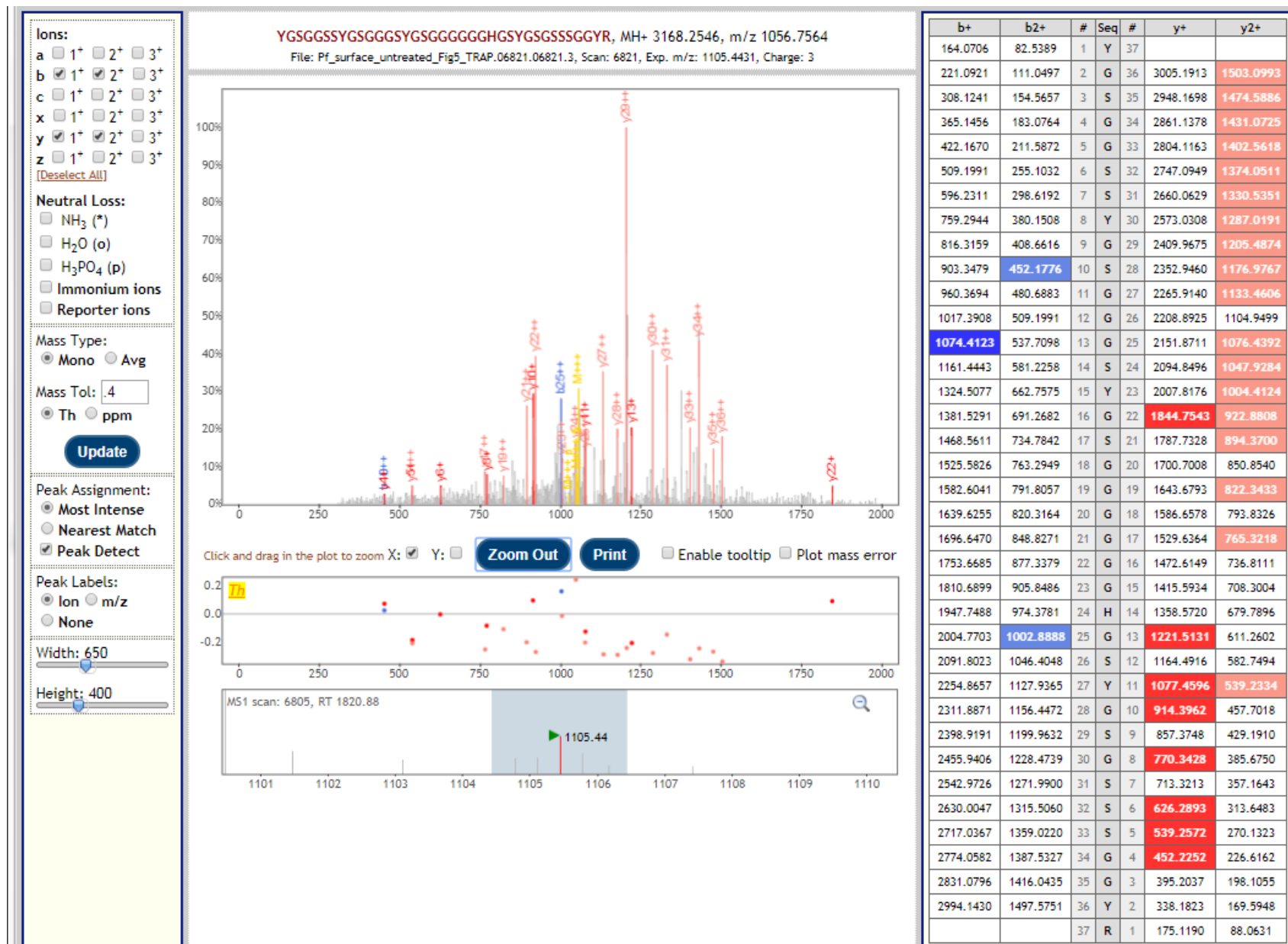


Figure S15b. A representative PSM identifying the same keratin peptide seen in Figure S15a, but spuriously identifying it as a semi-trypic fragment with neutral loss of 146.06. The semi-trypic fragment GS.YGS GGSYGS GGSYGS GGGGGHGSYGS GSSSGGYR has a mass loss of Gly-Ser = 144.05 Da. In addition, the +2H isotope peak 1105.44 *m/z* was incorrectly selected as the monoisotopic peak for this peptide. The total mass loss is then 144.05 + 2*1.0078 = 146.07, which is within the matching tolerance of the searched-for neutral loss of 146.06 Da for putative loss of O-Fuc. Because of the extensive y-ion coverage, the PSM still received a high score. The more likely correct annotation is shown in Figure S15c.

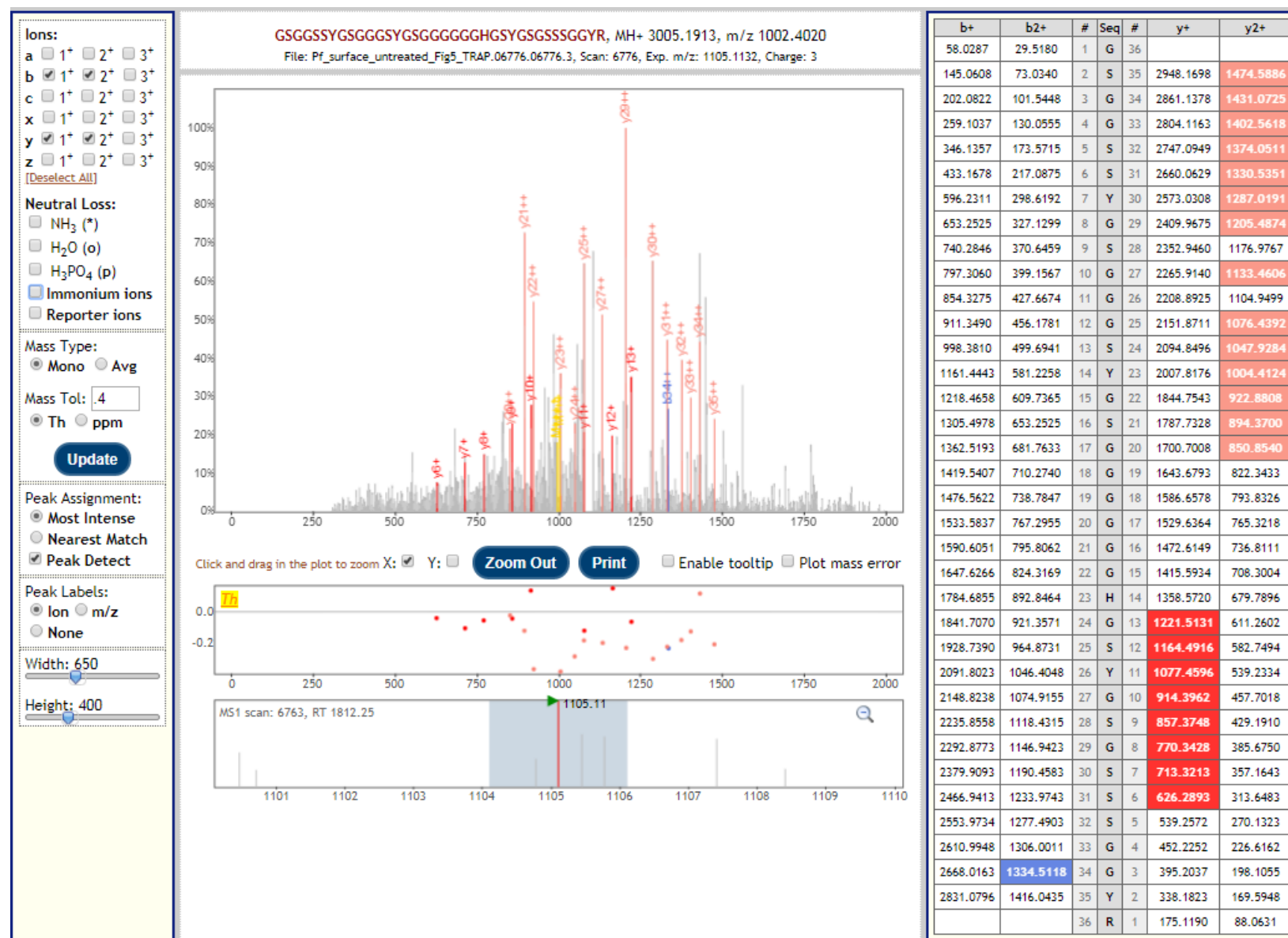


Figure S15d. A representative PSM identifying the same keratin peptide seen in Figure S15a, but spuriously identifying it as a semi-tryptic fragment with neutral loss of 308.11. The semi-tryptic fragment GSY.GSGGSSYSGGGSYSGGGGGHGSYSGSSSGGYR has a mass loss of Gly-Ser-Tyr = 307.12 Da. In addition, the +1H isotope peak 1105.11 *m/z* was incorrectly selected as the monoisotopic peak for this peptide. The total mass loss is then 307.12 + 1.0078 = 308.12, which is within the matching tolerance of the searched-for neutral loss of 308.11 Da for putative loss of O-Fuc-Glc. Because of the extensive y-ion coverage, the PSM still received a high score. The correct annotation is shown in Figure S15e.

