

Supporting Information

Memory of Chirality in Bromoalkyne Carbocyclization: Applications in Asymmetric Total Synthesis of Hasubanan Alkaloids

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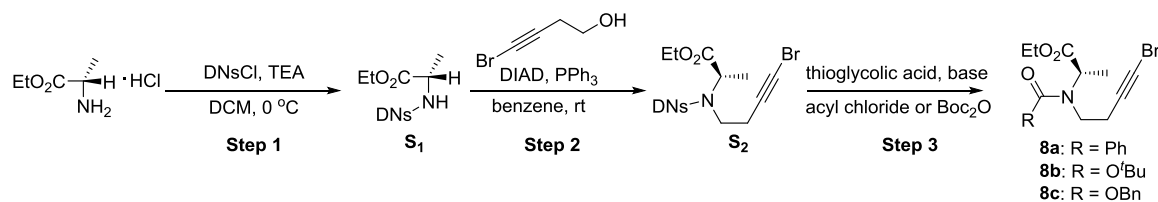
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1. Methods and Materials

All of the chemicals were of reagent grade and were used as purchased. All of the reactions were performed under an inert atmosphere consisting of dry nitrogen using distilled dry solvents. The reactions were monitored by thin layer chromatography (TLC) analysis using silica gel 60 F-254 TLC plates. The compound spots were visualized using UV light (254 nm). The melting points were measured by using a Buchi B-540 melting point apparatus without correction. Flash column chromatography was carried out on silica gel (230–400 mesh). The optical rotations were measured by using sodium light (D line 589.3 nm), and the values are given as specific optical rotation with exact temperature, concentration ($c/(10 \text{ mg/mL})$) and solvent. ^1H NMR (800, 600, 500, or 400 MHz) and ^{13}C NMR (200, 150, 125, or 100 MHz) spectra were referenced to Me_4Si (0 ppm), residual CHCl_3 (^1H NMR $\delta = 7.24 \text{ ppm}$, ^{13}C NMR $\delta = 77.16 \text{ ppm}$). The splitting patterns are indicated as s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet) and brs (broad singlet) for the ^1H NMR data. The IR spectra were measured on a Fourier Transform Infrared spectrometer. The high-resolution mass spectra (HRMS) were recorded using fast atom bombardment (FAB) or quadrupole time-of-flight (Q-TOF). HPLC was performed on an Agilent 1200 series or Agilent 1000 series instrument with a UV detector and CHIRALCEL OD–H column ($0.46 \times 25 \text{ cm}$).

2. General Procedures

Procedures for the preparation of 8



Step 1: 2,4-Dinitrobenzenesulfonyl chloride (DNsCl) (2.86 g, 10.7 mmol, 1.10 eq) was added to L-alanine ethyl ester hydrochloride (1.50 g, 9.76 mmol) in DCM (40 mL) at 0 °C. Then, TEA (3.00 mL, 21.5 mmol, 2.20 eq) in DCM (20 mL) was slowly added into the mixture with a syringe pump for more than 30 min. After the reaction mass was maintained between 0 °C–20 °C for 5 h, the reaction was quenched with H₂O (30 mL), warmed to room temperature, and extracted with DCM (3×30 mL). The combined organic layers were washed with brine (20 mL), dried over Na₂SO₄ and concentrated in vacuo. The brown residue was purified by flash column chromatography (Hexane:EtOAc = 3:1) to yield **S**₁ (2.75 g, 7.91 mmol, 81%) as a yellow oil.

Rf 0.40 (Hexane:EtOAc = 2:1), light yellow oil.

¹H NMR (400 MHz, CDCl₃) δ = 8.65 (d, *J* = 2.2 Hz, 1H), 8.49 (dd, *J* = 8.6, 2.2 Hz, 1H), 8.25 (d, *J* = 8.6 Hz, 1H), 6.18 (s, 1H), 4.22 (brs, 1H), 4.00–3.94 (m, 2H), 1.46 (d, *J* = 7.2 Hz, 3H), 1.10 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 171.3, 149.6, 147.6, 139.4, 132.1, 127.1, 120.7, 61.9, 52.6, 19.2, 13.8.

IR (neat, cm⁻¹): 3326, 3106, 2989, 1733, 1538, 1348, 1172, 1133.

HRMS (FAB): calcd for C₁₁H₁₄N₃O₈S [M+H]⁺ 348.0502, found 348.0500.

[α]_D²⁰ -162.8 (c = 0.5, CHCl₃).

Step 2: To a stirred solution of the sulfonamide **S**₁ (842 mg, 2.43 mmol), 4-bromo-3-butyn-1-ol¹ (539 mg, 3.64 mmol, 1.50 eq), and PPh₃ (1.27 g, 4.85 mmol, 2.00 eq) in benzene (40 mL), DIAD (1.02 mL, 4.85 mmol, 2.00 eq) in benzene (15 mL) was added dropwise with a syringe pump for more than 15 min at room temperature under an inert atmosphere. After 1 h, the mixture was concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (Hexane:EtOAc = 5:1) to yield **S**₂ (1.09 g, 2.29 mmol, 94%) as a light yellow oil.

Rf 0.40 (Hexane:EtOAc = 3:1), light yellow oil.

¹H NMR (400 MHz, CDCl₃) δ = 8.49 (dd, *J* = 8.7, 2.3 Hz, 1H), 8.40 (d, *J* = 2.1 Hz, 1H), 8.30 (d, *J* = 8.7 Hz, 1H), 4.75 (q, *J* = 7.4 Hz, 1H), 4.11–3.99 (m, 2H), 3.63 (ddd, *J* = 15.5, 10.0, 5.4 Hz, 1H), 3.24 (ddd, *J* = 15.4, 9.9, 5.8 Hz, 1H), 2.73 (qd, *J* = 16.4, 5.3 Hz, 1H), 2.54 (qd, *J* = 16.5, 5.4 Hz, 1H), 1.57 (d, *J* = 7.4 Hz, 3H), 1.17 (t, *J* = 7.1 Hz, 3H).

¹³C NMR (100 MHz, CDCl₃) δ = 170.8, 149.7, 148.1, 137.9, 132.7, 126.0, 119.6, 76.2, 62.0, 56.9, 45.2, 41.4, 22.3, 16.9, 14.0.

IR (neat, cm⁻¹): 3101, 2987, 1736, 1555, 1540, 1353, 1157.

HRMS (FAB) C₁₅H₁₇BrN₃O₈S [M+H]⁺ 477.9920, found 477.9933.

[α]_D²⁰ +27.4 (c = 0.5, CHCl₃).

Step 3: For synthesis of benzoyl amide (8a): To a solution of **S**₂ (273 mg, 0.57 mmol) and thioglycolic acid (79.6 μ L, 1.14 mmol, 2.00 eq) in DCM (10 mL), TEA (241 μ L, 1.72 mmol, 3.00 eq) in DCM (4 mL) was added dropwise at room temperature under an inert atmosphere. The reaction was stirred until judged to be complete by TLC analysis. After quenching with sat. NaHCO₃ solution (10 mL), the mixture was extracted with DCM (2 \times 15 mL). The crude residue was filtered with a pad of silica and celite to yield pure secondary amine as a light yellow oil. TEA (161 μ L, 1.14 mmol, 2.00 eq) and benzoyl chloride (99.7 μ L, 0.86 mmol, 1.50 eq) were added to the above secondary amine in DCM (5 mL) at 0 $^{\circ}$ C. After 10 min, the reaction was quenched with H₂O (5 mL) and extracted with DCM (2 \times 10 mL). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography (Hexane:EtOAc = 3:1) to yield **8a** (173 mg, 0.49 mmol, 86%) as a colorless oil.

R_f 0.30 (Hexane:EtOAc = 3:1), colorless oil. Rotamers were observed, and the ratio was 2:3 from ¹H NMR.

¹H NMR (800 MHz, CDCl₃) δ = 7.39–7.36 (m, 5H), 4.44–4.41 (m, 1H), 4.22–4.15 (m, 2H), 3.75 (s, 0.5H), 3.48–3.43 (m, 0.9H), 3.22 (s, 0.6H), 2.67 (s, 1.1H), 2.39 (s, 0.9H), 1.59 (s, 1.2H), 1.43 (s, 1.8H), 1.25 (s, 3H).

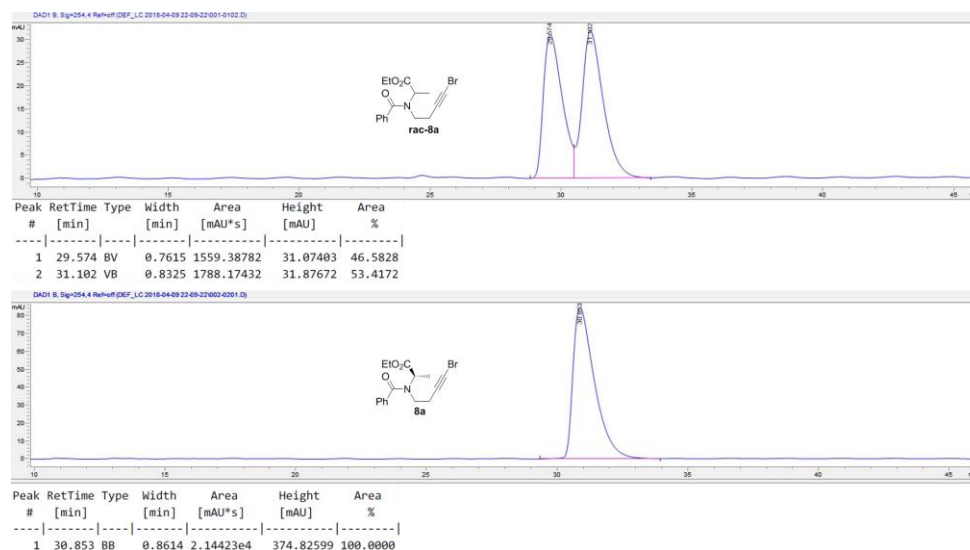
¹³C NMR (200 MHz, CDCl₃) δ = 172.6/171.8, 171.2, 136.0 (2C), 129.8/129.7, 128.6, 126.4 (2C), 77.7/76.4, 61.6/61.3, 57.6/55.0, 47.5/42.5, 40.9/39.8, 20.7/19.2, 16.1/14.9, 14.1.

IR (neat, cm⁻¹): 2984, 2938, 2344, 1736, 1639, 1219, 1075.

HRMS (FAB) calcd for C₁₆H₁₉BrNO₃ [M+H]⁺ 352.0548, found 352.0542.

[α]_D²⁰ -64.1 (c = 0.5, CHCl₃).

HPLC analysis Chiralpak OD-H (Hexane/*i*PrOH = 95/5, 0.5 mL/min, 254 nm, 25 $^{\circ}$ C); 30.9 min; >99% ee.



Step 3: For synthesis of Boc amide (8b): To a solution of **S**₂ (193 mg, 0.41 mmol) and thioglycolic acid (42.2 μ L, 0.62 mmol, 1.50 eq) in DCM (5 mL), DIPEA (141 μ L, 0.81 mmol, 2.00 eq) in DCM (2 mL) was added dropwise at room temperature under an inert atmosphere. After the starting material disappeared, DIPEA (141 μ L, 0.81 mmol, 2.00 eq) and Boc₂O (133 mg, 0.62 mmol, 1.50 eq) were added to the above complex at room temperature. The reaction mixture was stirred for another 12 h at room temperature before the addition of H₂O (10 mL), and extracted with DCM (3 \times 10 mL). The combined organic layers were washed with brine (10 mL), dried over Na₂SO₄ and

concentrated in vacuo. The crude residue was purified by flash column chromatography on silica gel (Hexane:EtOAc = 10:1) to yield **8b** (127 mg, 0.36 mmol, 90%) as a colorless oil.

Rf 0.50 (Hexane:EtOAc = 5:1), colorless oil. Rotamers were observed, and the ratio was 1:1 from ¹H NMR.

¹H NMR (600 MHz, CDCl₃) δ = 4.45–4.44 (m, 0.5H), 4.14–4.07 (m, 2.5H), 3.48–3.40 (m, 1H), 3.33–3.30 (m, 0.5H), 3.21–3.18 (m, 0.5H), 2.52–2.41 (m, 2H), 1.44–1.38 (m, 12H), 1.25–1.21 (m, 3H).

¹³C NMR (150 MHz, CDCl₃) δ = 172.2/172.1, 155.1/154.6, 80.6/80.5, 78.0/77.7, 61.1, 56.5/55.0, 46.1/44.8, 39.5/39.3, 28.3/28.2 (3C), 20.7/20.0, 16.1/15.5, 14.1.

IR (neat, cm⁻¹): 2980, 2939, 1740, 1696, 1367, 1163, 1073, 1035.

HRMS (FAB) calcd for C₁₄H₂₃BrNO₄ [M+H]⁺ 348.0810, found 348.0819.

[α]_D²⁰ -22.2 (c = 0.5, CHCl₃).

Step 3: For synthesis of Cbz amide 8c: To a solution of **S2** (500 mg, 1.05 mmol) and thioglycolic acid (146 μL, 2.10 mmol, 2.00 eq) in DCM (15 mL), TEA (441 μL, 3.15 mmol, 3.00 eq) in DCM (5 mL) was added dropwise at room temperature under an inert atmosphere. The reaction was stirred until judged to be complete by TLC analysis. After quenching with sat. NaHCO₃ solution (10 mL), the mixture was extracted with DCM (2×20 mL). The crude residue was filtered with a pad of silica and celite to yield pure secondary amine as a light yellow oil. NaHCO₃ (176 mg, 2.10 mmol, 2.00 eq) and benzyl chloroformate (149 μL, 1.05 mmol, 1.00 eq) were added to the above secondary amine in H₂O/acetone (1:1, 6 mL) at 0 °C. After 15 min, the reaction was quenched with H₂O (5 mL) and extracted with EtOAc (2×10 mL). The combined organic layers were washed with brine (5 mL), dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography (Hexane:EtOAc = 7:1) to yield **8c** (336 mg, 0.88 mmol, 84%) as a colorless oil.

Rf 0.50 (Hexane:EtOAc = 5:1), colorless oil. Rotamers were observed, and the ratio was 1.2:1 from ¹H NMR.

¹H NMR (400 MHz, CDCl₃) δ = 7.32–7.29 (m, 5H), 5.17–5.06 (m, 2H), 4.52 (q, *J* = 7.3 Hz, 0.55H), 4.32 (q, *J* = 7.1 Hz, 0.45H), 4.13 (q, *J* = 6.9 Hz, 1.1H), 4.08–3.93 (m, 0.9H), 3.56–3.48 (m, 1H), 3.39–3.25 (m, 1H), 2.59–2.39 (m, 2H), 1.45 (d, *J* = 7.2 Hz, 3H), 1.21 (t, *J* = 7.0 Hz, 1.6H), 1.12 (t, *J* = 7.0 Hz, 1.4H).

¹³C NMR (100 MHz, CDCl₃) δ = 171.93/171.86, 156.0/155.6, 136.5/136.3, 128.6/128.5 (2C), 128.3 (2C), 128.1/127.9, 77.9/77.5, 67.5, 61.3, 56.0/55.8, 46.1/44.8, 40.1/39.8, 20.8/20.0, 16.2/15.6, 14.2/14.1.

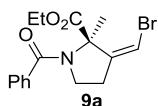
IR (neat, cm⁻¹): 2984, 2945, 1739, 1702, 1472, 1416, 1369, 1295, 1215, 1184, 1072, 1020.

HRMS (FAB) calcd for C₁₇H₂₁BrNO₄ [M+H]⁺ 382.0654, found 382.0652.

[α]_D²⁰ -16.1 (c = 0.5, CHCl₃).

MOC reaction procedure

To a solution of haloalkyne substrates (**8a–8c**, 0.10 mmol) in DMF (5 mL, 0.02 M), KO^tBu (0.15 mmol, 1.50 eq for **8a** and **8c**, 2.00 eq for **8b**) was added at 0 °C. After 5 min, sat. NH₄Cl (5 mL) and H₂O (5 mL) were added, and the aqueous phase was extracted with EtOAc (4×20 mL). The combined organic phases were dried over Na₂SO₄ and concentrated by high-vacuum evaporation. The residue was purified by flash column chromatography on silica gel to afford analytically pure products **9a–9c**.



Prepared with **8a** according to the general MOC procedure in 78% yield.

Rf 0.30 (Hexane:EtOAc = 3:1), White solid (m.p. = 129–131°C)

¹H NMR (400 MHz, CDCl₃) δ = 7.44–7.35 (m, 5H), 6.14 (t, *J* = 1.9 Hz, 1H), 4.30–4.20 (m, 2H), 3.66–3.59 (m, 2H), 2.78–2.62 (m, 2H), 1.92 (s, 3H), 1.29 (t, *J* = 7.1 Hz, 3H).

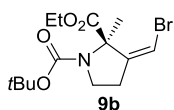
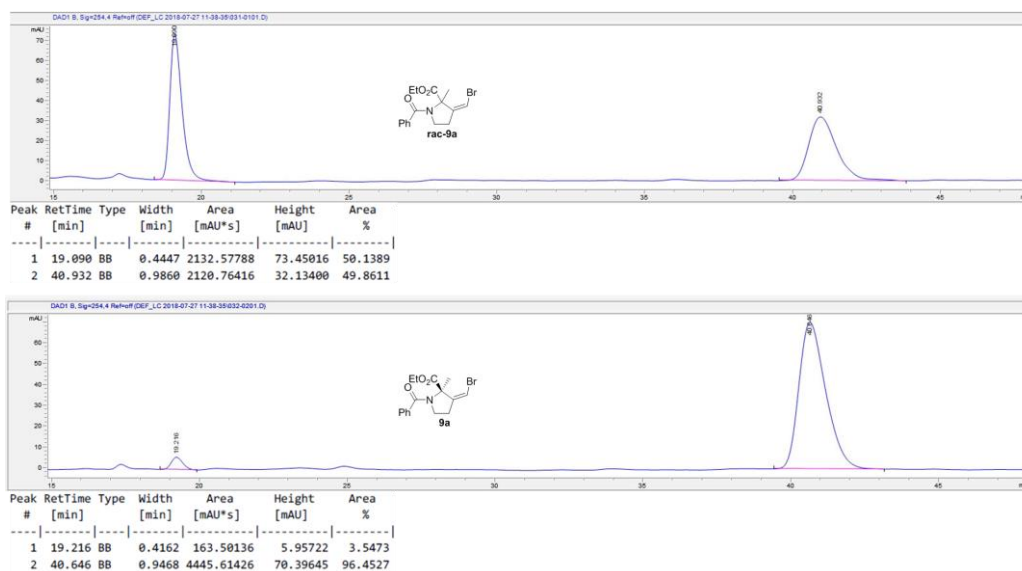
¹³C NMR (100 MHz, CDCl₃) δ = 169.4, 168.8, 145.5, 136.4, 130.0, 128.3 (2C), 126.8 (2C), 98.9, 69.2, 61.4, 48.5, 34.1, 18.7, 14.2.

IR (neat, cm⁻¹): 3062, 2981, 1746, 1632, 1405, 1250, 1023.

HRMS (FAB) calcd for C₁₆H₁₉BrNO₃ [M+H]⁺ 352.0548, found 352.0560.

[α]_D²⁰ -118.98 (c = 0.5, CH₂Cl₂).

HPLC analysis Chiralpak OD–H (Hexane/*i*PrOH = 85/15, 0.5 mL/min, 254 nm, 25 °C); 19.7 (minor), 40.9 (major) min; 93% ee.



Prepared with **8b** according to the general MOC procedure in 81% yield.

Rf 0.40 (Hexane:EtOAc = 5:1), colorless oil. Rotamers were observed, and the ratio was 1:2 from ¹H NMR.

¹H NMR (600 MHz, CDCl₃) δ = 6.05 (t, *J* = 2.1 Hz, 1H), 4.23–4.10 (m, 2H), 3.60–3.53 (m, 2H), 2.65–2.59 (m, 2H), 1.73 (s, 1H), 1.70 (s, 2H), 1.40 (s, 9H), 1.26–1.24 (m, 3H).

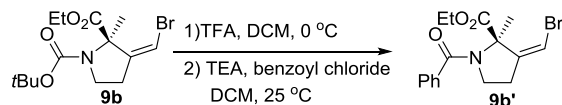
¹³C NMR (150 MHz, CDCl₃) δ = 170.34/170.25, 153.6/153.4, 147.4/146.4, 98.1, 80.8/80.1, 68.4/67.9, 61.4/61.3, 46.2/45.8, 33.0/32.3, 28.3/28.2 (3C), 20.2/19.6, 14.1.

IR (neat, cm⁻¹): 3072, 2979, 2937, 2845, 1751, 1698, 1385, 1367, 1249, 1163, 1114, 1056.

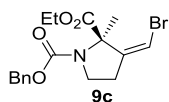
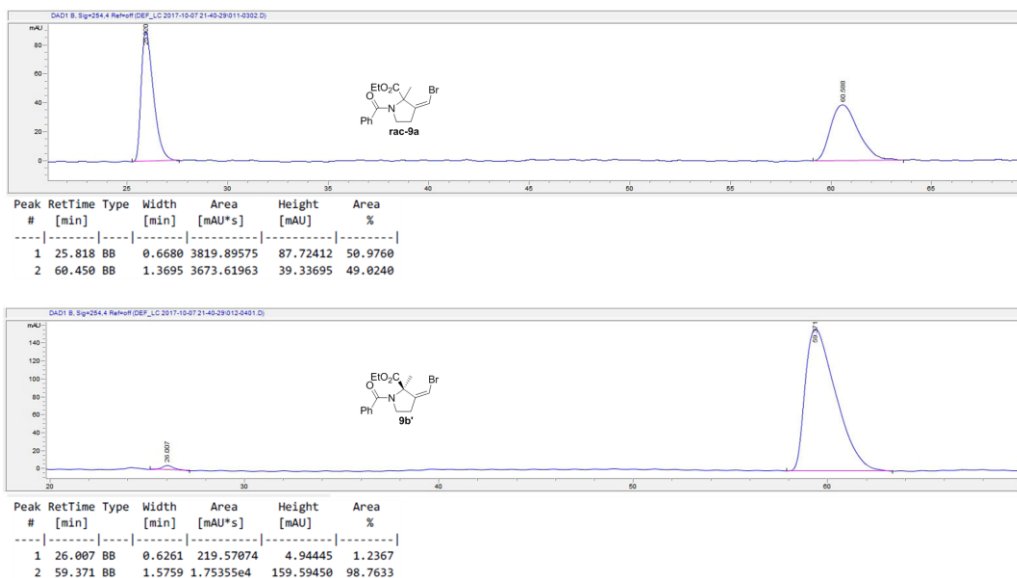
HRMS (FAB) calcd for C₁₄H₂₃BrNO₄ [M+H]⁺ 348.0810, found 348.0810.

[α]_D²⁰ -32.96 (c = 0.5, CHCl₃).

HPLC analysis Because **9b** can not be fully separated after trying a few chiral columns and conditions, we transform **9b** to **9b'** for HPLC analysis. Chiralpak OD-H (Hexane/*i*PrOH = 90/10, 0.5 mL/min, 254 nm, 25 °C); 26.0 (minor), 59.4 (major) min; 98% ee.



Trifluoroacetic acid (0.5 mL) was added to a solution of compound **9b** (16.5 mg, 0.047 mmol) in DCM (1.5 mL) and the mixture was stirred at room temperature for 30 min. The mixture was concentrated, dissolved in DCM (20 mL), washed with aq. NaHCO₃ and concentrated. The residue was redissolved in DCM (3 mL) and treated with TEA (27 μL, 0.19 mmol, 4.00 eq) and benzoyl chloride (12 μL, 0.095 mmol, 2.00 eq). The mixture was stirred at room temperature for 2 h, and diluted with DCM, and then, the organic phase was washed with water and concentrated. The residue was purified with flash purification by column chromatography on silica gel (Hexane:EtOAc = 3:1) to afford compound **9b'** (14.7 mg, 0.042 mmol, 88%) as a white solid.



Prepared with **8c** according to the general MOC procedure in 78% yield.

Rf 0.40 (Hexane:EtOAc = 5:1), colorless oil. Rotamers were observed, and the ratio was 1.3:1 from ¹H NMR.

¹H NMR (400 MHz, CDCl₃) δ = 7.34–7.24 (m, 5H), 6.08 (s, 1H), 5.14–5.06 (m, 2H), 4.18 (q, *J* = 7.2 Hz, 1H), 3.95–3.88 (m, 1H), 3.69–3.63 (m, 2H), 2.70–2.63 (m, 2H), 1.74 (d, *J* = 20.4 Hz, 3H), 1.21 (t, *J* = 7.0 Hz, 1.7H), 1.06 (t, *J* = 7.0 Hz, 1.3H).

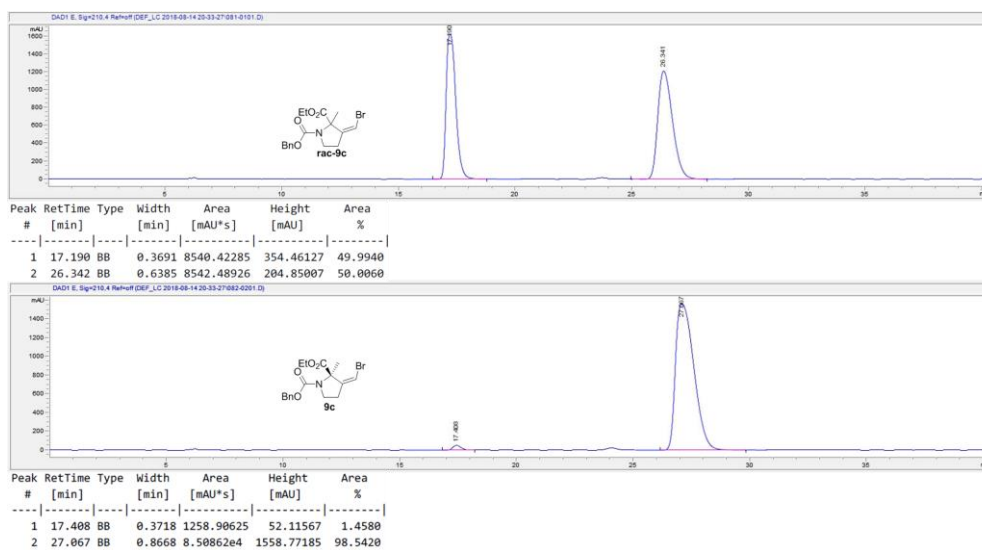
¹³C NMR (100 MHz, CDCl₃) δ = 170.4/170.3, 154.3/154.2, 147.3/146.3, 136.8/136.2, 128.7, 128.4, 128.3, 128.2, 128.1, 98.8/98.6, 69.1/68.3, 67.6/67.1, 61.7, 46.6/46.1, 33.1/32.4, 20.5/19.6, 14.2/14.1.

IR (neat, cm⁻¹): 2982, 2939, 2888, 1750, 1701, 1403, 1350, 1251, 1114, 1087, 1055.

HRMS (FAB) calcd for C₁₇H₂₁BrNO₄ [M+H]⁺ 382.0654, found 382.0652.

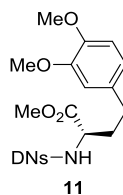
$[\alpha]_D^{20}$ -21.56 (c = 0.5, CHCl₃).

HPLC analysis Chiralpak OD–H (Hexane/*i*PrOH = 90/10, 0.5 mL/min, 210 nm, 25 °C); 17.4 (minor), 27.1 (major) min; 97% ee.



3. Total Synthesis of (–)-Runanine (2)

Methyl (*R*)-4-(3,4-dimethoxyphenyl)-2-((2,4-dinitrophenyl)sulfonamido)butanoate (**11**)



SOCl₂ (910 mL, 12.5 mmol, 1.50 eq) was slowly added to a cooled solution of the (*R*)-3,4-dimethoxy-homophenylalanine **10** (2.00 g, 8.36 mmol, 98% ee) in dry methanol (30 mL). The mixture was refluxed for 5 h and left at room temperature under stirring for 2 h. Methanol was then removed and the crude product was washed with toluene three times to give (*R*)-3,4-dimethoxy-homophenylalanine methyl ester hydrochloride as a white solid. 2,4-Dinitrobenzenesulfonyl chloride (2.45 g, 9.20 mmol, 1.10 eq) was added to (*R*)-3,4-dimethoxy-homophenylalanine methyl ester hydrochloride in DCM (50 mL) at 0 °C. Then, TEA (2.58 mL, 18.4 mmol, 2.20 eq) in DCM (20 mL) was added into the mixture slowly with a syringe pump for more than 30 min. After 5 h at 0 °C – 20 °C under an inert atmosphere, the reaction was quenched with H₂O (30 mL), warmed to room temperature, and extracted with DCM (2 × 50 mL). The combined organic layers were washed with brine (20 mL), dried over Na₂SO₄ and concentrated in vacuo. The brown residue was purified by flash column chromatography on silica gel (Hexane:EtOAc = 2:1) to yield **11** (3.41 g, 6.94 mmol, 83% overall yield) as a light yellow oil.

Rf 0.20 (Hexane:EtOAc = 2:1), light yellow oil.

¹H NMR (400 MHz, CDCl₃) δ = 8.70 (d, *J* = 2.2 Hz, 1H), 8.46 (dd, *J* = 8.6, 2.2 Hz, 1H), 8.18 (d, *J* = 8.6 Hz, 1H), 6.76 (d, *J* = 7.8 Hz, 1H), 6.69–6.67 (m, 2H), 6.16 (d, *J* = 9.2 Hz, 1H), 4.45 (td, *J* = 9.0 Hz, 1H), 3.84 (s, 3H), 3.82 (s, 3H), 3.47 (s, 3H), 2.76–2.63 (m, 2H), 2.21–2.13 (m, 1H), 2.05–1.96 (m, 1H).

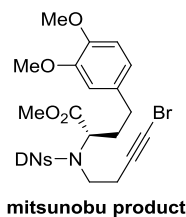
¹³C NMR (100 MHz, CDCl₃) δ = 171.4, 149.7, 148.8, 147.7, 147.5, 139.4, 132.1, 132.0, 127.1, 120.9, 120.3, 111.9, 111.3, 56.2, 55.8, 55.7, 52.6, 34.5, 30.8.

IR (neat, cm⁻¹): 3309, 3102, 2953, 2935, 2837, 1738, 1537, 1513, 1347, 1259, 1234, 1165, 1100, 1025.

HRMS (FAB) calcd for C₁₉H₂₁N₃O₁₀S [M]⁺ 483.0948, found 483.0946.

[α]_D²⁰ +70.36 (c = 0.5, CHCl₃).

Methyl (*R*)-2-((*N*-(4-bromobut-3-yn-1-yl)-2,4-dinitrophenyl)sulfonamido)-4-(3,4-dimethoxyphenyl)butanoate (mitsunobu product)



PPh₃ (1.18 g, 4.51 mmol, 2.00 eq) and 4-bromo-3-butyn-1-ol (0.50 g, 3.39 mmol, 1.50 eq) were added to a solution of sulfonamide **11** (1.09 g, 2.26 mmol) in benzene (40 mL) at room temperature. DIAD (0.90 mL, 4.51 mmol, 2.00 eq) in benzene (15 mL) was added slowly to the mixture with a syringe pump for more than 20 min under an inert

atmosphere. After 1 h, the mixture was concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (Hexane:EtOAc = 3:1) to yield **mitsunobu product** (1.31 g, 2.14 mmol, 95%) as a yellow oil.

Rf 0.30 (Hexane:EtOAc = 2:1), yellow oil.

¹H NMR (400 MHz, CDCl₃) δ = 8.46 (dd, J = 8.7, 2.2 Hz, 1H), 8.39 (d, J = 2.1 Hz, 1H), 8.20 (d, J = 8.8 Hz, 1H), 6.80 (d, J = 7.9 Hz, 1H), 6.74–6.71 (m, 2H), 4.61 (dd, J = 9.7, 4.8 Hz, 1H), 3.87 (s, 3H), 3.85 (s, 3H), 3.65 (ddd, J = 15.5, 10.2, 5.3 Hz, 1H), 3.59 (s, 3H), 3.30 (ddd, J = 15.6, 10.1, 5.6 Hz, 1H), 2.84 (ddd, J = 16.3, 10.4, 5.6 Hz, 1H), 2.77–2.68 (m, 2H), 2.55 (ddd, J = 16.4, 10.4, 5.9 Hz, 1H), 2.41–2.32 (m, 1H), 2.03–1.94 (m, 1H).

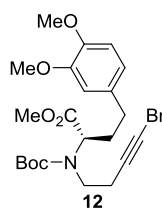
¹³C NMR (100 MHz, CDCl₃) δ = 170.8, 149.7, 149.0, 148.1, 147.6, 137.3, 132.8, 132.4, 125.9, 120.3, 119.6, 111.9, 111.4, 76.1, 60.9, 55.91, 55.88, 52.7, 45.7, 41.6, 32.0, 31.8, 22.1.

IR (neat, cm⁻¹): 3101, 2953, 2936, 2837, 1739, 1554, 1537, 1514, 1349, 1257, 1236, 1160, 1137, 1107, 1026.

HRMS (FAB) calcd for C₂₃H₂₄BrN₃O₁₀S [M]⁺ 613.0366, found 613.0375.

[α]_D²⁰ -38.72 (c = 0.5, CHCl₃).

Methyl (*R*)-2-((4-bromobut-3-yn-1-yl)(*tert*-butoxycarbonyl)amino)-4-(3,4-dimethoxyphenyl)butanoate (**12**)



DIPEA (506 μ L, 2.90 mmol, 2.00 eq) in DCM (5 mL) was added dropwise to a solution of **mitsunobu product** (0.89 g, 1.45 mmol) and thioglycolic acid (156 μ L, 2.18 mmol, 1.50 eq) in DCM (25 mL) at room temperature under an inert atmosphere. After the starting material disappeared, DIPEA (506 μ L, 2.90 mmol, 2.00 eq) and Boc₂O (634 mg, 0.62 mmol, 1.50 eq) were added to the above complex at room temperature. The reaction was stirred until judged to be complete by TLC analysis. The complex was quenched with H₂O (20 mL) and extracted with DCM (3 \times 30 mL). The combined organic layers were washed with brine (10 mL), dried over Na₂SO₄ and concentrated, the solvent evaporated and the crude residue was purified by flash column chromatography on silica gel (Hexane:EtOAc = 10:1) to yield **12** (630 mg, 1.30 mmol, 90%) as a colorless oil.

Rf 0.60 (Hexane:EtOAc = 2:1), colorless oil. Rotamers were observed, and the ratio was 1:1 from ¹H NMR.

¹H NMR (400 MHz, CDCl₃) δ = 6.79–6.77 (m, 1H), 6.72–6.70 (m, 2H), 4.40 (m, 0.5H), 3.96 (m, 0.5H), 3.85 (s, 3H), 3.83 (s, 3H), 3.68 (s, 3H), 3.58–3.55 (m, 0.5H), 3.42 (m, 0.5H), 3.21–3.14 (m, 1H), 2.60–2.59 (m, 2H), 2.55–2.45 (m, 2H), 2.34–2.27 (m, 1H), 2.06 (brs, 1H), 1.46 (s, 4.5H), 1.38 (s, 4.5H).

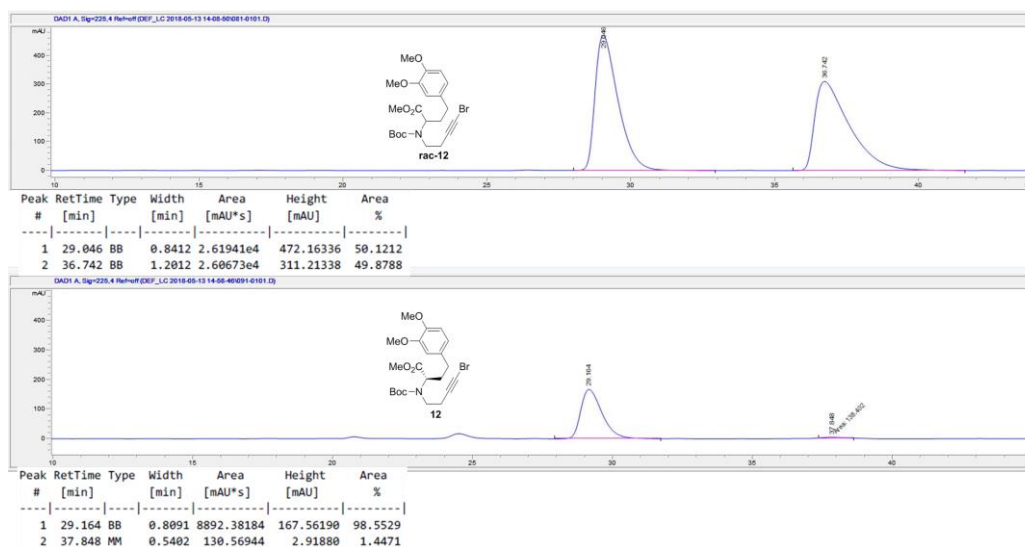
¹³C NMR (100 MHz, CDCl₃) δ = 172.2/172.0, 155.4/154.7, 148.9/147.4, 133.5/133.4, 120.2, 111.72/111.66, 111.3, 80.7, 77.8/77.5, 60.1/58.8, 56.1/56.0, 55.9/55.8 (2C), 52.1, 46.9/45.3, 39.6/39.3, 32.2/31.5, 32.1, 28.3/28.2 (3C), 20.4/19.6.

IR (neat, cm⁻¹): 2974, 2952, 2937, 2837, 1741, 1695, 1515, 1254, 1238, 1157, 1029.

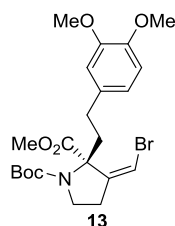
HRMS (FAB) calcd for C₂₂H₃₀BrNO₆ [M]⁺ 483.1256, found 483.1245.

[α]_D²⁰ +11.12 (c = 0.5, CHCl₃).

HPLC analysis Chiralpak OD–H (Hexane/*i*PrOH = 95/5, 0.5 mL/min, 225 nm, 25 °C); 37.8 (minor), 29.2 (major) min; 97% ee.



1-(*Tert*-butyl) 2-methyl (*S,Z*)-3-(bromomethylene)-2-(3,4-dimethoxyphenethyl)pyrrolidine-1,2-dicarboxylate (13**)**



To a solution of compound **12** (500 mg, 1.05 mmol) in DMF (51.5 mL, 0.02 M), KO^tBu (234 mg, 2.10 mmol, 2.00 eq) was added in three portions at 0 °C. After 10 min, The reaction was quenched by the addition of sat. NH₄Cl (25 mL) and concentrated in vacuo. The residue was dissolved in EtOAc (15 mL) and water (15 mL), and extracted with EtOAc (3×20 mL). The combined organic phases were dried over Na₂SO₄ and concentrated under reduced pressure. After purification with flash column chromatography over silica gel (Hexane:EtOAc = 3:1), and compound **13** (397 mg, 0.84 mmol, 79%) was obtained as a colorless oil.

R_f 0.30 (Hexane:EtOAc = 3:1), colorless oil. Rotamers were observed, and the ratio was 1:2 from ¹H NMR.

¹H NMR (400 MHz, CDCl₃) δ = 6.77–6.75 (m, 1H), 6.70–6.68 (m, 2H), 6.17 (s, 1H), 3.83 (s, 3H), 3.82 (s, 3H), 3.70 (s, 3H), 3.66–3.60 (m, 2H), 2.90–2.82 (m, 1H), 2.75–2.69 (m, 1.4H), 2.62–2.52 (m, 1.6H), 2.46–2.35 (m, 2H), 1.43–1.41 (m, 9H).

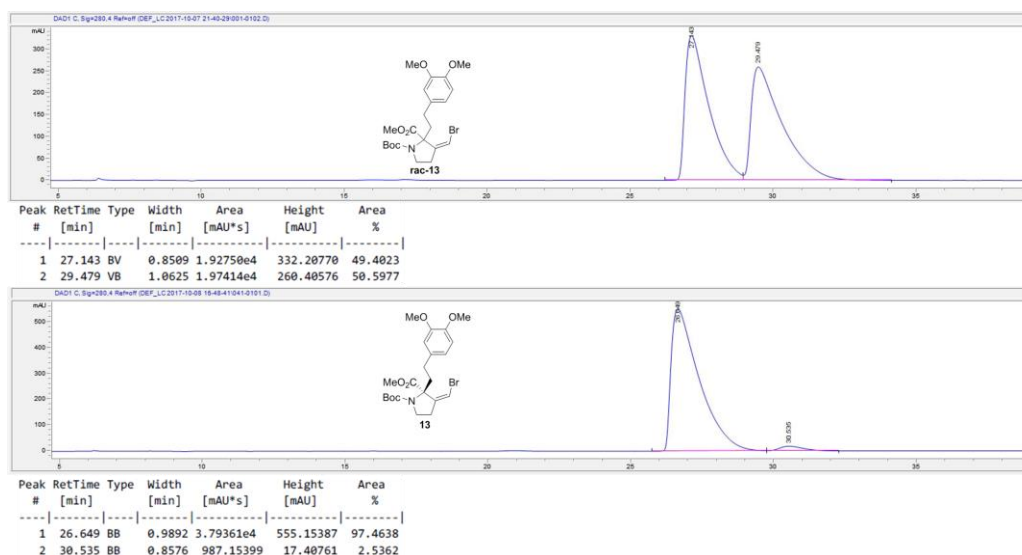
¹³C NMR (125 MHz, CDCl₃) δ = 170.9/170.6, 153.8/153.5, 148.9, 147.3, 145.3/144.4, 134.2, 120.1, 111.7/111.3, 99.1, 81.0/80.3, 71.8/71.3, 55.9, 55.8, 52.3, 46.7/46.5, 34.1/33.8, 33.0/32.6, 30.0, 29.7, 28.3 (3C).

IR (neat, cm⁻¹): 3068, 2975, 2950, 2836, 1750, 1695, 1513, 1454, 1383, 1231, 1158, 1137, 1028.

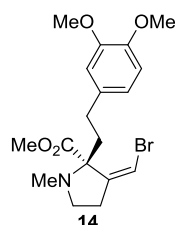
HRMS (FAB) calcd for C₂₂H₃₀BrNO₆ [M]⁺ 483.1256, found 483.1263.

[α]_D²⁰ -10.96 (c = 0.5, CHCl₃).

HPLC analysis Chiralpak OD–H (Hexane/*i*PrOH = 95/5, 0.5 mL/min, 280 nm, 25 °C); 30.5 (minor), 26.6 (major) min; 95% ee.



Methyl (*S,Z*)-3-(bromomethylene)-2-(3,4-dimethoxyphenethyl)-1-methylpyrrolidine-2-carboxylate (**14**)



To a solution of compound **13** (470 mg, 0.97 mmol) in DCM (6 mL), trifluoroacetic acid (2 mL) was added, and the mixture was stirred at room temperature for 1 h. The mixture was quenched by aq. NaHCO₃, extracted by DCM and concentrated in vacuo. The residue was redissolved in methanol (5 mL) and treated with formaldehyde (0.65 mL, 9.73 mmol, 10.0 eq, 37% in H₂O) and sodium cyanoborohydride (557 mg, 2.91 mmol, 3.00 eq). The mixture was stirred at room temperature for 1 h and diluted with DCM (25 mL), and then, the organic phase was washed with water and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (Hexane:EtOAc = 4:1), and compound **14** (0.35 g, 0.88 mmol, 91%) was obtained as a colorless oil.

Rf 0.40 (Hexane:EtOAc = 3:1), colorless oil.

¹H NMR (400 MHz, CDCl₃) δ = 6.77–6.72 (m, 3H), 6.10 (t, J = 1.7 Hz, 1H), 3.85 (s, 3H), 3.83 (s, 3H), 3.69 (s, 3H), 3.07–3.02 (m, 1H), 2.88–2.82 (m, 1H), 2.60 (d, J = 9.6 Hz, 2H), 2.54–2.46 (m, 2H), 2.34 (dd, J = 12.5, 9.2 Hz, 1H), 2.25 (s, 3H), 1.97 (dd J = 12.6, 9.8 Hz, 1H).

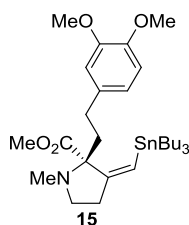
¹³C NMR (100 MHz, CDCl₃) δ = 171.8, 148.7, 147.6, 147.0, 135.3, 120.2, 111.9, 111.1, 98.3, 74.2, 55.9, 55.8, 53.2, 51.3, 34.8, 34.3, 32.2, 29.6.

IR (neat, cm⁻¹): 2946, 2834, 2789, 1728, 1513, 1451, 1258, 1218, 1155, 1029, 1002.

HRMS (FAB) calcd for C₁₈H₂₅BrNO₄ [M+H]⁺ 398.0967, found 398.0970.

[α]_D²⁰ -10.34 (c = 0.5, CHCl₃).

Methyl (*S,Z*)-2-(3,4-dimethoxyphenethyl)-1-methyl-3-((tributylstannyl)methylene)pyrrolidine-2-carboxylate (**15**)



To a solution of compound **14** (215 mg, 0.54 mmol) in diethyl ether (10 mL), *n*BuLi (0.51 mL, 0.81 mmol, 1.50 eq, 1.6 M in hexane) was added at -78 °C. After stirring for 30 min, tributyltin chloride (435 μ L, 1.63 mmol, 3.00 eq) in diethyl ether (3 mL) was added slowly at -78 °C and the mixture was allowed to warm to room temperature for 3 h. The mixture was quenched with H₂O (5 mL) and extracted with EtOAc (3 \times 20 mL). The combined organic layers were washed with brine (20 mL), dried over Na₂SO₄ and concentrated. The residue was purified by flash column chromatography on silica gel (Hexane:EtOAc = 5:1) to yield **15** (223 mg, 0.37 mmol, 68%) as a colorless oil.

Rf 0.20 (Hexane:EtOAc = 10:1), colorless oil.

¹H NMR (500 MHz, CDCl₃) δ = 6.76–6.69 (m, 3H), 5.84 (s, 1H), 3.84 (s, 3H), 3.83 (s, 3H), 3.65 (s, 3H), 3.00–2.96 (m, *J* = 7.9 Hz, 1H), 2.89–2.84 (m, 2H), 2.75–2.68 (m, 1H), 2.56 (dd, *J* = 15.5, 6.3 Hz, 1H), 2.39 (td, *J* = 13.3, 5.0 Hz, 1H), 2.25 (s, 3H), 2.14 (td, *J* = 13.3, 3.8 Hz, 1H), 1.99 (td *J* = 13.2, 4.8 Hz, 1H), 1.65–1.59 (m, 1H), 1.46–1.33 (m, 6H), 1.31–1.20 (m, 6H), 0.91–0.77 (m, 15H).

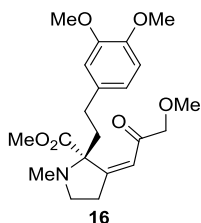
¹³C NMR (125 MHz, CDCl₃) δ = 173.9, 161.0, 148.7, 146.9, 135.6, 120.1, 120.0, 111.8, 111.2, 73.6, 55.9, 55.7, 52.6, 50.9, 37.2, 35.2, 34.4, 29.2, 29.1, 29.0, 28.9, 27.8, 27.6, 27.4, 27.1, 26.8, 17.5, 13.64, 13.58, 10.7.

IR (neat, cm⁻¹): 2954, 2922, 2851, 1725, 1613, 1514, 1462, 1259, 1218, 1156, 1140, 1075, 1032, 1004.

HRMS (FAB) calcd for C₃₀H₅₀NO₄Sn [M-H]⁻ 608.2762, found 608.2768.

[α]_D²⁰ -15.3 (*c* = 0.5, CHCl₃).

Methyl (S,Z)-2-(3,4-dimethoxyphenethyl)-3-(3-methoxy-2-oxopropylidene)-1-methylpyrrolidine-2-carboxylate (16)



To a deoxygenated solution (3 \times freeze – pump – thaw) of the vinylstannane compound **15** (170 mg, 0.28 mmol), PCy₃·HBF₄ (10.3 mg, 0.028 mmol, 0.10 eq), DIPEA (4.90 μ L, 0.028 mmol, 0.10 eq), and Pd₂(dba)₃ (12.8 mg, 0.014 mmol, 0.05 eq) in toluene (5 mL), methoxyacetyl chloride (32.4 μ L, 0.56 mmol, 2.00 eq) was added dropwise. The reaction mixture was heated at 50 °C for 12 h and then cooled and diluted with EtOAc (20 mL). The mixture was washed with 3% aq. NH₄OH (5 mL) and water and then concentrated in vacuo. The residue was purified by flash column chromatography over silica gel (Hexane:EtOAc = 1:1), and compound **16** (80 mg, 0.20 mmol, 74%) was obtained as a yellow oil.

Rf 0.10 (Hexane:EtOAc = 2:1), yellow oil.

¹H NMR (400 MHz, CDCl₃) δ = 6.72 (d, J = 7.9 Hz, 1H), 6.64–6.61 (m, 2H), 6.43 (s, 1H), 3.97 (d, J = 0.9 Hz, 2H), 3.82 (s, 3H), 3.81 (s, 3H), 3.64 (s, 3H), 3.38 (s, 3H), 3.09–3.04 (m, 1H), 2.79–2.72 (m, 2H), 2.70–2.54 (m, 2H), 2.50–2.43 (m, 1H), 2.29 (s, 3H), 2.24–2.04 (m, 2H).

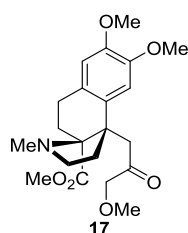
¹³C NMR (100 MHz, CDCl₃) δ = 196.1, 170.4, 165.6, 148.7, 147.0, 135.2, 120.0, 116.8, 111.7, 111.1, 78.0, 74.0, 59.2, 55.9, 55.8, 51.8, 51.0, 35.6, 35.3, 31.2, 29.9.

IR (neat, cm⁻¹): 2946, 2834, 2789, 1735, 1624, 1514, 1452, 1420, 1340, 1260, 1230, 1156, 1141, 1105, 1029.

HRMS (FAB) calcd for C₂₁H₃₀NO₆ [M+H]⁺ 392.2073, found 392.2072.

[α]_D²⁰ -12.66 (c = 0.5, CHCl₃).

Methyl (3a*S*,9b*S*)-7,8-dimethoxy-9b-(3-methoxy-2-oxopropyl)-3-methyl-1,2,3,4,5,9b-hexahydro-3a*H*-benzo[*e*]indole-3a-carboxylate (17)



To a solution of compound **16** (80 mg, 0.20 mmol) in DCM (4 mL), TfOH (90.0 μ L, 1.02 mmol, 5.00 eq) was added dropwise at 0 °C. After 10 min, the mixture was quenched with aq. NaHCO₃ (5 mL), and then extracted with DCM (20 mL). The organic layer was washed with brine (10 mL), dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (Hexane:EtOAc = 2:1), and compound **17** (66.3 mg, 0.17 mmol, 83%) was obtained as a yellow oil.

Rf 0.10 (Hexane:EtOAc=2:1), yellow oil.

¹H NMR (600 MHz, CDCl₃) δ = 6.74 (s, 1H), 6.49 (s, 1H), 3.81 (s, 3H), 3.79 (s, 3H), 3.72 (s, 3H), 3.57 (q, J = 24.1 Hz, 2H), 3.22 (s, 3H), 2.94 (td, J = 7.6, 7.5 Hz, 1H), 2.84–2.80 (m, 1H), 2.72–2.66 (m, 1H), 2.70 (d, J = 14.2 Hz, 1H), 2.62–2.57 (m, 1H), 2.53 (dd, J = 16.3, 3.4 Hz, 1H), 2.46 (d, J = 14.2 Hz, 1H), 2.37 (ddd, J = 12.4, 8.7, 3.2 Hz, 1H), 2.24 (s, 3H), 2.12 (td, J = 13.8, 5.5 Hz, 1H), 2.04 (dd, J = 14.6, 3.7 Hz, 1H).

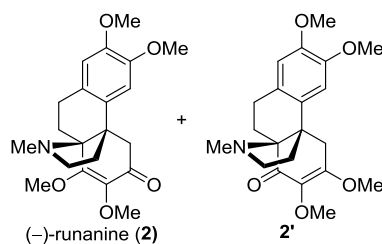
¹³C NMR (150 MHz, CDCl₃) δ = 207.0, 174.2, 147.2, 147.1, 132.3, 127.9, 111.2, 110.8, 78.8, 73.6, 59.0, 56.0, 55.7, 51.4, 51.1, 50.4, 47.5, 37.1, 34.8, 24.8, 23.3.

IR (neat, cm⁻¹): 2939, 2838, 2790, 1719, 1516, 1450, 1357, 1234, 1204, 1134, 1077, 1029, 1016.

HRMS (FAB) calcd for C₂₁H₃₀NO₆ [M+H]⁺ 392.2073, found 392.2071.

[α]_D²⁰ +172.12 (c = 0.5, CHCl₃).

(–)-runanine (2) and (–)-runanine isomer 2'



To a solution of compound **17** (39 mg, 0.10 mmol) in THF (3 mL), KO^tBu (0.15 mL, 0.15 mmol, 1.50 eq, 1.0 M in THF) was added dropwise at 0 °C. After 15 min, the mixture was quenched with sat. NH₄Cl (2 mL) and H₂O (2 mL), and then extracted with DCM and *i*PrOH (2:1, 30 mL) and concentrated in vacuo. The residue was redissolved in MeOH and CH₃CN (1:3, 2 mL) and treated with DIPEA (26.0 μL, 0.15 mmol, 1.50 eq) and (trimethylsilyl)diazomethane (0.25 mL, 0.15 mmol, 1.50 eq, 0.6 M in hexane). The mixture was stirred at room temperature for 12 h, and diluted with DCM (20 mL), and then, the organic phase was washed with water and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (Hexane:EtOAc = 1:2 for **2'**; DCM:MeOH = 20:1 for **2**) to afford the (-)-runanine (**2**) and (-)-runanine isomer **2'** as oil (26.0 mg, 0.07 mmol, 70%, **2**:**2'** = **2**:1).

For (-)-runanine (**2**):

Rf 0.10 (Hexane:EtOAc = 1:2), colorless oil.

¹H NMR (500 MHz, CDCl₃) δ = 6.65 (s, 1H), 6.48 (s, 1H), 4.06 (s, 3H), 3.81 (s, 3H), 3.80 (s, 3H), 3.62 (s, 3H), 2.99 (d, *J* = 15.9 Hz, 1H), 2.83–2.79 (m, 1H), 2.77–2.72 (m, 1H), 2.60 (d, *J* = 16.0 Hz, 1H), 2.55–2.54 (m, 1H), 2.51 (s, 3H), 2.18–2.13 (m, 1H), 2.06–1.94 (m, 2H).

¹³C NMR (125 MHz, CDCl₃) δ = 193.8, 165.4, 147.8, 147.1, 138.0, 134.5, 126.9, 111.0, 110.4, 67.2, 60.7, 60.6, 56.1, 55.8, 51.3, 48.5, 47.9, 37.1, 36.2, 25.2, 23.0.

IR (neat, cm⁻¹): 2934, 2849, 2797, 1668, 1603, 1516, 1451, 1334, 1241, 1211, 1145, 1119, 1062, 1051, 1018.

HRMS (FAB) calcd for C₂₁H₂₈NO₅ [M+H]⁺ 374.1967, found 374.1974.

[α]_D²⁵ -245 (c = 0.2, CHCl₃). Natural^[4]: [α]_D¹⁸ -400 (c = 0.8, CHCl₃). Synthetic (by Herzon)^[5]: [α]_D²⁵ -265 (c = 0.2, CHCl₃).

For (-)-runanine isomer **2'**:

Rf 0.15 (Hexane:EtOAc = 1:2), colorless oil.

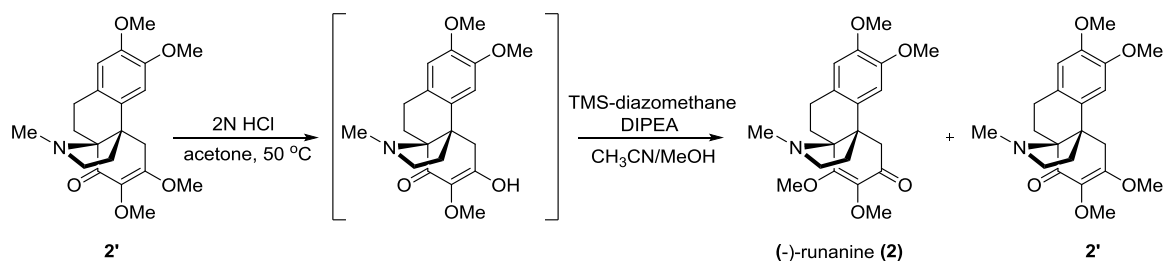
¹H NMR (600 MHz, CDCl₃) δ = 6.71 (s, 1H), 6.50 (s, 1H), 3.95 (s, 3H), 3.85 (s, 3H), 3.81 (s, 3H), 3.60 (s, 3H), 2.96 (d, *J* = 16.9 Hz, 1H), 2.81–2.79 (m, 2H), 2.74–2.68 (m, 1H), 2.70 (d, *J* = 17.5 Hz, 1H), 2.54–2.51 (m, 1H), 2.50 (s, 3H), 2.17 (t, *J* = 7.6 Hz, 2H), 1.98–1.60 (m, 1H), 1.90–1.85 (m, 1H).

¹³C NMR (150 MHz, CDCl₃) δ = 195.7, 160.1, 147.7, 147.3, 136.1, 134.2, 127.9, 111.4, 110.0, 68.2, 60.7, 58.3, 56.3, 55.8, 51.8, 47.8, 38.1, 37.7, 35.5, 25.4, 21.5.

IR (neat, cm⁻¹): 2930, 2851, 2832, 2793, 1657, 1611, 1513, 1450, 1353, 1337, 1252, 1210, 1144, 1124, 1014.

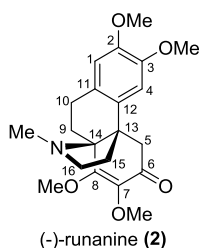
HRMS (FAB) calcd for C₂₁H₂₈NO₅ [M+H]⁺ 374.1967, found 374.1971.

[α]_D²⁰ -217.6 (c = 0.5, CHCl₃).



A solution of the (-)-runanine isomer **2'** (6.9 mg, 0.018 mmol) in 2N HCl (1 mL) and acetone (1 mL) was heated to 50 °C for 15 h. The mixture was cooled to room temperature, extracted with DCM/*i*PrOH (2:1, 15 mL) and concentrated. The residue was redissolved in MeOH and CH₃CN (1:3, 2 mL) and treated with DIPEA (9.5 μL, 0.054 mmol, 3.00 eq) and (trimethylsilyl)diazomethane (90 μL, 0.054 mmol, 3.00 eq, 0.6 M in hexane). The mixture was stirred at room temperature for 12 h and diluted with DCM (10 mL), and then, the organic phase was washed with water and concentrated. The residue was purified by flash column chromatography on silica gel (Hexane:EtOAc = 1:2 for **2'**; DCM:MeOH = 20:1 for **2**) to afford the (-)-runanine (**2**) and (-)-runanine isomer **2'** (5.5 mg, 0.014 mmol, 80%, **2**:**2'** = **2**:1).

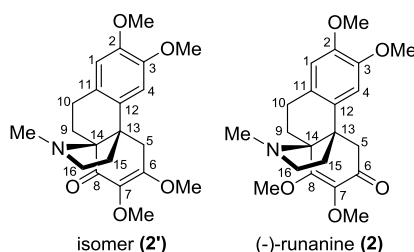
Table S1. Comparison of natural and synthetic (-)-runanine (**2**).



¹ H NMR				¹³ C NMR	
position	² Natural (300 MHz, CDCl ₃)	³ Herzon (500 MHz, CDCl ₃)	Synthetic (500 MHz, CDCl ₃)	³ Herzon (125MHz, CDCl ₃)	Synthetic (125 MHz, CDCl ₃)
1	6.47 (s, 1H)	6.47 (s, 1H)	6.47 (s, 1H)	111.0	111.0
2				147.1	147.1
3				147.8	147.8
4	6.64 (s, 1H)	6.67 (s, 1H)	6.64 (s, 1H)	110.4	110.4
5	3.00 (d, 13.2) 2.60 (d, 13.2)	3.00 (d, 16.0) 2.62 (d, 16.0)	3.00 (d, 15.9) 2.60 (d, 16.0)	48.5	48.5
6				193.8	193.8
7				138.0	138.0
8				165.4	165.4
9	2.8 (m)	2.22-2.15 (m) 1.99-1.93 (m)	2.18-2.13 (m) 2.00-1.94 (m)	23.0	23.0
10	2.8 (m)	2.80-2.71 (m) 2.57-2.52 (m)	2.77-2.71 (m) 2.57-2.51 (m)	25.2	25.2
11				126.9	126.9
12				134.5	134.5
13				47.9	47.9

14				67.2	67.2
15	2.0 (m)	2.22-2.15 (m) 2.08-2.02 (m)	2.18-2.13 (m) 2.06-2.00 (m)	37.1	37.1
16	2.0 (m)	2.86-2.82 (m) 2.80-2.71 (m)	2.83-2.79 (m) 2.77-2.71 (m)	51.2	51.3
C ₂ OCH ₃	3.79 (s)	3.82 (s)	3.79 (s)	55.8	55.8
C ₃ OCH ₃	3.80 (s)	3.84 (s)	3.81 (s)	56.1	56.1
C ₇ OCH ₃	3.61 (s)	3.65 (s)	3.62 (s)	60.7	60.7
C ₈ OCH ₃	4.05 (s)	4.09 (s)	4.06 (s)	60.6	60.6
NCH ₃	2.51 (s)	2.53 (s)	2.51 (s)	36.2	36.2

Table S2. Comparison of synthetic (–)-runanine (**2**) and its isomer **2'**.

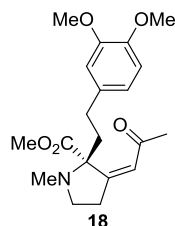


¹ H NMR			¹³ C NMR	
position	Isomer (2') (600 MHz, CDCl ₃)	Runanine (2) (500 MHz, CDCl ₃)	Isomer (2') (600 MHz, CDCl ₃)	Runanine (2) (125 MHz, CDCl ₃)
1	6.50 (s, 1H)	6.47 (s, 1H)	111.4	111.0
2			147.3	147.1
3			147.7	147.8
4	6.70 (s, 1H)	6.64 (s, 1H)	110.0	110.4
5	2.96 (d, 16.9) 2.70 (d, 16.9)	3.00 (d, 15.9) 2.60 (d, 16.0)	35.5	48.5
6			160.1	193.8
7			136.1	138.0
8			195.7	165.4
9	2.17 (t, 7.6, 2H)	2.18-2.13 (m) 2.00-1.94 (m)	21.5	23.0
10	2.74-2.70 (m) 2.54-2.51 (m)	2.77-2.71 (m) 2.57-2.51 (m)	25.4	25.2
11			127.9	126.9
12			134.2	134.5
13			47.8	47.9
14			68.2	67.2
15	1.98-1.96 (m) 1.90-1.85 (m)	2.18-2.13 (m) 2.06-2.00 (m)	38.1	37.1
16	2.81-2.79 (m, 2H)	2.83-2.79 (m) 2.77-2.71 (m)	51.8	51.3
C ₂ OCH ₃	3.81 (s)	3.79 (s)	55.8	55.8
C ₃ OCH ₃	3.85 (s)	3.81 (s)	56.3	56.1

C ₇ OCH ₃	3.60 (s)	3.62 (s)	60.7	60.7
C ₆ OCH ₃	3.95		58.3	
C ₈ OCH ₃		4.06 (s)		60.6
NCH ₃	2.50 (s)	2.51 (s)	37.7	36.2

4. Formal Synthesis of (–)-8-Demthoxyrunanine (**4**) and (–)-Cepharatine D (**21**)

Methyl (S,Z)-2-(3,4-dimethoxyphenethyl)-1-methyl-3-(2-oxopropylidene)pyrrolidine-2-carboxylate (**18**)



To a deoxygenated solution (3 × freeze – pump – thaw) of the vinylstannane compound **15** (76.1 mg, 0.12 mmol), PCy₃·HBF₄ (4.61 mg, 0.012 mmol, 0.10 eq), DIPEA (2.24 μL, 0.012 mmol, 0.10 eq), and Pd₂(dba)₃ (5.71 mg, 0.006 mmol, 0.05 eq) in toluene (5 mL), acetyl chloride (17.8 μL, 0.25 mmol, 2.00 eq) was added dropwise. The reaction mixture was heated at 50 °C for 12 h and then cooled and diluted with EtOAc (20 mL). The mixture was washed with 3% aq. NH₄OH (5 mL) and water and then concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (Hexane:EtOAc = 1:1), and compound **18** (31.7 mg, 0.087 mmol, 70%) was obtained as a yellow oil.

Rf 0.30 (Hexane:EtOAc = 1:1), yellow oil.

¹H NMR (500 MHz, CDCl₃) δ = 6.73–6.71 (m, 1H), 6.64–6.62 (m, 2H), 6.29 (s, 1H), 3.82 (s, 3H), 3.81 (s, 3H), 3.64 (s, 3H), 3.07–3.04 (m, 1H), 2.75–2.62 (m, 3H), 2.54–2.52 (m, 1H), 2.42 (td, *J* = 13.0, 5.1 Hz, 1H), 2.28 (s, 3H), 2.19 (td, *J* = 12.8, 5.1 Hz, 1H), 2.13 (s, 3H), 2.06 (td, *J* = 13.0, 3.7 Hz, 1H).

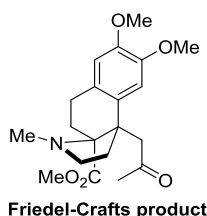
¹³C NMR (125 MHz, CDCl₃) δ = 196.7, 170.6, 163.0, 148.7, 147.0, 135.3, 121.3, 120.0, 111.7, 111.1, 73.7, 55.9, 55.8, 51.8, 50.9, 35.3, 35.2, 31.2, 31.1, 29.9.

IR (neat, cm⁻¹): 2946, 2834, 2788, 1734, 1691, 1626, 1513, 1226, 1155, 1028.

HRMS (FAB) calcd for C₂₀H₂₈NO₅ [M+H]⁺ 362.1967, found 362.1974.

[α]_D²⁰ -19.84 (c = 0.5, CHCl₃).

Methyl (3a*S*,9b*S*)-7,8-dimethoxy-3-methyl-9b-(2-oxopropyl)-1,2,3,4,5,9b-hexahydro-3a*H*-benzo[*e*]indole-3a-carboxylate (Friedel-Crafts product)



To a solution of compound **18** (147 mg, 0.41 mmol) in DCM (5 mL), TfOH (180 μL, 2.03 mmol, 5.00 eq) was added dropwise at 0 °C. After 10 min, the mixture was quenched with aq. NaHCO₃ (5 mL), and then extracted with DCM (20 mL). The organic layer was washed with brine (10 mL), dried over Na₂SO₄ and concentrated in vacuo. The residue was purified by flash column chromatography on silica gel (Hexane:EtOAc = 2:1), and **Friedel-Crafts product** (124 mg, 0.35 mmol, 85%) was obtained as a colorless oil.

Rf 0.50 (Hexane:EtOAc = 1:1), colorless oil.

¹H NMR (400 MHz, CDCl₃) δ = 6.80 (s, 1H), 6.49 (s, 1H), 3.81 (s, 3H), 3.79 (s, 3H), 3.71 (s, 3H), 2.93 (td, J = 7.8, 7.6 Hz, 1H), 2.85–2.74 (m, 2H), 2.72–2.65 (m, 1H), 2.58–2.51 (m, 3H), 2.37 (ddd, J = 12.4, 8.7, 3.3 Hz, 1H), 2.23 (s, 3H), 2.12–2.00 (m, 2H), 1.81 (s, 3H).

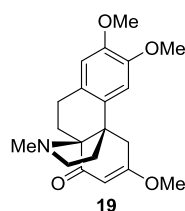
¹³C NMR (100 MHz, CDCl₃) δ = 208.3, 174.2, 147.1, 147.0, 132.7, 127.9, 111.0, 110.9, 73.5, 56.0, 55.6, 52.5, 51.3, 51.1, 50.5, 37.2, 34.8, 32.7, 24.8, 23.3.

IR (neat, cm⁻¹): 2947, 2838, 2789, 1716, 1514, 1449, 1356, 1232, 1204, 1134, 1078, 1014.

HRMS (FAB) calcd for C₂₀H₂₈NO₅ [M+H]⁺ 362.1967, found 362.1962.

[α]_D²⁰ +236.46 (c = 0.5, CHCl₃).

(4b*S*,8a*S*)-2,3,6-Trimethoxy-11-methyl-9,10-dihydro-8a,4b-(epiminoethano)phenanthren-8(5*H*)-one (19)



To a solution of the **Friedel-Crafts product** (39 mg, 0.11 mmol) in THF (3 mL), KO^tBu (0.16 mL, 0.16 mmol, 1.50 eq, 1.0 M in THF) was added dropwise at 0 °C. After 15 min, the mixture was quenched with sat. NH₄Cl (2 mL) and H₂O (2 mL), extracted with DCM/*i*PrOH (2:1, 30 mL) and concentrated in vacuo. The residue was redissolved in methanol (3 mL) and treated with TiCl₄ (8 drops, 1 M solution in DCM). The mixture was stirred at room temperature for 12 h. After quenching with sat. NaHCO₃, the reaction was extracted with DCM (2×10 mL). The residue was purified by flash column chromatography on silica gel (Hexane:EtOAc = 1:1) to afford compound **19** (30.0 mg, 0.09 mmol, 81%) as a colorless oil.

R_f 0.15 (Hexane:EtOAc = 1:1), colorless oil.

¹H NMR (600 MHz, CDCl₃) δ = 6.71 (s, 1H), 6.50 (s, 1H), 5.33 (s, 1H), 3.84 (s, 3H), 3.80 (s, 3H), 3.63 (s, 3H), 2.95 (d, J = 16.5 Hz, 1H), 2.89–2.87 (m, 1H), 2.78 (td, J = 9.5, 4.3 Hz, 1H), 2.74–2.69 (m, 1H), 2.69 (d, J = 17.4 Hz, 1H), 2.56–2.51 (m, 1H), 2.47 (s, 3H), 2.17–2.07 (m, 3H), 1.80 (ddd, J = 13.1, 7.6, 5.3 Hz, 1H).

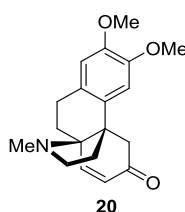
¹³C NMR (150 MHz, CDCl₃) δ = 199.7, 174.9, 147.7, 147.3, 134.1, 128.1, 111.4, 110.1, 101.9, 68.0, 56.3, 55.9, 55.8, 52.0, 49.3, 39.8, 38.3, 35.6, 25.8, 21.0.

IR (neat, cm⁻¹): 2934, 2834, 2793, 1643, 1614, 1513, 1445, 1376, 1253, 1201, 1134, 1014.

HRMS (FAB) calcd for C₂₀H₂₆NO₄ [M+H]⁺ 344.1862, found 344.1858.

[α]_D²⁰ -284.36 (c = 0.5, CHCl₃).

(4b*S*,8a*R*)-2,3-dimethoxy-11-methyl-9,10-dihydro-8a,4b-(epiminoethano)phenanthren-6(5*H*)-one (20)



To a solution of compound **19** (41.4 mg, 0.12 mmol) in DCM (3 mL), diisobutylaluminium hydride (0.30 mL, 0.30 mmol, 2.50 eq, 1 M solution in Hexane) was added dropwise at -78 °C. After 30 min, the mixture was quenched with brine (2 mL), extracted with DCM (2×10 mL) and concentrated in vacuo. The residue was redissolved in diethyl ether (5 mL) and treated with 2N HCl (1 mL) at 0 °C. After 10 min, the reaction was quenched with sat. NaHCO₃, and then extracted with EtOAc (2×10 mL). The residue was purified by flash column chromatography on silica gel (Hexane:EtOAc = 1:1) to afford compound **20** (21.7 mg, 0.07 mmol, 58%) as a colorless oil.

Rf 0.20 (Hexane:EtOAc = 1:1), colorless oil.

¹H NMR (500 MHz, CDCl₃) δ = 6.82 (d, J = 10.4 Hz, 1H), 6.66 (s, 1H), 6.51 (s, 1H), 6.12 (d, J = 10.4 Hz, 1H), 3.82 (s, 3H), 3.81 (s, 3H), 2.95–2.84 (m, 1H), 2.89 (d, J = 16.3 Hz, 2H), 2.57–2.53 (m, 1H), 2.54 (td, J = 16.4 Hz, 1H), 2.43 (s, 3H), 2.43–2.41 (m, 1H), 2.28–2.22 (m, 1H), 2.04–1.96 (m, 2H), 1.78–1.71 (m, 1H).

¹³C NMR (125 MHz, CDCl₃) δ = 198.8, 150.3, 147.9, 147.1, 135.6, 129.9, 126.3, 111.1, 110.5, 63.2, 56.1, 55.8, 51.6, 49.5, 48.3, 36.2, 33.3, 25.1, 24.6.

IR (neat, cm⁻¹): 2929, 2833, 2790, 1678, 1513, 1452, 1255, 1207, 1140, 1067.

HRMS (FAB) calcd for C₁₉H₂₄NO₃ [M+H]⁺ 314.1756, found 314.1757.

[α]_D²⁰ -413.96 (c = 0.5, CHCl₃).

All data obtained are in full agreement with those by Reisman.⁴

5. Computational Study

Calculation of the nucleophilicity of the oxygen

Theoretical background of Fukui function

Local softness is therefore one of the widely used local density functional descriptors to compare reactivity at different sites within one molecule. Local softness is given by

$$s(r) = Sf(r)$$

where S is global softness and $f(r)$ is the Fukui function.⁵ The global softness can be approximated as

$$S = 1 / (\text{IP} - \text{EA})$$

where IP and EA are the ionization potential and electron affinity respectively, of the chemical species.

The Fukui function describes the variation of the electronic density ($\rho(r)$) upon changing the number of electrons (N) in the system.⁶

$$f(r) = [\partial \rho(r) / \partial N]_{r(r)}$$

Thus, Fukui functions for nucleophilic and electrophilic attack on an atom, k , in an N electron system was introduced by Yang and Mortier⁷ as

$$f_k^+ = \rho_k(N+1) - \rho_k(N) \text{ (nucleophilic attack)}$$

$$f_k^- = \rho_k(N) - \rho_k(N-1) \text{ (electrophilic attack)}$$

where $\rho_k(N+1)$, $\rho_k(N)$ and $\rho_k(N-1)$ are the electronic populations on atom k in the $N+1$, N and $N-1$ electron systems. These functions can be condensed to the nuclei by using an atomic charge partitioning scheme, such as Mulliken⁸ population analysis. Therefore, the local softness for atom k can be written as

$$s_k^+ = [\rho_k(N+1) - \rho_k(N)]S$$

$$s_k^- = [\rho_k(N) - \rho_k(N-1)]S$$

Local softness is used as reactivity index, however not always provide the correct reactivity trends. So new parameters are devised by S. Pal,⁹ which are “relative electrophilicity” and “relative nucleophilicity”.

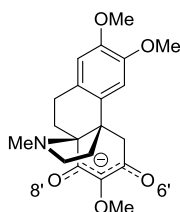
$$\text{Relative electrophilicity} = s_k^+ / s_k^-$$

$$\text{Relative nucleophilicity} = s_k^- / s_k^+$$

These two relative values indicate the preferable reactive sites in the reaction.

Geometry optimization and energy minimization of 1,3-diketone intermediate

The computational energy minimization of **1,3-diketone intermediate** was performed using the DMol3 program in Material Studio 2018. The Fukui function, ionization potential and electron affinity were also calculated by using DMol3 program.¹⁰ In these calculations, we employed generalized gradient approximation (GGA) in the Perdew-Burke-Ernzerhof (PBE)¹¹ form as well as a double numerical polarized (DNP) basis sets. To calculate the Fukui function values, a Mulliken partitioning scheme was employed.



1,3-diketone intermediate

Table S3. Cartesian coordinates of the **1,3-diketone intermediate**.**1,3-diketone intermediate**

Atom	X	Y	Z	Atom	X	Y	Z
C	-1.44658	-1.30985	-0.48253	C	1.433279	-0.91594	-0.08688
C	-1.0042	-0.09347	0.062277	C	0.773348	1.480819	-0.72642
C	-1.9735	0.852379	0.455541	H	0.422832	1.24684	-1.7429
C	-3.3415	0.61793	0.329819	H	0.208883	2.363804	-0.40501
C	-3.77677	-0.60179	-0.22485	C	2.895483	-0.49081	-0.45639
C	-2.82671	-1.53582	-0.61595	C	3.208537	0.867756	-0.7143
H	-1.63551	1.795087	0.870977	C	2.24346	1.905889	-0.81728
H	-3.18433	-2.47595	-1.03221	O	2.486854	3.115133	-1.03575
O	-5.12216	-0.91423	-0.33146	O	4.53024	1.196394	-1.01706
O	-4.32472	1.504309	0.713482	C	5.412405	1.149206	0.089702
C	-5.83486	-0.18775	-1.33159	H	6.401371	1.444736	-0.28246
H	-5.39688	-0.36365	-2.32476	H	5.100873	1.858464	0.873957
H	-6.86004	-0.56962	-1.31429	H	5.470789	0.136215	0.509364
H	-5.83917	0.886906	-1.11704	C	0.822484	0.631075	1.696426
C	-3.92309	2.733659	1.294743	H	1.504649	1.485261	1.736416
H	-3.31987	3.332633	0.599063	H	-0.07676	0.896576	2.260604
H	-4.84732	3.268172	1.527671	C	1.49789	-0.63133	2.268713
H	-3.35241	2.573667	2.21971	H	2.558551	-0.4278	2.514652
C	-0.47079	-2.38231	-0.91172	H	1.01583	-0.99868	3.18688
H	-0.86804	-2.91367	-1.78847	N	1.370415	-1.63114	1.214813
H	-0.38024	-3.12548	-0.10719	C	2.158334	-2.82871	1.451549
C	0.482507	0.285565	0.206253	H	2.058878	-3.52532	0.614476
C	0.908731	-1.80554	-1.225	H	3.233409	-2.6297	1.584708
H	1.636161	-2.6	-1.42176	H	1.76833	-3.32075	2.356874
H	0.861042	-1.19532	-2.13762	O	3.741705	-1.42287	-0.48647

Table S4. Molecular energy of **1,3-diketone intermediate**.

Compound	Hartree (Ha) ^a	Ionization potential (eV)	Electron affinity (eV)	Global softness (eV ⁻¹) ^b
1,3-diketone intermediate	-1206.446295	2.459881997	-3.544065212	-3.544065212

^a1Ha = 627.509391 kcal/mol. ^bGlobal softness = 1 / (Ionization potential – Electron affinity).**Table S5.** Molecular property of **1,3-diketone intermediate**.

Atom	Fukui function (–)	Fukui function (+)	Local softness (S–) ^a	Local softness (S+)	Relative nucleophilicity ^b
O-8'	0.151	0.026	0.02515	0.00433	5.81
O-6'	0.148	0.037	0.02465	0.00616	4.00

^aLocal softness = Global softness x Fukui function. ^bRelative nucleophilicity = Local softness (S–) / Local softness (S+).

6. Crystal Structure of 9a

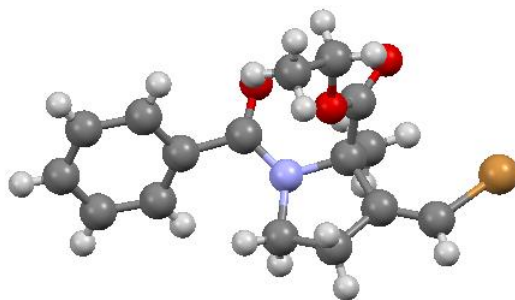


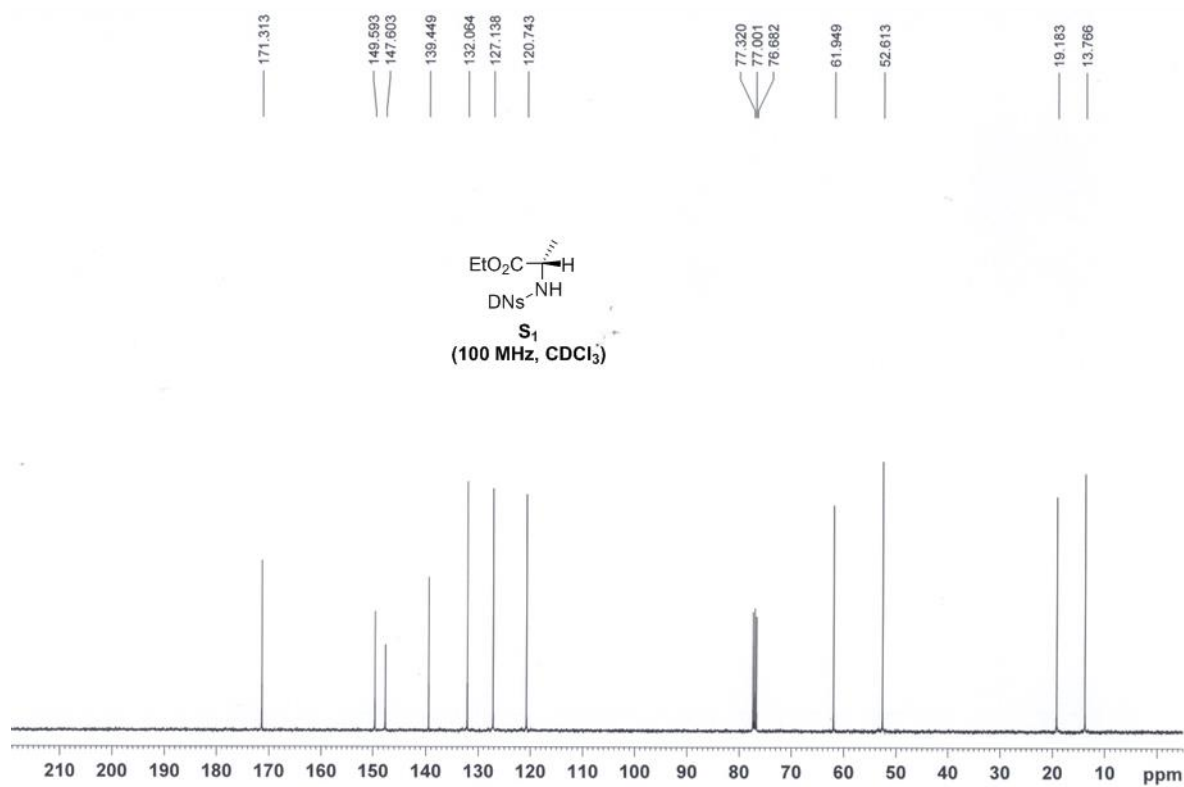
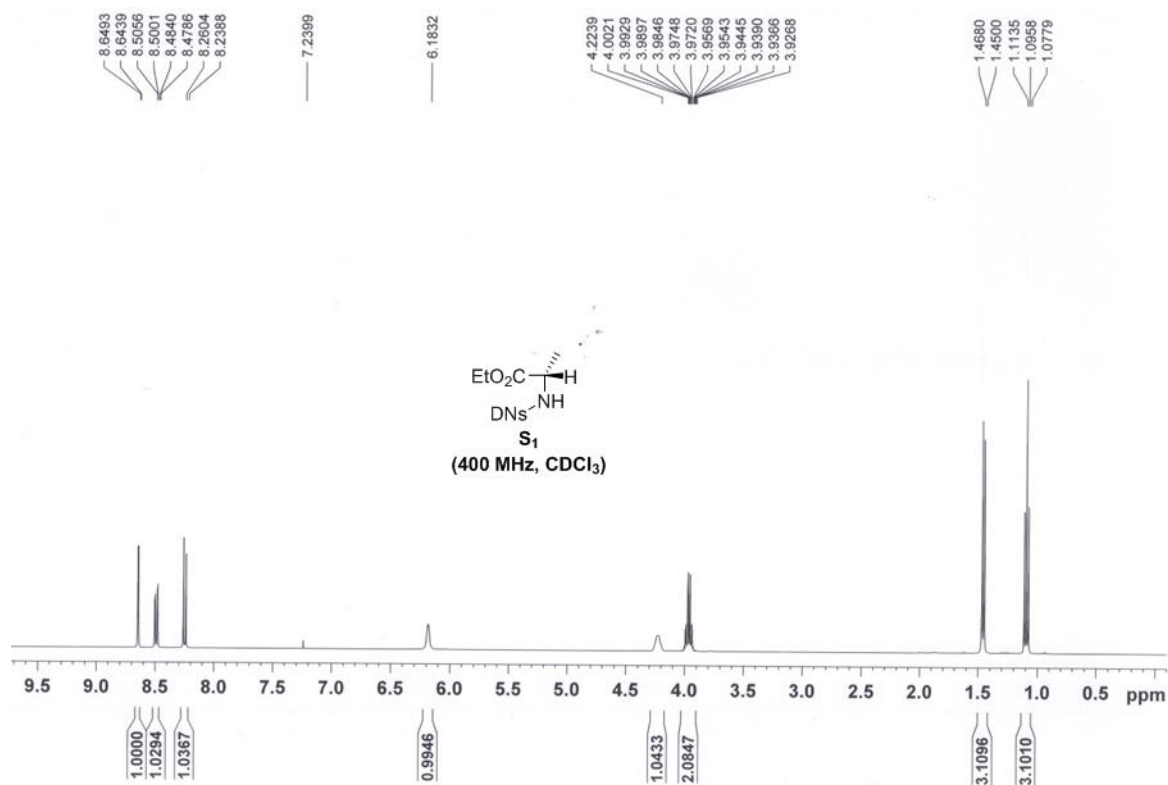
Table S6. Crystal data and structure refinement for **9a**.

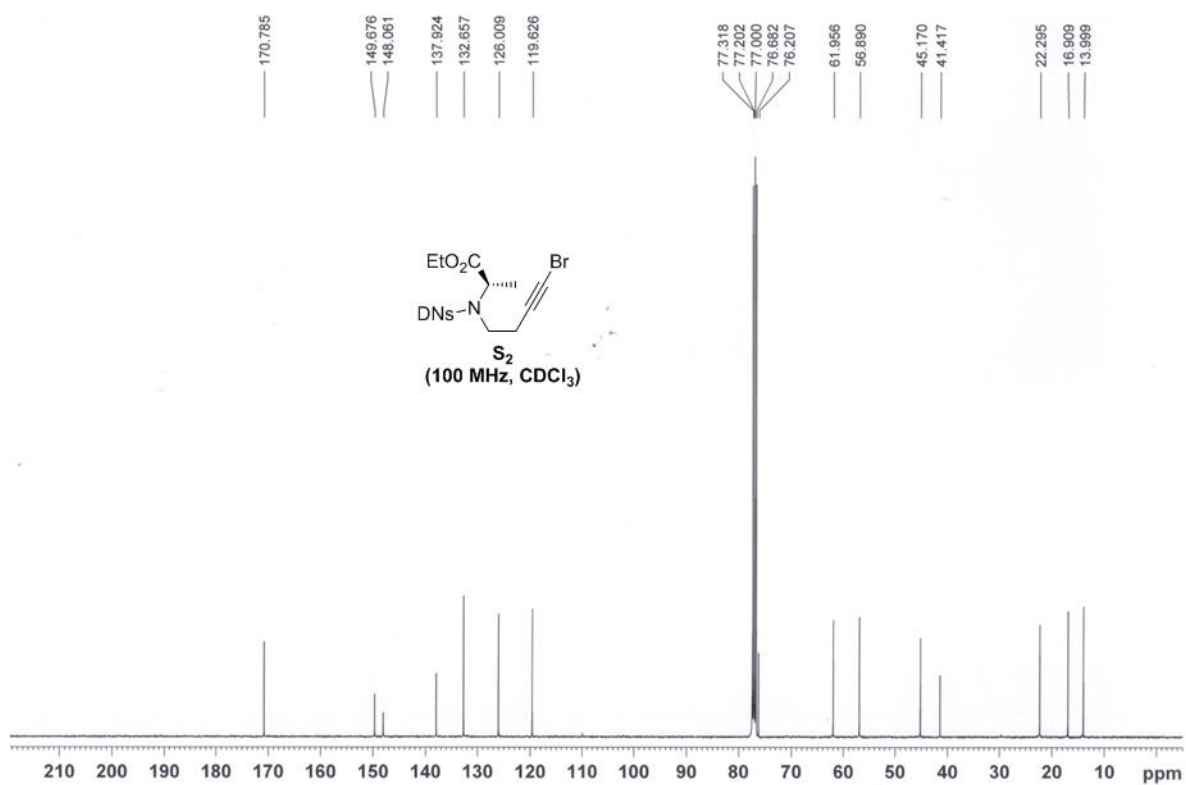
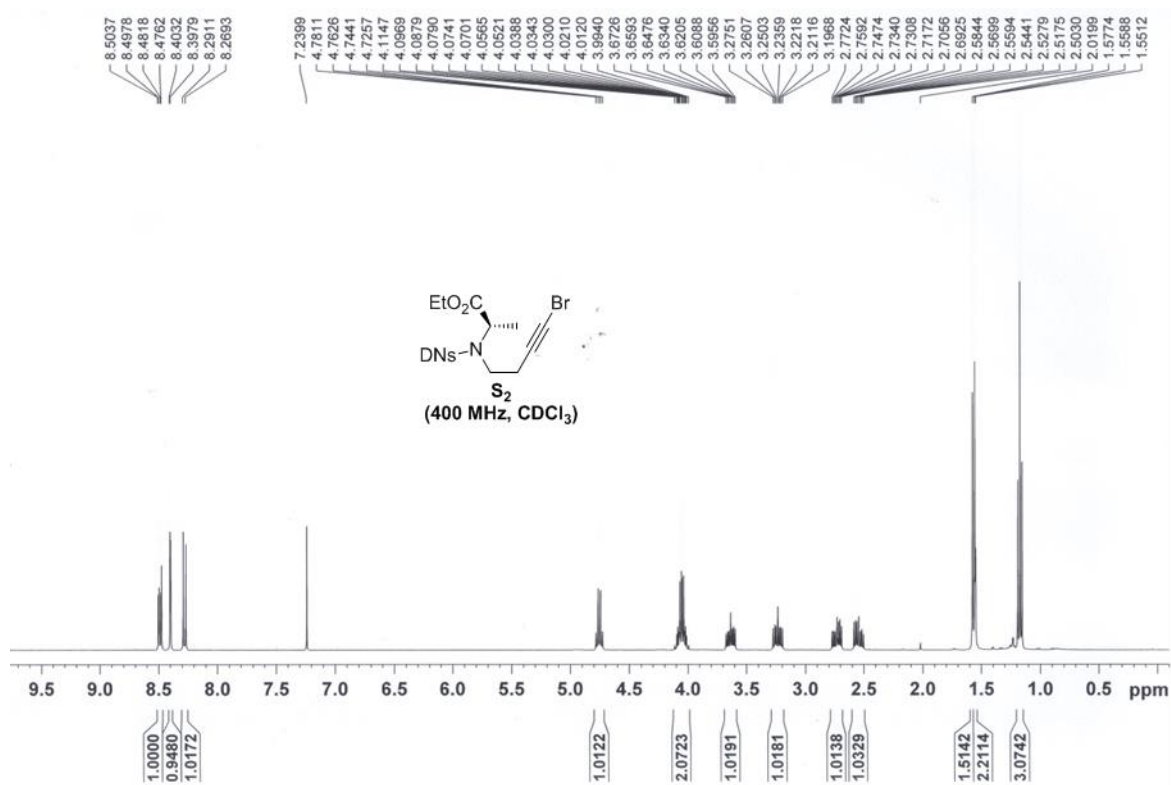
Identification code	9a
Empirical formula	C ₁₆ H ₁₈ BrO ₃ N
Formula weight	352.23
Temperature/K	293.37(10)
Crystal system	orthorhombic
Space group	P2 ₁ 2 ₁ 2 ₁
a/Å	7.27280(10)
b/Å	9.2844(2)
c/Å	24.0663(4)
α/°	90
β/°	90
γ/°	90
Volume/Å ³	1625.04(5)
Z	4
ρ _{calc} /g/cm ³	1.4396
μ/mm ⁻¹	3.528
F(000)	719.2
Crystal size/mm ³	0.518 × 0.261 × 0.193
Radiation	Cu Kα (λ = 1.54184)
2θ range for data collection/°	7.34 to 147.22
Index ranges	-9 ≤ h ≤ 8, -10 ≤ k ≤ 11, -17 ≤ l ≤ 29
Reflections collected	5835
Independent reflections	3190 [R _{int} = 0.0106, R _{sigma} = 0.0117]
Data/restraints/parameters	3190/0/192
Goodness-of-fit on F ²	1.048
Final R indexes [I ≥ 2σ (I)]	R ₁ = 0.0272, wR ₂ = 0.0715
Final R indexes [all data]	R ₁ = 0.0276, wR ₂ = 0.0720
Largest diff. peak/hole / e Å ⁻³	0.17/-0.45
Flack parameter	0.023(18)

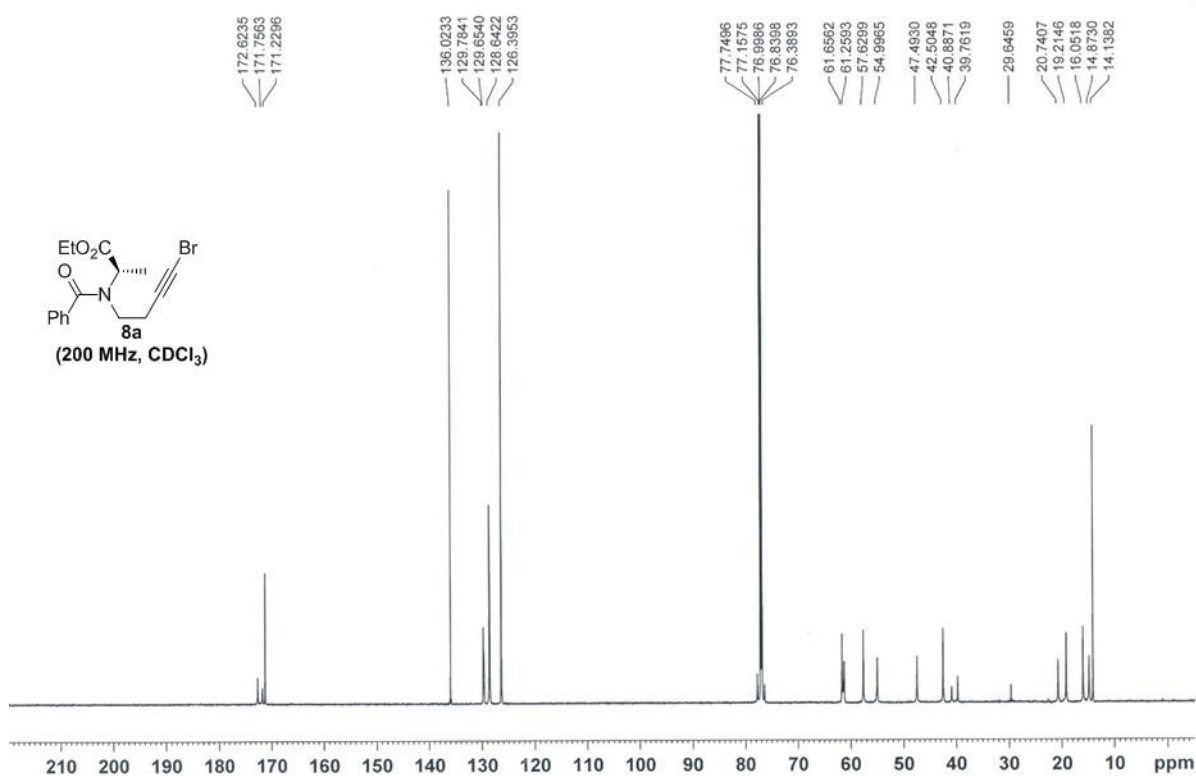
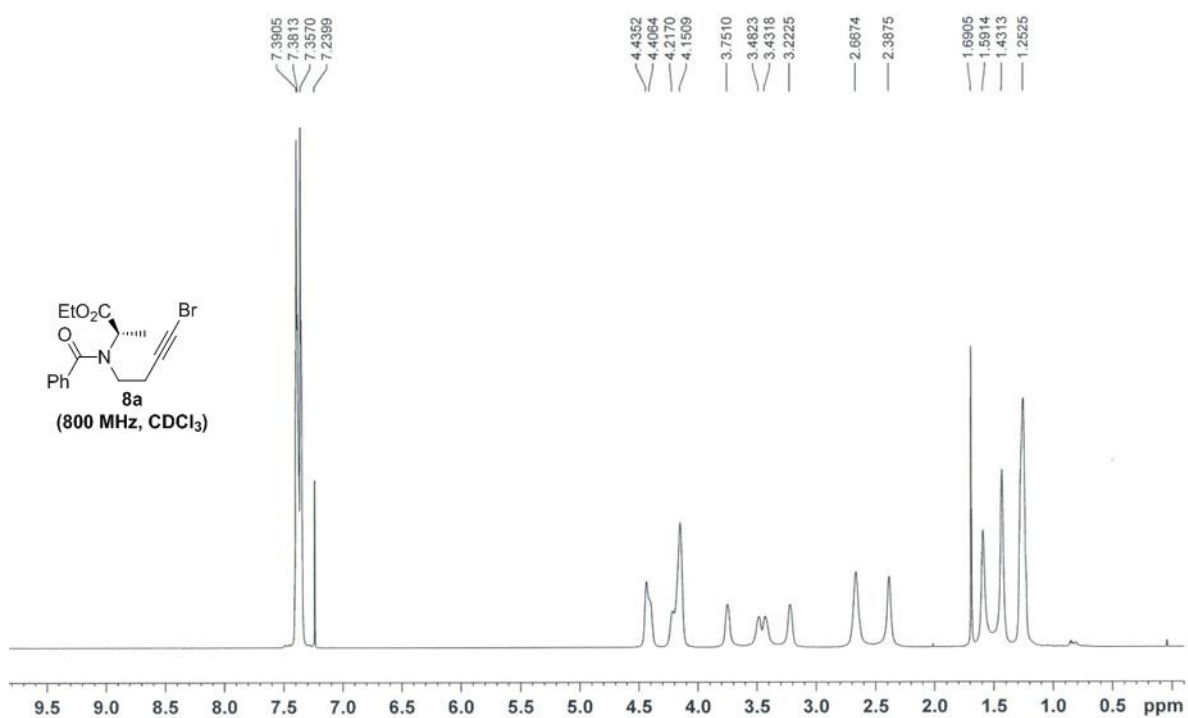
7. Reference

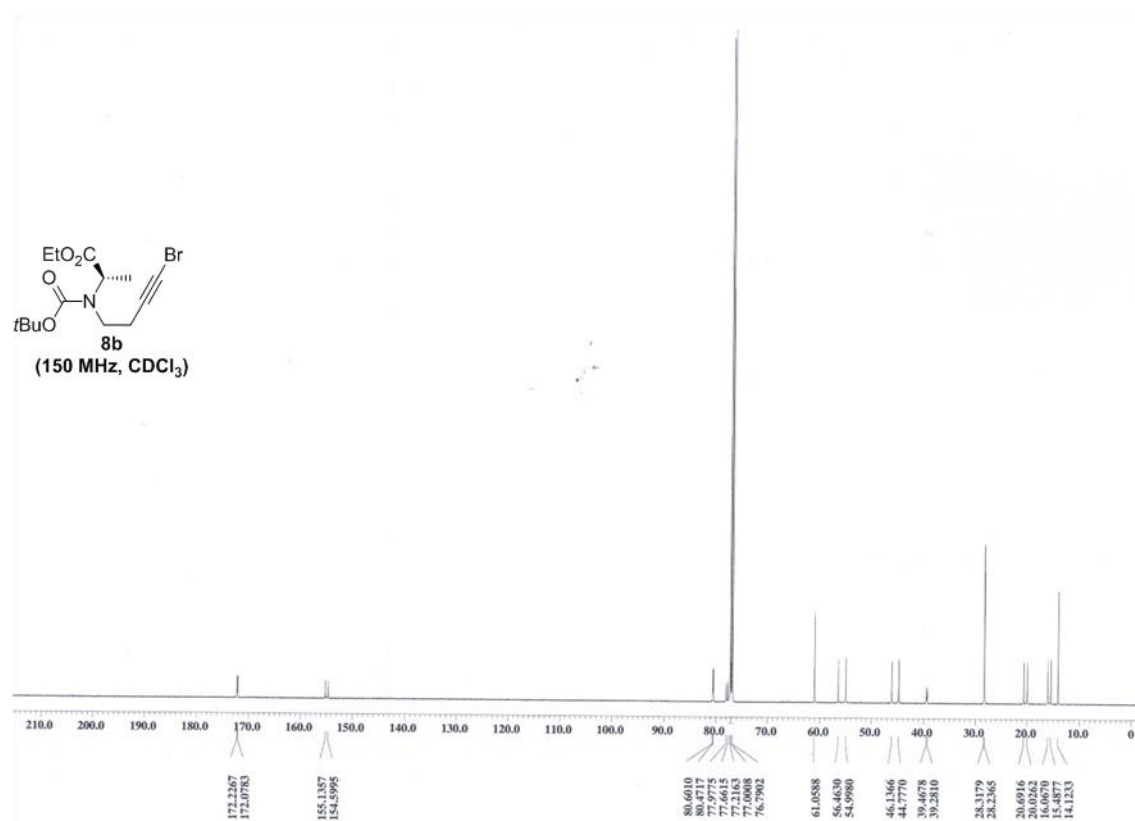
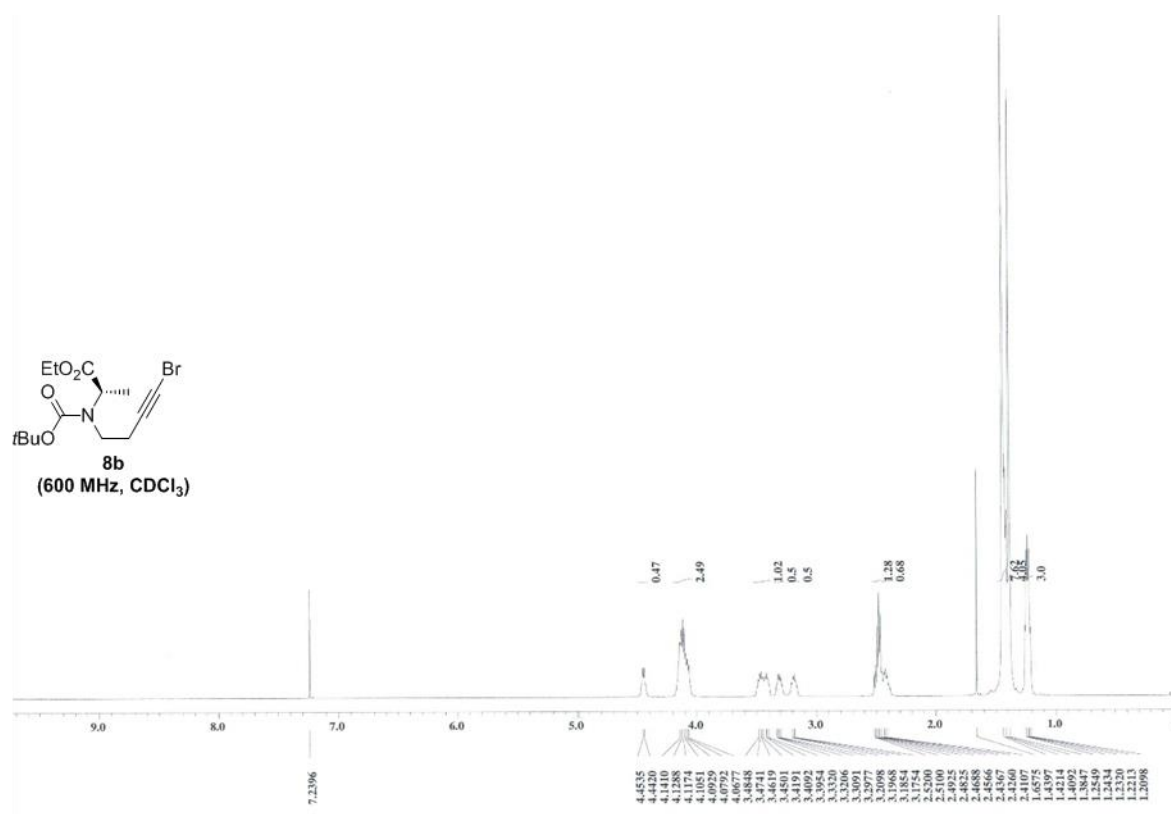
1. Cherney, A. H.; Reisman, S. E. *J. Am. Chem. Soc.* **2014**, *136*, 14365.
2. Zhi-Da, M.; Ge, L.; Guang-Xi, X.; Iinuma, M.; Tanaka, T.; Mizuno, M. *Phytochemistry* **1985**, *24*, 3084.
3. Herzon, S. B.; Calandra, N. A.; King, S. M. *Angew. Chem. Int. Ed.* **2011**, *50*, 8863.
4. Chuang, K. V.; Navarro, R.; Reisman, S. E. *Angew. Chem. Int. Ed.* **2011**, *50*, 9447.
5. Yang, W.; Parr, R. G. *Proc. Natl. Acad. Sci. U.S.A.* **1985**, *82*, 6723.
6. Parr, R. G.; Yang, W. *J. Am. Chem. Soc.* **1984**, *106*, 4049.
7. Yang, W.; Mortier, W. J. *J. Am. Chem. Soc.* **1986**, *108*, 5708.
8. Mulliken, R. S. *J. Chem. Phys.* **1955**, *23*, 1833.
9. Roy, R. K.; Krishnamurti, S.; Geerlings, P.; Pal, S. *J. Phys. Chem. A* **1998**, *102*, 3746.
10. (a) Delley, B. *J. Chem. Phys.* **1990**, *92*, 508. (b) Delley, B. *J. Chem. Phys.* **2000**, *113*, 7756.
11. Perdew, J. P.; Burke, K.; Ernzerhof, M. *Phys. Rev. Lett.* **1996**, *77*, 3865.

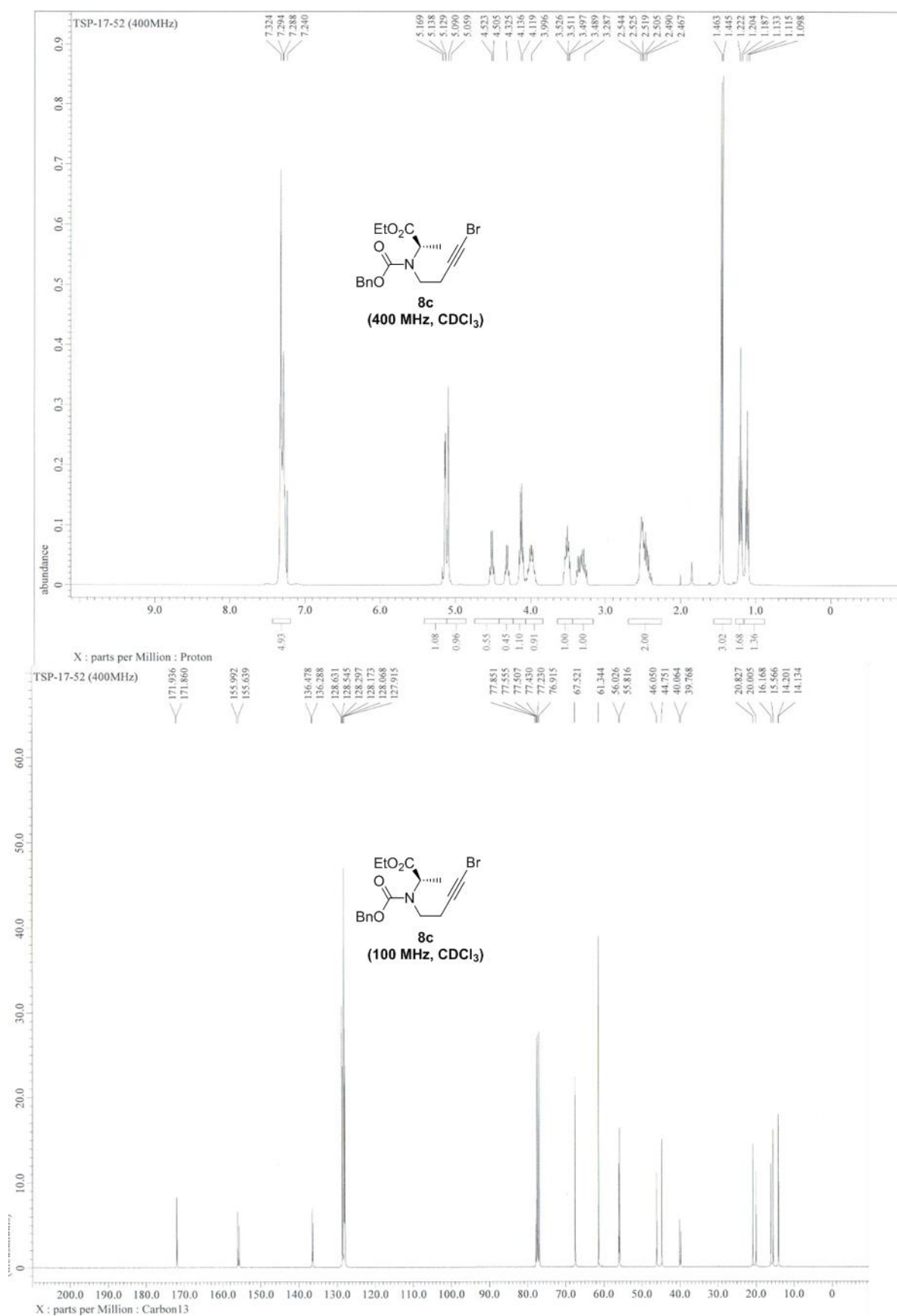
8. ^1H NMR and ^{13}C NMR Spectra

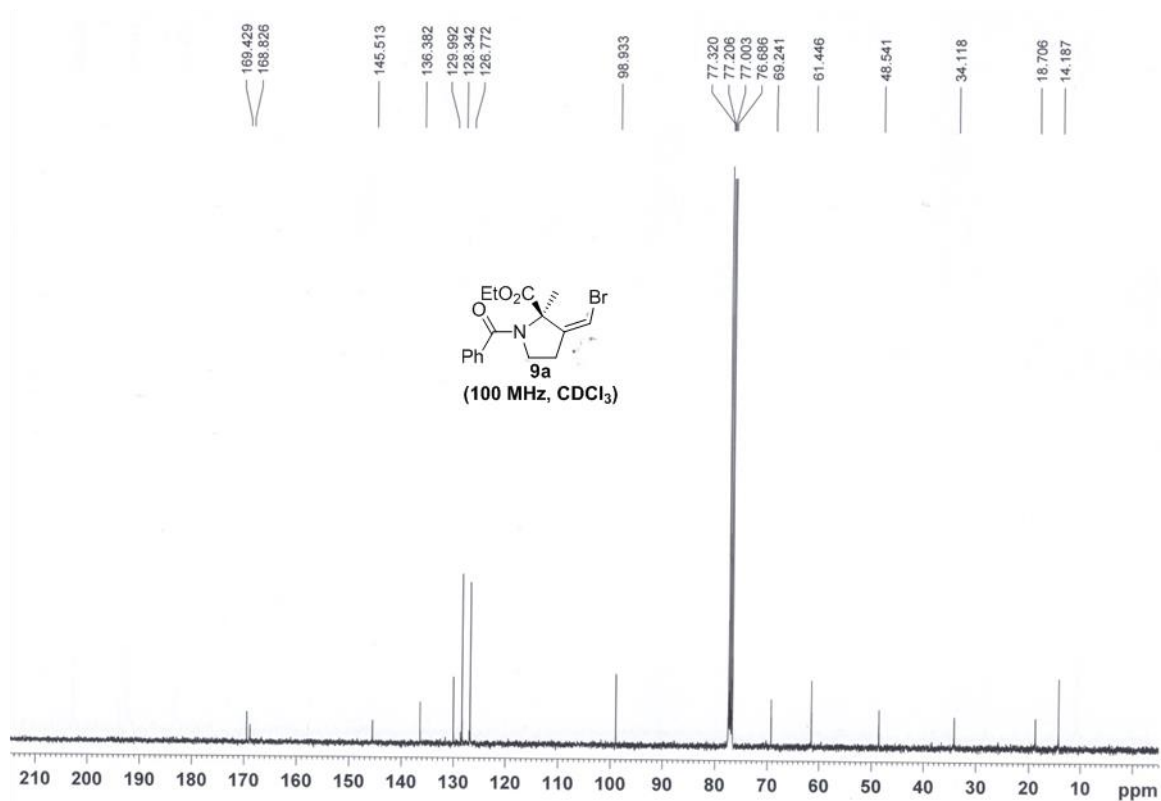
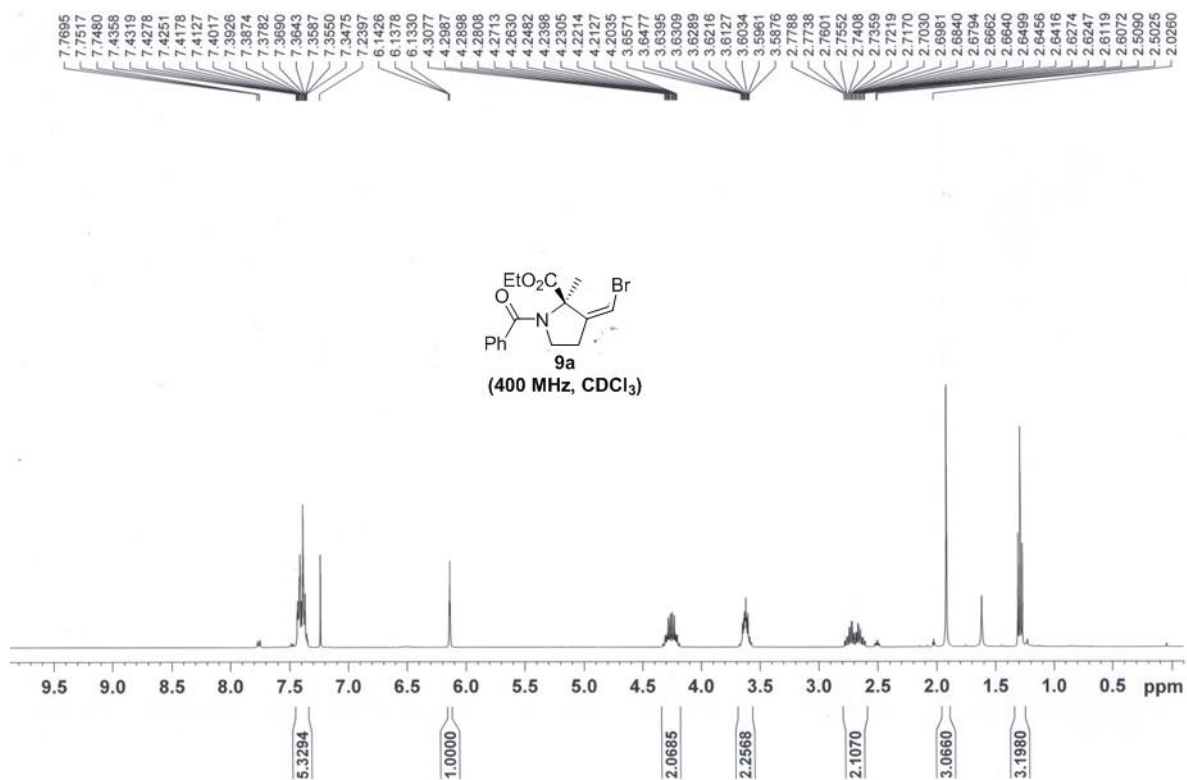


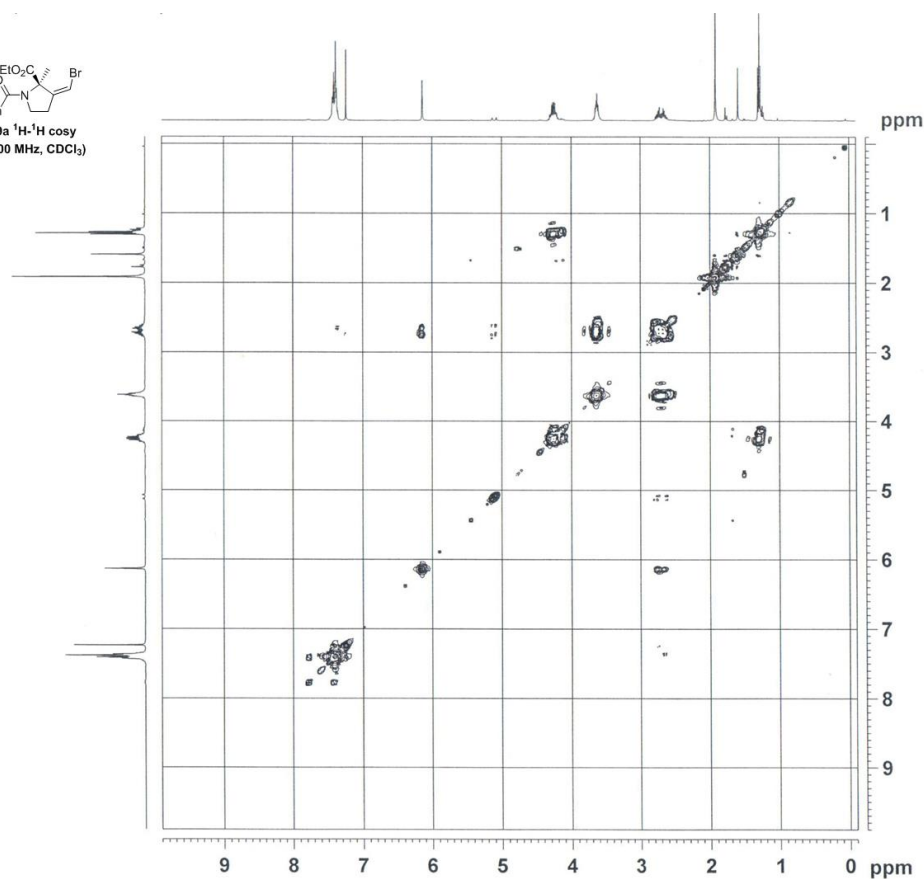
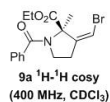












Current Data Parameters
NAME 15-115
EXPNO 11
PROCNO 1

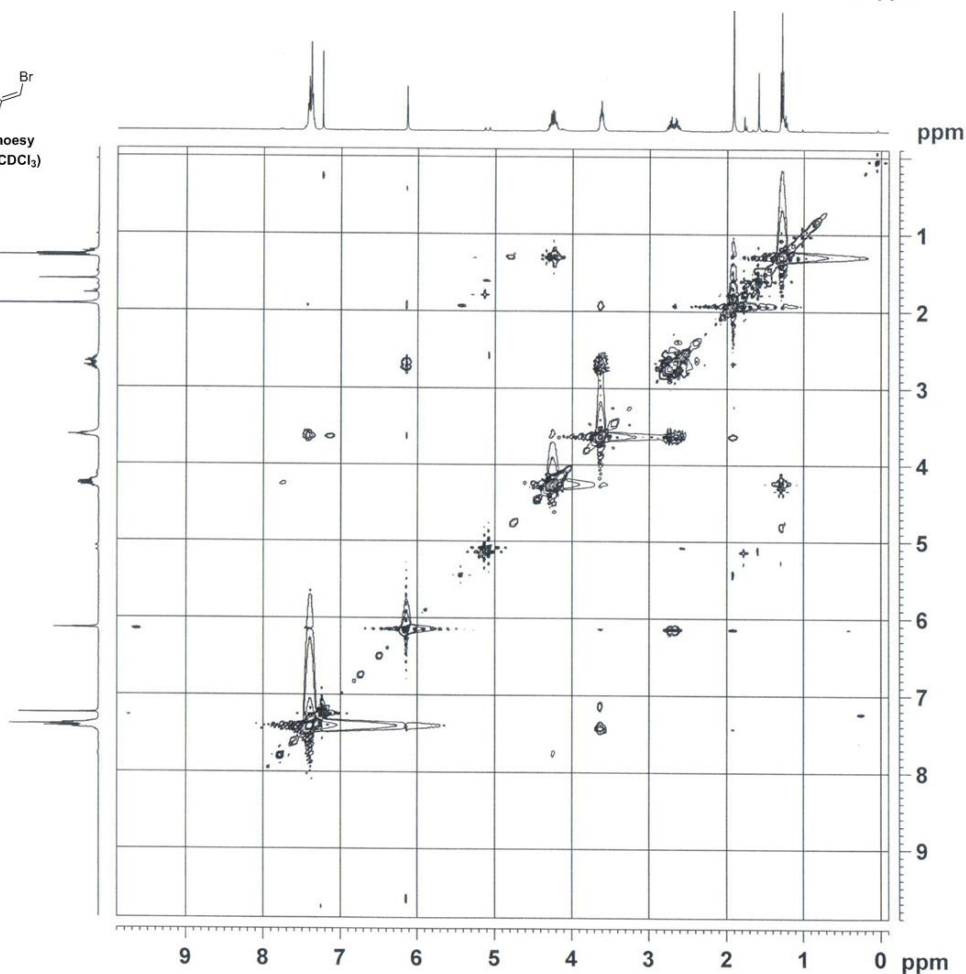
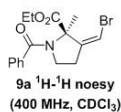
F2 - Acquisition Parameters
Date_ 20180412
Time_ 11.14
INSTRUM spect
PROBHD 5 mm Multinucl
PULPROG cosy445
TD 2048
SOLVENT CDCl_3
NS 12
DS 16
SWH 5592.841 Hz
FIDRES 2.730880 Hz
AQ 0.1831412 sec
RG 256
DW 89.400 usec
DE 6.00 usec
TE 298.0 K
D0 0.00000300 sec
D1 2.00000000 sec
IN0 0.00017880 sec
MCREST 0.00000000 sec
MCWRK 2.00000000 sec

===== CHANNEL f1 =====
NUC1 ^1H
P1 11.90 usec
PL1 5.50 dB
SFO1 400.1320191 MHz

F1 - Acquisition parameters
ND0 1
TD 256
SFO1 400.132 MHz
FIDRES 21.847036 Hz
SW 13.977 ppm
FMODE QF

F2 - Processing parameters
SI 1024
SF 400.1300162 MHz
WDW SINE
SSB 0
LB 0.00 Hz
GB 0
PC 1.00

F1 - Processing parameters
SI 1024
MC2 QF
SF 400.1300161 MHz
WDW SINE
SSB 0
LB 0.00 Hz
GB 0



Current Data Parameters
NAME 15-115
EXPNO 14
PROCNO 1

F2 - Acquisition Parameters
Date_ 20180412
Time_ 13.09
INSTRUM spect
PROBHD 5 mm Multinucl
PULPROG noesyph
TD 2048
SOLVENT CDCl_3
NS 16
DS 16
SWH 5592.841 Hz
FIDRES 2.730880 Hz
AQ 0.1831412 sec
RG 228.1
DW 89.400 usec
DE 6.00 usec
TE 298.0 K
D0 0.00007425 sec
D1 2.00000000 sec
D8 1.00000000 sec
IN0 0.00017880 sec
MCREST 0.00000000 sec
MCWRK 1.00000000 sec
STICNT 128

===== CHANNEL f1 =====
NUC1 ^1H
P1 11.90 usec
PL1 5.50 dB
SFO1 400.1320191 MHz

F1 - Acquisition parameters
ND0 1
TD 256
SFO1 400.132 MHz
FIDRES 21.847036 Hz
SW 13.977 ppm
FMODE States-TPPI

F2 - Processing parameters
SI 2048
SF 400.1300175 MHz
WDW SINE
SSB 2
LB 0.00 Hz
GB 0
PC 1.00

F1 - Processing parameters
SI 1024
MC2 States-TPPI
SF 400.1300147 MHz
WDW SINE
SSB 2
LB 0.00 Hz
GB 0

