Supporting information

Methods

Effect of simulated intestinal media on the detection of H₂O₂ by the AAP-DCHBS-HRP method

Solutions of H₂O₂ at final concentrations up to 15 µM were prepared in different media by diluting

a commercial solution of H₂O₂. AAP (0.1 mM final concentration), DCHBS (1 mM final

concentration), HRP (3.6 units/mL, final concentration) were then added and incubated for 15 min

at 25 °C. For each sample, absorbances at 515 nm and at 750 nm were read. The differences were

calculated and subtracted by the value calculated in the sample not containing H₂O₂.

Production of H₂O₂ by DAO incubated with pancreatin

DAO (0.15 mg of solid/mL) was incubated for 24 h at 37 °C in SIF pH 6.8 containing 1% pancreatin

Aliquots were withdrawn, supplemented or not with 2000 units/mL bovine liver catalase and

incubated for 15 min at 37 °C. Samples were then diluted to a final concentration of 0.6 x10⁻³ mg

of solid/mL into a solution containing AAP, DCHBS, HRP at the concentrations indicated above and

incubated for 15 min at 25 °C. Absorbance at 515 nm of the adduct generated from the reaction of

DCHBS with AAP (oxidized by H₂O₂ in the presence of HRP) was read, and after subtraction of the

background absorbance (at 750 nm), it was corrected with the value obtained in the absence of

DAO; the content of H_2O_2 was calculated using 2.6×10^4 M⁻¹cm⁻¹ as the extinction coefficient.

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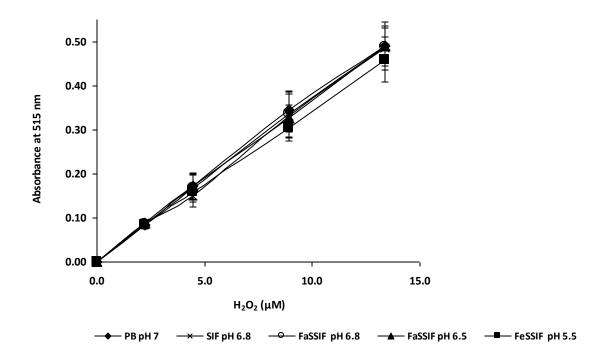
Figure captions

Figure S1: Effect of simulated intestinal media on the detection of H_2O_2 by the AAP-DCHBS-HPR method. Solutions of H_2O_2 were prepared at the indicated final concentrations in PB at pH 7 (\blacklozenge - \blacklozenge), SIF at pH 6.8 (x-x), FaSSIF at pH 6.8 (\blacklozenge - \blacklozenge), FaSSIF at pH 6.5 (\blacktriangle - \spadesuit), FeSSIF at pH 5.5 (\blacksquare - \blacksquare). AAP, DCHBS, HRP were then added and absorbance recorded and corrected as reported in Material and Methods (means +/- SD, n=3 different experiments).

Figure S2: Michaelis-Menten kinetics for the oxidation of benzylamine by DAO in different simulated intestinal media. DAO (0.18 mg of solid/mL final concentration) was added to PB at pH 7 (♦-♦), SIF at pH 6.8 (x-x), FAS SIF at pH 6.8 (•-•), FAS SIF at pH 6.5 (▲-▲), FES SIF at pH 5.5 (■-■) at 25 °C in the presence of benzylamine and the formation of benzaldehyde measured (means +/-SD, n=3 different experiments).

Figure S3: Effect of CA on DAO activity in the presence of proteases. DAO (0.15 mg of solid/mL final concentration) was incubated at 37 °C in SIF at pH 6.8 alone (control); in the presence of 3 mM CA; in the presence of 1% pancreatin; in the presence of 3 mM CA and 1% pancreatin; in the presence of 1% trypsin; in the presence of 3 mM CA and 1% trypsin (from the darkest to lightest shade). At the indicated times, aliquots were withdrawn, diluted in the incubation buffer and the activity measured by the AAP-DCHBS-HRP method. Reported values of DAO activity refer to the enzymatic activity in the incubation media (means +/- SD, n=3 different experiments).

Figure S4: Production of H₂O₂ during the incubation of pancreatin with DAO. DAO (0.15 mg of solid/mL final concentration) was incubated at 37 °C in SIF pH 6.8 containing 1% pancreatin. After 24 h of the incubation samples were diluted in SIF at pH 6.8 to a final concentration of 0.6x10⁻³ mg of solid/mL, then added with AAP, DCHBS, HRP and H₂O₂ concentration measured. Samples not incubated with bovine liver catalase (dark shade); samples incubated for 15 min with bovine liver catalase (2000 units/mL) (light shade) before dilution and addition of AAP, DCHBS and HRP. Reported values refer to DAO activity in the incubation media (means +/- SD, n=3 different experiments).



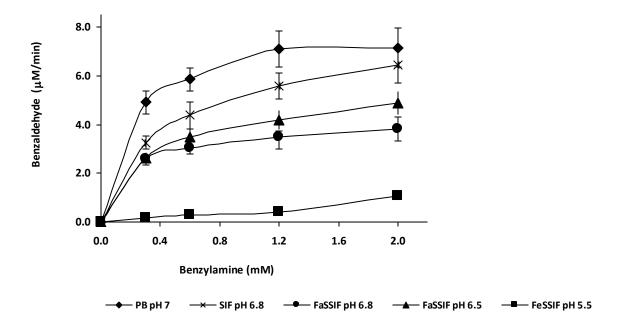
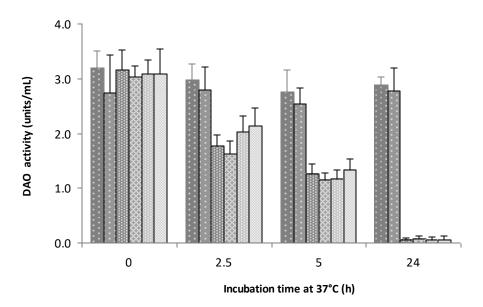


Fig. S2



■ Control ■ CA 3mM ■ 1% pancreatin ■ CA + 1% pancreatin ■ 1% trypsin ■ CA + 1% trypsin

